

# DEVELOPMENT AND VALIDATION OF Sawit, AN OIL PALM GROWTH AND YIELD MODEL

By

**CHEAH SEE SIANG** 

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

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Chair Faculty : Christopher Teh Boon Sung, PhD : Agriculture

A study was undertaken to develop and validate Sawit, an oil palm growth and yield model that considers weather variables, planting densities and soil textures. Sawit consists of five components: meteorology, photosynthesis, energy balance, water balance, and crop growth. The meteorology component was parameterized using daily weather data recorded in oil palm plantations. The photosynthesis component was parameterized through leaf measurement on young to mature palms, where photosynthetic parameters were measured. Measurements of trunk and canopy heights, as well as the relationship between oil palm leaf stomatal conductance with photosynthetically active radiation and air vapour pressure deficit, were taken to parameterize the energy balance component using the Shuttleworth-Wallace model. These measurements were used to model evapotranspiration and leaf temperature. Water flow in the soil was modelled following the 'tipping bucket' system using Darcy's law. The crop growth component was parameterized based on the OPSIM model. Different ages of oil palms were destructively sampled to determine the dry matter partitioning and respiration coefficients. Sawit was validated by oil palm planting-density experiments in six different sites: Rengam, Merlimau, Kerayong, Sungai Buloh, Sabrang, and Seri Intan. The model showed good agreement in predicting oil palm growth and yield parameters. However, the accuracy of the simulation varied considerably between sites and parameters. The yield simulation was considered sufficiently accurate for Merlimau and Sungai Buloh, with the refined index of agreement  $(d_r)$  equal to 0.76 and 0.74, respectively, and the normalized mean absolute error (NMAE) equal to 0.19 and 0.20, respectively. However, the yield was simulated as less than satisfactory for Rengam, Kerayong, Sabrang and Seri Intan. Their  $d_r$  values were small (0.52-0.56), but NMAE (0.13-0.25) were comparable to Merlimau and Sungai Buloh. The simulation of vegetative dry matter production was good for Rengam ( $d_r =$ 0.78, NMAE = 0.20), Kerayong ( $d_r$  = 0.71, NMAE = 0.14) and Sabrang ( $d_r$  = 0.76, NMAE = 0.11), but satisfactory for Merlimau ( $d_r = 0.62$ , NMAE = 0.23) and Sungai Buloh ( $d_r = 0.64$ , NMAE = 0.21). Total dry matter production was simulated sufficiently accurate with  $d_r$  ranging from 0.71-0.78 and NMAE ranging from 0.07-0.17 across all sites and planting densities. Rachis ( $d_r = 0.68$ -0.86, NMAE = 0.11-0.19), fronds ( $d_r$  = 0.65-0.83, NMAE = 0.11-0.20) and trunk ( $d_r$ = 0.69-0.82, NMAE = 0.24-0.39) were simulated more accurately than pinnae ( $d_r$ = 0.43-0.73, NMAE = 0.18-0.30) across all sites and planting densities except for the simulation of trunk biomass in Sungai Buloh ( $d_r = 0.38$ , NMAE = 0.77). The leaf area index was simulated sufficiently accurate for Merlimau, Rengam, and Seri Intan, with  $d_r$  ranging from 0.78-0.84 and NMAE ranging from 0.03-0.16. In contrast, the leaf area index was simulated as less than satisfactory for Kerayong, Sungai Buloh, and Sabrang, with  $d_r$  ranging from 0.57-0.62 and NMAE ranging from 0.13-0.17. The simulation of trunk height was especially good for Merlimau, Kerayong, Seri Intan and Sabrang, with large  $d_r$  values (0.87-0.93) and small NMAE values (0.06-0.12). However, trunk height was simulated satisfactory for Rengam ( $d_r = 0.67$ , NMAE = 0.39) and Sungai Buloh ( $d_r = 0.69$ , NMAE = 0.32). In addition, Sawit effectively simulated the impacts of El Niño event on oil palm yield. It also accounted for the influence of soil textures, rainfall, planting densities, and meteorological factors on water deficits. However, the simulation errors increased with increasing planting density due to insufficient characterization of microclimate conditions and plant water uptake under dense oil palm canopies, and higher variability of measurements for higher planting densities. In conclusion, an oil palm model called Sawit was developed and has been parameterized to simulate the growth and yield of oil palms under the influence of weather conditions, planting densities and soil textures. Improving the representation of oil palm microclimate and plant water uptake under dense canopies, incorporating fruiting activity, and refining the trunk's dry matter partitioning mechanism could enhance Sawit's accuracy.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

## PEMBANGUNAN DAN PENGESAHAN Sawit, MODEL PERTUMBUHAN DAN HASIL KELAPA SAWIT

Oleh

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Satu kajian telah dijalankan untuk membangunkan dan mengesahkan Sawit, sebuah model pertumbuhan dan hasil kelapa sawit yang mengambil kira pembolehubah cuaca, kepadatan penanaman, dan tekstur tanah. Pembangunan model ini terdiri daripada lima komponen: kesan meteorologi, kadar fotosintesis, kadar keseimbangan tenaga, kadar keseimbangan air, dan kadar pertumbuhan tanaman. Kesan meteorologi berparameterkan data cuaca harian yang direkodkan di ladang kelapa sawit. Kadar fotosintesis diukur pada daun muda dan matang di pokok sawit. Kadar keseimbangan tenaga dikira dengan menggunakan model Shuttleworth-Wallace, yang mana ketinggian batang serta kanopi sawit dan hubungan antara konduktiviti stomata daun yang aktif dalam julat spektrum cahaya untuk fotosintesis (photosynthetically active radiation) dengan defisit air akibat pengewapan diambilkira untuk menilai kadar evapotranspirasi dan suhu daun. Pergerakan air dalam tanah dinilai menggunakan sistem 'tipping bucket' yang berasaskan Darcy law. Kadar pertumbuhan tanaman diukur dengan menggunakan model OPSIM. Pokok sawit yang berlainan umur telah disampel secara destruktif untuk penentuan berat bahagian-bahagian pokok dan pekali kadar respirasi. Sawit telah disahkan oleh eksperimen kepadatan penanaman kelapa sawit di enam lokasi berbeza: Rengam, Merlimau, Kerayong, Sungai Buloh, Sabrang dan Seri Intan. Sawit menunjukkan persetujuan yang baik dalam meramalkan parameter pertumbuhan dan hasil kelapa sawit. Walau bagaimanapun, ketepatan simulasi berbeza dengan ketara antara lokasi dan parameter. Hasil telah disimulasikan dengan cukup tepat untuk Merlimau dan Sungai Buloh, dengan indeks persetujuan yang ditapis  $(d_r)$  bersamaan dengan 0.76 dan 0.74, masing-masing, dan ralat min mutlak ternormal (NMAE) bersamaan dengan 0.19 dan 0.20, masing-masing. Bagaimanapun, hasil disimulasikan sebagai kurang

memuaskan bagi Rengam, Kerayong, Sabrang dan Seri Intan. Nilai d<sub>r</sub> mereka adalah kecil (0.52-0.56), tetapi NMAE (0.13-0.25) adalah setanding dengan Merlimau dan Sungai Buloh. Simulasi pengeluaran bahan kering vegetatif adalah baik untuk Rengam ( $d_r = 0.78$ , NMAE = 0.20), Kerayong ( $d_r = 0.71$ , NMAE = 0.14) dan Sabrang ( $d_r$  = 0.76, NMAE = 0.11), tetapi memuaskan untuk Merlimau ( $d_r$  = 0.62, NMAE = 0.23) dan Sungai Buloh ( $d_r$  = 0.64, NMAE = 0.21). Jumlah pengeluaran bahan kering telah disimulasikan dengan cukup tepat dengan *d<sub>r</sub>* antara 0.71-0.78 dan NMAE antara 0.07-0.17 merentasi semua lokasi dan kepadatan penanaman. Rachis ( $d_r$  = 0.68-0.86, NMAE = 0.11-0.19), pelepah  $(d_r = 0.65 - 0.83, \text{NMAE} = 0.11 - 0.20)$  dan batang  $(d_r = 0.69 - 0.82, \text{NMAE} = 0.24 - 0.39)$ lebih tepat daripada yang disimulasikan ( $d_r = 0.43-0.73$ , NMAE = 0.18-0.30) merentasi semua lokasi dan kepadatan tanaman kecuali simulasi biojisim batang di Sungai Buloh ( $d_r = 0.38$ , NMAE = 0.77). Indeks luas daun telah disimulasikan dengan cukup tepat untuk Merlimau, Rengam, dan Seri Intan, dengan  $d_r$  antara 0.78-0.84 dan NMAE antara 0.03-0.16. Sebaliknya, indeks luas daun disimulasikan sebagai kurang memuaskan untuk Kerayong, Sungai Buloh, dan Sabrang, dengan  $d_r$  antara 0.57-0.62 dan NMAE antara 0.13-0.17. Simulasi ketinggian batang sangat baik untuk Merlimau, Kerayong, Seri Intan dan Sabrang, dengan nilai  $d_r$  besar (0.87-0.93) dan nilai NMAE kecil (0.06-0.12). Walau bagaimanapun, ketinggian batang telah disimulasikan memuaskan untuk Rengam ( $d_r = 0.67$ , NMAE = 0.39) dan Sungai Buloh ( $d_r = 0.69$ , NMAE = 0.32). Selain itu, Sawit berjaya mensimulasikan kesan kejadian El Niño terhadap hasil kelapa sawit. Sawit juga memperhitungkan pengaruh tekstur tanah, taburan hujan, kepadatan penanaman, dan faktor-faktor meteorologi terhadap defisit air. Walau bagaimanapun, kesilapan simulasi meningkat dengan peningkatan kepadatan penanaman disebabkan kurangnya pencirian keadaan mikroiklim dan penyerapan air tumbuhan di bawah kanopi kelapa sawit yang padat, serta keragaman pengukuran yang lebih tinggi untuk kepadatan penanaman yang lebih tinggi. Secara kesimpulannya, model kelapa sawit yang bernama Sawit telah dibangunkan dan diparameterkan untuk mensimulasikan pertumbuhan dan hasil kelapa sawit di bawah pengaruh keadaan cuaca, kepadatan penanaman, dan tekstur tanah. Peningkatan dalam representasi mikroiklim kelapa sawit dan penyerapan air tumbuhan di bawah kanopi kelapa sawit yang padat, penyertaan aktiviti pembungaan, dan penyempurnaan mekanisme pemartitionan bahan kering batang boleh meningkatkan ketepatan Sawit.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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# LIST OF ABBREVIATIONS

A <sub>max</sub>	Maximum light-saturated photosynthesis rate
AET	Actual evapotranspiration
ASW	Available soil water content
BAR	Bunch CH <sub>2</sub> O requirement
BARMAX	Maximum bunch CH <sub>2</sub> O requirement
CH <sub>2</sub> O	Carbohydrate
FASW	Fractional available soil water content
FvCB	Farquhar-von Caemmerer-Berry
GDM	Reproductive dry matter production
L	Leaf area index
LUE	Light use efficiency
gst	Stomatal conductance
MAE	Mean absolute error
МРОВ	Malaysia Palm Oil Board
NMAE	Normalized mean absolute error
NMBE	Normalized mean bias error
PAR PD	Photosynthetically active radiation planting density
PET	Potential evapotranspiration
PNGOPRA	Papua New Guinea Oil Palm Research Association

RH	Relative humidity
RMSE	Root mean square error
SLA	Specific leaf area
TPU	Triose-phosphate untilization
TDM	Total dry matter production
USDA	United States Department of Agriculture
USDA-FAS	United States Department of Agriculture-Foreign Agriculture Service
VDMP	Vegetative dry matter production
VPD	Vapour pressure deficit
WUE	Water use efficiency
YAP	Year after field planting

G

## LIST OF SYMBOLS USED BY SAWIT MODEL

Symbol	Description	Unit
$\alpha_1$	fraction of PAR absorbed (0.8)	-
$\alpha_2$	extinction coefficient for wind speed	-
β	solar elevation or height (from horizontal)	radians
δ	solar declination	radians
Δ	slope of the saturated vapor pressure curve	mbar K-1
φ	solar azimuth (angle clockwise from North)	radians
$\phi_p$	soil porosity	-
γ	psychometric constant (0.658)	mbar K <sup>-1</sup>
Γ*	CO <sub>2</sub> compensation point	µmol mol <sup>-1</sup>
$\lambda_1$	site latitude	radians
$\lambda_2$	latent heat of vaporization of water (2454000)	J kg-1
$\lambda_3$	slope of the logarithmic suction-soil moisture curve	-
$\lambda ET$	total latent heat flux density	W m <sup>-2</sup>
$\lambda ET_c$	latent heat flux density of crop	W m <sup>-2</sup>
$\lambda ET_s$	latent heat flux density of soil	W m <sup>-2</sup>
$\Lambda_{canopy}$	gross canopy assimilation rate of $\text{CO}_2$ for the given hour	µmol CO <sub>2</sub> m <sup>-2</sup> ground s <sup>-1</sup>
$\Lambda_{sl/sh}$	gross leaf assimilation rate of CO <sub>2</sub> for sunlit (sl) or shaded (sh) leaves	µmol CO <sub>2</sub> m <sup>-2</sup> leaf s <sup>-1</sup>
π	pi constant (3.1428571)	-

 $\bigcirc$ 

θ	solar inclination (from vertical)	radians
$ heta_i$	volumetric water content for soil layer <i>i</i>	m <sup>3</sup> m <sup>-3</sup>
$ heta_{s}$	saturated volumetric water content	m <sup>3</sup> m <sup>-3</sup>
$\theta_{33}$	volumetric water content at field capacity	m <sup>3</sup> m <sup>-3</sup>
$ heta_{1500}$	volumetric water content at permanent wilting point	m <sup>3</sup> m <sup>-3</sup>
$ heta_{root}$	volumetric water content in the root zone	m <sup>3</sup> m <sup>-3</sup>
$ heta_{cr,root}$	critical volumetric water content in the root zone	m <sup>3</sup> m <sup>-3</sup>
$\theta_{s,root}, \\ \theta_{1500,root}$	volumetric water content at saturation and permanent wilting point in the root zone, respectively	m <sup>3</sup> m <sup>-3</sup>
$\Theta_i$	soil water content for soil layer <i>I</i> , expressed as water depth	m
$\rho c_p$	volumetric heat capacity of air (1221.09)	J m <sup>-3</sup> K <sup>-1</sup>
σ	Stefan-Boltzmann constant (5.67 x 10 <sup>-8</sup> )	$W m^{-2} K^{-4}$
$ au_1$	hour angle	radians
$ au_2$	CO <sub>2</sub> / O <sub>2</sub> specificity factor	µmol µmol-1
$ au_3$	soil tortuosity	-
$ au_{dr, \alpha}$	penetration function for direct solar radiation (corrected for scattering)	-
ω	canopy clustering coefficient	-
ω <sub>0</sub>	canopy clustering coefficient when the sun is at zenith	-
$\psi_e$	air entry suction	m
Age	tree age	days
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А	total energy supply	W m <sup>-2</sup>
$A_c$	fraction of energy available to the plant	W m <sup>-2</sup>
$A_s$	fraction of energy available to the soil	W m <sup>-2</sup>
С	fractional clay content in soil	- 0
$C_a$	ambient CO <sub>2</sub> concentration in air	µmol mol-1
Ci	intercellular CO <sub>2</sub> concentration	µmol mol-1
CVF, CVF <sub>2</sub>	conversion from CH <sub>2</sub> O to DM weight	kg DM kg <sup>-1</sup> CH <sub>2</sub> O
d	zero plane displacement	m
d <sub>root</sub>	rooting depth	m
dg <sub>root</sub>	rooting depth growth rate	m day-1
D	vapor pressure deficit	mbar
$D_0$	vapor pressure deficit at the mean canopy flow	mbar
Dleaf	leaf vapour pressure deficit	mbar
$D_{min}$	minimum vapour pressure deficit	mbar
$D_{m,v}$	vapor diffusion coefficient in air (24.7 x $10^{-6}$ )	$m^2 s^{-1}$
DL	day length	hour
$DM_x$	fraction of DM to plant part <i>x</i> , where <i>x</i> is for pinnae, rachis, trunk, and roots	-
ea	air vapor pressure	mbar
$e_m$	quantum efficiency or quantum yield	µmol µmol-1

$e_{s[[T]]}$	function returning the saturated vapor pressure at air temperature $T$	mbar
Ea	actual soil evaporation	m day-1
Eiso,sl/sh	emitted isoprene concentration by sunlit (sl) or shaded (sh) leaves	g C <sub>5</sub> H <sub>8</sub> g <sup>-1</sup> leaf s <sup>-1</sup>
$ET_c$	potential transpiration	m day-1
$ET_s$	potential evaporation	m day-1
fD	reduction to maximum stomatal conductance due to vapour pressure deficit	
<i>f</i> par	reduction to maximum stomatal conductance due to PAR	
fwater	reduction to maximum stomatal conductance due to water stress	
gst	stomatal conductance	m s <sup>-1</sup>
gst <sub>max</sub>	maximum stomatal conductance	m s <sup>-1</sup>
G	soil (ground) heat flux density	W m <sup>-2</sup>
G <sub>gen</sub>	assimilates available to generative organs growth	kg CH <sub>2</sub> O palm <sup>-1</sup> day <sup>-1</sup>
Ggrowth	assimilates available to growth respiration	kg CH <sub>2</sub> O palm <sup>-1</sup> day <sup>-1</sup>
Gmaleflo, Gimmbunch, Gmatbunch	growth rate of male flowers, immature bunches, and mature bunches, respectively	kg DM day-1
G <sub>x</sub>	growth rate of plant part <i>x</i> , where <i>x</i> is for pinnae, rachis, trunk, and roots	kg DM palm <sup>-1</sup> day <sup>-1</sup>
$G_{death,leaves}$	death rate of both pinnae and rachis due to natural causes	kg DM palm <sup>-1</sup> day <sup>-1</sup>

	$G_{death,roots}$	death rate of roots due to natural causes	kg DM palm <sup>-1</sup> day <sup>-1</sup>
	h	full tree height (trunk + canopy)	m
	h <sub>canopy</sub>	canopy height	m
	h <sub>trunk</sub>	trunk height	m
	h' <sub>trunk</sub>	rate of growth for trunk height	m day-1
	$H_i$	soil hydraulic gradient for soil layer <i>i</i>	m
	$H_c$	sensible heat flux density of crop	W m-2
	$H_{g,i}$	gravity head for soil layer <i>i</i>	m
	$H_{m,i}$	matric suction head for soil layer <i>i</i>	m
	$H_s$	sensible heat flux density of soil	W m <sup>-2</sup>
	Ic	solar constant	W m <sup>-2</sup>
	I <sub>df</sub>	diffuse solar irradiance	W m <sup>-2</sup>
	I <sub>dr</sub>	direct solar irradiance	W m <sup>-2</sup>
	I <sub>et</sub>	extraterrestrial solar irradiance	W m <sup>-2</sup>
	I <sub>sl/sh</sub>	total solar irradiance on sunlit (sl) or shaded (sh) leaves	W m <sup>-2</sup>
	I <sub>t</sub>	total solar irradiance	W m <sup>-2</sup>
	I <sub>df,d</sub>	daily diffuse solar irradiance	J m <sup>-2</sup> day <sup>-1</sup>
	I <sub>dr,d</sub>	daily direct solar irradiance	J m <sup>-2</sup> day <sup>-1</sup>
	I <sub>et,d</sub>	daily extraterrestrial solar irradiance	J m <sup>-2</sup> day <sup>-1</sup>
	I <sub>t,d</sub>	daily total solar irradiance	J m <sup>-2</sup> day <sup>-1</sup>
	k	von Karman's constant (0.4)	-

	$k_{df}$	canopy extinction coefficient for diffuse solar radiation (corrected for canopy clustering)	-
	k <sub>dr</sub>	canopy extinction coefficient for direct radiation	-
	K <sub>θ,i</sub>	soil hydraulic conductivity for soil layer <i>i</i>	m day-1
	$\overline{K}_{\Theta,i}$	logarithmic mean of hydraulic conductivity for soil layer <i>i</i> and ( <i>i</i> -1)	m day-1
	$K_c$	Michaelis-Menten constant for CO <sub>2</sub>	µmol mol-1
	Ko	Michaelis-Menten constant for O <sub>2</sub>	µmol mol-1
	K <sub>s,i</sub>	saturated hydraulic conductivity for soil layer <i>i</i>	m day-1
	$K_T$	sky clearness index	<u>\$</u>
	$l_1$	dry soil layer thickness	m
	$l_2$	air mixing height	m
	<i>l</i> <sub>3</sub>	mean pinnae length	m
	$l_m$	mean distance between pinnae	m
	L	leaf area index	m² leaf m² ground
	L <sub>cr</sub>	critical leaf area index	m² leaf m² ground
	L <sub>sh</sub>	shaded leaf area index	m² leaf m²² ground
	L <sub>sl</sub>	sunlit leaf area index	m² leaf m² ground
	L <sub>maxPD</sub>	maximum leaf area index for a given <i>PD</i> planting density (palms ha <sup>-1</sup> )	m² leaf m² ground

$M_{total}$ , $M'_{total}$	total maintenance respiration requirement with and without temperature correction, respectively	kg CH <sub>2</sub> O palm <sup>-1</sup> day <sup>-1</sup>
$M_x$	maintenance requirement for plant part <i>x,</i> where <i>x</i> is for pinnae, rachis, trunk, roots, organs, metabolic, and total	kg CH2O palm <sup>-1</sup> day <sup>-1</sup>
M <sub>c,x</sub>	maintenance coefficient for plant part $x$ , where $x$ is for pinnae, rachis, trunk, and roots	kg CH <sub>2</sub> O kg <sup>-1</sup> DM
MAX[x1, x2,xn], MIN[x1,x 2,xn]	functions to return the maximum and minimum values of $x_1, x_2,, x_n$ , respectively	
n	extinction coefficient for eddy diffusivity	-
N <sub>x</sub>	fraction by weight of nitrogen content in plant part $x$ , where $x$ is for pinnae, rachis, trunk, and roots.	
Oa	ambient concentration of O <sub>2</sub> in air	µmol mol-1
ОМ	organic matter content in soil	%
р	surface albedo (reflection)	-
$P_a$	atmospheric pressure (101)	kPa
PD	planting density	palms ha-1
P <sub>g,d</sub>	rainfall (above canopies) for the given day of year	mm
$\overline{P_{g,yr}}$	average rainfall in the year	mm
P <sub>net,d</sub>	net rainfall (below canopies) for the given day of year	mm
$PAR_{max}$	maximum PAR	W m <sup>-2</sup>
$q_i$	water flux into soil layer <i>i</i>	m day-1

$\widehat{q}_i$	net water flux into soil layer <i>i</i>	m day-1
$Q_{df}$	diffuse PAR component (above canopies)	µmol m <sup>-2</sup> leaf s <sup>-1</sup>
$Q_{dr}$	direct PAR component (above canopies)	µmol m <sup>-2</sup> leaf s <sup>-1</sup>
$Q_{sh}$	total PAR absorbed by shaded leaves	µmol m <sup>-2</sup> leaf s <sup>-1</sup>
$Q_{sl}$	total PAR absorbed by sunlit leaves	µmol m <sup>-2</sup> leaf s <sup>-1</sup>
Q10,ξ	relative change in a parameter $\xi$ for every 10 °C change	·
$\bar{Q}_{p,df}$	mean diffuse component of PAR (within canopies)	µmol m <sup>-2</sup> ground s <sup>-1</sup>
$Q_{p,dr}$	unintercepted direct component of PAR (with scattering component) (within canopies)	µmol m <sup>-2</sup> leaf s <sup>-1</sup>
$Q_{p,dr,\alpha}$	PAR scattered component only (within canopies)	µmol m <sup>-2</sup> ground s <sup>-1</sup>
$Q_{p,dr,dr}$	unintercepted direct component of PAR (without scattering component) (within canopies)	µmol m <sup>-2</sup> leaf s <sup>-1</sup>
r∆Ca	annual rate of change for ambient CO <sub>2</sub> concentration	µmol mol <sup>-1</sup> yr <sup>-1</sup>
$r_{\Delta Pg}$	annual rate of change for rainfall	mm yr <sup>-1</sup>
r⊿RH	annual rate of change for relative humidity	% yr-1
rдs	annual rate of change for sunshine hours	hour yr-1
۴∆Tmax	annual rate of change for maximum air temperature	°C yr <sup>-1</sup>
rΔTmin	annual rate of change for minimum air temperature	°C yr <sup>-1</sup>
$r_{\Delta u}$	annual rate of change for wind speed	m s <sup>-1</sup> yr <sup>-1</sup>

$R_n$	net solar radiation	W m <sup>-2</sup>
$R_{nL}$	net longwave radiation	W m-2
$R_{D,e}$	reduction factor for evaporation	-
$R_{D,t}$	reduction factor for transpiration	
RH	relative humidity for the given hour	%
$RH_d$	average relative humidity for the given day of year	%
S <sub>d</sub>	sunshine hours for the given day of year	hour
$S_i$	thickness of soil layer <i>i</i>	m
S	fractional sand content in soil	-
$S_i$	cumulative thickness of soil layer <i>i</i>	m
SLA	specific leaf area	$m^2 g^{-1}$
t <sub>c</sub>	the fraction of net radiation as soil heat flux under full canopies (0.05)	-
$t_d$	day of year	-
$t_h$	local solar time	hour
ts	the fraction of net radiation as soil heat flux for bare soil (no canopies) (0.315)	-
t <sub>sr</sub>	time of sunrise	hour
$t_{ss}$	time of sunset	hour
T <sub>air</sub>	air temperature	°C
$T_a$	actual plant transpiration	m day-1
T <sub>avg</sub>	average air temperature	°C

	$T_b$	base temperature for crop growth	°C
	$T_f$	temperature of canopy	°C
	T <sub>a,i</sub>	root extraction of water by roots in soil layer $i$	m day-1
	$T_{dew}$	dew point temperature	°C
	$T_{dew,cal}$	calibrated dew point temperature	°C
	$T_{max}$	maximum air temperature	°C
	$T_{min}$	minimum air temperature for the given hour	°C
	T <sub>set</sub>	air temperature at sunset	°C
	$\mathcal{U}^*$	friction velocity	m s <sup>-1</sup>
	и	wind speed at the current hour	m s <sup>-1</sup>
	$\mathcal{U}_A$	highest wind speed in the year	m s <sup>-1</sup>
	$\mathcal{U}_d$	mean wind speed for the given day of year	m s <sup>-1</sup>
	$u_h$	wind speed at canopy height <i>h</i>	m s <sup>-1</sup>
	U <sub>max</sub> , U <sub>min</sub>	maximum and minimum wind speed for the given day, respectively	m s <sup>-1</sup>
	<del>u<sub>yr</sub></del>	average wind speed in the year	m s <sup>-1</sup>
	v <sub>c</sub>	Rubisco-limited rate of CO <sub>2</sub> assimilation	µmol m <sup>-2</sup> leaf s <sup>-1</sup>
	vq	light-limited rate of CO2 assimilation	µmol m <sup>-2</sup> leaf s <sup>-1</sup>
	$v_s$	sink-limited rate of CO <sub>2</sub> assimilation	µmol m <sup>-2</sup> leaf s <sup>-1</sup>
	$V_{\rm cmax}$	Rubisco capacity rate (200)	µmol m <sup>-2</sup> leaf s <sup>-1</sup>
	$VDM_d$	daily VDM requirement	kg DM palm <sup>-1</sup> day <sup>-1</sup>

VDM <sub>yr</sub>	annual VDM requirement	kg DM palm <sup>-1</sup> yr <sup>-1</sup>
VDM <sub>max,PD</sub>	maximum annual VDM for the given planting density	kg DM palm <sup>-1</sup> yr <sup>-1</sup>
w	mean pinnae width	m
W <sub>x</sub>	dry weight of plant part <i>x</i> , where <i>x</i> is for pinnae, rachis, trunk, roots, male flowers ( <i>maleflo</i> ), immature bunches ( <i>immbunch</i> ), and mature bunches ( <i>matbunch</i> ).	kg DM palm-1
X <sub>c</sub>	correction for mineral content in all plant parts (2.0)	
X <sub>x</sub>	fraction by weight of mineral content in plant part <i>x</i> , where <i>x</i> is for pinnae, rachis, trunk, and roots.	-
$Z_0$	surface roughness length	m
Zi	depth from soil surface to the middle of soil layer <i>i</i>	m
Zr	reference height	m
$Z_{s0}$	roughness length of soil surface	m

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#### CHAPTER 1

#### **INTRODUCTION**

#### 1.1 Research Background

Oil palm (*Elaeis guineensis* Jacq.) is one of the world's major vegetable oil crops. Global consumption of palm oil over the last two decades has tripled, and annual consumption increased from 24.99 million tonnes in 2001 (USDA, 2006) to 74.90 million tonnes in 2020 (USDA, 2021) due to the extensive use of palm oil in a wide range of consumer products, and production of biofuel (Koh and Ghazoul, 2008). Malaysia, the second largest producer and exporter of palm oil in the world (USDA, 2021), has benefited considerably from the worldwide growing demand for palm oil. The area planted with oil palm had expanded from 540,000 hectares in 1960 to 5.87 million hectares in 2020 (MPOB, 2021), with Sarawak having the largest planted area and closely followed by Sabah (Figure 1.1). Peninsular Malaysia has the largest oil palm planted area in Malaysia, with 2.74 million hectares or 46.7% of the total planted area. In 2021, Malaysia exported 15.56 million metric tonnes of palm oil from a total production of 18.12 million. Together with other palm-based products, the Malaysian palm oil industry brought in RM108.52 billion of export earnings to the country (MPOB, 2022), signifying the economic importance of oil palm to the Malaysian economy.



Figure 1.1: Distribution (%) of oil palm planted area in Malaysia, based on the total planted area of 5.87 million hectares recorded in 2020 (MPOB, 2021).

Oil palm requires an optimum mean temperature of 24-33 °C (Corley and Tinker, 2016; Paterson et al., 2015). Furthermore, it grows well with at least 5 hours of sunshine hours per day or equivalent solar radiation of 15-17 MJ m<sup>-2</sup> day<sup>-1</sup> (Corley and Tinker, 2016; Rhebergen et al., 2016) and requires an evenly distributed annual rainfall of 2000-2500 mm (Corley and Tinker, 2016; Bakoumé et al., 2013). Oil palm is, however, sensitive to water stress, and the production of palm oil is affected by the length and severity of drought (Oettli et al., 2018; Caliman and Southwood, 1998). Therefore, extreme weather events such as El Niño, which brings much lower rainfall, could significantly impact palm oil production through lower fruit bunch formation. This was most notable in 2015/2016, when an extreme El Niño event blanketed Malaysia and Indonesia (Lim et al., 2017), causing severe water stress to oil palms (Oettli et al., 2018; Azlan *et al.*, 2016). As a result, palm oil production in Malaysia reduced as much as 11% in 2015 and 5% in 2016 compared to production in 2014, the year preceding the onset of El Niño (USDA-FAS, 2021). Such adverse impact of El Niño on palm oil yield underpinned the vulnerability of oil palm to the vagaries of climatic changes.

Global warming, mainly driven by rising atmospheric CO<sub>2</sub> concentration, is projected to continue in the years ahead regardless of efforts to reduce its emissions (IPCC, 2018). Under a warmer climate, Malaysia will experience drier and more extreme weather in the years ahead (Kwan et al., 2013), and weather anomalies such as El Niño will become more frequent and intense (Wang et al., 2019; Cai et al., 2018, 2014; Keupp et al., 2017). El Niño coupled with land-use change such as converting forest to oil palm, can change the partition of surface energy balance and increase sensible heat fluxes, which in turn could lead to higher surface temperature (Ma'rufah et al., 2021). Mean air temperature in Malaysia has increased at a rate of 0.14–0.25 °C per decade (NRE, 2015), and it is expected to increase further in tandem with the projected increase in global mean air temperature of at least 1.7 °C by the end of the 21st century (Loh et al., 2016). Furthermore, the rainfall patterns in Malaysia are projected to vary substantially in space and time, with a tendency for reduced rainfall from December to May by 20-40% (Loh et al., 2016). Thus, the imminent threat of climate change on palm oil production in Malaysia is real. Rising air temperature coupled with frequent water deficit will stress oil palm (Oettli et al., 2018) and, in turn, reduce palm oil production. Areas under oil palm cultivation in Malaysia might reduce substantially under future warmer and drier climates (Paterson et al., 2017, 2015).

However, elevated atmospheric  $CO_2$  concentration is beneficial to C3 plants like oil palms because photosynthesis of C3 plants will be enhanced under elevated atmospheric  $CO_2$  concentration (Ainsworth and Long, 2005). Thus, the adverse effects of heat and water stresses might be moderated by the increased rate of  $CO_2$  assimilation. For example, oil palm was more tolerant to drought during the 2015/2016 El Niño event under high solar irradiance. However, increased  $CO_2$  concentration did not increase the photosynthesis rate under reduced solar irradiance (Stiegler *et al.*, 2019). On the other hand, oil palm seedlings (Jaafar and Ibrahim, 2012; Ibrahim *et al.*, 2010) and mature oil palm (Henson and Harun, 2005a, b) have shown increased growth after exposure to elevated  $CO_2$  concentration a result of increased rates of  $CO_2$  assimilation and lower transpiration rates. Therefore, we must understand how oil palm responds to weather variables so that crop management adaptation strategies can be developed to mitigate the detrimental impacts of weather irregularities on oil palm.

Given the complex interactions between weather variables and crop production system, crop models have been increasingly used to understand how weather variables affect crop production and to assist in the development of crop management adaptation strategies such as planting densities, cultivar selection, irrigation, fertiliser regimes and crop rotations (Challinor *et al.*, 2018; Asseng *et al.*, 2014; Rosenzweig *et al.*, 2014). Although the crop model is a simplified mathematical representation of a complex crop-soil-atmosphere system, it integrates essential biophysical functions that drive crop growth, such as meteorology, photosynthesis, respiration, transpiration, phenology, soil evaporation, and uptake of nutrients and water (Asseng *et al.*, 2014; Teh, 2006). As such, making it an effective means to study crop performance across diverse environments and management practices. Rival (2017) pointed out that breeding oil palm for climate change will require multidisciplinary research, and modelling responses of oil palm to changing climate is paramount in developing climate-resilient oil palm genotypes and crop management practices.

Many oil palm models such as OPSIM (van Kraalingen, 1989 *et al.*; Gerristma, 1988), SIMPALM (Dufrêne *et al.*, 1990), GPHOT, GPHOT2, OPLFSIM3, OPRODSIM (Henson, 2009, 2006b, 2000, 1989), WaNuLCAS (van Noordwijk *et al.*, 2011), ECOPALM (Combres *et al.*, 2012), PALMSIM (Hoffmann *et al.*, 2014), APSIM-Oil Palm (Huth *et al.*, 2014), CLM-Palm (Fan *et al.*, 2015), CLIMEX-Oil Palm (Paterson *et al.*, 2015) and ORCHIDEE–MICT–OP (Xu *et al.*, 2020) have been developed especially over the last two decades to meet different objectives of studies. These oil palm models differ in regard to model structure, parameterization of biophysical processes and model outputs. Though these models have been applicable, there are several key features that make them less versatile in terms of simulating the responses of oil palm to weather irregularities and crop management practices.

### 1.2 Problem Statement

Key features that make the current oil palm models less versatile in terms of simulating the responses of oil palm to weather irregularities and crop management practices are:

First, the prevailing approach is to use daily or monthly weather data to model physiological processes, such as photosynthesis, transpiration, and respiration. Nonetheless, according to Ephrath *et al.* (1996), the use of daily or monthly weather data frequently results in deviations when modelling crop responses to weather variables. This is because physiological processes immediately respond to weather variables, and daily or monthly weather data may not suffice to account for these instantaneous responses compared to hourly weather data. A potential solution to address this issue is to simulate the hourly patterns of weather variables based on available daily weather data. However, this approach has not yet been implemented in major oil palm growing regions in Malaysia.

Second, except for CLM-Palm (Fan et al., 2015) and ORCHIDEE-MICT-OP (Xu et al., 2020), all other oil palm models used oil palm's radiation use efficiency (RUE) function to convert the intercepted radiation into gross assimilates. Though this RUE function is useful but is empirical and hence less robust in simulating photosynthesis responses to rising atmospheric CO<sub>2</sub> concentration (Streck et al., 2012) and other environmental variables (Wu et al., 2016). In contrast, the biochemical photosynthesis model of Farquhar et al. (1980) is more mechanistic and has been used extensively to predict responses of photosynthesis to environmental variables (Xu et al., 2020; Fan et al., 2015; Medlyn et al., 2005; Cramer et al., 2001). However, this biochemical photosynthesis model requires the temperature dependency of Rubisco kinetics, such as maximum Rubisco carboxylation rate  $(V_{cmax})$ , electron transport rate (J), leaf day respiration rate in the light  $(R_d)$ , CO<sub>2</sub> photocompensation point in the absence of leaf day respiration ( $\Gamma^*$ ) and mesophyll conductance to CO<sub>2</sub> ( $g_m$ ) (von Caemmerer, 2000). These parameters and their temperature dependency are, however, largely unknown for oil palm, although Xu et al. (2020), Nugroho (2018), Meijide et al. (2017) and Fan et al. (2015) had made some measurements to estimate the  $V_{cmax}$  of oil palm.

Third, simulations of evapotranspiration or transpiration of oil palm by most of oil palm models either use the Penman-Monteith model (Monteith, 1973) or a simple transpiration efficiency (TE) coefficient (Huth *et al.*, 2014). Nevertheless, this approach appears inadequate to model the microclimate environment within and under the oil palm canopies. The transpiration efficiency coefficient is an empirical value which might vary seasonally and from place to place, depending on weather conditions and management practices (Tfwala *et al.*, 2021;

Bennie *et al.*, 1997). Furthermore, it only accounts for water transpired from oil palm but ignores soil evaporation which is inadequate as soil evaporation under oil palm canopies can be substantial (Röll *et al.*, 2015). On the other hand, the Penman-Monteith model only simulates heat fluxes from either soil or crop but not both simultaneously.

In contrast, the Shuttleworth-Wallace model (Shuttleworth and Wallace, 1985) extends the Penman-Monteith model but allows simultaneous simulation of heat fluxes from both soil and crop. This has the advantage of getting a more accurate simulation of soil evaporation, especially during wet periods when the soil moisture content is high (Fisher et al., 2005). The Shuttleworth-Wallace model has done well in many studies (Yan and Oue, 2011; Iritz *et al.*, 1999; Vörösmarty *et al.*,1998; Farahani and Bausch, 1995). However, like its predecessor, the main drawback is the requirement to measure its parameters (Farahani and Ahuja, 1996) which has not been done for oil palm.

Fourth, the first oil palm model, OPSIM (van Kraalingen et al., 1989; van Kraalingen, 1985), used respiration coefficients to simulate the maintenance respiration of oil palm. These coefficients were calculated from nitrogen and mineral content determined in different plant parts of oil palm. This approach is touted to be more mechanistic (Penning de Vries, 1975, 1972) and was adopted by SIMPALM (Dufrêne et al., 1990), OPRODSIM (Henson, 2009) and PALMSIM (Hoffmann et al., 2014). The maintenance respiration coefficients used in the SIMPALM (Dufrêne et al., 1990) and PALMSIM (Hoffmann et al., 2014) were computed from nitrogen and mineral contents determined from a single tenera x dura (L2T x D10D) oil palm progeny grown in Ivory Coast. On the other hand, OPSIM (van Kraalingen et al., 1989; van Kraalingen, 1985) and OPRODSIM (Henson, 2009) determined their maintenance respiration coefficients using nitrogen and mineral contents reported by Ng et al. (1968), which were primarily obtained from dura oil palm grown under good growing conditions in Malaysia. These maintenance respiration coefficients are rather dated and may not apply today for the current high-yielding tenera oil palm.

Fifth, the growth of oil palm can be separated into vegetative and reproductive growth. Vegetative growth consists of leaf, trunk, and root growth (Corley and Tinker, 2016). However, the methods used to simulate vegetative growth are primarily empirical, based on the vegetative dry matter requirement of oil palm (Henson, 2009; Dufrêne *et al.*, 1990; van Kraalingen, 1985) or phytomer phenology of oil palm (Xu *et al.*, 2020; Fan *et al.*, 2015; Combres *et al.*, 2013). Therefore, to minimize errors in simulating vegetative growth, it would be necessary to use the updated data on phytomer phenology or dry matter requirement and partitioning derived from the current tenera oil palm.

Sixth, planting density is a crucial crop management strategy that drives palm oil yield (Palat *et al.*, 2012; Nazeeb *et al.*, 2008; Nazeeb *et al.*, 1989; Breure, 1988; Corley, 1973). Although commercial oil palm plantations generally plant between 128 and 148 palms per hectare but planting densities higher than this range have been explored to increase palm oil yield (Palat *et al.*, 2012; Breure, 2010; Nazeeb *et al.*, 2008; Breure *et al.*, 1990; Nazeeb *et al.*, 1989; Breure and Corley, 1983). Changes in planting density will affect interception of solar irradiance by oil palm canopies, microclimate within and under oil palm canopies and soil water balance, consequently affecting the growth and yield of oil palm. Having able to use a crop model to characterize the interactions between planting density and environmental variables will give better insight into how planting density can be managed to drive palm oil production. However, except for OPRODSIM (Henson, 2009), the present oil palm models cannot essentially simulate the growth and yield of oil palm grown under different planting densities.

### 1.3 Objectives of Research

Given the drawbacks of the current oil palm models, the present study was undertaken to develop and validate a versatile and more mechanistic oil palm model that assesses the impacts of weather irregularities and crop management practices on the growth and yield of oil palm specifically under Malaysian environment and management. To achieve this overall objective, the following specific objectives were undertaken:

- i. To parameterise the meteorological models for hourly estimations of air temperature, relative humidity, and solar irradiance.
- ii. To parameterise the biochemical model of C3 leaf photosynthesis through photosynthesis measurements on different ages of tenera oil palm in the fields.
- iii. To parameterise maintenance respiration and dry matter partitioning coefficients using nitrogen and mineral contents and dry matter production data from destructive measurements on different ages of current tenera oil palm.
- iv. To parameterise the Shuttleworth-Wallace model through measurements on stomatal conductance, trunk, and canopy height of different ages of tenera oil palm.
- v. To develop a new oil palm growth and yield model and validate its accuracy against observed data from six oil palm planting density trials conducted in Malaysia.
- vi. To examine the impact of weather anomalies such as El Niño on oil palm growth and yield through simulations.

## 1.4 Limitation of the Study

The present study only focuses on developing an oil palm growth and yield model that is applicable to the Dura x Pisifera oil palm progeny. In addition, the present study does not examine the effects of La Niña and fertiliser on oil palm growth and yield through simulations.

### 1.5 Structure of the PhD Thesis

The thesis consists of seven chapters, with the first chapter outlining the research background and objectives. In Chapter 2, the growth and yield of oil palm and their simulation methods are reviewed. Chapters 3 through 5 have been published in peer-reviewed journals, while Chapter 6 appears as a book chapter. Chapter 7 concludes the thesis by summarizing the research findings and discussing how they relate to the research objectives. Additionally, recommendations for future research are provided.

# 1.5.1 Chapter 3

The third chapter is published by Cheah See Siang, Christopher Teh Boon Sung, Mohd Razi Ismail and Mohd Rafii Yusop (2020). **Modelling hourly air temperature, relative humidity and solar irradiance over several major oil palm growing areas in Malaysia**. *Journal of Oil Palm Research* Vol. 32(1): 34–49. In this chapter, a study was undertaken with the aim to assess the precision of various meteorological models in estimating hourly air temperature, relative humidity, and solar irradiance values across several significant oil palm cultivation regions in Malaysia. The validated meteorological models were used to generate hourly estimations of these weather variables, which were then used as inputs for the oil palm growth and yield model.

## 1.5.2 Chapter 4

The fourth chapter is published as S.S. Cheah and C.B.S. Teh (2020). **Parameterization of the Farquhar-von Caemmerer-Berry C3 photosynthesis model for oil palm**. *Photosynthetica* 58(3): 769-779. This research was undertaken to determine critical model parameters of the Farquhar-von Caemmerer-Berry (FvCB) C3 biochemical photosynthesis model for oil palm. The key FvCB model parameters such as photocompensation point ( $\Gamma^*$ ), mesophyll conductance ( $g_m$ ), maximum rates of Rubisco carboxylation ( $V_{cmax}$ ) and electron transport ( $J_{max}$ ), triose phosphate utilization (TPU) and their temperature dependencies between

25 and 40°C in oil palm were determined. The derived values of these parameters were then used to parameterize the photosynthesis module of the oil palm growth and yield model.

# 1.5.3 Chapter 5

The fifth chapter is published as Cheah See Siang, Siti Aishah Abd Wahid and Christopher Teh Boong Sung (2022). **Standing biomass, dry matter production, and nutrient demand of tenera oil palm**. *Agronomy* 12, 426. This research was undertaken to determine the aboveground vegetative biomass, dry matter production and partitioning, and nutrient contents in different plant parts of tenera oil palms grown under current crop management standards in Malaysia. The results were used to determine dry matter partitioning coefficients and maintenance respiration coefficients of oil palm.

## 1.5.4 Chapter 6

The sixth chapter was published as Christopher Teh Boon Sung and Cheah See Siang (2018). **Modelling crop growth and yield in palm oil cultivation**. In *Achieving sustainable cultivation of oil palm*. Volume 1: Introduction, breeding and cultivation techniques. Alain Rival (ed.) Burleigh Dodds Science Publishing, Cambridge, UK. This book chapter describes the development of a new oil palm growth and yield model called PySawit, and its validation using data collected from an oil palm planting density trial. Since the publication of PySawit in 2018, further improvements have been made and the oil palm growth and yield model has been renamed as Sawit. Additional trial data collected from six different locations in Malaysia were used to further validate Sawit. Further development of Sawit is described and additional validations are discussed in Chapter 6.

#### REFERENCES

- Adam, H., Jouannic, S., Escoute, J., Duval, Y., Verdeil, J. and Tregear, W. 2005. Reproductive developmental complexity in the African oil palm (*Elaeis guineensis*, Arecaceae). American Journal of Botany, 92(110: 1836-1852.
- Ainsworth, E. and Long, S. 2005. What have we learned from 15 years of Free-Air CO<sub>2</sub> Enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy properties and plant production to rising CO<sub>2</sub>. The New phytologist, 165: 351–71.
- Ainsworth, E. and Rogers, A. 2007. The response of photosynthesis and stomatal conductance to rising [CO<sub>2</sub>]: mechanisms and environmental interactions. Plant, Cell & Environment, 30: 258–270.
- Ali, A.A., Nugroho, B., Moyano, F.E., Brambach, F., Jenkins, M.W., Pangle, R., Stiegler, C., Blei, E., Cahyo, A.N., Olchev, A., et al. 2021. Using a Bottom-Up Approach to Scale Leaf Photosynthetic Traits of Oil Palm, Rubber, and Two Coexisting TropicalWoody Species. Forests, 2021 (12): 359.
- Ali, M.H., Lee, T., Kwok, C., Eloubaidy, A.F. 2000. Modelling evaporation and evapotranspiration under temperature change in Malaysia. Pertanika Journal of Science and Technology, 8: 191–204.
- Alonso, A., Pérez, P., Martinez-Carrasco, R. 2009. Growth in elevated CO<sub>2</sub> enhances temperature response of photosynthesis in wheat. Physiologia Plantarum, 135: 109-120.
- Amthor, J.S. 1984. The role of maintenance respiration in plant growth. Plant, Cell & Environment, 7(8): 561-569.
- Amthor, J.S. 2000. The McCree-de Wit-Penning de Vries-Thornley respiration paradigms: 30 years later. Annals of Botany, 86: 1-20.
- Angstrom, A. 1924. Solar and terrestrial radiation. Quaterly Journal of the Royal Meteoralogical Society, 50: 121.
- Asseng, S., Ewert, F., Martre, P., Rötter, R.P., Lobell, D.B., Cammarano, D., Kimball, B.A., Ottman, M.J., Wall, G.W., White, J.W., et al. 2015. Rising temperatures reduce global wheat production. Nature Climate Change, 5: 143.
- Asseng, S., Zhu, Y., Basso, B., Wilson, T., Cammarano, D. and van Neal, K. 2014. Simulation modeling: applications in cropping systems A2- Alfen. In: Encyclopedia of Agriculture and Food Systems. Academic Press, Oxford, pp. 102–112.

- Atkin, O.K., Evans, J.R., Ball, M.C Lambers, H. and Pons, T.L. 2000. Leaf respiration of snow gum in the light and dark. Interactions between temperature and irradiance. Plant Physiology, 122: 915-923.
- Augustine, C. and Nnabuchi, M. 2009. Correlation of cloudiness index with clearness index for four selected cities in Nigeria. The Pacific Journal of Science and Technology, 10: 568-573.
- Awal, M.A. 2006. Image-based measurement of leaf area index and radiation interception for modelling of oil palm, PhD Thesis, Universiti Putra Malaysia.
- Awal, M.A. 2008. Assessment of gap fraction by hemispherical photograph in oil palm plantation. Suranaree Journal of Science and Technology, 15: 233-241.
- Awal, M.A., Wan Ishak, W.I. and Bockari-Gevao, S.M. 2010. Determination of leaf area index for oil palm plantation using hemispherical photography technique. Pertanika Journal of Science and Technology, 18(1): 23 32.
- Awal, MA., Ishak, W., Endan, J. and Haniff, M. 2004. Determination of specific leaf area and leaf area-laf mass relationship in oil palm plantation. Asian Journal of Plant Sciences, 3(3): 264-268.
- Ayub G., Smith R.A., Tissue D.T. *et al.* 2011. Impacts of drought on leaf respiration in darkness and light in Eucalyptus saligna exposed to industrial-age atmospheric CO<sub>2</sub> and growth temperature. New Phytologist, 190: 1003-1018.
- Azlan, A.H., Lee, C.T., Soh, K.Y., Selvaraja, S., Rohan, R., Arffin, I. and Palaniappan, S. 2016. Impact of El Niño on palm oil production. The Planter, 92(1088): 789-806.
- Babst, F., Bouriaud, O., Poulter, B., Trouet, V., Girardin, M.P., Frank, D.C. 2019. Twentieth century redistribution in climatic drivers of global tree growth. Science Advances 5: eaat4313.
- Bakoumé, C., Shahbudin, N., Yacob, S., Siang, C. S., and Thambi, M. N. 2013. Improved method for estimating soil moisture deficit in oil palm (*Elaeis guineensis* Jacq.) areas with limited climatic data. Journal of Agricultural Science, 5: 57–65.
- Barnard, H.R. and Ryan M.G. 2003. A test of the hydraulic limitation hypothesis in fast-growing *Eucalyptus saligna*. Plant, Cell & Environment, 26: 1-11.
- Bauerle, W.L., Weston, D.J., Bowden, J.D., Dudley, J.B. and Toler, J.E. 2004. Leaf absorptance of photosynthetically active radiation in relation to chlorophyll

meter estimates among woody plant species. Scientia Horticulturae, 101: 169-178.

- Bennie, A.T.P., van Rensburg, L.D., Strydom, M.G. and Du Preez, C.C. 1997. Response of crops on pre-programmed deficit irrigation. Water Research Commission Report no. 423/1/97, Pretoria, South Africa.
- Bernacchi, C.J., Pimentel, C. and Long, S.P. 2003. *In vivo* temperature response functions of parameters required to model RuBP-limited photosynthesis. Plant, Cell & Environment, 26: 1419-1430.
- Bernacchi, C.J., Portis, A.R., Nakano, H., von Caemmerer, S. and Long, S.P. 2002. Temperature response of mesophyll conductance. Implication for the determination of Rubisco enzyme kinetics and for limitations to photosynthesis in vivo. Plant Physiology, 130: 1992–1998.
- Bernacchi, C.J., Singsaas, E.L. Pimentel, C., Portis Jr, A.R. and Long, S.P. 2001. Improved temperature response functions for models of Rubisco-limited photosynthesis. Plant, Cell & Environment, 24: 253–259.
- Betti M., Bauwe H., Busch, F.A., Fernie, A.R., Keech, O., Levey, M., Ort, D.R., Parry, M.A., Sage, R., Timm, S., Walker, B. and Weber, A.P. 2016. Manipulating photorespiration to increase plant productivity: recent advances and perspectives for crop improvement. Journal of Experimental Botany, 67: 2977-2988.
- Bittelli, M., Campbell, G.S., and Tomeu, F. 2015. Soil physics with Python. Transport in the soil-plant-atmosphere system. Oxford UK: Oxford University Press.
- Brenner, A.J. 1996. "Microclimate modifications in agroforestry," in Tree Crop Interactions: a Physiological Approach, eds C. K. Ong and P. Huxley (Wallingford: CAB International/ ICRAF), 157–187.
- Breshears, D.D., Adams, H.D., Eamus, D., McDowell, N.G., Law, D.J., Will, R.E., Williams, A.P., Zou, C.B. 2013. The critical amplifying role of increasing atmospheric moisture demand on tree mortality and associated regional dieoff. Frontiers in Plant Science, 4: 266.
- Breure, C.J. 1982. Factors affecting yield and growth of oil palm teneras in West New Britain. Oléagineux, 37: 213-227.
- Breure, C.J. 1985. Relevant factors associated with crown expansion in oil palm (*Elaeis guineensis* Jacq.). Euphytica, 34: 161–175.
- Breure, C.J. 1988. The effect of different planting densities on yield trends in oil palm. Experimental Agriculture, 24: 37–52.

- Breure, C.J. 1994. Development of leaves in oil palm (*Elaeis guineensis* Jacq) and the determination of leaf opening rates. Experimental Agriculture, 30: 467-472.
- Breure, C.J., 2003. The search for yield in oil palm: basic principles. In: Oil palm: management for large and sustainable yields, (Ed. by Fairhurst, T., Härdter, R.).1<sup>st</sup> ed. PPI/PPIC-IPI, Singapore, pp. 59-98.
- Breure, C.J. 2010. Rate of leaf expansion: a criterion for identifying oil palm (*Elaeis guineensis* Jacq.) types suitable for planting at high densities. NJAS Wageningen Journal of Life Sciences, 57: 141-147
- Breure, C.J. and Corley, R.H.V. 1983. Selection of oil palms for high density planting. Euphytica, 32: 177-186.
- Breure, C.J. and Corley, R.H.V. 1992. Fruiting activity, growth and yield of oil palm. II. Observations in untreated populations. Experimental Agriculture, 28: 111-121.
- Breure, C.J. and Menendez, T. 1990. The determination of bunch yield components in the development of inflorescences in oil Palm (*Elaeis Guineensis*). Experimental Agriculture, 26: 99-115.
- Breure, C.J. and Powell, M.S. 1988. The one-shot method of establishing growth parameters in oil palm. In: Proceedings of the 1987 International Oil Palm Conference. Progress and prospects, (Ed. by Halim, H.A., Chew, P.S. and Wood, B.J.), pp203-209, Palm Oil Research Institute of Malaysia, Kuala Lumpur.
- Breure, C.J. and Siregar, M.M. 2020. Selection of oil palm male parents for optimal planting density estimated from mature crown surface. Journal of Oil Palm Research, 32(2): 191-200.
- Breure, C.J., Menendez, T. and Powell, M.S. 1990. The effect of planting density on the yield components of oil palm (*Elaeis guineensis* Jacq.). Experimental Agriculture, 26: 117-124.
- Bristow, K.L. and Campbell, G.S. 1984. On the relationship between incoming solar radiation and daily maximum and minimum temperature. Agricultural and Forest Meteorology, 31(2): 159-66.
- Broekmans, A.F.M. 1957. Growth, flowering and yield of the oil palm in Nigeria. Journal of West African Institute for Oil Palm Research, 2: 187-220.
- Brutsaert, W. (1982). Evaporation into the Atmosphere. Theory, History and Applications. Dordrecht, The Netherlands: D. Reidel Publishing Company.

- Buckley, T.N. and Diaz-Espejo, A. 2015. Reporting estimates of maximum potential electron transport rate. New Phytologist, 205: 14-17.
- Bunce J.A. 2008. Acclimation of photosynthesis to temperature in *Arabidopsis thaliana* and *Brassica oleracea*. Photosynthetica, 46: 517-524.
- Burgess, A.J., Retkute, R., Preston, S.P., Jensen, O.E., Pound, M.P., Pridmore, T.P. and Murchie, E.H. 2016. The 4-Dimensional Plant: Effects of Wind-Induced Canopy Movement on Light Fluctuations and Photosynthesis. Frontier in Plant Science, 7:1392.
- Cai, W., Borlace, S., Lengaigne, M., van Resch, P., Collins, M., Vecchi, G., and Jin, F.F. 2014. Increasing frequency of extreme El Niño events due to greenhouse warming. Nature Climate Change, 4: 111–116.
- Cai, W., Wang, G., Dewitte, B., Wu, L., Santoso, A., Takahashi, K., and McPhaden, M. 2018. Increased variability of eastern Pacific El Niño under greenhouse warming. Nature, 564: 201–206.
- Caliman, J.P. and Southworth, A. 1998. Effect of drought and haze on the performance of oil palm. In: Proc. 1998 Int. Oil Palm Conf. 'Commodity of the past, today and the future' (Ed. by A. Jatmika *et al.*), pp. 250-274, Indonesian Oil Palm Research Institute, Medan, Indonesia.
- Campbell, G.S. 1994. Soil physics with basic transport models for soil-plant systems. Developments in Soil Science 14. Elsevier Science B.V., Amsterdam, The Netherlands.
- Campbell, G.S. and Norman, J.M. 1998. An introduction to environmental biophysics. 2<sup>nd</sup> Edition. Springer-Verlag, New York.
- Carr, M.K.V. 2011. The water relations and irrigation requirements of oil palm (*Elaeis guineensis*): a review. Experimental Agriculture, 47: 629-652.
- Challinor, A.J. and Wheeler, T.R. 2008. Crop yield reduction in the tropics under climate change: processes and uncertainties. Agricultural and Forest Meteorology, 148: 343–356.
- Challinor, A.J., Müller, C., Asseng, S., Deva, C., Nicklin, K.J., Wallach, D., Vanuytrecht, E., Whitfield, S., Ramirez-Vilegas, J. and Koehler, A. 2018. Improving the use of crop models for risk assessment and climate change adaptation. Agricultural Systems, 159: 296-306.
- Chan, K.W. 1979. Irrigation of oil palm in Malaysia. In: Proc. Symp. Water in Malaysian agriculture (Ed. by Pushparajah, E.), pp. 103-116, Malaysian Society of Soil Science, Kuala Lumpur.

- Chan, K.W. 1991. Predicting oil palm yield potential based upon solar radiation. Paper presented at 2<sup>nd</sup> National Seminar on Agrometeorology, Meteorological Department Malaysia, Petaling Jaya.
- Choudhury, B.J. and Monteith, J.L. 1988. A four-layer model for the heat budget of homogeneous land surfaces. Quarterly Journal of the Royal Meteorological Society, 114: 373-398.
- Chuah, D.G.S. and Lee, S.L. 1981. Solar radiation estimates in Malaysia. Solar Energy, 26: 33-40.
- Collatz, G.T., Ball, J.T., Grivet, C. and Berry, J.A. 1991. Physiological and environmental - regulation of stomatal conductance, photosynthesis and transpiration: a model that includes a laminar boundary layer. Agricultural and Forest Meteorology, 54: 107-136.
- Combres, J.C., Pallas, B., Rouan, L., Mialet-Serra, I., Caliman, J.P., Braconnier, S., Soulié, J.C. and Dingkuhn, M. 2012. Simulation of inflorescence dynamics in oil palm and estimation of environment-sensitive phenological phases: A model based analysis. Functional Plant Biology, 40: 263–79.
- Corley, R.H.V. 1973. Effects of plant density on growth and yield of oil palm. Experimental Agriculture, 9: 169-180
- Corley, R.H.V. 1976a. Photosynthesis and productivity. In: Oil palm research (Ed. by R.H.V. Corley, J.J. Hardon and B.J. Wood), Elsevier, Amsterdam, pp. 55-76.
- Corley, R.H.V. 1976b. Inflorescence abortion and sex differentiation. In: Oil palm research (Ed. by Corley, R.H.V., Hardon, J.J. and Wood, B.J.), Elsevier, Amsterdam, pp. 37-54.
- Corley, R.H.V. 1976c. Effects of severe leaf pruning on oil palm, and its possible use for selection purposes. Malaysia Agriculture Research and Development Institute Research Bulletin, 4(2): 23-28.
- Corley, R.H.V. 1977. Oil palm yield components and yield cycles. In: International developments in oil palm (Ed. by Earp, D.A. & Newall, W.), Incorporated Society of Planters, Kuala Lumpur, pp. 116-129
- Corley R.H.V. 1986. Oil palm. In: CRC Handbook of fruit set and development (Ed. by Monselise, S.P.), CRC Press, Boca Raton, Florida, pp. 253-259
- Corley, R.H.V. and Breure, C.J. 1992. Fruiting activity, growth and yield of oil palm. I. Effects of fruit removal. Experimental Agriculture, 28: 99-109.

- Corley, R.H.V. and Donough, C.R. 1992. Potential yield of oil palm clones the importance of planting density. In: Proceedings of the workshop on yield potential in the oil palm (Ed. by Rao, V., Henson, I.E. & Rajanaidu, N.), pp. 58-70, International Society for Oil Palm Breeders, Kuala Lumpur, pp.58-70.
- Corley, R.H.V. and Gray, B.S. 1976. Yield and yield components. In: Developments in Crop Science. 1. Oil Palm Research (Ed. by Corley, R.H.V., Hardon J.J., Woods, B.), Elsevier Science, Netherlands, pp. 77-86.
- Corley, R.H.V. and Hong, T.K. 1982. Irrigation of oil palms in Malaysia. In: The oil palm in agriculture in the eighties, Vol. 2 (Ed. By Pushparajah, E. and Chew, P.S.), Incorporated Society of Planters, Kuala Lumpur, pp. 343-346
- Corley, R.H.V. and Mok, C.K. 1972. Effects of nitrogen, phosphorus, potassium and magnesium on growth of the oil palm. Experimental Agriculture, 8: 347-353.
- Corley, R.H.V. and Tinker, P.B. 2003. The Oil Palm, 4<sup>th</sup> Edn. Chichester: Wiley Blackwell Publishing.
- Corley, R.H.V. and Tinker, P.B. 2016. The Oil Palm, 5<sup>th</sup> Edn. Chichester: Wiley Blackwell Publishing.
- Corley, R.H.V., Gray, B.S. and Ng, S.K. 1971a. Productivity of the oil palm (Elaeis guineensis Jacq.) in Malaysia. Experimental Agriculture, 7: 129-136.
- Corley, R.H.V., Hardon, J.J. and Tan, G.Y. 1971b. Analysis of growth of the oil palm (*Elaeis guineensis* Jacq.) I. Estimation of growth parameters and application in breeding. Euphytica, 20: 307-315.
- Corley, R.H.V., Ng, M. and Donough, C.R. 1995. Effects of defoliation on sex differentiation in oil palm clones. Experimental Agriculture, 31: 177-189.
- Cui, J., Davanture, M., Zivy, M., Lamade, E. and Tcherkez, G. 2019. Metabolic responses to potassium availability and waterlogging reshape respiration and carbon use efficiency in oil palm. New Phytologist, 223: 310-322.
- Cunningham, S.C. 2005. Photosynthetic responses to vapour pressure deficit in temperature and tropical evergreen rainforest trees of Australia. Oecologia, 142: 521-528.
- Cramer, W., Bondeau, A., Woodward, F,I., Prentice, I.C., Betts, R.A., Brovkin, V., Cox, P.M., Fisher, V., Foley, J.A., Friend, A.D. *et al.* 2001. Global response of terrestrial ecosystem structure and function to CO<sub>2</sub> and climate change: results from six dynamic global vegetation models. Global Change Biology, 7: 357-373

- Cronquist, A. 1981. An integrated system of classification of flowering plants. Columbia University Press, New York.
- Dai, A.G. 2013. Increasing drought under global warming in observations and models. Nature Climate Change, 3: 52–58.
- de Langre, E. 2008. Effects of wind on plants. Annual Review of Fluid Mechanics, 40: 141-168.
- de Wit, C.T. and Goudriaan, J. 1978. Simulation of ecological processes. Simulation Monographs, Pudoc, Wagenungen, The Netherlands.
- de Wit, C.T. and Penning de Vries, F.W.T. 1982. L'Analyse des systemes de production primaire [Analysis of primary production systems]. In: F. W. T. Penning de Vries and M. A. Djiteye (Eds.), Une etude des sols, des vegetations et de l'exploitation de cette ressource naturelle [A study of the soils, vegetations and exploitation of this natural resource], Agricultural Research Reports 918. Wageningen, The Netherlands: Pudoc, pp. 20–27.
- de Wit, C.T., Goudriaan, J. and Van Laar, H.H. 1978. Simulation of assimilation, respiration and transpiration of crops. Pudoc, Wageningen, The Netherlands.
- Delzon, S., Bosc, A., Cantet, L. and Loustau, D. 2005. Variation of the photosynthetic capacity across a chronosequence of maritime pine correlates with needle phosphorus concentration. Annals of Forest Science, 62: 537-543.
- Desa, M.N. and Rakhecha, P.R. 2006. Deriving the highest persisting monthly 24-hour dew points in Malaysia for the estimation of PMP. In: Climate Variability and Change–Hydrological Impacts (Proceedings of the Fifth FRIEND World Conference, Havana, Cuba, November 2006), IAHS Publication, 308, 2006.
- Desmarest, J. 1967. Essai d'irrigation sur jeune palmeraie industrielle. Oléagineux, 22: 441-447.
- Di Marco, G., Manes, F., Tricoli, D. and Vitale, E. 1990. Fluorescence parameters measured concurrently with net photosynthesis to investigate chloroplastic CO<sub>2</sub> concentration in leaves of *Quercus ilex* L. Journal of Plant Physiology, 136: 538-543.
- Díaz-Espejo, A., Walcroft, A.S., Fernandez, J.E., Hafidi, B., Palomo, M.J. and Girón, I.F. 2006. Modeling photosynthesis in olive leaves under drought conditions. Tree Physiology, 26: 1445-1456.

- Dimas, F.A., Gilani, S.I.H. and Aris, M.S. 2011. Hourly solar radiation estimation using ambient temperature and relative humidity data. International Journal of Environmental Science and Development, 2: 188-93.
- Dolmat, M., Hamdan, A.B., Zulkifli, H. and Ahmad Tarmizi, M. 1996. Fertiliser requirement of oil palm on peat—An update. In: Proceeding of the 1996 PORIM International Palm Oil Congress, Agriculture Conference, (Ed. by Ariffin, D., Mohd Basri, W., Mohd Tayeb, D., Paranjothy, K., Rajanaidu, N., Cheah, S.C., Chang, K.W., Ravigadevi, S.), pp. 153-169, Palm Oil Research Institure of Malaysia, Kuala Lumpur.
- Donough, C.R., Cahyo, A., Oberthür, T., Ruli, W., Gerendas, J. and Gatot, A.R. 2014. Improving nutrient management of oil palms on sandy soils in Kalimantan using the 4R concept of IPNI. In: Proceedings of the International Oil Palm Conference, Bali, Indonesia, 17–19 June 2014.
- Dufrêne, E. 1989. Photosynthese, consommation en eau et modelisation de la production chez le palmier a huile (*Elaeis guineensis* Jacq.). Thesis, University of Paris-Sud, Orsay.
- Dufrêne, E. and Saugier, B. 1993. Gas exchange of oil palm in relation to light, vapour pressure deficit, temperature and leaf age. Functional Ecology, 7: 97-104.
- Dufrêne, E., Ochs, R. and Saugier, B. 1990. Oil palm photosynthesis and productivity linked to climatic factors. Oléagineux, 45: 345-55.
- Dufrêne, E., Dubos, B., Rey, H., Quencez, P. and Saugier, B. 1992. Changes in evapotranspiration from an oil palm stand (*Elaeis guineensis* Jacq.) exposed to seasonal soil water deficits. Acta Ecologia, 13: 299-314.
- Duursma R.A. 2015. Plantecophys An R Package for Analysing and Modelling Leaf Gas Exchange Data. PLoS ONE, 10 (11): e0143346.
- Ehleringer, J. and Pearcy, R.W. 1983. Variation in Quantum Yield for CO2 Uptake among C3 and C4 Plants. Plant Physiology, 73 (3): 555-559.
- Ephrath, J.E., Goudriaan, J. and Marani, A. 1996. Modelling diurnal patterns of air temperature, radiation, wind speed and relative humidity by equations from daily characteristics. Agricultural Systems, 51: 377-93.
- Ethier, G.J. and Livingston, N.J. 2004. On the need to incorporate sensitivity to CO<sub>2</sub> transfer conductance into the Farquhar-von Caemmerer-Berry leaf photosynthesis model. Plant, Cell & Environment, 27: 137-153.
- Fan, Y., Roupsard, O., Bernoux, M., Le Maire, G., Panferov, O., Kotowska, M. M. and Knohl, A. 2015. A sub-canopy structure for simulating oil palm in the

Community Land Model (CLM-Palm): phenology, allocation and yield. Geoscientific Model Development, 8: 3785-3800.

- Farahani, H.J. and Ahuja, L.R. 1996. Evapotranspiration modeling of partial canopy/residue-covered fields. Transactions of the ASAE, 39: 2051-2064.
- Farahani, H.J. and Bausch, W.C. 1995. Performance of evapotranspiration models for maize: Bare soil to closed canopy. Transactions of the ASAE, 38(4): 1049-1059.
- Farmanta, Y. 2014. Analyses of rainfall interception in immature and mature oil palm plantation In: Proceedings of International Seminar on the Land Reclamation Technology for Sustainable Land Use (IS-LRT4SLU), (Ed. by Margarettha, M.P., Wiwaha Anas, S., Aras Melin, S.P.), pp. 126-133, Jambi, Indonesia.
- Farquhar, G.D. and Wong, S.C. 1984. An empirical model of stomatal conductance. Australia Journal of Plant Physiology, 11: 191-210.
- Farquhar, G.D., von Caemmerer, S. and Berry, J.A. 1980. A biochemical model of photosynthetic CO<sub>2</sub> assimilation in leaves of C<sub>3</sub> species. Planta, 149: 78-90.
- Ferwerda, J.D. 1977. Oil palm. In: Ecophysiology of tropical crops (Ed. by P.d.T. Alvim and T.T. Kozlowski), pp. 351-383, Academic Press, London.
- Fisher, J.B., Debiase, T.A., Qi, Y., Xu, M. and Goldstein, A.H. 2005. Evapotranspiration models compared on a Sierra Nevada forest ecosystem. Environmental Modelling & Software, 20: 783-796.
- Floyd, R.B. and Braddock, R.D. 1984. A simple method for fitting average diurnal temperature curves. Agricultural and Forest Meteorology, 32: 107-119.
- Foong, S.F. 1982. An improved weather-based model for estimating oil palm fruit yield. In: The oil palm in agriculture in the eighties, Vol. 1 (Ed. by Pushparajah, E. and Chew P.S.), pp. 327-350, Incorporated Society of Planters, Kuala Lumpur.
- Foong, S.F. 1993. Potential evapotranspiration, potential yield and leaching losss of oil palm. In: Proceedings of the 1991 PORIM International Palm Oil Conference, Agriculture, (Ed. by Basiron, Y. *et al.*), pp. 105-119, Palm Oil Research Institute of Malaysia, Kuala Lumpur.
- Foong, F.S. 1999. Impact of moisture on potential evapotranspiration, growth and yield of oil palm. In: Proceedings of the 1999 PORIM International Palm Oil Conference, Agriculture, (Ed. By Arifin, D., Chan, K.W. and Sharifah, S.R.S.A.), pp. 265-287, Palm Oil Research Institute of Malaysia, Kuala Lumpur.

- Foster, H.L. and Prabowo, N.E. 1996. Yield response of oil palm to P fertilisers on different soils in North Sumatra. Paper presented at Conf. 'Sustainability of oil palm plantations-agronomic and environmental perspectives', International Society of Oil Palm, Kuala Lumpur.
- Foster, H.L. and Prabowo, N.E. 2002. Overcoming the limitations of foliar diagnosis in oil palm. In: Proceedings of the 2002 International Oil Palm Conference, Bali, Indonesia, 8–12 July 2002; Indonesian Oil Palm Research Institute: Sumatera Utara, Indonesia.
- Foster, H.L., Ghazali, M.Z. and Tayeb Dolmat, M. 1984. The available water holding capacity of oil palm soils in Malaysia. Palm Oil Reseach Institute of Malaysia Bulletin, 8: 1-9.
- Gardiner, B., Berryd, P. and Mouliae, B. 2016. Review: Wind impacts on plant growth, mechanics and damage. Plant Science, 245: 94-118.
- Genty, B., Briantais, J. and Baker, N.R. 1989. The relationship between the quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. Biochimica et Biophysica Acta, 990: 87-92.
- Germer, J. and Sauerborn, J. 2004. Solar radiation below the oil palm (*Elaeis guineensis* Jacq.) canopy and its impact on the undergrowth species composition. Planter, 80: 13-27.
- Gerritsma, W. 1988. Simulation of oil palm yield. Report. Department of Theoretical Production Ecology. Wageningen, The Netherlands: Wageningen Agricultural University.
- Gerritsma, W. and Soebagyo, F.X. 1999. An analysis of the growth of leaf area of oil palms in Indonesia. Experimental Agriculture, 35: 293-308.
- Gerritsma, W. and Wessel, M. 1997. Oil palm: domestication achieved? Netherlands Journal of Agricultural Science, 45: 463-475.
- Goh, K.J. 2000. Climatic requirements of the oil palm for high yields. In: Managing oil palm for high yields: agronomic principles (Ed. by K.J. Goh), pp. 1-17, Malaysian Society of Soil Science and Param Agricultural Surveys, Kuala Lumpur.
- Goh, K.J., Chew, P.S. and Kee, K.K. 1994. K Nutrition for Mature Oil Palm in Malaysia, IPI Research Topics No.17; International Potash Institute: Basel, Switzerland.
- Goh, K.J., Chew, P.S., Teoh, K.C. 1997a. Dry matter production and nutrient budget in the mature oil palm (*Elaeis guineensis* Jacq.) agroecosystem: III N budget. AAR Confidential Report, 3: 39.

- Goh, K.J., Chew, P.S., Teoh, K.C. 1997b. Dry matter production and nutrient budget in the mature oil palm (*Elaeis guineensis* Jacq.) agroecosystem: IV P budget. AAR Confidential Report, 4: 30.
- Goh, K., Gan, H., Kee, K., Chew, P., Teoh, K. 2007. Boron Nutrition and Boron Application in Crops. In: Advances in Plant and Animal Boron Nutrition, (Ed. by Xu, F.), Springer: Dordrecht, The Netherlands.
- Gong, X.Y., Tcherkez, G., Wenig, J., Schäufele, R. and Schnyder, H. 2018. Determination of leaf respiration in the light: comparison between an isotopic disequilibrium method and the Laisk method. New Phytologist, 218: 1371-1382.
- Goudriaan, J. 1977. Crop micrometeorology: a simulation study. Simulation Monograph. Pudoc, Wageningen.
- Goudriaan, J. and van Laar, H.H. 1978. Calculation of daily totals of the gross CO2 assimilation of leaf canopies. Netherlands Journal of Agricultural Science, 26: 373-382.
- Goudriaan, J. and van Laar, H.H. 1994. Modeling potential crop growth processes. A textbook with exercise. Current issues in production ecology. Netherlands, Kluwer Academic.
- Gray, B.S. 1969. A study of the influence of genetic, agronomic and environmental factors on the growth, flowering and bunch production of the oil palm on the West coast of West Malaysia. Ph.D. Thesis, University of Aberdeen.
- Grossiord, C., Buckley, T.N., Cernusak, L.A., Novick, K.A., Poulter, B., Siegwolf, R.T.W., Sperry, J.S. and McDowell, N.G. 2020. Plant responses to rising vapor pressure deficit. New Phytologist, 226: 1550-1566.
- Gu, L.H. and Sun, Y. 2014. Artefactual responses of mesophyll conductance to CO<sub>2</sub> and irradiance estimated with the variable *J* and online isotope discrimination methods. Plant, Cell & Environment, 37: 1231-1249.
- Gueymard, C.A. 2014. Impact of on-site atmospheric water vapor estimation methods on the accuracy of local solar irradiance predictions. Solar Energy, 101: 74-82.
- Gurmit, S. 1988. Zinc nutrition of oil palms on peat soils. In: Proc. 1987 Int. Oil Palm Conf. Progress and prospects (Ed. by A. Halim Hassan *et al.*), pp. 321-328, Palm Oil Research Institute of Malaysia, Kuala Lumpur.
- Han, Q., Kawasaki, T., Nakano, T. and Chiba, Y. 2008. Leaf-age effects on seasonal variability in photosynthetic parameters and its relationships with

leaf mass per area and leaf nitrogen concentration within a *Pinus densiflora* crown. Tree Physiology, 28: 551-558.

- Hansen, F.V. 1993. Surface roughness lengths. ARL Technical Report, U.S. Army, White Sands Missile Range, NM 88002-5501.
- Hanson, D.T., Stutz, S.S. and Boyer, J.S. 2016. Why small fluxes matter: the case and approaches for improving measurements of photosynthesis and (photo) respiration. Journal of Experimental Botany, 67: 3027-3039.
- Hardon, J.J., Williams C.N. and Watson, I. 1969. Leaf area and yield in the oil palm in Malaya. Experimental Agriculture, 5: 25-32.
- Harley, P.C. and Sharkey, T.D. 1991. An improved model of C<sub>3</sub> photosynthesis at high CO<sub>2</sub>: reversed O<sub>2</sub> sensitivity explained by lack of glycerate reentry into the chloroplast. Photosynthesis Research, 27: 169-178.
- Harley, P.C., Weber, J.A. and Gates, D.M. 1985. Interactive effects of light, leaf temperature, CO<sub>2</sub> and O<sub>2</sub> on photosynthesis in soybean. Planta, 165: 249-263.
- Harley, P.C., Loreto, F., Di Marco, G. and Sharkey, T.D. 1992a. Theoretical considerations when estimating the mesophyll conductance to CO<sub>2</sub> flux by analysis of the response of photosynthesis to CO<sub>2</sub>. Plant Physiology, 98: 1429-1436.
- Harley, P.C., Thomas, R.B., Reynolds, J.F. and Strain, B.R. 1992b. Modelling photosynthesis of cotton grown in elevated CO<sub>2</sub>. Plant, Cell & Environment, 15: 271-282.

Hartley, C.W.S. 1988. The oil palm. 3rd edition, Longman, London/New York.

- Hayawin, Z.N., Astimar, A.A., Nur Rashyeda, R., Nor Faizah, J., Idris, J., Ravi Menon, N., Syirat Z.B., Ropandi, M. and Hamzah, A. 2016. Influence of frond, stem and roots of oil palm seedlings in vermicompost from oil palm biomass. Journal of Oil Palm Research, 28(4): 479-484.
- Henry, P. 1958. Croissance et développement chez *Elaeis guineensis* Jacq. de la germination a la première floraison. Revue générale de botanique, 66: 5-34.
- Henson, I.E. 1989. Modelling Gas Exchange, Yield and Conversion Efficiency. Workshop on Productivity of Oil Palm. Bangi, Malaysia: Palm Oil Research Institute of Malaysia.
- Henson, I.E. 1991. Age-related changes in stomatal and photosynthetic characteristics of leaves of oil palm (*Elaeis guineensis* Jacq.). Elaeis, 3: 336-348.

- Henson, I.E. 1992. Carbon assimilation, respiration and productivity of young oil palm (*Elaeis guineensis*). Elaeis, 4: 51-59.
- Henson, I.E. 1995a. Photosynthesis, dry matter production and yield of oil palm under light-limiting conditions. In: Proceedings of the 1993 PORIM International Palm Oil Congress - Update and Vision (Agriculture), (Ed. by Jalani, S., Ariffin, D., Rajanaidu, N., Mohd Tayeb, D., Paranjothy, K., Mohd Basri, W., Henson, I.E. and Chan, C.K.), pp. 525-541, Palm Oil Research Institute of Malaysia, Kuala Lumpur.
- Henson, I.E. 1995b. Carbon assimilation, water use and energy balance of an oil palm plantation assessed using micrometeorological techniques. In: Proceedings of the 1993 PORIM International Palm Oil Congress - Update and Vision, Agriculture, (Ed. by Jalani, S., Ariffin, D., Rajanaidu, N., Mohd Tayeb, D., Paranjothy, K., Mohd Basri, W., Henson, I.E. and Chan, C.K.), pp. 137-158, Palm Oil Research Institute of Malaysia, Kuala Lumpur.
- Henson, I.E. 1998. Notes on oil palm productivity. II. An empirical model of canopy photosynthesis based on radiation and atmospheric vapour pressure deficit. Journal of Oil Palm Research, 10: 25-28.
- Henson, I.E. 2000. Modelling the effects of 'haze' on oil palm productivity and yield. Journal of Oil Palm Research, 12: 123-34.
- Henson, I.E. 2004. Estimating maintenance respiration of oil palm. Oil Palm Bulletin, 48: 1-10.
- Henson, I.E. 2005. OPRODSIM, a versatile, mechanistic simulation model of oil palm dry matter production and yield. In: Proceedings of the PIPOC 2005: agriculture, biotechnology and sustainability, pp. 801–832, Malaysian Palm Oil Board, Kuala Lumpur.
- Henson, I.E. 2006a. Modelling vegetative dry matter production of oil palm. Oil Palm Bulletin, 52: 25-47.
- Henson, I.E. 2006b. OPFLSIM3 An improved oil palm seasonal flowering and yield simulation model. Oil Palm Bulletin, 53: 1-24.
- Henson, I.E. 2007. Modelling the effects of physiological and morphological characters on oil palm growth and productivity. Oil Palm Bulletin, 54: 1-26.
- Henson, I.E. 2009. Modelling dry matter production, partitioning and yield of oil palm. OPRODSIM: A mechanistic simulation model for teaching and research. Technical manual and users' guide. Kuala Lumpur, Malaysia: Malaysian Palm Oil Board.

- Henson, I.E. and Chai, S.H. 1997. Analysis of oil palm productivity. II. Biomass, distribution, productivity and turnover of the root system. Elaeis, 9: 78-92.
- Henson, I.E. and Chang, K.C. 1990. Evidence for water as a factor limiting performance of field palms in West Malaysia. In: Proceedings of 1989 International Palm Oil Development Conference, Agriculture, (Ed. by Jalani, B.S. *et al.*), pp. 487-498, Palm Oil Research Institute of Malaysia, Kuala Lumpur.
- Henson, I.E. and Chang, K.C. 2000. Oil palm productivity and its component processes. In: Advances in oil palm research, Vol. I (Ed. by Basiron, Y., Jalani, B.S. & Chan, K.W.), Malaysian Palm Oil Board, Kuala Lumpur, pp.97-145.
- Henson, I.E. and Chang, K.C. 2007. Modelling oil palm nutrient demand, nutrient turnover and nutrient balance. MPOB Technology, 30: 1-66.
- Henson, I.E. and Harun, M.H. 2005a. Carbon dioxide enrichment in oil palm canopies and its possible influence on photosynthesis. Oil Palm Bulletin, 51: 10-19.
- Henson, I.E. and Harun, M.H. 2005b. The influence of climatic conditions on gas and energy exchanges above a young oil palm stand in North Kedah, Malaysia. Journal of Oil Palm Research, 17: 73-91.
- Henson, I.E. and Mohd Tayeb, D. 2003. Physiological analysis of an oil palm density trial on a peat soil. Journal of Oil Palm Research, 15: 1-27.
- Henson, I.E., Chang, K.C. and Harun M.H. 1994. Physiology. In: Palm Oil Research Institute of Malaysia Scientific Report, 7: 204-265.
- Henson, I.E., Chang, K.C., Mustakim, S.N.A, Chai, S.H., Hasnuddin, M.Y. and Zakaria, A. 1999. The oil palm trunk as a carbohydrate reserve. Journal of Oil Palm Research, 11: 98-113.
- Hoffmann, M.P., Castaneda Vera, A., van Wijk, M.T., Giller, K.E., Oberthür, T., Donough, C. and Whitbreadand, A.M. 2014. Simulating potential growth and yield of oil palm (*Elaeis guineensis*) with PALMSIM: Model description, evaluation and application. Agricultural Systems, 131: 1-10.
- Hong, T.K. and Corley, R.H.V. 1976. Leaf temperature and photosynthesis of a tropical C3 plant, Elaeis guineensis. Malaysia Agriculture Reserach and Development Institute Research Bulletin, 4(1): 16-20.
- Hubbard, R.M., Bond, B.J. and Ryan, M.G. 1999. Evidence that hydraulic conductance limits photosynthesis in old *Pinus ponderosa* trees. Tree Physiology, 19: 165-172.

- Huth, N.I., Banabas, M., Nelson, P.N. and Webb, M.J. 2014. Development of an oil palm cropping systems model: lessons learned and future directions. Environmental Modelling and Software, 62: 411-419.
- Ibrahim, M.H., Jaafar, H.Z.E., Harun, M.H. and Yusop, M.R. 2010. Changes in growth and photosynthetic patterns of oil palm (*Elaeis guineensis* Jacq.) seedlings exposed to short-term CO<sub>2</sub> enrichment in a closed top chamber. Acta Physiologiae Plantarum, 32: 305-313.
- IPCC, 2018. Global Warming of 1.5°C. An IPCC Special Report on the impacts of global warming of 1.5°C above pre-industrial levels and related global greenhouse gas emission pathways, in the context of strengthening the global response to the threat of climate change, sustainable development, and efforts to eradicate poverty (Ed. by Masson-Delmotte, V., P. Zhai, H.O. Pörtner, D. Roberts, J. Skea, P.R. Shukla, A. Pirani, W. Moufouma-Okia, C. Péan, R. Pidcock, S. Connors, J.B.R. Matthews, Y. Chen, X. Zhou, M.I. Gomis, E. Lonnoy, T. Maycock, M. Tignor, and T. Waterfield). Geneva, Switzerland.
- Iritz, Z., Anders, L., Martti, H., Achim, G. and Kellner, E. 1999. Test of a modified Shuttleworth-Wallace estimate of boreal forest evaporation. Agricultural and Forest Meteorology. 98-99
- Jaafar, H. Z. and Ibrahim, M. H. 2012. Photosynthesis and quantum yield of oil palm juveniles to elevated carbon dioxide. In: Advances Photosynthesis Fundamental Aspects, (Ed. by Najafpour, M.M.), InTechPubl, Rijeka, Croatia, 321-340.
- Jacquemard, J.C. 1979. Contribution to the study of the height growth of the stems of *Elaeis guineensis* Jacq. Study of the L2T x D10D cross. Oléagineux, 34: 492-497.

Jacquemard, J.C. 1998. Oil palm. Macmillan Education Ltd, London.

- Jacquemard, J.C. and Baudouin, L. 1988. Contribution to the study of oil palm vertical growth – a descriptive model. In: Proceedings of the 1987 International Oil Palm/Palm Oil Conference, Agriulture, (Ed. by Abdul Halim, H., Chew, P.S., Wood, B.J. & Pushparajah, E.), Palm Oil Research Institute of Malaysia, Bangi, pp. 657-665.
- Jarvis, P.G. 1976. The interpretation of the variations in leaf water potential and stomatal conductance found in canopies in the field. Philosophical Transactions of the Royal Society of London, 273(927): 593-610.
- John, M.K., Chuah, H.H. and Neufeld, J.H. 1975. Application of improved azomethine-H method to the determination of boron in soils and plants. Analytical Letter, 8: 559-568.

- Jourdan, C. and Rey, H. 1997. Architecture and development of the oil palm (*Elaeis guineensis* Jacq.) root system. Plant and Soil, 189: 33-48.
- June, T., Evans, J.R. and Farquhar, G.D. 2004. A simple new equation for the reversible temperature dependence of photosynthetic electron transport: a study on soybean leaf. Functional Plant Biology, 31: 275-283.
- Kallarackal, J. 1996. Water relations and photosynthesis of the oil palm in Peninsular India. KFRI Research Report 110. Kerala Forest Research Institute, Peechi, Thrissur.
- Kallarackal, J., Jeyakumar, P. and George, S.J. 2004. Water use of irrigated oil palm at three different arid locations in Peninsular India. Journal of Oil Palm Research, 16: 45-53.
- Kaplanis, S. and Kaplani, E. 2007. A model to predict expected mean and stochastic hourly global solar radiation values. Renewable Energy, 32:1414–25.
- Kee, K.K. and Chew, P.S. 1993. Oil palm responses to nitrogen and drip irrigation in a wet monsoonal climate in Peninsular Malaysia. In: Proc. 1991 Int. Palm Oil Conf. Agriculture (Ed. By Y. Basiron *et al.*), pp. 321-335, Palm Oil Research Institute of Malaysia, Kuala Lumpur.
- Kee, K.K. and Chew, P.S. 1996. Nutrient loss through surface runoff and soil erosion implications for improved fertiliser efficiency in mature oil palms.
  In: Proc. PORIM Int. Oil Palm Congr., pp. 153–169, Palm Oil Research Institute of Malaysia, Kuala Lumpur.
- Keupp, L., Pollinger, F., and Paeth, H. 2017. Assessment of future ENSO changes in a CMIP3/CMIP5 multi-model and multi-index framework. International Journal of Climatology, 37: 3439-3451.
- Khalid, H., Zin, Z.Z. and Anderson, J.M. 1999a. Quantification of oil palm biomass and nutrient value in a mature plantation. I. Above-ground biomass. Journal of Oil Palm Research, 11: 23-32.
- Khalid, H., Zin, Z.Z. and Anderson, J.M. 1999b. Quantification of oil palm biomass and nutrient value in a mature plantation. II. Belowground biomass. Journal of Oil Palm Research, 11(1): 63-71.
- Khalid, H., Zin, Z.Z. and Anderson, J.M. 2000. Decomposition processes and nutrient release patterns of oil palm residues. Journal of Oil Palm Research, 12(1): 46-63.

- Khatib, T., Mohamed, A., Sopian, K. and Mahmoud, M. 2012. Solar Energy Prediction for Malaysia Using Artificial Neural Networks. International Journal of Photoenergy, 1-16.
- Kimball, B.A. and Bellamy, L.A. 1986. Generation of diurnal solar radiation, temperature, and humidity patterns. Energy in Agriculture, 5(3): 185-197.
- Koh, L. P. and Ghazoul, J. 2008. Biofuels, bodiversity, and people: Understanding the conflicts and finding opportunities. Biological Conservation, 141: 2450-2460.
- Kositsup, B., Kasemsap, P., Thanisawanyangkura, S., Chairungsee, N., Satakhun, D., Teerawatanasuk, K., Ameglio, T. and Thaler, P. 2010. Effect of leaf age and position on light-saturated CO<sub>2</sub> assimilation rate, photosynthetic capacity, and stomatal conductance in rubber trees. Photosynthetica, 48: 67-78.
- Kropff, M.J. 1993. Mechanisms of competition for light. In: Modelling crop-weed interactions. (Ed. by Kropff, M.J. and van Laar, H.H.), pp. 33-61. CAB International (in association with International Rice Research Institute), Wallingford.
- Kumar, S. and Ranjith, A.M. 2015. Studies on inflorescence production and pollination in oil palm. Progressive Horticulture, 47: 194.
- Kusnu, M., Siahaan, M.M. and Poeloengan, Z. 1996. Effects of N, P, K, and Mg fertilizer on the growth and yield of oil palm on Typic Paleudult. In: Sustainability of Oil Palm Plantations. Agronomic and Environmental Perspectives. Kuala Lumpur, 27–28 September. ISOPA, PORIM.
- Kustas, W.P. and Daughtry, C.S.T. 1990. Estimation of the soil heat flux/net radiation ratio from spectral data. Agricultural and Forest Meteorology, 49: 205-223.
- Kustas, W.P. and Norman, J.M. 1999. Evaluation of soil and vegetation heat flux predictions using a simple two-source model with radiometric temperatures for partial canopy cover. Agricultural and Forest Meteorology, 94: 13-29.
- Kwan, B.K.W. 1994. The effect of planting density on the first fifteen years of growth and yield of oil palm in Sabah. Technical Bulletin No. 11. Department of Agriculture, Sabah.
- Kwan, M.S., Tangang, F. and Liew, J. 2013. Projected changes of future climate extremes in Malaysia. Sains Malaysiana 42 (8): 1051-1059.
- Laisk, A. 1977. Kinetics of Photosynthesis and Photorespiration in C<sub>3</sub> Plants. Nauka, Moscow, Russia.

- Laisk, A. and Loreto, F. 1996. Determining photosynthetic parameters from leaf CO<sub>2</sub> exchange and chlorophyll fluorescence (ribulose-1,5-bisphosphate carboxylase/ oxygenase specificity factor, dark respiration in the light, excitation distribution between photosystems, alternative electron transport rate, and mesophyll diffusion resistance. Plant Physiology, 110: 903-912.
- Lamade, E., Djegui, N. and Leterme, P. 1996. Estimation of carbon allocation to the roots from soil respiration measurements of oil palm. Plant and Soil, 181: 329-339.
- Law, B.E, Sun, O.J., Campbell, J., Van Tuyl, S. and Thornton, P.E. 2003. Changes in carbon storage and fluxes in a chronosequence of ponderosa pine. Global Change Biology, 9: 510-524.
- Lee, C.H. 1999. Yield potential of Golden Hope DxP oil palm planting materials. In: Proceedings of the 1996 Seminar Sourcing of oil palm planting materials for local and overseas joint ventures (Ed. by Rajanaidu, N. & Jalani, B.S.), Palm Oil Research Institute of Malaysia, Kuala Lumpur, pp. 106-117.
- Legros, S., Mialet-Serra, I., Caliman, J.P., Clement-Vidal, A., Siregar, F.A., Widiastuti, L., Jourdan, C. and Dingkuhn, M. 2006. Carbohydrates reserves in 9 years old oil palm: Nature, distribution and seasonal changes. In: Proceedings of the International Oil Palm Conference on Optimum Use of Resources: Challenges and Opportunities for Sustainable Oil Palm Development, Bali, Indonesia, 19–23 June 2006.
- Lewis, K., Rumpang, E., Kho, L.K., McCalmont1, J., Teh, Y.A., Gallego-Sala, A. and Hill, T.C. 2020. An assessment of oil palm plantation aboveground biomass stocks on tropical peat using destructive and non-destructive methods. Scientific Report, 10: 2230.
- Li, R., Reddy, V.A., Jin, J., Rajan, C., Wang, Q., Yue, G., Lim, C.H., Chua, N.H., Ye, J. and Sarojam, R. 2017. Comparative transcriptome analysis of oil palm flowers reveals an EAR-motif-containing R2R3-MYB that modulates phenylpropene biosynthesis. BMC Plant Biology, 17: 219.
- Liau, S.S. and Ahmad Alwi, 1995. Defoliation and crop loss in young oil palm. In: Proceedings of the 1993 PORIM International Palm Oil Conference, Agriculture, (Ed. by Jalani, B.S. *et al.*), Oil Palm Research Institure of Malaysia, Kuala Lumpur, pp. 408–427.
- Lim, K.H., Goh, K.J., Kee, K.K. and Henson, I.E. 2011. Climatic requirements of oil palm. In: Agronomic Principles and Practices of Oil Palm Cultivation, (Ed. by Goh, K.J., Chiu, S.B., Paramananthan, S.), pp. 3–48, Agricultural Crop Trust: Petaling Jaya, Malaysia.
- Lim, Y.K., Kovach, R.M., Pawson, S. and Vernieres, G. 2017. The 2015/16 El Niño event in context of the MERRA-2 reanalysis: a comparison of the tropical Pacific with 1982/83 and 1997/98. Journal of Climate, 30: 4819-4842.
- Lobell, D.B., Hammer, G.L., McLean, G., Messina, C., Roberts, M.J., Schlenker, W. 2013. The critical role of extreme heat for maize production in the United States. Nature Climate Change, 3: 497.
- Lobell, D.B., Schlenker, W., Costa-Roberts, J. 2011. Climate trends and global crop production since 1980. Science, 333: 616–620.
- Loh, J.L., Tangang, F., Juneng, L., David, H. and Lee, D.I. 2016. Projected rainfall and temperature changes over Malaysia at the end of the 21st century based on PRECIS modelling system. Asia-Pacific Journal of Atmospheric Sciences, 52: 191-208.
- Long, S.P. and Bernacchi, C.J. 2003. Gas exchange measurements, what can they tell us about the underlying limitations to photosynthesis? Procedures and sources of errors. Journal of Experimental Botany 54: 2393–2401.
- Loriaux, S.D., Avenson, T.J., Welles, J.M., McDermitt, D.K., Eckles, R.D., Riensche, B. and Genty, B. 2013. Closing in on maximum yield of chlorophyll fluorescence using a single multiphase flash of sub-saturating intensity. Plant, Cell & Environment, 36: 1755-1770.
- Maene, L.M., Thong, K.C., Ong, T.S. and Mokhtaruddin, A.M. 1979. Surface wash under mature oil palm. In: Proceeding of the Symposium of Water in Agriculture in Malaysia (Ed. by Pushparajah, E.), Malaysian Society of Soil Science, Kuala Lumpur, pp. 204-216.
- Manter, D.K. and Kerrigan, J. 2004. A/C<sub>i</sub> curve analysis across a range of woody plant species: influence of regression analysis parameters and mesophyll conductance. Journal of Experimental Botany, 55: 2581-2588.
- Massman, W.J. 1987. A comparative study of some mathematical models of the mean wind structure and aerodynamic drag of plant canoipies. Boundary-Layer Meteorology, 40: 179–197.
- Massman, W.J. 1997. An analytical one-dimensional model of momentum transfer by vegetation of arbitrary structure. Boundary-Layer Meteorology, 83: 407-421.
- Matejovic, I. 1993. Determination of carbon, hydrogen, and nitrogen in soils by automated elemental analysis (dry combustion method). Communication in Soil Science and Plant Analysis, 24: 17-18.

- Ma'rufah, U., June, T., Faqih, A., Ali, A.A., Stiegler, C. and Knohl, A. 2021. Implication of land use change to biogeophysical and biogeochemical processes in Jambi, Indonesia: Analysed using CLM5. Terrestrial, Atmospheric and Oceanic Sciences, 32(2): 203-215.
- McDowell, N.G., Allen, C.D., Anderson-Teixeira, K., Brando, P., Brienen, R., Chambers, J., Christoffersen, B., Davies, S., Doughty, C., Duque, A., et al. 2018. Drivers and mechanisms of tree mortality in moist tropical forests. New Phytologist, 219: 851–869.
- McDowell, N.G., Phillips, N., Lunch, C., Bond, B.J. and Ryan, M.G. 2002. An investigation of hydraulic limitation and compensation in large, old Douglas-fir trees. Tree Physiology, 22: 763-774.
- Medlyn, B.E., Berbigier, P., Clement, R., Grelle, A., Loustau, D., Linder, S., Wingate, L., Jarvis, P.G., Sigurdsson, B.D. and McMurtrie, R.E. 2005. Carbon balance of coniferous forests growing in contrasting climates: model-based analysis. Agricultural and Forest Meteorology 131: 97-124
- Meijide, A., Röll, A., Fan, Y., Herbst, M., Niu, FuRong., Tiedemann, F., June, T., Rauf, A., Hölscher, D. and Knohl, A. 2017. Controls of water and energy fluxes in oil palm plantations: Environmental variables and oil palm age. Agricultural and Forest Meteorology, 239: 71-85.

Miyazaki, T. 2005. Water flow in soils. CRC Press, Boca Raton, FL.

- Monteith, J.L. 1973. Principles of environmental physics. Edward Arnold, London.
- Moriasi, D.N., Arnold, J.G., Van Liew, M.W., Bingner, R.L., Harmel, R.D., Veith, T.L. 2007. Model evaluation guidelines for systematic quantification of accuracy in watershed simulations. Transaction of the American Society of Agricultural and Biological Engineers, 50: 885e900.
- Morice, C.P., Kennedy, J.J., Rayner, N.A., Winn, J.P., Hogan, E., Killick, R.E., Dunn, R.J.H., Osborn, T.J., Jones, P.D. and Simpson, I.R. 2021. An updated assessment of near-surface temperature change from 1850: the HadCRUT5 dataset. Journal of Geophysical Research (Atmospheres).
- Monzon, J.P., Jabloun, M., Cock, J., Caliman, J.P., Couëdel, A., Donough, C.R., Philip Ho, V.V., Lim, Y.L., Mathews, J., Oberthür, T., Prabowo, N.E., Edreira, J.I.R., Sidhu, M., Slingerland, M.A., Sugianto, H. and Grassini, P. 2022. Influence of weather and endogenous cycles on spatiotemporal yield variation in oil palm. Agricultural and Forest Meteorology, 314: 1-10.
- Monzon, J.P., Slingerland, M.A., Rahutomo, S., Agus, F., Oberthür, T., Andrade, J.F., Couëdel, A., Edreira, J.I.R., Hekman, W., van den Beuken, R., Hidayat,

F., Pradiko, I., Purwantomo, D.K.G., Donough, C.R., Sugianto, H., Lim, Y.L., Farrell, T. and Grassini, P. 2021. Fostering a climate-smart intensification for oil palm. Nature Sustainability, 4: 595–601.

- Moualeu-Ngangue, D.P., Chen, T.W. and Stützel, H. 2017. A new method to estimate photosynthetic parameters through net assimilation rateintercellular space CO<sub>2</sub> concentration (*A*-*C*<sub>i</sub>) curve and chlorophyll fluorescence measurements. New Phytologist, 213: 1543-1554.
- MPOB. 2021. Overview of the Malaysian oil palm industry 2020. Malaysia Palm Oil Board, Bangi, Malaysia.
- MPOB. 2022. Overview of the Malaysian oil palm industry 2021. Malaysia Palm Oil Board, Bangi, Malaysia.
- Najihah, T.S., Ibrahim, M.H., Mohd Zain, N.A., Nulit R. and Megat Wahab P.E. 2020. Activity of the oil palm seedlings exposed to a different rate of potassium fertilizer under water stress condition. AIMS Environmental Science, 7(1): 46-68.
- Nazeeb, M. 2000. Investigations on optimal planting density for oil palm (*Elaeis guineensis* Jacq.) in Peninsular Malaysia. PhD Thesis. University Malaya.
- Nazeeb. M., Loong, S.G., Goh, K.H. and Wood, B.J. 1989. Trials on planting oil palms at high initial density with later thinning. In: Proceedings of the 1989 PORIM International Palm Oil Development Conference, Agriculture, (Ed. by Jalani, S., Zawawi, Z., Paranjothy, K., Ariffin, D., Rajanaidu, N., Cheah, S.C., Basri, M., Henson, I.E. and Tayeb, M.), Palm Oil Research Institure of Malaysia, Kuala Lumpur, pp. 199-214.
- Nazeeb, M., Tang, M.K., Loong, S.G. and Barakbah, S.S. 2008. Variable density plantings for oil palms (*Elaies guineensis*) in Peninsular Malaysia. Journal of Oil Palm Research, 20: 61-90.
- Ng, S.K., Tan, Y.P., Chen, E. and Cheong, S.P. 1974. Nutritional complexes of oil palms planted on peat soil in Malaysia. II. Preliminary results of copper sulphate treatments. Oléagineux, 29: 445-456.
- Ng, S.K. and Thamboo, S. 1967. Nutrient contents of oil palms in Malaya. I. Nutrients required for reproduction: Fruit bunches and male inflorescence. The Malaysian Agriculrural Journal, 46: 3-45.
- Ng, S.K., Thamboo, S. and de Souza, P. 1968. Nutrient contents of oil palms in Malaya. II Nutrients in vegetative tissues. The Malaysian Agricultural Journal, 46: 332-391.

- Nelson, P.N., Banabas, M., Scotter, D.R. and Webb, M.J. 2006. Using soil water depletion to measure spatial distribution of root activity in oil palm (*Elaeis guineensis* Jacq.) plantations. Plant and Soil, 286: 109-121.
- NRE, 2015. Malaysia Biennial Update Report to the UNFCC. Ministry of Natural Resources and Environment Malaysia, Putrajaya, Malaysia.
- Norhayati, A., Norhazela, S. and Cheah, S.S. 2019. Impact of palm biomass removal during replanting on oil palm replants. Poster paper presented at the 2019 MPOB international palm oil congress and exhibition (PIPOC), 19-21 November 2019, Kuala Lumpur.
- Nouy, B., Baudouin, L., Djegui, N. and Omore, A. 1999. Oil palm under limiting water supply conditions. Plantations, Recherche, Développement, 6: 31-45.
- Novick, K.A., Ficklin, D.L., Stoy, P.C., Williams, C.A., Bohrer, G., Oishi, A.C., Papuga, S.A., Blanken, P.D., Noormets, A., Sulman, B.N., *et al.* 2016. The increasing importance of atmospheric demand for ecosystem water and carbon fluxes. Nature Climate Change, 6: 1023.
- Nugroho, B. 2018. Leaf gas exchange measurement under land use changes in Jambi, Indonesia. Dissertation, Georg-August-Universität Göttingen.
- Nur Amanina, S., Haniff Harun, M., Mohd Roslan, M.N. and Md Hasnudin, M.Y. 2011. Growth and photosynthesis of oil palm under elevated carbon dioxide. Paper presented at Int. Palm Oil Congr. 'Palm oil: fortifying and energising the World', Malaysian Palm Oil Board, Kuala Lumpur, 15–17 November.
- Ochs, R. and Daniel, C. 1976. Research on techniques adapted to dry regions. In: Oil palm research (Ed. by Corley, R.H.V., Hardon, J.J. & Wood, B.J.), Elsevier, Amsterdam, pp. 315-330.
- Oettli, P., Behera, S. K. and Yamagata, T. 2018. Climate based predictability of oil palm tree yield in Malaysia, Scientific Reports, 8: 2271.
- Oleson, K., Lawrence, D.M., Bonan, G.B., Drewniak, B., Huang, M., Koven, C. D., Levis, S., Li, F., Riley, W.J., Subin, Z.M., Swenson, S.C., Thornton, P.E. *et al.* 2013. Technical description of version 4.5 of the Community Land Model (CLM) (No. NCAR/TN-503+STR). NCAR Technical Note, Boulder, Colorado.
- Ollivier, J., Flori, A., Cochard, B., Amblard, P., Turnbull, N., Syahputra, I., Suryana, E., Lubis, Z., Surya, E.; Sihombing, E. and Gasselin, D.T. 2017. Genetic variation in nutrient uptake and nutrient use efficiency of oil palm. Journal of Plant Nutrition, 40: 558-573.

- Olivin, J. 1968. Etude pour la localisation d'un bloc industriel de palmiers à huile. II Les critèrie de jugement. Oléagineux, 23: 499-504.
- Olivin, J. 1986. Study for the siting of a commercial oil palm plantation. Oléagineux, 41: 113-118.
- Padfield, R., Hansen, S., Davies, Z.G., *et al.* 2019. Co-producing a research agenda for sustainable palm oil. Frontier in Forest and Global Change, 2: 13.
- Paine, C.E.T., Marthews, T.R., Vogt, D.R., Purves, D., Rees, M., Hector, A. and Turnbull, L.A. 2012. How to fit nonlinear plant growth models and calculate growth rates: An update for ecologists. Methods in Ecology and Evolution, 3: 245-256.
- Palat, T., Chayawat, N. and Corley, R.H.V. 2012. Maximising oil palm yield by high density planting and thinning. Planter, 88: 241-256.
- Palat, T., Chayawat, N., Clendon, J.H. and Corley, R.H.V. 2008. A review of 15 years of oil palm irrigation research in Southern Thailand. Planter, 84: 537-546.
- Paramananthan, S. 2000a. Soils of Malaysia Their Characteristics and Identification; Academy of Sciences: Kuala Lumpur, Malaysia, Volume I, pp. 436-437.
- Paramananthan, S. 2000b. Soil requirements of oil palm for high yields. In: Managing Oil Palm for High Yields: Agronomic Principles (Ed. by Goh, K.J.), Malaysian Society of Soil Science and Param Agricultural Surveys: Kuala Lumpur, Malaysia, pp. 18-38.
- Paramananthan, S. 2003a. Best practices for oil palm cultivation land selection and management. Planter, 79: 311-323.
- Paramananthan, S. 2003b. Peat soils of Malaysia: Their extent, characteristics, mapping and classification. Planter, 89: 737-757.
- Paterson, R.R.M., Kumar, L., Shabani, F., and Lima, N. 2017. World climate suitability projections to 2050 and 2100 for growing oil palm. The Journal of Agricultural Science, 155: 659-702.
- Paterson, R.R.M., Kumar, L., Taylor, S. and Lima, N. 2015. Future climate effects on suitability for growth of oil palms in Malaysia and Indonesia. Scientific Reports, 5: 14457.
- Peisker, M. and Apel, H. 2001. Inhibition by light of CO<sub>2</sub> evolution from dark respiration: comparison of two gas exchange methods. Photosynthesis Research, 70: 291-298.

- Penning de Vries, F.W.T. 1972. Respiration and growth. In: Crop processes in controlled environments (Ed. by Rees, A.R. *et al.*), Academic Press, London, pp. 327-346.
- Penning de Vries, F.W.T. 1975. The cost of maintenance processes in plant cells. Annals of Botany, 39: 77-92.
- Penning de Vries, F.W.T., van Laar, H.H. and Chardon, M.C.M. 1983. Bioenergetics of growth of seeds, fruits, and storage organs. In: Potential productivity of field crops under different environments. International Rice Research Institute, Los Baños Laguna, Philippines, pp. 37-59.
- Phillips, N., Bond, B.J., McDowell, N.G., Ryan, M.G. and Schauer, A. 2003. Leaf area compounds height-related hydraulic costs of water transport in Oregon White Oak trees. Functional Ecology, 17: 832-840.
- PNGOPRA. 2019. Annual Research Report. Papua New Guinea Oil Palm Research Association, Kimbe, Papua New Guinea.
- Pons, T.L. and Welschen, R.A. 2003. Midday depression of net photosynthesis in the tropical rainforest tree *Eperua grandiflora*: contributions of stomatal and internal conductances, respiration and Rubisco functioning. Tree Physiology, 23: 937-947.
- Pons, T.L., Flexas, J., von Caemmerer, S., Evans, J.R., Genty, B., Ribas-Carbo, M. and Brugnoli, E. 2009. Estimating mesophyll conductance to CO<sub>2</sub>: methodology, potential errors, and recommendations. Journal of Experimental Botany, 60: 2217-2234.
- Prabowo, N.E. and Foster, H.L. 1998. Variation in oil and kernel extraction rates of palms in North Sumatra due to nutritional and climatic factors. In: Proceeding of the 1998 International Oil Palm Conference, Commodity of the past, today and the future (Ed. by Jatmika, A. *et al.*), Indonesian Oil Palm Research Institute, Medan, pp. 275-286.
- Prabowo, N.E., Foster, H.L. and Silalahi, A.J. 2006. Recycling oil palm bunch nutrients. In: Proceedings of the Oil Palm Conference, Bali, Indonesia, 19–23 June 2006.
- Purvis, C. 1956. The root system of the oil palm: its distribution, morphology and anatomy. Journal of West Afica Institute for Oil Palm Research, 1(4): 61-82.
- R Core Team. 2013. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL http://www.R-project.org/.

- Rajanaidu, N. and Kushairi, A. 2006. Oil palm planting materials and their yield potential. In: Proceedings of the International Society of Oil Palm Breeders Symposium: Yield Potential in Oil Palm II, Phuket, Thailand, 27–28 November 2006.
- Rajaratnam, J.A. 1976. Micronutrients. In: Oil Palm Research. (Developments in Crop Science I), (Ed. by Corley, R.H.V., Hardon, J.J. and Wood, B.J.), Elsevier, Amsterdam, pp. 263-270.
- Ramos, H., Meschiatti, M., de Matos, R. and Blain, G. 2018. On the performance of three indices of agreement: An easy-to-use R-code for calculating the Willmott indices. Bragantia, 77(2): 394-403.
- Rankine, I.R. and Fairhurst, T.H. 1999. Field handbook oil palm series Vol. 2 Immature. Potash and Phosphate Institute, Singapore.
- Rao, V. and Law, I.H. 1998. The problem of poor fruit set in parts of East Malaysia. Planter, 74: 463-483.
- Rao, V., Rajanaidu, N., Kushairi, A. and Jalani, S. 1992. Density effect in the oil palm. In: Proceedings in the ISOP International Workshop of Yield Potential in the Oil Palm, International Society for Oil Palm Breeders and PORIM, Bangi, pp. 71-79.
- Rees, A.R. 1964. The apical organization and phyllotaxis of the oil palm. Annals of Botany, 28: 57-69.
- Rees, A.R. and Tinker, P.B. 1963. Dry matter production and nutrient content of plantation oil palms in Nigeria. I. Growth and dry-matter production. Plant and Soil, 19: 19-32.
- Rey, H., Quencez, P., Dufrêne, E. and Dubos, B. 1998. Oil palm water profiles and water supplies in Côte d'Ivoire. Plantations, Recherche, Développement, 5: 47-57.
- Rhebergen, T., Fairhurst, T., Zingore, S., Fisher, M., Oberthür, T. and Whitbread, A. 2016. Climate, soil and land-use based land suitability evaluation for oil palm production in Ghana. European Journal of Agronomy, 81: 1-14.
- Rival, A. 2017. Breeding the oil palm (*Elaeis guineensis* Jacq.) for climate change. Oilseeds and fats Crops and Lipids, 24(1) D017: 1-7.
- Robertson, G.W. and Foong, S.F. 1977. Weather-based yield forecasts for oil palm fresh fruit bunches. In: International developments in oil palm, (Ed. by Earp, D.A. and Newall, W.), Incorporated Society of Planters, Kuala Lumpur, pp. 695-709.

- Röll, A., Niu, F., Meijide, A., Hardanto, A., Hendrayanto, Knohl, A. and Hölscher, D. 2015. Transpiration in an oil palm landscape: effects of palm age. Biogeoscuences, 12: 5619-5633.
- Rosenzweig, C., Elliott, J., Deryng, D., Ruane, A.C., Müller, C., Arneth, A., Boote, K.J., Folberth, C., Glotter, M., Khabarov, N., Neumann, K., Piontek, F., Pugh, T.A., Schmid, E., Stehfest, E., Yang, H. and Jones, J.W. 2014. Assessing agricultural risks of climate change in the 21st century in a global gridded crop model intercomparison. Proceedings of the National Academy of Sciences, 111: 3268-3273.
- Ruer, P. 1967. Morphologie et anatomie du système radiculaire du palmier à huile. Oléagineux, 22: 595-599.
- Ruth, D.W. and Chant, R.E. 1976. The relationship of diffuse radiation to total radiation in Canada. Solar Energy, 18: 153-154.
- Saxton, K.E. and Rawls, W.J. 2006. Soil water characteristic estimates by texture and organic matter for hydrologic solutions. Soil Science Society of America Journal, 70: 1569-1578.
- Saxton, K.E., Rawls, W.J., Romberger, J.S. and Papendick, R.I. 1986. Estimating generalized soil water characteristics from texture. Transaction ASAE 50:1031–1035.
- Santanello, J.A. and Friedl, M.A. 2003. Diurnal covariation in soil heat flux and net radiation. Journal of Applied Meteorology, 42: 851-862.
- Seager, R., Hooks, A., Williams, A.P., Cook, B., Nakamura, J., Henderson, N. 2015. Climatology, variability, and trends in the U.S. vapor pressure deficit, an important fire-related meteorological quantity. Journal of Applied Meteorology and Climatology, 54: 1121–1141.
- Setyobudi, H., Lihanaswan, S. and Wanasuria, S. 1998. Iron deficiency in mature oil palms in Riau, Sumatra. In: Proceedings of the 1998 International Oil Palm Conference - Commodity of the past, today and the future (Ed. by Jatmika, A. *et al.*), Indonesian Oil Palm Research Institute, Medan, pp. 363-369
- SDPR, 2019. Annual Report. Sime Darby Plantation Research. Carey Island, Selangor, Malaysia.
- Sharkey, T.D. 2015. Commentary: what gas exchange data can tell us about photosynthesis. Plant, Cell & Environment, 39: 1161-1163.
- Sharkey, T.D., Stitt, M., Heineke, D., Gerhardt, R., Raschke, K. and Heldt, H.W. 1986. Limitation of photosynthesis by carbon metabolism. II O<sub>2</sub> insensitive

CO<sub>2</sub> uptake results from limitation of triose phosphate utilization. Plant Physiology, 81: 1123–1129.

- Shavalipour, A., Hakemzadeh, M.H., Sopian, K., Mohamed Haris, S. and Zaidi, S.H. 2013. New Formulation for the Estimation of Monthly Average Daily Solar Irradiation for the Tropics: A Case Study of Peninsular Malaysia. International Journal of Photoenergy, 1e174671–6.
- Shanmuganathan, S. and Narayanan, A. 2012. Modelling the climate change effects on Malaysia's oil palm yield. IEEE Symposium on E-learning, E-management and E-services (IS3E): 1-6.
- Shanmuganathan, S., Narayanan, A. and Mohamed, M. 2014. A hybrid approach to modelling the climate change effects on Malaysia's oil palm yield at the regional scale. In: Recent advances on soft computing and data mining (Ed. by Herawan, T., Ghazali, R., and Deris, M.M.), pp. 335–345, Cham, Switzerland: Springer.
- Shen, L. and Chen, Z. 2007. Critical review of the impact of tortuosity on diffusion. Chemical Engineering Science, 62: 3748-3755.
- Shuttleworth, W.J. and Wallace, J.S. 1985. Evaporation from sparse crops an energy combination theory. Quarterly Journal of the Royal Meteorological Society, 111: 839-855.
- SIRIM. 1980. SIRIM Recommended Methods for Plants Analysis, 1<sup>st</sup> ed.; SIRIM: Shah Alam, Malaysia.
- Siwar, C., Ahmed, F. and Begum, R.A. 2013. Climate change, agriculture and food security issues: Malaysian perspective. Journal of Food, Agriculture and Environment, 11(2): 1118-1123.
- Skillman, J.B. 2008. Quantum yield variation across the three pathways of photosynthesis: not yet out of dark. Journal of Experimental Botany, 59(7): 1647-1661.
- Smith, B.G. 1989. The effects of soil water and atmospheric vapour pressure deficit on stomatal behaviour and photosynthesis in the oil palm. Journal of Experimental Botany, 40: 647-651.
- Smith, V.C. and Ennos, A.R. 2003. The effects of air flow and stem flexure on the mechanical and hydraulic properties of the stems of sunflowers Helianthus annuus L. Journal of Experimental Botany, 54: 845-849.
- Sparnaaij, L.D. 1960. The analysis of bunch production in the oil palm. Journal of West Africa Institute for Oil Palm Research, 3: 109-180.

- Squire, G.R. 1984. Light interception, productivity and yield of oil palm. Internal report, Palm Oil Research Institute of Malaysia.
- Squire, G.R. 1990. The physiology of tropical crop production. CAB International, Wallingford.
- Squire, G.R. and Corley, R.H.V. 1987. Oil palm. In: Tree crop physiology (Ed. by M.R. Sethuraj and A.S. Raghavendra), pp.141-167, Elsevier Science Publishers, Amsterdam.
- Stewart, J.B. 1988. Modelling surface conductance of pine forest. Agricultural and Forest Meteorology, 43: 19-35.
- Stiegler, C., Meijide, A., Fan, Y., Ashween Ali, A., June, T. and Knohl, A. 2019. El Niño–Southern Oscillation (ENSO) event reduces CO<sub>2</sub> uptake of an Indonesian oil palm plantation. Biogeosciences, 16: 2873-2890.
- Sopian, K., Hj. Othman, M.Y. and Wirsat, A. 1995. The wind energy potential of Malaysia. Renewable Energy, 6(8): 1005-1016.
- Streck, N.A., Rosa, H.T., Walter, L.C., Silva, M.R. and Uhlmann, L.O. 2012. CO<sub>2</sub>response function of radiation use efficiency in rice for climate change scenarios. Pesquisa Agropecuária Brasileira, 47(7): 879-885.
- Subronto, M. and Pamin, K. 1991. Efisiensi pengalihan enerji pada tanaman kelapa sawit. Bulletin Perkebunan, 22: 33-49.
- Sun, Y., Gu, L., Dickinson, R.E., Pallardy, S.G., Baker, J., Cao, Y., DaMatta, F.M., Dong, X., Ellsworth, D., Van Goethem, D., Jensen, A.M., Law, B.E., Loos, R., Martins, S.C., Norby, R.J., Warren, J., Weston, D. and Winter, K. 2014. Asymmetrical effects of mesophyll conductance on fundamental photosynthetic parameters and their relationships estimated from leaf gas exchange measurements. Plant, Cell & Environment, 37: 978-994.
- Suresh, K. and Nagamani, C. 2006. Variations in photosynthetic rate and associated parameters with age of oil palm leaves under irrigation. Photosynthetica, 44: 309-311.
- Suresh, K., Kumar, M.K., Kanitha, D.L., Lakshimi, R.P. and Kumar, K.S. 2012. Variations in photosynthetic parameters and leaf water potential in oil palm grown under two different moisture regimes. Indian Journal of Plant Physiology, 17 (3 & 4): 233-240.
- Syahrinudin. 2005. The Potential of Oil Palm and Forest Plantations for Carbon Sequestration on Degraded Land in Indonesia; Ecology and Development Series 28; Cuvillier: Gottingen, Germany.

- Szeicz, G. and Long, I.F. 1969. Surface resistance of crop canopies. Water Resources Research, 5: 622-633.
- Tailliez, B. 1971. The root system of the oil palm on the San Alberto plantation in Colombia. Oléagineux, 26: 435-448.
- Tan, Y.P. and Ng, S.K. 1977. Spacing for oil palms on coastal clays in Peninsular Malaysia. In: International Developments in Oil Palm. Proceedings of the Malaysian International Agricultural Oil Palm Conference, (Ed. by Earp, D.A. and Newall, W.), Incorporated Society of Planters, Kuala Lumpur, pp. 183-191
- Tang, C.K. and Chin, N. 2012. Chapter 2: Malaysia's weather data. In: Building Energy Efficiency Technical Guideline for Passive Design.
- Tangang, F.T. and Juneng, L. 2007. Climate variability, climate change and extreme weather events in Malaysia. National Seminar on Socio-Economic Impacts of Extreme Weather and Climate Change: Putrajaya.
- Tarmizi Mohamad, A. and Tayeb Dolmat, M. 2006. Nutrient demands of Tenera oil palm planted on inland soils of Malaysia. Journal of Oil Palm Research, 18: 204-209.
- Taub, D.R. and Wang, X. 2018. Why are nitrogen concentrations in plant tissues lower under elevated CO<sub>2</sub>? A critical examination of the hypotheses. Journal of Integrative Plant Biology, 50: 1365-1374.
- Tcherkez G., Gauthier P., Buckley T.N., Busch, F.A., Barbour, M.M., Bruhn, D., Heskel, M.A., Gong, X.Y., Crous, K.Y., Griffin, K., Way, D., Turnbull, M., Adams, M.A., Atkin, O.K., Farquhar, G.D. and Cornic, G. 2017. Leaf day respiration: low CO<sub>2</sub> flux but high significance for metabolism and carbon balance. New Phytologist, 216: 986-1001.
- Teh, C.B.S. 2006. Introduction to mathematical modeling of crop growth. How the equations are derived and assembled into a computer program, Brown Walker Press, Boca Raton, Florida, USA.
- Teh, C.B.S. 2012. Effect of climate change on oil palm yield in Malaysia: Some simulations. http://www.christopherteh.com/blog/2012/09/cc-oil-palm/.
- Tenhunen J.D., Weber J.A., Yocum C.S. and Gates, D.M. 1976. Development of a photosynthesis model with an emphasis on ecological applications. Oecologia, 26: 101-119.
- Teoh, K.C. and Chew, P.S. 1988. Potassium in the oil palm eco-system and some implications to manuring practice. In: Proceedings of the 1987 International

Oil Palm Conference Progress and Prospects, (Ed. by Halim Hassan, A.), Palm Oil Research Institute of Malaysia, Kuala Lumpur, pp. 277-286.

- Tfwala, C.M., Mengistu, A.G., Haka, I.B.U., van Rensburg, L.D. and Du Preez, C.C. 2021. Seasonal variations of transpiration efficiency coefficient of irrigated wheat. Heliyon, 7(2): 1-8.
- Tinker, P.B. and Smilde, K.W. 1963. Dry-matter production and nutrient content of plantation oil palms in Nigeria. II. Nutrient content. Plant and Soil, 19: 350-363.
- Urban, O., Klem, K., Holišová, P., Šigut, L., Šprtová, M., Teslová-Navrátilová, P., and Grace, J. 2014. Impact of elevated CO<sub>2</sub> concentration on dynamics of leaf photosynthesis in *Fagus sylvatica* is modulated by sky conditions. Environmental Pollution, 185: 271–280.
- USDA. 2006. Oilseeds: world markets and trade, June 2006. USDA Foreign Agricultural Service, Washington, D.C.
- USDA. 2021. Oilseeds: world markets and trade, June 2021. USDA Foreign Agricultural Service, Washington, D.C.
- USDA-FAS. 2021. Production, Supply and Distribution Online: Oilseeds. USDA Foreign Agricultural Service, Washington, D.C., URL: http://apps.fas.usda.gov/psdonline/psdDownload.aspx
- van Keulen, H. and Seligman, N.G. 1987. Simulation of water use, nitrogen and growth of a spring wheat crop. Simulation Monographs. Pudoc, Wageningen.
- van Keulen, H., Penning de Vries, F.W.T. and Drees, E.M. 1982. A summary model for crop growth. In: Simulation of Plant Growth and Crop Production. Simulation Monographs (Ed. by Penning de Vries, F.W.T. and van Laar, H.H.), Pudoc, Wageningen, pp. 87-94.
- van Kraalingen, D.W.G. 1985. Simulation of oil palm growth and yield. Doctoral Thesis, Department of Theoretical Production Ecology, Agricultural University, Wageningen.
- van Kraalingen, D.W.G., Breure C.J. and Spitters C.J.T. 1989. Simulation of oil palm growth and yield. Agricultural and Forest Meteorology, 46: 227-244.
- van Noordwijk, M., Lusiana, B., Villamor, G., Purnomo, H. and Dewi, S. 2011. Feedback loops added to four conceptual models linking land change with driving forces and actors. Ecology and Society, 16(1): r1.

- Varley, J.A. 1966. Automatic methods for the determination of nitrogen, phosphorus and potassium in plant material. Analyst, 91: 119-126.
- Villar, R., Held, A.A., Merino, J. 1994. Comparison of methods to estimate dark respiration in the light in leaves of two woody species. Plant Physiology, 105: 167-172.
- von Caemmerer, S. 2000. Biochemical Models of Leaf Photosynthesis, Vol. 2. Collingwood, ON: CSIRO Publishing.
- von Caemmerer, S. and Evans, J.R. 1991. Determination of the average partia1 pressure of CO<sub>2</sub> in chloroplasts from leaves of several C<sub>3</sub> plants. Australia Journal of Plant Physiology, 18: 287-305.
- von Caemmerer, S. and Evans, J.R. 2015. Temperature responses of mesophyll conductance differ greatly between species. Plant, Cell & Environment, 38: 629-637.
- von Caemmerer, S., Evans, J.R., Hudson, G.S. and Andrews, T.J. 1994. The kinetics of ribulose-1,5-bisphosphate carboxylase/oxygenase *in vivo* inferred from measurements of photosynthesis in leaves of transgenic tobacco. Planta, 195: 88-97.
- von Uexkull, H.R., Henson, I.E. and Fairhurst, T.H. 2003. Canopy management to optimize yield. In: Oil palm: management for large and sustainable yields (Ed. by Fairhurst, T.H. & Härdter, R.), Potash & Phosphate Institute of Canada (ESEAP), Singapore, pp. 163-180.
- Vörösmarty, C.J., Federer, C.A. and Schloss, A.L. 1998. Potential evaporation functions compared on US watersheds: Possible implications for globalscale water balance and terrestrial ecosystem modeling. Journal of Hydrology, 207 (3–4): 147-169.
- Waichler, S.R. and Wigmosta, M.S. 2003. Development of hourly meteorological values from daily data and significance to hydrological modeling at H.J. Andrews Experimental Forest. Journal of Hydrometeorology, 4: 251-263.
- Waite, P.A., Schuldt, B., Link, R.M., Breidenbach, N., Triadiati, T., Hennings, N., Saad, A. and Leuschner, C. 2019. Soil moisture regime and palm height influence embolism resistance in oil palm. Tree Physiology, 39(10): 1696-1712.
- Walker, B.J. and Ort, D.R. 2015. Improved method for measuring the apparent CO<sub>2</sub> photocompensation point resolves the impact of multiple internal conductances to CO<sub>2</sub> to net gas exchange. Plant, Cell & Environment, 38: 2462-2474.

- Walker, B.J., Ariza, L.S., Kaines, S., Badger, M.R. and Cousins, A.B. 2013. Temperature response of *in vivo* Rubisco kinetics and mesophyll conductance in *Arabidopsis thaliana*: comparisons to *Nicotiana tabacum*. Plant, Cell & Environment, 36: 2108-2119.
- Walker, B.J., Orr, D.J., Carmo-Silva, E., Parry, M.A.J., Bernacchi, C.J. and Ort, D.R. 2017. Uncertainty in measurements of the photorespiratory CO<sub>2</sub> compensation point and its impact on models of leaf photosynthesis. Photosynthesis Research, 132: 245-255.
- Wang, B., Luo, X., Yang, Y.M., Sun, W., Cane, M.A., Cai, W., Yeh, S.W. and Liu, J. 2019. Historical change of El Niño properties sheds light on future changes of extreme El Niño. PNAS, 116 (45): 22512-22517.
- Wann, M. Yen, D. and Gold, H.J. 1985. Evaluation and calibration of three models for daily cycle of air temperature. Agricultural and Forest Meteorology, 34: 121-128.
- Warren, C.R. 2008. Does growth temperature affect the temperature response of photosynthesis and internal conductance to CO<sub>2</sub>? A test with *Eucalyptus regnans*. Tree Physiology, 28: 11-19.
- Warren, C.R. and Dreyer, E. 2006. Temperature response of photosynthesis and internal conductance to CO<sub>2</sub>: results from two independent approaches. Journal of Experimental Botany, 57: 3057-3067.
- Weise, S.E., Carr, D.J., Bourke, A.M., Hanson, D.T., Swarthout, D. and Sharkey, T.D. 2015. The arc mutants of Arabidopsis with fewer large chloroplasts have a lower mesophyll conductance. Photosynthesis Research, 124: 117-126.
- Wilkerson, G.G., Jones, J.W., Boote, K.J., Ingram, K.T. and Mishoe, J.W. 1983. Modeling soybean growth for crop management. Transactions of the ASAE, 26: 63–73.
- Williams, A.P., Allen, C.D., Macalady, A.K., Griffin, D., Woodhouse, C.A., Meko, D.M., Swetnam, T.W., Rauscher, S.A., Seager, R., Grissino-Mayer, H.D., et al. 2013. Temperature as a potent driver of regional forest drought stress and tree mortality. Nature Climate Change Change, 3: 292–297.
- Williams, A.P., Seager, R., Berkelhammer, M., Macalady, A.K., Crimmins, M.A., Swetnam, T.W., Trugman, A.T., Buenning, N., Hryniw, N., McDowell, N.G., et al. 2014. Causes and implications of extreme atmospheric moisture demand during the record-breaking 2011 wildfire season in the southwestern United States. Journal of Applied Meteorology and Climatology, 53: 2671–2684.

- Willmott, C. and Matsuura, K. 2005. Advantages of the Mean Absolute Error (MAE) over the Root Mean Square Error (RMSE) in assessing average model performance. Climate Research, 30: 79-82.
- Willmott, C., Robeson, S.M. and Maysuura, K. 2012. A refined index of model performance. International Journal of Climatology, 32: 2088-2094.
- Wise, R.R., Olson, A.J., Schrader, S.M. and Sharkey, T.D. 2004. Electron transport is the functional limitation of photosynthesis in field grown Pima cotton plants at high temperature. Plant, Cell & Environment, 27: 717-724.
- Woittiez, L.S., van Wijk, M.T., Slingerland, M., van Noordwijk, M., Giller, K.E. 2017. Yield gaps in oil palm: A quantitative review of contributing factors. European Journal of Agronomy 83: 57-77.
- Wu, A., Song, Y., van Oosterom, E.J. and Hammer, G.L. 2016. Connecting Biochemical Photosynthesis Models with Crop Models to Support Crop Improvement. Frontier in Plant Science, 7: 1518.
- Wullschleger, S.D. 1993. Biochemical limitations to carbon assimilation in  $C_3$  plants a retrosprective analysis of the  $A/C_i$  curves from 109 species. Journal of Experimental Botany, 44: 907-920.
- Xu, Y., Yu, L., Cials, P., Yu, L., Li, W., Chen, X., Zhang, H., Yue, C., Kanniah, K., Cracknell, A.P. and Gong, P. 2020. Oil palm modelling in the global landsurface model ORCHIDEE-MICT. Geoscientific Model Development, pp. 1-32.
- Yan, H. and Oue, H. 2011. Application of the two-layer model for predicting transpiration from the rice canopy and water surface evaporation beneath the canopy. Journal of Agricultural Meteorology, 67(3): 89-97.
- Yang, K. and Koike, T. 2002. Estimating surface solar radiation from upper-air humidity. Solar Energy, 72: 177-18.
- Yang, J.T., Preiser, A.L., Li, Z., Weise, S.E. and Sharkey, T.D. 2016. Triose phosphate use limitation of photosynthesis: short-term and long-term effects. Planta, 243: 687-698.
- Yang, J., Yang, J., Liu, S. and Hoogenboom, G. 2014. An evaluation of the statistical methods for testing the performance of crop models with observed data. Agricultural Systems, 127: 81-89.
- Yin, X. and van Laar, H.H. 2005. Crop systems dynamics. An ecophysiological simulation model for genotype-by-environment intreactions. Wageningen Academic Publishers, The Netherlands.

- Yu, S., Eder, B., Dennis, R., Chu, S.H. and Schwartz, S.E. 2006. New unbiased symmetric metrics for evaluation of air quality models. Atmospheric Science Letters, 7: 26–34.
- Yuan, Z., Liu, W., Niu, S. and Wan, S. 2007. Plant nitrogen dynamics and nitrogen-use strategies under altered nitrogen seasonality and competition. Annals of Botany, 100: 821-830.
- Zaharah, A.R., Sharifuddin H.A.H., Ahmad Sahali, M. and Mohd Hussein, M.S. 1989. Fertilizer placement studies in mature oil palm using isotope technique. Planter, 65: 384-388.
- Zhao, C., Liu, B., Piao, S., Wang, X., Lobell, D.B., Huang, Y., Huang, M., Yao, Y., Bassu, S., Ciais, P., et al. 2017. Temperature increase reduces global yields of major crops in four independent estimates. Proceedings of the National Academy of Sciences, USA 114: 9326–9331.
- Zuidema, P.A., Leffelaar, P.A., Gerritsma, W., Mommer, I. and Anten, N.P.R. 2005. A physiological production model for cocoa (*Theobroma cacao*): model presentation, validation and application. Agricultural systems, 84: 195-225.