



**PROFILING OF PARTHENIUM (*Parthenium hysterophorus* L.) ACTIVE
COMPOUNDS AND ALLELOPATHIC SUPPRESSION OF CROPS AND
WEEDS IN BAMBARA GROUNDNUT (*Vigna subterranea* L.) CULTIVATION**

By

H. M KHAIRUL BASHAR

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

September 2022

FP 2022 69

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

To my pretty father, Md. Fazlur Rahman, late mother, Mst. Rokeya Begam, my father in-law, Md. Abdur Rahim Mia, Mother-in-law, Tara Banu and my beloved wife Ferdoushi Rahaman, my adorable son Zubair Bin Bashar, my daughter Ayesha Al Humayera and Fatima Tuz Zohra who always kept praying for me day and night and sacrifice to achieve my goal. Specially thanks to all of my family members

and

To all of my friends who supported me all these years.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

PROFILING OF PARTHENIUM (*Parthenium hysterophorus* L.) ACTIVE COMPOUNDS AND ALLELOPATHIC SUPPRESSION OF CROPS AND WEEDS IN BAMBARA GROUNDNUT (*Vigna subterranea* L.) CULTIVATION

By

H. M KHAIRUL BASHAR

September 2022

Chairman : Professor Dato' Abdul Shukor bin Juraimi, PhD
Faculty : Agriculture

Parthenium hysterophorus L. is a noxious herb with pharmaceutical properties and has allelopathic potentiality to retards the growth of many species. Hence, research was undertaken with the aim to identified phytotoxic compounds and determine the possibility of using aerial parts (leaf, stem and flower) of *P. hysterophorus* as a potential bioherbicide to control weed in Bambara groundnut field. The experiment was conducted in laboratory and glasshouse condition at University Putra Malaysia (UPM) using complete randomize design (CRD) and randomize complete block design (RCBD) accordingly. Germination of seeds and seedling growth of selected crops and weed species were tested with six concentrations of methanol, hexane, acetone, and water extracts (25, 50, 75, 100, and 150 g L⁻¹) and distilled water as the control. The results showed that the phytotoxicity effects were significantly higher by leaf extract and methanol solvent compared to other treatments. The methanol extract of *P. hysterophorus* leaf sprayed as post-emergence applications on bambara groundnut (*V. subterranean* L.), maize (*Z. mays*), *D. sanguinalis*, *E. indica*, *A. conyzoides*, *C. iria*, *E. hirta*, *C. difformis* showed that the efficacy of the treatments were in the following trend; leaf>flower>stem and methanol>acetone>water>hexane. The liquid chromatography-mass spectrometry (LC-MS) analysis exposed the presence of 67 and 196 compounds in methanol, 60 and 191 in hexane, 75 and 77 in acetone, and 52 and 184 from water in both positive and negative polarity analysis. Seven phytotoxic derivatives were identified from the methanol solvents of leaf and flower but two and three compounds in acetone, hexane, and water extract respectively. On the other hand, high-performance liquid chromatography (HPLC) analysis confirmed methanol with leaf extracts contained a higher number and amount of chemical compounds than did those of the other solvent extracts. Caffeic acid, chlorogenic acid, and parthenin (highest allelopathic activity) were detected in the highest concentrations in the methanol leaf extract, but this compound was not found in the other extracts. *P. hysterophorus* methanol leaf extract administered to the test species at doses of 100 g L⁻¹ showed a reduction in photosynthesis rate (22-83.37%), stomatal conductance (17-120.40%), transpiration

(27.41%), chlorophyll content (13-143.87%), and carotenoid content (15-117.72%) at 6 hours after spray (HAS). The activities of antioxidant enzymes, malondialdehyde (MDA)(183-596%), Proline content (150-500%), superoxide dismutase (SOD)(76-95%), catalase (CAT)(98-208%), and peroxidase (POD)(94-212%) were also increased with the same concentrations at 72 HAS in different weeds. Present findings confirm that the methanol leaf extract of *P. hysterophorus* can disrupt the physiological and biochemical mechanism of target weeds. For field evaluation, treatment of *P. hysterophorus* leaf methanol extract (75, 100, and 150 g L⁻¹) along with five chemical herbicides [chemical name % a.i. (Fusilade Forte 150 EC), chemical name % a.i. (Prowl® 3.3 EC), chemical name % a.i. (Hextar Diuron 80wp), glyphosate 41% a.i. (Roundup®) and glufosinate-ammonium 13.5% a.i. (Basta®)] at 1 L ha⁻¹, no herbicide treatment and weeding manually was done. It was observed that *P. hysterophorus* leaf methanol extract at the rate of 150 g L⁻¹ and all chemical herbicides killed Bambara groundnut and most weeds. However, Bambara groundnut yielded the most when sprayed with *P. hysterophorus* leaf methanol extract at a rate of 100 g L⁻¹ followed by manual (hand weeding). As a result, 100 g L⁻¹ of *P. hysterophorus* leaf methanol extract could be sprayed to manage weeds without reducing Bambara groundnut yield. Based on these findings, *P. hysterophorus* leaf methanol extract can be utilized as a natural post-emergent herbicide in Bambara groundnut fields to suppress weeds, as well as for the development of novel herbicides based on phytotoxic chemicals generated by this plant.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PROFIL PARTHENIUM (*Parthenium hysterophorus* L.) SEBATIANYA AKTIF DAN PENINDASAN ALELOPATIK TANAMAN DAN RUMPAI DALAM PENANAMAN KACANG BAMBARA (*Vigna subterranea* L.)

Oleh

H. M. KHAIRUL BASHAR

September 2022

Pengerusi : Profesor Dato' Abdul Shukor bin Juraimi, PhD
Fakulti : Pertanian

Penggunaan racun rumpai sintetik yang berterusan dan sembarangan telah menyebabkan pencemaran alam sekitar dan pembentukan rumpai rintang herbisida. Tumbuhan alelopati sebagai racun herba semula jadi boleh menjadi penyelesaian baru untuk mengurangkan pergantungan pada racun rumpai sintetik di samping menambah baik pengurusan rumpai tanaman. Rumpai berbahaya mempunyai potensi untuk menyampaikan alelokimia dan mempengaruhi organisma di sekelilingnya. Terdapat sedikit maklumat yang tersedia tentang aktiviti alelopati spesies rumpai berbahaya Malaysia. Penemuan rumpai berbahaya alelopati yang tidak diketahui di Malaysia boleh menyediakan jalan untuk membangunkan racun herba semula jadi yang baharu. Beberapa eksperimen telah dijalankan di makmal dan rumah kaca untuk mencapai matlamat berikut; i) untuk menilai aktiviti alelopati 30 spesies rumpai berbahaya di Malaysia ii) untuk menentukan kesannya di makmal dan rumah kaca; iii) untuk mengenal pasti bahan alelopati rumpai tersebut; iv) untuk menentukan keberkesanan berbanding dengan racun rumpai komersial, v) untuk menilai kesan ke atas perubahan fisiologi dan biokimia rumpai. Eksperimen telah dijalankan di Makmal Teknologi Benih, Sains Tanaman, dan Ladang 15, Universiti Putra Malaysia. Reka bentuk rawak lengkap (CRD) dengan lima replikasi dan reka bentuk blok lengkap rawak (RCBD) dengan empat replikasi telah disusun atur untuk eksperimen makmal dan rumah kaca. Eksperimen terhadap kesan ekstrak metanol 30-spesies rumpai Malaysia yang berbahaya (9 famili) terhadap kadar kemandirian benih dan pertumbuhan anak benih padi angin (*Oryza sativa* f. *spontanea* Roshev) di makmal telah dijalankan. Lima kepekatan (6.25, 12.5, 25, 50 dan 100 g L⁻¹) ekstrak dibandingkan dengan kawalan (air suling). *Parthenium hysterophorus* L., *Cleome rutidosperma* DC. dan *Borreria alata* (Aubl.) DC. telah terbukti mempunyai alelopati yang kuat berbanding dengan ekstrak lain yang diuji. Kesan fitotoksik kepekatan ekstrak *P. hysterophorus*, *C. rutidosperma* dan *B. alata* telah disiasat selanjutnya dan dibandingkan dengan kadar survival dan pertumbuhan tanaman (*Zea mays* L., *Oryza sativa* L., *Abelmoschus esculentus* L. Moench, *Amaranthus gangeticus* L.) dan rumpai (padi angin, *Echinochloa colona* L. Link.,

Euphorbia hirta L. dan *Ageratum conyzoides* L.) di dalam suasana makmal dan rumah kaca. Tanaman-tanaman yang diuji didapati kurang sensitif kepada *C. rutidosperma* dan *B. alata* berbanding ekstrak *P. hysterophorus* dalam kedua-dua keadaan (makmal dan rumah kaca). *Ageratum conyzoides*, *E. hirta*, *A. esculentus* dan *A. gangeticus* kebanyakannya dapat dicerderakan oleh ekstrak *P. hysterophorus* pada 100 g L^{-1} dalam rumah kaca. Analisis LC-QTOF-MS/MS telah mengesahkan kehadiran sebatian fenolik (flavonoid, fenol, kumarin, asid karboksilik, asid benzoik), terpenoid, alkaloid, asid amino, asid lemak, piperazine, benzofuran, indole, amina, azoles, asid sulfonik dan sebatian lain yang tidak diketahui dalam *P. hysterophorus*, *C. rutidosperma* dan *B. alata*. Keputusan tersebut menunjukkan bahawa ekstrak metanol *C. rutidosperma* dan *B. alata* mempunyai sedikit bahan fitotoksik yang diketahui berbanding *P. hysterophorus*. Kajian perbandingan telah dijalankan antara ekstrak *P. hysterophorus* ($20, 40, \text{ dan } 80 \text{ g L}^{-1}$), racun herba sintetik (glifosat dan glufosinate-ammonium pada kadar 2 L ha^{-1}) sebagai kawalan positif dan tiada rawatan (kawalan negatif) pada *A. conyzoides*, padi angin dan *Cyperus iria*. Tiada perbezaan ketara diperolehi apabila ekstrak *P. hysterophorus* (80 g L^{-1}) dan racun herba sintetik (glyphosate dan glufosinate-ammonium) diberikan pada *A. conyzoides*. Tindak balas sifat fisiologi dan biokimia *A. conyzoides*, padi angin dan *C. iria* juga telah disiasat dengan memberikan semburan dedaun *P. hysterophorus* pada kepekatan yang berbeza ($20, 40 \text{ dan } 60 \text{ g L}^{-1}$). Sifat fisiologi dan biokimia *A. conyzoides* didapati lebih sensitif terhadap *P. hysterophorus*, terutamanya pada kepekatan yang tinggi (60 g L^{-1}). Hasil kajian ini mengesahkan bahawa *P. hysterophorus* mempunyai kesan herbisida yang ketara untuk kawalan rumpai. Oleh itu, rumpai ini berpotensi untuk digunakan sebagai bioherbisida untuk mengawal rumpai di Malaysia.

ACKNOWLEDGEMENTS

First and foremost, I am most grateful to Allah S.W.T for giving me the strength and courage to complete the writing of this research thesis within the period given. I am eternally grateful to my Supervisor, Professor Dato' Dr. Abdul Shukor Juraimi for his advice, motivation, enthusiasm and proper guidance throughout the period of the thesis. I am very much indebted to him for his unwavering support, guidance and valuable advice in completing this thesis. Without his counsel, this thesis would not be able to achieve its objective. May Allah repays his deeds with liberal blessings.

With a great deal of luck, I got an excellent Supervisory Committee. I owe an immense debt to the rest of my supervisory committee members for their advice, Dr. Muhammad Saiful Ahmad-Hamdani, Dr. Md Kamal Uddin, Dr. Norhayu Asib, and Professor Dr. Md. Parvez Anwar (BAU, Bangladesh) who provided insightful comments, critical review and friendly supervision.

I would like to express my deep sense of respect and immense gratitude to all the Professor and Lecturer in Department of Crop Science and Department of Plant Protection, Faculty of Agriculture, Universiti Putra Malaysia (UPM), especially Professor Dr. S. M. Rezaul Karim, IOI chair, for their assistance, good teaching and cooperation which led to the smooth running of this study. I consider it an honour to work with all the administrative and technical staffs of the Universiti Putra Malaysia for their full cooperation and for providing me the assistance, especially during the entire implementation of my experiments. Mr. Mohd Yonos Bin Abdul Wahab, Mr. Md. Azhar Othman and Mr. Mohd Norhaizan Saliudin, I will never forget the warm and kind help you extended to me.

Last but not least, many thanks to my family and friends for sharing useful information to help with this period. I wish to sincerely acknowledge Ministry of Agriculture (MoA), of the People's Republic of Bangladesh, Bangladesh Agricultural Research Council (NATP Phase-II Project, BARC), Bangladesh Agricultural Research Institute (BARI) (research grant: vote number 6282506), for providing financial support and Universiti Putra Malaysia for assistance. I would like to apologize for any inconvenience caused throughout this postgraduate study period.

I certify that a thesis examination committee has met on 22 September 2022 to conduct the final examination of H M Khairul Bashar on his Doctor of Philosophy thesis entitled “Profiling of *Parthenium (Parthenium hysterophorus)* Active Compounds and Allelopathic Suppression of Crops and Weeds in Bambara groundnut (*Vigna subterranea* L.) Cultivation” in accordance with Universiti Pertanian Malaysia (Higher Degree) act 1980 and Universiti Pertanian Malaysia (Higher Degree) regulations 1981. The committee recommends that the candidate be awarded the relevant degree.

Members of the Examination Committee were as follows:

Khiarulmazmi Ahmad, PhD
Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Norida Mazlan, PhD
Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Zulkefly Sulaiman, PhD
Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Farooq Anwar, PhD
Professor
Department of Chemistry
University of Sargodha
Pakistan
(External Examiner)

ZURIATI AHMAD ZUKARNAIN, PhD
Professor Ts. and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfillment of the requirements for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Abdul Shukor bin Juraimi, PhD

Professor Dato'
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Muhammad Saiful bin Ahmad-Hamdani, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Md. Kamal Uddin, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Norhayu binti Asib, PhD

Senior Lecturer, Ts.
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Md. Parvez Anwar, PhD

Professor
Faculty of Agriculture
Bangladesh Agricultural University,
Bangladesh
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 12 January 2023

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____

Date: _____

Name and Matric No: H M Khairul Bashar

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: Professor Dato'
Dr. Abdul Shukor bin Juraimi

Signature: _____
Name of Member
of Supervisory
Committee: Associate Professor
Dr. Muhammad Saiful bin Ahmad-Hamdani

Signature: _____
Name of Member
of Supervisory
Committee: Associate Professor
Dr. Md Kamal Uddin

Signature: _____
Name of Chairman
of Supervisory
Committee: Ts. Dr. Norhayu Asib

Signature: _____
Name of Member
of Supervisory
Committee: Professor
Dr. Md. Parvez Anwar

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	vii
LIST OF TABLES	xvi
LIST OF FIGURES	xx
LIST OF ABBREVIATIONS	xxiii
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	4
2.1 The importance of Bambara groundnut	4
2.1.1 Pharmacognostical description of Bambara groundnut	5
2.1.2 A botanical description of Bambara groundnut and taxonomic tree	5
2.1.3 Cultivar selection	7
2.1.4 Cultivation procedure	7
2.2 Weed problems in Bambara groundnut fields	7
2.2.1 Critical period of crop weed competition and its effect on growth components	8
2.2.2 Weed community in the Bambara groundnut field	9
2.2.3 Yield loss due to weeds	11
2.3 Weed control problems	11
2.4 Parthenium overview	12
2.4.1 Family and genus of Parthenium	12
2.4.2 A botanical description of Parthenium and its distribution	13
2.4.3 The beneficial effects of <i>P. hysterophorus</i>	14
2.4.3.1 Antidiabetic effects	14
2.4.3.2 Antioxidant activity	14
2.4.3.3 Antitumor activity	15
2.4.3.4 Antimicrobial activity	15
2.4.3.5 Larvicidal effects	17
2.4.3.6 Parthenium compost	17
2.4.3.7 Herbicidal effects	17
2.4.3.8 Allelopathic effects	18
2.4.3.9 Pesticidal effects	18
2.4.3.10 Heavy metal and dye removal	18
2.4.3.11 Other economic benefits	18
2.5 Herbicidal effects of Parthenium extracts	19
2.5.1 Methanol extracts	19
2.5.2 Ethanol extracts	20
2.5.3 Hexane extracts	20

2.5.4	Acetone extracts	20
2.5.5	Aqueous extracts	21
2.6	Chemical constituents of Parthenium plants and their mechanisms	21
2.7	Allelopathic composites	22
2.8	Phytochemistry	23
2.9	Allelopathy	26
2.9.1	How allelochemicals are released from the donor	26
2.9.2	Mode of action of allelochemicals	29
2.9.3	The Role of allelopathy in weed management and control	33
2.9.4	Herbicidal potential of allelochemicals'	33
3	ALLELOPATHIC EFFECT OF VARIOUS PARTS OF <i>Parthenium hysterophorus</i> ON DIFFERENT CROPS AND WEEDS	34
3.1	Introduction	34
3.2	Materials and Methods	35
3.2.1	Phytotoxic interference of various parts of Parthenium extracts with different solvents	35
3.2.1.1	Site description	35
3.2.1.2	Layout of experimental design and treatment arrangements	35
3.2.1.3	Collection of Parthenium weed and other test plant seeds	37
3.2.1.4	Extract preparation	37
3.2.1.5	Germination and growth bioassays	38
3.2.1.6	Data collection	38
3.2.1.7	Statistical analysis	39
3.3	Results	39
3.3.1	Allelopathic activity of leaf, stem and flower extracts of <i>P. hysterophorus</i>	39
3.3.1.1	Inhibitory effect of methanol extracts of the areal parts of Parthenium on different plant species	39
3.3.1.2	Inhibitory effect of hexane extracts of the areal parts of Parthenium on different crops and weeds	52
3.3.1.3	Inhibitory effect of acetone extracts of the areal parts of Parthenium on different crops and weeds	68
3.3.1.4	Inhibitory effect of water extracts of the areal parts of Parthenium on different plant species	81
3.3.1.5	Comparison between extract solvents in inhibitory effects on indicator plants	94
3.3.1.6	Comparison between different solvents with aerial parts extracts of <i>P. hysterophorus</i> in response to initial growth parameters of test plants	94

3.4	Discussion	111
3.5	Conclusion	117
4	POST-EMERGENT APPLICATION OF LEAF METHANOL EXTRACT OF <i>Parthenium hysterophorus</i> AS AN HERBICIDE TO CONTROL WEEDS UNDER GLASS HOUSE CONDITION	118
4.1	Introduction	118
4.2	Materials and methods	119
4.2.1	Effects of post-emergent application of <i>P. hysterophorus</i> leaf methanol extract on the test species	119
4.2.1.1	Experimental site	119
4.2.1.2	Layout of Experimental design and treatment arrangements	119
4.2.1.3	Plant materials and method	120
4.2.1.4	Data Collection	121
4.2.1.5	Statistical analysis	121
4.3	Results	121
4.3.1	Phytotoxic effect of Parthenium methanol extract on different crops and weeds under glasshouse condition	121
4.3.1.1	Effect on plant height and root length of crops	121
4.3.1.2	Effect on leaf area, total chlorophyll, fresh and dry weight of weeds	122
4.3.1.3	Effect on plant height and root length of weeds	122
4.3.1.4	Effect on leaf area, total chlorophyll content, total fresh and dry weight of weeds	123
4.4	Discussion	128
4.5	Conclusion	131
5	CHROMATOGRAPHY ANALYSIS: IDENTIFICATION AND QUANTIFICATION OF PHYTOTOXIC SUBSTANCES IN LEAF, STEM, AND FLOWER OF <i>Parthenium hysterophorus</i>	132
5.1	Introduction	132
5.2	Materials and Methods	133
5.2.1	Experimental site	133
5.2.2	Sample preparation	133
5.2.3	Identification of phytotoxic compounds from <i>P. hysterophorus</i> leaves, stems, and flowers through LC-MS analysis	133
5.2.4	Quantification of phytotoxic compounds through HPLC analysis	134
5.2.4.1	Chemicals	134
5.2.4.2	Preparation of stock solutions and doses	134
5.2.4.3	Analysis of compounds	134
5.3	Results	135
5.3.1	LC-MS analysis	135
5.3.1.1	Identified compounds from LC-MS analysis of methanol extract of Parthenium	135

5.3.1.2	Identified compounds from LC-MS analysis of hexane extract of Parthenium	135
5.3.1.3	Identified compounds from LC-MS analysis in the acetone extract of Parthenium	138
5.3.1.4	Identified compounds from LC-MS analysis in the water extract of Parthenium	138
5.3.2	HPLC analysis	141
5.4	Discussion	145
5.5	Conclusion	148
6	HERBICIDAL EFFECT OF PHYTOTOXIC COMPOUNDS PRESENT IN <i>Parthenium hysterophorus</i> ON <i>Eleusine indica</i> AND <i>Digitaria sanguinalis</i>	149
6.1	Introduction	149
6.2	Materials and methods	150
6.2.1	Site description	150
6.2.2	Layout of experimental design and treatment arrangements	150
6.2.3	Plant materials and compounds	151
6.2.4	Bioassay	151
6.2.5	Data collection	152
6.2.6	Statistical analysis	152
6.3	Results	152
6.3.1	Allelopathic effects of the phytochemicals	152
6.3.1.1	Effects on germination and early growth of <i>D. sanguinalis</i>	153
6.3.1.2	Comparison between phytochemicals in their effects on growth parameters	157
6.3.1.3	Cluster analysis and valuation of principal component analysis	158
6.3.1.4	Germination and early growth of <i>E. indica</i> treated with detected allelochemicals	159
6.3.1.5	Comparison of phytochemicals in their effects on examined initial growth parameters	164
6.3.1.6	Cluster analysis and assessment of principal component analysis	165
6.4	Discussion	166
6.5	Conclusion	168
7	DETERMINATION OF PARTHENIUM WEED EXTRACT HERBICIDAL MODE-OF-ACTION AGAINST PLANT GROWTH	170
7.1	Introduction	170
7.2	Materials and methods	171
7.2.1	Experimental site	171
7.2.2	Experimental treatments and design	171
7.2.3	Plant materials and methods	172
7.2.4	Data collections	173

7.2.4.1	Determination of photosynthetic rate, stomatal conductance and transpiration	173
7.2.4.2	Estimation of chlorophyll pigments	173
7.2.4.3	Measurement of malondialdehyde (MDA) content	174
7.2.4.4	Measurement of proline content	174
7.2.4.5	Estimation of enzymes	175
7.2.5	Statistical analysis	177
7.3	Results	177
7.3.1	Effect on chlorophyll-a content of the crops and weeds	177
7.3.2	Effect on chlorophyll-b content of the crops and weeds	177
7.3.3	Effect on total chlorophyll and carotenoid content of the crops and weeds	181
7.3.4	Effect on photosynthesis rate of the tested crops and weeds	181
7.3.5	Effect on stomatal conductance of the tested crops and weeds	185
7.3.6	Effect on transpiration rate of the tested crops and weeds	185
7.3.7	Effect on malondialdehyde content of the tested crops and weeds	191
7.3.8	Effect on proline content of the tested crops and weeds	191
7.3.9	Effect on superoxide dismutase of the tested crops and weeds	196
7.3.10	Effect on catalase activity of the tested crops and weeds	196
7.3.11	Effect on peroxidase activity of the tested crops and weeds	196
7.4	Discussion	203
7.5	Conclusion	205
8	EVALUATION OF LEAF EXTRACT OF <i>Parthenium hysterophorus</i> ON THE EMERGENCE AND GROWTH OF BAMBARA GROUNDNUT AND ASSOCIATED WEEDS IN GLASSHOUSE CONDITION	206
8.1	Introduction	206
8.2	Materials and methods	208
8.2.1	Site description	208
8.2.2	Layout of experimental design and treatments arrangement	208
8.2.3	Extraction procedure	209
8.2.4	Field soil properties	209
8.2.5	Crop management practices	209
8.2.6	Plant Materials and Methods	210
8.2.7	Data collections	210
8.2.7.1	Weed density and dry weight	211
8.2.7.2	Yield of Bambara groundnut	211
8.2.8	Statistical analysis	212
8.3	Results	212

8.3.1	Efficacy of Parthenium leaf methanol extract in comparison with commercial herbicides on different weeds growth	212
8.3.2	Effects on the growth and yield of Bambara groundnut plants	217
8.4	Discussion	224
8.5	Conclusion	226
9	GENERAL DISCUSSION, CONCLUSION AND FUTURE RECOMMENDATIONS	227
9.1	General discussion and conclusion	227
9.2	Future recommendations	229
	RERERENCES	230
	APPENDICES	270
	BIODATA OF STUDENT	335
	LIST OF PUBLICATIONS	336

LIST OF TABLES

Table		Page
2.1	Common weeds in Bambara groundnut fields	10
2.2	Beneficial effects of <i>Parthenium hysterophorus</i> weeds	16
2.3	Allelochemicals of <i>Parthenium hysterophorus</i> and their mechanism of action	23
2.4	Main groups and constituents of <i>P. hysterophorus</i> plant	25
2.5	Mode of action of some allelopathic compounds	31
3.1	Effect of leaves, stem and flower methanol extracts of <i>Parthenium hysterophorus</i> on germination, radicle and hypocotyl length of different crops	41
3.2	Percent inhibition in germination, radicle and hypocotyl length of different crops due to <i>Parthenium</i> methanol extracts	44
3.3	The half inhibitory effect of <i>Parthenium</i> methanol extracts, their sensitivity to different crop species	49
3.4	Similarity among the indicator plants	54
3.5	Effect of leaves, stem and flower extracts of <i>Parthenium hysterophorus</i> with hexane on germination, radicle and hypocotyl length of different crops	57
3.6	Percent inhibition on germination, radicle and hypocotyl length of different crops due to <i>Parthenium hysterophorus</i> hexane extract	60
3.7	The half inhibitory effect of <i>Parthenium</i> hexane extracts on initial growth parameters of test plants	64
3.8	The similarity among the indicator plants in response to leaf, stem and flower extracts of <i>Parthenium</i>	66
3.9	Effect of leaf, stem and flower extracts of <i>Parthenium hysterophorus</i> with acetone on germination, radicle and hypocotyl length of different crops	70
3.10	Percent inhibition in germination, radicle and hypocotyl growth of different crops due to <i>Parthenium hysterophorus</i> extract with acetone	73

3.11	Sensitivity of acetone extracts on initial growth parameters of test plants	77
3.12	Showing the similarity among the indicator plants	78
3.13	Effect of leaf, stem and flower extracts of <i>Parthenium hysterophorus</i> with water on germination, radicle and hypocotyl length of different crops	83
3.14	Percent inhibition in germination, radicle and hypocotyl length of different crops due to water extracts of parthenium leaf, stem and flower	86
3.15	Sensitivity of water extracts Parthenium plant parts on initial growth parameters of test plants	90
3.16	Similarity in response to phytotoxicity among the indicator plants	91
3.17	Germination and early growth inhibition (%) in the test plants treated with different solvents of <i>P. hysterophorus</i> leaf extracts	96
3.18	Germination and early growth inhibition (%) in the test plants treated with different solvents of <i>P. hysterophorus</i> stem extracts	99
3.19	Germination and early growth inhibition (%) of test plants treated with different solvents of <i>P. hysterophorus</i> flower extracts	102
3.20	Comparison between different solvents with leaf extracts of <i>P. hysterophorus</i> in response sensitivity of examined initial growth parameters and plants	105
3.21	Comparison between different solvents with stem extracts of <i>P. hysterophorus</i> in response to initial growth parameters of test plants	107
3.22	Comparison between different solvents with flower extracts of <i>P. hysterophorus</i> in response to initial growth parameters of test plants	109
4.1	Effect of <i>P. hysterophorus</i> with methanol extract on plant height (cm), leaf area (cm ²), root length (cm), total chlorophyll (SPAD), total fresh weight (g pot ⁻¹) and total dry weight (g pot ⁻¹) of <i>Vigna subterranea</i> and <i>Zea mays</i>	124
4.2	Effect of <i>P. hysterophorus</i> with methanol extract on plant height (cm), leaf area (cm ²), root length (cm), total chlorophyll (SPAD), total fresh weight (g pot ⁻¹) and total dry weight (g pot ⁻¹) of some weeds	125
5.1	Phenolic derivatives found in methanol extract of <i>Parthenium hysterophorus</i> aerial parts through positive and negative polarity analysis	137

5.2	Phenolic derivatives found in hexane extract of <i>Parthenium hysterophorus</i> aerial parts through positive and negative polarity analysis	139
5.3	Phenolic derivatives found in the acetone extracts of <i>Parthenium hysterophorus</i> aerial parts through positive and negative polarity analysis	140
5.4	Phenolic derivatives found from water extract of <i>Parthenium hysterophorus</i> different parts through positive and negative polarity analysis	142
5.5	Major phytotoxic compounds and their quantity (ppm) detected from tested extracts of <i>P. hysterophorus</i>	144
6.1	Germination (%) of <i>D. sanguinalis</i> treated with selected phytochemicals	154
6.2	Root length (cm) of <i>D. sanguinalis</i> treated with selected phytochemicals	155
6.3	Shoot length (cm) of <i>D. Sanguinalis</i> treated with detected allelochemicals	156
6.4	Inhibitory effect of phytotoxic compounds, sensitivity of examined initial growth parameters of <i>D. sanguinalis</i>	157
6.5	Germination (%) of <i>E. indica</i> treated with the phytochemicals	161
6.6	Root length (cm) of <i>E. indica</i> treated with the phytochemicals	162
6.7	Shoot length (cm) of <i>E. indica</i> treated with the phytochemicals	163
6.8	Inhibitory effect of phytotoxic compounds, sensitivity of examined initial growth parameters of <i>E. indica</i>	164
7.1	Response of chlorophyll-a content and chlorophyll-b of some crops and weeds to a foliar spray of <i>P. hysterophorus</i> leaf extract	178
7.2	Response of total chlorophyll content and carotenoids of some crops and weeds to a foliar spray of <i>P. hysterophorus</i> leaf extract	182
7.3	Response of photosynthesis rate, stomatal conductance and transpiration rate of some crops and weeds to a foliar spray of <i>P. hysterophorus</i> leaf extract	186
7.4	Response of malondialdehyde and proline content of some crops and weeds to a foliar spray of <i>P. hysterophorus</i> leaf extract	193

7.5	Response of superoxide dismutase, catalase, and peroxidase activity of some crops and weeds to a foliar spray of <i>P. hysterophorus</i> leaf extract	197
8.1	Injury rating scale	211
8.2	Effect of Parthenium leaf methanol extract on the visual injury and plant height of different weeds	213
8.3	Effect of Parthenium leaf methanol extract on the fresh weight and dry weight of different weeds.	214
8.4	Effect of Parthenium leaf methanol extract on the plant height, fresh weight and dry weight of Bambara groundnut	215
8.5	Effect of Parthenium leaf methanol extract on the number of nuts plant ⁻¹ , pod weight plant ⁻¹ and seed yield plant ⁻¹ of Bambara groundnut	220
8.6	Effect of Parthenium leaf methanol extract on the seed yield m ⁻² , 100-seed weight and seed yield of Bambara groundnut	222

LIST OF FIGURES

Figure		Page
2.1	Traditional local delicacies in different African countries prepared from Bambara groundnut	4
2.2	Bambara groundnut field, vegetative stage and seed	6
2.3	The different plant parts and rosette stage of <i>P. hysterophorus</i>	13
2.4	The life cycle of Parthenium weed	13
2.5	Economic benefits of <i>P. hysterophorus</i>	19
2.6	(Four) main pathways of allelochemical releasing: volatilization (V), leaching (L), descomposition (D) and root exudation (E)	27
2.7	Possible mechanisms of allelochemicals release and transformation	28
2.8	The flow diagram of mode of action of herbicide	29
3.1	Dendrogram showing the mean EC ₅₀ values of seed germination, radicle and hypocotyle length of all indicator plants treated with the leaf, stem and flower extracts of <i>P. hysterophorus</i> with methanol revealed by non-overlapping (SAHN) UPGMA method	50
3.2a	Principal component analysis (PCA)-2D graphical relationship among the indicator plants treated with Parthenium leaf, stem and flower with methanol extract based on Euclidian distance	51
3.2b	Principal component analysis (PCA)-3D graphical relationship among the indicator plants treated with Parthenium leaf, stem and flower with methanol extract based on Euclidian distance	52
3.3	Dendrogram showing the mean EC ₅₀ values of seed germination, radicle and hypocotyle length of all indicator plants treated with the leaf, stem and flower extracts of <i>P. hysterophorus</i> with hexane revealed by non-overlapping (SAHN) UPGMA method	65
3.4a	Principal component analysis (PCA)-2D graphical relationship among the indicator plants treated with Parthenium leaf, stem and flower with methanol extract based on Euclidian distance	66
3.4b	Principal component analysis (PCA)-3D graphical relationship among the indicator plants treated with Parthenium leaf, stem and flower with methanol extract based on Euclidian distance	67

3.5	Dendrogram showing the mean EC ₅₀ values of seed germination, radicle and hypocotyle length of all indicator plants treated with the leaf, stem and flower extracts of <i>P. hysterophorus</i> with Acetone revealed by non-overlapping (SAHN) UPGMA method	78
3.6a	Principal component analysis (PCA)-2D graphical relationship among the indicator plants treated with Parthenium leaf, stem and flower with Acetone extract based on Euclidian distance	79
3.6b	Principal component analysis (PCA)-3D graphical relationship among the indicator plants treated with Parthenium leaf, stem and flower with Acetone extract based on Euclidian distance	79
3.7	Dendrogram showing the mean EC ₅₀ values of seed germination, radicle and hypocotyle length of all indicator plants treated with the leaf, stem and flower extracts of <i>P. hysterophorus</i> with Water revealed by non-overlapping (SAHN) UPGMA method	91
3.8a	Principal component analysis (PCA)-2D graphical relationship among the indicator plants treated with Parthenium leaf, stem and flower with water extract based on Euclidian distance	92
3.8b	Principal component analysis (PCA)-3D graphical relationship among the indicator plants treated with Parthenium leaf, stem and flower with water extract based on Euclidian distance	92
6.1	Dendrogram showing the mean EC ₅₀ values of seed germination, root, and hypocotyl length of <i>D. sanguinalis</i> exposed with the identified allelochemicals revealed by non-overlapping (SAHN) UPGMA method	158
6.2a	Based on Euclidian distance, principal component analysis (PCA)-2D graphical relationship among the discovered allelochemicals (eigenvector)	158
6.2b	Based on Euclidian distance, principal component analysis (PCA)-3D graphical relationship among the discovered allelochemicals (eigenvalue)	159
6.3	Dendrogram showing the mean EC ₅₀ values of seed sprouting, root and hypocotyl length of <i>E. indica</i> treated with the identified allelochemicals revealed by non-overlapping (SAHN) UPGMA method.	165
6.4a	Based on Euclidian distance, principal component analysis (PCA)-2D graphical relationship among the discovered allelochemicals (eigenvector)	166

6.4b	Based on Euclidian distance, principal component analysis (PCA)-3D graphical relationship among the discovered allelochemicals (eigenvalue)	166
7.1	Malondialdehyde (MDA) content (% increase compared with control) of test crops and weeds treated with <i>P. hysterophorus</i> leaf methanol extract concentrations at the different exposure times. Values with the same letter among the tested crops and weeds at the same extract concentrations are not significantly different at $p>0.005$ by Tukeys HSD	192
7.2	Proline content (% increase compared with control) of test crops and weeds treated with <i>P. hysterophorus</i> leaf methanol extract concentrations at the different exposure times. Values with the same letter among the tested crops and weeds at the same extract concentrations are not significantly different at $p>0.005$ by Tukeys HSD	195
7.3	Activities of superoxide dismutase (SOD) (% increase compared with control) of test crops and weeds treated with <i>P. hysterophorus</i> leaf methanol extract concentrations at the different exposure times. Values with the same letter among the tested crops and weeds at the same extract concentrations are not significantly different at $p>0.005$ by Tukeys HSD	200
7.4	Activities of catalase (CAT) (% increase compared with control) of test crops and weeds treated with <i>P. hysterophorus</i> leaf methanol extract concentrations at the different exposure times. Values with the same letter among the tested crops and weeds at the same extract concentrations are not significantly different at $p>0.005$ by Tukeys HSD	201
7.5	Activities of peroxidase (POD) (% increase compared with control) of test crops and weeds treated with <i>P. hysterophorus</i> leaf methanol extract concentrations at the different exposure times. Values with the same letter among the tested crops and weeds at the same extract concentrations are not significantly different at $p>0.005$ by Tukeys HSD	202

LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
FW	Fresh weight
DW	Dry weight
cm	Centimetre
CRD	Completely randomize design
G	Gram
H	Hour
ha ⁻¹	Hectare
Kg	Kilogram
L	Liter
RT	Retention time
m	Meter
mg/L	Milligram per liter
ml	Milliliter
mm	Millimeter
ppm	Part per millions
ppt	Part per thousands
SD	Standard Division
Sp.	Species
T	Treatment
UPM	Universiti Putra Malaysia
USA	United States of America
°C	Degree Celsius
%	Percentage

<	Less than
>	More than
SOD	Superoxide dismutase
POD	Peroxidase
CAT	Catalase
MDA	Malondialdehyde
HAS	Hours after spray
ROS	Reactive oxygen species
RL	Root length
SL	Shoot length
Ger	Germination

CHAPTER 1

INTRODUCTION

Bambara groundnut (*Vigna subterranea* L.) is an indigenous African legume crop primarily grown for its seeds. It is becoming popular as a food crop in rural areas of many countries in Africa (Jeklin et al., 2011). As human food, it is considered as a complete food crop since it has sufficient levels of the N, P, K macronutrients. Compared to intensively grown legumes such as groundnut and soybean, it has a high carbohydrate content (55%), low oil levels (5-6%), and a protein content of 22% (Oyeleke et al., 2012), with higher concentrations of essential amino acids like methionine and lysine (Mazahib et al., 2013). Bambara groundnut is a crop similar in morphology and growth habit to groundnut (*Arachis hypogaea* L.) but with high level of nutritional qualities and is drought tolerant (Feldman et al., 2019).

Parthenium (*Parthenium hysterophorus* L.) weed is a mystic annual invasive plant prevalent in several tropical and sub-tropical countries (Cowie et al., 2020), threatening natural ecosystems and agroecosystems in over 30 countries worldwide (Adkins & Shabbir, 2014). *Parthenium hysterophorus* is a plant which has positive and negative impacts on human being. This plant first come out in Australia and later spread to the whole world (Khan & Fahad, 2020). Parthenium weed is now spread from India to most of its neighbouring countries. It is suspected that this weed spread to Nepal, Bhutan, Bangladesh and Pakistan through vehicles or as a contaminant of seed lots (Shabbir et al., 2012). This plant is a strongly allelopathic species (Singh et al., 2005) and this trait is important in its invasion and persistence in a wide range of native and non-native ecosystems. Parthenium weed contains a number of potential allelochemicals within its aerial parts (e.g. leaves) and in its roots, which can inhibit the germination and growth of a wide range of plant species, including native plants, as well as various crops and pasture species (Singh et al., 2005).

Allelopathy is a mechanism in which the chemical produced by the plant may increase or decrease the plant growth (Sturm et al., 2016). Allelopathy is considered as an effective, economical and environment friendly weed management approach. The release of allelopathic compounds from leaves, flowers, seeds, stems and roots of living and decomposing plant materials can influence weed density and growth (Sathishkumar et al., 2020). The harmful chemicals released by allelopathic plants are known as allelochemicals. Some allelochemicals change the amount of chlorophyll production in a plant and it can be used to control weeds in agricultural systems by using allelopathic crops for intercropping, crop rotation, or mulching (Rehman et al., 2019). The allelopathic compounds can be used as natural herbicides and other pesticides; they are less disruptive to the global ecosystem than synthetic agrochemicals (Shahena et al., 2020).

Khan et al., (2012) remarked the inhibited seed germination and the seedling growth of wheat cultivar due to allelopathic effects of root, stem and leaf extracts of parthenium.

Hassan et al., (2018) reported that with increasing concentration of Parthenium weed extract, the germination, root length and shoot length of weeds decreased significantly. Leaves of Parthenium may be a source of natural weedicide which can help to control invasive plants (Maharjan et al., 2007). (Msafiri et al., 2013) revealed that significant allelopathic effects of aqueous extract of *P. hysterophorus* leaf and seed on seed germination, roots and shoot length, fresh and dry weight of crops. Aqueous extract of *P. hysterophorus* and *D. alba* decreased the seed germination, shoot length, fresh and dry biomass of different crops (Afridi & Khan, 2015). The higher concentration of Parthenium leaf extract decreased the photosynthetic pigments, carbohydrate, and enzymes in *Vigna aconitifolia* L. (Patil Bhimarao & Khade Hemlata, 2017). Seed germination and growth performance of chickpea were low at high concentrations of leaf extract of Parthenium (Shikha & Jha, 2016).

With the increasing trend of world population and decreasing available resources, weed management to protect crops is even more important as well as a challenging job. Weed management can be done mechanically, chemically or biologically. Mechanical weed control is very expensive, chemical weed control leads to environmental pollution and results in the evolution of herbicide resistance in weeds (Sharma, Kumar, et al., 2019). Biological weed control especially using the mechanism of allelopathy is well-adapted and a promising science (Petit et al., 2018). Shafiq et al., (2020) conducted an experiment with different parts of Parthenium on seed germination and biomasses of *Cicer arietinum* L and found that aqueous extracts of root, leaves and flowers significantly reduced the germination and seedling growth. Devi et al., (2014) investigated the allelopathic effect of Parthenium on growth and productivity of *Z. mays* L. and its phytochemical screening they observed that increasing concentration of *P. hysterophorus* whole plant extract significantly inhibited the germination, shoot length, root length, seedling length and SVI of crops. Hassan et al., (2018) recorded allelopathic effect of Parthenium extract and residue on some agronomic crops and weeds. There is inadequate data with respect to the effect of Parthenium plant extract on the growth and seed quality of Bambara groundnut.

Bambara groundnut is a new crop for Malaysia and Bangladesh and there is no information available on the allelopathic suppression of weeds on Bambara groundnut. Another one, the studies on profiling of active compounds and physio-biochemical responses of Parthenium allelochemicals are scarce. An accurate and sustainable weed control is compulsory for maintaining food security throughout the world. Weeds caused a 30 percent yield loss on average, but inadequate management circumstances resulted in yield losses of up to 60 percent (Mhlanga et al., 2016). In case of Bambara groundnut yield losses of 35 to 80 percent, as well as lower pod and haulm yields, according to Priya et al., (2013).

The overuse of chemical herbicides not only causes the environmental pollution but also poses many other threats to human health. Most importantly, herbicides, with few exceptions, cannot be applied in fields in which crops are being grown organically. The use of bio-herbicides may serve to keep weed infestations low and improve the crop productivity across the globe. This technique will help to reduce the dependence on chemical herbicide, reducing the chances of herbicide resistance development in weeds,

reduce health hazards, environmental damage and strengthen national economy as well. Hence, exploring allelopathic potential of Parthenium, a combination of the laboratory and green house trials to investigate the allelopathic suppression of weeds by Parthenium in Bambara groundnut is a new avenue of research. This research is enriching the data pool and provides information to future study. With these ends in views the studies were conducted with the following objectives:

1. To evaluate pre- and post-herbicidal effects of various parts of Parthenium extracts in different solvents on weeds and crops.
2. To identify and quantify potential phytotoxic compounds in different parts of Parthenium weed under different solvents by HPLC analysis.
3. To evaluate phytotoxic potential of the identified compounds on seed germination and initial growth of weeds.
4. To determine the mode of action of Parthenium allelochemical in inhibiting plant growth.
5. To develop potential formulation of Parthenium extract and to compare its efficacy to commercial herbicides for weed control in Bambara groundnut.

REFERENCES

- Abbas, M. N., Rana, S. A., Mahmood-Ul-Hassan, M., Rana, N., & Iqbal, M. (2013). Phytochemical constituents of weeds: baseline study in mixed crop zone agroecosystem. *Pakistan Journal of Weed Science Research*, 19(2), 213–218.
- Abbas, T., Ahmad, A., Kamal, A., Nawaz, M. Y., Jamil, M. A., Saeed, T., Abid, M. A., Ali, H. A., & Ateeq, M. (2021). Ways to use allelopathic potential for weed management: a review. *International Journal Food Science and Agriculture*, 5(3), 492–498.
- Abbas, T., Tanveer, A., Khaliq, A., Safdar, M. E., & Nadeem, M. A. (2014). Allelopathic effects of aquatic weeds on germination and seedling growth of wheat. *Herbologia*, 14(2), 11–25.
- Abd-Elgawad, A. M., El Gendy, A. E. N. G., Assaeed, A. M., Al-Rowaily, S. L., Alharthi, A. S., Mohamed, T. A., Nassar, M. I., Dewir, Y. H., & Elshamy, A. I. (2021). Phytotoxic effects of plant essential oils: A systematic review and structure-activity relationship based on chemometric analyses. *Plants*, 10(1), 1–16.
- Abdel-Farid, I. B., Massoud, M. S., Al-Enazy, Y., Abdel Latef, A. A. H., Jahangir, M., & Gomaa, N. H. (2021). Allelopathic potential of *Haloxylon persicum* against wheat and black mustard with special reference to its phytochemical composition and antioxidant activity. *Agronomy*, 11(2), 289–293.
- Abdulkerim-Ute, J., & Legesse, B. (2016). *Parthenium hysterophorus* L: Distribution, impact, and possible mitigation measures in Ethiopia. *Tropical and Subtropical Agroecosystems*, 19(1), 61–72.
- Abhilasha, D., Quintana, N., Vivanco, J., & Joshi, J. (2008). Do allelopathic compounds in invasive *Solidago canadensis* sl restrain the native European flora? *Journal of Ecology*, 96(5), 993–1001.
- Abu Bakar, F. I., Abu Bakar, M. F., Abdullah, N., Endrini, S., & Fatmawati, S. (2020). Optimization of Extraction Conditions of Phytochemical Compounds and Anti-Gout Activity of *Euphorbia hirta* L. (Ara Tanah) Using Response Surface Methodology and Liquid Chromatography-Mass Spectrometry (LC-MS) Analysis. *Evidence-Based Complementary and Alternative Medicine*, 2020, 1–13.
- Achatz, M., & Rillig, M. C. (2014). Arbuscular mycorrhizal fungal hyphae enhance transport of the allelochemical juglone in the field. *Soil Biology and Biochemistry*, 78, 76–82.
- Adelakun, O. E., Oyelade, O. J., Ade-Omowaye, B. I. O., Adeyemi, I. A., & Van de Venter, M. (2009). Chemical composition and the antioxidative properties of Nigerian Okra Seed (*Abelmoschus esculentus* Moench) Flour. *Food and Chemical Toxicology*, 47(6), 1123–1126.

- Adeleke, O. R., Adiamo, O. Q., & Fawale, O. S. (2018). Nutritional, physicochemical, and functional properties of protein concentrate and isolate of newly-developed Bambara groundnut (*Vigna subterrenea* L.) cultivars. *Food Science & Nutrition*, 6(1), 229–242.
- Adkins, S., & Shabbir, A. (2014). Biology, ecology and management of the invasive parthenium weed (*Parthenium hysterophorus* L.). *Pest Management Science*, 70(7), 1023–1029.
- Adler, M. J., & Chase, C. A. (2007). Comparison of the allelopathic potential of leguminous summer cover crops: Cowpea, sunn hemp, and velvetbean. *HortScience*, 42(2), 289–293.
- Aebi, H. E. (1984). Catalase in vitro. *Methods of Enzymatic Analysis*, 105, 121–126.
- Afridi, R. A., & Khan, M. A. (2015). Comparative effect of water extract of *Parthenium hysterophorus*, *Datura alba*, *Phragmites australis* and *Oryza sativa* on weeds and wheat. *Sains Malaysiana*, 44(5), 693–699.
- Ahmad, J., Bagheri, R., Bashir, H., Affan Baig, M., Al-Huqail, A., Ibrahim, M. M., & Irfan Qureshi, M. (2018). Organ-Specific Phytochemical Profiling and Antioxidant Analysis of *Parthenium hysterophorus* L. *BioMed Research International*, 2018, 1–11.
- Ahn, J. K., & Chung, I. M. (2000). Allelopathic potential of rice hulls on germination and seedling growth of barnyardgrass. *Agronomy Journal*, 92(6), 1162–1167.
- Ajeigbe, H. A., Waliyar, F., Echekwu, C. A., Ayuba, K., Motagi, B. N., Eniayeju, D., & Inuwa, A. (2014). A Farmer's guide to groundnut production in Nigeria. *Patancheru*, 502(324), 36.
- Ajmal, M., Rao, R. A. K., Ahmad, R., & Khan, M. A. (2006). Adsorption studies on *Parthenium hysterophorus* weed: Removal and recovery of Cd(II) from wastewater. *Journal of Hazardous Materials*, 135(1–3), 242–248.
- Akemo, M. C., Regnier, E. E., & Bennett, M. A. (2000). Weed suppression in spring-sown rye (*Secale cereale*): Pea (*Pisum sativum*) cover crop mixes. *Weed Technology*, 545–549.
- Alam, M. A., Juraimi, A. S., Rafii, M. Y., Hamid, A. A., Aslani, F., & Hakim, M. A. (2016). Salinity-induced changes in the morphology and major mineral nutrient composition of purslane (*Portulaca oleracea* L.) accessions. *Biological Research*, 49(1), 1–19.
- Algangdaby, M. M., & Salama, M. (2018). Management of the noxious weed; *Medicago polymorpha* L. via allelopathy of some medicinal plants from Taif region, Saudi Arabia. *Saudi Journal of Biological Sciences*, 25(7), 1339–1347.
- Algangdaby, M. M., Salama, M., & El-Darier, S. M. (2018). Management of the noxious weed; *Medicago polymorpha* L. via allelopathy of some medicinal plants from

Taif region, Saudi Arabia. *Saudi Journal of Biological Sciences*, 25(7), 1339–1347.

- Alshareef, B. B., Mohamed, P., & Alaib, A. (2019). The third international conference on basic sciences and their applications investigation of Allelopathic potential of *Acacia nilotica* L. *The Third International Conference on Basic Sciences and Their Applications*, 1(1), 476–486.
- Amador-Vargas, S., Dominguez, M., León, G., Maldonado, B., Murillo, J., & Vides, G. L. (2014). Leaf-folding response of a sensitive plant shows context-dependent behavioral plasticity. *Plant Ecology*, 215(12), 1445–1454.
- Amarowicz, R., Cwalina-Ambroziak, B., Janiak, M. A., & Bogucka, B. (2020). Effect of N fertilization on the content of phenolic compounds in Jerusalem artichoke (*Helianthus tuberosus* L.) tubers and their antioxidant capacity. *Agronomy*, 10(8), 1215.
- Ambika, S. R. (2013). Multifaceted attributes of allelochemicals and mechanism of allelopathy. In *Allelopathy* (pp. 389–405). Springer.
- Ames, B. N., Shigenaga, M. K., & Hagen, T. M. (1993). Oxidants, antioxidants, and the degenerative diseases of aging. *Proceedings of the National Academy of Sciences*, 90(17), 7915–7922.
- Amini, S., Azizi, M., Joharchi, M. R., Shafei, M. N., Moradinezhad, F., & Fujii, Y. (2014). Determination of allelopathic potential in some medicinal and wild plant species of Iran by dish pack method. *Theoretical and Experimental Plant Physiology*, 26(3), 189–199.
- Anaya, A. L., Waller, G. R., Owuor, P. O., Friedman, J., Chou, C. H., Suzuki, T., Arroyo-Estrada, J. F., & Cruz-Ortega, R. (2002). The role of caffeine in the production decline due to autotoxicity in coffee and tea plantations. *Allelopathy: From Molecules to Ecosystems*. Enfield: Science Publishers Inc, 71–91.
- Ani, O., Onu, O., Okoro, G., & Uguru, M. (2018). Overview of biological methods of weed control. In *Biological approaches for controlling weeds* (pp. 1–12).
- Anjum, T., & Bajwa, R. (2007). Field appraisal of herbicide potential of sunflower leaf extract against *Rumex dentatus*. *Field Crops Research*, 100(2–3), 139–142.
- Ansari, M. A., Choudhury, B. U., Roy, S. S., Sharma, S. K., Saraswat, P. K., Mishra, R. K., Singh, I. M., Singh, A. L., Lal, B., & Prakash, N. (2021). Evaluation of groundnut (*Arachis hypogaea*) cultivars for destabilized ecosystem of north eastern hill region. *Legume Research-An International Journal*, 44(10), 1240–1246.
- Anteneh, N., & Esayas, M. (2011). Allelopathic effects of *Parthenium hysterophorus* L. aqueous extracts on soybean (*Glycine max* L.) and haricot bean (*Phaseolus vulgaris* L.) seed germination, shoot and root growth and dry matter production. *Journal of Applied Botany and Food Quality*, 84(2), 219–222.

- Anwar, M. F., Yadav, D., Kapoor, S., Chander, J., & Samim, M. (2015). Comparison of antibacterial activity of Ag nanoparticles synthesized from leaf extract of *Parthenium hysterophorus* L in aqueous media and gentamicin sulphate: in-vitro. *Drug Development and Industrial Pharmacy*, 41(1), 43–50.
- Apurva, P., Sinha, S. K., & Thakur, P. C. (2010). Composting an obnoxious weed, *Parthenium hysterophorus* L., with the help of a millipede, *Harpaghe haydeniana*. *Asian Journal of Experimental Biological Sciences*, 1(2), 337–343.
- Arafat, Y., Khalid, S., WenXiong, L., ChangXun, F., Sadia, S., Ali, N., & Azeem, S. (2015). Allelopathic evaluation of selected plants extract against broad and narrow leaves weeds and their associated crops. *Academia Journal of Agricultural Research*, 3(10), 226–234.
- Asaduzzaman, M., & Asao, T. (2020). Autotoxicity in strawberry under recycled hydroponics and its mitigation methods. *The Horticulture Journal*, 89(2), 124–137.
- Ashebir, B., Sharma, J. J., & Lisanwork, N. (2012). Allelopathic effects of aqueous extracts and plant residues of *Parthenium hysterophorus* L. on kabuli chickpea and sesame. *Ethiopian Journal of Weed Management*, 5, 13–26.
- Asiwe, J. (2021). Effects of row spacing and weed infestation on yield of bambara groundnut, *Vigna subterranea* (L.) Verdc. in South Africa. *Tropical Agriculture*, 97(4).
- Aslam, F., Khaliq, A., Matloob, A., Tanveer, A., Hussain, S., & Zahir, Z. A. (2017). Allelopathy in agro-ecosystems: a critical review of wheat allelopathy-concepts and implications. *Chemoecology*, 27(1), 1–24.
- Aslani, F., Juraimi, A. S., Ahmad-Hamdani, M. S., Hashemi, F. S. G., Alam, M. A., Hakim, M. A., & Uddin, M. K. (2016). Effects of *Tinospora tuberculata* leaf methanol extract on seedling growth of rice and associated weed species in hydroponic culture. *Journal of Integrative Agriculture*, 15(7), 1521–1531.
- Aslani, F., Juraimi, A. S., Ahmad-Hamdani, M. S., Omar, D., Alam, M. A., Hakim, M. A., & Uddin, M. K. (2013). Allelopathic effects of Batawali (*Tinospora tuberculata*) on germination and seedling growth of plants. *Research on Crops*, 14, 1222–1231.
- Aslani, F., Juraimi, A. S., Ahmad-Hamdani, M. S., Omar, D., Alam, M. A., Hashemi, F. S. G., Hakim, M. A., & Uddin, M. K. (2014). Allelopathic effect of methanol extracts from *Tinospora tuberculata* on selected crops and rice weeds. *Acta Agriculturae Scandinavica Section B: Soil and Plant Science*, 64(2), 165–177.
- Aziz, A. et al. (2021). Exploring the herbicidal potential of some weed species by using two distinct extraction methods. *Agricultural and Biological Research*, 37(1), 88–92.

- Baar, J., Ozinga, W. A., Sweers, I. L., & Kuyper, T. W. (1994). Stimulatory and inhibitory effects of needle litter and grass extracts on the growth of some ectomycorrhizal fungi. *Soil Biology and Biochemistry*, 26(8), 1073–1079.
- Badgajar, S. B., Patel, V. V., Bandivdekar, A. H., & Mahajan, R. T. (2014). Traditional uses, phytochemistry and pharmacology of *Ficus carica*: A review. *Pharmaceutical Biology*, 52(11), 1487–1503.
- Bajwa, A. A., Weston, P. A., Gurusinghe, S., Latif, S., Adkins, S. W., & Weston, L. A. (2020). Toxic potential and metabolic profiling of two Australian biotypes of the invasive plant parthenium weed (*Parthenium hysterophorus* L.). *Toxins*, 12(7), 447.
- Barkosky, R. R., & Einhellig, F. A. (2003). Allelopathic interference of plant-water relationships by para-hydroxybenzoic acid. *Botanical Bulletin of Academia Sinica*, 44, 53–58.
- Barkosky, R. R., Einhellig, F. A., & Butler, J. L. (2000). Caffeic acid-induced changes in plant-water relationships and photosynthesis in leafy spurge *Euphorbia esula*. *Journal of Chemical Ecology*, 26(9), 2095–2109.
- Barnes, J., Anderson, L. A., & Phillipson, J. D. (2002). *Herbal medicines: a guide for healthcare professionals* (2nd editio). Pharmaceutical Press, London, UK. pp. 530.
- Bashar, H. M., Juraimi, A. S., Ahmad-Hamdani, M. S., Uddin, M. K., Asib, N., Anwar, M., & Rahaman, F. (2021). A Mystic Weed, *Parthenium hysterophorus*: Threats, Potentials and Management. *Agronomy*, 11(8), 1514.
- Batish, D. R., Kaur, S., Singh, H. P., & Kohli, R. K. (2009). Nature of interference potential of leaf debris of *Ageratum conyzoides*. *Plant Growth Regulation*, 57(2), 137–144.
- Batish, D. R., Lavanya, K., Singh, H. P., & Kohli, R. K. (2007). Phenolic allelochemicals released by *Chenopodium murale* affect the growth, nodulation and macromolecule content in chickpea and pea. *Plant Growth Regulation*, 51(2), 119–128.
- Batish, D. R., Singh, H. P., & Kaur, S. (2001). Crop allelopathy and its role in ecological agriculture. *Journal of Crop Production*, 4(2), 121–161.
- Batish, D. R., Singh, H. P., Kaur, S., Kohli, R. K., & Yadav, S. S. (2008). Caffeic acid affects early growth, and morphogenetic response of hypocotyl cuttings of mung bean (*Phaseolus aureus*). *Journal of Plant Physiology*, 165(3), 297–305.
- Batish, D. R., Singh, H. P., Kohli, R. K., Saxena, D. B., & Kaur, S. (2002). Allelopathic effects of parthenin against two weedy species, *Avena fatua* and *Bidens pilosa*. *Environmental and Experimental Botany*, 47(2), 149–155.

- Batish, D. R., Singh, H. P., Rana, N., & Kohli, R. K. (2006). Assessment of allelopathic interference of *Chenopodium album* through its leachates, debris extracts, rhizosphere and amended soil. *Archives of Agronomy and Soil Science*, 52(6), 705–715.
- Batish, D. R., Singh, H. P., Setia, N., Kaur, S., & Kohli, R. K. (2002). Phytotoxic effect of Parthenium residues on the selected soil properties and growth of chickpea and radish. *Weed Biology and Management*, 2(2), 73–78.
- Batish, D. R., Singh, H. P., Setia, N., Kaur, S., & Kohli, R. K. (2006). Effect of 2-benzoxazolinone (BOA) on seedling growth and associated biochemical changes in mung bean (*Phaseolus aureus*). *Zeitschrift Fur Naturforschung - Section C Journal of Biosciences*, 61(9–10), 709–714.
- Baziramakenga, R., Leroux, G. D., & Simard, R. R. (1995). Effects of benzoic and cinnamic acids on membrane permeability of soybean roots. *Journal of Chemical Ecology*, 21(9), 1271–1285.
- Baziramakenga, R., Leroux, G. D., Simard, R. R., Nadeau, P., Baziramakenga, R., Leroux, G. D., Simard, R. R., & Nadeau, P. (1997). Allelopathic effects of phenolic acids on nucleic acid and protein levels in soybean seedlings. *Canadian Journal of Botany*, 75(3), 445–450.
- Baziramakenga, R., Simard, R. R., & Leroux, G. D. (1994). Effects of benzoic and cinnamic acids on growth, mineral composition, and chlorophyll content of soybean. *Journal of Chemical Ecology*, 20(11), 2821–2833.
- Belel, M. D., Halim, R. A., Rafii, M. Y., & Saud, H. M. (2014). Intercropping of corn with some selected legumes for improved forage production: A review. *Journal of Agricultural Science*, 6(3), 48.
- Belz, R. G., Reinhardt, C. F., Foxcroft, L. C., Hurle, K., & Singh, I. (2007). Residue allelopathy in *Parthenium hysterophorus* L.-Does parthenin play a leading role? *Crop Protection*, 26(3), 237–245.
- Bertin, C., Weston, L. A., Huang, T., Jander, G., Owens, T., Meinwald, J., & Schroeder, F. C. (2007). Grass roots chemistry: meta-tyrosine, an herbicidal nonprotein amino acid. *Proceedings of the National Academy of Sciences*, 104(43), 16964–16969.
- Bertolino, L. T., Caine, R. S., & Gray, J. E. (2019). Impact of stomatal density and morphology on water-use efficiency in a changing world. *Frontiers in Plant Science*, 10, 225.
- Bhadoria. (2011). Allelopathy: a natural way towards weed management. *Journal of Experimental Agriculture International*, 1(1), 7–20.
- Bhagawati, R., Paul, S., & Dambale, A. S. (2020). Utility potential of *Parthenium hysterophorus* for its strategic management. In *Advances in Agronomy* (Issue December, pp-1-136).

- BHAT, M. A. (2017). *Evaluation of allelopathic effect of aqueous leaf extract of dried leaves of Tecoma stans L on seed germination and biochemical changes in Vigna radiata L (green gram)*. A Dissertation submitted to Pondicherry University in partial fulfillment of the requirements for award of the degree of Masters of Science in Ecology and Environmental Sciences. Pondicherry University, Pondich. pp-1-50.
- Bhavyasree, P. G., & Xavier, T. S. (2022). Green synthesised copper and copper oxide based nanomaterials using plant extracts and their application in antimicrobial activity. *Current Research in Green and Sustainable Chemistry*, 5, 100249.
- Blumenthal, M. (2003). The ABC clinical guide to herbs. *Annals of Internal Medicine*, 139(6), 534.
- Bogatek, R., Oracz, K., & Gniazdowska, A. (2005). Ethylene and ABA production in germinating seeds during allelopathy stress. *Fourth World Congress in Allelopathy*, Warsaw Agricultural University, Nowoursynowska 159.
- Bonny, B. S., Adjoumani, K., Seka, D., Koffi, K. G., Kouonon, L. C., Koffi, K. K., & Bi, I. A. Z. (2019). Agromorphological divergence among four agro-ecological populations of Bambara groundnut (*Vigna subterranea* (L.) Verdc.) in Côte d'Ivoire. *Annals of Agricultural Sciences*, 64(1), 103–111.
- Boon, H., & Smith, M. (2004). The complete natural medicine guide to the 50 most common medicinal herbs. *Choice Reviews Online*, 42(01), 42.
- Braga, T. M., Rocha, L., Chung, T. Y., Oliveira, R. F., Pinho, C., Oliveira, A. I., Morgado, J., & Cruz, A. (2020). Biological activities of gedunin—A limonoid from the Meliaceae family. *Molecules*, 25(3), 493.
- Brennan, E. B., & Smith, R. F. (2005). Winter cover crop growth and weed suppression on the central coast of California. *Weed Technology*, 19(4), 1017–1024.
- Brink, M., Belay, G., & De Wet, J. M. J. (2006). Plant resources of tropical Africa. *Cereals and Pulses Prota Foundation/Backhuys Publishers/CTA, Wageningen*, 58(2), 129–133.
- Burrill, L. C., Cárdenas, J., & Locatelli, E. (1976). Field manual for weed control research. *International Plant Protection Center, Oregon State University, Corvallis, Oregon, USA*, 63 pp.
- Carballido, J., & Agüera, J. (2013). Field sprayer for inter- and intra-row weed control : performance and labor savings. *Spanish Journal of Agricultural Research*, 11(3), 642–651.
- Caverzan, A., Passaia, G., Rosa, S. B., Ribeiro, C. W., Lazzarotto, F., & Margis-Pinheiro, M. (2012). Plant responses to stresses: role of ascorbate peroxidase in the antioxidant protection. *Genetics and Molecular Biology*, 35(4), 1011–1019.

- Chakraborty, S., Rao, P. S., & Mishra, H. N. (2014). Effect of pH on enzyme inactivation kinetics in high-pressure processed pineapple (*Ananas comosus* L.) puree using response surface methodology. *Food and Bioprocess Technology*, 7(12), 3629–3645.
- Chaudhry, S., & Sidhu, G. P. S. (2021). Climate change regulated abiotic stress mechanisms in plants: a comprehensive review. *Plant Cell Reports*, 1–31.
- Chauhan, B. S. (2013). Strategies to manage weedy rice in Asia. *Crop Protection*, 48, 51–56.
- Chauhan, B. S., & Johnson, D. E. (2008). Germination Ecology of Goosegrass (*Eleusine indica*): An Important Grass Weed of Rainfed Rice. *Weed Science*, 56(5), 699–706.
- Cheng, H. H. (1992). A conceptual framework for assessing allelochemicals in the soil environment. In *Allelopathy* (pp. 21–29). Springer.
- Chiapusio, G., Pellissier, F., & Gallet, C. (2004). Uptake and translocation of phytochemical 2-benzoxazolinone (BOA) in radish seeds and seedlings. *Journal of Experimental Botany*, 55(402), 1587–1592.
- Chibarabada, T. P., Modi, A. T., & Mabhaudhi, T. (2017). Expounding the value of grain legumes in the semi-and arid tropics. *Sustainability*, 9(1), 60.
- Chin, D. Van. (2001). Biology and management of barnyardgrass, red sprangletop and weedy rice. *Weed Biology and Management*, 1(1), 37–41.
- Chinedu, S. N., & Nwinyi, C. O. (2012). Proximate analysis of *Sphenostylis stenocarpa* and *Voadzeia subterranean* consumed in South-Eastern Nigeria. *Journal of Agricultural Biotechnology and Sustainable Development*, 4(1), 1–6.
- Chopra, N., Tewari, G., Tewari, L. M., Upreti, B., & Pandey, N. (2017). Allelopathic effect of *Echinochloa colona* L. and *Cyperus iria* L. weed extracts on the seed germination and seedling growth of rice and soyabean. *Advances in Agriculture*, 2017, 1–5.
- Chou, C.-H. (1980). Allelopathic researches in the subtropical vegetation in Taiwan. *Comparative Physiology and Ecology*, 5(4), 222–234.
- Chuah, T. S., & Lim, W. K. (2015). Assessment of phytotoxic potential of oil palm leaflet, rachis and frond extracts and powders on goosegrass (*Eleusine indica* (L.) Gaertn.) germination, emergence and seedling growth. *Malaysian Applied Biology*, 44(2), 75–84.
- Chuah, T. S., & Lim, W. K. (2021). Combination ratio affects synergistic activity of oil palm frond residue and S-metolachlor on goosegrass (*Eleusine Indica*). *Pakistan Journal of Botany*, 53(4), 1473–1477.

- Clewis, S. B., Everman, W. J., Jordan, D. L., & Wilcut, J. W. (2007). Weed management in North Carolina peanuts (*Arachis hypogaea*) with S-metolachlor, diclosulam, flumioxazin, and sulfentrazone systems. *Weed Technology*, 21(3), 629–635.
- Cowie, B. W., Byrne, M. J., Witkowski, E. T. F., Strathie, L. W., Goodall, J. M., & Venter, N. (2020). Parthenium avoids drought: Understanding the morphological and physiological responses of the invasive herb *Parthenium hysterophorus* to progressive water stress. *Environmental and Experimental Botany*, 171, 103945.
- da Silva, R. F., Bressan, R. T., Zilli, B. M., Pilatti, M. A., de Souza, S. N. M., & Santos, R. F. (2016). Allelopathic effect of aqueous extract of fresh leaf castor beans (*Ricinus communis* L.) applied to the beginning stage of soy (*Glycine max* L.) and safflower (*Carthamus tinctorius* L.). *African Journal of Biotechnology*, 15(49), 2787–2793.
- Dąbrowski, P., Baczevska-Dąbrowska, A. H., Kalaji, H. M., Goltsev, V., Paunov, M., Rapacz, M., Wójcik-Jagła, M., Pawluśkiewicz, B., Bąba, W., & Brestic, M. (2019). Exploration of chlorophyll a fluorescence and plant gas exchange parameters as indicators of drought tolerance in perennial ryegrass. *Sensors*, 19(12), 2736.
- Dahiya, S., Kumar, S., Khedwal, R. S., & Jakhar, S. R. (2017). Allelopathy for sustainable weed management. *Journal of Pharmacognosy and Phytochemistry*, 6, 832–837.
- Dalton, B. R., Blum, U., & Weed, S. B. (1989). Differential Sorption of Exogenously Applied Ferulic, p-Coumaric, p-Hydroxybenzoic, and Vanillic Acids in Soil. *Soil Science Society of America Journal*, 53(3), 757–762.
- Das, A., Raychaudhuri, U., & Chakraborty, R. (2012). Effect of freeze drying and oven drying on antioxidant properties of fresh wheatgrass. *International Journal of Food Sciences and Nutrition*, 63(6), 718–721.
- Das, B., Jangilia, P., Srilathaa, M., Kashannaa, J., & Dasb, R. (2013). Pseudoguaianolides from a collection of the flowers of *Parthenium hysterophorus* Linn.(Compositae). *Journal of Organic and Biomolecular Chemistry*, 1, 195–200.
- Das, B., Mahender, G., Rao, Y. K., Ramesh, C., Venkateswarlu, K., Ravikumar, K., Geethangili, M., & Tzeng, Y. (2006). Pseudoguaianolides from the flowers of *Parthenium hysterophorus*. *Helvetica Chimica Acta*, 89(2), 285–290.
- Das, B., Salvanna, N., Kumar, R. A., & Das, R. (2020). Our Phytochemical Research on *Parthenium hysterophorus*. *Mini-Reviews in Organic Chemistry*, 17(7), 843–854.
- Dass, A., Shekhawat, K., Choudhary, A. K., Sepat, S., Rathore, S. S., Mahajan, G., & Chauhan, B. S. (2017). Weed management in rice using crop competition-a review. *Crop Protection*, 95, 45–52.

- Datta, S., & Saxena, D. B. (2001). Pesticidal properties of parthenin (from *Parthenium hysterophorus*) and related compounds. *Pest Management Science*, 57(1), 95–101.
- Dayan, F. E., Owens, D. K., & Duke, S. O. (2012). Rationale for a natural products approach to herbicide discovery. *Pest Management Science*, 68(4), 519–528.
- De Albuquerque, M. B., Dos Santos, R. C., Lima, L. M., Melo Filho, P. D. A., Nogueira, R. J. M. C., Da Câmara, C. A. G., & Ramos, A. D. R. (2011). Allelopathy, an alternative tool to improve cropping systems. A review. *Agronomy for Sustainable Development*, 31(2), 379–395.
- de Oliveira Júnior, R. G., Bonnet, A., Braconnier, E., Groult, H., Prunier, G., Beaugard, L., Grougnet, R., da Silva Almeida, J. R. G., Ferraz, C. A. A., & Picot, L. (2019). Bixin, an apocarotenoid isolated from *Bixa orellana* L., sensitizes human melanoma cells to dacarbazine-induced apoptosis through ROS-mediated cytotoxicity. *Food and Chemical Toxicology*, 125, 549–561.
- Demissie, A. G., Ashenafi, A., Arega, A., Etenash, U., Kebede, A., & Tigist, A. (2013). Effect of *Parthenium hysterophorus* L. on germination and elongation of onion (*Allium cepa*) and bean (*Phaseolus vulgaris*). *Research Journal of Chemical and Environmental Sciences*, 1(2), 17–21.
- Desoky, E.-S. M., Merwad, A.-R., Abo El-Maati, M. F., Mansour, E., Arnaout, S. M. A. I., Awad, M. F., Ramadan, M. F., & Ibrahim, S. A. (2021). Physiological and biochemical mechanisms of exogenously applied selenium for alleviating destructive impacts induced by salinity stress in bread wheat. *Agronomy*, 11(5), 926.
- Devi, O. I., Dutta, B. K., & Zea, L. (2012). Allelopathic Effect of the Aqueous Extract of *Parthenium hysterophorus* and *Chromolaena Odorata* on the Seed Germination and Seedling Vigour of *Zea mays* L. In vitro. *Academic Journal of Plant Sciences*, 5(4), 110–113.
- Devi, Y. N., Dutta, B. K., Sagolshemcha, R. S., & Irabanta, N. (2014). Allelopathic effect of *Parthenium hysterophorus* L. on growth and productivity of *Zea mays* L. and its phytochemical screening. *International Journal of Current Microbiology and Applied Sciences*, 3(7), 837–846.
- Dey, P., Chandra, S., Chatterjee, P., & Bhattacharya, S. (2012). Allelopathic Potential of Aerial Parts from *Mikania scandens* (L.) Willd. *World Journal of Agricultural Sciences*, 8(2), 203–207.
- Dhawan, D., & Gupta, J. (2017). Comparison of Different Solvents for Phytochemical Extraction Potential from *Datura metel* Plant Leaves. *International Journal of Biological Chemistry*, 11(1), 17–22.
- Dhileepan, K. (2007). Biological control of parthenium (*Parthenium hysterophorus*) in Australian rangeland translates to improved grass production. *Weed Science*, 55(5), 497–501.

- Dhileepan, K., Callander, J., Shi, B., & Osunkoya, O. O. (2018). Biological control of parthenium (*Parthenium hysterophorus*): the Australian experience. *Biocontrol Science and Technology*, 28(10), 970–988.
- Di Mola, I., Rouphael, Y., Ottaiano, L., Duri, L. G., Mori, M., & De Pascale, S. (2017). Assessing the effects of salinity on yield, leaf gas exchange and nutritional quality of spring greenhouse lettuce. *International Symposium on New Technologies for Environment Control, Energy-Saving and Crop Production in Greenhouse and Plant 1227*, 479–484.
- Dilipkumar, M., Chuah, T. S., Goh, S. S., & Sahid, I. (2020). Weed management issues, challenges, and opportunities in Malaysia. *Crop Protection*, 134, 104347.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S., & Ju, Y.-H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Linnophila aromatica*. *Journal of Food and Drug Analysis*, 22(3), 296–302.
- Dodia, D. A., Patel, I. S., & Patel, G. M. (2010). Botanical pesticides for pest management. In *Wiley* (pp. 181–193).
- Doganlar, Z. B. (2012). Quizalofop-p-ethyl-induced phytotoxicity and genotoxicity in *Lemna minor* and *Lemna gibba*. *Journal of Environmental Science and Health - Part A Toxic/Hazardous Substances and Environmental Engineering*, 47(11), 1631–1643.
- Dogra, K. S., & Sood, S. K. (2012). Phytotoxicity of *Parthenium hysterophorus* residues towards growth of three native plant species (*Acacia catechu* willd, *Achyranthes aspera* L. and *Cassia tora* L.) in Himachal Pradesh, India. *International Journal of Plant Physiology and Biochemistry*, 4(5), 105–109.
- Eche, C. O., Vabi, M. B., Ekefan, E. J., Ajeigbe, H. A., & Ocholi, F. A. (2017). Evaluation of different fertilizer sources for the management of aflatoxin contamination in groundnut (*arachis hypogaea* l.) in the southern guinea savannah agro-ecological zone of Nigeria. *International Journal of Advanced Research*, 5(11), 967–976.
- Einhellig, F. A., Galindo, J. C. G., Molinillo, J. M. G., & Cutler, H. G. (2004). Mode of allelochemical action of phenolic compounds. In *Allelopathy: Chemistry and mode of action of allelochemicals* (pp. 217–238). CRC Press Boca Raton, FL.
- El-Mergawi, R. A., & Al-Humaid, A. I. (2019). Searching for natural herbicides in methanol extracts of eight plant species. *Bulletin of the National Research Centre*, 43(1), 1–6.
- El Omari, B., & El Ghachtouli, N. (2021). Arbuscular mycorrhizal fungi-weeds interaction in cropping and unmanaged ecosystems: a review. *Symbiosis*, 83(3), 279–292.

- Elhaak, M. A., Ahmed, F. A., Kashlana, M. E., & Saad-Allah, K. M. (2014). Diversity of *Varthemia candicans* phytochemicals in response to growth habitat. *Asian Journal of Microbiology, Biotechnology and Environmental Sciences*, 5(4), 264–278.
- Elsheikh, E. A. E., El-Keblawy, A., Mosa, K. A., Okoh, A. I., & Saadoun, I. (2021). Role of Endophytes and Rhizosphere Microbes in Promoting the Invasion of Exotic Plants in Arid and Semi-Arid Areas: A Review. *Sustainability*, 13(23), 13081.
- Evans, H. C. (1997). *Parthenium hysterophorus*. a review of its weed status and the possibilities for biological control. *Biocontrol News and Information*, 18(3), 89–98.
- Evans, J. A., Williams, A., Hager, A. G., Mirsky, S. B., Tranel, P. J., & Davis, A. S. (2018). Confronting herbicide resistance with cooperative management. *Pest Management Science*, 74(11), 2424–2431.
- Farooq, M., Bajwa, A. A., Cheema, S. A., & Cheema, Z. A. (2013). Application of allelopathy in crop production. *International Journal of Agriculture and Biology*, 15(6), 1367–1378.
- Faustino, L. I., Bulfe, N. M. L., Pinazo, M. A., Monteoliva, S. E., & Graciano, C. (2013). Dry weight partitioning and hydraulic traits in young *Pinus taeda* trees fertilized with nitrogen and phosphorus in a subtropical area. *Tree Physiology*, 33(3), 241–251.
- Feldman, A. B., Ho, W. K., Massawe, F., & Mayes, S. (2019). Bambara groundnut is a climate-resilient crop: How could a drought-tolerant and nutritious legume improve community resilience in the face of climate change? In *Sustainable Solutions for Food Security: Combating Climate Change by Adaptation* (pp. 150–167). Springer International Publishing.
- Feng, G., Chen, M., Ye, H.-C., Zhang, Z.-K., Li, H., Chen, L.-L., Chen, X.-L., Yan, C., & Zhang, J. (2019). Herbicidal activities of compounds isolated from the medicinal plant *Piper sarmentosum*. *Industrial Crops and Products*, 132, 41–47.
- Ferguson, J. J., Rathinasabapathi, B., & Chase, C. A. (2013). Allelopathy: How Plants Suppress Other Plants. *Edis*, 3(3), 1–5.
- Filippou, P., Bouchagier, P., Skotti, E., & Fotopoulos, V. (2014). Proline and reactive oxygen/nitrogen species metabolism is involved in the tolerant response of the invasive plant species *Ailanthus altissima* to drought and salinity. *Environmental and Experimental Botany*, 97, 1–10.
- Franco, D. M., Silva, E. M., Saldanha, L. L., Adachi, S. A., Schley, T. R., Rodrigues, T. M., Dokkedal, A. L., Nogueira, F. T. S., & de Almeida, L. F. R. (2015). Flavonoids modify root growth and modulate expression of SHORT-ROOT and HD-ZIP III. *Journal of Plant Physiology*, 188, 89–95.

- Friebe, A., Roth, U., Kück, P., Schnabl, H., & Schulz, M. (1997). Effects of 2, 4-dihydroxy-1, 4-benzoxazin-3-ones on the activity of plasma membrane H⁺-ATPase. *Phytochemistry*, 44(6), 979–983.
- Fritz, D., Bernardi, A. P., Haas, J. S., Ascoli, B. M., Bordignon, S. A. de L., & Von Poser, G. (2007). Germination and growth inhibitory effects of *Hypericum myrianthum* and *H. polyanthemum* extracts on *Lactuca sativa* L. *Revista Brasileira de Farmacognosia*, 17(1), 44–48.
- Galindo, J. C. G., Hernández, A., Dayan, F. E., Tellez, M. R., Macías, F. A., Paul, R. N., & Duke, S. O. (1999). Dehydrozalanin C, a natural sesquiterpenolide, causes rapid plasma membrane leakage. *Phytochemistry*, 52(5), 805–813.
- García, S., Abadín, J., Couto-Vázquez, A., Martín, A., & González-Prieto, S. J. (2020). Bioavailability of macro-and micro-nutrients chemically extracted in acidic soils for wheat. *Journal of Plant Nutrition and Soil Science*, 183(6), 705–717.
- Garko, M. S., Adulmuminin, M. Y., Saad, A. M., & Abba, U. U. (2018). Growth and development of bambara groundnut (*Vigna subterranea* L.) as affected by phosphorus levels and weed control method in Sudan Savanna, Nigeria. *FUDMA JAAT*, 4(1), 24–31.
- Gerrano, A. S., Eifediyi, E. K., Labuschagne, M., Ogedegbe, F. O., & Hassen, A. I. (2021). Production Practices of Bambara Groundnut. In *Food and Potential Industrial Applications of Bambara Groundnut* (pp. 7–25). Springer.
- Ghareib, H. R. A., Abdelhamed, M. S., & Ibrahim, O. H. (2010). Antioxidative effects of the acetone fraction and vanillic acid from *Chenopodium murale* on tomato plants. *Weed Biology and Management*, 10(1), 64–72.
- Gianessi, L. P. (2013). The increasing importance of herbicides in worldwide crop production. *Pest Management Science*, 69(10), 1099–1105.
- Gill, S. S., & Tuteja, N. (2010). Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology and Biochemistry*, 48(12), 909–930.
- Gniazdowska, A., & Bogatek, R. (2005). Allelopathic interactions between plants. Multi site action of allelochemicals. *Acta Physiologiae Plantarum*, 27(3), 395–407.
- Gong, D. H., Wang, G. Z., Si, W. T., Zhou, Y., Liu, Z., & Jia, J. (2018). Effects of salt stress on photosynthetic pigments and activity of ribulose-1, 5-bisphosphate carboxylase/oxygenase in *Kalidium foliatum*. *Russian Journal of Plant Physiology*, 65(1), 98–103.
- Grisi, P. U., Gualtieri, S. C. J., Ranal, M. A., & Santana, D. G. (2012). Allelopathic interference of *Sapindus saponaria* root and mature leaf aqueous extracts on diaspore germination and seedling growth of *Lactuca sativa* and *Allium cepa*. *Brazilian Journal of Botany*, 35(1), 1–9.

- Grossmann, K., Hutzler, J., Tresch, S., Christiansen, N., Looser, R., & Ehrhardt, T. (2012). On the mode of action of the herbicides cinmethylin and 5-benzoyloxymethyl-1, 2-isoxazolines: putative inhibitors of plant tyrosine aminotransferase. *Pest Management Science*, 68(3), 482–492.
- Gulzar, A., & Siddiqui, M. B. (2014). Allelopathic effect of aqueous extracts of different part of *Eclipta alba* (L.) Hassk. on some crop and weed plants. *Journal of Agricultural Extension and Rural Development*, 6(1), 55–60.
- Gulzar, A., Siddiqui, M. B., & Bi, S. (2016). Phenolic acid allelochemicals induced morphological, ultrastructural, and cytological modification on *Cassia sophera* L. and *Allium cepa* L. *Protoplasma*, 253(5), 1211–1221.
- Guo, Y., Kim, K.-U., Yoder, J. I., & Shin, D. (2011). Parasitic plants as a new target plant for screening rice allelopathic potential. *Journal of Life Sciences*, 5(9), 201–210.
- Gupta, N., Martin, B. M., Metcalfe, D. D., & Rao, P. V. S. (1996). Identification of a novel hydroxyproline-rich glycoprotein as the major allergen in Parthenium pollen. *Journal of Allergy and Clinical Immunology*, 98(5), 903–912.
- Gupta, A. Sen, Heinen, J. L., Holaday, A. S., Burke, J. J., & Allen, R. D. (1993). Increased resistance to oxidative stress in transgenic plants that overexpress chloroplastic Cu/Zn superoxide dismutase. *Proceedings of the National Academy of Sciences*, 90(4), 1629–1633.
- Gurib-Fakim, A., Sewraj, M. D., Gueho, J., & Dulloo, E. (1996). Medicinal plants of Rodrigues. *International Journal of Pharmacognosy*, 34(1), 2–14.
- Han, C.-M., Pan, K.-W., Wu, N., Wang, J.-C., & Li, W. (2008). Allelopathic effect of ginger on seed germination and seedling growth of soybean and chive. *Scientia Horticulturae*, 116(3), 330–336.
- Hao, W., Ren, L., Ran, W., & Shen, Q. (2010). Allelopathic effects of root exudates from watermelon and rice plants on *Fusarium oxysporum* f. sp. niveum. *Plant and Soil*, 336(1), 485–497.
- Haq, R. A., Hussain, M., Cheema, Z. A., Mushtaq, M. N., & Farooq, M. (2010). Mulberry leaf water extract inhibits bermudagrass and promotes wheat growth. *Weed Biology and Management*, 10(4), 234–240.
- Harrathi, J., Attia, H., Neffati, M., Hosni, K., Marzouk, B., Lachâal, M., & Karray-Bouraoui, N. (2013). Salt effects on shoot growth and essential oil yield and composition in safflower (*Carthamus tinctorius* L.). *Journal of Essential Oil Research*, 25(6), 482–487.
- Hasan, M., Ahmad-Hamdani, M. S., Rosli, A. M., & Hamdan, H. (2021). Bioherbicides: An eco-friendly tool for sustainable weed management. *Plants*, 10(6), 1–21.

- Hasan, M., Uddin, M. K., Muda Mohamed, M. T., & Kee Zuan, A. T. (2018). Nitrogen and phosphorus management for bambara groundnut (*Vigna subterranea*) production-A review. *Legume Research: An International Journal*, 41(4), 483–489.
- Hasanuzzaman, M., Nahar, K., Anee, T. I., & Fujita, M. (2017). Exogenous silicon attenuates cadmium-induced oxidative stress in *Brassica napus* L. by modulating AsA-GSH pathway and glyoxalase system. *Frontiers in Plant Science*, 8, 1061.
- Hassan, G., Rashid, H. U., Amin, A., Khan, I. A., & Shehzad, N. (2018). Allelopathic effect of *Parthenium hysterophorus* on germination and growth of some important crops and weeds of economic importance. *Planta Daninha*, 36(0), 1–11.
- Hassan, S. M., & Ghareib, H. R. (2009). Bioactivity of *Ulva lactuca* L. acetone extract on germination and growth of lettuce and tomato plants. *African Journal of Biotechnology*, 8(16), 3832–3838.
- Hayat, S., Hayat, Q., Alyemeni, M. N., Wani, A. S., Pichtel, J., & Ahmad, A. (2012). Role of proline under changing environments: a review. *Plant Signaling & Behavior*, 7(11), 1456–1466.
- Holt, J. S., Welles, S. R., Silvera, K., Heap, I. M., Heredia, S. M., Martinez-Berdeja, A., Palenscar, K. T., Sweet, L. C., & Ellstrand, N. C. (2013). Taxonomic and life history bias in herbicide resistant weeds: implications for deployment of resistant crops. *PLoS ONE*, 8(9), 71916.
- Hong, N. H., Xuan, T. D., Eiji, T., Hiroyuki, T., Mitsuhiro, M., & Khanh, T. D. (2003). Screening for allelopathic potential of higher plants from Southeast Asia. *Crop Protection*, 22(6), 829–836.
- Hoppin, J. A. (2014). Pesticides and respiratory health: where do we go from here? *Occupational and Environmental Medicine*, 71(2), 80–80.
- Hordiei, K. R., Gontova, T. M., Serbin, A. G., Kotov, A. G., & Kotova, E. E. (2019). Study of phenolic compounds in the Feverfew Herb by TLC and HPLC methods. *Ukrainian Biopharmaceutical Journal*, 3(60), 64–70.
- Huihui, Z., Xin, L., Zisong, X., Yue, W., Zhiyuan, T., Meijun, A., Yuehui, Z., Wenxu, Z., Nan, X., & Guangyu, S. (2020). Toxic effects of heavy metals Pb and Cd on mulberry (*Morus alba* L.) seedling leaves: photosynthetic function and reactive oxygen species (ROS) metabolism responses. *Ecotoxicology and Environmental Safety*, 195(0), 110469.
- Hussain, M. I., & Reigosa, M. J. (2011). Allelochemical stress inhibits growth, leaf water relations, PSII photochemistry, non-photochemical fluorescence quenching, and heat energy dissipation in three C3 perennial species. *Journal of Experimental Botany*, 62(13), 4533–4545.

- Iderawumi, A. M., & Friday, C. E. (2018). Characteristics effects of weed on growth performance and yield of maize (*zea mays*). *Biomedical Journal*, 1, 4.
- Ijarotimi, O. S., & Esho, T. R. (2009). Comparison of nutritional composition and anti-nutrient status of fermented, germinated and roasted bambara groundnut seeds (*vigna subterranea*). *British Food Journal*, 111(4), 376–386.
- Imatomi, M., Novaes, P., & Gualtieri, S. C. J. (2013). Interspecific variation in the allelopathic potential of the family Myrtaceae. *Acta Botanica Brasilica*, 27(1), 54–61.
- Inderjit, W., & DA, K. (n.d.). The Ecosystem and Evolutionary Contexts of Allelopathy. *Trends Ecology and Evolution*, 26(12), 655–662.
- Ishnava, K. B., Chauhan, J. B., Garg, A. A., & Thakkar, A. M. (2012). Antibacterial and phytochemical studies on *Calotropis gigantea* (L.) R. Br. latex against selected cariogenic bacteria. *Saudi Journal of Biological Sciences*, 19(1), 87–91.
- Islam, A., Hasan, M. M., Yeasmin, S., Abedin, M. A., Kader, M. A., Rashid, M. H. O., & Anwar, M. P. (2019). Bioassay screening of sawdust obtained from selected tropical tree species for allelopathic properties and their field performance against paddy weeds. *Fundamental and Applied Agriculture*, 4(0), 906–915.
- Islam, A. K. M., & Kato-Noguchi, H. (2014). Phytotoxic activity of *Ocimum tenuiflorum* extracts on germination and seedling growth of different plant species. *The Scientific World Journal*, 2014, 1–8.
- Ismail, B. S., & Siddique, M. A. B. (2011). The inhibitory effect of grasshopper's cyperus (*Cyperus iria* L.) on the seedling growth of five Malaysian rice varieties. *Tropical Life Sciences Research*, 22(1), 81.
- Ivanescu, B., Miron, A., & Corciova, A. (2015). Sesquiterpene Lactones from Artemisia Genus: Biological Activities and Methods of Analysis. *Journal of Analytical Methods in Chemistry*, 2015, 1–21.
- Jabran, K. (2017). Manipulation of allelopathic crops for weed control. In *Springer Briefs in Plant Science*. Springer nature switzerland (pp. 50-87).
- Jabran, K., Cheema, Z. A., Farooq, M., & Hussain, M. (2010). Lower doses of pendimethalin mixed with allelopathic crop water extracts for weed management in canola (*Brassica napus*). *International Journal of Agriculture and Biology*, 12(3), 335–340.
- Jabran, K., Mahajan, G., Sardana, V., & Chauhan, B. S. (2015). Allelopathy for weed control in agricultural systems. *Crop Protection*, 72, 57–65.
- Jat, R. S., Meena, H. N., Singh, A. L., Surya, J. N., & Misra, J. B. (2011). Weed management in groundnut (*Arachis hypogaea* L.) in India-a review. *Agricultural Reviews*, 32(3), 155–171.

- Javaid, A., & Anjum, T. (2005). *Parthenium hysterophorus* L. - A noxious alien weed. *Pakistan Journal of Weed Science Research*, 11(3–4), 81–87.
- Javaid, A., Shafique, S. S., & Shafique, S. S. (2011). Management of *Parthenium hysterophorus* (Asteraceae) by *Withania somnifera* (Solanaceae). *Natural Product Research*, 25(4), 407–416.
- Javaid, A., Shafique, S., & Shafique, S. (2007). Causes of rapid spread of *Parthenium hysterophorus* L. in Pakistan and possible control measures-a review. *Pakistan Journal of Botany*, 39, 2611–2618.
- Javaid, A., Shafique, S., & Shafique, S. (2008). Herbicidal activity of *Datura metel* L. against *Phalaris minor* Retz. *Pakistan Journal of Weed Science Research*, 14(3–4), 209–220.
- Jayanth, K. P., & Visalakshy, P. N. G. (1994). Dispersal of the parthenium beetle *Zygogramma bicolorata* (Chrysomelidae) in India. *Biocontrol Science and Technology*, 4(3), 363–365.
- Jeklin, A. (2016). Assessing the morphological variation and characterising the proteins of bambara groundnut (*Vigna Subterranea* L. Verdc). *PhD Diss, Department of Biotechnology, Faculty of Applied and Computer Sciences, Vaal University of Technology, July*, 1–23.
- Jeklin, A., Vurayai, R., Emongor, V., & Moseki, B. (2011). Physiological responses of bambara groundnut (*Vigna subterranea* L. Verdc) to short periods of water stress during Different developmental stages. *Asian Journal of Agricultural Sciences*, 3(July), 1–23.
- Jensen, L. B., Courtois, B., Shen, L., Li, Z., Olofsdotter, M., & Mauleon, R. P. (2001). Locating genes controlling allelopathic effects against barnyardgrass in upland rice. *Agronomy Journal*, 93(1), 21–26.
- Jhala, A., Rathod, P. H., Patel, K. C., & Van, P. D. (2005). Growth and yield of groundnut (*Arachis hypogaea* L.) as influenced by weed management practices and Rhizobium inoculation. *Communications in Agricultural and Applied Biological Sciences*, 70(3), 493–500.
- Jiang, A., Zuo, J., Zheng, Q., Guo, L., Gao, L., Zhao, S., Wang, Q., & Hu, W. (2019). Red LED irradiation maintains the postharvest quality of broccoli by elevating antioxidant enzyme activity and reducing the expression of senescence-related genes. *Scientia Horticulturae*, 251, 73–79.
- Jideani, V. A., & Jideani, A. I. O. (2021). Ethnonutritional and ethnomedicinal uses of bambara groundnut. In *Bambara groundnut: Utilization and Future Prospects* (pp. 49–60). Springer.
- Johnson, D. E. (1998). *Applied Multivariate Methods for Data Analysis*, 26– 27. Pacific Grove, Cal.: Brooks/Cole.

- Johra, F. T., Bepari, A. K., Bristy, A. T., & Reza, H. M. (2020). A mechanistic review of β -carotene, lutein, and zeaxanthin in eye health and disease. *Antioxidants*, 9(11), 1046.
- Jolly, R. J. H., & Sanderson, D. J. (1997). A Mohr circle construction for the opening of a pre-existing fracture. *Journal of Structural Geology*, 19(6), 887–892.
- Joshi, V. K., Joshi, A., & Dhiman, K. S. (2017). The Ayurvedic Pharmacopoeia of India, development and perspectives. *Journal of Ethnopharmacology*, 197, 32–38.
- Kamal, J. (2011). Impact of allelopathy of sunflower (*Helianthus annuus* L.) roots extract on physiology of wheat (*Triticum aestivum* L.). *African Journal of Biotechnology*, 10(65), 14465–14477.
- Kanaujiya, D. K., Ansari, N., Dubey, S. K., Chaudhary, K., & Kishor, K. (2019). Allelopathic effect of the liquid extract of *Parthenium hysterophorus*, on microorganism. *International Journal of Advanced Research in Microbiology and Immunology*, 1(1), 1–5.
- Kanchan, S. D. (1980). Allelopathic effects of *Parthenium hysterophorus* L: exudation of inhibitors through roots [of beans]. *Plant and Soil (Netherlands)*, 53(1), 27–35.
- Kapoor, D., Tiwari, A., Sehgal, A., Landi, M., Brestic, M., & Sharma, A. (2019). Exploiting the allelopathic potential of aqueous leaf extracts of *Artemisia absinthium* and *Psidium guajava* against *Parthenium hysterophorus*, a widespread weed in India. *Plants*, 8(12), 552.
- Kaptsou, K. G., Njintang, Y. N., Nguemtchouin, M. M. G., Scher, J., Hounhouigan, J., & Mbofung, C. M. (2015). Physicochemical and micro-structural properties of flours, starch and proteins from two varieties of legumes: bambara groundnut (*Vigna subterranea*). *Journal of Food Science and Technology*, 52(8), 4915–4924.
- Kar, A., Choudhary, B. K., & Bandyopadhyay, N. G. (1999). Preliminary studies on the inorganic constituents of some indigenous hypoglycaemic herbs on oral glucose tolerance test. *Journal of Ethnopharmacology*, 64(2), 179–184.
- Karim, S. M. R. (2013). Obnoxious environmental pollutant, the parthenium weed: a possibility of ecological disaster in Malaysia. In *In: Joint seminar on Natural Disasters: Experiences in Malaysia and Indonesia* (Issue Faculty of Earth Science, Universiti Malaysia Kelantan, Jeli Campus, Malaysia.).
- Karim, S. M. R., Nurzafirah, Z., & Norhafizah, M. Z. (2017). Weed seed bank of *Parthenium hysterophorus* L. in Batang Kali, Selangor, Malaysia. *Pertanika Journal of Tropical Agricultural Science*, 40(4), 565–576.
- Kasar, B. A., & Chavhan, K. R. (2010). Effects of pendimethalin and different doses of ALS (Aceto lactate synthase) inhibitor on growth and yield of groundnut. *Crop Research (Hisar)*, 39(1/2/3), 65–67.

- Kaur, G., Kumar, S., Thakur, P., Malik, J. A., Bhandhari, K., Sharma, K. D., & Nayyar, H. (2011). Involvement of proline in response of chickpea (*Cicer arietinum* L.) to chilling stress at reproductive stage. *Scientia Horticulturae*, 128(3), 174–181.
- Kaur, S., Kaur, R., & Chauhan, B. S. (2018). Understanding crop-weed-fertilizer-water interactions and their implications for weed management in agricultural systems. *Crop Protection*, 103, 65–72.
- Kaur, S., Singh, H. P., Mittal, S., Batish, D. R., & Kohli, R. K. (2010). Phytotoxic effects of volatile oil from *Artemisia scoparia* against weeds and its possible use as a bioherbicide. *Industrial Crops and Products*, 32(1), 54–61.
- Kaya, A., & Yigit, E. (2014). The physiological and biochemical effects of salicylic acid on sunflowers (*Helianthus annuus*) exposed to flurochloridone. *Ecotoxicology and Environmental Safety*, 106, 232–238.
- Khaket, T. P., Aggarwal, H., Jodha, D., Dhanda, S., & Singh, J. (2015). *Parthenium hysterophorus* in current scenario: A toxic weed with industrial, agricultural and medicinal applications. *Journal of Plant Sciences*, 10(2), 42.
- Khaket, T. P., Singh, M., Dhanda, S., Singh, T., & Singh, J. (2012). Biochemical characterization of consortium compost of toxic weeds *Parthenium hysterophorus* and *Eichhornia crassipe*. *Bioresource Technology*, 123, 360–365.
- Khan, H., Marwat, K. B., Hassan, G., & Khan, M. A. (2012). Chemical control of *Parthenium hysterophorus* L. at different growth stages in non-cropped area. *Pakistan Journal of Botany*, 44(5), 1721–1726.
- Khan, M. B., Ahmad, M., Hussain, M., Jabran, K., Farooq, S., & Waqas-Ul-Haq, M. (2012). Allelopathic plant water extracts tank mixed with reduced doses of atrazine efficiently control *Trianthema portulacastrum* L. *Zea Mays*, 339–346.
- Khan, N., & Fahad, S. (2020). Economic review of *Parthenium Hysterophorus* L. plant in the world. *SSRN Electronic Journal*, 1–20.
- Khan, N., George, D., Shabbir, A., & Adkins, S. W. (2019). Suppressive plants as weed management tool: Managing *Parthenium hysterophorus* under simulated grazing in Australian grasslands. *Journal of Environmental Management*, 247, 224–233.
- Khan, N., Naveed, K., Hussain, Z., & Alam Khan, S. (2012). Assessment of allelopathic effects of *Parthenium* (*Parthenium hysterophorus* L.) plant parts on seed germination and seedling growth of wheat (*Triticum aestivum* L.) cultivars. *Pakistan Journal of Weed Science Research*, 18(1).
- Khan, R. A., Ahmed, M., Khan, M. R., Yasir, M., Muhammad, B., & Khan, R. (2011). Nutritional investigation and biological activities of *Parthenium hysterophorus*. *African Journal of Pharmacy and Pharmacology*, 5(18), 2073–2078.

- Khan, R. A., Khan, M. R., Sahreen, S., & Bokhari, J. (2010). Antimicrobial and phytotoxic screening of various fractions of *Sonchus asper*. *African Journal of Biotechnology*, 9(25), 3883–3887.
- Khan, Z. U., Aisikaer, G., Khan, R. U., Bu, J., Jiang, Z., Ni, Z., & Ying, T. (2014). Effects of composite chemical pretreatment on maintaining quality in button mushrooms (*Agaricus bisporus*) during postharvest storage. *Postharvest Biology and Technology*, 95, 36–41.
- Khanh, T. D., Hong, N. H., Nhan, D. Q., Kim, S. L., Chung, I. M., & Xuan, T. D. (2006). Assessment of weed-suppressing potential among rice (*Oryza sativa* L.) landraces against the growth of Barnyardgrass (*Echinochloa crus-galli* P. Beauv) in field condition. *Journal of Agronomy and Crop Science*, 192(6), 427–433.
- Kishor, P., Ghosh, A. K., Surendra, S., & Maurya, B. R. (2010). Potential use of *Parthenium* (*Parthenium hysterophorus* L.) in agriculture. *Asian Journal of Agricultural Research*, 4(4), 220–225.
- Kohli, R. K., Kumari, A., & Saxena, D. B. (1985). Auto and teleotoxicity of *Parthenium hysterophorus* L. *Acta Universitatis Agriculturae Brno*, 33, 253–263.
- Kohli, R. K., & Rani, D. (1994). Exhibition of allelopathy by *Parthenium hysterophorus* L. in agroecosystems. *Tropical Ecology*, 35(2), 295–307.
- Korav, S., Dhaka, A., Singh, R., & Chandramohan, R. (2018). A study on crop weed competition ... preview & related info. *Journal of Pharmacognosy and Phytochemistry*, 7(4), 3235–3240.
- Kordali, S., Cakir, A., Akcin, T. A., Mete, E., Akcin, A., Aydin, T., & Kilic, H. (2009). Antifungal and herbicidal properties of essential oils and n-hexane extracts of *Achillea gypsicola* Hub-Mor. and *Achillea biebersteinii* Afan. (Asteraceae). *Industrial Crops and Products*, 29(2–3), 562–570.
- Korres, N. E., Burgos, N. R., Travlos, I., Vurro, M., Gitsopoulos, T. K., Varanasi, V. K., Duke, S. O., Kudsk, P., Brabham, C., Rouse, C. E., & Salas-Perez, R. (2019). New directions for integrated weed management: Modern technologies, tools and knowledge discovery. In *Advances in Agronomy* (1st ed., Vol. 155). Elsevier Inc.
- Kua, Y. L., & Gan, S. (2019). Natural deep eutectic solvent (NADES) as a greener alternative for the extraction of hydrophilic (polar) and lipophilic (non-polar) phytonutrients. *Key Engineering Materials*, 797, 20–28.
- Kua, Y. L., Gan, S., Morris, A., & Ng, H. K. (2016). Ethyl lactate as a potential green solvent to extract hydrophilic (polar) and lipophilic (non-polar) phytonutrients simultaneously from fruit and vegetable by-products. *Sustainable Chemistry and Pharmacy*, 4, 21–31.

- Kulig, D., Matysiak, M., Baldovská, S., Štefániková, J., Maruniaková, N., Mňahončáková, E., Árvay, J., Galbavý, D., & Kolesárová, A. (2021). Screening of polyphenolic compounds from traditional medicinal herbs. *Journal of Microbiology, Biotechnology and Food Sciences*, 2021, 487–491.
- Kumar, M., & Kumar, S. (2010). Effect of *Parthenium hysterophorus* ash on growth and biomass of *Phaseolus mungo*. *Academia Arena*, 2(1), 98–102.
- Kumar, N. S. (2009). Effect of plant density and weed management practices on production potential of groundnut (*Arachis hypogaea* L.). *Indian Journal of Agricultural Research*, 43(1), 57–60.
- Kumar, S., Chashoo, G., Saxena, A. K., & Pandey, A. K. (2013). *Parthenium hysterophorus*: A probable source of anticancer, antioxidant and anti-HIV agents. *BioMed Research International, Hindawi*, 2013, 1–11.
- Kumar, S., Mishra, A., & Pandey, A. K. (2013). Antioxidant mediated protective effect of *Parthenium hysterophorus* against oxidative damage using in vitro models. *BMC Complementary and Alternative Medicine*, 13(1), 1–9.
- Kumar, S., Singh, A. P., Nair, G., Batra, S., Seth, A., Wahab, N., & Warikoo, R. (2011). Impact of *Parthenium hysterophorus* leaf extracts on the fecundity, fertility and behavioural response of *Aedes aegypti* L. *Parasitology Research*, 108(4), 853–859.
- Kushwaha, V. B., & Maurya, S. (2012). Biological utilities of *Parthenium hysterophorus*. *Journal of Applied and Natural Science*, 4(1), 137–143.
- Kwinda, M. (2021). Weed profiling fields of herbicide tolerant maize in the Mthatha region, Eastern Cape Province M Kwinda. *Master of Science in Botany at the North-West University, Phillipines*, July, 1–146.
- Labudda, M. (2013). *Lipid peroxidation as a biochemical marker for oxidative stress during drought. An effective tool for plant breeding.*
- Ladhari, A., Omezzine, F., Dellagreca, M., Zarrelli, A., & Haouala, R. (2013). Phytotoxic activity of *Capparis spinosa* L. and its discovered active compounds. *Allelopathy Journal*, 32(2), 175–190.
- Ladhari, A., Omezzine, F., Rinez, A., & Haouala, R. (2011). Phytotoxicity of *Daphne Gnidium* L. Occurring in Tunisia. *World Academy of Science, Engineering and Technology*, 59, 1534–1537.
- Laitonjam, W. S. (2012). Natural antioxidants (NAO) of plants acting as scavengers of free radicals. In *Studies in natural products chemistry* (Vol. 37, pp. 259–275). Elsevier.
- Lalita, K. A. (2018). Review on a weed *Parthenium hysterophorus* (L.). *International Journal of Current Research and Academic Review*, 10, 23.

- Latif, S., Chiapusio, G., & Weston, L. A. (2017). Allelopathy and the Role of Allelochemicals in Plant Defence. In G. B. T.-A. in B. R. Becard (Ed.), *How Plants Communicate with their Biotic Environment* (Vol. 82, pp. 19–54). Academic Press.
- Lee, L. J., & Ngim, J. (2000). A first report of glyphosate-resistant goosegrass (*Eleusine indica* (L) Gaertn) in Malaysia. *Pest Management Science*, 56(4), 336–339.
- Lestari, S. A. D., Melati, M., & Purnamawati, H. (2016). Penentuan Dosis Optimum Pemupukan N, P, dan K pada Tanaman Kacang Bogor [*Vigna subterranea* (L.) Verdcourt]. *Indonesian Journal of Agronomy*, 43(3), 193.
- Levizou, E. F. I., Karageorgou, P., Psaras, G. K., & Manetas, Y. (2002). Inhibitory effects of water soluble leaf leachates from *Dittrichia viscosa* on lettuce root growth, statocyte development and graviperception. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 197(2), 152–157.
- Li, B., Olson, E., Perugini, A., & Zhong, W. (2011). Simultaneous enhancements in damping and static dissipation capability of polyetherimide composites with organosilane surface modified graphene nanoplatelets. *Polymer*, 52(24), 5606–5614.
- Li, L., Wang, Q., Yang, Y., Wu, G., Xin, X., & Aisa, H. A. (2012). Chemical components and antidiabetic activity of essential oils obtained by hydrodistillation and three solvent extraction methods from *Carthamus tinctorius* L. *Acta Chromatographica*, 24(4), 653–665.
- Li, S., Peng, F., Xiao, Y., Gong, Q., Bao, Z., Li, Y., & Wu, X. (2020). Mechanisms of high concentration valine-mediated inhibition of peach tree shoot growth. *Frontiers in Plant Science*, 11(October), 1–13.
- Lichtenthaler, H. K., & Buschmann, C. (2001). Chlorophylls and carotenoids: Measurement and characterization by UV-VIS spectroscopy. *Current Protocols in Food Analytical Chemistry*, 1(1), F4-3.
- Lin, D., Xiao, M., Zhao, J., Li, Z., Xing, B., Li, X., Kong, M., Li, L., Zhang, Q., Liu, Y., Chen, H., Qin, W., Wu, H., & Chen, S. (2016). An overview of plant phenolic compounds and their importance in human nutrition and management. *Molecules*, 21(10), 1374.
- Lin, D., & Xing, B. (2007). Phytotoxicity of nanoparticles: inhibition of seed germination and root growth. *Environmental Pollution*, 150(2), 243–250.
- Lokajová, V., Bačkorová, M., & Bačkor, M. (2014). Allelopathic effects of lichen secondary metabolites and their naturally occurring mixtures on cultures of aposymbiotically grown lichen photobiont *Trebouxia erici* (Chlorophyta). *South African Journal of Botany*, 93, 86–91.
- López-Luna, J., Camacho-Martínez, M. M., Solís-Domínguez, F. A., González-Chávez, M. C., Carrillo-González, R., Martínez-Vargas, S., Mijangos-

- Ricardez, O. F., & Cuevas-Díaz, M. C. (2018). Toxicity assessment of cobalt ferrite nanoparticles on wheat plants. *Journal of Toxicology and Environmental Health, Part A*, 81(14), 604–619.
- Macías, F. A., Marín, D., Oliveros-Bastidas, A., Varela, R. M., Simonet, A. M., Carrera, C., & Molinillo, J. M. (2003). Allelopathy as a new strategy for sustainable ecosystems development. *Biological Sciences in Space*, 17(1), 18–23.
- Macías, F. A., Mejías, F. J. R., & Molinillo, J. M. G. (2019). Recent advances in allelopathy for weed control: from knowledge to applications. *Pest Management Science*, 75(9), 2413–2436.
- Mahala, A. G., & Mohammed, A. A. A. (2010). Nutritive evaluation of bambara groundnut (*Vigna subterranean*) pods, seeds and hull as animal feeds. *Journal of Applied Science Research*, 6(5), 383–386.
- Maharjan, S., Shrestha, B., & Jha, P. (2007). Allelopathic effects of aqueous extract of leaves of *Parthenium hysterophorus* L. on seed germination and seedling growth of some cultivated and wild herbaceous species. *Scientific World*, 5(5), 33–39.
- Mahdavia, F., Saharkhiz, M. J., & Karami, A. (2017). Defensive response of radish seedlings to the oxidative stress arising from phenolic compounds in the extract of peppermint (*Mentha × piperita* L.). *Scientia Horticulturae*, 214, 133–140.
- Majola, N. G., Gerrano, A. S., & Shimelis, H. (2021). Bambara groundnut (*Vigna subterranea* [L.] Verdc.) production, utilisation and genetic improvement in Sub-Saharan Africa. *Agronomy*, 11(7), 1345.
- Makhaye, G., Mofokeng, M. M., Tesfay, S., Aremu, A. O., Van Staden, J., & Amoo, S. O. (2021). Influence of plant biostimulant application on seed germination. *Biostimulants for Crops from Seed Germination to Plant Development*, 109–135.
- Mao, R., Shabbir, A., & Adkins, S. (2021). *Parthenium hysterophorus*: A tale of global invasion over two centuries, spread and prevention measures. *Journal of Environmental Management*, 279, 111751.
- Marchiosi, R., dos Santos, W. D., Constantin, R. P., de Lima, R. B., Soares, A. R., Finger-Teixeira, A., Mota, T. R., de Oliveira, D. M., Foletto-Felipe, M. de P., & Abrahao, J. (2020). Biosynthesis and metabolic actions of simple phenolic acids in plants. *Phytochemistry Reviews*, 19, 865–906.
- Marwat, S. K., Fazal-ur-Rehman, & Khan, I. U. (2015). Ethnobotanical importance and phytochemical constituents of *Parthenium* weed (*Parthenium hysterophorus* L.) – A Review. *Plant Science Today*, 2(2), 77–81.
- Mazahib, A. M., Nuha, M. O., Salawa, I. S., & Babiker, E. E. (2013). Some nutritional attributes of bambara groundnut as influenced by domestic processing. *International Food Research Journal*, 20(3), 1165.

- Meena, R. K., Dutt, B., Kumar, R., & Sharma, K. R. (2017). Phyto-chemistry of congress grass (*Parthenium hysterophorus* L.) and harmful and beneficial effect on human and animals: A review. *International Journal of Chemical Studies*, 5(4), 643–647.
- Mehla, N., Sindhi, V., Josula, D., Bisht, P., & Wani, S. H. (2017). An introduction to antioxidants and their roles in plant stress tolerance. In *Reactive oxygen species and antioxidant Systems in Plants: role and regulation under abiotic stress* (pp. 1–23). Springer.
- Mehmood, K., Asif, H. M., Bajwa, R., Shafique, S., & Shafique, S. (2011). Phytotoxic potential of bark extracts of *Acacia Nilotica* and *Syzygium cumini* against *Parthenium hysterophorus*. *Pakistan Journal of Botany*, 43(6), 3007–3012.
- Meksawat, S., & Pornprom, T. (2010). Allelopathic effect of itchgrass (*Rottboellia cochinchinensis*) on seed germination and plant growth. *Weed Biology and Management*, 10(1), 16–24.
- Menon, R. G., Chien, S. H., & Hammond, L. L. (1989). Comparison of Bray I and Pi tests for evaluating plant-available phosphorus from soils treated with different partially acidulated phosphate rocks. *Plant and Soil*, 114(2), 211–216.
- Merga, B., & Ahmed, A. (2019). A review on agricultural problems and their management in Ethiopia. *Turkish Journal of Agriculture-Food Science and Technology*, 7(8), 1189–1202.
- Mersie, W., & Singh, M. (1987). Allelopathic effect of parthenium (*Parthenium hysterophorus* L.) extract and residue on some agronomic crops and weeds. *Journal of Chemical Ecology*, 13(7), 1739–1747.
- Mhlanga, B., Chauhan, B. S., & Thierfelder, C. (2016). Weed management in maize using crop competition: A review. *Crop Protection*, 88, 28–36.
- Mijangos Ricárdez, O. F., Ruiz-Jiménez, J., Lagunez-Rivera, L., & Luque De Castro, M. D. (2011). Fast ultrasound-assisted extraction of polar (phenols) and nonpolar (lipids) fractions in *Heterotheca inuloides* Cass. *Phytochemical Analysis*, 22(6), 484–491.
- Mirmostafae, S., Azizi, M., & Fujii, Y. (2020). Study of allelopathic interaction of essential oils from medicinal and aromatic plants on seed germination and seedling growth of lettuce. *Agronomy*, 10(2).
- Mishra, A. (2015). Allelopathic properties of *Lantana camara*. *International Research Journal of Basic and Clinical Studies*, 3(1), 13–28.
- Mogote, G. J. (2011). *The herbicidal potential of Tagetes minuta and Tagetes patula extracts and their residues on the germination and growth of some crop plants*. Master of Science (Plant Physiology and Biochemistry). University of NAIROBI.

- Mohamadi, N., & Rajaie, P. (2009). Effects of aqueous eucalyptus (*E. camadulensis* Labill) extracts on seed germination, seedling growth and physiological responses of *Phaseolus vulgaris* and *Sorghum bicolor*. *Research Journal of Biological Sciences*, 4(12), 1292–1296.
- Mominul Islam, A. K. M., Hasan, M., Musha, M. M. H., Uddin, M. K., Juraimi, A. S., & Anwar, M. P. (2018). Exploring 55 tropical medicinal plant species available in Bangladesh for their possible allelopathic potentiality. *Annals of Agricultural Sciences*, 63(1), 99–107.
- Motmainna et al. (2021a). Phytochemical constituents and allelopathic potential of *Parthenium hysterophorus* L. In comparison to commercial herbicides to control weeds. *Plants*, 10(7), 1–16.
- Motmainna, M., Juraimi, A. S., Uddin, M. K., Asib, N. B., Islam, A. K. M. M., & Hasan, M. (2021b). Bioherbicidal properties of *Parthenium hysterophorus*, *Cleome rutidosperma* and *Borreria alata* extracts on selected crop and weed species. *Agronomy*, 11(4), 53–68.
- Mousavi, S. S., Karami, A., Haghighi, T. M., Alizadeh, S., & Maggi, F. (2021). Phytotoxic potential and phenolic profile of extracts from *scrophularia striata*. *Plants*, 10(1), 1–18.
- Msafiri, C., Tarimo, M., & Ndakidemi, P. (2013). Allelopathic effects of *Parthenium hysterophorus* on seed germination, seedling growth, fresh and dry mass production of *Alysicarpus glumaceae* and *Chloris gayana*. *American Journal of Research Communication*, 1(11), 190–205.
- Musco, A., Panuccio, M. R., & Sidari, M. (2001). The effect of phenols on respiratory enzymes in seed germination. *Plant Growth Regulation*, 35(1), 31–35.
- Mushtaq, W., Ain, Q., & Siddiqui, M. B. (2018). Screening of allelopathic activity of the leaves of *Nicotiana plumbaginifolia* Viv. on some selected crops in Aligarh, Uttar Pradesh, India. *International Journal of Photochemistry and Photobiology*, 2(1), 1–4.
- Mushtaq, W., Ain, Q., Siddiqui, M. B., & Hakeem, K. R. (2019). Cytotoxic allelochemicals induce ultrastructural modifications in *Cassia tora* L. and mitotic changes in *Allium cepa* L.: a weed versus weed allelopathy approach. *Protoplasma*, 256(3), 857–871.
- Mushtaq, W., & Siddiqui, M. B. (2018). Allelopathy in Solanaceae plants. *Journal of Plant Protection Research*, 58(1).
- Mushtaq, W., Siddiqui, M. B., & Hakeem, K. R. (2020). Mechanism of action of allelochemicals. In *Allelopathy* (pp. 61–66). Springer.
- Muzell Trezzi, M., Vidal, R. A., Balbinot Junior, A. A., von Hertwig Bittencourt, H., & da Silva Souza Filho, A. P. (2016). Allelopathy: driving mechanisms governing its activity in agriculture. *Journal of Plant Interactions*, 11(1), 53–60.

- Mwangi, H. W. (2017). Integration of cover crop technology and imidazolinone resistant maize on weeds and maize yield in two ASAL areas of Kenya. In *Doctoral dissertation* (pp. 1–182). Faculty of Agriculture, JKUAT.
- Nadeem, M. A., Aziz, A., & Aziz, A. (2020). Allelopathic effects of aqueous extracts of *Carthamus tinctorius* L. on emergence and seedling growth of *Echinochloa crusgalli* L. *Pakistan Journal of Weed Science Research*, 26(3), 367–379.
- Nadir, S., Xiong, H.-B., Zhu, Q., Zhang, X.-L., Xu, H.-Y., Li, J., Dongchen, W., Henry, D., Guo, X.-Q., & Khan, S. (2017). Weedy rice in sustainable rice production. A review. *Agronomy for Sustainable Development*, 37(5), 1–14.
- Nandini, S. V. (2016). Isolation and screening of native root nodulating bacteria (*Vigna subterranea* (L.) Verdc). In *PhD dissertation, University of Agricultural Sciences, GKVK, Bengaluru, India* (pp. 1–80).
- Nargotra, P., Sharma, V., & Bajaj, B. K. (2019). Consolidated bioprocessing of surfactant-assisted ionic liquid-pretreated *Parthenium hysterophorus* L. biomass for bioethanol production. *Bioresource Technology*, 289(June), 121611.
- Narwal, S. S. (2018). Allelopathic plants: *Parthenium hysterophorus* L. *Allelopathy Journal*, 11(2), 151–170.
- Narwal, S. S., Palaniraj, R., Sati, S. C., Kadian, H. S., & Dahiya, D. S. (2003). Allelopathic plants: *Parthenium hysterophorus* L. *Allelopathy Journal*, 11(2), 151–170.
- Navie, S. C., McFadyen, R. E., Panetta, F. D., & Adkins, S. W. (1996). The biology of Australian weeds: *Parthenium hysterophorus* L. *Plant Protection Quarterly*, 11(2), 76–88.
- Negi, B., Bargali, S. ., Bargali, K., & Khatri, K. (2020). Allelopathic Interference of *Ageratum conyzoides* L. against Rice Varieties. *Current Agriculture Research Journal*, July, 69–76.
- Netsere, A. (2015). Allelopathic effects of aqueous extracts of an invasive alien weed *Parthenium hysterophorus* L. on maize and sorghum seed germination and seedling growth. *Journal of Biology, Agriculture and Healthcare*, 5(1), 120–124.
- Nguyen, Bajwa, A. A., Navie, S., O'donnell, C., & Adkins, S. (2017). *Parthenium* weed (*Parthenium hysterophorus* L.) and climate change: the effect of CO₂ concentration, temperature, and water deficit on growth and reproduction of two biotypes. *Environmental Science and Pollution Research*, 24(11), 10727–10739.
- Nguyen, D. T. C., Tran, T. Van, Kumar, P. S., Din, A. T. M., Jalil, A. A., & Vo, D.-V. N. (2022). Invasive plants as biosorbents for environmental remediation: a review. *Environmental Chemistry Letters*, 1–31.

- Nie, X.-P., Liu, B.-Y., Yu, H.-J., Liu, W.-Q., & Yang, Y.-F. (2013). Toxic effects of erythromycin, ciprofloxacin and sulfamethoxazole exposure to the antioxidant system in *Pseudokirchneriella subcapitata*. *Environmental Pollution*, *172*, 23–32.
- Nigatu, L., Hassen, A., Sharma, J., & Adkins, S. W. (2010). Impact of *Parthenium hysterophorus* on grazing land communities in north-eastern Ethiopia. *Weed Biology and Management*, *10*(3), 143–152.
- Niranjan, A., Mishra, S., Lehri, A., Amla, D. V, Upadhyay, R. S., & Nautiyal, C. S. (2013). Identification and quantification of heterologous compounds parthenin and organic acids in *Parthenium hysterophorus* L. using HPLC-PDA-MS-MS. *Analytical Letters*, *46*(1), 48–59.
- Noga, M., Sucharski, F., Suder, P., & Silberring, J. (2007). A practical guide to nano-LC troubleshooting. *Journal of Separation Science*, *30*(14), 2179–2189.
- NTSYS-pc, N. T., & Taxonomy, N. (2005). Multivariate Analysis System, version 2.2. *Exeter Software: Setauket, NY, USA*.
- Ogwu, M. C., Ahana, C. M., & Osawaru, M. E. (2018). Sustainable food production in Nigeria: a case study for Bambara Groundnut (*Vigna subterranea* (L.) Verdc. Fabaceae). *Journal of Energy and Natural Resource Management (JENRM)*, *1*(1), 125.
- Oli, S., Chopra, N., Tewari, L. M., Mohan, B., Pandey, N., Bharti, M., Bohra, N., & Tewari, G. (2018). Phytotoxic effect of the extracts of *Parthenium hysterophorus* L. on the germination, seedling growth and biomass of some agricultural crops. *G-Journal of Environmental Science and Technology*, *5*(4), 40–45.
- Olofsdotter, M., Jensen, L. B., & Courtois, B. (2002). Improving crop competitive ability using allelopathy—an example from rice. *Plant Breeding*, *121*(1), 1–9.
- Omar, S. A., Elsheery, N. I., Kalaji, H. M., Xu, Z.-F., Song-Quan, S., Carpentier, R., Lee, C.-H., & Allakhverdiev, S. I. (2012). Dehydroascorbate reductase and glutathione reductase play an important role in scavenging hydrogen peroxide during natural and artificial dehydration of *Jatropha curcas* seeds. *Journal of Plant Biology*, *55*(6), 469–480.
- Onyilagha, J. C., & Islam, S. (2009). Flavonoids and other polyphenols of the cultivated species of the genus *Phaseolus*. *International Journal of Agricultural Biology*, *11*, 231–234.
- Ouedraogo, M., Ouedraogo, J. T., Tignere, J. B., Bilma, D., Dabire, C. B., & Konate, G. (2008). Characterization and evaluation of accessions of Bambara groundnut (*Vigna subterranea* (L.) Verdcourt) from Burkina Faso. *Sciences & Nature*, *5*(2), 191–197.

- Ouedraogo, M., Zagre M'bi, B., Liu, F., Ortiz, R., & Jørgensen, S. T. (2013). Timing of mounding for bambara groundnut affects crop development and yield in a rainfed tropical environment. *Acta Agriculturae Scandinavica, Section B–Soil & Plant Science*, 63(4), 370–375.
- Oviedo, P., Herrera, O., & Caluff, M. G. (2012). National list of invasive and potentially invasive plants in the Republic of Cuba-2011. *Bissea: Boletín Sobre Conservación de Plantas Del Jardín Botánico Nacional de Cuba.*, 6(Special Issue No. 1), 22–96.
- Owen, M. D. K. (2016). Diverse approaches to herbicide-resistant weed management. *Weed Science*, 64(S1), 570–584.
- Oyeleke, G. O., Afolabi, O., & Isola, A. D. (2012). Some quality characteristics and carbohydrate fractions of bambara groundnut (*Vigna subterranea* L.) seed flour. *IOSR Journal of Applied Chemistry*, 2(4), 16–19.
- Padma, S., & Deepika, S. (2013). Phytochemical screening and in vitro antifungal investigation of *Parthenium hysterophorus* extracts against *Alternaria alternate*. *International Research Journal of Pharmacy*, 4(7), 190–193.
- Pan, L., Li, X., Yan, Z., Guo, H., & Qin, B. (2015). Phytotoxicity of umbelliferone and its analogs: Structure–activity relationships and action mechanisms. *Plant Physiology and Biochemistry*, 97, 272–277.
- Pandey, A. K. (2007). Anti-staphylococcal activity of a pan-tropical aggressive and obnoxious weed *Parthenium hysterophorus*: an in vitro study. *National Academy Science Letters*, 30(11/12), 383–386.
- Pandey, R. A., Gole, A. R., Sankpal, R. V., Jadav, P. V., Waghmode, M. S., & Patil, N. N. (2019). Bioactive potential of *Parthenium hysterophorus* and cytotoxicity assay of parthenin. *International Journal of Pharmacy and Biological Sciences*, 9(3), 296–313.
- Pandiyan, K., Tiwari, R., Rana, S., Arora, A., Singh, S., Saxena, A. K., & Nain, L. (2014). Comparative efficiency of different pretreatment methods on enzymatic digestibility of *Parthenium* sp. *World Journal of Microbiology and Biotechnology*, 30(1), 55–64.
- Pareek, A., Suthar, M., Rathore, G. S., & Bansal, V. (2011). Feverfew (*Tanacetum parthenium* L.): A systematic review. *Pharmacognosy Reviews*, 5(9), 103.
- Paris-Palacios, S., Mosleh, Y. Y., Almohamad, M., Delahaut, L., Conrad, A., Arnoult, F., & Biagianti-Risbourg, S. (2010). Toxic effects and bioaccumulation of the herbicide isoproturon in *Tubifex tubifex* (Oligocheate, Tubificidae): a study of significance of autotomy and its utility as a biomarker. *Aquatic Toxicology*, 98(1), 8–14.
- Parsons, W. T., & Cuthbertson, E. G. (1992). Weeds of Natural Ecosystems. *Noxious Weeds of Australia*. Inkata Press, Melbourne, Victoria, Australia, 173–175.

- Parsons, W. T., Parsons, W. T., & Cuthbertson, E. G. (2001). *Noxious weeds of Australia*. CSIRO publishing. 2nd edition. Inkata Press.
- Patel, S. (2011). Harmful and beneficial aspects of *Parthenium hysterophorus*: an update. *Biotech*, 1(1), 1–9.
- Patel, V. S., Chitra, V., Prasanna, P. L., & Krishnaraju, V. (2008). Hypoglycemic effect of aqueous extract of *Parthenium hysterophorus* L. in normal and alloxan induced diabetic rats. *Indian Journal of Pharmacology*, 40(4), 183.
- Pati, U. K., & Chowdhury, A. (2015). Study of *Parthenium hysterophorus* L. extracts (First clean-up fractions) on seed germination behaviour in search of bioactive fractions for preparation of bioherbicide formulations. *International Letters of Natural Sciences*, 49(March), 69–80.
- Pati, U. K., & Chowdhury, A. (2016). Study of *Parthenium hysterophorus* L. Extracts (First Clean-Up Fractions) on Seed Germination Behaviour in Search of Bioactive Fractions for Preparation of Bioherbicide Formulations . *Published by SciPress Ltd, Switzerland*, 49(March), 69–80.
- Patil, P. K., Sharma, A., Kaur, I., & Cannoo, D. S. (2019). Correlation study among the extraction techniques, phytochemicals, and antioxidant activity of *Nepeta spicata* aerial part. *Biocatalysis and Agricultural Biotechnology*, 20, 101275.
- Patil Bhimarao, J., & Khade Hemlata, N. (2017). Allelopathic effect of *Parthenium hysterophorus* L. on photosynthetic pigments and biochemical constituents of *Vigna aconitifolia* L. *International Journal of Life Sciences*, 5(4), 677–682.
- Paudel, V. R. (2010). *Parthenium hysterophorus* L., a noxious invasive weed. *Botanica Orientalis: Journal of Plant Science*, 6, 85–92.
- Paul, V., Sharma, L., Kumar, R., Pandey, R., & Meena, R. C. (2017). Estimation of chlorophylls/photosynthetic pigments—their stability is an indicator of crop plant tolerance to abiotic stresses. *Manual of ICAR Sponsored Training Programme for Technical Staff of ICAR Institutes on “physiological Techniques to Analyze the Impact of Climate Change on Crop Plants*, 8, 14–129.
- Pavlovic, D., Nikolic, B., Djurovic, S., Waisi, H., Andjelkovic, A., & Marisavljevic, D. (2014). Chlorophyll as a measure of plant health: Agroecological aspects. *Pesticidi i Fitomedicina*, 29(1), 21–34.
- Pérez, F. J. (1990). Allelopathic effect of hydroxamic acids from cereals on *Avena sativa* and *A. fatua*. *Phytochemistry*, 29(3), 773–776.
- Pérez, M. B., Calderon, N. L., & Croci, C. A. (2007). Radiation-induced enhancement of antioxidant activity in extracts of rosemary (*Rosmarinus officinalis* L.). *Food Chemistry*, 104(2), 585–592.

- Petersen, J., Belz, R., Walker, F., & Hurle, K. (2001). Weed suppression by release of isothiocyanates from turnip-rape mulch. *Agronomy Journal*, 93(1), 37–43.
- Petit, S., Cordeau, S., Chauvel, B., Bohan, D., Guillemin, J.-P., & Steinberg, C. (2018). Biodiversity-based options for arable weed management. A review. *Agronomy for Sustainable Development*, 38(5), 1–21.
- Pickett, J. A., Aradottir, G. I., Birkett, M. A., Bruce, T. J. A., Hooper, A. M., Midega, C. A. O., Jones, H. D., Matthes, M. C., Napier, J. A., Pittchar, J. O., Smart, L. E., Woodcock, C. M., Khan, Z. R., B, P. T. R. S., Birkett, M. A., Bruce, T. J. A., Pickett, J. A., Aradotti, G. I., Hooper, A. M., ... Khan, Z. R. (2014). Delivering sustainable crop protection systems via the seed : exploiting natural constitutive and inducible defence pathways. *Philosophical Transactions of the Royal Society*, B 369:20120281.
- Priya, R., Chinnusamy, C., Manickasundaram, P., & Babu, C. (2013). A review on weed management in groundnut (*Arachis hypogea* L.). *International Journal of Agricultural Science and Research (IJASR)*, 3(1), 163–171.
- Qasem, J. R., & Foy, C. L. (2001). Weed allelopathy, its ecological impacts and future prospects: a review. *Journal of Crop Production*, 4(2), 43–119.
- Qureshi, M. I., Abdin, M. Z., Ahmad, J., & Iqbal, M. (2013). Effect of long-term salinity on cellular antioxidants, compatible solute and fatty acid profile of Sweet Annie (*Artemisia annua* L.). *Phytochemistry*, 95, 215–223.
- Radosevich, S. R., Holt, J. S., & Ghersa, C. M. (2007). *Ecology of weeds and invasive plants: relationship to agriculture and natural resource management*. John Wiley & Sons. (pp. 1-454).
- Raguindin, P. F., Itodo, O. A., Stoyanov, J., Dejanovic, G. M., Gamba, M., Asllanaj, E., Minder, B., Bussler, W., Metzger, B., & Muka, T. (2021). A systematic review of phytochemicals in oat and buckwheat. *Food Chemistry*, 338, 127982.
- Rahman, M. A. (2021). Cover crops' effect on soil quality and soil health. In *Cover Crops and Sustainable Agriculture* (pp. 124–146). CRC Press.
- Rahman, M., Pudasainee, D., & Gupta, R. (2017). Review on chemical upgrading of coal: Production processes, potential applications and recent developments. *Fuel Processing Technology*, 158, 35–56.
- Rajendiran, K. (2005). Mitodepressive effects of aqueous extracts of *Parthenium hysterophorus* L. on *Vigna radiata* (L.) Wilczek. *GEOSCIENCE JOURNAL*, 32(4), 237.
- Rajput, R. L., & Kasana, B. S. (2020). Integration of chemical and cultural methods for weed control management in soybean (*Glycin max.* l). *Legume Research*, 43(1), 122–125.

- Ramos, A., Rivero, R., Victoria, M. C., Visozo, A., Piloto, J., & Garcia, A. (2001). Assessment of mutagenicity in *Parthenium hysterophorus* L. *Journal of Ethnopharmacology*, 77(1), 25–30.
- Ramos, A., Rivero, R., Visozo, A., Piloto, J., & García, A. (2002). Parthenin, a sesquiterpene lactone of *Parthenium hysterophorus* L. is a high toxicity clastogen. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 514(1–2), 19–27.
- Rana, S. S. (2016). Cropping System. In *Department of Agronomy, College of Agriculture, CSK Himachal Pradesh Krishi Vishvavidyalaya, Palampur, 80* (Issue October, pp. 1–92).
- Rao, M. V., Paliyath, G., & Ormrod, D. P. (1996). Ultraviolet-B-and ozone-induced biochemical changes in antioxidant enzymes of *Arabidopsis thaliana*. *Plant Physiology*, 110(1), 125–136.
- Rashid, M. D. (2014). Performance of groundnut under different plant population and weed management. In *Doctoral dissertation*, Department of Agronomy.
- Rasouli, H., Farzaei, M. H., Mansouri, K., Mohammadzadeh, S., & Khodarahmi, R. (2016). Plant cell cancer: may natural phenolic compounds prevent onset and development of plant cell malignancy? A literature review. *Molecules*, 21(9), 1104.
- Razzaq, A., Cheema, Z. A., Jabran, K., Hussain, M., Farooq, M., & Zafar, M. (2012). Reduced herbicide doses used together with allelopathic sorghum and sunflower water extracts for weed control in wheat. *Journal of Plant Protection Research*, 52(2).
- Reddy, C. (2018). A study on crop weed competition in field crops. *Journal of Pharmacognosy and Phytochemistry*, 7(4), 3235–3240.
- Reddy, P. S., Jogeswar, G., Rasineni, G. K., Maheswari, M., Reddy, A. R., Varshney, R. K., & Kishor, P. B. K. (2015). Proline over-accumulation alleviates salt stress and protects photosynthetic and antioxidant enzyme activities in transgenic sorghum [*Sorghum bicolor* (L.) Moench]. *Plant Physiology and Biochemistry*, 94, 104–113.
- Reeves, D. W., Price, A. J., & Patterson, M. G. (2005). Evaluation of three winter cereals for weed control in conservation-tillage nontransgenic cotton. *Weed Technology*, 19(3), 731–736.
- Rehman, S., Shahzad, B., Bajwa, A. A., Hussain, S., Rehman, A., Cheema, S. A., Abbas, T., Ali, A., Shah, L., Adkins, S., & Li, P. (2019). Utilizing the allelopathic potential of *Brassica species* for sustainable crop production: A review. *Journal of Plant Growth Regulation*, 38(1), 343–356.

- Reigosa, M. J., & Pazos-Malvido, E. (2007). Phytotoxic effects of 21 plant secondary metabolites on *Arabidopsis thaliana* germination and root growth. *Journal of Chemical Ecology*, 33(7), 1456–1466.
- Rezaei, F., Jamei, R., & Heidari, R. (2017). Evaluation of the phytochemical and antioxidant potential of aerial parts of Iranian *Tanacetum parthenium*. *Pharmaceutical Sciences*, 23(2), 136–142.
- Rice, E. L. (2012). *Allelopathy*. Department of Botany and Microbiology. The university of Oklahoma. Norman, Oklahoma, UK. Academic press.
- Roy, D. C., & Shaik, M. M. (2013). Toxicology, phytochemistry, bioactive compounds and pharmacology of *Parthenium hysterophorus*. *Journal of Medicinal Plants Studies*, 1(3), 126–141.
- Safdar, M. E., Aslam, A., Qamar, R., Ali, A., Javaid, M. M., Hayyat, M. S., & Raza, A. (2021). Allelopathic effect of prickly chaff flower (*Achyranthes aspera* L.) used as a tool for managing noxious weeds. *Asian Journal of Agriculture and Biology*, 2021(3), 1–10.
- Sahrawat, A., Sharma, J., Rahul, S. N., Tiwari, S., & Rai, D. V. (2018). *Parthenium hysterophorus*: Current status and its possible effects on mammals-A review. *International Journal of Current Microbiology and Applied Sciences*, 7(11), 3548–3557.
- Saidi, I., Ayouni, M., Dhieb, A., Chtourou, Y., Chaïbi, W., & Djebali, W. (2013). Oxidative damages induced by short-term exposure to cadmium in bean plants: protective role of salicylic acid. *South African Journal of Botany*, 85, 32–38.
- Saini, A., Aggarwal, N. K., Sharma, A., Kaur, M., & Yadav, A. (2014). Utility potential of *Parthenium hysterophorus* for its strategic management. *Advances in Agriculture, Hindawi*, 2014, 1–16.
- Samanta, A., Das, G., & Das, S. K. (2011). Roles of flavonoids in plants. *Carbon*, 100(6), 12–35.
- Samuels, L., Kunst, L., & Jetter, R. (2008). Sealing plant surfaces: cuticular wax formation by epidermal cells. *Annual Review of Plant Biology*, 59, 683–707.
- Sánchez-Moreiras, A. M., de la Peña, T. C., & Reigosa, M. J. (2008). The natural compound benzoxazolin-2 (3H)-one selectively retards cell cycle in lettuce root meristems. *Phytochemistry*, 69(11), 2172–2179.
- Sanchez-Moreiras, A. M., Martinez, A., Gonzalez, L., Pellisier, F., & Regiosa, M. J. (2004). Mode of action of the hydroxamic acid BOA and other related compounds. In *In Allelopathy*. (pp. 239–252). CRC Press, Boca Raton, FL.
- Sathishkumar, A., Srinivasan, G., Subramanian, E., & Rajesh, P. (2020). Role of allelopathy in weed management: A review. *Agricultural Reviews*, 41(4), 380–386.

- Scavo, A., Abbate, C., & Mauromicale, G. (2019). Plant allelochemicals: agronomic, nutritional and ecological relevance in the soil system. *Plant and Soil*, 442(1–2), 23–48.
- Scavo, A., & Mauromicale, G. (2020). Integrated weed management in herbaceous field crops. *Agronomy*, 10(4), 466.
- Schandry, N., & Becker, C. (2020). Allelopathic plants: models for studying plant–interkingdom interactions. *Trends in Plant Science*, 25(2), 176–185.
- Schymanski, E. L., Jeon, J., Gulde, R., Fenner, K., Ruff, M., Singer, H. P., & Hollender, J. (2014). Identifying small molecules via high resolution mass spectrometry: communicating confidence. *Environmental Science and Technology*, 48(4), 2097–2098.
- Scognamiglio, M., Abrosca, B. D., Esposito, A., & Fiorentino, A. (2015). Metabolomics: an unexplored tool for allelopathy studies. *Journal of Allelochemical Interactions*, 1(January), 9–23.
- Scognamiglio, M., D'Abrosca, B., Esposito, A., Pacifico, S., Monaco, P., & Fiorentino, A. (2013). Plant growth inhibitors: allelopathic role or phytotoxic effects? Focus on Mediterranean biomes. *Phytochemistry Reviews*, 12(4), 803–830.
- Scognamiglio, M., Esposito, A., D'Abrosca, B., Pacifico, S., Fiumano, V., Tsafantakis, N., Monaco, P., & Fiorentino, A. (2012). Isolation, distribution and allelopathic effect of caffeic acid derivatives from *Bellis perennis* L. *Biochemical Systematics and Ecology*, 43, 108–113.
- Shabbir, A. (2014). Chemical control of *Parthenium hysterophorus* L. *Pakistan Journal of Weed Science Research*, 20(1), 1–10.
- Shabbir, A., Dhileepan, K., & Adkins, S. W. (2012). Spread of parthenium weed and its biological control agent in the Punjab, Pakistan. *Pakistan Journal of Weed Science Research*, 18(Special Issue), 581–588.
- Shafiq, F., Irfan, S., & Khan Shakir, S. (2020). Comparative Allelopathic Effects of Different Parts of *Parthenium hysterophorus* L. on Seed Germination and Biomasses of *Cicer arietinum* L. *Original Text Journal of Stress Physiology & Biochemistry*, 16(1), 64–75.
- Shahena, S., Rajan, M., Chandran, V., & Mathew, L. (2020). Controlled release herbicides and allelopathy as sustainable alternatives in crop production. In *Controlled Release of Pesticides for Sustainable Agriculture* (pp. 237–252). Springer.
- Sharma, A., Kapoor, D., Wang, J., Shahzad, B., Kumar, V., Bali, A. S., Jasrotia, S., Zheng, B., Yuan, H., & Yan, D. (2020). Chromium bioaccumulation and its impacts on plants: an overview. *Plants*, 9(1), 100.

- Sharma, A., Khare, S. K., & Gupta, M. N. (2002). Enzyme-assisted aqueous extraction of peanut oil. *Journal of the American Oil Chemists' Society*, 79(3), 215–218.
- Sharma, A., Kumar, V., Shahzad, B., Tanveer, M., Sidhu, G. P. S., Handa, N., Kohli, S. K., Yadav, P., Bali, A. S., & Parihar, R. D. (2019). Worldwide pesticide usage and its impacts on ecosystem. *SN Applied Sciences*, 1(11), 1–16.
- Sharma, A., Shahzad, B., Kumar, V., Kohli, S. K., Sidhu, G. P. S., Bali, A. S., Handa, N., Kapoor, D., Bhardwaj, R., & Zheng, B. (2019). Phytohormones regulate accumulation of osmolytes under abiotic stress. *Biomolecules*, 9(7), 285.
- Sharma, A., Singh, H. P., Batish, D. R., & Kohli, R. K. (2019). Chemical profiling, cytotoxicity and phytotoxicity of foliar volatiles of *Hyptis suaveolens*. *Ecotoxicology and Environmental Safety*, 171, 863–870.
- Sharma, M., & Devkota, A. (2018). Allelopathic influences of *Artemisia Dubia* Wall. ex. Besser on seed germination and seedling vigor of *Parthenium hysterophorus* L. *Journal of Institute of Science and Technology*, 22(2), 117–128.
- Sharma, S., Jat, R. A., & Sagarka, B. K. (2015). Effect of weed-management practices on weed dynamics, yield, and economics of groundnut (*Arachis hypogaea*) in black calcareous soil. *Indian Journal of Agronomy*, 60(2), 312–317.
- Shekhawat, K., Rathore, S. S., Dass, A., Das, T. K., Mahajan, G., & Chauhan, B. S. (2017). Weed menace and management strategies for enhancing oilseed brassicas production in the Indian sub-continent: A review. *Crop Protection*, 96, 245–257.
- Sheteawi, S. A., & Tawfik, K. M. (2007). Interaction effect of some biofertilizers and irrigation water regime on mung bean (*Vigna radiata*) growth and yield. *Citeseer*, 3(3), 251–262.
- Shikha, R., & Jha, A. K. (2016). Allelopathic effect of leaf extract of *Parthenium hysterophorus* L. on Seed Germination and Growth of *Cicer aeritinum* L. *International Journal of Science and Research (IJSR)*, 5(3), 652–655.
- Shin, H.-Y., Shim, S.-H., Ryu, Y.-J., Yang, J.-H., Lim, S.-M., & Lee, C.-G. (2018). Lipid extraction from *Tetraselmis sp. microalgae* for biodiesel production using hexane-based solvent mixtures. *Biotechnology and Bioprocess Engineering*, 23(1), 16–22.
- Singh. (2005). Phytotoxic effects of *Parthenium hysterophorus* residues on three Brassica species. *Weed Biology and Management*, 5(3), 105–109.
- Singh, A., Singh, D., & Singh, N. B. (2015). Allelopathic activity of *Nicotiana plumbaginifolia* at various phenological stages on sunflower. *Allelopathy Journal*, 36(2), 315–325.

- Singh, & Batish. (2005). Phytotoxic effects of *Parthenium hysterophorus* residues on three Brassica species. *Weed Biology and Management*, 5(3), 105–109.
- Singh, I. (2020). *Parthenium hysterophorus* a threat or beneficial weed and management : A review. *Indian Journal of Natural Sciences*, 11(63), 28719–28729.
- Singh, P., Batish, D., Kaur, S., Setia, N., & Kohli, R. (2005). Effects of 2-benzoxazolinone on the germination, early growth and morphogenetic response of mung bean (*Phaseolus aureus*). *Annals of Applied Biology*, 147(3), 267–274.
- Singh, P. K. (2021). An analysis regarding several aspects of weeds. *ACADEMICIA: An International Multidisciplinary Research Journal*, 11(11), 610–616.
- Singh, R., Chaurasia, S., Gupta, A. D., & Soni, P. (2014). Studies on effect of *Parthenium hysterophorus* plants extract on germination and growth in certain pulses. *Scholarly Journal of Agriculture Science*, 4(4), 235–239.
- Singh, R. K., Kumar, S., Kumar, S., & Kumar, A. (2008). Development of parthenium based activated carbon and its utilization for adsorptive removal of p-cresol from aqueous solution. *Journal of Hazardous Materials*, 155(3), 523–535.
- Singh, V., & Agrawal, K. K. (2020). Agriculture & Food: September 2020 monthly online magazine. In *e-NEWSLETTER*. (Vol. 2, Issue 9, p. 682).
- Siyar, S., Majeed, A., Muhammad, Z., Ali, H., & Inayat, N. (2019). Allelopathic effect of aqueous extracts of three weed species on the growth and leaf chlorophyll content of bread wheat. *Acta Ecologica Sinica*, 39(1), 63–68.
- Smith, R. G., Ryan, M. R., & Menalled, F. D. (2011). Direct and indirect impacts of weed management practices on soil quality. *Wiley*, 16802, 275–286.
- Smykal, P., Coyne, C. J., Ambrose, M. J., Maxted, N., Schaefer, H., Blair, M. W., Berger, J., Greene, S. L., Nelson, M. N., & Besharat, N. (2015). Legume crops phylogeny and genetic diversity for science and breeding. *Critical Reviews in Plant Sciences*, 34(1–3), 43–104.
- Sodaeizadeh, H., Rafieiolhossaini, M., Havlík, J., & van Damme, P. (2009). Allelopathic activity of different plant parts of *Peganum harmala* L. and identification of their growth inhibitors substances. *Plant Growth Regulation*, 59(3), 227–236.
- Sohal, S. K., Rup, P. J., Kaur, H., Kumari, N., & Kaur, J. (2002). Evaluation of the pesticidal potential of the congress grass, *Parthenium hysterophorus* Linn. on the mustard aphid, *Lipaphis erysimi* (Kalt.). *Journal of Environmental Biology*, 23(1), 15–18.
- Song, B., Chuah, T., Tam, S. M., & Olsen, K. M. (2014). Malaysian weedy rice shows its true stripes: wild *Oryza* and elite rice cultivars shape agricultural weed evolution in Southeast Asia. *Molecular Ecology*, 23(20), 5003–5017.

- Soujanya Philomina, N., & Rao, J. V. S. (1999). Allelopathic effects of some weeds on germination of bajra. *Journal of Environmental Biology*, 20(4), 325–328.
- Stoll, M. E., Price, A. J., & Jones, J. R. (2006). Cover crop extract effects on radish radicle elongation. *28th Southern Conservation Tillage Conference*, 28, 184–186.
- Strategy, A. W. (2017). *Invasive Plants and Animals Committee 2016, Australian Weeds Strategy 2017 to 2027, Australian Government Department of Agriculture and Water Resources, Canberra. Invasive*. Department of Agriculture and Water Resources. Canberra. pp. 1-48.
- Sturm, D. J., Kunz, C., & Gerhards, R. (2016). Inhibitory effects of cover crop mulch on germination and growth of *Stellaria media* (L.) Vill., *Chenopodium album* L. and *Matricaria chamomilla* L. *Crop Protection*, 90, 125–131.
- Suhairi, T. A. S. T. M., Jahanshiri, E., & Nizar, N. M. M. (2018). Multicriteria land suitability assessment for growing underutilised crop, bambara groundnut in Peninsular Malaysia. *IOP Conference Series: Earth and Environmental Science*, 169(1).
- Suman, A., Shahi, H. N., Singh, P., & Gaur, A. (2002). Allelopathic influence of *Vigna mungo* (black gram) seeds on germination and radical growth of some crop plants. *Plant Growth Regulation*, 38(1), 69–74.
- Sunohara, Y., Shirai, S., Wongkantrakorn, N., & Matsumoto, H. (2010). Sensitivity and physiological responses of *Eleusine indica* and *Digitaria adscendens* to herbicide quinclorac and 2,4-D. *Environmental and Experimental Botany*, 68(2), 157–164.
- Swanton, C. J., Nkoa, R., & Blackshaw, R. E. (2015). Experimental methods for crop–weed competition studies. *Weed Science*, 63(SP1), 2–11.
- Tabaglio, V., Gavazzi, C., Schulz, M., & Marocco, A. (2008). Alternative weed control using the allelopathic effect of natural benzoxazinoids from rye mulch. *Agronomy for Sustainable Development*, 28(3), 397–401.
- Tadić, V., Živković, J., Bigović, D., & Žugić, A. (2019). Variation of parthenolide and phenolic compounds content in different parts of *Tanacetum parthenium* (L.) Schulz Bip., Asteraceae during 18 months storage. *Lekovite Sirovine*, 39(39), 35–39.
- Tafese Bezuneh, T. (2015). Phytochemistry and antimicrobial activity of *Parthenium hysterophorus* L.: A review. *Science Journal of Analytical Chemistry*, 3(3), 30.
- Taha, R. S., Seleiman, M. F., Alotaibi, M., Alhammad, B. A., Rady, M. M., & Mahdi, A. (2020). Exogenous potassium treatments elevate salt tolerance and performances of *Glycine max* L. by boosting antioxidant defense system under actual saline field conditions. *Agronomy*, 10(11), 1741.

- Talakal, T. S., Dwivedi, S. K., & Sharma, S. R. (1995). In vitro and in vivo therapeutic activity of *Parthenium hysterophorus* against *Trypanosoma evansi*. *Indian Journal of Experimental Biology*, 33(11), 89.
- Tan, X. L., Azam-Ali, S., Goh, E. V., Mustafa, M., Chai, H. H., Ho, W. K., Mayes, S., Mabhaudhi, T., Azam-Ali, S., & Massawe, F. (2020). Bambara Groundnut: An Underutilized Leguminous Crop for Global Food Security and Nutrition. *Frontiers in Nutrition*, 7(December), 1–16.
- Tarinezhad, A., Sabouri, A., & Mohammadi, S. A. (2005). Statistical software NTSYS PC application in plant breeding. *The 7th Conference of Iran Statistics*.
- Tefera, T. (2002). Allelopathic effects of *Parthenium hysterophorus* extracts on seed germination and seedling growth of *Eragrostis tef*. *Journal of Agronomy and Crop Science*, 188(5), 306–310.
- Telussa, I., & Nurachman, Z. (2019). Dynamics of β -carotene and fucoxanthin of tropical marine *Navicula* sp. as a response to light stress conditions. *Algal Research*, 41, 101530.
- Temegne, N. C., Dooh, J. P. N., Nbandah, P., Ntsomboh-Ntsefong, G., Taffouo, V. D., & Youmbi, E. (2020). Cultivation and utilization of Bambara groundnut (*Vigna subterranea* (L.) Verdc.), a neglected plant in Cameroon. *Asian Plant Research Journal*, 4, 9–21.
- Tessema, S. S., & Tura, A. M. (2018). Allelopathic property of Parthenin on seed germination and seedling growth of wheat (*Triticum aestivum*) and barley (*Hordeum vulgare*). *International Journal of Chemical and Biochemical Sciences, IJCBS*, 14(2018), 23–27.
- Thandapani, K., Kathiravan, M., Namasivayam, E., Padiksan, I. A., Natesan, G., Tiwari, M., Giovanni, B., & Perumal, V. (2018). Enhanced larvicidal, antibacterial, and photocatalytic efficacy of TiO₂ nanohybrids green synthesized using the aqueous leaf extract of *Parthenium hysterophorus*. *Environmental Science and Pollution Research*, 25(11), 10328–10339.
- Thusoo, S., Gupta, S., Sudan, R., Kour, J., Bhagat, S., Hussain, R., & Bhagat, M. (2014). Antioxidant activity of essential oil and extracts of *Valeriana jatamansi* Roots. *BioMed Research International*, 2014, 1–5.
- Tian, Z., Shen, G., Yuan, G., Song, K., Lu, J., & Da, L. (2020). Effects of *Echinochloa crusgalli* and *Cyperus difformis* on yield and eco-economic thresholds of rice. *Journal of Cleaner Production*, 259, 120807.
- Tiku, A. R., Jeanine Vélez-Gavilán, Asad Shabbir, & Chris Parker. (2014). *Parthenium hysterophorus* (Parthenium weed). In *Biological Control of Weeds in Australia*. Springer International Publishing (Issue 2012, pp. 845–868).
- Trujillo, A. N., González, L. B., Morales, C. G., Guerrero, A. R., Sosa, F. C., & Zuñiga, M. E. E. (2017). Phenolic compounds and parthenolide production from in vitro

- cultures of *Tanacetum parthenium*. *Revista Mexicana de Ingeniería Química*, 16(2), 371–383.
- Tuan, V. D., Hilger, T., MacDonald, L., Clemens, G., Shiraishi, E., Vien, T. D., Stahr, K., & Cadisch, G. (2014). Mitigation potential of soil conservation in maize cropping on steep slopes. *Field Crops Research*, 156, 91–102.
- Turk, M. A., & Tawaha, A. M. (2003). Allelopathic effect of black mustard (*Brassica nigra* L.) on germination and growth of wild oat (*Avena fatua* L.). *Crop Protection*, 22(4), 673–677.
- Upadhyay, R., Jha, A., Singh, S. P., Kumar, A., & Singh, M. (2015). Appropriate solvents for extracting total phenolics, flavonoids and ascorbic acid from different kinds of millets. *Journal of Food Science and Technology*, 52(1), 472–478.
- Venkataiah, B., Ramesh, C., Ravindranath, N., & Das, B. (2003). Charminarone, a seco-pseudoguaianolide from *Parthenium hysterophorus*. *Phytochemistry*, 63(4), 383–386.
- Verdeguer, M., Blázquez, M. A., & Boira, H. (2009). Phytotoxic effects of *Lantana camara*, *Eucalyptus camaldulensis* and *Eriosephalus africanus* essential oils in weeds of Mediterranean summer crops. *Biochemical Systematics and Ecology*, 37(4), 362–369.
- Verma, K. K., Song, X.-P., Zeng, Y., Guo, D.-J., Singh, M., Rajput, V. D., Malviya, M. K., Wei, K.-J., Sharma, A., & Li, D.-P. (2021). Foliar application of silicon boosts growth, photosynthetic leaf gas exchange, antioxidative response and resistance to limited water irrigation in sugarcane (*Saccharum officinarum* L.). *Plant Physiology and Biochemistry*, 166, 582–592.
- Vijayalakshmi, P., Vijayalakshmi, K. M., & Nanda Kumar, N. V. (1999). Depolarizing neuromuscular junctional blocking action of *Parthenium hysterophorus* leaf extracts in rat. *Phytotherapy Research*, 13(5), 367–370.
- von Hertwig Bittencourt, H., Trezzi, M. M., Teixeira, S. D., da Silva Bonome, L., de Vargas, A. G., da Silva Souza Filho, A., & Oldoni, T. C. (2018). Chemical ecology of *Eragrostis plana* helps understanding of the species' invasiveness in an agroecosystem community. *Crop and Pasture Science*, 69(10), 1050–1060.
- Vurayai, R., Emongor, V., & Moseki, B. (2011). Effect of water stress imposed at different growth and development stages on morphological traits and yield of bambara groundnuts (*Vigna subterranea* L. Verdc). *American Journal of Plant Physiology*, 6(1), 17–27.
- Wakjira, M. (2009). Allelopathic effects of *Parthenium hysterophorus* L. on germination and growth of onion. *Allelopathy Journal*, 24(2), 351–362.

- Wakjira, M., Berecha, G., & Bulti, B. (2005). Allelopathic effects of *Parthenium hysterophorus* extracts on seed germination and seedling growth of lettuce. *Tropical Science*, 45(4), 159–162.
- Wakjira, M., Berecha, G., & Tulu, S. (2009). Allelopathic effects of an invasive alien weed *Parthenium hysterophorus* L. compost on lettuce germination and growth. *African Journal of Agricultural Research*, 4(11), 1325–1330.
- Wang, P., Zhang, X., & Kong, C. (2013). The response of allelopathic rice growth and microbial feedback to barnyardgrass infestation in a paddy field experiment. *European Journal of Soil Biology*, 56, 26–32.
- Wang, X., Wang, J., Zhang, R., Huang, Y., Feng, S., Ma, X., Zhang, Y., Sikdar, A., & Roy, R. (2018). Allelopathic effects of aqueous leaf extracts from four shrub species on seed germination and initial growth of *Amygdalus pedunculata* pall. *Forests*, 9(11), 1–14.
- Wani, A. S., Hayat, S., Ahmad, A., & Tahir, I. (2017). Efficacy of brassinosteroid analogues in the mitigation of toxic effects of salt stress in *Brassica juncea* plants. *Journal of Environmental Biology*, 38(1), 27.
- Xie, G., Zhou, J., & Yan, X. (2011). *Encyclopedia of traditional Chinese medicines: Molecular structures, pharmacological activities, natural sources and applications*. 2nd edition, Ashgate, England, 2003. Springer.
- Yadav, A., & Garg, V. K. (2011). Recycling of organic wastes by employing *Eisenia fetida*. *Bioresource Technology*, 102(3), 2874–2880.
- Yang, C.-M., Lee, C.-N., & Chou, C.-H. (2002). Effects of three allelopathic phenolics on chlorophyll accumulation of rice (*Oryza sativa*) seedlings: Inhibition of supply-orientation. *Botanical Bulletin of Academia Sinica*, 43(4), 299–304.
- Yeomans, J. C., & Bremner, J. M. (1988). A rapid and precise method for routine determination of organic carbon in soil. *Communications in Soil Science and Plant Analysis*, 19(13), 1467–1476.
- Yoneya, K., & Takabayashi, J. (2014). Plant–plant communication mediated by airborne signals: ecological and plant physiological perspectives. *Plant Biotechnology*, 14–827.
- Yu, G., Jiang, Y., Xu, L., & Li, G. Y. (2015). Multi-objective energy-efficient resource allocation for multi-RAT heterogeneous networks. *IEEE Journal on Selected Areas in Communications*, 33(10), 2118–2127.
- Zeng, R. Sen. (2014). Allelopathy - The solution is indirect. *Journal of Chemical Ecology*, 40(6), 515–516.
- Zhang, D., Zhang, J., Yang, W., & Wu, F. (2010). Potential allelopathic effect of *Eucalyptus grandis* across a range of plantation ages. *Ecological Research*, 25(1), 13–23.

Zhou, B., Kong, C. H., Li, Y. H., Wang, P., & Xu, X. H. (2013). Crabgrass (*Digitaria sanguinalis*) allelochemicals that interfere with crop growth and the soil microbial community. *Journal of Agricultural and Food Chemistry*, 61(22), 5310–5317.

Zimdahl, R. L. (2018). *Fundamentals of weed science*. 5th edition, Academic press in Elsevier (pp. 1-735).

Zohaib, A., Abbas, T., & Tabassum, T. (2016). Weeds cause losses in field crops through allelopathy. *Notulae Scientia Biologicae*, 8(1), 47–56.

