

UNIVERSITI PUTRA MALAYSIA

IMMUNOMODULATORY ACTIVITY OF MANNHEIMIA HAEMOLYTICA A2 LIPOPOLYSACCHARIDE

AZLINA BT MOHD SALIM

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By

AZLINA BT MOHD SALIM

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By

AZLINA BT MOHD.SALIM

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Chairman: Associate Professor Daud Ahmad Israf Ali, Ph.D.

Faculty : Medicine and Health Science

Bacterial lipopolysaccharides (LPS) are endotoxins. However, there ample evidence to support its role in immunomodulation. In fact LPS is commonly used as a B cell mitogen. The activity of LPS from different genera of bacteria varies considerably which makes generalisation of characteristics difficult. Despite their lethal consequences, the LPS of *Mannheimia haemolytica* A2 have not been tested for immunomodulatory activity. Therefore, the objective of this study is to investigate the adjuvant properties of LPS from *Mannheimia haemolytica* A2 upon peripheral and mucosal immunity toward protein antigen.

The experimental results indicate that, the group of mice that received 10 µg LPS in oil induced anti-BSA IgG response in serum and pulmonary antibody response when administered intraperitoneally. However, in the IgG and IgA intestinal antibody response, mice received 10 µg LPS without oil showed significantly high compared to controls

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(p≤0.05). This indicates that LPS can be used as oral adjuvant to enhance response at mucosal level. For the oral immunisation, the specific anti-OVA IgG titre in serum and IgA levels in the intestinal fluid were significantly high in group that received 500 µg LPS. The oral administration of 500 µg LPS also increased the number of anti-OVA ASC (antigen secreting cell) observed in spleen, mesentric lymph node and Peyer's patch. Hence, LPS from *Mannheimia haemolytica* A2 does stimulate peripheral and mucosal immune responses to an unrelated antigen.



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AKTIVITI IMUNOMODULATOR MEMBRAN LIPOPOLISAKARIDA DARIPADA BAKTERIA MANNHEIMIA HAEMOLYTICA A2

Oleh

AZLINA BT MOHD.SALIM

September 2003

Pengerusi: Profesor Madya Daud Ahmad Israf Ali, Ph.D.

Fakulti : Perubatan dan Sains Kesihatan

Lipopolisakarida (LPS) dalam bakteria merupakan sejenis endotoksin. Walaubagaimanapun, terdapat bukti yang menyatakan bahawa LPS mempunyai peranan sebagai imunomodulator di mana LPS biasanya bertindak sebagai mitogen dalam sel B. Aktiviti LPS daripada pelbagai genera bakteria sangat meluas sehingga agak sukar untuk membuat pengkelasan. Selain daripada mempunyai sifat-sifat toksin, ujikaji terhadap aktiviti imunomodulator masih belum dijalankan terutama LPS dari jenis *Mannheimia haemolytica* A2. Oleh yang demikian, di dalam eksperimen ini, beberapa ujikaji telah

sama ada LPS dari jenis *Mannheimia haemolytica* A2 boleh bertindak sebagai 'adjuvant' di dalam sistem periperal dan mukosa terhadap antigen.

Hasil kajian menunjukkan bahawa, kumpulan mencit yang diberi suntikan intraperitoneal campuran 10 µg LPS dan minyak menghasilkan antibodi anti-BSA IgG yang tinggi dalam serum dan

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ekstrak pulmonari. Walau bagaimanapun, di dalam ekstrak intestin kandungan antibodi IgG dan IgA telah menunjukkan peningkatan yang baik daripada kumpulan mencit yang disuntik dengan 10 µg LPS tanpa minyak (p≤0.05) berbanding kawalan. Ini mununjukkan bahawa LPS boleh diberi secara oral untuk merangsang pembentukkan antibodi di dalam sistem mukosa. Keputusan eksperimen menunjukkan bahawa 500 µg LPS yang diberi secara oral kepada kumpulan mencit telah meningkatkan antibodi anti-OVA IgG dalam serum dan IgA dalam ekstrak intestin. Selain daripada meningkatkan paras antibodi, bilangan sel-sel limfoid (anti-OVA ASC) juga meningkat terutama di dalam organ limpa, sel-sel mesentrik dan Peyer's patch. Keputusan ujikaji jelas menunjukkan bahawa LPS dari bakteria jenis *Mannheimia haemolytica* A2 boleh merangsang tindakbalas imun di dalam sistem periperal dan mukosa terhadap antigen.



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TABLE OF CONTENTS

				Page
ABSTRACT ABSTRAK ACKNOWLEDGEMENTS APPROVAL DECLARATION			ii iv vi vii ix	
LIST		GURES	ATIONS	xiii xiv xviii
2101	OT THE			244111
СНА	PTER			
1	INTR	ODUCI	TION	1
2	LITER 2.1	Adjuv 2.1.1 2.1.2 2.1.3 2.1.4 2.1.5	E REVIEW ants Monophosphoryl lipid A (MPL) Titermax® Saponins 2.1.3.1 Structure of Quillaja Saponins Liposome Muramyl Dipeptide (MPD) and derivatives Bacterial Lipopolysaccharides	4 4 6 7 8 9 10
		2.1.8 c 2.1.9	Mechanism of LPS adjuvanticity Cholera toxin (CT) E. Coli heat-labile enterotoxin (LT) Oral immunisation	17 20 21 22
3			f Lipopolysaccharides (LPS) from haemolytica A2	26
	3.1	Materi 3.1.1 3.1.2	Brain Heart Infusion (BHI) broth preparation	26 27 27 27
		3.1.3	Bacteria strains and growth conditions Lipopolysaccharides extraction	28 28



		3.1.5	Protein Assay	29
		3.1.6	Gel preparation	30
			3.1.6.1 Separating gel	30
			3.1.6.2 Stacking gel	30
		3.1.7	Sodium Dodecyl Sulphate	
			Polyacrylamide gel Electrophoresis	
			(SDS-PAGE)	30
		3.1.8	,	31
	3.2	Result	S	32
		3.2.1	Protein estimation	32
		3.2.2	Silver staining	33
	3.3	Discus	9	34
			. C I DOLL	
4			at of anti-BSA immune response	
			toneal immunisation with BSA	
			red with Mannheimia	
	haen	nolytica P	A2 LPS	36
		Introd	uction	36
	4.1		als and Method	
	4.1			37
		4.1.1		37
			Animals	37
		4.1.3		37
		4 1 4	complex	20
			Immunisation protocol	38
			Sample collection	38
		4.1.6	2	
			(ELISA) for determination of serum	
			antibodies	40
		4.1.7	2	
			(ELISA) for determination of	
			intestinal and	41
		4.1.8	Enzyme linked immunosorbent assay	
			(ELISA) for determination of	
			coproantibodies	43
		4.1.9	Statistical analysis	43
	4.2	Result		44
		4.2.1	Serum antibody response	44
		4.2.2	Intestinal antibody response	44
		4.2.3	Pulmonary antibody response	45
		4.2.4	Faecal IgA response	45
	4.3	Discus	sion	58



	5	Enhancement of anti-OVA immune responses via oral boosting with OVA co-administered with				
			_	aemolytica A2 LPS	61	
			T / 1		<i>C</i> 1	
		- 1		uction	61	
		5.1	5.1.1	ials and Methods Mice	63	
					63	
				Immunisation	63	
				Sample collection	63	
			3.1.4	In-vitro restimulation of lymphocytes	66	
			5.1.5	Enzyme linked immunosorbent assay	00	
			0.1.0	(ELISA) to measure specific IgG		
				responses	66	
			5.1.6	Enzyme linked immunosorbent assay		
			0.110	(ELISA) to measure specific IgA		
				responses	67	
			5.1.7	Enzyme linked immunospot assay		
				(ELISPOT) to measure antibody		
				secreting cells	69	
			5.1.8	Statistical analysis	70	
		5.2	Result	S	71	
			5.2.1	- L	71	
			5.2.2	<i>5</i> 1	71	
			5.2.3	Faecal IgA response	76	
			5.2.4	3 3 1	76	
			5.2.5	Ŷ O		
		- 0	ъ.	(ASCs) in SP, PP and MLN	76	
		5.3	Discus	ssion	82	
	6	GENE	RAL DI	SCUSSION AND CONCLUSION	85	
					00	
		ERENCES				
		NDICE			99	
	BIOD	ATA O	F THE	AUTHOR	115	



LIST OF TABLES

		Page
Table 2.1	Side effect of FIA (Rajesh et al., 1993)	6
Table 2.2	Biological activities of lipid A or LPS	16
Table 4.1	EXPERIMENTAL DESIGN	39
Table 5.1	EXPERIMENTAL DESIGN	65
Table 6.1	Data summarised the adjuvant effects of LPS from <i>Mannheimia haemolytica</i> A2. The group of mice that received 10 μg LPS in oil for IP and 500 μg for oral immunisation showed significantly high in antibody titres (p≤0.05)	86



LIST OF FIGURES

		Page
Figure 2.1	Gram-negative bacteria and cell membrane (Nakano & Matsuura, 1995)	14
Figure 3.1	Detection of lipopolysaccharides by silver stained followed by Coomasie blue staining. Samples were analysed on 12% SDS-polyacrylamide gel electrophoresis. Lane 1, 10 µg <i>S. typhimurium</i> LPS smear (control); lane 2, 10 µg <i>E. coli</i> 0111:B4 LPS smear (control); lanes 3, 4 and 5, <i>M. haemolytica</i> LPS smears at 10, 5 and 1µg respectively. The molecular weight protein markers (kDa) were just to detect whether there was a protein contamination on the gel and LPS samples	33
Figure 4.1(a)	Peripheral anti-BSA IgG antibody responses in mice. Mice were immunised with BSA-FCA ■, BSA 1 µg LPS in oil ★, BSA 10 µg LPS in oil ★ and 100 µg BSA as a control ◆	46
Figure 4.1(b)	Peripheral anti-BSA IgG antibody responses in mice. Mice were immunised with BSA-FCA ■, BSA 1 µg LPS without oil ▲, BSA 10 µg LPS without oil △ and 100 µg BSA as a control ◆	47
Figure 4.2(a)	Peripheral anti-BSA IgA antibody responses in mice. Mice were immunised with BSA-FCA \square , BSA 1 μ g LPS in oil \spadesuit , BSA 10 μ g LPS in oil \triangle and 100 μ g BSA as a control $*$	48
Figure 4.2(b)	Peripheral anti-BSA IgA antibody responses in mice. Mice were immunised with BSA-FCA □, BSA 1 µg LPS without oil ▲, BSA 10 µg LPS without oil ■ and 100 µg BSA as a control ★	49



		Page
Figure 4.3(a)	Peripheral anti-BSA IgM antibody responses in mice. Mice were immunised with BSA-FCA \spadesuit , BSA 1 μ g LPS in oil \diamondsuit , BSA 10 μ g LPS in oil \triangle and 100 μ g BSA as a control $*$	50
Figure 4.3(b)	Peripheral anti-BSA IgM antibody responses in mice. Mice were immunised with BSA-FCA ◆, BSA 1 µg LPS without oil ♦, BSA 10 µg LPS without oil ▲ and 100 µg BSA as a control ★	51
Figure 4.4	Intestinal anti-BSA IgA antibody responses of mice immunised with 100 μg BSA with LPS or without LPS. Mice receiving 10 μg LPS showed significant increase compared with BSA/Freund's and other groups (p≤0.05)	52
Figure 4.5	Intestinal anti-BSA IgG antibody responses of mice immunised with 100 μg BSA with LPS or without LPS. Mice receiving 10 μg LPS showed significant increase compared with BSA/Freund's and other groups (p≤0.05)	53
Figure 4.6	Pulmonary anti-BSA IgG antibody responses of mice immunised with 100 μg BSA with LPS or without LPS. Mice receiving 10 μg LPS showed significant increase compared with group that received $1\mu g$ LPS with or without oil (p≤0.05)	54
Figure 4.7	Pulmonary anti-BSA IgA antibody responses of mice immunised with 100 µg BSA with LPS or without LPS. The result showed that there was no significant different among all groups	55
Figure 4.8(a)	Feacal extract anti-BSA IgA antibody responses of mice immunised with BSA-FCA ■, BSA-1 µg LPS in oil △, BSA-10 µg LPS in oil ★ 100 µg BSA ♠ and PBS ♠	56



		Page
Figure 4.8(b)	Feacal extract anti-BSA IgA antibody responses of mice immunised with BSA-FCA ■, BSA-1 µg LPS without oil △, BSA-10 µg LPS without oil ★, 100 µg BSA ▲ and PBS ◆	57
Figure 5.1	Peripheral anti-OVA IgG antibody responses in mice. Mice were immunised with 1 mg OVA for ip and 10 mg OVA for oral with or without LPS as described in Table 5.1	72
Figure 5.2	Peripheral anti-OVA IgA antibody responses in mice. Mice were immunised with 1 mg OVA for ip and 10 mg OVA for oral with or without LPS as described in Table 5.1	73
Figure 5.3	Intestinal anti-OVA IgA antibody responses in mice immunised with 10 mg OVA with or without LPS. The concentration of IgA in intestinal washing were significantly increased in mice that received 500 µg LPS compared with control (p≤0.05)	74
Figure 5.4	Intestinal anti-OVA IgG antibody responses in mice immunised with 10 mg OVA with 10 μ g CT were higher than all treated groups but no significant difference between controls and group that received 500 μ g LPS	75
Figure 5.5	Feacal extract anti-OVA IgA antibody responses in mice. Mice were immunised with 10 mg OVA orally with 100 µg LPS, OVA-500 µg LPS, OVA-CT and OVA alone as a control	77
Figure 5.6	Pulmonary anti-OVA IgG antibody responses in mice. Mice were immunised with 10 mg OVA orally with 100 µg LPS, OVA-500 µg LPS, OVA-CT and OVA alone as a control	78



		Page
Figure 5.7	Anti-OVA IgA and IgG secreting cells/10 ⁶ cells in spleen (SP) from mice immunised orally with 10 mg OVA-100 µg LPS, OVA-500 µg LPS, OVA-CT and OVA alone as a control	79
Figure 5.8	Anti-OVA IgA and IgG secreting cells/10 ⁶ cells in Peyer's patch (PP) from mice immunised orally with 10 mg OVA-100 µg LPS, OVA-500 µg LPS, OVA-CT and OVA alone as a control	80
Figure 5.9	Anti-OVA IgA and IgG secreting cells/10 ⁶ cells in Mesentric lymph node (MLN) from mice immunised orally with 10 mg OVA-100 µg LPS, OVA-CT and OVA alone as a control	81



LIST OF ABBREVIATIONS

FCA Freund's complete adjuvant FIA Freund's incomplete adjuvant

LPS Lipopolysaccharides
DAG 2,3 -dideoxy-D-glucose
OMPs Outer membrane proteins
BSA Bovine serum albumin

OVA Ovalbumin

MPL Monophosphoryl lipid A MDP Muramyl dipeptide CWS Cell wall skeleton

EAU Experimental autoimmune ureoretinitis

CTL Cytotoxic T lymphocyte
TNF Tumour necrosis factor

IL Interleukin IFN Interferon

APC Antigen presenting cell

CT Cholera toxin

SDS-PAGE Sodium dodecyl sulphate polyacrylamide gel

electrophoresis

BHI Brain heart infusion

TEMED N-N-N'-N'-tetramethylethylenediamine

TNP Trinitropenyl hepten
PBS Phosphate buffered saline
PMSF Phenylmethylsulfonyl fluoride

ELISA Enzyme linked immunosorbent assay

TMB Tetramethylbenzidine LT Heat-labile enterotoxin

GALT Gut-associate lymphoid tissues

UPM Universiti Putra Malaysia

IP Intraperitoneal

HRPO Horseradish peroxidase

ELISPOT Enzyme linked immunospot assay

AEC Amino ethyl carbazole



CHAPTER I

INTRODUCTION

Immunomodulators are compounds that can cause either suppression or potentiation of the immune system. Immunomodulators are used to stimulate immune responses to vaccines. Adjuvants are generally considered to be materials that are added to vaccines with the intent of potentiating the immune response so that a greater amount of antibody is produced. The most common adjuvants that are safe for human use are aluminium hydroxide, aluminium phosphate and calcium phosphate (Edelman, 1980). Adjuvants that have been used for research purposes are oil emulsions which were first used by Le Moignic and Pinoy (1916), who found that a suspension of killed Salmonella typhimurium in mineral oil increased the immune response. Freund's complete adjuvant (FCA), a water mineral oil emulsion with killed mycobacterium is used extensively to augment the immune response of laboratory animals for experimental purposes. Freund's incomplete adjuvant (FIA) which is water in oil emulsion without mycobacterium is used successfully in veterinary rabies, parainfluenza and Newcastle disease. There are also other

other

bacteria

and

their

products.

from

adjuvants

derived



They play an important role in the immune response. The common bacterial used as adjuvants mainly for experimental purposes include mycobacterium, *Corynebacterium parvum* or *Corynebacterium granulosum*, *Bordetella pertussis* and lipopolysacharides (Rajesh *et al.*, 1993).

Lipopolysacharides (LPS) are common bacterial products that can be used as adjuvants. It is a constituent of the outer membrane of gramnegative bacteria that contains 2,3-dideoxy-D-glucose (DAG), an amino sugar attached to a lipid A backbone. LPS has long been known to be a potent modulator of immune reactions such as B cell mitogenicity, immunogenicity, polyclonal antibody synthesis, adjuvanticity, macrophage activation, monokine secretion and regulation of IgA responses (Qureshi et al., 1990). LPS also induces regression of spontaneous, induced and transplanted tumours in laboratory animals. Its toxicity which involves pyrogenicity, tumour necrosis and lethality has limited exploration of its therapeutic potential in man. However, studies have shown that the toxic compound of the LPS molecule can be cleaved chemically (Ribi et al., 1986). The dose of LPS and the time of exposure will determine the resultant effects that will be seen in the host. It has been shown to effect on B cell but not on T lymphocytes. LPS has been suggested to bind non-specifically to the cell surface followed by intercalation of its hydrophobic lipid A moiety into the plasma membrane (Jacobs, 1992). Others have implicated the involvement of specific receptors on the surface (Tahiri & Chaby, 1990). Recently, it has



been reported that the existence of membrane-associated LPS and lipid A-specific binding proteins on responsive cells, including macrophages, lymphocytes and B cells (Hara Kuge *et al.*, 1990).

There are many gram-negative bacteria that have been used as a source of LPS, particularly members of the enterobacteriaceae {Salmonella, Shigella, Escherichia, Proteus, Pseudomonas, Klebsiella, Mannheimia (Pasteurella). Mannheimia haemolytica serotype A2 is the most commonly occurring serotype associated with pneumonic pasteurellosis in cattle and sheep. Immunological studies have been done to determine responses in the respiratory tract using killed M. haemolytica A2 (Zamri et al., 1999). Although experimental vaccines against pneumonic pasteurellosis have been done including both live and killed bacteria (Purdy et al., 1986), research has attempted to identify important immunogenic components of M. haemolytica such as iron-regulated outer-membrane proteins (OMPs) (Gilmor et al., 1991), carbohydrate-protein subunits (Lesley et al., 1985) and LPS (Confer et al., 1986). The immunomodulatory properties of LPS from M. haemolytica A2 have not been studied despite being an important pathogen of livestock. Therefore, the objectives of the study are:-

- 1. To extract and purify LPS from Mannheimia haemolytica A2.
- 2. To determine the immunostimulatory effect of *M. haemolytica* A2 LPS upon peripheral and mucosal immunity toward model protein antigens such as bovine serum albumin (BSA) and ovalbumin (OVA).



CHAPTER 2

LITERATURE REVIEW

2.1 Adjuvants

The response to an immunogen can be enhanced by the use of adjuvants. Adjuvants are compound that potentiate the immune response when mixed and administered with antigen. They enhance and prolong antibody production and increase effector cell counts (Coleman et al., 1992). From this definition, it is obvious that adjuvants are useful and necessary components of the development of vaccines. An adjuvant must be able to increase both amount and duration of antibody and T helper cell response. The mode of action of adjuvants was summarised by Chedid et al., (1975). First, the formation of a depot of antigen at the site of inoculation which is slowly released. Secondly, the presentation of antigen to immunocompetent cells and the production of different lymphokines such as various interleukins and tumour necrosis factor.

It has long been known that many adjuvants are surface-active agents (Allison, 1979). The importance of surface activity in adjuvanticity has been confirmed with modern adjuvants. Saponins, lipid A and muramyl dipeptide (MDP) derivatives are all surface active



in that they consist of discrete hydrophilic and hydrophobic domains. With the use of adjuvants, less antigen is required, thus reducing vaccine production cost (Rajesh et al., 1993). For many years there has been a search to find adjuvants with the ability to potentiate the immune response but with minimal side effects. Freund's complete adjuvant (FCA) is one of the most potent oil adjuvants described so far and used extensively to augment the immune response of laboratory animals for experimental purposes. FCA as originally formulated was too toxic to be used in human. The water in mineral oil emulsion without Mycobacteria is known as Freund's incomplete adjuvant (FIA). FIA was used successfully in a number of veterinary vaccines including foot-andmouth disease, rabies, parainfluenza and Newcastle disease. FIA has been used in human, particularly with influenza and killed poliomyelitis vaccines by enhancing the immunogenicity of the vaccines (Salk, 1977). However, FIA is unsuitable for routine human application because of previously reported side effects (Table 2.1). From these classical adjuvants, there are many alternative adjuvant that have been used for many years such as monophosphoryl lipid A (MPL), Titermax[®], saponins, liposome, muramyl dipeptide (MDP), lipopolysaccharides (LPS), cholera toxin and heat labile enterotoxin.



Table 2.1: Side effect of FIA (Rajesh et al., 1993)

Local reaction

Sterile abscesses and cysts Granulomas Inflammation

Carcinogenicity

Oil induced neoplasm in mice Arlacel A induced carcinogenicity in mice

2.1.1 Monophosphoryl lipid A (MPL)

MPL is a non-toxic lipid A with the lack of phosphate on the reducing end of the disaccharide backbone which has 1000-fold less toxic than endotoxin (Ribi, 1984). MPL has immunostimulatory activities such as B cell mitogenicity, activation of macrophages, colonystimulating activity and the induction of interleukin-1 (IL-1) production by human monocytes (Rudbach & Cantrell, 1990). Tomai *et al.*, (1987) showed that MPL has an adjuvant activity with poly-L-lysine antigen in LPS-hyporesponsive C3H/HeJ and C57BL/10ScN mice and in aging BALB/c mice, which have a low capacity to produce antibody, when examined by a modified haemolytic plaque assay. They also reported an adjuvant effect to sheep red blood cell (SRBC) antigen in young and ageing mice (Johnson *et al.*, 1987). Schneerson *et al.*, (1991) also reported that both the primary and secondary serum antibody responses to the capsular polysaccharides (CP) antigens in mice were stimulated

