

# **UNIVERSITI PUTRA MALAYSIA**

# CARBON FLUX ANALYSIS OF LIPID BIOSYNTHESIS PATHWAYS IN OIL PALM (ELAEIS GUINEENSIS JACQ. TENERA)

EMILY QUEK MING POH

FPSK (M) 2002 3

### CARBON FLUX ANALYSIS OF LIPID BIOSYNTHESIS PATHWAYS IN OIL PALM (ELAEIS GUINEENSIS JACQ. TENERA)

By

### EMILY QUEK MING POH

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Science

March 2002

### DEDICATION

I dedicate this thesis to my parents and my family.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

### CARBON FLUX ANALYSIS OF LIPID BIOSYNTHESIS PATHWAYS IN OIL PALM (ELAEIS GUINEENSIS JACQ. TENERA)

By

### EMILY QUEK MING POH

#### March 2002

### Chairman: Associate Professor Ong King Kok, Ph.D.

Faculty: Medicine and Health Sciences

The aim of this study was to investigate the carbon flux through lipid biosynthesis pathways in the oil palm (*Elaeis guineensis* Jacq. *tenera*) using metabolic control analysis (MCA). Three types of tissue from oil palm, namely liquid culture, plastid and mesocarp were used. The results showed mesocarp tissue was the most suitable tissue for carbon flux analysis because it incorporated the radioactive precursors mainly into triacylglycerol (TAG). Further analysis on different stages of fruit development was carried out using mesocarp tissue at 12, 15 and 20 weeks after anthesis (WAA). It was confirmed that 20-WAA mesocarp tissue was the best stage of fruit development for metabolic flux studies because it reflected biosynthesis of storage lipid. Three modes of radiolabel introduction into the oil palm fruits were investigated, namely injecting the radiolabel into the fruits still attached to the palm, injecting the radiolabel into the loose fruit and injecting the radiolabel into incubation mixture containing mesocarp tissue slices. The level of radioactivity in fruits attached to the palm was lower than the other two modes of radiolabel introduction. Carbon flux of lipid biosynthesis pathways was modulated by temperature and the inhibitor 2-bromoctanoate. Radiolabels [1-<sup>14</sup>C]



acetate and [U-<sup>14</sup>C] glycerol were used to monitor the carbon flux through the lipid biosynthesis pathways. Temperature caused a constraint in the distribution of radioactivity at the level of diacylglycerol acyltransferase (DAGAT). Therefore, DAGAT may be a regulatory enzyme. 2-Bromooctanoate inhibited the carbon flux of lipid biosynthesis pathways. The overall results of MCA suggested that the control of carbon flux in the oil palm may be distributed over two blocks of the lipid metabolic pathway, namely the fatty acid biosynthesis block and TAG formation block. Acetyl-CoA carboxylase (ACCase) plays an important role in fatty acid biosynthesis block while DAGAT plays an important role in the TAG formation block. The molecular structure of ACCase was investigated using immunoblotting with streptavidin and screening of ACCase gene in 15-WAA oil palm mesocarp cDNA library. Immunoblots with streptavidin showed the presence of large molecular weight (approximately 180 kDa) multifunctional ACCase and smaller molecular weight (approximately 58 kDa) multisubunit ACCase in oil palm mesocarp. Screening for ACCase gene in 15-WAA oil palm mesocarp cDNA library showed several strong signals corresponding to the putative  $\beta$ -carboxyl transferase subunit of ACCase.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

### ANALISIS FLUKS KARBON TAPAK JALAN BIOSINTESIS LIPID DI DALAM POKOK SAWIT (*ELAEIS GUINEENSIS* JACQ. *TENERA*)

Oleh

### **EMILY QUEK MING POH**

**Mac 2002** 

Pengerusi: Profesor Madya Ong King Kok, Ph.D.

Fakulti: Perubatan dan Sains Kesihatan

Matlamat kajian ini adalah untuk mengkaji fluks karbon melalui tapak jalan biosintesis lipid di dalam pokok sawit (*Elaeis guineensis* Jacq. *tenera*) dengan menggunakan analisis kawalan metabolik (MCA). Tiga jenis tisu daripada pokok sawit iaitu kultur cecair, plastid dan mesokarpa telah digunakan. Hasil menunjukkan bahawa tisu mesokarpa merupakan tisu yang sangat sesuai untuk analisis fluks karbon kerana ia menukarkan prekursor radioaktif kebanyakannya ke dalam triasilgliserol (TAG). Analisis lanjutan ke atas peringkat perkembangan buah yang berlainan telah dilakukan dengan menggunakan tisu mesokarpa pada 12, 15 dan 20 minggu selepas pendebungaan (WAA). Tisu mesokarpa pada 20 WAA telah disahkan sebagai peringkat perkembangan buah yang terbaik untuk kajian fluks metabolik kerana ia mengimbas biosintesis penyimpanan lipid. Tiga cara kemasukan penanda radioaktif ke dalam buah sawit iaitu menyuntik penanda radioaktif ke dalam buah yang dipetik dan menyuntik penanda radioaktif ke dalam larutan eraman yang mengandungi hirisan tisu mesokarpa telah dikaji. Aras aktiviti radioaktif dalam buah yang masih di pokok adalah lebih



rendah berbanding dengan dua cara kemasukan penanda radioaktif yang lain. Fluks karbon dalam tapak jalan biosintesis lipid telah dimodulasikan oleh suhu dan perencat 2-bromooktanoat. Penanda radioaktif [1-14C] asetat dan [U-14C] gliserol telah digunakan di dalam kajian ini untuk memantau fluks karbon melalui tapak jalan biosintesis lipid. Suhu menyebabkan rencatan dalam penyebaran aktiviti radioaktif pada peringkat diasilgliserol asiltransferase (DAGAT). Oleh itu, DAGAT mungkin merupakan enzim pengawalatur. 2-Bromooktanoat merencat fluks karbon dalam tapak jalan biosintesis lipid. Hasil keseluruhan MCA mencadangkan kawalan fluks karbon dalam buah sawit disebarkan melalui dua blok dalam tapak jalan metabolik lipid iaitu blok biosintesis asid lemak dan blok pembentukan TAG. Asetil-CoA karboksilase (ACCase) memainkan peranan penting dalam blok biosintesis asid lemak manakala DAGAT memainkan peranan penting dalam blok pembentukan TAG. Stuktur molekular ACCase telah dikaji dengan menggunakan pemblotan imuno dengan streptavidin dan penyaringan gen ACCase dalam koleksi cDNA 15-WAA mesokarpa sawit. Pemblotan imuno dengan streptavidin menunjukkan kehadiran protein berberat molekul besar (kira-kira 180 kDa) iaitu ACCase pelbagai-fungsi dan protein berberat molekul kecil (kira-kira 58 kDa) iaitu ACCase pelbagai-subunit di dalam mesokarpa sawit. Penyaringan gen ACCase dalam koleksi cDNA 15-WAA mesokarpa sawit telah menunjukkan beberapa signal yang menyamai subunit  $\beta$ -karboksil transferase ACCase.



### ACKNOWLEDGEMENTS

I would like to express my sincere appreciation and million thanks to my supervisors, Associate Professor Ong King Kok, Professor Khor Hun Teik and Dr Ravigadevi Sambanthamurthi for their suggestion, advice, support and guidance throughout my project.

My appreciation and gratitude go to my parents and my family for their constant support, love and patient throughout my graduate study. My sincere thanks and gratitude are also extended to all the staff of Metabolics Laboratory especially Ms. Jane Sonia, Mr. Andy Yip, En. Jamil, En. Rahim, Pn. Jabariah and Pn. Siti Hasnah for their help towards the success of this project and also to my friends for their support.



# TABLE OF CONTENTS

	Page
DEDICATION	ii
ABSTRACT	iii
ABSTRAK	v
ACKNOWLEDGEMENTS	vii
APPROVAL	viii
DECLARATION	x
LIST OF TABLES	xiv
LIST OF FIGURES	xv
LIST OF ABBREVIATIONS	xviii

# CHAPTER

1	INT	RODUCTION	1
2	LII	TERATURE REVIEW	4
	2.1	Oil Palm	4
	2.2	Palm Oil	7
	2.3	Plant Lipids	8
		2.3.1 Classification of Lipids	8
		2.3.2 Functions of Lipids	10
	2.4	Fatty Acid Biosynthesis	10
		2.4.1 Acyl Carrier Protein (ACP)	13
		2.4.2 Acetyl-CoA	14
		2.4.3 Acetyl-CoA Carboxylase (ACCase)	15
		2.4.4 Fatty Acid Synthase (FAS)	16
		2.4.5 Desaturase	17
		2.4.6 Acyl-ACP Thioesterase	18
	2.5	Triacylglycerol Biosynthesis	18
	2.6	The Kennedy Pathway	19
	2.7	Metabolic Control Analysis (MCA)	21
	2.8	Metabolic Engineering	26
3	MA	TERIALS AND METHODS	28
	3.1	Chemicals and Reagents	28
	3.2	Tagging of Oil Palm Fruits	30
	3.3	Experimental Fruits	30
	3.4	Oil Palm Liquid Cultures	30
	3.5	Preparation of Mesocarp Tissue Slices	31
	3.6	Isolation of Crude Plastids	31
	3.7	Bio-Rad Protein Assay	32
	3.8	Radioactive Incorporation	32
		3.8.1 [1- <sup>14</sup> C] Acetate Incorporation in Liquid Cultures	33
		3.8.2 [1- <sup>14</sup> C] Acetate Incorporation in Plastids	33
		3.8.3 $[1^{-14}C]$ Acetate or $[U^{-14}C]$ Glycerol Incorporation in	
		Mesocarp Tissue Slices	34



3	3.9	Mode of Radiolabel Uptake into the Oil Palm Fruits	34
3	3.10	Determination of pH Optimum	35
- 3	3.11	Time Course Experiment of Incorporation of Radiolabel	35
3	3.12	Optimisation of MES-NaOH Concentration	36
3	3.13	Effect of Temperature	36
3	3.14	Inhibition with 2-Bromooctanoate	36
		3.14.1 Preparation of 2-Bromooctanoate Solution	36
		3.14.2 Incubation with 2-Bromooctanoate	37
3	3.15	Analysis of Radioactive Incorporation Products	37
3	3.16	Thin Layer Chromatography (TLC)	38
		3.16.1 Preparation of TLC Plates	38
		3.16.2 Application of Samples onto TLC Plate	39
		3.16.3 Separation of Lipid Classes	39
		3.16.4 Detection of Lipids	39
		3.16.5 Determination of Radioactivity of Separated Lipids	40
3	3.17	Separation of Acyl-ACPs and Acyl-CoAs	40
3	3.18	Liquid Scintillation Counting	41
3	3.19	Extraction of Proteins	41
3	3.20	Acetyl-CoA Carboxylase (ACCase) Assay	42
3	3.21	Sodium Dodecyl Sulphate - Polyacrylamide Gel Electrophoresis	
		(SDS-PAGE)	42
		3.21.1 Preparation of SDS-PAGE Gel	43
		3.21.2 Electrophoresis Conditions for SDS-PAGE	44
		3.21.3 Development of SDS-PAGE Gel	44
3	3.22	Western Blotting	44
		3.22.1 Preparation for Western Blotting	45
		3.22.2 Assembly of the Western Blotting Unit	45
3	3.23	Immunoblotting with Streptavidin	46
		3.23.1 Preparation of Detection Solution	47
		3.23.2 Immunoblotting	47
3	3.24	Preliminary ACCase Gene Isolation	48
		3.24.1 Preparation of ACCase Probe	48
		3.24.2 Screening of ACCase Gene in 15-WAA Mesocarp cDNA	
		Library	54
		3.24.3 Storage of Transformed Cells	57
3	3.25	Statistical Analysis	57
	RES	ULTS AND DISCUSSION	58
2	4.1	Conditions for Incorporation of Radiolabel in Crude Plastids	58
2	4.2	Determination of Suitable Tissue	60
4	4.3	Mode of Radiolabel Uptake into the Oil Palm Fruits	66
4	4.4	Determination of pH Optimum	68
4	4.5	Time Course Experiment of Incorporation of Radiolabel	71
4	4.6	Optimisation of MES-NaOH Concentration	73
4	4.7	Effect of Temperature on Lipid Biosynthesis in Oil Palm Mesocarp	
		Tissue	73
		4.7.1 Effect of Temperature on the Incorporation of Radioactivity	
		into Total Lipids in Oil Palm Mesocarp Tissue	73



		4.7.2	Effect of Temperature on the Distribution of Radiolabel into	
			Lipid Classes in Oil Palm Mesocarp Tissue	76
		4.7.3	Effect of Temperature on Acyl-ACP and Acyl-CoA Pools in	
			Oil Palm Mesocarp Tissue	78
	4.8	Manipu	alation of Lipid Biosynthesis in Oil Palm Mesocarp Tissue by	
		the Inh	ibitor 2-Bromooctanoate	83
		4.8.1	Effect of 2-Bromooctanoate on the Incorporation of $[1^{-14}C]$	
			Acetate into Total Lipids in Oil Palm Mesocarp Tissue	83
		4.8.2	Effect of 2-Bromooctanoate on the Incorporation of $[1^{-14}C]$	
			Acetate into Lipid Classes in Oil Palm Mesocarp Tissue	85
		4.8.3	Effect of 2-Bromooctanoate on Acyl-ACP and Acyl-CoA	
			Pools in Oil Palm Mesocarp Tissue	85
		4.8.4	Effect of 2-Bromooctanoate on the Incorporation of $[U^{-14}C]$	
			Glycerol into Total Lipids in Oil Palm Mesocarp Tissue	87
		4.8.5	Effect of 2-Bromooctanoate on the Distribution of $[U^{-14}C]$	
			Glycerol into Lipid Classes in Oil Palm Mesocarp Tissue	87
	4.9	Applica	ation of Metabolic Control Analysis (MCA) on the Lipid	
		Biosyn	thesis Pathway in Oil Palm Mesocarp Tissue	91
	4.10	Acety	I-CoA Carboxylase (ACCase)	97
		4.10.1	Sodium Dodecyl Sulphate – Polyacrylamide Gel	07
		4 1 0 2	Electrophoresis (SDS-PAGE)	97
		4.10.2	Dulining with Streptavidin	99
	4 1 1	4.10.3	Preliminary ACCase Gene Isolation	102
	4.11	Future	e Studies	104
5	CO	NCLUS	ION	105
5		CLU		100
REFE	REN	ICES		107
APPE	NDI	X 1		117
VITA	E			119



# LIST OF TABLES

Table		Page
2.1	Lipid classification (King, 1996).	9
4.1	The percentage (%) incorporation of radioactivity into total lipids in the plastid without and with cofactors, ACP, ATP, NADH and NADPH	59
4.2	The percentage (%) incorporation of $[1-^{14}C]$ acetate into total lipids in the plastid, liquid culture and mesocarp tissue system.	60
4.3	The percentage (%) distribution of $[1-^{14}C]$ acetate into lipid classes in the plastid, liquid culture and mesocarp tissue system.	63
4.4	The percentage (%) distribution of $[1-^{14}C]$ acetate into lipid classes in 12-WAA, 16-WAA and 20-WAA oil palm mesocarp tissue.	64
4.5	The percentage (%) incorporation of radioactivity into total lipids in three different modes of radiolabel uptake into 20-WAA oil palm fruits.	67
4.6	The percentage (%) distribution of $[1-^{14}C]$ acetate into lipid classes in three different modes of radiolabel uptake into 20-WAA oil palm fruits.	68
4.7	The calculation of elasticity coefficients and flux control coefficients of the lipid biosynthesis pathway of the oil palm mesocarp tissue affected by temperature perturbation (see Appendix 1).	94



## LIST OF FIGURES

Figure		Page
2.1	Oil palm fruit.	5
2.2	Fatty acid biosynthesis pathway in plants.	12
2.3	The Kennedy pathway in plants.	19
2.4	Top-down metabolic control analysis (TDCA).	25
2.5	Bottom-up metabolic control analysis (BUCA).	25
3.1	A 25-ml conical flask containing the $CO_2$ trap and the radioactive incubation mixture.	33
3.2	Assembly of the Western blotting unit.	46
4.1	The separation of radiolabelled lipids by TLC. 1: Radiolabelled lipids extracted from mesocarp tissue. 2: Radiolabelled lipids extracted from plastid. 3: Radiolabelled lipids extracted from liquid cultures. 4: Lipid standards (Sigma).	62
4.2	Optimisation of pH for the incorporation of radioactivity.	70
4.3	The percentage (%) incorporation of radioactivity measured at one-hour intervals.	72
4.4	The percentage (%) incorporation of radioactivity for various concentrations of MES-NaOH buffer.	74
4.5	The percentage (%) incorporation of radioactivity into total lipids at various temperatures.	75
4.6	The percentage (%) incorporation of radioactivity into lipid classes at various temperatures.	77
4.7	The separation of the acyl-ACPs and acyl-CoAs through the SEP-PAK $C_{18}$ column (Waters).	80
4.8	Effect of various temperatures on the incorporation of radioactivity into acyl-ACPs in 20-WAA oil palm mesocarp tissue.	81
4.9	Effect of various temperatures on the incorporation of radioactivity into acvl-CoAs in 20-WAA oil palm mesocarp tissue.	81



4.10	Effect of various concentrations of 2-bromooctanoate on the incorporation of $[1-^{14}C]$ acetate into total lipids in 20-WAA oil palm mesocarp tissue.	84
4.11	Effect of various concentrations of 2-bromooctanoate on the distribution of $[1-^{14}C]$ acetate into lipid classes in 20-WAA oil palm mesocarp tissue.	86
4.12	Effect of various concentrations of 2-bromooctanoate on the incorporation of radioactivity into acyl-ACPs in 20-WAA oil palm mesocarp tissue.	88
4.13	Effect of various concentrations of 2-bromooctanoate on the incorporation of radioactivity into acyl-CoAs in 20-WAA oil palm mesocarp tissue.	88
4.14	Effect of various concentrations of 2-bromooctanoate on the incorporation of $[U^{-14}C]$ glycerol into total lipids in 20-WAA oil palm mesocarp tissue.	89
4.15	Effect of various concentrations of 2-bromooctanoate on distribution of [U- <sup>14</sup> C] glycerol into lipid classes in 20-WAA oil palm mesocarp tissue.	90
4.16	Top-down approach of MCA on lipid biosynthesis pathway.	93
4.17	(a) SDS-PAGE using standard gel to separate the crude proteins extracted from 15-WAA mesocarp. The standard gel consisted of a separating gel (12%) and a stacking gel (4%). Well 1: crude proteins extracted from 15-WAA mesocarp, and well 2: high range protein molecular weight markers (Bio-Rad). (b) Graph of log <sub>10</sub> molecular weight against distance of migration.	98
4.18	SDS-PAGE using gradient gel to separate the crude proteins extracted from 15-WAA mesocarp. The gradient gel consisted of a separating gel (5-20%) and a stacking gel (4%). Well 1 and 2: crude proteins extracted from 15-WAA mesocarp, and well 3: high range biotin-labelled calibration proteins (Boehringer Mannheim).	98
4.19	Immunoblotting of separated proteins with streptavidin. The highlighted bands indicate the biotin-containing proteins. These proteins were detected using chemiluminescence blotting kit (Boehringer Mannheim). Well 1: high range biotin-labelled calibration proteins (Boehringer Mannheim), and well 2: crude proteins extracted from 15-WAA	

mesocarp.

101



- Gel electrophoresis of restriction enzyme digestion products on 1.2% 4.20 agarose. Well 1: marker 100 bp DNA ladder, well 2: uncut plasmid DNA, well 3: plasmid DNA digested with Nco I, well 4: uncut plasmid DNA, well 5: plasmid DNA digested with Hind III and Bg II, well 6: uncut plasmid DNA, well 7: plasmid DNA digested with Hind III and Bg II, and well 8: marker 1 kbp DNA ladder.
- 103
- Screening of ACCase gene in 15-WAA mesocarp cDNA library. 4.21 Several strong signals corresponding to putative  $\beta$ -ct subunit of ACCase were obtained.

103



# LIST OF ABBREVIATIONS

.

ACCase	Acetyl-CoA carboxylase
ACP	Acyl carrier protein
ADP	Adenosine biphosphate
AMP	Adenosine monophosphate
ANOVA	Analysis of variance
ATP	Adenosine triphosphate
BC	Biotin carboxylase
BCCP	Biotin carboxyl carrier protein
BCS	Biodegradable counting scintillant
BSA	Bovine serum albumin
BUCA	Bottom-up metabolic control analysis
C12:0	Lauric acid
C16:0	Palmitic acid
C18:0	Stearic acid
C18:1	Oleic acid
C18:2	Linoleic acid
C18:3	Linolenic acid
C16:0-ACP	Palmitoyl-ACP
C18:0-ACP	Stearoyl-ACP
C18:1-ACP	Oleoyl-ACP
C <sub>16</sub>	16 carbons
C <sub>18</sub>	18 carbons
Ci	Curie
CoA	Coenzyme A
CO <sub>2</sub>	Carbon dioxide
cpm	Counts per min
CPO	Crude palm oil
СТ	Carboxyl transferase
DAG	Diacylglycerol
DAGAT	Diacylglycerol acyltransferase



DNA	Deoxyribonucleic acid
EDTA	Ethylenediaminetetraacetic acid
FAS	Fatty acid synthase
FFA	Free fatty acid
HCl	Hydrochloric acid
HCO <sub>3</sub> <sup>-</sup>	Ion bicarbonate
HEPES	N-[2-Hydroxylethyl]piperazine-N'-2-ethanesulphonic acid
$H_2SO_4$	Sulphuric acid
IPTG	Isopropylthio-β-D-galactoside
KAS	β-Ketoacyl-ACP synthetase
KCl	Potassium chloride
kDa	Kilo Dalton
KHCO <sub>3</sub>	Potassium bicarbonate
КОН	Potassium hydroxide
LB	Luria-Bertani
LPC	Lysophosphatidylcholine
MAG	Monoacylglycerol
MgCl <sub>2</sub>	Magnesium chloride
MgSO <sub>4</sub>	Magnesium sulphate
$MnCl_2$	Manganous chloride
MCA	Metabolic control analysis
MES	2-[N-morpholino]ethanesulphonic acid
NaCl	Sodium chloride
NADH	Nicotinamide adenine dinucleotide reduced form
NADPH	Nicotinamide adenine dinucleotide phosphate reduced form
$NaH^{14}CO_3$	Sodium [ <sup>14</sup> C] bicarbonate
NaOH	Sodium hydroxide
PAGE	Polyacrylamide gel electrophoresis
pfu	Plaque forming unit
pН	Hydrogen potential
РКО	Palm kemel oil



PL	Phospholipid
POD	Peroxidase
$\mathbf{P}_{i}$	Inorganic phosphate
$\mathbf{PP}_{i}$	Pyrophosphate
ppm	Part per million
PVDF	Polyvinylidene difluoride
SD	Standard deviation
SDS	Sodium dodecyl sulphate
sn	Stereospecific number
SOC	Sodium citrate
SPSS	Statistical package for social sciences
SSC	Sodium-citrate saline
TAG	Triacylglycerol
TBS	Tris-buffered saline
TBST	TBS-Tween 20
TCA	Tricarboxylic acid
TDCA	Top-down metabolic control analysis
TLC	Thin layer chromatography
V	Volt
var.	Variety
v/v	Volume/volume
WAA	Weeks after anthesis
w/v	Weight/volume
X-gal	5-Bromo-4-chloro-3-indolyl-β-D-galactoside



### CHAPTER 1

### **INTRODUCTION**

Recent advances in biotechnology, such as in genomics, proteomics, DNA microarray and bioinformatics, have enabled plants to be modified to produce novel products. These novel products include biodegradable plastics (Poirier *et al.*, 1992), antibodies (Hiatt *et al.*, 1989; Ma and Hein, 1995) and interferon (De Zoeten *et al.*, 1989).

Palm oil has become an important edible oil over the last few decades with a production of about 18.2 million tonnes in 1996-2000 from 3.7 million tonnes in 1976-1980 when it supplied a mere 7.1% of the world's oils and fats (Basiron, 2000). It now contributes 23% to the world's oils and fats production, making it the second-most produced oil after soybean oil (Gunstone, 2001). The oil palm is the highest-yielding oil crop in the world, surpassing the coconut (the next highest-yielding oil crop) by about 50-100% and other oil crops by even more (Basiron, 2000).

Malaysia is the largest producer of palm oil with 60% of the world production (Chow, 1997). Palm oil is predicted to become the major oil in the world by 2012 (Oil World, 1999), but the stiff competition from other oils has made it necessary to diversify its use. In addition, novel higher value-added products can be made producible by the oil palm by employing biotechnology methods such as recombinant DNA technology and genomics (Cheah, 1999). However, this requires a detailed understanding of the basic plant metabolism.



Plant lipid biosynthesis has been much studied in recent years (Browse and Somerville, 1991; Harwood, 1999; Ohlrogge and Jaworski, 1997; Slabas and Fawcett, 1992) but there remains much more to be learnt. To manipulate the oil palm for novel oils such as high oleate (Cheah, 1997) would require substantially more information on the regulation of lipid biosynthesis.

As metabolic pathways are under multi-step control, unraveling a single step or an individual enzyme is insufficient to understand the entire metabolism. Indeed, the converse is needed – to have an overall picture of the metabolic pathway before the particular steps can be understood. This can be achieved by applying the metabolic control analysis (MCA).

In most plants, including the oil palm, the storage lipids are mainly triacylglycerols (TAGs) (Harwood, 1980; Murphy, 1993). The metabolic pathways to TAG involve acetyl-CoA as the immediate carbon source, and information on the metabolic flux will be useful in modeling the carbon flux through pathways. This work was therefore to investigate the carbon flux in lipid biosynthesis by the oil palm using a radiolabel. It is hoped that the information gained may be useful in diverting the carbon to the formation of higher-value products by genetic manipulation.

To increase the production of a metabolite(s), it is necessary to manipulate a reaction, or a set of reactions, over another. However, the manipulation may still not result in production of the metabolite(s) if the necessary control mechanisms are not in place. Therefore, a comprehensive understanding of the entire cellular metabolism is needed.



Currently, attempts are being made to design cellular metabolism in order to maximize output of the desired metabolites. But the requisite prelude to this is quantification of the metabolite flux by MCA.

The objectives of this research were to:

- 1) Investigate carbon flux through the oil palm lipid biosynthesis pathway(s).
- 2) Apply MCA for a better understanding of the overall quantitative control structure of the lipid biosynthesis pathway(s).

#### CHAPTER 2

#### LITERATURE REVIEW

#### 2.1 Oil Palm

Oil palm is a perennial oil-bearing crop that has an economic life of about 25 years (Gascon *et al.*, 1989; Basiron, 2001). It begins to bear fruit two to three years after planting (Basiron, 2001). It is a unique crop that yields two types of oil, crude palm oil (CPO) from the mesocarp of the fruit and palm kernel oil (PKO) from the seed or kernel. These two oils have different physical and chemical properties. CPO contains mainly palmitic acid (C16:0) and oleic acid (C18:1), the two most common fatty acids in nature while PKO contains mainly lauric acid (C12:0).

The oil palm fruit is a sessible drupe which is usually oval in shape. The length of the fruit is 2.5 to 5.0 cm and 2.5 cm in diameter. It may weigh from 3 to 30 g (Godin and Spensley, 1971; Gascon *et al.*, 1989).

The oil palm fruit consists of the mesocarp, the shell or endocarp and the kernel as shown in Figure 2.1. The mesocarp or pulp of the ripe oil palm fruit is yellowishorange in colour. It is oily and fibrous (Vaughan, 1970). The outer layer of oil palm is called exocarp or epicarp. It shows variation in colour through yellow, orange, red, brown and black according to the variety (Cobley and Steele, 1976).

