



**EVALUATION OF POTENTIAL PROBIOTIC *Bacillus velezensis* FS26
AGAINST AQUACULTURE PATHOGENS AND ITS ABILITY TO UTILISE
PREBIOTICS FROM *Arthrospira platensis* A1**

By

MUHAMAD FIRDAUS SYAHMI BIN SAM-ON

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of
Philosophy**

October 2023

FBSB 2023 2

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

**EVALUATION OF POTENTIAL PROBIOTIC *Bacillus velezensis* FS26
AGAINST AQUACULTURE PATHOGENS AND ITS ABILITY TO UTILISE
PREBIOTICS FROM *Arthospira platensis* A1**

By

MUHAMAD FIRDAUS SYAHMI BIN SAM-ON

October 2023

Chair : Professor Shuhaimi bin Mustafa, PhD
Faculty : Biotechnology and Biomolecular Sciences

Pathogenic bacteria such as *Vibrio* spp. and *Aeromonas* spp. frequently cause detrimental effects on aquaculture production. Antibiotics were previously utilised to treat infections, but this led to antibiotic-resistant bacteria emerging in the environment. Probiotics and prebiotics were introduced to boost the host's microbiota, disease protection, health condition, growth efficiency, feed consumption, stress response and overall vigour. Synbiotic is another approach that should be evaluated intensively to solve the infections problems. However, probiotic bacteria need to show the ability to utilise prebiotic for synbiotic preparation. Therefore, this study aims to evaluate the potential probiotic of selected bacteria against aquaculture pathogens and capability to use prebiotic from *Arthospira platensis* A1. *Bacillus velezensis* FS26 (Genome accession number: JAOPEO000000000; 16S rRNA accession number: MZ960133) was isolated from giant freshwater prawn and showed good probiotic properties against pathogens in aquaculture through *in vitro* and *in silico* studies. Agar well diffusion assay exhibited the capability of this bacterium through diameter of inhibition against the pathogenic *Aeromonas hydrophila* LMG13658, *A. veronii* clone DK-A.veronii-27, *Vibrio campbellii* PKGL21, *V. alginolyticus* PKS15, *V. parahaemolyticus* PKK24 at 23.7, 25, 30, 13.3 and 12.3 mm, respectively. This bacterium is regarded as harmless based on susceptibility towards 13 antibiotics, γ -haemolytic activity in blood agar and *in silico* prediction for the lack of 96 antibiotic resistance genes, pathogenic genes and virulence factors. Additionally, *B. velezensis* FS26 has shown the capability to utilise commercial prebiotics such as lactulose, raffinose and inulin through *in silico* and *in vitro* investigation. The bacterial genome predicted six enzymes that could hydrolyse glycosidic bonds in prebiotics, such as α -galactosidase, invertase, endolevanase, β -2,6-fructan-6-levanbiohydrolase and levansucrase. Moreover, using prebiotics from aquaculture sources such as microalgae could enhance the palatability of the synbiotic. Hence, water-soluble nondigestible polysaccharide was extracted from *Arthospira platensis* A1 as a prebiotic for *B. velezensis*

FS26. The structure of water-soluble polysaccharides demonstrated the presence of α -glycosidic bonds at wavelength 860 nm and water-soluble compounds at wavelength between 300 to 400 nm through FTIR and UV-VIS spectroscopy analysis. In the outcome, water-soluble nondigestible polysaccharide extracted using autoclave exhibited a promising prebiotic activity towards potential probiotic *B. velezensis* FS26 by significantly enhance the bacterial growth on enriched and minimal media at 0.6×10^9 CFU/mL and 7.1×10^9 CFU/mL, compared to control. In conclusion, the findings demonstrate the capability of probiotic *B. velezensis* FS26 to inhibit the aquaculture pathogens and utilise prebiotic from *A. platensis* A1.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENILAIAN POTENSI PROBIOTIK *Bacillus velezensis* FS26 TERHADAP
PATOGEN AKUAKULTUR DAN KEUPAYAANNYA UNTUK
MEMANFAATKAN PREBIOTIK DARI *Arthospira platensis* A1**

Oleh

MUHAMAD FIRDAUS SYAHMI BIN SAM-ON

Okttober 2023

Pengerusi : Profesor Shuhaimi bin Mustafa, PhD
Fakulti : Bioteknologi dan Sains Biomolekul

Bakteria patogen seperti *Vibrio* spp. dan *Aeromonas* spp. sering menyebabkan kesan buruk terhadap pengeluaran akuakultur. Antibiotik sebelum ini digunakan untuk merawat jangkitan, tetapi ini menyebabkan bakteria yang tahan terhadap antibiotik muncul dalam alam sekitar. Probiotik dan prebiotik diperkenalkan untuk meningkatkan mikrobiota hos, perlindungan terhadap penyakit, keadaan kesihatan, kecekapan pertumbuhan, pengambilan makanan, respons tekanan, dan kesegaran keseluruhan. Pendekatan Synbiotic adalah satu lagi pendekatan yang perlu dievaluasi secara intensif untuk menyelesaikan masalah jangkitan. Walau bagaimanapun, bakteria probiotik perlu menunjukkan keupayaan untuk menggunakan prebiotik dalam persediaan synbiotic. Oleh itu, kajian ini bertujuan untuk menilai potensi probiotik bakteria terpilih terhadap patogen akuakultur dan keupayaannya menggunakan prebiotik dari *Arthospira platensis* A1. *Bacillus velezensis* FS26 (Nombor akses genom: JAOP000000000; Nombor akses 16S rRNA: MZ960133) diasingkan dari udang air tawar raksasa dan menunjukkan sifat probiotik yang baik terhadap patogen dalam akuakultur melalui kajian *in vitro* dan *in silico*. Ujian penyebaran sumuran agar menunjukkan keupayaan bakteria ini melalui diameter perintangan terhadap *Aeromonas hydrophila* patogen LMG13658, klon A. veronii DK-A.veronii-27, *Vibrio campbellii* PKGL21, *V. alginolyticus* PKS15, *V. parahaemolyticus* PKK24 pada 23.7, 25, 30, 13.3, dan 12.3 mm, masing-masing. Bakteria ini dianggap tidak berbahaya berdasarkan kerentanannya terhadap 13 antibiotik, aktiviti γ -hemolitik dalam agar darah, dan ramalan *in silico* untuk ketidaaan 96 gen rintangan antibiotik, gen patogen, dan faktor virulensi. Tambahan pula, *B. velezensis* FS26 telah menunjukkan keupayaan untuk menggunakan prebiotik komersial seperti laktulosa, rafinosa, dan inulin melalui penyelidikan *in silico* dan *in vitro*. Genom bakteria meramalkan beberapa enzim yang boleh menghidrolisis ikatan glikosida dalam prebiotik, seperti α -galaktosidase, invertase, endolevanase, β -2,6-fructan-6-levanbiohydrolase, dan levansucrase. Selain itu,

menggunakan prebiotik dari sumber akuakultur seperti mikroalga dapat meningkatkan kesedapan sinbiotik. Oleh itu, polisakarida tak larut dalam air telah diekstrak dari *Arthrospira platensis* A1 sebagai prebiotik untuk *B. velezensis* FS26. Struktur polisakarida tak larut dalam air menunjukkan kehadiran ikatan α -glikosida pada panjang gelombang 860 nm dan sebatian larut dalam air pada panjang gelombang antara 300 hingga 400 nm melalui analisis spektroskopi FTIR dan UV-VIS. Hasilnya, polisakarida tak larut dalam air yang diekstrak menggunakan autoklaf menunjukkan aktiviti prebiotik yang menjanjikan terhadap pertumbuhan bakteria *B. velezensis* FS26 yang berpotensi pada media yang diperkaya dan minimal pada 0.6×10^9 CFU/mL dan 7.1×10^9 CFU/mL, berbanding dengan kawalan. Penemuan ini menunjukkan keupayaan probiotik *B. velezensis* FS26 untuk menghalang patogen akuakultur dan menggunakan prebiotik dari *A. platensis* A1.

ACKNOWLEDGEMENTS

Foremost, I want to express my sincerest gratitude to Prof Dr Shuhaimi Mustafa, my supervisor, for his constant support, guidance, and encouragement throughout my PhD journey. His mentorship, insights, and expertise have been invaluable in navigating the complexities of my research. I am deeply appreciative of my supervisory committee members, Associate Prof Dr Termizi Yusof and Dr Shahrizim Zulkifly, for providing insightful feedback and valuable input that have helped me to shape and refine my research work. I extend my gratitude to Dr Amalia Mohd Hashim for her assistance in reviewing my manuscripts and providing constructive feedback, which has significantly improved the quality of my work.

My parents, Sam-on bin Yaacob and Norfairuzzah binti Johan, have been a constant source of unwavering support, love, and encouragement throughout my academic journey. Their sacrifices and dedication have been a significant motivation for me. My siblings, Along, Abang Fahem, Iqah, Azhar, Qirah, Syasya, and Faiq, have been pillars of support, encouragement, and understanding through the ups and downs of my PhD journey. The love and joy brought by my cute nephews, Uwais, Hayyan, and Uzair, have been a constant source of happiness and inspiration.

To my dearest Ku Nur Azwa Ku Aizuddin, words cannot express the depth of my gratitude for your unwavering love, support, and encouragement throughout my PhD journey. Your belief in me has been the brightest light on even the darkest of days, and your presence in my life has been a constant source of motivation and inspiration. I am forever grateful for your unwavering commitment to our relationship and for being my rock during this challenging time. Thank you for being my partner, my best friend, and my soulmate.

I am deeply grateful to my friend and colleague at FamTech Laboratory for their unwavering support and encouragement during my PhD journey. Their kindness, expertise, and unwavering dedication have been a tremendous asset to my research, and their friendship has been a source of comfort and inspiration throughout the challenges of the past three years. I am truly blessed to have such a remarkable colleague and friend, and I will always cherish the memories we have made together. Thank you for being an essential part of my journey.

Lastly, I express my sincere appreciation to all who have contributed to my PhD journey, including my professors, colleagues, friends, and family members. Without their support and encouragement, this achievement would not have been possible. Thank you all !!!

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Shuhaimi bin Mustafa, PhD

Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Mohd Termizi bin Yusof, PhD

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

Shahrizim bin Zulkifly, PhD

Senior Lecturer

Faculty of Science

Universiti Putra Malaysia

(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 14 December 2023

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xviii
 CHAPTER	
1 INTRODUCTION	1
1.1 Background	1
1.2 Problem statement	3
1.3 Hypothesis	3
1.4 Objective	3
2 LITERATURE REVIEW	5
2.1 Global Aquaculture Production	5
2.2 Aquaculture in Malaysia	5
2.3 Impact of the aquaculture industry	6
2.4 Disease in aquaculture	6
2.5 Diseases management approaches for aquaculture	9
2.5.1 Vaccines	9
2.5.2 Bacteriophages	10
2.5.3 Antibiotics	10
2.6 Propose disease management approaches in aquaculture	11
2.6.1 Probiotics	12
2.6.2 Prebiotics	18
2.6.3 Synbiotics	22
2.6.4 Postbiotics	25
3 ISOLATION AND EVALUATION OF POTENTIAL PROBIOTICS OF SELECTED BACTERIA FROM THE GUT OF GIANT FRESHWATER PRAWN	28
3.1 Introduction	28
3.2 Materials and Methods	29
3.2.1 Materials	29
3.2.2 Isolation of bacteria from the gut of giant freshwater prawn	31
3.2.3 Identification of bacterial isolates	31
3.2.4 Antagonism assay of bacterial isolates	33

3.2.5	Cell Adhesion test of bacterial isolates	33
3.2.6	pH and bile salt tolerance tests of bacterial isolates	35
3.2.7	Safety assessment of bacterial isolates	35
3.2.8	Statistical analysis	36
3.3	Results and Discussion	36
3.3.1	Isolation of bacteria from the gut of giant freshwater prawn	36
3.3.2	Identification of the bacterial isolates	37
3.3.3	Antimicrobial activity of bacterial isolates against aquaculture pathogens	42
3.3.4	Cell Adhesion test of bacterial isolates	46
3.3.5	pH and bile tolerance test of bacterial isolates	50
3.3.6	Safety assessment of bacterial isolates	55
3.4	Conclusion	57
4	IN SILICO PREDICTIONS OF PROBIOTIC MARKERS AND SECONDARY METABOLITES OF SELECTED BACTERIA AGAINST AQUACULTURE PATHOGENS	58
4.1	Introduction	58
4.2	Materials and Methods	59
4.2.1	Materials	59
4.2.2	Genomic DNA extraction and sequencing of <i>Bacillus velezensis</i> FS26	59
4.2.3	<i>Bacillus velezensis</i> FS26 genome annotation	60
4.2.4	Average nucleotide identity of <i>Bacillus velezensis</i> FS26 genome	60
4.2.5	Prediction of secondary metabolites in the <i>Bacillus velezensis</i> FS26 genome	61
4.2.6	Prediction of probiotic marker genes in the <i>Bacillus velezensis</i> FS26 genome	61
4.2.7	Safety analysis of <i>Bacillus velezensis</i> FS26 genome	61
4.3	Results and Discussion	62
4.3.1	Comprehensive genome analysis of <i>Bacillus velezensis</i> FS26	62
4.3.2	Average nucleotide identity of <i>Bacillus velezensis</i> FS26 genome	63

4.3.3	Prediction of secondary metabolites in the <i>Bacillus velezensis</i> FS26 genome	65
4.3.4	Prediction of probiotic marker genes in the <i>Bacillus velezensis</i> FS26 genome	70
4.3.5	Safety analysis of <i>Bacillus velezensis</i> FS26 genome	72
4.4	Conclusion	74
5	DETERMINATION OF PREBIOTIC UTILISATION CAPABILITY OF POTENTIAL PROBIOTIC OF SELECTED BACTERIA THROUGH <i>IN SILICO</i> AND <i>IN VITRO</i> APPROACHES	75
5.1	Introduction	75
5.2	Materials and Methods	77
5.2.1	Materials	77
5.2.2	Analysis of carbohydrate-active enzymes from <i>Bacillus velezensis</i> FS26 genome	77
5.2.3	Evaluation of prebiotics utilising capability of <i>Bacillus velezensis</i> FS26	77
5.2.4	Growth Analysis of <i>Bacillus velezensis</i> FS26	78
5.2.5	Statistical Analysis	78
5.3	Results and Discussion	78
5.3.1	<i>In silico</i> analysis of carbohydrate-active enzyme (CAZy) in <i>Bacillus velezensis</i> FS26 genome	78
5.3.2	Evaluation of the prebiotic utilisation and growth capability of <i>Bacillus velezensis</i> FS26 using <i>in vitro</i> approaches	83
5.4	Conclusion	91
6	PREBIOTIC ACTIVITY OF WATER-SOLUBLE NONDIGESTIBLE POLYSACCHARIDE EXTRACTED FROM MICROALGAE <i>ARTHROSPIRA PLATENSIS</i> A1 TOWARDS SELECTED BACTERIA	92
6.1	Introduction	92
6.2	Materials and Methods	93
6.2.1	Materials	93
6.2.2	Identification of <i>Arthrosipa platensis</i> A1	93
6.2.3	Extraction of water-soluble polysaccharide	94

6.2.4	Digestibility of water-soluble polysaccharide	95
6.2.5	FTIR and UV-VIS spectra of water-soluble polysaccharide	95
6.2.6	Prebiotic activity water-soluble polysaccharide towards <i>Bacillus velezensis</i> FS26	96
6.2.7	Statistical Analysis	96
6.3	Results and Discussion	97
6.3.1	Identification of <i>Arthospira platensis</i> A1	97
6.3.2	Extraction of water-soluble polysaccharide	99
6.3.3	Digestibility of water-soluble polysaccharide	101
6.3.4	FTIR and UV-VIS spectra of water-soluble polysaccharide	102
6.3.5	Prebiotic activity water-soluble polysaccharide towards <i>Bacillus velezensis</i> FS26	105
6.4	Conclusion	113
7	CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	114
7.1	Conclusion	114
7.2	Recommendation for future research	115
REFERENCES		116
APPENDICES		163
BIODATA OF STUDENT		173
LIST OF PUBLICATIONS		174

LIST OF TABLES

Table		Page
2.1	The probiotics list and their effect towards aquaculture	15
2.2	The prebiotics list and their effect towards aquaculture	21
2.3	The synbiotics list and their effect towards aquaculture	24
2.4	The postbiotics list and their effect towards aquaculture	27
3.1	List of aquaculture pathogens	30
3.2	List of antibiotics tested for bacterial isolates	30
3.3	Microscopic, morphology and biochemical analysis of bacterial isolates	38
3.4	Molecular identification of bacterial isolates	41
3.5	Agar well diffusion assay of bacterial isolates against aquaculture pathogens	45
3.6	Percentage of bile salt tolerance (%) for bacterial isolates	55
3.7	Safety assessment of bacterial isolates	56
4.1	<i>Bacillus velezensis</i> genomes from NCBI database	60
4.2	Genome assembly and annotation of <i>Bacillus velezensis</i> FS26	63
4.3	List of the putative gene clusters encoding for secondary metabolites by antiSMASH analysis in <i>Bacillus velezensis</i> FS26 genome	65
4.4	Probiotic marker of <i>Bacillus velezensis</i> FS26 using Prokka analysis	71
4.5	Safety analysis of <i>Bacillus velezensis</i> FS26 genome	73

5.1	Enzymes predicted for cleavage the α -glycosidic bonds in carbohydrate from the <i>Bacillus velezensis</i> FS26 genome	80
5.2	Enzymes predicted for cleavage the β -glycosidic bonds in carbohydrate from the <i>Bacillus velezensis</i> FS26 genome	82
6.1	Extraction analysis of water-soluble polysaccharides from <i>Arthrosphaera platensis</i> A1	100

LIST OF FIGURES

Figure		Page
3.1	Phylogenetic tree of bacterial isolates	39
3.2	Agar-well diffusion assay for bacterial isolates against <i>Aeromonas hydrophila</i> LMG13658	43
3.3	Auto-aggregation (%) of bacterial isolates	46
3.4	Co-aggregation (%) of bacterial isolates	47
3.5	Cell surface hydrophobicity (%) of bacterial isolates	49
3.6	pH tolerance test of bacterial isolates	50
4.1	A circular graphical display of the distribution of the genome annotations for <i>Bacillus velezensis</i> FS26 with subsystems and gene frequencies	62
4.2	Heatmap of average nucleotide identity (ANI) between 11 <i>Bacillus velezensis</i> genomes and <i>Bacillus velezensis</i> FS26 genome	64
4.3	Classification of antimicrobial peptides predicted by antiSMASH analysis on <i>Bacillus velezensis</i> FS26 genome	66
4.4	Bacilysin biosynthesis of <i>Bacillus velezensis</i> FS26 genome	68
4.5	Heatmap of comparison of secondary metabolites between 11 <i>Bacillus velezensis</i> genomes and <i>Bacillus velezensis</i> FS26	69
5.1	Prediction of carbohydrate-active enzyme (CAZy) families in <i>Bacillus velezensis</i> FS26 genome	79
5.2	Colour spectra of prebiotic utilisation assay of <i>Bacillus velezensis</i> FS26	85
5.3	pH reading of prebiotic utilisation assay of <i>Bacillus velezensis</i> FS26	88

5.4	Growth analysis of <i>Bacillus velezensis</i> FS26	90
6.1	Microscopic observation of the <i>Arthrosphaera platensis</i> A1	97
6.2	Phylogenetic tree of microalgae <i>Arthrosphaera platensis</i> A1	98
6.3	Extraction of water-soluble polysaccharide from <i>Arthrosphaera platensis</i> A1 using autoclave and hot water	100
6.4	Degree of hydrolysis for crude polysaccharide extracted from <i>Arthrosphaera platensis</i> A1 using artificial human gastric juice	101
6.5	FTIR analysis of crude polysaccharide extracted from <i>Arthrosphaera platensis</i> A1	103
6.6	UV-VIS spectroscopy analysis of crude polysaccharide extracted from <i>Arthrosphaera platensis</i> A1	104
6.7	Colour spectra of crude polysaccharide extracted from <i>Arthrosphaera platensis</i> A1 utilisation assay	107
6.8	pH reading of crude polysaccharide extracted from <i>Arthrosphaera platensis</i> A1 utilisation assay	109
6.9	Prebiotic activity of crude polysaccharide extracted from <i>Arthrosphaera platensis</i> A1 on <i>Bacillus velezensis</i> FS26	112

LIST OF ABBREVIATIONS

kg	Kilogram
k	Kilo
g	Gram
bp	Base pair
min	Minute
s	Second
h	Hour
d	Day
°C	Degree Celsius
sp.	Species (one)
spp.	Species (two or more)
rpm	Rotation per minute
DNA	Deoxyribonucleic acid
RNA	Ribonucleic acid
PCR	Polymerase chain reaction
CFU	Colony forming unit
OD	Optical density
L	Litter
µL	Microliter
mL	Milliliter
mg	Milligram
USD	United States dollar
UK	United Kingdom
FAO	The Food and Agriculture Organization of the United Nations

%	Percent
mg/mL	Milligram per millilitre
SD	Standard deviation
nm	Nanometer
CAZy	Carbohydrate-active enzymes
antiSMASH	The Antibiotics and Secondary Metabolite Analysis Shell
ANI	Average nucleotide identity
V	Voltage
α	Alpha
β	Beta
γ	Gamma
\pm	Plus-minus sign
<	Less than
>	More than

CHAPTER 1

INTRODUCTION

1.1 Background

Aquaculture is a critical sector that must be retained for stabilising global food security. This industry has contributed over 179 million metric tonnes of fish production, valued at roughly USD 250 billion. FAO (2022) reported that nearly 156 million metric tonnes of fish are consumed by humans, providing 20.5 kg of aquaculture products per person. Moreover, FAO (2016) and FAO (2018) reported that Malaysia increased aquaculture production by ranking eleventh globally in 2016 as opposed to the fifteenth in 2014. Furthermore, Malaysia made USD 714.1 million in 2017 by exporting significant quantities of prawns and sashimi tuna, according to FAO (2019). Among several types of the aquaculture industry, prawn farming has shown significant growth and new technology has contributed to making production more efficient (El-Sayed et al., 2021; Emerenciano et al., 2022). Nonetheless, the industry frequently faces significant obstacles, such as pathogenic infection, which hinder productivity.

Studies by Zorriehzahra et al. (2016) and Nakayama et al. (2009) stated that pathogenic *Vibrio* is a bacterial genus that caused a massive loss in aquaculture. Tan et al. (2017) reported that *Vibrio parahaemolyticus* affects various types of seafood, especially shellfish. The worst impact of this bacterium is transferable to humans, causing gastrointestinal tract infections (Tan et al., 2017). According to Kavitha et al. (2018), *Aeromonas* spp., *Pseudomonas* spp., *Acinetobacter* spp., and *Streptococcus* spp. are also regarded as pathogens in aquaculture. Study has shown the use of antibiotics, vaccination and bacteriophage as disease management in aquaculture. Even though antibiotics have broad-spectrum activity against pathogens, the emergence of bacteria that are resistant to antibiotics has impacted the environment and exacerbated the problem (Balcazar et al., 2006). Whereas vaccination and bacteriophage are often specific in combat with pathogens. Therefore, probiotics, prebiotics, synbiotics, and postbiotics are proposed as safe and broad activity toward pathogens.

Probiotics are "live microorganisms that, when administered in adequate amounts, confer a health benefit on the host" (Hill et al., 2014). Suresh (2023) and Biswas et al. (2023) stated that *Bacillus subtilis*, *B. coagulans*, and *B. licheniformis* are among the most studied *Bacillus* species as probiotics. *Bacillus subtilis*, according to Cutting (2011), is a probiotic that

can contribute to the host's health benefits. Applying this *Bacillus* species in shrimp cultures has shown promising effects in enhancing the activity of digestive enzymes such as lipase, protease, and amylase. This probiotic also enhances shrimp survivability, according to Buruiană et al. (2014). Similarly, Tank et al. (2018) reported that *Bacillus subtilis* isolated from white-leg shrimp could suppress the growth of pathogenic *Vibrio harveyi*, *V. parahaemolyticus* and *V. vulnificus* in shrimp cultures.

Prebiotics are "substrate that is selectively used by host microorganisms to confer a health benefit" (Gibson et al., 2017). Commercial prebiotic such as lactulose, raffinose and inulin have shown a positive effect towards the growth and disease resistance for aquaculture (Madreseh et al., 2019; Karimi et al., 2020; Ajdari et al., 2022). Moreover, a study by Ramadhan et al. (2019) reported that prebiotics is an effective supplement in aquaculture. Research on microalgae as a prebiotic and an enhancer for probiotic growth is ongoing. *Isochrysis galbana* is an example of marine microalgae that displays promising prebiotic properties due to its ability to boost the growth of lactic acid bacteria and its contents, which comprise highly soluble and insoluble fibre (De Jesus et al., 2016). Lee et al. (2003) reported that *Spirulina (Arthrospira) platensis* contains oligosaccharides as a bioactive compound that improves the prawn's immune system and disease resistance. Additionally, Tayag et al. (2010) demonstrated that *Arthrospira platensis* could produce C-phycocyanin, which improved immunity and disease endurance in the white-leg shrimp.

According to Gibson et al. (2017), synbiotics are "a mixture comprising live microorganisms and substrate(s) selectively utilised by host microorganisms that confers a health benefit on the host". The prebiotic metabolism by probiotics will generate different short-chain fatty acids, which regard as postbiotics, including acetate, lactate, propionate, butyrate, and valerate, which benefits the hosts' health and inhibit the pathogens' growth (Pan et al., 2009; Markowiak-Kope and Liewska, 2020; Holmes et al., 2022). Probiotic bacteria must be able to utilise prebiotics by hydrolysing the glycosidic linkages to create a synbiotic. Using genome sequences to predict glycosidase hydrolyse families with the dbcan2 program is a cutting-edge technology for synbiotic preparation (Zhang et al., 2018). However, due to the limitation of the study, *in vitro* validation is needed to verify the enzyme production.

Apart from synbiotics, probiotics can also produce a variety of postbiotics, including bacteriocin, polymyxin, antibiotics, and lantibiotics, which are efficient antimicrobial agents against infections, according to Chalasani et al. (2015). Postbiotics are the "preparation of inanimate microorganisms

and/or their components that confers a health benefit on the host" (Salminen et al., 2021). Nowadays, the postbiotic can be forecast by utilising the genome sequence of the microbes. The Antibiotics and Secondary Metabolite Analysis Shell (antiSMASH) pipeline has emerged as the first tool to identify biosynthetic loci for the complete range of known secondary metabolite compound classes. The antiSMASH website can predict a variety of secondary metabolites, known as postbiotics, in bacterial and fungal genomes, including polyketides, non-ribosomal peptides, terpenes, aminoglycosides, aminocoumarins, indolocarbazoles, lantibiotics, bacteriocins, nucleosides and beta-lactam (Medema et al., 2011).

1.2 Problem statement

Pathogenic bacteria cause infections in aquatic animals and reduce production in the aquaculture industry. Previously, antibiotics had been used to solve this infection problem, but antibiotic-resistant bacteria emerged. A single-used probiotic and prebiotic is another approach but is inadequate to solve the problems, unlike symbiotic. However, probiotic bacteria need to show the ability to utilise prebiotic for symbiotic preparation. Hence, the effectiveness of probiotic bacteria against aquaculture pathogens and ability to utilise prebiotic from microalgae is an approach that should be evaluated intensively to overcome aquaculture diseases.

1.3 Hypothesis

Selected bacterial isolates from the gut of the giant freshwater prawn exhibit potential probiotics. Then, the selected bacterial genome foreseen good probiotic markers for aquaculture together with secondary metabolites genes against aquaculture pathogens. The selected bacterial isolate can utilise prebiotics through *in silico* and *in vitro* approaches. Thus, Microalgae *Arthrospira platensis* A1 exhibit the potential prebiotic and symbiotic with selected bacterial isolates against aquaculture pathogens.

1.4 Objective

The main objective of this study is to evaluate the potential probiotic of selected bacterial isolates against aquaculture pathogens and capability to use prebiotic from *Arthrospira platensis* A1.

The specific objectives of this study are as follows:

1. to isolate and characterise the selected bacteria from the gut of giant freshwater prawns as potential probiotics.
2. to analyse the general genome characteristics and probiotic markers from the selected bacteria with secondary metabolites genes predicted against aquaculture pathogens.
3. to investigate the ability of the potential probiotic of selected bacteria to utilise prebiotics using *in silico* and *in vitro* techniques.
4. to examine the water-soluble nondigestible polysaccharide extracted from *Arthrosphaera platensis* A1 and its prebiotic effect towards the selected bacteria.

REFERENCES

- Ababouch, L., Nguyen, K. A. T., Castro de Souza, M., and Fernandez-Polanco, J. (2023). Value chains and market access for aquaculture products. *Journal of the World Aquaculture Society*, 54, 527-553.
- Abasubong, K. P., Li, X. F., Adjoumani, J. J. Y., Jiang, G. Z., Desouky, H. E., and Liu, W. B. (2022). Effects of dietary xylooligosaccharide prebiotic supplementation on growth, antioxidant and intestinal immune-related genes expression in common carp *Cyprinus carpio* fed a high-fat diet. *Journal of Animal Physiology and Animal Nutrition*, 106(2), 403-418.
- Abbasiliasi, S., Tan, J. S., Ibrahim, T. A. T., Ramanan, R. N., Vakhshiteh, F., Mustafa, S., Ling, T. C., Abdul Rahim, R., and Ariff, A. B. (2012). Isolation of *Pediococcus acidilactici* Kp10 with ability to secrete bacteriocin-like inhibitory substance from milk products for applications in food industry. *BMC Microbiology*, 12(1), 260. <https://doi.org/10.1186/1471-2180-12-260>.
- Abdel-Latif, H. M., El-Ashram, S., Sayed, A. E. D. H., Alagawany, M., Shukry, M., Dawood, M. A., and Kucharczyk, D. (2022). Elucidating the ameliorative effects of the cyanobacterium *Spirulina (Arthrospira platensis)* and several microalgal species against the negative impacts of contaminants in freshwater fish: a review. *Aquaculture*, 554, 738155. <https://doi.org/10.1016/j.aquaculture.2022.738155>.
- Abdolnabi, S., Ina-Salwany, M. Y., Daud, H. M., Mariana, S. D., and Abdelhadi, Y. M. (2015). Pathogenicity of *Aeromonas hydrophila* in giant freshwater prawn *Macrobrachium rosenbergii*, cultured in East Malaysia. *Iranian Journal of Fisheries Science*, 14(1), 232-245.
- Abraham, E. P., Callow, D., and Gilliver, K. (1946). Adaptation of *Staphylococcus aureus* to growth in the presence of certain antibiotics. *Nature*, 158(4023), 818-821.
- Abriouel, H., Pérez Montoro, B., Casimiro-Soriguer, C. S., Pérez Pulido, A. J., Knapp, C. W., Caballero Gómez, N., Castillo-Giterrez, S., Estudillo-Martinez, M. D., Galvez, A., and Benomar, N. (2017). Insight into potential probiotic markers predicted in *Lactobacillus pentosus* MP-10 genome sequence. *Frontiers in Microbiology*, 8, 891. <https://doi.org/10.3389/fmicb.2017.00891>.
- Abumourad, I. M. K., Abbas, W. T., Awaad, E. S., Authman, M. M. N., El-Shafei, K., Sharaf, O. M., Ibrahim, G. A., Sadek, Z. I., and El-Sayed, H. S. (2013). Evaluation of *Lactobacillus plantarum* as a probiotic in

- aquaculture: emphasis on growth performance and innate immunity. *Journal of Applied Sciences Research*, 9(1), 572-582.
- Adel, M., Nayak, S., Lazado, C. C., and Yeganeh, S. (2016). Effects of dietary prebiotic GroBiotic®-A on growth performance, plasma thyroid hormones and mucosal immunity of great sturgeon, *Huso huso* (Linnaeus, 1758). *Journal of Applied Ichthyology*, 32(5), 825-831.
- Agung, L.A., and Yuhana M. (2015). Application of micro-encapsulated probiotic *Bacillus* NP5 and prebiotic mannan oligosaccharide (MOS) to prevent streptococcosis on tilapia *Oreochromis niloticus*. *Research Journal of Microbiology*, 10(12), 571. <http://dx.doi.org/10.3923/jm.2015.571.581>.
- Aich, N., Ahmed, N., and Paul, A. (2018). Issues of antibiotic resistance in aquaculture industry and its way forward. *International Journal of Current Microbiology and Applied Sciences*, 7(8), 26-41.
- Ajdar, A., Ghafarifarsani, H., Hoseinifar, S. H., Javahery, S., Narimanizad, F., Gatphayak, K., and Van Doan, H. (2022). Effects of dietary supplementation of primaLac, inulin, and biomin imbo on growth performance, antioxidant, and innate immune responses of common carp (*Cyprinus carpio*). *Aquaculture Nutrition*, 2022, 1-13. <https://doi.org/10.1155/2022/8297479>.
- Akhter, N., Wu, B., Memon, A. M., and Mohsin, M. (2015). Probiotics and prebiotics associated with aquaculture: a review. *Fish and Shellfish Immunology*, 45(2), 733-741.
- Akmal, M., Rahimi-Midani, A., Hafeez-ur-Rehman, M., Hussain, A., and Choi, T. J. (2020). Isolation, characterisation, and application of a bacteriophage infecting the fish pathogen *Aeromonas hydrophila*. *Pathogens*, 9(3), 215. <https://doi.org/10.3390/pathogens9030215>.
- Akter, M. N., Hashim, R., Sutriana, A., Siti Azizah, M. N., and Asaduzzaman, M. (2019). Effect of *Lactobacillus acidophilus* supplementation on growth performances, digestive enzyme activities and gut histomorphology of striped catfish (*Pangasianodon hypophthalmus* Sauvage, 1878) juveniles. *Aquaculture Research*, 50(3), 786-797.
- Albarracin, L., Raya Tonetti, F., Fukuyama, K., Suda, Y., Zhou, B., Baillo, A., Fadda, S., Saavedra, L., Kurata, S., Hebert, E. M., Kitazawa, H., and Villena, J. (2022). Genomic characterization of *Lactiplantibacillus plantarum* strains possessing differential antiviral immunomodulatory activities. *Bacteria*, 1(3), 136-160.
- Al-Sheraji, S.H., Ismail, A., Manap, M.Y., Mustafa, S., Yusof, R.M., and Hassan, F.A. (2013). Prebiotics as functional foods: a review. *Journal of Functional Foods*, 5(4), 1542-1553.

- Al-Sum, A. B., and Al-Dhabi, N. A. (2014). Isolation of bacteriophage from *Mentha* species in Riyadh, Saudi Arabia. *Journal of Pure and Applied Microbiology*, 8(2), 945-949.
- Al-Taee, A. M. R., Khamees, N. R., and Al-Shammari, N. A. H. (2017). *Vibrio* species isolated from farmed fish in Basra city in Iraq. *Journal of Aquaculture Research and Development*, 8(2), 1000472. <https://doi.org/10.4172/2155-9546.1000472>.
- Amal, M. N. A., Koh, C. B., Nurliyana, M., Suhaiba, M., Nor-Amalina, Z., Santha, S., Diyana-Nadhirah, K. P., Yusof, M. T., Ina-Salwany, M. Y., and Zamri-Saad, M. (2018). A case of natural co infection of tilapia lake virus and *Aeromonas veronii* in a Malaysian red hybrid tilapia (*Oreochromis niloticus* × *O. mossambicus*) farm experiencing high mortality. *Aquaculture*, 485, 12–16.
- Amenyogbe, E., Chen, G., Wang, Z., Huang, J., Huang, B., and Li, H. (2020). The exploitation of probiotics, prebiotics and synbiotics in aquaculture: present study, limitations and future directions: a review. *Aquaculture International*, 28, 1017–1041.
- Amoah, K., Huang, Q. C., Tan, B. P., Zhang, S., Chi, S. Y., Yang, Q. H., Liu, H. Y., and Dong, X. H. (2019). Dietary supplementation of probiotic *Bacillus coagulans* ATCC 7050, improves the growth performance, intestinal morphology, microflora, immune response, and disease confrontation of Pacific white shrimp, *Litopenaeus vannamei*. *Fish and Shellfish Immunology*, 87, 796–808.
- Amorim, C., Silvério, S. C., Cardoso, B. B., Alves, J. I., Pereira, M. A., and Rodrigues, L. R. (2020). *In vitro* assessment of prebiotic properties of xylooligosaccharides produced by *Bacillus subtilis* 3610. *Carbohydrate Polymers*, 229, 115460. <https://doi.org/10.1016/j.carbpol.2019.115460>
- Amorim, C., Silvério, S.C., Cardoso, B.B., Alves, J.I., Pereira, M.A., and Rodrigues, L.R. (2020). *In vitro* fermentation of raffinose to unravel its potential as prebiotic ingredient. *LWT-Food Science and Technology*, 126, 109322. <https://doi.org/10.1016/j.lwt.2020.109322>.
- Ang, C. Y., and Lai, T. M. (2019). Isolation and characterisation of probiotic bacteria from the gastrointestinal tract of pond-cultured *Litopenaeus vannamei* in Tuaran, Sabah. *International Journal of Aquatic Science*, 10(1), 60-73.
- Ang, C. Y., Sano, M., Dan, S., Leelakriangsak, M., and Lal, T. M. (2020). Postbiotics applications as infectious disease control agent in aquaculture. *Biocontrol Science*, 25(1), 1-7.

- Anggraeni, A.A. (2022). Mini-Review: The potential of raffinose as a prebiotic. In IOP Conference Series: Earth and Environmental Science (Vol. 980, No. 1, p. 012033). IOP Publishing.
- Anisha, G. S., John, R. P., and Prema, P. (2009). Biochemical and hydrolytic properties of multiple thermostable α -galactosidases from *Streptomyces griseoalbus*: obvious existence of a novel galactose-tolerant enzyme. *Process Biochemistry*, 44(3), 327-333.
- Anjur, N., Sabran, S. F., Daud, H. M., and Othman, N. Z. (2021). An update on the ornamental fish industry in Malaysia: *Aeromonas hydrophila*-associated disease and its treatment control. *Veterinary World*, 14(5), 1143-1152.
- Anokyewaa, M. A., Amoah, K., Li, Y., Lu, Y., Kuebutornye, F. K., Asiedu, B., and Seidu, I. (2021). Prevalence of virulence genes and antibiotic susceptibility of *Bacillus* used in commercial aquaculture probiotics in China. *Aquaculture Reports*, 21, 100784. <https://doi.org/10.1016/j.aqrep.2021.100784>
- Anwar, M., Mros, S., McConnell, M., and Bekhit, A. E. D. A. (2021). Effects of extraction methods on the digestibility, cytotoxicity, prebiotic potential and immunomodulatory activity of taro (*Colocasia esculenta*) water-soluble non-starch polysaccharide. *Food Hydrocolloids*, 121, 107068. <https://doi.org/10.1016/j.foodhyd.2021.107068>
- Arfatahery, N., Davoodabadi, A., and Abedimohtasab, T. (2016). Characterization of toxin genes and antimicrobial susceptibility of *Staphylococcus aureus* isolates in fishery products in Iran. *Scientific Reports*, 6, 34216. <https://doi.org/10.1038/srep34216>.
- Arfatahery, N., Mirshafiey, A., Abedimohtasab, T. P., and ZeinolabediniZamani, M. (2015). Study of the prevalence of *Staphylococcus aureus* in marine and farmed shrimps in Iran aiming the future development of a prophylactic vaccine. *Procedia in Vaccinology*, 9, 44-49.
- Ashith, V. S., Deepthi, C. V., Joseph, R. M., Ramaswamy, A., and Vivekanandhan, G. (2019). Detection of hemolytic activity of *Aeromonas* sp. isolated from water samples of coastal area, Kochi. *Biotechnological Research*, 5(2), 9-15.
- Avendaño-Herrera, R. (2021). Salmon aquaculture, *Piscirickettsia salmonis* virulence, and one health: Dealing with harmful synergies between heavy antimicrobial use and piscine and human health comment on. *Aquaculture*, 532, 736062. <https://doi.org/10.1016/j.aquaculture.2020.736062>.

- Azmi, A. F. M. N., Mustafa, S., Hashim, D. M., and Manap, Y. A. (2012). Prebiotic activity of polysaccharides extracted from *Gigantochloa levis* (Buluh beting) shoots. *Molecules*, 17(2), 1635-1651.
- Azra, M. N., Okomoda, V. T., Tabatabaei, M., Hassan, M., and Ikhwanuddin, M. (2021). The contributions of shellfish aquaculture to global food security: assessing its characteristics from a future food perspective. *Frontiers in Marine Science*, 8, 654897. <https://doi.org/10.3389/fmars.2021.654897>.
- Balcázar, J. L., and Rojas-Luna, T. (2007). Inhibitory activity of probiotic *Bacillus subtilis* UTM 126 against *Vibrio* species confers protection against vibriosis in juvenile shrimp (*Litopenaeus vannamei*). *Current Microbiology*, 55, 409-412.
- Balderas-Ruiz, K. A., Bustos, P., Santamaria, R. I., González, V., Cristiano-Fajardo, S. A., Barrera-Ortíz, S., Mezo-Villalobos, M., Aranda-Ocampo, S., Guevara-Garcia, A. A., Galindo, E., and Serrano-Carreón, L. (2020). *Bacillus velezensis* 83 a bacterial strain from mango phyllosphere, useful for biological control and plant growth promotion. *Amb Express*, 10(1), 163. <https://doi.org/10.1186/s13568-020-01101-8>
- Bao, Y., Zhang, Y., Zhang, Y., Liu, Y., Wang, S., Dong, X., Wang, Y., and Zhang, H. (2010). Screening of potential probiotic properties of *Lactobacillus fermentum* isolated from traditional dairy products. *Food Control*, 21(5), 695-701.
- Barer, M. R., and Irving, W. L. (Eds.). (2018). Medical Microbiology E-Book: A Guide to Microbial Infections: Pathogenesis, Immunity, Laboratory Investigation and Control. Leicester, United Kingdom. Elsevier Health Sciences.
- Barría, A., Trinh, T. Q., Mahmuddin, M., Benzie, J. A., Chadag, V. M., and Houston, R. D. (2020). Genetic parameters for resistance to Tilapia Lake Virus (TiLV) in Nile tilapia (*Oreochromis niloticus*). *Aquaculture*, 522, 735126. <https://doi.org/10.1016/j.aquaculture.2020.735126>
- Bastardo, A., Ravelo, C., Castro, N., Calheiros, J., and Romalde, J. L. (2012). Effectiveness of bivalent vaccines against *Aeromonas hydrophila* and *Lactococcus garvieae* infections in rainbow trout *Oncorhynchus mykiss* (Walbaum). *Fish and Shellfish Immunology*, 32(5), 756-761.
- Becking, T., Kiselev, A., Rossi, V., Street-Jones, D., Grandjean, F., and Gaulin, E. (2022). Pathogenicity of animal and plant parasitic *Aphanomyces* spp. and their economic impact on aquaculture and agriculture. *Fungal Biology Reviews*, 40, 1-18.

- Bello-López, J. M., Cabrero-Martínez, O. A., Ibáñez-Cervantes, G., Hernández-Cortez, C., Pelcastre-Rodríguez, L. I., Gonzalez-Avila, L. U., and Castro Escarpulli, G. (2019). Horizontal gene transfer and its association with antibiotic resistance in the genus *Aeromonas* spp. *Microorganisms*, 7(9), 363. <https://doi.org/10.3390/microorganisms7090363>.
- Bentzon-Tilia, M., Sonnenschein, E. C., and Gram, L. (2016). Monitoring and managing microbes in aquaculture—towards a sustainable industry. *Microbial Biotechnology*, 9(5), 576-584.
- Bird, K., Boopathy, R., Nathaniel, R., and LaFleur, G. (2019). Water pollution and observation of acquired antibiotic resistance in *Bayou Lafourche*, a major drinking water source in Southeast Louisiana, USA. *Environmental Science and Pollution Research*, 26(33), 34220-34232.
- Biswas, S., Kim, M. H., Baek, D. H., and Kim, I. H. (2023). Probiotic mixture (*Bacillus subtilis* and *Bacillus licheniformis*) a potential in-feed additive to improve broiler production efficiency, nutrient digestibility, caecal microflora, meat quality and to diminish hazardous odour emission. *Journal of Animal Physiology and Animal Nutrition*, 107(4), 1065-1072.
- Boris, S., Suarez, J. E., and Barbes, C. (1997). Characterization of the aggregation promoting factor from *Lactobacillus gasseri*, avaginal isolate. *Journal of Applied Microbiology*, 83(4), 413-420.
- Borowitzka, M. A. (2018). Biology of microalgae. In *Microalgae in health and disease prevention* (pp. 23-72). Western Australia, Australia. Academic Press.
- Boyd, C. E., and Gross, A. (1998). Use of probiotics for improving soil and water quality in aquaculture ponds. *Advances in Shrimp Biotechnology*, 101-105.
- Briones-Hidrovo, A., Quinteiro, P., and Dias, A. C. (2023). Investigating the environmental sustainability of a seabass and seabream aquaculture system in Portugal based on life cycle and nexus approaches. *Science of The Total Environment*, 890, 164195. <https://doi.org/10.1016/j.scitotenv.2023.164195>.
- Bull, M., Plummer, S., Marchesi, J., and Mahenthiralingam, E. (2013). The life history of *Lactobacillus acidophilus* as a probiotic: a tale of revisionary taxonomy, misidentification and commercial success. *FEMS Microbiology Letters*, 349(2), 77-87.
- Buntin, N., Hongpattarakere, T., Ritari, J., Douillard, F.P., Paulin, L., Boeren, S., Shetty, S. A., and de Vos, W.M. (2017). An inducible operon is involved in inulin utilisation in *Lactobacillus plantarum* strains, as

- revealed by comparative proteogenomics and metabolic profiling. *Applied and Environmental Microbiology*, 83(2), e02402-16. <https://doi.org/10.1128/AEM.02402-16>.
- Buruiană, C. T., Profr, A.G., and Vizireanu, C. (2014). Effects of probiotic *Bacillus* species in aquaculture- an overview. *Annals of the University Dunarea de Jos of Galati*, 38, 9-17.
- Butucel, E., Balta, I., McCleery, D., Marcu, A., Stef, D., Pet, I., Callaway, T., Stef, L., and Corcionivoschi, N. (2022). The prebiotic effect of an organic acid mixture on *Faecalibacterium prausnitzii* metabolism and its anti-pathogenic role against *Vibrio parahaemolyticus* in shrimp. *Biology*, 12(1), 57. <https://doi.org/10.3390/biology12010057>.
- Bwadi, B. E., Mustafa, F. B., Ali, M. L., and Bhassu, S. (2018). Spatial analysis of water quality and its suitability in farming giant freshwater prawn (*Macrobrachium rosenbergii*) in Negeri Sembilan region, Peninsular Malaysia. *Singapore Journal of Tropical Geography*, 40(1), 71-91.
- Cabello, F. C. (2006). Heavy use of prophylactic antibiotics in aquaculture: a growing problem for human and animal health and for the environment. *Environmental Microbiology*, 8(7), 1137-1144.
- Cantarel, B. L., Coutinho, P. M., Rancurel, C., Bernard, T., Lombard, V., and Henrissat, B. (2009). The Carbohydrate-Active EnZymes database (CAZy): an expert resource for glycogenomics. *Nucleic Acids Research*, 37(1), D233-D238.
- Cao, Y., Zhang, Y., Lan, W., and Sun, X. (2021). Characterisation of vB_VpaP_MGD2, a newly isolated bacteriophage with biocontrol potential against multidrug-resistant *Vibrio parahaemolyticus*. *Archives of Virology*, 166(2), 413-426.
- Chaiklahan, R., Chirasuwan, N., Triratana, P., Tia, S., and Bunnag, B. (2014). Effect of extraction temperature on the diffusion coefficient of polysaccharides from Spirulina and the optimal separation method. *Biotechnology and Bioprocess Engineering*, 19, 369-377.
- Chaillou, S., Lucquin, I., Najjari, A., Zagorec, M., and Champomier-Verges, M. C. (2013). Population genetics of *Lactobacillus sakei* reveals three lineages with distinct evolutionary histories. *PLoS One*, 8(9), e73253. <https://doi.org/10.1371/journal.pone.0073253>.
- Chang, Y., Yin, C., Peng, H., and Shi, Y. (2020). Differentially proteomic analysis of the hemocytes against *Aeromonas hydrophila* infection in oriental river prawn *Macrobrachium nipponense* by iTRAQ approach. *Fish and Shellfish Immunology*, 104, 324-336.

- Chauhan, A., and Singh, R. (2019). Isolation and evaluation of putative probiotic strains from different teleost to prevent *Pseudomonas aeruginosa* infection in *Cyprinus carpio*. *Aquaculture Research*, 50(12), 3616-3627.
- Chen, J., Sun, R., Pan, C., Sun, Y., Mai, B., and Li, Q. X. (2020). Antibiotics and food safety in aquaculture. *Journal of Agricultural and Food Chemistry*, 68(43), 11908-11919.
- Chen, S. C., Lin, Y. D., Liaw, L. L., and Wang, P. C. (2001). *Lactococcus garvieae* infection in the giant freshwater prawn *Macrobrachium rosenbergii* confirmed by polymerase chain reaction and 16S rDNA sequencing. *Diseases of Aquatic Organisms*, 45(1), 45-52.
- Chen, W. W., Romano, N., Ebrahimi, M., and Natrah, I. (2017). The effects of dietary fructooligosaccharide on growth, intestinal short chain fatty acids level and hepatopancreatic condition of the giant freshwater prawn (*Macrobrachium rosenbergii*) post-larvae. *Aquaculture*, 469, 95-101.
- Chen, X. H., Koumoutsi, A., Scholz, R., Eisenreich, A., Schneider, K., Heinemeyer, I., Morgenstern, B., Voss, B., Hess, W. R., Reva, O., Junge, H., Voight, B., Jungblut, P. R., Vater, J., Sussmuth, R., Liesegang, H., Strittmatter, A., Gottschalk, G., and Borriis, R. (2007). Comparative analysis of the complete genome sequence of the plant growth-promoting bacterium *Bacillus amyloliquefaciens* FZB42. *Nature Biotechnology*, 25(9), 1007-1014.
- Cheng, Z. X., Ma, Y. M., Li, H., and Peng, X. X. (2014). N-acetylglucosamine enhances survival ability of tilapias infected by *Streptococcus iniae*. *Fish and Shellfish Immunology*, 40(2), 524-530.
- Chiew, I. K. M. (2017). Resistant starch from underutilised legumes as prebiotic and its effect on the growth of *Danio rerio* and *Lates calcarifer* (Doctoral dissertation, University of Nottingham Malaysia Campus, Selangor, Malaysia). Retrieved from <https://core.ac.uk/download/pdf/159994068.pdf> on 14th November 2020.
- Chikkerur, J., Samanta, A.K., Kolte, A.P., Dhali, A., and Roy, S. (2020). Production of short chain fructo-oligosaccharides from inulin of chicory root using fungal endoinulinase. *Applied Biochemistry and Biotechnology*, 191(2), 695-715.
- Church, D. L., Cerutti, L., Gürtler, A., Griener, T., Zelazny, A., and Emmer, S. (2020). Performance and application of 16S rRNA gene cycle sequencing for routine identification of bacteria in the clinical microbiology laboratory. *Clinical Microbiology Reviews*, 33(4), e00053-19. <https://doi.org/10.1128/cmr.00053-19>.

- Cleveland, J., Montville, T. J., Nes, I. F., and Chikindas, M. L. (2001). Bacteriocins: safe, natural antimicrobials for food preservation. *International Journal of Food Microbiology*, 71(1), 1-20.
- CLSI. (2015). Verification of commercial microbial identification and antimicrobial susceptibility testing systems—first edition: m52-ed1. Wayne, pa: Clinical and laboratory standards institute.
- Coleman, D. J., Camus, A. C., Martínez-López, B., Yun, S., Stevens, B., and Soto, E. (2018). Effects of temperature on *Veronaea botryosa* infections in white sturgeon *Acipenser transmontanus* and fungal induced cytotoxicity of fish cell lines. *Veterinary Research*, 49(1), 1-9.
- Collado, M. C., Meriluoto, J., and Salminen, S. (2008). Adhesion and aggregation properties of probiotic and pathogen strains. *European Food Research and Technology*, 226, 1065-1073.
- Cutting, S. M. (2011). *Bacillus* probiotics. *Food Microbiology*, 28(2), 214-220.
- da Silva Menezes, B., Rossi, D. M., Squina, F., and Ayub, M. A. Z. (2018). Xylooligosaccharides production by fungi cultivations in rice husk and their application as substrate for lactic acid bacteria growth. *Bioresource Technology Reports*, 2, 100-106.
- Daghlas, S. A., and Mohiuddin, S. S. (2019). Biochemistry, Glycogen. StatPearls Publishing LLC.
- Dahech, I., Ayed, H. B., Belghith, K. S., Belghith, H., and Mejdoub, H. (2013). Microbial production of levanase for specific hydrolysis of levan. *International Journal of Biological Macromolecules*, 60, 128-133.
- Dan, T., Liu, W., Song, Y., Xu, H., Menghe, B., Zhang, H., and Sun, Z. (2015). The evolution and population structure of *Lactobacillus fermentum* from different naturally fermented products as determined by multilocus sequence typing (MLST). *BMC Microbiology*, 15, 107. <https://doi.org/10.1186/s12866-015-0447-z>.
- Dar, G. H., Dar, S. A., Kamili, A. N., Chishti, M. Z., and Ahmad, F. (2016). Detection and characterisation of potentially pathogenic *Aeromonas sobria* isolated from fish *Hypophthalmichthys molitrix* (Cypriniformes: Cyprinidae). *Microbial Pathogenesis*, 91, 136-140.
- Das, S., Dash, H. R., Mangwani, N., Chakraborty, J., and Kumari, S. (2014). Understanding molecular identification and polyphasic taxonomic approaches for genetic relatedness and phylogenetic relationships of microorganisms. *Journal of Microbiological Methods*, 103, 80-100.

- Das, S., Mondal, K., and Haque S. (2017). A review on application of probiotic, prebiotic and synbiotic for sustainable development of aquaculture. *Journal of Entomology and Zoology Studies*, 5(2), 422-429.
- Davani-Davari, D., Negahdaripour, M., Karimzadeh, I., Seifan, M., Mohkam, M., Masoumi, S. J., Berenjian, A., and Ghasemi, Y. (2019). Prebiotics: definition, types, sources, mechanisms, and clinical applications. *Foods*, 8(3), 92-118.
- Dawood, M. A. O., Eweedah, N. M., Moustafa, E. M., and Shahin, M. G. (2019). Synbiotic effects of *Aspergillus oryzae* and β-Glucan on growth and oxidative and immune responses of Nile tilapia, *Oreochromis niloticus*. *Probiotics and Antimicrobial Proteins*, 12(1), 172-183.
- Dawood, M. A., and Koshio, S. (2016). Recent advances in the role of probiotics and prebiotics in carp aquaculture: a review. *Aquaculture*, 454, 243-251.
- de Campos, C. M., Zanuzzo, F. S., Gimbo, R. Y., Favero, G. C., Soares, M. P., Pilarski, F., and Urbinati, E. C. (2022). Dietary inulin modulated the cortisol response and increased the protection against pathogens in juvenile pacu (*Piaractus mesopotamicus*). *Aquaculture Research*, 53(3), 860-869.
- De Jesus, M. R., de Morais, A., and de Morais, R. (2016). Emergent sources of prebiotics: seaweeds and microalgae. *Marine Drugs*, 14(2), 27. <https://doi.org/10.3390/md14020027>.
- Deeseenthum, S., Leelavatcharamas, V., and Brookes, J. D. (2007). Effect of feeding *Bacillus* sp. as probiotic bacteria on growth of giant freshwater prawn (*Macrobrachium rosenbergii* de Man). *Pakistan Journal of Biological Sciences: PJBS*, 10(9), 1481-1485.
- Del Valle, J. C., Bonadero, M. C., and Gimenez, A. V. F. (2023). *Saccharomyces cerevisiae* as probiotic, prebiotic, synbiotic, postbiotics and parabiotics in aquaculture: an overview. *Aquaculture*, 569, 739342.
- Deleu, M., Paquot, M., and Nylander, T. (2008). Effect of Fengycin, a lipopeptide produced by *Bacillus subtilis*, on model biomembranes. *Biophysical Journal*, 94(7), 2667-2679.
- Denis, M., Ramasamy, S. M., Doss, B. S., and Thayappan, K. (2016). Calcium dependent lectin in the serum of the marine crab *Atergatis subdentatus* (De Hann, 1835). *Journal of Modern Biotechnology*, 5(5), 1-5.

Department of Fisheries Malaysia. (2020). Malaysian fishing industry scenario. Retrieved on from <https://www.dof.gov.my/index.php/pages/view/42> 31st October 2020.

Deris, Z. M., Iehata, S., Ikhwanuddin, M., Sahimi, M. B. M. K., Do, T. D., Sorgeloos, P., Sung, Y. Y., and Wong, L. L. (2020). Immune and bacterial toxin genes expression in different giant tiger prawn, *Penaeus monodon* post-larvae stages following AHPND-causing strain of *Vibrio parahaemolyticus* challenge. *Aquaculture Reports*, 16, 100248. <https://doi.org/10.1016/j.aqrep.2019.100248>.

Di Stefano, M., Miceli, E., Gotti, S., Missanelli, A., Mazzocchi, S., and Corazza, G.R. (2007). The effect of oral α -galactosidase on intestinal gas production and gas-related symptoms. *Digestive Diseases and Sciences*, 52(1), 78-83.

Di, J., Chu, Z., Zhang, S., Huang, J., Du, H., and Wei, Q. (2019). Evaluation of the potential probiotic *Bacillus subtilis* isolated from two ancient sturgeons on growth performance, serum immunity and disease resistance of *Acipenser dabryanus*. *Fish and Shellfish Immunology*, 93, 711-719.

Ding, T., Sun, H., Pan, Q., Zhao, F., Zhang, Z., and Ren, H. (2020). Isolation and characterization of *Vibrio parahaemolyticus* bacteriophage vB_VpaS_PG07. *Virus Research*, 286, 198080. <https://doi.org/10.1016/j.virusres.2020.198080>

Doan, C.T., Tran, T.N., Nguyen, T.T., Tran, T.P.H., Nguyen, V.B., Tran, T.D., Nguyen, A. D., and Wang, S.L. (2021). Production of sucrolytic enzyme by *Bacillus licheniformis* by the bioconversion of Pomelo Albedo as a carbon source. *Polymers*, 13(12), 1959. <https://doi.org/10.3390/polym13121959>.

Dolganyuk, V., Sukhikh, S., Kalashnikova, O., Ivanova, S., Kashirskikh, E., Prosekov, A., Michaud, P., and Babich, O. (2023). Food proteins: potential resources. *Sustainability*, 15(7), 5863. <https://doi.org/10.3390/su15075863>.

Dong, Y., Yang, Y., Liu, J., Awan, F., Lu, C., and Liu, Y. (2018). Inhibition of *Aeromonas hydrophila*-induced intestinal inflammation and mucosal barrier function damage in crucian carp by oral administration of *Lactococcus lactis*. *Fish and Shellfish Immunology*, 83, 359-367.

Drula, E., Garron, M. L., Dogan, S., Lombard, V., Henrissat, B., and Terrapon, N. (2022). The carbohydrate-active enzyme database: functions and literature. *Nucleic acids Research*, 50(D1), D571-D577.

- Eckel, V. P., Ziegler, L. M., Vogel, R. F., and Ehrmann, M. (2020). *Bifidobacterium tibiigranuli* sp. nov. isolated from homemade water kefir. *International Journal of Systematic and Evolutionary Microbiology*, 70(3), 1562-1570.
- Einarsson, Á., and Óladóttir, Á. D. (2020). *Fisheries and Aquaculture: The Food Security of the Future*. London Wall, United Kingdom. Academic Press.
- Elain, A., Nkounkou, C., Le Fellic, M., and Donnart, K. (2020). Green extraction of polysaccharides from *Arthrospira platensis* using high pressure homogenization. *Journal of Applied Phycology*, 32(3), 1719-1727.
- El-Sayed, A. F. M. (2021). Use of biofloc technology in shrimp aquaculture: a comprehensive review, with emphasis on the last decade. *Reviews in Aquaculture*, 13(1), 676-705.
- Eltanahy, E., and Torky, A. (2021). Microalgae as cell factories: food and feed-grade high-value metabolites. *Microalgal Biotechnology: Recent Advances, Market Potential, and Sustainability*; Royal Society of Chemistry: Cambridge, UK, 1-35.
- Emam, A. M., and Dunlap, C. A. (2020). Genomic and phenotypic characterization of *Bacillus velezensis* AMB-y1; a potential probiotic to control pathogens in aquaculture. *Antonie van Leeuwenhoek*, 113(12), 2041-2052.
- Emerenciano, M. G., Rombenso, A. N., Vieira, F. D. N., Martins, M. A., Coman, G. J., Truong, H. H., Noble, T. H., and Simon, C. J. (2022). Intensification of penaeid shrimp culture: an applied review of advances in production systems, nutrition and breeding. *Animals*, 12(3), 236. <https://doi.org/10.3390/ani12030236>.
- Escobedo-Bonilla, C. M., Alday-Sanz, V., Wille, M., Sorgeloos, P., Pensaert, M. B., and Nauwynck, H. J. (2008). A review on the morphology, molecular characterisation, morphogenesis and pathogenesis of white spot syndrome virus. *Journal of Fish Diseases*, 31(1), 1-18.
- Escobedo-Fregoso, C., Quiroz-Guzmán, E., Mendoza-Carrion, G., and Peña-Rodríguez, A. (2021). Effect of dietary prebiotic inulin and probiotic *Bacillus subtilis* and *Lactobacillus* sp., on the intestinal microbiota of white shrimp *Litopenaeus vannamei*. *Biotechnia*, 23(3), 50-57.
- Fan, B., Blom, J., Klenk, H. P., and Borris, R. (2017). *Bacillus amyloliquefaciens*, *Bacillus velezensis*, and *Bacillus siamensis* form an “operational group *B. amyloliquefaciens*” within the *B. subtilis* species complex. *Frontiers in Microbiology*, 8, 22. <https://doi.org/10.3389/fmicb.2017.00022>.

- Fan, B., Wang, C., Song, X., Ding, X., Wu, L., Wu, H., Gao, X., and Boriss, R. (2018). *Bacillus velezensis* FZB42 in 2018: the gram-positive model strain for plant growth promotion and biocontrol. *Frontiers in Microbiology*, 9, 2491. <https://doi.org/10.3389/fmicb.2018.02491>.
- FAO (2016). Contributing to food security and nutrition for all. The State of World Fisheries and Aquaculture. Rome. Itali (pp. 200), FAO.
- FAO (2018). Meeting the sustainable development goals. The State of World Fisheries and Aquaculture. Sustainability in action. Rome. Itali (pp. 227), FAO.
- FAO (2019). Fishery and aquaculture country profiles. Malaysia. *Food and Agriculture Organization of the United Nations*. Retrieved from <http://www.fao.org/fishery/facp/MYS/en> on 30th October 2020.
- FAO. (1997). Aquaculture development. FAO technical guidelines for responsible fisheries. 1020–5292, No.5. FAO, *Fisheries Department*, Rome, 4.
- FAO. (2020). Sustainability in action. The State of World Fisheries and Aquaculture. Rome. Itali (pp. 244), FAO.
- FAO. (2022). The State of World Fisheries and Aquaculture 2022. Towards Blue Transformation. Rome, FAO. <https://doi.org/10.4060/cc0461en>.
- Feng, Y., Bai, M., Geng, Y., Chen, D., Huang, X., Ouyang, P., Guo, H., Zuo, Z., Huang, C., and Lai, W. (2021). The potential risk of antibiotic resistance of *Streptococcus iniae* in sturgeon cultivation in Sichuan, China. *Environmental Science and Pollution Research*, 28(48), 69171-69180.
- Flores, M. L. (2011). The use of probiotic in aquaculture: an overview. *International Research Journal of Microbiology*, 2(12), 471-478.
- Foo, S. M., Eng, W. W. H., Lee, Y. P., Gui, K., and Gan, H. M. (2017). New sequence types, of *Vibrio parahaemolyticus* isolated from a Malaysian aquaculture pond, as revealed, by whole-genome sequencing. *Genome Announcements*, 5(19), e00302-17. <https://doi.org/10.1128/genomea.00302-17>.
- Fox, J. D., and Robyt, J. F. (1991). Miniaturization of three carbohydrate analyses using a microsample plate reader. *Analytical Biochemistry*, 195(1), 93-96.
- Fry, B., Zhu, T., Domach, M. M., Koepsel, R. R., Phalakornkule, C., and Ataai, M. M. (2000). Characterization of growth and acid formation in a *Bacillus subtilis* pyruvate kinase mutant. *Applied and Environmental Microbiology*, 66(9), 4045-4049.

- Fuandila, N. N., Widanarni, W., and Yuhana, M. (2019). Growth performance and immune response of prebiotic honey fed pacific white shrimp *Litopenaeus vannamei* to *Vibrio parahaemolyticus* infection. *Journal of Applied Aquaculture*, 32(3), 221-235.
- Ganzle, M.G. 2011. Lactose and oligosaccharides | lactose: derivatives. *Encyclopedia of Dairy Sciences*, 2, 202-208.
- Gao, X. Y., Liu, Y., Miao, L. L., Li, E. W., Hou, T. T., and Liu, Z. P. (2017). Mechanism of anti-*Vibrio* activity of marine probiotic strain *Bacillus pumilus* H2, and characterization of the active substance. *AMB Express*, 7, 23. <https://doi.org/10.1186/s13568-017-0323-3>.
- García-Bernal, M., Medina-Marrero, R., Rodríguez-Jaramillo, C., Marrero-Chang, O., Campa-Córdova, Á. I., Medina-García, R., and Mazón-Suástegui, J. M. (2018). Probiotic effect of *Streptomyces* spp. on shrimp (*Litopenaeus vannamei*) postlarvae challenged with *Vibrio parahaemolyticus*. *Aquaculture Nutrition*, 24(2), 865-871.
- Garlock, T., Asche, F., anderson, J., Bjørndal, T., Kumar, G., Lorenzen, K., Ropicki, A., Smith, M. D., and Tveterås, R. (2019). A global blue revolution: aquaculture growth across regions, species, and countries. *Reviews in Fisheries Science and Aquaculture*, 28(1), 107-116.
- Gewaily, M. S., Shukry, M., Abdel-Kader, M. F., Alkafafy, M., Farrag, F. A., Moustafa, E. M., Doan, H. V., Abd-Elghany, M. F., Abdelhamid, A. F., Eltanahy, A., and Dawood, M. A. (2021). Dietary *Lactobacillus plantarum* relieves Nile tilapia (*Oreochromis niloticus*) juvenile from oxidative stress, immunosuppression, and inflammation induced by deltamethrin and *Aeromonas hydrophila*. *Frontiers in Marine Science*, 8, 621558. <https://doi.org/10.3389/fmars.2021.621558>.
- Ghafarifarsani, H., Rashidian, G., Bagheri, T., Hoseinifar, S. H., and Van Doan, H. (2021). Study on growth enhancement and the protective effects of dietary prebiotic inulin on immunity responses of rainbow trout (*Oncorhynchus mykiss*) fry infected with *Aeromonas hydrophila*. *Annals of Animal Science*, 21(2), 543-559.
- Ghosh, S., Sinha, A., and Sahu, C. (2007). Isolation of putative probiotics from the intestines of Indian major carps. *Israeli Journal of Aquaculture*, 59(3), 127-132.
- Gibson, G. R., Hutkins, R., Sanders, M. E., Prescott, S. L., Reimer, R. A., Salminen, S. J., Karen, S., Stanton, C., Swanson, K. S., Cani, P. D., Verbeke, K., and Reid, G. (2017). Expert consensus document: The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of

- prebiotics. *Nature Reviews Gastroenterology and Hepatology*, 14(8), 491-502.
- Goh, Y.J., Lee, J.H., and Hutzins, R.W. (2007). Functional analysis of the fructooligosaccharide utilisation operon in *Lactobacillus paracasei* 1195. *Applied and Environmental Microbiology*, 73(18), 5716-5724.
- Golmakani, M. T., Soleimanian-Zad, S., Alavi, N., Nazari, E., and Eskandari, M. H. (2019). Effect of Spirulina (*Arthrospira platensis*) powder on probiotic bacteriologically acidified feta-type cheese. *Journal of Applied Phycology*, 31, 1085-1094.
- González-Jaramillo, L. M., Aranda, F. J., Teruel, J. A., Villegas-Escobar, V., and Ortiz, A. (2017). Antimycotic activity of Fengycin C biosurfactant and its interaction with phosphatidylcholine model membranes. *Colloids and Surfaces B: Biointerfaces*, 156, 114-122.
- Grosshagauer, S., Kraemer, K., and Somoza, V. (2020). The true value of Spirulina. *Journal of Agricultural and Food Chemistry*, 68(14), 4109-4115.
- Guarino, P. L., M., Altomare, A., Emerenziani, S., Di Rosa, C., Ribolsi, M., Balestrieri, P., Lovino, P., Rocchi, G., and Cicala, M. (2020). Mechanisms of action of prebiotics and their effects on gastrointestinal disorders in adults. *Nutrients*, 12(4), 1037-1060.
- Guerreiro, I., Olivia-Teles, A., and Enes, P. (2017). Prebiotics as functional ingredients: focus on Mediterranean fish aquaculture. *Reviews in Aquaculture*, 10(4), 800-832.
- Guo, Y., Xing, Z., He, Z., Zhao, X., and Ye, S. (2020). Honokiol inhibits *Vibrio harveyi* hemolysin virulence by reducing its haemolytic activity. *Aquaculture Research*, 51(1), 206-214.
- Hamza, F., Kumar, A. R., and Zinjarde, S. (2018). Efficacy of cell free supernatant from *Bacillus licheniformis* in protecting *Artemia salina* against *Vibrio alginolyticus* and *Pseudomonas gessardii*. *Microbial Pathogenesis*, 116, 335-344.
- Han, Z. L., Yang, M., Fu, X. D., Chen, M., Su, Q., Zhao, Y. H., and Mou, H. J. (2019). Evaluation of prebiotic potential of three marine algae oligosaccharides from enzymatic hydrolysis. *Marine Drugs*, 17(3), 173. <https://doi.org/10.3390%2Fmd17030173>.
- Haniastuti, T., Puspasari, T. A., Hakim, E. R., and Tandelilin, R. T. (2023). Potential effect of giant freshwater prawn shell nano chitosan in inhibiting the development of *Streptococcus mutans* and *Streptococcus sanguinis* biofilm *in vitro*. *International Journal of Dentistry*, 2023, 8890750. <https://doi.org/10.1155/2023/8890750>.

- Hasheminya, S. M., and Dehghannya, J. (2020). Novel ultrasound-assisted extraction of kefiran biomaterial, a prebiotic exopolysaccharide, and investigation of its physicochemical, antioxidant and antimicrobial properties. *Materials Chemistry and Physics*, 243, 122645. <https://doi.org/10.1016/j.matchemphys.2020.122645>.
- Hayatgheib, N., Moreau, E., Calvez, S., Lepelletier, D., and Pouliquen, H. (2020). A review of functional feeds and the control of *Aeromonas* infections in freshwater fish. *Aquaculture International*, 28, 1083-1123.
- Hegde, A., Kabra, S., Basawa, R. M., Khile, D. A., Abbu, R. U. F., Thomas, N. A., Manickam, N. B., and Raval, R. (2023). Bacterial diseases in marine fish species: current trends and future prospects in disease management. *World Journal of Microbiology and Biotechnology*, 39(11), 317. <https://doi.org/10.1007/s11274-023-03755-5>.
- Heo, S., Kim, J. H., Kwak, M. S., Sung, M. H., and Jeong, D. W. (2021). Functional annotation genome unravels potential probiotic *Bacillus velezensis* strain KMU01 from traditional Korean fermented kimchi. *Foods*, 10(3), 563. <https://doi.org/10.3390%2Ffoods10030563>.
- Hersi, M. A., Genc, E., Pipilos, A., and Keskin, E. (2023). Effects of dietary synbiotics and biofloc meal on the growth, tissue histomorphology, whole-body composition and intestinal microbiota profile of Nile tilapia (*Oreochromis niloticus*) cultured at different salinities. *Aquaculture*, 570, 739391. <https://doi.org/10.1016/j.aquaculture.2023.739391>.
- Hill, C., Guarner, F., Reid, G., Gibson, G. R., Merenstein, D. J., Pot, B., Morelli, L., Canani, R. B., Flint, G. H., Salminen, S., Calder, P. C., and Sanders, M. E. (2014). Expert consensus document: The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nature reviews Gastroenterology and hepatology*, 11, 506-514.
- Holmes, Z. C., Villa, M. M., Durand, H. K., Jiang, S., Dallow, E. P., Petrone, B. L., Silberman, J. D., Lin, P. H., and David, L. A. (2022). Microbiota responses to different prebiotics are conserved within individuals and associated with habitual fiber intake. *Microbiome*, 10, 114. <https://doi.org/10.1186/s40168-022-01307-x>.
- Hong, T., Yin, J. Y., Nie, S. P., and Xie, M. Y. (2021). Applications of infrared spectroscopy in polysaccharide structural analysis: progress, challenge and perspective. *Food Chemistry*: X, 12, 100168. <https://doi.org/10.1016/j.fochx.2021.100168>.

- Hong, W., Zhang, M., and Cheng, J. X. (2022). Rapid determination of antimicrobial susceptibility by SRS single-cell metabolic imaging. In *Stimulated Raman Scattering Microscopy* (pp. 445-461). Elsevier.
- Hoque, F., Jawahar Abraham, T., Nagesh, T. S., and Kamilya, D. (2019). *Pseudomonas aeruginosa* FARP 72 offers protection against *Aeromonas hydrophila* infection in *Labeo rohita*. *Probiotics and Antimicrobial Proteins*, 11, 973-980.
- Hoseinifar, S. H., Sun, Y. Z., Wang, A., and Zhou, Z. (2018). Probiotics as means of diseases control in aquaculture, a review of current knowledge and future perspectives. *Frontiers in Microbiology*, 9, 2429. <https://doi.org/10.3389/fmicb.2018.02429>.
- Hoseinifar, S.H., Sharifian, M., Vesaghi, M.J., Khalili, M., and Esteban, M. Á. (2014). The effects of dietary xylooligosaccharide on mucosal parameters, intestinal microbiota and morphology and growth performance of Caspian white fish (*Rutilus frisii kutum*) fry. *Fish and Shellfish Immunology*, 39(2), 231-236.
- Hu, W., Yu, X., Jin, D., Zhai, F., Zhou, P., Ali, K. T., Cui, J., Wang, P., Liu, X., Sun, Y., Yi, G., and Xia, L. (2021). Isolation of a new *Streptomyces virginiae* W18 against fish pathogens and its effect on disease resistance mechanism of *Carassius auratus*. *Microbial Pathogenesis*, 161, 105273. <https://doi.org/10.1016/j.micpath.2021.105273>.
- Hu, X., Yang, H. L., Yan, Y. Y., Zhang, C. X., Ye, J. D., Lu, K. L., Hu, L. H., Zhang, J. J., Ruan, L., and Sun, Y. Z. (2019). Effects of fructooligosaccharide on growth, immunity and intestinal microbiota of shrimp (*Litopenaeus vannamei*) fed diets with fish meal partially replaced by soybean meal. *Aquaculture Nutrition*, 25(1), 194-204.
- Hu, Y., Chen, W., Shen, Y., Zhu, B., and Wang, G. X. (2019). Synthesis and antiviral activity of coumarin derivatives against infectious hematopoietic necrosis virus. *Bioorganic and Medicinal Chemistry Letters*, 29(14), 1749-1755.
- Hua, P., Yu, Z., Xiong, Y., Liu, B., and Zhao, L. (2018). Regulatory efficacy of *Spirulina platensis* protease hydrolyzate on lipid metabolism and gut microbiota in high-fat diet-fed rats. *International Journal of Molecular Sciences*, 19(12), 4023. <https://doi.org/10.3390/ijms19124023>.
- Huang, C. H., Chen, C. C., Liou, J. S., Lee, A. Y., Blom, J., Lin, Y. C., Huang, L., and Watanabe, K. (2020). Genome-based reclassification of *Lactobacillus casei*: emended classification and description of the species *Lactobacillus zeae*. *International Journal of Systematic and Evolutionary Microbiology*, 70(6), 3755-3762.

- Huang, H. T., Hu, Y. F., Lee, B. H., Huang, C. Y., Lin, Y. R., Huang, S. N., Chen, Y. Y., Chang, J. J., and Nan, F. H. (2022). Dietary of *Lactobacillus paracasei* and *Bifidobacterium longum* improve non-specific immune responses, growth performance, and resistance against *Vibrio parahaemolyticus* in *Penaeus vannamei*. *Fish and Shellfish Immunology*, 128, 307-315.
- Huang, H., Zhou, P., Chen, P., Xia, L., Hu, S., Yi, G., Lu, J., Yang, S., Xie, J., Peng, J., and Ding, X. (2020). Alteration of the gut microbiome and immune factors of grass carp infected with *Aeromonas veronii* and screening of an antagonistic bacterial strain (*Streptomyces flavotrichini*). *Microbial Pathogenesis*, 143, 104092. <https://doi.org/10.1016/j.micpath.2020.104092>.
- Huang, Y., Ma, F. T., and Ren, Q. (2020). Function of the MOB kinase activator-like 1 in the innate immune defense of the oriental river prawn (*Macrobrachium nipponense*). *Fish and Shellfish Immunology*, 102, 440-448.
- Huang, Y., Zhang, H., Ben, P., Duan, Y., Lu, M., Li, Z., and Cui, Z. (2018). Characterization of a novel GH36 α-galactosidase from *Bacillus megaterium* and its application in degradation of raffinose family oligosaccharides. *International Journal of Biological Macromolecules*, 108, 98-104.
- Huyen, T. T., Trang, H. P., Thi-Ngan, N., Dinh-Thanh, B., Quoc, L. P. T., and Nam, T. N. (2023). The responsibility of C-terminal domain in the thermolabile haemolysin activity of *Vibrio parahaemolyticus* and inhibition treatments by *Phellinus* sp. extracts. *Fisheries and Aquatic Sciences*, 26(3), 204-215.
- Ina-Salwany, M. Y., Al-saari, N., Mohamad, A., Mursidi, F. A., Mohd-Aris, A., Amal, M. N. A., Kasai, H., Mino, S., Sawabe, T., and Zamri-Saad, M. (2019). Vibriosis in fish: a review on disease development and prevention. *Journal of Aquatic Animal Health*, 31(1), 3-22.
- Ina-Salwany, M. Y., Hishammuddin, H., Zulperi, Z., Salema, M., Karim, M., and Natrah, F. M. I. (2015). Elucidating the probiotic potential of Malaysian *Paenibacillus pabuli* against *Vibrio alginolyticus* in Artemia culture. *Asian Journal of Agricultural Research*, 9(5), 223-236.
- Ismail, S. A., El-Sayed, H. S., and Fayed, B. (2020). Production of prebiotic chitooligosaccharide and its nano/microencapsulation for the production of functional yoghurt. *Carbohydrate polymers*, 234, 115941. <https://doi.org/10.1016/j.carbpol.2020.115941>.
- Jahari, M. A., Mustafa, S., Roslan, M. A. H., Abd Manap, Y., Lamasudin, D. U., and Jamaludin, F. I. (2018). The effects of synbiotics and

- probiotics supplementation on growth performance of red hybrid tilapia, *Oreochromis mossambicus* x *Oreochromis niloticus*. *Journal of Biochemistry, Microbiology and Biotechnology*, 6(1), 5-9.
- Jahari, M. A., Mustafa, S., Roslan, M. A. H., Lamasudin, D. U., and Abd Manap, Y. (2019). Encapsulation of *Lactobacillus plantarum* with mannan and sodium alginate improves its cell production. *Journal of Biochemistry, Microbiology and Biotechnology*, 7(1), 17-22.
- Jahromi, M. F., Liang, J. B., Abdullah, N., Goh, Y. M., Ebrahimi, R., and Shokryazdan, P. (2016). Extraction and characterization of oligosaccharides from palm kernel cake as prebiotic. *BioResources*, 11(1), 674-695.
- Jain, C., Rodriguez-R, L. M., Phillippe, A. M., Konstantinidis, K. T., and Aluru, S. (2018). High throughput ANI analysis of 90K prokaryotic genomes reveals clear species boundaries. *Nature communications*, 9, 5114. <https://doi.org/10.1038/s41467-018-07641-9>.
- James, C., and Natalie, S. (2014). *Microbiology. A laboratory manual*. Pearson Education. Delhi, India.
- Jatmiko, Y. D., Howarth, G. S., and Barton, M. D. (2017, November). Assessment of probiotic properties of lactic acid bacteria isolated from Indonesian naturally fermented milk. In AIP conference proceedings (Vol. 1908, No. 1, p. 050008). AIP Publishing LLC.
- Jia, E., Zheng, X., Cheng, H., Liu, J., Li, X., Jiang, G., Liu, W., and Zhang, D. (2019). Dietary fructooligosaccharide can mitigate the negative effects of immunity on Chinese mitten crab fed a high level of plant protein diet. *Fish and Shellfish Immunology*, 84, 100-107.
- Jiang, Y., Zi, W., Pei, Z., and Liu, S. (2018). Characterization of polysaccharides and their antioxidant properties from *Plumula nelumbinis*. *Saudi Pharmaceutical Journal*, 26(5), 656-664.
- Jimoh, W. A., Kamarudin, M. S., Sulaiman, M. A., and Dauda, A. B. (2019). Assessment of prebiotic potentials in selected leaf meals of high dietary fibre on growth performance, body composition, nutrient utilisation and amylase activities of a tropical commercial carp fingerlings. *Aquaculture Research*, 50(11), 3401-3411.
- Johnson, E. M., Jung, D. Y. G., Jin, D. Y. Y., Jayabalan, D. R., Yang, D. S. H., and Suh, J. W. (2019). Bacteriocins as food preservatives: challenges and emerging horizons. *Critical Reviews in Food Science and Nutrition*, 58, 2743-2767.
- Ju, L., Pan, Z., Zhang, H., Li, Q., Liang, J., Deng, G., Yu, M., and Long, H. (2019). New insights into the origin and evolution of α -amylase

- genes in green plants. *Scientific Reports*, 9, 4929. <https://doi.org/10.1038/s41598-019-41420-w>.
- Julius, O. O. (2023). Climate SMART Best Practices in Aquaculture and Fisheries with Specific Emphasis on Sierra Leone. In *Emerging Sustainable Aquaculture Innovations in Africa* (pp. 459-475). Singapore: Springer Nature Singapore.
- Kalidas, N.R., Saminathan, M., Ismail, I.S., Abas, F., Maity, P., Islam, S.S., Manshoor, N., and Shaari, K. (2017). Structural characterisation and evaluation of prebiotic activity of oil palm kernel cake mannanoligosaccharides. *Food Chemistry*, 234, 348-355.
- Karimi, M., Paknejad, H., Hoseinifar, S. H., Shabani, A., and Mozanzadeh, M. T. (2020). The effects of dietary raffinose on skin mucus immune parameters and protein profile, serum non-specific immune parameters and immune related genes expression in common carp (*Cyprinus carpio* L.). *Aquaculture*, 520, 734525. <https://doi.org/10.1016/j.aquaculture.2019.734525>.
- Karimi, M., Paknejad, H., Hoseinifar, S. H., Shabani, A., and Mozanzadeh, M. T. (2020). The effects of dietary raffinose on skin mucus immune parameters and protein profile, serum non-specific immune parameters and immune related genes expression in common carp (*Cyprinus carpio* L.). *Aquaculture*, 520, 734525. <https://doi.org/10.1016/j.aquaculture.2019.734525>.
- Kautsky, K. (2013). The Labour Revolution (Routledge Revivals). Routledge. London, United Kingdom, (pp. 1-287). Taylor and Francis Group.
- Kavitha, M., Raja, M., and Perumal, P. (2018). Evaluation of probiotic potential of *Bacillus* spp. isolated from the digestive tract of freshwater fish *Labeo calbasu* (Hamilton, 1822). *Aquaculture Reports*, 11, 59–69.
- KEGG (2007). *Bacillus velezensis* FZB42. BlastKOALA. Retrieved from on 13th September 2022 from https://www.genome.jp/kegg-bin/show_organism?org=bay.
- Kewcharoen, W., and Srisapoome, P. (2019). Probiotic effects of *Bacillus* spp. from Pacific white shrimp (*Litopenaeus vannamei*) on water quality and shrimp growth, immune responses, and resistance to *Vibrio parahaemolyticus* (AHPND strains). *Fish and Shellfish Immunology*, 94, 175-189.
- Keysami, M. A., Saad, C. R., Sijam, K., Daud, H. M., and Alimon, A. R. (2007). Effect of *Bacillus subtilis* on growth development and survival of larvae *Macrobrachium rosenbergii* (de Man). *Aquaculture Nutrition*, 13(2), 131-136.

- Khullar, G., Det-udom, R., Prombutar, P., and Prakitchaiwattana, C. (2022). Probiogenomic analysis and safety assessment of *Bacillus* isolates using omics approach in combination with *in-vitro*. *LWT-Food Science and Technology*, 159, 113216. <https://doi.org/10.1016/j.lwt.2022.113216>.
- Kim, J. A., Bayo, J., Cha, J., Choi, Y. J., Jung, M. Y., Kim, D. H., and Kim, Y. (2019). Investigating the probiotic characteristics of four microbial strains with potential application in feed industry. *PLoS One*, 14(6), e0218922. <https://doi.org/10.1371/journal.pone.0218922>.
- Kim, S. S., Kim, K. I., Yoo, H. K., Han, Y. S., Jegal, M. E., Byun, S. G., Lim, H. J., Park, J. S., and Kim, Y. J. (2021). Differential virulence of infectious hematopoietic necrosis virus (IHNV) isolated from salmonid fish in Gangwon Province, Korea. *Fish and Shellfish Immunology*, 119, 490-498.
- Kim, Y. O., Mahboob, S., Viayaraghavan, P., Biji, D., Al-Ghanim, K. A., Al-Misned, F., Ahmed, Z., Kwon, J. T., Na, S. W., and Kim, H. J. (2020). Growth promoting activity of *Penaeus indicus* by secondary metabolite producing probiotic bacterium *Bacillus subtilis* isolated from the shrimp gut. *Journal of King Saud University-Science*, 32(2), 1641-1646.
- Kosasih, W., Pudjiraharti, S., Ratnaningrum, D., and Priatni, S. (2015). Preparation of inulin from dahlia tubers. *Procedia Chemistry*, 16, 190-194.
- Kötzler, M. P., Hancock, S. M., and Withers, S. G. (2014). Glycosidases: functions, families and folds. *Encyclopedia of Life Sciences*. Wiley.
- Kowalczyk, J.E., Benoit, I., and de Vries, R.P. (2014). Regulation of plant biomass utilisation in *Aspergillus*. *Advances in Applied Microbiology*, 88, 31-56.
- Krausova, G., Hyrslova, I., and Hynstova, I. (2019). *In vitro* evaluation of adhesion capacity, hydrophobicity, and auto-aggregation of newly isolated potential probiotic strains. *Fermentation*, 5(4), 100. <https://doi.org/10.3390/fermentation5040100>.
- Krishnaveni, G., Vignesh, S., Vidhyalakshmi, N., Vijay, V., and Ramesh, U. (2021). Effects of dietary supplementation of *Lactobacillus fermentum* URLP18 on growth, innate immunity and survival against *Aeromonas hydrophila* ATCC 7966 challenge in freshwater fish *Cyprinus carpio* (common carp). *Aquaculture Research*, 52(3), 1160-1176.
- Kuebutornye, F. K. A., Abarike, E. D., and Lu, Y. (2019). A review on the application of *Bacillus* as probiotics in aquaculture. *Fish and Shellfish Immunology*, 87, 820-828.

- Kuebutornye, F. K., Lu, Y., Abarike, E. D., Wang, Z., Li, Y., and Sakyi, M. E. (2020). *In vitro* assessment of the probiotic characteristics of three *Bacillus* species from the gut of Nile tilapia, *Oreochromis niloticus*. *Probiotics and Antimicrobial Proteins*, 12, 412-424.
- Kumar, R., Huang, J. Y., Ng, Y. S., Chen, C. Y., and Wang, H. C. (2022). The regulation of shrimp metabolism by the white spot syndrome virus (WSSV). *Reviews in Aquaculture*, 14(3), 1150-1169.
- Lafontaine, G. M. F., Fish, N.M., and Connerton, I. F. (2020). *In vitro* evaluation of the effects of commercial prebiotic GOS and FOS products on human colonic caco-2 cells. *Nutrients*, 12(5), 1281. <https://doi.org/10.3390/nu12051281>.
- Le Roy, K., Lammens, W., Verhaest, M., De Coninck, B., Rabijns, A., Van Laere, A., and Van den Ende, W. (2007). Unraveling the difference between invertases and fructan exohydrolases: a single amino acid (Asp-239) substitution transforms *Arabidopsis* cell wall invertase1 into a fructan 1-exohydrolase. *Plant Physiology*, 145(3), 616-625.
- Lee, J. M., Jang, W. J., Hasan, M. T., Lee, B. J., Kim, K. W., Lim, S. G., Han, H. S., and Kong, I. S. (2018a). Characterisation of a *Bacillus* sp. isolated from fermented food and its symbiotic effect with barley β -glucan as a biocontrol agent in the aquaculture industry. *Applied Microbiology and Biotechnology*, 103(3), 1429-1439.
- Lee, S., Katya, K., Hamidoghli, A., Hong, J., Kim, D. J., and Bai, S. C. (2018c). Synergistic effects of dietary supplementation of *Bacillus subtilis* WB60 and mannanoligosaccharide (MOS) on growth performance, immunity and disease resistance in Japanese eel, *Anguilla japonica*. *Fish and Shellfish Immunology*, 83, 283-291.
- Lee, Y. K., Chew, P. F., Soh, B. S., and Tham, L. Y. (2003). Enhancing phagocytic activity of hemocytes and disease resistance in the prawn *Penaeus merguiensis* by feeding *Spirulina platensis*. *Journal of Applied Phycology*, 15(4), 279–287.
- Letchumanan, V., Loo, K. Y., Law, J. W. F., Wong, S. H., Goh, B. H., Ab Mutalib, N. S., and Lee, L. H. (2019). *Vibrio parahaemolyticus*: the protagonist of foodborne diseases. *Progress In Microbes and Molecular Biology*, 2, 1. <https://doi.org/10.36877/pmmmb.a0000029>.
- Li, J., Wu, Z. B., Zhang, Z., Zha, J. W., Qu, S. Y., Qi, X. Z., Wang, G. X., and Ling, F. (2019). Effects of potential probiotic *Bacillus velezensis* K2 on growth, immunity and resistance to *Vibrio harveyi* infection of hybrid grouper (*Epinephelus lanceolatus*♂ × *E. fuscoguttatus*♀). *Fish and Shellfish Immunology*, 93, 1047-1055.
- Li, M., Seo, S., and Karboune, S. (2015). *Bacillus amyloliquefaciens* levansucrase-catalyzed the synthesis of fructooligosaccharides,

- oligolevan and levan in maple syrup-based reaction systems. *Carbohydrate Polymers*, 133, 203-212.
- Li, Q., Wang, W., Zhu, Y., Chen, Y., Zhang, W., Yu, P., Mao, G., Zhao, T., Feng, W., Yang, L., and Wu, X. (2017). Structural elucidation and antioxidant activity a novel se-polysaccharide from Se-enriched *Grifola frondosa*. *Carbohydrate Polymers*, 161, 42-52.
- Li, R., Tang, N., Jia, X., Nirasawa, S., Bian, X., Zhang, P., and Cheng, Y. (2020). Isolation, physical, structural characterisation and *in vitro* prebiotic activity of a galactomannan extracted from endosperm splits of Chinese Sesbania cannabina seeds. *International Journal of Biological Macromolecules*, 162, 1217-1226.
- Li, X., Gao, X., Zhang, S., Jiang, Z., Yang, H., Liu, X., Jiang, Q., and Zhang, X. (2020a). Characterization of a *Bacillus velezensis* with antibacterial activity and inhibitory effect on common aquatic pathogens. *Aquaculture*, 523, 735165. <https://doi.org/10.1016/j.aquaculture.2020.735165>.
- Li, Y. H., and Wang, H. P. (2017). Advances of genotyping-by-sequencing in fisheries and aquaculture. *Reviews in Fish Biology and Fisheries*, 27(3), 535-559.
- Lim, L. L. (2020). The socioeconomic impacts of COVID-19 in Malaysia: policy review and guidance for protecting the most vulnerable and supporting enterprises. Malaysia, (pp. 104). International Labour Organization.
- Lim, S. M., Yoon, M. Y., Choi, G. J., Choi, Y. H., Jang, K. S., Shin, T. S., Park, H. W., Yu, N. H., Kim, Y. H., and Kim, J. C. (2017). Diffusible and volatile antifungal compounds produced by an antagonistic *Bacillus velezensis* G341 against various phytopathogenic fungi. *The Plant Pathology Journal*, 33(5), 488-498.
- Liu, G., Deng, Y., Ma, H., Jiang, J., Feng, J., and Guo, Z. (2022). Antibacterial characterization of *Bacillus velezensis* LG37 and mining of genes related to biosynthesis of antibacterial substances. *Israeli Journal of Aquaculture-Bamidgeh*, 74, 12. <https://doi.org/10.46989/001c.57533>.
- Liu, H., Wang, S., Cai, Y., Guo, X., Cao, Z., Zhang, Y., Liu, S., Yuan, W., Zhu, W., Zheng, Y., Xie, Z., Guo, W., and Zhou, Y. (2017). Dietary administration of *Bacillus subtilis* HAINUP40 enhances growth, digestive enzyme activities, innate immune responses and disease resistance of tilapia, *Oreochromis niloticus*. *Fish and Shellfish Immunology*, 60, 326-333.
- Liu, J., Cheng, J., Huang, M., Shen, C., Xu, K., Xiao, Y., Pan, W., and Fang, Z. (2021a). Identification of an invertase with high specific activity

for raffinose hydrolysis and its application in soymilk treatment. *Frontiers in Microbiology*, 12, 646801. <https://doi.org/10.3389/fmicb.2021.646801>.

- Liu, J., Shi, L., Ma, X., Jiang, S., Hou, X., Li, P., Ma, T., and Han, B. (2023). Characterization and anti-inflammatory effect of selenium-enriched probiotic *Bacillus amyloliquefaciens* C-1, a potential postbiotics. *Scientific Reports*, 13, 14302. <https://doi.org/10.1038/s41598-023-40988-8>.
- Liu, J., Zhang, X., Zhang, J., Yan, M., Li, D., Zhou, S., Feng, J., and Liu, Y. (2022). Research on extraction, structure characterisation and immunostimulatory activity of cell wall polysaccharides from *Sparassis latifolia*. *Polymers*, 14(3), 549. <https://doi.org/10.3390/polym14030549>.
- Liu, X., Yang, Q., He, Z., and Yao, S. (2021b). Efficacy and safety of inulin supplementation for functional constipation: a systematic review protocol. *BMJ open*, 11(4), e042597. <http://dx.doi.org/10.1136/bmjopen-2020-042597>.
- Liu, Y., Štefanič, P., Miao, Y., Xue, Y., Xun, W., Zhang, N., Shen, Q., Zhang, R., Xu, Z., and Mandic-Mulec, I. (2022). Housekeeping gene *gyrA*, a potential molecular marker for *Bacillus* ecology study. *AMB Express*, 12, 133. <https://doi.org/10.1186/s13568-022-01477-9>.
- Loh, J. Y., Chan, H. K., Yam, H. C., In, L. L. A., and Lim, C. S. Y. (2019). An overview of the immunomodulatory effects exerted by probiotics and prebiotics in grouper fish. *Aquaculture International*, 28, 729-750.
- Lone, S. A., and Manohar, S. (2018). *Saprolegnia parasitica*, a lethal oomycete pathogen: demands to be controlled. *Journal of Infection and Molecular Biology*, 6(2), 36-44.
- Loo, K., Letchumanan, V., Law, J. W., Pusparajah, P., Goh, B., Ab Mutalib, N. S., He, Y. W., and Lee, L. (2020). Incidence of antibiotic resistance in *Vibrio* spp. *Reviews in Aquaculture*, 12, 2590-2608.
- Lordan, C., Thapa, D., Ross, R. P., and Cotter, P. D. (2019). Potential for enriching next generation health-promoting gut bacteria through prebiotics and other dietary components. *Gut Microbes*, 1-20. <https://doi.org/10.1080/19490976.2019.1613124>.
- Lu, E., Yeung, M., and Yeung, C.K. (2018). Comparative analysis of lactulose and fructooligosaccharide on growth kinetics, fermentation, and antioxidant activity of common probiotics. *Food and Nutrition Sciences*, 9(03), 161. <https://doi.org/10.4236/fns.2018.93013>.

- Lu, J., Bu, X., Xiao, S., Lin, Z., Wang, X., Jia, Y., Wang, X., Qin, J. G., and Chen, L. (2019). Effect of single and combined immunostimulants on growth, anti-oxidation activity, non-specific immunity and resistance to *Aeromonas hydrophila* in Chinese mitten crab (*Eriocheir sinensis*). *Fish and Shellfish Immunology*, 93, 732-742.
- Madreseh, S., Ghaisari, H. R., and Hosseinzadeh, S. (2019). Effect of lyophilized, encapsulated *Lactobacillus fermentum* and lactulose feeding on growth performance, heavy metals, and trace element residues in rainbow trout (*Oncorhynchus mykiss*) tissues. *Probiotics and Antimicrobial Proteins*, 11, 1257-1263.
- Magouz, F. I., Bassuini, M. I., Khalafalla, M. M., Abbas, R., Sewilam, H., Aboelenin, S. M., Soliman, M. M., Amer, A. A., Soliman, A. A., Doan, H. V., and Dawood, M. A. (2021). Mannan oligosaccharide enhanced the growth rate, digestive enzyme activity, carcass composition, and blood chemistry of Thinlip grey mullet (*Liza ramada*). *Animals*, 11(12), 3559. <https://doi.org/10.3390/ani11123559>.
- Mahdhi, A., Chakroun, I., Espinosa-Ruiz, C., Messina, C. M., Arena, R., Majdoub, H., Santulli, A., Mzoughi, R., and Esteban, M. A. (2020). Dietary administration effects of exopolysaccharide from potential probiotic strains on immune and antioxidant status and nutritional value of European sea bass (*Dicentrarchus labrax* L.). *Research in Veterinary Science*, 131, 51-58.
- Mamun, M. A. A., Nasren, S., Rathore, S. S., Sidiq, M. J., Dharmakar, P., and Anjusha, K. V. (2019). Assessment of probiotic in aquaculture: functional changes and impact on fish gut. *Microbiology Research Journal International*, 29(1), 1-10. <http://dx.doi.org/10.9734/mrji/2019/v29i130156>.
- Marimuthu, K. (2019). A short review on induced spawning and seed production of African catfish *Clarias gariepinus* in Malaysia. *IOP conference series: Earth and Environmental Science*, 348, 012134. <http://dx.doi.org/10.1088/1755-1315/348/1/012134>.
- Marín-Manzano, M. D. C., Hernandez-Hernandez, O., Diez-Municipio, M., Delgado-Andrade, C., Moreno, F. J., and Clemente, A. (2020). Prebiotic properties of non-fructosylated α -galactooligosaccharides from pea (*Pisum sativum* L.) using infant fecal slurries. *Foods*, 9(7), 921. <https://doi.org/10.3390/foods9070921>.
- Markowiak-Kopeć, P., and Śliżewska, K. (2020). The effect of probiotics on the production of short-chain fatty acids by human intestinal microbiome. *Nutrients*, 12(4), 1107. <https://doi.org/10.3390/nu12041107>.

- Martínez-Núñez, M. A. (2016). Nonribosomal peptides synthetases and their applications in industry. *Sustainable Chemical Processes*, 4, 13. <https://doi.org/10.1186/s40508-016-0057-6>.
- Martins, G. N., Ureta, M. M., Tymczyszyn, E. E., Castilho, P. C., and Gomez-Zavaglia, A. (2019). Technological aspects of the production of fructo and galacto-oligosaccharides. Enzymatic synthesis and hydrolysis. *Frontiers in Nutrition*, 6, 78. <https://doi.org/10.3389/fnut.2019.00078>.
- McFarland, L. V., Evans, C. T., and Goldstein, E. J. (2018). Strain-specificity and disease-specificity of probiotic efficacy: a systematic review and metaanalysis. *Frontiers in Medicine*, 5, 124. <https://doi.org/10.3389/fmed.2018.00124>.
- Medema, M. H., Blin, K., Cimermancic, P., De Jager, V., Zakrzewski, P., Fischbach, M. A., Weber, T., Takano, E., and Breitling, R. (2011). AntiSMASH: rapid identification, annotation and analysis of secondary metabolite biosynthesis gene clusters in bacterial and fungal genome sequences. *Nucleic Acids Research*, 39(2), 339-346.
- Mehrabi, Z., Firouzbakhsh, F., Rahimi-Mianji, G., and Paknejad, H. (2020). Immunity and growth improvement of rainbow trout (*Oncorhynchus mykiss*) fed dietary nettle (*Urtica dioica*) against experimental challenge with *Saprolegnia parasitica*. *Fish and Shellfish Immunology*, 104, 74-82.
- Melkonian, E. A., and Schury, M. P. (2019). Biochemistry, anaerobic glycolysis. Michigan, United States. StatPearls Publishing
- Meng, C., Liu, L. K., Li, D. L., Gao, R. L., Fan, W. W., Wang, K. J., Wang, H. C., and Liu, H. P. (2020). White spot syndrome virus benefits from endosomal trafficking, substantially facilitated by a valosin-containing protein, to escape autophagic elimination and propagate in the crustacean *Cherax quadricarinatus*. *Journal of Virology*, 94(24), e01570-20. <https://doi.org/10.1128/jvi.01570-20>.
- Mensink, M. A., Frijlink, H. W., van der Voort Maarschalk, K., and Hinrichs, W. L. (2015). Inulin, a flexible oligosaccharide I: review of its physicochemical characteristics. *Carbohydrate Polymers*, 130, 405-419.
- Michael, E. T., Amos, S. O., and Hussaini, L. T. (2014). A review on probiotic application in aquaculture. *Fisheries and Aquaculture Journal*, 5(4), 1. <https://doi.org/10.4172/2150-3508.1000111>.
- Miller, G. L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical Chemistry*, 31(3), 426-428.

- Mínguez, C., Webster, P.G., and Villa, M. (2016). Effect of a prebiotic supplementation of mannan oligosaccharide on growth traits and mortality of rainbow trout (*Oncorhynchus mykiss*). *Aquaculture, Aquarium, Conservation and Legislation*, 9(6), 1260-1264.
- Miranda, C. D., Godoy, F. A., and Lee, M. R. (2018). Current status of the use of antibiotics and the antimicrobial resistance in the Chilean salmon farms. *Frontiers in Microbiology*, 9, 1284. <https://doi.org/10.3389/fmicb.2018.01284>.
- Mishra, S. S., Das, R., Choudhary, P., Debbarma, J., and Sahoo, S. N. (2017). Present status of fisheries and impact of emerging diseases of fish and shellfish in Indian aquaculture. *Journal of Aquatic Research and Marine Sciences*, 1, 5-26.
- Mladenović, K. G., Grujović, M. Ž., Nikodijević, D., and Čomić, L. R. (2020). The hydrophobicity of enterobacteria and their co-aggregation with *Enterococcus faecalis* isolated from Serbian cheese. *Bioscience of Microbiota, Food and Health*, 39(4), 227-233.
- Modesto, M., Satti, M., Watanabe, K., Puglisi, E., Morelli, L., Huang, C. H., Liou, J. S., Miyashita, M., Tamura, T., Saito, S., Mori, K., Huang, L., Sciailla, P., Sandri, C., Spiezzio, C., Vitali, F., Cavalieri, D., Perpetuini, G., Tofala, R., Bonetti, A., and Mattarelli, P. (2019). Characterization of *Bifidobacterium* species in feaces of the Egyptian fruit bat: description of *B. vespertilionis* sp. nov. and *B. rousettii* sp. nov. *Systematic and Applied Microbiology*, 42(6), 126017. <https://doi.org/10.1016/j.syapm.2019.126017>.
- Mohamad, A., Zamri-Saad, M., Amal, M. N. A., Al-Saari, N., Monir, M. S., Chin, Y. K., and Md Yasin, I. S. (2021). Vaccine efficacy of a newly developed feed-based whole-cell polyvalent vaccine against vibriosis, streptococcosis and motile aeromonad septicemia in Asian Seabass, *Lates calcarifer*. *Vaccines*, 9(4), 368. <https://doi.org/10.3390/vaccines9040368>.
- Mohamad, N., Manan, H., Sallehuddin, M., Musa, N., and Ikhwanuddin, M. (2020). Screening of Lactic Acid Bacteria isolated from giant freshwater prawn (*Macrobrachium rosenbergii*) as potential probiotics. *Aquaculture Reports*, 18, 100523. <https://doi.org/10.1016/j.aqrep.2020.100523>.
- Mohammadian, T., Jangaran-Nejad, A., Mesbah, M., Shirali, T., Malekpouri, P., and Tabandeh, M. R. (2019). Effect of *Lactobacillus casei* on innate immunity responses and *Aeromonas hydrophila* resistance in shabot, *Tor grypus*. *Probiotics and Antimicrobial Proteins*, 12(1), 224-235.
- Mohd Yazid, S. H., Mohd Daud, H., Azmai, M. N. A., Mohamad, N., and Mohd Nor, N. (2021). Estimating the economic loss due to vibriosis

in net-cage cultured asian seabass (*Lates calcarifer*): evidence from the East Coast of Peninsular Malaysia. *Frontiers in Veterinary Science*, 8, 1111. <https://doi.org/10.3389/fvets.2021.644009>.

- Mohd-Aris, A., Saad, M. Z., Daud, H. M., Yusof, M. T., and Ina-Salwany, M. Y. (2019). *Vibrio harveyi* protease deletion mutant as a live attenuated vaccine candidate against vibriosis and transcriptome profiling following vaccination for *Epinephelus fuscoguttatus*. *Aquaculture International*, 27(1), 125-140.
- Molaei, H., and Jahanbin, K. (2018). Structural features of a new water-soluble polysaccharide from the gum exudates of *Amygdalus scoparia* Spach (Zedo gum). *Carbohydrate Polymers*, 182, 98-105.
- Møller, M., Goh, Y., Viborg, A., Andersen, J., Klaenhammer, T., Svensson, B., and Abou Hachem, M. (2014). Recent insight in α -glucan metabolism in probiotic bacteria. *Biologia*, 69(6), 713-721.
- Mondal, H., Thomas, J., Chandrasekaran, N., and Mukherjee, A. (2023a). Isolation and identification of *Ichthyophonus hoferi* from fishes. In *Aquaculture Microbiology* (pp. 69-72). New York, NY: Springer US.
- Mondal, H., Thomas, J., Chandrasekaran, N., and Mukherjee, A. (2023b). Isolation and identification of *Branchiomyces demigrans* from fishes. In *Aquaculture Microbiology* (pp. 73-76). New York, NY: Springer US.
- Mongkolthanaruk, W. (2012). Classification of *Bacillus* beneficial substances related to plants, humans and animals. *Journal of Microbiology and Biotechnology*, 22(12), 1597-1604.
- Moreno, J., and Peinado, R. (2012). Chapter 6- Sugars: structure and classification. Enological chemistry. Academic Press.
- Mubarak, Z., and Soraya, C. (2018). The acid tolerance response and pH adaptation of *Enterococcus faecalis* in extract of lime *Citrus aurantiifolia* from Aceh Indonesia. *F1000Research*, 7, 287. <https://doi.org/10.12688%2Ff1000research.13990.2>.
- Mudannayake, D. C., Silva, K. F. S. T., Wimalasiri, K. M. S., and Ajlouni, S. (2016). *In vitro* prebiotic properties of partially purified *Asparagus Falcatus* and *Taraxacum Javanicum* inulins. *Journal of Food and Nutritional Disorders*, 5, 6. <https://doi.org/10.4172/2324-9323.1000213>.
- Nagao, T., Adachi, K., Sakai, M., Nishijima, M., and Sano, H. (2001). Novel Macrolactins as antibiotic lactones from a marine bacterium. *The Journal of Antibiotics*, 54(4), 333-339.
- Nair, A. V., Antony, M. L., Praveen, N. K., Sayooj, P., Swaminathan, T. R., and Vijayan, K. K. (2021). Evaluation of *in vitro* and *in vivo* potential

- of *Bacillus subtilis* MBTDCMFRI Ba37 as a candidate probiont in fish health management. *Microbial Pathogenesis*, 152, 104610. <https://doi.org/10.1016/j.micpath.2020.104610>.
- Nakayama, T., Lu, H., and Nomura, N. (2009). Inhibitory effects of *Bacillus* probionts on growth and toxin production of *Vibrio harveyi* pathogens of shrimp. *Letters in Applied Microbiology*, 49(6), 679-684.
- Narmada, S. (2022). Molecular characterization, virulence gene profiling and multidrug resistance profiles of *Aeromonas hydrophila* isolated from raw fish samples collected in and around Tirupati, Andhra Pradesh, India. *The Pharma Innovation Journal*, 11(5), 1669-1673.
- Nazarudin, M. F., Yusoff, F., Idrus, E. S., and Paiko, M. A. (2020). Brown *Sargassum polycystum* as dietary supplement exhibits prebiotic potentials in asian sea bass *Lates calcarifer* fingerlings. *Aquaculture Reports*, 18, 100488. <https://doi.org/10.1016/j.aqrep.2020.100488>.
- Nedaei, S., Noori, A., Valipour, A., Khanipour, A. A., and Hoseinifar, S. H. (2019). Effects of dietary galactooligosaccharide enriched commercial prebiotic on growth performance, innate immune response, stress resistance, intestinal microbiota and digestive enzyme activity in Narrow clawed crayfish (*Astacus leptodactylus* Eschscholtz, 1823). *Aquaculture*, 499, 80-89.
- Nedaei, S., Noori, A., Valipour, A., Khanipour, A. A., and Hoseinifar, S. H. (2023). Dietary effects of *Lactobacillus plantarum* combined with galactooligosaccharide on immunological and biochemical parameters, gut microbiota, digestive enzyme activity, body composition, and stress resistance in narrow-clawed crayfish, *Pontastacus leptodactylus* (eschscholtz, 1823). *Aquaculture Nutrition*, 2023, 3345916. <https://doi.org/10.1155/2023/3345916>.
- Negash, A. W., and Tsehai, B. A. (2020). Current applications of bacteriocin. *International Journal of Microbiology*, 2020, 4374891. <https://doi.org/10.1155/2020/4374891>.
- Nehlah, R., Ina-Salwany, M. Y., and Zulperi, Z. (2016). Antigenicity analysis and molecular characterisation of two outer membrane proteins of *Vibrio alginolyticus* strain VA2 as vaccine candidates in tiger grouper culture. *Journal of Biological Sciences*, 16, 1-11.
- Nguyen, T. L., and Kim, D. H. (2018). Genome-wide comparison reveals a probiotic strain *Lactococcus lactis* WFLU12 isolated from the gastrointestinal tract of olive flounder (*Paralichthys olivaceus*) harboring genes supporting probiotic action. *Marine Drugs*, 16(5), 140. <https://doi.org/10.3390/md16050140>.

- Niccolai, A., Shannon, E., Abu-Ghannam, N., Biondi, N., Rodolfi, L., and Tredici, M. R. (2019). Lactic acid fermentation of *Arthrospira platensis* (Spirulina) biomass for probiotic based products. *Journal of Applied Phycology*, 31, 1077-1083.
- Nourmohammadi, N., Soleimanian-Zad, S., and Shekarchizadeh, H. (2020). Effect of Spirulina (*Arthrospira platensis*) microencapsulated in alginate and whey protein concentrate addition on physicochemical and organoleptic properties of functional stirred yogurt. *Journal of the Science of Food and Agriculture*, 100(14), 5260-5268.
- O'Quin, K. E., Schulte, J. E., Patel, Z., Kahn, N., Naseer, Z., Wang, H., Conte, M. A., and Carleton, K. L. (2012). Evolution of cichlid vision via trans-regulatory divergence. *BMC Evolutionary Biology*, 12, 251. <https://doi.org/10.1186/1471-2148-12-251>.
- Okey, I. B., Gabriel, U. U., and Deekae, S. N. (2018). The use of synbiotics (prebiotic and probiotic) in aquaculture development. *Sumerianz Journal of Biotechnology*, 1(2), 51-60.
- Okochi, M., Sugita, T., Asai, Y., Tanaka, M., and Honda, H. (2017). Screening of peptides associated with adhesion and aggregation of *Lactobacillus rhamnosus* GG *in vitro*. *Biochemical Engineering Journal*, 128, 178-185.
- Oliveira, F. C., Soares, M. P., Oliveira, B. P. N., Pilarski, F., and de Campos, C. M. (2022). Dietary administration of *Bacillus subtilis*, inulin and its synbiotic combination improves growth and mitigates stress in experimentally infected *Pseudoplatystoma reticulatum*. *Aquaculture Research*, 53(12), 4256-4265.
- Ortega, C., Irgang, R., Valladares-Carranza, B., Collarte, C., and Avendaño-Herrera, R. (2020). First identification and characterisation of *Lactococcus garvieae* isolated from rainbow trout (*Oncorhynchus mykiss*) cultured in mexico. *Animals*, 10(9), 1609. <https://doi.org/10.3390/ani10091609>.
- Ortiz, A., and Sansinenea, E. (2020). Macrolactin antibiotics: Amazing natural products. *Mini Reviews in Medicinal Chemistry*, 20(7), 584-600.
- Özcengiz, G., and Öğülür, İ. (2015). Biochemistry, genetics and regulation of Bacilysin biosynthesis and its significance more than an antibiotic. *New Biotechnology*, 32(6), 612-619.
- Pan, X. D., Chen, F. Q., Wu, T. X., Tang, H. G., and Zhao, Z. Y. (2009). Prebiotic oligosaccharides change the concentrations of short-chain fatty acids and the microbial population of mouse bowel. *Journal of Zhejiang University – Science B*, 10, 258-263.

- Panase, A., Thirabunyanon, M., Promya, J., and Chitmanat, C. (2023). Influences of *Bacillus subtilis* and fructooligosaccharide on growth performances, immune responses, and disease resistance of Nile tilapia, *Oreochromis niloticus*. *Frontiers in Veterinary Science*, 9, 1094681. <https://doi.org/10.3389/fvets.2022.1094681>.
- Panigrahi, A., Kiron, V., Puangkaew, J., Kobayashi, T., Satoh, S., and Sugita, H. (2005). The viability of probiotic bacteria as a factor influencing the immune response in rainbow trout *Oncorhynchus mykiss*. *Aquaculture*, 243(1-4), 241-254.
- Pastorino, P., Vela Alonso, A. I., Colussi, S., Cavazza, G., Menconi, V., Mugetti, D., Righetti, M., Barbero, R., Zuccaro, G., Fernandez-Garayzabal, J. F., Dondo, A., Acutis, P. L., and Prearo, M. (2019). A summer mortality outbreak of Lactococciosis by *Lactococcus garvieae* in a raceway system affecting farmed rainbow trout (*Oncorhynchus mykiss*) and brook trout (*Salvelinus fontinalis*). *Animals*, 9(12), 1043. <https://doi.org/10.3390/ani9121043>.
- Patel, P. S., Huang, S., Fisher, S., Pirnik, D., Aklonis, C., Dean, L., Meyers, E., Fernandes, P., and Mayerl, F. (1995). Bacillaene, a novel inhibitor of prokaryotic protein synthesis produced by *Bacillus subtilis*: production, taxonomy, isolation, physico-chemical characterization and biological activity. *The Journal of Antibiotics*, 48(9), 997-1003.
- Pawar, N. A., Prakash, C., Kohli, M. P. S., Jamwal, A., Dalvi, R. S., Devi, B. N., Singh, S. K., Gupta, S., Lende, S. R., Sontakke, S. D., Gupta, S. and Jadhao, S. B. (2023). Fructooligosaccharide and *Bacillus subtilis* symbiotic combination promoted disease resistance, but not growth performance, is additive in fish. *Scientific Reports*, 13, 11345. <https://doi.org/10.1038/s41598-023-38267-7>.
- Pereira, J. Q., Ritter, A. C., Cibulski, S., and Brandelli, A. (2019). Functional genome annotation depicts probiotic properties of *Bacillus velezensis* FTC01. *Gene*, 713, 143971. <https://doi.org/10.1016/j.gene.2019.143971>.
- Phéllippé, M., Gonçalves, O., Thouand, G., Cogne, G., and Laroche, C. (2019). Characterization of the polysaccharides chemical diversity of the cyanobacteria *Arthrospira platensis*. *Algal Research*, 38, 101426. <https://doi.org/10.1016/j.algal.2019.101426>.
- Phengnoin, P., Charoenwongpaiboon, T., Wangpaiboon, K., Klaewkla, M., Nakapong, S., Visessanguan, W., Ito, K., Pichyangkura, R., and Kuttiyawong, K. (2020). Levansucrase from *Bacillus amyloliquefaciens* KK9 and its Y237S variant producing the high bioactive levan-type fructooligosaccharides. *Biomolecules*, 10(5), 692. <https://doi.org/10.3390/biom10050692>.

- Pietrzak, E., Mazurkiewicz, J., and Slawinska, A. (2020). Innate immune responses of skin mucosa in common carp (*Cyprinus carpio*) fed a diet supplemented with galactooligosaccharides. *Animals*, 10(3), 438. <https://doi.org/10.3390/ani10030438>.
- Poblete, E. G., Drakeford, B. M., Ferreira, F. H., Barraza, M. G., and Failler, P. (2019). The impact of trade and markets on Chilean Atlantic salmon farming. *Aquaculture International*, 27, 1465-1483.
- Poolsawat, L., Li, X., Xu, X., Rahman, M. M., Boonpeng, N., and Leng, X. (2021). Dietary xylooligosaccharide improved growth, nutrient utilisation, gut microbiota and disease resistance of tilapia (*Oreochromis niloticus* x *O. aureus*). *Animal Feed Science and Technology*, 275, 114872. <https://doi.org/10.1016/j.anifeedsci.2021.114872>.
- Poolsawat, L., Li, X., Yang, H., Yang, P., Kabir Chowdhury, M. A., Yusuf, A., and Leng, X. (2020). The potentials of fructooligosaccharide on growth, feed utilisation, immune and antioxidant parameters, microbial community and disease resistance of tilapia (*Oreochromis niloticus* x *O. aureus*). *Aquaculture Research*, 51(11), 4430-4442.
- Porras-Domínguez, J. R., Ávila-Fernández, Á., Rodríguez-Alegría, M. E., Miranda-Molina, A., Escalante, A., González-Cervantes, R., Olvera, C., and Munguía, A. L. (2014). Levan-type FOS production using a *Bacillus licheniformis* endolevanase. *Process Biochemistry*, 49(5), 783-790.
- Powthong, P., Jantrapanukorn, B., Suntornthiticharoen, P., and Laohaphatanalert, K. (2020). Study of prebiotic properties of selected banana species in Thailand. *Journal of Food Science and Technology*, 57(7), 2490-2500.
- Preena, P. G., Swaminathan, T. R., Kumar, V. J. R., and Singh, I. S. B. (2020). Antimicrobial resistance in aquaculture: a crisis for concern. *Biologia*, 75, 1497-1517.
- Priyodip, P., and Balaji, S. (2019). A preliminary study on probiotic characteristics of *Sporosarcina* spp. for poultry applications. *Current Microbiology*, 76(4), 448-461.
- Puthiyedathu, S. K., Angel, J. R. J., Thirugnanamurthy, S., Suresh, S., Nathamuni, S., Raja, R. A., Kumar, S., Tomy, S., Dayal, J. S., Paran, B. C., Sawant, P. B., Shashi, S. M., Kumar, C. N., and KonduSamy, A. (2022). Effect of dietary C-Phycocyanin on growth, survival, haematology, immune response, gut microbiome and disease resistance of Pacific white shrimp, *Penaeus vannamei*. *Aquaculture Research*, 53(17), 6292-6309.

- Putri, F. N. (2019). Growth performance of white shrimp *Litopenaeus vannamei* fed with various dosages of prebiotic honey. In *IOP Conference Series: Earth and Environmental Science* (Vol. 278, No. 1, p. 012079). IOP Publishing.
- Qamar, T.R., Syed, F., Nasir, M., Rehman, H., Zahid, M.N., Liu, R.H., and Iqbal, S. (2016). Novel combination of prebiotics galacto-oligosaccharides and inulin-inhibited aberrant crypt foci formation and biomarkers of colon cancer in wistar rats. *Nutrients*, 8(8), 465.
- Ra, S.H., Renchinkhand, G., Park, M.G., Kim, W.S., Paik, S.H., and Nam, M.S. (2018). Hydrolysis of non-digestible components of soybean meal by α -galactosidase from *Bacillus coagulans* NRR1207. *Journal of Life Science*, 28(11), 1347-1353.
- Rahman, M. M., Paul, S. I., Akter, T., Tay, A. C. Y., Foysal, M. J., and Islam, M. T. (2020). Whole genome sequence of *Bacillus subtilis* WS1A, a promising fish probiotic strain isolated from marine sponge of the Bay of Bengal. *Microbiology Resource Announcements*, 9(39), e00641-20. <https://doi.org/10.1128/mra.00641-20>.
- Rahmani, N., Kahar, P., Lisdiyanti, P., Lee, J., Prasetya, B., Ogino, C., and Kondo, A. (2019). GH-10 and GH-11 Endo-1,4- β -xylanase enzymes from *Kitasatospora* sp. produce xylose and xylooligosaccharides from sugarcane bagasse with no xylose inhibition. *Bioresource Technology*, 272, 315-325.
- Rahmatullah, M., Ariff, M., Kahieshesfandiari, M., Daud, H. M., Zamri-Saad, M., Sabri, M. Y., Amal, M. N. A., and Ina Salwany, M. Y. (2017). Isolation and pathogenicity of *Streptococcus iniae* in cultured red hybrid tilapia in Malaysia. *Journal of Aquatic Animal Health*, 29(4), 208-213.
- Rahmi, D., Gustilatov, M., and Utami, D.A.S. (2020). Immune responses and resistance of white shrimp *Litopenaeus vannamei* administered *Bacillus* sp. NP5 and honey against white spot syndrome virus infection. *Jurnal Akuakultur Indonesia*, 19(2), 118-130.
- Rajavel, M., Mitra, A., and Gopal, B. (2009). Role of *Bacillus subtilis* *BacB* in the synthesis of Bacilysin. *Journal of Biological Chemistry*, 284(46), 31882-31892.
- Ramadhani, D. E., Widanarni, W., and Sukenda, S. (2019). Microencapsulation of probiotics and its applications with prebiotic in Pacific white shrimp larvae through *Artemia* sp. *Jurnal Akuakultur Indonesia*, 18(2), 130-140.
- Ray, A. K., Ghosh, K., and Ringø, E. (2012). Enzyme-producing bacteria isolated from fish gut: a review. *Aquaculture Nutrition*, 18(5), 465-492.

- Raza, W., Hussain, Q., and Shen, Q. (2012). Production, regulation and transportation of Bacillibactin in *Bacillus subtilis*. *Journal of the Chemical Society of Pakistan*, 34, 996-1002.
- Reid, G., McGroarty, J. A., Angotti, R., and Cook, R. L. (1988). *Lactobacillus* inhibitor production against *Escherichia coli* and coaggregation ability with uropathogens. *Canadian Journal of Microbiology*, 34(3), 344-351.
- Ricigliano, V. A., and Simone-Finstrom, M. (2020). Nutritional and prebiotic efficacy of the microalga *Arthrospira platensis* (Spirulina) in honeybees. *Apidologie*, 51(5), 898-910.
- Ringø, E., and Song, S. K. (2015). Application of dietary supplements (synbiotics and probiotics in combination with plant products and β glucans) in aquaculture. *Aquaculture Nutrition*, 22(1), 4-24.
- Ringø, E., Doan, H. V., Lee, S. H., Soltani, M., Hoseinifar, S. H., Harikrishnan, R., and Song, S. K. (2020). Probiotics, lactic acid bacteria and bacilli: interesting supplementation for aquaculture. *Journal of Applied Microbiology*, 129, 116-136.
- Ringø, E., Olsen, R. E., Gifstad, T. Ø., Dalmo, R. A., Amlund, H., Hemre, G. I., and Bakke, A. M. (2010). Prebiotics in aquaculture: a review. *Aquaculture Nutrition*, 16(2), 117-136.
- Rodger, H. D. (2016). Fish disease causing economic impact in global aquaculture. In *Fish vaccines* (pp. 1-34). Springer, Basel.
- Rodriguez-Estrada, U., Satoh, S., Haga, Y., Fushimi, H., and Sweetman, J. (2009). Effects of single and combined supplementation of *Enterococcus faecalis*, mannan oligosaccharides and polyhydroxybutyrate acid on growth performance and immune response of rainbow trout, *Oncorhynchus mykiss*. *Aquaculture Science*, 57, 609-617.
- Rodriguez-Estrada, U., Satoh, S., Haga, Y., Fushimi, H., and Sweetman, J. (2013). Effects of inactivated *Enterococcus faecalis* and mannan oligosaccharide and their combination on growth, immunity, and disease protection in rainbow trout. *North American Journal of Aquaculture*, 75(3), 416-428.
- Romano, N., Kanmani, N., Ebrahimi, M., Chong, C. M., Teh, J. C., Hoseinifar, S. H., Amin, S. M. N., Kamarudin, M. S., and Kumar, V. (2018a). Combination of dietary pre gelatinized starch and isomalto oligosaccharides improved pellet characteristics, subsequent feeding efficiencies and physiological status in African catfish, *Clarias gariepinus*, juveniles. *Aquaculture*, 484, 293-302.

Romero, J., Ringø, E., and Merrifield, D. L. (Eds.). (2014). Aquaculture nutrition: gut health, probiotics and prebiotics, (pp. 75-100). John Wiley and Sons.

Rosenberg, M., Gutnick, D., and Rosenberg, E. (1980). Adherence of bacteria to hydrocarbons: a simple method for measuring cell-surface hydrophobicity. *FEMS Microbiology Letters*, 9(1), 29-33.

Ruiz-Moyano, S., Gonçalves dos Santos, M. T. P., Galván, A. I., Merchán, A. V., González, E., Cordoba, M. D. G., and Benito, M. J. (2019). Screening of autochthonous lactic acid bacteria strains from artisanal soft cheese: Probiotic characteristics and prebiotic metabolism. *LWT-Food Science and Technology*, 114, 108388. <https://doi.org/10.1016/j.lwt.2019.108388>.

Sahandi, J., Jafaryan, H., Soltani, M., and Ebrahimi, P. (2019). The use of two *Bifidobacterium* strains enhanced growth performance and nutrient utilisation of Rainbow Trout (*Oncorhynchus mykiss*) fry. *Probiotics and antimicrobial proteins*, 11(3), 966-972.

Sahoo, T. K., Jena, P. K., Patel, A. K., and Seshadri, S. (2014). Bacteriocins and their applications for the treatment of bacterial diseases in aquaculture: a review. *Aquaculture Research*, 47(4), 1013-1027.

Saliba, R., Mizrahi, A., Gauthier, P. D. P., Alban, L. M., Zahar, J. R., and Pilmiss, B. (2022). Antimicrobial stewardship program: reducing antibiotic's spectrum of activity is not the solution to limit the emergence of multidrug-resistant bacteria. *Antibiotics*, 11(1), 70. <https://doi.org/10.3390/antibiotics11010070>.

Salminen, S., Collado, M. C., Endo, A., Hill, C., Lebeer, S., Quigley, E. M., and Vinderola, G. (2021). The International Scientific Association of Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of postbiotics. *Nature Reviews Gastroenterology and Hepatology*, 18(9), 649-667.

Samson, J. S., Choresca, J. C. H., and Quiazon, K. M. A. (2020). Selection and screening of bacteria from African nightcrawler, *Eudrilus eugeniae* (Kinberg, 1867) as potential probiotics in aquaculture. *World Journal of Microbiology and Biotechnology*, 36, 16. <https://doi.org/10.1007/s11274-019-2793-8>.

Sánchez-Ortiz, A. C., Luna-González, A., Campa-Córdova, Á. I., Escamilla-Montes, R., del Carmen Flores-Miranda, M., and Mazón-Suástequi, J. M. (2015). Isolation and characterization of potential probiotic bacteria from pustulose ark (*Anadara tuberculosa*) suitable for shrimp farming. *Latin American Journal of Aquatic Research*, 43(1), 123-136.

- Sanders, M.E., Merenstein, D.J., Reid, G., Gibson, G.R., and Rastall, R.A. (2019). Probiotics and prebiotics in intestinal health and disease: from biology to the clinic. *Nature Reviews Gastroenterology and Hepatology*, 16(10), 605-616.
- Sandoval-Sierra, J. V., Latif-Eugenin, F., Martín, M. P., Zaror, L., and Dieguez-Uribeondo, J. (2014). Saprolegnia species affecting the salmonid aquaculture in Chile and their associations with fish developmental stage. *Aquaculture*, 434, 462-469.
- Sanjaya, Y., and Trinaya, C. (2019, November). Isolation and identification of potential culturable probiotics bacteria from intestine of *Anguilla bicolor*. In Journal of Physics: Conference Series (Vol. 1280, No. 2, p. 022005). IOP Publishing.
- Santos, L., and Ramos, F. (2018). Antimicrobial resistance in aquaculture: current knowledge and alternatives to tackle the problem. *International Journal of Antimicrobial Agents*, 52(2), 135-143.
- Sarkar, P., Raju, V. S., Kuppusamy, G., Rahman, M. A., Elumalai, P., Harikrishnan, R., Arshad, A., and Arockiaraj, J. (2022). Pathogenic fungi affecting fishes through their virulence molecules. *Aquaculture*, 548, 737553. <https://doi.org/10.1016/j.aquaculture.2021.737553>.
- Seemann, T. (2014). Prokka: rapid prokaryotic genome annotation. *Bioinformatics*, 30(14), 2068-2069.
- Senapin, S., Shyam, K. U., Meemetta, W., Rattanarojpong, T., and Dong, H. T. (2018). Inapparent infection cases of tilapia lake virus (TiLV) in farmed tilapia. *Aquaculture*, 487, 51-55.
- Shahcheraghi, S. H., Ayatollahi, J., and Lotf, M. (2015). Applications of *Bacillus subtilis* as an important bacterium in medical sciences and human life. *Tropical Journal of Medical Research*, 18, 1. <http://dx.doi.org/10.4103/1119-0388.152530>.
- Shehata, M. G., El Sohaimy, S. A., El-Sahn, M. A., and Youssef, M. M. (2016). Screening of isolated potential probiotic lactic acid bacteria for cholesterol lowering property and bile salt hydrolase activity. *Annals of Agricultural Sciences*, 61(1), 65-75.
- Shi, L. (2016). Bioactivities, isolation and purification methods of polysaccharides from natural products: a review. *International Journal of Biological Macromolecules*, 92, 37-48.
- Sieiro, C., Areal-Hermida, L., Pichardo-Gallardo, Á., Almuñá-González, R., De Miguel, T., Sánchez, S., Sánchez-Perez, A., and Villa, T. G. (2020). A hundred years of bacteriophages: can phages replace

- antibiotics in agriculture and aquaculture?. *Antibiotics*, 9(8), 493. <https://doi.org/10.3390%2Fantibiotics9080493>.
- Silvério, S.C., Macedo, E.A., Teixeira, J.A., and Rodrigues, L.R. (2018). New β -galactosidase producers with potential for prebiotic synthesis. *Bioresource Technology*, 250, 131-139.
- Sivalingam, K. M. (2020). Isolation, identification and evaluation of *Spirulina platensis* for its effect on seed germination of groundnut (*Arachis hypogaea* L.), Wolaita Sodo, Southern Ethiopia. *Journal of Algal Biomass Utilization*, 11(2), 34-42.
- Soltani, M., Ghosh, K., Hoseinifar, S. H., Kumar, V., Lymbery, A., Roy, S., and Ringø, E. (2019). Genus *Bacillus*, promising probiotics in aquaculture: aquatic animal origin, bio-active components bioremediation and efficacy in fish and shellfish. *Reviews in Fisheries Science and Aquaculture*, 27(3), 331-379.
- Song, R., Li, H., Kang, Z., Zhong, R., Wang, Y., Zhang, Y., Qu, G., and Wang, T. (2021). Surface plasma induced elimination of antibiotic-resistant *Escherichia coli* and resistance genes: antibiotic resistance, horizontal gene transfer, and mechanisms. *Separation and Purification Technology*, 275, 119185. <https://doi.org/10.1016/j.seppur.2021.119185>.
- Song, S. K., Beck, B. R., Kim, D., Park, J., Kim, J., Kim, H. D., and Ringø, E. (2014). Prebiotics as immunostimulants in aquaculture: a review. *Fish and Shellfish Immunology*, 40(1), 40-48.
- Soto, E., Fast, M. D., Purcell, S. L., Coleman, D. D., Yazdi, Z., Kenelty, K., Yun, S., and Camus, A. (2022). Expression of immune markers of white sturgeon (*Acipenser transmontanus*) during *Veronaea botryosa* infection at different temperatures. *Comparative Biochemistry and Physiology Part D: Genomics and Proteomics*, 41, 100950. <https://doi.org/10.1016/j.cbd.2021.100950>.
- Stannek-göbel, L., Stefan P., Thomas B., Claudia B., and Stella M. (2022). "Compositions containing Bacillaene producing bacteria or preparations thereof." U.S. Patent Application 17/429, 905, filed June 2, 2022.
- Stephen, J., Mukherjee, S., Lekshmi, M., and Kumar, S. H. (2023). Diseases and Antimicrobial Use in Aquaculture. In *Handbook on Antimicrobial Resistance: Current Status, Trends in Detection and Mitigation Measures* (pp. 1-23). Singapore: Springer Nature Singapore.
- Stratev, D., and Odeyemi, O. A. (2017). An overview of motile *Aeromonas* septicaemia management. *Aquaculture International*, 25(3), 1095-1105.

- Sun, Y., Wang, G., Peng, K., Huang, Y., Cao, J., Huang, W., Chen, B., and Hu, J. (2019). Effects of dietary xylooligosaccharides on growth performance, immunity and *Vibrio alginolyticus* resistance of juvenile *Litopenaeus vannamei*. *Aquaculture Research*, 50(1), 358-365.
- Sun, Z., Liu, W., Song, Y., Xu, H., Yu, J., Bilige, M., Zhang, H., and Chen, Y. (2015). Population structure of *Lactobacillus helveticus* isolates from naturally fermented dairy products based on multilocus sequence typing. *Journal of Dairy Science*, 98(5), 2962-2972.
- Surachetpong, W., and Sirikanchana, K. (2020). Orthomyxovirosis (tilapia lake virus). In climate change and infectious fish diseases (pp. 142-156). Wallingford, United Kingdom. CABI.
- Suresh, A. (2023). Oral microbial shift induced by probiotic *Bacillus coagulans* along with its clinical perspectives. *Journal of Oral Biology and Craniofacial Research*, 13(3), 398-402.
- Sutriana, A., Hashim, R., Akter, M. N., and Nor, S. A. M. (2018). Galactooligosaccharide and a combination of yeast and β-glucan supplements enhance growth and improve intestinal condition in striped catfish *Pangasianodon hypophthalmus* fed soybean meal diets. *Fisheries Science*, 84(3), 523-533.
- Talagrand-Reboul, E., Jumas-Bilak, E., and Lamy, B. (2017). The social life of *Aeromonas* through biofilm and quorum sensing systems. *Frontiers in Microbiology*, 8, 37. <https://doi.org/10.3389/fmicb.2017.00037>.
- Tan, C. W., Malcolm, T. T. H., Kuan, C. H., Thung, T. Y., Chang, W. S., Loo, Y. Y., Premarathne, J. M. K. J. K., Ramzi, O. B., Norshafawati, M. F. S., Yusralimuna, N., Rukayadi, Y., Nakaguchi, Y., Nishibuchi, M., and Radu, S. (2017). Prevalence and antimicrobial susceptibility of *Vibrio parahaemolyticus* isolated from short mackerels (*Rastrelliger brachysoma*) in Malaysia. *Frontiers in Microbiology*, 8, 1087. <https://doi.org/10.3389/fmicb.2017.01087>.
- Tan, C. W., Rukayadi, Y., Hasan, H., Abdul-Mutalib, N. A., Jambari, N. N., Hara, H., Thung, T. Y., Lee, E., and Radu, S. (2021). Isolation and characterisation of six *Vibrio parahaemolyticus* lytic bacteriophages from seafood samples. *Frontiers in Microbiology*, 12, 616548. <https://doi.org/10.3389/fmicb.2021.616548>.
- Tank, P. R., Vadher, K. H., and Patel, M. P. (2018). Isolation of probiotic bacteria from gastrointestinal tract of pacific white shrimp *Litopenaeus vannamei* and antibacterial activity of probiotic bacteria against *Vibrio* spp. *Journal of Entomology and Zoology Studies*, 6, 974-978.

- Tarrah, A., Pakroo, S., Corich, V., and Giacomini, A. (2020). Whole-genome sequence and comparative genome analysis of *Lactobacillus paracasei* DTA93, a promising probiotic lactic acid bacterium. *Archives of Microbiology*, 202(7), 1997-2003.
- Tepaamornde, S., Chantarasakha, K., Kingcha, Y., Chaiyapechara, S., Phromson, M., Sriariyanun, M., Kirschke, C. P., Huang, L., and Visessanguan, W. (2018). Effects of *Bacillus aryabhattachi* TBRC8450 on vibriosis resistance and immune enhancement in Pacific white shrimp, *Litopenaeus vannamei*. *Fish and Shellfish Immunology*, 86, 4-13.
- Thankappan, B., Ramesh, D., Ramkumar, S., Natarajaseenivasan, K., and Anbarasu, K. (2015). Characterization of *Bacillus* spp. from the gastrointestinal tract of *Labeo rohita*- towards to identify novel probiotics against fish pathogens. *Applied Biochemistry and Biotechnology*, 175, 340-353.
- Tilwani, Y. M., Lakra, A. K., Domdi, L., Jha, N., and Arul, V. (2023). Preparation, physicochemical characterization, and *in vitro* biological properties of selenium nanoparticle synthesized from exopolysaccharide of *Enterococcus faecium* MC-5. *BioNanoScience*, 13(2), 413-425.
- Touraki, M., Karamanlidou, G., Koziotis, M., and Christidis, I. (2013). Antibacterial effect of *Lactococcus lactis* subsp. *lactis* on *Artemia franciscana* nauplii and *Dicentrarchus labrax* larvae against the fish pathogen *Vibrio anguillarum*. *Aquaculture International*, 21, 481-495.
- Toutiaee, S., Mojgani, N., Harzandi, N., Moharrami, M., and Mokhberosafa, L. (2022). *In vitro* probiotic and safety attributes of *Bacillus* spp. isolated from bee bread, honey samples and digestive tract of honeybees *Apis mellifera*. *Letters in Applied Microbiology*, 74(5), 656-665.
- Trabelsi, L., M'sakni, N. H., Ben Ouada, H., Bacha, H., and Roudesli, S. (2009). Partial characterization of extracellular polysaccharides produced by cyanobacterium *Arthrospira platensis*. *Biotechnology and Bioprocess Engineering*, 14, 27-31.
- Truc, L. N. T., Thanh, T. N., Hong, T. T. T., Van, D. P., Tuyet, M. V.T., Trong, N. N., Cong, M. P., Ngoc, D. C., and Quoc, P. T. (2021). Effects of feed mixed with lactic acid bacteria and carbon, nitrogen, phosphorus supplied to the water on the growth and survival rate of white leg shrimp (*Penaeus vannamei*) infected with acute hepatopancreatic necrosis disease caused by *Vibrio parahaemolyticus*. *Biology*, 10(4), 280. <https://doi.org/10.3390/biology10040280>.

- Tsilingiri, K., and Rescigno, M. (2013). Postbiotics: what else?. *Beneficial Microbes*, 4(1), 101-107.
- Tzortzis, G., Jay, A.J., Baillon, M.L.A., Gibson, G.R., and Rastall, R.A. (2003). Synthesis of α -galactooligosaccharides with α -galactosidase from *Lactobacillus reuteri* of canine origin. *Applied Microbiology and Biotechnology*, 63(3), 286-292.
- United States Department of Agriculture. (2018). Guarding against threats to fish health. *AgResearch Magazine*, 66. Retrieved from <https://agresearchmag.ars.usda.gov/2018/jan/health/> on 12 Jun 2023.
- van Pijkeren, J. P., Canchaya, C., Ryan, K. A., Li, Y., Claesson, M. J., Sheil, B., Steidler, L., O'Mahony, L., Fitzgerald, G. F., Sinderen, D. V., and O'Toole, P. W. (2006). Comparative and functional analysis of sortase-dependent proteins in the predicted secretome of *Lactobacillus salivarius* UCC118. *Applied and Environmental Microbiology*, 72(6), 4143-4153.
- Vanittanakom, N., Loeffler, W., Koch, U., and Jung, G. (1986). Fengycin-a novel antifungal lipopeptide antibiotic produced by *Bacillus subtilis* F-29-3. *The Journal of Antibiotics*, 39(7), 888-901.
- Varghese, N. J., Mukherjee, S., Ivanova, N., Konstantinidis, K. T., Mavrommatis, K., Kyripides, N. C., and Pati, A. (2015). Microbial species delineation using whole genome sequences. *Nucleic Acids Research*, 43(14), 6761-6771.
- Vega, R., and Zuniga-Hansen, M. E. (2014). A new mechanism and kinetic model for the enzymatic synthesis of short-chain fructooligosaccharides from sucrose. *Biochemical Engineering Journal*, 82, 158-165.
- Ventura, M., Turroni, F., and van Sinderen, D. (2015). *Bifidobacteria* of the human gut: our special friends. In diet-microbe interactions in the gut (pp. 41-51). Academic Press.
- Vinderola, G., Capellini, B., Villarreal, F., Suárez, V., Quibroni, A., and Reinheimer, J. (2008). Usefulness of a set of simple *in vitro* tests for the screening and identification of probiotic candidate strains for dairy use. *LWT-Food Science and Technology*, 41(9), 1678-1688.
- Wan, D., Wu, Q., and Kuča, K. (2021). Spirulina. In Nutraceuticals (pp. 959-974). Kentucky, United Stated. Academic Press.
- Wan, Y. J., Hong, T., Shi, H. F., Yin, J. Y., Koev, T., Nie, S. P., Gilbert, R. G., and Xie, M. Y. (2021). Probiotic fermentation modifies the structures of pectic polysaccharides from carrot pulp. *Carbohydrate*

- Polymers*, 251, 117116. <https://doi.org/10.1016/j.carbpol.2020.117116>.
- Wang, A., Ran, C., Wang, Y., Zhang, Z., Ding, Q., Yang, Y., Olsen, E. R., Ringo, E., Bindelle, J., and Zhou, Z. (2018). Use of probiotics in aquaculture of China- a review of the past decade. *Fish and Shellfish Immunology*, 86, 734-755.
- Wang, B., Liu, Q., Huang, Y., Yuan, Y., Ma, Q., Du, M., Cai, T., and Cai, Y. (2018). Extraction of polysaccharide from Spirulina and evaluation of its activities. *Evidence-Based Complementary and Alternative Medicine*, 2018, 3425615. <https://doi.org/10.1155/2018/3425615>.
- Wang, J., Zhang, D., Wang, Y., Liu, Z., Liu, L., and Shi, C. (2021). Probiotic effects of the *Bacillus velezensis* GY65 strain in the mandarin fish, *Siniperca chuatsi*. *Aquaculture Reports*, 21, 100902. <https://doi.org/10.1016/j.aqrep.2021.100902>.
- Wang, L. T., Lee, F. L., Tai, C. J., and Kasai, H. (2007). Comparison of *gyrB* gene sequences, 16S rRNA gene sequences and DNA–DNA hybridization in the *Bacillus subtilis* group. *International Journal of Systematic and Evolutionary Microbiology*, 57(8), 1846-1850.
- Wang, X., Farnell, Y.Z., Peebles, E.D., Kiess, A.S., Wamsley, K.G.S., and Zhai, W. (2016). Effects of prebiotics, probiotics, and their combination on growth performance, small intestine morphology, and resident *Lactobacillus* of male broilers. *Poultry Science*, 95(6), 1332-1340.
- Wang, Y. B., Li, J. R., and Lin, J. (2008). Probiotics in aquaculture: challenges and outlook. *Aquaculture*, 281(1), 1-4.
- Wang, Y. X., Xin, Y., Yin, J. Y., Huang, X. J., Wang, J. Q., Hu, J. L., ... and Nie, S. P. (2022). Revealing the architecture and solution properties of polysaccharide fractions from *Macrolepiota albuminosa* (Berk.) Pegler. *Food Chemistry*, 368, 130772. <https://doi.org/10.1016/j.foodchem.2021.130772>.
- Wang, Y., Liu, Y., Yu, H., Zhou, S., Zhang, Z., Wu, D., Yan, M., Tang, Q., and Zhang, J. (2017). Structural characterisation and immuno-enhancing activity of a highly branched water-soluble β -glucan from the spores of *Ganoderma lucidum*. *Carbohydrate Polymers*, 167, 337-344.
- Wang, Y., Wang, J., Bai, D., Wei, Y., Sun, J., Luo, Y., Zhao, J., Liu, Y., and Wang, Q. (2020). Synergistic inhibition mechanism of pediocin PA-1 and L-lactic acid against *Aeromonas hydrophila*. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1862(10), 183346. <https://doi.org/10.1016/j.bbamem.2020.183346>.

- Wang, Y., Zhang, H., Zhang, L., Liu, W., Zhang, Y., Zhang, X., and Sun, T. (2010). *In vitro* assessment of probiotic properties of *Bacillus* isolated from naturally fermented congee from Inner Mongolia of China. *World Journal of Microbiology and Biotechnology*, 26(8), 1369-1377.
- Wanna, W., Surachat, K., Kaitimonchai, P., and Phongdara, A. (2021). Evaluation of probiotic characteristics and whole genome analysis of *Pediococcus pentosaceus* MR001 for use as probiotic bacteria in shrimp aquaculture. *Scientific Reports*, 11, 18334. <https://doi.org/10.1038/s41598-021-96780-z>.
- Wee, W. C., Mok, C. H., Romano, N., Ebrahimi, M., and Natrah, I. (2018). Dietary supplementation use of *Bacillus cereus* as quorum sensing degrader and their effects on growth performance and response of Malaysian giant river prawn *Macrobrachium rosenbergii* juvenile towards *Aeromonas hydrophila*. *Aquaculture Nutrition*, 24(6), 1804-1812.
- Wegh, C. A., Geerlings, S. Y., Knol, J., Roeselers, G., and Belzer, C. (2019). Postbiotics and their potential applications in early life nutrition and beyond. *International Journal of Molecular Sciences*, 20(19), 4673. <https://doi.org/10.3390%2Fijms20194673>.
- Wiercigroch, E., Szafraniec, E., Czamara, K., Pacia, M. Z., Majzner, K., Kochan, K., Kaczor, A., Baranska, M., and Malek, K. (2017). Raman and infrared spectroscopy of carbohydrates: a review. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 185, 317-335.
- Wongputtisin, P., Ramaraj, R., Unpaprom, Y., Kawaree, R., and Pongtrakul, N. (2015). Raffinose family oligosaccharides in seed of Glycine max cv. Chiang Mai60 and potential source of prebiotic substances. *International Journal of Food Science and Technology*, 50(8), 1750-1756.
- Wu, L., Wu, H., Chen, L., Xie, S., Zang, H., Borriss, R., and Gao, X. (2014). Bacilysin from *Bacillus amyloliquefaciens* FZB42 has specific bactericidal activity against harmful algal bloom species. *Applied and Environmental Microbiology*, 80(24), 7512-7520.
- Xi, D. (2011). Application of probiotics and green tea extract in post-harvest processes of Pacific oysters (*Crassostrea gigas*) for reducing *Vibrio parahaemolyticus* and extending shelf life. Master thesis. Oregon State University. Corvallis, Oregon.
- Xiong, Q., Liu, D., Zhang, H., Dong, X., Zhang, G., Liu, Y., and Zhang, R. (2020). Quorum sensing signal autoinducer-2 promotes root colonization of *Bacillus velezensis* SQR9 by affecting biofilm

- formation and motility. *Applied Microbiology and Biotechnology*, 104(16), 7177-7185.
- Xu, H., Su, Y., Zhang, L., Tian, T., Xu, R., Sun, H., ... and Yu, D. (2022). Effects of dietary galactooligosaccharide on growth, antioxidants, immunity, intestinal morphology and disease resistance against *Aeromonas hydrophila* in juvenile hybrid sturgeon (*Acipenser baerii*♀ × *A. schrenckii*♂). *Aquaculture Reports*, 23, 101097. <https://doi.org/10.1016/j.aqrep.2022.101097>.
- Yan, J., Han, Z., Qu, Y., Yao, C., Shen, D., Tai, G., ... and Zhou, Y. (2018). Structure elucidation and immunomodulatory activity of a β-glucan derived from the fruiting bodies of *Amillariella mellea*. *Food Chemistry*, 240, 534-543.
- Yang, K., Zhang, Y., Cai, M., Guan, R., Neng, J., Pi, X., and Sun, P. (2020). *In vitro* prebiotic activities of oligosaccharides from the by-products in *Ganoderma lucidum* spore polysaccharide extraction. *RSC Advances*, 10(25), 14794-14802.
- Yao, L., Wang, C., Li, G., Xie, G., Jia, Y., Wang, W., Liu, S., Xu, T., Luo, K., Zhang, Q., and Kong, J. (2022). Identification of *Fusarium solani* as a causal agent of black spot disease (BSD) of Pacific white shrimp, *Penaeus vannamei*. *Aquaculture*, 548, 737602. <https://doi.org/10.1016/j.aquaculture.2021.737602>.
- Yasmin, I., Saeed, M., Khan, W. A., Khaliq, A., Chughtai, M. F. J., Iqbal, R., Tehseen, S., Naz, S., Liaqat, A., Mehmood, T., Ahsan, S., and Tanweer, S. (2020). *In vitro* probiotic potential and safety evaluation (hemolytic, cytotoxic activity) of *Bifidobacterium* strains isolated from raw camel milk. *Microorganisms*, 8(3), 354. <https://doi.org/10.3390/microorganisms8030354>.
- Yi, Y., Zhang, Z., Zhao, F., Liu, H., Yu, L., Zha, J., and Wang, G. (2018). Probiotic potential of *Bacillus velezensis* JW: antimicrobial activity against fish pathogenic bacteria and immune enhancement effects on *Carassius auratus*. *Fish and Shellfish Immunology*, 78, 322-330.
- Yilmaz, M., Soran, H., and Beyatli, Y. (2006). Antimicrobial activities of some *Bacillus* spp. strains isolated from the soil. *Microbiological Research*, 161(2), 127-131.
- Yukgehraish, K., Kumar, P., Sivachandran, P., Marimuthu, K., Arshad, A., Paray, B. A., and Arockiaraj, J. (2020). Gut microbiota metagenomics in aquaculture: factors influencing gut microbiome and its physiological role in fish. *Reviews in Aquaculture*, 12(3), 1903-1927.

- Zahura, U. A., Chowdhury, M. B. R., and Faruk, M. A. R. (2004). Fungal infection in freshwater fishes of Mymensingh Bangladesh. *Indian Journal of Fisheries*, 51(1), 61-68.
- Zainul, R. (2016). Isolation and molecular identification of freshwater microalgae in Maninjau Lake West Sumatra. *Der Pharmacia Lettre*, 8(20), 177-187.
- Zajicek, P., Corbin, J., Belle, S., and Rheault, R. (2023). Refuting marine aquaculture myths, unfounded criticisms, and assumptions. *Reviews in Fisheries Science and Aquaculture*, 31(1), 1-28.
- Zakaria, Z. H., MohdYaminudin, N. J., Md Yasin, I. S., Mohdlkhsan, N. F. and Abd Karim, M. M. (2019). Evaluation of *Enterobacter* sp. Strain G87 as potential probiont against *Vibrio harveyi* infection in *Artemia Nauplii* and Asian seabass (*Lates calcarifer*) Larvae. *Tropical Agricultural Science*, 42(4), 1251-1262.
- Zaki, M. A., Ashour, M., Heneash, A. M., Mabrouk, M. M., Alprol, A. E., Khairy, H. M., Nour, A. M., Mansour, A. T., Hassanien, H. A., Gaber, A., and Elshobary, M. E. (2021). Potential Applications of native cyanobacterium isolate (*Arthrospira platensis* NIOF17/003) for biodiesel production and utilization of its byproduct in marine rotifer (*Brachionus plicatilis*) production. *Sustainability*, 13(4), 1769. <https://doi.org/10.3390/su13041769>.
- Zanollo, V., Biondi, N., Niccolai, A., Abiusi, F., Adessi, A., Rodolfi, L., and Tredici, M. R. (2022). Protein, phycocyanin, and polysaccharide production by *Arthrospira platensis* grown with LED light in annular photobioreactors. *Journal of Applied Phycology*, 34(3), 1189-1199.
- Zendeboodi, F., Khorshidian, N., Mortazavian, A. M., and da Cruz, A. G. (2020). Probiotic: conceptualisation from a new approach. *Current Opinion in Food Science*, 32, 103-123.
- Zhang, D. F., Xiong, X. L., Wang, Y. J., Gao, Y. X., Ren, Y., Wang, Q., and Shi, C. B. (2021). *Bacillus velezensis* WLYS23 strain possesses antagonistic activity against hybrid snakehead bacterial pathogens. *Journal of Applied Microbiology*, 131(6), 3056-3068.
- Zhang, D. X., Kang, Y. H., Zhan, S., Zhao, Z. L., Jin, S. N., Chen, C., Zhang, L., Shen, J. Y., Wang, C. F., Wang, G. Q., Shan, X. F., and Qian, A. D. (2019). Effect of *Bacillus velezensis* on *Aeromonas veronii*-induced intestinal mucosal barrier function damage and inflammation in crucian carp (*Carassius auratus*). *Frontiers in Microbiology*, 10, 2663. <https://doi.org/10.3389/fmicb.2019.02663>.
- Zhang, D., Qiang, R., Zhou, Z., Pan, Y., Yu, S., Yuan, W., Cheng, J., Wang, J., Zhao, D., Zhu, J., and Yang, Z. (2022). Biocontrol and action mechanism of *Bacillus subtilis* lipopeptides' Fengycins against

- Alternaria solani* in potato as assessed by a transcriptome analysis. *Frontiers in Microbiology*, 13, 861113. <https://doi.org/10.3389/fmicb.2022.861113>.
- Zhang, H., Yohe, T., Huang, L., Entwistle, S., Wu, P., Yang, Z., Busk, P. K., Xu, Y., and Yin, Y. (2018). dbCAN2: a meta server for automated carbohydrate-active enzyme annotation. *Nucleic Acids Research*, 46(W1), W95-W101.
- Zhang, J., Hu, Y., Sun, Q., Li, X., and Sun, L. (2021). An inactivated bivalent vaccine effectively protects turbot (*Scophthalmus maximus*) against *Vibrio anguillarum* and *Vibrio harveyi* infection. *Aquaculture*, 544, 737158. <https://doi.org/10.1016/j.aquaculture.2021.737158>.
- Zhang, L., Hu, Y., Duan, X., Tang, T., Shen, Y., Hu, B., Liu, A., Chen, H., Li, C., and Liu, Y. (2018). Characterisation and antioxidant activities of polysaccharides from thirteen boletus mushrooms. *International Journal of Biological Macromolecules*, 113, 1-7.
- Zhang, S., Yang, J., Henning, S. M., Lee, R., Hsu, M., Grojean, E., Pisegna, R., Ly, A., Heber, D., and Li, Z. (2017). Dietary pomegranate extract and inulin affect gut microbiome differentially in mice fed an obesogenic diet. *Anaerobe*, 48, 184-193.
- Zhang, X., Sun, J., Han, Z., Chen, F., Lv, A., Hu, X., Sun, X., Qi, H., and Guo, Y. (2021). *Vibrio parahaemolyticus* alters the community composition and function of intestinal microbiota in Pacific white shrimp, *Penaeus vannamei*. *Aquaculture*, 544, 737061. <https://doi.org/10.1016/j.aquaculture.2021.737061>.
- Zhang, Z. H., Chen, M., Xie, S. W., Chen, X. Q., Liu, Y. J., Tian, L. X., and Niu, J. (2020). Effects of dietary xylooligosaccharide on growth performance, enzyme activity and immunity of juvenile grass carp, *Ctenopharyngodon idellus*. *Aquaculture Reports*, 18, 100519. <https://doi.org/10.1016/j.aqrep.2020.100519>.
- Zheng, S. C., Xu, J. Y., and Liu, H. P. (2019). Cellular entry of white spot syndrome virus and antiviral immunity mediated by cellular receptors in crustaceans. *Fish and Shellfish Immunology*, 93, 580-588.
- Zhou, L., Li, H., Qin, J. G., Wang, X., Chen, L., Xu, C., and Li, E. (2020). Dietary prebiotic inulin benefits on growth performance, antioxidant capacity, immune response and intestinal microbiota in Pacific white shrimp (*Litopenaeus vannamei*) at low salinity. *Aquaculture*, 518, 734847. <https://doi.org/10.1016/j.aquaculture.2019.734847>.
- Zhou, M., Liu, F., Yang, X., Jin, J., Dong, X., Zeng, K. W., Liu, D., Zhang, Y., Ma, M., and Yang, D. (2018). Bacillibactin and bacillomycin analogues with cytotoxicities against human cancer cell lines from

- marine *Bacillus* sp. PKU-MA00093 and PKU-MA00092. *Marine Drugs*, 16(1), 22. <https://doi.org/10.3390/md16010022>.
- Zhu, J., Ren, S., and Peng, J. (2012). Optimization of polysaccharide extraction from *Spirulina platensis* by cell freeze-thaw cooperated with hot water extraction and deproteinization. *Food Science*, 33, 111-116.
- Zhu, Y. T., Yue, S. M., Li, R. T., Qiu, S. X., Xu, Z. Y., Wu, Y., and Li, Y. (2021). Prebiotics inulin metabolism by lactic acid bacteria from young rabbits. *Frontiers in Veterinary Science*, 8, 719927. <https://doi.org/10.3389/fvets.2021.719927>.
- Zokaeifar, H., Babaei, N., Saad, C. R., Kamarudin, M. S., Sijam, K., and Balcazar, J. L. (2014). Administration of *Bacillus subtilis* strains in the rearing water enhances the water quality, growth performance, immune response, and resistance against *Vibrio harveyi* infection in juvenile white shrimp, *Litopenaeus vannamei*. *Fish and Shellfish Immunology*, 36(1), 68-74.
- Zokaeifar, H., Balcazar, J. L., Saad, C. R., Kamarudin, M. S., Sijam, K., Arshad, A., and Nejat, N. (2012). Effects of *Bacillus subtilis* on the growth performance, digestive enzymes, immune gene expression and disease resistance of white shrimp, *Litopenaeus vannamei*. *Fish and Shellfish Immunology*, 33(4), 683-689.
- Zommara, M., El-Ghaish, S., Haertle, T., Chobert, J. M., and Ghanimah, M. (2023). Probiotic and technological characterization of selected *Lactobacillus* strains isolated from different egyptian cheeses. *BMC microbiology*, 23, 160. <https://doi.org/10.1186/s12866-023-02890-1>.
- Zorriehzahra, M. J., and Banaederakhshan, R. (2015). Early mortality syndrome (EMS) as new emerging threat in shrimp industry. *Advances in Animal and Veterinary Sciences*, 3(2S), 64-72.
- Zorriehzahra, M. J., Delshad, S. T., Adel, M., Tiwari, R., Karthik, K., Dhama, K., and Lazado, C. C. (2016). Probiotics as beneficial microbes in aquaculture: an update on their multiple modes of action: a review. *Veterinary Quarterly*, 36(4), 228-241.
- Zubaidah, A., and Yuhana, M. (2015). Encapsulated synbiotic dietary supplementation at different dosages to prevent vibriosis in white shrimp, *Litopenaeus vannamei*. *HAYATI Journal of Biosciences*, 22(4), 163-168.
- Zulkhairi Amin, F. A., Sabri, S., Ismail, M., Chan, K. W., Ismail, N., Mohd Esa, N., Mohd Lila, M. Z., and Zawawi, N. (2020). Probiotic properties of *Bacillus* strains isolated from stingless bee (*Heterotrigona itama*) honey collected across Malaysia.