



**SIZE SEPARATION OF UNCOATED AND POLYETHYLENE GLYCOL
COATED SILVER NANOPARTICLES USING DENSITY GRADIENT
CENTRIFUGATION**

By

AMIRAH SHAFILLA BINTI MOHAMAD KASIM

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree Master of Science

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May 2021

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Purification of silver nanoparticles (AgNPs) is essential for biomedical field, where high purity and application are difficulty to have compatible size ranges of AgNPs as drug delivery. AgNPs in different size ranges, such as 1-20 nm, 20-40 nm, 40-60 nm, and 60-80 nm, would size uniformity AgNPs are required for drug delivery. Currently, the limitations in biomedical improve drug delivery efficacy. However, AgNPs with a wide range of sizes have disadvantage such as being toxic to health cells. Therefore, surface-bound chemical modification using coating agents such as polyethylene glycol (PEG) and purification procedures are required to generate smaller size ranges of AgNPs, which will also give a better understanding on the role of AgNPs' physical properties in biomedical applications. In this study, uncoated AgNPs were produced via chemical and biological synthesis and then coated with PEG for its stability. UV-Visible spectrophotometer, high resolution-transmission electron microscope (HR-TEM), Fourier-transform infrared spectroscopy (FTIR), and dynamic light scattering (DLS) were used to characterise uncoated and PEG-coated AgNPs from chemical and biological synthesis for size, shape and morphology. The size distribution of PEG-coated AgNPs from both synthesis methods are slightly bigger around 1 – 3 nm compared to the uncoated AgNPs from both synthesis methods. Further, the shape of PEG-coated AgNPs from both synthesis methods revealed predominant spherical shape (86.86%). It showed that PEG-coated AgNPs from both synthesis methods was homogenous in term of size, shape and morphology compared to the uncoated AgNPs from both synthesis methods. In addition, a purification method which is density-gradient centrifugation (DGC) was carried out on the both uncoated and PEG-coated AgNPs from both synthesis methods by using sucrose at concentration ranging from 10% to 50% (w/v). Based on the results from UV-Visible spectrophotometer and HR-TEM, the size ranges of uncoated and PEG-coated AgNPs from both synthesis methods after purification was smaller than the crude one. In comparison to uncoated AgNPs from both synthesis methods, PEG-coated

AgNPs from both synthesis methods obtained desired separated size ranges. Hence, the PEG- coated AgNPs from both synthesis methods are ideal for drug delivery in biomedical applications.



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sebagai memenuhi keperluan untuk Master Sains

**PENGELASAN SAIZ NANOZARAH ARGENTUM YANG TIDAK DILAPIS
DAN YANG DILAPIS DENGAN POLIETILENA GLIKOL MENGGUNAKAN
SENTRIFUGASI KECERUNAN KETUMPATAN**

Oleh

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Penulenan nanopartikel argentum (AgNPs) sangat penting bagi bidang bioperubatan, di mana ketulenan tinggi dan keseragaman saiz AgNPs diperlukan sebagai penghantaran ubat. Pada masa kini, pembatasan dalam aplikasi bioperubatan adalah kesukaran untuk mendapatkan julat saiz AgNPs yang sesuai sebagai penghantaran ubat. AgNPs dalam julat saiz yang berbeza, seperti 1-20 nm, 20-40 nm, 40-60 nm, dan 60-80 nm, akan meningkatkan keberkesanan penghantaran ubat. Walau bagaimanapun, AgNPs dengan pelbagai saiz mempunyai kelemahan seperti toksik pada sel yang sihat. Oleh itu, pengubahsuaian kimia untuk permukaan yang terikat menggunakan agen salutan seperti polietilena glikol (PEG) dan prosedur penulenan diperlukan untuk menghasilkan julat saiz AgNPs yang seragam, yang juga akan memberikan pemahaman yang lebih baik mengenai peranan sifat fizikal AgNPs dalam aplikasi bioperubatan. Dalam kajian ini, AgNPs yang tidak disaluti dihasilkan melalui sintesis kimia dan biologi dan kemudian disaluti dengan PEG untuk kestabilannya. Spektrofotometer, resolusi tinggi mikroskop elektron transmisi (HR-TEM), spektroskopi inframerah transformasi fourier, dan taburan cahaya dinamik (DLS) digunakan untuk menilai sifat AgNPs yang tidak disaluti dan disaluti PEG dari sintesis kimia dan biologi untuk saiz, bentuk, dan morfologi. Taburan saiz AgNPs bersalut PEG dari kedua-dua kaedah sintesis mempunyai saiz yang lebih besar sedikit, sekitar 1 - 3 nm berbanding dengan AgNPs yang tidak disaluti dari kedua-dua kaedah sintesis. Seterusnya, bentuk AgNPs yang disaluti PEG dari kedua-dua kaedah sintesis tersebut menunjukkan bentuk sfera yang dominan (86.86%). Ini menunjukkan bahawa AgNPs yang disaluti PEG dari kedua-dua kaedah sintesis adalah seragam dari segi saiz, bentuk, dan morfologi berbanding AgNPs yang tidak disaluti dari kedua-dua kaedah sintesis. Tambahan pula, kaedah penulenan yang merupakan sentrifugasi kecerun

ketumpatan (DGC) dilakukan pada kedua-dua AgNPs yang tidak disaluti dan disaluti PEG dari kedua-dua kaedah sintesis dengan menggunakan sukrosa pada kepekatan antara 10% hingga 50% (w / v). Berdasarkan keputusan daripada spektrofotometer dan HR-TEM, julat saiz AgNPs yang tidak disaluti dan disaluti PEG dari kedua-dua kaedah sintesis setelah penulenan, lebih kecil daripada yang mentah. Sebagai perbandingan dengan AgNPs yang tidak disaluti dari kedua-dua kaedah sintesis, AgNPs yang disaluti PEG dari kedua-dua kaedah sintesis mempunyai julat saiz yang diinginkan. Maka, AgNPs yang disaluti PEG dari kedua-dua kaedah sintesis sangat sesuai digunakan sebagai penghantaran ubat dalam aplikasi bioperubatan.



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LIST OF ABBREVIATIONS

ρ	Density
γ	Gamma
$^{\circ}\text{C}$	Degree celsius
%	Percentage
Ag+	Silver ion
AFM	Atomic force microscope
AGE	Agarose gel electrophoresis
AgNO ₃	Silver nitrate
AgNPs	Silver nanopartilces
ANVA	Analysis of variance
BET	Brunauer-emmet-teller
CFF	Cross flow-filtration
D_{BET}	Average diameter of spherical nanoparticles
DGC	Density gradient centrifugation
DLS	Dynamic light scattering
DNA	Deoxyribonucleic acid
EDTA	Ethylenediaminetetraacetic acid
FTIR	Fourier transform infrared spectroscopy
g	Gravitaional force
HR-TEM	High resolution transmission electron microscope
kV	Kilovolt
M	Molar
mg	Milligram

mL	Milliliter
μL	Microliter
μm	Micrometer
mm	Millimeter
mV	Millivolt
M	Molar
NaBH ₄	Sodium borohydride
nm	Nanometer
NPs	Nanoparticles
PdI	Polydispersity index
PEG	Polyethylene glycol
ROS	Reactive oxygen species
RZC	Rate zonal centrifugation
<i>S</i>	Specific surface area
S.D	Standard deviation
SEC	Size exclusion chromatography
SPR	Surface plasmon resonance
TBE	Tris-borate-EDTA
TEM	Transmission electron microscope
ZP	Zeta potential
XRD	X-ray diffraction

CHAPTER 1

INTRODUCTION

1.1 Background of study

Nanoparticles (NPs) can be defined differently, depending on the types of materials, fields, and applications (Ng et al., 2013). However, the particles in the size range from 1 nm to 100 nm are generally considered to be NPs (Hosokawa et al., 2012). Additionally, NPs have different physical and chemical properties from their bulk materials; these properties are affording the exploitation of NPs for various applications (Ardani et al., 2017). The NPs with smaller size have larger specific surface area; thus, the total surface area of a particle is inversely proportional to its diameter (Cataxo, 2011; Hasan, 2015). Silver nanoparticles (AgNPs) are one of the important metallic nanoparticles that have received great attention because of their usage in biomedical application (Ahmed et al., 2016). AgNPs have been intensely studied due to their intrinsic unique properties (i.e., optical behaviour, conductivity, chemical stability, and catalytic activity) (Singh et al., 2015).

Two methods, biological and chemical, could be used to synthesise AgNPs. Chemical synthesis of AgNPs are typically generated by chemical reduction and obtained an effective yield; however, they can lead to the limitations (i.e., usage of hazardous chemicals, high cost, and increased energy consumption) (Srikar et al., 2016). AgNPs can also be prepared using biological sources (e.g., plant extract, bacteria, and fungi); this approach is therefore more environmentally-friendly and cost-effective (Ajitha et al., 2014). However, plant extract-based synthesis appears to be the most appealing approach due to the easier subsequent extraction process compared to microbial routes, which require aseptic conditions for cultivation and laborious work in maintaining the cells (Awwad et al., 2013). In addition, the usage of plant extract may also offer many benefits, including high source availability, eco-friendliness, safety to handle, and containing a wide range of plant metabolites (Kulkarni et al., 2011). In addition, the synthesised AgNPs are preferable compared to the commercial AgNPs due to the long-term effects of commercial AgNPs exposure on human physiology and has given high negative impact as their release into the environment (Stensberg et al., 2011).

The biggest challenge associated with AgNPs synthesis is their instability and susceptibility to agglomeration intrinsic characteristics, which are resulting in the formation of larger-size AgNPs (Ardani et al., 2017). Additionally, Sarkar et al (2005) highlighted that polymers (e.g., polysaccharides, polyacrylamide, and PEG) and ligands (e.g., citrates, amines, peptides, and lipids) are widely used as capping agents for the surface modification because these substances can control the rate of reduction of metal ions and the aggregation process of the metal clusters. However, eventhough the synthesised AgNPs from chemical and

biological methods has been coated with polymer, these will still lead to the wide size range and shapes of NPs (Suresh et al., 2015).

Based on the disadvantages of unseparated synthesised AgNPs, it is important to obtain well-separated size and shape of NPs for characterisation and also for biomedical application (Suresh et al., 2015). In previous study, spherical shaped AgNPs with size range 50 – 100 nm were tested towards pathogen bacteria (e.g., *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, and *Enterococcus faecalis*) by agar diffusion method and showed as good potential antibacterial agents (Ivanova et al., 2018). Other researcher has been tested spherical shape AgNPs with size range below 20 nm towards *Lactobacillus salivarius* as potential antibacterial agents, to be further used in dental implantology (Ghiuță and Cristea, 2020).

Purification of NPs into smaller sizes and similar shapes may not only enhance the NPs condition, but also provide ways to recognise which physical properties of NPs can be useful for a drug delivery vector (Robertson et al., 2016). The synthesis of NPs produced polydisperse mixtures, however the potential used of these NPs in biomedical application required monodispersed populations (Akbulut et al., 2012). In order to obtain homogeneous NPs populations, post-synthetic separation methods (i.e., density gradient centrifugation, gel electrophoresis and size exclusion chromatography) is required (Shin et al. 2013).

In this study, the uncoated AgNPs were synthesised by chemical and biological methods and then coated with PEG for its stability. Then, the uncoated and PEG-coated AgNPs from chemical and biological synthesis were characterised in terms of their size, shape, surface charge, and morphology using UV-Visible spectrophotometer, high resolution-transmission electron microscope (HR-TEM), Fourier-transform infrared spectroscopy (FTIR), and dynamic light scattering (DLS). Furthermore, a purification method which is density gradient centrifugation (DGC) was conducted for both uncoated and PEG-coated AgNPs from chemical and biological synthesis to distinguish the size range of AgNPs. Multiple layers of a common solvent which was sucrose in water with different densities in ascending order of its concentration was applied in DGC (Mace et al., 2012). Shin et al. (2013) claimed that DGC was one of the effective purification methods used to separate gold nanoparticles, resulting in small distributions of its diameters, shapes and aggregation state.

Generally, this research aimed at characterising the uncoated and PEG-coated AgNPs from chemical and biological synthesis and to evaluate the feasibility of DGC for the size separation of uncoated and PEG-coated AgNPs from chemical and biological synthesis. Development of size classification technique to enhance the properties of sub-micrometer particles has also been reported (Mori, 2015). Nevertheless, reports on the purification of uncoated and PEG-coated AgNPs from chemical and biological synthesis and the characterisation of homogenous nanoparticles are scarce. It is hypothesised that, DGC has the

ability to separate size of synthesised AgNPs according to the different concentrations of solvent used. The NPs separation was proven by optical confirmation, spectrophotometric and transmission electron microscopy measurements (Suresh et al., 2015).

1.2 Specific objectives

Hence, the specific objectives of this research were:

1. To characterise the uncoated and PEG-coated AgNPS, synthesised by chemical and biological methods.
2. To evaluate the effect of size ranges within the uncoated and PEG-coated AgNPS from different synthesis methods using density gradient centrifugation (DGC).

REFERENCES

- Ahmad, T., Wani, I. A., Manzoor, N., Ahmed, J., & Asiri, A. M. (2013). Biosynthesis, structural characterization and antimicrobial activity of gold and silver nanoparticles. *Colloids and Surfaces B: Biointerfaces*, 107, 227-234.
- Ahmadi, T. S., Wang, Z. L., Green, T. C., Henglein, A., & El-Sayed, M. A. (1996). Shape-controlled synthesis of colloidal platinum nanoparticles. *Science*, 272(5270), 1924-1925.
- Ahmed, S., Ahmad, M., Swami, B. L., & Ikram, S. (2016). A review on plants extract mediated synthesis of silver nanoparticles for antimicrobial applications: a green expertise. *Journal of Advanced Research*, 7(1), 17-28.
- Ajitha, B., Reddy, Y. A. K., & Reddy, P. S. (2014). Biogenic nano-scale silver particles by *Tephrosia purpurea* leaf extract and their inborn antimicrobial activity. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 121, 164-172.
- Akbulut, O., Mace, C. R., Martinez, R. V., Kumar, A. A., Nie, Z., Patton, M. R., & Whitesides, G. M. (2012). Separation of nanoparticles in aqueous multiphase systems through centrifugation. *Nano Letters*, 12(8), 4060-4064.
- Alfadul, S. M., & Elneshwy, A. A. (2010). Use of nanotechnology in food processing, packaging and safety—Review. *African Journal of Food, Agriculture, Nutrition and Development*, 10, 2719–2739.
- Anuj, S. A., & Ishnava, K. B. (2013). Plant mediated synthesis of silver nanoparticles using dried stem powder of *Tinospora cordifolia*, its antibacterial activity and its comparison with antibiotics. *International Journal of Pharmacy and Biological Sciences*, 4, 849-863.
- Ardani, H. K., Imawan, C., Handayani, W., Djuhana, D., Harmoko, A., & Fauzia, V. (2017). Enhancement of the stability of silver nanoparticles synthesized using aqueous extract of *Diospyros discolor* Willd. leaves using polyvinyl alcohol. *Conference Series: Materials Science and Engineering*, 188(1) 012056.
- Asharani, P. V., Low Kah Mun, G., Hande, M. P., & Valiyaveetil, S. (2009). Cytotoxicity and genotoxicity of silver nanoparticles in human cells. *American Chemical Society Nano*, 3(2), 279-290.
- Awwad, A. M., Salem, N. M., & Abdeen, A. O. (2013). Green synthesis of silver nanoparticles using carob leaf extract and its antibacterial activity. *International Journal of Industrial Chemistry*, 4(1), 29.

- Balaji, D. S., Basavaraja, S., Deshpande, R., Mahesh, D. B., Prabhakar, B. K., & Venkataraman, A. (2009). Extracellular biosynthesis of functionalized silver nanoparticles by strains of cladosporium cladosporioides fungus. *Colloids and Surfaces B: Biointerfaces*, 68, 88-92.
- Baraton, M. I. (2002). Surface analysis of semiconducting nanoparticles by FTIR spectroscopy. *Nanocrystalline Metals and Oxides*, 165-187.
- Basavaraja, S., Balaji, S. D., Lagashetty, A., Rajasab, A. H., & Venkataraman, A. (2008). Extracellular biosynthesis of silver nanoparticles using the fungus fusarium semitectum. *Materials Research Bulletin*, 43, 1164-1170.
- Bedi, P., & Kaur, A. (2015). An overview on uses of ZnO NPs. *World Journal of Pharmacy and Pharmaceutical Sciences*, 4(12), 1177-1196.
- Behera, O. (2011). Synthesis and characterization of ZnO NPs of various sizes and applications in biological systems. *Doctoral dissertation*.
- Behera, S., & Debata, A. (2011). Biomedical applications of silver nanoparticles. *Journal of Asian Scientific Research*, 1(1), 27.
- Bhainsa, K. C., & D'souza, S. F. (2006). Extracellular biosynthesis of silver nanoparticles using the fungus aspergillus fumigates. *Colloids and Surfaces B: Biointerfaces*, 47, 160-164.
- Bhatia, S. (2016). NPs types, classification, characterization, fabrication methods and drug delivery applications. *Natural Polymer Drug Delivery Systems*, 33-93.
- Bhattacharjee, S. (2016). DLS and zeta potential—what they are and what they are not. *Journal of Controlled Release*, 235, 337-351.
- Bouwmeester, H., Dekkers, S., Noordam, M. Y., Hagens, W., Bulder, A. S., Heer, P. M., & Sips, A. (2007). *Health Impact of Nanotechnologies in Food Production*, 14.
- Brown, S. D., Utturkar, S. M., Klingeman, D. M., Johnson, C. M., Martin, S. L., Land, M. L., & Pelletier, D. A. (2012). Twenty-one genome sequences from Pseudomonas species and 19 genome sequences from diverse bacteria isolated from the rhizosphere and endosphere of Populus deltoides. *Journal of Bacteriology*, 194, 5991-5993.
- Burduşel, A. C., Gherasim, O., Grumezescu, A. M., Mogoantă, L., Ficai, A., & Andronescu, E. (2018). Biomedical applications of silver nanoparticles: An up-to-date overview. *Nanomaterials*, 8(9), 681.
- Cartaxo, A. L. P. (2011). NPs types and properties—understanding these promising devices in the biomedical area. *Doctoral Dissertation*.

- Chandran, S. P., Chaudhary, M., Pasricha, R., Ahmad, A., & Sastry, M. (2006). Synthesis of gold nanotriangles and silver nanoparticles using Aloe vera plant extract. *Biotechnology Progress*, 22(2), 577-583.
- Chaudhry, Q., & Castle, L. (2011). Food applications of nanotechnologies: an overview of opportunities and challenges for developing countries. *Trends in Food Science and Technology*, 22(11), 595-603.
- Chu H., Kim H. J., Kim J. S., Kim M. S., Yoon B. D., Park H. J., & Kim C. Y. (2012). A nanosized Ag-silica hybrid complex prepared by γ -irradiation activates the defense response in Arabidopsis. *Radiation Physics and Chemistry*, 81, 180–184.
- Chung, S. J., Leonard, J. P., Nettleship, I., Lee, J. K., Soong, Y., Martello, D. V., & Chyu, M. K. (2009). Characterization of ZnO nanoparticle suspension in water: effectiveness of ultrasonic dispersion. *Powder Technology*, 194(1-2), 75-80.
- Coussens, C., & Goldman, L. (2005). Implications of Nanotechnology for Environmental Health Research. *The National Academies Press*.
- Das, S. K., Khan, M. M. R., Guha, A. K., Das, A. K., & Mandal, A. B. (2012). Silver-nanobiohybrid material: synthesis, characterization and application in water purification. *Bioresource Technology*, 124, 495-499.
- Deepak, V., Umamaheshwaran, P. S., Guhan, K., Nanthini, R. A., Krithiga, B., Jaithoon, N. M. H., & Gurunathan, S. (2011). Synthesis of gold and silver nanoparticles using purified URAK. *Colloids and Surfaces B: Biointerfaces*, 86, 353-358.
- Desai, R., Mankad, V., Gupta, S. K., & Jha, P. K. (2012). Size distribution of silver nanoparticles: UV-visible spectroscopic assessment. *Nanoscience and Nanotechnology Letters*, 4(1), 30-34.
- Dowling, A. P. (2004). Development of nanotechnologies. *Materials Today*, 7(12), 30-35.
- Dubey, S. P., Lahtinen, M., & Sillanpää, M. (2010). Tansy fruit mediated greener synthesis of silver and gold nanoparticles. *Process Biochemistry*, 45, 1065-1071.
- Durán, N., Durán, M., De Jesus, M. B., Seabra, A. B., Fávaro, W. J., & Nakazato, G. (2016). Silver nanoparticles: A new view on mechanistic aspects on antimicrobial activity. *Nanomedicine: Nanotechnology, Biology and Medicine*, 12(3), 789-799.
- Elavazhagan, T., & Arunachalam, K. D. (2011). Memecylon edule leaf extract mediated green synthesis of silver and gold nanoparticles. *International Journal of Nanomedicine*, 6, 1265.

- El Badawy, A. M., Scheckel, K. G., Suidan, M., & Tolaymat, T. (2012). The impact of stabilization mechanism on the aggregation kinetics of silver nanoparticles. *Science of the Total Environment*, 429, 325-331.
- El-Nour, K. M. A., Eftaiha, A. A., Al-Warthan, A., & Ammar, R. A. (2010). Synthesis and applications of silver nanoparticles. *Arabian Journal of Chemistry*, 3(3), 135-140.
- Englebienne, P., Hoonacker, A. V., & Verhas, M. (2003). Surface plasmon resonance: Principles, methods and applications in biomedical sciences. *Journal of Spectroscopy*, 17, 255-273.
- Evanoff, D. D., & Chumanov, G. (2005). Synthesis and optical properties of silver nanoparticles and arrays. *ChemPhysChem*, 6(7), 1221-1231.
- Faure, B., Salazar-Alvarez, G., Ahniyaz, A., Villaluenga, I., Berriozabal, G., De Miguel, Y. R., & Bergström, L. (2013). Dispersion and surface functionalization of oxide NPs for transparent photocatalytic and UV-protecting coatings and sunscreens. *Science and Technology of Advanced Materials*, 14(2), 023001.
- Firdhouse, M. J., & Lalitha, P. (2012). Green synthesis of silver nanoparticles using the aqueous extract of *Portulaca oleracea* (L.). *Asian Journal of Pharmaceutical and Clinical Research*, 6(1), 92-94.
- Garcia Esparza, A. T. (2011). Size controlled synthesis of transition metal nanoparticles for catalytic applications. *Doctoral Dissertation*.
- Geetha, N., Geetha, T. S., Manonmani, P., & Thiyagarajan, M. (2014). Green synthesis of silver nanoparticles using *Cymbopogon Citratus* (Dc) Stapf. Extract and its antibacterial activity. *Australian Journal of Basic and Applied Sciences*, 8(3), 324-31.
- Geethalakshmi, R., & Sarada, D. V. L. (2010). Synthesis of plant-mediated silver nanoparticles using *Trianthema decandra* extract and evaluation of their antimicrobial activities. *International Journal of Engineering Science and Technology*, 2(5), 970-975.
- Ghiuță, I., & Cristea, D. (2020). Silver nanoparticles for delivery purposes. *Nanoengineered Biomaterials for Advanced Drug Delivery*, 347.
- Ghormade, V., Deshpande, M. V., & Paknikar, K. M. (2011). Perspectives for nano-biotechnology enabled protection and nutrition of plants. *Biotechnology Advances*, 29(6), 792-803.
- Ghosh, S., Patil, S., Ahire, M., Kitture, R., Kale, S., Pardesi, K., & Chopade, B. A. (2012). Synthesis of silver nanoparticles using *Dioscorea bulbifera* tuber extract and evaluation of its synergistic potential in combination with antimicrobial agents. *International Journal of Nanomedicine*, 7, 483.

- Gnanajobitha, G., Paulkumar, K., Vanaja, M., Rajeshkumar, S., Malarkodi, C., Annadurai, G., & Kannan, C. (2013). Fruit-mediated synthesis of silver nanoparticles using *Vitis vinifera* and evaluation of their antimicrobial efficacy. *Journal of Nanostructure in Chemistry*, 3(1), 67.
- Gogoi, S. J. (2013). Green synthesis of silver nanoparticles from leaves extract of ethnomedicinal plants-*Pogostemon benghalensis* (B) O. Ktz. *Advances in Science and Research*, 4, 274-278.
- Gondwal, M., & Pant, G. J. N. (2013). Biological evaluation and green synthesis of silver nanoparticles using aqueous extract of *Calotropis procera*. *International Journal of Pharmaceutical and Biology Sciences*, 4(4).
- Govindaraju, K., Kiruthiga, V., Kumar, V. G., & Singaravelu, G. (2009). Extracellular synthesis of silver nanoparticles by a marine alga, *Sargassum wightii* Grevilli and their antibacterial effects. *Journal of Nanoscience and Nanotechnology*, 9(9), 5497-5501.
- Guarrotxena, N., & Braun, G. (2012). Ag-nanoparticle fractionation by low melting point agarose gel electrophoresis. *Journal of Nanoparticle Research*, 14(10), 1199.
- Gurunathan, S., Han, J. W., Kwon, D. N., & Kim, J. H. (2014). Enhanced antibacterial and anti-biofilm activities of silver nanoparticles against Gram-negative and Gram-positive bacteria. *Nanoscale Research Letters*, 9(1), 373.
- Hahm, J. I., & Lieber, C. M. (2004). Direct ultrasensitive electrical detection of DNA and DNA sequence variations using nanowire nanosensors. *Nano Letters*, 4(1), 51-54.
- Hamouda, R. A., Yousuf, W. E., Abdeen, E. E., & Mohamed, A. (2019). Biological and Chemical Synthesis of Silver Nanoparticles: Characterization, MIC and Antibacterial Activity against Pathogenic Bacteria. *Journal of Chemical and Pharmaceutical Research*, 11, 1–12.
- Hanauer, M., Pierrat, S., Zins, I., Lotz, A., & Sönnichsen, C. (2007). Separation of NPs by gel electrophoresis according to size and shape. *Nano Letters*, 7(9), 2881-2885.
- Hancock, J. M. (2013). Formation and analysis of ZnO NPs and ZnO hexagonal prisms and optical analysis of cadmium selenide NPs. *Doctoral Dissertation*.
- Hasan, S. (2015). A review on NPs: their synthesis and types. *Research Journal of Recent Sciences*, 2277, 2502.
- Hasenoehrl, C., Alexander, C. M., Azzarelli, N. N., & Dabrowiak, J. C. (2012). Enhanced detection of gold NPs in agarose gel electrophoresis. *Electrophoresis*, 33(8), 1251-1254.

- Hosokawa, M., Nogi, K., Naito, M., & Yokoyama, T. (2012). Structural control of nanoparticles. *Nanoparticle Technology Handbook (Second Edition)*, 51-112.
- Huang, Y., Chen, S., Bing, X., Gao, C., Wang, T., & Yuan, B. (2011). Nanosilver migrated into food-simulating solutions from commercially available food fresh containers. *Packaging Technology and Science*, 24(5), 291-297.
- Ingle, A., Rai, M., Gade, A., & Bawaskar, M. (2009). *Fusarium solani*: A novel biological agent for the extracellular synthesis of silver nanoparticles. *Journal of Nanoparticle Research*, 11, 2079-2085.
- Ivanova, N., Gugleva, V., Dobрева, M., Pehlivanov, I., Stefanov, S., & Andonova, V. (2018). Silver nanoparticles as multi-functional drug delivery systems. *Nanomedicines*.
- Jimenez, M. S., Luque-Alled, J. M., Gomez, T., & Castillo, J. R. (2016). Evaluation of agarose gel electrophoresis for characterization of silver NPs in industrial products. *Electrophoresis*, 37(10), 1376-1383.
- Jo, Y. K., Kim, B. H., & Jung, G. (2009). Antifungal activity of silver ions and nanoparticles on phytopathogenic fungi. *Plant Disease*, 93(10), 1037-1043.
- Kalimuthu, K., Babu, R. S., Venkataraman, D., Bilal, M., & Gurunathan, S. (2008). Biosynthesis of silver nanocrystals by bacillus licheniformis. *Colloids and Surfaces B: Biointerfaces*, 65, 150-153.
- Kalishwaralal, K., Deepak, V., Pandian, S. R. K., Kottaisamy, M., Barathmanikant, S., Kartikeyan, B., & Gurunathan, S. (2010). Biosynthesis of silver and gold nanoparticles using *brevibacterium casei*. *Colloids and Surfaces B: Biointerfaces*, 77, 257-262.
- Kannan, R. R. R., Strik, W. A., & Staden, V. (2013). Synthesis of silver nanoparticles using the seaweed *codium capitatum* P.C. Silva (Chlorophyceae). *South African Journal of Botany*, 86, 1-4.
- Karakoti, A. S., Das, S., Thevuthasan, S., & Seal, S. (2011). PEGylated inorganic nanoparticles. *Angewandte Chemie International Edition*, 50(9), 1980-1994.
- Kasprowicz, M. J., Koziół, M., & Gorczyca, A. (2010). The effect of silver nanoparticles on phytopathogenic spores of *Fusarium culmorum*. *Canadian Journal of Microbiology*, 56(3), 247-253.
- Kathiraven, T., Sundaramanickam, A., Shanmugam, N., & Balasubramanian, T. (2015). Green synthesis of silver nanoparticles using marine algae *Caulerpa racemosa* and their antibacterial activity against some human pathogens. *Applied Nanoscience*, 5(4), 499-504.

- Kathiresan, K., Manivannan, S., Nabeel, M. A., & Dhivya, B. (2009). Studies on silver nanoparticles synthesized by marine fungus, penicillium fellutanum isolated from coastal mangrove sediment. *Colloids and Surfaces B: Biointerfaces*, 71, 133-137.
- Kaviya, S., Santhanalakshmi, J., & Viswanathan, B. (2011). Green synthesis of silver nanoparticles using Polyalthia longifolia leaf extract along with D-sorbitol: study of antibacterial activity. *Journal of Nanotechnology*.
- Kesharwani, J., Yoon, K. Y., Hwang, J., & Rai, M. (2009). Phytofabrication of silver nanoparticles by leaf extract of Datura metel: hypothetical mechanism involved in synthesis. *Journal of Bionanoscience*, 3(1), 39-44.
- Kim, S. W., Jung, J. H., Lamsal, K., Kim, Y. S., Min, J. S., & Lee, Y. S. (2012). Antifungal effects of silver nanoparticles (AgNPs) against various plant pathogenic fungi. *Mycobiology*, 40(1), 53-58.
- Kim, S. W., Kim, K. S., Lamsal, K., Kim, Y. J., Kim, S. B., Jung, M., & Lee, Y. S. (2009). An in vitro study of the antifungal effect of silver nanoparticles on oak wilt pathogen Raffaelea sp. *Journal of Microbiology and Biotechnology*, 19(8), 760-764.
- Kittler, S., Greulich, C., Diendorf, J., Koller, M., & Epple, M. (2010). Toxicity of silver nanoparticles increases during storage because of slow dissolution under release of silver ions. *Chemistry of Materials*, 22(16), 4548-4554.
- Knoppe, S., Boudon, J., Dolamic, I., Dass, A., & Burgi, T. (2011). Size exclusion chromatography for semipreparative scale separation of Au₃₈ (SR) 24 and Au₄₀ (SR) 24 and larger clusters. *Analytical Chemistry*, 83(13), 5056-5061.
- Kowalczyk, B., Lagzi, I., & Grzybowski, B. A. (2011). Nanoseparations: strategies for size and/or shape-selective purification of NPs. *Current Opinion in Colloid and Interface Science*, 16(2), 135-148.
- Krishnaraj, C., Jagan, E.G., Rajasekar, S., Selvakumar, P., Kalaichelvan, P.T., & Mohan, N. (2010). Synthesis of silver nanoparticles using *Acypha indica* extracts and its antibacterial activity against water borne pathogens. *Colloids and Surfaces B: Biointerfaces*, 76, 50-56.
- Kuang, Y., Song, S., Liu, X., Li, M., Cai, Z., Luo, L., & Sun, X. (2014). Solvent switching and purification of colloidal nanoparticles through water/oil Interfaces within a density gradient. *Nano Research*, 7(11), 1670-1679.
- Kulkarni, A. P., Srivastava, A. A., Harpale, P. M., & Zunjarrao, R. S. (2011). Plant mediated synthesis of silver nanoparticles-tapping the unexploited sources. *Journal of Natural Product and Plant Resources*, 1(4), 100-107.

- Kumar, S. A., Peter, Y. A., & Nadeau, J. L. (2008). Facile biosynthesis, separation and conjugation of gold nanoparticles to doxorubicin. *Nanotechnology*, 19(49), 495101.
- Kumar, S., & Barth, A. (2010). Following enzyme activity with infrared spectroscopy. *Sensors*, 10(4), 2626-2637.
- Kumar, S., Daimary, R. M., Swargiary, M., Brahma, A., Kumar, S., & Singh, M. (2013). Biosynthesis of silver nanoparticles using Premna herbacea leaf extract and evaluation of its antimicrobial activity against bacteria causing dysentery. *International Journal of Pharmacy and Biological Sciences*, 4(4), 378-84.
- Kumar, P. V., Pammi, S. V. N., Kollu, P., Satyanarayana, K. V. V., & Shameem, U. (2014). Green synthesis and characterization of silver nanoparticles using Boerhaavia diffusa plant extract and their anti-bacterial activity. *Industrial Crops and Products*, 52, 562-566.
- Kumar, V., & Yadav, S. K. (2011). Synthesis of Stable, Polyshaped Silver, and Gold Nanoparticles Using Leaf Extract of Lonicera japonica L. *International Journal of Green Nanotechnology*, 3(4), 281-291.
- Kvítek, L., Panáček, A., Soukupova, J., Kolář, M., Večeřová, R., Pucek, R., & Zbořil, R. (2008). Effect of surfactants and polymers on stability and antibacterial activity of silver nanoparticles (NPs). *The Journal of Physical Chemistry C*, 112(15), 5825-5834.
- Lamsa, K., Kim, S. W., Jung, J. H., Kim, Y. S., Kim, K. S., & Lee, Y. S. (2011). Inhibition effects of silver nanoparticles against powdery mildews on cucumber and pumpkin. *Mycobiology*, 39(1), 26-32.
- Liu, J., He, S., Zhang, Z., Cao, J., Lv, P., He, S., & Joyce, D. C. (2009). Nano-silver pulse treatments inhibit stem-end bacteria on cut gerbera cv. Ruikou flowers. *Postharvest Biology and Technology*, 54(1), 59-62.
- Liu, J. F., Yu, S. J., Yin, Y. G., & Chao, J. B. (2012). Methods for separation, identification, characterization and quantification of silver nanoparticles. *Trends in Analytical Chemistry*, 33, 95-106.
- Mace, C. R., Akbulut, O., Kumar, A. A., Shapiro, N. D., Derda, R., Patton, M. R., & Whitesides, G. M. (2012). Aqueous multiphase systems of polymers and surfactants provide self-assembling step-gradients in density. *Journal of the American Chemical Society*, 134(22), 9094-9097.
- Manno, D., Filippo, E., Di Giulio, M., & Serra, A. (2008). Synthesis and characterization of starch-stabilized Ag nanostructures for sensors applications. *Journal of Non-Crystalline Solids*, 354(52-54), 5515-5520.

- Mariselvam, R., Ranjitsingh, A. J. A., Nanthini, A. U. R., Kalirajan, K., Padmalatha, C., & Selvakumar, P. M. (2014). Green synthesis of silver nanoparticles from the extract of the inflorescence of *Cocos nucifera* (Family: Arecaceae) for enhanced antibacterial activity. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 129, 537-541.
- Marsalek, R. (2014). Particle size and zeta potential of ZnO. *APCBEE Procedia*, 9, 13-17.
- Masurkar, S. A., Chaudhari, P. R., Shidore, V. B., & Kamble, S. P. (2011). Rapid biosynthesis of silver nanoparticles using *Cymbopogon citratus* (lemongrass) and its antimicrobial activity. *Nano-Micro Letters*, 3(3), 189-194.
- McGillicuddy, E., Murray, I., Kavanagh, S., Morrison, L., Fogarty, A., Cormican, M., & Morris, D. (2017). Silver nanoparticles in the environment: Sources, detection and ecotoxicology. *Science of the Total Environment*, 575, 231-246.
- McNeil, S. E. (2005). Nanotechnology for the biologist. *Journal of Leukocyte Biology*, 78(3), 585-594.
- Mehta, N., Basu, S., & Kumar, A. (2016). Separation of zinc oxide nanoparticles in water stream by membrane filtration. *Journal of Water Reuse and Desalination*, 6(1), 148-155.
- Mekhamer, W. K. (2010). The colloidal stability of raw bentonite deformed mechanically by ultrasound. *Journal of Saudi Chemical Society*, 14(3), 301-306.
- Min, J. S., Kim, K. S., Kim, S. W., Jung, J. H., Lamsal, K., Kim, S. B., & Lee, Y. S. (2009). Effects of colloidal silver nanoparticles on sclerotium-forming phytopathogenic fungi. *The Plant Pathology Journal*, 25(4), 376-380.
- Mishra, S., Shimpi, N. G., & Sen, T. (2013). The effect of PEG encapsulated silver nanoparticles on the thermal and electrical property of sonochemically synthesized polyaniline/silver nanocomposite. *Journal of Polymer Research*, 20(1), 49.
- Mishra, S., & Singh, H. B. (2015). Biosynthesized silver nanoparticles as a nanoweapon against phytopathogens: exploring their scope and potential in agriculture. *Applied Microbiology and Biotechnology*, 99(3), 1097-1107.
- Mohd Yusof, H., Rahman, A., Mohamad, R., & Zaidan, U. H. (2020). Microbial Mediated Synthesis of Silver Nanoparticles by *Lactobacillus Plantarum* TA4 and its Antibacterial and Antioxidant Activity. *Applied Sciences*, 10, 6973.

- Mondal, N. K., Chowdhury, A., Dey, U., Mukhopadhyaya, P., Chatterjee, S., Das, K., & Datta, J. K. (2014). Green synthesis of silver nanoparticles and its application for mosquito control. *Asian Pacific Journal of Tropical Disease*, 4, 204-210.
- Mondal, S., Roy, N., Laskar, R. A., Sk, I., Basu, S., Mandal, D., & Begum, N. A. (2011). Biogenic synthesis of Ag, Au and bimetallic Au/Ag alloy nanoparticles using aqueous extract of mahogany (*Swietenia mahogani* JACQ.) leaves. *Colloids and Surfaces B: Biointerfaces*, 82(2), 497-504.
- Mori, Y. (2015). Size-selective separation techniques for nanoparticles in liquid. *KONA Powder and Particle Journal*, 32, 102-114.
- Moussa, S. H., Tayel, A. A., Alsohim, A. S., & Abdallah, R. R. (2013). Botryticidal activity of nanosized silver-chitosan composite and its application for the control of gray mold in strawberry. *Journal of Food Science*, 78(10), 1589-1594.
- Mukherjee, P., Ahmad, A., Mandal, D.D., Senapati, S., Sainkar, S.R., Khan, M.I., Parishcha, R., Ajaykumar, P.V., Alam, M., Kumar, R., & Sastry, M. (2001). Fungus-Mediated Synthesis of Silver Nanoparticles and Their Immobilization in the Mycelial Matrix: A Novel Biological Approach to Nanoparticle Synthesis. *Nano Letters*, 1, 515-519.
- Mukunthan, K. S., Elumalai, E. K., Patel, T. N., & Murty, V. R. (2011). *Catharanthus roseus*: a natural source for the synthesis of silver nanoparticles. *Asian Pacific Journal of Tropical Biomedicine*, 1(4), 270-274.
- Muzamil, M., Khalid, N., Aziz, M. D., & Abbas, S. A. (2014). Synthesis of silver nanoparticles by silver salt reduction and its characterization. *Conference Series: Materials Science and Engineering*, 60(1), 012034.
- Nabikhan, A., Kandasamy, K., Raj, A., & Alikunhi, N. M. (2010). Synthesis of antimicrobial silver nanoparticles by callus and leaf extracts from saltmarsh plant, *Sesuvium portulacastrum* L. *Colloids and Surfaces B: Biointerfaces*, 79(2), 488-493.
- Nabiyouni, G., Barati, A., & Saadat, M. (2011). Surface adsorption of polyethylene glycol and polyvinyl alcohol with variable molecular weights on zinc oxide nanoparticles. *Iranian Journal of Chemical Engineering*, 8(1), 20-30.
- Nair, R., Varghese, S. H., Nair, B. G., Maekawa, T., Yoshida, Y., & Kumar, D. S. (2010). Nanoparticulate material delivery to plants. *Plant Science*, 179(3), 154-163.
- Nakkala, J. R., Mata, R., Gupta, A. K., & Sadras, S. R. (2014). Biological activities of green silver nanoparticles synthesized with *Acorous calamus* rhizome extract. *European Journal of Medicinal Chemistry*, 85, 784-794.

- Nanda, A., & Saravanan, M. (2009). Biosynthesis of silver nanoparticles from *Staphylococcus aureus* and its antimicrobial activity against MRSA and MRSE. *Nanomedicine: Nanotechnology, Biology and Medicine*, 5(4), 452-456.
- Narayanan, K. B., & Park, H. H. (2014). Antifungal activity of silver nanoparticles synthesized using turnip leaf extract (*Brassica rapa* L.) against wood rotting pathogens. *European Journal of Plant Pathology*, 140(2), 185-192.
- Natarajan, R. K., John Nayagam, A. A., Gurunagarajan, S., Muthukumar, Ekambaram, N., & Manimaran, A. (2013). *Elaeagnus indica* mediated green synthesis of silver nanoparticles and its potent toxicity against human pathogens. *Global Journal of Pharmacology*, 7, 222-231.
- Naveen, K. S. H., Kumar, G., Karthik, L., & Rao, K. V. B. (2010). Extracellular biosynthesis of silver nanoparticles using the filamentous fungus *penicillium* sp. *Archives of Applied Science Research*, 2, 161-167.
- Ng, L. Y., Mohammad, A. W., Leo, C. P., & Hilal, N. (2013). Polymeric membranes incorporated with metal/metal oxide nanoparticles: a comprehensive review. *Desalination*, 308, 15–33.
- Novak, J. P., Nickerson, C., Franzen, S., & Feldheim, D. L. (2001). Purification of molecularly bridged metal nanoparticle arrays by centrifugation and size exclusion chromatography. *Analytical chemistry*, 73(23), 5758-5761.
- Otsuka, H., Nagasaki, Y., & Kataoka, K. (2003). PEG-coated nanoparticles for biological and pharmaceutical applications. *Advanced Drug Delivery Reviews*, 55(3), 403-419.
- Pal, A., Shah, S., & Devi, S. (2009). Microwave-assisted synthesis of silver nanoparticles using ethanol as a reducing agent. *Materials Chemistry and Physics*, 114(2-3), 530-532.
- Panáček, A., Kolář, M., Večeřová, R., Pucek, R., Soukupova, J., Kryštof, V., & Kvítek, L. (2009). Antifungal activity of silver nanoparticles against *Candida* spp. *Biomaterials*, 30(31), 6333-6340.
- Panigrahi, T. (2013). Synthesis and characterization of silver nanoparticles using leaf extract of *Azadirachta indica*, *Doctoral Dissertation*.
- Parak, W. J., Pellegrino, T., Micheel, C. M., Gerion, D., Williams, S. C., & Alivisatos, A. P. (2003). Conformation of oligonucleotides attached to gold nanocrystals probed by gel electrophoresis. *Nano Letters*, 3(1), 33-36.
- Park H. J., Kim S. H., Kim S. J., & Choi S. H. (2006). A new composition of nanosized silica-silver for control of various plant diseases. *The Plant Pathology Journal*, 22, 295–302.

- Perni, S., Hakala, V., & Prokopovich, P. (2014). Biogenic synthesis of antimicrobial silver nanoparticles capped with L-cysteine. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 460, 219-224.
- Perrault, S. D., & Chan, W. C. (2009). Synthesis and surface modification of highly monodispersed, spherical gold nanoparticles of 50– 200 nm. *Journal of the American Chemical Society*, 131(47), 17042-17043.
- Prasad, T. N. V. K. V., & Elumalai, E. K. (2011). Biofabrication of Ag nanoparticles using Moringa oleifera leaf extract and their antimicrobial activity. *Asian Pacific Journal of Tropical Biomedicine*, 1(6), 439.
- Prema, P. (2011). Chemical mediated synthesis of silver nanoparticles and its potential antibacterial application. *Progress in Molecular and Environment Bioengineering-from Analysis and Modeling to Technology Applications*, 6, 151–166.
- Pyrz, W. D., & Buttrey, D. J. (2008). Particle size determination using TEM: a discussion of image acquisition and analysis for the novice microscopist. *Langmuir*, 24(20), 11350-11360.
- Rahimi-Nasrabadi, M., Pourmortazavi, S. M., Shandiz, S. A. S., Ahmadi, F., & Batooli, H. (2014). Green synthesis of silver nanoparticles using Eucalyptus leucoxylon leaves extract and evaluating the antioxidant activities of extract. *Natural Product Research*, 28(22), 1964-1969.
- Rai, M., Yadav, A., & Gade, A. (2009). Silver nanoparticles as a new generation of antimicrobials. *Biotechnology Advances*, 27(1), 76-83.
- Rajakumar, G., & Rahuman, A.A. (2011). Larvicidal activity of synthesized silver nanoparticles using eclipta prostrata leaf extract against filariasis and malaria vectors. *Acta Tropica*, 118, 196-203.
- Rajathi, K., & Sridhar, S. (2013). Green synthesized silver nanoparticles from the medicinal plant Wrightia tinctoria and its antimicrobial potential. *International Journal of ChemTech Research*, 5, 1707-1713.
- Rajeshkumar, S., & Bharath, L. V. (2017). Mechanism of plant-mediated synthesis of silver nanoparticles—a review on biomolecules involved, characterisation and antibacterial activity. *Chemico-Biological Interactions*, 273, 219-227.
- Rasmussen, J. W., Martinez, E., Louka, P., & Wingett, D. G. (2010). ZnO NPs for selective destruction of tumor cells and potential for drug delivery applications. *Expert Opinion on Drug Delivery*, 7(9), 1063-1077.
- Raut, R. W., Mendhulkar, V. D., & Kashid, S. B. (2014). Photosensitized synthesis of silver nanoparticles using Withania somnifera leaf powder and silver nitrate. *Journal of Photochemistry and Photobiology B: Biology*, 132, 45-55.

- Reddy, N. J., Vali, D. N., Rani, M., & Rani, S. S. (2014). Evaluation of antioxidant, antibacterial and cytotoxic effects of green synthesized silver nanoparticles by Piper longum fruit. *Materials Science and Engineering: C*, 34, 115-122.
- Reidy, B., Haase, A., Luch, A., Dawson, K. A., & Lynch, I. (2013). Mechanisms of silver nanoparticle release, transformation and toxicity: a critical review of current knowledge and recommendations for future studies and applications. *Materials*, 6(6), 2295-2350.
- Robertson, J. D., Rizzello, L., Avila-Olias, M., Gaitzsch, J., Contini, C., Magoñ, M. S., & Battaglia, G. (2016). Purification of NPs by size and shape. *Scientific Reports*, 6, 27494.
- Robinson, I. (2012). Nanoparticle structure by coherent XRD. *Journal of the Physical Society of Japan*, 82(2), 021012.
- Rodríguez-León, E., Iñiguez-Palomares, R., Navarro, R. E., Herrera-Urbina, R., Tánori, J., Iñiguez-Palomares, C., & Maldonado, A. (2013). Synthesis of silver nanoparticles using reducing agents obtained from natural sources (*Rumex hymenosepalus* extracts). *Nanoscale Research Letters*, 8(1), 318.
- Rohman, A., & Man, Y. C. (2010). Fourier transform infrared (FTIR) spectroscopy for analysis of extra virgin olive oil adulterated with palm oil. *Food Research International*, 43(3), 886-892.
- Rolim, W. R., Pelegriño, M. T., de Araújo Lima, B., Ferraz, L. S., Costa, F. N., Bernardes, J. S., & Seabra, A. B. (2019). Green tea extract mediated biogenic synthesis of silver nanoparticles: Characterization, cytotoxicity evaluation and antibacterial activity. *Applied Surface Science*, 463, 66-74.
- Roopan, S. M., Madhumitha, G., Rahuman, A. A., Kamaraj, C., Bharathi, A., & Surendra, T. V. (2013). Low-cost and eco-friendly phyto-synthesis of silver nanoparticles using *Cocos nucifera* coir extract and its larvicidal activity. *Industrial Crops and Products*, 43, 631-635.
- Rout, A. N. A. N. D. I. N. I., Jena, P. K., Parida, U. K., & Bindhani, B. K. (2013). Green synthesis of silver nanoparticles using leaves extract of *Centella asiatica* L. for studies against human pathogens. *International Journal of Pharmacy and Biological Sciences*, 4(4), 661-74.
- Rupiasih, N. N., Aher, A., Gosavi, S., & Vidyasagar, P. B. (2015). Green synthesis of silver nanoparticles using latex extract of *Thevetia peruviana*: a novel approach towards poisonous plant utilization. *Recent Trends in Physics of Material Science and Technology*, 1-10.

- Sadeghi, B., Rostami, A., & Momeni, S. S. (2015). Facile green synthesis of silver nanoparticles using seed aqueous extract of *Pistacia atlantica* and its antibacterial activity. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, *134*, 326-332.
- Sadeghi, B., & Gholamhoseinpoor, F. (2015). A study on the stability and green synthesis of silver nanoparticles using *Ziziphora tenuior* (Zt) extract at room temperature. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, *134*, 310-315.
- Santhoshkumar, T., Rahuman, A. A., Rajakumar, G., Marimuthu, S., Bagavan, A., Jayaseelan, C., & Kamaraj, C. (2011). Synthesis of silver nanoparticles using *Nelumbo nucifera* leaf extract and its larvicidal activity against malaria and filariasis vectors. *Parasitology Research*, *108*(3), 693-702.
- Sarkar, A., Kapoor, S., & Mukherjee, T. (2005). Preparation, characterization, and surface modification of silver nanoparticles in formamide. *The Journal of Physical Chemistry B*, *109*(16), 7698-7704.
- Shafaghat, A. (2015). Synthesis and characterization of silver nanoparticles by phytosynthesis method and their biological activity. *Synthesis and Reactivity in Inorganic, Metal-Organic, and Nano-Metal Chemistry*, *45*(3), 381-387.
- Shaligram, N. S., Bule, M., Bhambure, R., Singhal, R. S., Singh, S. K., Szakacs, G., & Pandey, A. (2009). Biosynthesis of silver nanoparticles using aqueous extract from the compactin producing fungal strain. *Process Biochemistry*, *44*(8), 939-943.
- Shameli, K., Bin Ahmad, M., Jazayeri, S. D., Sedaghat, S., Shabanzadeh, P., Jahangirian, H., & Abdollahi, Y. (2012). Synthesis and characterization of polyethylene glycol mediated silver nanoparticles by the green method. *International Journal of Molecular Sciences*, *13*(6), 6639-6650.
- Shankar, S. S., Ahmad, A., & Sastry, M. (2003). Geranium leaf assisted biosynthesis of silver nanoparticles. *Biotechnology Progress*, *19*(6), 1627-1631.
- Shin, Y. J., Ringe, E., Personick, M. L., Cardinal, M. F., Mirkin, C. A., Marks, L. D., & Hersam, M. C. (2013). Centrifugal shape sorting and optical response of polyhedral gold nanoparticles. *Advanced Materials*, *25*(29), 4023-4027.
- Sikder, M., Lead, J. R., Chandler, G. T., & Baalousha, M. (2018). A rapid approach for measuring silver nanoparticle concentration and dissolution in seawater by UV-Vis. *Science of The Total Environment*, *618*, 597-607.

- Singh, A., Jain, D., Upadhyay, M. K., Khandelwal, N., & Verma, H. N. (2010). Green synthesis of silver nanoparticles using *Argemone mexicana* leaf extract and evaluation of their antimicrobial activities. *Digest Journal of Nanomaterials and Biostructures*, 5(2), 483-489.
- Singh, B. P., Menchavez, R., Takai, C., Fuji, M., & Takahashi, M. (2005). Stability of dispersions of colloidal alumina particles in aqueous suspensions. *Journal of Colloid and Interface Science*, 291(1), 181-186.
- Singh, P., Kim, Y. J., Singh, H., Wang, C., Hwang, K. H., Farh, M. E. A., & Yang, D. C. (2015). Biosynthesis, characterization, and antimicrobial applications of silver nanoparticles. *International Journal of Nanomedicine*, 10, 2567.
- Sondi, I., Goia, D. V., & Matijević, E. (2003). Preparation of highly concentrated stable dispersions of uniform silver nanoparticles. *Journal of Colloid and Interface Science*, 260(1), 75-81.
- Srikar, S. K., Giri, D. D., Pal, D. B., Mishra, P. K., & Upadhyay, S. N. (2016). Green synthesis of silver nanoparticles: a review. *Green and Sustainable Chemistry*, 6(01), 34.
- Steinigeweg, D., Schütz, M., Salehi, M., & Schlücker, S. (2011). Gold Nanoparticles: Fast and Cost-Effective Purification of Gold Nanoparticles in the 20–250 nm Size Range by Continuous Density Gradient Centrifugation. *Small*, 7(17), 2406-2406.
- Stensberg, M. C., Wei, Q., McLamore, E. S., Porterfield, D. M., Wei, A., & Sepúlveda, M. S. (2011). Toxicological studies on silver nanoparticles: challenges and opportunities in assessment, monitoring and imaging. *Nanomedicine*, 6(5), 879-898.
- Sun, C., Sze, R., & Zhang, M. (2006). Folic acid-PEG conjugated superparamagnetic nanoparticles for targeted cellular uptake and detection by MRI. *Journal of Biomedical Materials Research Part A: An Official Journal of The Society for Biomaterials, The Japanese Society for Biomaterials, and The Australian Society for Biomaterials and The Korean Society for Biomaterials*, 78(3), 550-557.
- Sun, Q., Cai, X., Li, J., Zheng, M., Chen, Z., & Yu, C. P. (2014). Green synthesis of silver nanoparticles using tea leaf extract and evaluation of their stability and antibacterial activity. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 444, 226-231.
- Sun, X., Tabakman, S. M., Seo, W. S., Zhang, L., Zhang, G., Sherlock, S., & Dai, H. (2009). Separation of NPs in a density gradient: FeCo@C and gold nanocrystals. *Angewandte Chemie International Edition*, 48(5), 939-942.

- Sun, S., Zeng, H., Robinson, D. B., Raoux, S., Rice, P. M., Wang, S. X., & Li, G. (2004). Monodisperse mfe₂o₄ (m=fe, co, mn) nanoparticles. *Journal of the American Chemical Society*, 126(1), 273-279.
- Sunita, D., Tambhale, D., Parag, V., & Adhyapak, A. (2014). Facile green synthesis of silver nanoparticles using Psoralea corylifolia. seed extract and their in-vitro antimicrobial activities. *International Journal of Pharmacy and Biological Sciences*, 5(1), 457-67.
- Suresh, G., Gunasekar, P. H., Kokila, D., Prabhu, D., Dinesh, D., Ravichandran, N., & Siva, G. V. (2014). Green synthesis of silver nanoparticles using Delphinium denudatum root extract exhibits antibacterial and mosquito larvicidal activities. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 127, 61-66.
- Suresh, A. K., Pelletier, D. A., Moon, J. W., Phelps, T., & Doktycz, M. J. (2015). Size-separation of silver NPs using sucrose gradient centrifugation. *Journal of Chromatography and Separation Techniques*, 6(5).
- Surugau, N., & Urban, P. L. (2009). Electrophoretic methods for separation of NPs. *Journal of Separation Science*, 32(11), 1889-1906.
- Sweeney, S. F., Woehrl, G. H., & Hutchison, J. E. (2006). Rapid purification and size separation of gold nanoparticles via diafiltration. *Journal of the American Chemical Society*, 128(10), 3190-3197.
- Ulug, B., Turkdemir, M. H., Cicek, A., & Mete, A. (2015). Role of irradiation in the green synthesis of silver nanoparticles mediated by fig (Ficus carica) leaf extract. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 135, 153-161.
- Vigneshwaran, N., Nachane, R. P., Balasubramanya, R. H., & Varadarajan, P. V. (2006). A novel one-pot 'green' synthesis of stable silver nanoparticles using soluble starch. *Carbohydrate Research*, 341(12), 2012-2018.
- Vijayaraghavan, K., Nalini, S.P.K., Prakash, N.U., & Madhankumar, D. (2012). One step green synthesis of silver nano/micro particles using extracts of Trahyspermum ammi and Papaver somniferum. *Colloids and Surfaces B: Biointerfaces*, 94, 114-117.
- Vijaykumar, P.P.N., Pammi, S.V.N., Kollu, P., Satyanarayana, K.V.V., & Shameem, U. (2014). Green synthesis and characterization of silver nanoparticles using boerhaavia diffusa plant extract and their antibacterial activity. *Industrial Crops and Products*, 52, 562-566.
- Wang, W. N., Tarafdar, J. C., & Biswas, P. (2013). Nanoparticle synthesis and delivery by an aerosol route for watermelon plant foliar uptake. *Journal of Nanoparticle Research*, 15(1), 1417.

- Wei, G. T., & Liu, F. K. (1999). Separation of nanometer gold particles by size exclusion chromatography. *Journal of Chromatography A*, 836(2), 253-260.
- Wei, G. T., Liu, F. K., & Wang, C. C. (1999). Shape separation of nanometer gold particles by size-exclusion chromatography. *Analytical Chemistry*, 71(11), 2085-2091.
- Wei, L., Lu, J., Xu, H., Patel, A., Chen, Z. S., & Chen, G. (2015). Silver nanoparticles: synthesis, properties, and therapeutic applications. *Drug Discovery Today*, 20(5), 595-601.
- Whitfield, P., & Mitchell, L. (2004). XRD analysis of NPs: Recent developments, potential problems and some solutions. *International Journal of Nanoscience*, 3(06), 757-763.
- Xu, X., Caswell, K. K., Tucker, E., Kabisatpathy, S., Brodhacker, K. L., & Scrivens, W. A. (2007). Size and shape separation of gold NPs with preparative gel electrophoresis. *Journal of Chromatography A*, 1167(1), 35-41.
- Zargar, M., Hamid, A. A., Bakar, F. A., Shamsudin, M. N., Shameli, K., Jahanshiri, F., & Farahani, F. (2011). Green synthesis and antibacterial effect of silver nanoparticles using *Vitex negundo* L. *Molecules*, 16(8), 6667-6676.
- Zhang, Y., Kohler, N., & Zhang, M. (2002). Surface modification of superparamagnetic magnetite nanoparticles and their intracellular uptake. *Biomaterials*, 23(7), 1553-1561.
- Zhang, Y., Yang, D., Kong, Y., Wang, X., Pandoli, O., & Gao, G. (2010). Synergetic antibacterial effects of silver nanoparticles@ aloe vera prepared via a green method. *Nano Biomedicine and Engineering*, 2(4), 252-257.
- Zhang, M., Zhang, K., Gusseme, B.D., Verstraete, W., & Field, R. (2014). The Antibacterial and Anti-Biofouling Performance of Biogenic Silver Nanoparticles by *Lactobacillus fermentum*. Biofouling. *The Journal of Bioadhesion and Biofilm Research*, 30, 347-357.
- Zonnoz, N.F., & Salouti, M. (2011). Extracellular biosynthesis of silver nanoparticles using cell filtrate of *Streptomyces* sp. ERI-3. *Scientia Iranica*, 18, 1631-1635.
- Zuas, O., Hamim, N., & Sampora, Y. (2014). Bio-synthesis of silver nanoparticles using water extract of *Myrmecodia* pendan (Sarang Semut plant). *Materials Letters*, 123, 156-159.