



***In Vitro* PRODUCTION AND ASSESSMENT OF SALT TOLERANT LINES
OF MALAYSIAN *Indica* RICE (*Oryza sativa* L.) CV. MARDI SIRAJ 297**

By

NOORHAZIRA BINTI SIDEK

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of
Philosophy**

May 2022

FS 2022 46

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

My beloved family – En Sidek bin Idris, Pn. Kamariah Awang, Nurul Izzaty Sidek, Mohd Khalil Salleh and Imran Harith bin Mohd Khalil.

May this journey humbles me,
and benefit others.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

***In Vitro* PRODUCTION AND ASSESSMENT OF SALT TOLERANT LINES OF MALAYSIAN *Indica* RICE (*Oryza sativa* L.) CV. MARDI SIRAJ 297**

By

NOORHAZIRA BINTI SIDEK

May 2022

Chair : Rosimah binti Nulit, PhD
Faculty : Science

As most of other economically important crops, rice (*Oryza sativa* L.) is sensitive to salinity. Due to emerging salinity impacts on global food security, different strategies have been implemented including the development of salt tolerant varieties to minimize yield loss. Therefore, this study was conducted to produce salt tolerant lines of an importantly grown local rice cultivar, MARDI Siraj 297 through *in vitro* callus selection. The first objective of this study was to optimize the embryogenic callus induction medium. Sterilized MARDI Siraj 297 seeds were inoculated on MS basal medium supplemented with 0 to 3.5 mg/L 2,4-dichlorophenoxyacetic acid (2,4-D) and 0 to 0.5 mg/L kinetin (Kin) for 35 days. The MS medium supplemented with 2.0 mg/L 2,4-D and 0.2 mg/L Kin exhibited the maximum response in all callus growth parameters evaluated. Hence this combination was selected as the optimum medium for embryogenic calli proliferation in the subsequent experiments. The second objective was to produce, screen and select the salt tolerant calli. Embryogenic calli were treated in selection medium containing 0 to 150 mM NaCl for 5 months, followed by screening and selection of salt tolerant variants using morphology and biochemical markers. Normal callus morphology was observed in NaCl concentration up to 75 mM. The biochemical profile of these surviving calli showed that the salt tolerant lines had significantly higher content of proline, total soluble sugar, catalase activity, ascorbate peroxidase activity and K^+/Na^+ ratio compared to the non-tolerant control. Meanwhile, reduction of protein content and elevated MDA production was observed in these salt tolerant calli with increasing salinity level. The third objective involves the salt tolerance enhancement of the selected calli by supplementation of salicylic acid (SA) as phytoprotectant in the growth medium. The addition of 1.0 mM SA reduced the morphological injury while maximized the regeneration frequency and number of shoots as compared to the non-SA-treated calli. In the fourth objective, the regenerated salt tolerant plants (R_0) were acclimatized. The evaluation of growth and agronomic traits found that 6 tolerant lines derived from 25 and 75 mM NaCl

were morphologically normal and able to produce seeds of the first generation of salt tolerant lines (R_1) while 2 lines derived from 100 mM NaCl were sterile. In the final objective, the comparison of agronomic traits between the R_1 salt tolerant lines and control plant showed that the salt tolerant lines exhibited significantly improved agronomic traits, lower stress susceptibility index (SSI) and higher stress tolerance index (STI) in different salinity level during germination, vegetative and reproductive stage. In summary, *in vitro* selective salinity pressure in this study has successfully produced the R_0 , R_1 and R_2 generation of MARDI Siraj 297 salt tolerant lines. These established salt tolerant lines have potential to be utilized by farmers in salinity affected rice field areas. This study also provides a reliable protocol for the establishment of salt tolerant rice lines through tissue culture selection.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENGHASILAN DAN PENILAIAN GALUR TAHAN GARAM BAGI PADI
Indica MALAYSIA (*Oryza sativa* L.) CV. MARDI SIRAJ 297 SECARA *In*
*Vitro***

Oleh

NOORHAZIRA BINTI SIDEK

Mei 2022

Pengerusi : Rosimah Nulit, PhD
Fakulti : Sains

Seperti kebanyakan tanaman utama yang lain, padi (*Oryza sativa* L.) adalah sensitif terhadap kemasinan. Berikutan impak kemasinan yang semakin meluas terhadap jaminan makanan global, pelbagai strategi telah dilaksanakan termasuklah pembangunan varieti tahan garam bagi mengurangkan kehilangan hasil. Oleh itu, kajian ini dilaksanakan bertujuan untuk membangun galur padi tahan garam bagi kultivar tempatan yang penting iaitu MARDI Siraj 297 melalui pemilihan kalus. Objektif pertama dalam kajian ini adalah untuk mengoptimalkan medium bagi induksi kalus embriogenik. Biji benih MARDI Siraj 297 diinokulasi ke dalam medium MS yang ditambah dengan 0 hingga 3.5 mg/L 2,4-dikloroasetik asid (2,4-D) dan 0 hingga 0.5 mg/L kinetin (Kin) selama 35 hari. Medium MS yang ditambah dengan 2.0 mg/L 2,4-D dan 0.2 mg/L kinetin menunjukkan kesan maksimum terhadap kesemua parameter pertumbuhan kalus. Oleh itu, kombinasi ini telah dipilih sebagai medium yang optimum untuk proliferasi kalus embriogenik bagi eksperimen seterusnya. Objektif kedua adalah untuk menghasilkan, menyaring dan memilih kalus yang tahan garam. Kalus embriogenik dikultur di dalam medium pemilihan yang mengandungi 0 hingga 150 mM NaCl selama 5 bulan, diikuti dengan saringan dan pemilihan varian tahan garam menggunakan ciri morfologi dan penanda biokimia. Morfologi kalus yang normal dapat diperhatikan di dalam kepekatan NaCl sehingga 75 mM. Profil biokimia bagi kalus *in vitro* yang masih hidup menunjukkan bahawa galur tahan garam tersebut mempunyai kandungan prolina, jumlah gula terlarut, aktiviti katalase, aktiviti askorbat peroksida dan nisbah K^+/Na^+ yang lebih tinggi berbanding kawalan yang tidak toleran. Sementara itu, penurunan kandungan protin dan penghasilan MDA yang tinggi dapat diperhatikan di dalam kalus tahan garam apabila tahap kemasinan meningkat. Objektif ketiga adalah untuk mempertingkatkan toleransi garam pada kalus yang telah dipilih dengan penambahan asid salisilik (SA) sebagai fitopelindung di dalam medium pertumbuhan. Penambahan 1.0 mM SA dapat mengurangkan kecederaan morfologi serta memaksimumkan frekuensi regenerasi dan bilangan pucuk

berbanding kalus yang tidak dirawat dengan SA. Di dalam objektif keempat, anak pokok yang diregenerasi dari kalus (R_0) telah diaklimatisasi. Penilaian terhadap pertumbuhan dan ciri agronomi bagi anak pokok tahan garam (R_0) mendapati bahawa 6 galur tahan garam yang terhasil dari medium 25 dan 75 mM NaCl adalah subur dan normal dari segi morfologi dan berjaya menghasilkan biji benih tahan garam generasi pertama (R_1) manakala 2 galur dari medium 100 mM adalah mandul. Dalam objektif terakhir, perbandingan ciri agronomi di antara generasi pertama (R_1) galur tahan garam dan pokok kawalan menunjukkan bahawa galur tahan garam mempunyai ciri agronomi yang lebih baik, indeks kerentanan tekanan (SSI) yang rendah serta indeks ketahanan tekanan (STI) yang tinggi sewaktu peringkat percambahan, vegetatif dan pembiakan. Secara ringkasnya, tekanan kemasinan selektif *in vitro* dalam kajian ini telah berjaya menghasilkan galur tahan garam generasi R_0 , R_1 dan R_2 bagi MARDi Siraj 297. Galur padi tahan garam yang dihasilkan ini berpotensi untuk digunakan oleh pesawah di sawah padi yang terjejas dengan kemasinan. Kajian ini juga menyediakan protokol bagi penghasilan galur padi tahan garam melalui kaedah pemilihan kultur tisu.

ACKNOWLEDGEMENTS

The utmost appreciation to my respected and dedicated supervisor, Prof Madya Dr. Rosimah binti Nulit, as well as my resourceful co-supervisors, Prof Yap Chee Kong, Dr. Christina Yong Seok Yien and Dr Rogayah binti Sekeli (MARDI). Special gratitude to the staffs of Biology Department, Faculty of Science, UPM for technical support and finally to the Ministry of Higher Education Malaysia for the scholarship (SLAB).



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Rosimah binti Nulit, PhD

Associate Professor
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Yap Chee Kong, PhD

Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

Christina Yong Seok Yien, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Member)

Rogayah Sekeli, PhD

Principle Research Officer
Biotechnology and Nanotechnology Research Centre
Malaysian Agricultural Research and Development Institute (MARDI)
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 14 December 2023

TABLE OF CONTENTS

		Page
	ABSTRACT	i
	ABSTRAK	iii
	ACKNOWLEDGEMENTS	v
	APPROVAL	vi
	DECLARATION	viii
	LIST OF TABLES	xiv
	LIST OF FIGURES	xvi
	LIST OF ABBREVIATIONS	xix
	CHAPTER	
1	INTRODUCTION	
	1.1 Background of study	1
	1.2 Problem statement	2
	1.3 Significance of study	3
	1.4 Objectives of study	3
2	LITERATURE REVIEW	
	2.1 Rice origins and diversity	4
	2.1.1 Malaysian rice varieties	5
	2.2.1 Rice growth and morphology	6
	2.2 Global rice production area and status	6
	2.3 Salinity impacts on crops productivity	8
	2.3.1 World distribution of saline soil occurrence	8
	2.3.2 Salinity threats to rice fields in Malaysia	11
	2.3.3 Causes of salinity	11
	2.3.4 Effects of salinity on rice growth	13
	2.3.5 Plant response mechanisms towards salinity stress	15
	2.4 Strategies to improve salinity tolerance in rice	20
	2.4.1 Application of phytoprotectant	21
	2.4.2 Conventional breeding	25
	2.4.3 Marker assisted breeding	25
	2.4.4 Transgenic approach	26
	2.4.5 Genome editing	26
	2.4.6 <i>In vitro</i> selection of somaclonal variants	27
3	MATERIALS AND METHODS / METHODOLOGY	
	3.1 Plant material	32
	3.2 Seed sterilization	32
	3.3 Medium optimization for embryogenic callus induction of MARDI Siraj 297	32
	3.4 Proliferation of embryogenic callus of MARDI Siraj 297	34

3.5	Production and screening of salt tolerant calli of MARDI Siraj 297	34
3.5.1	Callus morphology	35
3.5.2	Proline	35
3.5.3	Total soluble sugar (TSS)	35
3.5.4	Total protein	36
3.5.5	Catalase (CAT) activity	36
3.5.6	Ascorbate peroxidation (APX) activity	37
3.5.7	Lipid peroxidation (LPX)	37
3.5.8	K ⁺ /Na ⁺ ratio	37
3.6	Enhancement and selection of salt tolerant calli using salicylic acid (SA) as phytoprotectant	38
3.7	Regeneration of salt tolerant calli of MARDI Siraj 297	38
3.8	Acclimatization and growth of R ₀ (regenerant) salt tolerant lines of MARDI Siraj 297	38
3.9	Comparison of growth and agronomic traits between R ₁ (first generation) salt tolerant lines and control in salinity at different growth stages	40
3.9.1	Germination stage	40
3.9.2	Vegetative stage	43
3.9.3	Reproductive stage	44
3.10	Statistical analysis	46
4	RESULTS	
4.1	Medium optimization for embryogenic callus induction of MARDI Siraj 297	49
4.1.1	Callus growth	49
4.1.2	Callus morphology and somatic embryogenesis of MARDI Siraj 297	54
4.1.3	Regeneration ability of embryogenic callus of MARDI Siraj 297	56
4.2	Production, screening and selection of salt tolerant calli of MARDI Siraj 297	56
4.2.1	Callus morphology	57
4.2.2	Biochemical content	59
4.3	Enhancement and selection of salt tolerant calli using salicylic acid (SA) as phytoprotectant	63
4.3.1	Morphological response of salt treated calli in SA treatment	63
4.3.2	Regeneration of salt tolerant lines from callus	64
4.4	Acclimatization, phenotypic variation and growth of the regenerated salt tolerant line plants (R ₀)	68
4.4.1	Acclimatization and phenotypic variation of the regenerated salt tolerant line plants (R ₀)	68
4.4.2	The growth of acclimatized R ₀ (regenerants) salt tolerant lines	71

4.5	Comparison of growth and agronomic traits between first generation (R_1) salt tolerant lines and control plant in salinity condition at different growth stages	74
4.5.1	Growth performance of first generation (R_1) salt tolerant lines and control plant in salinity during germination stage	74
4.5.2	Growth performance of R_1 salt tolerant lines and control plant in salinity during vegetative stage	83
4.5.3	Agronomic traits of R_1 salt tolerant lines and control plants in salinity during reproductive stage	89
5	DISCUSSION	
5.1	Medium optimization for embryogenic callus induction of MARDI Siraj 297	96
5.2	Production, screening and selection of salt tolerant callus of MARDI Siraj 297	98
5.2.1	The effects of salinity on callus morphology	98
5.2.2	The effects of salinity on compatible solutes of salt tolerant calli	99
5.2.3	The effect of salinity on antioxidant activity of the salt tolerant calli	102
5.3	The effect of salicylic acid (SA) as phytoprotectant to enhance the tolerance level of the selected salt tolerant calli	103
5.4	Acclimatization, phenotypic variation and growth of the regenerated salt tolerant lines (R_0)	105
5.5	Comparison of growth and agronomic traits between first generation of salt tolerant lines (R_1) and control plant under salinity stress at different growth stages	107
5.5.1	Comparison of germination traits between first generation (R_1) salt tolerant lines and control plant under salinity stress during germination stage	107
5.5.2	Growth performance of first generation (R_1) salt tolerant lines and control plant in salinity during vegetative stage	109
5.5.3	Growth performance of first generation (R_1) salt tolerant lines and control plant in salinity during reproductive stage	111

6	CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	
6.1	Conclusion	114
6.2	Recommendations for future research	115
	REFERENCES	117
	APPENDICES	150
	BIODATA OF STUDENT	200
	PUBLICATION	201



LIST OF TABLES

Table		Page
2.1	MARDI Siraj 297 agronomic characteristic and resistance to rice blast	6
2.2	Soil salinity classification and its respective crop response	8
2.3	Osmolyte distribution in selected salt tolerant plant families	19
2.4	Reaction mechanisms of major ROS scavenging enzymatic antioxidants	20
2.5	Phytoprotectant used in rice treated in different salinity levels	22
2.6	Improved salinity tolerance rice cultivars developed using conventional breeding technique	25
2.7	Major families of transcript factors that regulate salinity tolerance in rice	26
3.1	Combination of 2,4-D and Kinetin treatments for embryogenic callus induction	32
3.2	Salt tolerant enhancement medium composition	38
3.3	Modified standard evaluation score (SES) of visual salt injury in rice	44
4.1	The effect of different 2,4-D and Kin concentrations of callus growth of MARDI Siraj 297	51
4.2	Regeneration frequency and number of shoots per explants of salt tolerant plantlets (R_0) from different SA enhancement medium	66
4.3	Successfully acclimatized R_0 salt tolerant lines	68
4.4	Growth and agronomic traits of acclimatized R_0 (regenerants) salt tolerant lines and control plants	73
4.5	The effect of lines and salinity level on final germination percentage (FGP), germination index (GI), germination rate index (GRI) and mean germination time (MGT) in R_1 salt tolerant lines and control plant	75

4.6	The interaction between salinity levels and lines on various seedling growth parameters and vigor of salt tolerant lines and control during germination stage	77
4.7	Tolerance indices of salt tolerant lines and control plants at different salinity levels during germination stage	80
4.8	The interaction between salinity levels and lines on growth parameters of salt tolerant lines (R_1) and control plant during vegetative stage	84
4.9	Tolerance indices of salt tolerant lines and control plants at different salinity levels during vegetative stage	88
4.10	The interaction between salinity treatments and lines on agronomic traits of R_1 salt tolerant lines and control plant during reproductive stage	90
4.11	Tolerance indices of salt tolerant lines and control at high salinity level during reproductive stage	95

LIST OF FIGURES

Figure		Page
2.1	Evolutionary pathway of two cultivated rice species	4
2.2	Developmental stages of the rice plant	6
2.3	Rice production quantities by country in 2019	7
2.4	Global distribution of the change in likelihood (θ) of surface soils (0 to 30 cm) with an $EC_e \geq 4 \text{ dSm}^{-1}$ in year 2000 to 2018, relative to the 1981 - 1999	10
2.5	Dry land salinity processes	12
2.6	Variation in the salt tolerance level of cereal species, in comparison with the salt sensitive species (<i>Arabidopsis thaliana</i> L. Heynh.) and salt tolerant halophyte species (tall wheatgrass and saltbush) based on percentage of dry matter increase in saline soil	13
2.7	Detrimental effects of salinity on plant growth	15
2.8	Integration of various 'omics-based' approaches in dissecting out the salinity tolerance mechanisms in rice.	16
2.9	Plant system adaptation in saline soil	17
2.10	Management strategies for saline soil improvement	21
2.11	Biological functions of phytohormones in the salt stress response regulation in plants	23
2.12	Mechanism of somaclonal variation in <i>in vitro</i> plants resulting from oxidative burst during culture	28
2.13	Different approaches employed to create somaclonal variants	29
3.1	The overview of MARDI Siraj 297 salt tolerant lines development	31
3.2	Morphological distinction between embryogenic and non-embryogenic rice calli	34
3.3	Acclimatization of R_0 salt tolerant plants	39
3.4	The measured growth and agronomic parameters of R_0 (regenerants) salt tolerant line plants	40

3.5	Seed germination test at different salinity level in the controlled growth room	41
3.6	Rice seedlings of salt tolerant lines and control at day 14 (2 - 3 leaf stage) in different salinity levels	43
3.7	Rice seedlings transferred into paddy soil at day 7 after germination	45
4.1	Overview of the parameters assessed in the study	48
4.2	Morphogenic response of rice calli at 35 days on MS medium supplemented with different concentrations of 2,4-D and Kin	54
4.3	Progressive development of somatic embryogenesis in MARDI Siraj 297 rice callus	55
4.4	Regeneration of embryogenic calli in MARDI Siraj 297 from the optimized callus induction medium	56
4.5	Morphology of MARDI Siraj 297 calli after 5 months in selection medium containing NaCl under 20X magnification	58
4.6	The total protein content, proline content, total soluble sugar and K ⁺ /Na ⁺ ratio of salt tolerant calli treated in different salinity level	60
4.7	Catalase activity, ascorbate peroxidase activity, and lipid peroxidation in callus treated under different salinity level	62
4.8	Morphology of rice calli after 1 month on salinity tolerance enhancement medium containing salicylic acid (SA)	63
4.9	Regeneration stages of salt tolerant plantlets (R ₀) from calli	65
4.10	The healthy growing regenerated salt tolerant line plantlets (R ₀) were ready for acclimatization	67
4.11	Acclimatized R ₀ salt tolerant lines at 115 days (harvest)	69
4.12	The spikelet of R ₀ salt tolerant line plants during maturity showed ripened rice grains, except for the N100L1 and N100L2	70
4.13	The florets of R ₀ show the presence of complete number of stigma (2) and anther (6) in control plants and all salt	71

	tolerant lines (including in N100L1 and N100L2 which failed to produce grains)	
4.14	The seedlings morphology on tenth day after germination at different salinity levels demonstrated that the salt tolerant lines showed higher shoot, root and seedling length compared to control plant	82
4.15	Morphology of control plant and R ₁ salt tolerant lines in different salinity level during vegetative stage	87
4.16	The morphology of R ₁ salt tolerant lines and control plant grown in different treatment at day 95 after sowing in non-saline and high salinity	93
4.17	Panicle length of salt tolerant lines and control plant in non-saline and high salinity (10 dSm ⁻¹)	94

LIST OF ABBREVIATIONS

ANOVA	analysis of variance
APX	ascorbate peroxidase
CAT	catalase
CIM	callus induction medium
CRD	complete randomized design
dH ₂ O	Distilled water
DMRT	Duncan's Multiple Range Test
dSm ⁻¹	deciSiemens per meter
EC	electrical conductivity (unit = dS m ⁻¹)
g	relative centrifugal force
ha	hectare
H ₂ O ₂	hydrogen peroxide
MDA	malondialdehyde
mM	millimolar
MS	Murashige & Skoog
N	normality (unit for solution concentration)
NaCl	sodium chloride
NADP ⁺	nicotinamide adenine dinucleotide phosphate
NAD(P)H	nicotinamide adenine dinucleotide phosphate hydrogen
PGR	plant growth regulator
RCBD	randomized complete block design
ROS	reactive oxygen species
R ₀	regenerated salt tolerant plantlets (from callus)
R ₁	first generation of salt tolerant lines

SA	salicylic acid
TSS	total soluble sugars
v/v	volume/ volume
w/v	weight/ volume
ϵ	absorption coefficient



CHAPTER 1

INTRODUCTION

1.1 Background of study

Rice (*Oryza sativa* L.) is the world's most important food crop that feeds about 4 billion people around the globe with an estimation of 25% increase in demand between 2010 to 2030 (IRRI, 2019). However, rice production in most regions has progressively been affected by various abiotic stress such as drought, salinity and flood, which are expected to worsen due to climate change (Mandal et al., 2018; Pareek et al., 2020; Dar et al., 2021). In Malaysia, the impact of climate change such as temperature increase and soil degradation has imposed severe threats to rice production (Firdaus et al., 2020), whereby salinity became one of the main contributors for soil degradation and productivity loss in cultivable lands (Machado & Serralheiro, 2017; Raoufi et al., 2021).

Salinity induced various major responses in rice plants in terms of morphology, physiology, biochemical and agronomic attributes (Riaz et al., 2019; Irakoze et al., 2020; Razzaq et al., 2020; Dramalis et al., 2021). Dissolved salt in soil water exerts two phases of growth response in plants, as revealed by Munns & Tester (2008). The first phase is known as osmotic effect whereby it reduces plants ability for water uptake, followed by ionic effect that interferes with transpiration and causes cell injury in the transpiring leaves (Lefevre et al., 2001; Munns, 2005). These two events hamper cellular metabolisms, accelerate senescence, interrupt source-sink relationship and finally impair plant growth and development (Rahman et al., 2017). In rice, increased severity of salinity effect occurs during seedling and early vegetative phase (Amirjani, 2010; Krishnamurthy et al., 2016) and later during reproductive stage (Reddy et al., 2017; Sen et al., 2017).

In order to survive, plants are endowed with various mechanisms to surge their tolerance during this stressful condition (Borsani et al., 2003), which includes the activation of antioxidant enzymes (Kim et al., 2018) and enhanced accumulation of compatible solutes such as amino acids and soluble sugars for osmotic adjustment (Chen & Murata, 2002). Previous studies by Zeng & Shannon (2000) and Reddy et al. (2017) demonstrated that certain physiological and morphological parameters serve as reliable indicators for salinity tolerance evaluation in rice. Thus, assessing the cumulative effect of these morpho-physiological traits can help to build a comprehensive protocol for salinity tolerant selection and elicit the underlying mechanisms involved in order to develop salt tolerant varieties (Kakar et al., 2019)

1.2 Problem statement

Similar to the majority of crop species, rice, including the Malaysian varieties are categorized as a glycophyte or salt sensitive plant (Green et al., 2017). The MARDI Siraj 297 which is the majorly grown local variety, was also recognized as susceptible to salinity level as low as 4 dSm^{-1} (Sazali et al., 2021). Therefore, salinity could pose significant threat to the country's rice supply, since 47.9% of the paddy fields are cultivated with this variety (Rahim et al., 2021). In Malaysia, sea water intrusion into the paddy fields during high tide phenomenon had been reported in northern states of Malaysia (Perak, Kedah and Perlis), with an estimated destruction of 35 hectares of productive rice cultivation areas, including the area where MARDI Siraj 297 is cultivated (Rahman et al., 2021). Due to increased frequency of such event, it was expected that salinity occurrence will continue to worsen and cause a huge loss in this highly demanded variety (Aling, 2020).

In addition, studies on climate change impacts in Malaysia demonstrated that few factors contributing to salinity such as temperature increase, precipitation variability, intensified use of nitrogen fertilizers and rising seawater level also posed severe threats to paddy productivity in the main rice granary areas of Peninsular Malaysia including MADA (Kedah), IADA (Pulau Pinang), KADA (Kelantan), Perak and Selangor (Herman et al., 2015; Sazali et al., 2021). These factors are inevitably contribute to the increased salinity level in agricultural lands including rice field especially in the coastal line areas and therefore compromising the nation's food security (Jamaluddin et al., 2018).

Due to the emerging impacts of salinity towards plant productivity in Malaysia and worldwide (Pareek et al., 2020; Rahman et al., 2021), different strategies have been implemented to mitigate this problem including the development of salt tolerant cultivars (Mandal et al., 2018) and application of various phytoprotectant such as salicylic acid (Jini & Joseph, 2017; Rahman et al., 2017). Salicylic acid is regarded as a key phytohormone involved in plant systematic acquired resistance (Kim et al., 2018), with ability to regulate many physiological and biochemical responses including modulation of other endogenous hormones and antioxidant activities (Janda et al., 2006; Yusuf et al., 2013).

Research on salt tolerant rice cultivars have been conducted through various approaches including conventional breeding, transgenic manipulation and *in vitro* induced mutation. However, each of them imposed its own limitations (Reddy et al., 2017; Chen et al., 2021). Conventional breeding which depends on wild germplasms for salt tolerant traits mostly caused reduction in agronomic characters such as light-sensitivity, decreased yield and poor grain quality, making it difficult for breeders to introduce them into domesticated varieties (Das et al., 2015). In molecular breeding, limited progress in the research was mainly due to limited genetic resources with sufficient tolerance level and lack of reliable salinity tolerance genes with huge effects (Chen et al., 2021). Despite numerous salinity-responsive genes have been identified in rice, none of them have been

successfully incorporated into commercial germplasm so far (Kotula et al., 2020; Liu et al., 2020). Therefore, tissue culture selection was opted in this study due to its feasibilities such as faster development compared to conventional breeding, high frequency of trait changes, possibility of obtaining novel variants and allowing the use of large population of cells for selection purpose (Deepthi, 2018).

1.3 Significance of study

To date, Malaysia has yet to develop its own salt tolerant rice cultivar. Although few studies have successfully regenerate *in vitro* salt tolerant rice (Kalhori et al., 2017; Atabaki et al., 2018), the performance of these lines in salinity at different growth stages has not been evaluated. At present, rice farmers in stress-prone areas depend on cultivating local low yielding traditional cultivars and landraces (Dar et al., 2021) while some others left their paddy fields uncultivated during stressful seasons (Ismail et al., 2013), thereby causing significant loss in the rice supply. Hence, this study aims to develop salt tolerant rice lines that could be used by farmers in salt affected lands in order to ensure sustainable rice production for the future.

1.4 Objectives of study

Previous findings showed that *in vitro* selection is likely to have a significant role in the recovery of stable somaclonal variants with improved stress tolerance. This study was therefore looked at the feasibility of producing salt tolerant rice lines of MARDI Siraj 297 through *in vitro* selection and evaluates the role of salicylic acid in enhancing salt tolerance ability of these lines. Hence, this study was conducted to achieve the following objectives:

1. To optimize embryogenic callus induction from MARDI Siraj 297 seeds.
2. To produce, screen and select the salt tolerant callus of MARDI Siraj 297 using morphological and biochemical markers.
3. To enhance salt tolerance traits of MARDI Siraj 297 callus by supplementation of salicylic acid as phytoprotectant.
4. To acclimatize the regenerants (R_0) salt tolerant lines of MARDI Siraj 297.
5. To compare the growth and agronomic traits between first generation (R_1) salt tolerant lines and control plant of MARDI Siraj 297 at different growth stages.

REFERENCES

- Aala, W. F., & Gregorio, G. B. (2019). Morphological and molecular characterization of novel salt-tolerant rice germplasms from the Philippines and Bangladesh. *Rice Science*, 26(3), 178–188. <https://doi.org/10.1016/j.rsci.2018.09.001>
- Abdul-Baki, A. A., & Anderson, J. D. (1973). Vigor determination in soybean seed by multiple criteria. *Crop Science*, 13(6), 630–633. <https://doi.org/https://doi.org/10.2135/cropsci1973.0011183X001300060013x>
- Abdullah, Z., Khan, M. A., & Flowers, T. J. (2001). Causes of sterility in seed set of rice under salinity stress. *Journal of Agronomy and Crop Science*, 187(1), 25–32. <https://doi.org/10.1046/j.1439-037X.2001.00500.x>
- Abdullah, Zaibunnisa, Khan, M. A., & Flowers, T. J. (2002). Causes of sterility in rice under salinity stress. In R. Ahmad & K. A. Malik (Eds.), *Prospects for Saline Agriculture* (1st ed., pp. 177–187). Springer, Dordrecht. https://doi.org/10.1007/978-94-017-0067-2_19
- Abiri, R., Maziah, M., Shaharuddin, N. A., Yusof, Z. N. B., Atabaki, N., Hanafi, M. M., Sahebi, M., Azizi, P., Kalhori, N., & Valdiani, A. (2017). Enhancing somatic embryogenesis of Malaysian rice cultivar MR219 using adjuvant materials in a high-efficiency protocol. *International Journal of Environmental Science & Technology*, 14, 1091–1108. <https://doi.org/10.1007/s13762-016-1221-y>
- Abrol, I. P., Yadav, J. S. P., & Massoud, F. I. (1988). Saline soils and their management. In *Salt-Affected Soils and their Management*. Food and Agriculture Organization of the United Nations. <http://www.fao.org/3/x5871e/x5871e00.htm#Contents>
- Acosta-Moto, J. R., Ortuno, M. F., Bernal-Vicente, A., Diaz-Vivancos, P., Jesus Sanchez-Blanco, M., & Antonio Hernandez, J. (2017). Plant responses to salt stress: Adaptive mechanisms. *Agronomy*, 7(1), 18. <https://doi.org/10.3390/agronomy7010018>
- Acosta-Motos, J. R., Penella, C., Hernandez, J. A., Diaz-Vivancos, P., Sanchez-Blanco, M. J., Navarro, J. M., Gomez-Bellot, M. J., & Barba-Espin, G. (2020). Towards a sustainable agriculture: Strategies involving phytoprotectants against salt stress. *Agronomy*, 10, 194. <https://doi.org/10.3390/agronomy10020194>
- Aebi, H. (1984). Catalase *in vitro*. *Methods in Enzymology*, 105, 121–126. [https://doi.org/10.1016/S0076-6879\(84\)05016-3](https://doi.org/10.1016/S0076-6879(84)05016-3)
- Agastian, P., Kingsley, S., & Vivekanandan, M. (2000). Effect of salinity on photosynthesis and biochemical characteristics in mulberry genotypes.

- Ahanger, M. A., Qin, C., Begum, N., Maodong, Q., Dong, X. X., El-Esawi, M., El-Sheikh, M. A., Alatar, A. A., & Zhang, L. (2019). Nitrogen availability prevents oxidative effects of salinity on wheat growth and photosynthesis by up-regulating the antioxidants and osmolytes metabolism, and secondary metabolite accumulation. *BMC Plant Biology*, 19, 479. <https://doi.org/10.1186/s12870-019-2085-3>
- Ahmad, F., Singh, A., & Kamal, A. (2020). Osmoprotective role of sugar in mitigating abiotic stress in plants. In Aryadeep Roychoudhury & Durgesh Kumar Tripathi (Eds.), *Protective Chemical Agents in the Amelioration of Plant Abiotic Stress: Biochemical and Molecular Perspectives* (pp. 53–70). John Wiley & Sons Ltd. <https://doi.org/10.1002/9781119552154.ch3>
- Ahmad, P., Ahanger, M. A., Alam, P., Alyemeni, M. N., Wijaya, L., Ali, S., & Ashraf, M. (2019). Silicon (Si) supplementation alleviates NaCl toxicity in mung bean (*Vigna radiata* L. Wilczek) through the modifications of physio-biochemical attributes and key antioxidant enzymes. *Journal of Plant Growth Regulation*, 38, 70–82. <https://doi.org/https://doi.org/10.1007/s00344-018-9810-2>
- Ahmad, P., Azooz, M. M., & Prasad, M. N. V. (2013). *Ecophysiology and responses of plants under salt stress* (P. Ahmad, M. M. Azooz, & M. N. V. Prasad (eds.)). Springer New York Heidelberg Dordrecht. <https://doi.org/10.1007/978-1-4614-4747-4>
- Al-Forkan, M., Rahim, M. A., Chowdhury, T., Akter, P., & Khaleda, L. (2005). Development of highly in vitro callogenesis and regeneration system for some salt tolerant rice (*Oryza sativa* L.) cultivars of Bangladesh. *Biotechnology*, 4(3), 230–234. <https://doi.org/10.3923/biotech.2005.230.234>
- Al-Khateeb, S. A., Al-Khateeb, A. A., Sattar, M. N., & Mohmand, A. S. (2020). Induced in vitro adaptation for salt tolerance in date palm (*Phoenix dactylifera* L.) cultivar Khalas. *Biological Research*, 53, 37. <https://doi.org/10.1186/s40659-020-00305-3>
- Alhasnawi, A. N., Zain, C. R. C. M., Kadhimi, A. A., Isahakb, A., Mohamad, A., Ashraf, M. F., & Yusoff, W. M. W. (2016). Applications of polysaccharides (β -glucan) for physiological and biochemical parameters for evaluation rice tolerance under salinity stress at seedling stage. *Journal of Crop Science and Biotechnology*, 19(5), 353–362. <https://doi.org/10.1007/s12892-016-0009-4>
- Aling, Y. D. (2020, November 8). MARDI perlukan 6 tahun hasilkan benih padi sah. *METRO TV*. <https://www.hmetro.com.my/mutakhir/2020/11/639733/mardi-perlukan-6-tahun-hasilkan-benih-padi-sah-metrotv>

- Allison, L. E., Bernstein, L., Bower, C. A., Brown, J. W., Fireman, M., Hatcher, J. T., Hayward, H. E., Pearson, G. A., Reeve, R. C., Richards, L. A., & Wilcox, L. V. (1954). *Diagnosis and improvement of salinity and alkali soils* (L. A. Richards (ed.)). United States Department of Agriculture.
- Alsahli, A., Mohamed, A. K., Alaraidh, I., Al-Ghamdi, A., Al-Watban, A., El-Zaidy, M., & Alzahrani, S. M. (2019). Salicylic acid alleviates salinity stress through the modulation of biochemical attributes and some key antioxidants in wheat seedlings. *Pakistan Journal of Botany*, *51*(5), 1551–1559. [https://doi.org/10.30848/PJB2019-5\(12\)](https://doi.org/10.30848/PJB2019-5(12))
- Amako, K., Chen, G. X., & Asada, K. (1994). Separate assays specific for ascorbate peroxidase and guaiacol peroxidase and for the chloroplastic and cytosolic isozymes of ascorbate peroxidase in plants. *Plant and Cell Physiology*, *35*(3), 497–504. <https://doi.org/10.1093/oxfordjournals.pcp.a078621>
- Amini, F., & Ehsanpour, A. A. (2005). Soluble proteins, proline, carbohydrates and Na⁺/K⁺ changes in two tomato (*Lycopersicon esculentum* Mill.) cultivars under in vitro salt stress. *American Journal of Biochemistry and Biotechnology*, *1*(4), 204–208. <https://doi.org/https://doi.org/10.3844/ajbbbsp.2005.204.208>
- Amirjani, M. R. (2010). Effect of NaCl on some physiological parameters of rice. *European Journal of Biological Sciences*, *3*(1), 6–16. <https://doi.org/https://doi.org/10.1080/03235408.2011.559034>
- Anjum, N. A., Sharma, P., Gill, S. S., Hasanuzzaman, M., Khan, E. A., Kachhap, K., Mohamed, A. A., Thangavel, P., Devmanjuri Devi, G., Vasudhevan, P., Sofo, A., Khan, N. A., Narayan Misra, A., Lukatkin, A. S., Pal Singh, H., Pereira, E., Tuteja, N., & Narayan Misra misraan, A. (2016). Catalase and ascorbate peroxidase-representative H₂O₂-detoxifying heme enzymes in plants. *Environmental Science and Pollution Research*, *23*, 19002–19029. <https://doi.org/10.1007/s11356-016-7309-6>
- Asfaw, K. G., & Danno, F. I. (2011). Effects of salinity on days to heading (DTH), days from heading to maturity (DHTM) and days to maturity (DTM) of tef (*Eragrostis tef* Zucc. Trotter) accessions and varieties in Ethiopia. *Asian Journal of Agricultural Sciences*, *3*(4), 250–256.
- Ashraf, M., & Foolad, M. R. (2007). Roles of glycine betaine and proline in improving plant abiotic stress resistance. *Environmental and Experimental Botany*, *59*, 206–216. <https://doi.org/https://doi.org/10.1016/j.envexpbot.2005.12.006>
- Aslam, R., Bostan, N., Maria, M., & Safdar, W. (2011). A critical review on halophytes: Salt tolerant plants. *Journal of Medicinal Plants Research*, *5*(33), 7108–7118. <https://doi.org/10.5897/JMPRx11.009>
- Atabaki, N., Nulit, R., Kalhori, N., Lasumin, N., Sahebi, M., & Abidiri, R. (2018).

In vitro selection and development of Malaysian salt-tolerant rice (*Oryza sativa* L. cv. MR263) under salinity. *Acta Scientific Agriculture*, 2(8), 8–17.

Azimi, M. R. (2021, November 10). 5,849 hektar sawah di Kedah ditenggelami banjir. *Utusan Malaysia*. <https://www.utusan.com.my/terkini/2021/11/5849-hektar-sawah-di-kedah-ditenggelami-banjir/>

Azooz, M. M., Youssef, A. M., & Ahmad, P. (2011). Evaluation of salicylic acid (SA) application on growth, osmotic solutes and antioxidant enzyme activities on broad bean seedlings grown under diluted seawater. *International Journal of Plant Physiology and Biochemistry*, 3(14), 253–264. <https://doi.org/10.5897/ijppb11.052>

Bado, S., Forster, B. P., Ghanim, A. M. A., Jankowicz-Cieslak, J., Berthold, G., & Luxiang, L. (2016). Protocols for pre-field screening of mutants for salt tolerance in rice, Wheat and barley. In S. Bado, B. P. Forster, A. M. A. Ghanim, J. Jankowicz-Cieslak, G. Berthold, & L. Luxiang (Eds.), *Protocols for Pre-Field Screening of Mutants for Salt Tolerance in Rice, Wheat and Barley*. Springer International Publishing. <https://doi.org/10.1007/978-3-319-26590-2>

Bashir, M. A., Silvestri, C., Ahmad, T., Hafiz, I. A., Abbasi, N. A., Manzoor, A., Cristofori, V., & Rugini, E. (2020). Osmotin: A cationic protein Leads to improve biotic and abiotic stress tolerance in plants. *Plants*, 9, 992. <https://doi.org/10.3390/plants9080992> www.mdpi.com/journal/plants

Bates, L. S. (1973). Rapid determination of free proline for water-stress studies. *Plant and Soil*, 39, 205–207.

Berita Harian. (2019, August 26). Sawah padi makin susut. *Berita Harian*. <https://www.bharian.com.my/berita/nasional/2019/08/600332/sawah-padi-makin-susut>

Bhatia, S. (2015). Plant tissue culture. In *Modern Applications of Plant Biotechnology in Pharmaceutical Sciences* (3rd ed., pp. 31–107). Academic Press. <https://doi.org/http://dx.doi.org/10.1016/B978-0-12-802221-4.00002-9>

Binte Mostafiz, S., & Wagiran, A. (2018). Efficient callus induction and regeneration in selected indica rice. *Agronomy*, 8(5). <https://doi.org/10.3390/agronomy8050077>

Borsani, O., Valpuesta, V., & Botella, M. A. (2003). Developing salt tolerant plants in a new century: A molecular biology approach. *Plant Cell, Tissue and Organ Culture*, 73, 101–115. <https://doi.org/https://doi.org/10.1023/A:1022849200433>

Bose, J., Rodrigo-Moreno, A., & Shabala, S. (2014). ROS homeostasis in halophytes in the context of salinity stress tolerance. *Journal of Experimental Botany*, 65(5), 1241–1257. <https://doi.org/10.1093/jxb/ert430>

- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72, 248–254.
- Brar, D. S., & Jain, S. M. (1998). Somaclonal variation: Mechanism and applications in crop improvement. In S. M. Jain, D. S. Brar, & B. S. Ahloowalia (Eds.), *Somaclonal Variation and Induced Mutations in Crop Improvement*. (pp. 15–37). Springer, Dordrecht. https://doi.org/https://doi.org/10.1007/978-94-015-9125-6_2
- Briens, M., & Larher, F. (1982). Osmoregulation in halophytic higher plants: A comparative study of soluble carbohydrates, polyols, betaines and free proline. *Plant, Cell & Environment*, 5(4), 287–292. <https://doi.org/10.1111/1365-3040.EP11572682>
- Bui, E. N. (2017). Causes of soil salinization, sodification, and alkalinization. In *Oxford Research Encyclopedia of Environmental Science*. Oxford University Press. <https://doi.org/10.1093/acrefore/9780199389414.013.264>
- Butcher, K., Wick, A. F., Desutter, T., Chatterjee, A., & Harmon, J. (2016). Soil salinity: A threat to global food security. *Agronomy Journal*, 108(6), 2189–2200. <https://doi.org/10.2134/agronj2016.06.0368>
- Campino, I. D. L., Lopez, N. M., & Hiencaapie, A. M. V. (2020). Selection of somaclonal variants of maracuya (*Passiflora edulis* var Flavicarpa. Deneger) tolerant to water deficit. *Revista Colombiana de Biotecnología*, 22(2), 44–52. <https://doi.org/10.15446/REV.COLOMB.BIOTE.V22N2.79623>
- Che Omar, S., Shaharudin, A., & Tumin, S. A. (2019). The status of the paddy and rice industry in Malaysia. In *Khazanah Research Institute*. http://www.krinstitute.org/assets/contentMS/img/template/editor/20190409_RiceReport_Full Report_Final.pdf
- Chen, T. H., & Murata, N. (2002). Enhancement of tolerance of abiotic stress by metabolic engineering of betaines and other compatible solutes. *Current Opinion in Plant Biology*, 5, 250–257. [https://doi.org/https://doi.org/10.1016/s1369-5266\(02\)00255-8](https://doi.org/https://doi.org/10.1016/s1369-5266(02)00255-8)
- Chen, Tianxiao, Shabala, S., Niu, Y., Chen, Z.-H., Shabala, L., Meinke, H., Venkataraman, G., Pareek, A., X, J., & Zhou, M. (2021). Molecular mechanisms of salinity tolerance in rice. *The Crop Journal*, 9, 506–520. <https://doi.org/https://doi.org/10.1016/j.cj.2021.03.005>
- Chen, Tingting, Zhao, X., Zhang, C., Yang, Y., Feng, B., Zhang, X., Fu, G., & Tao, L. (2017). Application of salicylic acid improves filling of inferior grains of rice during late maturity under mild cold stress. *Crop Science*, 57(4), 2130–2142. <https://doi.org/10.2135/cropsci2016.11.0941>

- Cheng, Y. W., Kong, X. W., Wang, N., Wang, T. T., Chen, J., & Shi, Z. Q. (2020). Thymol confers tolerance to salt stress by activating anti-oxidative defense and modulating Na⁺ homeostasis in rice root. *Ecotoxicology and Environmental Safety*, 188(October 2019), 109894. <https://doi.org/10.1016/j.ecoenv.2019.109894>
- Childs, N. W. (2021). *Rice Outlook: May 2021*. <https://www.ers.usda.gov/webdocs/outlooks/101196/rcs-21d.pdf?v=6409.4>
- Childs, N. W., & Lebeau, B. (2021). *Rice Outlook: December 2021*. <https://downloads.usda.library.cornell.edu/usda-esmis/files/dn39x152w/6q183m76f/x059d9866/rcs-21k.pdf>
- Chinnusamy, V., Jagendorf, A., & Zhu, J.-K. (2005). Understanding and improving salt tolerance in plants. *Crop Science*, 45(2), 437–448. <https://doi.org/10.2135/CROPSCI2005.0437>
- Colomba, E. L., Grunberg, K., Griffa, S., Ribotta, A., Mroginski, L., & Biderbost, E. (2006). The effect of genotype and culture medium on somatic embryogenesis and plant regeneration from mature embryos of fourteen apomictic cultivars of buffel grass (*Cenchrus ciliaris* L.). *Grass and Forage Science*, 61(1), 2–8. <https://doi.org/10.1111/j.1365-2494.2006.00499.x>
- Dalila, Z. D., Jaafar, H., & Manaf, A. A. (2013). Effects of 2,4-D and kinetin on callus induction of *Barringtonia racemosa* leaf and endosperm explants in different types of basal media. *Asian Journal of Plant Sciences*, 12(1), 21–27. <https://doi.org/10.3923/ajps.2013.21.27>
- Dar, M. H., Bano, D. A., Waza, S. A., Zaidi, N. W., Majid, A., Shikari, A. B., Ahangar, M. A., Hossain, M., Kumar, A., & Singh, U. S. (2021). Abiotic stress tolerance - Progress and pathways of sustainable rice production. *Sustainability*, 13(4), 2078. <https://doi.org/10.3390/SU13042078>
- Das, K., Roychoudhury, A., & Anjum, N. A. (2014). Reactive oxygen species (ROS) and response of antioxidants as ROS-scavengers during environmental stress in plants. *Frontiers in Environmental Science*, 2, 53. <https://doi.org/10.3389/fenvs.2014.00053>
- Das, P., Nutan, K. K., Singla-Pareek, S. L., & Pareek, A. (2015). Understanding salinity responses and adopting “omics-based” approaches to generate salinity tolerant cultivars of rice. *Frontiers in Plant Science*, 6, 712. <https://doi.org/10.3389/fpls.2015.00712>
- Decros, G., Baldet, P., Beauvoit, B., Stevens, R., Flandin, A., Colombié, S., Gibon, Y., & Pétriacoq, P. (2019). Get the balance right: ROS homeostasis and redox signalling in fruit. *Frontiers in Plant Science*, 10, 1091. <https://doi.org/10.3389/fpls.2019.01091>
- Deepthi, V. P. (2018). Somaclonal variation in micropropagated banana.

Advances in Plants & Agriculture Research, 8(6), 624–627.
<https://doi.org/https://doi.org/10.15406/apar.2018.08.00395>

- Dehnavi, A. R., Zahedi, M., Ludwiczak, A., Perez, S. C., & Piernik, A. (2020). Effect of salinity on seed germination and seedling development of sorghum (*Sorghum bicolor* L. Moench) genotypes. *Agronomy*, 10(6), 859. <https://doi.org/10.3390/agronomy10060859>
- Demiral, T., & Turkan, I. (2005). Comparative lipid peroxidation, antioxidant defense systems and proline content in roots of two rice cultivars differing in salt tolerance. *Environmental and Experimental Botany*, 53(3), 247–257. <https://doi.org/10.1016/j.envexpbot.2004.03.017>
- DOSM. (2020). *Padi*. https://www.dosm.gov.my/v1/uploads/files/3_Time_Series/Malaysia_Time_Series_2019/09_Padi.pdf
- Dramalis, C., Katsantonis, D., & Koutroubas, S. D. (2021). Rice growth, assimilate translocation and grain quality in response to salinity under Mediterranean conditions. *AIMS Agriculture and Food*, 6(1), 255–272. <https://doi.org/10.3934/AGRFOOD.2021017>
- Dunand, R., & Saichuk, J. (2014). Rice growth and development. In *Louisiana Rice Production Handbook* (Vol. 2321, pp. 41–53). Louisiana State University AgCenter.
- El-Tayeb, M. A. (2005). Response of barley grains to the interactive effect of salinity and salicylic acid. *Plant Growth Regulation*, 45, 215–224. <https://doi.org/10.1007/s10725-005-4928-1>
- Esa, N., Puteh, A., Mat, M., Ismail, R., & Yusop, M. R. (2020). Increasing yield of susceptible and resistant rice blast cultivars using silicon fertilization. *Indonesian Journal of Agricultural Science*, 21(2), 49. <https://doi.org/10.21082/ijas.v21n2.2020.p49-58>
- Esechie, H. A. (1994). Interaction of salinity and temperature on the germination of sorghum. *Journal of Agronomy and Crop Science*, 172(3), 194–199. <https://doi.org/10.1111/j.1439-037X.1994.tb00166.x>
- FAO. (2020). *FAOSTAT - Crops and livestock products*. <http://www.fao.org/faostat/en/#data/QCL/visualize>
- Farhat, S., Jain, N., Singh, N., Sreevathsa, R., Dash, P. K., Rai, R., Yadav, S., Kumar, P., Sarkar, A. K., Jain, A., Singh, N. K., & Rai, V. (2019). CRISPR-Cas9 directed genome engineering for enhancing salt stress tolerance in rice. *Seminars in Cell & Developmental Biology*, 96, 91–99. <https://doi.org/10.1016/J.SEMCDB.2019.05.003>
- Fatikhasari, Z., & Rachmawati, D. (2020). Growth and oxidative defense response to silicon application on rice (*Oryza sativa* L. 'Sembada Merah') under salinity stress. *AIP Conference Proceedings*, 2260(030021), 1–9.

- Feher, A., Pasternak, T. P., & Dudits, D. (2003). Transition of somatic plant cells to an embryogenic state. *Plant Cell, Tissue and Organ Culture*, 74, 201–228. <https://doi.org/10.1023/A>
- Ferdosi, M. F. H., Shoaib, A., Habib, S., & Khan, K. A. (2021). Modulation of salt-induced stress impact in *Gladiolus grandiflorus* L. by exogenous application of salicylic acid. *Scientific Reports*, 11, 15597. <https://doi.org/10.1038/s41598-021-95243-9>
- Firdaus, R. B. R., Leong Tan, M., Rahyla Rahmat, S., & Senevi Gunaratne, M. (2020). Paddy, rice and food security in Malaysia: A review of climate change impacts. *Cogent Social Sciences*, 6(1), 1818373. <https://doi.org/10.1080/23311886.2020.1818373>
- Flowers, T., & Colmer, T. (2008). Salinity tolerance in halophytes. *New Phytologist*, 179, 945–963. <https://doi.org/10.1111/j.1469-8137.2008.02531.x>
- Flowers, T., Troke, P. F., & Yeo, A. R. (1977). The mechanism of salt tolerance in halophytes. *Annual Review of Plant Physiology*, 28(1), 89–121. <https://doi.org/10.1146/ANNUREV.PP.28.060177.000513>
- Fogliatto, S., Serra, F., Patrucco, L., Milan, M., & Vidotto, F. (2019). Effect of different water salinity levels on the germination of imazamox-resistant and sensitive weedy rice and cultivated rice. *Agronomy*, 9, 658. <https://doi.org/10.3390/agronomy9100658>
- Foolad, M. R., Subbiah, P., & Zhang, L. (2007). Common QTL affect the rate of tomato seed germination under different stress and nonstress conditions. *International Journal of Plant Genomics*, 2007, 97386. <https://doi.org/10.1155/2007/97386>
- Fricke, W. (2020). Energy costs of salinity tolerance in crop plants: night-time transpiration and growth. *New Phytologist*, 225, 1152–1165. <https://doi.org/10.1111/nph.15773>
- Gaj, M. (2004). Factors influencing somatic embryogenesis induction and plant regeneration with particular reference to *Arabidopsis thaliana* L. Heynh. *Plant Growth Regulation*, 43, 27–47. <https://doi.org/http://dx.doi.org/10.1023/B:GROW.0000038275.29262.fb>
- Garcia, C., Furtado de Almeida, A. A., Costa, M., Britto, D., Valle, R., Royaert, S., & Marelli, J. P. (2019). Abnormalities in somatic embryogenesis caused by 2,4-D: An overview. *Plant Cell, Tissue and Organ Culture*, 137(2), 193–212. <https://doi.org/10.1007/s11240-019-01569-8>
- Ge, S., Sang, T., Lu, B.-R., & Hong, D.-Y. (1999). Phylogeny of rice genomes with emphasis on origins of allotetraploid species. *Proceedings of the National Academy of Sciences*, 96(25), 14400–14405. <https://doi.org/10.1073/PNAS.96.25.14400>

- Gemes, K., Poór, P., Sulyok, Z., Szepesi, Á., Szabó, M., & Tari, I. (2008). Role of salicylic acid pre-treatment on the photosynthetic performance of tomato plants (*Lycopersicon esculentum* Mill. L. cvar. Rio Fuego) under salt stress. *Acta Biologica Szegediensis*, 52(1), 161–162.
- Gerona, M. E. B., Deocampo, M. P., Egdane, J. A., Ismail, A. M., & Dionisio-Sese, M. L. (2019). Physiological responses of contrasting rice genotypes to salt stress at reproductive stage. *Rice Science*, 26(4), 207–219. <https://doi.org/10.1016/j.rsci.2019.05.001>
- Ghassemi, F., Jakeman, A. J., & Nix, H. A. (1995). *Salinisation of land and water resources: Human causes, extent, management and case studies*. CAB International. <https://www.cabdirect.org/cabdirect/abstract/19976767459>
- Ghosh, B., Nasim, A. M., & Gantait, S. (2016). Response of rice under salinity stress: A review update. *Rice Research: Open Access*, 4(2), 1–8. <https://doi.org/10.4172/2375-4338.1000167>
- Gill, S. S., & Tuteja, N. (2010). Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology and Biochemistry*, 48(12), 909–930. <https://doi.org/10.1016/j.plaphy.2010.08.016>
- Gomez, J. F., Talle, B., & Wilson, Z. A. (2015). Anther and pollen development: A conserved developmental pathway. *Journal of Integrative Plant Biology*, 57, 876–891. <https://doi.org/10.1111/jipb.12425>
- Gou, W., Zheng, P., Zheng, P., Wang, K., Zhang, L., & Akram, N. A. (2016). Salinity-induced callus browning and re-differentiation, root formation by plantlets and anatomical structures of plantlet leaves in two *Malus* species. *Pakistan Journal of Botany*, 48(4), 1393–1398. <https://inis.iaea.org/search/searchsinglerecord.aspx?recordsFor=SingleRecord&RN=47116025>
- Grattan, S. R., Zeng, L., Shannon, M. C., & Roberts, S. R. (2002). Rice is more sensitive to salinity than previously thought. *California Agriculture*, 56(6), 189–198. <https://doi.org/10.3733/ca.v056n06p189>
- Green, T. G. A., Sancho, L. G., Pintado, A., Saco, D., Martin, S., Arroniz-Crespo, M., Casermeiro, M. A., Caravaca, M. T. de la C., Cameron, S., & Rozzi, R. (2017). Sodium chloride accumulation in glycophyte plants with cyanobacterial symbionts. *AoB Plants*, 9(6), plx053. <https://doi.org/https://doi.org/10.1093/aobpla/plx053>
- Gregorio, G. B., Senadhira, D., & Mendoza, R. D. (1997). Screening rice for salinity tolerance. In *Development* (22, Vol. 22). http://rkb.irri.org/ricebreedingcourse/documents/Screening_manual.pdf
- Guo, Y., Wu, W., Liu, Y., Wu, Z., Geng, X., Zhang, Y., Bryant, C. R., & Fu, Y. (2020). Impacts of climate and phenology on the yields of early mature rice

in china. *Sustainability*, 12(23), 1–16. <https://doi.org/10.3390/su122310133>

- Gupta, B., & Huang, B. (2014). Mechanism of salinity tolerance in plants: Physiological, biochemical, and molecular characterization. *International Journal of Genomics*, 2014, 701596. <https://doi.org/https://doi.org/10.1155/2014/701596>
- Hameg, R., Arteta, T. A., Landin, M., Gallego, P. P., & Barreal, M. E. (2020). Modeling and optimizing culture medium mineral composition for in vitro propagation of *Actinidia arguta*. *Frontiers in Plant Science*, 11, 554905. <https://doi.org/10.3389/FPLS.2020.554905>
- Hanin, M., Ebel, C., Ngom, M., Laplaze, L., & Masmoudi, K. (2016). New insights on plant salt tolerance mechanisms and their potential use for breeding. *Frontiers in Plant Science*, 7, 1787. <https://doi.org/10.3389/fpls.2016.01787>
- Hannachi, S., Werbrouck, S., Bahrini, I., Abdelgadir, A., Siddiqui, H. A., Christine, M., Labeke, V., Llugany, M., Roussos, P. A., & Cheema, M. (2021). Obtaining salt stress-tolerant eggplant somaclonal variants from *in vitro* selection. *Plants*, 10, 2539. <https://doi.org/10.3390/plants10112539>
- Hare, P. D., Cress, W. A., & Van Staden, J. (1998). Dissecting the roles of osmolyte accumulation during stress. *Plant, Cell and Environment*, 21(6), 535–553. <https://doi.org/10.1046/J.1365-3040.1998.00309.X>
- Hasanuzzaman, M., Borhannuddin Bhuyan, M. H. M., Anee, T. I., Parvin, K., Nahar, K., Al Mahmud, J., & Fujita, M. (2019). Regulation of ascorbate-glutathione pathway in mitigating oxidative damage in plants under abiotic stress. *Antioxidants*, 8, 384. <https://doi.org/10.3390/antiox8090384>
- Hasanuzzaman, M., Borhannuddin Bhuyan, M. H. M., Zulfiqar, F., Raza, A., Mohsin, S. M., Al Mahmud, J., Fujita, M., & Fotopoulos, V. (2020). Reactive oxygen species and antioxidant defense in plants under abiotic stress: Revisiting the crucial role of a universal defense regulator. *Antioxidants*, 9, 681. <https://doi.org/10.3390/antiox9080681>
- Hasanuzzaman, M., Hossain, M. A., Silva, J. A. T. da, & Fujita, M. (2012). Plant response and tolerance to abiotic oxidative stress: Antioxidant defense is a key factor. In M. M. Venkateswarlu B., Shanker A., Shanker C. (Ed.), *Crop Stress and its Management: Perspectives and Strategies* (pp. 261–315). Springer New York LLC. <https://doi.org/10.1007/978-94-007-2220-0>
- Hasegawa, P. M., Bressan, R. A., Zhu, J.-K., & Bohnert, H. J. (2000). Plant cellular and molecular responses to high salinity. *Annual Review of Plant Physiology and Plant Molecular Biology*, 51, 463–499. <https://doi.org/10.1146/ANNUREV.ARPLANT.51.1.463>
- Hasnan, H. A. (2021, November 23). Parlimen: Sektor agromakanan berdepan 5 cabaran utama - MAFI. *Astro Awani*. <https://www.astroawani.com/berita->

- Hassani, A., Azapagic, A., & Shokri, N. (2020). Predicting long-term dynamics of soil salinity and sodicity on a global scale. *Proceedings of the National Academy of Sciences of the United States of America*, 117(52), 33017–33027. <https://doi.org/10.1073/PNAS.2013771117>
- Heath, R. L., & Packer, L. (1968). Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. *Archives of Biochemistry and Biophysics*, 125(1), 189–198. [https://doi.org/10.1016/0003-9861\(68\)90654-1](https://doi.org/10.1016/0003-9861(68)90654-1)
- Herman, T., Warsi, A., Murchie, E., & Warsi, A. A. (2015). Rice production and climate change: A case study of Malaysian rice. *Pertanika Journal of Tropical Agricultural Science*, 38(3), 321–328. [http://www.pertanika.upm.edu.my/resources/files/Pertanika PAPERS/JTAS Vol. 38 \(3\) Aug. 2015/02 JTAS 0670-2014 \(Short Comm\).pdf](http://www.pertanika.upm.edu.my/resources/files/Pertanika_PAPERS/JTAS_Vol.38_(3)_Aug.2015/02_JTAS_0670-2014_(ShortComm).pdf)
- Hiei, Y., Ohta, S., Komari, T., & Kumashiro, T. (1994). Efficient transformation of rice (*Oryza sativa* L.) mediated by *Agrobacterium* and sequence analysis of the boundaries of the T-DNA. *The Plant Journal*, 6(2), 271–282. <https://doi.org/10.1046/J.1365-313X.1994.6020271.X/FORMAT/PDF>
- Hoang, Thi M.L., Moghaddam, L., Williams, B., Khanna, H., Dale, J., & Mundree, S. G. (2015). Development of salinity tolerance in rice by constitutive-overexpression of genes involved in the regulation of programmed cell death. *Frontiers in Plant Science*, 6(March), 1–14. <https://doi.org/10.3389/fpls.2015.00175>
- Hoang, Thi My Linh, Tran, T. N., Kieu, T., Nguyen, T., Williams, B., Wurm, P., Bellairs, S., & Mundree, S. (2016). Improvement of salinity stress tolerance in rice: Challenges and opportunities. *Agronomy*, 6(4), 54. <https://doi.org/10.3390/agronomy6040054>
- Hossain, M. A., Razi Ismail, M., Kamal Uddin, M., Islam, M. ., & Ashrafuzzaman, M. (2013). Efficacy of ascorbate-glutathione cycle for scavenging H₂O₂ in two contrasting rice genotypes during salinity stress. *Australian Journal of Crop Science*, 7(12), 1801–1808.
- IADA. (2015). *Kaedah penggunaan manual tanaman padi*. Kawasan Pembangunan Pertanian Bersepadu. <http://iadapp.gov.my/Web/Page/Default.aspx?Page=KaedahTanamanPadi-2>
- Ijaz, B., Formentin, E., Ronci, B., Locato, V., Barizza, E., Hyder, M. Z., Lo Schiavo, F., & Yasmin, T. (2019). Salt tolerance in *indica* rice cell cultures depends on a fine tuning of ROS signalling and homeostasis. *PLOS ONE*, 14(4), e0213986. <https://doi.org/10.1371/journal.pone.0213986>

- Ikeuchi, M., Sugimoto, K., & Iwase, A. (2013). Plant callus: Mechanisms of induction and repression. *Plant Cell*, 25(9), 3159–3173. <https://doi.org/10.1105/tpc.113.116053>
- Iraokeze, W., Prodjimoto, H., Nijimbere, S., Rufyikiri, G., & Lutts, S. (2020). NaCl and Na₂SO₄ salinities have different impact on photosynthesis and yield-related parameters in rice (*Oryza sativa* L.). *Agronomy*, 10, 864. <https://doi.org/10.3390/agronomy10060864>
- Irigoyen, J. J., Einerich, D. W., & Sanchez-Diaz, M. (1992). Water stress induced changes in concentrations of proline and total soluble sugars in nodulated alfalfa (*Medicago sativa*) plants. *Physiologia Plantarum*, 84(1), 55–60. <https://doi.org/10.1111/j.1399-3054.1992.tb08764.x>
- IRRI. (2019). Race for impact. In *Annual Report 2019*. http://books.irri.org/AR2019_content.pdf
- Ismail, A. M., & Horie, T. (2017). Genomics, physiology and molecular breeding approaches for improving salt tolerance. *The Annual Review of Plant Biology*, 68, 405–434. <https://doi.org/10.1146/annurev-arplant-042916>
- Ismail, A. M., Singh, U. S., Singh, S., Dar, M. H., & Mackill, D. J. (2013). The contribution of submergence-tolerant (Sub1) rice varieties to food security in flood-prone rainfed lowland areas in Asia. *Field Crops Research*, 152, 83–93. <https://doi.org/10.1016/j.fcr.2013.01.007>
- Ivushkin, K., Bartholomeus, H., Bregt, A. K., Pulatov, A., Kempen, B., & De Sousa, L. (2019). Global mapping of soil salinity change. *Remote Sensing of Environment*, 231, 111260. <https://doi.org/10.1016/j.rse.2019.111260>
- Jamali, B., Eshghi, S., & Shahidi-Rad, K. (2015). Growth and fruit characteristics of strawberry cv. Selva as affected by different application timing of salicylic acid under saline conditions. *International Journal of Fruit Science*, 15(3), 339–352. <https://doi.org/10.1080/15538362.2015.1015761>
- Jamaluddin, U. A., Lim, C. S., & Pereira, J. J. (2018). Implikasi perubahan iklim terhadap zon pesisir pantai di Kuala Selangor, Malaysia. *Bulletin of the Geological Society of Malaysia*, 66, 107–119. http://ancst.org/apn/wp-content/uploads/2019/03/702001-101749-PDF_2.pdf
- James, R. A., Blake, C., Byrt, C. S., & Munns, R. (2011). Major genes for Na⁺ exclusion, Nax1 and Nax2 (wheat HKT1;4 and HKT1;5), decrease Na⁺ accumulation in bread wheat leaves under saline and waterlogged conditions. *Journal of Experimental Botany*, 62(8), 2939–2947. <https://doi.org/https://doi.org/10.1093/jxb/err003>
- Jamil, A., Riaz, S., Ashraf, M., & Foolad, M. R. (2011). Gene expression profiling of plants under salt stress. *Critical Reviews in Plant Sciences*, 30(5), 435–458. <https://doi.org/10.1080/07352689.2011.605739>

- Jamil, M., Lee, D. B., Jung, K. Y., Ashraf, M., Lee, S. C., & Rha, E. S. (2006). Effect of salt (NaCl) stress on germination and early seedling growth of four vegetables species. *Pure and Applied Biology*, 7(2), 273–282. <https://doi.org/10.19045/bspab.2018.700218>
- Janda, T., Horvath, E., Szalai, G., & Paldi, E. (2006). Role of salicylic acid in the induction of abiotic stress tolerance. In S. Hayat & A. Ahmad (Eds.), *Salicylic Acid: A Plant Hormone* (pp. 91–150). Springer. https://doi.org/10.1007/3-540-30223-9_6
- Jimenez, Victor M., & Thomas, C. (2005). Participation of plant hormones in determination and progression of somatic embryogenesis. *Plant Cell Monographs*, 2, 103–118. https://doi.org/10.1007/7089_034
- Jimenez, Victor M, & Bangerth, F. (2001). Endogenous hormone levels in explants and in embryogenic and non-embryogenic cultures of carrot. *Physiologia Plantarum*, 111, 389–395.
- Jimenez, Victor M. (2005). Involvement of plant hormones and plant growth regulators on *in vitro* somatic embryogenesis. *Plant Growth Regulation*, 47, 91–110. <https://doi.org/10.1007/s10725-005-3478-x>
- Jini, D., & Joseph, B. (2017a). Physiological mechanism of salicylic acid for alleviation of salt stress in rice. *Rice Science*, 24(2), 97–108. <https://doi.org/10.1016/j.rsci.2016.07.007>
- Jini, D., & Joseph, B. (2017b). Salicylic acid mediated salt tolerance at different growth stages of *Oryza sativa* L. and its effect on salicylic acid biosynthetic pathway genes. *BioTechnology: An Indian Journal*, 13(2), 134. <https://www.tsijournals.com/articles/salicylic-acid-mediated-salt-tolerance-at-different-growth-stages-of-oryza-sativa-l-and-its-effect-on-salicylic-acid-biosynthetic-.html>
- Jones, A. M. P., & Saxena, P. K. (2013). Inhibition of phenylpropanoid biosynthesis in *Artemisia annua* L.: A novel approach to reduce oxidative browning in plant tissue culture. *PLoS ONE*, 8(10), e76802. <https://doi.org/https://dx.doi.org/10.1371/journal.pone.0076802>
- Kader, A., & Lindberg, S. (2010). Cytosolic calcium and pH signaling in plants under salinity stress. *Plant Signaling & Behavior*, 5(3), 233–238. <https://doi.org/10.4161/psb.5.3.10740>
- Kakar, N., Jumaa, S. H., Redoña, E. D., Warburton, M. L., & Reddy, K. R. (2019). Evaluating rice for salinity using pot-culture provides a systematic tolerance assessment at the seedling stage. *Rice*, 12(1), 57. <https://doi.org/10.1186/s12284-019-0317-7>
- Kalhuri, N., Nulit, R., Go, R., Zulkifly, S., Azizi, P., & Abiri, R. (2017). Selection, characterizations and somatic embryogenesis of Malaysian salt-tolerant rice (*Oryza sativa* cv. MR219) through callogenesis. *International Journal*

of *Agriculture and Biology*, 19(1), 157–163.
<https://doi.org/10.17957/IJAB/15.0258>

- Kanawapee, N., Sanitchon, J., Srihaban, P., & Theerakulpisut, P. (2013). Physiological changes during development of rice (*Oryza sativa* L.) varieties differing in salt tolerance under saline field condition. *Plant and Soil*, 370, 89–101. <https://doi.org/10.1007/s11104-013-1620-5>
- Karimian, R., Lahouti, M., & Davarpanah, S. J. (2014). Effects of different concentrations of 2,4-D and kinetin on callogenesis of *Taxus revifolia* Nutt. *Journal of Applied Biotechnology Reports*, 1(4), 167–170.
- Karthikeyan, A., Pandian, S. T. K., & Ramesh, M. (2009). High frequency plant regeneration from embryogenic callus of a popular indica rice (*Oryza sativa* L.). *Physiology and Molecular Biology of Plants*, 15(4), 371–375. <https://doi.org/10.1007/s12298-009-0042-6>
- Karthikeyan, A., Thevar, S., Pandian, K., & Ramesh, M. (2009). AD Physiol. Mol. Biol. Plants. *Physiology and Molecular Biology of Plants*, 15(4), 371–375.
- Kerepesi, I., & Galiba, G. (2000). Osmotic and salt stress-induced alteration in soluble carbohydrate content in wheat seedlings. *Crop Science*, 40, 482–487. <https://doi.org/https://doi.org/10.2135/cropsci2000.402482x>
- Khalvandi, M., Siosemardeh, A., Roohi, E., & Keramati, S. (2021). Salicylic acid alleviated the effect of drought stress on photosynthetic characteristics and leaf protein pattern in winter wheat. In *Heliyon* (Vol. 7, Issue 1). <https://doi.org/10.1016/j.heliyon.2021.e05908>
- Khan, M. S. A., Hamid, A., Salahuddin, A. B. M., Quasem, A., & Karim, M. A. (1997). Effect of sodium chloride on growth, photosynthesis and mineral ions accumulation of different types of rice (*Oryza sativa* L.). *Journal of Agronomy and Crop Science*, 179(3), 149–161. <https://doi.org/10.1111/j.1439-037X.1997.tb00511.x>
- Khan, M. S., Akther, T., Ali, D. M., & Hemalatha, S. (2019). An investigation on the role of salicylic acid alleviate the saline stress in rice crop (*Oryza sativa* (L)). *Biocatalysis and Agricultural Biotechnology*, 18, 101027. <https://doi.org/10.1016/j.bcab.2019.101027>
- Khan, S., Javed, M. A., Jahan, N., & Manan, F. A. (2016). A short review on the development of salt tolerant cultivars in rice. *International Journal of Public Health Science*, 5(2), 201–212. <https://doi.org/10.11591/ijphs.v5i2.4786>
- Khatun, S., Rizzo, C. A., & Flowers, T. J. (1995a). Genotypic variation in the effect of salinity on fertility in rice. *Plant and Soil*, 173(2), 239–250. <https://doi.org/10.1007/BF00011461>
- Khatun, S., Rizzo, C., & Flowers, T. (1995b). Genotypic variation in the effect of salinity on fertility in rice. In *Plant and Soil* (Vol. 173). Kluwer Academic

Publishers. <https://link.springer.com/content/pdf/10.1007/BF00011461.pdf>

- Khush, G. S. (1997). Origin, dispersal, cultivation and variation of rice. *Plant Molecular Biology*, 35, 25–34. <https://doi.org/https://doi.org/10.1023/A:1005810616885>
- Kibria, M. G., Hossain, M., Murata, Y., & Hoque, A. (2017). Antioxidant defense mechanisms of salinity tolerance in rice genotypes. *Rice Science*, 24(3), 155–162. <https://doi.org/10.1016/j.rsci.2017.05.001>
- Kim, Y.-H., Khan, A. L., Kim, D.-H., Lee, S.-Y., Kim, K.-M., Waqas, M., Jung, H.-Y., Shin, J.-H., Kim, J.-G., & Lee, I.-J. (2014). Silicon mitigates heavy metal stress by regulating P-type heavy metal ATPases, *Oryza sativa* low silicon genes, and endogenous phytohormones. *BMC Plant Biology*, 14(1), 1–13. <https://doi.org/10.1186/1471-2229-14-13>
- Kim, Y., Mun, B. G., Khan, A. L., Waqas, M., Kim, H. H., Shahzad, R., Imran, M., Yun, B. W., & Lee, I. J. (2018). Regulation of reactive oxygen and nitrogen species by salicylic acid in rice plants under salinity stress conditions. *PLoS ONE*, 13(3), 1–20. <https://doi.org/10.1371/journal.pone.0192650>
- Kobata, T. (2013). Spikelet Sterility is Associated with a Lack of Assimilate in High-Spikelet-Number Rice. *Article in Agronomy Journal*. <https://doi.org/10.2134/agronj2013.0115>
- Kolachevskaya, O. O., Myakushina, Y. A., Getman, I. A., Lomin, S. N., Deyneko, I. V., Deigraf, S. V., & Romanov, G. A. (2021). Hormonal regulation and crosstalk of auxin/cytokinin signaling pathways in potatoes in vitro and in relation to vegetation or tuberization stages. *International Journal of Molecular Sciences*, 22(15), 8207. <https://doi.org/10.3390/IJMS22158207/S1>
- Konar, S., Karmakar, J., Ray, A., Adhikari, S., & Bandyopadhyay, T. K. (2018). Regeneration of plantlets through somatic embryogenesis from root derived calli of *Hibiscus sabdariffa* L. (Roselle) and assessment of genetic stability by flow cytometry and ISSR analysis. *PLoS ONE*, 13(8), 1–17. <https://doi.org/10.1371/journal.pone.0202324>
- Kotula, L., Garcia Caparros, P., Zörb, C., Colmer, T. D., & Flowers, T. J. (2020). Improving crop salt tolerance using transgenic approaches: An update and physiological analysis. *Plant Cell and Environment*, 43(12), 2932–2956. <https://doi.org/10.1111/PCE.13865>
- Krishna, H., Alizadeh, M., Singh, D., Singh, U., Chauhan, N., Eftekhari, M., & Kishan Sadh, R. (2016). Somaclonal variations and their applications in horticultural crops improvement. 3 *Biotech*, 6, 54. <https://doi.org/10.1007/s13205-016-0389-7>
- Krishnamurthy, S. L., Pundir, P., Singh Warraich, A., Rathor, S., Lokeshkumar, B. M., Singh, N. K., & Sharma, P. C. (2020). Introgressed Saltol QTL lines

improves the salinity tolerance in rice at seedling stage. *Frontiers in Plant Science*, 11, 833. <https://doi.org/10.3389/fpls.2020.00833>

Krishnamurthy, S. L., Sharma, P. C., Sharma, S. K., Batra, V., Kumar, V., & Rao, L. V. S. (2016). Effect of salinity and use of stress indices of morphological and physiological traits at the seedling stage in rice. *Indian Journal of Experimental Biology*, 54(12), 843–850.

Krishnan, S. R. S., & Siril, E. A. (2017). Auxin and nutritional stress coupled somatic embryogenesis in *Oldenlandia umbellata* L. *Physiology and Molecular Biology of Plants*, 23(2), 471–475. <https://doi.org/10.1007/s12298-017-0425-z>

Larher, F., J, H., & GR, S. (1977). 3-Dimethylsulfoniumpropanoic acid from *Spartina anglica*. *Phytochemistry*, 14, 205–207.

Larkin, P. J., & Scowcroft, W. R. (1981). Somaclonal variation - a novel source of variability from cell cultures for plant improvement. *Theoretical and Applied Genetics*, 60, 197–214. <https://doi.org/10.1007/BF02342540>

Le Bris, M. (2017). Hormones in growth and development. In *Reference Module in Life Sciences* (pp. 364–369). Elsevier. <https://doi.org/10.1016/b978-0-12-809633-8.05058-5>

Lefevre, I., Gratia, E., & Lutts, S. (2001). Discrimination between the ionic and osmotic components of salt stress in relation to free polyamine level in rice (*Oryza sativa*). *Plant Science*, 161(5), 943–952. [https://doi.org/10.1016/S0168-9452\(01\)00485-X](https://doi.org/10.1016/S0168-9452(01)00485-X)

Li, R., Li, M., Ashraf, U., Liu, S., & Zhang, J. (2019). Exploring the relationships between yield and yield-related traits for rice varieties released in China from 1978 to 2017. *Frontiers in Plant Science*, 10, 543. <https://doi.org/10.3389/fpls.2019.00543>

Lin, Y., & Zhang, Q. (2005). Optimising the tissue culture conditions for high efficiency transformation of indica rice. *Plant Cell Reports*, 23, 540–547. <https://doi.org/10.1007/s00299-004-0843-6>

Liu, J., Yang, Z., Li, W., Yu, J., & Huang, B. (2013). Improving cold tolerance through *in vitro* selection for somaclonal variations in seashore paspalum. *Journal of the American Society for Horticultural Science*, 138(6), 452–460. <https://doi.org/10.21273/jashs.138.6.452>

Liu, L., Xia, W., Li, H., Zeng, H., Wei, B., Han, S., & Yin, C. (2018). Salinity inhibits rice seed germination by reducing α -amylase activity via decreased bioactive gibberellin content. *Frontiers in Plant Science*, 9, 275. <https://doi.org/10.3389/fpls.2018.00275>

Liu, M., Yu, H., Ouyang, B., Shi, C., Demidchik, V., Hao, Z., Yu, M., & Shabala, S. (2020). NADPH oxidases and the evolution of plant salinity tolerance.

Plant Cell and Environment, 43(12), 2957–2968.
<https://doi.org/10.1111/PCE.13907>

- Liu, Z., Lin, S., Shi, J., Yu, J., Zhu, L., Yang, X., Zhang, D., & Liang, W. (2017). Rice No Pollen 1 (NP1) is required for anther cuticle formation and pollen exine patterning. *The Plant Journal*, 91, 263–277.
<https://doi.org/10.1111/tj.13561>
- Lokhande, V. H., & Suprasanna, P. (2012). Prospects of halophytes in understanding and managing abiotic stress tolerance. In *Environmental Adaptations and Stress Tolerance of Plants in the Era of Climate Change* (pp. 29–56). Springer, New York, NY. https://doi.org/10.1007/978-1-4614-0815-4_2
- Ludwiczak, A., Osiak, M., Cárdenas-Pérez, S., Lubińska-Mielińska, S., & Piernik, A. (2021). Osmotic stress or ionic composition: Which affects the early growth of crop species more? *Agronomy*, 11(3), 435.
<https://doi.org/10.3390/agronomy11030435>
- Ma, N. L., Lah, A. C., Kadir, W. A., Mustaqim, N., Mohamad, Rahmat, Z., Ahmad, A., Datt Lam, S., & Razi Ismail, M. (2018). Susceptibility and tolerance of rice crop to salt threat: Physiological and metabolic inspections. *PLoS ONE*, 13(2), e0192732. <https://doi.org/10.1371/journal.pone.0192732>
- Ma, X., Feng, F., Wei, H., Mei, H., Xu, K., Chen, S., Li, T., Liang, X., Liu, H., & Luo, L. (2016). Genome-wide association study for plant height and grain yield in rice under contrasting moisture regimes. *Frontiers in Plant Science*, 7, 1801. <https://doi.org/10.3389/fpls.2016.01801>
- Ma, Y., Dias, M. C., & Freitas, H. (2020). Drought and salinity stress responses and microbe-induced tolerance in plants. *Frontiers in Plant Science*, 11, 591911. <https://doi.org/10.3389/fpls.2020.591911>
- Machado, R. M. A., & Serralheiro, R. P. (2017). Soil salinity: Effect on vegetable crop growth. Management practices to prevent and mitigate soil salinization. *Horticulturae*, 3(2), 30.
<https://doi.org/10.3390/horticulturae3020030>
- MADA. (2020). *Paddy industry development program*. https://www.mada.gov.my/?page_id=13761&lang=en
- Madson, D. (2019). *Saltwater invades rice fields, damages crops*. Yale Climate Connections. <https://yaleclimateconnections.org/2019/02/saltwater-invades-rice-fields-damages-crops/>
- MAFI. (2021a). *Dasar Agromakanan Negara 2.0*. <https://www.mafi.gov.my/documents/20182/361765/Ringkasan+Eksekutif+Dasar+Agromakanan+Negara++2.0+2021-2030.pdf/5d91f8c3-df74-47fa-85bc-85cbf4bf703a>

- MAFI. (2021b). *MAFI mengalu-alukan pembangunan benih padi baharu IS21 oleh Agensi Nuklear Malaysia, MOSTI*. <https://www.mafi.gov.my/documents/20182/269754/SIARAN+MEDIA+MAFI+VARIETI+BENIH+PADI+%2820+NOVEMBER+2021%29-min.pdf/48681d83-e1c5-425f-9901-6dae0a537b7d>
- Mahdavian, K. (2018). Effect of salicylic acid spray on the leaf ascorbate, proline and quercetin content in ultra-violet stressed pepper seedlings. *Journal of Plant Process and Function*, 6(22), 55–62.
- Mahesh, H. B., Shirke, M. D., Singh, S., Rajamani, A., Hittalmani, S., Wang, G. L., & Gowda, M. (2016). *Indica* rice genome assembly, annotation and mining of blast disease resistance genes. *BMC Genomics*, 17(1), 1–12. <https://doi.org/10.1186/S12864-016-2523-7>
- Mandal, S., Raju, R., Kumar, A., Kumar, P., & Sharma, P. C. (2018). Current status of research, technology response and policy needs of salt-affected soils in India - A review. *Journal of the Indian Society of Coastal Agricultural Research*, 36(2), 40–53.
- Manful, J. T., & Graham-Acquaah, S. (2016). African rice (*Oryza glaberrima*): A brief history and its growing importance in current rice breeding efforts. *Encyclopedia of Food Grains: Second Edition*, 1–4, 140–146. <https://doi.org/10.1016/B978-0-12-394437-5.00016-4>
- MARDI. (2016). *MARDI Siraj 297: Varieti padi berhasil tinggi*. MARDI; Malaysian Agriculture Research and Development Institute. <http://padipedia.mardi.gov.my/resources/documents/slider3-document.pdf>
- MARDI. (2021, October 15). MARDI henti jual benih padi MR 219. *Berita Harian*. <https://www.bharian.com.my/berita/nasional/2021/10/876638/mardi-henti-jual-benih-padi-mr-219>
- Mariani, T. S., Miyake, H., & Takeoka, Y. (1998). Changes in surface structure during direct somatic embryogenesis in rice scutellum observed by scanning electron microscopy. *Plant Production Science*, 1(3), 223–231. <https://doi.org/10.1626/pp.s.1.223>
- Mastuti, R., Munawarti, A., & Firdiana, E. R. (2017). The combination effect of auxin and cytokinin on in vitro callus formation of *Physalis angulata* L. - A medicinal plant. *AIP Conference Proceedings*, 1908(November). <https://doi.org/10.1063/1.5012721>
- Mattioli, R., Palombi, N., Funck, D., & Trovato, M. (2020). Proline accumulation in pollen grains as potential target for improved yield stability under salt stress. *Frontiers in Plant Science*, 11, 582877. <https://doi.org/10.3389/fpls.2020.582877>
- Matysik, J. A., Bhalu, B., & Mohanty, P. (2002). Molecular mechanisms of quenching of reactive oxygen species by proline under stress in plants.

- Mendez-Hernandez, H. A., Ledezma-Rodriguez, M., Avilez-Montalvo, R. N., Juarez-Gomez, Y. L., Skeete, A., Avilez-Montalvo, J., De-La-Pena, C., & Loyola-Vargas, V. M. (2019). Signaling overview of plant somatic embryogenesis. *Frontiers in Plant Science*, 10. <https://doi.org/10.3389/fpls.2019.00077>
- Michalczuk, L., Ribnicky, D. M., Cooke, T. J., & Cohen, J. D. (1992). Regulation of indole-3-acetic acid biosynthetic pathways in carrot cell cultures. *Plant Physiology*, 100, 1346–1353.
- Miguel, C., & Marum, L. (2011). An epigenetic view of plant cells cultured in vitro: Somaclonal variation and beyond. *Journal of Experimental Botany*, 62(11), 3713–3725. <https://doi.org/10.1093/jxb/err155>
- Mishra, S., Singh, B., Panda, K., Singh, B. P., Singh, N., Misra, P., Rai, V., & Singh, N. K. (2016). Association of SNP haplotypes of HKT family genes with salt tolerance in Indian wild rice germplasm. *Rice*, 9(1), 15. <https://doi.org/10.1186/s12284-016-0083-8>
- Misra, N., & Misra, R. (2012). Salicylic acid changes plant growth parameters and proline metabolism in *Rauwolfia serpentina* leaves grown under salinity stress. *American-Eurasian Journal of Agriculture & Environmental Sciences*, 12(12), 1601–1609. <https://doi.org/10.5829/idosi.aejaes.2012.12.12.1919>
- Mittler, R. (2017). ROS are good. *Trends in Plant Science*, 22(1), 11–19. <https://doi.org/10.1016/j.tplants.2016.08.002>
- Mittova, V., Volokita, M., Guy, M., & Tal, M. (2000). Activities of SOD and the ascorbate-glutathione cycle enzymes in subcellular compartments in leaves and roots of the cultivated tomato and its wild salt-tolerant relative *Lycopersicon pennellii*. *Physiologia Plantarum*, 110, 42–51. <https://doi.org/10.1034/j.1399-3054.2000.110106.x>
- Mohd Din, A. R. J., Iliyas Ahmad, F., Wagiran, A., Abd Samad, A., Rahmat, Z., & Sarmidi, M. R. (2016a). Improvement of efficient in vitro regeneration potential of mature callus induced from Malaysian upland rice seed (*Oryza sativa* cv. Panderas). *Saudi Journal of Biological Sciences*, 23(1), S69–S77. <https://doi.org/10.1016/j.sjbs.2015.10.022>
- Moller, I. M., Jensen, P. E., & Hansson, A. (2007). Oxidative modifications to cellular components in plants. *Annual Review of Plant Biology*, 58, 459–481. <https://doi.org/10.1146/annurev.arplant.58.032806.103946>
- Mondal, S., Borromeo, T. H., Diaz, M. G. Q., Amas, J., Rahman, M. A., Thomson, M. J., & Gregorio, G. B. (2019). Dissecting QTLs for reproductive stage salinity tolerance in rice from BRR1 dhan 47. *Plant Breeding and Biotechnology*, 7(4), 302–312. <https://doi.org/https://doi.org/10.9787/PBB.2019.7.4.302>

- Mostafiz, S. B. (2019). *Analysis of embryogenic callus induction and regeneration of indica rice variety of Malaysia* [Universiti Teknologi Malaysia]. <http://eprints.utm.my/id/eprint/81396/>
- Moukhtari, A. El, Cabassa-Hourton, C., Farissi, M., & Savouré, A. (2020). How does proline treatment promote salt stress tolerance during crop plant development? *Frontiers in Plant Science*, *11*, 1127. <https://doi.org/10.3389/FPLS.2020.01127>
- Munns, R. (2005). Genes and salt tolerance: Bringing them together. *New Phytologist*, *167*(3), 645–663. <https://doi.org/10.1111/J.1469-8137.2005.01487.X>
- Munns, R., & Gilliam, M. (2015). Salinity tolerance of crops - what is the cost? *New Phytologist*, *208*(3), 668–673. <https://doi.org/10.1111/NPH.13519>
- Munns, R., Passioura, J. B., Colmer, T. D., & Byrt, C. S. (2020). Osmotic adjustment and energy limitations to plant growth in saline soil. *New Phytologist*, *225*(3), 1091–1096. <https://doi.org/10.1111/nph.15862>
- Munns, R., & Tester, M. (2008). Mechanisms of salinity tolerance. *Annual Review of Plant Biology*, *59*, 651–681. <https://doi.org/10.1146/annurev.arplant.59.032607.092911>
- Murashige, T., & Skoog, F. (1962). A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiologia Plantarum*, *15*, 473–497. http://priede.bf.lu.lv/grozs/AuguFiziologijas/Augu_audu_kulturas_MAG/literatura/03_Murashige_Skoog1962.pdf
- Mutlu, S., Atici, O., Nalbantoglu, B., & Mete, E. (2016). Exogenous salicylic acid alleviates cold damage by regulating antioxidative system in two barley (*Hordeum vulgare* L.) cultivars. *Frontiers in Life Science*, *9*(2), 99–109. <https://doi.org/10.1080/21553769.2015.1115430>
- Mwando, E., Han, Y., Tolera Angessa, T., Zhou, G., Hill, C. B., Zhang, X.-Q., & Li, C. (2020). Genome-wide association study of salinity tolerance during germination in barley (*Hordeum vulgare* L.). *Frontiers in Plant Science*, *11*, 118. <https://doi.org/10.3389/fpls.2020.00118>
- Naeem, M., Basit, A., Ahmad, I., Mohamed, H. I., & Wasila, H. (2020). Effect of salicylic acid and salinity stress on the performance of tomato plants. *Gesunde Pflanzen*, *72*(4), 393–402. <https://doi.org/10.1007/s10343-020-00521-7>
- Naliwajski, M., & Skłodowska, M. (2021). The relationship between the antioxidant system and proline metabolism in the leaves of cucumber plants acclimated to salt stress. *Cells*, *10*(3), 1–15. <https://doi.org/10.3390/cells10030609>

- Nasim Ali, M., Yeasmin, L., Gantait, S., Goswami, R., & Chakraborty, S. (2014). Screening of rice landraces for salinity tolerance at seedling stage through morphological and molecular markers. *Physiology and Molecular Biology of Plants*, 20(4), 411–423. <https://doi.org/10.1007/s12298-014-0250-6>
- Negrao, S., Schmöckel, S. M., & Tester, M. (2017). Evaluating physiological responses of plants to salinity stress. *Annals of Botany*, 119(1), 1–11. <https://doi.org/10.1093/aob/mcw191>
- Niedz, R. P., & Evens, T. J. (2007). Regulating plant tissue growth by mineral nutrition. *In Vitro Cellular & Developmental Biology*, 43, 370–381. <https://doi.org/DOI 10.1007/s11627-007-9062-5>
- Niroula, R. K., Pucciariello, C., Ho, V. T., Novi, G., Fukao, T., & Perata, P. (2012). SUB1A-dependent and-independent mechanisms are involved in the flooding tolerance of wild rice species. *The Plant Journal*, 72, 282–293. <https://doi.org/10.1111/j.1365-313X.2012.05078.x>
- Nivas, S., & DSouza, L. (2014). Genetic fidelity in micropropagated plantlets of *Anacardium occidentale* L. (cashew) an important fruit tree. *International Journal of Science Research*, 3(11), 2142–2146.
- Nouri, H., Chavoshi Borujeni, S., Alaghmand, S., Anderson, S. J., Sutton, P. C., Parvazian, S., & Beecham, S. (2018). Soil salinity mapping of urban greenery using remote sensing and proximal sensing techniques; the case of Veale Gardens within the Adelaide parklands. *Sustainability*, 10, 2826. <https://doi.org/10.3390/su10082826>
- Orchard, T. J. (1977). Estimating the parameters of plant seedling emergence. *Seed Science and Technology*. <https://agris.fao.org/agris-search/search.do?recordID=US201302496360>
- Osman, H. S., & Salim, B. B. M. (2016). Influence of exogenous application of some phytoprotectants on growth, yield and pod quality of snap bean under NaCl salinity. *Annals of Agricultural Sciences*, 61(1), 1–13. <https://doi.org/10.1016/J.AOAS.2016.05.001>
- Pal, M., Szalai, G., Kovacs, V., Gondor, O. K., & Janda, T. (2013). Salicylic acid-mediated abiotic stress tolerance. In *Salicylic Acid* (pp. 183–247). Springer, Dordrecht. https://doi.org/10.1007/978-94-007-6428-6_10
- Pandey, S., & Chakraborty, D. (2015). Salicylic acid and drought stress response: Biochemical to molecular crosstalk. In B. N. Tripathi & M. Muller (Eds.), *Stress Responses in Plants: Mechanisms of Toxicity and Tolerance* (pp. 247–265). Springer, Cham. https://doi.org/10.1007/978-3-319-13368-3_10
- Pareek, A., Dhankher, O. P., & Foyer, C. H. (2020). Mitigating the impact of climate change on plant productivity and ecosystem sustainability. *Journal of Experimental Botany*, 71(2), 451–456. <https://doi.org/10.1093/jxb/erz518>

- Parvaiz, A., Latef, A. A. A., Hashem, A., Abd Allah, E. F., Gucel, S., & Tran, L. S. P. (2016). Nitric oxide mitigates salt stress by regulating levels of osmolytes and antioxidant enzymes in chickpea. *Frontiers in Plant Science*, 7(MAR2016), 347. <https://doi.org/10.3389/FPLS.2016.00347/BIBTEX>
- Passioura, J. (1988). Root signals control leaf expansion in wheat seedlings growing in drying soil. *Australian Journal of Plant Physiology*, 15, 687–693. <https://doi.org/10.1071/pp9900149>
- Pasternak, T. P., Prinsen, E., Ayaydin, F., Miskolczi, P., Potters, G., Asard, H., Onckelen, H. A. Van, Dudits, D., & Fehér, A. (2002). The role of auxin, pH, and stress in the activation of embryogenic cell division in leaf protoplast-derived cells of alfalfa. *Plant Physiology*, 129, 1807–1819. <https://doi.org/10.1104/pp.000810>
- Pillai, S. E., Kumar, C., Patel, H. K., & Sonti, R. V. (2018). Overexpression of a cell wall damage induced transcription factor, OsWRKY42, leads to enhanced callose deposition and tolerance to salt stress but does not enhance tolerance to bacterial infection. *BMC Plant Biology*, 18, 177. <https://doi.org/10.1186/s12870-018-1391-5>
- Platten, J. D., Egdane, J. A., & Ismail, A. M. (2013). Salinity tolerance, Na⁺ exclusion and allele mining of HKT1;5 in *Oryza sativa* and *O. glaberrima*: Many sources, many genes, one mechanism? *BMC Plant Biology*, 13, 32. <https://doi.org/10.1186/1471-2229-13-32>
- Poeaim, A., Poeaim, S., Poraha, R., Pongjaroenkit, S., & Pongthongkam, P. (2016). Optimization for callus induction and plant regeneration from mature seeds of Thai rice variety: Nam Roo (*Oryza sativa* L.). *Bioengineering and Bioscience*, 4(5), 95–99. <https://doi.org/10.13189/bb.2016.040504>
- Pukacka, S., Ratajczak, E., & Kalemba, E. (2009). Non-reducing sugar levels in beech (*Fagus sylvatica*) seeds as related to withstanding desiccation and storage. *Journal of Plant Physiology*, 166(13), 1381–1390. <https://doi.org/10.1016/j.jplph.2009.02.013>
- Qin, H., Li, Y., & Huang, R. (2020). Advances and challenges in the breeding of salt-tolerant Rice. *International Journal of Molecular Sciences*, 21(21), 8385. <https://doi.org/10.3390/ijms21218385>
- Qun, S., Jian-Hua, W., & Bao-Qi, S. (2007). Advances on seed vigor physiological and genetic mechanisms. *Agricultural Sciences in China*, 6(9), 1060–1066.
- Rad, H. E., Aref, F., Rezaei, M., Amiri, E., & Khaledian, M. R. (2011). The effects of salinity at different growth stage on rice yield. *Ecology, Environment and Conservation*, 17(2), 111–117.
- Rahim, H., Amin, E. E. E. A., & Mat, M. Z. (2021). *Penilaian tahap penggunaan*

teknologi padi MARDI di Muda Agricultural Development Authority (MADA) (Vol. 23). <http://ebuletin.mardi.gov.my/buletin/23/Hairazi.pdf>

- Rahman, A., Nahar, K., Mahmud, J. Al, Hasanuzzaman, M., Hossain, M. S., & Fujita, M. (2017). Salt stress tolerance in rice: Emerging role of exogenous phytoprotectants. In J. Li (Ed.), *Advances in International Rice Research* (pp. 139–174). Intech Open. <https://doi.org/10.5772/67098>
- Rahman, S. N. A., Rahman, M. H. A., Razak, M. S. F. A., Othman, M. R., & Rahim, N. I. A. (2021). *Varieti padi toleran kepada tekanan air masin sebagai sumber genetik padi bagi menangani isu perubahan iklim di Malaysia* (Vol. 26). Buletin Teknologi MARDI.
- Raja, V., Majeed, U., Kang, H., Andrabi, K. I., & John, R. (2017). Abiotic stress: Interplay between ROS, hormones and MAPKs. *Environmental and Experimental Botany*, 137, 142–157. <https://doi.org/10.1016/j.envexpbot.2017.02.010>
- Rajan, R. P., & Singh, G. (2021). A review on application of somaclonal variation in important horticulture crops. *Plant Cell Biotechnology and Molecular Biology*, 22(35–36), 161–175. <https://doi.org/https://www.ikpress.org/index.php/PCBMB/article/view/6319>
- Rajendran, K., Tester, M., & Roy, S. J. (2009). Quantifying the three main components of salinity tolerance in cereals. *Plant Cell and Environment*, 32, 237–249. <https://doi.org/10.1111/j.1365-3040.2008.01916.x>
- Ramakrishna, D., & Shasthree, T. (2016). High efficient somatic embryogenesis development from leaf cultures of *Citrullus colocynthis* (L.) Schrad for generating true type clones. *Physiology and Molecular Biology of Plants*, 22(2), 279–285. <https://doi.org/10.1007/s12298-016-0357-z>
- Ramli, A. (2019, January 25). Kegagalan petani urus sawah jejas hasil: MARDI. *Berita Harian*. <https://www.pressreader.com/malaysia/berita-harian-malaysia/20190125/281784220301216>
- Rana, M. M., Takamatsu, T., Baslam, M., Kaneko, K., Itoh, K., Harada, N., Sugiyama, T., Ohnishi, T., Kinoshita, T., Takagi, H., & Mitsui, T. (2019). Salt tolerance improvement in rice through efficient SNP marker-assisted selection coupled with speed-breeding. *International Journal of Molecular Sciences*, 20(10). <https://doi.org/10.3390/IJMS20102585>
- Raoufi, A., Salehi, H., Rahemi, M., Shekafandeh, A., & Khalili, S. (2021). In vitro screening: The best method for salt tolerance selection among pistachio rootstocks. *Journal of the Saudi Society of Agricultural Sciences*, 20(3), 146–154. <https://doi.org/10.1016/J.JSSAS.2020.12.010>
- Rattana, K., & Bunnag, S. (2015). Differential salinity tolerance in calli and shoots of four rice cultivars. *Asian Journal of Plant Sciences*, 7(1), 48–60.

<https://doi.org/https://dx.doi.org/10.3923/ajcs.2015.48.60>

- Raven, J. A. (1985). Regulation of pH and generation of osmolarity in vascular plants: A cost-benefit analysis in relation to efficiency of use of energy, nitrogen and water. *New Phytologist*, 101, 25–77.
- Razzaq, A., Ali, A., Safdar, L. Bin, Zafar, M. M., Rui, Y., Shakeel, A., Shaukat, A., Ashraf, M., Gong, W., & Yuan, Y. (2020). Salt stress induces physiochemical alterations in rice grain composition and quality. *Journal of Food Science*, 85(1), 14–20. <https://doi.org/10.1111/1750-3841.14983>
- Reddy, I. N. B. L., Kim, B. K., Yoon, I. S., Kim, K. H., & Kwon, T. R. (2017). Salt tolerance in rice: Focus on mechanisms and approaches. *Rice Science*, 24(3), 123–144. <https://doi.org/10.1016/j.rsci.2016.09.004>
- Reddy, I. N. B. L., Kim, S. M., Kim, B. K., Yoon, I. S., & Kwon, T. R. (2017). Identification of rice accessions associated with K⁺/Na⁺ ratio and salt tolerance based on physiological and molecular responses. *Rice Science*, 24(6), 360–364. <https://doi.org/10.1016/j.rsci.2017.10.002>
- Rengasamy, P. (2010). Soil processes affecting crop production in salt-affected soils. *Functional Plant Biology*, 37(7), 613–620. <https://doi.org/10.1071/FP09249>
- Rengasamy, P., Chittleborough, D., & Helyar, K. (2003). Root-zone constraints and plant-based solutions for dryland salinity. *Plant and Soil*, 257, 249–260. <https://doi.org/https://doi.org/10.1023/A:1027326424022>
- Reyes-Olalde, J. I., Zúñiga-Mayo, V. M., Marsch-Martínez, N., & de Folter, S. (2017). Synergistic relationship between auxin and cytokinin in the ovary and the participation of the transcription factor SPATULA. *Plant Signaling & Behavior*, 12(10), e1376158. <https://doi.org/10.1080/15592324.2017.1376158>
- Reza, M. N., Na, I. S., Baek, S. W., & Lee, K. H. (2019). Rice yield estimation based on K-means clustering with graph-cut segmentation using low-altitude UAV images. *Biosystems Engineering*, 177(2018), 109–121. <https://doi.org/10.1016/j.biosystemseng.2018.09.014>
- Rhodes, D., Nadolska-Orczyk, A., & Rich, P. J. (2002). Salinity, osmolytes and compatible solutes. In *Salinity: Environment - Plants - Molecules* (pp. 181–204). Springer, Dordrecht. https://doi.org/10.1007/0-306-48155-3_9
- Riaz, M., Arif, M. S., Ashraf, M. A., Mahmood, R., Yasmeen, T., Shakoor, M. B., Shahzad, S. M., Ali, M., Saleem, I., Arif, M., & Fahad, S. (2019). A comprehensive review on rice responses and tolerance to salt stress. In M. Hasanuzzaman, M. Fujita, K. Nahar, & J. K. Biswas (Eds.), *Advances in rice research for abiotic stress tolerance* (pp. 133–158). Woodhead Publishing. <https://doi.org/https://dx.doi.org/10.1016/B978-0-12-814332-2.00007-1>

- Runyan, C. W., & Odorico, P. D. (2010). Ecohydrological feedbacks between salt accumulation and vegetation dynamics: Role of vegetation-groundwater interactions. *Water Resources Research*, 46, W11561. <https://doi.org/10.1029/2010WR009464>
- Ryu, H., & Cho, Y.-G. (2015). Plant hormones in salt stress tolerance. *Journal of Plant Biology*, 58(3), 147–155. <https://doi.org/10.1007/S12374-015-0103-Z>
- Saad, A., Othman, O., Azlan, S., Alias, I., & Habibudin, H. (2004). Impact and contribution of resistant varieties in rice pest management in Malaysian modern rice farming. *Modern Rice Farming: International Rice Conference*.
- Safdar, H., Amin, A., Ahafiq, Y., Ali, A., Yasin, R., Shoukat, A., Hussan, M. U., & Sarwar, M. I. (2019). A review: Impact of salinity on plant growth. *Nature and Science*, 17(1), 34–40. <https://doi.org/10.7537/marsnsj170119.06>
- Sah, S. K., Kaur, A., & Sandhu, J. S. (2014). High frequency embryogenic callus induction and whole plant regeneration in *japonica* rice cv. Kitaake. *Rice Research*, 2(2), 125. <https://doi.org/10.4172/jrr.1000125>
- Sahab, S., Suhani, I., Srivastava, V., Chauhan, P. S., Singh, R. P., & Prasad, V. (2021). Potential risk assessment of soil salinity to agroecosystem sustainability: Current status and management strategies. *Science of The Total Environment*, 764, 144164. <https://doi.org/10.1016/j.scitotenv.2020.144164>
- Sahi, C., Singh, A., Kumar, K., Blumwald, E., & Grover, A. (2006). Salt stress response in rice: Genetics, molecular biology, and comparative genomics. *Functional & Integrative Genomics*, 6, 263–284. <https://doi.org/10.1007/s10142-006-0032-5>
- Sahoo, K. K., Tripathi, A. K., Pareek, A., Sopory, S. K., & Singla-Pareek, S. L. (2011). An improved protocol for efficient transformation and regeneration of diverse *indica* rice cultivars. *Plant Methods*, 7(1), 49. <https://doi.org/10.1186/1746-4811-7-49>
- Sangeetha, R. (2013). Effect of salinity induced stress and its alleviation on the activity of amylase in the germinating seeds of *Zea mays*. *International Journal of Basic and Life Sciences*, 1(1), 1–9.
- Sarkar, R. K., Mahata, K. R., & Singh, D. P. (2013). Differential responses of antioxidant system and photosynthetic characteristics in four rice cultivars differing in sensitivity to sodium chloride stress. *Acta Physiologiae Plantarum*, 35(10), 2915–2926. <https://doi.org/10.1007/s11738-013-1322-x>
- Sazali, S. A., Mohd Shukor Nordin, Shahari, R., Noraziyah Abd. Aziz Shamsudin, Mohd Rafii Yusop, Mohd Shahril Firdaus Ab Razak, Rahiniza Kamaruzaman, & Mohd Syahmi Salleh. (2021). Susceptibility of Malaysian rice (*Oryza sativa* L.) cultivar to saline water submergence based on the

morphological traits. *Journal Of Agrobiotechnology*, 12(2), 47–55. <https://doi.org/10.37231/jab.2021.12.2.257>

Scaranari, C., Leal, P. A. M., & Mazzafera, P. (2009). Shading and periods of acclimatization of micropropagated banana plantlets cv. Grande Naine. *Scientia Agricola*, 66(3), 331–337. <https://doi.org/10.1590/s0103-90162009000300008>

Scott, S. J., Jones, R. A., & Williams, W. A. (1984). Review of data analysis methods for seed germination. *Crop Science*, 24(6), 1192–1199. <https://doi.org/10.2135/cropsci1984.0011183x002400060043x>

Sengupta, S., & Majumder, A. L. (2009). Insight into the salt tolerance factors of a wild halophytic rice, *Porteresia coarctata*: A physiological and proteomic approach. *Planta*, 229, 911–929. <https://doi.org/10.1007/s00425-008-0878-y>

Senguttuvel, P., Sravanraju, N., Padmavathi, G., Sundaram, R. M., Madhav, S., Hariprasad, A. S., Kota, S., Bhadana, V. P., Subrahmanyam, D., Subbarao, L. V., Brajendra, & Ravindrababu, V. (2016). Identification and quantification of salinity tolerance through salt stress indices and variability studies in rice (*Oryza sativa* L.). *Sabroo Journal of Breeding and Genetics*, 48(2), 172–179.

Shahbaz, M., & Ashraf, M. (2013). Improving salinity tolerance in cereals. In *Critical Reviews in Plant Sciences* (Vol. 32, pp. 237–249). Springer, Dordrecht. <https://doi.org/10.1080/07352689.2013.758544>

Shahid, S. A., Zaman, M., & Heng, L. (2018). Soil salinity: Historical perspectives and a world overview of the problem. In *Guideline for Salinity Assessment, Mitigation and Adaptation Using Nuclear and Related Techniques* (pp. 43–53). Springer International Publishing. https://doi.org/10.1007/978-3-319-96190-3_2

Shakirova, F. M. (2007). Role of hormonal system in the manifestation of growth promoting and antistress action of salicylic acid. In S. Hayat & A. Ahmad (Eds.), *Salicylic Acid: A Plant Hormone* (pp. 69–89). Springer.

Shakirova, Farida M, Sakhabutdinova, A. R., Bezrukova, M. V, Fatkhutdinova, R. A., & Fatkhutdinova, D. R. (2003). Changes in the hormonal status of wheat seedlings induced by salicylic acid and salinity. *Plant Science*, 164(3), 317–322. [https://doi.org/https://doi.org/10.1016/S0168-9452\(02\)00415-6](https://doi.org/https://doi.org/10.1016/S0168-9452(02)00415-6)

Sharma, A., Rana, C., Singh, S., & Katoch, V. (2016). Soil salinity: Causes, effects and management in cucurbits. In M. Pessaraki (Ed.), *Handbook of Cucurbits: Growth, Cultural Practices, and Physiology* (Issue February, pp. 419–439). CRC Press.

Sharma, P., Jha, A. B., Dubey, R. S., & Pessaraki, M. (2012). Reactive oxygen

- species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. *Journal of Botany*, 2012, 217037. https://doi.org/10.1007/978-1-4614-1533-6_100084
- Shekar, S., Sankepally, R., & Singh, B. (2016). Optimization of regeneration using differential growth regulators in *indica* rice cultivars. *3 Biotech*, 6, 19. <https://doi.org/10.1007/s13205-015-0343-0>
- Shen, R. S., Jian, Y. L., Lee, Y. I., Huang, K. L., & Miyajima, I. (2016). Histological observation of the somatic embryogenesis in *Dieffenbachia* "Anna." *Journal of the Faculty of Agriculture, Kyushu University*, 61(1), 1–6.
- Shrivastava, P., & Kumar, R. (2015). Soil salinity: A serious environmental issue and plant growth promoting bacteria as one of the tools for its alleviation. *Saudi Journal of Biological Sciences*, 22(2), 123–131. <https://doi.org/10.1016/j.sjbs.2014.12.001>
- Signorelli, S. (2016). The fermentation analogy: A point of view for understanding the intriguing role of proline accumulation in stressed plants. *Frontiers in Plant Science*, 7, 1339. <https://doi.org/10.3389/fpls.2016.01339>
- Signorelli, S., Dans, P. D., Coitiño, E. L., Borsani, O., & Monza, J. (2015). Connecting proline and γ -aminobutyric acid in stressed plants through non-enzymatic reactions. *PLoS ONE*, 10(3), e0115349. <https://doi.org/10.1371/journal.pone.0115349>
- Singh, B. P., Jayaswal, P. K., Singh, B., Singh, P. K., Kumar, V., Mishra, S., Singh, N., Panda, K., & Singh, N. K. (2015). Natural allelic diversity in OsDREB1F gene in the Indian wild rice germplasm led to ascertain its association with drought tolerance. *Plant Cell Reports*, 34, 993–1004. <https://doi.org/10.1007/s00299-015-1760-6>
- Singh, R. K., Kota, S., & Flowers, T. J. (2021a). Salt tolerance in rice: Seedling and reproductive stage QTL mapping come of age. *Theoretical and Applied Genetics*, 134(11), 3495–3533. <https://doi.org/10.1007/s00122-021-03890-3>
- Singh, R. K., Kota, S., & Flowers, T. J. (2021b). Salt tolerance in rice: Seedling and reproductive stage QTL mapping come of age. *Theoretical and Applied Genetics*, 134, 3495. <https://doi.org/10.1007/S00122-021-03890-3>
- Singh, Sanjay, Sengar, R. S., Kulshreshtha, N., Datta, D., Tomar, R. S., Rao, V. P., Garg, D., & Ojha, A. (2015). Assessment of multiple tolerance indices for salinity stress in bread wheat (*Triticum aestivum* L.). *Journal of Agricultural Science*, 7(3), 45–57. <https://doi.org/10.5539/JAS.V7N3P49>
- Singh, Shilpy, Kumar, A., Rana, V., Kumar, A., & Rakesh Sharma, V. (2018a). In vitro callus induction and plant regeneration in basmati rice (*Oryza sativa* L.) varieties. *Journal of Pharmacognosy and Phytochemistry*, SP5, 65–69. www.agricoop.nic.in;

- Sinha, R., & Khare, S. K. (2014). Protective role of salt in catalysis and maintaining structure of halophilic proteins against denaturation. *Frontiers in Microbiology*, 5, 165. <https://doi.org/10.3389/FMICB.2014.00165>
- Skoog, F., & Miller, C. O. (1957). Chemical regulation of growth and organ formation in plant tissues cultured in vitro. *Symposia of the Society for Experimental Biology*, 11, 118–130. <http://www.ncbi.nlm.nih.gov/pubmed/13486467>
- Slama, I., Abdelly, C., Bouchereau, A., Flowers, T., & Savoure, A. (2015). Diversity, distribution and roles of osmoprotective compounds accumulated in halophytes under abiotic stress. *Annals of Botany*, 115, 433–447. <https://doi.org/https://doi.org/10.1093/aob/mcu239>
- Smirnoff, N., & Cumbes, Q. J. (1989). Hydroxyl radical scavenging activity of compatible solutes. *Phytochemistry*, 28(4), 1057–1060. [https://doi.org/10.1016/0031-9422\(89\)80182-7](https://doi.org/10.1016/0031-9422(89)80182-7)
- Sofa, A., Scopa, A., Nuzzaci, M., & Vitti, A. (2015). Ascorbate peroxidase and catalase activities and their genetic regulation in plants subjected to drought and salinity stresses. *International Journal of Molecular Sciences*, 16(6), 13561–13578. <https://doi.org/10.3390/ijms160613561>
- Sofy, M. R., Seleiman, M. F., Alhammad, B. A., Alharbi, B. M., & Mohamed, H. I. (2020). Minimizing adverse effects of Pb on maize plants by combined treatment with jasmonic, salicylic acids and proline. *Agronomy*, 10(5), 699. <https://doi.org/10.3390/agronomy10050699>
- Solis, C. A., Yong, M. T., Vinarao, R., Jena, K., Holford, P., Shabala, L., Zhou, M., Shabala, S., & Chen, Z.-H. (2020). Back to the wild: On a quest for donors toward salinity tolerant rice. *Frontiers in Plant Science*, 11, 323. <https://doi.org/10.3389/fpls.2020.00323>
- Souki, H. (2015). *Prestasi dan ciri-ciri biji padi varieti tempatan yang ditanam di Kota Belud, Sabah* (Vol. 8). [http://ebuletin.mardi.gov.my/buletin/08/Prestasi biji padi.pdf](http://ebuletin.mardi.gov.my/buletin/08/Prestasi%20biji%20padi.pdf)
- Sun, B., Huang, T., & Fu, C. (2018). Identification and genetic characterization of a novel tillering dwarf semi-sterile mutant tdr1 in rice. *American Journal of Plant Sciences*, 9, 2545–2554. <https://doi.org/10.4236/ajps.2018.913184>
- Sun, K., Hunt, K., & Hauser, B. A. (2004). Ovule abortion in Arabidopsis triggered by stress. *Plant Physiology*, 135(4), 2358–2367. <https://doi.org/10.1104/pp.104.043091>
- Sunian, E., Solihen Jamal, M., Amri Saidon, S., Mokhtar, A., Kamaruzaman, R., Ramli, A., Hashim, S., Ramachandran, K., Norsuha Misman, S., Fitri Masarudin, M., Mohamad Saad, M., Naim Fadzli Abd Rani, M., Mohd Yusob, S., Hosni, H., Amzah, B., Esa, N., Rahim, H., & Mohd Zuki dan

- Hamidah Mohd Sarif Pengenalan, Z. (2021). Varieti padi baharu MARDI: MR 315. In *Buletin Teknologi MARDI* (Vol. 23). <http://ebuletin.mardi.gov.my/buletin/23/Elixon.pdf>
- Sweetlove, L. J., & Moller, I. M. (2009). Oxidation of proteins in plants - Mechanisms and consequences. *Advances in Botanical Research*, 52, 1–23. [https://doi.org/10.1016/S0065-2296\(10\)52001-9](https://doi.org/10.1016/S0065-2296(10)52001-9)
- Szabados, Laszlo, & Savoure, A. (2009). Proline: A multifunctional amino acid. *Trends in Plant Science*, 15(2), 89–97. <https://doi.org/10.1016/j.tplants.2009.11.009>
- Tam, D. M., & Lang, N. T. (2003). *In vitro* selection for salt tolerance in rice. *Omonrice*, 11, 68–73.
- Tang, Y., Liu, K., Bao, X., Zhi, Y., Wu, Q., Guo, Y., Yin, X., Zeng, L., Li, J., Zhang, J., He, W., Liu, W., Wang, Q., Jia, C., & Li, Z. (2019). Overexpression of a MYB family gene, OsMYB6, increases drought and salinity stress tolerance in transgenic rice. *Frontiers in Plant Science*, 10, 168. <https://doi.org/10.3389/fpls.2019.00168>
- Thitisaksakul, M., Tananuwong, K., Shoemaker, C. F., Chun, A., Tanadul, O., Labavitch, J. M., & Beckles, D. M. (2015). Effects of timing and severity of salinity stress on rice (*Oryza sativa* L.) yield, grain composition and starch functionality. *Journal of Agriculture and Food Chemistry*, 63, 2296–2304. <https://doi.org/10.1021/jf503948p>
- Torun, H., Novák, O., Mikulík, J., Pěňčík, A., Strnad, M., & Ayaz, F. A. (2020). Timing-dependent effects of salicylic acid treatment on phytohormonal changes, ROS regulation, and antioxidant defense in salinized barley (*Hordeum vulgare* L.). *Scientific Reports*, 10(1). <https://doi.org/10.1038/s41598-020-70807-3>
- Tran Thi Huong Sen, Phan Thi Phuong Nhi, & Trinh Thi Sen. (2017). Salinity effect at seedling and flowering stages of some rice lines and varieties (*Oryza sativa* L.). *Journal of Agricultural Science and Technology A*, 7(10), 32–39. <https://doi.org/10.17265/2161-6256/2017.10.005S>
- Tucuch-Haas, C. J., Alcántar-González, G., Trejo-Téllez, L. I., Volke-Haller, H., Salinas-Moreno, Y., & Larqué-Saavedra, A. (2017). Effect of salicylic acid on growth, nutritional status and performance of maize (*Zea mays*). *Agrociencia*, 51, 771–781.
- Ucarli, C. (2021). Effects of salinity on seed germination and early seedling stage. In *Abiotic Stress in Plants* (pp. 1–21). IntechOpen. <https://doi.org/10.5772/intechopen.93647>
- Umar, J., Aliyu, A., Shehu, K., & Abubakar, L. (2018). A study on effects of salinity on growth and yield of tomato genotype (*Solanum lycopersicum*). *Sustainable Food Production*, 3, 16–24.

<https://doi.org/10.18052/www.scipress.com/SFP.3.16>

- Vaidyanathan, H., Sivakumar, P., Chakrabarty, R., & Thomas, G. (2003). Scavenging of reactive oxygen species in NaCl-stressed rice (*Oryza sativa* L.) - Differential response in salt-tolerant and sensitive varieties. *Plant Science*, 165(6), 1411–1418. <https://doi.org/10.1016/J.PLANTSCI.2003.08.005>
- Vennapusa, A. R., Vemanna, R. S., Reddy, B. . R., Babitha, K. C., Kiranmai, K., Nareshkumar, A., & Sudhakar, C. (2015). An efficient callus induction and regeneration protocol for a drought tolerant rice *indica* genotype AC39020. *Journal of Plant Sciences*, 3(5), 248. <https://doi.org/10.11648/j.jps.20150305.11>
- Verma, D., Joshi, R., Shukla, A., & Kumar, P. (2011). Protocol for *in vitro* somatic embryogenesis and regeneration of rice (*Oryza sativa* L.). *Indian Journal of Experimental Biology*, 49(12), 958–963.
- Verslues, P. E., Agarwal, M., Katiyar-Agarwal, S., Zhu, J., & Zhu, J.-K. (2006). Methods and concepts in quantifying resistance to drought, salt and freezing, abiotic stresses that affect plant water status. *The Plant Journal*, 45, 523–539. <https://doi.org/10.1111/j.1365-313X.2005.02593.x>
- Verslues, P. E., & Sharma, S. (2010). Proline metabolism and its implications for plant-environment interaction. In *The Arabidopsis Book* (Vol. 8, p. e0140). American Society of Plant Biologists. <https://doi.org/10.1199/tab.0140>
- Vijayalakshmi, T., Vijayakumar, A. S., Kiranmai, K., Nareshkumar, A., & Sudhakar, C. (2016). Salt stress induced modulations in growth, compatible solutes and antioxidant enzymes response in two cultivars of safflower (*Carthamus tinctorius* L. cultivar TSF1 and cultivar SM) differing in salt tolerance. *American Journal of Plant Sciences*, 7, 1802–1819. <https://doi.org/10.4236/ajps.2016.713168>
- Visarada, K. B. R. S., Sailaja, M., & Sarma, N. P. (2002). Effect of callus induction media on morphology of embryogenic calli in rice genotypes. *Biologia Plantarum*, 45(4), 495–502. <https://doi.org/10.1023/A:1022323221513>
- Voleti, S. R., Seeds, N., Banda, S., & Subbarao, L. V. (2013). Spikelet dominance and sterility in rice (*Oryza sativa* L.) genotypes: A mechanism of survival? *Indian Journal of Plant Physiology*, March. <https://doi.org/10.1007/s40502-013-0005-3>
- Vondrakova, Z., Eliasova, K., Fischerova, L., & Vagner, M. (2011). The role of auxins in somatic embryogenesis of *Abies alba*. *Central European Journal of Biology*, 6(4), 587–596. <https://doi.org/10.2478/s11535-011-0035-7>
- Wan, Q., Hongbo, S., Zhaolong, X., Jia, L., Dayong, Z., & Yihong, H. (2017). Salinity tolerance mechanism of Osmotin and Osmotin-like proteins: A promising candidate for enhancing plant salt tolerance. *Current Genomics*,

18(6), 553. <https://doi.org/10.2174/1389202918666170705153045>

- Wang, K., Zhang, L., Gao, M., Lv, L., Zhao, Y., Zhang, L., Li, B., Han, M., & Alva, A. K. (2013). Influence of salt stress on growth and antioxidant responses of two *Malus* species at callus and plantlet stages. *Pakistan Journal of Botany*, 45(2), 375–381.
- Wang, X., Gao, Y., Yan, Q., & Chen, W. (2016). Salicylic acid promotes autophagy via NPR3 and NPR4 in *Arabidopsis* senescence and innate immune response. *Acta Physiologiae Plantarum*, 38, 241. <https://doi.org/https://doi.org/10.1007/s11738-016-2257-9>
- Wang, Y. H., & Irving, H. R. (2011). Developing a model of plant hormone interactions. *Plant Signaling & Behavior*, 6(4), 494. <https://doi.org/10.4161/PSB.6.4.14558>
- Wang, Z., Wang, J., Bao, Y., Wang, F., & Zhang, H. (2010). Quantitative trait loci analysis for rice seed vigor during the germination stage. *Journal of Zhejiang University- Science B (Biomedicine & Biotechnology)*, 11(12), 958–964. <https://doi.org/https://dx.doi.org/10.1631/2Fjzus.B1000238>
- Wani, M. A., Khan, F. U., Nazki, I. T., Din, A., Iqbal, S., & Qadri, Z. A. (2019). Influence of various treatments on pre and post germination properties of *Callistephus chinensis* (L.) Nees cv. Powderpuff. *Bangladesh Journal of Botany*, 48(3), 449–455. <https://doi.org/10.3329/BJB.V48I3.47724>
- Wani, S. H., Sanghera, G. S., & Gosal, S. S. (2011). An efficient and reproducible method for regeneration of whole plants from mature seeds of a high yielding *Indica* rice (*Oryza sativa* L.) variety PAU 201. *New Biotechnology*, 28(4), 418–422. <https://doi.org/10.1016/j.nbt.2011.02.006>
- Watanabe, S., Kojima, K., Ide, Y., & Sasaki, S. (2000). Effects of saline and osmotic stress on proline and sugar accumulation in *Populus euphratica* in vitro. *Plant Cell, Tissue and Organ Culture*, 63(3), 199–206. <https://doi.org/10.1023/A:1010619503680>
- Wei, X., & Huang, X. (2019). Origin, taxonomy, and phylogenetics of rice. In *Rice: The First Crop Genome* (pp. 1–29). AACC International Press. <https://doi.org/10.1186/s12284-016-0087-4>
- Xu, N., Chu, Y., Chen, H., Li, X., Wu, Q., Jin, L., Wang, G., & Huang, J. (2018). Rice transcription factor OsMADS25 modulates root growth and confers salinity tolerance via the ABA-mediated regulatory pathway and ROS scavenging. *PLOS Genetics*, 14(10), e1007662. <https://doi.org/10.1371/journal.pgen.1007662>
- Yang, G., Xing, Y., Li, S., Ding, J., Yue, B., Deng, K., Li, Y., & Zhu, Y. (2006). Molecular dissection of developmental behavior of tiller number and plant height and their relationship in rice (*Oryza sativa* L.). *Hereditas*, 143(2006), 236–245. <https://doi.org/10.1111/j.2006.0018-0661.01959.x>

- Yang, Y., Zhu, K., Xia, H., Chen, L., & Chen, K. (2014). Comparative proteomic analysis of *indica* and *japonica* rice varieties. *Genetics and Molecular Biology*, 37(4), 652–661. <https://doi.org/10.1590/S1415-47572014005000015>
- Yeo, A. R. (1983). Salinity resistance: Physiologies and prices. *Physiologia Plantarum*, 58(2), 214–222. <https://doi.org/10.1111/J.1399-3054.1983.TB04172.X>
- Yoon, J. Y., Hamayun, M., Lee, S.-K., & Lee, I.-J. (2009). Methyl jasmonate alleviated salinity stress in soybean. *Journal of Crop Science and Biotechnology*, 12(2), 63–68. <https://doi.org/https://doi.org/10.1007/s12892-009-0060-5>
- Yoshida, S., Forno, D. A., Cock, J. H., & Gomez, K. A. (1976). Laboratory Manual for Physiological Studies of Rice. In *The International Rice Research Institute* (Vol. 53, Issue 9). http://books.irri.org/9711040085_content.pdf
- Yusuf, M., Hayat, S., Alyemini, M. N., Fariduddin, Q., & Ahmad, A. (2013). Salicylic acid: Physiological roles in plants. In *Salicylic Acid* (pp. 15–30). Springer, Dordrecht. https://doi.org/10.1007/978-94-007-6428-6_2
- Zafar, K., Sedeek, K. E. M., Rao, G. S., Khan, M. Z., Amin, I., Kamel, R., Mukhtar, Z., Zafar, M., Mansoor, S., & Mahfouz, M. M. (2020). Genome Editing Technologies for Rice Improvement: Progress, Prospects, and Safety Concerns. *Frontiers in Genome Editing*, 0, 5. <https://doi.org/10.3389/FGEEED.2020.00005>
- Zafar, S. A., Patil, S. B., Uzair, M., Fang, J., Zhao, J., Guo, T., Yuan, S., Uzair, M., Luo, Q., Shi, J., Schreiber, L., & Li, X. (2020). Degenerated panicle and partial sterility 1 (DPS1) encodes a cystathionine β -synthase domain containing protein required for anther cuticle and panicle development in rice. In *New Phytologist* (Vol. 225, Issue 1, pp. 356–375). <https://doi.org/10.1111/nph.16133>
- Zakaria, B. K., Kumar, S. A., & Yusof, M. F. M. (2015, October 2). Penerobosan air masin ancam ekonomi negara. *Berita Harian*.
- Zavattieri, M. A., Frederico, A. M., Lima, M., Sabino, R., & Arnholdt-Schmitt, B. (2010). Induction of somatic embryogenesis as an example of stress-related plant reactions. *Electronic Journal of Biotechnology*, 13(1). <https://doi.org/10.2225/vol13-issue1-fulltext-4>
- Zelm, E. van, Zhang, Y., & Testerink, C. (2020). Salt tolerance mechanisms of plants. *Annual Review of Plant Biology*, 71, 403–433. <https://doi.org/10.1146/annurev-arplant-050718>
- Zeng, L., & Shannon, M. C. (2000). Salinity Effects on Seedling Growth and Yield Components of Rice. *Crop Science*, 40(4), 996–1003. <https://doi.org/10.2135/CROPSCI2000.404996X>

- Zhang, A., Liu, Y., Wang, F., Li, T., Chen, Z., Kong, D., Bi, J., Zhang, F., Luo, X., Wang, J., Tang, J., Yu, X., Liu, G., & Luo, L. (2019). Enhanced rice salinity tolerance via CRISPR/Cas9-targeted mutagenesis of the OsRR22 gene. *Frontiers in Genome Editing*, 39, 47. <https://doi.org/10.1007/S11032-019-0954-Y>
- Zhang, D., Wang, Z., Wang, N., Gao, Y., Liu, Y., Wu, Y., Bai, Y., Zhang, Z., Lin, X., Ou, Y. D. X., Xu, C., & Liu, B. (2014). Tissue Culture-Induced Heritable Genomic Variation in Rice, and Their Phenotypic Implications. *PLoS ONE*, 9(5), e96879. <https://doi.org/https://doi.org/10.1371/journal.pone.0096879>
- Zhang, G., Xu, N., Chen, H., Wang, G., & Huang, J. (2018). OsMADS25 regulates root system development via auxin signalling in rice. *The Plant Journal*, 95, 1004–1022. <https://doi.org/10.1111/tpj.14007>
- Zhang, K., Su, J., Xu, M., Zhou, Z., Zhu, X., Ma, X., Hou, J., Tan, L., Zhu, Z., Cai, H., Liu, F., Sun, H., Gu, P., Li, C., Liang, Y., Zhao, W., Sun, C., & Fu, Y. (2020). A common wild rice-derived BOC1 allele reduces callus browning in *indica* rice transformation. *Nature Communications*, 11, 443. <https://doi.org/10.1038/s41467-019-14265-0>
- Zhang, Y., Lan, H., Shao, Q., Wang, R., Chen, H., Tang, H., Zhang, H., Huang, J., & Zhang, J. (2016). An A20/AN1-type zinc finger protein modulates gibberellins and abscisic acid contents and increases sensitivity to abiotic stress in rice (*Oryza sativa*). *Journal of Experimental Botany*, 67(1), 315–326. <https://doi.org/10.1093/jxb/erv464>
- Zhao, C., Zhang, H., Song, C., Zhu, J.-K., & Shabala, S. (2020). Mechanisms of plant responses and adaptation to soil salinity. *The Innovation*, 1(1), 100017. <https://doi.org/10.1016/j.xinn.2020.100017>
- Zhao, X., Wang, W., Zhang, F., Deng, J., Li, Z., & Fu, B. (2014). Comparative metabolite profiling of two rice genotypes with contrasting salt stress tolerance at the seedling stage. *PLoS ONE*, 9(9), e108020. <https://doi.org/10.1371/journal.pone.0108020>
- Zhifang, G., & Loescher, W. H. (2003). Expression of a celery mannose 6-phosphate reductase in *Arabidopsis thaliana* enhances salt tolerance and induces biosynthesis of both mannitol and a glucosyl-mannitol dimer. *Plant, Cell and Environment*, 26(2), 275–283. <https://doi.org/10.1046/j.1365-3040.2003.00958.x>
- Zhou, W., Lv, T., Yang, Z., Wang, T., Fu, Y., Chen, Y., Hu, B., & Ren, W. (2017). Morphophysiological mechanism of rice yield increase in response to optimized nitrogen management. *Scientific Reports*, 7(1), 1–11. <https://doi.org/10.1038/s41598-017-17491-y>
- Zhu, N., Cheng, S., Liu, X., Du, H., Dai, M., Zhou, D. X., Yang, W., & Zhao, Y. (2015). The R2R3-type MYB gene OsMYB91 has a function in coordinating plant growth and salt stress tolerance in rice. *Plant Science*, 236, 146–156.