



UPM
UNIVERSITI PUTRA MALAYSIA
BERILMU BERBAKTI

**GENETIC DIVERSITY AND DEVELOPMENT OF HIGH FIBER YIELD
KENAF (*Hibiscus cannabinus* L.) MUTANT HYBRIDS THROUGH DIALLEL
CROSSES**

By

MD. AL-MAMUN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

August 2022

IPTSM 2022 9

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATIONS

This thesis is dedicated to my father and mother, who infused in me the spirit of studying and whose blessing inspired me to achieve higher life goals. I also dedicated it to the people of my hometown for inspiring me to continue my education.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

**GENETIC DIVERSITY AND DEVELOPMENT OF HIGH FIBER YIELD
KENAF (*Hibiscus cannabinus* L.) MUTANT HYBRIDS THROUGH DIALLEL
CROSSES**

By

MD AL-MAMUN

August 2022

Chairman : Professor Mohd Rafii bin Yusop, PhD
Institute : Tropical Agriculture and Food Security

Kenaf (*Hibiscus cannabinus* L.) is an economically important fiber crop globally for multipurpose industrial uses such as paper making, car interior components and building boards. However, commercial cultivation of this fiber crop in Malaysia has limitations due to lack of superior varieties with high fiber yield. This study was conducted to assess the genetic diversity among kenaf mutants based on agro-morphological traits and molecular markers diversity, analyze the combining ability of the selected parents and hybrids, and estimate heterosis and the nature of gene actions. To achieve these objectives, field trials and molecular diversity works were conducted at Universiti Putra Malaysia (UPM), Serdang, Selangor, Malaysia, between 2019 and 2021. Data were collected on the eight qualitative and 15 quantitative traits of the 31 kenaf genotypes. The analysis of variance over two evaluation seasons showed significant differences among genotypes for all traits except for stem top diameter. According to principal component analysis (PCA), the first five PCA accounted for 85.16% variation between genotypes based on a correlation matrix of all the quantitative traits. The mutant lines derived from V36 parental variety were grouped into six clusters, indicating a wide range of genetic variability. Path coefficient analysis revealed the maximum contribution of fresh stem weight without leaves and pod to fiber yield, followed by plant height. For molecular characterization, 10 out of the 72 expressed sequence tags - simple sequence repeat (EST-SSR) primers produced clear polymorphic bands with a mean value of 5.2 alleles per primer. These EST-SSR primers were found to be extremely informative, suggesting that among the 31 genotypes were genetically diverse. The polymorphism information content (PIC) value ranged from 0.531 to 0.737, with a mean of 0.610, indicating that the genotypes are genetically diverse. Shannon index estimation varied from 0.982 to 1.515 among 31 genotypes, whereas 0.123 to 0.405 was recorded within the early, intermediate and mid-late flowering groups. Genetic differentiation ranged from 0.67 to 1.0 alongside the average gene flow (N_m) of 0.024. The analysis of molecular variance (AMOVA) revealed 76% variation within three flowering groups., while among the groups recorded was 24%. Based on the unweighted pair group method with arithmetic means (UPGMA) dendrogram the evaluated kenaf mutants were

clustered into five major groups based on EST-microsatellites data. Combining phenotypic and molecular data in the selection of parents for hybridization activities would yield a more accurate summary and the cluster analysis identified eight mutant lines (G5, G9, G13, G21, G22, G24, G30 and G31) and one inbred (G28) that were selected for hybrid crossing using a half diallel method. The genotypes were chosen as parental sources from each heterotic group at the genetic and phenotypic levels on genetic distances and several important agronomic traits. Thirty-six hybrids were successfully produced from nine morphologically distinct kenaf genotypes using half diallel mating design. Field trials were carried out in two environments to evaluate the yield performance of 36 F₁ hybrid crosses and nine parental lines using seven qualitative and 15 quantitative attributes. From the combined data of the two environments, the combining ability analysis revealed significant general combining ability (GCA) for all traits except stem top diameter, and significant specific combining ability (SCA) for all traits except plant height and stem top diameter. The result indicated that additive and non-additive gene actions were involved in the genetic control of the traits. The magnitude of GCA variance was considerably higher than that of SCA variance except for stem top diameter, node number, and fresh stem weight with leaves and pod, showing that additive gene action predominates for these traits. The parental lines P₁ (G5: ML5), P₃ (G13: ML36-10) and P₄ (G21: ML36-24) had outstanding general combiners for fiber yield and yield-related traits. Most traits showed over-dominant gene action, except for plant height, stem middle diameter, stem top diameter, days to first flowering, dry stick weight, dry fiber weight and 1000 seed weight, which showed a partial dominance. Considering specific combining ability and heterosis values, the crosses P₁ (G5) × P₄ (G21), P₁ (G5) × P₉ (G31), P₂ (G9) × P₃ (G13), P₂ (G9) × P₅ (G22), P₄ (G21) × P₆ (G24), P₄ (G21) × P₇ (G28), P₄ (G21) × P₉ (G31), P₅ (G22) × P₈ (G30), and P₇ (G28) × P₉ (G31) were high heterotic response for fiber yield, stick yield, number of pods per plant and smaller seed size. The nine hybrids are recommended for further evaluation at multi-location trials to select several superior hybrids with high fiber yield for commercial cultivation in Malaysia.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia Sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KEPELBAGAIAN GENETIK DAN PEMBANGUNAN HIBRID MUTAN
KENAF (*Hibiscus cannabinus* L.) BERHASIL FIBER TINGGI MELALUI
KACUKAN DIALLEL**

Oleh

MD AL-MAMUN

Ogos 2022

Pengerusi : Profesor Mohd Rafii bin Yusop, PhD
Institut : Pertanian Tropika dan Sekuriti Makanan

Kenaf (*Hibiscus cannabinus* L.) ialah tanaman fiber yang penting dari segi ekonomi di peringkat global untuk kegunaan industri pelbagai guna seperti pembuatan kertas, komponen dalaman kereta dan papan bangunan. Walau bagaimanapun, penanaman secara komersial tanaman fiber ini di Malaysia mempunyai kekangan kerana kekurangan varieti unggul dengan hasil fiber yang tinggi. Kajian ini telah dijalankan untuk menilai kepelbagaian genetik antara mutan kenaf berdasarkan ciri-ciri agro-morfologi dan kepelbagaian penanda molekular, menganalisis keupayaan gabungan induk dan hibrid terpilih, serta menganggarkan heterosis dan kesan tindakan gen. Untuk mencapai objektif ini, kajian di lapangan dan percubaan kepelbagaian molekul telah dijalankan di Universiti Putra Malaysia (UPM), Serdang, Selangor, Malaysia, antara 2019 dan 2021. Data telah dicerap untuk lapan ciri kualitatif dan 15 ciri kuantitatif ke atas 31 genotip kenaf. Analisis varians sepanjang dua musim penilaian menunjukkan perbezaan yang signifikan antara genotip untuk semua ciri kecuali diameter atas batang. Menurut analisis komponen utama (PCA), lima PCA pertama menyumbang 85.16% variasi antara genotip berdasarkan matriks korelasi semua ciri-ciri kuantitatif. Titisan mutan yang terhasil dari varieti induk V36 telah diklusterkan kepada enam kelompok, menunjukkan kepelbagaian genetik yang luas antara mutan tersebut. Analisis pekali laluan menunjukkan sumbangan maksimum berat batang segar tanpa daun dan buah terhadap hasil fiber, dan diikuti dengan tinggi pokok. Untuk pencirian molekul, 10 daripada 72 primer penanda jujukan terekspres – jujukan ringkas berulang (EST-SSR) menghasilkan jalur polimorfik yang jelas dengan nilai min 5.2 alel bagi setiap penanda. Penanda EST-SSR ini didapati sangat informatif, yang menunjukkan bahawa di kalangan 31 genotip mempunyai kepelbagaian genetik. Nilai kandungan maklumat polimorfisma (PIC) adalah antara 0.531 hingga 0.737, dengan nilai min 0.610 yang menunjukkan genotip tersebut mempunyai kepelbagaian genetik. Anggaran indeks Shannon berbeza dari 0.982 hingga 1.515 dalam kalangan 31 genotip, manakala 0.123 hingga 0.405 direkodkan dalam kumpulan berbunga awal, pertengahan dan pertengahan akhir. Pembezaan genetik dengan julat antara 0.67 hingga 1.0, beserta 0.024 aliran gen purata (Nm) telah direkodkan. Analisis varians molekul (AMOVA) memberikan 76% variasi di dalam tiga kumpulan berbunga.,

manakala antara kumpulan pula yang direkodkan ialah 24%. Berdasarkan kaedah dendrogram kumpulan pasangan tak berwajaran dengan cara aritmetik (UPGMA), mutan kenaf yang dinilai telah diklusterkan kepada lima kumpulan utama berdasarkan data EST-microsatellites. Gabungan data fenotip dan molekul dalam pemilihan induk untuk aktiviti hibridisasi akan menghasilkan rumusan yang lebih tepat dan analisis kluster telah mengenalpasti lapan titisan mutan (G5, G9, G13, G21, G22, G24, G30 and G31) dan satu inbred (G28) yang telah dipilih untuk hibrid menggunakan kaedah separa dialel. Genotip tersebut telah dipilih sebagai sumber induk daripada setiap kumpulan heterotik di peringkat genetik dan fenotip bagi jarak genetik dan beberapa ciri agronomik yang penting. Tiga puluh enam kacukan telah berjaya dihasilkan daripada sembilan genotip kenaf yang berbeza secara morfologi menggunakan reka bentuk mengawan separuh dialel. Lain kajian lapangan telah dijalankan di dua persekitaran bagi menilai hasil prestasi kacukan 36 hibrid F₁ dan sembilan titisan induk menggunakan tujuh ciri kualitatif dan 15 ciri kuantitatif. Berdasarkan kepada gabungan data di dua persekitaran, analisis keupayaan bergabung mendapati keupayaan bergabung am (GCA) adalah signifikan untuk semua ciri kecuali diameter batang atas, manakala keupayaan bergabung khusus (SCA) pula signifikan untuk semua ciri kecuali ketinggian tumbuhan dan diameter batang atas. Keputusan ini menunjukkan bahawa tindakan gen aditif dan bukan-aditif terlibat dalam kawalan genetik bagi ciri-ciri tersebut. Magnitud varians GCA adalah jauh lebih tinggi daripada varians SCA kecuali diameter batang atas, bilangan nod, dan berat batang segar dengan daun dan buah, menunjukkan bahawa tindakan gen aditif yang mendominasi ciri-ciri ini. Induk P₁ (G5: ML5), P₃ (G13: ML36-10) and P₄ (G21: ML36-24) mempunyai keupayaan bergabung am yang tinggi untuk hasil fiber dan ciri komponen hasil yang lain. Kebanyakan ciri menunjukkan tindakan gen dominan lampau, kecuali ketinggian pokok, diameter batang tengah, diameter batang atas, hari berbunga pertama, berat batang kering, berat fiber kering dan berat 1000 biji, yang menunjukkan dominan separa. Mengambil kira keupayaan bergabung khusus dan nilai heterosis, hibrid P₁ (G5) × P₄ (G21), P₁ (G5) × P₉ (G31), P₂ (G9) × P₃ (G13), P₂ (G9) × P₅ (G22), P₄ (G21) × P₆ (G24), P₄ (G21) × P₇ (G28), P₄ (G21) × P₉ (G31), P₅ (G22) × P₈ (G30) dan P₇ (G28) × P₉ (G31) didapati memberikan nilai heterotik yang tinggi terhadap hasil fiber, hasil kayu, bilangan buah setiap tumbuhan dan saiz benih yang lebih kecil. Sembilan hibrid ini disyorkan untuk penilaian selanjutnya di percubaan berbilang lokasi bagi memilih beberapa hibrid unggul dengan hasil fiber yang tinggi untuk penanaman komersial di Malaysia.

ACKNOWLEDGEMENTS

I would like to express my appreciation and gratitude to the Almighty Allah, the most merciful for his strengths and blessing in completing this study. My sincere gratitude goes to Prof. Dr. Mohd Rafii bin Yusop, the chairman of my supervising committee, for his patience, encouragement, constant understanding, support, benevolence, excitement, and direction during my studies. I would like to further extend my gratitude to my supervisory committee members, Dr. Azizah binti Misran, Dr. Zulkarami bin Berahim, and Dr. Zaiton binti Ahmad, for their guidance and suggestions throughout my research. I am greatly thankful to the Universiti Putra Malaysia for giving me the opportunity to pursue my PhD program in Malaysia. I would like to express my heartfelt and deep appreciation to the Ministry of Agriculture of the People's Republic of Bangladesh, the Bangladesh Agriculture Research Council, and the Bangladesh Jute Research Institute for providing me with sufficient funding and other resources. I am grateful to the UPM International Students Association and the Bangladeshi Students Union of Malaysia for allowing me to serve as an organizer and for their continued support and encouragement during my studies. I'm also grateful to Nuclear Malaysia's management and personnel for all their prompt responses and helpful advice throughout my experiments. I would like to special thanks to the staffs from the Institute of Tropical Agriculture and Food Security, Universiti Putra Malaysia for their excellent technical support and all of Prof. Rafii's students, as well as Bangladeshi students, are my colleagues and friends for making my time as a postgraduate student so enjoyable. I also would like to express my sincere thanks to Dr. Yussuf Oladosu Adeniyi and Mr. Md Mahmudul Hasan Khan for their constructive critique, which helped make this study a success in some way. Finally, I would like to offer my heartfelt thanks to my dearest parents, my lovely wife, sisters, and children who always believed in me and made me believe in myself to perform to my maximum ability.

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Mohd Rafii bin Yusop, PhD

Professor
Institute of Tropical Agriculture and Food Security
Universiti Putra Malaysia
(Chairman)

Azizah binti Misran, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Zulkarami bin Berahim, PhD

Senior Research Officer
Institute of Tropical Agriculture and Food Security
Universiti Putra Malaysia
(Member)

Zaiton binti Ahmad, PhD

Senior Research Officer
Malaysian Nuclear Agency
Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 10 November 2022

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in Rule 41 in Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____
Name of
Chairman of
Supervisory
Committee: Professor Dr. Mohd Rafii bin Yusop

Signature: _____
Name of
Member
of Supervisory
Committee: Dr. Azizah binti Misran

Signature: _____
Name of
Member
of Supervisory
Committee: Dr. Zulkarami bin Berahim

Signature: _____
Name of
Member
of Supervisory
Committee: Dr. Zaiton binti Ahmad

TABLE OF CONTENTS

		Page
ABSTRACT		i
ABSTRAK		iii
ACKNOWLEDGEMENTS		v
APPROVAL		vi
DECLARATION		viii
LIST OF TABLES		xiii
LIST OF FIGURES		xvi
LIST OF APPENDICES		xviii
LIST OF ABBREVIATIONS		xix
CHAPTER		
1	INTRODUCTION	1
	1.1 General introduction	1
	1.2 Problem Statement	2
	1.3 Research objectives	3
	1.3.1 Main Objective	3
	1.3.2 Specific Objectives	3
 2	LITERATURE REVIEW	 4
	2.1 Kenaf (<i>Hibiscus cannabinus L.</i>)	4
	2.1.1 Taxonomy and botanical description of kenaf	4
	2.1.2 Origin and distribution	5
	2.1.3 Morphological characteristics	6
	2.1.4 Cultivation and agronomic practices	7
	2.1.5 Commercial potentials of kenaf	8
	2.1.6 Growing of kenaf in Malaysia	10
	2.2 Genetic diversity of kenaf	11
	2.2.1 Correlation and Path Coefficient Analysis	11
	2.2.2 Heritability and genetic advance	13
	2.3 History of plant mutagenesis	14
	2.3.1 Mutation Breeding	15
	2.3.2 Practical considerations in induced crop mutations	16
	2.3.3 Mutation breeding approach for developing mutants	16
	2.3.4 Impact of mutant cultivars	18
	2.4 Concept of Molecular Breeding	20
	2.4.1 Expressed sequence tag SSRs (EST-SSR)	20
	2.4.2 Application of EST-SSR marker for kenaf breeding	21
	2.5 Concept of Diallel Mating Design	21
	2.5.1 Kenaf improvement through Diallel Mating Design	22
	2.5.2 Heterosis and hybrid kenaf breeding	23

3	GENETIC VARIABILITY, CORRELATION AND PHENOTYPIC PATH COEFFICIENT ANALYSIS FOR HIGH FIBER YIELD IN KENAF MUTANTS UNDER TROPICAL CONDITIONS	25
3.1	Introduction	25
3.2	Materials and Methods	26
3.2.1	Experiment location	26
3.2.2	Plant materials	26
3.2.3	Experimental design and field layout	27
3.2.4	Crop husbandry	27
3.2.5	Data collection	27
3.2.6	Statistical analysis	29
3.3	Results and Discussion	34
3.3.1	Qualitative variation	34
3.3.2	Morpho-physiological and yield characters	38
3.3.3	Mean performance of genotypes over two seasons	40
3.3.4	Genetic analysis, broad-sense heritability, and genetic advance	43
3.3.5	Multivariate statistical analysis	45
3.4	Conclusion	56
4	MOLECULAR CHARACTERIZATION AND GENETIC DIVERGENCE AMONG KENAF MUTANTS AND INBRED LINES USING EST-SSR MARKERS, AS WELL AS SELECTION OF PARENTAL INBRED LINES FOR HIGH FIBER YIELD	57
4.1	Introduction	57
4.2	Materials and Methods	58
4.2.1	Plant materials	58
4.2.2	Molecular marker	58
4.2.3	DNA extraction and quantification	59
4.2.4	PCR amplification	59
4.2.5	Gel electrophoresis	60
4.2.6	Band scoring	60
4.2.7	Statistical analysis	60
4.3	Results and Discussion	61
4.3.1	Analysis of polymorphisms in the EST-SSR markers	61
4.3.2	Genetic diversity in kenaf flowering group populations	63
4.3.3	Band patterns across populations	64
4.3.4	Analysis of molecular variance (AMOVA) using EST-SSR markers	65
4.3.5	Genetic relationship among the accessions	66
4.3.6	Evaluation of genetic relationships among 31 kenaf genotypes	68
4.3.7	Comparison of two distance measures used to assess genetic diversity	71
4.3.8	Selection of genotypes for future improvement	73
4.4	Conclusion	74

5	ESTIMATION OF COMBINING ABILITIES AND HETEROSIS VALUES FOR HYBRID KENAF SELECTION BASED ON YIELD PERFORMANCE	75
5.1	Introduction	75
5.2	Materials and Methods	76
5.2.1	Experimental location	76
5.2.2	Breeding materials	76
5.2.3	Development of F ₁ using half diallel mating design	76
5.2.4	Stages for crossing	77
5.2.5	Experimental design and field layout	77
5.2.6	Crop husbandry	77
5.2.7	Data collection	78
5.2.8	Statistical analysis	78
5.3	Results and discussion	86
5.3.1	Qualitative variation	86
5.3.2	Variation among all genotypes for quantitative traits	91
5.3.3	Mean performance of genotypes over two environments	92
5.3.4	Estimation of heterosis effect of kenaf hybrids for fiber yield components	97
5.3.5	Combining ability analysis of kenaf hybrids for fiber yield components	102
5.3.6	V _r -W _r regression analysis	116
5.3.7	Correlation among fiber yield components	129
5.4	Conclusion	132
6	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	133
6.1	Summary	133
6.2	General Conclusion	138
6.3	Recommendations for Future Research	138
	REFERENCES	140
	APPENDICES	164
	BIODATA OF STUDENT	182
	LIST OF PUBLICATIONS	183

LIST OF TABLES

Table	Page	
2.1	Prices of Malaysian kenaf fiber in November 2021	8
2.2	Statistics of kenaf dried stem/seed production by state of Malaysia from the annual publications of the National Kenaf and Tobacco Board (NKTB)	10
3.1	List of 31 kenaf genotypes including 28 mutant lines derived from V36 variety through acute and chronic gamma irradiation	26
3.2	List of quantitative traits studied from 31 kenaf genotypes	28
3.3	Key out of ANOVA table for individual seasons	29
3.4	Growth and mature stage characteristics of 31 kenaf genotypes	36
3.5	Analysis of variance for 15 characters for the 31 kenaf genotypes studied over two seasons	39
3.6	Fiber yield components of kenaf genotypes planted in two growing seasons	41
3.7	Estimation of genetic parameters of 15 quantitative traits of 31 germplasm of kenaf over two seasons	44
3.8	Estimation of principal component of 15 quantitative traits of 31 kenaf genotypes	46
3.9	Component loadings of the five principal components of 15 quantitative traits from 31 kenaf lines	47
3.10	Grouping of 31 kenaf genotypes according to cluster analysis	48
3.11	Estimates of phenotypic (indicated in bold), genotypic and environment correlation coefficients for 12 traits in 31 kenaf genotypes	50
3.12	Phenotypic path analysis was used to investigate the direct (diagonal) and indirect effects of eleven traits on dry fiber yield in 31 kenaf genotypes	53
3.13	First order and second-order relationships	54
3.14	Fiber yield per plant's second-order component	55
4.1	List of kenaf genotypes used in this study	58

4.2	Characteristics of 10 EST-SSR markers used to identify 31 kenaf genotypes	59
4.3	Genetic information revealed by EST-SSR markers amplified among 31 kenaf genotypes	62
4.4	Estimation of genetic diversity among three flowering groups of the 31 kenaf genotypes	64
4.5	Determination of the inter-population genetic variances by using AMOVA	66
4.6	Distribution of 31 kenaf genotypes under diverse clusters using EST-SSRs data	67
4.7	Genetic identity (above diagonal) and genetic distance (below diagonal) values among 31 kenaf genotypes	69
4.8	Five higher and lower Nei's (1972) genetic distance (D) between pairs of kenaf genotypes were determined using EST-SSR markers	71
4.9	Comparison between clustering on morphological and molecular parameters	72
5.1	Salient features of nine selected kenaf genotypes used as parents for diallel cross	76
5.2	ANOVA table key-out to test the effects of environments, blocks within environment, genotypes, and genotypes by environments in the combined analysis	78
5.3	ANOVA table for diallel analysis using Griffing's Method 2 on data from kenaf traits and environments	80
5.4	Hayman's diallel ANOVA after Morley Jones modification for kenaf traits and environments	82
5.5	Characteristics of selected parents' growth stages and the F ₁ kenaf population	87
5.6	Combined analysis of variance was performed for 15 quantitative traits of nine parents and their crosses over two environments	92
5.7	Mean performance of nine parents and their hybrids for 15 quantitative traits in kenaf	93
5.8	Estimates of mid-parent and better parent heterosis for 15 quantitative traits of 36 crosses in kenaf over two environments	99

5.9	Mean squares in ANOVA across two environments for 15 quantitative traits from 9×9 diallel cross (Griffing's Method 2)	103
5.10	Estimates of general combining ability effect for 15 quantitative traits of kenaf	106
5.11	Estimates of specific combining ability effects for 15 quantitative traits of kenaf	108
5.12	Mean squares of analysis of variance for 15 quantitative traits of nine parents and their crosses over two environments	113
5.13	Components of genetic variance (Hayman, 1954b) for all nine parents and their crosses over two environments for all traits studied	115
5.14	Homogeneity tests for hypothesis validity were performed on 15 traits of kenaf in 9×9 half diallel's analysis	116
5.15	Correlation coefficient among 15 quantitative traits of nine parents and their crosses studied at combined in two environments	131

LIST OF FIGURES

Figure		Page
2.1	Kenaf and related fibers produced worldwide from 2017/2018	6
2.2	Photograph shows the kenaf inner and outer stem portions, as well as a schematic of the kenaf parts	6
2.3	General flowchart of cultivation procedure of kenaf	8
2.4	Path diagram and factor coefficients influencing yield component	13
2.5	Mutation breeding procedure for obtaining new superior varieties	17
2.6	Distribution of officially released mutant crop varieties based on the continents	19
2.7	Distribution of officially released mutant crop varieties based on crop types	19
3.1	Some qualitative variables of kenaf genotypes	37
3.2	Some qualitative characters' variation (a) Stem color, (b) Leaf shape, (c) Leaf color, (d) Petiole color, (e) Flower color, (f) Pod shape, (g) Seed shape, and (h) Seed coat color	37
3.3	Clustering pattern of the quantitative traits at dissimilarity coefficient of 0.21	45
3.4	PCA - 2D graphical association exhibiting six heterotic groupings generated from 31 kenaf genotypes' quantitative traits	46
3.5	PCA biplot with cases loading of 31 kenaf genotypes using XLSTAT	47
3.6	The influence of first-order on second-order components and the latter on yield per hectare is depicted in a path diagram	52
4.1	Microsatellite profiles of 31 kenaf genotypes at locus KU89646262 (A) and KU896456 (B); M: molecular wt. marker (100 bp DNA ladder)	62
4.2	Banding patterns among the population of the three days to flowering groups using EST-SSRs data set	65

4.3	Pie chart showing the percentage of inter-population molecular variances by using AMOVA	65
4.4	Unweighted pair group method of arithmetic mean (UPGMA) cluster tree shows five heterotic groups derived from 31 kenaf genotypes	67
5.1	Photographs of the parents' and F ₁ pop's stem coloration (70 DAS)	88
5.2	Photographs of the parents' and F ₁ pop's leaf shapes and coloration	89
5.3	Photographs of the parents and F ₁ populations pod shapes and colors	90
5.4	Qualitative variation in the kenaf F ₁ population (a) Stem color, (b) Leaf shape, (c) Leaf color (lamina), (d) Petiole color, (e) Pod shape, (f) Seed shape, and (g) Seed coat color	90
5.5 (a)	W _r /V _r graphs for Plant height	118
5.5 (b)	W _r /V _r graphs for Stem base diameter	118
5.5 (c)	W _r /V _r graphs for Core diameter	119
5.5 (d)	W _r /V _r graphs for Stem middle diameter	120
5.5 (e)	W _r /V _r graphs for Stem top diameter	121
5.5 (f)	W _r /V _r graphs for Nodes number	121
5.5 (g)	W _r /V _r graphs for Days to first flowering	122
5.5 (h)	W _r /V _r graphs for Days to 50% flowering	123
5.5 (i)	W _r /V _r graphs for Fresh stem weight with leaves and pod	124
5.5 (j)	W _r /V _r graphs for Fresh stem weight without leaves and pod	124
5.5 (k)	W _r /V _r graphs for Dry stick weight	125
5.5 (l)	W _r /V _r graphs for Dry fiber weight	126
5.5 (m)	W _r /V _r graphs for Number of pods per plant	127
5.5 (n)	W _r /V _r graphs Number of seeds per pod	127
5.5 (o)	W _r /V _r graph for 1000 seeds weight	128

LIST OF APPENDICES

Appendix		Page
A1	Descriptors and descriptor states in kenaf	164
A2	Schematic representation of development of kenaf hybrids through diallel cross	166
A3	Meteorological data of the study location in five seasons	167
A4	Field evaluation for 31 kenaf genotypes repeatedly over two seasons	168
A5	ANOVA of 15 morpho-physiological traits of first field expt. of kenaf	169
A6	ANOVA of 15 morpho-physiological traits of second field expt. of kenaf	169
B1	Details of the selected 71 SSR markers used in the study (chromosome number, position, their sequences, annealing temperature and expected range of allele)	170
B2	Flow chart showing processes leading to genotyping (Southern blot analysis)	172
B3	A typical nanoDrop spectrophotometer result to determine the purity, concentration of DNA sample	172
B4	Typical nanoDrop spectrophotometer result taken and formulae for calculation of DNA working solution	173
C1	Photographs of the pollination and emasculation of kenaf hybrids developed by diallel cross	174
C2	Over two environments, nine parents and 36 F ₁ s were evaluated in the field	175
C3	Photographs of the parents' and F ₁ populations' leaf shape and stem pigmentation	176
C4	Comparison of photographs showing stem pigmentation and leaf shape of parents and F ₁ population	177
C5	Nine mutant hybrids showed promising heterotic responses and desirable SCA effects for fiber yield and yield contributing traits	181

LIST OF ABBREVIATIONS

AFLP	Amplified Fragment Length Polymorphism
AMOVA	Analysis of Molecular Variance
ANOVA	Analysis of Variance
Avg. He	Average heterozygosity
Bfb	Black with few brownish
BJRI	Bangladesh Jute Research Institute
Bp	Base pairs
BP	Mean value of better parent
CC	Completely cream with white stigma
CD	Core diameter
Cpc	Cream with purple Centre
CTAB	Cetyltrimethylammonium Bromide
CV	Coefficient of variation
D50%F	Days to 50% flowering
DAS	Days after sowing
DF	Degrees of freedom
DFW	Dry fiber weight
DNA	Deoxyribonucleic acid
DNase	Deoxyribonuclease
dNTP	Deoxy nucleotide triphosphate
DSW	Dry stick weight
DFFF	Days to first flowering
EDTA	Ethylenediaminetetraacetic acid
EMS	Error Means Square

EST	Expressed Sequence Tag
FAO	Food and Agriculture Organization of the United Nations
Fst	Genetic differentiation
FW1	Fresh stem weight with leaves and pod
FW2	Fresh stem weight without leaves and pod
G × E	Genotype × environmental interaction
GCA	General Combining Ability
Grp	Green with reddish patches
Gy	Gray
H ₁	Dominance
H ₂	Dominance × dominance
h ²	Overall dominance
h ² _B	Heritability in broad sense
h ² _N	Heritability in narrow sense
HCl	Hydrochloric acid
He	Expected heterozygosity
Ho	Observed heterozygosity
IAEA	International Atomic Energy Agency
IJO	International Jute Organization
LD ₅₀	50% lethal dose
LSD	Least Significant Difference
M ₁	First generation after mutagenic treatment
M ₂	Second generation after mutagenic treatment
M ₃	Third generation after mutagenic treatment
M ₄	Fourth generation after mutagenic treatment

M ₅	Fifth generation after mutagenic treatment
M ₆	Sixth generation after mutagenic treatment
M ₇	Seventh generation after mutagenic treatment
mm	Millimeter (s)
mM	MilliMolar
MNA	Malaysian Nuclear Agency
MP	Mean value of two parental involves in F ₁
MVD	Mutant Variety Database
Na	Average observed number of alleles
Ne	Average effective number of alleles
NF	Number of pods per plant
ng	Nano gram
Nm	Estimate of gene flow among populations
nm	Nano meter
NN	Number of nodes
NS	Non-significant
NS	Number of seeds per pod
OA Mean	Overall mean
PC	Principal component
PCA	Principal component analysis
PCO	Principal co-ordinate analysis
PCR	Polymerase Chain Reaction
PH	Plant height
PIC	Polymorphism Information Content
RaGb	Reddish above greenish below

RAPD	Random Amplified Polymorphic DNA
RCBD	Randomized Complete Block Design
S.O.V	Source of variation
SAS	Statistical Analysis System
SBD	Stem base diameter
SDS	Sodium dodecyl sulphate
SE	Standard error
SI	Shannon's information index
SMD	Stem middle diameter
SSR	Simple sequence repeat
STD	Stem top diameter
SW	1000 seeds weight
Taq	Thermus aquaticus
TBE	Tris borate EDTA
TE	Tris/EDTA
TR	Trace reddish
Tris	Tris (hydroxymethyl) amino methane
uHe	Unbiased expected heterozygosity
UPGMA	Unweighted pairs group method of arithmetic mean
UPM	Universiti Putra Malaysia
UrLg	Upper surface light reddish but lower surface green
UV	Ultraviolet
μl	Microliter
μM	Micromolar

CHAPTER 1

INTRODUCTION

1.1 General introduction

Kenaf (*Hibiscus cannabinus* L.) is an industrial crop belonging to the Malvaceae family and the genus *Hibiscus*. There are over 200 species in this family including hibiscus (*Hibiscus hibiscum* L.), hollyhock (*Althaea rosea*), cotton (*Gossypium hirsutum* L.), and okra (*Hibiscus esculentus*) that are commercially grown in several countries, including Malaysia. Kenaf is a dicot plant with two distinct fiber types: short core fibers, which make up 70 to 75% of the dry weight of the stalk, and long bark fibers, which make up the remaining 25 to 30% of the fiber component (Wong et al., 2008; Lips et al., 2009). Kenaf is a multipurpose crop that produces biomass for energy and natural fiber for industrial uses, owing to its height growth and fiber content (Dauda et al., 2013). The protein content of kenaf leaves suggests that it could be used as a high-protein base feed for livestock (Hollowell et al., 1997). Because of increased concerns about global warming and the rising cost of petroleum-based products, kenaf has been praised as the fiber crop of the twenty-first century, with a short breeding season of four months (Saba et al., 2015a).

Kenaf is one of the most essential crops cultivated for its smooth fiber production. It has exceptional air permeability, antibacterial properties, and excellent biological qualities such as salinity tolerance, drought resistance, wide adaptability, and high yield. The key factor for effective use of this natural fiber material is to ensure the sustenance of high-quality raw materials to meet the growing global demands. Hence, understanding the genetic basis for kenaf planting materials is a prerequisite for a successful breeding program.

Genetic variability is a dynamic property of germplasm, and it can be estimated using molecular, biochemical, and morphological assessment (Oladosu et al., 2015). Among the three methods, morphological evaluation is the most cost-effective and readily quantifiable, making it the best alternative for crop improvement programs. In comparison to conventional breeding, mutation induction is the best way to develop new varieties with desirable traits within a short period (Oladosu et al., 2016). Correlation coefficient studies between characters are instrumental in determining effective breeding procedures (Tulu, 2014). Similarly, selection and recommendation for commercial cultivation are only proper after comprehensive information and research of available germplasm.

Simple sequence repeats (SSRs) or Microsatellites have shown clear superiority over other molecular markers due to high reproducibility and abundance, easy scoring, co-dominant nature, and extensive coverage. SSRs are classified into genomic SSRs (gSSRs) and expressed sequence tag SSRs (EST-SSRs) based on their source. Genomic SSRs are usually related to non-coding parts, while EST-SSRs are derived from the

genome expressed parts (Li et al., 2004). Germplasm evaluation using SSRs derived from ESTs may improve the role of genetic markers by analyzing variations in transcribed and known function genes (Datta et al., 2021). Therefore, EST-SSRs are among the important genetic markers for marker-assisted breeding, high-density genetic mapping, and analysis of genetic diversity (Li et al., 2016). In addition, the transcribed region of the genome contains EST-SSRs, which may be relatively well conserved and represent a better relationship between species or varieties.

Compared to alternative mating designs, the diallel analysis is an effective method for screening parents for hybrid production. General combining ability (GCA) refers to a parent's average performance in a series of crossings. Parents with a high GCA effect have additive gene action, but they do not always have a favorable SCA in their combination (Santha et al., 2017). Meanwhile, determining the sort of gene action that affects the phenotypes of interest using SCA estimation is useful in genetic research. A high SCA identifies non-additive gene action (Virmani, 2003). SCA and GCA data aid in selecting hybrids and parents for successful breeding (Patel et al., 2013). Therefore, strong hybrids are produced by parents who have good general combining ability (Shattuck et al., 1993).

1.2 Problem Statement

As part of the ASEAN Free Trade Area (AFTA), the Malaysian government encouraged kenaf planting to replace tobacco to reduce tobacco import duties by 2010 (Kamal, 2014). Despite its wide distribution in tropical countries, kenaf productivity in Malaysia is low due to a lack of high-yielding varieties with fiber production of around 5 to 10 tons per hectare and research into developing new kenaf varieties is still lacking (Sani et al., 2017). Developing new kenaf varieties that produce high biomass is vital for effective kenaf production (Al-Mamun and Saha, 2017). Thus, breeding kenaf in Malaysia is essential for producing high fiber content and adaptable growth in the local climates (Sani et al., 2017).

In some countries, such as China, Russia, and Thailand, kenaf hybrids are cultivated commercially, contributing to increasing production (Liu, 2005). In Malaysia, hybrid kenaf technology is considered novel and more research is required to assess genetics, agronomy, and crop management. A hybrid kenaf breeding program is required to develop a high-yielding, stable performance hybrid kenaf for the Malaysian environment. The availability of high-quality jute and kenaf seed would help ensure fiber quality to a large extent (Al-Mamun et al., 2017). As a result, developing locally adapted hybrid kenaf seeds is a viable alternative for enhancing national kenaf yield and increasing kenaf producer income.

1.3 Research objectives

1.3.1 Main Objective

To create stable kenaf hybrids with high fiber yield for commercial cultivation.

1.3.2 Specific Objectives

- i. To determine the genetic variability, heritability, and paths of influence among various yield components of selected kenaf mutants and inbred lines for high fiber yield.
- ii. To analyze the phylogenetic relationship and genetic divergence among kenaf mutants and inbred lines using molecular and agronomic characterizations to select potential parents for heterotic hybridization.
- iii. To estimate the nature of gene action and the magnitude of heterosis, assess the combining ability of selected parents and their hybrids.
- iv. To select stable and high yielding kenaf hybrids for dual-purpose (fiber and seeds) commercial cultivars in Malaysia's hot climate.

REFERENCES

- Abaza, G. M. S. M., Awaad, H. A., Attia, Z. M., Abdel-lateif, K. S., Gomaa, M. A., Abaza, S. M. S. M. and Mansour, E. (2020). Inducing potential mutants in bread wheat using different doses of certain physical and chemical mutagens. *Plant Breeding and Biotechnology* 8(3): 252-264.
- Abd El-Satar, M. A., Fahmy, R. M. and Hassan, T. H. A. (2015). Genetic control of sunflower seed yield and its components under different edaphic and climate conditions. *The 9th Plant Breeding International Conference* September 2015, Egyptian Journal of Plant Breeding 19(5): 103–123.
- Abu, F., Mat Taib, C. N., Mohd Moklas, M. A. and Mohd Akhir, S. (2017). Antioxidant properties of crude extract, partition extract, and fermented medium of *Dendrobium sabin* flower. *Evidence-Based Complementary and Alternative Medicine* 2017.
- Abu Sin, M. (2019). Genetics and combining ability of corn (*Zea mays* L.) genotypes for forage utilization. Doctoral thesis, Universiti Putra Malaysia.
- Agbolade, O., Nazri, A., Yaakob, R., Ghani, A. A. and Cheah, Y. K. (2019). 3-Dimensional facial expression recognition in human using multi-points warping. *BMC Bioinformatics* 20(1): 1-15.
- Agrobase, T. M. (2000). Agronomix Software Inc., 171 Waterloo Street Winnipeg, Manitoba, R3N0S4, Canada.
- Ahmad, F., Ahmad, Z., Hassan, A. A., Ariffin, S., Noordin, N., Salleh, S., Hussein, S., Akil, M., Sani, M. Z., Harun, A. R. and Rahim, K. A. (2018). A review on gamma greenhouse as a chronic gamma irradiation facility for plant breeding and improvement program. *Jurnal Sains Nuklear Malaysia* 30(1): 8-18.
- Ahmad, S. H., Rasid, R., Bonnia, N. N., Zainol, I., Mamun, A. A., Bledzki, A. K. and Beg, M. D. H. (2011). Polyester-kenaf composites: effects of alkali fiber treatment and toughening of matrix using liquid natural rubber. *Journal of Composite Materials* 45(2): 203-217.
- Ahmad, Z., Abu Hassan, A., Salleh, S., Ariffin, S., Shamsudin, S., and Basiran, M. N. (2012). Improvement of Malaysian ornamental plants through induced mutation. *Pertanika Journal of Tropical Agricultural Sciences* 35(3): 631-636.
- Aisha, A. H., Rafii, M. Y., Rahim, H. A., Juraimi, A. S., Misran, A., and Oladosu, Y. (2017). Radio-sensitivity test of acute gamma irradiation of two variety of chili pepper chili Bangi 3 and chili Bangi 5. *International Journal of Scientific and Technology Research* 7(12): 3-8.
- Aifen, T., Jianmin, Q., and Peiqing, L. (2008). Cluster analysis and evaluation of elite kanaf germplasm based on principal components. *Scientia Agricultura Sinica*, 41(9): 2859-2867.

- Akil, H., Omar, M. F., Mazuki, A. M., Safiee, S. Z. A. M., Ishak, Z. M., and Bakar, A. A. (2011). Kenaf fiber reinforced composites: A review. *Materials and Design* 32(8-9): 4107-4121.
- Akinrotimi, C. A., and Okocha, P. I. (2018). Evaluations of genetic divergence in kenaf (*Hibiscus cannabinus* L.) genotypes using agro-morphological characteristics. *Journal of Plant Sciences and Agricultural Research* 2(12): 2167-0412.
- Akter, N. (2009). Genetic Analysis of Fibre and Seed Yield in Tossa Jute (*Corchorus olitorius* L.) (Doctoral dissertation, PhD. Thesis. Department of Genetics and Plant Breeding. Bangladesh Agricultural University, Mymensingh, Bangladesh).
- Alexopoulou, E., Christou, M., Mardikis, M. and Chatziathanassiou, A. (2000). Growth and yields of kenaf varieties in central Greece. *Industrial Crop and Products* 11 (2-3):163-72.
- Alexopoulou, E., Christou, M., Nicholaou, A. and Mardikis, M. (2004). BIOKENAF: a network for industrial products and biomass for energy from kenaf. In *Biomass for energy, industry, and climate protection. Proceedings of the 2nd World Biomass Conference* 10(14): 2040-2043.
- Alexopoulou, E., Papatheohari, Y., Picco, D., Di Virgilio, N. and Monti, A. (2013). Crop management. In *Kenaf: A Multi-Purpose Crop for Several Industrial Applications* 59-82. Springer, London.
- Aljane, F. and Ferchichi, A. (2007). Characterization and evaluation of six cultivars of caprifig (*Ficus carica* L.) in Tunisia. *Plant Genetic Resources Newsletter* 151: 22-26.
- Al-Mamun, M. and Saha. C. K. (2017). BJRI udbavito Pat, Kenaf o Mesta Phosoler Procholito Jaat. Bangladesh Jute Research Institute, Dhaka, Bangladesh. 2017, 1-30.
- Al-Mamun, M., Saha, C. K., Mostofa, M. G., Miah, A. and Hossain, M. Z. (2017). Identification of suitable varieties for seed production of jute in non-traditional areas of Bangladesh. *Bangladesh Journal of Plant Breeding and Genetics* 30(1): 33-38.
- Almasy, L. and Blangero, J. (2010). Variance component methods for analysis of complex phenotypes. *Cold Spring Harbor Protocols* (5): 77.
- Alza, J. O. and Fernandez-Martinez, J. M. (1997). Genetic analysis of yield and related traits in sunflower (*Helianthus annuus* L.) in dryland and irrigated environments. *Euphytica* 95(2): 243-251.
- Aminah, A., Wong, C. C. and Hashim, G. M. (2006). Production potential of kenaf for forage and fibre on BRIS under smallholder production systems. In *Fourth Technical Review Meeting on the National Kenaf Research Project, MARDI*: 15-20.

- Anuar, H. and Zuraida, A. (2011). Improvement in mechanical properties of reinforced thermoplastic elastomer composite with kenaf bast fibre. *Composites Part B: Engineering* 42(3): 462-465.
- Aslam, M., Kashif, S. Z., Yousaf, U. and Asghar, H. (2019). Mutation Breeding: Is it supplementing the Genetic Erosion? Types of mutations: *Journal of Agriculture and Basic Sciences* 4(4): 40-56.
- Assefa, K., Ketema, S., Tefera, H., Nguyen, H. T., Blum, A., Ayele, M., Bai, G., Simane, B. and Kefyalew, T. (1999). Diversity among germplasm lines of the Ethiopian cereal tef [*Eragrostis tef* (Zucc.) Trotter]. *Euphytica* 106(1): 87-97.
- Ayadi, R., Hanana, M., Mzid, R., Hamrouni, L., Khouja, M. L. and Salhi Hanachi, A. (2017). *Hibiscus cannabinus* L. – kenaf: a review paper. *Journal of Natural Fibers* 14(4): 466-484
- Azhar, M. and Ahsanulkhalique, A. W. (2014). Gamma greenhouse: a chronic facility for crops improvement and agrobiotechnology. In *AIP Conference Proceedings* 1584(1): 32-37. American Institute of Physics.
- Bahtoe, A., Zargari, K. and Baniani, E. (2012). An investigation on fiber production of different kenaf (*Hibiscus cannabinus* L.) genotypes. *World Applied Sciences Journal* 16(1): 63-66.
- Baker, R. J. (1978). Issues in diallel analysis. *Crop Science* 18(4): 533-536.
- Baldwin, B. S. and Graham, J. W. (2006). Population density and row spacing effects on dry matter yield and bark content of kenaf (*Hibiscus cannabinus* L.). *Industrial Crops and Products* 23(3): 244-248.
- Balogun, M. O., Raji, J. A. and Akande, S. R. (2008). Morphological characterization of 51 kenaf (*Hibiscus cannabinus* L.) accessions in Nigeria. *Revista Científica UDO Agrícola* 8(1): 23-28.
- Banerjee, S., Das, M., Mir, R. R., Kundu, A., Topdar, N., Sarkar, D., Sinha, M. K., Balyan, H. S. and Gupta, P. K. (2012). Assessment of genetic diversity and population structure in a selected germplasm collection of 292 jute genotypes by microsatellite (SSR) markers. *Molecular Plant Breeding* 3:11-25
- Basri, M. H. A., Abdu, A., Junejo, N., Hamid, H. A. and Ahmed, K. (2014). Journey of kenaf in Malaysia: A Review. *Scientific Research and Essays* 9(11): 458-470.
- Basu, N. C. and Chakravarty, K. (1971). Study on growth and flowering in mesta (*Hibiscus cannabinus* L.). *Indian Agriculture* 15(1/2): 169-174.
- Behmaram, R., Saleh, G., Foroughi, M., Noori, Z., Malar Panandam, J. and Harun, J. (2014). Genetic control of fiber yield and quality in kenaf (*Hibiscus cannabinus* L.). *Iranian Journal of Genetics and Plant Breeding* 3(1): 41-31.

- Beyaz, R. and Yildiz, M. (2017). The use of gamma irradiation in plant mutation breeding. *Plant Engineering* 33-46.
- Bhattacharjee, A. K., Mukherjee, Nirmalendu, Dutta, A. N. and Goswami, K. K. (1987). Suitability of rained kenaf (*Hibiscus Canabinus*) as the source of raw materials for newsprints. *Jute Development Journal* 7(2): 27-29.
- Bhor, T. J., Pacharne, D. P. and Wagh, R. S. (2020). Genetic variability, correlation and path analysis studies in Jute (*Corchorus olitorius*) germplasm lines. *Journal of Pharmacognosy and Phytochemistry* 9(5): 359-364.
- Biswas, S. K., Islam, S. N., Sarker, M. D. H., Moniruzzaman, M. and Tareq, M. Z. (2018). Genetic variability, heritability and genetic advance for yield related characters of tossa jute (*Corchorus olitorius*) genotypes. *Journal of Bioscience and Agriculture Research* 17(01): 1416-1421.
- Boćanski, J., Srećkov, Z. and Nastasić, A. (2009). Genetic and phenotypic relationship between grain yield and components of grain yield of maize (*Zea mays* L.). *Genetika* 41(2): 145-154.
- Bondari, K. (1990). Path analysis in agricultural research, in Conference on Applied Statistics in Agriculture, Vol. 14, (Manhattan).
- Botstein, D., White, R. L., Skolnick, M. and Davis, R. W. (1980). Construction of a genetic linkage map in man using restriction fragment length polymorphisms. *American Journal of Human Genetics* 32(3): 314-331.
- Bruce, A. B. (1910). The Mendelian theory of heredity and the augmentation of vigor. *Science* 32(827): 627-628.
- Bukenya-Ziraba R. (2004). *Solanum anguivi* Lam. In: Gruben GJH and Denton OA (eds.) *Plant Resources of Tropical Africa 2. Vegetables*. Wageningen: PROTA Foundations/Backhuys Publishers/CTA.
- Carberry, P. S., Muchow, R. C., Williams, R., Sturtz, J. D. and McCown, R. L. (1992). A simulation model of kenaf for assisting fibre industry planning in northern Australia. I. General introduction and phenological model. *Australian Journal of Agricultural Research* 43(7): 1501-1513.
- Chahal, G. S. and Gosal, S. S. (2002). *Principles and procedures of plant breeding: Biotechnological and conventional approaches*. Narosa Publishing House, New Delhi, India p. 604
- Chakravarti, S. K., Singh, S., Ram, C. N., Vishwakarma, M. K. and Verma, G. S. (2017). Mutagenic effects of gamma rays and EMS in M₁ and M₂ generations in two traditional genotypes of aromatic rice (*Oryza sativa* (L.)). *International Journal of Agricultural and Statistical Sciences* 13(2), 537–543.

- Chang, S., Lee, U., Hong, M. J., Jo, Y. D. and Kim, J. B. (2020). High-throughput phenotyping (HTP) data reveal dosage effect at growth stages in *Arabidopsis thaliana* irradiated by gamma rays. *Plants* 9(5): 557.
- Chaudhury, S. K. and Sasmal, B. C. (1992). Heterosis and combining ability for fibre strength in tossa jute (*Corchorus olitorius* L.) [in India]. *Bangladesh Journal of Botany* 21 (2): 213-218.
- Chen, J., Zhou, H., Xie, W., Xia, D., Gao, G., Zhang, Q., Wang, G., Lian, X., Xiao, J. and He, Y. (2019). Genome-wide association analyses reveal the genetic basis of combining ability in rice. *Plant Biotechnology Journal* 17(11): 2211-2222.
- Chen, M. X., Wei, C. L., Qi, J. M., Chen, X. B., Su, J. G., Li, A. Q., Tao, A. F. and Wu, W. R. (2011). Genetic linkage map construction for kenaf using SRAP, ISSR and RAPD markers. *Plant Breeding* 130(6): 679-687.
- Cheng, Z., Lu, B. R., Baldwin, B. S., Sameshima, K. and Chen, J. K. (2002). Comparative studies of genetic diversity in kenaf (*Hibiscus cannabinus* L.) varieties based on analysis of agronomic and RAPD data. *Hereditas* 136(3): 231-239.
- Christie, B. R. and Shattuck, V. I. (1992). The diallel cross: design, analysis, and use for plant breeders. *Plant Breeding Reviews* 9(1): 9-36.
- Chukwu, S. C., Rafii, M. Y., Ramlee, S. I., Ismail, S. I., Oladosu, Y., Okporie, E., Onyishi, G., Utobo, E., Ekwu, L., Swaray, S. and Jalloh, M. (2019). Marker-assisted selection and gene pyramiding for resistance to bacterial leaf blight disease of rice (*Oryza sativa* L.). *Biotechnology and Biotechnological Equipment* 33(1): 440-455.
- Chukwu, S. C., Rafii, M. Y., Ramlee, S. I., Ismail, S. I., Oladosu, Y., Muhammad, I. I., Musa, I., Ahmed, M., Jatto, M. I. and Yusuf, B. R. (2020). Recovery of recurrent parent genome in a marker-assisted backcrossing against rice blast and blight infections using functional markers and SSRs. *Plants* 9(11): 1411.
- CIAT (Centro Internacional de Agricultura Tropical). 2007. Annual report 2007: Outcome line SBA-1: Improved beans for the developing world. Cali, Colombia. 210–216.
- Coetzee, R. (2004). Characterization of kenaf (*Hibiscus cannabinus* L.) cultivars in South Africa. Faculty of Natural and Agricultural Sciences, University of the Free State, Bloemfontein, South Africa, 135 pp (Doctoral dissertation, University of the Free State).
- Coetzee, R., Labuschagne, M. T. and Hugo, A. (2008). Fatty acid and oil variation in seed from kenaf (*Hibiscus cannabinus* L.). *Industrial Crops and Products* 27(1): 104-109.

- Collard, B. C., Jahufer, M. Z. Z., Brouwer, J. B. and Pang, E. C. K. (2005). An introduction to markers, quantitative trait loci (QTL) mapping and marker-assisted selection for crop improvement: the basic concepts. *Euphytica* 142(1): 169-196.
- Cordell, H. J. (2002). Epistasis: what it means, what it doesn't mean, and statistical methods to detect it in humans. *Human Molecular Genetics* 11(20): 2463-2468.
- Cosentino, S. L., Copani, V., Patanè, C., Mantineo, M. and D'Agosta, G. M. (2008). Agronomic, energetic, and environmental aspects of biomass energy crops suitable for Italian environments. *Italian Journal of Agronomy* 3(2): 81-96.
- Cox, T. S., Murphy, J. P. and Rodgers, D. (1986). Changes in genetic diversity in the red winter wheat regions of the United States. *Proceedings of the National Academy of Sciences* 83(15): 5583-5586.
- Crane, J. C. and Acuña, J. B. (1945). Effect of plant spacing and time of planting on seed yield of kenaf, *Hibiscus cannabinus* L. *Journal of the American Society of Agronomy* 37(12): 969-975.
- Crow, J. F. (2001). Heterosis. In S. Brenner and J. H. Miller (Eds.), *Encyclopedia of Genetics* 933.
- Curtis, M. (2012). DNA repair pathways and genes in plant. *Plant Mutation Breeding and Biotechnology* 57-69.
- Cyprien, M. and Kumar, V. (2011). Correlation and path coefficient analysis of rice cultivars data. *Journal of Reliability and Statistical Studies* 119-131.
- Datta, D. R., Yusop, M. R., Misran, A., Jusoh, M., Oladosu, Y., Arolu, F., Haque, A. and Sulaiman, N. M. (2021). Genetic diversity in eggplant (*Solanum melongena* L.) germplasm from three secondary geographical origins of diversity using SSR markers. *Biocell* 45(5): 1393.
- Dauda, S. M., Ahmad, D., Khalina, A. and Othman, J. (2013). Performance evaluation of a tractor mounted kenaf harvesting machine. *Academic Research International* 4(2):70-81.
- de Vries, H. (1901–1903). *Die mutationstheorie. Vol. I and II.* Leipzig (Germany): Verlag von Veit and Company.
- Dempsey, J. M. (1963). *Long vegetable fiber development in south Vietnam and other Asian countries 1957-1962.* Operation Mission, Saigon.
- Dempsey, J. M. (1975). *Kenaf, Fiber Crops,* The University Presses of Gainesville, Gainesville, FL, 203-302.
- Denis, D. and Legerski, J. (2006). Causal modeling and the origins of path analysis. *Theory and Science*, 7(1), 2-10.

- Dhillon, B. S., Singh, A. K., Lather, B. P. S. and Srinivasan, G. (2004). Advances in hybrid breeding methodology. In *Plant Breeding* 419-450. Springer, Dordrecht.
- Dudley, J. W. and Moll, R. H. (1969). Interpretation and use of estimates of heritability and genetic variances in plant breeding 1. *Crop Science* 9(3): 257-262.
- Echekwu, C. A. and Showemimo, F. A. (2004). Genetic, phenotypic, and environmental variances and character associations in Kenaf. *African Crop Science Journal* 12(4): 321-326.
- Edeerozey, A. M., Akil, H. M., Azhar, A. B. and Ariffin, M. Z. (2007). Chemical modification of kenaf fibers. *Materials Letters* 61(10): 2023-2025.
- El-Esawi, M. A., Germaine, K., Bourke, P. and Malone, R. (2016). Genetic diversity and population structure of *Brassica oleracea* germplasm in Ireland using SSR markers. *Comptes Rendus Biologies* 339(3-4): 133-140.
- Eleweanya, N. P, Uguru, M. I, Ene-Obong, E. E. and Okocha, P. I. (2005). Correlation and path coefficient analysis of grain field related characters in maize (*Zea mays* L) under umudike conditions of southeastern Nigeria. *Agro-science* 4(1): 24-28.
- Esnault, M. A., Legue, F. and Chenal, C. (2010). Ionizing radiation: advances in plant response. *Environmental and Experimental Botany* 68(3): 231-237.
- Eujayl, I., Sorrells, M. E., Baum, M., Wolters, P. and Powell, W. (2002). Isolation of EST-derived microsatellite markers for genotyping the A and B genomes of wheat. *Theoretical and Applied Genetics* 104(2): 399-407.
- Falconer, D.S. (1960). Introduction to quantitative genetics. Oliver and Boyd Ltd., Edinburgh, 1-140.
- Falconer, D. S. and Mackay, T. F. C. (1996). Introduction to quantitative genetics, Ed 4. Longmans Green, Harlow, Essex, UK.
- Falusi, O. A. (2005). Inheritance of Stem Pigmentation in Two Local Varieties of *Hibiscus sabdariffa* in Nigeria. *African Journal of Plant Science and Biotechnology* 2(2): 107-108.
- FAO/IAEA-MVD, (2020). Food and agriculture organization of the United Nations/International atomic energy agency Mutant variety database. Rome, Italy.
- Faruq, G., Alamgir, M. A., Rahman, M. M., Motior, M. R., Zakaria, H. P., Marchalina, B. and Mohamed, N. A. (2013). Morphological characterization of kenaf (*Hibiscus cannabinus* L.) in Malaysian tropical environment using multivariate analysis. *JAPS, Journal of Animal and Plant Sciences* 23(1): 60-67.

- Faruq, G., Rahman, M. M., Zabed, H. and Latif, A. (2015). Assessment of genetic variation in different kenaf (*Hibiscus cannabinus* L.) genotypes using morpho-agronomic traits and RAPD markers. *International Journal of Agriculture and Biology* 17(3): 507–514.
- Field, A. (2013). *Discovering statistics using IBM SPSS statistics*. sage.
- Forster, B. P. and Shu, Q. Y. (2012). Plant mutagenesis in crop improvement: basic terms and applications. *Plant Mutation Breeding and Biotechnology* 9-20.
- Gaurav, K., Gill, B. S. and Sohu, R. S. (2007). Heterosis and combining ability analysis for plant and seed characters in upland cotton (*Gossypium hirsutum* L.). *Journal of Cotton Research and Development* 21(1): 12-15.
- Gaurav, N., Sivasankari, S., Kiran, G. S., Ninawe, A. and Selvin, J. (2017). Utilization of bioresources for sustainable biofuels: a review. *Renewable and Sustainable Energy Reviews* 73: 205-214.
- Ghosh, T. and Chakravarty, K. (1970). Growing *Hibiscus sabdariffa* (HS Mesta) for fiber. *Jute Bull* 33(3): 1-5.
- Golam, F., Alamgir, M. A., Rahman, M. M., Subha, B. and Motior, M. R. (2011). Evaluation of genetic variability of kenaf (*Hibiscus cannabinus* L.) From different geographic origins using morpho-agronomic traits and multivariate analysis. *Australian Journal of Crop Science* 5(13): 1882-1890.
- Gomez, K. A. and Gomez, A. A. (1984). *Statistical procedures for agricultural research*. John Wiley and Sons, Singapore, 533-561.
- Goyal, S., Wani, M. R., Laskar, R. A., Raina, A. and Khan, S. (2020). Performance evaluation of induced mutant lines of black gram (*Vigna mungo* L.) Hepper. *Acta Fytotechn Zootechn* 23(2): 70-77.
- Gray, L. N., Collavino, N. G., Simón, G. E. and Mariotti, J. A. (2006). Diallelic analysis of genetic effects determining days to flowering in kenaf. *Industrial Crops and Products* 23(2): 194-200.
- Griffing, B. R. (1956). Concept of general and specific combining ability in relation to diallel crossing systems. *Australian Journal of Biological Sciences* 9(4): 463-493.
- Guang, H., Defang, L. and Anguo, C. (2009). Genetic diversity analysis of 44 shares of *Hibiscus cannabinus* L. germplasm resources using ISSR molecular marker. *Agricultural Science and Technology* 1: 284-289.
- Hallauer, A. R. (1999). Heterosis What Have We Learned? What Have We Done? Where Are We Headed? *Genetics and exploitation of heterosis in crops* 483-492.
- Hamidon, M. H., Sultan, M. T., Ariffin, A. H. and Shah, A. U. (2019). Effects of fibre treatment on mechanical properties of kenaf fibre reinforced composites: a review. *Journal of Materials Research and Technology* 8(3): 3327-3337.

- Hase, Y., Satoh, K., Seito, H. and Oono, Y. (2020). Genetic consequences of acute/chronic gamma and carbon ion irradiation of *Arabidopsis thaliana*. *Frontiers in Plant Science* 11: 336.
- Haseena, K., Jesmin, A., Islam, M. S., Sajib, A. A., Nadim, A. and Samiul, H. (2008). Microsatellite markers for determining genetic identities and genetic diversity among jute cultivars. *Australian Journal of Crop Science* 1(3): 97-107.
- Hassan, K. M., Bhuyan, M. I., Islam, M. K., Hoque, M. F. and Monirul, M. (2018). Performance of some jute and allied fiber varieties in the southern part of Bangladesh. *International Journal of Advanced Geosciences* 6(1): 117-121.
- Hayman, B. I. (1954a). The theory and analysis of diallel crosses. *Genetics* 39: 789-809.
- Hayman, B. I. (1954b). The analysis of variance of diallel tables. *Biometrics* 10(2): 235-244.
- Hayman, B. I. (1958). The theory and analysis of diallel crosses. II. *Genetics* 43(1): 63-85.
- Heliyanto, B., Hossain, M. and Basak, S. L. (1998). Genetic evaluation of several kenaf (*Hibiscus cannabinus* L.) germplasm through diallel crossing. *Indonesian Journal of Crop Sciences* 13(1): 15-22.
- Ho, W. M., Ang, L. H. and Lee, D. K. (2008). Assessment of Pb uptake, translocation and immobilization in kenaf (*Hibiscus cannabinus* L.) for phytoremediation of sand tailings. *Journal of Environmental Sciences* 20(11): 1341-1347.
- Hollowell, J. E. (1997). Nutritional and yield evaluation of kenaf (*Hibiscus cannabinus* L.) as a potential high-quality forage for the southeastern United States. Mississippi State University. ProQuest Dissertations Publishing, 1386042.
- Hong, M. J., Kim, D. Y., Ahn, J. W., Kang, S. Y., Seo, Y. W. and Kim, J. B. (2018). Comparison of radiosensitivity response to acute and chronic gamma irradiation in colored wheat. *Genetics and Molecular Biology* 41(3): 611-623.
- Hossain, M. D., Hanafi, M. M., Jol, H. and Hazandy, A. H. (2011). Growth, yield and fiber morphology of kenaf (*Hibiscus cannabinus* L.) grown on sandy bris soil as influenced by different levels of carbon. *African Journal of Biotechnology* 10(50): 10087-10094.
- Hossain, M. D., Hanafi, M. M., Saleh, G., Foroughi, M., Behmaram, R. and Noori, Z. (2012). Growth, photosynthesis and biomass allocation of different kenaf (*Hibiscus cannabinus* L.) accessions grown on sandy soil. *Australian Journal of Crop Science* 6(3): 480-487.
- Hussain, A., Zafar, Z. U., Athar, H. U. R., Farooq, J., Ahmad, S. and Nazeer, W. (2019). Assessing gene action for hypoxia tolerance in cotton (*Gossypium hirsutum* L.). *Agronomia Mesoamericana* 30(1): 51-62.

- Ibrahim, M. M. and Hussein, R. M. (2006). Variability, heritability and genetic advance in some genotypes of roselle (*Hibiscus sabdariffa* L.). *World Journal of Agricultural Sciences* 2(3): 340-345.
- Iqbal, A. M., Nehvi, F. A., Wani, S. A., Qadir R. and Dar, Z. A. (2007). Combining ability analysis for yield and yield related traits in maize (*Zea mays* L.). *International Journal of Plant Breeding and Genetics* 1(2): 101-105.
- Islam, M. S., Uddin, M. N., Haque, M. M. and Islam, M. N. (2001). Path coefficient analysis for some fibre yield related traits in white jute (*Corchorus capsularis* L.). *Pakistan Journal of Biological Sciences* 4(1): 47-49.
- Islam, M. S., Nasreen, A. L., Begum, S. E. and Haque, S. A. (2004). Correlated response and path analysis in Tossa jute (*Corchorus olitorius* L.). *Bangladesh Journal of Botany* 33(2): 99-102.
- Ismail, N. A., Rafii, M. Y., Mahmud, T. M. M., Hanafi, M. M. and Miah, G. (2019). Genetic diversity of torch ginger (*Etilingera elatior*) germplasm revealed by ISSR and SSR markers. *BioMed Research International* 2019: 1–14.
- Jakhar, D. S., Singh, R. and Kumar, A. (2017). Studies on path coefficient analysis in maize (*Zea mays* L.) for grain yield and its attributes. *International Journal of Current Microbiology and Applied Sciences* 6(4): 2851-2856.
- Jaradat, A. A. (2016). Breeding oilseed crops for climate change. In *Breeding Oilseed Crops for Sustainable Production* 421-472. Academic Press.
- Jeong, S. W., Kwon, S. J., Ryu, J., Kim, J. B., Ahn, J. W., Kim, S. H., Jo, Y. D., Choi, H. I., Im, S. B. and Kang, S. Y. (2017). Development of EST-SSR markers through de novo RNA sequencing and application for biomass productivity in kenaf (*Hibiscus cannabinus* L.). *Genes and Genomics* 39(10): 1139-1156.
- Jianmin, Q., Youyu, C., Ruiyang, Z., Lihui, L., Kangjing, L., Jianmei, W. and Pingping, F. (2005). Genetic effects and heterosis analysis for yield and quality traits in kenaf (*Hibiscus cannabinus* L.). *Zuo wu xue bao* 31(4): 469-475.
- Jinks, J. L. (1954). The analysis of continuous variation in a diallel cross of *Nicotiana rustica* varieties. *Genetics* 39(6): 767.
- Jinks, J. L. (1956). The F₂ and backcross generations from a set of diallel crosses. *Heredity* 10(1): 1-30.
- Jones, M. D., Puentes, C. and Suarez, R. (1955). Isolation of Kenaf for Seed Increase 1. *Agronomy Journal* 47(6): 256-257.
- Jones, R. M. (1965). Analysis of variance of the half diallel table. *Heredity* 20(1): 117-121.
- Johnson, H. W., Robinson, H. F. and Comstock, R. E. (1955). Genotypic and phenotypic correlations in soyabean and their implication. *Agronomy Journal* 47: 477-483.

- Juliana, A. H., Paridah, M. T., Rahim, S., Azowa, I. N. and Anwar, U. M. K. (2012). Properties of particleboard made from kenaf (*Hibiscus cannabinus* L.) as function of particle geometry. *Materials and Design* 34: 406-411.
- Kabré, V. N., Kiébré, Z., Kiébré, M., Sawadogo, B., Traoré, R. E., Sawadogo, N. and Bationo-Kando, P. (2018). Farmers' Knowledge and Interest Traits of *Hibiscus cannabinus* Grown in Burkina Faso. *Journal of Agriculture and Ecology Research International* 1-8.
- Kamal, I. B. (2014). Kenaf for bio composite: an overview. *Journal of Science and Technology* 6(2): 41-66.
- Kandil, A. A., Shareif, A. E., Abo-Zaied, T. A. and Moussa, A. G. (2012). Multivariate analysis of some economic characters in flax. *Pakistan Journal of Biological Sciences* 15(2): 85-91.
- Kang, S., Kwon, S., Jeong, S., Kim, J., Kim, S. and Ryu, J. (2016). New cultivar 'pole' of Kenaf (*Hibiscus cannabinus* L.) that can be harvested in Korea. *Korean Journal of Breeding Science* 48(3): 349-354.
- Karimi, S., Tahir, P. M., Karimi, A., Dufresne, A. and Abdul khani, A. (2014). Kenaf bast cellulosic fibers hierarchy: a comprehensive approach from micro to nano. *Carbohydrate Polymers* 101: 878-885.
- Karmakar, P. G., Hazra, S. K., Sinha, M. K. and Chaudhury, S. K. (2008). Breeding for quantitative traits and varietal development in jute and allied fibre crops. *Jute and Allied Fibre Updates* 57-75.
- Kashiani, P., Saleh, G., Abdullah, S. N. and Abdullah, N. A. P. (2008). Performance, heritability correlation studies on nine advance sweet corn inbred lines. Proceeding of the 10th Symposium of Malaysian Society of Applied Biology, Nov. 6-8, 2008. Malaysia, 48-49.
- Khalil, H. A., Yusra, A. I., Bhat, A. H. and Jawaid, M. (2010). Cell wall ultrastructure, anatomy, lignin distribution, and chemical composition of Malaysian cultivated kenaf fiber. *Industrial Crops and Products* 31(1): 113-121.
- Khan, M. M. H., Rafii, M. Y., Ramlee, S. I., Jusoh, M. and Mamun, A. (2020). Genetic variability, heritability, and clustering pattern exploration of Bambara groundnut (*Vigna subterranea* L. Verde) accessions for the perfection of yield and yield-related traits. *BioMed Research International* 2020.
- Kharkwal, M. C. (2012). A brief history of plant mutagenesis. *Plant Mutation Breeding and Biotechnology*. CABI, Wallingford, 21-30.
- Khatun, R. 2007. Improvement of fibre yield and associated traits in white jute (*Corchorus capsularis* L.). (Doctoral dissertation, Dhaka University, Bangladesh).

- Khatun, R., Hossain, M. A., Rashid, M. H., Bhuiyan, M. S. H. and Al-Mamun, M. (2007). Correlation and regression between fibre yield and other plant characters in tossa jute. *International Journal of Biology and Biotechnology* 4(4):399-401.
- Khursheed, S., Raina, A., Laskar, R. A. and Khan, S. (2018). Effect of gamma radiation and EMS on mutation rate: their effectiveness and efficiency in faba bean (*Vicia faba* L.). *Caryologia* 71(4): 397–404.
- Kimura, M. and Crow, J. F. (1964). The number of alleles that can be maintained in a finite population. *Genetics* 49(4): 725-738.
- Kobayashi, Y. (1991). In S. Arai (Ed.), Kenaf - Useful paper resource for environmental protection, 20-21.
- Kumar, D., Agrawal, R. C. and Begum, T. (2008). Analysis for identification of distinct and uniform extant jute (*Corchorus olitorius* L. and *C. capsularis* L.) varieties. *Seed Research* 36(1): 121-134.
- Kumar, K., Prasad, Y., Mishra, S. B., Pandey, S. S. and Kumar, R. (2013). Study on genetic variability, correlation and path Analysis with grain yield and yield attributing traits in green gram [*Vigna Radiata* (L) Wilczek]. *The Bioscan* 8(4): 1551-1555.
- Kwofie, J. (2015). Intra-Specific Hybridization and Performance Evaluation of Hybrid Lines of Roselle (*Hibiscus Sabdariffa* L.) (Doctoral dissertation, University of Ghana).
- Lakshmana, D., Biradar, B. D. and Ravikumar, R. L. (2009). Genetic variability studies for quantitative traits in a pool of restorers and maintainers lines of pearl millet (*Pennisetum glaucum* L.). *Karnataka Journal of Agricultural Sciences* 22(4): 881-882.
- Lagoda, P. J. L. (2012). Effects of radiation on living cells and plants. *Plant Mutation Breeding and Biotechnology* 123-134.
- Le Gouis, J., Beghin, D., Heumez, E. and Pluchard, P. (2002). Diallel analysis of winter wheat at two nitrogen levels. *Crop Science* 42(4): 1129-1134.
- Lembaga Kenaf and Tembakau Negara (2021).
- Liao, X., Zhao, Y., Kong, X., Khan, A., Zhou, B., Liu, D., Kashif, M. H., Chen, P., Wang, H. and Zhou, R. (2018). Complete sequence of kenaf (*Hibiscus cannabinus*) mitochondrial genome and comparative analysis with the mitochondrial genomes of other plants. *Scientific Reports* 8(1): 1-13.
- Li, D., Chen, A. and Gong, Y. (2000). Studies on super hybrid kenaf. Proc. 2000 *International Kenaf Symposium*, 13–14 Oct. 2000, Hiroshima, Japan. Tokyo: Japan Kenaf Association.

- Li, H., Li, D., Chen, A., Tang, H., Li, J. and Huang, S. (2016). Characterization of the kenaf (*Hibiscus cannabinus*) global transcriptome using illumina paired-end sequencing and development of EST-SSR markers. *PLoS ONE* 11(3), e0150548.
- Li, Y. C., Korol, A. B., Fahima, T. and Nevo, E. (2004). Microsatellites within genes: structure, function, and evolution. *Molecular Biology and Evolution* 21(6): 991-1007.
- Lips, S. J., de Heredia, G. M. I., den Kamp, R. G. O. and van Dam, J. E. (2009). Water absorption characteristics of kenaf core to use as animal bedding material. *Industrial Crops and Products* 29(1): 73-79.
- Liu, Y. (2005). Diallel and stability analysis of kenaf (*Hibiscus cannabinus* L.) in South Africa (Doctoral dissertation, University of the Free State).
- Liu, Y. and Labuschagne, M. T. (2009). The influence of environment and season on stalk yield in kenaf. *Industrial Crops and Products* 29(2-3): 377-380.
- Lundqvist, U., Franckowiak, J. D. and Forster, B. P. (2012). Mutation categories. *Plant mutation breeding and biotechnology*, 47-55.
- Lynch, M. and Walsh, B. (1998). *Genetics and Analysis of Quantitative Traits*. Sinauer Associates, Sunderland, MA.
- Maiti, R. K. (1997). *World Fiber Crops*. Science Publishers Inc. New Hampshire, USA, 41-61.
- Massey, P. and Nautiyal, M. (2020). Studies on induction of genetic variation through seed mutation in cowpea (*Vigna unguiculata* L. Walp.) by gamma irradiation. *International Journal of Chemical Studies* 8(1): 796-800.
- Mat Daham, M. (2015). Prospects of commercialize mechanized kenaf production and processor Proceedings of the National Conference Agricultural and Food Mechanization, Malaysia Agricultural Research and Mechanization Institute (MARDI). 23-25 June, 64-66.
- Mat Sulaiman, N. N., Rafii, M. Y., Duangjit, J., Ramlee, S. I., Phumichai, C., Oladosu, Y., Datta, D. R. and Musa, I. (2020). Genetic variability of eggplant germplasm evaluated under open field and glass house cropping conditions. *Agronomy* 10(3): 436.
- Mather, K and J. L. Jinks. (1971). *Biometrical genetics*. 2nd edition. Chapman and Hall, London.
- Matzinger, D. F., Sprague, G. F. and Cockerham, C. C. (1959). Diallel Crosses of Maize in Experiments Repeated over Locations and Years 1. *Agronomy Journal* 51(6): 346-350.

- Mazuki, A. A. M., Akil, H. M., Safiee, S., Ishak, Z. A. M. and Bakar, A. A. (2011). Degradation of dynamic mechanical properties of pultruded kenaf fiber reinforced composites after immersion in various solutions. *Composites Part B: Engineering* 42(1): 71-76.
- Mohammad, S. M. (2003). Diallel analysis for estimating combining ability of quantitatively inherited traits in upland cotton. *Asian Journal of Plant Sciences* 2(11): 853-857.
- Mohammed, M., Rozyanty, A. R., Osman, A. F., Adam, T., Hashim, U., Mohammed, A. M., Noriman, N. Z., Dahham, O. S. and Betar, B. O. (2017). The Weathering Effect in Natural Environment on Kenaf Bast Filled Unsaturated Polyester Composite and Integration of Nano Zinc particle for Water Repellent. *Micro and Nanosystems* 9(1): 16-27.
- Mohd Rifaat, A. H. (2008), Kenaf ganti tembakau, Berita Harian, online: <http://www.mtib.gov.my/repository/stayinform/kenaf-ganti-tembakau.pdf>, accessed on 15th August 2012.
- Mohd Zulmadi, S., Faiz, A., Mustapha, A., Zaiton, A., Affrida, A. H., Abdul Rahim, H. and Mohd Rafii, Y. (2017). Gamma-radiation mutagenesis of *Hibiscus cannabinus* L. V36 variety: Radiosensitivity study, phenotypic characterization, and multivariate analysis to explain variation among selected M₁ progenies, *Jurnal Sains Nuklear Malaysia* 29(2): 21-32.
- Mohndiratta, P. D. (1968). General and specific combining ability studies and graphical analysis in wheat (*Triticum aestivum* L.). M. Sc. (Ag.) Thesis, Punjab Agricultural University, Ludhiana, India.
- Moll, R. H. and Stuber, C. W. (1974). Quantitative genetics-empirical results relevant to plant breeding. In *Advances in agronomy* 26: 277-313. Academic Press.
- Monnahan, P. J. and Kelly, J. K. (2015). Epistasis is a major determinant of the additive genetic variance in *Mimulus guttatus*. *PLoS Genetics* 11(5): e1005201.
- Monti, A. (2013). Kenaf: A Multi-purpose Crop for Several Industrial Applications: New Insights from the Bio kenaf Project Springer Science and Business Media, Technology and Engineering, UK.
- Mostofa, M. G., Islam, M. R., Alam, A. T. M. M., Ali, S. M. and Mollah, M. A. F. (2002). Genetic variability, heritability, and correlation studies in kenaf (*Hibiscus cannabinus* L.). *Journal of Biological Sciences* 2(6): 422-424.
- Mostofa, M. G., Rahman, L. and Hussain, M. M. (2011). Combining ability for yield and yield contributing characters in Kenaf (*Hibiscus cannabinus* L.). *Bangladesh Journal of Plant Breeding and Genetics* 24(1): 01-06.
- Mostofa, M. G., Rahman, L. and Ghosh, R. K. (2013). Genetic Analysis of some Important Seed Yield Related Traits in Kenaf (*Hibiscus cannabinus* L.). *Agriculture and Natural Resources* 47(2): 155-165.

- Mukewar, A. M., Zope, J. S. and Lahane, P. S. (1997). Combining ability analysis in kenaf (*Hibiscus sabdariffa* L.) *Indian Journal of Genetics and Plant Breeding* 57(2): 161-162.
- Mullainathan, L. and Aruldoss, T. (2015). Effect of Gamma Rays in Induced Morphological Mutants on M₂ Generation of Chilli (*Capsicum annum* L.) Var K 1. *International Letters of Natural Sciences* 30: 19–24.
- Myint, K. A., Amiruddin, M. D., Rafii, M. Y., Abd Samad, M. Y., Ramlee, S. I., Yaakub, Z. and Oladosu, Y. (2019). Genetic diversity and selection criteria of MPOB-Senegal oil palm (*Elaeis guineensis* Jacq.) germplasm by quantitative traits. *Industrial Crops and Products* 139, 111558.
- Myint, K. A., Yaakub, Z., Rafii, M. Y., Oladosu, Y., Samad, M. Y. A., Ramlee, S. I., Mustafa, S., Arolu, F., Abdullah, N., Marjuni, M. and Amiruddin, M. D. (2021). Genetic Diversity Assessment of MPOB-Senegal Oil Palm Germplasm Using Microsatellite Markers. *BioMed Research International*, 2021.
- Nagatomi, S. and Degi, K. (2009). Mutation breeding of chrysanthemum by gamma field irradiation and in vitro culture. Food and Agriculture Organization of United Nations, Rome, 258-261.
- Nawaz, M. A., Yang, S. H., Rehman, H. M., Baloch, F. S., Lee, J. D., Park, J. H. and Chung, G. (2017). Genetic diversity and population structure of Korean wild soybean (*Glycine soja* Sieb. and Zucc.) inferred from microsatellite markers. *Biochemical Systematics and Ecology* 71: 87-96.
- Nayak, B. K. and Baisakh, B. (2008). Character association and path analysis in tossa jute (*Corchorus olitorius* L.). *Environment and Ecology* 26(1A): 361.
- Nei, M. (1972). Genetic distance between populations. *The American Naturalist* 106(949): 283-292.
- Nei, M. (1987). Molecular evolutionary genetics. Columbia university press, New York.
- Nei, M. and Li, W. H. (1979). Mathematical model for studying genetic variation in terms of restriction endonucleases. *Proceedings of the National Academy of Sciences* 76(10): 5269-5273.
- NTSYS-pc, N. T. and Taxonomy, N. (2005). Multivariate Analysis System, version 2.2. Exeter Software: Setauket, NY, USA.
- Ogunniyan, D. J. (2016). Assessment of genetic divergence in kenaf (*Hibiscus cannabinus* L) genotypes using agro-botanical characteristics and multivariate analysis. *SABRAO Journal of Breeding and Genetics* 48(1): 61-71.
- Oladosu, Y., Rafii, M. Y., Abdullah, N., Abdul Malek, M., Rahim, H. A., Hussin, G., Abdul Latif, M. and Kareem, I. (2014). Genetic variability and selection criteria in rice mutant lines as revealed by quantitative traits. *The Scientific World Journal* 2014: 1–12.

- Oladosu, Y., Rafii, M. Y., Abdullah, N., Malek, M. A., Rahim, H. A., Hussin, G., Ismail, M. R., Latif, M. A. and Kareem, I. (2015). Genetic variability and diversity of mutant rice revealed by quantitative traits and molecular markers. *Agrociencia* 49(3): 249-266.
- Oladosu, Y., Rafii, M. Y., Abdullah, N., Hussin, G., Ramli, A., Rahim, H. A., Miah, G. and Usman, M. (2016). Principle and application of plant mutagenesis in crop improvement: a review. *Biotechnology and Biotechnological Equipment* 30(1): 1-16.
- Olawale, A. S. and Oluwatoyin, B. M. (2019). Genetic studies of fibre yield-related traits and days to anthesis in some kenaf (*Hibiscus cannabinus* L.) accessions. *Journal of Agricultural Sciences* 64(1): 9-19.
- Olawamide, D. O. and Fayeun, L. S. (2020). Correlation and path coefficient analysis for yield and yield components in late maturing pro-vitamin A synthetic maize (*Zea mays* L.) breeding lines. *Journal of Experimental Agriculture International* 64-72.
- Olayiwola, M. O., Ajala, S. O., Ariyo, O. J., Ojo, D. K. and Gedil, M. (2021). Heterotic grouping of tropical maize inbred lines and their hybrid performance under stem borer infestation and low soil nitrogen condition in West and Central Africa. *Euphytica* 217(1), 1-22.
- Olkin, I. and Sampson, A.R. (2001). Multivariate Analysis: Overview. Published in International Encyclopedia of the Social and Behavioral Sciences, 2001. Electronically.
- Omar, S. R., Ahmed, O. H., Saamin, S. and Majid, N. M. A., (2008), Gamma radiosensitivity study on chili (*Capsicum annum*), *American Journal of Applied Sciences* 5(2): 67-70.
- Osman, M., Golam, F., Saberi, S., Abdul Majid, N., Nagoor, N. H. and Zulqarnain, M. (2011). Morpho-agronomic analysis of three roselle (*Hibiscus sabdariffa* L.) mutants in tropical Malaysia. *Australian Journal of Crop Science* 5(10): 1150-1156.
- Pace, S., Piscioneri, I. and Settanni, I. (1998). Heterosis and combining ability in a half diallel cross of kenaf (*Hibiscus cannabinus* L.) in south Italy. *Industrial Crops and Products* 7(2-3): 317-327.
- Paradiž, J., Škrk, J. and Druškovič, B. (1992). Cytogenetic effects of ionizing radiation on meristem. *Acta Pharmaceutica* 42: 397-401.
- Paridah, M. T., Abdelrhman, A. H. and Shahwahid, M. (2017). Cost benefit analysis of kenaf cultivation for producing fiber in Malaysia. *Arabian Journal of Business and Management Review* 7(5).

- Patanè, C. and Sortino, O. (2010). Seed yield in kenaf (*Hibiscus cannabinus* L.) as affected by sowing time in South Italy. *Industrial Crops and Products* 32(3): 381-388.
- Patel, J. P., Singh, U., Kashyap, S. P., Singh, D. K., Goswami, A., Tiwari, S. K. and Singh, M. (2013). Combining ability for yield and other quantitative traits in eggplant (*Solanum melongena* L.). *Vegetable Science* 40(1): 61-64.
- Pathirana, R. (2011). Plant mutation breeding in agriculture. *Plant Sciences Reviews* 6(32): 107-126.
- Patil, R. C. and Thombre, M. V. (1980). Heterosis and combining ability studies in *Hibiscus cannabinus* L. *Journal of Maharashtra Agricultural Universities* 5(2): 123-126.
- Patil, R. C. and Thombre, M. V. (1981). Graphic and variance components analysis of five quantitative characters in *Hibiscus cannabinus* L. *Journal of Maharashtra Agricultural Universities* 6(3): 221-224.
- Peakall, R. O. D. and Smouse, P. E. (2006). GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and research. *Molecular Ecology Notes* 6(1): 288-295.
- Peikun, H. (1987). The preliminary study of chromosome configuration of kenaf. *China's Fibre Crops* 1: 1-249.
- Pervin, T. N. (2012). Path coefficient analysis for fibre yield related traits in deshi jute (*Corchorus capsularis* L.). *International Research Journal of Applied Life Sciences* 1(3): 72-77.
- Pham, T. D., Geleta, M., Bui, T. M., Bui, T. C., Merker, A. and Carlsson, A. S. (2011). Comparative analysis of genetic diversity of sesame (*Sesamum indicum* L.) from Vietnam and Cambodia using agro-morphological and molecular markers. *Hereditas* 148(1): 28-35.
- Pirchner, F. (1964). Populations genetik in der Tierzucht. Hamburg und Berlin.
- Quamruzzaman, A. K. M., Rashid, M. A., Ahmad, S., Rahman, M. M. and Sultana, N. A. (2007). Combining ability estimates in nine eggplant varieties. *Pakistan Journal of Science and Industrial Research* 50(1):55-59.
- Rahman, L., Molla, M. R., Sultana, S., Islam, M. N., Ahmed, N. U., Rahman, M. S. and Nazim-ud-Dowla, M. (2007). Plant varieties of Bangladesh: Morphological and molecular characterization. *Seed Wing, Ministry of Agriculture, Government of the People's Republic of Bangladesh* 1: 486.
- Rai, B. (1979). Heterosis Breeding. Agro-biological publications. Azad Nagar, New Delhi, India.

- Raina, A., Laskar, R., Khursheed, S., Amin, R., Tantray, Y., Parveen, K. and Khan, S. (2017). Role of Mutation Breeding in Crop Improvement- Past, Present and Future. *Asian Research Journal of Agriculture* 2(2): 1–13.
- Raina, A., Laskar, R. K., Jahan, R., Amin, R., Khursheed, S., Wani, M. R. and Nisa, S. K. (2018). Mutation breeding for crop improvement. Introduction to Challenges and Strategies to Improve Crop Productivity in Changing Environment. *Enriched Publications. PVT. LTD, New Delhi* 303-317.
- Raina, A. and Khan, S. (2020). Mutagenic effectiveness and efficiency of gamma rays and sodium azide in M₂ generation of Cowpea [*Vigna unguiculata* (L.) Walp.]. *BioRxiv*.
- Raji, J. A. (2007). Intercropping kenaf and cowpea. *African Journal of Biotechnology* 6(24): 2807-2809.
- Rakoczy-Trojanowska, M. and Bolibok, H. (2004). Characteristics and a comparison of three classes of microsatellite-based markers and their application in plants. *Cellular and Molecular Biology Letters* 9(2): 221-238.
- Ratner, B. (2009). The correlation coefficient: Its values range between+ 1/- 1, or do they? *Journal of Targeting, Measurement, and Analysis for Marketing* 17(2): 139-142.
- R Core Team (2020). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
- Rodriguez, F., Alvarado, G., Pacheco, A., Burgueño, A. and Crossa, J. (2018). Analysis of genetic designs with R for Windows, version 5.0. CIMMYT Research Data and Software Repository, V14.
- Rohlf, F. J. (1992). NTSYS-pc. Numerical Taxonomy and multivariate analysis system: version 2.02e. Applied Biostatistics, New York. Spice board of India.
- Rohlf, F. J. and Fisher, D. R. (1968). Tests for hierarchical structure in random data sets. *Systematic Biology*, 17(4): 407-412.
- Rojas, B. A. and Sprague, O. F. (1952). A Comparison of Variance Components in Corn Yield Trials: III. General and Specific Combining Ability and Their Interaction with Locations and Years 1. *Agronomy Journal* 44(9): 462-466.
- Rowell, R. M., Sanadi, A. R., Caulfield, D. F. and Jacobson, R. E. (1997). Utilization of natural fibers in plastic composites: problems and opportunities. *Lignocellulosic-plastics Composites* 13: 23-51.
- Roy Chowdhury, R. and Tah, J. (2013). Mutagenesis - A potential approach for crop improvement. *Crop Improvement*, 149-187. Springer, Boston, MA.

- Ryu, J., Ha, B. K., Kim, D. S., Kim, J. B., Kim, S. H. and Kang, S. Y. (2013). Assessment of growth and seed oil composition of kenaf (*Hibiscus cannabinus* L.) germplasm. *Journal of Crop Science and Biotechnology* 16(4): 297-302.
- Ryu, J., Kwon, S. J., Ahn, J. W., Jo, Y. D., Kim, S. H., Jeong, S. W., Lee, M. K., Kim, J. B. and Kang, S. Y. (2017a). Phytochemicals and antioxidant activity in the kenaf plant (*Hibiscus cannabinus* L.). *Journal of Plant Biotechnology* 44(2): 191-202.
- Ryu, J., Kwon, S. J., Kim, D. G., Lee, M. K., Kim, J. M., Jo, Y. D., Kim, S. H., Jeong, S.W., Kang, K. Y., Kim, S. W., Kim, J. B. and Kang, S. Y. (2017b). Morphological characteristics, chemical and genetic diversity of kenaf (*Hibiscus cannabinus* L.) genotypes. *Journal of Plant Biotechnology* 44(4): 416-430.
- Saba, N., Paridah, M. T., Jawaid, M., Abdan, K. and Ibrahim, N. A. (2015a). Potential utilization of kenaf biomass in different applications. *Agricultural Biomass Based Potential Materials*, 1-34. Springer, Cham.
- Saba, N., Jawaid, M., Hakeem, K. R., Paridah, M. T., Khalina, A. and Allothman, O. Y. (2015b). Potential of bioenergy production from industrial kenaf (*Hibiscus cannabinus* L.) based on Malaysian perspective. *Renewable and Sustainable Energy Reviews* 42: 446-459.
- Saba, N., Paridah, M. T., Abdan, K. and Ibrahim, N. A. (2016). Dynamic mechanical properties of oil palm nano filler/kenaf/epoxy hybrid nanocomposites. *Construction and Building Materials* 124: 133-138.
- Sabri, R. S., Rafii, M. Y., Ismail, M. R., Yusuff, O., Chukwu, S. C. and Hasan, N. A. (2020). Assessment of agro-morphologic performance, genetic parameters and clustering pattern of newly developed blast resistant rice lines tested in four environments. *Agronomy* 10(8): 1098.
- Sani, M. Z., Ahmad, F., Akil, M., Ahmad, Z., Hassan, A. A., Harun, A. R. and Yusop, M. R. (2017). Gamma-radiation mutagenesis of *Hibiscus cannabinus* L. V36 variety: Radiosensitivity study, phenotypic characterization, and multivariate analysis to explain variation among selected M₁ progenies. *Jurnal Sains Nuklear Malaysia* 29(2): 21-32.
- Santha, S., Vaithilingam, R., Karthikeyan, A. and Jayaraj, T. (2017). Combining ability analysis and gene action of grain quality traits in rice (*Oryza sativa* L.) using line \tilde{A} - tester analysis. *Journal of Applied and Natural Science* 9(2): 1236-1255.
- Sao, A. and Mehta, N. (2010). Heterosis and inbreeding depression for fruit yield and its components in brinjal (*Solanum melongena* L.). *Dirasat Agricultural Sciences* 37(1): 36-45.
- Sarif, H. M., Rafii, M. Y., Ramli, A., Oladosu, Y., Musa, H. M., Rahim, H. A., Zuki, Z. M. and Chukwu, S. C. (2020). Genetic diversity and variability among pigmented rice germplasm using molecular marker and morphological traits. *Biotechnology and Biotechnological Equipment* 34(1): 747-762.

- Sarker, U., Rasul, M. G. and Mian, M. A. K. (2002). Heterosis and combining ability in rice. *Bangladesh Journal of Plant Breeding and Genetics* 15(1): 17-26.
- Sarker, U. and Mian, M. A. K. (2002). Line \times tester analysis for yield and its components in rice (*Oryza sativa* L.). *Journal of the Asiatic Society of Bangladesh* 28 (1): 71-81.
- Sawarkar, A. S., Yumnam, S. O., Patil, S. G. and Mukherjee, S. (2014). Correlation and path coefficient analysis of yield and its attributing traits in tossa jute (*Corchorus olitorius* L.). *The Bioscan* 9(2): 883-87.
- Sellers, T. (1999). Kenaf properties, processing, and products. Mississippi State University, Starkville, Mississippi. 197, 381–382, 386
- Sen, T. and Reddy, H. J. (2011). Application of sisal, bamboo, coir and jute natural composites in structural upgradation. *International Journal of Innovation, Management and Technology* 2(3): 186-191.
- Senapati, S., Ali, M. N. and Sasmal, B. G. (2006). Genetic variability, heritability and genetic advance in *Corchorus* sp. *Environmental and Ecological Statistics* 24: 1-3.
- Shaibu, A. S., Badu-Apraku, B. and Ayo-Vaughan, M. A. (2021). Enhancing Drought Tolerance and Striga hermonthica Resistance in Maize Using Newly Derived Inbred Lines from the Wild Maize Relative, *Zea diploperennis*. *Agronomy* 11(1), 177.
- Shao B. F., Weng C. J., Qui Y. D. and Zhang X. X. (1993). Quantitative analysis on the genetic and correlative characteristics in jute and Kenaf. *China's Fibre Crops* 2: 7–11.
- Sharma, J. R. (2006). Statistical and biometrical techniques in plant breeding. *New Age Internationa*, 61(4): 391-392.
- Sharma, H. K., Choudhary, S. B., Kumar, A. A., Maruthi, R. T. and Pandey, S. K. (2017). Combining ability and heterosis analysis for fibre yield and quality parameters in roselle (*Hibiscus sabdariffa* L.). *Journal of Applied and Natural Science* 9(4): 2502-2506.
- Shattuck, V. I., Christie, B. and Corso, C. (1993). Principles for Griffing's combining ability analysis. *Genetica* 90(1): 73-77.
- Shikazono, N., Yokota, Y., Kitamura, S., Suzuki, C., Watanabe, H., Tano, S. and Tanaka, A. (2003). Mutation rate and novel to mutants of *Arabidopsis thaliana* induced by carbon ions. *Genetics* 163(4): 1449-1455.
- Shirasawa, K., Hirakawa, H., Nunome, T., Tabata, S. and Isobe, S. (2016). Genome-wide survey of artificial mutations induced by ethyl methane sulfonate and gamma rays in tomato. *Plant Biotechnology Journal* 14(1): 51-60.

- Shu, Q. Y., Forster, B. P. and Nakagawa, H. (2012). Plant Mutation Breeding and Biotechnology. CAB International and FAO.
- Shukla, G. K. and Singh, D. P. (1967). Studies on heritability correlation and discriminant function selection in jute. *Indian Journal of Genetics and Plant Breeding* 27(2): 220.
- Shull, G. H. (1908). The composition of a field of maize. *Journal of Heredity* (1): 296-301.
- Shull, G. H. (1914). Duplicate genes for capsule-form in *Bursa bursa-pastoris*. *Z Indukt Abstamm Vererbungsl* 12(1): 97-149.
- Siepe, T., Ventrella, D. and Lapenta, E. (1997). Evaluation of genetic variability in a collection of *Hibiscus cannabinus* (L.) and *Hibiscus* spp (L.). *Industrial Crops and Products* 6(3-4): 343-352.
- Singh, B. P. (2010). Industrial crops and uses. CAB International, CABI Head Office, Nosworthy Way, Wallingford, Oxfordshire OX10 8DE, UK.
- Singh, B. P., Singh, H. P. and Obeng, E. (2013). "Elephant grass," in Biofuel Crops: Production, Physiology and Genetics, ed. B. P. Singh (Fort Valley, GA: CAB International), 271–291.
- Singh, R. K. and Chaudhary, B. D. (1977). Biometrical methods in quantitative genetic analysis. Kalyani Publishers, New Delhi, India.
- Sivasubramanian, S. and Menon, P. M. (1973). Genotypic and phenotypic variability in rice. *Madras Agriculture Journal* 60(9/13): 1093-1096.
- Sobhan, M. A. (1993). Heritability of Fibre, Fruit and Seed Yield in *Hibiscus sabdariffa* L (Doctoral dissertation, PhD Thesis. Department of Botany, Dhaka University. Dhaka, Bangladesh).
- Srayya, S. and Kumar, B. S. (2015). Evaluation on the mechanical properties of hybrid composites (kenaf, E-glass, jute). *International Journal of Latest Trend in Engineering and Technology* 5: 164-170.
- Steel, R. G. and Torrie, J. H. (1980). Principles and procedures of statistics: a biometrical approach, 2: 137-139. New York: McGraw-Hill.
- Su, J., Chen, A. and Lin, J. (2004). Genetic diversity, evaluation, and utilization of kenaf germplasm in China. *Plant Fibre and Products* 26(1): 5-9.
- Tao, A., Qi, J., Li, A., Fang, P. P., Lin, L. H., Wu, J. M. and Wu, W. R. (2005). The analysis of genetic diversity and relationship of elite kenaf germplasm based on inter-simple sequence repeats. *Acta Agronomica Sinica* 31(12): 1668.

- Tharazi, I., Sulong, A. B., Muhamad, N., Haron, C. H. C., Tholibon, D., Ismail, N. F., Radzi, M. K. F. M. and Razak, Z. (2017). Optimization of hot press parameters on tensile strength for unidirectional long kenaf fiber reinforced polylactic-acid composite. *Procedia Engineering* 184: 478-485.
- Thombre, M. V. and Patil, R. C. (1985). Genetic architecture of raw fibre yield and its components in *Hibiscus cannabinus* L. *Journal of Maharashtra Agricultural Universities* 10(3): 307-310.
- Thorat, I. H. and Gawande, V. L. (2021). Selection of elite parents and crosses for seed yield and its components using combining ability analysis over the three environments in safflower (*Carthamus tinctorious* L.). *Indian Journal of Genetics and Plant Breeding* 81(1): 139-143.
- Tomkowiak, A., Bocianowski, J., Kwiatek, M. and Kowalczewski, P. Ł. (2020). Dependence of the heterosis effect on genetic distance, determined using various molecular markers. *Open Life Sciences* 15(1): 1-11.
- Tuhina-Khatun, M., Hanafi, M. M., Rafii Yusop, M., Wong, M. Y., Salleh, F. M. and Ferdous, J. (2015). Genetic variation, heritability, and diversity analysis of upland rice (*Oryza sativa* L.) genotypes based on quantitative traits. *Biomed Research International* 2015: 1-7.
- Tulu, B. N. (2014). Correlation and path coefficients analysis studies among yield and yield related traits of quality protein maize (QPM) inbred lines. *International Journal of Plant Breeding and Crop Science* 1(2): 006-017.
- Usman, M. G., Rafii, M. Y., Martini, M. Y., Oladosu, Y. and Kashiani, P. (2016). Genotypic character relationship and phenotypic path coefficient analysis in chili pepper genotypes grown under tropical condition. *Journal of the Science of Food and Agriculture* 97: 1164-1171.
- Usman, M. G., Rafii, M. Y., Martini, M. Y., Oladosu, Y. and Kashiani, P. (2017). Genotypic character relationship and phenotypic path coefficient analysis in chili pepper genotypes grown under tropical condition. *Journal of the Science of Food and Agriculture* 97(4): 1164-1171.
- Verma, R. C. and Khah, M. A. (2016). Assessment of gamma rays induced cytotoxicity in common wheat (*Triticum aestivum* L.). *Cytologia* 81(1): 41-45.
- Virmani, S. S., Sun, Z. X., Mou, T. M., Jauhar, A. A. and Mau, C. X. (2003) Two-line hybrid rice breeding manual. Los Baños, Philippines: International Rice Research Institute.
- Voulgaridis, E., Passialis, C. and Grigoriou, A. (2000). Anatomical characteristics and properties of kenaf stems (*Hibiscus cannabinus*). *IAWA Journal* 21(4): 435-442.
- Wang, H. and Deng, X. W. (2018). Development of the “third-generation” hybrid rice in China. *Genomics, Proteomics and Bioinformatics* 16(6): 393-396.

- Warner, R. M. and Erwin, J. E. (2003). Effect of plant growth retardants on stem elongation of *Hibiscus* species. *HortTechnology* 13(2): 293-296.
- Webber, C., Harbans, L. and Venita, K. (2002) Kenaf production: fiber, feed and seed. In: Janick J, Whipkey A (eds) Trends in new crops and new uses, ASHS Press, Alexandria 13: 327–339.
- Wilson, F. D. (2003). Kenaf history and botany. *Economic Botany* 18: 80-91.
- Wilson, F. D. and Menzel, M. Y. (1964). Kenaf (*Hibiscus cannabinus*), roselle (*Hibiscus sabdariffa*). *Economic Botany* 18(1): 80-91.
- Wong, C. C., Mat Daham, M. D., Abdul Aziz, A. M. and Abdullah, O. (2008). Kenaf germplasm introductions and assessment of their adaptability in Malaysia. *Journal of Tropical Agriculture and Food Science* 36(1): 1-19.
- Wong, Y. H., Tan, W. Y., Tan, C. P., Long, K. and Nyam, K. L. (2014). Cytotoxic activity of kenaf (*Hibiscus cannabinus* L.) seed extract and oil against human cancer cell lines. *Asian Pacific Journal of Tropical Biomedicine* 4: 510-515.
- Wood, I. (2003). Kenaf: the forgotten fiber crop. The Australian New Crops Newsletter, 10.
- Wray, N. and Visscher, P. (2008). Estimating trait heritability. *Nature Education*, 1(1): 29.
- Wright, A. J. (1985). Diallel designs, analyses, and reference populations *Heredity*, 54(3): 307-311.
- Wright, S. (1921). Correlation, and causation. *Journal of Agricultural Research* 20(7): 557–585.
- Wynne, J. C., Emery, D. A. and Rice, P. W. (1970). Combining ability estimates in *Arachis hypogaea* L. II. field performance of F₁ hybrids 1. *Crop Science* 10(6): 713-715.
- Xu, Z. (1990). Genetic Evaluation of IJO Collected Kenaf (*Hibiscus cannabinus* L.) Germplasm. MSc. Thesis. Bidhan Chandra Krishi Viswavidyalaya. West Bengal, India.
- Yang, Z., Jin, L., Zhu, H., Wang, S., Zhang, G. and Liu, G. (2018). Analysis of epistasis among QTLs on heading date based on single segment substitution lines in rice. *Scientific Reports* 8(1): 1-8.
- Yeh, F. C. and Boyle, T. J. B. (1997). Simple genetic analysis of co-dominant and dominant markers and quantitative traits. *Belgeum Journal of Botany* 130: 229-253.

- Yeh, F. C., Yang, R.-C., Boyle, T. B. J., Ye, Z.-H. and Mao, J. X. (1999). POPGENE, the user-friendly shareware for population genetic analysis, Molecular Biology and Biotechnology Centre, University of Alberta, Edmonton, Canada.
- Youcai, G., Anping, G. and Weijie, L. (1998). Analysis in genetic components and combining ability for hybrid F₁ and their parents of kenaf. *Zhongguo Mazuo*, 20(3): 1-6.
- Yu, H. and Yu, C. (2007). Study on microbe retting of kenaf fiber. *Enzyme Microb. Technol.* 40: 1806-1809.
- Yunus, M. F., Aziz, M. A., Kadir, M. A., Daud, S. K. and Rashid, A. A., (2013), In vitro mutagenesis of *Etilingera elatior* (Jack) and early detection of mutation using RAPD markers, *Turkish Journal of Biology* 37: 716-725.
- Zhang, Y. and Kang, M. S. (1997). DIALLEL-SAS: A SAS program for Griffing's diallel analyses. *Agronomy Journal* 89(2): 176-182.
- Zhang, Y. and Kang, M. S. (2003). DIALLEL-SAS: a program for Griffing's diallel methods. In: Kang, M.S. (Ed.), *Handbook of Formulas and Software for Plant Geneticists and Breeders*. Haworth Press Inc., New York, NY, 1-19.
- Zhang, G. Q., Qi, J. M., Zhang, X. C., Fnag, P. P., Su, J. G., Tao, A. F., Tao, L. A. N., Wu, W. R. and Liu, A. M. (2011). A genetic linkage map of kenaf (*Hibiscus cannabinus* L.) based on SRAP, ISSR and RAPD markers. *Agricultural Sciences in China* 10(9): 1346-1353.
- Zhang, L., Wan, X., Xu, J., Lin, L. and Qi, J. 2015. De novo assembly of kenaf (*Hibiscus cannabinus*) transcriptome using Illumina sequencing for gene discovery and marker identification. *Molecular breeding* 35(10): 1-11.
- Zhao, Y., Chen, P., Liao, X., Zhou, B., Liao, J., Huang, Z., Kong, X. and Zhou, R. (2013). A comparative study of the atp9 gene between a cytoplasmic male sterile line and its maintainer line and further development of a molecular marker specific for male sterile cytoplasm in kenaf (*Hibiscus cannabinus* L.). *Molecular Breeding* 32(4): 969-976.
- Zheng, L., Li, F., Hao, J. and Li, G. (1995). Fluorescence probe studies of bis (2-ethylhexyl) sodium sulfosuccinate (AOT) and AOT/cetyltrimethylammonium bromide (CTAB) systems. *Colloids and Surfaces A: Physicochemical and Engineering Aspects* 98(1-2): 11-18.
- Zhou, R. (2002). Discovery of male sterile plants in kenaf (*Hibiscus cannabinus* L.). *Scientia Agricultura Sinica* 2:212.
- Zhou, R., Wu, Z., Jiang, F. L. and Liang, M. (2015). Comparison of gSSR and EST-SSR markers for analyzing genetic variability among tomato cultivars (*Solanum lycopersicum* L.). *Genetics and Molecular Research* 14(4); 13184-13194.