



**UNIVERSITI PUTRA MALAYSIA**

***POTENTIAL OF ACACIA GUM AS A PREBIOTIC  
INGREDIENT THROUGH *In Vitro* STUDIES  
USING HUMAN FAECAL MICROBIOTA***

**MUHAMAD HANIF RAWI**

**FSPM 2020 7**



**POTENTIAL OF ACACIA GUM AS A PREBIOTIC INGREDIENT THROUGH  
*In Vitro* STUDIES USING HUMAN FAECAL MICROBIOTA**

By

**MUHAMAD HANIF BIN RAWI**

**Thesis Submitted to the School of Graduate Studies, Universiti  
Putra Malaysia, in Fulfilment of the Requirements for the Degree of  
Doctor of Philosophy.**

**May 2020**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**POTENTIAL OF ACACIA GUM AS A PREBIOTIC INGREDIENT THROUGH  
*In Vitro* STUDIES USING HUMAN FAECAL MICROBIOTA**

By

**MUHAMAD HANIF BIN RAWI**

May 2020

**Chair : Shahrul Razid Sarbini, PhD**  
**Faculty: Agriculture and Food Sciences, Bintulu Campus**

There is a growing consensus of acacia gum, it is currently marketed as functional food to improve overall human health particularly the gut. This natural product of Acacia tree is the branched polysaccharide gummy exudate made up of complex biopolymer of arabinose and galactose monosaccharides. It is widely used in African countries and in the Middle East as a traditional medicine. Recently, the consumption of acacia gum has been related to potential health benefits. In terms of its potential prebiotics properties, this may be particular relevant to treat stomach disease. Thus, in this research commercially available acacia gum was investigated *in vitro* to assess its prebiotic potential. The studied acacia gum was from two different species; *Acacia senegal* and *A. seyal*. The experiment employed an *in vitro* batch culture using colon model to mimic the distal part of human large intestine. This batch culture system allows rapid response in monitoring performance of acacia gum fermented by beneficial colonic microbes such as *bifidobacteria* and *lactobacilli*. Inoculation of gut microbiota from human faeces into the model start the fermentation system, continuously administered with automated pH-controller maintained at 37°C. Samples taken at 0, 6, 12, and 24 hours of incubation were brought for short chain fatty acids (SCFA) analysis using HPLC and bacterial enumeration via fluorescent *in situ* hybridization (FISH) methods. Experiment were repeated four times with fructo-oligosaccharides (FOS) as control. Results showed that acacia gum significantly promoting *Bifidobacteria* proliferation at the end of fermentation to the same extend benefit of that in FOS while significantly inhibit the growth of pathogenic *Clostridium histolyticum* group which can be an important target group commonly associate with gut dysbiosis. The metabolites analysed for acetate, propionate, and butyrate showed a similar trend as FOS ( $p > 0.05$ ). Further evaluation of AG was observed in the independent enrichment culture study of acacia gum. Five bacteria strains were isolated ought to be gum-fermenting bacteria of which based on 16s RNA sequence result matched to *Escherichia fergusonii*. Here, isolated *Escherichia fergusonii* shows butyrate producing activity while fermenting AG. This effects may have related to the

lowering pH regards to organic acid production. Finally, further investigation to evaluate the isolates and probiotic bacteria associated in a co-culture environment. The symbiotic combination led to increase acetate and overall SCFA production. This effect was more apparent than with the isolates, or probiotic alone treatments. In monoculture of probiotic, it showed that *Bifidobacteria longum* alone does not thrive in acacia gum medium whereas co-culture of probiotics and isolated strain showed otherwise. In conclusion, the potential of AG as a prebiotic has been explored. Furthermore, a conceptual relationship has been developed during the co-culture fermentation. These products could show great potential against gut dysbiosis, as acacia gum could promote native gut-probiotics, through the modulation of microbial population and SCFA production especially butyrate, therefore lead to improve overall human health.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**POTENSI GAM AKASIA SEBAGAI RAMUAN PREBIOTIK MELALUI  
PENILAIAN *In Vitro* MIKROBIOTA TINJA MANUSIA.**

Oleh

**MUHAMAD HANIF BIN RAWI**

Mei 2020

**Pengerusi : Shahrul Razid Sarbini, PhD**  
**Fakulti : Sains Pertanian dan Makanan, Kampus Bintulu**

Kini masyarakat boleh mendapatkan gam akasia yang diperakui ramai sebagai makanan berfungsi untuk kesihatan menyeluruh termasuk usus manusia. Produk semula jadi ini adalah rembesan gam pokok Akasia dan merupakan polisakarida bercabang yang terdiri daripada biopolimer kompleks monosakarida, arabinosa dan galaktosa. Sejak dahulu lagi, gam akasia telah digunakan secara meluas di negara Afrika dan di timur tengah sebagai ubat tradisional. Kini, penggunaannya adalah berkaitan dengan pemakanan untuk manfaat kesihatan. Dari segi potensinya sebagai prebiotik, ia khusus untuk merawat penyakit perut. Penyelidikan ini bertujuan untuk menilai gam akasia komersial ini sebagai sumber prebiotik menerusi kajian model kolon *in vitro*. Gam akasia yang digunakan adalah dari dua spesies iaitu *Acacia senegal* dan *Acacia seyal*. Eksperimen ini mengguna pakai sistem kultur kumpulan *in vitro* menerusi model kolon untuk meniru bahagian usus besar (distal) manusia. Sistem kultur kumpulan ini membolehkan maklum balas yang segera dalam memantau prestasi gam akasia yang ditapai oleh mikrob kolon bermanfaat seperti *bifidobakteria* dan *laktobasili*. Sistem penapaian ini dimulakan dengan inokulasi mikrobiota dari tinja manusia ke dalam model usus. Sistem ini dilengkapi dengan kawalan pH automatik dan dikekalkan pada suhu 37°C. Sampel yang diambil pada tempoh 0, 6, 12, dan 24 jam inkubasi, diproses untuk analisa kandungan asid lemak rantai pendek (SCFA) menggunakan sistem HPLC dan perubahan populasi bakteria melalui penghibridan *in situ* pendafluor (FISH). Eksperimen diulang sebanyak empat kali dan frukto-oligosakarida (FOS) diguna sebagai kawalan. Hasil kajian menunjukkan gam akasia menggalakkan pertumbuhan *Bifidobakteria* dengan ketara seperti dalam FOS, dalam masa yang sama menghalang pertumbuhan patogen *Clostridium histolyticum*, yang mana kumpulan sasaran ini sering dikaitkan dengan punca penyakit berkaitan usus. SCFA (asetat, propionat, dan butirat ) yang dianalisis menunjukkan kadar pengeluaran yang sama seperti FOS ( $p > 0.05$ ). Penilaian gam akasia selanjutnya diperhatikan melalui kajian kultur gam akasia yang berasingan. Menerusi kajian ini, lima bakteria dipencil mempunyai keupayaan untuk menapai

gam akasia. Bakteria tersebut adalah dikenal pasti daripada spesies *Escherichia fergusonii*. Di sini, *Escherichia fergusonii* yang dipencil menunjukkan aktiviti kadar penghasilan butirat yang tinggi semasa proses penapaian gam akasia. Kesan ini boleh dikaitkan dengan penurunan pH hasil pengeluaran asid organik. Akhirnya, siasatan lanjut dibuat untuk menilai *Escherichia fergusonii* dan probiotik dalam kaedah kokultur. Gabungan kokultur menunjukkan peningkatan asetat dan SCFA secara menyeluruh. Kesan tersebut adalah lebih ketara berbanding dengan rawatan *Escherichia fergusonii*, atau probiotik secara berasingan. Berdasarkan monokultur probiotik, *Bifidobacteria longum* secara sendiri tidak berupaya menggunakan gam akasia manakala melalui kokultur probiotik bersama *Escherichia fergusonii* menunjukkan keupayaan yang positif. Kesimpulannya, potensi gam akasia sebagai prebiotik telah diterokai. Tambahan pula, satu konsep hubungan telah ditunjukkan semasa kokultur. Produk ini boleh menunjukkan potensi yang besar terhadap kesihatan usus. Kesimpulannya, gam akasia dapat menggalakkan aktiviti probiotik, melalui modifikasi populasi mikrob dan penghasilan SCFA, sekali gus dapat meningkatkan kesihatan manusia secara menyeluruh.

## ACKNOWLEDGEMENTS

I would like to take this opportunity to express the deepest appreciation to all the people who have involved and accompanied me for the past five years I have spent in Universiti Putra Malaysia Bintulu Sarawak Campus.

First and foremost, special thanks to my supervisor, Assoc. Prof. Dr. Shahrul Razid Sarbini for his immense guidance and generosity in sharing knowledge throughout my research. As the supervisor, he has always encouraged and helped students to develop creative and independent thinking, which I believe are the most important assets not only for the scientific career but also in life. He is such an excellent scientist with distinctives persona and scientific wisdom in research and teaching, as well as positive motivation and passion in life. I am also appreciated his patience, and, more importantly, we admired how much he is caring and being responsible for his students. It would be a lifetime memory for me to grow up as a young scientist under his supervision and graduate as his first PhD. student.

I would like to thank my committee members; Assoc. Prof. Dr. Patricia King Jie Hung and Prof. Dr. Amin Ismail, for their in-prompt feedback and I always enjoyed in their enthusiastic discussion in the meeting and appreciated their valuable comments for my research projects. I would like to acknowledge all the collaborators I have been worked with; Prof. Dr. Aminah Abdullah for her kindness to provide the samples in the studies and Prof. Dr. Abdul Salam Babji for granted me with HPLC system for SCFA analysis.

I would also like to express my deepest appreciation to my colleagues, Siti Maisarah Mohd Nahsri, Tan Hui Yan, Chai Lee Ling, Kathleen Michelle, Nur Bazilah Burhan, Dayang Marshitah Abang Bajury, Siti Aisyah Zaman, Siti Aminah Abdullah, Mayrilyn Solo Thompson, and Morven Mundi, who are always readily to offer their help; to collect samples, extract SCFA, countless number of PCR reactions and gel electrophoresis, especially during those weekends.

Thanks to my friends, Nathaniel Mikal, Ali Maru, and Hamad Mohamad Salah Hamad Ahallil, my heartfelt thanks to them for always motivate me. My thanks also go to those who supported me in any way during my research, to all students, professors and staffs in Crop Science department, as well as Noor Hakim Ahmad as the newly appointed coordinator for Research and Postgraduate Unit of whom is a brother that someone would ever wish for.

Besides, I would like to thank our laboratory assistants, Miss Georgina Sylvia Niwin and Miss Siti Aziah Kushairi for their assistance and cooperation throughout my research. Without the help from all these people, I would not have been able to complete the dissertation.



Last but not least, I would like to thank my parents, Rawi Mohd Ali and, my step mother, Rodzilah Ibrahim for their selfless love and endless support in my PhD journey. To all my siblings; Mohd Faiz Rawi, Mohd Asyraf Rawi, Najwa Rawi, Mohd Fatihi Ikram Rawi and Muhammad Herman Rawi, for their warmest welcome bestowed upon me during my sleepover to every getaway break whenever I need. Indeed, I have the best support group beyond my families.

**Dedicated to my dearest mother,**

This dissertation is in memory of my mother, who inspired me to come to Sarawak and pursue PhD. She would always cheer me up during those tough days. We shared so many laughs and tears together until our family lost her after my second year. I devoted her fortitude of seven years of difficult battle against kidney terminal disease. Her sufferings were very great at times of which would not worthy to be compared with mine. It would take the whole universe to rattle her stoicism, it was edifying for me being around her. Although today she would not be able to read my dissertation, I know she would be very proud of her son making such a big accomplishment!

I certify that a Thesis Examination Committee has met on 5 May 2020 to conduct the final examination of Muhamad Hanif bin Rawi on his thesis entitled "Potential of Acacia Gum as a Prebiotic Ingredient Through *In Vitro* Studies using Human Faecal Microbiota" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

**Nor Mariah bt Adam, PhD**

Professor Ir.  
Faculty of Agriculture and Food Science (Bintulu Campus)  
Universiti Putra Malaysia  
(Chairman)

**Anjas Asmara @ Ab. Hadi bin Samsudin, PhD**

Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Internal Examiner)

**Osumanu Haruna Ahmed, PhD**

Professor  
Faculty of Agriculture and Food Science (Bintulu Campus)  
Universiti Putra Malaysia  
(Internal Examiner)

**Anuraga Jayanegara, PhD**

Associate Professor  
Faculty of Animal Science  
Bogor Agricultural University  
Indonesia  
(External Examiner)

---

**ZURIATI AHMAD ZUKARNAIN, PhD**

Professor Ts. and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 02 March 2021

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Shahrul Razid Sarbini, PhD**

Associate Professor  
Faculty of Agriculture and Food Sciences  
Universiti Putra Malaysia, Bintulu Campus  
(Chairman)

**Patricia King Jie Hung, PhD**

Associate Professor  
Faculty of Agriculture and Food Sciences  
Universiti Putra Malaysia, Bintulu Campus  
(Member)

**Amin Ismail, PhD**

Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 11 March 2021

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: 29 MARCH 2021

Name and Matric No.: Muhamad Hanif Bin Rawi (GS44869)

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature : \_\_\_\_\_  
Name of Chairman of  
Supervisory : Assoc. Prof. Dr. Shahrul Razid Sarbini  
Committee

Signature : \_\_\_\_\_  
Name of Member of  
Supervisory : Assoc. Prof. Dr. Patricia King Jie Hung  
Committee

Signature : \_\_\_\_\_  
Name of Member of  
Supervisory : Prof. Dr. Amin Ismail  
Committee

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vii
<b>DECLARATION</b>	ix
<b>LIST OF TABLES</b>	xiv
<b>LIST OF FIGURES</b>	xv
<b>LIST OF ABBREVIATIONS</b>	xvii
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
1.1 Study Background	1
1.2 Problem Statement	2
1.3 Significance of Study	3
1.4 Aims and Objectives	4
1.5 Scope and Limitations	5
1.6 Thesis Layout	6
<b>2 LITERATURE REVIEW</b>	<b>7</b>
2.1 Probiotic and Prebiotic	7
2.2 Acacia Gum	8
2.2.1 Production of Acacia Gum	9
2.2.2 Application of Acacia Gum in Various Industries	10
2.2.3 Composition of Acacia Gum	11
2.2.4 Health Benefit of Acacia Gum	13
2.3 Role of Gut Microbes in Overall Human Health	13
2.3.1 Physiological Significance of Colon Fermentation	17
2.3.2 Other End-products	19
2.4 Acacia Gum as a Prebiotic	20
2.5 Methods of <i>in vitro</i> Digestion	22
2.6 Methods of <i>in vitro</i> Fermentation	23
2.6.1 Batch System	24
2.6.2 Dynamic System	24
2.7 Substrates-species Specific Fermentation	25
2.7.1 Amylolytic Bacteria	25
2.7.2 Butyrogenic Bacteria	26
2.7.3 Methanogenic Bacteria	28
2.8 Factors Affecting the Efficacy of <i>in vitro</i> Fermentation	29
2.9 Prebiotic evaluation Based on Short Chain Fatty Acid Production and Gut Bacterial Enumeration	32
2.9.1 Fluorescence <i>in situ</i> Hybridization	32
2.9.2 High Performance Liquid Chromatography	33

2.10	Anaerobic Enrichment Culture	34
<b>3</b>	<b>MATERIALS AND METHODS</b>	<b>36</b>
3.1	Flowchart of Study	36
3.2	Substrates and Chemicals	36
3.3	Proximate Nutritional Analysis of Acacia Gum	37
3.3.1	Crude Protein Content	37
3.3.2	Fat Content	37
3.3.3	Ash Content	38
3.3.4	Moisture Content	38
3.3.5	Total Carbohydrate Content	38
3.4	<i>In vitro</i> Digestion	39
3.4.1	Determination of Digestibility	40
3.4.2	Determination of Reducing Sugar Content	40
3.4.3	Determination of Total Sugar Content	40
3.4.4	Determination of Carbohydrate Size	40
3.5	<i>In vitro</i> Fermentation	41
3.5.1	Faecal Sample Preparation	41
3.5.2	Colon Model Fermentation	41
3.6	Prebiotic Evaluation	42
3.6.1	Microbiological Analysis	42
3.6.2	Organic Acids Analysis	44
3.7	Anaerobic Enrichment Culture	44
3.7.1	Chemical and Molecular Ecological Analyses	45
3.7.2	Polymerase Chain Reaction	45
3.8	Isolation of Acacia Gum Fermenting Bacteria	46
3.9	Gram Staining	46
3.10	Probiotic Co-Culture Fermentation	46
3.11	Statistical Analysis	47
<b>4</b>	<b>RESULTS AND DISCUSSIONS</b>	<b>48</b>
4.1	Proximate Analysis for Acacia Gum	48
4.2	Carbohydrate Assay	49
4.2.1	Total Carbohydrate Content	50
4.2.2	Total Reducing sugar Content	51
4.2.3	Degree of Polymerisation of Acacia Gum	53
4.3	Prebiotic Evaluation	53
4.3.1	Colonic Bacterial Enumeration	54
4.3.2	Short Chain Fatty Acids Production	58
4.4	Acacia Gum Fermenting Bacteria	63
4.5	Monoculture Fermentation of Acacia Gum by Isolated Strains	67
4.6	Homology Analysis of Isolates	69
4.7	Co-Culture Fermentation of Acacia Gum with Probiotic	72
4.8	Cross Feeding of Gut Microbiota	75

<b>5</b>	<b>CONCLUSION</b>	<b>77</b>
<b>REFERENCES</b>		<b>79</b>
<b>APPENDICES</b>		<b>120</b>
<b>BIODATA OF STUDENT</b>		<b>127</b>
<b>LIST OF PUBLICATIONS</b>		<b>128</b>





## LIST OF TABLES

Table		Page
2.1	Fibre substrates of butyrogenic bacteria in human colon.	27
3.1	Treatments for acacia gum and controls used this study.	37
3.2	16S rRNA oligonucleotide probes used in present study.	43
4.1	Proximate analysis of AG1 and AG2 for its content (%) of total crude protein, fat, moisture, ash and carbohydrate.	48
4.2	Mean values of bacterial population ( $\log_{10}$ cells/mL batch culture fluid) in the colon model at 0, 6, 12 and 24 h fermentation inoculated with faecal microbiota. (n = 4)	55
4.3	Total SCFA concentration at respective fermentation time and acetate:propionate ratio.	59
4.4	List of the sequences that showed similarity with the <i>Enterobacteriaceae</i> group.	71

## LIST OF FIGURES

Figure		Page
2.1	Different colour and size of acacia gum.	10
2.2	The propose structure of complex acacia gum polysaccharide. A = arabinosyl; filled circles = 3-linked galactose (galactose attached); open circle 6-linked galactose (galactose or glucuronic acid attached or end group); R1 = rhamnose-glucuronic acid; R2 = galactose-3arabinose; R3 = arabinose-3arabinose-3arabinose.	12
2.3	Pathways that are recognised for short chain fatty acids formation by representative bacterial genera and species from the human colon bacterial cross-feeding. Substrate is non-digestable carbohydrates that are converted to pyruvate before continuing to different pathways. As shown in dotted arrow, some bacteria reverse utilise lactate to produce propionate ( <i>Veilonella</i> spp.) and butyrate ( <i>Anaerostipes</i> spp and <i>E. hallii</i> ) via CoA transferase pathway (Besten <i>et al.</i> , 2013). The bacterial species listed are not exhaustive, only as reviewed from studies in culture isolates of dominant species and metagenomic analyses. PEP, phosphoenolpyruvate; DHAP, dihydroxyacetonephospate.	18
2.4	The illustrative representation of lactic acid production from amylaceous starch based materials in the human colon.	26
2.5	Cross feeding mechanism of the human colonic microbiota.	28
2.6	Illustrative diagram of methanogenesis and production of methane gas by methanogens in human gastrointestinal tract.	29
3.1	Overall flow chart of the study.	36
4.1	Total carbohydrate content (mg/mL) of fructo-oligosaccharides and acacia gum.	51
4.2	Total reducing sugar content (mg/mL) of fructo-oligosaccharides and acacia gum.	52
4.3	Concentration of short chain fatty acids against time.	60
4.4	Concentration of short chain fatty acids against time.	61

4.5	Concentration of short chain fatty acids against time.	62
4.6	Concentration of short chain fatty acids against time.	63
4.7	Short chain fatty acids concentration during enrichment culture on acacia gum. (■, acetate; ▲, propionate; ●, butyrate). Transfer of culture to a fresh medium was performed 48 h and 96 h.	64
4.8	Average molar proportion of acetate, propionate and butyrate in every stage (48 h). n=8	65
4.9	Trend analysis of the concentration of short chain fatty acids of the monoculture by different isolates strains. (■, acetate; ▲, propionate; ●, butyrate).	67
4.10	Trend analysis of the concentration of short chain fatty acids for (a) monoculture of <i>Bifidobacteria longum</i> ; (b) monoculture of <i>Lactobacillus casei</i> Shirota; (c) co-culture of <i>Bifidobacteria longum</i> with Strain 3: <i>Escherichia fergusonii</i> ; (d) co-culture of <i>Lactobacillus casei</i> Shirota with Strain 3: <i>Escherichia fergusonii</i> ; (e) co-culture of <i>Bifidobacteria longum</i> with Strain 4: <i>Escherichia fergusonii</i> ; (f) co-culture of <i>Lactobacillus casei</i> Shirota with Strain 4: <i>Escherichia fergusonii</i> . (■, acetate; ▲, propionate; ●, butyrate).	73

## LIST OF ABBREVIATIONS

SCFA	Short Chain Fatty Acid
FOS	Fructo-Oligosaccharides
MW	Molecular Weight
HPLC	High Performance Liquid Chromatography
FISH	Fluorescence <i>in situ</i> Hybridization
DP	Degree of Polymerisation
LAB	Lactic Acid Bacteria
BCFA	Branched Chain Fatty Acid



## CHAPTER 1

### INTRODUCTION

#### 1.1 Study Background

Acacia gum (AG) is a soluble secretion found on the trunks and branches of *Acacia senegal* and *Acacia seyal* trees (Mariod *et al.*, 2018). The exudate secreted as a result of subsequent injuries onto the bark of the trees. Acacia gum is among the hydrocolloids that has been exploited in various food and non-food applications. Having exceptionally low viscosity and strong adhesive made acacia gum as the best source of soluble dietary fibre because it can boost fibre levels in foods or beverages without modifying the final viscosity of food products (Samuel *et al.*, 2018).

Most consumers may ever set their hands onto soft drinks containing acacia gum without even knowing it. As the matter of fact, more than three quarter of industrial applications of this hydrocolloid are for emulsification, encapsulation, coating, and confectionaries (Roberfroid, *et al.*, 2010). It is made up of 95% long chain complex polysaccharides, mainly from highly branched galactan polymers, side chains of galactose and/or arabinose; and rhamnose or glucuronic acid as termination residues (Nakov, 2016). With this in mind, looking at its carbohydrate-based structure and functions, acacia gum is an excellent prospect to be commercialised as prebiotic.

Prebiotic is substrate that selectively nourished the host microorganisms in returns to bring health benefits to the host. Some strict criterions needed to be met before any substrates are known as prebiotic. Numbers of test have to be performed where the propose substrate should be; resistant to digestive activity from the upper gastrointestinal gut, fermentable by colonic microbiota and selectively promoting the growth and/or activity of health promoting bacteria commonly *Bifidobacterium* and *Lactobacillus* (Roberfroid, *et al.*, 2010). These bacteria are known to ferment established prebiotic compound i.e. FOS, GOS, and inulin to produce beneficial metabolites known as short chain fatty acids (SCFA) mainly acetate, propionate and butyrate (Pourabedin, Guan, and Zhao, 2015). These SCFA may exert direct effect to host health by lowering luminal pH as most enteric pathogens does not grow well in low pH. Butyrate especially contribute towards the energy requirement by epithelial cells, thus, improving intestinal motility.

These promoting bacteria also been exploited for its good property as probiotic. Probiotic as defined by FDA and WHO (2002) is the live microorganisms which when administered in adequate amounts confer a health benefit to the host. For example, probiotic known to act via a variety of means, including modulation of immune function, association with native gut microbiota, and interacting with the host epithelial cell like gut barrier integrity and enzyme formation. *Lactobacillus*

spp. and *Bifidobacterium* spp. are the most common probiotic bacteria that have been utilized by the food industry (Min *et al.*, 2018), but others such as *Saccharomyces cerevisiae (boulardii)*, *Enterococcus*, *Bacillus*, and *Escherichia* are also applied. They are mainly saccharolytic metabolism that are capable of carbohydrate utilisation. Therefore, study in the importance of the species specific among particular groups might uncover the health-promoting conditions (Calame *et al.*, 2008).

Years ago, health wellness awareness arose from weight management and superficial disease-relating concern, but now it started to move to digestive health. People nowadays are more aware on probiotic products in their daily food intake. Consumers are more than aware that food can influence health, and there is a clear relationship between the food we consume and how it affects our body.

## 1.2 Problem Statement

Colonic bacteria metabolised range of carbohydrate hydrolysing enzymes in the production of hydrogen, methane, carbon dioxide, SCFA (mainly acetate, propionate and butyrate), and lactate. This gut flora obtained their energy through fermentation of food supplied from the upper gut (Cherbut, 2003).

Several studies have claimed that dysbiosis, which is the imbalance and unhealthy condition of colon microbiota diversity lead to disorders such as obesity (Turnbaugh *et al.*, 2009), metabolic complications, immunity dysregulation (Kamal *et al.*, 2018), changes in energy and hormone regulation and even irregular inflammatory mechanism (Ali *et al.*, 2020). Dysbiosis in the gut can reflect changes in host systems, typically in metabolic and immune process. Thus, causing diseases such as inflammatory bowel disease (IBD) (Opstelten *et al.*, 2016), psoriatic arthritis (Scher *et al.*, 2015), type 1 and type 2 diabetes (de Goffau *et al.*, 2013; Lambeth *et al.*, 2015), atopic eczema (Wang *et al.*, 2008), coeliac disease (Manichanh *et al.*, 2006; Schippa *et al.*, 2010) and arterial stiffness (Menni *et al.*, 2018).

As to counteract the issue studies showed that a healthy diet, lifestyle and habit is the key toward a good human gut. Consequently, a healthy diet is another direct key towards healthy gut diversity. Apart from having molecule rich in sugar complex, prebiotic have to be fermentable by means to escape the digestion and absorption from the upper gastrointestinal tract before progress into the large colon. The knowledge of these properties is important to evaluate efficiency of different acacia gum species and unique composition of polysaccharides as prebiotic. Over the last decades, investigation on AG for its health promoting properties were studied and shown positive effect like regulation of autoimmune disease (Kamal *et al.*, 2018), reduce duodenal inflammation in mice (Ali *et al.*, 2020), anti-ulcerogenic on gastric mucosal injury in rats (Taha *et al.*, 2020), methane gas mitigation in ruminant (Adejero *et al.*, 2019), and also topical treatment of skin lesions of kwashiorkor children (Ali, 2018).

Studies made on acacia gum showed that AG plays the role of prebiotic in the human colon due to its ability to resist human upper gastrointestinal digestion (Phillips, 1998; Kishimoto, 2006). Acacia gum is readily can be used in its natural form. Historically, acacia gum has been commonly consumed by African citizen to treat stomach relating illness. It is also claimed that AG helps in enhancing the absorption of sodium and water in the small intestine into the bloodstream which prevent diarrhoea and dehydration condition (Codipillu and Wapnir, 2004; Rehman *et al.*, 2003; Wapnir *et al.*, 1997). The fermentation of AG in the colon contributing in the production of beneficial metabolites, short chain fatty acids (SCFA) such as acetate, butyrate and propionate in major portion (Kaddam and Kaddam, 2020). The strict demand of healthy food among modern people has made acacia gum that contains large amount of dietary fibre to fulfil the requirement of healthy food.

Although prebiotic effects of AG were previously studied *in vitro* and *in vivo* including humans, but there is an inadequate evidence as some of the researches were not consistent which makes it difficult to scrutinize. Several efforts made as to incorporating AG with probiotic and/or yogurt products (Ghafarloo *et al.*, 2019; Chundakkattumalayil *et al.*, 2019; Talin *et al.*, 2018) that also studied to have promising outcomes. While there is study reported that AG fibre showed bifidogenic effects, which indicating that bifidobacteria can utilised the complex structure of AG (Cherbut *et al.*, 2003).

On the other hand, as reported by Rossi *et al.* (2005), most *Bifidobacterium* species are not capable to utilise longer chain inulin but prefer shorter chain FOS substrates. Furthermore, different observation in mixed faecal cultures as other faecal bacterial species first degrade inulin to oligomers providing preferable chain length for bifidobacterial (Rossi *et al.*, 2005). The capacity to metabolise carbohydrates relies on the availability of functional transport system and intracellular metabolic pathways of gut microbiota (Buntin *et al.*, 2017). In *Lactobacillus acidophilus* and *Bifidobacterium breve*, ABC transporters have been observed to be involved in FOS uptake (Ventura *et al.*, 2007). Therefore, this study addressing the possible pathways of AG fermentation by colonic bacteria. It was done to investigate the uncertain findings on AG fermentation by the group of *Bifidobacteria* spp. describing its capabilities.

### 1.3 Significance of Study

Today, acacia gum is sought after for its properties as thickener, stabiliser, emulsifier in food products and used in pharmaceutical products as a demulcent (Duke, 2018). The applications of acacia gum tracked to be found since Ancient Egyptian time. Other than for traditional medicine (pain-reliever base) and in everyday applications, Egyptians known for their embalming procedure where they perfected the mummification process using acacia gum as adhesive (Scott, *et al.*, 2004; Scott *et al.*, 2009).

Hardened acacia gum on branch is fragile, fragrance-free and generally tasteless crystals. There are only two species (out of thousands) of acacia gum that are deemed for commercial uses, *Acacia senegal* and *Acacia seyal* (Abuarra *et al.*, 2014). The former is considered the best quality, and made majority for global trade due to low tannin traces and superior emulsifier (Egadu *et al.*, 2007). While the latter produces lower grade gum, *A. seyal* gum with lower market price is being discovered with further uses and thus boosted its value. The uses of *A. seyal* in 'health beverages' as a fermentable has given it a value in its own right (William and Phillips, 2009).

In this study, both species of acacia gum were tested and evaluate for its prebiotic potential. Thus, assessment made should not limit within the perspective of industrial functions only, instead, under a fair consideration, either species could have the same potential and be a good prebiotic candidate. Although many studies have shown some difference between composition of *A. senegal* and *A. seyal* (Vanloot *et al.*, 2012), but not many of those data reporting on the prebiotic aspect of the gums (Musa *et al.*, 2017). The finding from this research can be benefit to further understand current food products, since our usual meal may have incorporated with acacia gum (E414), added as stabilizer ingredients such as in candies, chewing gums, ice cream salad-dressing, bread, cereals and cola drinks.

#### **1.4 Aims and Objectives**

This research aims to investigate the impact of commercially available acacia gum as a prebiotic source through *in vitro* colon model study. Therefore, in this study work *in vitro* approaches have been used to investigate digestibility of AG, changes in gut microbiota and SCFA production. The experiment includes *in vitro* digestion and fermentation of acacia gum in a custom build colon model which mimic the distal part of human colon. Moreover, investigation on the putative primary degrader of acacia gum based on enrichment culture was used to generate more input on the relationship of different microbial species in gut.

The main objectives of this study are:

- a. To investigate the nutritional content and digestibility of acacia gum.
- b. To determine the physiological significance of prebiotic effects of acacia gum on modulating gut microbiota and its fermentation products, short chain fatty acids in relation with the gut microbiota.
- c. To isolate and identify the gum fermenting bacteria through employment of enrichment culture fermentation.
- d. To demonstrate the conceptual relationship in co-culture fermentation of acacia gum between probiotic strain with isolated gum fermenting bacteria.



## 1.5 Scope and Limitation

At least two decades of gut research has passed, lots of the unknown before to the scientists, now becoming clearer. The realisation of the relationship is far more complex and becoming more ambiguous; what does it really mean by the gut environment of having mutualists, commensals and pathogens? Dominant microorganisms inhabit the colon are generally from the phyla *Firmicutes* and *Bacteroidetes* (Walker *et al.*, 2011). Together with the host immune system, gut microbiota contributes significantly as barrier and prevention of invasion of pathogenic bacteria in the gastrointestinal tract. We have known that the microbiota within the colon expressed no straightforward interactions with host metabolic pathway (Tremaroli and Backhead, 2012). At the most basic notion, the relationship is highly mutualistic where the host provides shelter together with the food supply, while the bacteria in return assist in digestion of complex food components apart from supplying vitamin B and K to the host. But among those advance research, many were confirmed the link between diet we consumed and changes on gut microbiota always occurred upon substrate intervention (Ríos-Covián *et al.*, 2016; David *et al.*, 2014).

The fermentation of AG resulted in bacterial proliferation (May *et al.*, 1994). The increase population of colonic bacteria thus promotes the production of beneficial metabolites, i.e. the short chain fatty acids (SCFA) that play an important role in many physiological effects. Based on the study, observation was made based on selective agar medium. During those years the best possible methods to evaluate was using culture dependent techniques. Thus, having selective media that only targeting beneficial bacteria was not sufficient to actually verify the fermentability of AG by colonic bacteria. In addition, it is reported from the study of fermentation in cecum of rats and human that AG can be break down in the colon (Ross *et al.*, 1984; Walter *et al.*, 1988; Phillips, 1998; Phillips, 1998). While this study was evaluated in vivo, there is no direct proof to claimed the findings since the parameters was only evaluate the post-administration of candidate after being fed with AG during the study. Whereas, animal study of Kishimoto *et al.*, in 2006 has found that the predominant microbes responsible for AG fermentation in and contributes in production of propionate is *Prevotella ruminicola*-like bacterium. While the research was investigating pooled cecal inoculum of pig shown isolated bacteria was the predominant species of the animal. In contrast, sstudied using human and porcine faecal inoculum with 2% of acacia gum managed to isolate *Bifidobacterium longum* and *Bacteroides ovatus*, *Bacteroides oris*, *Bacteroides buccae* and *Prevotella ruminicola*-like bacterium (Wyatt *et al.*, 1986). Although the research concluded those results in human faecal, it turns out that presumptive measures led to their conclusions that based only on culture plating techniques but not molecular based DNA sequencing, which only emerged in 1970s that could possibly not available at that time in their lab. Therefore, the traditional detection of many uncultured species assigned to this genus may reflect current taxonomic limitations rather than a biological signal. FISH technique has limit detection  $1 \times 10^6$ , enabling bacteria to be accurately enumerated above this point.

The chemical compositions of AG slightly varied based on origin, sources, climate, season, tree age and species (William and Phillips, 2009). High carbohydrate content within this complex carbohydrate serves as the basic principle to evaluate the prebiotic potential of acacia gum. Species difference of AG were taken into account to see how far will the species affecting the fermentation performance in the colon. Thus, the evaluation of SCFA production and bacteria composition changes due to input of AG as substrate are therefore the foundation for the benefits of human colon health. The limitation in simulating the interaction between host and gut microbes in a fermentation system may as well be one of the criteria condition of a fermentation system. Hence, through this work, it is possible to enhance the understanding for prebiotic applications of acacia gum in various industries particularly nutraceuticals. While FOS had been profoundly studied as prebiotic to regulate and selectively stimulate bacteria population in the colon (Tuohy *et al.*, 2001; Palframan *et al.*, 2002; Hidalgo *et al.*, 2012; Cueva *et al.*, 2013).

Despite that, this current study is the first to evaluate the prebiotic potential in one continuous flow of from the *in vitro* digestion to the fermentation using the colon model. Apart from adding new insights towards the old knowledge about the prebiotic potential of AG, this study also opens up the new perspectives on substrate-species relationship in the metabolism of food products. In tandem with the advance of technologies and knowledges, the study aims to look at the cross feeding feature in gut microbiota to better understand how such prebiotic products could leverage research in the applications or real-world situation.

## **1.6 Thesis Layout**

This thesis comprises of five chapters, Chapter 1 will introduce the main subjects and a brief background on the topic of the research. Followed by Literature Review in Chapter 2 is the previous research and findings on the subject matter. Also together with the types of available methods that usually was used and can be implement for this type of study. More detail and comprehensive elaborations on the Materials and Methodologies is featured in Chapter 3. Next, in chapter 4 is the parts for Results and Discussions that subdivided into several parts according to it respective objectives. Lastly, the in chapter 5, this research concluded by highlighting the outcomes and whether the objectives were fulfilled.

## REFERENCES

- Aachary, A. A. and Prapulla, S. G. (2009). Value addition to spent osmotic sugar solution (SOS) by enzymatic conversion to fructooligosaccharides (FOS), a low calorie prebiotic. *Innovative Food Science and Emerging Technologies*, 10(2), 284–288. doi:10.1016/j.ifset.2008.11.013
- Abdel Nour, H. O. (1999). Gum Arabic in Sudan: production and socio-economic aspects. In *International Expert Meeting on Medicinal, Culinary and Aromatic Plants in the Near East, Cairo (Egypt), 19-21 May 1997*. Retrieved from fao.org/docrep/x5402e/x5402e12.htm. (9 December 2017).
- Abdin, A. A. and Saeid, E. M. (2008). An experimental study on ulcerative colitis as a potential target for probiotic therapy by *Lactobacillus acidophilus* with or without “olsalazine”. *Journal of Crohn's and Colitis*, 2(4), 296-303.
- Abell, G. C., Conlon, M. A., and McOrist, A. L. (2006). Methanogenic archaea in adult human faecal samples are inversely related to butyrate concentration. *Microbial Ecology in Health and Disease*, 18(3-4), 154-160.
- Abuarra, A., Hashim, R., Bauk, S., Kandaiya, S., and Tousi, E. T. (2014). Fabrication and characterization of gum arabic bonded *Rhizophora* spp. particleboards. *Materials and Design*, 60, 108–115.
- Adejoro, F. A., Hassen, A., and Thantsha, M. S. (2019). Characterization of starch and gum arabic-maltodextrin microparticles encapsulating acacia tannin extract and evaluation of their potential use in ruminant nutrition. *Asian-Australasian Journal of Animal Sciences*, 32(7), 977-987. doi:10.5713/ajas.18.0632
- Agrawal, N., Minj, D. K., and Rani, K. (2015). Estimation of total carbohydrate present in dry fruits. *IOSR Journal of Environmental Science, Toxicology and Food Technology (IOSR-JESTFT)*, 1(6), 24–27.
- Assaf, S., Phillips, G., and Williams, P. (2005). Studies on acacia exudate gums. Part I: The molecular weight of gum exudate. *Food Hydrocolloids*, 19(4), 647-660. doi:10.1016/j.foodhyd.2004.09.002
- Al-Assaf, S., Phillips, G. O., Aoki, H., and Sasaki, Y. (2007). Characterization and properties of Acacia senegal (L.) Willd. var. senegal with enhanced properties (Acacia (sen) SUPER GUM™): Part 1—Controlled maturation of Acacia senegal var. senegal to increase viscoelasticity, produce a hydrogel form and convert a poor into a good emulsifier. *Food Hydrocolloids*, 21(3), 319-328. doi:10.1016/j.foodhyd.2006.04.011

- Ali, B. H., Za'Abi, M. A., Suleimani, Y. A., Manoj, P., Ali, H., Ribeiro, D. A., and Nemmar, A. (2020). Gum arabic reduces inflammation, oxidative, and nitrosative stress in the gastrointestinal tract of mice with chronic kidney disease. *Naunyn-Schmiedeberg's Archives of Pharmacology*. doi:10.1007/s00210-020-01844-y
- Ali, I. A. (2018). Use of Acacia Gum in the Treatment of Skin Lesions of Two Children With Kwashiorkor. *Gum Arabic*, 221-228. doi:10.1016/b978-0-12-812002-6.00018-x
- Altuntas, E. and Erkol, M. (2009). The effect of moisture content on colour characteristics of walnuts. *International Journal of Food Engineering*, 5(2). doi: 10.2202/1556-3758.1577.
- Amaretti, A., Bernardi, T., Tamburini, E., Zanoni, S., Lomma, M., Matteuzzi, D. and Rossi, M. (2007). Kinetics and metabolism of *Bifidobacterium adolescentis* MB 239 growing on glucose, galactose, lactose, and galactooligosaccharides. *Applied and Environmental Microbiology*, 73(11), 3637-3644.
- Andersen, J. M., Barrangou, R., Hachem, M. A., Lahtinen, S. J., Goh, Y. J., Svensson, B., and Klaenhammer, T. R. (2013). Transcriptional analysis of oligosaccharide utilization by *Bifidobacterium lactis* BI-04. *BMC genomics*, 14(1), 312. doi: 10.1186/1471-2164-14-312
- Anderson, D. M. W. and Eastwood, M. A. (1989). The safety of gum arabic as a food additive and its Energy Value as an ingredient: a brief review. *Journal of Human Nutrition and Dietetics*, 2(3), 137-144. doi:10.1111/j.1365-277x.1989.tb00017.x
- Anderson, D. M. W. and McDougall, F. J. (1987). Degradative studies of gum arabic (*Acacia senegal* (L.) Willd.) with special reference to the fate of the amino acids present. doi: 10.1080/02652038709373633
- Anderson, D. M. W. and Stoddart, J. F. (1966a). Studies on uronic acid materials: Part XV. The use of molecular-sieve chromatography in studies on *Acacia senegal* gum (Gum Arabic). *Carbohydrate Research*, 2(2), 104-114.
- Anderson, D. M. W., Hirst, E., and Stoddart, J. F. (1966b). Studies on uronic acid materials. Part XVII. Some structural features of *Acacia senegal* gum (gum arabic). *Journal of The Chemical Society C: Organic*, 1959. doi:10.1039/j39660001959
- Anderson, D. M. W., Hirst, E., Rahman S., and Stainsby, G. (1967). Studies, on uronic acid materials: Part XVIII. Light-scattering studies on some molecular-weight fractions from *Acacia senegal* gum. *Carbohydrate Research* 3(3), 308-317. doi: 10.1016/S0008-6215(00)82206-5

- Anderson, D. M. W., Millar, J. R. A., and Weiping, W. (1991). Gum Arabic(*Acacia senegal*):Unambiguous identification by<sup>13</sup>C-NMR spectroscopy as an adjunct to the revised JECFA specification, and the application of<sup>13</sup>C-NMR spectra for regulatory/legislative purposes. *Food Additives and Contaminants*, 8(4), 405–421. doi:10.1080/02652039109373991
- Annison, G., Trimble, R. P., and Topping, D. L. (1995). Feeding Australian Acacia gums and gum arabic leads to non-starch polysaccharide accumulation in the cecum of rats. *The Journal of Nutrition*, 125(2), 283-292.
- Aphibanthammakit, C., Nigen, M., Gaucel, S., Sanchez, C., and Chaliar, P. (2018). Surface properties of *Acacia senegal* vs *Acacia seyal* films and impact on specific functionalities. *Food Hydrocolloids*, 82, 519-533.
- Arpaia, N., Campbell, C., Fan, X., Dikiy, S., van der Veeken, J., Deroos, P., ... and Rudensky, A. Y. (2013). Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. *Nature*, 504(7480), 451-455. doi: 10.1038/nature12726
- Arumugam, M., Raes, J., Pelletier, E., Le Paslier, D., Yamada, T., Mende, D. R., ... and Bertalan, M. (2011). Enterotypes of the human gut microbiome. *Nature*, 473(7346), 174-180. doi: 10.1038/nature09944
- Asoodeh, A., Chamani, J., and Lagzian, M. (2010). A novel thermostable, acidophilic  $\alpha$ -amylase from a new thermophilic "*Bacillus sp. Ferdowsicus*" isolated from Ferdows hot mineral spring in Iran: Purification and biochemical characterization. *International Journal of Biological Macromolecules*, 46(3), 289–297.
- Awad, S., Rabah, A., Ali, H., and Mahmoud, T. (2018). Acacia Seyal Gums in Sudan: Ecology and Economic Contribution. *Gum Arabic*, 3-11. doi:10.1016/b978-0-12-812002-6.00001-4
- Awad, S. S., Rabah, A. A., Ali, H. I., and Mahmoud, T. E. (2017). *Acacia seyal* gums in Sudan: a review. *University of Khartoum Engineering Journal*.
- Azad, M., Kalam, A., Sarker, M., and Wan, D. (2018). Immunomodulatory effects of probiotics on cytokine profiles. *BioMed Research International*, 2018.
- Azzaoui K., Hammouti B., Lamhamdi A., Mejdoubi E., and Berrabah M. (2015). The gum Arabic in the southern region of Morocco. *Morocco Journal of Chemistry*, 3(1):99–107
- Babiker, R., Merghani, T. H., Elmusharaf, K., Badi, R. M., Lang, F., and Saeed, A. M. (2012). Effects of gum arabic ingestion on body mass index and body fat percentage in healthy adult females: two-arm randomized,

placebo controlled, double-blind trial. *Nutrition Journal*, 11(1), 111.  
doi:10.1186/1475-2891-11-111

- Balal, M.E., Salih, N.K.E., and Abdel, Magid, T.D. (2014). Ethno-botany of natural forest of Nuba Mountains. *Journal of Forest Production and Industry*. 3(1):13-19: South Kordofan State, Sudan. ISSN: 2325-4513, ISSN 2325-453X
- Balqis, U., Hambal, M., Admi, M., Safika, Meutia, N., Sugito, ... and Abrar, M. (2018). *Escherichia fergusonii* identified in preputial swabs from healthy Aceh cattle by phylogenetic 16S rRNA analysis. *Malaysian Journal of Microbiology*, 14(3), 229-235.
- Bansal, S., Mangal, M., Sharma, S. K., and Gupta, R. K. (2016). Non-dairy based probiotics: A healthy treat for intestine. *Critical Reviews in Food Science and Nutrition*, 56(11), 1856–1867.
- Barbosa, T. M., Serra, C. R., Ragione, R. M. L., Woodward, M. J., and Henriques, A. O. (2005). Screening for bacillus isolates in the broiler gastrointestinal tract. *Applied and Environmental Microbiology*, 71(2), 968–978. doi:10.1128/aem.71.2.968-978.2005
- Barcenilla, A., Pryde, S. E., Martin, J. C., Duncan, S. H., Stewart, C. S., Henderson, C., and Flint, H. J. (2000). Phylogenetic relationships of butyrate-producing bacteria from the human gut. *Applied Environmental Microbiology*, 66(4), 1654-1661.
- Barrangou, R., Altermann, E., Hutkins, R., Cano, R., and Klaenhammer, T. R. (2003). Functional and comparative genomic analyses of an operon involved in fructooligosaccharide utilization by *Lactobacillus acidophilus*. *Proceedings of the National Academy of Sciences*, 100(15), 8957-8962. doi: 10.1073/pnas.1332765100
- Barroso, E., Cueva, C., Peláez, C., Martínez-Cuesta, M. C., and Requena, T. (2015). The computer-controlled multicompartmental dynamic model of the gastrointestinal system SIMGI. *The Impact of Food Bioactives on Health*, 319.
- Beck, J. V. (1971). [8] Enrichment culture and isolation techniques particularly for anaerobic bacteria. In *Methods in enzymology* (Vol. 22, pp. 57-64). Academic Press.
- Belenguer, A., Duncan, S. H., Calder, A. G., Holtrop, G., Louis, P., Lobley, G. E., and Flint, H. J. (2006). Two routes of metabolic cross-feeding between *Bifidobacterium adolescentis* and butyrate-producing anaerobes from the human gut. *Applied Environmental Microbiology*, 72(5), 3593-3599.

- Bereswill, S., Muñoz, M., Fischer, A., Plickert, R., Haag, L. M., Otto, B., ... and Heimesaat, M. M. (2010). Anti-inflammatory effects of resveratrol, curcumin and simvastatin in acute small intestinal inflammation. *PLoS one*, 5(12).
- Bernalier-Donadille, A. (2010). Fermentative metabolism by the human gut microbiota. *Gastroenterologie Clinique et Biologique*, 34, S16–S22.
- Bhanwar, S. and Ganguli, A. (2014).  $\alpha$ -amylase and  $\beta$ -galactosidase production on potato starch waste by *Lactococcus lactis* subsp *lactis* isolated from pickled yam. *Journal of Scientific and Industrial Research (JSIR)*, 324-330.
- Blaut, M. (2002). Relationship of prebiotics and food to intestinal microflora. *European Journal of Nutrition*, 41(0), 1–1. doi:10.1007/s00394-002-1102-7
- Bockmühl, D., Jassoy, C., Nieveler, S., Scholtyssek, R., Wadle, A., and Waldmann-Laue, M. (2007). Prebiotic cosmetics: an alternative to antibacterial products. *International Journal of Cosmetic Science*, 29(1), 63–64. doi:10.1111/j.1467-2494.2007.00355\_2.x
- Boisen, S. and Eggum, B. O. (1991). Critical evaluation of *in vitro* methods for estimating digestibility in simple-stomach animals. *Nutrition Research Reviews*, 4(1), 141–162. doi:10.1079/nrr19910012
- Bolca, S., Van de Wiele, T., and Possemiers, S. (2013). Gut metabolites govern health effects of dietary polyphenols. *Current Opinion in Biotechnology*, 24(2), 220-225.
- Bone, E., Tamm, A., and Hill, M. (1976). The production of urinary phenols by gut bacteria and their possible role in the causation of large bowel cancer. *The American Journal of Clinical Nutrition*, 29(12), 1448-1454.
- Bourriaud, C., Robins, R. J., Martin, L., Kozłowski, F., Tenailleau, E., Cherbut, C., and Michel, C. (2005). Lactate is mainly fermented to butyrate by human intestinal microfloras but inter-individual variation is evident. *Journal of Applied Microbiology*, 99(1), 201-212.
- Brockhaus, M., Dettinger, H. M., Kurz, G., Lehmann, J., and Wallenfels, K. (1979). Participation of HO-2 in the cleavage of  $\beta$ -d-galactosides by the  $\beta$ -d-galactosidase from *E. coli*. *Carbohydrate Research*, 69(1), 264-268.
- Brodkorb, A., Egger, L., Alminger, M., Alvito, P., Assunção, R., Ballance, S., . . . Recio, I. (2019). INFOGEST static *in vitro* simulation of gastrointestinal food digestion. *Nature Protocols*. doi:10.1038/s41596-018-0119-1

- Brown, A. J., Goldsworthy, S. M., Barnes, A. A., Eilert, M. M., Tcheang, L., Daniels, D., ... and Pike, N. B. (2003). The Orphan G protein-coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids. *Journal of Biological Chemistry*, 278(13), 11312-11319.
- Buntin, N., Hongpattarakere, T., Ritari, J., Douillard, F. P., Paulin, L., Boeren, S., ... and de Vos, W. M. (2017). An inducible operon is involved in inulin utilization in *Lactobacillus plantarum* strains, as revealed by comparative proteogenomic and metabolic profiling. *Applied Environmental Microbiology*, 83(2), e02402-16. doi: 10.1128/aem.02402-16
- Calame, W., Thomassen, F., Hull, S., Viebke, C., and Siemensma, A. D. (2011). Evaluation of satiety enhancement, including compensation, by blends of gum arabic. a methodological approach. *Appetite*, 57(2), 358-364.
- Calame, W., Weseler, A. R., Viebke, C., Flynn, C., and Siemensma, A. D. (2008). Gum arabic establishes prebiotic functionality in healthy human volunteers in a dose-dependent manner. *British Journal of Nutrition*, 100(6), 1269-1275. doi: 10.1017/S0007114508981447.
- Cantu-Jungles, T. M., Ruthes, A. C., El-Hindawy, M., Moreno, R. B., Zhang, X., Cordeiro, L. M., Hamaker, B. R., and Iacomini, M. (2018). *In vitro* fermentation of *Cookeina speciosa* glucans stimulates the growth of the butyrogenic *Clostridium* cluster XIVa in a targeted way. *Carbohydrate Polymers*, 183, 219–229.
- Chambers, E. S., Viardot, A., Psichas, A., Morrison, D. J., Murphy, K. G., Zac-Varghese, S. E., ... and Blundell, J. E. (2015). Effects of targeted delivery of propionate to the human colon on appetite regulation, body weight maintenance and adiposity in overweight adults. *Gut*, 64(11), 1744-1754.
- Chaucheyras-Durand, F. and Durand, H. (2010). Probiotics in animal nutrition and health. *Beneficial Microbes*, 1(1), 3–9. doi:10.3920/bm2008.1002
- Chen, W. J. L., Anderson, J. W., and Jennings, D. (1984). Propionate may mediate the hypocholesterolemic effects of certain soluble plant fibers in cholesterol-fed rats. *Proceedings of the Society for Experimental Biology and Medicine*, 175(2), 215-218.
- Cherbut, C. (2003). Motor effects of short-chain fatty acids and lactate in the gastrointestinal tract. *Proceedings of the Nutrition Society*, 62(1), 95–99. doi:10.1079/pns2002213
- Cherbut, C., Ferrier, L., Rozé, C., Anini, Y., Blottière, H., Lecannu, G., and Galmiche, J. P. (1998). Short-chain fatty acids modify colonic motility through nerves and polypeptide YY release in the rat. *American Journal*



of *Physiology-Gastrointestinal and Liver Physiology*, 275(6), G1415-G1422.

- Cherbut, C., Michel, C., Raison, V., Kravtchenko, T., and Severine, M. (2003). Acacia gum is a bifidogenic dietary fibre with high digestive tolerance in healthy humans. *Microbial Ecology in Health and Disease*, 15(1). doi: 10.3402/mehd.v15i1.7977
- Chretien, M., Chikamai, B., Loktari, P. E., Ngichili, J., Loupa, N., Odee, D., and Lesueur, D. (2008). The current situation and prospects for gum arabic in Kenya: a promising sector for pastoralists living in arid lands. *International Forestry Review*, 10(1), 14-22.
- Chundakkattumalayil, H. C., Kumar, S., Narayanan, R., and Raghavan, K. T. (2019). Role of *L. plantarum* KX519413 as Probiotic and Acacia Gum as Prebiotic in Gastrointestinal Tract Strengthening. *Microorganisms*, 7(12), 659. doi:10.3390/microorganisms7120659
- Churms, S. C., Merrifield, E. H., and Stephen, A. M. (1983). Some new aspects of the molecular structure of Acacia senegal gum (gum arabic). *Carbohydrate Research*, 123(2), 267–279. doi:10.1016/0008-6215(83)88483-3
- Codipilly, C. N. and Wapnir, R. A. (2004). Proabsorptive action of gum arabic in isotonic solutions orally administered to rats. II. Effects on solutes under normal and secretory conditions. *Digestive Diseases and Sciences*, 49(9), 1473-1478.
- Cohn, M. and Monod, J. (1951). Purification et propriétés de la  $\beta$ -galactosidase (lactase) d'*Escherichia coli*. *Biochimica et Biophysica Acta*, 7, 153-174.
- Coico, R. (2006). Gram staining. *Current Protocols in Microbiology*, (1), A-3C. doi:10.1002/9780471729259.mca03cs00
- Coles, L., Moughan, P., and Darragh, A. (2005). *In vitro* digestion and fermentation methods, including gas production techniques, as applied to nutritive evaluation of foods in the hindgut of humans and other simple-stomached animals. *Animal Feed Science and Technology*, 123-124, 421–444. doi:10.1016/j.anifeedsci.2005.04.021
- Collado, M. C., Gueimonde, M., Hernandez, M., Sanz, Y., and Salminen, S. (2005). Adhesion of selected *Bifidobacterium* strains to human intestinal mucus and the role of adhesion in enteropathogen exclusion. *Journal of Food Protection*, 68(12), 2672-2678. doi:10.4315/0362-028x-68.12.2672

- Cook, S. and Sellin, J. (1998). Review article: short chain fatty acids in health and disease. *Alimentary Pharmacology and Therapeutics*, 12(6), 499–507.
- Counotte, G. H. and Prins, R. A. (1981). Regulation of lactate metabolism in the rumen. *Veterinary Research Communications*, 5(2), 101-115.
- Counotte, G. H. M., Lankhorst, A., and Prins, R. A. (1983). Role of DL-lactic acid as an intermediate in rumen metabolism of dairy cows. *Journal of Animal Science*, 56(5), 1222-1235.
- Couto, S. R. and Sanromán, M. A. (2006). Application of solid-state fermentation to food industry—a review. *Journal of Food Engineering*, 76(3), 291–302.
- Craeyveld, V. V., Swennen, K., Dornez, E., Wiele, T. V. D., Marzorati, M., Verstraete, W., ... and Courtin, C. M. (2008). Structurally different wheat-derived arabinoxylooligosaccharides have different prebiotic and fermentation properties in rats. *The Journal of Nutrition*, 138(12), 2348–2355. doi:10.3945/jn.108.094367
- Cueva, C., Sánchez-Patán, F., Monagas, M., Walton, G. E., Gibson, G. R., Martín-Álvarez, P. J., ... and Moreno-Arribas, M. V. (2013). *In vitro* fermentation of grape seed flavan-3-ol fractions by human faecal microbiota: changes in microbial groups and phenolic metabolites. *FEMS Microbiology Ecology*, 83(3), 792-805.
- Cummings, J. H. (1995). Short chain fatty acids. In Glenn R Gibson and G. T. Macfarlane (Eds.), *Human Colonic Bacteria: Role in Nutrition, Physiology and Pathology* (pp. 101–130). Boca Raton, Florida: CRC Press Inc.
- Cummings, J. and Macfarlane, G. (1991). The control and consequences of bacterial fermentation in the human colon. *Journal of Applied Bacteriology*, 70(6), 443–459. doi:10.1111/j.1365-2672.1991.tb02739.x
- Cummings, J. H. (1981). Short chain fatty acids in the human colon. *Gut*, 22(9), 763–779.
- Cummings, J. H., Hill, M. J., Bone, E. S., Branch, W. J., and Jenkins, D. J. A. (1979). The effect of meat protein and dietary fibre on colonic function and metabolism II. Bacterial metabolites in faeces and urine. *The American Journal of Clinical Nutrition*, 32(10), 2094-2101.
- Daims, H., Brühl, A., Amann, R., Schleifer, K.-H., and Wagner, M. (1999). The domain-specific probe EUB338 is insufficient for the detection of all bacteria: development and evaluation of a more comprehensive probe

set. *Systematic and Applied Microbiology*, 22(3), 434–444.  
doi:10.1016/s0723-2020(99)80053-8

Daims, H., Stoecker, K., and Wagner, M. (2004). Fluorescence *in situ* hybridization for the detection of prokaryotes. In *Molecular microbial ecology* (pp. 208–228). Taylor and Francis.

David, L. A., Maurice, C. F., Carmody, R. N., Gootenberg, D. B., Button, J. E., Wolfe, B. E., ... and Biddinger, S. B. (2014). Diet rapidly and reproducibly alters the human gut microbiome. *Nature*, 505(7484), 559–563. doi: 10.1038/nature12820

de Goffau, M. C., Luopajarvi, K., Knip, M., Ilonen, J., Ruohtula, T., Härkönen, T., ... and Vaarala, O. (2013). Faecal microbiota composition differs between children with  $\beta$ -cell autoimmunity and those without. *Diabetes*, 62(4), 1238–1244.

Degnan, B. A., Macfarlane, S., Quigley, M. E., and Macfarlane, G. T. (1997). Starch utilization by *Bacteroides ovatus* isolated from the human large intestine. *Current Microbiology*, 34(5), 290–296.

den Besten, G., van Eunen, K., Groen, A. K., Venema, K., Reijngoud, D. J., and Bakker, B. M. (2013). The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *Journal of Lipid Research*, 54(9), 2325–2340. doi: 10.1194/jlr.r036012

Diamant, M., Blaak, E. E., and De Vos, W. M. (2011). Do nutrient–gut–microbiota interactions play a role in human obesity, insulin resistance and type 2 diabetes?. *Obesity Reviews*, 12(4), 272–281. doi: 10.1111/j.1467-789x.2010.00797.x

Dimidi, E., Christodoulides, S., Fragkos, K. C., Scott, S. M., and Whelan, K. (2014). The effect of probiotics on functional constipation in adults: a systematic review and meta-analysis of randomized controlled trials. *The American Journal of Clinical Nutrition*, 100(4), 1075–1084. doi:10.3945/ajcn.114.089151

Dominika, Š., Arjan, N., Karyn, R. P., and Henryk, K. (2011). The study on the impact of glycated pea proteins on human intestinal bacteria. *International Journal of Food Microbiology*, 145(1), 267–272.

Dubois, M., Gilles, K. A., Hamilton, J. K., Rebers, P. T., and Smith, F. (1956). Colorimetric method for determination of sugars and related substances. *Analytical Chemistry*, 28(3), 350–356.

Duke, J. A. (2018). *Handbook of Medicinal Herbs Herbal Reference Library*. Milton: CRC Press.

- Duncan, S. H., Barcenilla, A., Stewart, C. S., Pryde, S. E., and Flint, H. J. (2002). Acetate utilization and butyryl coenzyme A (CoA): acetate-CoA transferase in butyrate-producing bacteria from the human large intestine. *Applied Environmental Microbiology*, 68(10), 5186-5190.
- Duncan, S. H., Louis, P., and Flint, H. J. (2004). Lactate-utilizing bacteria, isolated from human faeces, that produce butyrate as a major fermentation product. *Applied Environmental Microbiology*, 70(10), 5810-5817. doi: 10.1128/aem.70.10.5810-5817.2004
- Duncan, S. H., Russell, W. R., Quartieri, A., Rossi, M., Parkhill, J., Walker, A. W., and Flint, H. J. (2016). Wheat bran promotes enrichment within the human colonic microbiota of butyrate-producing bacteria that release ferulic acid. *Environmental Microbiology*, 18(7), 2214–2225.
- Eckburg, P. B., Bik, E. M., Bernstein, C. N., Purdom, E., Dethlefsen, L., Sargent, M., Gill, S. R., Nelson, K. E., and Relman, D. A. (2005). Diversity of the human intestinal microbial flora. *Science*, 308(5728), 1635–1638.
- Egadu, S. P., Mucunguzi, P., and Obua, J. (2007). Uses of tree species producing gum arabic in Karamoja, Uganda. *African Journal of Ecology*, 45(s1), 17–21. doi: 10.1111/j.1365-2028.2007.00732.x
- ElKhawad, H.E. (2008). Gum arabic processing and marketing in Sudan. M.Sc. thesis in Chemical Engineering, University of Khartoum
- Erra-Pujada, M., Debeire, P., Duchiron, F., and O'Donohue, M. J. (1999). The type II pullulanase of *Thermococcus hydrothermalis*: molecular characterization of the gene and expression of the catalytic domain. *Journal of Bacteriology*, 181(10), 3284-3287.
- Everard, A., Lazarevic, V., Derrien, M., Girard, M., Muccioli, G. G., Neyrinck, A. M., ... and Cani, P. D. (2011). Responses of gut microbiota and glucose and lipid metabolism to prebiotics in genetic obese and diet-induced leptin-resistant mice. *Diabetes*, 60(11), 2775–2786. doi:10.2337/db11-0227
- Falony, G., Calmeyn, T., Leroy, F., and De Vuyst, L. (2009). Coculture fermentations of *Bifidobacterium* species and *Bacteroides thetaiotaomicron* reveal a mechanistic insight into the prebiotic effect of inulin-type fructans. *Applied Environmental Microbiology*, 75(8), 2312-2319.
- FAO/WHO. (2002). *Guidelines for The Evaluation of Probiotics in Food*. World Health Organization (WHO) London, Ontario, Canada.
- Farmer, J. J., Fanning, G. R., Davis, B. R., O'hara, C. M., Riddle, C., Hickman-Brenner, F. W., ... and Brenner, D. J. (1985). *Escherichia fergusonii* and

*Enterobacter tayloreae*, two new species of *Enterobacteriaceae* isolated from clinical specimens. *Journal of Clinical Microbiology*, 21(1), 77-81.

- Fässler, C., Arrigoni, E., Venema, K., Hafner, V., Brouns, F., and Amadò, R. (2006). Digestibility of resistant starch containing preparations using two *in vitro* models. *European Journal of Nutrition*, 45(8), 445-453.
- Flint, H. J. (2006). The significance of prokaryote diversity in the human gastrointestinal tract. In *Prokaryotic Diversity: Mechanisms and Significance* (p. 65 p). Cambridge University Press.
- Flint, H. J., Duncan, S. H., Scott, K. P., and Louis, P. (2015). Links between diet, gut microbiota composition and gut metabolism. *Proceedings of The Nutrition Society*, 74(1), 13-22.
- Floch, M. H. and Hong-Curtiss, J. (2002). Probiotics and functional foods in gastrointestinal disorders. *Current Treatment Options in Gastroenterology*, 5(4), 311–321.
- Flourie, B., Etanchaud, F., Florent, C., Pellier, P., Bouhnik, Y., and Rambaud, J. C. (1990). Comparative study of hydrogen and methane production in the human colon using caecal and faecal homogenates. *Gut*, 31(6), 684–685.
- Food and Agriculture Organization (FAO) of the United Nations. (1990). Specifications for identity and purity of certain food additives (Food and Nutrition Paper No. 49). FAO, Rome, Italy, 23-25.
- Ford, A. C., Quigley, E. M. M., Lacy, B. E., Lembo, A. J., Saito, Y. A., Schiller, L. R., ... and Moayyedi, P. (2014). Efficacy of prebiotics, probiotics, and synbiotics in irritable bowel syndrome and chronic idiopathic constipation: systematic review and meta-analysis. *American Journal of Gastroenterology*, 109(10), 1547–1561. doi:10.1038/ajg.2014.202
- Forgetta, V., Rempel, H., Malouin, F., Vaillancourt Jr, R., Topp, E., Dewar, K., and Diarra, M. S. (2012). Pathogenic and multidrug-resistant *Escherichia fergusonii* from broiler chicken. *Poultry Science*, 91(2), 512-525.
- Forster, S. C., Kumar, N., Anonye, B. O., Almeida, A., Viciani, E., Stares, M. D., Dunn, M., Mkandawire, T. T., Zhu, A., Shao, Y., and Pike, L. J. (2019). A human gut bacterial genome and culture collection for improved metagenomic analyses. *Nature Biotechnology*, 37(2), 186.
- Fossi, B. T. and Tavea, F. (2013). Application of amylolytic *Lactobacillus fermentum* 04BBA19 in fermentation for simultaneous production of thermostable alpha-amylase and lactic acid. In *Lactic Acid Bacteria-R and D for Food, Health and Livestock Purposes*. IntechOpen.

- Frost, G. S., Walton, G. E., Swann, J. R., Psichas, A., Costabile, A., Johnson, L. P., ... and Barraclough, T. G. (2014). Impacts of plant-based foods in ancestral hominin diets on the metabolism and function of gut microbiota *in vitro*. *MBio*, 5(3), e00853-14.
- Fukuda, S., Toh, H., Hase, K., Oshima, K., Nakanishi, Y., Yoshimura, K., ... and Taylor, T. D. (2011). Bifidobacteria can protect from enteropathogenic infection through production of acetate. *Nature*, 469(7331), 543-547. doi: 10.1038/nature09646
- Fuller, M. F. (1991). *In vitro digestion for pigs and poultry*. CAB International (CABI).
- Furusawa, Y., Obata, Y., Fukuda, S., Endo, T. A., Nakato, G., Takahashi, D., ... and Takahashi, M. (2013). Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature*, 504(7480), 446-450. doi: 10.1038/nature13041
- Gaastra, W., Kusters, J. G., Van Duijkeren, E., and Lipman, L. J. A. (2014). *Escherichia fergusonii*. *Veterinary microbiology*, 172(1-2), 7-12.
- Gänzle, M. G. and Follador, R. (2012). Metabolism of oligosaccharides and starch in lactobacilli: a review. *Frontiers in Microbiology*, 3. doi:10.3389/fmicb.2012.00340
- Garrett, W. S. and Onderdonk, A. B. (2010). *Bacteroides, Prevotella, Porphyromonas, and Fusobacterium Species (and Other Medically Important Anaerobic Gram-Negative Bacilli)*. Mandell, Douglas, and Bennetts Principles and Practice of Infectious Diseases, 3111–3119. doi:10.1016/b978-0-443-06839-3.00247-2
- Gashua I. B. (2016) An Investigation of the Molecular Structure, Composition and Biophysical Properties of Gum Arabic (Ph.D. thesis), University of Wolverhampton.
- Gaudier, E., Jarry, A., Blottiere, H. M., De Coppet, P., Buisine, M. P., Aubert, J. P., ... and Hoebler, C. (2004). Butyrate specifically modulates MUC gene expression in intestinal epithelial goblet cells deprived of glucose. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 287(6), G1168-G1174. doi: 10.1152/ajpgi.00219.2004
- Geurts, L., Neyrinck, A., Delzenne, N., Knauf, C., and Cani, P. (2014). Gut microbiota controls adipose tissue expansion, gut barrier and glucose metabolism: novel insights into molecular targets and interventions using prebiotics. *Beneficial Microbes*, 5(1), 3–17. doi:10.3920/bm2012.0065
- Ghfarloo, M. H., Jouki, M., and Tabari, M. (2019). Production and characterization of synbiotic Doogh, a yogurt-based Iranian drink by gum

arabic, ginger extract and *B. bifidum*. *Journal of Food Science and Technology*, 57(3), 1158-1166. doi:10.1007/s13197-019-04151-4

Gibson G. R. and Roberfroid, M. B. (2008). *Handbook of Prebiotics*. Boca Raton (FL): CRC Press.

Gibson, G. and Wang, X. (1994). Regulatory effects of *Bifidobacteria* on the growth of other colonic bacteria. *Journal of Applied Bacteriology*, 77(4), 412–420. doi:10.1111/j.1365-2672.1994.tb03443.x

Gibson, G. R. (2004). Fibre and effects on probiotics (the prebiotic concept). *Clinical Nutrition Supplements*, 1(2), 25–31. doi:10.1016/j.clnu.2004.09.005

Gibson, G. R. and Roberfroid, M. B. (1995). Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. *The Journal of Nutrition*, 125(6), 1401–1412. doi:10.1093/jn/125.6.1401

Gibson, G. R., Cummings, J. H., and Macfarlane, G. T. (1988). Use of a three-stage continuous culture system to study the effect of mucin on dissimilatory sulfate reduction and methanogenesis by mixed populations of human gut bacteria. *Applied Environmental Microbiology*, 54(11), 2750-2755.

Gibson, G. R., Hutkins, R., Sanders, M. E., Prescott, S. L., Reimer, R. A., Salminen, S. J., ... and Verbeke, K. (2017). Expert consensus document: the International scientific association for probiotics and prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nature Reviews Gastroenterology and Hepatology*, 14(8), 491.

Gibson, G. R., Willems, A., Reading, S., and Collins, M. D. (1996). Fermentation of non-digestible oligosaccharides by human colonic bacteria. *Proceedings of the Nutrition Society*, 55(3), 899–912. doi:10.1079/pns19960087

Gill, C. I. R. and Rowland, I. R. (2002). Diet and cancer: assessing the risk. *British Journal of Nutrition*, 88(S1), s73-s87. doi: 10.1079/bjn2002632

Goh, Y. J., Lee, J. H., and Hutkins, R. W. (2007). Functional analysis of the fructooligosaccharide utilization operon in *Lactobacillus paracasei* 1195. *Applied Environmental Microbiology*, 73(18), 5716-5724. doi: 10.1128/aem.00805-07

Goh, Y. J., Zhang, C., Benson, A. K., Schlegel, V., Lee, J. H., and Hutkins, R. W. (2006). Identification of a putative operon involved in fructooligosaccharide utilization by *Lactobacillus paracasei*. *Applied*

*Environmental Microbiology*, 72(12), 7518-7530. doi: 10.1128/aem.00877-06

Gullón, B., Gullón, P., Sanz, Y., Alonso, J. L., and Parajó, J. (2011). Prebiotic potential of a refined product containing pectic oligosaccharides. *LWT - Food Science and Technology*, 44(8), 1687–1696. doi:10.1016/j.lwt.2011.03.006

Haddad, P. R. and Jackson, P. E. (1990). *Ion chromatography* (Vol. 46). Elsevier.

Hamer, H. M., De Preter, V., Windey, K., and Verbeke, K. (2012). Functional analysis of colonic bacterial metabolism: relevant to health?. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 302(1), G1-G9. doi: 10.1152/ajpgi.00048.2011

Hamer, H. M., Jonkers, D. M. A. E., Venema, K., Vanhoutvin, S. A. L. W., Troost, F. J., and Brummer, R. J. (2008). The role of butyrate on colonic function. *Alimentary Pharmacology and Therapeutics*, 27(2), 104-119. doi: 10.1111/j.1365-2036.2007.03562.x

Havenaar, R. (2011). Intestinal health functions of colonic microbial metabolites: a review. *Beneficial Microbes*, 2(2), 103-114. doi: 10.3920/bm2011.0003

Hayisama-Ae, W., Kantachote, D., Bhongsuwan, D., Nokkaew, U., and Chaiyasut, C. (2014). A potential synbiotic beverage from fermented red seaweed (*Gracilaria fisheri*) using *Lactobacillus plantarum* DW12. *International Food Research Journal*, 21(5).

Henningsson, Å., Björck, I., and Nyman, M. (2001). Short-chain fatty acid formation at fermentation of indigestible carbohydrates. *Food and Nutrition Research*, 45, 165–168.

Hidalgo, M., Oruna-Concha, M. J., Kolida, S., Walton, G. E., Kallithraka, S., Spencer, J. P., and de Pascual-Teresa, S. (2012). Metabolism of anthocyanins by human gut microflora and their influence on gut bacterial growth. *Journal of Agricultural and Food Chemistry*, 60(15), 3882-3890.

Hijova, E. and Chmelarova, A. (2007). Short chain fatty acids and colonic health. *BratislavskéLekárske Listy*, 108(8), 354.

Hillman, E. T., Lu, H., Yao, T., and Nakatsu, C. H. (2017). Microbial ecology along the gastrointestinal tract. *Microbes and Environments*, ME17017.

Hinz, S. W., Pastink, M. I., van den Broek, L. A., Vincken, J. P., and Voragen, A. G. (2005). *Bifidobacterium longum* endogalactanase liberates galactotriose from type I galactans. *Applied Environmental*



*Microbiology*, 71(9), 5501-5510. doi: 10.1128/aem.71.9.5501-5510.2005

- Holden, H. M., Rayment, I., and Thoden, J. B. (2003). Structure and function of enzymes of the Leloir pathway for galactose metabolism. *Journal of Biological Chemistry*, 278(45), 43885-43888. doi: 10.1074/jbc.r300025200
- Hosseini, E., Grootaert, C., Verstraete, W., and Van de Wiele, T. (2011). Propionate as a health-promoting microbial metabolite in the human gut. *Nutrition Reviews*, 69(5), 245-258.
- Hove, H. and Mortensen, P. B. (1995). Influence of intestinal inflammation (IBD) and small and large bowel length on faecal short-chain fatty acids and lactate. *Digestive Diseases and Sciences*, 40(6), 1372-1380.
- Howard, M. D., Gordon, D. T., Garleb, K. A., and Kerley, M. S. (1995). Dietary fructooligosaccharide, xylooligosaccharide and gum arabic have variable effects on cecal and colonic microbiota and epithelial cell proliferation in mice and rats. *The Journal of nutrition*, 125(10), 2604-2609.
- Huber, R. E. and Gaunt, M. T. (1983). Importance of hydroxyls at positions 3, 4, and 6 for binding to the "galactose" site of  $\beta$ -galactosidase (*Escherichia coli*). *Archives of Biochemistry and biophysics*, 220(1), 263-271.
- Huber, R. E. and Hurlburt, K. L. (1986). Reversion reactions of  $\beta$ -galactosidase (*Escherichia coli*). *Archives of Biochemistry and Biophysics*, 246(1), 411-418.
- Hughes, S. (2008). The effect of cereal cell wall fractions on fermentation by the human gut microbiota; with a focus upon health promoting effects (Doctoral dissertation), University of Reading.
- Hwang, L., Low, K., Khoshini, R., Melmed, G., Sahakian, A., Makhani, M., Pokkunuri, V., and Pimentel, M. (2010). Evaluating breath methane as a diagnostic test for constipation-predominant IBS. *Digestive Diseases and Sciences*, 55(2), 398-403.
- Ibrahim, M. A., Kohn, N., and Wapnir, R. A. (2004). Proabsorptive effect of gum arabic in isotonic solutions orally administered to rats: effect on zinc and other solutes. *The Journal of Nutritional Biochemistry*, 15(3), 185-189.
- Ibrahim, O. B., Osman, M. E., Hassan, E. A. (2013). Characterization and simple fractionation of *Acacia senegal*. *Journal of Chemical Acta*. (2), 11-17.

- Idris, O., and Haddad, G. (2011). Gum Arabic's (Gum Acacia's) Journey from Tree to End User. *Gum Arabic Special Publication*, 3-17. doi:10.1039/9781849733106-00003
- Iraporda, C., Errea, A., Romanin, D. E., Cayet, D., Pereyra, E., Pignataro, O., Sirard, J. C., Garrote, G. L., Abraham, A. G., and Rumbo, M. (2015). Lactate and short chain fatty acids produced by microbial fermentation downregulate proinflammatory responses in intestinal epithelial cells and myeloid cells. *Immunobiology*, 220(10), 1161-1169. doi: 10.1016/j.imbio.2015.06.004.
- Ishaq, S. L., Moses, P. L., and Wright, A.-D. G. (2016). The pathology of methanogenic archaea in human gastrointestinal tract disease. *The Gut Microbiome - Implications for Human Disease*. doi: <https://doi.org/10.5772/64637>
- Islam, A., Phillips, G., Slijivo, A., Snowden, M., and Williams, P. (1997). A review of recent developments on the regulatory, structural and functional aspects of gum arabic. *Food Hydrocolloids*, 11(4), 493–505. doi:10.1016/s0268-005x(97)80048-3
- Jang, S.-I., Kim, J.-H., Youn, Y. H., Park, H., Lee, S. I., and Conklin, J. L. (2010). Relationship between intestinal gas and the development of right colonic diverticula. *Journal of Neurogastroenterology and Motility*, 16(4), 418.
- Janssen, P. H. (2010). Influence of hydrogen on rumen methane formation and fermentation balances through microbial growth kinetics and fermentation thermodynamics. *Animal Feed Science and Technology*, 160(1–2), 1–22.
- Jenkins, D. J., Kendall, C. W., and Vuksan, V. (1999). Inulin, oligofructose and intestinal function. *The Journal of Nutrition*, 129(7), 1431S-1433S.
- Ji, G. -E., Han, H.-K., and Yun, S.-W. (1992). Isolation of amylolytic *Bifidobacterium* sp. Int-57 and characterization of amylase. *Journal of Microbiology and Biotechnology*, 2(2), 85–91.
- Joseleau, J. P. and Ullmann, G. (1985). A relation between starch metabolism and the synthesis of gum arabic. *Bull Int Group Study Mimosoideae*, 13, 46-54.
- Juers, D. H., Matthews, B. W., and Huber, R. E. (2012). LacZ  $\beta$ -galactosidase: structure and function of an enzyme of historical and molecular biological importance. *Protein Science*, 21(12), 1792-1807.
- Kaddam, L. A., and Kaddam, A. S. (2020). Effect of Gum Arabic (Acacia Senegal) on C-reactive protein level among Sickle Cell Anemia patients. doi:10.21203/rs.2.23398/v2

- Kafeshani, M. (2017). The gut microbiome, diet, and chronic kidney disease. *Journal of Preventive Epidemiology*, 2(1).
- Kalina, U., Koyama, N., Hosoda, T., Nuernberger, H., Sato, K., Hoelzer, D., ... and Böcker, U. (2002). Enhanced production of IL-18 in butyrate-treated intestinal epithelium by stimulation of the proximal promoter region. *European Journal of Immunology*, 32(9), 2635-2643. doi: 10.1002/1521-4141(200209)32:9<2635::aid-immu2635>3.0.co;2-n
- Kamal, E., Kaddam, L. A., Dahawi, M., Osman, M., Salih, M. A., Alagib, A., and Saeed, A. (2018). Gum Arabic Fibers Decreased Inflammatory Markers and Disease Severity Score among Rheumatoid Arthritis Patients, Phase II Trial. *International Journal of Rheumatology*, 2018, 1-6. doi:10.1155/2018/4197537
- Kappelman, M. D., Rifas-Shiman, S. L., Kleinman, K., Ollendorf, D., Bousvaros, A., Grand, R. J., and Finkelstein, J. A. (2007). The prevalence and geographic distribution of Crohn's disease and ulcerative colitis in the United States. *Clinical Gastroenterology and Hepatology*, 5(12), 1424-1429.
- Kim, G., Deepinder, F., Morales, W., Hwang, L., Weitsman, S., Chang, C., Gunsalus, R., and Pimentel, M. (2012). *Methanobrevibacter smithii* is the predominant methanogen in patients with constipation-predominant IBS and methane on breath. *Digestive Diseases and Sciences*, 57(12), 3213–3218.
- Kim, H. J., Kim, H. Y., Lee, S. Y., Seo, J. H., Lee, E., and Hong, S. J. (2013). Clinical efficacy and mechanism of probiotics in allergic diseases. *Korean Journal of Pediatrics*, 56(9), 369.
- King, S., Glanville, J., Sanders, M. E., Fitzgerald, A., and Varley, D. (2014). Effectiveness of probiotics on the duration of illness in healthy children and adults who develop common acute respiratory infectious conditions: a systematic review and meta-analysis. *British Journal of Nutrition*, 112(1), 41–54. doi:10.1017/s0007114514000075
- Kishimoto, A., Ushida, K., Phillips, G. O., Ogasawara, T., and Sasaki, Y. (2006). Identification of intestinal bacteria responsible for fermentation of gum arabic in pig model. *Current Microbiology*, 53(3), 173-177.
- Koeth, R. A., Wang, Z., Levison, B. S., Buffa, J. A., Org, E., Sheehy, B. T., ... and Smith, J. D. (2013). Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nature Medicine*, 19(5), 576. doi: 10.1038/nm.3145

- Komeda, Y. (1988). Isolation of fla-lacZ fusions in *Escherichia coli* K-12: most fusions result in soluble beta-galactosidase. *Journal of Bacteriology*, 170(4), 1980-1983.
- Korakli, M., Ganzle, M. and Vogel, R. (2002). Metabolism by *Bifidobacteria* and lactic acid bacteria of polysaccharides from wheat and rye and exopolysaccharides produced by *Lactobacillus sanfranciscensis*. *Journal of Applied Microbiology*, 92(5), 958-965. doi:10.1046/j.1365-2672.2002.01607.x
- Kravtchenko, T. (1998). The use of acacia gum as a source of soluble dietary fibre. *Gums and Stabilisers for The Food Industry* 9, 413-420. doi:10.1533/9781845698362.6.413
- Kunkel, D., Basseri, R. J., Makhani, M. D., Chong, K., Chang, C., and Pimentel, M. (2011). Methane on breath testing is associated with constipation: a systematic review and meta-analysis. *Digestive Diseases and Sciences*, 56(6), 1612-1618.
- Kuo, S.-M. (2013). Rhe interplay between fibre and the intestinal microbiome in the inflammatory response. *Advances in Nutrition*, 4(1), 16-28. doi:10.3945/an.112.003046
- La Rosa, S. L., Kachrimanidou, V., Buffetto, F., Pope, P. B., Pudlo, N. A., Martens, E. C., Rastall, R. A., Gibson, G. R., and Westereng, B. (2019). Wood-derived dietary fibres promote beneficial human gut microbiota. *MSphere*, 4(1), e00554-18.
- Lambeth, S. M., Carson, T., Lowe, J., Ramaraj, T., Leff, J. W., Luo, L., ... and Shah, V. O. (2015). Composition, diversity and abundance of gut microbiome in prediabetes and type 2 diabetes. *Journal of Diabetes and Obesity*, 2(3), 1.
- Latimer, G. W. (2019). *Official methods of analysis of AOAC International*. Rockville, MD: AOAC International.
- Lau, J. T., Whelan, F. J., Herath, I., Lee, C. H., Collins, S. M., Bercik, P., and Surette, M. G. (2016). Capturing the diversity of the human gut microbiota through culture-enriched molecular profiling. *Genome Medicine*, 8(1), 72.
- LeBlanc, J. G., Chain, F., Martín, R., Bermúdez-Humarán, L. G., Courau, S., and Langella, P. (2017). Beneficial effects on host energy metabolism of short-chain fatty acids and vitamins produced by commensal and probiotic bacteria. *Microbial Cell Factories*, 16(1), 79. doi: <https://doi.org/10.1186/s12934-017-0691-z>

- Lederberg, J. (1950). The beta-d-galactosidase of *Escherichia coli*, strain K-12. *Journal of Bacteriology*, *60*(4), 381.
- Lee, J. H., Lee, S. K., Park, K. H., Hwang, I. K., and Ji, G. E. (1999). Fermentation of rice using amylolytic *Bifidobacterium*. *International Journal of Food Microbiology*, *50*(3), 155–161.
- Lee, S. K., Kim, Y. B., and Ji, G. E. (1997). Note: purification of amylase secreted from *Bifidobacterium adolescentis*. *Journal of Applied Microbiology*, *83*(3), 267–272.
- Lemenih, M. (2005). Production and marketing of gums and gum resins in Ethiopia. *Production and Marketing of Gum Resins: Frankincense, Myrrh and Opoponax*, 55-70.
- Lepp, P. W., Brinig, M. M., Ouverney, C. C., Palm, K., Armitage, G. C., and Relman, D. A. (2004). Methanogenic archaea and human periodontal disease. *Proceedings of The National Academy of Sciences*, *101*(16), 6176–6181.
- Levitt, M. D. and Bond, J. H. (1970). Volume, composition, and source of intestinal gas. *Gastroenterology*, *59*(6), 921–929.
- Levitt, M. D., Gibson, G., and Christl, S. U. (1995). Gas metabolism in the large intestine. In Gibson, G. R., and Macfarlane, G. T. *Human Colonic Bacteria: Role in Nutrition, Physiology and Pathology* (pp. 131-154). Boca Raton, Florida: CRC Press Inc.
- Ley, R. E., Turnbaugh, P. J., Klein, S., and Gordon, J. I. (2006). Microbial ecology: human gut microbes associated with obesity. *Nature*, *444*: 1022–1023.
- Lilly, D. M. and Stillwell, R. H. (1965). Probiotics: growth-promoting factors produced by microorganisms. *Science*, *147*(3659), 747–748. doi:10.1126/science.147.3659.747
- Liu, J., Kandasamy, S., Zhang, J., Kirby, C. W., Karakach, T., Hafting, J., ... and Prithiviraj, B. (2015). Prebiotic effects of diet supplemented with the cultivated red seaweed *Chondrus crispus* or with fructo-oligo-saccharide on host immunity, colonic microbiota and gut microbial metabolites. *BMC Complementary and Alternative Medicine*, *15*(1). doi:10.1186/s12906-015-0802-5
- Lobel, L. and Garrett, W. S. (2019). Butyrate makes macrophages “Go Nuclear” against bacterial pathogens. *Immunity*, *50*(2), 275-278.
- Loeffler, R. T., Sinnott, M. L., Sykes, B. D., and Withers, S. G. (1979). Interaction of the lacZ  $\beta$ -galactosidase of *Escherichia coli* with some  $\beta$ -d-

- galactopyranoside competitive inhibitors. *Biochemical Journal*, 177(1), 145-152.
- Lopez-Torrez, L., Nigen, M., Williams, P., Doco, T., and Sanchez, C. (2015). *Acacia senegal* vs. *Acacia seyal* gums—Part 1: composition and structure of hyperbranched plant exudates. *Food Hydrocolloids*, 51, 41-53. doi: 10.1016/j.foodhyd.2015.04.019.
- Losada, M. and Olleros, T. (2002). Towards a healthier diet for the colon: the influence of fructooligosaccharides and *Lactobacilli* on intestinal health. *Nutrition Research*, 22(1-2), 71–84. doi:10.1016/s0271-5317(01)00395-5
- Louis, P. and Flint, H. J. (2009). Diversity, metabolism and microbial ecology of butyrate-producing bacteria from the human large intestine. *FEMS Microbiology Letters*, 294(1), 1–8.
- Louis, P. and Flint, H. J. (2017). Formation of propionate and butyrate by the human colonic microbiota. *Environmental Microbiology*, 19(1), 29-41.
- Louis, P., Duncan, S. H., McCrae, S. I., Millar, J., Jackson, M. S., and Flint, H. J. (2004). Restricted distribution of the butyrate kinase pathway among butyrate-producing bacteria from the human colon. *Journal of Bacteriology*, 186(7), 2099-2106.
- Louis, P., Hold, G. L., and Flint, H. J. (2014). The gut microbiota, bacterial metabolites and colorectal cancer. *Nature Reviews Microbiology*, 12(10), 661-672. doi: 10.1038/nrmicro3344
- Louis, P., Scott, K., Duncan, S. and Flint, H. (2007). Understanding the effects of diet on bacterial metabolism in the large intestine. *Journal of Applied Microbiology*, 102(5), 1197–1208. doi:10.1111/j.1365-2672.2007.03322.x
- Louis, P., Young, P., Holtrop, G., and Flint, H. J. (2010). Diversity of human colonic butyrate-producing bacteria revealed by analysis of the butyryl-CoA: acetate CoA-transferase gene. *Environmental Microbiology*, 12(2), 304-314.
- Lu, K., Abo, R. P., Schlieper, K. A., Graffam, M. E., Levine, S., Wishnok, J. S., ... and Fox, J. G. (2014). Arsenic exposure perturbs the gut microbiome and its metabolic profile in mice: an integrated metagenomics and metabolomics analysis. *Environmental Health Perspectives*, 122(3), 284-291.
- Macfarlane, G. T. and Englyst, H. N. (1986). Starch utilization by the human large intestinal microflora. *Journal of Applied Bacteriology*, 60(3), 195–201.

- Macfarlane, G. T. and Gibson, G. R. (1997). Carbohydrate fermentation, energy transduction and gas metabolism in the human large intestine. In *Gastrointestinal Microbiology* (pp. 269–318). Springer.
- Macfarlane, G. T., Gibson, G. R., and Cummings, J. H. (1992). Comparison of fermentation reactions in different regions of the human colon. *Journal of Applied Bacteriology*, 72(1), 57-64.
- Macfarlane, G. T., Macfarlane, S., and Gibson, G. R. (1998). Validation of a three-stage compound continuous culture system for investigating the effect of retention time on the ecology and metabolism of bacteria in the human colon. *Microbial Ecology*, 35(2), 180-187.
- Macfarlane, S. and Macfarlane, G. T. (2003). Regulation of short-chain fatty acid production. *Proceedings of the Nutrition Society*, 62(1), 67-72.
- Macia, L., Tan, J., Vieira, A. T., Leach, K., Stanley, D., Luong, S., ... and Binge, L. (2015). Metabolite-sensing receptors GPR43 and GPR109A facilitate dietary fibre-induced gut homeostasis through regulation of the inflammasome. *Nature Communications*, 6, 6734. doi: 10.1038/ncomms7734
- Mackie, R. I. and Gilchrist, F. M. (1979). Changes in lactate-producing and lactate-utilizing bacteria in relation to pH in the rumen of sheep during stepwise adaptation to a high-concentrate diet. *Applied Environmental Microbiology*, 38(3), 422-430.
- Magee, E. A., Richardson, C. J., Hughes, R., and Cummings, J. H. (2000). Contribution of dietary protein to sulfide production in the large intestine: an *in vitro* and a controlled feeding study in humans. *The American Journal of Clinical Nutrition*, 72(6), 1488-1494.
- Maheux, A. F., Boudreau, D. K., Bergeron, M. G., and Rodriguez, M. J. (2014). Characterization of *Escherichia fergusonii* and *Escherichia albertii* isolated from water. *Journal of Applied Microbiology*, 117(2), 597-609.
- Maifreni, M., Frigo, F., Bartolomeoli, I., Innocente, N., Biasutti, M., and Marino, M. (2013). Identification of the Enterobacteriaceae in Montasio cheese and assessment of their amino acid decarboxylase activity. *Journal of Dairy Research*, 80(1), 122-127.
- Mandalari, G., Nueno-Palop, C., Bisignano, G., Wickham, M., and Narbad, A. (2008). Potential prebiotic properties of almond (*Amygdalus communis* L.) seeds. *Applied and Environmental Microbiology*, 74(14), 4264–4270.
- Manichanh, C., Rigottier-Gois, L., Bonnaud, E., Gloux, K., Pelletier, E., Frangeul, L., ... and Roca, J. (2006). Reduced diversity of faecal microbiota in

- Crohn's disease revealed by a metagenomic approach. *Gut*, 55(2), 205-211.
- Mao, B., Gu, J., Li, D., Cui, S., Zhao, J., Zhang, H., and Chen, W. (2018). Effects of different doses of fructooligosaccharides (FOS) on the composition of mice faecal microbiota, especially the Bifidobacterium composition. *Nutrients*, 10(8), 1105.
- Mariod, A. A. (2018). Gum arabic: Structure, properties, application and economics. London, United Kingdom: Academic Press.
- Martínez, I., Lattimer, J. M., Hubach, K. L., Case, J. A., Yang, J., Weber, C. G., Louk, J. A., Rose, D. J., Kyureghian, G., and Peterson, D. A. (2013). Gut microbiome composition is linked to whole grain-induced immunological improvements. *The ISME Journal*, 7(2), 269.
- May, T., Mackie, R. I., Fahey, G. C., Cremin, J. C., and Garleb, K. A. (1994). Effect of fibre source on short-chain fatty acid production and on the growth and toxin production by *Clostridium difficile*. *Scandinavian journal of gastroenterology*, 29(10), 916-922.
- McBurney, M. I. and Thompson, L. U. (1987). Effect of human faecal inoculum on *in vitro* fermentation variables. *British Journal of Nutrition*, 58(2), 233–243. doi:10.1079/bjn19870091
- McCarthy, R. E., Pajreau, M., and Salyers, A. A. (1988). Role of starch as a substrate for *Bacteroides vulgatus* growing in the human colon. *Applied and Environmental Microbiology*, 54(8), 1911–1916.
- McCleary, B. V. (2011). The evolution of dietary fibre definitions and methods and the role of AACC International. *Cereal Foods World*, 56(3), 103.
- Mcfarland, L. (2015). Deciphering meta-analytic results: a mini-review of probiotics for the prevention of paediatric antibiotic-associated diarrhoea and *Clostridium difficile* infections. *Beneficial Microbes*, 6(2), 189–194. doi:10.3920/bm2014.0034
- McKay, L. F., Holbrook, W. P., and Eastwood, M. A. (1982). Methane and hydrogen production by human intestinal anaerobic bacteria. *Acta Pathologica Microbiologica Scandinavica Series B: Microbiology*, 90(1–6), 257–260.
- Mee, K. A. and Gee, D. L. (1997). Apple fibre and gum arabic lowers total and low-density lipoprotein cholesterol levels in men with mild hypercholesterolemia. *Journal of the American Dietetic Association*, 97(4), 422–424. doi:10.1016/s0002-8223(97)00106-5



- Menni, C., Lin, C., Cecelja, M., Mangino, M., Matey-Hernandez, M. L., Keehn, L., ... and Chowieńczyk, P. (2018). Gut microbial diversity is associated with lower arterial stiffness in women. *European Heart Journal*, 39(25), 2390-2397.
- Meyer, D. and Stasse-Wolthuis, M. (2009). The bifidogenic effect of inulin and oligofructose and its consequences for gut health. *European Journal of Clinical Nutrition*, 63(11), 1277–1289. doi:10.1038/ejcn.2009.64
- Miller-Fleming, L., Olin-Sandoval, V., Campbell, K., and Ralser, M. (2015). Remaining mysteries of molecular biology: the role of polyamines in the cell. *Journal of Molecular Biology*, 427(21), 3389-3406. doi: 10.1016/j.jmb.2015.06.020
- Miller, T. L. and Wolin, M. (1979). Fermentations by saccharolytic intestinal bacteria. *The American Journal of Clinical Nutrition*, 32(1), 164–172.
- Miller, T. L. and Wolin, M. J. (1996). Pathways of acetate, propionate, and butyrate formation by the human fecal microbial flora. *Applied and environmental microbiology*, 62(5), 1589–1592. doi:10.1128/aem.62.5.1589-1592.1996
- Min, M., Bunt, C. R., Mason, S. L., and Hussain, M. A. (2018). Non-dairy probiotic food products: An emerging group of functional foods. *Critical Reviews in Food Science and Nutrition*, 1–16.
- Minekus, M., Smeets-Peeters, M., Bernalier, A., Marol-Bonnin, S., Havenaar, R., Marteau, P., ... and Fonty, G. (1999). A computer-controlled system to simulate conditions of the large intestine with peristaltic mixing, water absorption and absorption of fermentation products. *Applied Microbiology and Biotechnology*, 53(1), 108-114.
- Miquel, S., Martín, R., Bridonneau, C., Robert, V., Sokol, H., Bermúdez-Humarán, L. G., ... and Langella, P. (2014). Ecology and metabolism of the beneficial intestinal commensal bacterium *Faecalibacterium prausnitzii*. *Gut Microbes*, 5(2), 146-151.
- Miquel, S., Martin, R., Rossi, O., Bermudez-Humaran, L. G., Chatel, J. M., Sokol, H., ... and Langella, P. (2013). *Faecalibacterium prausnitzii* and human intestinal health. *Current Opinion in Microbiology*, 16(3), 255-261.
- Mirghani, M. E., Elnour, A. A., Kabbashi, N., Alam, M. Z., Musa, K. H., and Abdullah, A. (2018). Determination of antioxidant activity of gum arabic: An exudation from two different locations. *Science Asia*, 44(3), 179. doi:10.2306/scienceasia1513-1874.2018.44.179
- Mohamed, R. E., Gadour, M. O., and Adam, I. (2015). The lowering effect of Gum Arabic on hyperlipidemia in Sudanese patients. *Frontiers in physiology*, 6, 160.

- Moro, T., Rasmussen, H., and Hamaker, B. (2019). Potential of prebiotic butyrogenic fibres in Parkinson's disease. *Frontiers in Neurology*, 10, 663.
- Morrison, D. J. and Preston, T. (2016). Formation of short chain fatty acids by the gut microbiota and their impact on human metabolism. *Gut Microbes*, 7(3), 189-200.
- Mortensen, P. B. and Clausen, M. R. (1996). Short-chain fatty acids in the human colon: relation to gastrointestinal health and disease. *Scandinavian Journal of Gastroenterology*, 31(sup216), 132–148.
- Moses, P., Ishaq, S., Gupta, K., Maurer, S., and Wright, A.-D. (2015). Biodiversity of human gut methanogens varies with concentration of exhaled breath methane. *American Journal of Gastroenterology*, 110.
- Moundras, C., Behr, S. R., Demigné, C., Mazur, A., and Rémésy, C. (1994). Fermentable polysaccharides that enhance faecal bile acid excretion lower plasma cholesterol and apolipoprotein E-rich HDL in rats. *The Journal of Nutrition*, 124(11), 2179-2188.
- Musa, H. H., Ahmed, A. A., and Musa, T. H. (2017). Chemistry, biological, and pharmacological properties of gum arabic. *Reference Series in Phytochemistry Sweeteners*, 1–18. doi: 10.1007/978-3-319-54528-8\_11-1
- Nakamura, J., Kubota, Y., Miyaoka, M., Saitoh, T., Mizuno, F., and Benno, Y. (2002). Comparison of four microbial enzymes in *Clostridia* and *Bacteroides* isolated from human feces. *Microbiology and Immunology*, 46(7), 487-490.
- Nakov, G., Georgieva, D., Ivanova, N., Damyanova, S., Stamatovska, V., and Necinova, L. (2016). Prebiotic effects of inulin and acacia gum. *Food and Environment Safety Journal*, 14(2).
- Neyrinck, A. M., Possemiers, S., Verstraete, W., De Backer, F., Cani, P. D., and Delzenne, N. M. (2012). Dietary modulation of *Clostridial* cluster XIVa gut bacteria (*Roseburia spp.*) by chitin–glucan fibre improves host metabolic alterations induced by high-fat diet in mice. *The Journal of Nutritional Biochemistry*, 23(1), 51–59.
- Ng, K. L. and Haddad, P. R. (2000). ACIDS| Liquid Chromatography.
- Nielsen, S. S. (2017). Total Carbohydrate by Phenol-Sulfuric Acid Method. *Food Analysis Laboratory Manual Food Science Text Series*, 137-141. doi:10.1007/978-3-319-44127-6\_14

- Norat, T. and Riboli, E. (2001). Meat consumption and colorectal cancer: a review of epidemiologic evidence. *Nutrition Reviews*, 59(2), 37-47. doi: 10.1111/j.1753-4887.2001.tb06974.x
- O'Connell-Motherway, M., Kinsella, M., Fitzgerald, G. F., and van Sinderen, D. (2013). Transcriptional and functional characterization of genetic elements involved in galacto-oligosaccharide utilization by *Bifidobacterium breve* UCC 2003. *Microbial Biotechnology*, 6(1), 67-79. doi: 10.1111/1751-7915.12011
- Ohashi, Y., Harada, K., Tokunaga, M., Ishihara, N., Okubo, T., Ogasawara, Y., ... Fujisawa, T. (2012). Faecal fermentation of partially hydrolyzed guar gum. *Journal of Functional Foods*, 4(1), 398-402. doi:10.1016/j.jff.2011.09.007
- Olano-Martin, E., Mountzouris, K. C., Gibson, G. R. and Rastall, R. A. (2000). *In vitro* fermentability of dextran, oligodextran and maltodextrin by human gut bacteria. *British Journal of Nutrition*, 83(3), 247-255. doi: 10.1017/S0007114500000325.
- Onderdonk, A. B. and Garrett, W. S. (2014). Gas gangrene and other clostridium-associated diseases. In *Mandell, Douglas and Bennett's Principles and Practice of Infectious Diseases* (Vol. 2, pp. 2768-2772). doi: https://doi.org/10.1016/B978-1-4557-4801-3.00248-4
- Opstelten, J. L., Plassais, J., van Mil, S. W., Achouri, E., Pichaud, M., Siersema, P. D., ... and Cervino, A. C. (2016). Gut microbial diversity is reduced in smokers with Crohn's disease. *Inflammatory Bowel Diseases*, 22(9), 2070-2077.
- Orwa, C., Mutua, A., Kindt, R., Jamnadass, R. and Anthony, S. (2009). *Agroforestry Database: A Tree Reference and Selection Guide Version 4.0*. Kenya: World Agroforestry
- Ouwehand, A. C., Tiihonen, K., Saarinen, M., Putaala, H., and Rautonen, N. (2008). Influence of a combination of *Lactobacillus acidophilus* NCFM and lactitol on healthy elderly: intestinal and immune parameters. *British Journal of Nutrition*, 101(3), 367-375. doi:10.1017/s0007114508003097
- Palframan, R. J., Gibson, G. R., and Rastall, R. A. (2002). Effect of pH and dose on the growth of gut bacteria on prebiotic carbohydrates *in vitro*. *Anaerobe*, 8(5), 287-292.
- Pandey, A. (2003). Solid-state fermentation. *Biochemical Engineering Journal*, 13(2-3), 81-84.

- Parkar, S. G., Trower, T. M., and Stevenson, D. E. (2013). Faecal microbial metabolism of polyphenols and its effects on human gut microbiota. *Anaerobe*, 23, 12–19.
- Parks, D. H., Chuvochina, M., Waite, D. W., Rinke, C., Skarshewski, A., Chaumeil, P. A., and Hugenholtz, P. (2018). A standardized bacterial taxonomy based on genome phylogeny substantially revises the tree of life. *Nature Biotechnology*, 36(10), 996-1004.
- Pasumarthi, R., Chandrasekaran, S., and Mutnuri, S. (2013). Biodegradation of crude oil by *Pseudomonas aeruginosa* and *Escherichia fergusonii* isolated from the Goan coast. *Marine Pollution Bulletin*, 76(1-2), 276-282.
- Paul, A. A. and Southgate, D. A. (1985). *The composition of foods*, Elsevier.
- Payne, A. N., Zihler, A., Chassard, C., and Lacroix, C. (2012). Advances and perspectives in *in vitro* human gut fermentation modelling. *Trends in Biotechnology*, 30(1), 17–25.
- Petrova, P., Petrov, K., and Stoyancheva, G. (2013). Starch-modifying enzymes of lactic acid bacteria—structures, properties, and applications. *Starch-Stärke*, 65(1–2), 34–47.
- Phillips, A. O. and Phillips, G. O. (2011). Biofunctional behaviour and health benefits of a specific gum arabic. *Food Hydrocolloids*, 25(2), 165-169.
- Phillips, G. O. (1998). Acacia gum (Gum Arabic): A nutritional fibre; metabolism and calorific value. *Food Additives and Contaminants*, 15(3), 251–264. doi:10.1080/02652039809374639
- Phillips, G. O. and Williams, P. (2001). Tree exudate gums: natural and versatile food additives and ingredients. *Food Ingredients and Analysis International*, 26.
- Pimentel, M., Mayer, A. G., Park, S., Chow, E. J., Hasan, A., and Kong, Y. (2003). Methane production during lactulose breath test is associated with gastrointestinal disease presentation. *Digestive Diseases and Sciences*, 48(1), 86–92.
- Pitt, P., De Bruijn, K. M., Beeching, M. F., Goldberg, E., and Blendis, L. M. (1980). Studies on breath methane: the effect of ethnic origins and lactulose. *Gut*, 21(11), 951-954.
- Pittman, K. A. and Bryant, M. P. (1964). Peptides and other nitrogen sources for growth of *Bacteroides ruminicola*. *Journal of Bacteriology*, 88(2), 401-410.

- Pokusaeva, K., Fitzgerald, G. F., and van Sinderen, D. (2011). Carbohydrate metabolism in Bifidobacteria. *Genes and Nutrition*, 6(3), 285-306.
- Pons, T., Olmea, O., China, G., Beldarraín, A., Márquez, G., Acosta, N., ... and Valencia, A. (1998). Structural model for family 32 of glycosyl-hydrolase enzymes. *Proteins: Structure, Function, and Bioinformatics*, 33(3), 383-395. doi: 10.1002/(sici)1097-0134(19981115)33:3<383::aid-prot7>3.0.co;2-r
- Pool-Zobel, B., Van Loo, J., Rowland, I., and Roberfroid, M. B. (2002). Experimental evidences on the potential of prebiotic fructans to reduce the risk of colon cancer. *British Journal of Nutrition*, 87(S2), S273-S281.
- Postler, T. S. and Ghosh, S. (2017). Understanding the holobiont: how microbial metabolites affect human health and shape the immune system. *Cell Metabolism*, 26(1), 110-130. doi: 10.1016/j.cmet.2017.05.008
- Pourabedin, M., Guan, L., and Zhao, X. (2015). Xylo-oligosaccharides and virginiamycin differentially modulate gut microbial composition in chickens. *Microbiome*, 3(1). doi: 10.1186/s40168-015-0079-4
- Pratt, C. W. and Cornely, K. (2004). In *Essential biochemistry* (p. 626). Wiley Hoboken, NJ.
- Rabin, M., Uhlenbeck, O. C., Steffensen, D. M., and Mangel, W. F. (1984). Chromosomal sites of integration of simian virus 40 DNA sequences mapped by *in situ* hybridization in two transformed hybrid cell lines. *Journal of Virology*, 49(2), 445-451. doi:10.1128/jvi.49.2.445-451.1984
- Rahim A. H. A. (2006). Economic analysis of deforestation-the case of gum belt in Sudan. (Unpublished doctoral dissertation). Wageningen: Wageningen University,.
- Rahim, A. H., van Ierland, E. C., and Wesseler, J. (2007). Economic incentives for abandoning or expanding gum arabic production in Sudan. *Forest Policy and Economics*, 10(1-2), 36-47.
- Rajilić-Stojanović, M. and deVos, W. M. (2014). The first 1000 cultured species of the human gastrointestinal microbiota. *FEMS Microbiology Reviews*. 38, 996-1047.
- Ramirez-Farias, C., Slezak, K., Fuller, Z., Duncan, A., Holtrop, G., and Louis, P. (2008). Effect of inulin on the human gut microbiota: stimulation of *Bifidobacterium adolescentis* and *Faecalibacterium prausnitzii*. *British Journal of Nutrition*, 101(4), 541-550. doi: 10.1017/s0007114508019880

- Ramsay, A. G., Scott, K. P., Martin, J. C., Rincon, M. T., and Flint, H. J. (2006). Cell-associated  $\alpha$ -amylases of butyrate-producing *Firmicute* bacteria from the human colon. *Microbiology*, 152(11), 3281-3290.
- Randall, R. C., Phillips, G. O., and Williams, P. A. (1989). Fractionation and characterization of gum from *Acacia senegal*. *Food Hydrocolloids*, 3(1), 65-75. doi:10.1016/s0268-005x(89)80034-7
- Readers Digest Association. (1986). *Magic and Medicine of Plants*. Pleasantville, New York.
- Reddy, G., Altaf, M. D., Naveena, B. J., Venkateshwar, M., and Kumar, E. V. (2008). Amylolytic bacterial lactic acid fermentation—a review. *Biotechnology Advances*, 26(1), 22–34.
- Rehman, K. U., Codipilly, C. N., and Wapnir, R. A. (2004). Modulation of small intestinal nitric oxide synthase by gum arabic. *Experimental Biology and Medicine*, 229(9), 895-901.
- Rehman, K. U., Wingertzahn, M. A., Teichberg, S., Harper, R. G., and Wapnir, R. A. (2003). Gum arabic (GA) modifies paracellular water and electrolyte transport in the small intestine. *Digestive Diseases and Sciences*, 48(4), 755-760.
- Rehman, K., Wingertzahn, M. A., Harper, R. G., and Wapnir, R. A. (2001). Proabsorptive action of gum arabic: regulation of nitric oxide metabolism in the basolateral potassium channel of the small intestine. *Journal of Paediatric Gastroenterology and Nutrition*, 32(5), 529-533.
- Reichardt, N., Duncan, S. H., Young, P., Belenguer, A., Leitch, C. M., Scott, K. P., ... and Louis, P. (2014). Phylogenetic distribution of three pathways for propionate production within the human gut microbiota. *The ISME journal*, 8(6), 1323-1335. doi: 10.1038/ismej.2014.48
- Reid, G. (2014). Modulating the vaginal microbiome: the need for a bridge between science and practice. *Seminars in Reproductive Medicine*, 32(01), 028–034. doi:10.1055/s-0033-1361820
- Renard, D., Lavenant-Gourgeon, L., Lapp, A., Nigen, M., and Sanchez, C. (2014). Enzymatic hydrolysis studies of arabinogalactan-protein structure from *Acacia* gum: The self-similarity hypothesis of assembly from a common building block. *Carbohydrate Polymers*, 112, 648-661. doi: 10.1016/j.carbpol.2014.06.041.
- Renard, D., Lavenant-Gourgeon, L., Ralet, M., and Sanchez, C. (2006). *Acacia senegal* Gum: Continuum of Molecular Species Differing by Their Protein to Sugar Ratio, Molecular Weight, and Charges. *Biomacromolecules*, 7(9), 2637-2649. doi:10.1021/bm060145j

- Reznikoff, W. S. and Miller, J. H. (Eds.). (1978). *The operon*. Cold Spring Harbor Laboratory.
- Ridaura, V. K., Faith, J. J., Rey, F. E., Cheng, J., Duncan, A. E., Kau, A. L., ... and Muehlbauer, M. J. (2013). Gut microbiota from twins discordant for obesity modulate metabolism in mice. *Science*, *341*(6150), 1241214. doi:10.1126/science.1241214
- Rigottier-Gois, L., Rochet, V., Garrec, N., Suau, A. and Doré, J. (2003). Enumeration of bacteroides species in human faeces by fluorescent in situ hybridisation combined with flow cytometry using 16s rRNA probes. *Systematic and Applied Microbiology*, *26*(1), 110–118. doi:10.1078/072320203322337399
- Ríos-Covián, D., Ruas-Madiedo, P., Margolles, A., Gueimonde, M., de los Reyes-Gavilán, C. G., and Salazar, N. (2016). Intestinal short chain fatty acids and their link with diet and human health. *Frontiers in Microbiology*, *7*, 185. doi: 10.3389/fmicb.2016.00185
- Roberfroid, M. B. (2005). Introducing inulin-type fructans. *British Journal of Nutrition*, *93*(S1), S13–S25.
- Roberfroid, M., Gibson, G. R., Hoyles, L., McCartney, A. L., Rastall, R., Rowland, I., Wolvers, D., Watzal, B., Szajewska, H., Stahl, B., and Meheust, A. (2010). Prebiotic effects: metabolic and health benefits. *British Journal of Nutrition*, *104*(S2), S1–S63. doi: 10.1017/s0007114510003363
- Robertson, J. A., Ryden, P., Botham, R. L., Reading, S., Gibson, G., and Ring, S. G. (2001). Structural properties of diet-derived polysaccharides and their influence on butyrate production during fermentation. *Lwt - Food Science and Technology*, *34*(8), 567–573. doi:10.1006/fstl.2001.0816
- Rocklin, R. D. (1991). Detection in ion chromatography. *Journal of Chromatography A*, *546*, 175–187.
- Rodrigues, D., Walton, G., Sousa, S., Rocha-Santos, T. A., Duarte, A. C., Freitas, A. C., and Gomes, A. M. (2016). *In vitro* fermentation and prebiotic potential of selected extracts from seaweeds and mushrooms. *Lwt- Food Science and Technology*, *73*, 131–139. doi:10.1016/j.lwt.2016.06.004
- Roediger, W. E. (1980). Role of anaerobic bacteria in the metabolic welfare of the colonic mucosa in man. *Gut*, *21*(9), 793–798.
- Rose, D. J., Venema, K., Keshavarzian, A., and Hamaker, B. R. (2010). Starch-entrapped microspheres show a beneficial fermentation profile and decrease in potentially harmful bacteria during *in vitro* fermentation in

faecal microbiota obtained from patients with inflammatory bowel disease. *British Journal of Nutrition*, 103(10), 1514-1524.

- Ross, A. H. M., Eastwood, M. A., Brydon, W. G., Busuttill, A., McKay, L. F., and Anderson, D. M. (1984). A study of the effects of dietary gum arabic in the rat. *British Journal of Nutrition*, 51(1), 47-56.
- Rossi, M., Corradini, C., Amaretti, A., Nicolini, M., Pompei, A., Zanoni, S., and Matteuzzi, D. (2005). Fermentation of fructooligosaccharides and inulin by *Bifidobacteria*: a comparative study of pure and faecal cultures. *Applied Environmental Microbiology*, 71(10), 6150-6158. doi: 10.1128/aem.71.10.6150-6158.2005
- Rubin, D. C., Shaker, A., and Levin, M. S. (2012). Chronic intestinal inflammation: inflammatory bowel disease and colitis-associated colon cancer. *Frontiers in Immunology*, 3, 107. doi:10.3389/fimmu.2012.00107
- Rumney, C. J. and Rowland, I. R. (1992). *In vivo* and *in vitro* models of the human colonic flora. *Critical Reviews in Food Science and Nutrition*, 31(4), 299-331.
- Russell, W. R., Gratz, S. W., Duncan, S. H., Holtrop, G., Ince, J., Scobbie, L., ... and Duthie, G. G. (2011). High-protein, reduced-carbohydrate weight-loss diets promote metabolite profiles likely to be detrimental to colonic health. *The American Journal of Clinical Nutrition*, 93(5), 1062-1072.
- Russell, W. R., Hoyles, L., Flint, H. J., and Dumas, M. E. (2013). Colonic bacterial metabolites and human health. *Current Opinion in Microbiology*, 16(3), 246-254.
- Ryan, S. M., Fitzgerald, G. F., and van Sinderen, D. (2005). Transcriptional regulation and characterization of a novel  $\beta$ -fructofuranosidase-encoding gene from *Bifidobacterium breve* UCC2003. *Applied Environmental Microbiology*, 71(7), 3475-3482. doi: 10.1128/aem.71.7.3475-3482.2005
- Sadasivam, S. and Manickam, A. (2005). Phenol-sulphuric acid method for total carbohydrate. *Biochemical Methods*.
- Sagay, G. A., Igboanugo, A. B. I., Imarhiagbe, E. O., and Mesike, C. S. (2007). Status of gum Arabic exploitation techniques in Nigeria. *Conference proceedings, Twenty First Annual National Conference of Farm Management Association of Nigeria* (pp. 189–192). College of Agricultural Science, Olabisi Onabanjo University.
- Salazar, N., Binetti, A., Gueimonde, M., Alonso, A., Garrido, P., Del Rey, C. G., González, C., Ruas-Madiedo, P., and Clara, G. (2011). Safety and



intestinal microbiota modulation by the exopolysaccharide-producing strains *Bifidobacterium animalis* IPLA R1 and *Bifidobacterium longum* IPLA E44 orally administered to Wistar rats. *International Journal of Food Microbiology*, 144(3), 342-351.

- Salazar, N., Ruas-Madiedo, P., Kolida, S., Collins, M., Rastall, R., Gibson, G., and Clara, G. (2009). Exopolysaccharides produced by *Bifidobacterium longum* IPLA E44 and *Bifidobacterium animalis* subsp. lactis IPLA R1 modify the composition and metabolic activity of human faecal microbiota in pH-controlled batch cultures. *International Journal of Food Microbiology*, 135(3), 260-267.
- Salminen, S., Bouley, C., Boutron, M. C., Cummings, J. H., Franck, A., Gibson, G. R., ... and Rowland, I. (1998). Functional food science and gastrointestinal physiology and function. *British Journal of Nutrition*, 80(S1), S147-S171.
- Salonen, A., Lahti, L., Salojärvi, J., Holtrop, G., Korpela, K., Duncan, S. H., Date, P., Farquharson, F., Johnstone, A. M., and Lobley, G. E. (2014). Impact of diet and individual variation on intestinal microbiota composition and fermentation products in obese men. *The ISME Journal*, 8(11), 2218.
- Salyers, A. A. (1979). Energy sources of major intestinal fermentative anaerobes. *The American Journal of Clinical Nutrition*, 32(1), 158–163.
- Samuel, N. K., Symon, M. M., and Mary, O. (2018). Preparation and analysis of goat milk mozzarella cheese containing soluble fibre from *Acacia senegal* var. *kerensis*. *African Journal of Food Science*, 12(3), 46–53. doi: 10.5897/ajfs2017.1652
- Sanders, M. E., Lenoir-Wijnkoop, I., Salminen, S., Merenstein, D. J., Gibson, G. R., Petschow, B. W., ... and Pot, B. (2014). Probiotics and prebiotics: prospects for public health and nutritional recommendations. *Annals of the New York Academy of Sciences*, 1309(1), 19–29. doi:10.1111/nyas.12377
- Sanders, M. E., Merenstein, D. J., Reid, G., Gibson, G. R., and Rastall, R. A. (2019). Probiotics and prebiotics in intestinal health and disease: From biology to the clinic. *Nature Reviews Gastroenterology and Hepatology*, 1. <https://doi.org/10.1038/s41575-019-0173-3>
- Sanoja, R. R., Morlon-Guyot, J., Jore, J., Pintado, J., Juge, N., and Guyot, J. P. (2000). Comparative characterization of complete and truncated forms of *Lactobacillus amylovorus*  $\alpha$ -amylase and role of the C-terminal direct repeats in raw-starch binding. *Applied and Environmental Microbiology*, 66(8), 3350–3356.

- Sanz, M. L., Côté, G. L., Gibson, G. R. and Rastall, R. A. (2005). Prebiotic properties of alternansucrase maltose-acceptor oligosaccharides. *Journal of Agricultural and Food Chemistry*, 53(15), 5911-5916.
- Sanz, M. L., Polemis, N., Morales, V., Corzo, N., Drakoularakou, A., Gibson, G. R., and Rastall, R. A. (2005). *In vitro* investigation into the potential prebiotic activity of honey oligosaccharides. *Journal of Agricultural and Food Chemistry*, 53(8), 2914-2921.
- Sarbini, S. R. and Rastall, R. A. (2011). Prebiotics: metabolism, structure, and function. *Functional Food Reviews*, 3(30), 93–106.
- Sarbini, S. R., Kolida, S., Gibson, G. R., and Rastall, R. A. (2013). *In vitro* fermentation of commercial  $\alpha$ -gluco-oligosaccharide by faecal microbiota from lean and obese human subjects. *British Journal of Nutrition*, 109(11), 1980-1989.
- Sarbini, S. R., Kolida, S., Naeye, T., Einerhand, A., Brison, Y., Remaud-Simeon, M., M., Monsan, P., Gibson, G. R., and Rastall, R. A. (2011). *In vitro* fermentation of linear and alpha-1,2-branched dextrans by the human faecal microbiota. *Applied and Environmental Microbiology*, 77(15), 5307–5315. doi:10.1128/AEM.02568-10
- Saulnier, D. M., Molenaar, D., de Vos, W. M., Gibson, G. R., and Kolida, S. (2007). Identification of prebiotic fructooligosaccharide metabolism in *Lactobacillus plantarum* WCFS1 through microarrays. *Applied Environmental Microbiology*, 73(6), 1753-1765. doi: 10.1128/aem.01151-06
- Sayar, S., Jannink, J. L., and White, P. J. (2007). Digestion residues of typical and high- $\beta$ -glucan oat flours provide substrates for *in vitro* fermentation. *Journal of Agricultural and Food Chemistry*, 55(13), 5306-5311.
- Scheppach, W., Luehrs, H., and Menzel, T. (2001). Beneficial health effects of low-digestible carbohydrate consumption. *British Journal of Nutrition*, 85(S1), S23-S30.
- Scher, J. U., Ubeda, C., Artacho, A., Attur, M., Isaac, S., Reddy, S. M., ... and Manasson, J. (2015). Decreased bacterial diversity characterizes the altered gut microbiota in patients with psoriatic arthritis, resembling dysbiosis in inflammatory bowel disease. *Arthritis and Rheumatology*, 67(1), 128-139.
- Schippa, S., Iebba, V., Barbato, M., Di Nardo, G., Totino, V., Checchi, M. P., ... and Conte, M. P. (2010). A distinctive 'microbial signature' in celiac paediatric patients. *BMC Microbiology*, 10(1), 175.

- Scott, D. A., Dodd, L. S., Furihata, J., Tanimoto, S., Keeney, J., Schilling, M. R., and Cowan, E. (2004). An ancient Egyptian cartonnage broad collar - Technical examination of pigments and binding media. *Studies in Conservation*, 49(3), 177-192.
- Scott, D. A., Warmlander, S., Mazurek, J., and Quirke, S. (2009). Examination of some pigments, grounds and media from Egyptian cartonnage fragments in the Petrie Museum, University College London. *Journal of Archaeological Science*, 36(3), 923-932.
- Scott, K. P., Martin, J. C., Duncan, S. H., and Flint, H. J. (2014). Prebiotic stimulation of human colonic butyrate-producing bacteria and *Bifidobacteria*, *in vitro*. *FEMS Microbiology Ecology*, 87(1), 30-40. doi: 10.1111/1574-6941.12186
- Serpa, J., Caiado, F., Carvalho, T., Torre, C., Gonçalves, L. G., Casalou, C., Lamosa, P., Rodrigues, M., Zhu, Z., Lam, E. W., and Dias, S. (2010). Butyrate-rich colonic microenvironment is a relevant selection factor for metabolically adapted tumour cells. *Journal of Biological Chemistry*, 285(50), 39211-39223.
- Shin, J. E. N., Maradufu, A., Marion, J., and Perlin, A. S. (1980). Specificity of  $\alpha$ - and  $\beta$ -D-galactosidase towards analogs of D-galactopyranosides modified at C-4 or C-5. *Carbohydrate Research*, 84(2), 328-335.
- Shin, N. R., Lee, J. C., Lee, H. Y., Kim, M. S., Whon, T. W., Lee, M. S., and Bae, J. W. (2014). An increase in the *Akkermansia spp.* population induced by metformin treatment improves glucose homeostasis in diet-induced obese mice. *Gut*, 63(5), 727-735.
- Shin, W., Wu, A., Massidda, M., Foster, C., Thomas, N., Lee, D. W., ... and Kim, H. J. (2019). A robust longitudinal co-culture of obligate anaerobic gut microbiome with human intestinal epithelium in an anoxic-oxic interface-on-a-chip. *Frontiers in Bioengineering and Biotechnology*, 7, 13.
- Siddig, N., Osman, M., Alassaf, S., Phillips, G., and Williams, P. (2005). Studies on acacia exudate gums, part IV. Distribution of molecular components in relation to. *Food Hydrocolloids*, 19(4), 679-686. doi:10.1016/j.foodhyd.2004.09.005
- Singh, N., Gurav, A., Sivaprakasam, S., Brady, E., Padia, R., Shi, H., ... and Lee, J. R. (2014). Activation of Gpr109a, receptor for niacin and the commensal metabolite butyrate, suppresses colonic inflammation and carcinogenesis. *Immunity*, 40(1), 128-139. doi: 10.1016/j.immuni.2013.12.007
- Sinnott, M. L. and Withers, S. G. (1974). The  $\beta$ -galactosidase-catalysed hydrolyses of  $\beta$ -d-galactopyranosyl pyridinium salts. Rate-limiting

generation of an enzyme-bound galactopyranosyl cation in a process dependent only on aglycone acidity. *Biochemical Journal*, 143(3), 751-762.

- Slavin, J. (2013). Fiber and Prebiotics: Mechanisms and Health Benefits. *Nutrients*, 5(4), 1417–1435. doi:10.3390/nu5041417
- Smerilli, M., Neureiter, M., Wurz, S., Haas, C., Frühauf, S., and Fuchs, W. (2015). Direct fermentation of potato starch and potato residues to lactic acid by *Geobacillus stearothermophilus* under non-sterile conditions. *Journal of Chemical Technology and Biotechnology*, 90(4), 648–657.
- Smith, P. M., Howitt, M. R., Panikov, N., Michaud, M., Gallini, C. A., Bohlooly-Y, M., ... and Garrett, W. S. (2013). The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. *Science*, 341(6145), 569-573. doi: 10.1126/science.1241165
- Sokol, H., Pigneur, B., Watterlot, L., Lakhdari, O., Bermúdez-Humarán, L. G., Gratadoux, J. J., ... and Grangette, C. (2008). *Faecalibacterium prausnitzii* is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients. *Proceedings of the National Academy of Sciences*, 105(43), 16731-16736.
- Sokol, H., Seksik, P., Furet, J. P., Firmesse, O., Nion-Larmurier, I., Beaugerie, L., ... Doré, J. (2009). Low counts of *Faecalibacterium prausnitzii* in colitis microbiota. *Inflammatory Bowel Diseases*, 15(8), 1183–1189.
- Sriram, M. I., Gayathiri, S., Gnanaselvi, U., Jenifer, P. S., Raj, S. M., and Gurunathan, S. (2011). Novel lipopeptide biosurfactant produced by hydrocarbon degrading and heavy metal tolerant bacterium *Escherichia fergusonii* KLU01 as a potential tool for bioremediation. *Bioresource Technology*, 102(19), 9291-9295.
- Stevani, J., Grivet, J. P., Hannequart, G., and Durand, M. (1991). Glucose and lactate catabolism by bacteria of the pig large intestine and sheep rumen as assessed by <sup>13</sup>C nuclear magnetic resonance. *Journal of Applied Bacteriology*, 71(6), 524-530.
- Stevenson, A., Buchanan, C. J., Abia, R. and Eastwood, M. A. (1997). A simple *in vitro* fermentation system for polysaccharides—the effects of fermenter fluid surface area/fluid volume ratio and amount of substrate. *Journal of The Science of Food and Agriculture*, 73(1), 101-105.
- Stone-Dorshow, T. and Levitt, M. D. (1987). Gaseous response to ingestion of a poorly absorbed fructo-oligosaccharide sweetener. *The American Journal of Clinical Nutrition*, 46(1), 61-65.

- Sun, H., Chen, Y., Cheng, M., Zhang, X., Zheng, X., and Zhang, Z. (2018). The modulatory effect of polyphenols from green tea, oolong tea and black tea on human intestinal microbiota *in vitro*. *Journal of Food Science and Technology*, 55(1), 399–407. doi:10.1007/s13197-017-2951-7
- Taha, M. S., El-Sherbiny, E. M., and Osman, H. F. (2020). Anti-ulcerogenic activity of Gum Arabic in gastric mucosal injury induced by ethanol in male albino rats. *Applied Physiology, Nutrition, and Metabolism*, 1-6. doi:10.1139/apnm-2018-0233
- Takagi, R., Sasaki, K., Sasaki, D., Fukuda, I., Tanaka, K., Yoshida, K. I., Kondo, A., and Osawa, R. (2016). A single-batch fermentation system to simulate human colonic microbiota for high-throughput evaluation of prebiotics. *PLoS One*, 11(8). doi: https://doi.org/10.1371/journal.pone.0160533
- Takemura, N., Hagio, M., Ishizuka, S., Ito, H., Morita, T., and Sonoyama, K. (2010). Inulin prolongs survival of intragastrically administered *Lactobacillus plantarum* No. 14 in the gut of mice fed a high-fat diet. *The Journal of Nutrition*, 140(11), 1963-1969. doi: 10.3945/jn.110.128082
- Talib, M. A., Rayis, O. A., Konozy, E. H., and Salih, M. A. (2018). Effect of Gum Arabic (Prebiotic) on Physicochemical and Organoleptic Properties of Yogurt (Probiotic). *Gum Arabic*, 167-171. doi:10.1016/b978-0-12-812002-6.00014-2
- Tanaka, K., Chikara, H., Hu, W., and Hasebe, K. (1999). Separation of carboxylic acids on a weak acidic cation-exchange resin by ion-exclusion chromatography. *Journal of Chromatography A*, 850(1–2), 187–196.
- Tannock, G. W., Wilson, C. M., Loach, D., Cook, G. M., Eason, J., Otoole, P. W., ... Lawley, B. (2011). Resource partitioning in relation to cohabitation of *Lactobacillus* species in the mouse forestomach. *The ISME Journal*, 6(5), 927–938. doi:10.1038/ismej.2011.161
- Teichberg, S., Wingertzahn, M. A., Moyse, J., and Wapnir, R. A. (1999). Effect of gum arabic in an oral rehydration solution on recovery from diarrhoea in rats. *Journal of Pediatric Gastroenterology and Nutrition*, 29(4), 411-417.
- Ten Bruggencate, S. J., Bovee-Oudenhoven, I. M., Lettink-Wissink, M. L., Katan, M. B., and van der Meer, R. (2006). Dietary fructooligosaccharides affect intestinal barrier function in healthy men. *The Journal of Nutrition*, 136(1), 70-74. doi:10.1093/jn/136.1.70
- ter Steege J. C., Forget, P. P., and Buurman, W. A. (1999). Oral spermine administration inhibits nitric oxide-mediated intestinal damage and levels of systemic inflammatory mediators in a mouse endotoxin model. *Shock*

(Augusta, Ga.), 11(2), 115-119. doi: 10.1097/00024382-199902000-00008

- Terpend, K., Possemiers, S., Daguet, D., and Marzorati, M. (2013). Arabinogalactan and fructo-oligosaccharides have a different fermentation profile in the Simulator of the Human Intestinal Microbial Ecosystem (SHIME®). *Environmental Microbiology Reports*, 5(4), 595-603.
- Thapar, N. and Sanderson, I. R. (2004). Diarrhoea in children: an interface between developing and developed countries. *The Lancet*, 363(9409), 641-653.
- Tischer, C. (2002). The free reducing oligosaccharides of gum arabic: Aids for structural assignments in the polysaccharide. *Carbohydrate Polymers*, 47(2), 151-158. doi:10.1016/s0144-8617(01)00173-4
- Tremaroli, V. and Bäckhed, F. (2012). Functional interactions between the gut microbiota and host metabolism. *Nature*, 489(7415), 242-249. doi: 10.1038/nature11552
- Tremaroli, V., Karlsson, F., Werling, M., Ståhlman, M., Kovatcheva-Datchary, P., Olbers, T., ... and Bäckhed, F. (2015). Roux-en-Y gastric bypass and vertical banded gastroplasty induce long-term changes on the human gut microbiome contributing to fat mass regulation. *Cell metabolism*, 22(2), 228-238. doi: 10.1016/j.cmet.2015.07.009
- Triantafyllou, K., Chang, C., and Pimentel, M. (2014). Methanogens, methane and gastrointestinal motility. *Journal of Neurogastroenterology and Motility*, 20(1), 31.
- Trindade, M. I., Abratt, V. R., and Reid, S. J. (2003). Induction of sucrose utilization genes from *Bifidobacterium lactis* by sucrose and raffinose. *Applied and Environmental Microbiology*, 69(1), 24-32. doi:10.1128/aem.69.1.24-32.2003
- Tsujikawa, Y., Nomoto, R., and Osawa, R. (2013). Difference in degradation patterns on inulin-type fructans among strains of *Lactobacillus delbrueckii* and *Lactobacillus paracasei*. *Bioscience of Microbiota, Food and Health*, 32(4), 157-165. doi: 10.12938/bmfh.32.157
- Tulung, B., Rémésy, C., and Demigné, C. (1987). Specific effects of guar gum or gum arabic on adaptation of cecal digestion to high fibre diets in the rat. *The Journal of Nutrition*, 117(9), 1556-1561.
- Tuncil, Y. E., Thakkar, R. D., Marcia, A. D. R., Hamaker, B. R., and Lindemann, S. R. (2018). Divergent short-chain fatty acid production and succession

of colonic microbiota arise in fermentation of variously-sized wheat bran fractions. *Scientific Reports*, 8(1), 16655.

- Tuohy, K. M., Kolida, S., Lustenberger, A. M., and Gibson, G. R. (2001). The prebiotic effects of biscuits containing partially hydrolysed guar gum and fructo-oligosaccharides—a human volunteer study. *British Journal of Nutrition*, 86(3), 341-348.
- Turnbaugh, P. J., Bäckhed, F., Fulton, L., and Gordon, J. I. (2008). Diet-induced obesity is linked to marked but reversible alterations in the mouse distal gut microbiome. *Cell Host and Microbe*, 3(4), 213-223. doi: 10.1016/j.chom.2008.02.015
- Turnbaugh, P. J., Hamady, M., Yatsunencko, T., Cantarel, B. L., Duncan, A., Ley, R. E., ... and Egholm, M. (2009). A core gut microbiome in obese and lean twins. *Nature*, 457(7228), 480-484.
- Turnbaugh, P. J., Ley, R. E., Mahowald, M. A., Magrini, V., Mardis, E. R., and Gordon, J. I. (2006). An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature*, 444(7122), 1027.
- Ummadi, M. and Curic-Bawden, M. (2008). Use of protein hydrolysates in industrial starter culture fermentations. *Protein Hydrolysates in Biotechnology*, 91–114. doi:10.1007/978-1-4020-6674-0\_6
- Ushida, K. and Hoshi, S. (2002). <sup>13</sup>C-NMR studies on lactate metabolism in a porcine gut microbial ecosystem. *Microbial Ecology in Health and Disease*, 14(4), 242-247.
- Valdes, A. M., Walter, J., Segal, E., and Spector, T. D. (2018). Role of the gut microbiota in nutrition and health. *Bmj*, 361, k2179.
- Van Craeyveld, V., Swennen, K., Dornez, E., Van de Wiele, T., Marzorati, M., Verstraete, W., ... and De Ketelaere, B. (2008). Structurally different wheat-derived arabinoxylooligosaccharides have different prebiotic and fermentation properties in rats. *The Journal of Nutrition*, 138(12), 2348-2355.
- Van de Wiele, T., Van den Abbeele, P., Ossieur, W., Possemiers, S., and Marzorati, M. (2015). The simulator of the human intestinal microbial ecosystem (SHIME®). In *The Impact of Food Bioactives on Health* (pp. 305-317). Springer, Cham.
- Van den Abbeele, P., Belzer, C., Goossens, M., Kleerebezem, M., De Vos, W. M., Thas, O., ... and Van de Wiele, T. (2013). Butyrate-producing *Clostridium* cluster XIVa species specifically colonize mucins in an *in vitro* gut model. *The ISME journal*, 7(5), 949-961.

- Van den Abbeele, P., Gérard, P., Rabot, S., Bruneau, A., El Aidy, S., Derrien, M., ... and Van de Wiele, T. (2011). Arabinoxylans and inulin differentially modulate the mucosal and luminal gut microbiota and mucin-degradation in humanized rats. *Environmental Microbiology*, 13(10), 2667-2680.
- Van den Broek, L. A., Hinz, S. W., Beldman, G., Vincken, J.-P., and Voragen, A. G. (2008). *Bifidobacterium* carbohydrases-their role in breakdown and synthesis of (potential) prebiotics. *Molecular Nutrition and Food Research*, 52(1), 146– 163.
- Van Duynhoven, J., Vaughan, E. E., Jacobs, D. M., Kemperman, R. A., Van Velzen, E. J., Gross, G., ... and Westerhuis, J. A. (2011). Metabolic fate of polyphenols in the human superorganism. *Proceedings of The National Academy of Sciences*, 108(Supplement 1), 4531-4538.
- Vanloot, P., Dupuy, N., Guiliano, M., and Artaud, J. (2012). Characterisation and authentication of *A. senegal* and *A. seyal* exudates by infrared spectroscopy and chemometrics. *Food Chemistry*, 135(4), 2554-2560.
- van Zanten, G. C., Knudsen, A., Röytiö, H., Forssten, S., Lawther, M., Blennow, A., ... and Jespersen, L. (2012). The effect of selected synbiotics on microbial composition and short-chain fatty acid production in a model system of the human colon. *PloS one*, 7(10).
- Velikova, P. V., Blagoeva, G. I., Gotcheva, V. G., and Petrova, P. M. (2014). Novel Bulgarian *Lactobacillus* strains ferment prebiotic carbohydrates. *Journal of BioScience and Biotechnology*.
- Vella, M. N., Stratton, L. M., Sheeshka, J., and Duncan, A. M. (2013). Exploration of functional food consumption in older adults in relation to food matrices, bioactive ingredients and health. *Journal of Nutrition in Gerontology and Geriatrics*, 32(2), 122–144.
- Ventura, M., O'Connell-Motherway, M., Leahy, S., Moreno-Munoz, J. A., Fitzgerald, G. F., and van Sinderen, D. (2007). From bacterial genome to functionality; case bifidobacteria. *International Journal of Food Microbiology*, 120(1-2), 2-12. doi: 10.1016/j.ijfoodmicro.2007.06.011
- Verbeken, D., Dierckx, S., and Dewettinck, K. (2003). Exudate gums: occurrence, production, and applications. *Applied Microbiology and Biotechnology*, 63(1), 10–21. doi:10.1007/s00253-003-1354-z
- Vergin, F. (1954). Antibiotics and probiotics. *Hippokrates*, 25(4), 116–119.
- Visek, W. J. (1978). Diet and cell growth modulation by ammonia. *The American Journal of Clinical Nutrition*, 31(10), S216-S220.



- Vital, M., Howe, A. C., and Tiedje, J. M. (2014). Revealing the bacterial butyrate synthesis pathways by analysing (meta) genomic data. *MBio*, 5(2), e00889–14.
- Volpi, E. V. and Bridger, J. M. (2008). FISH glossary: an overview of the fluorescence *in situ* hybridization technique. *BioTechniques*, 45(4), 385–409. doi:10.2144/000112811
- Wagner, M., Horn, M., and Daims, H. (2003). Fluorescence *in situ* hybridisation for the identification and characterisation of prokaryotes. *Current Opinion in Microbiology*, 6(3), 302–309. doi:10.1016/s1369-5274(03)00054-7
- Walker, A. W., Duncan, S. H., Louis, P., and Flint, H. J. (2014). Phylogeny, culturing, and metagenomics of the human gut microbiota. *Trends in Microbiology*, 22(5), 267–274.
- Walker, A. W., Ince, J., Duncan, S. H., Webster, L. M., Holtrop, G., Ze, X., ... and Louis, P. (2011). Dominant and diet-responsive groups of bacteria within the human colonic microbiota. *The ISME journal*, 5(2), 220-230. doi: 10.1038/ismej.2010.118
- Wallenfels, K. and Weil, R. (1972). Beta-galactosidase. In: Boyer, P. (Ed.). *The Enzymes VII*, Academic Press, New York.
- Walter, D. J., Eastwood, M. A., Brydon, W. G., and Elton, R. A. (1988). Fermentation of wheat bran and gum arabic in rats fed on an elemental diet. *British Journal of Nutrition*, 60(2), 225–232. doi:10.1079/bjn19880094
- Walter, J. (2008). Ecological role of lactobacilli in the gastrointestinal tract: implications for fundamental and biomedical research. *Applied and Environmental Microbiology*, 74(16), 4985–4996. doi:10.1128/aem.00753-08
- Walton, G. E. (2006). The bacterial metabolism of dietary prebiotics and the potential for protection against colorectal cancer (Doctoral dissertation), University of Reading.
- Wang, J. F., Zhu, Y. H., Li, D. F., Wang, Z. and Jensen, B. B. (2004). *In vitro* fermentation of various fibre and starch sources by pig faecal inocula. *Journal of Animal Science*, 82(9), 2615-2622.
- Wang, K., Yin, J., Shen, D. and Li, N. (2014). Anaerobic digestion of food waste for volatile fatty acids (VFAs) production with different types of inoculum: effect of pH. *Bioresource Technology*, 161, 395–401.

- Wang, M., Karlsson, C., Olsson, C., Adlerberth, I., Wold, A. E., Strachan, D. P., ... and Coates, A. R. (2008). Reduced diversity in the early faecal microbiota of infants with atopic eczema. *Journal of Allergy and Clinical Immunology*, 121(1), 129-134.
- Wang, S. L., Liang, Y. C., and Liang, T. W. (2011). Purification and characterization of a novel alkali-stable  $\alpha$ -amylase from *Chryseobacterium taeanense* TKU001, and application in antioxidant and prebiotic. *Process Biochemistry*, 46(3), 745-750.
- Wapnir, R. A., Teichberg, S., Go, J. T., Wingertzahn, M. A., and Harper, R. G. (1996). Oral rehydration solutions: enhanced sodium absorption with gum arabic. *Journal of the American College of Nutrition*, 15(4), 377-382.
- Wapnir, R. A., Wingertzahn, M. A., Moyse, J. E. F. F. R. E. Y., and Teichberg, S. A. U. L. (1997). Gum arabic promotes rat jejunal sodium and water absorption from oral rehydration solutions in two models of diarrhoea. *Gastroenterology*, 112(6), 1979-1985.
- Weaver, G. A., Krause, J. A., Miller, T. L., and Wolin, M. J. (1986). Incidence of methanogenic bacteria in a sigmoidoscopy population: an association of methanogenic bacteria and diverticulosis. *Gut*, 27(6), 698-704.
- Weiss, A. T. A., Lübke-Becker, A., Krenz, M., and van der Grinten, E. (2011). Enteritis and septicemia in a horse associated with infection by *Escherichia fergusonii*. *Journal of Equine Veterinary Science*, 31(7), 361-364.
- Willemsen, L. E. M., Koetsier, M. A., Van Deventer, S. J. H., and Van Tol, E. A. F. (2003). Short chain fatty acids stimulate epithelial mucin 2 expression through differential effects on prostaglandin E1 and E2 production by intestinal myofibroblasts. *Gut*, 52(10), 1442-1447. doi: 10.1136/gut.52.10.1442
- Williams, G. R. (1990). The processing of gum arabic to give improved functional properties. *Gums and Stabilisers for the Food Industry*, 5, 37-40.
- Williams, P. A., and Phillips, G. O. (2009). Gum arabic. In *Handbook of Hydrocolloids* (pp. 252-273). Woodhead Publishing.
- Williams, P. A., Phillips, G. O., and Stephen, A. M. (1990). Spectroscopic and molecular comparisons of three fractions from *Acacia senegal* gum. *Food Hydrocolloids*, 4(4), 305-311.
- Wingertzahn, M. A., Teichberg, S., and Wapnir, R. A. (2001). Stimulation of non-sodium-dependent water, electrolyte, and glucose transport in rat small

intestine by gum arabic. *Digestive Diseases and Sciences*, 46(5), 1105-1112.

- Wrapp, P., La Ragione, R. M., Best, A., Reichel, R., Anjum, M. F., Mafura, M., and Woodward, M. J. (2009). Characterisation of *Escherichia fergusonii* isolates from farm animals using an *Escherichia coli* virulence gene array and tissue culture adherence assays. *Research in Veterinary Science*, 86(1), 27-35.
- Wrzosek, L., Miquel, S., Noordine, M. L., Bouet, S., Chevalier-Curt, M. J., Robert, V., ... and Langella, P. (2013). *Bacteroides thetaiotaomicron* and *Faecalibacterium prausnitzii* influence the production of mucus glycans and the development of goblet cells in the colonic epithelium of a gnotobiotic model rodent. *BMC biology*, 11(1), 61. doi: 10.1186/1741-7007-11-61
- Wyatt, G. M., Bayliss, C. E., and Holcroft, J. D. (1986). A change in human faecal flora in response to inclusion of gum arabic in the diet. *British Journal of Nutrition*, 55(2), 261-266.
- Younes, H., Garleb, K., Behr, S., Rémésy, C., and Demigné, C. (1995). Fermentable fibers or oligosaccharides reduce urinary nitrogen excretion by increasing urea disposal in the rat cecum. *The Journal of Nutrition*, 125(4), 1010-1016.
- Zhang, M., Caragine, T., Wang, H., Cohen, P. S., Botchkina, G., Soda, K., ... and Tracey, K. J. (1997). Spermine inhibits proinflammatory cytokine synthesis in human mononuclear cells: a counterregulatory mechanism that restrains the immune response. *The Journal of Experimental Medicine*, 185(10), 1759-1768. doi: 10.1084/jem.185.10.1759
- Zhu, S., Ashok, M., Li, J., Li, W., Yang, H., Wang, P., ... and Wang, H. (2009). Spermine protects mice against lethal sepsis partly by attenuating surrogate inflammatory markers. *Molecular Medicine*, 15(7), 275-282. doi: 10.2119/molmed.2009.00062
- Zhu, Y., Liu, X., and Yang, S. T. (2005). Construction and characterization of pta gene-deleted mutant of *Clostridium tyrobutyricum* for enhanced butyric acid fermentation. *Biotechnology and Bioengineering*, 90(2), 154-166.
- Zigová, J., Šturdík, E., Vandák, D., and Schlosser, Š. (1999). Butyric acid production by *Clostridium butyricum* with integrated extraction and pertraction. *Process Biochemistry*, 34(8), 835-843.

## BIODATA OF STUDENT

The student named Muhamad Hanif bin Rawi (GS44869) was born in Kota Bharu, Kelantan on 9<sup>th</sup> of December 1992. He is currently residing at Kg. Huda Telok Baru, Lundang, 15200 Kota Bharu, Kelantan. He was a former student of UPM Bintulu Sarawak Campus and graduated his bachelor with first class honour in Bachelor of Bioindustry, year 2015. He is currently a PhD candidate of UPM for the degree of Doctor of Philosophy in Food Biotechnology.

