

## YIELD IMPROVEMENT AND GENOTYPE BY ENVIRONMENT INTERACTION OF SELECTED BAMBARA GROUNDNUT GENOTYPES DEVELOPED THROUGH PURELINE SELECTION AND MOLECULAR APPROACHES



MD MAHMUDUL HASAN KHAN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of Philosophy

September 2022

**IPTSM 2022 8** 

# COPYRIGHT

All material contained within the thesis, including, without limitation, text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of the material may only be made with the express prior written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia

 $\mathbf{C}$ 



## DEDICATIONS

This thesis is dedicated to my parents (*Mahmuda Rahman and Md Matiur Rahman Khan*) for their boundless love, care, support, and sacrifice throughout my study life. I also dedicated it to my beloved family (Sarmin Akter, Anamta Manha, & Abdullah-al-Muhtady Khan). Likewise, I would like to devote my efforts to the people of the People's Republics of Bangladesh.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

## YIELD IMPROVEMENT AND GENOTYPE BY ENVIRONMENT INTERACTION OF SELECTED BAMBARA GROUNDNUT GENOTYPES DEVELOPED THROUGH PURELINE SELECTION AND MOLECULAR APPROACHES

By

#### MD MAHMUDUL HASAN KHAN

September 2022

Chairman: Professor Mohd Rafii Yusop, PhD Institute: Tropical Agriculture and Food Security

As a crop for the new millennium, Bambara groundnut (Vigna subterranea [L.] Verdc.) is considered as a "miracle lucrative" legume for Asia and Africa because of its drought resilience, excellent nutritional profiles, and significant versatile uses. Due to minute floral biology and cleistogamous reproductive system, its potential for improvement through hybridization breeding is limited. However, addressing genetic diversity and pure line selection, along with molecular breeding issues are the most imperative and viable methods for its improvement, as a newly introduced legume in Malaysia. The progress of this underexploited legume has been limited owing to a lack of extensive research, marginalization, inadequate knowledge, and readily available research funding, among other concerns. In this point of view, a comprehensive breeding program is carried out. The main objective of this research work was to assess morphological variation considering 14 qualitative and 27 quantitative traits as well as advancing the generation via pure-line selection, genomic DNA analysis using inter-simple sequence repeat (ISSR) markers, genotype by environment (GEI) effects on yield components, and yield stability assessment to discover the superior variety. The plant materials were sourced from the GenBank of the Institute of Tropical Agriculture and Food Security, Universiti Putra Malaysia (UPM), Serdang, Selangor. The field experiment adopted a randomized complete block design (RCBD) with three replications. The recorded data were subjected to analysis of variance, correlation, principal component analysis, clustering pattern, and heatmap analysis, as well as genetic parameters estimation. Analysis of variance revealed significant differences at  $p \le 0.05$  or  $p \le 0.01$  signifying a degree of variability present in evaluated traits for all generations of evaluation (S<sub>0</sub> to  $S_5$ ). The result showed a downward trend in values of coefficient of variation for yield trait as 55.15%, 30.18%, 19.37%, 18.74%, and 17.53% for selfed generation of  $S_0$ ,  $S_1$ ,  $S_2$ ,  $S_3$ , and  $S_4$ , respectively indicating that pure line selection is effective for this legume improvement based on yield trait. Additionally, in all rounds of evaluated generation (So- $S_4$ ), yield component traits such as total number of pods, fresh pod weight (g), hundred seed weight (g), dry pod weight (g), dry seed weight (g), and harvest index (%) had a positive and highly significant correlation  $(0.75 \le r \le 1.00; p \le 0.01)$  with yield trait. Selection based on these traits for this legume improvement may lead to higher success due to their estimated high heritability and genetic gain values of Hb  $\ge$  60% and GA  $\ge$ 20%, respectively. In terms of molecular characterization, a set of 32 ISSR markers on forty-four accessions from S4 selfed generation, revealed 97.64% polymorphism with an average calculated polymorphic information content (PIC) was 0.243, Nei's genetic diversity (h = 0.248), and Shannon's information index (I = 0.395), which indicate the presence of a low to moderate level of genetic variability among the accessions. UPMGA phylogenetic tree clustered the entire genotypes into 3 major clusters and 6 sub-clusters that were also proved by admixture (Bayesian model) analysis which assembled genotypes into 3 distinct genetic components (membership likelihood Q > 0.60) based on best  $\Delta K = 3$ . Within the population, estimated variation was higher as 75% compared to among the population of 25% with gene flow of Nm = 0.9229 indicating that higherlevel genetic variation exists within the population. The total variation of 26.51% was revealed by principal component analysis (PCA) based on ISSR data, where PC1 (13.92%) > PC2 (12.59%). The path-coefficient analysis explored that dry pod weight had the highest direct effect on yield trait. GGE biplot represents the variation of the first two principal components, PC1 (94.97%) and PC2 (3.11%) for yield trait. The source of variation namely, genotypes (22.40%), seasons (31.13%), and locations (14.02%) were found to be the most significant causes of yield heterogeneity for overall  $G + E + G \times E$ variation. The GGE biplot and AMMI revealed that the three winning lines S5G1 (MaikP12-18), S5G3 (Maik P6-18), and S5G5 (JataP1-18) appear as superior lines and in addition, based on ideal genotype ranking, line MaikP12-18 (S5G1) was highly stable, with a high mean yield across the environments. Conclusively, these three superior lines namely, S5G1 (MaikP12-18), S5G3 (Maik P6-18), and S5G5 (JataP1-18) are highly recommended for further large-scale evaluation before release as newly developed varieties for commercial cultivation in Malaysia.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

## PENAMBAHBAIKAN HASIL DAN INTERAKSI GENOTIP DENGAN PERSEKITARAN KE ATAS GENOTIP TERPILIH KACANG BAMBARA YANG DIBANGUNKAN MELALUI PENDEKATAN PEMILIHAN TITISAN TULEN DAN MOLEKULAR

Oleh

#### MD MAHMUDUL HASAN KHAN

September 2022

Pengerusi: Profesor Mohd Rafii Yusop, PhD Institut: Pertanian Tropika dan Keselamatan Makanan

Sebagai tanaman untuk alaf baharu, kacang Bambara (Vigna subterranea [L.] Verdc.) dianggap sebagai kekacang "keajaiban lumayan" untuk Asia dan Afrika kerana daya tahan kemarau, profil pemakanan yang sangat baik dan kegunaan serba boleh yang signifikan. Disebabkan oleh biologi bunganya yang kecil dan sistem pembiakan kleistogami, potensinya untuk diperbaiki melalui pembiakbakaan penghidridan adalah terhad. Walau bagaimanapun, bagi menangani isu-isu kepelbagaian genetik dan pemilihan titisan tulen, bersama dengan pembiakbakaan molekular adalah kaedah yang paling penting dan berdaya maju untuk penambahbaikannya, sebagai kekacang yang baru diperkenalkan di Malaysia. Kemajuan kacang Bambara yang kurang dieksploitasi ini adalah terhad disebabkan oleh kekurangan penyelidikan yang meluas, peminggiran dan pengetahuan yang tidak mencukupi serta pembiayaan penyelidikan yang terhad, adalah antara kebimbangan yang wujud. Berdasar pandangan ini, satu program pembiakan yang komprehensif telah dijalankan. Objektif utama penyelidikan ini adalah untuk menilai variasi morfologi 14 ciri kualitatif dan 27 ciri kuantitatif serta memajukan generasi seterusnya melalui pemilihan titisan tulen, analisis DNA genomik menggunakan penanda molekul antara jujukan berulang mudah (ISSR), kesan interkasi genotip dengan persekitaran (GEI) ke atas komponen hasil, dan seterusnya penilaian kestabilan hasil untuk menbangunkan varieti unggul. Bahan tanaman kajian ini diperoleh daripada GenBank Institut Pertanian Tropika dan Sekuriti Makanan (ITAFoS), Universiti Putra Malaysia (UPM), Serdang, Selangor. Eksperimen di lapangan menggunakan reka bentuk blok lengkap rawak (RCBD) dengan tiga replikasi. Data yang direkodkan tertakluk kepada analisis varians, korelasi, analisis komponen utama, corak pengklusteran dan analisis peta haba, serta penganggaran parameter genetik. Analisis varians merekodkan perbezaan ketara pada  $p \le 0.05$  atau  $p \le 0.01$  yang menunjukkan wujud kepelbagaian dalam ciri yang dinilai untuk semua generasi yang dinilai (S<sub>0</sub> hingga  $S_5$ ). Keputusan merekodkan corak penurunan dalam nilai pekali variasi untuk ciri hasil iaitu masing-masing 55.15%, 30.18%, 19.37%, 18.74% dan 17.53% untuk generasi swakacuk S<sub>0</sub>, S<sub>1</sub>, S<sub>2</sub>, S<sub>3</sub> dan S<sub>4</sub>, menunjukkan bahawa pemilihan titisan tulen adalah berkesan untuk pembaikan tanaman ini berdasarkan ciri hasil. Tambahan pula, di dalam semua genarasi yang dinilai (S<sub>0</sub>-S<sub>4</sub>), didapati ciri- ciri komponen hasil seperti jumlah bilangan lenggai, berat lenggai segar (g), berat seratus biji (g), berat lenggai kering (g), berat biji kering (g), dan indeks penuaian (%) mempunyai korelasi positif yang sangat signifikan  $(0.75 \le r \le 1.00; p \le 0.01)$  dengan ciri hasil. Pemilihan berdasarkan ciri-ciri tersebut untuk pembiakbakaan kekacang ini akan memberikan kejayaan yang tinggi kerana mempunyai anggaran nilai kebolehwarisan dan kemajuan genetik tinggi iaitu masingmasing iaitu Hb  $\geq$  60% dan GA  $\geq$  20%. Dari segi pencirian molekul, satu set 32 penanda ISSR keatas empat puluh empat aksesi daripada generasi swa-kacuk S<sub>4</sub>, menunjukkan 97.64% polimorfisme dengan anggaran purata kandungan maklumat polimorfik (PIC) adalah 0.243, kepelbagaian genetik Nei (h = 0.248), dan Shannon indeks (I = 0.395), yang menunjukkan kewujudan tahap kebolehubahan genetik yang rendah hingga sederhana di kalangan aksesi tersebut. Pokok filogenetik UPMGA mengklusterkan keseluruhan genotip kepada 3 kluster utama dan 6 subkluster yang turut disahkan oleh analisis admixture (model Bayesian) yang mengklusterkan genotip kepada 3 komponen genetik yang berbeza (kebarangkalian keahlian Q > 0.60) berdasarkan  $\Delta K = 3$  terbaik. Dalam populasi, anggaran variasi adalah lebih tinggi iaitu 75% berbanding dengan di antara populasi iaitu 25% dengan aliran gen Nm = 0.9229 menunjukkan aras variasi genetik lebih tinggi di dalam populasi ini. Dari analisis komponen utama (PCA) yang berdasarkan data ISSR, didapati jumlah variasi sebanyak 26.51% di mana PC1 (13.92%) > PC2 (12.59%). Analisis pekali laluan mendapati bahawa berat lenggai kering mempunyai kesan langsung tertinggi ke atas ciri hasil. Biplot GGE mewakili variasi dua komponen utama pertama, PC1 (94.97%) dan PC2 (3.11%) untuk ciri hasil. Sumber variasi iaitu, genotip (22.40%), musim (31.13%) dan lokasi (14.02%) didapati menjadi peyumbang heterogeniti hasil yang paling signifikan bagi variasi keseluruhan G + E +  $G \times E$ . Biplot GGE dan AMMI mendapati bahawa tiga titisan pemenang iaitu S5G1 (MaikP12-18), S5G3 (Maik P6-18) dan S5G5 (JataP1-18) muncul sebagai titisan unggul, dan tambahan pula berdasarkan kedudukan genotip ideal, titisan MaikP12-18 (S5G1) adalah sangat stabil, dengan purata hasil yang tinggi merentasi persekitaran. Secara konklusifnya, tiga titisan unggul ini iaitu, S5G1 (MaikP12-18), S5G3 (Maik P6-18), dan S5G5 (JataP1-18) adalah sangat disyorkan untuk penilaian berskala besar selanjutnya sebelum diistiharkan sebagai varieti baru yang dibangunkan untuk penanaman komersial di Malaysia.

#### ACKNOWLEDGEMENTS

In the name of almighty **ALLAH**, the holy and the most merciful to whom I owe the sense of purpose and strength enabled me to conduct this study with which without His blessing and amazing grace, it wouldn't have been a reality.

My honest gratitude goes to the chairman of my supervisory committee **Prof. Dr. Mohd Rafii Yusop**, for his patience, encouragement, tireless understanding, support, benevolence, enthusiasm to help, and guidance throughout the study period and during the analysis. I am deeply indebted to my supervisory committee members, namely **Dr. Shairul Izan Ramlee** and **Dr. Mashitah Jusoh**, for their tireless understanding, support, and constructive criticism during my study. I really enjoyed every of my moment with you all.

I would like to extend my deepest and sincere appreciation to the Ministry of Agriculture (MoA), Bangladesh Agricultural Research Council (BARC- Project of NATP Phase-II), Bangladesh Agricultural Research Institute (BARI) of The People's Republic of Bangladesh. Thankfulness also goes to Universiti Putra Malaysia (UPM), Malaysia who supported my candidature.

I am exceedingly grateful to all family members for their love, motivation, encouragement, and prayers during my absence for this study. My unsurpassed parents, I remain deeply grateful to you for providing me the freedom to pursue my academic dream; I'm truly thankful to you for discomforting your salves to offer me the best. To all my siblings namely Waziha Tasnim Subha, Ahmed Zawad, Anamta Manha, Arham Tahmid Zarif, and Abdulla Muhtady as well as my brother Mr. Md Mahmudur Rahman and sister Saidunnesa Zenith thank you very much for their standing firm for the success of my studies. I am grateful to Mr. Md Matiur Rahman Khan, Mahmuda Rahman, Saleha Majid, Sarmin Akter, ABM Ilias Molla and Bibi Aysha for their especial Du'a, support, and encouragement throughout the study period.

I am equally appreciative to staff of research field and laboratory of ITAFoS, UPM for all the timely response and necessary assistance given to me throughout my experiment. I am grateful to my colleagues and friends, all Prof. Mohd Rafii Yusop students and all staff of the Institute of Tropical Agriculture and Food Security, UPM, thank you very much for making my life as a post-graduate student a delightful experience. Thanks to Dr. Yusuff Oladosu, and Md Al-Mamun for your incredible friendship and all those I couldn't mention who contributed to the success of this study in one way or the other.

Finally, the patience and prayers of my parents, spouse, kids, and loved ones during my absence are highly acknowledged. I thank you for your prayer, support, encouragement, and motivation.

I certify that a Thesis Examination Committee has met on **02 September**, **2022** to conduct the final examination of **Md Mahmudul Hasan Khan** on his thesis entitled "Yield improvement and genotype by environment interaction of selected Bambara groundnut (*Vigna subterranea* L. verdc.) genotypes developed through pureline selection and molecular approaches" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The Committee recommends that the student be awarded the degree of Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

## DATO ABDUL SHUKOR JURAIMI, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

## NOOR AZMI SHAHARUDDIN, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Internal Examiner)

## AHMED OSUMANU HARUNA, PhD

Professor Institute of Ecosystem Science Borneo Universiti Putra Malaysia (Kampus Bintulu Sarawak) (Internal Examiner)

## NAQIB ULLAH KHAN, PhD

Professor Department of Plant Breeding and Genetics The University of Agriculture Peshawar, Pakistan (External Examiner)

## ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 10 November 2022

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

## Mohd Rafii Bin Yusop, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

## Shairul Izan Binti Ramlee, PhD

Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Member)

## Mashitah Binti Jusoh, PhD

Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Member)

## ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 10 November 2022

## **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_

Date: \_\_\_\_

Name and Matric No: Md Mahmudul Hasan Khan

# **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision.
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: Name of Chairman of Supervisory Committee:	Professor Dr. Mohd Rafii Bin Yusop
Signature: Name of Member of Supervisory Committee:	Dr. Shairul Izan Binti Ramlee
Signature: Name of Member of Supervisory Committee:	Dr. Mashitah Binti Jusoh

# TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xvi
LIST OF FIGURES	XX
LIST OF APPENDICES	xxiv
LIST OF ABBREVIATIONS	xxvii

CHAPTER			
1	INTRO	DUCTION	1
	1.1	General introduction	1
	1.2	Problems statement	2
	1.3	Significance of the study	2
	1.4	Research hypothesis	3
	1.5	Research objectives	3
2	LITER	ATURE REVIEW	4
_	2.1	Introductory of Bambara groundnut	4
		2.1.1 Nomenclature and taxonomy	4
		2.1.2 Centre of origin and distribution	5
		2.1.3 Botanical description of Bambara	-
		groundnut	5
		2.1.4 Agronomic attributes of Bambara	6
		groundnut	0
		2.1.5 Bambara groundnut's resiliency to climate change	8
		2.1.6 Suitable soil type for Bambara groundnut	9
		farming	
	2.2	Bambara groundnut's role to mitigate the nutritional	9
		gap	
	2.3	Yield potential and present production scenario	10
	2.4	Genetic diversity and its improvement	13
	2.5	Research and farming of Bambara groundnut in Malaysia	14
	2.6	Concept and genetic background of pure-line selection	15
	2.7	Rank Summation Index (RSI): selection of elite	16
	2.8	Inbreeding depression and its mechanism	16
	2.9	Genetic parameter analysis and heritability	16
	2.10	Clustering pattern and principal component analysis	17
	2.11	Correlation and path coefficient analysis	18
	2.12	Basic concept of molecular marker	19

	2.12.1	Molecular tools: Inter simple sequence	20
		repeat (ISSR)	20
	2.12.2	Molecular marker application for	20
		Bambara groundnut improvement	20
2.13	Genotype b	y Environment interaction (GEI)	21
	2.13.1	Cross and non-cross over interaction	21
2.14	Stability an	alysis	22
	2.14.1	Univariate stability analysis	22
	2.14.2	Multivariate stability analysis	24
	2.14.2.1	Biplot approaches: GGE and AMMI	24
	2.14.2.1.1	Mega environment assessment	24
	2.14.2.1.2	Genotype assessment	25
	2.14.2.1.3	Test environment assessment	25
2.15	AMMI and	GGE biplot application for crop	26
2.15	improveme	nt	20

## **GENETIC ANALYSIS AND PURE-LINE SELECTION OF BAMBARA GROUNDNUT (Vigna subterranea [L.] Verdc.) ACCESSIONS FOR HIGH YIELD REVEALED BY QUALITATIVE AND QUANTITATIVE TRAITS**

27

3.1	Introduc	tion	27
3.2	Material	s and methods	28
	3.2.1	Experimental location	28
	3.2.2	Plant materials	28
	3.2.3	General theme of current research program	20
		for pure line selection	29
	3.2.4	Experimental design and crop management	30
	3.2.4.1	Soil characteristics of research area	30
	3.2.5	Data collection	32
	3.2.6	Statistical analysis	33
	3.2.6.1	Analysis of variance (ANOVA) estimation	33
	3.2.6.2	Correlation matrix analysis	33
	3.2.6.3	Estimation of genetic parameters	33
	3.2.6.4	Multivariate analysis	35
	3.2.6.5	Selection measure: Rank Summation Index	35
		(RSI)	55
	3.2.6.5	Estimation of inbreeding depression (ID%)	35
3.3	Results a	and Discussion	36
3.3.1	Selfed g	eneration zero (S <sub>0</sub> )	36
	3.3.1.1	Assessment of quantitative traits	36
3.3.2	Selfed g	eneration one $(S_1)$	42
	3.3.2.1	Assessment of qualitative traits	42
	3.3.2.2	Assessment of quantitative traits	45
	3.3.2.3	Correlation matrix analysis	48
	3.3.2.4	Estimation of genetic parameters	51
	3.3.2.4.1	Variance components, heritability, and	51
		genetic advance	51
	3.3.2.5	Assessment of clustering pattern	54
	3.3.2.6	Assessment of principal components (PCs)	57

3

xi

	3.3.2.7	Selection based on Rank Summation Index	59
222	Calfad as	(RSI)	<b>C</b> 0
3.3.3	Selled ge	$\frac{1}{2} = \frac{1}{2} \left( \frac{1}{2} \right)$	60
224	5.5.5.1	Assessment of quantitative traits	60
3.3.4	Selfed ge	neration three $(S_3)$	63
	3.3.4.1	Assessment of quantitative traits	63
3.3.5	Selfed ge	neration $S_3 + S_4$ (pool analysis)	66
	3.3.5.1	Assessment of quantitative traits	66
	3.3.5.2	Correlation matrix analysis	72
	3.3.5.3	Estimation of genetic parameters	76
	3.3.5.3.1	Variance components, heritability, and	76
		genetic advance	10
	3.3.5.4	Assessment of clustering pattern	81
	3.3.5.5	Assessment of principal component (PCs)	84
	3.3.5.5.1	PCA biplot analysis	86
	3.3.5.6	Estimation of inbreeding depression (ID%)	89
	3.3.5.7	Selection based on Rank Summation Index	04
		(RSI)	94
3.4	Summary		96
MOLE	CULAR I	DIVERSITY, PHYLOGENETIC	
LINKA	GE AND	ADMIXTURE MAPPING OF	
SELEC	TED BAN	MBARA GROUNDNUT (Vigna	98
subterra	nea [L.] y	verdc.) ACCESSIONS USING ISSR	
MARK	ERS SYS	TEM	
4.1	Introduct	ion	98
4.2	Materials	and Methods	99
	4.2.1	Genetic materials	99
	4.2.2	Genomic DNA extraction and	
		quantification	99
	423	PCR amplification	100
	424	Scoring of ISSR hand	102
	425	Statistical analysis	102
	1.2.5	Genetic diversity banding pattern and	102
	4.2.3.1	frequency analysis	102
	4.2.5.2	Phylogenetic relationship analysis	103
	4.2.5.3	Marker efficiency analysis (MEA)	103
	4.2.5.4	Genetic structure and admixture analysis	104
4.3	Results an	nd Discussion	105
	4.3.1	Quantification of polymorphism by ISSR	
		primers	105
	4.3.2	Marker efficiency analysis (MEA)	110
	4.3.3	Genetic diversity among the population	112
	4.3.4	Banding pattern and heterozygosity	114
	4.3.5	Molecular variance among and within the	
		populations	115
	436	Genetic differentiation and geneflow	
	1.5.0	setimation among a setimations	116
		eshmanon among populations	
	137	Nei's population genetic distance vs	
	4.3.7	Nei's population genetic distance vs.	118
	4.3.7	Nei's population genetic distance vs. PhiPT value analysis	118

G

	4.3.9	Nei's genetic distance among the	120
	4.3.10	Phylogenetic relationship among the	121
		accessions	121
	4.3.11	Heatmap analysis based on ISSR banding profile	122
	4.3.12	Ordination: Principal component analysis (PCA)	124
	4.3.13	PCA biplot analysis: loading of variable and sample	127
	4.3.14	Genetic structure analysis and admixture mapping	129
	4.3.15	Fixation index ( $F_{ST}$ values) analysis	133
4 4	Congrue	ence of morphological and molecular	120
4.4	characte	rization	130
4.5	Summar	У	140
INS	IGHT INTO	GENOTYPE × ENVIRONMENT	
INT	ERACTION	NEFFECTS ON GROWTH AND YIELD	
CO	MPONENTS	S OF SELECTED BAMBARA	142
GR	OUNDNUT	(Vigna subterranea [L.] VERDC.)	
AC	CESSIONS	OVER MULTI-ENVIRONMENTS	
5.1	Introduc	tion	142
5.2	Material	s and Methods	143
	5.2.1	Planting materials	143
	5.2.2	Description of the study environment and	144
		location	144
	5.2.3	Experimental design and crop management	145
	5.2.4	Data collection	145
	5.2.5	Statistical analysis	145
	5.2.5.1	Analysis of variance (ANOVA)	145
	5.2.5.2	Estimation of variance components, heritability, and genetic advance	147
5.3	Results a	and Discussion	148
	5.3.1	Analysis of variance, mean performance,	
		and variance component for individual environment	148
	5.3.1.1	Analysis of variance for vegetative traits	148
	5.3.1.2	Varianace components analysis for vegetetive traits	152
	5.3.1.3	Anlysis of variance for yield and yield contributed traits	157
	5.3.1.4	Varianace components analysis for yield and yield contributed traits	160
5 2	Pooled a	analysis of variance, mean performance, and	140
5.5.	<sup>2</sup> variance	components for two locations two seasons.	108
	5.3.2.1	Combined variance analysis for vegetetive traits	168
	5.3.2.2	Combined variance analysis for yield and yield components	171

(C)

xiii

5.3.3	Estimati	on of correlation matrix between traits	178
	5.3.3.1	Correlation between vegetetive and yield	170
		components	1/8
	5.3.3.2	Correlation between yield and yield	170
		components	1/9
	5.3.3.3	Correlation networking and abundance	183
		(richness) analysis	105
531	Variance	e component analysis for pooled seasons (2	186
5.5.4	location	s 2 seasons)	100
	5.3.4.1	Estimation of phenotypic coefficient of	
		variation (PCV) and genotypic coefficient	186
		of variation (GCV)	
	5.3.4.2	Estimation of heritability	187
5 4	5.3.4.3	Estimation of genetic advance	18/
5.4	Summar	y	190
ратн	-COEFFI	CIENT ANALYSIS AND	
CORR	ELATIO	N STUDY FOR GROWTH AND YIELD	
COM	ONENTS	S OF SELECTED BAMBARA	
GROU	NDNUT	(Vigna subterranea [L.] Verdc.)	191
ACCE	SSIONS	OVER MULTI-ENVIRONMENT	
TRIAI	LS (METs	;)	
6.1	Introduc	tion	191
6.2	Material	s and Methods	192
	6.2.1	Plant materials, experimental design, and	102
		plant husbandry	192
	6.2.2	Data collection	193
	6.2.3	Statistical analysis	193
6.3	Results a	and Discussion	196
	6.3.1	Correlation analysis: genotypic, phenotypic	196
		and environmental correlation	170
	6.3.2	Direct and indirect effects of vegetative	200
	600	traits on pod yield	202
	6.3.3	Two stage relationships estimation	203
	6.3.3.1	Effects of 1 <sup>st</sup> order components association	203
	6222	on 2 <sup>nd</sup> order components	
	0.3.3.2	on pod vield	205
64	Summar	v	206
0.4	Summa	<i>y</i>	200
GENO	TYPE × 1	ENVIRONMENT INTERACTION AND	
STAB	LITY AN	VALYSIS FOR YIELD AND YIELD	
COMI	ONENTS	S OF SELECTED BAMBARA	207
GROU	INDNUT	(Vigna subterranea [L.] Verdc.)	207
GENO	TYPES 1	TESTED UNDER MULTI-	
ENVI	RONMEN	ITS IN MALAYSIA	
7.1	Introduc	tion	207
7.2	Material	s and Methods	209
	7.2.1	Plant materials, experimental design, and	209
		plant husbandry	_0/
	7.2.2	Data collection	209

		7.2.3	Statistical analysis	209
	7.3	Results a	nd Discussion	210
	7.3.1	Biplot pa	ttern for elucidation of multivariate analysis	210
		7.3.1.1	Polygon view of GGE biplot ('which-won- where' pattern)	211
		7.3.1.2	GGE biplot pattern of 'mean vs. stability' analysis and ideal genotype assessment	213
		7.3.1.3	Genotype ranking: best and ideal genotypes assessment	216
		7.3.1.4	'Descriminitiveness vs. Representativeness' pattern of GGE biplot	217
		7.3.1.5	Environment assessment: relatedness among the test environments	221
		7.3.1.6	Additive main effects and multiplicative interaction: AMMI 1	224
		7.3.1.7	Additive main effects and multiplicative interaction: AMMI 2	225
	7.3.2	Evaluatio	on of genotypes	228
		7.3.2.1	Mean performance and comparison of genotypes	228
	7.3.3	Methods	of univariate stability statistics	230
	7.3. <mark>4</mark>	Univaria correlatio	te stability measures by Spearman's rank	235
	7.4	Summary	y	236
	GENE	RAL CON	ICLUSIONS, AND	
8	RECO	MMEND	ATIONS FOR FUTURE RESEARCH	237
	8.1	General	discussion	237
	8.2	General	conclusion	241
	8.3	Recomm	endation for future research	242
REFEREN	CES			243
APPENDIC	APPENDICES		261	
BIODATA	OF STUL	DENT		305
LIST OF P	UBLICA	LIONS		306
LIST OF CONFERENCE AND AWARD RECEIVED 30				307

6

xv

# LIST OF TABLES

Table		Page
3.1	The line-up of Bambara groundnut accessions of $S_0$ selfed generation.	28
3.2	Meteorological data from the research sites across planting seasons.	31
3.3	Twenty-seven quantitative and fourteen qualitative traits measured according to IPGRI, IITA, BAMNET (2000).	32
3.4	Outline of ANOVA table and EMS for individual trail.	33
3.5	Analysis of variance for 27 quantitative traits in $S_0$ generation of Bambara groundnut.	39
3.6	The mean and mean comparison of vegetative traits of $S_0$ generation of Bambara groundnut accessions.	40
3.7	The mean and mean comparison of yield and its contributing traits of $S_0$ generation of Bambara groundnut accessions.	41
3.8	Analysis of variance for 27 quantitative traits in $S_1$ generation of Bambara groundnut.	47
3.9	Pearson's correlation matrix (r) for 16 yield reflecting quantitative traits in $S_1$ selfed generation of Bambara groundnut accessions.	50
3.10	Assessment of variance components, heritability, and genetic advance of $S_1$ selfed generation of Bambara groundnut.	53
3.11	Relative proportion of average grand yield for five clusters revealed by cluster analysis of $S_1$ generation of Bambara groundnut.	57
3.12	Principal component analysis and Shannon–Weaver diversity index for quantitative traits of $S_1$ generation of Bambara groundnut.	58
3.13	Analysis of variance for 27 quantitative traits in $S_2$ generation of Bambara groundnut.	62
3.14	Analysis of variance for 27 quantitative traits in $S_3$ generation of Bambara groundnut.	65
3.15	Mean square and coefficient of variation for 27 traits of $S_3$ and $S_4$ generation revealed by ANOVA.	70
3.16	Mean performance, standard deviation, and range for 27 traits of $S_3$ and $S_4$ generation revealed by ANOVA.	71
3.17	Pearson's correlation matrix (r) for 16 yield reflecting quantitative traits in pool, $S_3$ , and $S_4$ selfed generation of Bambara groundnut accessions.	74

G

3.18	Assessment of variance components, heritability, and genetic advance of pool, $S_3$ and $S_4$ selfed generation of Bambara groundnut.	79
3.19	Relative proportion of average grand yield for four groups of $S_4$ generation based on clustering pattern revealed by NTSYS program.	82
3.20	Principal component analysis and Shannon–Weaver diversity index for quantitative traits of $S_4$ generation of Bambara groundnut.	85
3.21	Estimates of inbreeding depression (ID%) considering $S_3$ and $S_4$ selfed generations for major yield contributing traits.	91
3.22	The rank summation index (RSI) scores of characteristics that are positively strong and significantly linked with yield per hectare based on highest to lowest ranking order.	95
4.1	List of the selected thirty-two ISSR primers used in this study and their properties such as sequences, base pair, GC content, melting, and annealing temperature.	101
4.2	The details banding profile and polymorphism revealed by 32 ISSR primers of 44 <i>V. subterranea</i> accessions.	107
4.3	Efficacy of primer polymorphism calculated with iMEC of Bambara groundnut genotypes.	112
4.4	Genetic diversity parameters within 11 populations over the loci of <i>V. subteranea</i> species	113
4.5	Estimated banding pattern across eleven <i>V. subterranea</i> populations using 32 ISSR assay.	114
4.6	Analysis of molecular variance (AMOVA) among and within populations	116
4.7	Nei's genetic differentiation analysis of 11 Bambara nut populations.	118
4.8	Nei's genetic distance (above diagonal) and pairwise PhiPT values (below diagonal) among 11 populations.	118
4.9	Estimation of eigenvalues, and percentage of total variation contributed by principal component (PCs).	125
4.10	Allele-frequency divergence among pops (Net nucleotide distance), computed using point estimates of 'P' through Structure analysis.	134
4.11	Genotypes and variables differentiation based molecular and morphological clustering pattern.	139
5.1	The listed of 30 selected Bambara groundnut accession used in this study.	143
5.2	Environmental description of the experimental site.	144
5.3	Selective physio-chemical soil properties of the experimental site.	144

	5.4	Outline of ANOVA table and EMS for individual environment.	145
	5.5	Outline of ANOVA table and EMS for genotype by environments for individual trait.	146
	5.6	Outline of ANOVA table and EMS for pooled locations and seasons.	147
	5.7	Mean performance for vegetative traits of individual environment.	153
	5.8	Mean square ANOVA and genetic components estimation for DTE, D50%F, and DTM in an individual environment.	154
	5.9	Mean square ANOVA and genetic components estimation for PH, NB, and NS in an individual environment.	154
	5.10	Mean square ANOVA and genetic components estimation for NP, NL, and NNS in an individual environment.	155
	5.11	Mean square ANOVA and genetic components estimation for IL, BFW, and BDW in an individual environment.	155
	5.12	Mean square ANOVA and genetic variance component of genotype by environment interaction for vegetative traits	156
	5.13	Mean square ANOVA and genetic components estimation for TNP, NMP, and NIP in an individual environment.	164
	5.14	Mean square ANOVA and genetic components estimation for FPW, DPW, and PL in an individual environment.	164
	5.15	Mean square ANOVA and genetic components estimation for PW, NSP, and DSW in an individual environment.	165
	5.16	Mean square ANOVA and genetic components estimation for SL, SW, and HSW in an individual environment.	165
	5.17	Mean square ANOVA and genetic components estimation for Shell%, HI, & Yield in an individual environment.	166
	5.18	Mean square ANOVA and genetic variance component of genotype by environment interaction for yield traits.	167
	5.19	Mean performance for yield traits of individual environment.	168
	5.20	A pooled ANOVA for vegetative traits (DTE, D50%F, DTM, and PH) across the two seasons of two locations.	175
	5.21	A pooled ANOVA for vegetative traits (NB, NS, NP, and NL) across the two seasons of two locations.	175
	5.22	A pooled ANOVA for vegetative traits (NNS, IL, BFW, and BDW) across the two seasons of two locations.	176
	5.23	A pooled ANOVA for yield traits (TNP, NMP. NIP, FPW and DPW) across the two seasons of two locations.	176

5.24	A pooled ANOVA for yield traits (PL, PW, NSP, DSW, and SL) across the two seasons of two locations.	177
5.25	A pooled ANOVA for yield traits (SW, HSW, Shell%, HI, and Yiled) across the two seasons of two locations.	177
5.26	The Pearson's corelation coefficient (r) among the vegetative components across the two seasons two locations.	181
5.27	The Pearson's corelation coefficient (r) among the vegetative and yield components across the two seasons two locations.	181
5.28	Estimation of variance components, heritability, and genetic advance for 27 quantitative traits in two seasons two locations.	189
6.1	Phenotypic (above diagonal) and genotypic (bellow diagonal) correlation among 12 traits of <i>V. subterranea</i> genotypes.	199
6.2	Estimation of environmental correlation among 12 characters of V. <i>subterranea</i> genotypes.	199
6.3	Direct (diagonal) and indirect effects of 11 related traits on pod yield.	201
6.4	First order and second-order components relationship.	204
6.5	Effects of second-order components on yield per hectare.	205
7.1	Mean performance and their comparisons of <i>V. subterranea</i> genotypes for yield and its contributing traits over four environments (2 seasons 2 locations).	229
7.2	Mean and stability statistics for TNP and FPW of 30 Bambara nut genotypes over four environments (two seasons two locations).	233
7.3	Mean and stability statistics for HSW & Yield of 30 Bambara nut genotypes over four environments (two seasons two locations).	234
7.4	Spearman rank correlations among trait mean and univariate stability parameters for 30 Bambara groundnut genotypes tested in four environments.	235

 $\bigcirc$ 

# LIST OF FIGURES

Figure		Page
2.1	A typical Bambara groundnut seedling and plants with fresh pods.	6
2.2	A typical growth and development stage of Bambara groundnut.	7
2.3	Top six Bambara groundnut producing countries.	12
2.4	World Bambara groundnut producing countries and their contribution.	12
2.5	General steps for breeding through the pure-line selection based on Johannsen, (1903).	15
2.6	A basic path diagram with factor coefficients on the effect of yield components on yield	19
2.7	Interactions between genotypes and environments in terms of altering mean performance across contexts: (A) Additive model, (B) Divergence, (c) Convergence, and (D) Cross-over interaction.	22
3.1	Breeding scheme of evaluation as well as advancing of $S_1$ to $S_5$ selfed generation of Bambara groundnut.	29
3.2	The map of the study site at Universiti Putra Malaysia.	31
3.3	Graphical relationship of dry seed weight (DSW) and hundred seed weight (HSW) with yield (kgha <sup>-1</sup> ) for $S_0$ generation of Bambara groundnut.	37
3.4	Different morphotypes of Bambara groundnut landraces.	43
3.5	Some qualitative features of Bambara groundnut seeds.	43
3.6	Summary of the frequency distribution of fourteen qualitative traits of Bambara groundnut accessions.	44
3.7	Some vegetative variation of Bambara groundnut: a-c) first stem colour; d-f) growth habit; g-i) terminal leaflet shape.	44
3.8	Cluster analysis revealed a dendrogram for 150 Bambara groundnut accessions of $S_1$ generation based on the UPGMA method of SAHN clustering.	56
3.9	Graphical visualization of best 30 genotypes of $S_4$ generation based on yield and its strongly correlated traits such as dry pod weight (DPW) and hundred seed weight (HSW).	68
3.10	Clustering pattern for 44 Bambara groundnut accessions of S4 generation using UPMGA based on 27 morphological features.	83

 $\bigcirc$ 

3.11	Two-dimensional (2D) graph showing the relationship among Bambara groundnut accessions of $S_4$ generation using PCA revealed by NTSYS program.	87
3.12	PCA biplot represents the loading of 44 accessions and 27 variables of S4 generation of Bambara groundnut.	88
4.1	Inter Simple Sequence (ISSR) banding profiles of 44 <i>V. subterranea</i> accessions. The PCR product of a) the UBC 873 primer; b) ISSR 11 primer, and c) UBC 807 amplified with Agarose gel electrophoresis.	108
4.2	Inter Simple Sequence (ISSR) banding profiles of 44 <i>V. subterranea</i> accessions. The PCR product of a) UBC 808 primer, b) UBC 809, and b) UBC 841 primer amplified with agarose gel electrophoresis.	109
4.3	Scatter matrix plot showing relationship a) MI vs PIC, b) RP vs PIC, c) PIC vs EMR, and d) MI vs RP generated by ISSR assay.	111
4.4	Graphical display of banding pattern and heterozygosity of 11 Bambara groundnut populations.	115
4.5	Fractions of molecular variance regarding subdivided population.	116
4.6	The relationship of 11 V. subterranea populations based on Nei's original genetic measures revealed by UPMGA method.	119
4.7	The UPMGA phylogenetic tree illustrates the genetic inconsistency and visual relationship among <i>V. suberranea</i> genotypes based on Nei's genetic distance.	123
4.8	Heatmap plot based on ISSR binary data of 44 <i>V. subterranea</i> accessions constructed by Euclidian distance with Ward (unsquared distances) linkage method.	124
4.9	PCA (Euclidian's measure) case scores using the NTSYSpc program. (A). PCA two-dimensional display of genotypes; (B). three-dimensional (3D) graphical display of 44 genotypes generated by using NCSS 2021 based on ISSR data set.	126
4.10	PCA score plot (sample loading) (A); PCA variables loading (loci) (B); and PCA biplot representing the loci (red vector) and sample (accessions) loading (C) based on ISSR markers using JMP ver. 16 programs.	128
4.11	The population membership of the studied Bambara groundnut species group for a priori distinct number of $K = 1-10$ inferred by the STRUCTURE software (PRITCHARD LAB, CA, USA).	131
4.12	Structure harvester and Delta K value were elucidated using Evano et al., (2005) method and Bayesian model-based valuation of population structure for 44 <i>V. subterranea</i> accessions based on ISSR markers.	132

4.13	The STRUCTURE software (Pritchard Lab, CA, USA) is a based illustration of fixation index (Fst).	135
4.14	Heatmap and hierarchical clustering (double dendrogram) responses to morphological descriptors of Bambara groundnut landraces constructed using ClustVis tools (https://bio.tools/clustvis).	138
4.15	The result of the Mantel test uses a genetic distance matrix between morphological and molecular basis.	138
5.1	(A) Correlation network illustrates the relationship among traits (that meet p-value $=0.05$ ) of 30 accessions sampled from 11 populations. (B) Pattern search plot insight into the correlation abundance of the top 22 traits of the respective population.	184
5.2	Relative abundance sketching: (A) Stacked bar showing the richness of traits in each population and (B) Pai chart showing the richness percentages with a unique color code for each trait. In the "Stacked bar" each bar represents an individual population.	185
6.1	(A) Correlation cluster (heatmap); (B) Correlation (Spearman's) network depicts the direct relationship of tested characteristics.	198
6.2	Genotypic path coefficient diagram representing cause and effect relationships among quantitative traits and grain yield. Path diagram and coefficients of factors on the influence of first order on the second-order components and the latter on yield per hectare of <i>V. subterranea</i> genotypes.	202
7.1	"Which-won-where" pattern of GGE biplot polygon view displaying the $G+G\times E$ interaction effect of 30 Bambara groundnut genotypes in two seasons two locations for total number of pods (Pattern A), fresh pod weight (Pattern B), hundred seed weight (Pattern C), yield per hectare (Pattern D).	212
7.2	'Mean vs. stability' pattern of GGE biplot illustrating the $G+G\times E$ interaction effect of 30 Bambara groundnut genotypes in two seasons two locations for total number of pods (Pattern A), fresh pod weight (Pattern B), hundred seed weight (Pattern C), yield per hectare (Pattern D).	215
7.3	The GGE biplot 'genotypes ranking' pattern for genotype comparison with ideal genotype showing the $G+G\times E$ interaction effect of 30 Bambara groundnut genotypes in two seasons two locations for total number of pods (Pattern A), fresh pod weight (Pattern B), hundred seed weight (Pattern C), yield per hectare (Pattern D).	218
7.4	The GGE biplot 'Discriminativeness vs. Representativeness' pattern for genotype comparison with ideal genotype showing the $G+G\times E$ interaction effect of 30 Bambara groundnut genotypes in two seasons two locations for total number of pods (Pattern A), fresh pod	219

xxii

weight (Pattern B), hundred seed weight (Pattern C), yield per hectare (Pattern D).

The GGE biplot 'Environment ranking' pattern for environment comparison with ideal environment showing the G+G×E interaction

7.5 effect of 30 Bambara groundnut genotypes in two seasons two locations for total number of pods (Pattern A), fresh pod weight (Pattern B), hundred seed weight (Pattern C), yield per hectare (Pattern D).

The vector view of GGE biplot showing the relationship among environment (tested environment with the ideal environment) of 30

7.6 Bambara groundnut genotypes under two seasons two location for total no. of pods (Pattern A), fresh pod weight (Pattern B), hundred seed weight (Pattern C), yield per hectare (Pattern D).

The biplot 'AMMI 1' illustrated the trait main effect and first principal component (PC1) effects of both genotype and environment of 30 Bambara groundnut genotypes under two seasons two locations for total number of pods (Pattern A), fresh pod weight

7.7 two locations for total number of pods (Pattern A), fresh pod weight (Pattern B), hundred seed weight (Pattern C), yield per hectare (Pattern D).

The biplot 'AMMI 2' illustrated the first two principal components (PC1 and PC2) effects of genotype plus GE interaction effect of 30

7.8 Bambara groundnut genotypes under two seasons two locations for total number of pods (Pattern A), fresh pod weight (Pattern B), hundred seed weight (Pattern C), yield yield per hectare (Pattern D).

220

223

226

227

# LIST OF APPENDICES

А	ppendix		Page
	3.1	Overall breeding scheme of development high yielding variety (HYV) for <i>Vigna subterranea</i> (L) through pure line selection and molecular approach.	261
	3.2	The image of fifteen seed morphotypes used for S0 generation evaluation.	262
	3.3	The line-up of Bambara groundnut accessions used in the evaluation of $S_1$ generation.	263
	3.4	The line-up of Bambara groundnut accessions used in evaluation of $S_2$ generation.	264
	3.5	The line-up of Bambara groundnut accessions used in evaluation of $S_3$ generation.	264
	3.6	The line-up of Bambara groundnut accessions used in evaluation of $S_3 + S_4$ generation.	265
	3.7	Experimental view of the different growing seasons at Ladang 15.	266
	3.8	Experimental plot of pooled $(S3 + S4 \text{ selfed generation})$ study at Ladang 15 and harvesting of Bambara nut and counting of the total number of pods per plant.	266
	3.9	Measuring procedures of recorded qualitative traits based on IPGRI, IITA, BAMNET, (2000).	267
	3.10	Measuring procedures of recorded quantitative traits based on IPGRI, IITA, BAMNET, (2000).	268
	3.11	Measuring some phenotypic traits during the investigation.	334
	3.12	Measuring of (A) pod length (mm) (a-c) and pod width (mm) (d-f) and (B) seed length (mm) (a-c) and seed width (mm) (d-f) of Duna, Maikai, and Giiwa, respectively.	269
	3.13	Variation in color of the mature pod and double seeded pod of some accessions. Some fresh pod and leaves morphological variation:	269
	3.14	Phenotypic variation among selected promising lines at vegetative stage.	270
	3.15	Selected high yielding promising lines during field evaluation of research.	270
	3.16	Categorization of 150 Bambara groundnut accessions into different units by morphotypes.	271

3.17	Accessions, their traits mean, rank, and rank summation index (RSI) of the Bambara groundnut.	272
3.18	The means and mean comparison of 27 traits of $S_2$ generation of Bambara groundnut accessions.	275
3.19	The mean and mean comparison of 27 traits of $S_3$ generation of Bambara groundnut accessions.	277
3.20	The mean and mean comparison of 27 traits of $S_4$ generation of Bambara groundnut accessions.	279
3.21	Estimates of inbreeding depression (ID%) considering $S_3$ and $S_4$ selfed generations.	281
4.1	List of Bambara groundnut accessions, code, and population name.	283
4.2	Preparation of reagents for DNA extraction and details of DNA isolation protocol (Zheng, 1995) as well as DNA quantification.	283
4.3	Forty-four Bambara groundnut genomic DNA dilution given nanodrop spectrophotometric nucleic acid concentration and quality.	285
4.4	(A) Normal curves of Nanodrop spectrophotometry result from extracted genomic DNAs of the 44 selected Bambara groundnut lines. (B) Some activities of molecular study.	286
4.5	List of the 96 ISSR primers tested for quantification of polymorphism on 44 accessions of Bambara groundnut.	287
4.6	The full-length gels: a) UBC 873 primer; b) ISSR 11 primer, and c) UBC 807 amplified with Agarose gel electrophoresis.	289
4.7	The full-length gels: a) UBC 808, b) UBC 809, and c) UBC 841 primer amplified with Agarose gel electrophoresis.	289
4.8	Molecular genetic distance (Nei's original measures below diagonal) and morphological genetic distance (above diagonal) for 44 <i>V. subterranea</i> genotypes.	290
4.9	Estimation of Shannon diversity (H') index, eigenvalues, eigenvectors, and percentage of total variation contributed by principal component (PCs).	292
5.1	(A) Experimental plot for $G \times E$ interaction assessment of season one and season two at Ladang 10. (B) Experimental plot for $G \times E$ interaction assessment of season one and season two at Ladang 15.	293
5.2	Mean performance and comparison for DTE, D50%F, and DTM of Bambara groundnut genotypes tested in four environments.	294

5.3	Mean performance and comparison for PH, NB, and NS of Bambara groundnut genotypes tested in four environments.	295
5.4	Mean performance and comparison for NP, NL, and NNS of Bambara groundnut genotypes tested in four environments.	296
5.5	Mean performance and comparison for IL, BFW, and BDW of Bambara groundnut genotypes tested in four environments	297
5.6	Mean performance and comparison for TNP, NMP, and NIP of Bambara groundnut genotypes tested in four environments.	298
5.7	Mean performance and comparison for FPW, DPW, and PL of Bambara groundnut genotypes tested in four environments.	299
5.8	Mean performance and comparison for PW, NSP, and DSW of Bambara groundnut genotypes tested in four environments.	300
5.9	Mean performance and comparison for SL, SW, and HSW of Bambara groundnut genotypes tested in four environments.	301
5.10	Mean performance and comparison for Shell%, HI, and YLD of Bambara groundnut genotypes tested in four environments.	302
5.11	Mean comparison for vegetative traits of 30 Bambara groundnut genotypes tested in two seasons two locations.	303
5.12	Mean comparison for yield traits of 30 Bambara groundnut genotypes tested in two seasons across two locations.	304

 $\bigcirc$ 

# LIST OF ABBREVIATIONS

AEC	Average environment coordinate			
AMMI	Additive main effect and multiplicative interaction effect			
ANOVA	Analysis of variance			
AOCC	African Orphan Crops Consortium			
AMOVA	Analysis of molecular variance			
AFLP	Amplified Fragment Length Polymorphisms			
bi	Regression slope			
bp	Base pairs			
CCFF	Center for Crops for the Future			
CV	Coefficient of variation			
СТАВ	Cetyltrimethylammonium bromide (C19H42BrN)			
cm	Centimetre			
DARWin	Dissimilarity Analysis and Representation for windows			
DNA	Deoxyribonucleic acid			
dNTP	Deoxy nucleoside triphosphate			
DNMRT	Duncan's New Multiple Range Test			
EDTA	Ethylenediaminetetraacetic acid $(C_{10}H_{16}N_2O_8)$			
EMS	Expected mean square			
Fst	Fixation index			
GD	Genetic distance			
Gst	Gene differentiation			
GV & PV	Genotypic variance and phenotypic variance			
GA	Genetic advance			
GCV	Genotypic coefficient of variation			
G×E	Genotype $\times$ environmental interaction			

	GGE	Genotype main effects plus genotype by environmental interaction
	GGL	Genotype plus genotype by location interaction
	g	Gram
	h	Nei's (1973) gene diversity
	h <sup>2</sup> b	Broad sense heritability
	ha	Hectare
	H′	Shannon diversity index
	ISSR	Inter simple sequence repeat
	IITA	International Institute of Tropical Agriculture
	IPGRI	International Plant Genetic Resources Institute
	kg	Kilogram
	LSD	Least significant difference
	mm	Millimetre
	Mg	Milligram
	MVSP	Multivariate Statistical Package
	$MS_{\rm E}$	Mean square of error
	$MS_G$	Mean square of genotype
	METs	Multiple environment trials
	Min	Minute
	NTSYS	Numerical Taxonomy and Multivariate Analysis System
	n <sub>e</sub>	Effective number of alleles
	Nm	Gene flow or Genetic drift
	Na	Number of alleles
	NCSS	Number Cruncher Statistical Systems
	р	Probability
	PCV	Phenotypic coefficient of variation
	PC	Principal component

PCA	Principal	component	analysis
-----	-----------	-----------	----------

PopGene Population genetic analysis

- PCR Polymerase chain reaction
- REML Restricted Maximum Likelihood
- RAPD Random Amplified Polymorphic DNA
- RFLP Restriction fragment length polymorphism
- r Correlation of variation
- S<sup>2</sup>d Deviation from regression
- SSR Simple sequence repeat
- SAS Statistical analysis software
- SASG×E SAS genotype × environmental interaction analysis program
- SVD Singular value decomposition
- SVP Singular value partition
- SVP Singular value portioning
- StD Standard deviation
- SE Standard error
- SAS Statistical Analysis System
- TaqThermos aquatics
- TBE Tris EDTA buffer
- Tm Melting temperature
- Ta Annealing temperature
- UV Ultraviolet
- UPMGA Unweighted pair group method with arithmetic mean
- VERCOM Variance component
- WHC Ward hierarchal clustering
- Wi Wricke's ecovalence
- YSi Kang's yield stability statistics

- βi Perkins and Jinks beta
- σi2 Shukla's variance
- µg Microgram
- µl Microliter
- μm Micrometre
- $\sum$ RSI Rank summation index
- % Percentages
- °C Degree Celsius

#### **CHAPTER 1**

#### INTRODUCTION

#### 1.1 General introduction

Bambara groundnut (Vigna subterranea [L.] Verdc. Syn. Voandzeia subterranea [L.] Thouars ex DC. 2n = 2x = 22) belongs to an important taxon: the genus Vigna, under the member of Fabaceae or Leguminosae family and Faboideae subfamily (Heller et al., 1997). Bambara groundnut is widely cultivated as a subsistence crop, however, after peanut (Arachis hypogaea) and cowpea (Vigna unguiculata) it becomes Africa's third most prevalent legume in terms of consumption and production (Obidiebube et al., 2020). The crop contributes to increasing soil fertility profile by fixing atmospheric nitrogen and can withstand drought as well as thrive in poor soils where other legumes fail to survive (Minnaar-Ontong et al., 2021). Even though it is a consistent source of food for low-income people and sources of earning by selling excess Bambara groundnut regularly in local markets, as well as offering a good source of essential nutrients (Olanrewaju et al., 2021a). Consumption of agri-based food reduces mortality, resulting in coronary heart diseases so, Bambara groundnut can be an agri-based protein source for marginal people who are unaffordable to precious animal protein (Oludare et al., 2017). Bambara groundnut is a hardy crop that has been noted as a lucrative and nutritive food source when other food is under threat (Mbosso et al., 2020). Due to balanced nutritional profiles viz. carbohydrates (64.4%), protein (23.6%), oil (6.5%), fibre (5.5%), and a significant amount of minerals it is remarked as a "Complete Food" having the potentiality to mitigate nutritional gaps and ensure food safety (Lin Tan et al., 2020).

The main drawback of expansion of this legume cultivation is limited yield potential, which can be overcome by neglecting native cultivars with replacing the high-yielding modern cultivars through comprehensive research (Obidiebube et al., 2020). Evidence on the genetic relationship of a crop considering molecular and morphological markers is imperative for effective breeding schemes (Fatimah and Ardiarini, 2018). In literature, negligible evidence was found to use of ISSRs marker in Bambara groundnut. To identify genetically similar lines, ISSRs have potential by generating very repeatable and authentic bands on evaluated genotypes (Nilkanta et al., 2017). A set of stability statistical measures namely biplots (AMMI and GGE) are widely executed in different crops to graphically show the interrelationships between genotypes (G), environments (Olanrewaju et al., 2021b). Several studies, including Olanrewaju et al., (2021b) and Mogale, (2018), have successfully established GGE biplots-based genotype assessment to assess yield stability.

However, as a new crop in Malaysia, a gap in research at the genomic level, information on the crop's botany, modern farming techniques, economic value addition, and diversity of *V. subterranea* prompted the inauguration of this research.

## 1.2 Problems statement

Bambara groundnut is a highly nutritious and potentially lucrative crop in Malaysia, having the potential to provide a significant impact on world food and nutritional security (Belel, 2018). Globally, this under-researched legume received little attention by researchers and policymakers compared to the other pulses and legumes for its ultimate improvement (Muhammad et al., 2020). Despite its great yield potential, its existing yield is unimpressive, owing to a lack of systematic genetic improvement (Majola et al., 2021). The main point of low yield is the use of local landraces in addition to little or no interest in its genetic improvement, lack of improved varieties, traditional production technologies, lack of effective research and resources, less research by scientific personnel (Mohammed, 2019).

Due to cleistogamous floral behaviour, numerous attempts at hybridization have failed horribly, with fewer than 2% success (Muhammad et al., 2020). No highyielding cultivars have been released yet in Malaysia. However, this legume is successfully grown in some pockets area of Sik district Kedah, Malaysia using local cultivars. Little is known about the relationship between yield and its aiding traits, though, it was definitely insufficient to assist breeders in selecting successful cultivars. Despite the fact that numerous attempts have been made on pure line selection, molecular characterization, and GEI assessment in various legumes, however, a little or no work has been executed on this underexploited legume in Malaysia. Therefore, it is imperative to achieve genotypes that displayed outstanding performance in grain yield that can be used as breeding lines for the genetic improvement of the Bambara groundnut.

## 1.3 Significance of the study

The ever-increasing demand for food and nutrient consumption justifies the need to increase current yield by introducing high-yielding cultivars. So, this study focuses on the genetic divergence in various qualitative and quantitative variables to investigate the diversity across the genotypes and variables, subject to the selection of elite homogenous lines for future breeding programs with a specific goal of high yield. This research will be a significant endeavour toward promoting the Bambara groundnut molecular breeding strategy in Malaysia. Moreover, the current study will provide an idea of the efficiency of different stability analysis methods for selecting high-yielding superior and stable genotypes over a wide range of environments. Additionally, a better understanding of their morphological and molecular genetic relationship, as well as G×E interaction effects assessment will be beneficial for judicial utilization, maintenance, conservation, application of limited genetic resources.

2

## **1.4** Research hypothesis

The study hypothesised that, the research findings will deliver significantly unique *V*. *subterranea* genetic materials that can be announced as superior variety for large-scale commercial farming in different agroecological zone of Malaysia.

## 1.5 Research objectives

## Main objective:

To develop pure-lines and high-yielding superior stable genotypes of Bambara groundnut for commercial cultivation in Malaysia.

## **Specific objectives:**

- i. To evaluate genetic and agronomic performance as well as advancing the generation for development of high yielding homogenous lines through pure-line selection method.
- ii. To insight into molecular diversity, phylogenetic relationship, and admixture mapping (genetic structure) of selected lines through ISSR markers system.
- iii. To study the genotype  $\times$  environment interaction effect (GEI) on yield and yield components in addition to criteria selection for crop yield improvement.
- iv. To quantify the direct and indirect effects of growth and yield components on crop yield through the degree and nature of traits association as well as path-coefficient analysis.
- v. To identify the high-yielding superior and stable accessions using univariate and multivariate stability statistical approaches over multi-environment trials.

#### REFERENCES

- Abu, H. B., & Buah, S. S. J. (2011). Characterization of Bambara groundnut landraces and their evaluation by farmers in the Upper West Region of Ghana. *Journal of Developments in Sustainable Agriculture*, 6(1), 64-74.
- Ahmad, N. (2012). Bambara groundnut, the crop for the new millennium: Molecular Techniques to Improve the Resiliency of Bambara Groundnut, *Lamber Academic publishers*. Republic of Moldova, Chisinau-2068, str. A.Russo 15, of.61.
- Ahmad, N. S. (2013). Genetic analysis of plant morphology in Bambara groundnut (Vigna subterranea (L.) Verdc.) (*Doctoral dissertation, University of Nottingham*). ID 13150, http://eprints.nottingham.ac.uk/id/eprint/13150
- Ahmed, M. Z., Masoud, I. M., & Zedan, S. Z. (2018). Genetic Diversity and Relationship among Nine Cultivated Flax Genotypes (*Linum usitatissimum* L.) Based on SCoT Markers. *Sciences*, 8(04), 1480-1490.
- Al-Mamun, M., Rafii, M., Oladosu, Y., Misran, A. B., Berahim, Z., Ahmad, Z., Arolu, F. & Khan, M. H. (2020). Genetic diversity among kenaf mutants as revealed by qualitative and quantitative traits. *Journal of Natural Fibers*, 1-18.
- Allard, R.W. (1960). Principles of Plant Breeding. John Wiley and Sons Inc. New York, p. 2245.
- Aliyu, S., Massawe, F., & Mayes, S. (2016). Genetic diversity and population structure of Bambara groundnut (Vigna subterranea (L.) Verdc.): synopsis of the past two decades of analysis and implications for crop improvement programmes. *Genetic Resources and Crop Evolution*, 63(6),925-943. https://doi.org/10.1007/s10722-016-0406-z.
- Aliyu, S., & Massawe, F. J. (2013). Microsatellites based marker molecular analysis of Ghanaian bambara groundnut (Vigna subterranea (L.) Verdc.) landraces alongside morphological characterization. *Genetic Resources and Crop Evolution*, 60(2), 777-787. https://doi.org/10.1007/s1072 2-012-9874-y.
- Aliyu, S., Massawe, F., & Mayes, S. (2014). Beyond landraces: developing improved germplasm resources for underutilized species-a case for Bambara groundnut. *Biotechnology and Genetic Engineering Reviews*, 30(2), 127-141.
- Aliyu, S. H. (2017). "Soil genesis, classification and soil-crop suitability of Serdang and Malacca soil series in a Malaysian public university", Masters thesis, Universiti Putra Malaysia. <u>http://psasir.upm.edu.my/id/eprint/70320</u>
- Alansi, S., Tarroum, M., Al-Qurainy, F., Khan, S., & Nadeem, M. (2016). Use of ISSR markers to assess the genetic diversity in wild medicinal Ziziphus spina-christi (L.)
  Willd. collected from different regions of Saudi Arabia. *Biotechnology & Biotechnological Equipment*, 30(5), 942-947.

- Allendorf, F. W., Luikart, G., & Aitken, S. N. (2007). Conservation and the genetics of populations. *Mammalia*, 2007(2007), 189-197.
- Alkan, B. B., Atakan, C., & Akdi, Y. (2015). Visual analysis using biplot techniques of rainfall changes over Turkey. *Mapan*, *30*(1), 25-30.
- Alake, O. C., & Ayo-Vaughan, M. A. (2017). Genotypic variation and correlations between yield system traits and yield components in African landraces of bambara groundnut. *South African Journal of Plant and Soil*, 34(2), 125-137.
- Amadou, H. I., Bebeli, P. J., & Kaltsikes, P. J. (2001). Genetic diversity in Bambara groundnut (*Vigna subterranea* L.) germplasm revealed by RAPD markers. *Genome*, 44(6), 995-999.
- Aman, J., Bantte, K., Alamerew, S., & Sbhatu, D. B. (2020). Correlation and path coefficient analysis of yield and yield components of quality protein maize (*Zea mays* L.) hybrids at Jimma, western Ethiopia. *International Journal of Agronomy*, 2020. https://doi.org/10.1155/2020/9651537
- Ambrose, S. F. (2015). Genotype by environment interaction in groundnut genotypes for yield and other agronomic traits in two locations in Ghana (*Doctoral dissertation*).
- Amiryousefi, A., Hyvönen, J., & Poczai, P. (2018). iMEC: online marker efficiency calculator. *Applications in Plant Sciences*, 6(6), e01159.
- Araújo, L. B. R., Fiege, L. B. C., Silva, A. B. A., & Bertini, C. H. C. M. (2019). Genetic diversity in cowpea landraces analyzed by ISSR markers. *Genetics and Molecular Research*, 18(1): gmr18082.
- Arolu, I. W., Rafii, M. Y., Hanafi, M. M., Mahmud, T. M. M., & Latif, M. A. (2012). Molecular characterization of 'Jatropha curcas' germplasm using inter simple sequence repeat (ISSR) markers in Peninsular Malaysia. Australian Journal of Crop Science, 6(12), 1666-1673.
- Asibuo, J. Y., Forpoh, A. S., & Akromah, R. (2018). Genotype X environment interactions of groundnut (*Arachis hypogaea* L.) for pod yield. *Ecological Genetics* and Genomics, 7, 27-32.
- Asra, R., Syamsuardi, S., Mansyurdin, M., & Witono, J. R. (2014). The study of genetic diversity of Daemonorops draco (Palmae) using ISSR markers. *BIODIVERSITAS Journal of Biological Diversity*, 15(2).
- Atoyebi, J. O., Osilesi, O., Abberton, M. T., Adebawo, O., & Oyatomi, O. (2018). Quantification of selected anti-nutrients and bioactive compounds in African bambara groundnut (Vigna subterranea (L.) Verdc.). American Journal of Food and Nutrition. 6, 88–95.
- Azam-Ali, S. N., Sesay, A., Karikari, S. K., Massawe, F. J., Aguilar-Manjarrez, J., Bannayan, M., & Hampson, K. J. (2001). Assessing the potential of an underutilized crop–a case study using bambara groundnut. *Experimental Agriculture*, 37(4), 433-472.

- Azman, R. (2016). Growing Bambara Groundnut in Malaysia, bamyield programme manager, bamyield project update: nutritional profile of bambara groundnut and its potential for food product development in Malaysia. (http://www.cffresearch.orgs).
- Bamshaiye, O. M., Adegbola, J. A., & Bamishaiye, E. I. (2011). Bambara groundnut: an under-utilized nut in Africa. *Advances in Agricultural Biotechnology*, 1(1), 60-72.
- Bamnetwork. 2017, Bambara Groundnut. Crops. URL: http://www. Bambaragroundnut.org. (Accessed 5th January 2019).
- Basu, S., Roberts, J. A., Azam-Ali, S. N., & Mayes, S. (2007). Development of microsatellite markers for bambara groundnut (Vigna subterranea L. Verdc.)—an underutilized African legume crop species. *Molecular Ecology Notes*, 7(6), 1326-1328.
- Baye, T. M., Abebe, T., & Wilke, R. A. (2011). Genotype–environment interactions and their translational implications. *Personalized Medicine*, 8: 59–70.
- Becker, H.C., & Leon, J. 1988. Stability analysis in plant breeding. *Plant Breeding*. 101:1–23.
- Belel, M. D. (2018). Contributions of Growth and Physiological Traits to Yield of Seven Bambara nut Landraces Grown Under the Tropical Humid Climate of Malaysia. *International Journal of Agriculture Innovations and Research*, 7(1), 109–116.
- Belel, M. D., Halim, R. A., Rafii, M. Y., & Saud, H. M. (2014). Intercropping of Corn With Some Selected Legumes for Improved Forage Production : A Review. *Journal* of Agricultural Science, 6(3), 48–62.
- Beket, S. Ã., Dagou, S. E. K. A., Adjoumani, K., & SylvÃ, R. (2019). Evaluation of the diversity in qualitative traits of Bambara groundnut germplasm (*Vigna subterranea* (L.) Verdc.) of Cte dIvoire. *African Journal of Biotechnology*, 18(1), 23-36.
- Bonny, B. S., Adjoumani, K., Seka, D., Koffi, K. G., Kouonon, L. C., Koffi, K. K., & Bi, I. A. Z. (2019). Agromorphological divergence among four agro-ecological populations of Bambara groundnut (*Vigna subterranea* (L.) Verdc.) in Côte d'Ivoire. *Annals of Agricultural Sciences*, 64(1), 103-111.
- Botstein, D., White, R. L., Skolnick, M., & Davis, R. W. (1980). Construction of a genetic linkage map in man using restriction fragment length polymorphisms. *American Journal of Human Genetics*, *32*(3), 314.
- CRAN, 2014. The comprehensive R archive network. Comprehensive R Archive Network for the R programming language. [cited 2016 December 1). Available from:http: // cran.r-project.org/web/packages/available\_packages\_by name.html#available-packages-A.
- Crossa, J. (1990). Statistical analyses of multilocation trials. *Advances in Agronomy*, 44: 55–85.

- Darwin, C. (1876). *The Effects of Cross and Self Fertilisation in the Vegetable Kingdom: By Charles Darwin.* John Murray, Albemarle Street.
- Damfami, A., & Namo, O. A. T. (2020). Bambara groundnut (*Vigna subterranea* (L.) Verd.): A Review of its past, present, and future role in human nutrition. *Journal of Agriculture and Forest Meteorology Research*, 3(1), 274-281.
- Dansi, A., Vodouhè, R., Azokpota, P., Yedomonhan, H., Assogba, P., Adjatin, A., Loko, Y.L., Dossou-Aminon, I. and Akpagana, K.J.T.S.W.J. (2012). Diversity of the neglected and underutilized crop species of importance in Benin. *The Scientific World Journal*, 2012., Article ID 932947. 2012.
- Danyali, S. F., Razavi, F., Segherloo, A. E., Dehghani, H., & Sabaghpour, S. H. (2012). Yield Stability in Chickpea (*Cicer arietinum* L.) and Study Relationship among the univariate and multivariate stability Parameters. *Research in Plant Biology*, 2(3).
- De Riek, J., Calsyn, E., Everaert, I., Van Bockstaele, E., & De Loose, M. (2001). AFLP based alternatives for the assessment of distinctness, uniformity, and stability of sugar beet varieties. *Theoretical and Applied Genetics*, 103(8), 1254-1265. doi:10.1007/s001220.100710.
- Dell, M., Jones, B.F. & Olken, B.A. (2012) 'Temperature Shocks and Economic Growth: Evidence from the Last Half Century', American Economic Journal: Macroeconomics, 4(3), 66–95.
- Dia, M., Wehner, T.C, & Arellano, C. (2015). Analysis of genotype×environment interaction (G×E) using SAS programming. [accessed on 2019 December). http://cuke.hort.ncsu.edu/cucurbit/ wehner/software.html.
- Dos Santos, L. F., de Oliveira, E. J., dos Santos Silva, A., de Carvalho, F. M., Costa, J. L., & Pádua, J. G. (2011). ISSR markers as a tool for the assessment of genetic diversity in Passiflora. *Biochemical Genetics*, 49(7-8), 540-554.
- Du Petit Thours, L. M. A (1806). Genera nova Madagascariensis, p.23. Paris. Cited from Promoting the conservation and use of underutilized and neglected crops. 9. In: Proceedings of the Workshop on Conservation and Improvement of Bambara Groundnut (*Vigia subterranea* (L.) Verdc) Harare, Zambabwe. p.4.
- Earl, D. A. (2012). STRUCTURE HARVESTER: a website and program for visualizing STRUCTURE output and implementing the Evanno method. *Conservation Genetics Resources*, 4(2), 359-361.
- Eifediyi, E. K., Ahamefule, H. E., Izuogu, B. N., Ogedegbe, F. O., & Remison, S. U. (2020). Growth and yield of bambara groundnut (*Vigna subterranea* L. Verdc) as influenced by tillage and phosphorus application. *Rastenievadni Nauki*, 57(3), 59-70.
- Evanno, G., Regnaut, S., & Goudet, J. (2005). Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. *Molecular Ecology*, 14(8), 2611-2620.

- Eberhart, S.A., & Russell, W.A., (1966). Stability parameters for comparing varieties. *Crop Science*. 6: 36–40.
- Ezekannagha, E. (2020). Assessing the climatic suitability of Bambara groundnut as an underutilised crop to future climate projections in Sikasso and Ségou, Mali (Master's thesis, Faculty of Science).
- FAOSTAT, (2017) .http://www.fao.org/faostat/en/#data/QC, http://www.fao.org/faostat/en/#interactive\_download (accessed on 5\_3\_2019).
- FAOSTAT, (2020). Food and Agriculture Organization of the United Nations. Statistical Databases; FAOSTAT: Rome, Italy, 2020.
- Falconer, D.S. & Mackay, F.C. (1996). Introduction to Quantitative Genetics. Fourth ed. Longman, New York 464 p.
- Fatimah, S., & Ardiarini, N. R. (2018). Genetic diversity of Madurese bambara groundnut (Vigna subterranea L. Verdc.) lines based on morphological and RAPD markers. SABRAO Journal of Breeding & Genetics, 50(2).
- Finlay, K.W., & Wilkinson, G.N. (1963). The analysis of adaptation in a plant-breeding programme. *Crop and Pasture Science*, 14: 742–754.
- Frankham, R., Ballou, S. E. J. D., Briscoe, D. A., & Ballou, J. D. (2002). Introduction to conservation genetics. *Cambridge university press.* http://dx.doi.org/10.1017/CBO9780511808999.
- Francis, T.R., & Kannenberg, L.W., (1978). Yield stability studies in short–season maize. 1. A descriptive method for grouping genotypes. *Canadian Journal of Plant Science*, 58: 1029–1034.
- Zobel, R. W., & Gauch, H. G. (1996). AMMI analysis of yield trails. *Genotype by Environment Interaction. CRC Pub., Boca Raton, Florida*, 88-122.
- Gauch Jr, H. G., Piepho, H. P., & Annicchiarico, P. (2008). Statistical analysis of yield trials by AMMI and GGE: Further considerations. *Crop Science*, 48(3), 866-889.
- Gbaguidi, A. A., Dansi, A., Dossou-Aminon, I., Gbemavo, D. S. J. C., Orobiyi, A., Sanoussi, F., & Yedomonhan, H. (2018). Agromorphological diversity of local Bambara groundnut (*Vigna subterranea* (L.) Verdc.) collected in Benin. *Genetic Resources and Crop Evolution*, 65(4), 1159-1171.
- Gedif, M., & Yigzaw, D. (2014). Genotype by environment interaction analysis for tuber yield of potato (*Solanum tuberosum* L.) using a GGE biplot method in Amhara region, Ethiopia. *Agricultural Sciences*, 2014.
- Ghalmi, N., Malice, M., Jacquemin, J. M., Ounane, S. M., Mekliche, L., & Baudoin, J. P. (2010). Morphological and molecular diversity within Algerian cowpea (Vigna unguiculata (L.) Walp.) landraces. *Genetic Resources and Crop Evolution*, 57(3), 371-386.

- Godfray, H.C.J., Beddington, J.R., Crute, I.R., Haddad, L., Lawrence, D., Muir, J.F., Pretty, J., Robinson, S., Thomas, S.M. and Toulmin, C. (2010). Food security: the challenge of feeding 9 billion people. *Science*, 327(5967), 812-818.
- Gonné, S., Félix-Alain, W., & Benoît, K. B. (2013). Assessment of twenty bambara groundnut (Vigna subterranea (L.) Verdcourt) landraces using quantitative morphological traits. *International Journal of Plant Research*, 3(3), 39-45.
- Halimi, R. A., Barkla, B. J., Mayes, S., & King, G. J. (2019). The potential of the underutilized pulse bambara groundnut (Vigna subterranea (L.) Verdc.) for nutritional food security. *Journal of Food Composition and Analysis*, 77, 47-59.
- Hartl, D. L., Clark, A. G., & Clark, A. G. (1997). *Principles of Population Genetics* (Vol. 116). Sunderland, MA: Sinauer associates.
- Hashim, N., Rafii, M. Y., Oladosu, Y., Ismail, M. R., Ramli, A., Arolu, F., & Chukwu, S. (2021). Integrating Multivariate and Univariate Statistical Models to Investigate Genotype–Environment Interaction of Advanced Fragrant Rice Genotypes under Rainfed Condition. *Sustainability*, 13(8), 4555.
- Heller, J., Begemann, F. & Mushonga, J. (eds.). (1997). Bambara groundnut, Vigna subterranea (L.) Verdc., Promoting the Conservation and Use of Underutilized and Neglected Crops n.9, 166 p. ISBN: 978-92-9043-299-9, ISBN: 92-9043-299-3
- Holm, J. M., & Marioth, B. W. (1940). The Bambarra groundnut or Njugo bean. *Farming in South Africa*, *15*, 195-20
- Hillocks, R. J., Bennett, C., & Mponda, O. M. (2012). Bambara nut: A review of utilisation, market potential and crop improvement. *African Crop Science Journal*, 20(1).
- Ifo, S.A., Moutsambote, J.M., Koubouana, F., Yoka, J., Ndzai, S.F., Bouetou-Kadilamio, L.N.O., Mampouya, H., Jourdain, C., Bocko, Y., Mantota, A.B. and Mbemba, M., (2016). Tree species diversity, richness, and similarity in intact and degraded forest in the tropical rainforest of the Congo Basin: case of the forest of Likouala in the Republic of Congo. *International Journal of Forestry Research*, 2016.
- IPGRI, IITA, BAMNET. (2000). Descriptors for Bambara groundnut (Vigna subterranea). International Plant Genetic Resources Institute, Rome, Italy; International Institute of Tropical Agriculture, Ibadan, Nigeria. The International Bambara Groundnut Network, Germany. 2000; 57.
- Jideani, V. A., & Diedericks, C. F. (2014). Nutritional, therapeutic, and prophylactic properties of Vigna subterranea and Moringa oleifera. *Antioxidant-Antidiabetic Agents and Human Health*, *9*, 187-201.
- Johannsen, W. (1903) Om arvelighed i samfund og i rene linier. Oversigt over det Kongelige Danske Videnskabernes Selskabs Forhandlinger, vol. 3: 247-270. German ed. Erblichkeit in Populationen Und in Reinen Linien (1903) Gustav Fischer, Jena.

- Johnson, H. W, H. F. Robinson. & R. E. Comstock. (1955). Estimation of genetic and environmental variability in soybeans. Agronomy Journal, 47: 314–318.
- Jonah, P. M., Abimiku, O. E., & Adeniji, O. T. (2014). Multivariate analysis and character association on the growth and yield of Bambara groundnut in Mubi, Adamawa State, Nigeria. *International Journal of Management and Social Sciences Research*, 3(2), 58-65.
- Jonah, P. M., Aliyu, B., Adeniji, T. O., & Bello, D. (2012). Seasonal variation and Pearson correlation in yield and yield components in Bambara groundnut. World Journal of Agricultural Sciences, 8(1), 26-32.
- Kaur, G., Joshi, A., Jain, D., Choudhary, R., & Vyas, D. (2016). Diversity analysis of green gram (Vigna radiata (L.) Wilczek) through morphological and molecular markers. *Turkish Journal of Agriculture and Forestry*, 40(2), 229-240.
- Karikari, S. K., & Tabona, T. T. (2004). Constitutive traits and selective indices of Bambara groundnut (Vigna subterranea (L) Verdc) landraces for drought tolerance under Botswana conditions. *Physics and Chemistry of the Earth, Parts A/B/C*, 29(15-18), 1029-1034.
- Karikari, S. K. (2000). Variability between local and exotic Bambara groundnut landraces in Botswana. *African Crop Science Journal*, 8(2), 145-152.
- Karuppanapandian, T., Wang, H. W., Karuppudurai, T., Rajendhran, J., Kwon, M., Jang, C. S., ... & Kim, W. (2010). Efficiency of RAPD and ISSR markers in assessing genetic diversity and relationships in black gram (Vigna mungo L. Hepper) vari. *Canadian Journal of Plant Science*, 90(4), 443-452.
- Kang, M. S. (2002). Genotype-environment interaction: progress and prospects. *Quantitative Genetics, Genomics and Plant Breeding*, 221–243.
- Kang, M. S. (1993). Simultaneous selection for yield and stability in crop performance trials: Consequences for growers. *Agronomy Journal*, 85(3), 754-757.
- Kang, M. S., Miller, J. D., & Darrah, L. L. (1987). A note on relationship between stability variance and ecovalence. *Journal of Heredity*, 78(2), 107-107.
- Kaya, Y., Akçura, M., & Taner, S. (2006). GGE-biplot analysis of multi-environment yield trials in bread wheat. *Turkish Journal of Agriculture and Forestry*, 30(5), 325-337.
- Khan, M. M. H., Rafii, M. Y., Ramlee, S. I., Jusoh, M., & Mamun, A. (2020). Genetic variability, heritability, and clustering pattern exploration of Bambara groundnut (Vigna subterranea L. Verdc) accessions for the perfection of yield and yield-related traits. *BioMed research international*, 2020.
- Khan, M. M. H., Rafii, M. Y., Ramlee, S. I., Jusoh, M., & Al Mamun, M. (2021a). Genetic analysis and selection of Bambara groundnut (Vigna subterranea [L.] Verdc.) landraces for high yield revealed by qualitative and quantitative traits. *Scientific Reports*, *11*(1), 1-21.

- Khan, M. M. H., Rafii, M. Y., Ramlee, S. I., Jusoh, M., & Al-Mamun, M. (2021b). Bambara groundnut (Vigna subterranea L. Verdc): A crop for the new millennium, its genetic diversity, and improvements to mitigate future food and nutritional challenges. *Sustainability*, 13(10), 5530.
- Khan, M. M. H., Rafii, M. Y., Ramlee, S. I., Jusoh, M., Al Mamun, M., & Halidu, J. (2021c). DNA fingerprinting, fixation-index (Fst), and admixture mapping of selected Bambara groundnut (Vigna subterranea [L.] Verdc.) accessions using ISSR markers system. *Scientific reports*, 11(1), 1-23.
- Khan, M. M. H., Rafii, M. Y., Ramlee, S. I., Jusoh, M., & Al Mamun, M. (2021d). AMMI and GGE biplot analysis for yield performance and stability assessment of selected Bambara groundnut (Vigna subterranea L. Verdc.) genotypes under the multi-environmental trials (METs). *Scientific reports*, *11*(1), 1-17.
- Khan, M. M. H., Rafii, M. Y., Ramlee, S. I., Jusoh, M., & Al Mamun, M. (2022a). Pathcoefficient and correlation analysis in Bambara groundnut (Vigna subterranea [L.] Verdc.) accessions over environments. *Scientific reports*, 12(1), 1-12.
- Khan, M. M. H., Rafii, M. Y., Ramlee, S. I., Jusoh, M., Oladosu, Y., Al Mamun, M., & Khaliqi, A. (2022b). Unveiling Genetic Diversity, Characterization, and Selection of Bambara Groundnut (Vigna subterranea L. Verdc) Genotypes Reflecting Yield and Yield Components in Tropical Malaysia. *BioMed research international*, 2022.
- Khan, M. M. H., Rafii, M. Y., Ramlee, S. I., Jusoh, M., & Al Mamun, M. (2022c). Hereditary analysis and genotype× environment interaction effects on growth and yield components of Bambara groundnut (Vigna subterranea (L.) Verdc.) over multienvironments. *Scientific Reports*, 12(1), 1-19.
- Khaliqi, A., Rafii, M. Y., Mazlan, N., Jusoh, M., & Oladosu, Y. (2021). Genetic Analysis and Selection Criteria in Bambara Groundnut Accessions Based Yield Performance. *Agronomy*, *11*(8), 1634.
- Khaliqi, A. (2021). Genetic Inheritance and Yield Performance of Bambara Groundnut Genotypes Based on Seed Weight. *Afghanistan Research Journal*, 2(1), PP: 12-25.
- Kimaro, D., Melis, R., Sibiya, J., Shimelis, H., & Shayanowako, A. (2020). Analysis of Genetic Diversity and Population Structure of Pigeonpea [*Cajanus cajan* (L.) Millsp] Accessions Using SSR Markers. *Plants*, 9(12), 1643.
- Kopelman, N. M., Mayzel, J., Jakobsson, M., Rosenberg, N. A., & Mayrose, I. (2015). Clumpak: A program for identifying clustering modes and packaging population structure inferences across K. *Molecular Ecology Resources*, 15(5), 1179–1191.
- Kumar, A., Mishra, P., Singh, S. C., & Sundaresan, V. (2014). Efficiency of ISSR and RAPD markers in genetic divergence analysis and conservation management of *Justicia adhatoda* L., a medicinal plant. *Plant Systematics and Evolution*, 300(6), 1409-1420.
- Langat, C., Ombori, O., Leley, P., Karanja, D., Cheruiyot, R., Gathaara, M., & Masila, B. (2019). Genetic variability of agronomic traits as potential indicators of drought

tolerance in common beans (*Phaseolus vulgaris* L.). International Journal of Agronomy, 2019, Article ID 2360848. 2019.

- Lenka, D., & Mishra, B. (1973). Path coefficient analysis of yield in rice varieties. *Indian Journal of Agricultural Science*, 43(4), 376.
- Iezzoni, A. F., & Pritts, M. P. (1991). Applications of principal component analysis to horticultural research. *HortScience*, 26(4), 334-338.
- Lestari, S. A. D. A. D., Melati, M., & Purnamawati, H. (2015). Penentuan dosis optimum pemupukan n, p, dan k pada tanaman kacang Bogor [Vigna subterranea (L.) Verdcourt]. *Jurnal Agronomi Indonesia (Indonesian Journal of Agronomy)*, 43(3), 193-200.
- Lin Tan, X., Azam-Ali, S., Goh, E. V., Mustafa, M. A., Chai, H. H., Kuan Ho, W. & Massawe, F. (2020). Bambara groundnut: an underutilized leguminous crop for global food security and nutrition. *Frontiers in Nutrition*, 7, 276.
- Linnaeus, C. (1763). Species Plantarum. 2nd (Edn.), *Impensis Laurentii Salvii*, Stockholm 2: 785-1684.
- Lin, C. S., & Binns, M. R. (1994). Concepts and methods for analyzing regional trial data for cultivar and location selection. *Plant Breeding Reviews*, 12, 271-297.
- Mabhaudhi, T., Modi, A. T., & Beletse, Y. G. (2013). Growth, phenological and yield responses of a bambara groundnut (*Vigna subterranea* L. Verdc) landrace to imposed water stress: II. Rain shelter conditions. *Water Sa*, 39(2), 191-198.
- Mabhaudhi, T., Chibarabada, T. P., Chimonyo, V. G. P., & Modi, A. T. (2018). Modelling climate change impact: a case of bambara groundnut (Vigna subterranea). *Physics and Chemistry of the Earth, Parts A/B/C, 105,* 25-31.
- Majola, N. G., Gerrano, A. S., & Shimelis, H. (2021). Bambara Groundnut (Vigna subterranea [L.] Verdc.) Production, Utilisation and Genetic Improvement in Sub-Saharan Africa. Agronomy, 11(7), 1345.
- Makanda, I., Tongoona, P., Madamba, R., Icishahayo, D., & Derera, J. (2009). Path coefficient analysis of bambara groundnut pod yield components at four planting dates. *Research Journal of Agriculture and Biological Sciences*, 5(3), 287-292.
- Mannan, S. & Hameed, S. (2013). A molecular tool for differentiation of *Xanthomonas* oryzae pathovars isolated from rice. *Journal of Agricultural Research*, *51*(1), 1-7.
- Masindeni, D. R. (2006). *Evaluation of Bambara groundnut (Vigna subterranea) for yield stability and yield related characteristics* (Doctoral dissertation, University of the Free State), Bloemfontein, South Africa.
- Massawe, F. J., Roberts, J. A., Azam-Ali, S. N., & Davey, M. R. (2003). Genetic diversity in bambara groundnut (*Vigna subterranea* (L.) Verdc) landraces assessed by Random Amplified Polymorphic DNA (RAPD) markers. *Genetic Resources and Crop Evolution*, 50(7), 737-741.

- Massawe, F. J., Dickinson, M., Roberts, J. A., & Azam-Ali, S. N. (2002). Genetic diversity in bambara groundnut (*Vigna subterranea* (L.) Verdc) landraces revealed by AFLP markers. *Genome*, 45(6), 1175-1180.
- Massawe, F. J., Mwale, S. S., Azam-Ali, S. N., & Roberts, J. A. (2005). Breeding in bambara groundnut (*Vigna subterranea* (L.) Verdc.): strategic considerations. *African Journal of Biotechnology*, 4(6), 463-471.
- Mantel, N. (1967). The detection of disease clustering and a generalized regression approach. *Cancer research*, 27(2 Part 1), 209-220.
- Maunde, S. M., Tanimu, B., & Mahmud, M. (2015). Correlation and path coefficient analysis of yield characters of bambara (*Vigna subterranea* L. Verdc.). African Journal of Environmental Science and Technology, 9(1), 12-15.
- Mayes, S., Stadler, F., Ribas-Vargas, G., Basu, S., Massawe, F., Bonin, A. & Azam-Ali, S. (2015). Marker-assisted breeding in crops: wheat and bambara groundnut as two contrasting examples of progress. *International Society of Oil Palm Breeders, Kuala Lumpur*, 1-12.
- Mayes, S., Ho, W. K., Chai, H. H., Gao, X., Kundy, A. C., Mateva, K. I. & Azam-Ali, S. N. (2019). Bambara groundnut: an exemplar underutilised legume for resilience under climate change. *Planta*, 250(3), 803-820.
- Mbosso, C., Boulay, B., Padulosi, S., Meldrum, G., Mohamadou, Y., Berthe Niang, A. & Sidibé, A. (2020). Fonio and bambara groundnut value chains in mali: issues, needs, and opportunities for their sustainable promotion. *Sustainability*, 12(11), 4766.
- McDermott, J. M. & McDonald, B. A. (1993). Gene flow in plant pathosystems. *Annual Review of phytopathology*, *31*(1), 353-373.
- Misangu, R. N., Azmio, A., Reuben, S. O. W. M., Kusolwa, P. M., & Mulungu, L. S. (2007). Path coefficient analysis among components of yield in bambara groundnut (*Vigna subterranea* L. Verdc) landraces under screen house conditions. *Journal of Agronomy*. 6(2), 317-323.
- Minnaar-Ontong, A., Gerrano, A. S., & Labuschagne, M. T. (2021). Assessment of genetic diversity and structure of Bambara groundnut [*Vigna subterranea* (L.) verdc.] landraces in South Africa. *Scientific Reports*, 11(1), 1-9.
- MMD. (2020). Malaysian Meteorological Department, Ministry of Science, Technology and Innovation, Kuala Lumpur, Malaysia. www.met.gov.my, *Weather record for Pusat pertanian*, Serdang (2013-2020).
- Modi, A.T. (2013). Pre-germination hydration affects seed performance in Bambara groundnut. *Scientific Research and Essays*, 8: 940-945
- Mogale, T. E. (2018). Multi-Location Field Evaluation of Bambara Groundnut (*Vigna subterranean* (L) Verdc) for Agronomic Performance and Seed Protein (Doctoral dissertation).

- Mojena R. Hierarchical grouping methods and stopping rules: an evaluation. *The Computer Journal*. 1977; 20(4): 359-363.
- Mohammed MS. *Pre-breeding of Bambara Groundnut (Vigna subterranea [L.] Verdc.)* (Doctoral dissertation). University of KwaZulu-Natal, Durban, South Africa. 2014.
- Mohammadi, S. A., & Prasanna, B. M. (2003). Analysis of genetic diversity in crop plants—salient statistical tools and considerations. *Crop science*, 43(4), 1235-1248.
- Mohammed SM, Shimelis HA and Laing MD (2019) Genetic diversity of Bambara groundnut genotypes (*Vigna subterranea* [L.] Verdc.) revealed by SSR markers. Society for Underutilized Legumes, https://sulegumes.org/ e-ISSN: 2705-3776, *Journal of Underutilized Legumes*, 1 (1): 169 182.
- Mohammed, M. S., Shimelis, H. A., & Laing, M. D. (2020). Preliminary morphological characterization and evaluation of selected Bambara groundnut [*Vigna subterranea* (L.) Verdc.] genotypes for yield and yield related traits. *Legume Research-an International Journal*, 43(2), 157-164. DOI: 10.18805/LR-475.
- Molosiwa, O. O., Aliyu, S., Stadler, F., Mayes, K., Massawe, F., Kilian, A., & Mayes, S. (2015). SSR marker development, genetic diversity, and population structure analysis of Bambara groundnut [*Vigna subterranea* (L.) Verdc.] landraces. *Genetic Resources and Crop Evolution*, 62(8), 1225-1243.
- Molosiwa, O. O. (2012). Genetic diversity and population structure analysis of bambara groundnuts (Vigna subterranea (L.) Verdc.) landraces using morpho-agronomic characters and SSR markers (Doctoral dissertation, University of Nottingham, UK).
- Mondal, S., Sutar, S. R., & Badigannavar, A. M. (2009). Assessment of genetic diversity in cultivated groundnut (*Arachis hypogaea* L.) with differential responses to rust and late leaf spot using ISSR markers. *Indian Journal of Genetics and Plant Breedingt*, 69(3), 219-224.
- Mubaiwa, J., Fogliano, V., Chidewe, C., & Linnemann, A. R. (2017). Hard-to-cook phenomenon in bambara groundnut (*Vigna subterranea* (L.) Verdc.) processing: Options to improve its role in providing food security. *Food Reviews International*, 33(2), 167-194.
- Mulamba, N. N., & Mock, J. J. (1978). Improvement of yield potential of the ETO blanco maize (*Zea mays* L.) population by breeding for plant traits [Mexico]. *Egyptian Journal of Genetics and Cytology*.
- Muhammad, I., Rafii, M. Y., Ramlee, S. I., Nazli, M. H., Harun, A. R., Oladosu, Y. & Arolu, I. W. (2020). Exploration of bambara groundnut (*Vigna subterranea* (L.) Verdc.), an underutilized crop, to aid global food security: Varietal improvement, genetic diversity, and processing. *Agronomy*, 10(6), 766.
- Muhammad, I., Rafii, M. Y., Nazli, M. H., Ramlee, S. I., Harun, A. R., & Oladosu, Y. (2021). Determination of lethal (LD) and growth reduction (GR) doses on acute and chronic gamma-irradiated Bambara groundnut [*Vigna subterranea* (L.) Verdc.] varieties. *Journal of Radiation Research and Applied Sciences*, 14(1), 133-145.

- Mukakalisa, C., Kandawa-Schulz, M., & Mapaure, I. (2011, June). Genetic diversity in landraces of bambara groundnut found in Namibia using RAPD markers. In II International Symposium on Underutilized Plant Species: Crops for the Future-Beyond Food Security 979, 683-687.
- Murphy, S. E., Lee, E. A., Woodrow, L., Seguin, P., Kumar, J., Rajcan, I., & Ablett, G. R. (2009). Genotype× environment interaction and stability for isoflavone content in soybean. *Crop Science*, 49(4), 1313-1321.
- Musa, M., Massawe, F., Mayes, S., Alshareef, I., & Singh, A. (2016). Nitrogen fixation and N-balance studies on Bambara groundnut (Vigna subterranea L. Verdc) landraces grown on tropical acidic soils of Malaysia. *Communications in Soil Science and Plant Analysis*, 47(4), 533-542.
- Naik, U. (2015). Correlation and Path Coefficient Analysis Between Seed Yield and Its Component Characters in M4 and M5 Generations of Bambara Groundnut (Doctoral dissertation, *University of Agricultural Sciences*, GKVK).
- Nankar, A. N., Tringovska, I., Grozeva, S., Ganeva, D., & Kostova, D. (2020). Tomato phenotypic diversity determined by combined approaches of conventional and highthroughput tomato analyzer phenotyping. *Plants*, 9(2), 197.
- Nei, M. (1978). Estimation of average heterozygosity and genetic distance from a small number of individuals. *Genetics*, 89(3), 583-590.
- Ng, W. L., & Tan, S. G. (2015). Inter-simple sequence repeat (ISSR) markers are we doing it right. *ASM Science Journal*, *9*(1), 30-39.
- Ndiang, Z., Bell, J. M., Missoup, A. D., Fokam, P. E., & Amougou, A. (2012). Etude de la variabilité morphologique de quelques variétés de voandzou au Cameroun. *Journal of Applied Biosciences*, 60, 4394-4409.
- Nilkanta, H., Amom, T., Tikendra, L., Rahaman, H., & Nongdam, P. (2017). ISSR marker-based population genetic study of *Melocanna baccifera* (Roxb.) Kurz: a commercially important bamboo of Manipur, North-East India. *Scientifica*, 2017.
- Nnamani, C. V., Afiukwa, C. A., Oselebe, H. O., Igwe, D. O., Uhuo, C. A., Idika, K. O., & Oketa, C. N. (2019). Genetic diversity of some African yam bean accessions in Ebonyi State assessed using Inter Simple Sequence Repeat (ISSR) markers. *Journal of Underutilized Legumes*, 1, 20-33.
- NTSYS-pc NT, Taxonomy N, Multivariate Analysis System, version 2.2. Exeter Software: Setauket, NY, USA. 2005.
- Ntundu, W. H., Bach, I. C., Christiansen, J. L., & Andersen, S. B. (2004). Analysis of genetic diversity in bambara groundnut [*Vigna subterranea* (L.) Verdc] landraces using amplified fragment length polymorphism (AFLP) markers. *African Journal of Biotechnology*, 3(4), 220-225.

- Ntundu, W. H., Shillah, S. A., Marandu, W. Y. F., & Christiansen, J. L. (2006). Morphological diversity of bambara groundnut [*Vigna subterranea* (L.) Verdc.] landraces in Tanzania. *Genetic Resources and Crop Evolution*, 53(2), 367-378.
- Obidiebube, E. A., Eruotor, P. G., Akparaobi, S. O., Okolie, H., & Obasi, C. C. (2020). Evaluation of Bambara Groundnut (*Vigna Subterranea* (L) Verdc.) Varieties for Adaptation to Rainforest Agroecological Zone of Delta State, Nigeria. *Evaluation*, 7(1), 39-43.
- Odeigah, P. G. C., & Osanyinpeju, A. O. (1998). Evaluating the genetic biodiversity of Bambara groundnut accessions from Nigeria using SDS-polyacrylamide gel electrophoresis. *Genetic Resources and Crop Evolution*, 45(5), 451-458.
- Odongo, F. O., Oyoo, M. E., Wasike, V., Owuoche, J. O., Karanja, L., & Korir, P. (2015). Genetic diversity of Bambara groundnut (*Vigna subterranea* (L.) verdc.) landraces in Kenya using microsatellite markers. *African Journal of Biotechnology*, 14(4), 283-291.
- Ofori, K., Kumaga, F. K., & Tonyigah, A. (2006). Morphological characterization and agronomic evaluation of Bambara groundnut (*Vigna subterranea* (L.) Verdc.) germplasm in Ghana. *Plant Genetic Resources Newsletter (IPGRI/FAO)*.
- Okoli, E. E. (2021). Exploitation of rank summation index for the selection of 21 maize hybrids for green maize production in South-eastern Nigeria. *Journal of Bioscience and Biotechnology Discovery*, 6(2), 13-18.
- Onwubiko, N. C., Uguru, M. I., & Chimdi, G. O. (2019a). Estimates of genetic parameters in bambara groundnut {Vigna subterranea (L.) VERDC.}. Plant Breeding and Biotechnology, 7(4), 295-301.
- Onwubiko, N. C., Uguru, M. I., & Chimdi, G. O. (2019b). Selection for yield improvement in Bambara groundnut (*Vigna subterranea* L. Verdc.). *Journal of Plant Breeding and Genetics*, 7(2), 41-53.
- Olatunji, T. L., & Afolayan, A. J. (2019). Evaluation of genetic relationship among varieties of Capsicum annuum L. and Capsicum frutescens L. in West Africa using ISSR markers. *Heliyon*, 5(5), e01700.
- Oladosu, Y., Rafii, M. Y., Abdullah, N., Magaji, U., Miah, G., Hussin, G., & Ramli, A. (2017). Genotype× Environment interaction and stability analyses of yield and yield components of established and mutant rice genotypes tested in multiple locations in Malaysia. Acta Agriculturae Scandinavica, Section B—Soil & Plant Science, 67(7), 590-606.
- Oladosu, Y., Rafii, M. Y., Magaji, U., Abdullah, N., Miah, G., Chukwu, S. C., & Kareem, I. (2018). Genotypic and phenotypic relationship among yield components in rice under tropical conditions. *BioMed Research International*, 2018.
- Oladosu, Y., Rafii, M. Y., Abdullah, N., Abdul Malek, M., Rahim, H. A., Hussin, G., & Kareem, I. (2014). Genetic variability and selection criteria in rice mutant lines as revealed by quantitative traits. *The Scientific World Journal*, 2014.

- Olanrewaju, O. S., Oyatomi, O., Babalola, O. O., & Abberton, M. (2021a). Genetic Diversity and Environmental Influence on Growth and Yield Parameters of Bambara Groundnut. *Frontiers in Plant Science*, 12, 796352-796352.
- Olanrewaju, O. S., Oyatomi, O., Babalola, O. O., & Abberton, M. (2021b). GGE Biplot Analysis of Genotype× Environment Interaction and Yield Stability in Bambara Groundnut. *Agronomy*, 11(9), 1839.
- Oludare, A. J., Kioko, J. I., Akeem, A. A., Olumide, A. T., Justina, K. R., Adejoke, I. M., & Aderonke, O. (2017). Genetic Diversity In Bambara Groundnut (*Vigna Subterranean*) As Revealed By Molecular Weights of The Seeds'proteins. *Bangladesh Journal of Plant Breeding and Genetics*, 30(2), 19-28.
- Olukolu, B. A., Mayes, S., Stadler, F., Quat Ng, N., Fawole, I., Dominique, D., & Kole, C. (2012). Genetic diversity in Bambara groundnut (*Vigna subterranea* (L.) Verdc.) as revealed by phenotypic descriptors and DArT marker analysis. *Genetic Resources* and Crop Evolution, 59(3), 347-358.
- Oliveira, I. J. D., Zanotto, M. D., Krieger, M., & Vencovsky, R. (2012). Inbreeding depression in castor bean (*Ricinus communis* L.) progenies. *Crop Breeding and Applied Biotechnology*, 12(4), 269-276.
- Ouedraogo, M., Ouedraogo, J. T., Tignere, J. B., Bilma, D., Dabire, C. B., & Konate, G. (2008). Characterization and evaluation of accessions of Bambara groundnut (*Vigna subterranea* (L.) Verdcourt) from Burkina Faso. *Sciences & Nature*, 5(2), 191-197.
- Oumer, O. A., Dagne, K., Feyissa, T., Tesfaye, K., Durai, J., & Hyder, M. Z. (2020). Genetic diversity, population structure, and gene flow analysis of lowland bamboo [Oxytenanthera abyssinica (A. Rich.) Munro] in Ethiopia. Ecology and evolution, 10(20), 11217-11236.
- Oyiga, B. C., Uguru, M. I., & Aruah, C. B. (2010). Studies on the floral traits and their implications on pod and seed yields in bambara groundnut [*Vigna subterrenea* (L.) Verdc]. *Australian Journal of Crop Science*, 4(2), 91-97.
- Oyiga, B. C., & Uguru, M. I. (2011). Interrelationships among pod and seed yield traits in bambara groundnut (*Vigna subterranea* L. Verdc) in the derived savanna agroecology of south-eastern Nigeria under two planting dates. *International Journal of Plant Breeding*, 5(2), 106-111.
- Parajuli, A.; Yu, L.; Peel, M.; See, D.; Wagner, S.; Norberg, S.; Zhang, Z. Self-Incompatibility, Inbreeding Depression, and Potential to Develop Inbred Lines in Alfalfa: A Review. Preprints 2020, 2020110390 (doi: 10.20944/preprints 202011.0390.v1.
- Peakall, R. O. D., & Smouse, P. E. (2006). GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and research. *Molecular ecology notes*, 6(1), 288-295.

Pearson K. Correlation coefficient. In Royal Society Proceedings. 1895; 58: 214.

- Powell, W., Morgante, M., Andre, C., Hanafey, M., Vogel, J., Tingey, S., & Rafalski, A. (1996). The comparison of RFLP, RAPD, AFLP and SSR (microsatellite) markers for germplasm analysis. *Molecular Breeding*, 2(3), 225-238.
- Pranesh, H., Nandini, R., Chandra, K., Rangaiah, S. & Nagaraju, N. (2017). Character association and path analysis of yield and yield components in m3 generation of bambara groundnut (*Vigna subterranea* (L.) verdc.) treated with ethyl methane sulphonate (EMS). *International Journal of Pure & Applied Bioscience*. 5(3), 306– 311.
- Prevost, A., & Wilkinson, M. J. (1999). A new system of comparing PCR primers applied to ISSR fingerprinting of potato cultivars. *Theoretical and applied Genetics*, 98(1), 107-112.
- Pritchard, J. K., Wen, W., & Falush, D. (2010). Documentation for STRUCTURE software: Version 2. *University of Chicago, Chicago, IL*.
- Pritchard, J. K., Stephens, M., & Donnelly, P. (2000). Inference of population structure using multilocus genotype data Wu. *Genetics*, 155(2), 945-959.
- Ramzan, M., Sarwar, S., Kauser, N., Saba, R., Hussain, I., Shah, A. A., & Alwahibi, M. S. (2020). Assessment of Inter simple sequence repeat (ISSR) and simple sequence repeat (SSR) markers to reveal genetic diversity among Tamarix ecotypes. *Journal of King Saud University-Science*, 32(8), 3437-3446.
- Rathore, R. K., & Ram, M. (2013). Role of selection in improving cultivars. *Popular Kheti*, 1(2), 45-48.
- Rebrean, F., Fustos, A., Tăut, I., Szabo, K., Hârţa, M., Pamfil, D., Rebrean, M., & Sălăgean, T., (2019). Genetic diversity of Acer pseudoplatanus L. populations from Transylvania. *Brazilian Journal of Botany*, 42(4), pp.643-650.
- RStudio. RStudio: Integrated development environment for R (Computer software v0.98.1074). RStudio. 2014, [cited 2021 February 2019]. Available from: http://www.rstudio.org/
- Rungnoi, O., Suwanprasert, J., Somta, P., & Srinives, P. (2012). Molecular genetic diversity of Bambara groundnut (*Vigna subterranea* L. Verdc.) revealed by RAPD and ISSR marker analysis. *SABRAO Journal of Breeding & Genetics*, 44(1).
- Saleh, G., Yusop, M. R., & Yap, T. C. (1993). Inbreeding depression and heterosis in sweet corn varieties Manis Madu and Bakti-1. *Pertanika Journal of Tropical Agricultural Science*, *16*(3), 209-214.
- Salleh, S. B. (2019). Genotype by environment interaction and genotype stability analysis on advances blast resistance rice genotypes (Master of science dissertation), Universiti Putra Malaysia,
- Salazar-Laureles, M. E., Pérez-López, D. D. J., González-Huerta, A., Vázquez-García, L. M., & Valadez-Moctezuma, E. (2015). Genetic variability analysis of faba bean

accessions using Inter-simple sequence repeat (ISSR) markers. *Chilean Journal of Agricultural Research*, 75(1), 122-130.

- Shegro, A., Van Rensburg, W. J., & Adebola, P. O. (2013). Assessment of genetic variability in bambara groundnut (*Vigna subterrenea* L. Verdc.) using morphological quantitative traits. *Academia Journal of Agricultural Research*, 1(3), 045-051.
- Shiyam, J. O., Nkor, N. N., Binang, W. B., & Effa, E. B. (2016). Yield response of bambara groundnut (*Voandzeia subterrenea* L. Thours.) varieties to organomineral fertilizer in the coastal forest of southeastern Nigeria. *Scirea Journal of Agriculture*, 1, 91-106.
- Singh, R.K. & Chaudhary, B.D. (1985). *Biometrical methods in Quantitative Genetic analysis.* Kalyani Publishers, New Delhi India, pp 253-260
- Soluki, M., Ghanbari, A., Sadehi, A. A. V., & Kazemipour, A. (2008). Yield stability analysis in hulless barley (*Hordeum vulgare L.*). Asian Journal of Plant Science, 7(6), 589-593.
- Somta, P., Chankaew, S., Rungnoi, O., & Srinives, P. (2011). Genetic diversity of the Bambara groundnut (*Vigna subterranea* (L.) Verdc.) as assessed by SSR markers. *Genome*, 54(11), 898-910.
- Shukla, G. K. (1972). Some statistical aspects of partitioning genotype environmental components of variability. *Heredity*, 29: 237–245.
- Tessier, C., David, J., This, P., Boursiquot, J. M., & Charrier, A. (1999). Optimization of the choice of molecular markers for varietal identification in *Vitis vinifera* L. *Theoretical and Applied Genetics*, 98(1), 171-177.
- Temesgen, T., Keneni, G., Sefera, T., & Jarso, M. (2015). Yield stability and relationships among stability parameters in faba bean (*Vicia faba* L.) genotypes. *The Crop Journal*, 3(3), 258-268.
- Tian, B., Yang, H. Q., Wong, K. M., Liu, A. Z., & Ruan, Z. Y. (2012). ISSR analysis shows low genetic diversity versus high genetic differentiation for giant bamboo, *Dendrocalamus giganteus* (Poaceae: Bambusoideae), in China populations. *Genetic Resources and Crop Evolution*, 59(5), 901-908.
- Touré, Y., Koné, M., Tanoh, H. K., & Koné, D. (2012). Agromorphological and phenological variability of 10 Bambara groundnut [*Vigna subterranea* (L.) Verdc. (Fabaceae)] landraces cultivated in the Ivory Coast. *Tropicultura*, 30(4), 216-221.
- Uba, C. U., Oselebe, H. O., Tesfaye, A. A., & Abtew, W. G. (2021). Genetic diversity and population structure analysis of bambara groundnut (*Vigna subterrenea* L) landraces using DArT SNP markers. *PloS one*, *16*(7), e0253600.
- Unigwe, A. E., Gerrano, A. S., Adebola, P., Pillay, M., & Monrovia, L. (2016). Morphological variation in selected accessions of Bambara groundnut (*Vigna subterranea* L. Verdc) in South Africa. *Journal of Agricultural Science*, 8(11), 69-80.

- Usman, M. G., Rafii, M. Y., Ismail, M. R., Malek, M. A., & Abdul Latif, M. (2014). Heritability and genetic advance among chili pepper genotypes for heat tolerance and morphophysiological characteristics. *The Scientific World Journal*, 2014.
- Usman, M. G., Rafii, M. Y., Martini, M. Y., Oladosu, Y., & Kashiani, P. (2017). Genotypic character relationship and phenotypic path coefficient analysis in chili pepper genotypes grown under tropical condition. *Journal of the Science of Food and Agriculture*, 97(4), 1164-1171.
- Varshney, R. K., Chabane, K., Hendre, P. S., Aggarwal, R. K., & Graner, A. (2007). Comparative assessment of EST-SSR, EST-SNP and AFLP markers for evaluation of genetic diversity and conservation of genetic resources using wild, cultivated and elite barleys. *Plant Science*, 173(6), 638-649.
- Verdcourt, B. (1980). *The correct name for the Bambara groundnut*. Kew Bull. 35(3): 474.
- Virga, G., Licata, M., Consentino, B. B., Tuttolomondo, T., Sabatino, L., Leto, C., & La Bella, S. (2020). Agro-morphological characterization of sicilian chili pepper accessions for ornamental purposes. *Plants*, 9(10), 1400.
- Wachira, F., Ng'etich, W., Omolo, J., & Mamati, G. (2002). Genotype × environment interactions for tea yields. *Euphytica*, 127: 289–297.
- Welt, R. S., Litt, A., & Franks, S. J. (2015). Analysis of population genetic structure and gene flow in an annual plant before and after a rapid evolutionary response to drought. *AoB Plants*, 7.
- Wigglesworth, D. J. (1996). The potential for genetic improvement of bambara groundnut (*Vigna subterranea* L. Verdc.) in Botswana. In *Proceedings of the International Symposium on Bambara groundnut*, 23-25.
- Wright, S. (1942). Statistical genetics and evolution. *Bulletin of the American Mathematical Society*, 48(4), 223-246.
- Wright, S. (1921). Correlation and causation. Journal of Agricultural Research, 20:557–585.
- Wright, S. (1984). Evolution and the genetics of populations, volume 4: variability within and among natural populations (Vol. 4). *University of Chicago press*.
- Wricke, G. (1962). On a method of understanding the biological diversity in field research. *Zeitschrift für Pflanzenzüchtung*, 47: 92–146.
- Wu, W., Chen, F., Yeh, K., & Chen, J. (2019). ISSR analysis of genetic diversity and structure of plum varieties cultivated in southern China. *Biology*, 8(1), 2.
- Yakubu, H., Kwari, J. D., & Sandabe, M. K. (2010). Effect of phosphorus fertilizer on nitrogen fixation by some grain legume varieties in Sudano–Sahelian zone of North Eastern Nigeria. *Nigerian Journal of Basic and Applied Sciences*, 18(1), 44-49.

- Yan, W., & Kang, M.S. (2002). GGE biplot analysis: A graphical tool for breeders, geneticists, and agronomists. *CRC press*. New York, USA, pp. 71.
- Yan, W., Kang, M.S., Ma, B., Woods, S., & Cornelius, P.L. (2007). GGE biplot vs. AMMI analysis of genotype–by–environment data. *Crop Science*, 47: 643–653.
- Yan W. & Rajcan I. (2002). Biplot Analysis of Test Sites and Trait Relations of Soybean in Ontario. *Crop Science*, 42: 11–20.
- Yan, W., Hunt, L.A., Sheng, Q., & Szlavnics, Z. (2000). Cultivar evaluation and megaenvironment investigation based on the GGE biplot. Crop Science. 40: 597–605.
- Yan, W. (2001). GGE Biplot–A Windows application for graphical analysis of multi– environment trial data and other types of two–way data. Agronomy Journal. 93:1111–1118.
- Yeh, F. C., Yang, R. C., & Boyle, T. (1999). POPGENE version 1.32: Microsoft Windows-based freeware for population genetic analysis, quick user guide. *Center* for International Forestry Research, University of Alberta, Edmonton, Alberta, Canada, 1-29.
- Zarei, A., & Erfani-Moghadam, J. (2021). SCoT markers provide insight into the genetic diversity, population structure and phylogenetic relationships among three Pistacia species of Iran. *Genetic Resources and Crop Evolution*, 68(4), 1625-1643.
- Zheng, K. (1995). Rapid DNA isolation for marker assisted selection in rice breeding. *Rice Genetics Newsletter*, *12*, 255-258.
- Zimisuhara, B., Valdiani, A., Shaharuddin, N. A., Qamaruzzaman, F., & Maziah, M. (2015). Structure and principal components analyse reveal an intervarietal fusion in Malaysian mistletoe fig (*Ficus deltoidea* Jack) populations. *International Journal of Molecular Sciences*, 16(7), 14369-14394.