



ETHIOPATHOGENESIS OF CASEOUS LYMPHADENITIS IN GOATS

By

ZAID KHUDHUR MAHMOOD

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of
Philosophy**

April, 2015

FPV 2015 7

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial use of material may only be made with the expression, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

TO MY FAMILY WHO MADE MY PROFESSIONAL PROGRESS POSSIBLE;
TO MY INSTRUCTORS, TEACHERS, MENTORS WHO MADE IT A REALITY;
AND TO THE FELLOW RESEARCHERS WHO MAY BE ABLE TO USE THE
RESULTS OF THIS RESEARCH, I DEDICATE MY WORK TO ALL WITH
LOVE AND GRATITUDE.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
Fulfillment of the requirement for the degree of Doctor of Philosophy

ETHIOPATHOGENESIS OF CASEOUS LYMPHADENITIS IN GOATS

By

ZAID KHUDHUR MAHMOOD

April, 2015

Chairman: Professor Abd Wahid Haron, Ph.D.
Faculty: Veterinary Medicine

Caseous lymphadenitis (CLA) is an economically important disease of sheep and goats worldwide. However, paucity of literature on CLA research in goats in Malaysia led to this study to investigate the mechanism by which *Corynebacterium pseudotuberculosis* and its exotoxin, phospholipase D (PLD) adversely affects reproduction in mice and goats. This study was also designed to test the hypothesis that acute phase proteins, specifically, haptoglobin and serum amyloid A can be used as diagnostic biomarkers for CLA.

One hundred and five clinically healthy mice of both sexes aged between 10 to 12 weeks, weighing 25-35 g and twenty six clinically healthy crossbred Boer goats (13 bucks and 13 does) aged between 12 to 14 months, weighing 30-35 kg were used in this study. The does were non-pregnant, non-lactating and have been housed separately from the bucks. At the beginning of the experiments, two types of inoculum were used that involved live *C. pseudotuberculosis* and its exotoxin, PLD. The mice were divided into three equal groups of 35 mice each. The crossbred Boer goats were also divided into three groups, the first group consisted of 6 goats, the second group consisted of 10 goats and the third group consisted of 10 goats. The control groups 1 (mice and goats) were inoculated with sterile phosphate buffered saline (PBS 1 ml/mouse intra-peritoneal and 1 ml/goat subcutaneous) whereas group 2 were inoculated with live *C. pseudotuberculosis* (1x10⁹ cfu intra-peritoneal in mice and subcutaneous in goats) and group 3 were inoculated with PLD (1 ml/mouse intra-peritoneal and 1 ml/20 kg B.W. in goats). Mice were observed during the entire experiment, blood was collected into heparinized tubes for sex hormones analyses. Post mortem examination of gross lesions was conducted on ovaries, uterus, testes and epididymis and tissue samples were fixed for

histopathological examination. Goats were also observed during the entire experiment for clinical signs and serial blood collections were conducted. Blood was placed into heparinized, plain, EDTA and sodium citrate tubes for acute phase proteins (APPs), haematology and sex hormones analyses. Post mortem examination of gross lesions was conducted on lungs, heart, liver, spleen, kidneys and lymph nodes and tissue samples were fixed for histopathology and cellular changes were scored.

Mice inoculated with *C. pseudotuberculosis* showed significant decrease ($p < 0.05$) in testosterone concentration. Both inoculated groups showed no gross changes in the reproductive organs. However, microscopic examination revealed that *C. pseudotuberculosis* inoculation caused congestion, oedema, infiltration of inflammatory cells, degeneration and necrosis in the ovaries, uterus, testes and epididymis. In addition, mice in PLD inoculated group showed congestion, degeneration and necrosis in the ovaries, uterus, testes and epididymis as well as thrombosis in the ovaries and uterus only.

Goats inoculated with both *C. pseudotuberculosis* and PLD showed significant increase ($p < 0.05$) in haptoglobin concentration with significant decrease ($p < 0.05$) in albumin concentration. Goats also showed significant increase ($p < 0.05$) in body temperature, heart rate, respiratory rate during different times of the experiment post inoculation with both *C. pseudotuberculosis* and PLD. Moreover, body score showed significant decrease ($p < 0.05$) in both treated groups toward the end of the experiment. Additionally, the site of inoculation and the superficial lymph nodes showed abscess formation in *C. pseudotuberculosis* inoculated group only.

Corynebacterium pseudotuberculosis inoculated goats showed significant decrease ($p < 0.05$) in red blood cell count, haemoglobin, creatine phosphokinase concentrations and significant increase ($p < 0.05$) in neutrophils count. Both *C. pseudotuberculosis* and PLD inoculated goats showed significant increase ($p < 0.05$) in packed cell volume, mean corpuscular volume, white blood cells, lymphocytes, monocytes, eosinophils and basophils counts and aspartate transaminase, total bilirubin, calcium, creatinine, gamma glutamyltransferase, urea, total protein, globulin, lactate dehydrogenase concentrations, prothrombin time and activated partial thromboplastin time whilst both inoculated groups showed significant decrease ($p < 0.05$) in mean corpuscular haemoglobin concentration, neutrophils count, alanine transaminase and alkaline phosphatase concentrations. Inoculation with PLD showed significant decrease ($p < 0.05$) in creatine phosphokinase concentration.

Testosterone, progesterone, estrogen hormones, scrotal circumference and semen volume, pH, wave pattern, motility and concentration showed significant decrease ($p < 0.05$) post inoculation with *C. pseudotuberculosis* and PLD. In

addition, dead sperm and abnormal sperm percentage showed significant increase ($p < 0.05$) in both treated groups. Microscopic examination of the ovaries showed fibrous tissue formation post inoculation with *C. pseudotuberculosis*, whilst in PLD inoculated group the ovaries showed severe congestion, degeneration and necrosis of the stromal cells. In addition, the uterine tissues showed congestion, degeneration and necrosis post inoculation with both *C. pseudotuberculosis* and PLD. The testes showed irregular and shrinkage of seminiferous tubules with oedema, degeneration and necrosis of spermatogonia cells and Leydig cells post inoculation with both *C. pseudotuberculosis* and PLD. Similarly, the epididymis showed oedema, degeneration and necrosis of the lining epithelium of the epididymal tubules post inoculation with both *C. pseudotuberculosis* and PLD.

Corynebacterium pseudotuberculosis inoculated group showed gross changes in lungs represented by grey and red hepatization, deeper inside, spots of congestion and abscess formation. The heart appeared oedematous and the pericardium showed some degree of opacity. The liver showed multiple abscess formation on its surface. The kidneys appeared friable with opaque capsule whilst the spleen showed mosaic appearance. The lymph nodes appeared enlarged, fluctuating and filled with purulent pus. The PLD inoculated group also showed gross changes in the lung represented by some spots of grey hepatization with no abscess formation in its parenchyma. The heart showed no obvious gross changes. The liver appeared congested with white line along the edges of almost all the liver lobes. The kidneys appeared congested with opaque spots on its capsule. The spleen appeared friable with mosaic appearance. The lymph nodes showed no abscess formation; however, when cut, it revealed a rusty yellow discoloration. Microscopic examination of *C. pseudotuberculosis* inoculated group showed significant congestion, oedema, and infiltration of inflammatory cells, degeneration and necrosis in the lungs, heart, liver, kidneys and spleen. The lymph nodes also showed significant granuloma formation with micro-foci of abscess formation. The PLD inoculated group showed significant congestion, oedema, degeneration and necrosis. In addition, hepatic tissue showed significant increase ($p < 0.05$) in Kupffer cells number. The lymph nodes showed no abscess formation with some haemosiderin deposits.

Therefore, from this study, it can be concluded that *C. pseudotuberculosis* had detrimental effects on testosterone concentration and reproductive organs in mice. However, in goats, *C. pseudotuberculosis* and PLD had negative effects on testosterone, estrogen and progesterone hormone concentrations as well as semen quality and reproductive organs in both genders. Acute phase reactant, mainly, haptoglobin, showed significant sensitivity in both treated groups; this sensitivity may have potential as diagnostic biomarker for CLA in goats. Similarly, *C. pseudotuberculosis* resulted in classical clinical manifestation of CLA specifically abscess formation in superficial lymph nodes, yet, PLD, showed different clinical manifestation with no abscessation. In addition, *C. pseudotuberculosis* and PLD showed significant changes in all haematological

parameters, yet, PLD showed different response pattern which may indicate a different mechanism of action. The gross and cellular changes were of typical CLA lesions in *C. pseudotuberculosis* inoculated group whilst PLD inoculated group showed less gross changes; however, the cellular changes were severe, this may indicate or reflect the toxic effects of PLD on tissues.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

ETIOPATOGENSIS PENYAKIT BISUL NODUS LIMFA PADA KAMBING

Oleh

ZAID KHUDHUR MAHMOOD

April, 2015

Pengerusi: Profesor Abd Wahid Haron, Ph.D.
Fakulti: Perubatan Veterinar

Caseous limfadenitis (CLA) adalah penyakit biri-biri dan kambing di seluruh dunia yang penting secara ekonomi. Walau bagaimanapun, kekurangan bahan penerbitan mengenai CLA pada kambing di Malaysia membawa kepada kajian ini untuk menyiasat mekanisme *Corynebacterium pseudotuberculosis* dan eksotoksinya, fosfolipase D (PLD) menjejaskan pembiakan dalam mencit dan kambing. Kajian ini juga bertujuan untuk menguji hipotesis bahawa protein fasa akut, khususnya, haptoglobin dan serum amiloid A boleh digunakan sebagai penanda biologi diagnostik untuk CLA.

Kedua-dua jantina untuk satu ratus lima ekor mencit klinikal yang sihat, berusia antara 10 hingga 12 minggu, berat 25-35 g serta dua puluh enam ekor kacukan kambing Boer klinikal yang sihat (13 jantan dan 13 betina) berusia antara 12 hingga 14 bulan, berat 30-35 kg digunakan dalam kajian ini. Betina tidak hamil, tidak menyusu dan ditempatkan berasingan daripada pejantan. Di awal eksperimen, dua jenis inokulum terlibat yang digunakan adalah *C. pseudotuberculosis* hidup dan eksotoksinya, PLD. Mencit telah dibahagikan kepada tiga kumpulan yang berjumlah 35 ekor setiap kumpulan. Kacukan kambing Boer juga dibahagikan kepada tiga kumpulan. Kumpulan pertama terdiri daripada 6 ekor kambing, kumpulan kedua terdiri daripada 10 ekor kambing dan kumpulan ketiga terdiri daripada 10 ekor kambing. Kumpulan kawalan 1 (mencit dan kambing) telah disuntik dengan fosfat steril mesin penimbang (PBS 1 ml / mencit antara peritoneal dan 1 ml / subkutaneus kambing), kumpulan 2 telah disuntik dengan *C. pseudotuberculosis* hidup (1×10^9 cfu antara peritoneal dalam mencit dan subkutaneus di kambing) manakala kumpulan 3 telah disuntik dengan PLD (1 ml / mouse antara

peritoneal dan 1 ml / 20 kg berat badan di kambing). Mencit telah diperhatikan sepanjang eksperimen, darah dikumpulkan ke dalam tiub heparinized untuk analisis hormon seks. Bedah siasat luka kasar telah dijalankan ke atas ovari, rahim, testis dan epididymis, dan sampel tisu telah ditetapkan untuk pemeriksaan histopatologi. Kambing juga diperhatikan semasa keseluruhan eksperimen untuk tanda-tanda klinikal dan koleksi siri darah telah dijalankan. Darah telah diletakkan ke dalam heparinized, polos, EDTA dan tiub natrium sitrat untuk protein fasa akut (aplikasi), hematologi dan analisis hormone seks. Bedah siasat luka kasar telah dijalankan ke atas paru-paru, jantung, hati, limpa, buah pinggang dan kelenjar limfa, dan sampel tisu telah ditetapkan untuk histopatologi serta perubahan sel dijangkingan.

Mencit yang disuntik dengan *C. pseudotuberculosis* menunjukkan penurunan yang signifikan ($p < 0.05$) dalam kepekatan testosterone. Kedua-dua kumpulan suntikan tidak menunjukkan sebarang perubahan kasar dalam organ-organ pembiakan. Walau bagaimanapun, pemeriksaan mikroskopik menunjukkan bahawa penginokulasian *C. pseudotuberculosis* menyebabkan kesesakan, edema, kemasukan keradangan sel, degenerasi dan nekrosis dalam ovari, rahim, testis dan epididymis. Manakala, kumpulan suntikan PLD menunjukkan kesesakan, degenerasi dan nekrosis dalam ovari, rahim, testis dan epididymis dan trombosis dalam ovari dan rahim sahaja.

Kambing yang disuntik dengan kedua-dua *C. pseudotuberculosis* dan PLD menunjukkan peningkatan yang signifikan ($p < 0.05$) dalam kepekatan haptoglobin dan penurunan yang signifikan ($p < 0.05$) dalam kepekatan albumin. Kambing juga menunjukkan peningkatan yang signifikan ($p < 0.05$) dalam suhu badan, kadar denyutan jantung, kadar pernafasan dalam waktu yang berbeza selepas penginokulasian dengan kedua-dua *C. pseudotuberculosis* dan PLD. Selain itu, skor badan menunjukkan penurunan yang signifikan ($p < 0.05$) dalam kedua-dua kumpulan yang dirawat pada akhir eksperimen. Selain itu, tapak inokulasi dan nodus limfa dangkal menunjukkan pembentukan nanah di kumpulan suntikan *C. pseudotuberculosis* sahaja.

Kambing yang disuntik dengan *C. pseudotuberculosis* menunjukkan penurunan yang signifikan ($p < 0.05$) dalam kiraan sel darah merah, hemoglobin, kepekatan creatine phosphokinase serta peningkatan yang signifikan ($p < 0.05$) dalam pengiraan neutrofil. Kambing yang disuntik kedua-dua *C. pseudotuberculosis* dan PLD menunjukkan peningkatan yang signifikan ($p < 0.05$) dalam jumlah sel yang padat, bermakna jumlah korpuskel, kiraan sel-sel darah putih, limfosit, monosit, eosinofil dan basophil dan aspartate transaminase, jumlah bilirubin, kalsium, kreatinin, gamma glutamyltransferase, urea, jumlah protein, globulin, kepekatan dehidrogenase laktat, masa prothrombin dan diaktifkan separa masa tromboplastin manakala kedua-dua kumpulan menunjukkan penurunan disuntik signifikan ($p < 0.05$) dalam kepekatan hemoglobin korpuskel min, kiraan neutrofil, transaminase alanina dan kepekatan fosfatase alkali.

PengInokulasian dengan PLD menunjukkan penurunan yang signifikan ($p < 0.05$) dalam kepekatan creatine phosphokinase.

Hormon testosteron, progesteron, estrogen, lilitan besar buah zakar dan isipadu, pH, corak gelombang, motilitis dan kepekatan air mani menunjukkan penurunan yang signifikan ($p < 0.05$) selepas pengInokulasian dengan *C. pseudotuberculosis* dan PLD. Di samping itu, sperma mati dan peratusan sperma tidak normal menunjukkan peningkatan yang signifikan ($p < 0.05$) dalam kedua-dua kumpulan dirawat. Pemeriksaan mikroskopik ovari menunjukkan pembentukan tisu pecah selepas diInokulasi dengan *C. pseudotuberculosis*, manakala untuk kumpulan suntikan PLD, ovari menunjukkan kesesakan teruk, degenerasi dan nekrosis sel stromal. Di samping itu, tisu rahim menunjukkan kesesakan, degenerasi dan nekrosis selepas diInokulasi dengan kedua-dua *C. pseudotuberculosis* dan PLD. Testis menunjukkan ketidakaturan struktur dan pengecutan tubul seminiferous dengan edema, degenerasi dan nekrosis sel spermatogonia, dan sel-sel Leydig selepas diInokulasi dengan kedua-dua *C. pseudotuberculosis* dan PLD. Begitu juga epididimis yang menunjukkan edema, degenerasi dan nekrosis lapisan epitelium daripada bedah siasat tubul epididymal yang diInokulasi dengan kedua-dua *C. pseudotuberculosis* dan PLD.

Kumpulan yang disuntik *C. pseudotuberculosis* menunjukkan perubahan kasar dalam paru-paru diwakili oleh hepatization kelabu dan merah, yang lebih mendalam di tempat kesesakan dan pembentukan nanah. Jantung menjadi oedematous dan perikardium menunjukkan beberapa tahap kelegapan. Hati menunjukkan pelbagai pembentukan nanah di permukaannya. Buah pinggang menjadi rapuh dengan kapsul legap manakala limpa menunjukkan penampilan mozek. Nodus limfa menjadi besar, turun naik dan dipenuhi dengan nanah. Kumpulan suntikan PLD juga menunjukkan perubahan kasar dalam paru-paru yang diwakili oleh beberapa tempat di hepatization kelabu tanpa pembentukan nanah di parenchyma itu. Jantung tidak menunjukkan sebarang perubahan kasar yang jelas. Hati kelihatan sesak dengan garis putih hampir di sepanjang semua cuping hati. Buah pinggang kelihatan sesak dengan tempat legap pada kapsul itu. Limpa menjadi rapuh dengan penampilan mozek. Nodus limfa tidak menunjukkan sebarang pembentukan nanah; Walau bagaimanapun, apabila dipotong, ia memperlihatkan perubahan warna kuning berkarat. Pemeriksaan mikroskopik ke atas kumpulan yang disuntik oleh *C. pseudotuberculosis* menunjukkan kesesakan yang ketara, edema, dan penyusupan keradangan sel, degenerasi dan nekrosis dalam paru-paru, jantung, hati, buah pinggang dan limpa. Nodus limfa juga menunjukkan pembentukan granuloma signifikan dengan tumpuan mikro-pembentukan nanah. Kumpulan suntikan PLD menunjukkan kesesakan yang ketara, edema, degenerasi dan nekrosis. Di samping itu, tisu hepatik menunjukkan peningkatan yang signifikan ($p < 0.05$) dalam bilangan sel Kupffer. Nodus limfa juga menunjukkan pembentukan abses dengan beberapa deposit haemosiderin.

Daripada kajian ini, dapat disimpulkan bahawa *C. pseudotuberculosis* mempunyai kesan ke atas kepekatan testosteron dan organ-organ pembiakan pada mencit. Walau bagaimanapun, dalam kambing, *C. pseudotuberculosis* dan PLD mempunyai kesan negatif ke atas kepekatan hormone testosteron, estrogen progesteron dan kualiti air mani serta organ-organ pembiakan kedua-dua jantina. Bahan tindak balas fasa akut, terutamanya, haptoglobin, menunjukkan sensitiviti yang ketara dalam kedua-dua kumpulan yang dirawat; kepekaan ini mungkin mempunyai potensi sebagai penanda bio diagnostik untuk CLA dalam kambing. Begitu juga *C. pseudotuberculosis* yang menyebabkan manifestasi klinikal klasik CLA khusus untuk pembentukan nanah di nod limfa cetek, namun, PLD, menunjukkan manifestasi klinikal yang berbeza tanpa pembentukan nanah. Di samping itu, *C. pseudotuberculosis* dan PLD menunjukkan perubahan ketara dalam semua parameter hematologi, namun, PLD menunjukkan corak tindak balas yang berbeza yang mungkin menunjukkan mekanisme yang berbeza tindakan. Perubahan kasar dan selular adalah biasa bagi luka CLA dalam kumpulan suntikan *C. pseudotuberculosis* manakala kumpulan suntikan PLD menunjukkan kurang perubahan kasar; Walau bagaimanapun, perubahan sel adalah teruk dan ini mungkin mencerminkan kesan toksik PLD pada tisu.

ACKNOWLEDGEMENTS

I would give the praise to Allah almighty, the most beneficent the most merciful who keep inspiring me, guiding me and always looking after me and directing me toward the utmost goodness.

I also would like to express my sincere gratitude and appreciations to my supervisor Professor Dr. Abd Wahid Haron for the priceless guidance, supervision and the encouragement throughout the research period. Many thanks and gratitude also goes to the supervisory committee for their guidance, advice and supervision starting with Professor Dr. Abdul Aziz Saharee, Professor Dr. Jasni Sabri, Associate Professor Dr. Rosnina Hj Yusoff and last but never least Dr. Faez Firdaus Jesse Abdullah for his continuous support and help. Many thanks also go to Mr. Yap Keng Chee, Mr. Ganesanmurthi Perumal, Mr. Mohd Jefri Norsidin and Mr. Mohd Fahmi Mashuri. I would like to express my utmost appreciation and gratitude to Universiti Putra Malaysia, School of Graduate Studies and Ministry of Higher Education for giving me the opportunity to pursue this study.

My thanks and appreciations also go to Taman Pertanian Universiti, UPM, Faculty of Veterinary Medicine, UPM, Department of Veterinary Clinical Studies and Department of Microbiology and Pathology, Faculty of Veterinary Medicine for providing me with all the facilities pertaining to my research. This study was funded by the Research University Grant Scheme (RUGS), Universiti Putra Malaysia.

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	v
ACKNOWLEDGEMENTS	ix
APPROVAL	x
DECLARATION	xii
LIST OF TABLES	xviii
LIST OF FIGURES	xix
LIST OF ABBREVIATIONS	xxvii
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	4
2.1 History of <i>Corynebacterium pseudotuberculosis</i>	4
2.2 <i>Corynebacterium pseudotuberculosis</i> attributes	4
2.2.1 Organism characteristics	4
2.2.2 Biochemical characteristics	5
2.2.3 Chemical resistance	5
2.3 Virulence factors of <i>Corynebacterium pseudotuberculosis</i>	6
2.3.1 Phospholipase D (PLD)	6
2.3.2 Mycolic acid	7
2.4 Prevalence of caseous lymphadenitis	8
2.5 Sources of caseous lymphadenitis infection	8
2.5.1 Primary source of infection	8
2.5.2 Secondary source of infection	9
2.6 Zoonosis of <i>Corynebacterium pseudotuberculosis</i>	9
2.7 Pathogenesis of caseous lymphadenitis	9
2.7.1 Pathology of caseous lymphadenitis	10
2.7.2 Haematology	11
2.8 Diagnosis of caseous lymphadenitis disease	12
2.9 Acute phase proteins	13
2.10 Types of acute phase proteins	14
2.11 Pathology of acute phase proteins	14
2.12 Acute phase proteins in ruminants	15
2.12.1 Haptoglobin (Hp)	15
2.12.2 Serum amyloid A (SAA)	16
2.13 Reproductive performance	16
2.14 Physiology and endocrinology of reproduction in goat	17
2.15 Reproductive anatomy of the buck	17
2.16 Reproductive anatomy of the doe	18
2.17 Reproductive endocrinology of the buck	19
2.17.1 Hormones	19
2.17.2 Spermatogenesis	19
2.18 Reproductive endocrinology of the doe	20
2.18.1 Hormones	20
2.18.2 Folliculogenesis	20
2.19 Bacterial disruption of reproduction	20

2.20	Caseous lymphadenitis and reproduction	21
2.21	Treatment of caseous lymphadenitis	22
2.22	Vaccination against caseous lymphadenitis	22
3	REPRODUCTIVE HORMONES AND ORGANS OF MICE TREATED WITH <i>Corynebacterium pseudotuberculosis</i> AND PHOSPHOLIPASE D	24
3.1	Introduction	24
3.2	Materials and methods	25
	3.2.1 isolation and identification of <i>Corynebacterium pseudotuberculosis</i>	25
	3.2.2 Phospholipase D	25
	3.2.3 Mice	25
	3.2.3 Inocula of the mice	25
	3.2.4 Hormone analysis	26
	3.2.4.a Testosterone ELISA assay	26
	3.2.4.b Estradiol ELISA assay	26
	3.2.4.c Progesterone ELISA assay	27
3.3	Results	28
	3.3.1 Reproductive hormones	28
	3.3.2 Histopathology of the reproductive organs	29
3.4	Discussion and conclusion	41
4	CLINICAL SIGNS, BODY SCORES AND LYMPH NODES OF GOATS TREATED WITH <i>Corynebacterium pseudotuberculosis</i> AND PHOSPHOLIPASE D	44
4.1	Introduction	44
4.2	Materials and methods	45
	4.2.1 Goats	45
	4.2.2 Inocula of the goats	45
	4.2.3 Clinical signs	45
4.3	Results	47
	4.3.1 Temperature	50
	4.3.2 Heart rate	51
	4.3.3 Respiratory rate	52
	4.3.4 Body score	53
	4.3.5 Lymph nodes	54
4.4	Discussion and conclusion	55
5	HAEMOGRAM, COAGULATION TIME AND BLOOD BIOCHEMICAL PARAMETERS OF GOATS TREATED WITH <i>Corynebacterium pseudotuberculosis</i> AND PHOSPHOLIPASE D	58
5.1	Introduction	58
5.2	Materials and methods	59
	5.2.1 Haematological and blood biochemical analysis	59
5.3	Results	59
	5.3.1 Red blood cells count	60
	5.3.2 Haemoglobin concentration	60
	5.3.3 Packed cell volume	61

5.3.4	Mean corpuscular volume	62
5.3.5	Mean corpuscular haemoglobin concentration	63
5.3.6	White blood cells count	64
5.3.7	Neutrophil count	66
5.3.8	Lymphocyte count	67
5.3.9	Monocyte count	68
5.3.10	Eosinophil count	69
5.3.11	Basophil count	70
5.3.12	Alanine transaminase concentration	71
5.3.13	Alkaline phosphatase concentration	72
5.3.14	Aspartate transaminase concentration	73
5.3.15	Total bilirubin concentration	74
5.3.16	Calcium concentration	75
5.3.17	Creatinine concentration	76
5.3.18	Creatine phosphokinase concentration	77
5.3.19	Gamma glutamyltransferase concentration	78
5.3.20	Urea concentration	79
5.3.21	Total protein concentration	80
5.3.22	Globulin concentration	81
5.3.23	Lactate dehydrogenase concentration	82
5.3.24	Prothrombin time	83
5.3.25	Activated partial thromboplastin time	84
5.4	Discussion and conclusion	84
6	ACUTE PHASE PROTEINS OF GOATS TREATED WITH <i>Corynebacterium pseudotuberculosis</i> AND PHOSPHOLIPASE D	93
6.1	Introduction	93
6.2	Materials and methods	94
6.2.1	Acute phase proteins analyses	94
6.2.1.a	Estimation of haptoglobin	94
6.2.1.b	Estimation of serum amyloid A	94
6.3	Results	95
6.3.1	Haptoglobin concentration	95
6.3.2	Serum amyloid A concentration	95
6.3.3	Albumin concentration	96
6.4	Discussion and conclusions	97
7	REPRODUCTIVE PARAMETERS AND ORGANS OF GOATS TREATED WITH <i>Corynebacterium pseudotuberculosis</i> AND PHOSPHOLIPASE D	100
7.1	Introduction	100
7.2	Materials and methods	101
7.2.1	Hormone analysis	101
7.2.1.a	Estradiol RIA assay	101
7.2.1.b	Progesterone RIA assay	102
7.2.1.c	Testosterone RIA assay	102
7.2.2	Scrotal circumferences (SC)	102
7.2.3	Semen collection	102
7.2.4	Semen evaluation	103
7.2.4.1	Wave pattern	103

7.2.4.2	Sperm motility	103
7.2.4.3	Sperm concentration	103
7.2.4.4	Sperm live/dead percentage	103
7.2.4.5	Sperm morphology	103
7.3	Results	104
7.3.1	Estrogen concentration	104
7.3.2	Progesterone concentration	104
7.3.3	Testosterone concentration	105
7.3.4	Scrotal circumference (cm)	106
7.3.5	Semen volume	108
7.3.6	Semen pH	108
7.3.7	Semen wave pattern	109
7.3.8	Sperm motility	110
7.3.9	Sperm concentration	111
7.3.10	Dead/ live sperm	112
7.3.11	Abnormal sperm morphology	113
7.3.12	Histopathology of the reproductive organs	115
7.4	Discussion and conclusion	127
8	ORGANS HISTOLOGY AND GROSS PATHOLOGY OF GOATS TREATED WITH <i>Corynebacterium pseudotuberculosis</i> AND PHOSPHOLIPASE D	131
8.1	Introduction	131
8.2	Materials and methods	132
8.2.1	Post mortem examination	132
8.2.2	Histopathology	132
8.3	Results	133
8.3.1	The lung	133
8.3.2	The heart	139
8.3.3	The liver	144
8.3.4	The kidney	150
8.3.5	The spleen	155
8.3.6	The lymph nodes	160
8.4	Discussion and conclusion	170
9	GENERAL DISCUSSION AND CONCLUSION	177
	REFERENCES	183
	APPENDICIES	206
	BIODATA OF STUDENT	209
	LIST OF PUBLICATIONS	210

LIST OF TABLES

Table		Page
8.1	Lesion parameters determined in the tissues of treated goats.	133
8.2	Mean score of cellular changes in the lung of goats post inoculation with the <i>Corynebacterium pseudotuberculosis</i> and the phospholipase D.	137
8.3	Mean score of cellular changes in the heart of goats post inoculation with the <i>Corynebacterium pseudotuberculosis</i> and the phospholipase D.	142
8.4	Mean score of cellular changes in the liver of goats post inoculation with the <i>Corynebacterium pseudotuberculosis</i> and the phospholipase D.	147
8.5	score of cellular changes in the kidney of goats post inoculation with the <i>Corynebacterium pseudotuberculosis</i> and the phospholipase D.	153
8.6	Mean score of cellular changes in the spleen of goats post inoculation with the <i>Corynebacterium pseudotuberculosis</i> and the phospholipase D.	159
8.7	Mean score of cellular changes in the lymph nodes of goats post inoculation with the <i>Corynebacterium pseudotuberculosis</i> and the phospholipase D.	165

LIST OF FIGURES

Figure		Page
3.1	Flowchart of the experimental design in a mouse model.	28
3.2	Reproductive hormone concentrations in mice post inoculation with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=35 mice per group.	29
3.3	Ovary of mouse from control group, notice the ovum (white arrow), granulose cells (yellow arrow), cumulus oophorus (orange arrow), corona radiata (red arrow) and the antrum (black arrow); H&E 400X.	30
3.4	Ovary of mouse inoculated with <i>Corynebacterium pseudotuberculosis</i> shows infiltration of inflammatory cells (white arrow) into the lumen of the follicle. A developed ova (red arrow) surrounded by the antrum and multiple layers of corona radiata cells, the main source of estrogen hormone (yellow arrow); H&E 400X.	31
3.5	Ovary of mouse inoculated with phospholipase D shows generalized congestion (white arrow), thrombus formation (yellow arrow), degeneration and necrosis (red arrow) of the stromal cells; H&E 400X.	32
3.6	Uterus of mouse from control group, notice the uterine stromal cells (white arrow), uterine glands (yellow arrow) and the muscular layer (orange arrow); H&E 200X.	33
3.7	Uterus of mouse inoculated with <i>Corynebacterium pseudotuberculosis</i> shows severe congestion (white arrow), infiltration of inflammatory cells (yellow arrow), vacuolar degeneration (orange arrow) and necrosis (red arrow); H&E 400X.	34
3.8	Uterus of mouse inoculated with phospholipase D shows generalized congestion (white arrow), thrombus formation (yellow arrow), degeneration (orange arrow) and necrosis of the muscular cells (red arrow); H&E 400X.	35
3.9	Testes of mouse from control group, notice the spermatogenic cells (white arrow), spermatids (yellow arrow) and the Leydig cells (orange arrow); H&E 400X.	36
3.10	Testes of mouse inoculated with <i>Corynebacterium pseudotuberculosis</i> shows infiltration of inflammatory cells (white arrow), congestion (yellow arrow), degeneration (orange arrow) and necrosis of some spermatogonia (red arrow); H&E 400X.	37
3.11	Testes of mouse inoculated with phospholipase D shows congestion (white arrow), oedema (yellow arrow) and degeneration of some spermatogonia (orange arrow); H&E 200X.	38
3.12	Epididymis of mouse from control group, notice the epithelial cells of the epididymal tubules (white arrow) and the sperms (yellow arrow); H&E 200X.	39
3.13	Epididymus of mouse inoculated with <i>Corynebacterium pseudotuberculosis</i> show infiltration of inflammatory cells	40

	(white arrow), congestion (yellow arrow), degeneration (orange arrow) and necrosis of tubular epithelia (red arrow); H&E 200X.	
3.14	Epididymus of mouse inoculated with phospholipase D shows hemorrhage inside the lumen of the epididymal tubules (white arrow), degeneration (yellow arrow) necrosis of the epithelial cells of the epididymal tubules (orange arrow) and interstitial oedema (red arrow); H&E 200X.	41
4.1	Flowchart of experimental design in goat	46
4.2	Inoculation site of <i>Corynebacterium pseudotuberculosis</i> shows an open abscess with purulent discharge	48
4.3	Lumpy lesion on buck ear inoculated with phospholipase D	49
4.4	Eye swelling with purulent discharge in phospholipase D inoculated bucks	50
4.5	Temperature (oC) of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	51
4.6	Heart rate (bpm) of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	52
4.7	Respiratory rate (breath/min) of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	53
4.8	Body score of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	54
4.9	Affected lymph nodes in <i>Corynebacterium pseudotuberculosis</i> inoculated animals; picture on the left shows a parotid lymph node (arrow) and on the right shows submandibular lymph node (arrow).	55
5.1	Red blood cell count of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	60
5.2	Haemoglobin concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	61
5.3	Packed cell volume of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	62
5.4	Mean corpuscular volume of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	63
5.5	Mean corpuscular haemoglobin concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	64
5.6	White blood cell count of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	65
5.7	Neutrophil count of the goats post inoculated with	66

	<i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	
5.8	Lymphocyte count of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	67
5.9	Monocyte count of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	68
5.10	Eosinophil count of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	69
5.11	Basophil count of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	70
5.12	Alanine transaminase concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	71
5.13	Alkaline phosphatase concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	72
5.14	Aspartate transaminase concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	73
5.15	Total bilirubin concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	74
5.16	Calcium concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	75
5.17	Creatinine concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	76
5.18	Creatine phosphokinase concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	77
5.19	Gamma Glutamyltransferase concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	78
5.20	Urea concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	79
5.21	Total protein concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	80
5.22	Globulin concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	81
5.23	Lactate dehydrogenase concentration of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	82
5.24	Prothrombin time of the goats post inoculated with	83

	<i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	
5.25	Activated partial thromboplastin time of the goats post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	84
6.1	Serum haptoglobin concentration of the goats post inoculated with the <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	95
6.2	Serum amyloid A concentration of the goats post inoculated with the <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	96
6.3	Albumin concentration of the goats post inoculated with the <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=26.	97
7.1	Estrogen concentration of the does post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=13.	104
7.2	Progesterone concentration of the does post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=13.	105
7.3	Testosterone concentration of the bucks post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=13.	106
7.4	Scrotal circumference of the bucks post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=13.	107
7.5	Testes of the bucks showing their sizes at the end of experiment period (week 12). <i>Corynebacterium pseudotuberculosis</i> inoculated animals (A), phospholipase D inoculated animals (B) and the control (C).	107
7.6	Semen volume of the bucks post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=13.	108
7.7	Semen pH of the bucks post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=13.	109
7.8	Semen wave pattern of the bucks post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=13.	110
7.9	Sperm motility of the bucks post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=13.	111
7.10	Sperm concentration of the bucks post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=13.	112
7.11	Percentage of dead/live sperm of the bucks post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=13.	113
7.12	Percentage of abnormal sperm morphology of the bucks post inoculated with <i>Corynebacterium pseudotuberculosis</i> and phospholipase D; n=13.	114

7.13	Different types of abnormal sperm morphology from the <i>Corynebacterium pseudotuberculosis</i> inoculated bucks; Nigrosin-Eosin stain 1000X. Spiral head (A), tapered sperm (B), coiled tail and decapitated heads (C), double forms (D), distal protoplasmic droplet (E) and broken neck (F).	114
7.14	Different types of abnormal sperm morphology from the PLD inoculated bucks; Nigrosin-Eosin stain 1000X. Detached knobbed head (A), distal protoplasmic droplet with bent tail (B), sterilizing tail stump (C), thickened midpiece (D), proximal protoplasmic droplet with coiled tail, (E) and distal droplet with bent tail (F).	115
7.15	Ovary of doe from control group, notice the ovum (white arrow), granulosa cells (yellow arrow), cumulus oophorus (orange arrow), corona radiata (red arrow) and the antrum (black arrow); H&E 200X.	116
7.16	Ovary shows proliferation of fibroblasts in the stromal area (red arrow) 12 weeks post inoculation with the <i>Corynebacterium pseudotuberculosis</i> ; H&E 400X.	117
7.17	Ovary shows severe congestion (white arrow), degeneration and necrosis (red arrow) 12 weeks post inoculation with the phospholipase D; H&E 400X.	118
7.18	Uterus of doe from control group, notice the uterine stromal cells (white arrow) and uterine glands (yellow arrow); H&E 200X.	119
7.19	Uterus shows congestion (white arrow), degeneration and necrosis (red arrow) 12 weeks post inoculation with the <i>Corynebacterium pseudotuberculosis</i> ; H&E 400X.	120
7.20	Uterus shows congestion (white arrow), degeneration and necrosis (red arrow) 12 weeks post inoculation with the phospholipase D; H&E 400X.	121
7.21	Testes of buck from control group, notice the spermatogenic cells (white arrow), spermatids (yellow arrow) and the Leydig cells (orange arrow); H&E 400X.	122
7.22	Testes shows irregular and shrinkage of the seminiferous tubules (red arrow), oedema (orange arrow), degeneration and necrosis (yellow arrow), fewer spermatides inside the lumen of the seminiferous tubules (black arrow) and necrosis of Leydig cells (white arrow) 12 weeks post inoculation with the <i>Corynebacterium pseudotuberculosis</i> ; H&E 400X.	123
7.23	Testes shows irregular and shrinkage of the seminiferous tubules (red arrow), severe congestion (white arrow), oedema (orange arrow), degeneration and necrosis (yellow arrow), and fewer spermatides inside the lumen of the seminiferous tubules (black arrow) 12 weeks post inoculation with the phospholipase D; H&E 400X.	124
7.24	Epididymis of buck from control group, notice the epithelial cells of the epididymal tubules (white arrow) and the sperms (yellow arrow); H&E 200X.	125
7.25	Epididymus shows oedema (white arrow), degeneration and necrosis of the lining epithelia of the epididymal tubules (red	126

- arrow), and morphologically abnormal sperms (orange arrow) inside the epididymal tubules 12 weeks post inoculation with the *Corynebacterium pseudotuberculosis*; H&E 400X.
- 7.26 Epididymus shows degeneration and necrosis (red arrow) of the lining epithelia of the epididymal tubules, morphologically abnormal sperms (orange arrow) inside the epididymal tubules and oedema (white arrow) 12 weeks post inoculation with the phospholipase D; H&E 400X. 127
- 8.1 Lung show disseminated mosaic discoloration (arrow) and congestion deeper inside (arrows head) 12 weeks post inoculation with the *Corynebacterium pseudotuberculosis*. 134
- 8.2 Lung show a thick whitish and lumpy pus discharge (arrow) 12 weeks post inoculation with *Corynebacterium pseudotuberculosis*. 135
- 8.3 lung from control group, notice the normal bronchioles (white arrow), the lining epithelium (yellow arrow) and the alveolar tissue (red arrow); H&E; 200X. 136
- 8.4 Lung show oedema (white arrow), congestion (yellow arrow) microfoci of abscess formation (orange arrow), degeneration and Necrosis (red arrow) 12 weeks post inoculation with *Corynebacterium pseudotuberculosis*; H&E; 200X. 137
- 8.5 Lung show light (arrow) and dark (arrows head) spots of gray hepatization 12 weeks post inoculation with the phospholipase D. 138
- 8.6 Lung show oedema (white arrow), congestion (yellow arrow) infiltration of inflammatory cells (orange arrow), degeneration and necrosis (red arrow) and hyperplasia of bronchiolar epithelium (black arrow) 12 weeks post inoculation with the phospholipase D; H&E; 400X. 139
- 8.7 Heart show opacity of the pericardium (arrow) 12 weeks post inoculation with the *Corynebacterium pseudotuberculosis*. 140
- 8.8 Heart muscle from control group, notice the normal structure of the cardiac fibers (white arrow); H&E; 200X. 141
- 8.9 Heart muscle show oedema (white arrow), degeneration and necrosis (yellow arrow) and infiltration of inflammatory cells (orange arrow) 12 weeks post inoculation with *Corynebacterium pseudotuberculosis*; H&E; 400X. 142
- 8.10 Heart appeared normal 12 weeks post inoculation with the phospholipase D. 143
- 8.11 Heart muscles show oedema (red arrow), congestion (yellow arrow) degeneration and necrosis (orange arrow) and infiltration of inflammatory cells (black arrow) 12 weeks post inoculation with the phospholipase D; H&E; 400X. 144
- 8.12 Liver show multiple abscess formation (arrow). Notice the fibrosis surrounding the abscesses 12 weeks post inoculation with the *Corynebacterium pseudotuberculosis*. 145
- 8.13 Liver from control group, notice the central vein (yellow arrow), hepatocytes (orange arrow) and the hepatic sinusoids (red arrow); H&E; 400X. 146

- 8.14 Liver show oedema (yellow arrow), congestion (white arrow), infiltration of the inflammatory cells (orange arrow), degeneration and necrosis (red arrow) and presence of Kupffer cells (black arrow) 12 weeks post inoculation with the *Corynebacterium pseudotuberculosis*; H&E; 400X. 147
- 8.15 Liver appeared dark in colour and the gall bladder (arrow) was empty 12 weeks post inoculation with the phospholipase D. 148
- 8.16 Liver show whitish colour line (arrow) along the edges of the lobe 12 weeks post inoculation with the phospholipase D. it's believed that the white line is a zone of necrosis. 149
- 8.17 Liver show oedema (yellow arrow), congestion (orange arrow), haemorrhage (red arrow), degeneration and necrosis (green arrow) and Kupffer cells (black arrow) post inoculation with phospholipase D; H&E; 400X. 150
- 8.18 Kidney show the opacity of its capsule (arrow), 12 weeks post inoculation with the *Corynebacterium pseudotuberculosis*. 151
- 8.19 Kidney from control group, notice the normal structure of the glomerulus (red arrow) and the renal tubules (yellow arrow); H&E; 400X. 152
- 8.20 Kidney show oedema (white arrow), congestion (yellow arrow), mild haemorrhage (orange arrow), infiltration of inflammatory cells (red arrow), degeneration and necrosis (black arrow) 12 weeks post inoculation with the *Corynebacterium pseudotuberculosis*; H&E; 400X. 153
- 8.21 Kidney show some opaque spots (arrow) on its surface 12 weeks post inoculation with the phospholipase D. 154
- 8.22 Kidney show oedema (white arrow), congestion (yellow arrow), mild haemorrhage (orange arrow), infiltration of inflammatory cells (red arrow), degeneration and necrosis (green arrow) 12 weeks post inoculation with the phospholipase D; H&E; 400X. 155
- 8.23 Spleen show some mosaic discoloration (arrow) on its surface post inoculation with the *Corynebacterium pseudotuberculosis*. 156
- 8.24 Spleen show opaque capsule (arrow) and it revealed dark mosaic discoloration (arrow head) post inoculation with the phospholipase D. 157
- 8.25 Spleen from control group, notice the normal structure of the red pulp (white arrow), the white pulp (yellow arrow) and the trabecula (orange arrow); H&E; 200X. 158
- 8.26 Spleen show sever inflammatory reaction with high population of lymphocyte and macrophage aggregation (yellow arrow) 12 weeks post inoculation with the *Corynebacterium pseudotuberculosis*; H&E; 400X. 159
- 8.27 Spleen show mild inflammatory reaction with mild population of lymphocyte and macrophage aggregation (yellow arrow), oedema (orange arrow), degeneration and necrosis (red arrow) of the red pulp 12 weeks post inoculation with the phospholipase D; H&E; 400X. 160

- 8.28 Prescapular lymph node show congested medulla (arrow) and pus oozing from the cortex area (arrow head) 12 weeks post inoculation with the *Corynebacterium pseudotuberculosis*. 161
- 8.29 Prefemoral lymph node show a thick and whitish caseous discharge (arrow) 12 weeks post inoculation with the *Corynebacterium pseudotuberculosis*. 162
- 8.30 Prescapular lymph node show normal appearance (arrow) 12 weeks post inoculation with the phospholipase D. 163
- 8.31 Prescapular lymph node shows some rusty yellow colour inside the lymph node (arrow) post inoculation with phospholipase D. 164
- 8.32 Lymph node from control group, notice the cortex area (yellow arrow) and the medulla area (orange arrow); H&E; 200X. 166
- 8.33 Lymph node show oedema (yellow arrow), infiltration of vast numbers of lymphocytes and macrophages marking an area of microfoci of abscess formation (orange arrow) 12 weeks post inoculation with the *Corynebacterium pseudotuberculosis*; H&E; 400X. 167
- 8.34 Lymph node show haemorrhage (yellow arrow), degeneration and necrosis (orange arrow) of the lymphatic tissue 12 weeks post inoculation with the *Corynebacterium pseudotuberculosis*; H&E; 400X. 168
- 8.35 Lymph node show oedema (white arrow), congestion (yellow arrow), degeneration and necrosis (red arrow) of the lymphatic tissue 12 weeks post inoculation with the phospholipase D; H&E; 400X. 169
- 8.36 Lymph node show scattered haemosiderin (arrow) deposits in the lymphatic tissue 12 weeks post inoculation with the phospholipase D; H&E; 400X. 170

LIST OF ABBREVIATIONS

°C	Degree celsius
cfu	Colony forming unit
CLA	Caseous lymphadenitis
RIA	Radioimmunoassay
PLD	Phospholipase D
APP	Acute phase proteins
AGID	Agar gel immunodiffusion
ELISA	Enzyme Linked Immunosorbent Assay
S.C	Subcutaneous
I.V	Intravenous
IP	Intraperitoneal
DVS	Department of Veterinary Services
Hp	Haptoglobin
OIE	Office of International Epizootic
PBS	Phosphate Buffered Saline
SAA	Serum amyloid A
UPM	Universiti Putra Malaysia
VLSU	Veterinary Laboratories Service Unit

CHAPTER 1

INTRODUCTION

Small ruminant industry, especially goats, in Malaysia is the agricultural sector by the government to increase the goat population and subsequently production of meat. In 2012, the population of sheep and goats were estimated at 129,850 and 482,280 heads respectively (<http://www.dvs.gov.my>). Caseous lymphadenitis is a chronic disease caused by *Corynebacterium pseudotuberculosis*, a pathogen that infects small ruminant and characterized by abscess formation affecting one or more lymph nodes. However, *C. pseudotuberculosis* can also infect other animals such as cattle, horses, mules, camels, deer, alpacas, but rarely man with a wide range of diseases specific to each species (Yeruham et al., 1997; Connor et al., 2000; Williamson, 2001; Abou-Zaid, 2001; Dorella et al., 2006; Paton, 2010; Guimarães et al., 2011a). *Corynebacterium pseudotuberculosis*, less frequently can also cause orchitis, mastitis, stillbirth, abortion, arthritis, hepatitis, subcutaneous abscesses and pneumonia in small ruminants (Junior et al., 2006; Radostits et al., 2007; Fontaine and Baird, 2008; Paton, 2010).

Caseous lymphadenitis occurs in all major sheep and goat rearing areas across the globe especially in countries with a large sheep population such as Australia and New Zealand. Farmers suffer from heavy economic losses after affected carcasses were degraded and condemned at slaughter and during meat inspection in abattoirs as well as death among affected animals in the farms (Paton, 1990; Williamson, 2001). Despite the significant economic losses and potential zoonoses, the scientific world has yet to come up with an effective vaccine.

The goat's industry in Malaysia partially depends on smallholders contributing to increase the goat population and meat production. These farms are plagued by CLA, the main cause of financial losses to the farmers. However, a survey conducted in East Coast Economic Region (ECER) in Peninsular Malaysia showed that the prevalence of CLA among small ruminant populations in ECER was 11.12% (Abdinasir et al., 2012). The pathogen, *C. pseudotuberculosis* can be identified based on two virulence factors, phospholipase D (PLD) and mycolic acids. Phospholipase D is a potent exotoxin of *C. pseudotuberculosis* and it plays a key role in the development and pathogenesis of CLA (Baird and Fontaine, 2007).

Phospholipase D hydrolyses the sphingomyelin in mammalian cell membrane increasing the vascular permeability especially the endothelial layer, leading to plasma protein leakage from the blood into the surrounding tissue space and from there into the lymphatic system. This helps the dissemination of *C. pseudotuberculosis* from the primary infection location to other parts of the

animal's body (Guimarães et al., 2011a). The notion that PLD is a significant virulence factor has been tested when some isolates of *C. pseudotuberculosis* that had their *pld* genes encoding PLD removed were not capable of causing abscess formation in the lymph nodes of CLA infected sheep (McNamara et al., 1994).

Acute phase proteins (APPs), mainly haptoglobin (Hp) and serum amyloid A (SAA) are appropriate indispensable parameters that can be used objectively and rapidly to evaluate animal health and well-being at any time which make it valuable tools for screening the animal's health status (Murata et al., 2004; Lakota et al., 2011; Piñeiro et al., 2013). They are frequently measured for diagnostic and predictive purposes (Vreugdenhil et al., 1999; Lakota et al., 2011). Hence, quantification of APPs has been established for clinical and research purposes to provide information on APPs behavior and their response mechanisms (Eckersall et al., 1999).

Caseous lymphadenitis has an insidious effect on an animal's fertility with eventual reduction in the reproductive efficiency as well as meat and milk production. Goldberger et al (1981) have isolated *C. pseudotuberculosis* in raw milk of sheep infected with CLA. Some lambs have infected by CLA through suckling their infected dams (Baird et al., 2004).

Little is known about the effects of CLA on reproduction performance in goats (Williamson, 2001; Conner et al., 2000). The reproductive organs that are mainly affected are the epididymis and the mammary gland in sheep and goats (Unanian et al., 1985). Valli and Parry (1993) claimed that the testes and uterus in sheep can also be affected by CLA. Caseous lymphadenitis abscesses have been found in the inguinal and scrotal lymph nodes of rams, but not in the epididymis neither in the testes nor semen (Williamson and Nairn, 1980). The lack of the knowledge and the inconsistent reports in regard to CLA and its impact on reproduction of sheep and goats has led to the current study focusing on the reproductive characteristic of the goats during the course of CLA.

Generally, there is scanty information of the haemogram and APPs response during the course of CLA in goats. There is also no specific report on the hormonal profile (testosterone, progesterone, estrogen) and histopathological changes of reproductive and visceral organs in the goats challenged with *C. pseudotuberculosis* and its exotoxin, PLD. Up-to-date, there is no documentation, report or any citation on pathophysiological changes of the reproductive system in both male and female goats challenged with *C. pseudotuberculosis* and PLD. Thus, this study was designed with the intention for better understanding of the haematological, biochemical, histopathological and hormonal changes that may revolve in the challenged host (mice and goat). Consequently, this will assist to improve our knowledge regarding ethiopathogenesis of CLA in goats.

Problem statement:

1. Caseous lymphadenitis is a chronic disease and difficult to diagnose at an early stage. Acute phase proteins have a significant, sensitive and valid role as diagnostic biomarkers for subclinical, acute and chronic conditions. Therefore, it may be useful to incorporate in the CLA diagnosis in goats.
2. Caseous lymphadenitis also has an insidious effect on reproduction which eventually affects the productivity of the goat. Studies have been reporting CLA lesions in the reproductive organs which influenced the reproductive efficiency in both males and females.

Hypothesis:

Acute phase proteins are more useful for monitoring health than cytokines because the latter have short half-life (few hours) compare to acute phase reactants. Acute phase proteins may have role as biomarkers for chronic conditions because of their presence in circulation can persist beyond the immediate infection time and as the lesion become chronic.

Thus, the objectives of the current study were:

1. To estimate the sex hormones (testosterone, progesterone, estrogen) responses between the infection by *C.pseudotuberculosis* and PLD inoculation in mouse model.
2. To observe the clinical signs between infection by *C. pseudotuberculosis* and PLD inoculated goats.
3. To measure the changes in the haemogram between the infection by *C.pseudotuberculosis* and PLD inoculated goats.
4. To estimate the acute phase proteins responses between infection by *C. pseudotuberculosis* and PLD inoculated goats.
5. To estimate the sex hormones (testosterone, progesterone, estrogen) responses between the infection by *C.pseudotuberculosis* and PLD inoculated goats.
6. To evaluate the histopathological changes of visceral organs, reproductive organs (testes, epididymus, ovary, uterus) and lymph nodes between the infection by *C.pseudotuberculosis* and PLD inoculated goats.

REFERENCES

- Abdinasir, Y.O., Jesse, F.F A. and Abdul Aziz, B.S. (2012). Sero-Prevalence of Caseous Lymphadenitis Evaluated by Agar Gel Precipitation Test among Small Ruminant Flocks in East Coast Economic Regions in Peninsular Malaysia. *Journal of Animal and Veterinary Advances* 11 (19): 3474-3480.
- Abdullah, F.F.J, Osman, A.Y., Adamu, L., Zakaria, Z. and Abdullah, R. (2013). Acute phase protein profiles in calves following infection with whole cell, lipopolysaccharide and outer membrane protein extracted from *Pasteurella multocida* type B: 2. *Asian Journal of Animal Veterinary Advances* 8: 655-662. DOI: 10.3923/ajava.2013.655.662.
- Abou-Zaid, A.A. (2001). *Corynebacterium pseudotuberculosis* in buffaloes, and es and sheep. *Veterinary Medicine Journal of Giza* 49: 435-450.
- Adams, D.O. (1976). The granulomatous inflammatory response. *American Journal of Pathology* 84: 164-187.
- Alharbi, K.B. (2014). Effect of Dietary Urea and Sulphur in the Immune Response of Sheep Vaccinated Against Caseous Lymphadenitis. *Life Science Journal* 11 (5).
- Al-Saadoon, E.A., Al-Naama, L.M. and Hassan, J. (2003). Serum lactate dehydrogenase (LDH) activity is children with malignant diseases. *Bahrain Medical Bulletin* 25(2): 1-7.
- Alsemgeest, S.P., Taverne, M.A., Boosman, R., Van Der Weyden, B.C. and Gruys, E. (1993). Peripartum acute-phase protein serum amyloid-A concentration in plasma of cows and fetuses. *American Journal of Veterinary Research* 54: 164-167.
- Ansell, G.B. and Hawthorne, J.N. 1964. Catabolism. In: *Phospholipids*, Elsevier Publishing, Amsterdam, pp: 152-174.
- Ashfaq, M.K. and Campbell, S.G. (1979). A survey of caseous lymphadenitis and its etiology in goats in the United States. *Veterinary Medicine, Small Animal Clinician* 74: 1161-1165.

- Asselineau, J. and LaneLelle, G. (1998). Mycobacterial lipids: a historical perspective. *Frontiers in Bioscience* 3: e164-e174, www.bioscience.org.
- Attia, H. and Aziza, Eassa. (1997). Some investigation on an outbreak of broncho-pneumonia among sheep. *Zagazig Veterinary Journal* 25 (3): 6-14.
- Baird, G.J. and Fontaine, M.C. (2007). *Corynebacterium pseudotuberculosis* and its role in ovine caseous lymphadenitis. *Journal of Comparative Pathology* 137: 179-210.
- Baird, G., Syngé, B., and Dercksen, D. (2004). Survey of caseous lymphadenitis seroprevalence in British terminal sire sheep breeds. *Veterinary Record* 154 (16): 505-506.
- Barth, A.D. and Oko, R.J. 1989. *Abnormal morphology of bovine spermatozoa*. Iowa State University Press. Ames, Iowa, USA, pp: 285-287.
- Bastos, B.L., Loureiro, D., Raynal, J.T., Guedes, M.T., Vale, V.L.C., Moura-Costa, L.F., Guimarães, J.E., Azevedo, V., Portela, R.W. and Meyer, R. (2013). Association between haptoglobin and IgM levels and the clinical progression of caseous lymphadenitis in sheep. *BMC Veterinary Research* 9: 254.
- Bastos, B.L., Meyer, R., Guimaraes, J.E., Ayres, M.C., Guedes, M.T. and Moura-Costa, L.F. (2011). Haptoglobin and fibrinogen concentrations and leukocyte counts in the clinical investigation of caseous lymphadenitis in sheep. *Veterinary Clinical Pathology, American Society for Veterinary Clinical Pathology* 40 (4):496–503.
- Batey, R.G. (1986). Pathogenesis of caseous lymphadenitis in sheep and goats. *Australian Veterinary Journal* 63: 269-272.
- Biberstein, E.L., Knight, H.D. and Jang, s. (1971). Two biotypes of *Corynebacterium pseudotuberculosis*. *Veterinary Records* 89: 691-692.
- Braga W, Schul S, Nunez A, Pezo D and Franco E (2007). A primary *Corynebacterium pseudotuberculosis* low dose infection in alpacas

(Lama pacos) protects against a lethal challenge exposure. *Small Ruminant Research* 72: 81-86.

Bretzlaff, K. 1994. *Problems of reproduction of goats*. Proceeding of Small Ruminant in Short Course, Society of Theriogenology, Hastings, NE, p: 72.

Brogden, K.A., Glenn, J.S., East, N. and Audibert, F. (1995). A *Corynebacterium pseudotuberculosis* bacterin with muramyl dipeptide induced antibody titres, increases the time of onset, and decreases naturally occurring external abscesses in sheep and goats. *Small Ruminant Research* 19: 161-168.

Brogden, K.A. and Engen, R.L. (1990). Alterations in the phospholipid composition and morphology of ovine erythrocytes after intravenous inoculation of *Corynebacterium pseudotuberculosis*. *American Journal of Veterinary Research* 51: 874-877.

Brogden, K.A., Richard, L. Engen, J. Glenn, S. and Joseph, G. (1990). Changes in ovine erythrocyte morphology due to sphingomyelin degradation by *Corynebacterium pseudotuberculosis* phospholipase D. *Microbial Pathogenesis* (8) 2: 157-162.

Brown, C.C. and Olander, H.J. (1987). Caseous lymphadenitis of goats and sheep: a review. *Veterinary Bulletin* 57: 1-12.

Brown, C.C., Olander, H.J., Zometa, C. and Alves, S.F. (1986). Serodiagnosis of inapparent caseous lymphadenitis in goats and sheep, using the synergistic hemolysis-inhibition test. *American Journal of Veterinary Research* 47: 1461-1463.

Burtis, C. A., Ashwood, R. A., and Bruns, E. 2008. *Tietz fundamentals of clinical chemistry*. Saunders. Saint Louis.

Burrell, D.H. (1980). A simplified double immunodiffusion technique for detection of *Corynebacterium ovis* antitoxin. *Research in Veterinary Science* 28: 234-237.

Carr, D.W. and Acott, T.S. (1989). Intracellular pH regulates bovine sperm motility and protein phosphorylation. *Biology of Reproduction* 41: 907-920.

- Carne, H.R. (1940). The toxin of *Corynebacterium ovis*. *Journal of Pathology and Bacteriology* 51: 199-212. In: Baird, G.J. and Fontaine, M.C. (2007). *Corynebacterium pseudotuberculosis* and its role in ovine caseous lymphadenitis. *Journal of Comparative Pathology* 137: 179-210.
- Carne, H.R. and Onon, E.O. (1978). Action of *Corynebacterium ovis* exotoxin on the endothelial cells of blood vessels. *Nature* 271: 246-248.
- Ceciliania, f., Ceronb, J.J., Eckersallc, PD., and Sauerweind H. (2012). Acute phase proteins in ruminants. *Journal of Proteomics* 75: 4207–4231.
- Çetinkaya, B., Karahan, M., Atil, E., Kalin, R., De Baere, T. and Vaneechoutte, M. (2002). Identification of *Corynebacterium pseudotuberculosis* isolates from sheep and goats by PCR. *Veterinary Microbiology* 88: 75-83.
- Chacón, J. (2001). Assessment of Sperm Morphology in Zebu bulls Under Field Conditions in the Tropics. *Reproduction of Domestic Animal* 36: 91-98.
- Chandler, J.E., Painter, C.L., Adkison, R.W., Memon, M.A. and Hoyt, P.G. (1988). Semen quality characteristics of dairy goats. *Journal of Dairy Science* 71: 1638-1646.
- Chemineau, P., Gauthier, D., Poirier, J.C. and Saumande, J. (1982). Plasma levels of LH, FSH, prolactin, oestradiol-17 β and progesterone during natural and induced oestrus in the dairy goat. *Theriogenology* 17: 313–323.
- Chirino-Zarraga, C., Scaramelli, A., and Rey-Valerion, C. (2006). Bacteriological characterization of *Corynebacterium pseudotuberculosis* in Venezuelan goat flocks. *Small Ruminant Research* 65:170–175.
- Chng, W.J., Sum, C. and Kuperan, P. (2005). Causes of isolated prolonged activated partial thromboplastin time in an acute care general hospital. *Singapore Medical Journal* 46 (9): 450.
- Christie, R., Atkins, N.E. and Munch-Petersen, E. (1944). A note on a lytic phenomenon shown by group B streptococci. *Australian Journal of Experimental Biology and Medical Science* 22: 197-200. IN: Baird, G.J. and Fontaine, M.C. (2007). *Corynebacterium pseudotuberculosis* and its role in ovine caseous lymphadenitis. *Journal of Comparative Pathology* 137: 179-210.

- Clarridge, J.E. and Spiegel, C.A. 1995. *Corynebacterium and related organisms*. In: *Manual of Clinical Microbiology*, 6th Edit., E. J. Barron, Ed., American Society for Microbiology, Washington, pp: 357-370.
- Connor, K.M., Quirie, M.M., Baird, G. and Donachie, W. (2000). Characterization of United Kingdom isolates of *corynebacterium pseudotuberculosis* using pulsed-field gel electrophoresis. *Journal of Clinical Microbiology* 38: 2633-2637.
- Contri, A., Gloria, A., Robbe, D., Valorz, C., Wegher, L. and Carluccio, A. (2013). Kinematic study on the effect of pH on bull sperm function. *Animal Reproduction Science* 136 (4): 252-259.
- Coyle, M.B., Hollis, D.G. and Groman, N.B. 1985. *Corynebacterium spp. and other coryneform organisms*. In: *Manual of Clinical Microbiology*, 4th Edit., E. H. Lennette, A. Balows, W. J. Hausler and H. J. Shadomy, Eds, American Society for Microbiology, Washington, pp: 198-199.
- Cray, C., Zaias, J. and Altman, N.H. (2009). Acute phase response in animals: A review. *Comparative Medicine* 59: 517-526. PMID: 20034426.
- Coulter, G.H. and Kastelic, J.P. (1999). Management programs for developing beef bulls. In: *Current therapy in large animal Theriogenology*. 2ed: Agents (Robert YS and TR Walter, 2007.): Saunders, United States of America, pp: 221-228.
- Coulter, G.H. (1986). Puberty and postpubertal development of beef bulls. In: *Current therapy in large animal Theriogenology*. 2ed: Agents (Robert YS and TR Walter, 2007.): Saunders, United States of America, pp: 221-228.
- Cupps, P.T. 1991. *Reproduction in Domestic Animals*. 4th Edition, Academic Press Inc., San Diego, New York, Boston.
- da CA Sá, M., Veschi, J.L., Santos, G.B., Amanso, E.S., Oliveira, S.A., Veneroni-Gouveia, R.A.M.G. and Costa, M.M. (2013). Activity of disinfectants and biofilm production of *Corynebacterium pseudotuberculosis*. *Pesquisa Veterinaria Brasileira* 33 (11): 1319-1324.
- Davis, E.W. (1990). *Corynebacterium pseudotuberculosis* infection in animals. In: Paton, M. (2010). *The epidemiology and control of caseous*

lymphadenitis in Australian sheep flocks, PhD Thesis, Murdoch University.

- Dennis, S.M. and Bamford, V.W. (1966). The role of Corynebacteria in perinatal lamb mortality. *Veterinary Record* 79: 105-108.
- De Pauw, I.M.C., Van Soom, A., Mintiens, K., Verberckmoes, S. and de Kruijff, A. (2003). In vitro survival of bovine spermatozoa stored at room temperature under epididymal conditions. *Theriogenology* 59 (5): 1093-1107.
- Dercksen, D.P., Brinkhof, J.M.A. and Dekker-Nooren, T. (2000). A comparison of four serological tests for the diagnosis of caseous lymphadenitis in sheep and goats. *Veterinary Microbiology* 75: 167-175.
- Dercksen, D.P., ter Laak, E.A. and Schreuder, B.E. (1996). Eradication programme for caseous lymphadenitis in goats in The Netherlands. *Veterinary Record* 138: 237.
- Devendra, C. and Burns, M. 1983. *Goat production in the tropics*. 2nd Edition, Commonwealth Agricultural Bureau.
- Dhandapani, S.S., Manju, D., Vivekanandhan, S., Sharma, B.S. and Mahapatra, A.K. (2009). Prognostic value of admission serum albumin levels in patients with head injury. *Pan Arab Journal of Neurosurgery* 13: 60-65.
- Dinarello, C.A. (1999). Cytokines as endogenous pyrogens. *Journal of Infectious Diseases* (2) 179: S294-304.
- Dorella, F.A., Pacheco, L.G.C., Oliveira, S.C., Miyoshi, A., and Azevedo, V. (2006). *Corynebacterium pseudotuberculosis*: microbiology, biochemical properties, pathogenesis and molecular studies of virulence. *Veterinary Research* 37: 201-218.
- Drent, M., Cobben, N.A., Henderson, R.F., Jacobs, J. A., Wouters, E.F. and van Dieijen-Visser, M.P. (1996). BAL fluid LDH activity and LDH isoenzyme pattern in lipoid pneumonia caused by an intravenous injection of lamp oil. *European Respiratory Journal* 9 (11): 2416-2418.
- DVS. (2013). Department of Veterinary Services. *Statistics document* at: <http://www.dvs.gov.my>, pp: 1-2. Accessed September 2014.

- Eckersall, P.D. and Bell, R. (2010). Acute phase proteins: Biomarkers of infection and inflammation in veterinary medicine. *The Veterinary Journal* 185 (1): 23-27.
- Eckersall, P.D., Young, F.J., McComb, C., Hogarth, C.J., Safi, S., Weber, A., McDonald, T., Nolan, A.M. and Fitzpatrick, J.L. (2001). Acute phase proteins in serum and milk from dairy cows with clinical mastitis. *Veterinary Record* 148: 35-41.
- Eckersall, P.D., Lawson, F.P. and Bence, L. (2007). Acute phase protein response in an experimental model of ovine caseous lymphadenitis. *BMC Veterinary Research* 335-341.
- Eckersall, P.D., Duthie, S., Toussaint, M.J.M., Gruys, E., Heegaard, P., Alava, M., Lipperheide, C. and Madec, F. (1999). Standardization of diagnostic assays for animal acute phase proteins. *Advances in Veterinary Medicine* 41: 643-655.
- Ellis, J.A., Hawk, D.A., Mills, K.W. and Pratt, D.L. (1991). Antigen specificity and activity of ovine antibodies induced by immunization with *Corynebacterium pseudotuberculosis* culture filtrate. *Veterinary Immunology and Immunopathology* (28) 3: 303-316.
- Euzéby, J.P. (2005). List of Bacterial Names with Standing in Nomenclature. *Society for Systematic and Veterinary Bacteriology* <http://www.bacterio.coct.fr>.
- Fontaine, M.C. and Baird, G.J. (2008). Caseous lymphadenitis. *Small Ruminant Research* 76: 42-48.
- Fontaine, M.C., Baird, G., Connor, K.M., Rudge, K., Sales, J. and Donachie, W. (2006). Vaccination confers significant protection of sheep against infection with a virulent United Kingdom strain of *Corynebacterium pseudotuberculosis*. *Vaccine* 24: 5986-5996.
- Fraser, G. (1961). Haemolytic activity of *Corynebacterium ovis*. *Nature* 189: 246.
- Gamel, A.A. and Tartour, G. (1974). Haematological and plasma protein changes in sheep experimentally infected with *Corynebacterium pseudotuberculosis*. *Journal Comparative Pathology* 84: 477-483.

- Ghannoum, M.A. (2000). Potential role of phospholipases in virulence and fungal pathogenesis. *Clinical Microbiology* 13: 122-143.
- Gimenez, D. (2007). Reproductive management of goats and sheep. Alabama Cooperative Extension System. *Bulletin ANR* 1316.
- Givens, M.D. and Marley, M.S.D. (2008). Pathogens that cause infertility of bulls or transmission via semen. *Theriogenology* 70 (3): 504-507.
- Goldberger, A.C. Lipsky, B.A. and Plorde, J.J. (1981). Suppurative granulomatous lymphadenitis caused by *Corynebacterium ovis* (pseudotuberculosis). *American Journal of Clinical Pathology* 76: 486-490.
- González, F.H.D., Ruiperez, F.H., Sanches, J.M., Sanza, J.C. and Marinez-subield, S. (2010). Haptoglobin and serum amyloid A in subacute ruminal acidosis in goats. *Rev Med Vet Zoot* 57.
- Gordon, E.D. (2012). Progressive wasting diseases of ewes. In: *dairy sheep association of North America symposium*, pp: 64.
- Greyling, J.P.C. (2000). Reproduction traits in the Boer goat doe. *Small Ruminant Research* 36: 171-177.
- Gronlund, U., Hulten, C., Eckersall, P.D., Hogarth, C. and Waller, K.P. (2003). Haptoglobin and serum amyloid A in milk and serum during acute and chronic experimentally induced *Staphylococcus aureus* mastitis. *Journal of Dairy Research* 70: 379–386.
- Grooms, D.L. (2004). Reproductive consequences of infection with bovine viral diarrhea virus. *Veterinary Clinics of North America: Food Animal Practice* 20 (1): 5-19.
- Gruys, E., Obwolo, M.J. and Toussaint, M.J.M. (1994). Diagnostic significance of the major acute phase proteins in veterinary clinical chemistry: a review. *Veterinary Bulletin* 64: 1009–1018.
- Guilloteau, L., Pepin, M., Pardon, P. and Le Pape, A. (1990). Recruitment of 99m-technetium- or 111-indium-labelled polymorphonuclear leucocytes in experimentally induced pyogranulomas in lambs. *Journal of Leukocyte Biology* 48: 343-352.

- Guimarães, A., Carmo, F.B., Paulett, R.B., Seyffert, N., Ribeiro, D., Lage, A.P., Heinemann, M.B., Miyoshi, M., Azevedo, V. and Gouveia, A.M.G. (2011a). Caseous lymphadenitis: epidemiology, diagnosis, and control. *IIOAB Journal* (2) 2: 33-43.
- Guimarães, A.S., Carmo, F.B., Heinemann, M.B., Portela, R.W.D., Meyer, R., Lage, A.P., Seyffert, N., Miyoshi, A., Azevedo, V. and Gouveia, A.M.G. (2011b). High sero-prevalence of caeous lymphadenitis identified in slaughterhouse samples as a consequence of deficiencies in sheep farm management in the state of Minas Gerais, Brazil. *BMC Veterinary Research* 7: 68.
- Hafez, E.S.E. 1993. *Reproduction in farm animals*. 6th ed, Lea and Febiger, Philadelphia, P.A., pp: 571.
- Hall, P. and Cash, J. (2012). What is the Real Function of the Liver 'Function' Tests?. *The Ulster Medical Journal* 81(1): 30.
- Hard, G.C. (1975). Comparative toxic effect of the surface lipid of *Corynebacterium ovis* on peritoneal macrophages. *Infection and Immunity* 12: 1439-1449.
- Hassan, N.A., Al-Humainy, A.A., Bahobail, A.S. and Mansour, A.M.A. (2011). Bacteriological and pathological studies on caseous lymphadenitis in sheep in Saudi Arabia. *International Journal of Microbiology Research* 2 (1): 28-37.
- Hedger, M. and Hales, D. 2006. *Immunophysiology of the male reproductive tract*. Elsevier Academic Press, pp: 1195-1286.
- Heegaard, P.M., Godson, D.L., Toussaint, M.J., Tjornehoj, K., Larsen, L.E., Viuff, B. and Ronsholt, L. (2000). The acute phase response of haptoglobin and serum amyloid A (SAA) in cattle undergoing experimental infection with bovine respiratory syncytial virus. *Veterinary journal of Immunology and Immunopathology* 77:151–159.
- Heinrich, P.C., Castell, J.V. and Andus, T. (1990). Interleukin-6 and the acute phase response. *Biochemical Journal* 265: 621–636.
- Hirvonen, J., Hietarkopi, S. and Saloniemi, H. (1997). Acute phase response in emergency slaughtered cows. *Meat Science* 3: 249–257.
- Hodgson, A.L., Bird, P. and Nisbet, I.T. (1990). Cloning, nucleotide sequence, and expression in *Escherichia coli* of the phospholipase D gene from

- Corynebacterium pseudotuberculosis*. *Journal of bacteriology* (3) 172: 1256-1261.
- Horadagoda, N.U., Knox, K.M.G., Gibbs, H.A., Reid, S.W.J., Horadagoda, A., Edwards, S.E.R. and Eckersall, P.D. (1999). Acute phase proteins in cattle: discrimination between acute and chronic inflammation. *Veterinary Record* 144: 437-441.
- Huijgen, H.J., Sanders, G.T., Koster, R.W., Vreeken, J. and Bossuyt, P.M. (1997). The clinical value of lactate dehydrogenase in serum: a quantitative review. *European Journal of Clinical Chemistry and Clinical Biochemistry* 35(8): 569-579.
- Husband, A.J. and Watson, D.L. (1977). Immunological events in the popliteal lymph node of sheep following injection of live or killed *Corynebacterium ovis* into an afferent popliteal lymphatic duct. *Research in Veterinary Science* 22: 105-112.
- Ibtisam, M. A. (2008). Some clinicopathological and pathological studies of *C. ovis* infection in sheep. *Egyptian Journal of Comparative Pathology and Clinical Pathology* (21) 1: 327- 343.
- Ismail, A.A. and Hamid, Y.M.A. (1972). Studies on the effect of some chemical disinfectants used in veterinary practice in *Corynebacterium ovis*. *Journal of the Egyptian Veterinary Medical Association* 32: 195-202.
- Jain, N.C. 2000. *Schalm's Veterinary Hematology*. 6th ed., Lea and Febiger, Philadelphia, USA.
- Jesse, F.F.A., Randolph, P.S.S., Saharee, A.A., Wahid, A.H., Zamri-Saad, M., Jasni, S., Omar, A.R., Adamu, L. and Abdinasir, Y.O. (2013). Clinicopathological response of mice following oral route infection of *C. pseudotuberculosis*. *Journal of Agriculture and Veterinary Science (IOSR-JAVS)* 2 (2): 38-42.
- Jesse, F.F.A., Sang, S.L., Saharee, A.A. and Shahirudin, S. (2011). Pathological Changes in the Organs of Mice Model Inoculated with *Corynebacterium pseudotuberculosis* Organism. *Pertanika Journal of Tropical and Agricultural Science* 34: 145 – 149.
- Jesse, F.F.A., Azlan, C.M., Saharee, A.A., Murugaiyah, M., Noordin, M.M., Jasni, S., Ragavan, K., Hassan, M.D., Haron, A.W., Siti, K.B., Hazilawati, H. and Mahmud, T. (2008). *Control of caseous*

lymphadenitis (CLA) in goat at UPM farm. Proceeding: 20th Veterinary Association Malaysia. (VAM).

- Join-Lambert, O.F., Ouache, M., Canioni, D., Beretti, J.L., Blanche, S., Berche, P. and Kayal, S. (2006). *Corynebacterium pseudotuberculosis* necrotizing lymphadenitis in a twelve-year-old patient. *Pediatric Infectious Disease Journal* 25: 848-851.
- Jones, J.M. and Bavister, B.D. (2000). Acidification of intracellular pH in bovine spermatozoa suppresses motility and extends viable life. *Journal of Andrology* 21 (5): 616-624.
- Johnson, E.H., Vidal, C.E.S., Santa rosa, J. and Kass, P.H. (1993). Observations on goats experimentally infected with *Corynebacterium pseudotuberculosis*. *Small Ruminant Research* 12: 357-369.
- Judson, R. and Songer, J.G. (1991). *Corynebacterium pseudotuberculosis*: in-vitro susceptibility to 39 antimicrobial agents. *Veterinary Microbiology* 27: 145-150.
- Junior, J.P., Oliveira, A.A.F., Alves, F.S.F., Silva, L.B.G., Rabelo, S.S.A., and Mota, R.A. (2006). *Corynebacterium pseudotuberculosis* experimental infection of goats mammary gland. *Arquivos do Instituto Biologico, Sao Paulo* 73(4): 395-400.
- Kaba, J., Nowicki, M., Frymus, T., Nowicka, D., Witkowski, L., Szalus-Jordanow, O., Czopowicz, M. and Thrushfield, M. (2011). Evaluation of the risk factors influencing the spread of caseous lymphadenitis in goat herds. *Polish Journal of Veterinary Science* 14 (2): 231-237.
- Kaba, J., Kutschke, L. and Gerlach, G.F. (2001). Development of an ELISA for the diagnosis of *Corynebacterium pseudotuberculosis* infection in goats. *Veterinary Microbiology* 78: 155-163.
- Katoh, N. and Nakagawa, H. (1999). Detection of haptoglobin in the high-density lipoprotein and the very high-density lipoprotein fractions from sera of calves with experimental pneumonia and cows with naturally occurring fatty liver. *Journal of Veterinary Medical Science* 61: 119-124.
- Kent, J. (1992). Acute phase proteins: Their use in veterinary diagnosis. *British Veterinary Journal* 148: 279-282.

- Kessel, R.G. 1998. *Basic Medical Histology: The biology of Cells, Tissues and Organs*. 1st ed, Oxford University Press Incorporation, New York. Chap 22 and 23.
- Khuder, Z., Osman, A.Y., Jesse, F.F., Wahid, A., Saharee, A.A., Jasni, S., Yusoff, R. and Rasedee, A. (2012). Sex hormone profiles and cellular changes of reproductive organs of mice experimentally infected with *C. pseudotuberculosis* and its exotoxin phospholipase D (PLD). *Journal of Agricultural Veterinary Science (IOSR-JAVS)* (1): 3, 24-29.
- Kim, Y.J., Jang, B.K., Kim, E.S., Park, K.S., Cho, K.B. and Chung, W.J. (2012). Rapid normalization of alanine aminotransferase predicts viral response during combined peginterferon and ribavirin treatment in chronic hepatitis C patients. *Korean Journal of Hepatology* 18 (1): 41-7.
- Klein, B.G. 2007. *Cunningham's Textbook of Veterinary Physiology*. Elsevier Health Sciences.
- Komala, T.S., Ramlan, M., Yeoh, N.N, Surayani, A.R. and Sharifa Hamidah, S.M. (2008). A survey of caseous lymphadenitis in small ruminant farms from two districts in Perak, Malaysia-Kinta and Hilir Perak. *Tropical Biomedicine* 25 (3): 196-201.
- Korte, W., Clarke, S. and Lefkowitz, J.B. (2000). Short activated partial thromboplastin times are related to increased thrombin generation and an increased risk for thromboembolism. *American Journal of Clinical Pathology* 113 (1): 123-127.
- Kuria, J.K.N., Mbutia, P.G., Kang'ethe, E.K., and Wahome, R.G. (2001). Caseous lymphadenitis in goats: The pathogenesis, incubation period and serological response after experimental infection. *Veterinary Research Communication* 25: 89-97.
- Lakota, K., Zigon, P., Mrak-Poljsak, K., Rozman, B., Shoenfeld, Y. and Sodin-Semrl, S. (2011). Antibodies against acute phase proteins and their functions in the pathogenesis of disease: a collective profile of 25 different antibodies. *Autoimmunity reviews* 10 (12): 779-789.
- Lee, s. and Lynch, K.R. (2005). Brown recluse spider (*Loxosceles reclusa*) venom phospholipase D (PLD) generates lysophosphatidic acid (LPA). *Biochemistry Journal* 391: 317-323.
- Liberato, I.R.D.O., Lopes, E.P.D.A., Cavalcante, M.A.G.D.M., Pinto, T.C., Moura, I.F., and Loureiro Júnior, L. (2012). Liver enzymes in patients

with chronic kidney disease undergoing peritoneal dialysis and hemodialysis. *Clinics* 67 (2): 131-134.

Licari, L.G. and Jan, P.K. (2009). Thrombin physiology and pathophysiology. *Journal of Veterinary Emergency and Critical Care* 19 (1): 11-22.

Lossos, I.S., Breuer, R., Intrator, O. and Sonenblick, M. (1997). Differential diagnosis of pleural effusion by lactate dehydrogenase isoenzyme analysis. *CHEST Journal* 111(3): 648-651.

Luna, L.G. 1968. *Manual of Histologic Staining Methods; of the Armed Forces Institute of Pathology*. 3rd ed, Blakiston Division, McGraw-Hill, University of Michigan.

Marino, G., Catone, G., Barna, A., Russo, M. and Zanghì, A. (2009). Testicular Inflammatory Diseases in the Buck. *Journal of Comparative Pathology* 141 (4): 297.

Menzies, P.I., Muckle, C.A., Hwang, Y.T. and Songer, J.G. (1994). Evaluation of an enzyme-linked immunosorbent assay using an Escherichia coli recombinant phospholipase D antigen for the diagnosis of *Corynebacterium pseudotuberculosis* infection. *Small Ruminant Research* 13, 193–198.

McNamara, P.J., Bradley, G.A. and Songer, J.G. (1994). Targeted mutagenesis of the phospholipase D gene results in decreased virulence of *Corynebacterium pseudotuberculosis*. *Molecular Microbiology* 12: 921-930.

Mineiro, A.L.B.B., Bezerra, E.E.A., Vasconcellos, S.A., Costa, F.A.L. and Macedo, N.A. (2007). Leptospiral infection in bovine and its association with reproductive failure and climatic conditions. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia* 59 (5): 1103-1109.

Monga, M. and Roberts, J.A. (1994). Spermagglutination by Bacteria: Receptor-Specific Interactions. *Journal of andrology*. 15 (2): 151-156.

Muckle, C.A. and Gyles, C.L. (1986). Exotoxic activities of *Corynebacterium pseudotuberculosis*. *Current Microbiology* 13: 57-60.

Muckle, C.A. and Gyles, C.L. (1983). Relation of lipid content and exotoxin production to virulence of *Corynebacterium pseudotuberculosis* in mice. *American Journal of Veterinary Research* 44: 1149-1153.

- Murata, H., Shimada, N. and Yoshioka, M. (2004). Current research on acute phase proteins in veterinary diagnosis: an overview. *The Veterinary Journal* 168: 28-40.
- Musa, M. (1998). Hemolytic interactions of *dermatophilus congoleis*. *Zbi. Veterinar medicine* 39B (2):139-142.
- Nairn, M.E. and Robertson, J.P. (1974). *Corynebacterium pseudotuberculosis* infection of sheep: role of skin lesions and dipping fluids. *Australian Veterinary Journal* 50: 537-542.
- Nathwani, R.A., Kumar, S.R., Reynolds, T.B. and Kaplowitz, N. (2005). Marked elevation in serum transaminases: an atypical presentation of choledocholithiasis. *American Journal of Gastroenterology* 100 (2): 295-8.
- Netea, M.G., Kullberg, B.J. and Van der Meer, J.W. (2000). Circulating cytokines as mediators of fever. *Clinical Infectious Diseases* (5) 31: S178-S184.
- North, R. 2004. *Anatomy and Physiology of the Goat*. 2nd ed, Agafact A7.0.3, <http://www.agric.nsw.gov.au> .
- OIE-Terrestrial manual. (2009). Ovine epididymitis (*Brucella ovis*). Chapter 2. 7. 9.
OIE-World Organization for Animal Health. (2009). http://www.oie.int/hs2/sit_mald_cont.asp?c_mald=156andc_cont=6andannee=2004. Accessed 26 Sept.
- Ono, K., Ono, T. and Matsumata, T. (1995). The pathogenesis of decreased aspartate aminotransferase and alanine aminotransferase activity in the plasma of hemodialysis patients: the role of vitamin B6 deficiency. *Clinical Nephrology* 43(6): 405-8.
- O'Reilly, K.M., Green, L.E., Malone, F.E. and Medley, G.F. (2008). Parameter estimation and simulations of a mathematical model of *Corynebacterium pseudotuberculosis* transmission in sheep. *Preventive Veterinary Medicine* 83: 242-259.
- Osman, A.Y., Abdullah, F.F.J., Adamu, L., Abdullah, T.T., Azuan, M.H., Omar, A.R., Haron, A.W. and Saharee, A.A. (2014). The study of caseous lymphadenitis: dose dependent infection of *C. pseudotuberculosis* in

mouse model via oral inoculation. *Research Opinion in Animal Veterinary Science* 4 (3): 163-169.

- Osman, A.Y., Abdullah, F.F.J., Saharee, A.A., Haron, A.W., Sabri, I., and Abdullah, R. (2012). Haematological and Biochemical Alterations in Mice Following Experimental Infection with Whole Cell and Exotoxin (PLD) Extracted from *C. Pseudotuberculosis*. *Journal of Animal and Veterinary Advances* 11(24): 4660-4667.
- Othman, A.M., Jesse, F.F.A., Adamu, L., Abba, Y., Adza Rina, M.N., Saharee, A.A., Wahid, A.H. and Zamri-Saad, M. (2014). Changes in Serum Progesterone and Estrogen Concentrations in Non-Pregnant Boer does Following Experimental Infection with *Corynebacterium Pseudotuberculosis*. *Journal Veterinary Advances* 4 (5): 524-528.
- Ozkanlar, Y., Aktas, M.S., Kaynar, O., Ozkanlar, S. and Kireccl, E. (2012). Bovine respiratory disease in naturally infected calves: Clinical signs, blood gases and cytokine response. *Revue de Médica Vétérinaire* 163: 123-130.
- Pacheco, L.G.G., Pena, R.R. and Castro, T.L.P. (2007). Multiplex PCR assay for identification of *Corynebacterium pseudotuberculosis* from pure cultures and for rapid detection of this pathogen in clinical samples. *Journal of Medical Microbiology* 56: 1-7.
- Palmieri, C., Schiavi, E. and Salda, L.D. (2011). Congenital and acquired pathology of ovary and tubular genital organs in ewes: A review. *Theriogenology* 75 (3): 393-410.
- Parmentier, J. H., Pavicevic, Z. and Malik, K. U. (2006). ANG II stimulates phospholipase D through PKC. *American Journal of Physiology and Heart-Circulatory Physiology* 290: H46-H54.
- Paton, M.W. (2010). *The epidemiology and control of caseous lymphadenitis in australian sheep flocks*. PhD Thesis, Murdoch University.
- Paton, M.W., Collett, M.G., Pepin, M. and Bath, G.F. (2005). *Corynebacterium pseudotuberculosis* infections. In: *Infectious Diseases of Livestock*. 3rd Ed, J.A.W. Coetzer and R.C.Tustin, Eds, Oxford University Press Southern Africa, Cape Town, pp: 1917-1930.
- Paton, M., Rose, I., Hart, R., Sutherland, S., Mercy, A. and Ellis, T. (1996). Post-shearing management affects the sero-incidence of *Corynebacterium pseudotuberculosis* infection in sheep flocks. *Preventive Veterinary Medicine* 26: 275-284.

- Paton, M. (1990). Caseous lymphadenitis. *University of Sydney Post Graduate Committee in Veterinary Science, Proceedings No 141*: 149.
- Paton, M.W., Mercy, A.R., Sutherland, S.S. and Ellis, T.M. (1988). The influence of shearing and age on the incidence of caseous lymphadenitis in Australian sheep flocks. *Acta Veterinaria Scandinavica* 84(Suppl.): 101-103.
- Paule, B.J.A., Azevedo, V., Regis, L.F., Carminati, R., Bahia, C.R., Vale, V.L.C., Moura-Costa, L.F., Freire, S.M., Nascimento, I., Schaer, R., Goes, A.M. and Meyer, R. (2003). Experimental *Corynebacterium pseudotuberculosis* primary infection in goats: kinetics of IgG and interferon- γ production, IgG avidity and antigen recognition by Western blotting. *Veterinary immunology and immunopathology* 96 (3): 129-139.
- Peel, M.M., Palmer, G.G., Stacpoole, A.M. and Kerr, T.G. (1997). Human lymphadenitis due to *Corynebacterium pseudotuberculosis*: report of ten cases from Australia and review. *Clinical Infectious Diseases* 24: 185-191.
- Pepin, M., Seow, H.F. and Corner, L. (1997). Cytokine gene expression in sheep following experimental infection with various strains of *Corynebacterium pseudotuberculosis* differing in virulence. *Veterinary Research* 28: 149-163.
- Pepin, M., Paton, M. and Hodgson, A.L. (1994). Pathogenesis and epidemiology of *Corynebacterium pseudotuberculosis* infection in sheep. *Current Topics in Veterinary Research* 1: 63-82.
- Pepin, M., Fontaine, J.J., Pardon, P., Marly, J. and Parodi, A.L. (1991). Histopathology of the early phase during experimental *Corynebacterium pseudotuberculosis* infection in lambs. *Veterinary Microbiology* 29: 123-134.
- Pepin, M., Ardon P., Arly, J. and Antier, F. (1988). *Corynebacterium pseudotuberculosis* infection in adult ewes by inoculation in the external ear. *American Journal of Veterinary Research* 49 (4): 459-463.
- Pereira, M.H.C., Cooke, R.F., Alfieri, A.A. and Vasconcelos, J.L.M. (2013). Effects of vaccination against reproductive diseases on reproductive performance of lactating dairy cows submitted to AI. *Animal Reproduction Science* 137 (3): 156-162.

- Permi, H.S., Jayaprakash Shetty, K., Padma, S.K., Teerthanath, S., Mathias, M., and Kumar, S. (2012). A histopathological study of granulomatous inflammation. *Nitte University Journal of Health Science* 2 (1): 2249-7110.
- Petersen, H.H., Nielsen, J.P. and Heegaard, P.M.H. (2004). Application of acute phase protein measurement in veterinary clinical chemistry. *Veterinary Research* 35: 163–187.
- Peterhans, E., Greenland, T., Badiola, J., Harkiss, G., Bertoni, G., Amorena, B., Eliaszewicz, M., Juste, R.A., Kraßnig, R., Lafont, J.P., Lenihan, P., Pétursson, G., Pritchard, G., Thorley, J., Vitu, C., Mornex, J.F. and Pépin, M. (2004). Routes of transmission and consequences of small ruminant lentiviruses (SRLVs) infection and eradication schemes. *Veterinary Research* 35 (3): 257-274.
- Piñeiro, M., Morales, J., Vizcaino, E., Murillo, J. A., Klauke, T., Petersen, B., and Piñeiro, C. (2013). The use of acute phase proteins for monitoring animal health and welfare in the pig production chain: The validation of an immunochromatographic method for the detection of elevated levels of pig-MAP. *Meat Science* 95(3): 712-718.
- Piontkowski, M.D. and Shivers, D.W. (1998). Evaluation of a commercially available vaccine against *Corynebacterium pseudotuberculosis* for use in sheep. *Journal of the American Veterinary Medical Association* 212: 1765-1768.
- Prescott, J.F. and Muckle, C.A. (1986). *Corynebacterium*. In: Paton, M.W. (2010). *The epidemiology and control of caseous lymphadenitis in Australian sheep flocks*. PhD Thesis, Murdoch University.
- Pugh, D.G. and Baird, A.N. 2012. *Sheep and goat medicine*. 2nd ed, Saunders, an imprint of Elsevier Incorporation. Chap 8: Theriogenology of Sheep and Goats.
- Pugh, D.G. 2002. *Sheep and goat medicine*. 1st ed, Philadelphia, PA, W.B. Saunders Company.
- Pugh, D.G. (1997). Caseous lymphadenitis in small ruminants. *Proceedings of North American Veterinary Conference*. 11: 983.

- Quartuccio, M., Marino, G., Cristarella, S. and Zanghi, A. (2009). Ram Epididymitis: Case Reports. *Journal of Comparative Pathology* 141: (4): 297.
- Quinn, P.J., Carter, M.E., Markey, B. and Carter, G.R. (1994). *Corynebacterium* species and *Rhodococcus equi*. In: *Clinical Veterinary Microbiology*. Wolfe Publishing Company, London, pp: 137-143.
- Radostits, O.M., Gay, C.C., Hinchcliff, K.W. and Constable, P.D. 2007. A *textbook of the diseases of cattle, horses, sheep, pigs and goats*. Veterinary Medicine. 10: 2045-2050.
- Radostits, O.M., Gay, C.C., Blood, D.C. and Hinchcliff, K.W. 2000. *Caseous lymphadenitis in sheep and goats*. In: *Veterinary Medicine*, 9th Edit., W.B. Saunders, London, pp: 727-730.
- Ribeiro, D., Dorella, F.A., Pacheco, L.G.C., Seyffert, N. and de Paula Castro, T.L. (2013). Subclinical Diagnosis of Caseous Lymphadenitis Based on ELISA in Sheep from Brazil. *Journal of Bacteriology and Parasitology* 4: 170. doi:10.4172/2155-9597.1000170.
- Rina, A., Zamri-Saad, M., Jesse, F.F.A., Saharee, A.A., Haron, A.W. and Shahirudin, S. (2013). Clinical and pathological changes in goats inoculated *Corynebacterium pseudotuberculosis* by intradermal, intranasal and oral routes. *Online Journal of Veterinary Research* 17 (2): 73-83.
- Robert, Y.S. and Walter, T.R. 2007. *Current therapy in large animal Theriogenology*. 2nd ed. Saunders. United States of America.
- Roser, J.F. (1999). Subfertility and infertility in the stallion. *World Equine Veterinary Review* 4: 32-37.
- Russell, K.E. and Grindem, C.B. (2000). Secondary thrombocytopenia. In: *Schalm's veterinary hematology*. Philadelphia: Lippincott Williams and Wilkins.487-495.
- Saab, S., Martin, P., Brezina, M., Gitnick, G. and Yee, H.F. (1995). Serum alanine aminotransferase in hepatitis c screening of patients on hemodialysis. *American Journal of Kidney Diseases* 37(2): 308-15, <http://dx.doi.org/10.1053/ajkd.2001.21294>.
- Samols, D., Agrawal, A. and Kushner, I. (2009). Acute phase proteins. In: Lakota, K., Zigon, P., Mrak-Poljsak, K., Rozman, B., Shoenfeld, Y. and

- Sodin-Semrl, S. (2011). Antibodies against acute phase proteins and their functions in the pathogenesis of disease: a collective profile of 25 different antibodies. *Autoimmunity reviews* 10 (12): 779-789.
- Sanocka, D., Frańczek, M., Jeńdrzejczak, P., Szumała-Kańkol, A. and Kurpisz, M. (2004). Male genital tract infection: an influence of leukocytes and bacteria on semen. *Journal of Reproductive Immunology* 62 (1): 111-124.
- Santiago, L.B., Pinheiro, R.R., Alves, F.S.F., Souza dos Santos, V.W., de Sousa Rodrigues, A., Lima, A.M.C., Luiz de Oliveira, E. and de Albuquerque, F.H. (2013). *In vivo* evaluation of antiseptics and disinfectants on control of Caseous Lymphadenitis: clinical, haematological, serological and microbiological monitoring. *Arq. Inst. Biol., São Paulo* 80 (3): 273-280.
- Schoenian, 2005. Reproduction in the Ram. In: *Getting Started In The Meat Goat Business*. McKenzie-Jakes, Angela.
- Seebach, J.D., Morant, R., Ruegg, R., Seifer, B. and Fehr, J. (1997). The diagnostic value of the neutrophil left shift in predicting inflammatory and infectious diseases. *American Journal of Clinical Pathology* 107: 82-91.
- Selim, S.A., Ghoneim, M.E. and Mohamed, K.H.F. (2010). Vaccinal efficacy of genetically inactivated phospholipase D against caseous lymphadenitis in small ruminants. *International Journal of Microbiological Research* 1 (3): 129-136.
- Sette, L.H. and Lopes, E.P. (2014). Liver enzymes serum levels in patients with chronic kidney disease on hemodialysis: a comprehensive review. *Clinics* 69 (4): 271-278.
- Shamay, A., Homans, R., Fuerman, Y., Levin, I., Barash, H. and Silanikove, N. (2005). Expression of albumin in nonhepatic tissues and its synthesis by the bovine mammary gland. *Journal of Dairy Science* (2) 88: 69-76.
- Shamsuddin, M., Rodriguez-Martinez, H. and Larsson, B. (1993). Fertilizing capacity of bovine spermatozoa selected after swim up in hyaluronic acid containing medium. *Reproduction and Fertility Development* 5: 7-15.
- Shigidi, M.T.A. (1978). An indirect haemagglutination test for the sero-diagnosis of *Corynebacterium ovis* infection in sheep. *Research in Veterinary Science* 24: 57-60.

- Shum, W.W., Ruan, Y.C., Silva, N. and Breton, S. (2011). Establishment of Cell-Cell Cross Talk in the Epididymis: Control of Luminal Acidification. *Journal of andrology* 32 (6): 576-586.
- Silva, W.M., Seyffert, N., Santos, A.V., Castro, T.L., Pacheco, L.G., Santos, A.R., Ciprandi, A., Dorella, F.A., Andrade, H.M., Barh, D., Pimenta, A.M.C., Silva, A., Miyoshi, A. and Azevedo, V. (2013). Identification of 11 new exoproteins in *Corynebacterium pseudotuberculosis* by comparative analysis of the exoproteome. *Microbial pathogenesis* 61: 37-42.
- Sivananda, N., Ajesh, S., Manju, C., Sugapriya, P., Mukund Sudharshan MG., Maheswari, G., Priscilla, K., Palaniappan, C and Sekar, B. (2010). Cytotoxic effect of diphtheria toxin in mammalian cell lines. *Indian Journal of Science and Technology* 3: 1177-1179.
- Skalet, L.H. (1986). *Effects of age and season on the spermogram of Nubian male goats*. MSc Thesis, Tuskegee University.
- Slayer, A. and Witt, D. (1994). Virulence factors that promote colonization. In: Baird, G.J. and Fontaine, M.C. (2007). *Corynebacterium pseudotuberculosis* and its role in ovine caseous lymphadenitis. *Journal of Comparative Pathology* 137: 179-210.
- Smith, B.I., Donovan, G.A., Risco, C.A., Young, C.R. and Stanker, L.H. (1998). Serum haptoglobin concentrations in Holstein dairy cattle with toxic puerperal metritis. *Veterinary Record* 142: 83-85.
- Songer, J.G. (1997). Bacterial phospholipases and their role in virulence. *Trends Microbiology* 5: 156-160.
- Songer, J.G., Beckenbach, K., Marshall, M.M., Olson, G.B. and Kelley, L. (1988). Biochemical and genetic characterization of *Corynebacterium pseudotuberculosis*. *American Journal of Veterinary Research* 49: 221-226.
- Soucek, A. and Souckova, A. (2001). Toxicity of bacterial sphingomyelinases D. *Journal of Hygiene, Epidemiology, Microbiology and Immunology* 18:327-335.
- Sreenivasan, R.S., Krishna Moorthy, P., Deecaraman, M., Prakash, N. and Renganathan, N.G. (2010). Variations in the Enzyme Activity of Carbohydrate Metabolic Disorder on Cardiac Function. *European Journal of Applied Sciences* 2 (2): 62-69.

- Stoops, S.G., Renshaw, H.W. and Thilsted, J.P. (1984). Ovine caseous lymphadenitis: disease prevalence, lesion distribution, and thoracic manifestations in a population of mature culled sheep from western United States. *American Journal of Veterinary Research* 45: 557-561.
- Sumathi, D., Selvaraj, P., Nambi, A.P., Prathaban, S. and Enbavan, P.A. (2012). Assessment of prothrombin and Activated partial thromboplastin time in dogs. *Tamilnadu Journal of Veterinary and Animal Sciences* 8 (4): 38-40.
- Sutherland, S.S., Hart, R.A. and Buller, N.B. (1996). Genetic differences between nitrate-negative and nitrate positive *Corynebacterium pseudotuberculosis* strains using restriction fragment length polymorphisms. *Veterinary Microbiology* 49: 1-9.
- Tashjian, J.J. and Campbell, S.G. (1983). Interaction between caprine macrophages and *Corynebacterium pseudotuberculosis*: an electron microscopic study. *American Journal of Veterinary Research* 44: 690-693.
- Tourlomousis, P., Eckersall, P.D., Waterston, M. and Buncic, S. (2004). A comparison of acute phase protein measurements and meat inspection findings in cattle. *Food borne Pathogens and Disease* 1: 281-290.
- Titball, R.W. (1993). Bacterial phospholipases C. *Microbiology and Molecular Biology Reviews* 57: 347-366.
- Unanian, M.M., Felicianosilva, A.E.D., and Pant, K.P. (1985). Abscesses and caseous lymphadenitis in goats in tropical Semi-Arid north-east Brazil. *Tropical Animal Health and Production* 17: 57-62.
- Ulutas, P.A., Voyvoda, H., Ulutas, B. and Aypak, S. (2008). Haptoglobin, serum amyloid-A and ceruloplasmin concentrations in goats with mixed helminth infection. *Turkiye Parazitoloji Dergisi* 32(3):229-33.
- Valli, V.E.O. (1993). The hematopoietic system. In: Paton, M.W. (2010). *The Epidemiology and Control of Caseous Lymphadenitis in Australian Sheep Flocks*. PhD Thesis. Murdoch University.

- Valli, V.E.O. and Parry, B.W. (1993). Caseous lymphadenitis. In: Baird, G.J. and Fontaine, M.C. (2007). *Corynebacterium pseudotuberculosis* and its role in ovine caseous lymphadenitis. *Journal of Comparative Pathology* 137: 179-210.
- Vandevyver, S., Dejager, L., Vandenbroucke, R. E., and Libert, C. (2014). An acute phase protein ready to go therapeutic for sepsis. *EMBO molecular medicine* 6 (1): 2-3.
- Van Tonder, E.M. (1975). Notes on some disease problems in Angora goats in South Africa. *Veterinary Medical Review* 1/2:109.
- Van Deursen, J., Heerschap, A., Oerlemans, F., Ruitenbeek, W., Jap, P., Laak, H.T. and Wieringa, B. (1993). Skeletal muscles of mice deficient in muscle creatine kinase lack burst activity. *Cell* 74: 612-631.
- Vreugdenhil, A.C., Dentener, M.A., Snoek, A.M., Greve, J.W. and Buurman, W.A. (1999). Lipopolysaccharide binding protein and serum amyloid A secretion by human intestinal epithelial cells during the acute phase response. *Journal of Immunology* 163: 2792–2798.
- Wallace, J.M., Milne, J.S., Redmer, D.A. and Aitken, R.P. (2006). Effect of diet composition on pregnancy outcome in over nourished rapidly growing adolescent sheep. *British Journal of Nutrition* 96: 1060-1068.
- Washburn, K.E, Bissett, W.T, Waldron, D.F, and Fajt, V.R. (2013). Serologic and bacteriologic culture prevalence of *Corynebacterium pseudotuberculosis* infection in goats and sheep and use of Bayesian analysis to determine value of assay results for prediction of future infection. *Journal of the American Veterinary Medical Association* (242) 7: 997-1002.
- Weibel, J.C. (2011). *Corynebacterium Pseudotuberculosis: Cell Invasion and Intracellular Survival*. PhD Thesis, Universität Zürich.
- Wells, B., Innocent, G., Eckersall, P.D., McCulloch, E., Nisbet, A.J. and Burgess, S.T.G. (2013). Two major ruminant acute phase proteins, haptoglobin and serum amyloid A, as serum biomarkers during active sheep scab infestation. *Veterinary Research* 44:103.

- Williamson, L.H. (2001). Caseous lymphadenitis in small ruminants. *Veterinary Clinics of North America Food Animal Practice* 17: 359-371.
- Williamson, P. and Nairn, M.E. (1980). Lesions caused by *Corynebacterium pseudotuberculosis* in the scrotum of rams. *Australian Veterinary Journal* 56: 496.
- Yasuda, K., Okuda, K., Endo, N., Ishiwatari, Y., Ikeda, R. and Hayashi, H. (1995) Hypoaminotransferasemia in patients undergoing long-term hemodialysis: clinical and biochemical appraisal. *Gastroenterology* 109 (4): 1295-300, [http://dx.doi.org/10.1016/0016-5085\(95\)90591-X](http://dx.doi.org/10.1016/0016-5085(95)90591-X).
- Yeruham, I., Elad, D., Van Ham, M., Shpigel, N.Y. and Perl, S. (1997). *Corynebacterium pseudotuberculosis* infection in Israeli cattle: clinical and epidemiological studies. *Veterinary Record* 140: 423-427.
- Yoshioka, M., Watanabe, A., Shimada, N., Murata, H., Yokomizo, Y. and Nakajima, Y. (2002). Regulation of haptoglobin secretion by recombinant bovine cytokines in primary cultured bovine hepatocytes. *Domestic Animal Endocrinology* 23: 425-433.
- Yozwiak, M.L. and Songer, J.G. (1993). Effect of *Corynebacterium pseudotuberculosis* phospholipase D on viability and chemotactic responses of ovine neutrophils. *American Journal of Veterinary Research* 54: 392-397.
- Zaki, M.M. (1976). Relation between the toxogenicity and pyogenicity of *Corynebacterium ovis* in experimentally infected mice. *Research in Veterinary Science* 20: 197-200.
- Zaki, M.M. (1968). The application of a new technique for diagnosing *Corynebacterium ovis* infection. *Research Veterinary Science* 9: 489.
- Zavoshti, F.R., Khoojine, A.B.S., Helan, J.A., Hassanzadeh, B. and Heydari, A.A. (2012). Frequency of caseous lymphadenitis (CLA) in sheep slaughtered in an abattoir in Tabriz: comparison of bacterial culture and pathological study. *Comparative clinical pathology* 21 (5): 667-671.