



**MOLECULAR CHARACTERIZATION AND PATHOGENICITY STUDY OF  
SEROTYPE 8B FOWL ADENOVIRUS ISOLATED IN MALAYSIA**

By

**NUR SYAZANA BINTI SABARUDIN**

Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of  
**Master of Science**

November 2019

**IB 2021 22**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**MOLECULAR CHARACTERIZATION AND PATHOGENICITY STUDY OF SEROTYPE 8B FOWL ADENOVIRUS ISOLATED IN MALAYSIA**

By

**NUR SYAZANA BINTI SABARUDIN**

**November 2019**

**Chair : Profesor Abdul Rahman Omar, DVM, PhD  
Institute : Bioscience**

Inclusion body hepatitis (IBH) is an economic importance viral disease affecting broilers in many countries in this region including Malaysia. Depending on geographical locations, different serotypes of fowl adenovirus (FAdV) have been identified as the causative agent of IBH. The present study aimed to detect FAdV from suspected cases of IBH from commercial chicken farms in Malaysia based on amplification and sequencing of the hexon gene, to determine the phylogenetic tree of the identified FAdV isolates based on hexon gene sequence analysis, to determine the phylogenetic tree of serotype 8b FAdV isolate UPM/FAdV/420/2017 based on hexon and fiber gene sequence analysis and to determine the pathogenicity of serotype 8b FAdV isolate UPM/FAdV/420/2017 in specific-pathogen-free (SPF) chickens following inoculation of the virus via the oral route and intramuscular route. From the study, 16 cases of IBH from commercial chicken farms in Malaysia were detected positive for Fowl adenoviruses (FAdVs) by polymerase chain reaction (PCR). Result of phylogenetic analysis based on the hexon gene sequence confirmed the typing of all the 16 isolates into a single cluster of FAdV serotype 8b species E. One of the isolates, UPM/FAdV/420/2017 was opted for fiber gene sequencing and proceeded with phylogenetic analysis based on fiber gene sequence. The result of the fiber gene analysis supported the grouping of the isolate into FAdV species E. Subsequently, the pathogenicity of the isolate UPM/FAdV/420/2017 was characterized in 11-day old specific-pathogen-free (SPF) chickens following inoculation of the virus at 108.53 TCID<sub>50</sub> via oral and intramuscular (IM) routes of inoculation. Infected chickens showed clinical signs of depression, ruffled feathers, huddling, inappetence, diarrhea and gross lesions characteristic of IBH such as enlarged, pale-yellowish liver with necrotic foci. Chickens infected via the IM route showed significantly higher clinical score values ( $p<0.05$ ) and higher percent mortality than orally inoculated birds. Histopathologic examination of the liver in infected chickens showed the presence of intranuclear inclusion bodies with a higher degree of severity in IM-

infected groups ( $p<0.05$ ). The amount and duration of virus shedding differ significantly between the two infected groups ( $p<0.05$ ) with IM-inoculated chickens secreted higher amount of virus and the shedding last until the last sampling point at 28 dpi. Meanwhile, the amount of virus shedding of the oral group was lower and was detected until the 14-day pi. At day 5 post-inoculation, virus copy number (VCN) ( $> \log_{10} 9$ ) was recorded highest in the liver and gizzard irrespective of inoculation. Although other organs namely cecal tonsils, bursa, kidney, spleen, and thymus were detected positive for FAdV, no significant gross and microscopic lesions were detected in these organs except the liver. The antibody response against FAdV showed a statistically significant difference ( $p<0.05$ ) between oral and IM-inoculated groups. Higher values were recorded in IM-inoculated chickens throughout the experimental trial. Results of the study suggested that FAdV serotype-8b is a dominant serotype circulating in Malaysia and associated with outbreaks of IBH. This study also confirmed that Malaysian FAdV-8b isolate UPM/FAdV/420/2017 is a pathogenic virus and caused symptomatic IBH in SPF birds. The study approved that the extent of pathogenicity can be influenced by the route of inoculation. Further study is underway on the ability of inactivated FAdV vaccine in conferring protection against this virus.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PENCIRIAN MOLEKULAR DAN KAJIAN KEPAТОGENAN PENCILAN  
ADENO-UNGГAS SEROTIP 8B DI MALAYSIA**

Oleh

**NUR SYAZANA BINTI SABARUDIN**

**November 2019**

**Pengerusi : Profesor Abdul Rahman Omar, DVM, PhD**  
**Institut : Biosains**

Badan inklusi hepatitis (BIH) merupakan penyakit virus berkepentingan ekonomi yang menjelaskan ayam daging di banyak negara di rantau ini termasuklah Malaysia. Bergantung pada lokasi geografi, serotip virus adeno-unggas yang berbeza telah dikenalpasti sebagai agen penyebab bagi penyakit BIH. Kajian ini bertujuan untuk mengesan virus adeno-unggas dari kes yang disyaki BIH daripada ladang ayam komersial di Malaysia berdasarkan amplifikasi dan penjujukan gen hexon, untuk menentukan pokok filogenetik virus adeno-unggas serotip 8b dikenalpasti berdasarkan analisis gen hexon, untuk menentukan pokok filogenetik serotip 8b virus adeno-unggas pencilan UPM/FAdV/420/2017 berdasarkan analisis gen hexon dan gen fiber dan untuk menentukan kepatogenan serotip 8b virus adeno-unggas pencilan UPM/FAdV/420/2017 dalam ayam-bebas-patogen-spesifik (SPF) selepas inokulasi virus melalui laluan oral dan intra-otot. Dari kajian ini, 16 kes BIH daripada ladang ayam komersial di Malaysia telah dikesan positif virus adeno-unggas oleh tindak balas rantai polimerase (PCR). Keputusan analisis filogenetik berdasarkan jujukan gen hexon mengesahkan penjenisan kesemua pencilan ke dalam kelompok virus adeno-unggas serotip 8b spesis E. Salah satu daripada pencilan, UPM/FAdV/420/2017 telah dipilih untuk analisis jujukan gen fiber dan diteruskan dengan analisis pokok filogenetik berdasarkan jujukan gen fiber. Keputusan dari analisis menyokong pengelompokan pencilan ke dalam virus adeno-unggas spesis E. Kemudiannya, kepatogenan pencilan UPM/FAdV/420/2017 telah dicirikan dalam ayam-bebas-patogen-spesifik (SPF) yang berusia 11 hari selepas inokulasi virus pada dos 108.53 TCID<sub>50</sub> melalui laluan inokulasi oral dan intra-otot. Ayam yang dijangkiti virus menunjukkan gejala klinikal seperti depresi, bulu kusut, saling berkerumun, hilang selera makan, cirit birit dan tanda perubahan kasar pada organ yang menunjukkan ciri-ciri jangkitan badan inklusi hepatitis seperti bengkak dan kuning kepucatan pada tisu hepar berserta fokus nekrosis. Ayam yang dijangkiti melalui laluan intra-otot mencatat perbezaan ketara skor klinikal lebih tinggi ( $p<0.05$ ) dan mencatat peratusan kematian lebih tinggi berbanding burung yang dijangkit

secara oral. Pemeriksaan histopatologi tisu hepar ayam terjangkit mendapati kehadiran badan inklusi intra-nuklear pada tahap keparahan yang lebih tinggi dalam ayam yang dijangkit intra-otot ( $p<0.05$ ). Jumlah dan durasi penumpahan virus berbeza dengan ketara antara kedua-dua kumpulan yang dijangkiti ( $p<0.05$ ) dengan ayam yang dijangkit intra-otot menyebarkan jumlah virus yang lebih tinggi dan penumpahan virus kekal sehingga titik pensampelan terakhir iaitu pada hari ke-28 selepas jangkitan. Manakala, jumlah penumpahan virus bagi kumpulan oral adalah lebih rendah dan dikesan sehingga pada hari ke-14 selepas jangkitan. Pada hari kelima selepas jangkitan, jumlah kepulihan virus ( $> \log_{10} 9$ ) direkodkan paling tinggi dalam tisu hepar dan hempedal tanpa mengira laluan inokulasi. Walaupun organ lain seperti tonsil sekum, timus, bursa, ginjal dan limpa dikesan positif virus adeno-unggas, tiada lesi kasar dan lesi mikroskopik yang ketara dapat dikesan pada organ-organ ini melainkan pada tisu hepar. Gerak balas antibodi terhadap virus adeno-unggas menunjukkan perbezaan statistik yang ketara ( $p<0.05$ ) di antara kumpulan inokulasi oral dan intra-otot. Nilai yang lebih tinggi direkodkan dalam ayam dijangkit intra-otot sepanjang percubaan eksperimen. Keputusan kajian ini mencadangkan bahawa virus adeno-unggas serotip 8b adalah serotip dominan yang beredar di Malaysia dan dikaitkan dengan wabak IBH. Kajian ini juga mengesahkan bahawa virus adeno-unggas Malaysia serotip 8b pencilan UPM/FAdV/420/2017 adalah virus patogenik dan menyebabkan gejala BIH pada burung SPF. Kajian ini menyetujui bahawa skala kepatogenan boleh dipengaruhi oleh laluan jangkitan. Kajian lanjut masih dijalankan bagi melihat keupayaan vaksin nyah-aktif virus adeno-unggas dalam memberikan perlindungan bagi menentang virus ini.

## **ACKNOWLEDGEMENTS**

In the name of Allah, the Most Compassionate and Most Merciful. All praise and thanks to Almighty Allah, with His blessings giving me the strength and patience to finish this study.

First of all, I would like to express my gratitude to my awesome supervisor Prof. Dr. Abdul Rahman Omar for his endless supervision, support, kind words and most importantly for trusting me and giving me the opportunity to undertake this research. May Allah repay his kindness with all the goodness in the world and hereafter.

I also feel deeply indebted to research officer Dr. Tan Sheau Wei for her constant source of guidance and helps especially during those days when experiments went wrong, and I am facing technical difficulties and in dire need of consultation or troubleshooting. I can never thank her enough. To my co-supervisor Prof. Dr. Mohd Hair Bejo who has given me an insight and suggestions throughout this project, thank you so much.

To be honest, this research journey has become so much exciting as I have been surrounded by amazing souls like Najwa, Azizah and Kak Suw. No words can describe how grateful I am for their presence, myriad of help (physically and emotionally) and their incessant words of encouragement which helps to keep me moving. I cannot even imagine this journey without them. I also want to thank my friends Harmi, Azlini and Shikin for their ceaseless concerns and cheers that never failed to make my day. I will miss them all 3000.

To all the staffs, Kak Fiza, Nadia, Kak Nancy, Kak Lina and students in Institute of Bioscience, thank you so much for your helpfulness, services, understanding and making it favorable for me to complete this research work.

Not to forget, I want to thank the Ministry of Education Malaysia, for the financial aid given to me through MyBrainSc scholarship. This support has helped me to stay focused on my study and relieved my financial concerns. Thank you to Boehringer Ingelheim Animal Health for funding this project.

Finally, and most importantly, my utmost gratitude goes to my parents and siblings, whose love, affection and never-ending prayers have driven me all in the right way to complete this journey.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Abdul Rahman bin Omar, PhD**

Professor

Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Chairman)

**Mohd Hair bin Bejo, PhD**

Professor

Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean

School of Graduate Studies  
Universiti Putra Malaysia

Date: 14 January 2021

## **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Nur Syazana binti Sabarudin

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:

Name of Chairman of  
Supervisory  
Committee:

\_\_\_\_\_

Prof. Dr. Abdul Rahman bin Omar

Signature:

Name of Member of  
Supervisory  
Committee:

\_\_\_\_\_

Prof. Dr. Mohd Hair bin Bejo

## TABLE OF CONTENTS

	Page	
<b>ABSTRACT</b>	i	
<b>ABSTRAK</b>	iii	
<b>ACKNOWLEDGEMENTS</b>	v	
<b>APPROVAL</b>	vi	
<b>DECLARATION</b>	viii	
<b>LIST OF TABLES</b>	xiii	
<b>LIST OF FIGURES</b>	xiv	
<b>LIST OF ABBREVIATIONS</b>	xvi	
CHAPTER		
<b>1</b>	<b>INTRODUCTION</b>	1
<b>2</b>	<b>LITERATURE REVIEW</b>	4
2.1	Adenovirus overview	4
2.2	Fowl adenovirus	4
2.2.1	Taxonomy	4
2.2.2	Structure and proteins	9
2.2.3	Genome organization	12
2.2.4	Virus entry and replication	15
2.2.5	Antigenic properties	17
2.3	Fowl adenovirus infections in poultry	18
2.3.1	Clinical diseases in poultry	18
2.3.2	Virus transmission	21
2.3.3	Pathogenesis	22
2.3.4	Host immune response	23
2.3.5	Diagnosis	29
2.4	FAdV roles in the IBH	31
2.4.1	FAdV as the primary agent	31
2.4.2	FAdV as the secondary agent	31
2.4.3	Pathology of IBH	32
2.5	Control and prevention against FAdV	34
2.5.1	Biosecurity	34
2.5.2	Vaccination	34
2.6	Summary	35
<b>3</b>	<b>MATERIALS AND METHODS</b>	36
3.1	PCR detection and sequence analysis of the hexon gene of Malaysian FAdVs	36
3.1.1	Origin of samples	36
3.1.2	Propagation of FAdV isolates in SPF embryonated chicken eggs	36
3.1.3	Extraction of viral DNA	37

3.1.4	Polymerase chain reaction to amplify hexon gene	37
3.1.5	Gel electrophoresis	38
3.1.6	Hexon gene sequencing and phylogenetic analysis	38
3.2	Characterization of fiber gene of isolate UPM/FAdV/420/2017	41
3.2.1	Isolate UPM/FAdV/420/2017	41
3.2.2	Extraction of total DNA	41
3.2.3	Polymerase chain reaction to amplify fiber gene	41
3.2.4	Gel electrophoresis	41
3.2.5	Fiber gene sequencing and phylogenetic analysis	42
3.3	Pathogenicity study of FAdV isolate UPM/FAdV/420/2017 in SPF chickens	45
3.3.1	Animals	45
3.3.2	Ethics statement	45
3.3.3	Origin of the FAdV isolate	45
3.3.4	Virus propagation	46
3.3.5	FAdV adaptation in CEL cell culture	46
3.3.6	Experimental design	47
3.3.7	Clinical signs, gross lesions and histopathology	50
3.3.8	Measurement of antibodies	50
3.3.9	Quantitative PCR detection of viral DNA	50
3.3.10	FAdV copy number in cloacal swabs	51
3.3.11	FAdV copy number in tissues	51
3.3.12	Statistical analysis	51
<b>4</b>	<b>RESULTS</b>	<b>52</b>
4.1	Isolation and hexon gene analysis of Malaysian FAdVs	52
4.1.1	Virus isolation	52
4.1.2	Amplification of FAdV portion hexon gene sequences	55
4.1.3	Phylogenetic analysis based on L1 region of the hexon gene	55
4.1.4	Similarity matrix	58
4.2	Fiber gene analysis of isolate UPM/FAdV/420/2017	60
4.2.1	Amplification of UPM/FAdV/420/2017 fiber sequence	60
4.2.2	Analysis of UPM/FAdV/420/2017 fiber sequence	61
4.2.3	Phylogenetic analysis based on FAdV fiber gene	65
4.3	Pathogenicity study of Malaysian FAdV isolate UPM/FAdV/420/2017 in SPF chickens	67
4.3.1	Clinical sign and gross pathology	67
4.3.2	Histopathology	72
4.3.3	Antibody response	78

4.3.4	FAdV copy number in cloacal swabs	80
4.3.5	FAdV copy number in tissues	81
<b>5</b>	<b>DISCUSSION</b>	<b>82</b>
5.1	Isolation and hexon gene analysis of Malaysian FAdVs	82
5.2	Fiber gene analysis of isolate UPM/FAdV/420/2017	85
5.3	Pathogenicity study of Malaysian FAdV isolate UPM/FAdV/420/2017 in 11-day-old SPF chickens	87
<b>6</b>	<b>CONCLUSION AND FUTURE RECOMMENDATIONS</b>	<b>91</b>
<b>REFERENCES</b>		<b>93</b>
<b>APPENDICES</b>		<b>116</b>
<b>BIODATA OF STUDENT</b>		<b>121</b>
<b>PUBLICATION</b>		<b>122</b>

## LIST OF TABLES

<b>Table</b>		<b>Page</b>
2.1	The taxonomy of genus <i>Aviadenovirus</i>	6
2.2	FAdV nomenclature according to the different naming system	8
3.1	Primer pair used to detect FAdV and amplify hexon gene sequences	36
3.2	FAdV reference strains used for molecular characterization of the hexon gene	39
3.3	Primer sequences for amplification of FAdV-8b fiber gene	41
3.4	Reference strains used for molecular analysis of FAdV fiber gene	43
3.5	Primer-probe sequences used for qPCR	51
4.1	Description of Malaysian FAdVs isolated in this study	52
4.2	The number of embryonic deaths after inoculation with FAdV isolates at different passage.	53
4.3	Pairwise similarity matrix of the hexon nucleotide sequence	59
4.4	Pairwise similarity matrix of the fiber nucleotide sequence	64
4.5	Antibody responses in the infected SPF chickens after oral and IM route of inoculation	78
4.6	FAdV copy numbers in cloacal swabs of chickens infected with UPM/FAdV/420/2017 via oral and IM route of inoculation	80
4.7	FAdV copy numbers in tissues of chickens infected with UPM/FAdV/420/2017 at 5 dpi via oral and IM route of inoculation	81

## LIST OF FIGURES

<b>Figure</b>		<b>Page</b>
2.1	Adenovirus structure and proteins	9
2.2	Structure of hexon protein	10
2.3	A schematic layout of the genome organization for a different genus of Adenovirus	13
2.4	Illustration summary of AdV infection and replication	15
3.1	Experimental flow chart of the pathogenicity study	49
4.1	Embryonated chicken eggs inoculated with FAdV during 2nd passage	54
4.2	Phylogenetic analysis based on 593 bp hexon gene fragment containing the L1 region from 16 Malaysian FAdV isolates collected in this study	56
4.3	Phylogenetic analysis based on 187 amino acid residues corresponding to hexon gene fragment containing the L1 region from 16 Malaysian FAdV isolates collected in this study	57
4.4	Agarose gel electrophoresis of fiber gene PCR product from isolate UPM/FAdV/420/2017	60
4.5	The amino acid pairwise alignment of the fiber tail region as computed by ClustalW	61
4.6	The shaft region of isolate UPM/FAdV/420/2017 aligned with reference FAdV-8b	62

4.7	Phylogenetic tree based on 1094 bp of fiber gene comprising the tail, shaft and knob regions	66
4.8	Chickens from IM infected group at 5 dpi showed severe depression, ruffled feathers and diarrhea with yellow green feces.	67
4.9	Chicken from IM infected group at 5 dpi showed mild depression, ruffled feathers and diarrhea with yellow green feces.	68
4.10	Clinical scores of chickens after inoculation with UPM/FADV/420/2017 via the oral and intramuscular route of inoculation	69
4.11	The percentage mortality of chickens after inoculation with UPM/FAdV/420/2017 via oral and IM route	70
4.12	The gross lesion in the liver of sacrificed chickens at 3 dpi (A and B), 5 dpi (C and D) and 7 dpi (E and F)	71
4.13	Histopathology in liver tissues of orally inoculated chickens (H&E)	73
4.14	Histopathology in liver tissue of intramuscularly inoculated chicken (H&E) at 1 dpi showing minimal multifocal hepatitis.	74
4.15	Histopathology in liver tissues of intramuscularly inoculated chickens (H&E)	75
4.16	Histopathology in liver tissue of intramuscularly inoculated chicken (H&E) at day 10 pi showing bile duct proliferation	76
4.17	Histopathological liver lesion score of chickens after inoculation with UPM/FAdV/420/2017	77
4.18	ELISA antibody titer in chickens inoculated with isolate UPM/FAdV/420/2017	79

## LIST OF ABBREVIATIONS

aa	Amino acid
AdV	Adenovirus
AdV	Adenovirus
BLAST	Basic Local Alignment Search Tool
bp	Base pair
BSA	Bovine Serum Albumin
CEE	Chicken embryonated egg
CMI	Cell-mediated immunity
CPE	Cytopathic effect
DMEM	Dulbecco's Modified Eagle's Medium
DNA	Deoxyribonucleic acid
dpi	Day post-inoculation
dsDNA	Double-stranded deoxyribonucleic acid
ELISA	Enzyme-linked immunosorbent assay
FAdV	Fowl adenovirus
HAdV	Human adenovirus
HHS	Hydropericardium syndrome
IACUC	Institutional Animal Care and Use Committee
IBH	Inclusion body hepatitis
ICTV	International Committee on Taxonomy of Viruses
Ig	Immunoglobulin
IM	Intramuscular
INIB	Intranuclear inclusion bodies
ITR	Inverted terminal repeats
L1	Loop 1
LIVES	Laboratory of Vaccine and Immunotherapeutic
MEGA	Molecular Evolutionary Genetics Analysis
MHC	Major histocompatibility complex
NCBI	National Centre for Biotechnology Information
nt	Nucleotide

OIE	World Organization of Animal Health
ORF	Open reading frame
PAMPs	Pathogen associated molecular patterns
PBS	Phosphate buffered saline
PCR	Polymerase chain reaction
pi	Post inoculation
qPCR	Quantitative polymerase chain reaction
rpm	Revolutions per minute
SPF	Specific-pathogen free
TCID <sub>50</sub>	50% tissue culture infectious dose
TP	Terminal proteins
UPM	Universiti Putra Malaysia
VCN	Virus copy number
VN	Virus neutralization
w/v	Weight/volume

## CHAPTER 1

### INTRODUCTION

The poultry industry is one of the key components in the Malaysian livestock sector. Currently, production of local poultry is self-sufficient to meet the domestic demand and exporting enough to become the major net exporter of chickens among ASEAN nations. Nevertheless, the growth of the industry is continuously threatened by potential endemic viral disease of opportunistic pathogens. These infectious agents may act alone or in concert with other encouraging factors to cause a disease strikes which directly impact the profit and loss of the industry. One of the common viruses found in the local poultry farm is fowl adenovirus (FAdV). In Malaysia, the virus is first detected in the field in 2005 (Hair-Bejo, 2005) and since then, a steady reported case of FAdV has been reported for past 5 years, affecting primarily broilers and breeders (De la Torre et al., 2018; Marek et al., 2016; Xia et al., 2017).

FAdV is a non-enveloped double-stranded DNA virus which belongs to family Adenoviridae (Zhao et al., 2015). The virus has characteristics icosahedral shape with approximately 70-100 nm capsid diameter and formed by three major structural protein namely the hexon, fiber, and penton. Presently, FAdV can be classified into five species (FAdV A to E) as determined by its molecular structure and 12 serotypes (FAdV-1 to -8a, -8b to -11) based on virus neutralization test (Hess, 2000). Circulation of FAdV is widespread, covering most geographical location and appeared to infect not only chickens but also various kind of avian species (Adair & Fitzgerald, 2008). A feature of the virus epidemiology is the unusually large number of serotypes that can be isolated on a farm. In many cases, it is also common to isolate different FAdV serotypes from the same diseased bird which suggested a lack of cross-protection among serotypes (Pereira et al., 2014).

The pathogenic role of FAdV under field conditions remains unclear since the virus is ubiquitous in nature and can be isolated in both sick and healthy birds. Heterogeneity of FAdV serotypes in term of genetic makeup has led to a discrepancy in regard to disease behavior and producibility of clinical signs between different serotype or different isolates of similar serotype (Gaba et al., 2010). Previously, FAdV is indicated to cause disease when they act in concert with an immunosuppressive agent such as chicken anemia virus or mycotoxins (Toro et al., 2002). However, several studies have revealed emerging virulent strains of FAdV that able to elicit severe infections alone without immunosuppression as a co-factor. In this case, results of the disease often come with a high degree of mortality depending

on the virulence of the strains, susceptibility of the birds or animal-related management of the farm. Then again, there is no strong evidence to support the probability of synergistic effect toward the severity of the disease due to infection of mixed serotypes (Changjing et al., 2016).

Epidemiological control of FAdV is tricky because they can set up latent infections and caused the infected birds to become a lifetime carrier of the virus. Upon reactivation of the latent virus, birds can vertically transmit the FAdV through the embryonated eggs and chicks may shed the virus in feces upon hatching. Apparently, chicks can reactivate the virus once their maternal antibody declines and during this time, usually between four to six weeks, virus shedding is at maximum and cease when the birds develop local immunity. Presence of virus in excretions inevitably lead to horizontal transmission of FAdV in the flocks. In addition, lateral spread also occurs through direct contact with infected birds and contaminated fomites or personnel. Chances of aerosol transmission are minimal though and work at a very short distance (McFerran & Smyth, 2000).

In the field, FAdV has been isolated in various clinical conditions. They are most notable as primary pathogens of inclusion body hepatitis (IBH), hepatitis-hydropericardium syndrome (HHS) and avian gizzard erosion (AGE) (Schachner et al., 2017). Serotypes of FAdV isolated from these cases are varied and more often than not, particular FAdV serotypes are predisposed to certain clinical conditions and geographical locations (McFerran & Smyth, 2000). For example, IBH outbreaks in New Zealand are associated with serotype 1, 8 and 12 although in Korea the culprits mostly from serotype 3, 9 and 11 (Lim et al., 2011). Meanwhile, HHS is mostly implicated with FAdV belong to serotype 4 and disease outbreak is more widespread in Asia (Harrach, 2014).

Classically, detection of FAdV is achieved by virus isolation in embryonated eggs or cell culture. Since this virus replicates inside the nucleus, the presence of the virus in cell culture is identified by the demonstration of intranuclear inclusion bodies or manifestation of cytopathic effect. Further confirmation is made by immunofluorescence and electron microscopy validated by the presence of virus particles. Presently, the traditional routine diagnostic test has been superseded by molecular tools of PCR detection and sequencing of the FAdV hexon gene. PCR is a powerful molecular tool that allows highly specific and sensitive detection of the FAdV genome, with cost-benefit advantages. This method also robust and allow differentiation of species and serotypes of FAdV to replace the traditional serological method of virus neutralization test for serotyping of isolates. FAdV hexon gene is targeted because it is the longest gene and contains the hypervariable loop 1 (L1) region that holds the most sequence variability between serotypes and thus can accurately classify FAdV serotypes apart (Meulemans et al., 2004).

In Malaysia, FAdV has been implicated in sporadic outbreaks of IBH. The outbreaks occurred mostly in meat-producing chicken characterized with sudden onset of mortality that peaks after three to four days and gradually cease after day 6 (Norina et al., 2016). Liver of affected chickens is swollen, pale and presence of inclusion bodies in the hepatocytes. FAdV infections often resulted in a low degree of morbidity, but mortality rates can range between 5% to 10% and in severe cases may up to 30%. High mortality accompanied by poor growth performance of survived chickens in the infected house has an indirect effect on the sustainability of chicken-meat supply and turnover of the poultry industry as a whole. In an effort to put the FAdV's outbreaks under control, it is vital to identify the serotypes of the FAdV involved along with its pathogenic behaviour especially when vaccination to be implemented as a major approach against outbreaks. In Malaysia, proper epidemiological surveys on its FAdV's serotype distribution and its pathogenic characterization are still lacking which demand further exploration. The hypothesis of this study is, IBH cases in Malaysia is associated with a single FAdV serotype that is pathogenic in chickens. Findings from this experiment will provide updated information on Malaysian FAdV for development of control strategies to counter the FAdV outbreaks including the development of inactivated FAdV vaccine based on the circulating FAdV serotypes.

Therefore, the objectives of the present study were:

1. To detect FAdV from suspected cases of IBH from commercial chicken farms in Malaysia based on amplification and sequencing of the hexon gene.
2. To determine the phylogenetic tree of the identified FAdV isolates based on hexon gene sequence analysis.
3. To determine the phylogenetic tree of serotype 8b FAdV isolate UPM/FAdV/420/2017 based on hexon and fiber gene sequence analysis.
4. To determine the pathogenicity of serotype 8b FAdV isolate UPM/FAdV/420/2017 in specific-pathogen-free (SPF) chickens following inoculation of the virus via the oral route and intramuscular route.

## REFERENCES

- Abbas, A. K., Lichtman, A. H., & Pillai, S. (2000). *Cellular and molecular immunology* (4th ed.). New York: Elsevier.
- Abdul-Aziz, T. A., & Al-Attar, M. A. (1991). New syndrome in Iraqi chicks. *The Veterinary Record*, 129(12), 272.
- Abdul-Aziz, T. A., & Hasan, S. Y. (1995). Hydropericardium syndrome in broiler chickens: its contagious nature and pathology. *Research in Veterinary Science*, 59(3), 219–221.
- Abrescia, N. G. A., Cockburn, J. J. B., Grimes, J. M., Sutton, G. C., Diprose, J. M., Butcher, S. J., ... Bamford, J. K. H. (2004). Insights into assembly from structural analysis of bacteriophage PRD1. *Nature*, 432, 68–74.
- Adair, B. M. (2000). Immunopathogenesis of chicken anemia virus infection. *Developmental and Comparative Immunology*, 24, 247–255.
- Adair, B. M., & Fitzgerald, S. . (2008). Group I adenovirus infections. In Y. M. Saif, A. M. Fadly, J. R. Glisson, L. R. McDougald, L. K. Nolan, & D. E. Swayne (Eds.), *Diseases of Poultry* (12th ed., pp. 251–266). Iowa: Blackwell Publishing Professional.
- Adair, B. M., Mcferran, J. B., & Calvert, V. M. (1980). Development of a microtitre fluorescent antibody test for serological detection of adenovirus infection in birds. *Avian Pathology*, 9, 291–300. <https://doi.org/10.1080/03079458008418414>
- Adrian Mockett, A. P., & Cook, J. K. A. (1983). The use of an enzyme-linked imunosorbent assay to detect IgG antibodies to serotype-specific and group-specific antigens of fowl adenovirus serotypes 2, 3 and 4. *Journal of Virological Methods*, 7(5–6), 327–335.
- Afzal, M., & Sakkir, M. (1994). Survey of antibodies against various infectious disease agents in racing camels in Abu Dhabi, United Arab Emirates. *Revue Scientifique et Technique*, 13(3), 787–792.
- Ahmad, M., Burgess, G. W., Ahmad, M., & Burgess, G. W. (2001). Production and characterization of monoclonal antibodies to fowl adenoviruses. *Avian Pathology*, 30(5), 457–463. <https://doi.org/10.1080/03079450120078635>
- Akira, S., Uematsu, S., & Takeuchi, O. (2006). Pathogen recognition and innate immunity. *Cell*, 124, 783–801. <https://doi.org/10.1016/j.cell.2006.02.015>

- Alemnesh, W., Hair-Bejo, M., Aini, I., & Omar, A. R. (2012). Pathogenicity of fowl adenovirus in specific pathogen free chicken embryos. *Journal of Comparative Pathology*, 146(2–3), 223–229. <https://doi.org/10.1016/j.jcpa.2011.05.001>
- Alexander, H. S., Huber, P., Cao, J., & Krell, P. J. (1998). Growth characteristics of fowl adenovirus type 8 in a chicken hepatoma cell line. *Journal of Virological Methods*, 74, 9–14.
- Alexopoulou, L., Holt, A. C., Medzhitov, R., & Flavell, R. A. (2001). Recognition of double-stranded RNA and activation of NF- $\kappa$ B by Toll-like receptor 3. *Nature*, 413(732–738).
- Ali, S., Mahmood, M. S., Hussain, I., & Khan, M. N. (2015). Preparation and evaluation of lyophilized live attenuated vaccine of inclusion body hepatitis hydro-pericardium syndrome (IBH-HPS) against challenge in broiler chickens. *International Journal of Agriculture & Biology*, 17(3), 658–662. <https://doi.org/10.17957/IJAB/17.3.14.560>
- Aoshi, T., Koyama, S., Kobiyama, K., Akira, S., & Ishii, K. J. (2011). Innate and adaptive immune responses to viral infection and vaccination. *Current Opinion in Virology*, 1, 226–232. <https://doi.org/10.1016/j.coviro.2011.07.002>
- Athappily, F. K., Ramachandran, M., Rux, J. J., Cai, Z., & Burnett, R. M. (1994). The refined crystal structure of hexon, the major coat protein of adenovirus type 2, at 2.9 Å resolution. *Journal of Molecular Biology*, 242(4), 430–455.
- Avgousti, D. C., Fera, N. Della, & Otter, C. J. (2017). Adenovirus core protein VII downregulates the DNA damage response on the host genome. *Journal of Virology*, 91(20), 1–14.
- Balamurugan, V., Katari, J. M., Tiwari, A. K., Toroghi, R., & Jadhao, S. J. (2001). Development of sandwich elisa for the detection of fowl adenovirus 4 associated with hydropericardium syndrome in experimentally infected chicken. *Acta Virologica*, 45(2), 95–100.
- Balamurugan, V., & Kataria, J. M. (2004). The hydropericardium syndrome in poultry--a current scenario. *Veterinary Research Communications*, 28(2), 127–148.
- Barua, S., & Rai, A. (2004). Restriction enzyme analysis of fowl adenovirus 4 DNA. *Journal of Immunology and Immunopathology*, 6, 59–63.

- Benko, M., Harrach, B., Both, G. W., Russell, W. C., Adair, B. M., Adam, E., ... Wadell, G. (2005). Family adenoviridae. In C. M. Fauquet, M. A. Mayo, J. Maniloff, U. Desselberger, & L. A. Ball (Eds.), *Virus Taxonomy. Eighth report of the International Committee on Taxonomy of Viruses* (pp. 213–228). New York: Elsevier.
- Bon, A. Le, & Tough, D. F. (2002). Links between innate and adaptive immunity via type I interferon. *Current Opinion in Immunology*, 14, 432–436.
- Bruder, J. T., & Kovesdi, I. (1997). Adenovirus infection stimulates the Raf / MAPK signaling pathway and induces interleukin-8 expression. *Journal of Virology*, 71(1), 398–404.
- Burnett, R. M., Griitter, M. G., & White, J. L. (1985). The structure of the adenovirus capsid I . An envelope model of hexon at 6 Å resolution. *Journal of Molecular Biology*, 185(1), 105–123.
- Changjing, L., Haiying, L., Dongdong, W., Jingjing, W., Youming, W., Shouchun, W., ... Yanbo, Y. (2016). Characterization of fowl adenoviruses isolated between 2007 and 2014 in China. *Veterinary Microbiology*, 197, 62–67. <https://doi.org/10.1016/j.vetmic.2016.11.005>
- Cheema, A. H., Ahmad, J., & Afzal, M. (1989). An adenovirus infection of poultry in Pakistan. *Rev. Sci. Tech. Off. Int. Epiz.*, 8(3), 789–795.
- Chen, L., Yin, L., Zhou, Q., Li, Q., Luo, Y., Xu, Z., ... Cao, Y. (2018). Immunogenicity and protective efficacy of recombinant fiber-2 protein in protecting SPF chickens against fowl adenovirus 4. *Vaccine*, 36(9), 1203–1208. <https://doi.org/10.1016/j.vaccine.2018.01.028>
- Chen, S., Cheng, A., & Wang, M. (2013). Innate sensing of viruses by pattern recognition receptors in birds. *Veterinary Research* 2013, 44(82), 1–12.
- Choi, K. S., Kye, S. J., Kim, J. Y., Jeon, W. J., Lee, E. K., Park, K. Y., & Sung, H. W. (2012). Epidemiological investigation of outbreaks of fowl adenovirus infection in commercial chickens in Korea. *Poultry Science*, 91(10), 2502–2506.
- Christensen, N. H., & Saifuddin, M. (1989). A primary epidemic of inclusion body hepatitis in broilers. *Avian Diseases*, 33(4), 622–630.
- Chroboczek, J., & Jacrot, B. (1987). The sequence of adenovirus fiber: similarities and differences between serotypes 2 and 5. *Virology*, 161(2), 549–554.
- Clemmer, D. I. (1972). Age-associated changes in fecal excretion patterns of Strain 93 chick embryo lethal orphan virus in chicks. *Infection and Immunity*, 5(I), 60–64.
- Clemmer, D. I., & Ichinose, H. (1986). The cellular site of virus replication in the

- intestine of chicks infected with an avian adenovirus. *Arch Ges Virusforsch*, 25, 277–287.
- Coico, R., & Sunshine, G. (2015). *Immunology: A short course* (7th ed.). UK: John Wiley & Sons.
- Cook, J. K. A. (1968). Isolation of a CELO virus from fertile chicken eggs. *The Veterinary Record*, 82, 294.
- Cook, J. K. A. (1983). Fowl adenoviruses: Studies on aspects of the pathogenicity of six strains for 1 - day - old chicks. *Avian Pathology*, 12, 35–43. <https://doi.org/10.1080/03079458308436147>
- Cowen, B., Calnek, B. W., & Hitchner, S. B. (1977). Broad antigenicity exhibited by some isolates of avian adenovirus. *American Journal of Veterinary Research*, 38(7).
- Cowen, B., Lu, H., Weinstock, D., & Castro, E. (1996). Pathogenicity studies of fowl adenovirus isolated in several region in the world. In *Proceeding International Symposium on Adenovirus and Reovirus Infections in Poultry* (pp. 79–84). Rauschholzhausen, Germany.
- Cowen, B. S. (1988). Chicken embryo propagation of type I avian adenoviruses. *Avian Diseases*, 32, 347–352.
- Crawford-miksza, L., & Schnurr, D. P. (1996). Analysis of 15 adenovirus hexon proteins reveals the location and structure of seven hypervariable regions containing serotype-specific residues. *Journal of Virology*, 70(3), 1836–1844.
- Davis, J., Goodwin, M. A., Rosenberger, J. K., Lamichhane, C. M., & Snyder, D. B. (1991). Adenovirus hepatitis in chickens on three farms. In *Proc. 40th West. Poult. Dis. Conf* (p. 70). Alapulco, Mexico.
- Davison, A., Benko, M., & Harrach, B. (2003). Genetic content and evolution of adenoviruses. *Journal of General Virology*, 84, 2895–2908. <https://doi.org/10.1099/vir.0.19497-0>
- Davison, F., Magor, K. E., & Kaspers, B. (2008). Structure and evolution of avian immunoglobulins. In F. Davison, B. Kaspers, & K. A. Schat (Eds.), *Avian Immunology* (pp. 162–174). Amsterdam: Elsevier Ltd.
- Dawson, G. J., Orsi, L. N., Yates, V. J., Chang, P. W., & Pronovost, A. D. (1980). An enzyme-linked immunosorbent assay for detection of antibodies to avian adenovirus and avian adenovirus-associated virus in chickens. *Avian Diseases*, 24(2), 393–402.
- De la Torre, D., Nunez, L. F. N., & Parra, S. H. S. (2018). Molecular characterization of fowl adenovirus group I in commercial broiler chickens in Brazil. *VirusDisease*, 29(1), 83–88. [https://doi.org/10.1007/s13337-018-](https://doi.org/10.1007/s13337-018-018-)

- Deng, L., & Sharif, S. (2013). Oral inoculation of chickens with a candidate fowl adenovirus 9. *Clinical and Vaccine Immunology*, 20(8), 1189–1196. <https://doi.org/10.1128/CVI.00187-13>
- Domanska-Blicharz, K., Tomczyk, G., Smietanka, K., Kozaczynski, W., & Minta, Z. (2011). Molecular characterization of fowl adenoviruses isolated from chickens with gizzard erosions. *Poultry Science*, 90(5), 983–989. <https://doi.org/10.3382/ps.2010-01214>
- Doszpoly, A., Wellehan, J. F. X., Childress, A. L., Tarján, Z. L., & Kovács, E. R. (2013). Infection, genetics and evolution partial characterization of a new adenovirus lineage discovered in testudinoid turtles. *Infection, Genetics and Evolution*, 17, 106–112. <https://doi.org/10.1016/j.meegid.2013.03.049>
- Dutta, B., Deka, P., Gogoi, S. M., Sarmah, M., Bora, M. K., & Pathak, D. C. (2017). Pathology of inclusion body hepatitis hydropericardium syndrome (IBH-HPS) in broiler chicken. *International Journal of Chemical Studies*, 5(3), 458–461.
- Erf, G. F. (2004). Cell-mediated immunity in poultry. *Poultry Science*, 83, 580–590.
- Erny, K. M., Barr, D. A., & Fahey, K. J. (1991). Molecular characterization of highly virulent fowl adenoviruses associated with outbreaks of inclusion body hepatitis. *Avian Pathology*, 20(4), 597–606.
- Fadly, A. M., Riegle, B. J., Nazerian, K., & Stephens, E. A. (1980). Some observations on an adenovirus isolated from specific pathogen free chickens. *Poultry Science*, 59, 21–27.
- Fadly, A. M., & Winterfield, R. W. (1973). Isolation and some characteristics of an agent associated with inclusion body hepatitis, hemorrhages, and aplastic anemia in chickens. *Avian Diseases*, 17(1), 182–193.
- Fadly, A. M., Winterfield, R. W., & Olander, H. J. (1976). Role of the bursa of Fabricius in the pathogenicity of inclusion body hepatitis and infectious bursal disease viruses. *Avian Diseases*, 20(3), 467–477.
- Gaba, A., Pal, J. K., Parmar, H., & Prajapati, K. S. (2010). Isolation, identification and molecular characterization of inclusion body hepatitis virus. *Veterinary World*, 9(3), 415–417. <https://doi.org/http://dx.doi.org/10.5455/vetworld.2010.415-417>
- Gause, W. C., Urban Jr, J. F., & Stadecker, M. J. (2003). The immune response to parasitic helminths: Insights from murine models. *Trends in Immunology*, 24(5), 269–277. [https://doi.org/10.1016/S1471-4906\(03\)00101-7](https://doi.org/10.1016/S1471-4906(03)00101-7)

- Gerlach, H. (1994). Viruses. In B. W. Ritchie, G. J. Harrison, & L. R. Harrison (Eds.), *Avian Medicine: Principles and Application* (pp. 862–948). Florida: Wingers Publishing, Inc.
- Ghosh, S. S., Gopinath, P., & Ramesh, A. (2006). Adenoviral vectors: A promising tool for gene therapy. *Applied Biochemistry and Biotechnology*, 133, 9–29.
- Gombault, A., Baron, L., & Couillin, I. (2013). ATP release and purinergic signaling in NLRP3 inflammasome activation. *Frontiers in Immunology*, 3, 1–6. <https://doi.org/10.3389/fimmu.2012.00414>
- Gomis, A. S., Goodhope, R., Ojkic, D., Willson, P., Gomis, S., Goodhope, A. R., ... C. P. W. (2006). Inclusion Body Hepatitis as a Primary Disease in Broilers in Saskatchewan , Canada. *Avian Diseases*, 50, 550–555.
- Goodwin, M. A., Latimer, K. S., Resureccion, R. S., Miller, P. G., & Campagnoli, R. P. (1996). DNA in situ hybridization for the rapid diagnosis of massive necrotizing avian adenovirus hepatitis and pancreatitis in chicks. *Avian Diseases*, 40(4), 828–831.
- Gowthaman, V., Singh, S. D., Dhama, K., Barathidasan, R., Asok Kumar, M., Desingu, P. A., ... Ramakrishnan, M. A. (2012). Fowl adenovirus (FAdV) in India: Evidence for emerging role as primary respiratory pathogens in chickens. *Pakistan Journal of Biological Sciences*, 15(18), 900–903.
- Grafl, B., Prokofieva, I., Wernsdorf, P., Steinborn, R., & Hess, M. (2014). Infection with an apathogenic fowl adenovirus serotype-1 strain (CELO) prevents adenoviral gizzard erosion in broilers. *Veterinary Microbiology*, 172(1–2), 177–185. <https://doi.org/10.1016/j.vetmic.2014.05.020>
- Grgić, H., Krell, P. J., & Nagy, É. (2014). Comparison of fiber gene sequences of inclusion body hepatitis (IBH) and non-IBH strains of serotype 8 and 11 fowl adenoviruses. *Virus Genes*, 48(1), 74–80. <https://doi.org/10.1007/s11262-013-0995-y>
- Grgić, H., Philippe, C., Ojkic, D., & Nagy, E. (2006). Study of vertical transmission of fowl adenoviruses. *The Canadian Journal of Veterinary Research*, 70, 230–233.
- Grgić, H., Poljak, Z., Sharif, S., & Nagy, É. (2013). Pathogenicity and cytokine gene expression pattern of a serotype 4 fowl adenovirus isolate. *PLoS ONE*, 8(10), 1–10. <https://doi.org/10.1371/journal.pone.0077601>
- Grgić, H., Yang, D. H., Nagy, É., Grgić, H., Grgić, H., Yang, D. H., & Nagy, É. (2011). Pathogenicity and complete genome sequence of a fowl adenovirus serotype 8 isolate. *Virus Research*, 156(1–2), 91–97. <https://doi.org/10.1016/j.virusres.2011.01.002>
- Grimes, T. M. (2007). Inclusion body hepatitis of chickens-occurrence and

- control. In *Proceedings of the 56th Western Poultry Disease Conference* (pp. 42–46). Las Vegas, USA.
- Günes, A., Marek, A., Grafl, B., Berger, E., & Hess, M. (2012). Real-time PCR assay for universal detection and quantitation of all five species of fowl adenoviruses (FAdV-A to FAdV-E). *Journal of Virological Methods*, 183(2), 147–153. <https://doi.org/10.1016/j.jviromet.2012.04.005>
- Gupta, A., Ahmed, K. A., Ayalew, L. E., Popowich, S., Kurukulasuriya, S., Goonewardene, K., ... Gomis, S. (2017). Immunogenicity and protective efficacy of virus-like particles and recombinant fiber proteins in broiler-breeder vaccination against fowl adenovirus (FAdV)-8b. *Vaccine*, 35(20), 2716–2722. <https://doi.org/10.1016/j.vaccine.2017.03.075>
- Gupta, A., Popowich, S., Ojkic, D., Kurukulasuriya, S., Chow-Lockerbie, B., Gunawardana, T., ... Gomis, S. (2017). Inactivated and live bivalent fowl adenovirus (FAdV8b + FAdV11) breeder vaccines provide broad-spectrum protection in chicks against inclusion body hepatitis (IBH). *Vaccine*, 36(5), 744–750. <https://doi.org/10.1016/j.vaccine.2017.12.047>
- Hafez, H. M. (2011). Avian adenoviruses infections with special attention to inclusion body hepatitis/ hydropericardium syndrome and egg drop syndrome. *Pakistan Veterinary Journal*, 31(2), 85–92.
- Hair-Bejo, M. (2005). Inclusion body hepatitis in a flock of commercial broiler chickens. *Journal of Veterinary Malaysia*, 17(1), 23–26.
- Hall, T. A. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium Series.
- Harrach, B. (2014). Adenoviruses : General features. In *Reference Module in Biomedical Research* (pp. 1–10). Amsterdam: Elsevier Inc. <https://doi.org/10.1016/B978-0-12-801238-3.02523-X>
- Harrach, B., Benko, M., Both, G. W., Brown, M., Davison, A. J., Echavarria, M., ... Wadell, G. (2011). Adenoviridae. In A. M. Q. King, M. J. Adams, E. B. Carstens, & E. J. Lefkowitz (Eds.), *Virus Taxonomy: Classification and Nomenclature of Viruses. Ninth Report of the International Committee on Taxonomy of Viruses* (pp. 125–141). San Diego: Elsevier.

- Heidari, M., Sarson, A. J., Huebner, M., Sharif, S., Kireev, D., & Zhou, H. (2010). Marek's disease virus-induced immunosuppression: Array analysis of chicken immune response gene expression profiling. *Viral Immunology*, 23(3), 309–319.
- Helmboldt, C. F., & Frazier, M. N. (1963). Avian hepatic inclusion bodies of unknown significance. *Avian Diseases*, 7(4), 446–450.
- Henry, L. J., Xia, D. I., Wilke, M. E., Deisenhofer, J., & Gerard, R. D. (1994). Characterization of the knob domain of the adenovirus type fiber protein expressed in Escherichia coli. *Journal of Virology*, 68(8), 5239–5246.
- Hess, M. (2000). Detection and differentiation of avian adenoviruses: A review. *Avian Pathology*, 29(3), 195–206. <https://doi.org/10.1080/03079450050045440>
- Hess, M. (2013). Aviadenovirus infections. In J. R. Glisson, L. R. McDougald, L. Nolan, D. L. Suarez, & V. Nair (Eds.), *Diseases of Poultry* (13th ed., pp. 290–300). Ames: Wiley-Blackwell.
- Hess, M., Cuzange, A., Ruigrok, R. W. H., Chroboczek, J., & Jacrot, B. (1995). The avian adenovirus penton: Two fibers and one base. *Journal of Molecular Biology*, 252(4), 379–385. <https://doi.org/10.1006/jmbi.1995.0504>
- Hess, Michael, Blöcker, H., & Brandt, P. (1997). The complete nucleotide sequence of the egg drop syndrome virus: An intermediate between mastadenoviruses and aviadenoviruses. *Virology*, 238(1), 145–156. <https://doi.org/10.1006/viro.1997.8815>
- Hidalgo, P., Anzures, L., Hernández-mendoza, A., Guerrero, A., Wood, C. D., Valdés, M., ... Gonzalez, R. A. (2016). Morphological, biochemical, and functional study of viral replication compartments isolated from adenovirus-infected cells. *Journal of Virology*, 90(7), 3411–3427. <https://doi.org/10.1128/JVI.00033-16.Editor>
- Hoeben, R. C., & Uil, T. G. (2013). Adenovirus DNA replication. *Cold Spring Harbor Perspectives in Biology*, 1–12.
- Hong, S. S., Habib, N. A., Franqueville, L., Jensen, S., & Boulanger, P. A. (2003). Identification of adenovirus (Ad) penton base neutralizing epitopes by use of sera from patients who had received conditionally replicative Ad (Addl 1520) for treatment of liver tumors. *Journal of Virology*, 77(19), 10366–10375. <https://doi.org/10.1128/JVI.77.19.10366>

- Hoshino, K., Sugiyama, T., Matsumoto, M., Tanaka, T., Saito, M., Hemmi, H., ... Kaisho, T. (2006). IκB kinase-α is critical for interferon-α production induced by Toll-like receptors 7 and 9. *Nature*, 440, 949–953. <https://doi.org/10.1038/nature04641>
- Howell, J., MacDonald, D. W., & Christian, R. G. (1970). Inclusion body hepatitis in chickens. *The Canadian Veterinary Journal*, 11(5), 99–101.
- Itakura, C., Matsushita, S., & Goto, M. (1977). Fine structure of inclusion bodies in hepatic cells of chickens naturally affected with inclusion body hepatitis. *Avian Pathology*, 6(1), 19–32. <https://doi.org/10.1080/03079457708418209>
- Itakura, C., Yasuba, M., & Goto, M. (1974). Histopathological studies on inclusion body hepatitis in broiler chickens. *The Japanese Journal of Veterinary Science*, 36(4), 329–340.
- Janeway, C. J., Travers, P., & Walport, M. (2001). *Immunobiology: The immune system in health and disease* (5th ed.). New York: Garland Science.
- Jankovic, D., Liu, Z., & Gause, W. C. (2001). Th1- and Th2-cell commitment during infectious disease: asymmetry in divergent pathways. *Trends in Immunology*, 22(8), 450–457.
- Jie, H., Lian, L., Qu, L. J., Zheng, J. X., Hou, Z. C., Xu, G. Y., ... Yang, N. (2013). Differential expression of Toll-like receptor genes in lymphoid tissues between Marek's disease virus-infected and noninfected chickens. *Poultry Science*, 92, 645–654.
- Jordan, A. B., Gongora, V., Hartley, D., & Oura, C. (2018). A review of eight high-priority, economically important viral pathogens of poultry within the Caribbean region. *Veterinary Sciences*, 5(14), 1–12. <https://doi.org/10.3390/vetsci5010014>
- Juliana, M. A., Nurulfiza, I., Hair-Bejo, M., Omar, A. R., & Aini, I. (2014). Molecular characterization of fowl adenoviruses isolated from inclusion body hepatitis outbreaks in commercial broiler chickens in Malaysia. *Pertanika Journal of Tropical Agricultural Science*, 37(4), 483–497.
- Junnu, S., Lertwatcharasarakul, P., Jala, S., Phattanakulanan, S., Monkong, A., Kulprasertsri, S., ... Songserm, T. (2015). An inactivated vaccine for prevention and control of inclusion body hepatitis in broiler breeders. *Thai Journal of Veterinary Medicine*, 45(1), 55–62.

- Junnu, S., Lertwatcharasarakul, P., Jala, S., Phattanakunanan, S., Moonjit, P., & Songserm, T. (2014). Developing an indirect ELISA based on recombinant hexon protein for serological detection of inclusion body hepatitis in chickens. *The Journal of Veterinary Medical Science*, 76(2), 289–293. <https://doi.org/10.1292/jvms.13-0196>
- Kaisho, T., & Akira, S. (2006). Toll-like receptor function and signaling. *Journal of Allergy and Clinical Immunology*, 117(5), 979–987. <https://doi.org/10.1016/j.jaci.2006.02.023>
- Kawai, T., & Akira, S. (2010). The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors. *Nature Immunology*, 11(5), 373–384. <https://doi.org/10.1038/ni.1863>
- Kim, M. S., Lim, T. H., Lee, D. H., Youn, H. N., Yuk, S. S., Kim, B. Y., ... Song, C. S. (2014). An inactivated oil-emulsion fowl adenovirus serotype 4 vaccine provides broad cross-protection against various serotypes of fowl adenovirus. *Vaccine*, 32(28), 3564–3568. [https://doi.org/10.1007/978-3-319-41691-5\\_26](https://doi.org/10.1007/978-3-319-41691-5_26)
- Kohn, A. (1962). Gallus adeno-like virus in chickens. Studies on infection, excretion and immunity. *American Journal of Veterinary Research*, 23(562–568).
- Kotenko, S. V., Gallagher, G., Baurin, V. V., Lewis-antes, A., Shen, M., Shah, N. K., ... Donnelly, R. P. (2003). IFN- $\lambda$ s mediate antiviral protection through a distinct class II cytokine receptor complex. *Nature Immunology*, 4(1), 69–77. <https://doi.org/10.1038/ni875>
- Kovacs, E. R., & Benko, M. (2011). Complete sequence of raptor adenovirus 1 confirms the characteristic genome organization of siadenoviruses. *Infection, Genetics and Evolution*, 11, 1058–1065. <https://doi.org/10.1016/j.meegid.2011.03.021>
- Koyama, S., Ishii, K. J., Coban, C., & Akira, S. (2008). Cytokine innate immune response to viral infection. *Cytokine*, 43, 336–341. <https://doi.org/10.1016/j.cyto.2008.07.009>
- La Rosa, G., Iaconelli, M., Pourshaban, M., Luca, E., Valentini, P., Sica, S., ... Muscillo, M. (2011). Molecular characterization of adenovirus from clinical samples through analysis of the hexon and fiber genes. *Journal of General Virology*, 92(2), 412–420. <https://doi.org/10.1099/vir.0.023176-0>
- Leen, A. M., Sili, U., Vanin, E. F., Jewell, A. M., Xie, W., Vignali, D., ... Rooney, C. M. (2004). Conserved CTL epitopes on the adenovirus hexon protein expand subgroup cross-reactive and subgroup-specific CD8 T cells. *Blood*, 104(8), 2432–2441. <https://doi.org/10.1182/blood-2004-02-0646>. Supported
- Li, E., Stupack, D., Bokoch, G. M., & Nemerow, G. R. (1998). Adenovirus

- endocytosis requires actin cytoskeleton reorganization mediated by Rho family GTPases. *Journal of Virology*, 72(11), 8806–8812.
- Li, E., Stupack, D. G., Brown, S. L., Klemke, R., Schlaepfer, D. D., & Nemerow, G. R. (2000). Association of p130 CAS with phosphatidylinositol-3-OH kinase mediates adenovirus cell entry. *The Journal of Biological Chemistry*, 275(19), 14729–14735.
- Li, E., Stupack, D., Klemke, R., Cheresh, D. A., & Nemerow, G. R. (1998). Adenovirus endocytosis via αv integrins requires phosphoinositide-3-OH kinase. *Journal of Virology*, 72(3), 2055–2061.
- Li, X., & Tikoo, S. K. (2002). Genetic organization and sequence analysis of pVIII, fiber and early region 4 of bovine adenovirus type 7. *Virus Genes*, 25(1), 59–65.
- Lim, A. T., Lee, H., Lee, D., Lee, Y., Park, J., Lim, T., ... A, C. S. (2011). Identification and virulence characterization of fowl adenoviruses in Korea. *Avian Diseases*, 55(4), 554–560.
- Lim, T., Kim, B., Kim, M., Jang, J., Lee, D., Kwon, Y., ... Song, C. (2012). Outbreak of gizzard erosion associated with fowl adenovirus infection in Korea. *Poultry Science*, 91, 1113–1117.
- Louis, N., Fender, P., Barge, A., Kitts, P., & Chroboczek, J. (1994). Domain of adenovirus serotype 2 fiber. *Journal of Virology*, 68(6), 4104–4106.
- MacLachlan, N. J., & Dubovi, E. J. (2016). *Fenner's veterinary virology* (5th ed.). New York: Academic Press.
- Macpherson, I., McDougall, J. S., & Laursen-Jones, A. P. (1974). Inclusion body hepatitis in a broiler integration. *The Veterinary Record*, 95(13), 286–289.
- Male, D., Brostoff, J., Roth, D., & Roitt, I. (2012). *Immnology* (8th ed.). Philadelphia: Elsevier.
- Marek, A., Günes, A., Schulz, E., & Hess, M. (2010). Classification of fowl adenoviruses by use of phylogenetic analysis and high-resolution melting-curve analysis of the hexon L1 gene region. *Journal of Virological Methods*, 170(1–2), 147–154. <https://doi.org/10.1016/j.jviromet.2010.09.019>
- Marek, A., Kaján, G. L., Kosiol, C., Benkő, M., Schachner, A., & Hess, M. (2016). Genetic diversity of species fowl aviadenovirus d and fowl aviadenovirus E. *Journal of General Virology*, 97(9), 2323–2332. <https://doi.org/10.1099/jgv.0.000519>
- Marek, A., Kosiol, C., Harrach, B., Kaján, G. L., Schlötterer, C., & Hess, M. (2013). The first whole genome sequence of a fowl adenovirus B strain

enables interspecies comparisons within the genus Aviadenovirus.  
*Veterinary Microbiology*, 166(1–2), 250–256.  
<https://doi.org/10.1016/j.vetmic.2013.05.017>

Marek, A., Nolte, V., Schachner, A., Berger, E., Schlötterer, C., & Hess, M. (2012). Two fiber genes of nearly equal lengths are a common and distinctive feature of fowl adenovirus C members. *Veterinary Microbiology*, 156(3–4), 411–417.  
<https://doi.org/10.1016/j.vetmic.2011.11.003>

Martín, C. S. (2012). Latest insights on adenovirus structure and assembly. *Viruses*, 4, 847–877. <https://doi.org/10.3390/v4050847>

Mase, M., Nakamura, K., & Imada, T. (2010). Characterization of fowl adenovirus serotype 4 isolated from chickens with hydropericardium syndrome based on analysis of the short fiber protein gene. *Journal of Veterinary Diagnostic Investigation*, 22, 218–223.

Matczuk, A. K., Niczyporuk, J. S., Kuczkowski, M., Woźniakowski, G., Nowak, M., & Wieliczko, A. (2017). Whole genome sequencing of fowl aviadenovirus A - a causative agent of gizzard erosion and ulceration, in adult laying hens. *Infection, Genetics and Evolution*, 48, 47–53.  
<https://doi.org/10.1016/j.meegid.2016.12.008>

Mathis, J. M., Stoff-khalili, M. A., & Curiel, D. T. (2005). Oncolytic adenoviruses – selective retargeting to tumor cells. *Oncogene*, 24, 7775–7791.  
<https://doi.org/10.1038/sj.onc.1209044>

Matos, M., Dublecz, K., Grafl, B., Liebhart, D., & Hess, M. (2018). Pancreatitis is an important feature of broilers suffering from inclusion body hepatitis leading to dysmetabolic conditions with consequences for zootechnical performance. *Avian Diseases*, 62(1), 57–64.

Matos, M., Grafl, B., Liebhart, D., & Hess, M. (2016). The outcome of experimentally induced inclusion body hepatitis (IBH) by fowl aviadenoviruses (FAdVs) is crucially influenced by the genetic background of the host. *Veterinary Research*, 47, 1–10.  
<https://doi.org/10.1186/s13567-016-0350-0>

Mazaheri, A., Prusas, C., VoB, M., & Hess, M. (2003). Vertical transmission of fowl Adenovirus serotype 4 investigated in specified pathogen-free birds after experimental infection. *Arch. Geflügelk.*, 67(1), 6–10.

- Mazaheri, A., Prusas, C., Voß, M., & Hess, M. (1998). Some strains of serotype 4 fowl adenoviruses cause inclusion body hepatitis and hydropericardium syndrome in chickens. *Avian Pathology*, 27, 269–276. <https://doi.org/10.1080/03079459808419335>
- McCracken, R. M., McFerran, J. B., Evans, R. T., & Connor, T. J. (1976). Experimental studies on the aetiology of inclusion body hepatitis. *Avian Pathology*, 5(4), 325–339.
- McFerran, J. B. (1981a). Adenoviruses of vertebrate animals. In E. Kurstak & C. Kurstak (Eds.), *Comparative Diagnosis of Viral Diseases III* (pp. 102–154). New York: Academic Press.
- McFerran, J. B. (1981b). Immunity to adenoviruses. In M. E. Rose, L. N. Payne, & B. M. Freeman (Eds.), *Avian Immunology* (pp. 187–203). Edinburgh: British Poultry Science Ltd.
- McFerran, J. B. (1997). Group I adenovirus infections. In B. W. Calnek, H. J. Barnes, C. W. Beard, L. R. McDougald, & Y. M. Saif (Eds.), *Diseases of Poultry* (10th ed., pp. 607–620). Iowa: Iowa State University Press.
- McFerran, J. B. (1998). Adenoviruses. In D. E. Swayne, J. E. Glisson, M. W. Jackwood, J. E. Pearson, & W. M. Reed (Eds.), *A Laboratory Manual for the Isolation and Identification of Avian Pathogens* (4th ed., pp. 100–106). Pennsylvania: American Association of Avian Pathologists.
- McFerran, J. B., Adair, B., & Connor, T. J. (1975). Adenoviral antigens (CELO, QBV, GAL). *American Journal of Veterinary Research*, 36, 527–529.
- McFerran, J. B., & Adair, B. M. (2003). Group I adenovirus infections. In Y. M. Saif, H. J. Barnes, J. R. Glisson, A. M. Fadly, M. L. R. R., & D. E. Swayne (Eds.), *Diseases of Poultry* (11th ed., pp. 214–227). Iowa: Iowa State Press.
- McFerran, J. B., & Adair, B. M. C. (1977). Avian adenoviruses - a review. *Avian Pathology*, 6(3), 189–217. <https://doi.org/10.1080/03079457708418228>
- McFerran, J. B., McCracken, R. M., Connor, T. J., & Evans, R. T. (1976). Isolation of viruses from clinical outbreaks of inclusion body hepatitis. *Avian Pathology*, 5(4), 315–324. <https://doi.org/10.1080/03079457608418201>
- McFerran, J. B., & Smyth, J. A. (2000). Avian adenoviruses. *Revue Scientifique et Technique*, 19, 589–601.
- Mei, Y., & Wadell, G. (1995). Highly heterogenous fiber genes in the two closely related adenovirus genome types Ad35p and Ad34a. *Virology*, 206, 686–689.
- Mettifogo, E., Nuñez, L. F. N., Parra, S. H. S., Astolfi-ferreira, C. S., Ferreira, A.

- J. P., E. A. M., ... H. S. P. S. (2014). Fowl adenovirus group I as a causal agent of inclusion body hepatitis / hydropericardium syndrome (IBH/HPS) outbreak in brazilian broiler flocks. *Pesquisa Veterinária Brasileira*, 34(8), 733–737.
- Meulemans, G., Boschmans, M., & van den Berg, T. P Decaesstecker, M. (2001). Polymerase chain reaction combined with restriction enzyme analysis for detection and differentiation of fowl adenoviruses. *Avian Pathology*, 30, 655–660. <https://doi.org/10.1080/03079450120092143>
- Meulemans, G., Couvreur, B., Decaesstecker, M., Berg, T. P. Van Den, Meulemans, G., Couvreur, B., ... Boschmans, M. (2004). Phylogenetic analysis of fowl adenoviruses. *Avian Pathology*, 33(2), 164–170. <https://doi.org/10.1080/03079450310001652086>
- Molinier-Frenkel, V., Lengagne, R., Gaden, F., Hong, S., Choppin, J., Gahery-Segard, H., ... Guillet, J.-G. (2002). Adenovirus hexon protein is a potent adjuvant for activation of a cellular immune response. *Journal of Virology*, 76(1), 127–135. <https://doi.org/10.1128/JVI.76.1.127>
- Monreal, G. (1992). Adenoviruses and adeno-associated viruses of poultry. *Poultry Science Reviews*, 4, 1–27.
- Nakamura, K., Shoyama, T., Mase, M., Imada, T., & Yamada, M. (2003). Reproduction of hydropericardium syndrome in three-week-old cyclophosphamide-treated specific-pathogen-free chickens by adenoviruses from inclusion body hepatitis. *Avian Diseases*, 47(1), 169–174.
- Narat, M. (2003). Production of antibodies in chickens. *Food Technology and Biotechnology*, 41(3), 259–267.
- Nicklin, S. A., Wu, E., Nemerow, G. R., & Baker, A. H. (2005). The influence of adenovirus fiber structure and function on vector development for gene therapy. *Molecular Therapy*, 12(3), 384–393. <https://doi.org/10.1016/j.ymthe.2005.05.008>
- Niczyporuk, J. S., Samorek-Salamonowicz, E., & Czekaj, H. (2013). Analysis of adenovirus strains isolated from poultry in Poland. *Bulletin of the Veterinary Institut in Pulawy*, 57, 305–310.
- Norina, L., Norsharina, A., Nurnadiah, A. H., Redzuan, I., Ardy, A., & Nor-Ismaliza, I. (2016). Avian adenovirus isolated from broiler affected with inclusion body hepatitis. *Malaysian Journal of Veterinary Research*, 7(2), 121–126.

- Norrby, E., & Wadell, G. (1969). Immunological relationships between hexons of certain human adenoviruses. *Journal of Virology*, 4(5), 663–670.
- O'Neill, L. A. J., & Bowie, A. G. (2007). The family of five: TIR-domain-containing adaptors in Toll-like receptor signalling. *Nature Reviews Immunology*, 7, 353–364.
- Ojkic, D., Martin, E., Swinton, J., Vaillancourt, J. P., Boulianne, M., & Gomis, S. (2008). Genotyping of Canadian isolates of fowl adenoviruses. *Avian Pathology*, 37(1), 95–100. <https://doi.org/10.1080/03079450701805324>
- Ojkic, D., & Nagy, É. (2000). The complete nucleotide sequence of fowl adenovirus type 8. *Journal of General Virology*, 81, 1833–1837.
- Ojkic, D., & Nagy, É. (2003). Antibody response and virus tissue distribution in chickens inoculated with wild-type and recombinant fowl adenoviruses. *Vaccine*, 22, 42–48. [https://doi.org/10.1016/S0264-410X\(03\)00544-9](https://doi.org/10.1016/S0264-410X(03)00544-9)
- Okuda, Y., Ono, M., Shibata, I., & Sato, S. (2004). Pathogenicity of serotype 8 fowl adenovirus isolated from gizzard erosions of slaughtered broiler chickens. *The Journal of Veterinary Medical Science / the Japanese Society of Veterinary Science*, 66(12), 1561–1566. <https://doi.org/10.1292/jvms.66.1561>
- Olson, N. . (1951). A respiratory disease (bronchitis) of quail caused by a virus. *Veterinary Medicine*, 46(1), 22.
- Pallister, J., Wright, P. J., & Sheppard, M. (1996). A single gene encoding the fiber is responsible for variations in virulence in the fowl adenoviruses. *Journal of Virology*, 70(8), 5115–5122.
- Patterson, S., & Russell, W. C. (1983). Ultrastructural and immunofluorescence studies of early events in adenovirus-HeLa cell interactions. *Journal of General Virology*, 64, 1091–1099.
- Pereira, C. G., Marin, S. Y., Santos, B. M., Resende, J. S., Resende, M., & Gomes, A. M. (2014). Occurrence of Aviadenovirus in chickens from the poultry industry of Minas Gerais. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, 66(3), 801–808.
- Philippe, C., Grgi, H., Ojkic, D., & Nagy, E. (2007). Serologic monitoring of a broiler breeder flock previously affected by inclusion body hepatitis and testing of the progeny for vertical transmission of fowl adenoviruses. *Canadian Journal of Veterinary Research*, 71, 98–102.
- Philipson, L., & Petterson, R. F. (2004). The coxsackie-adenovirus receptor a new receptor in the immunoglobulin family involved in cell adhesion. *Current Topics in Microbiology and Immunology*, 273, 87–111.
- Pizzuto, M. S., Battisti, C. De, Marciano, S., Capua, I., & Cattoli, G. (2010).

Pyrosequencing analysis for a rapid classification of fowl adenovirus species. *Avian Pathology*, 39(5), 391–398. <https://doi.org/10.1080/03079457.2010.510499>

Popowich, S., Gupta, A., Chow-lockerbie, B., Ayalew, L., Ambrose, N., Ojkic, D., ... Gomis, S. (2018). Broad spectrum protection of broiler chickens against inclusion body hepatitis by immunizing their broiler breeder parents with a bivalent live fowl adenovirus vaccine. *Research in Veterinary Science*, 118, 262–269. <https://doi.org/10.1016/j.rvsc.2018.03.003>

Rajasekhar, R., & Roy, P. (2014). Recombinant hexon antigen based single serum dilution ELISA for rapid serological profiling against fowl adenovirus-4 causing hydropericardium syndrome in chickens. *Journal of Virological Methods*, 207, 121–127. <https://doi.org/10.1016/j.jviromet.2014.06.017>

Ramis, A., Latimer, K. S., Niagro, F. D., Campagnoli, R. P., Ritchie, B. W., & Pesti, D. (1994). Diagnosis of psittacine beak and feather disease ( PBFD ) viral infection , avian polyomavirus infection , adenovirus infection and herpesvirus infection in psittacine tissues using DNA in situ hybridization Diagnosis of psittacine beak and feather diseas. *Avian Pathology*, 23, 643–657. <https://doi.org/10.1080/03079459408419034>

Rasmussen, U. B., Schlesinger, Y., Pavirani, A., & Mehtali, M. (1995). Sequence analysis of the canine adenovirus 2 fiber-encoding gene. *Gene*, 159, 279–280.

Raue, R., Gerlach, H., & Müller, H. (2005). Phylogenetic analysis of the hexon loop 1 region of an adenovirus from psittacine birds supports the existence of a new psittacine adenovirus (PsAdV). *Archives of Virology*, 150(10), 1933–1943. <https://doi.org/10.1007/s00705-005-0578-x>

Raue, R., & Hess, M. (1998). Hexon based PCRs combined with restriction enzyme analysis for rapid detection and differentiation of fowl adenoviruses and egg drop syndrome virus. *Journal of Virological Methods*, 73, 211–217.

Reece, R. L., Barr, D. A., & Grix, D. C. (1985). An investigation of vertical transmission of a fowl adenovirus serotype 8. *Australian Veterinary Journal*, 62(4), 136–137.

Reece, R. L., Barr, D. A., Grix, D. C., Forsyth, W. M., Condon, R. J., & Hindmarsh, M. (1986). Observations on naturally occurring inclusion body hepatitis in Victorian chickens. *Australian Veterinary Journal*, 63(6), 201–202.

Roberts, M. M., White, J. L., Grotter, M. G., Burnetrt, R. M., Grutter, M. G., & Burnett, R. M. (1986). Three-dimensional structure of the adenovirus major coat protein hexon. *Science*, 232, 1148–1151.

- Rodríguez, J., Koga, Y., Alvarado, A., & Tinoco, R. (2014). Molecular characterization of Peruvian fowl adenovirus ( FAdV ) isolates. *Advances in Microbiology*, 4(August), 595–603.
- Roelvink, P. W., Lee, G. M., Einfeld, D. A., Kovacs, I., & Wickham, T. J. (1999). Identification of a conserved receptor-binding site on the fiber proteins of CAR-recognizing Adenoviridae. *Science*, 286, 1568–1571.
- Rosenberger, J. K., Eckroade, R. J., Klopp, S., & Krauss, W. C. (1974). Characterization of several viruses isolated from chickens with inclusion body hepatitis and aplastic anemia. *Avian Diseases*, 18(3), 399–409.
- Rosenberger, J. K., Klopp, S., Eckroade, R. J., & Krauss, W. C. (1975). The roles of the infectious bursal agent and several avian adenoviruses in the hemorrhagic-aplastic-anemia syndrome and gangrenous dermatitis. *Avian Diseases*, 19(4), 717–729.
- Roy, H., Bhardwaj, S., Babu, M., Jauhainen, S., Herzig, K., Bellu, A. R., ... Carmeliet, P. (2005). Adenovirus-mediated gene transfer of placental growth factor to perivascular tissue induces angiogenesis via upregulation of the expression of endogenous vascular endothelial growth factor-A. *Human Gene Therapy*, 16, 1422–1428.
- Roy, P., Koteeswaran, A., & Manickam, R. (1999). Efficacy of an inactivated oil emulsion vaccine against hydropericardium syndrome in broilers. *Veterinary Record*, 145, 458–459.
- Roy, P., Vairamuthu, S., Sakthivelan, S. M., & Purushothaman, V. (2004). Hydropericardium syndrome in Japanese quail (*Coturnix coturnix japonica*). *Veterinary Record*, 155, 273–275.
- Ruan, S., Zhao, J., Ren, Y., Feng, J., & Zhang, G. (2017). Phylogenetic analyses of fowl adenoviruses (FAdV) isolated in China and pathogenicity of a FAdV-8 isolate. *Avian Diseases*, 61(3), 353–357.
- Ruan, W., Wu, Y., & Zheng, S. J. (2012). Different genetic patterns in avian Toll-like receptor (TLR) 5 genes. *Molecular Biology Reports*, 39, 3419–3426. <https://doi.org/10.1007/s11033-011-1113-7>
- Rueckert, R. R. (1990). Picornaviridae and their replication. In B. N. Fields & D. M. Knipe (Eds.), *Virology* (2nd ed., pp. 507–548). New York: Raven Press.

- Russell, W. C. (2009). Adenoviruses : update on structure and function. *Journal of General Virology*, 90(2009), 1–20. <https://doi.org/10.1099/vir.0.003087-0>
- S, H. J., Mullis, K. G., & Engler, J. A. (1988). Characterization of the early region 3 and fiber genes of Ad7. *Virology*, 167(2), 545–553.
- Saifuddin, M., & Wilks, C. R. (1990a). Development of an enzyme-linked immunosorbent assay to detect and quantify adenovirus in chicken tissues. *Avian Diseases*, 34(2), 239–245.
- Saifuddin, M., & Wilks, C. R. (1990b). Reproduction of inclusion body hepatitis in conventionally raised chickens inoculated with a New Zealand isolate of avian adenovirus. *New Zealand Veterinary Journal*, 38(2), 62–65.
- Saifuddin, M., & Wilks, C. R. (1991a). Pathogenesis of an acute viral hepatitis: inclusion body hepatitis in the chicken. *Archives of Virology*, 116, 33–43.
- Saifuddin, M., & Wilks, C. R. (1991b). Vertical transmission of avian adenovirus associated with inclusion body hepatitis. *New Zealand Veterinary Journal*, 39(2), 50–52.
- Saifuddin, M., & Wilks, C. R. (1992). Effects of fowl adenovirus infection on the immune system of chickens. *Journal of Comparative Pathology*, 107(3), 285–294.
- Sambrook, J., Sleigh, M., Engler, J. A., & Broker, T. R. (1980). The evolution of the adenoviral genome. *Annals New York Academy of Sciences*, 426–452.
- Samuel, C. E. (2001). Antiviral actions of interferons. *Clinical Microbiology Reviews*, 14(4), 778–809. <https://doi.org/10.1128/CMR.14.4.778>
- Sandhu, B. S., Singh, H., & Singh, B. (1995). Pathological studies in broiler chicks fed aflatoxin or ochratoxin and inoculated with inclusion body hepatitis virus singly and in concurrence. *Veterinary Research Communications*, 19(1), 27–37.
- Schachner, A., Marek, A., Grafl, B., & Hess, M. (2016). Detailed molecular analyses of the hexon loop-1 and fibers of fowl aviadenoviruses reveal new insights into the antigenic relationship and confirm that specific genotypes are involved in field outbreaks of inclusion body hepatitis. *Veterinary Microbiology*, 186, 13–20. <https://doi.org/10.1016/j.vetmic.2016.02.008>

- Schachner, A., Marek, A., Jaskulska, B., Bilic, I., & Hess, M. (2014). Recombinant FAdV-4 fiber-2 protein protects chickens against hepatitis – hydropericardium syndrome (HHS). *Vaccine*, 32(9), 1086–1092. <https://doi.org/10.1016/j.vaccine.2013.12.056>
- Schachner, A., Matos, M., Grafl, B., & Hess, M. (2017). Fowl adenovirus (FAdV) induced diseases and strategies for their control – a review on the current global situation. *Avian Pathology*, 1–77. <https://doi.org/10.1080/03079457.2017.1385724>
- Schade, R., Calzado, E. G., Sarmiento, R., Chacana, P. A., Porankiewicz-Asplund, J., & Terzolo, H. R. (2005). Chicken egg yolk antibodies (IgY-technology): a review of progress in production and use in research and human and veterinary medicine. *Alternatives to Laboratory Animals*, 33(2), 129–154.
- Schonewille, E., Singh, A., Göbel, T. W., Gerner, W., Saalmüller, A., & Hess, M. (2008). Fowl adenovirus (FAdV) serotype 4 causes depletion of B and T cells in lymphoid organs in specific pathogen-free chickens following experimental infection. *Veterinary Immunology and Immunopathology*, 121, 130–139. <https://doi.org/10.1016/j.vetimm.2007.09.017>
- Senne, D. A. (1998). Virus propagation in embryonating eggs. In D. Swayne, J. R. Glisson, M. W. Jackwood, J. E. Pearson, & W. M. Reed (Eds.), *A Laboratory Manual for the Isolation and Identification of Avian Pathogens* (4th ed., pp. 235–240). Pennsylvania: American Association of Avian Pathologists.
- Shah, M. S., Ashraf, A., Khan, M. I., Rahman, M., Habib, M., Chugtai, M. I., & Qureshi, J. A. (2017). Fowl adenovirus : history , emergence , biology and development of a vaccine against hydropericardium syndrome. *Archives of Virology*, 162(1833–1843). <https://doi.org/10.1007/s00705-017-3313-5>
- Shane, S. M. (1996). Hydropericardium-hepatitis syndrome, the current world situation. *Zootecnica International, January*, 20–27.
- Sheela, R. R., Babu, U., Mu, J., Elankumaran, S., Bautista, D. A., Raybourne, R. B., ... Song, W. (2003). Immune responses against *Salmonella enterica* Serovar Enteritidis infection in virally immunosuppressed chickens. *Clinical and Diagnostic Laboratory Immunology*, 10(4), 670–679. <https://doi.org/10.1128/CDLI.10.4.670>
- Sheppard, M., Werner, W., & Johnson, M. A. (1998). DNA sequence of the fowl adenovirus serotype 10 short fiber gene. *DNA Sequence: The Journal of DNA Sequencing and Mapping.*, 8(6), 391–396.

- Signas, C., Akusjarvi, G., & Pettersson, U. (1985). Adenovirus 3 fiber polypeptide gene: implications for the structure of the fiber protein. *Journal of Virology*, 53(2), 672–678.
- Singh, A., Grewal, G. S., Maiti, N. K., & Oberoi, M. S. (2006). Effect of fowl adenovirus-1 (IBH isolate) on humoral and cellular immune competency of broiler chicks. *Comparative Immunology, Microbiology and Infectious Diseases*, 29(5–6), 315–321. <https://doi.org/10.1016/j.cimid.2006.08.001>
- Smyth, J. A., & McNulty, M. . (2008). Adenoviridae. In M. Pattison, P. F. McMullin, J. M. Bradbury, & D. Alexander (Eds.), *Poultry Diseases* (6th ed., pp. 367–381). UK: Butterworth Heinemann.
- Sohaimi, N. M., Bejo, M. H., Omar, A. R., Ideris, A., & Isa, N. M. (2018). Hexon and fiber gene changes in an attenuated fowl adenovirus isolate from Malaysia in embryonated chicken eggs and its infectivity in chickens. *Journal of Veterinary Science*, 19(6), 759–770.
- Stallwood, Y., Fisher, K. D., Gallimore, P. H., & Mautner, V. (2000). Neutralisation of adenovirus infectivity by ascitic fluid from ovarian cancer patients. *Gene Therapy*, 7, 637–643.
- Steer, P. A., Kirkpatrick, N. C., O'Rourke, D., & Noormohammadi, A. H. (2009). Classification of fowl adenovirus serotypes by use of high-resolution melting-curve analysis of the hexon gene region. *Journal of Clinical Microbiology*, 47(2), 311–321. <https://doi.org/10.1128/JCM.01567-08>
- Steer, P. A., & Noormohammadi, A. H. (2011). *Inclusion body hepatitis outbreaks in Australian meat breeder and broiler flocks*. Barton: Rural Industries Research and Development Corporation.
- Steer, P. A., Sandy, J. R., O'Rourke, D., Scott, P. C., Browning, G. F., Noormohammadi, A. H., ... Noormohammadi, A. H. (2015). Chronological analysis of gross and histological lesions induced by field strains of fowl adenovirus serotypes 1, 8b and 11 in one-day-old chickens. *Avian Pathology*, 44(2), 106–113. <https://doi.org/10.1080/03079457.2015.1007919>
- Stone, D., Liu, Y., Shayakhmetov, D., Li, Z., Ni, S., & Lieber, A. (2007). Adenovirus-platelet interaction in blood causes virus sequestration to the reticuloendothelial system of the liver. *Journal of Virology*, 81(9), 4866–4871. <https://doi.org/10.1128/JVI.02819-06>
- Sumida, S. M., Truitt, D. M., Angelique, A. C., Vogels, R., Custers, J. H. H. V., Addo, M., ... Barouch, D. H. (2005). Neutralizing antibodies to adenovirus serotype 5 vaccine vectors are directed primarily against the adenovirus hexon protein. *The Journal of Immunology*, 174, 7179–7185. <https://doi.org/10.4049/jimmunol.174.11.7179>
- Takaoka, A., Wang, Z., Choi, M. K., Yanai, H., Negishi, H., Ban, T., ...

- Taniguchi, T. (2007). DAI (DLM-1/ZBP1) is a cytosolic DNA sensor and an activator of innate immune response. *Nature*, 448, 501–506. <https://doi.org/10.1038/nature06013>
- Takeuchi, O., & Akira, S. (2009). Innate immunity to virus infection. *Immunological Reviews*, 227(1), 75–86. <https://doi.org/10.1111/j.1600-065X.2008.00737.x>.Innate
- Tamura, K., Stecher, G., & Kumar, S. (2016). MEGA7: Molecular Evolutionary Genetics Analysis (MEGA) software version 7.0. *Molecular Biology and Evolution* 33.
- Temperley, N. D., Berlin, S., Paton, I. R., Griffin, D. K., & Burt, D. W. (2008). Evolution of the chicken Toll-like receptor gene family: A story of gene gain and gene loss. *BMC Genomics*, 9(62), 1–12. <https://doi.org/10.1186/1471-2164-9-62>
- Thaci, B., Ulasov, I. V., Wainwright, D. A., & Lesniak, M. S. (2011). The challenge for gene therapy: Innate immune response to adenoviruses. *Oncotarget*, 2(3), 113–121.
- Toivanen, A., Toivanen, P., & Lassila, E. J. (1981). Ontogeny of the chicken lymphoid system. In M. E. Rose, L. N. Payne, & B. M. Freeman (Eds.), *Avian Immunology* (pp. 45–62). Edinburgh: British Poultry Science Ltd.
- Toogood, C. I. A., Crompton, J., & Hay, R. T. (1992). Antipeptide antisera define neutralizing epitopes on the adenovirus hexon. *Journal of General Virology*, 73, 1429–1435.
- Toro, A. H., González, C., Cerdá, L., Morales, M. A., Dooner, P., Salamero, M., ... Sa, M. (2002). Prevention of inclusion body hepatitis / hydropericardium syndrome in progeny chickens by vaccination of breeders with fowl adenovirus and chicken anemia virus. *Avian Diseases*, 46(3), 547–554. [https://doi.org/10.1637/0005-2086\(2002\)046\[0547:POIBHH\]2.0.CO;2](https://doi.org/10.1637/0005-2086(2002)046[0547:POIBHH]2.0.CO;2)
- Toro, H., Gonzalez, C., Cerdá, L., Hess, M., Reyes, E., & Geisse, C. (2000). Chicken anemia virus and fowl adenoviruses : Association to induce the inclusion body hepatitis / hydropericardium syndrome. *Avian Diseases*, 44(1), 51–58.
- Toro, H., Gonzalez, O., Escobar, C., Cerdá, L., Morales, M. A., & Gonzalez, C. (2001). Note- vertical induction of the inclusion body hepatitis / hydropericardium syndrome with fowl adenovirus and chicken anemia virus. *Avian Diseases*, 45, 215–222.

- van den Hurk, J. V., & van Drunen Littel-van den Hurk, S. (1988). Characterization of group II avian adenoviruses with a panel of monoclonal antibodies. *Canadian Journal of Veterinary Research*, 52, 458–467.
- Vrati, S., Boyle, D., Kocherhans, R., & Both, G. W. (1995). Sequence of ovine adenovirus homologs for 100K hexon assembly, 33K, pVIII, and fiber genes: early region E3 is not in the expected location. *Virology*, 209, 400–408.
- Walters, R. W., Freimuth, P., Moninger, T. O., Ganske, I., Zabner, J., & Welsh, M. J. (2002). Adenovirus fiber disrupts CAR-mediated intercellular adhesion allowing virus escape. *Cell*, 110, 789–799.
- Wang, J., Wang, J., Chen, P., Liu, L., & Yuan, W. (2017). Development of a TaqMan-based real-time PCR assay for rapid and specific detection of fowl aviadenovirus serotype 4. *Avian Pathology*, 46(3), 338–343. <https://doi.org/10.1080/03079457.2016.1278428>
- Waye, M. M. Y., & Sing, C. W. (2010). Anti-viral drugs for human adenoviruses. *Pharmaceuticals*, 3, 3343–3354. <https://doi.org/10.3390/ph3103343>
- Wells, R. J. H., Westbury, H. A., Harrigan, K. E., Coleman, G. D. C., & Beilharz, R. G. (1977). Epidemic adenovirus inclusions body hepatitis of the chicken in Australia. *Australian Veterinary Journal*, 53, 586–590.
- Wibowo, M., Jonas, M., Murwijati, T., Mahardika, G., Sahesty, A., Mahardika, B., ... Suryanggono, J. (2019). Epizootiology, clinical signs, and phylogenetic analysis of fowl adenovirus in chicken farms in Indonesia from 2018 to 2019. *Avian Diseases*.
- Wigley, P., Barrow, P., & Schat, K. A. (2014). The avian reproductive immune system. In K. A. Schat, B. Kaspars, & P. Kaiser (Eds.), *Avian Immunology* (2nd ed., pp. 265–274). Elsevier Ltd. <https://doi.org/10.1016/B978-0-12-396965-1.00015-7>
- Winterfield, R. W., Fadly, A. M., & Galina, A. M. (1973). Adenovirus infection and disease . I . Some characteristics of an isolate from chickens in Indiana. *American Association of Avian Pathologist*, 17(2), 334–342.
- Wold, W. S., & Horwitz, M. S. (2007). Adenoviruses. In D. M. Knipe & P. M. Howley (Eds.), *Fields Virology* (2nd ed., pp. 2395–2436). Philadelphia: Lippincott Williams and Wilkins.
- Wu, E., Pache, L., Seggern, D. J. Von, Mullen, T., Mikyas, Y., Stewart, P. L., & Nemerow, G. R. (2003). Flexibility of the adenovirus fiber is required for efficient receptor interaction. *Journal of Virology*, 77(13), 7225–7235. <https://doi.org/10.1128/JVI.77.13.7225>
- Wu, H., Han, T., Belousova, N., Krasnykh, V., Kashentseva, E., Dmitriev, I., ...

- Curiel, D. T. (2005). Identification of sites in adenovirus hexon for foreign peptide incorporation. *Journal of Virology*, 79(6), 3382–3390. <https://doi.org/10.1128/JVI.79.6.3382>
- Xia, J., Yao, K., Liu, Y., You, G., Li, S., Liu, P., ... Huang, Y. (2017). Isolation and molecular characterization of prevalent Fowl adenovirus strains in southwestern China during 2015 – 2016 for the development of a control strategy. *Emerging Microbes & Infections*, 6(103), 1–9. <https://doi.org/10.1038/emi.2017.91>
- Xie, Z., Luo, S., Fan, Q., Xie, L., Liu, J., Xie, Z., ... Wang, X. (2013). Detection of antibodies specific to the non-structural proteins of fowl adenoviruses in infected chickens but not in vaccinated chickens. *Avian Pathology*, 42(5), 491–496. <https://doi.org/10.1080/03079457.2013.829553>
- Xie, Z., Tang, Y., Fan, Q., Liu, J., Pang, Y., Xie, Z., ... Khan, M. I. (2011). Rapid detection of group I avian adenoviruses by a loop-mediated isothermal amplification. *Avian Diseases*, 55(4), 575–579.
- Yates, V. J., & Fry, D. E. (1957). Observations on a chicken embryo lethal orphan (CELO) virus. *American Journal of Veterinary Research*, 18(68), 657–660.
- Zachary, J. F. (2011). Mechanisms of microbial infections. In M. McGavin (Ed.), *Pathologic Basis of Veterinary Disease* (5th ed., pp. 196–200). Missouri: Elsevier.
- Zadravec, M., Slavec, B., Krapez, U., Juntes, P., Jursic, C. R., Benko, M., & Zorman, R. O. (2013). Inclusion body hepatitis (IBH) outbreak associated with Fowl adenovirus type 8b in broilers. *Acta Veterinaria*, 63(1), 101–110. <https://doi.org/10.2298/AVB1301101Z>
- Zhang, Yuanming, & Bergelson, J. M. (2005). Minireview: Adenovirus receptors. *Journal of Virological Methods*, 79(19), 12125–12131. <https://doi.org/10.1128/JVI.79.19.12125>
- Zhang, Yuhan, Liu, R., Tian, K., Wang, Z., Yang, X., Gao, D., ... Zhao, J. (2018). Fiber2 and hexon genes are closely associated with the virulence of the emerging and highly pathogenic fowl adenovirus 4. *Emerging Microbes & Infections*, 7, 1–10. <https://doi.org/10.1038/s41426-018-0203-1>
- Zhao, J., Zhong, Q., Zhao, Y., Hu, Y. X., & Zhang, G. Z. (2015). Pathogenicity and complete genome characterization of fowl adenoviruses isolated from chickens associated with inclusion body hepatitis and hydropericardium syndrome in China. *PLoS ONE*, 10(7), 1–14. <https://doi.org/10.1371/journal.pone.0133073>