



**SELECTION OF CULTURE CONDITIONS THROUGH MANIPULATION OF
PHYSICAL AND CHEMICAL PARAMETERS FOR PRODUCTION OF HIGH-
VALUE METABOLITES IN MARINE MICROALGA *Tetraselmis tetraathele*
(WEST) BUTCHER 1959 BIOMASS**

By

NURUL FARAHIN BINTI ABD WAHAB

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of
Doctor of Philosophy**

June 2021

IB 2022 5

All materials contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, are copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purpose from copyright holder. Commercial use materials may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Science has always been a passion of mine. Dedicating my life to improve the lives of others is very pleasing to me. Whatever I had gone through before has led me to be the person that I am today. Special dedication of this grateful feeling to ...

*my beloved father and mother,
Mr. Abd Wahab Ahmad and Mrs. Norsiah Harron;*

*my beloved son,
Muhammad Faiz Firdaus;*

my loving brothers and sisters; and

all whom I love

for their patience, unconditional love and encouragement throughout the course of this work.

I love you.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

SELECTION OF CULTURE CONDITIONS THROUGH MANIPULATION OF PHYSICAL AND CHEMICAL PARAMETERS FOR PRODUCTION OF HIGH-VALUE METABOLITES IN MARINE MICROALGA *Tetraselmis tetrathele* (WEST) BUTCHER 1959 BIOMASS

By

NURUL FARAHIN BINTI ABD WAHAB

June 2021

Supervisor: Assoc. Prof. Natrah Fatin Mohd Ikhsan, PhD
Institute: Bioscience

Microalgae cultivation is one of the crucial aspects in the commercialization of microalgae to produce a high amount of biomass and metabolites. In the aquaculture industry, microalgae biomass is used as feed and growth enhancers, and it is also considered a renewable and sustainable resource. Commercially, *Tetraselmis* sp. is one of the most commonly used microalgae in the aquaculture field. Although there are several studies on the benefits and culture conditions of *Tetraselmis* sp., little information is known about the understanding of theoretical and technical knowledge on mass culture, which can affect biomass productivity and the quality of microalgae biomass produced. The present study aims to obtain key insights of three growth factors considered as major contributors on the effect of microalgae growth: (1) ammonium nitrogen concentration (chemical parameter), (2) light intensity, and (3) culture temperature (physical parameters) for mass production of an indigenous species, *Tetraselmis tetrathele* under tropical conditions. Bubble column reactors (BCRs) were used to mimic indoor and outdoor conditions in enhancing the growth characteristics of cells, the effect of physiological processes, and the composition of metabolites. Overall, this study revealed that although the growth performance of *T. tetrathele* decreased under 35 °C, this indigenous species showed excellent self-adaptation capabilities to cope with high ammonium nitrogen (0.87 g L⁻¹) and varying light intensities (up to 1,500 μmol m⁻² s⁻¹) by protecting microalgae from photodamage. These characteristics have significant implications for the selection of optimal conditions when designing more efficient microalgae culture systems in tropical conditions. The knowledge obtained from this work can be useful in assessing the applicability of this strain culture and also enhancing the understanding of the physiology of microalgae to sustainably maximize microalgae cultivation. Besides, these findings are particularly useful for relevant stakeholders to efficiently expand commercialization by selecting

high-quality biomass production with specific metabolites of interest in *T. tetrahele*.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PEMILIHAN KEADAAN PENGKULTURAN MELALUI MANIPULASI
PARAMETER FIZIKAL DAN KIMIA UNTUK PENGHASILAN METABOLIT
TINGGI-NILAI DALAM BIOJISIM MIKROALGA MARIN *Tetraselmis*
tetrathele (WEST) BUTCHER 1959**

Oleh

NURUL FARAHIN BINTI ABD WAHAB

Jun 2021

Penyelia: Prof. Madya. Natrah Fatin Mohd Ikhsan, PhD
Institut: Biosains

Pembiakan mikroalga adalah salah satu aspek yang penting dalam pengkomersilan mikroalga untuk menghasilkan biojisim dan metabolit. Dalam industri akuakultur, biojisim mikroalga digunakan sebagai bahan makanan dan penggalak pertumbuhan, dan ia juga dianggap sebagai sumber boleh diperbaharui dan mapan. Secara komersilnya, *Tetraselmis* sp. adalah salah satu daripada spesies mikroalga yang sering digunakan dalam bidang akuakultur. Walaupun terdapat banyak kajian mengenai manfaat dan keadaan pengkulturan *Tetraselmis* sp., hanya terdapat sedikit maklumat mengenai pengetahuan teknikal dan teori terhadap kultur spesies tersebut yang boleh mempengaruhi produktiviti biojisim dan kualiti biojisim mikroalga yang terhasil. Kajian ini bertujuan mendapatkan pemahaman penting mengenai tiga faktor pertumbuhan yang dianggap sebagai penyumbang utama terhadap kesan pertumbuhan mikroalga: (1) kepekatan ammonia nitrogen (parameter kimia), (2) keamatan cahaya, dan (3) suhu kultur (parameter fizikal) untuk pengeluaran besar-besaran sejenis spesies tempatan, *Tetraselmis tetrathele* dalam keadaan tropika. Reaktor turus gelembung (BCRs) digunakan untuk menyerupai keadaan dalam dan luar untuk meningkatkan ciri-ciri pertumbuhan sel, kesan proses fisiologi, dan komposisi metabolit. Secara keseluruhannya, kajian ini menunjukkan bahawa walaupun prestasi pertumbuhan *T. tetrathele* berkurang di bawah suhu 35 °C, namun spesies tempatan ini menunjukkan keupayaan adaptasi sendiri untuk menghadapi tahap ammonia nitrogen yang tinggi (0.87 g L⁻¹) dan keamatan cahaya yang berubah-ubah (sehingga 1,500 μmol m⁻² s⁻¹) dengan melindungi mikroalga daripada kerosakan disebabkan oleh cahaya. Ciri-ciri ini mempunyai kesan yang signifikan terhadap pemilihan keadaan optimum apabila mereka bentuk sistem pengkulturan mikroalga yang lebih berkesan dalam keadaan tropika. Pengetahuan yang diperolehi daripada kajian ini berguna untuk menilai kebolegunaan kultur strain ini dan juga meningkatkan pemahaman mengenai fisiologi mikroalga untuk memaksimumkan pembiakan mikroalga

secara mapan. Selain itu, dapatan kajian ini sangat berguna untuk pihak berkepentingan yang berkaitan untuk mengembangkan pengkomersilan secara berkesan dengan memilih penghasilan biojisim berkualiti tinggi dengan metabolit khusus iaitu *T. tetrahele*.



ACKNOWLEDGEMENTS

Assalamualaikum. Praise to Allah (S.W.T) the all-powerful, the wisest, and the most merciful, and greetings to His Messenger, the Saviour of mankind, Prophet Muhammad (S.A.W), his family, Moslems present and past. To Allah the Almighty, belong all praise and glory. Without His will and blessing, I would not have succeeded in completing this thesis.

First and foremost, I would like to extend my deepest appreciation to my supervisors, Assoc. Prof. Dr. Natrah Fatin Mohd Ikhsan and Dr. Norio Nagao, for their generous guidance, brilliant discussion, advice, and endless support that significantly contributed towards the completion of this research. Their careful reviews and constructive criticism were crucially important for the completion of this thesis. I would also like to extend my sincerest gratitude to my co-supervisors, Prof. Dato' Dr. Mohamed Shariff Mohamed Din and Prof. Dr. Fatimah Md. Yusoff, for their constructive advice, valuable guidance, priceless comment, and invaluable advice throughout the entire course of this research.

In addition, I would like to extend my special gratitude to the staff from the Laboratory of Marine Biotechnology, Institute of Bioscience, Universiti Putra Malaysia and the Department of Aquaculture, Universiti Putra Malaysia, especially Ms. Fadzillah Abdul Razak and Ms. Norhafizah Roslan, for their friendship, support, and cooperation. I wish to convey my deepest gratitude to my friends, especially Ms. Balqis, Ms. Nursuhayati, Ms. Adibah, Mrs. Fareha, Mrs. Shariza, Mrs. Atikah, Mrs. Nawwar, Ms. Wahidah, Ms. Laisha, Mr. Yuki Imaizumi, Dr. Norulhuda Ramli, Dr. Armania Nurdin, and Dr. Raihanah Ridzuan, for their continuous love, support, and encouragement.

Besides that, my sincerest thanks to Japan Science and Technology Agency (JST) or Japan International Cooperation Agency (JICA), Science and Technology Research Partnership for Sustainable Development (SATREPS) through the project for Continuous Operation System for Microalgae Production Optimised for Sustainable Tropical Aquaculture (COSMOS) (Grant No. JPMJSA1509) and the Ministry of Education Malaysia (MOE) for the SATREPS-COSMOS Matching Fund for funding this research.

Finally, and yet the most important, I would like to express my heartiest appreciation to my parents, son, and family members for their endless support, understanding, unconditional love, and prayers throughout my study.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Natrah Fatin binti Mohd Ikhsan, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Fatimah binti Md. Yusoff, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Mohamed Shariff bin Mohamed Din, PhD

Professor Dato'
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Norio Nagao, PhD

Researcher,
Bluescientific Shinkamigoto Co., Ltd,
Japan
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 10 February 2022

Declaration by graduate student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustrations and citations have been duly referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institution;
- Intellectual property from the thesis and copyright of thesis are fully owned by Universiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtained from supervisor and the office of Deputy Vice-Cancellor (Research and Innovation) before thesis published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules, or any other material as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/ fabrication in the thesis, and Scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Nurul Farahin binti Abd Wahab

Declaration by Members of Supervisory Committee

This is to confirm that:

- The research conducted and the writing of this thesis was under our supervision.
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to:

Signature : _____

Name of Chairman of supervisory committee : Assoc. Prof. Dr. Natrah Fatin Mohd. Ikhsan

Signature : _____

Name of Member of supervisory committee : Prof. Dr. Fatimah Md. Yusoff

Signature : _____

Name of Member of supervisory committee : Prof. Dato' Dr. Mohamed Shariff Mohamed Din

Signature : _____

Name of Member of supervisory committee : Dr. Norio Nagao

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xiv
LIST OF APPENDICES	xxi
LIST OF SYMBOLS AND ABBREVIATIONS	xxii
CHAPTER	
1 INTRODUCTION	1
1.1 Background of Study	1
1.2 Problem Statement	2
1.3 Research Objectives	4
2 LITERATURE REVIEW	5
2.1 Microalgae	5
2.2 Photosynthesis in Microalgae	6
2.3 Culture Conditions Affecting Growth Performance and Metabolite Compositions in Microalgae	15
2.3.1 Nutrient Availability	16
2.3.2 Light	34
2.3.3 Temperature	41
2.3.4 Other Parameters	50
2.4 General Characteristics of <i>Tetraselmis tetrathele</i>	51
3 TOLERANCE OF <i>Tetraselmis tetrathele</i> TO HIGH AMMONIUM NITROGEN AND ITS EFFECT ON GROWTH RATE, CAROTENOIDS CONTENT AND FATTY ACIDS PRODUCTIVITY	53
3.1 Introduction	53
3.2 Materials and Methods	54
3.2.1 Microalga Culture and Media Preparation	54
3.2.2 Experimental Design of the Growth Conditions with High Level of Nitrogen Sources	55
3.2.3 Growth Parameter Analysis	56
3.2.4 Chlorophyll Fluorescence Analysis	57
3.2.5 Quantification of Cellular Photosynthetic Pigments	57

3.2.6	Fatty Acid Methyl Esters (FAME) analysis	58
3.2.7	Data Handling and Statistical Analysis	59
3.3	Results and Discussion	60
3.3.1	Growth Performance of <i>Tetraselmis tetrathele</i> under Different Concentrations of Ammonium	60
3.3.2	Maximum photosynthetic efficiency (F_v/F_m)	62
3.3.3	Influence of Ammonium Nitrogen Availability on Pigments and Fatty Acids Profiles	64
3.4	Conclusion	68
4	HIGH INTENSITY OF LIGHT: A POTENTIAL STIMULUS FOR MAXIMIZING BIOMASS BY INDUCING PHOTOSYNTHETIC ACTIVITY IN MARINE MICROALGA, <i>Tetraselmis tetrathele</i> (WEST) BUTCHER 1959	69
4.1	Introduction	69
4.2	Materials and Methods	70
4.2.1	Microalga Growth Media	70
4.2.2	Photobioreactor Setup	71
4.2.3	Experimental Design on Growth Conditions with Different Light Intensities	72
4.2.4	Growth Parameter Analysis	72
4.2.5	Calculation of Parameters	73
4.2.6	Chlorophyll Fluorescence Analysis	73
4.2.7	Quantification of Cellular Photosynthetic Pigments	74
4.2.8	Lipids and Fatty Acid Methyl Ester (FAME) Profiling	75
4.2.9	Statistical analysis	75
4.3	Results and Discussion	76
4.3.1	Effects of Different Light Intensities on Microalga Growth Rate	76
4.3.2	Characteristics of Chlorophyll Fluorescence of Photosynthetic Activity	86
4.3.3	Effects of Different Light Intensities on the Production of Pigments	89
4.3.4	Effects of Different Light Intensities on the Compositions of Lipids and Fatty acids	92
4.4	Conclusion	98
5	EFFECTS OF TEMPERATURE ON THE GROWTH RATE, PIGMENTS PRODUCTION, LIPIDS AND FATTY ACIDS PROFILES OF MARINE MICROALGA, <i>Tetraselmis tetrathele</i> (WEST) BUTCHER 1959 IN SEMI-CONTINUOUS CULTURE	99
5.1	Introduction	99
5.2	Materials and Methods	100

5.2.1	Microalga and Culture Medium	100
5.2.2	Experimental Design on Growth Conditions with Different Temperature	101
5.2.3	Growth Measurement	102
5.2.4	Chlorophyll Fluorescence Analysis	103
5.2.5	Extraction and Quantification of Photosynthetic Pigments	104
5.2.6	Evaluation of Lipids and Fatty Acid Methyl Ester (FAME) Profiling	105
5.2.7	Statistical Analysis	105
5.3	Results and Discussion	106
5.3.1	Effect of Temperature on the Growth Productivity of <i>Tetraselmis tetraathele</i>	106
5.3.2	Characteristics of Chlorophyll Fluorescence of Photosynthetic Activity	112
5.3.3	Effect of Temperature on the Production of Pigments	116
5.3.4	Effect of Temperature on the Compositions of Lipids and Fatty Acids	121
5.4	Conclusion	124
6	SUMMARY, GENERAL CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	125
6.1	Summary and General Conclusion	125
6.2	Recommendations for Future Research	126
	REFERENCES	127
	APPENDICES	148
	BIODATA OF STUDENT	150
	LIST OF PUBLICATIONS	151

LIST OF TABLES

Table		Page
2.1	Evaluation of ammonium nitrogen activities from microalgae	18
2.2	Effect of abiotic stress (light) on microalgae biomass and metabolites	36
2.3	Maximum dry-cell weight and specific growth rate achieved under temperature stress	43
2.4	Effect of temperature on microalgae metabolites	46
3.1	Different concentrations of nitrogen sources and pH in culture media at time 0	55
3.2	Fatty acid composition (% of total fatty acids) of <i>Tetraselmis tetrathele</i> grown at day 6 in different concentration of nitrogen sources and review on two marine microalgae species.	67
4.1	Maximum dry-cell weight achieved in various culture conditions of <i>Tetraselmis</i> sp.	82
4.2	Various studies on the adaptation of microalgae to high light intensity	85
4.3	Composition of fatty acids (% of total fatty acids) in <i>Tetraselmis tetrathele</i> under different light intensities	97
5.1	Effect of temperature on microalga metabolites	109
5.2	Composition of fatty acids (% of total fatty acids) of <i>Tetraselmis tetrathele</i> grown at different temperatures	124

LIST OF FIGURES

Figure		Page
2.1	General structure of photosynthetic apparatus.	7
2.2	A simplified diagram of the carotenoid biosynthesis pathway, beginning with the conversion of isopentenyl pyrophosphate (IPP) to geranyl-geranyl pyrophosphate (GGPP), with the most prevalent carotenoids highlighted as violaxanthin, lutein, and β -carotene. Phytoene synthase (PSY), phytoene desaturase (PDS), lycopene β -cyclase (LCYB), lycopene ϵ -cyclase (LCYE).	8
2.3	Except for the biosynthesis of astaxanthin, which is only found in a few species of microalgae, the biosynthetic pathway of carotenoid production is nearly identical in most green microalgae species and higher plants	9
2.4	Photochemical work in the thylakoid membrane	13
2.5	A schematic diagram of photosynthetic rate versus light intensity curve	35
2.6	a) Dividing cells of <i>Tetraselmis tetraethele</i> under light microscope (600x magnification). Photo taken using Nikon Eclipse E600, Nikon Corp., Tokyo, Japan. Scale bar: 0.01 mm; b) Scanning electron micrograph of <i>T. tetraethele</i> in 30 ppt. Photo taken using Leo 1455 VPSEM with Oxford Inca EDX. Scale bar: 2 μ m.	52
3.1	Schematic diagram of the cultivation system.	56
3.2	Time course for the growth performance of <i>Tetraselmis tetraethele</i> at different concentrations of ammonium nitrogen in terms of (a) dry-cell weight (g-dw L ⁻¹) and (b) cell number (cells mL ⁻¹).	61
3.3	(a) Time course of maximum quantum yield of PS II (F_v/F_m) during culture period of 144 hours and (b) relationship of relative F_v/F_m with the ratio of free ammonia concentration for <i>Tetraselmis tetraethele</i> in 48 hours.	63
3.4	Means and standard deviations of phototrophic pigment production of <i>Tetraselmis tetraethele</i> using different concentrations of ammonia with nitrate as the control.	65
4.1	Schematic diagram of the bubble column photobioreactor. Light was provided to the surface of the water bath at 12L:12D	71

4.2	(a) Specific growth rate, (b) dry-cell weight, and (c) cell number of <i>Tetraselmis tetrathele</i> under different light intensities.	78
4.3	(a) Specific light intensity per cell and (b) cellular dry weight of <i>Tetraselmis tetrathele</i> under different light intensities.	79
4.4	Observation of <i>Tetraselmis tetrathele</i> cultivation in bubble column reactor and under microscope observation (40x magnification) when exposed in different light intensities; (a) 300 $\mu\text{mol m}^{-2} \text{s}^{-1}$, (b) 1500 $\mu\text{mol m}^{-2} \text{s}^{-1}$, (c) 2500 $\mu\text{mol m}^{-2} \text{s}^{-1}$	80
4.5	The changes of maximum photosynthetic efficiency (F_v/F_m) with time under different light intensities.	87
4.6	Comparison of rapid light curve between (a) light intensity of 300 $\mu\text{mol m}^{-2} \text{s}^{-1}$ and (b) light intensity of 1500 $\mu\text{mol m}^{-2} \text{s}^{-1}$ on Day 0 and Day 20 during the stationary phase.	88
4.7	Chlorophyll a, (b) lutein, (c) β -carotene, (d) antheraxanthin, (e) zeaxanthin, and (f) violaxanthin pigments under different light intensities.	91
4.8	Total lipid content (% dry weight) of <i>Tetraselmis tetrathele</i> under different light intensities.	93
4.9	Distribution of saturated, monounsaturated, and polyunsaturated fatty acids under different light intensities.	96
5.1	Schematic diagram of the experimental setup.	102
5.2	Effect of temperature on the (a) dry weight and cell number and (b) specific growth rate of <i>Tetraselmis tetrathele</i> under semi-continuous cultivation.	107
5.3	Maximum photosynthetic efficiency (F_v/F_m) for <i>Tetraselmis tetrathele</i> under semi-continuous cultivation at different temperatures.	112
5.4	Cellular dry weight of <i>Tetraselmis tetrathele</i> at different temperatures.	113
5.5	Photomicrographs of <i>Tetraselmis tetrathele</i> grown at (a) 25°C during the stationary phase and (b) 35°C.	114

5.6	Comparison of rapid light curve (RLC) of <i>Tetraselmis tetrathele</i> cultured at (a) 25°C, (b) 30°C, and (c) 35°C from the initial day to stationary phase and on Day 4.	115
5.7	Chlorophyll <i>a</i> , (b) chlorophyll <i>b</i> , and (c) chlorophyll <i>c</i> ₃ contents in <i>Tetraselmis tetrathele</i> under semi-continuous cultivation at different temperatures.	117
5.8	Antheraxanthin, (b) zeaxanthin, and (c) violaxanthin contents in <i>Tetraselmis tetrathele</i> under semi-continuous cultivation at different temperatures.	119
5.9	(a) Lutein and (b) β -carotene contents in <i>Tetraselmis tetrathele</i> under semi-continuous cultivation at different temperatures.	120
5.10	Total lipid content (% dry weight) of <i>Tetraselmis tetrathele</i> under semi-continuous cultivation at different temperatures.	122
5.11	Distribution of saturated, monosaturated, and polyunsaturated fatty acids in <i>Tetraselmis tetrathele</i> under semi-continuous cultivation at different temperatures.	123

LIST OF APPENDICES

Appendix		Page
A	Molecular identification	148
B	The growth performance of <i>Tetraselmis tetraathele</i> at different concentrations of ammonium nitrogen. The cultures were grown in 1L of bubble column reactor in sterilized conditions	149



LIST OF SYMBOLS AND ABBREVIATIONS

cm	Centimeter
°C	Degree celsius
DNA	Deoxyribonucleic acid
<i>ETR</i>	Electron transport rate
<i>et al.</i>	<i>et alia</i>
GCMS	Gas Chromatography Mass Spectrometry
g	Gram
g-dw L ⁻¹	Gram dry weight per liter
gL ⁻¹	Gram per liter
HPLC	High Performance Liquid Chromatpgraphy
h	Hour
α	Light-limited slope of RLC
E_k	Light-saturation index
<i>ETR</i> _{max}	Light-saturated rate of RLC
L	Liter
F_m	Maximal fluorescence measured in darkness
F_m	Maximal fluorescence under light condition
F_v/F_m	Maximum quantum efficiency defined by $(F_m - F_0)/F_m$
μM	Micro molar
$\mu\text{mol m}^{-2} \text{s}^{-1}$	Micromole per square meter per second
mg	Milligram
mg L ⁻¹	Milligram per liter
mm	Millimeter
F	Minimum fluorescence under light condition

F_0	Minimum fluorescence measured in darkness
min	Minute
M	Molar
%	Percentage
PPFD	Photosynthetic photon flux density
PS I	Photosystem I
PS II	Photosystem II
±	Plus minus
PCR	Polymerase Chain Reaction
pH	Potentiometric hydrogen ion concentration
rETR	Relative electron transport rate
RLC	Rapid light curve that are measured by plotting the ETR or rETR against the actinic PPFD
s	Second
μ	Specific growth rate
SD	Standard deviation
UPM	Universiti Putra Malaysia

CHAPTER 1

INTRODUCTION

1.1 Background of study

The rising growth and life expectancy of the global population have increased the demand for energy, healthy food, water, drugs, and other resources. This has caught the attention of the United Nations Committee on Science, Technology, and Innovation to provide scientific grounding to tackle various sustainability challenges, in line with the 2030 Agenda for Sustainable Development, which includes 17 Sustainable Development Goals (SDGs) proposed during the general assembly in September 2015. Microalgae, hailed as the “Green Gold”, have emerged as a promising source for innovative and sustainable development to solve the global food and energy crisis (Wolkers *et al.*, 2011). Microalgae have gained global attention of academicians and engineers over the past half-century for numerous reasons. The increase in the global demand for microalgae-based products is projected to amount to USD 53.43 billion by 2026, as compared to USD 32.60 billion in 2017 (Rahman, 2020). The growing market demand has resulted in mass microalgae cultivation for use in several applications, such as food and animal feed production (Lim *et al.*, 2017), pharmaceuticals (Ambati *et al.*, 2019), wastewater treatment (Schulze *et al.*, 2017), and bioenergy production (Chia *et al.*, 2018; Qu *et al.*, 2020). Owing to the increasing market demand for microalgae, microalgae cultivation becomes one of the crucial aspects to be focused on for commercialization by producing a high amount of biomass and metabolites to fulfill the demands.

The environmental conditions, such as nutrients, light, and temperature, are some of the main basic requirements for microalgae growth, which subsequently affect the quality of biomass produced. Microalgae biomass consists of numerous beneficial compounds that are useful in various markets. Microalgae are known to produce carotenoids, which are responsible for light harvesting in photosynthetic metabolism. Carotenoids play an important role in alleviating certain cancers, premature aging, cardiovascular disease, and arthritis (Ambati *et al.*, 2019), and also as a coloring agent in chewing gums, candies, and beverages (Adarme-Vega *et al.*, 2012). Besides, carotenoids also have antiaging, antiobesity, and antioxidant properties, which are considered as better alternatives for synthetic compounds (Gong and Bassi, 2016). The specific content of metabolites is strain-dependent and can be heavily influenced by the culture conditions employed. During photosynthesis, the energy converted from sunlight is stored as lipid or carbohydrate within the algae, which is then extracted from algae for energy supply. For example, the lipid content of microalgae is usually in the range of 20%–50% of the cell dry weight, sometimes exceeding 50%, and can also be as high as 80% under certain conditions (Brindhadevi *et al.*, 2021; Japar *et al.*, 2021), thus reducing the requirement of

other resources for the production of the same amount of oil. Many marine microalgae are rich in eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA), and also other fatty acids classes. Fatty acids with 14–20 carbons are used for the production of biodiesel, and polyunsaturated fatty acids (PUFAs) with more than 20 carbons are used as health food supplements and as feed for early larvae of crustacean and fish, early and late larvae mollusk, brine shrimp, copepods, and rotifers, especially DHA and EPA (Adarme-Vega *et al.*, 2012; Sun *et al.*, 2018).

Among the local marine microalgae, *Tetraselmis* sp. is recognized as one of the few species of microalgae that has been explored extensively due to its high amount and quality of intracellular content, such as PUFAs for human consumption and aquaculture feed (Farhadian *et al.*, 2008), polysaccharides for antibiotic development research (Kermanshahi-pour *et al.*, 2014), high vitamin E content (Carballo-Cárdenas *et al.*, 2003), and antioxidant for pharmaceutical and cosmeceutical purposes (Farahin *et al.*, 2019). This genus is recognized as “novel food” and even approved for human consumption of biomass by the European Union (Mantecón *et al.*, 2019). In culturing purposes, this strain displays high potential for commercial-scale production of biomass due to its high growth rate, ability to grow in high ammonium nitrogen (Farahin *et al.*, 2021) and seawater with high salinity, and also to outcompete contaminants (Pereira *et al.*, 2016). Apart from elucidating the capabilities of microalgae with high-density cultures, a better understanding of the factors influencing microalgae growth and biomass must be acquired (Borowitzka and Vonshak, 2017).

1.2 Problem statement

The enhancement of cultivation conditions using various techniques contributes to the growth and production of numerous compounds in microalgae. The effect of abiotic stress on metabolites, such as lipid, fatty acids, and pigment production from microalgae and corresponding growth rates, has been considered in previous studies (Go *et al.*, 2012; Roleda *et al.*, 2013; Michels *et al.*, 2014a; Imaizumi *et al.*, 2016). Nitrogen in the form of either ammonium (NH_4^+) or nitrate (NO_3^-) is an essential nutrient for the growth of microalgae, which subsequently contributes to the biomass produced. Several previous studies pointed out that different strains of microalgae require different levels of nitrogen uptake (Raven *et al.*, 1992; Feng *et al.*, 2020). Meanwhile, ammonium, which is the most predominant source of nitrogen, exists in urban, agricultural, and aerobic digested effluents with various concentrations, ranging from the concentration as low as $0.01 \text{ gL}^{-1}\text{-N}$ to the concentration as high as $2.0 \text{ gL}^{-1}\text{-N}$ (de la Noüe *et al.*, 1992; Cai *et al.*, 2013; Reddy *et al.*, 2017). Furthermore, ammonium nitrogen at a certain level of concentration can be toxic and inhibits the productivity of microalgae. Thus, further elucidation of the ammonium nitrogen tolerance in microalgae is needed.

Besides, commercial-scale production of biomass from this microalgae has remained uneconomical due to the challenges of achieving high biomass productivity (Gonçalves *et al.*, 2016; Pereira *et al.*, 2018). Light is vital in microalgae growth because this source allows microalgae to produce biomass and metabolites, as well as fix inorganic carbon into organic molecules (Huerlimann *et al.*, 2010; Liu *et al.*, 2019). Microalgae growth depends heavily on the degree of light penetration, and different light intensities are required for different species and also culture cell density and depth. Several studies reported that some species of microalgae could tolerate very high light intensity. The differences in light intensity tolerance are due to different cellular concentrations; moreover, the thylakoid structure of each species has different compositions of light-harvesting pigments (Conceição *et al.*, 2020). However, high illumination of microalgae culture negatively affects its photosynthetic process due to photoinhibition, resulting in diluted culture with low biomass concentrations. Hence, to better solve the above-mentioned problems, it is essential to determine microalgae adaptability to varying light intensities to control microalgae growth, especially in mass culture.

Other than light, temperature is also recognized as another key factor that controls the photosynthetic rates of microalgae and autotrophic organisms, which are thermally sensitive (Davison, 1991; Veeramani and Santhanam, 2015). According to the National Oceanic and Atmospheric Administration (NOAA) (2020), the past five years recorded the highest temperature range, and the average global temperature is now about 1.2 °C above the preindustrial level (World Meteorological Organization, 2021). These climatological conditions have affected the yield and quality of microalgae biomass. Microalgae are highly susceptible to high temperature stress, which impairs their cell functions (Mathur *et al.*, 2014). Therefore, the ability to withstand and/or acclimate to the environmental temperature variation is essential for the adaptation and survival of microalgae.

Tetraselmis tetrathele (West) Butcher 1959, a marine green microalga within the *Chlorophyta* isolated from Port Dickson, Malaysia, is recognized as one of the few potential species that can produce a large amount and high quality of intracellular content, such as PUFAs for human consumption and aquaculture feed (Juario and Storch, 1984; Fábregas *et al.*, 2001; Farhadian *et al.*, 2008; Michels *et al.*, 2014b), polysaccharides for antibiotic development research (Kermanshahi-pour *et al.*, 2014), high content of vitamin E (Carballo-Cárdenas *et al.*, 2003), and antioxidant for pharmaceutical and cosmeceutical purposes (Farahin *et al.*, 2019). Only a few studies focused on the cultivation of the *Tetraselmis* genus for high biomass production and its relationship with photosynthetic performance under stress conditions. The irradiance curves assessed by the variable chlorophyll fluorescence method provide in situ reaction (Michels *et al.*, 2014b). Based on this information, the determination of the photosynthetic system capacity and energy captured for light energy processing can be performed. Thus, to maximize the biomass yield of this microalga, this study determined the effect of high ammonium nitrogen, high light intensity, and culture temperature on the growth rate, photosynthetic activity, and production of pigments and fatty acids of *T. tetrathele* without the limitation of

nutrients and carbon dioxide supplies. The knowledge obtained from this work could be useful in assessing the applicability of this strain culture under the abiotic stress mentioned and also enhancing the understanding of the physiology of microalga to sustainably maximize the production of microalgae cultivation.

1.3 Research objectives

The main objective of this study was to investigate the range of suitable culture conditions of *T. tetrahele* for its applications under tropical conditions. For this study, bubble column reactors were used to mimic indoor and outdoor conditions in enhancing the growth characterization of cells to study the effect of physiological processes and metabolite compositions.

The specific objectives of this study using *T. tetrahele* are as follows:

- 1) To evaluate the tolerance capability in high ammonium nitrogen on growth, physiological response, and metabolite production.
- 2) To determine the growth performance of the microalga and its metabolites under high light intensity.
- 3) To investigate the effect of different temperatures on the growth rate and metabolite compositions.

With respect to the first objective, this study specifically examined the growth rate and photosynthetic efficiency (F_v/F_m) of *T. tetrahele* in different $\text{NH}_4^+\text{-N}$ concentrations, and quantified the production of pigments and PUFAs profiles (with NH_4^+ and NO_3^- as nitrogen sources) under six-day batch cultures. Meanwhile, with respect to the second objective, this study determined the effect of high light intensity on the growth rate, photosynthetic performance, and production (i.e., pigments and fatty acids) of *T. tetrahele* in semi-continuous cultures. Thirdly, with respect to the final objective, this study proceeded to investigate the effect of temperature on the growth rate, photosynthetic and physiological effects, and compositions (i.e., pigments, lipids, and fatty acids) of *T. tetrahele* under semi-continuous cultivation at the temperature of 25– 35 °C.

REFERENCES

- Abiusi, F., Sampietro, G., Marturano, G., Biondi, N., Rodolfi, L., D'Ottavio, M. & Tredici, M.R. (2014). Growth, photosynthetic efficiency, and biochemical composition of *Tetraselmis suecica* F&M-M33 grown with LEDs of different colors. *Biotechnology and Bioengineering*, 111: 956–964.
- Adarme-Vega, T. C., Lim, D. K. Y., Timmins, M., Vernen, F., Li, Y. & Schenk, P. M. (2012). Microalgal biofactories: a promising approach towards sustainable omega-3 fatty acid production. *Microbial Cell Factories*, 11(1): 1–10.
- Ambati, R.R., Gogisetty, D., Aswathanarayana, R.G., Ravi, S., Bikkina, P.N., Bo, L. & Yuepeng, S. (2019). Industrial potential of carotenoid pigments from microalgae: Current trends and future prospects. *Critical Reviews in Food Science and Nutrition*, 59(12): 1880-1902.
- Andreotti, V., Solimeno, A., Rossi, S., Ficara, E., Marazzi, F., Mezzanotte, V. & García, J. (2020). Bioremediation of aquaculture wastewater with the microalgae *Tetraselmis suecica*: Semi-continuous experiments, simulation and photo-respirometric tests. *Science of The Total Environment*, 738: 139859.
- APHA (2012). Standard methods for the examination of water and wastewater. 22nd Edn. Washington, D.C. American Public Health Association.
- Asada, K. (2006). Production and scavenging of reactive oxygen species in chloroplasts and their functions. *Plant Physiology*, 141: 391-396.
- Azov, Y. & Goldman, J.C. (1982). Free ammonia inhibition of algal photosynthesis in intensive cultures. *Applied and Environmental Microbiology*, 43(4): 735–739.
- Baker, N.R. & Oxborough, K. (2004). *Chlorophyll fluorescence as a probe of photosynthetic productivity*, in: *Chlorophyll a fluorescence*. Springer Netherlands, pp. 65–82.
- Barahoei, M., Hatamipour, M.S. & Afsharzadeh, S. (2020). CO₂ capturing by *Chlorella vulgaris* in a bubble column photo-bioreactor: Effect of bubble size on CO₂ removal and growth rate. *Journal of CO₂ Utilization*, 37: 9-19.
- Bartley, G.E. (1995). Plant carotenoids: Pigments for photoprotection, visual attraction, and human health. *Plant Cell*, 7(7):1027-1038.
- Bazdar, E., Roshandel, R., Yaghmaei, S. & Mardanpour, M.M. (2018). The effect of different light intensities and light/dark regimes on the performance of photosynthetic microalgae microbial fuel cell. *Bioresource Technology*, 261: 350–360.

- Belkin, S. & Boussiba, S. (1991). Resistance of *Spirulina platensis* to ammonia at high pH values. *Plant and Cell Physiology*, 32: 953–958.
- Bernard, O. & Rémond, B. (2012). Validation of a simple model accounting for light and temperature effect on microalgal growth. *Bioresource Technology*, 123: 520–527.
- Bitá, C.E. & Gerats, T. (2013). Plant tolerance to high temperature in a changing environment: Scientific fundamentals and production of heat stress-tolerant crops. *Frontiers in Plant Science*, 4: 273.
- Blanchard, M., Pechenik, J.A., Giudicelli, E., Connan, J.P. & Robert, R. (2008). Competition for food in the larvae of two marine molluscs, *Crepidula fornicata* and *Crassostrea gigas*. *Aquatic Living Resources*. 21: 197–205.
- Blankenship, R.E., Tiede, D.M., Barber, J., Brudvig, G.W., Fleming, G., Ghirardi, M., Gunner, M.R., Junge, W., Kramer, D.M., Melis, A., Moore, T.A., Moser, C.C., Nocera, D.G., Nozik, A.J., Ort, D.R., Parson, W.W., Prince, R.C. & Sayre, R.T. (2011). Comparing photosynthetic and photovoltaic efficiencies and recognizing the potential for improvement. *Science*, 332(6031):805-809.
- Bligh, E.G. and Dyer, W.J. (1959). A rapid method of total lipid extraction and purification. *Canadian Journal Biochemistry and Physiology*. 37: 911–917.
- Bonente, G., Pippa, S., Castellano, S., Bassi, R. & Ballottari, M. (2012). Acclimation of *Chlamydomonas reinhardtii* to different growth irradiances. *The Journal of Biological Chemistry*, 287(8): 5833–5847.
- Borowitzka, M.A. (2018). The ‘stress’ concept in microalgal biology-homeostasis, acclimation and adaptation. *Journal of Applied Phycology*, 30: 2815-2825.
- Borowitzka, M.A. (2016). *Algal physiology and large-scale outdoor cultures of microalgae: The physiology of microalgae*. Springer International Publishing, pp: 601-652
- Borowitzka, M.A. (2013). High-value products from microalgae - their development and commercialisation. *Journal of Applied Phycology*, 25: 743–756.
- Borowitzka, M.A. & Vonshak, A. (2017). Scaling up microalgal cultures to commercial scale. *European Journal of Phycology*, 52: 407–418.
- Brindhadevi, K., Mathimani, T., Rene, E. R., Shanmugam, S., Chi, N. T. L. & Pugazhendhi, A. (2021). Impact of cultivation conditions on the biomass and lipid in microalgae with an emphasis on biodiesel. *Fuel*, 284: 119058.

- Cai, T., Park, S.Y., Racharaks, R. & Li, Y. (2013). Cultivation of *Nannochloropsis salina* using anaerobic digestion effluent as a nutrient source for biofuel production. *Applied Energy*, 108: 486–492.
- Camargo, J.A., Alonso, A. & Salamanca, A. (2005). Nitrate toxicity to aquatic animals: a review with new data for freshwater invertebrates. *Chemosphere*, 58: 1255-1267.
- Campbell, N.A & Reece, J.B. (2005). *Photosynthesis, in Biology*. Pearson Benjamin Cummings. United States, pp. 181 – 198.
- Carballo-Cárdenas, E.C, Tuan, P.M., Janssen, M. & Wijffels, R.H. (2003). Vitamin E (α -tocopherol) production by the marine microalgae *Dunaliella tertiolecta* and *Tetraselmis suecica* in batch cultivation. *Biomolecular Engineering*, 20: 139-147.
- Chen, S.Y., Pan, L.Y., Hong, M. & Lee, A.C. (2012). The effects of temperature on the growth of and ammonia uptake by marine microalgae. *Botanical Studies*, 53: 125–133.
- Chen, W. C., Hsu, Y. C., Chang, J. S., Ho, S. H., Wang, L. F. & Wei, Y. H. (2019). Enhancing production of lutein by a mixotrophic cultivation system using microalga *Scenedesmus obliquus* CWL-1. *Bioresource Technology*, 291: 121891.
- Chia, M.A., Lombardi, A.T., Melão, M.D.G.G. & Parrish, C.C. (2013). Effects of cadmium and nitrogen on lipid composition of *Chlorella vulgaris* (Trebouxiophyceae, Chlorophyta). *European Journal of Phycology*, 48: 1–11.
- Chia, S. R., Ong, H. C., Chew, K. W., Show, P. L., Phang, S. M., Ling, T. C., Nagarajan, D., Lee, D. J. & Chang, J. S. (2018). Sustainable approaches for algae utilisation in bioenergy production. *Renewable Energy*, 129: 838–852.
- Chini Zittelli, G., Rodolfi, L., Biondi, N. & Tredici, M.R. (2006). Productivity and photosynthetic efficiency of outdoor cultures of *Tetraselmis suecica* in annular columns. *Aquaculture*, 261: 932–943.
- Chisti, Y., (2007). Biodiesel from microalgae. *Biotechnology Advances*, 25(3): 294-306
- Chokshi, K., Pancha, I., Trivedi, K., George, B., Maurya, R., Ghosh, A. & Mishra, S. (2015). Biofuel potential of the newly isolated microalgae *Acutodesmus dimorphus* under temperature induced oxidative stress conditions. *Bioresource Technology*, 180: 162–171.
- Collos, Y. & Harrison, P.J. (2014). Acclimation and toxicity of high ammonium concentrations to unicellular algae. *Marine Pollution Bulletin*, 80: 8-23.

- Collos, Y. & Slawyk, G. (2008). ^{13}C and ^{15}N uptake by marine phytoplankton .I. influence of nitrogen source and concentration in laboratory cultures of diatoms. *Journal of Phycology*, 15(2): 186–190.
- Conceição, D., Lopes, R.G., Derner, R.B., Cella, H., do Carmo, A.P.B., Montes D'Oca, M.G., Petersen, R., Passos, M.F., Vargas, J.V.C., Galli-Terasawa, L.V. & Kava, V. (2020). The effect of light intensity on the production and accumulation of pigments and fatty acids in *Phaeodactylum tricorutum*. *Journal of Applied Phycology*, 32: 1017-1025.
- Converti, A., Casazza, A.A., Ortiz, E.Y., Perego, P. & Del Borghi, M. (2009). Effect of temperature and nitrogen concentration on the growth and lipid content of *Nannochloropsis oculata* and *Chlorella vulgaris* for biodiesel production. *Chemical Engineering and Processing: Process Intensification*, 48(6): 1146-1151.
- Converti, A., Scapazzoni, A.S., Lodi, A.A. & Carvalho, J.C.M. (2006). Ammonium and urea removal by *Spirulina platensis*, *Journal of Industrial Microbiology and Biotechnology*, 33(1): 8-16.
- Couso, I., Vila, M., Vigarà, J., Cordero, B.F., Vargas, M.Á., Rodríguez, H. & León, R. (2012). Synthesis of carotenoids and regulation of the carotenoid biosynthesis pathway in response to high light stress in the unicellular microalga *Chlamydomonas reinhardtii*. *European Journal of Phycology*. 47(3): 223–232.
- Cullen, J.J. & Davis, R.F. (2003). The blank can make a big difference in oceanographic measurements. *Limnology and Oceanography Bulletin*, 12(2): 29–35.
- Dai, G., Deblois, C., Liu, S., Juneau, P. & Qiu, B. (2008). Differential sensitivity of five cyanobacterial strains to ammonium toxicity and its inhibitory mechanism on the photosynthesis of rice-field cyanobacterium Ge-Xian-Mi (*Nostoc*). *Aquatic Toxicology*, 89(2): 113–121.
- Das, P., Thaher, M., AbdulQuadir, M., Khan, S., Chaudhary, A. & Al-Jabri, H. (2019). Long-term semi-continuous cultivation of a halo-tolerant *Tetraselmis* sp. using recycled growth media. *Bioresource Technology*. 276: 35-41.
- Das, P., Thaher, M.I., Hakim, M.A.Q.M.A., Al-Jabri, H.M.S.J. & Alghasal, G.S.H.S., (2016). A comparative study of the growth of *Tetraselmis* sp. in large scale fixed depth and decreasing depth raceway ponds. *Bioresource Technology*, 216: 114-120.
- Davison, I.R. (1991). Environmental effects on algal photosynthesis: Temperature. *Journal of Phycology*, 27(1): 2-8.

- de-Bashan, L.E., Trejo, A., Huss, V.A.R., Hernandez, J.P. & Bashan, Y. (2008). *Chlorella sorokiniana* UTEX 2805, a heat and intense, sunlight-tolerant microalga with potential for removing ammonium from wastewater. *Bioresource Technology*, 99(1): 4980-4989.
- de la Noüe, J., Laliberté, G. & Proulx, D. (1992). Algae and waste water. *Journal of Applied Phycology*, 4: 247-254.
- Di Lena, G., Casini, I., Lucarini, M. & Lombardi-Boccia, G. (2019). Carotenoid profiling of five microalgae species from large-scale production. *Food Research International*. 120: 810-818.
- Ding, G.T., Mohd Yasin, N.H., Takriff, M.S., Kamarudin, K.F., Salihon, J., Yaakob, Z. & Mohd Hakimi, N.I.N. (2020). Phycoremediation of palm oil mill effluent (POME) and CO₂ fixation by locally isolated microalgae: *Chlorella sorokiniana* UKM2, *Coelastrella* sp. UKM4 and *Chlorella pyrenoidosa* UKM7. *Journal of Water Process Engineering*, 35: 101202.
- Dittmann, K.K., Rasmussen, B.B., Melchiorsen, J., Sonnenschein, E.C., Gram, L. & Bentzon-Tilia, M. (2020). Changes in the microbiome of mariculture feed organisms after treatment with a potentially probiotic strain of *Phaeobacter inhibens*. *Applied and Environmental Microbiology*, 86: 1-36.
- Dongsansuk, A., Lütz, C. & Neuner, G. (2013). Effects of temperature and irradiance on quantum yield of PSII photochemistry and xanthophyll cycle in a tropical and a temperate species. *Photosynthetica*, 51: 13-21.
- Dortch, Q. (1990). The interaction between ammonium and nitrate uptake in phytoplankton. *Marine Ecology Progress Series*, 61: 183–201.
- Drath, M., Kloft, N., Batschauer, A., Marin, K., Novak, J. & Forchhammer, K. (2008). Ammonia triggers photodamage of photosystem II in the cyanobacterium *Synechocystis* sp. strain PCC 6803. *Plant Physiology*, 147: 206-215.
- Eismann, A.I., Perpetuo Reis, R., Ferreira da Silva, A. & Negrão Cavalcanti, D. (2020). *Ulva* spp. carotenoids: Responses to environmental conditions. *Algal Research*, 48: 101916.
- El Gamal, A. A. (2010). Biological importance of marine algae. *Saudi Pharmaceutical Journal*, 18(1): 1–25.
- Fabregas, J., Abalde, J., Herrero, C., Cabezas, B. & Veiga, M. (1984). Growth of the marine microalga *Tetraselmis suecica* in batch cultures with different salinities and nutrient concentrations. *Aquaculture*, 42: 207–215.

- Fábregas, J., Otero, A., Domínguez, A. & Patiño, M. (2001). Growth rate of the microalga *Tetraselmis suecica* changes the biochemical composition of *Artemia* species. *Marine Biotechnology*, 3: 256–263.
- Falkowski, P.G. & Raven, J.A. (2007). *Aquatic Photosynthesis*. In: STU-Stud. ed., Princeton University Press. pp. 21-132.
- Fan, H., Wang, K., Wang, C., Yu, F., He, X., Ma, J. & Li, X. (2020). A comparative study on growth characters and nutrients removal from wastewater by two microalgae under optimized light regimes. *Environmental Technology and Innovation*, 19: 100849.
- Farahin, A. W., Natrah, I., Nagao, N., Yusoff, F. M., Shariff, M., Banerjee, S., Katayama, T., Nakakuni, M., Koyama, M., Nakasaki, K. & Toda, T. (2021). Tolerance of *Tetraselmis tetrathele* to high ammonium nitrogen and its effect on growth rate, carotenoid, and fatty acids productivity. *Frontiers in Bioengineering and Biotechnology*, 9: 568776.
- Farahin, A.W., Yusoff, F.M., Basri, M., Nagao, N. & Shariff, M. (2019). Use of microalgae: *Tetraselmis tetrathele* extract in formulation of nanoemulsions for cosmeceutical application. *Journal of Applied Phycology*, 31(3): 1743–1752.
- Farahin, A. W., Yusoff, F. M., Nagao, N., Basri, M. & Shariff, M. (2016). Phenolic content and antioxidant activity of *Tetraselmis tetrathele* (West) Butcher 1959 cultured in annular photobioreactor. *Journal of Environmental Biology*, 37(4): 631–639.
- Farhadian, O., Yusoff, F.M. & Mohamed, S. (2008). Nutritional values of *Apocyclops dengizicus* (Copepoda: Cyclopoida) fed *Chaetoceros calcitrans* and *Tetraselmis tetrathele*. *Aquaculture Research*, 40(1): 74–82.
- Feng, P., Deng, Z., Hu, Z. & Fan, L. (2011). Lipid accumulation and growth of *Chlorella zofingiensis* in flat plate photobioreactors outdoors. *Bioresource Technology*, 102: 10577–10584.
- Feng, P., Xu, Z., Qin, L., Asraful Alam, M., Wang, Z. & Zhu, S. (2020). Effects of different nitrogen sources and light paths of flat plate photobioreactors on the growth and lipid accumulation of *Chlorella* sp. GN1 outdoors. *Bioresource Technology*, 301: 122762.
- Fu, W., Guomundsson, Ó., Paglia, G., Herjólfsson, G., Andrésón, Ó.S., Pálsson, B.O. & Brynjólfsson, S. (2013). Enhancement of carotenoid biosynthesis in the green microalga *Dunaliella salina* with light-emitting diodes and adaptive laboratory evolution. *Applied Microbiology and Biotechnology*, 97: 2395-2403.

- Gambelli, D., Alberti, F., Solfanelli, F., Vairo, D. & Zanoli, R. (2017). Third generation algae biofuels in Italy by 2030: A scenario analysis using Bayesian networks. *Energy Policy*, 103: 165–178.
- Geel, C., Versluis, W. & Snel, J.F.H. (1997). Estimation of oxygen evolution by marine phytoplankton from measurement of the efficiency of Photosystem II electron flow. *Photosynthesis Research*, 51: 61–70.
- Geider, R.J. (1987). Light and temperature dependence of the carbon to chlorophyll a ratio in microalgae and cyanobacteria: Implications for physiology and growth of phytoplankton. *The New Phytologist*, 106(1): 1-34.
- George, B., Pancha, I., Desai, C., Chokshi, K., Paliwal, C., Ghosh, T. & Mishra, S. (2014). Effects of different media composition, light intensity and photoperiod on morphology and physiology of freshwater microalgae *Ankistrodesmus falcatus* – A potential strain for biofuel production. *Bioresource Technology*. 171: 367–374.
- Gilbert, M., Domin, A., Becker, A. & Wilhelm, C. (2000). Estimation of primary productivity by chlorophyll a in vivo fluorescence in freshwater phytoplankton. *Photosynthetica*, 38: 111-126.
- Go, S., Lee, S.J., Jeong, G.T. & Kim, S.K. (2012). Factors affecting the growth and the oil accumulation of marine microalgae, *Tetraselmis suecica*. In: *Bioprocess and Biosystems Engineering*. Springer. pp. 145-150.
- Gonçalves, A.L., Pires, J.C.M. & Simões, M. (2016). The effects of light and temperature on microalgal growth and nutrient removal: An experimental and mathematical approach. *RSC Advances*. 6(27): 22896–22907.
- Gong, M. & Bassi, A. (2016). Carotenoids from microalgae: A review of recent developments. *Biotechnology Advances*, 34: 1396–1412.
- González-Fernández, C., Sialve, B., Bernet, N. & Steyer, J.P. (2012). Impact of microalgae characteristics on their conversion to biofuel. Part II: Focus on biomethane production. *Biofuels, Bioproducts and Biorefining*, 6(2): 205-218.
- Gorai, T., Katayama, T., Obata, M., Murata, A. & Taguchi, S. (2014). Low blue light enhances growth rate, light absorption, and photosynthetic characteristics of four marine phytoplankton species. *Journal of Experimental Marine Biology and Ecology*, 459: 87-95.
- Gordillo, F.J.L., Jiménez, C., Chavarría, J. & Xavier Niell, F. (2001). Photosynthetic acclimation to photon irradiance and its relation to chlorophyll fluorescence and carbon assimilation in the halotolerant green alga *Dunaliella viridis*. *Photosynthesis Research*, 68: 225-235.

- Goto, M., Nagao, N., Yusoff, F.M., Kamarudin, M.S., Katayama, T., Kurosawa, N., Koyama, M., Nakasaki, K. & Toda, T. (2018). High ammonia tolerance on growth rate of marine microalga *Chlorella vulgaris*. *Journal of Environmental Biology*, 39: 843-848.
- Grobbelaar, J.U. (2007). Algal Nutrition - Mineral Nutrition. In: Handbook of Microalgal Culture. Blackwell Publishing Ltd, Oxford, UK. pp. 95–115.
- Grotkjær, T., Bentzon-Tilia, M., D'Alvise, P., Dierckens, K., Bossier, P. & Gram, L. (2016). *Phaeobacter inhibens* as probiotic bacteria in non-axenic *Artemia* and algae cultures. *Aquaculture*, 462: 64–69.
- Grudzinski, W., Krzeminska, I., Luchowski, R., Nosalewicz, A. & Gruszecki, W.I. (2016). Strong-light-induced yellowing of green microalgae *Chlorella*: A study on molecular mechanisms of the acclimation response. *Algal Research*. 16: 245-254.
- Guedes, A.C., Meireles, L.A., Amaro, H.M. & Malcata, F.X. (2010). Changes in lipid class and fatty acid composition of cultures of *Pavlova lutheri*, in response to light intensity. *Journal of American Oil Chemist' Society*, 87: 791-801.
- Guillard, R. R. (1975). *Culture of phytoplankton for feeding marine invertebrates*. In: Culture of Marine Invertebrate Animals, Plenum Press, New York. pp. 26–60.
- Guillard, R. R. & Ryther, J. H. (1962). Studies of marine planktonic diatoms. *I. cyclotella nana* Hustedt, and *Detonula confervacea* (cleve) Gran. *Canadian Journal of Microbiology*, 8: 229–239.
- Hargreaves, J., Tucker, C. (2004). Ammonia dynamics in fish ponds. Southern Regional Aquaculture Centre.
- Hartig, P., Wolfstein, K., Lippemeier, S. & Colijn, F. (1998). Photosynthetic activity of natural microphytobenthos populations measured by fluorescence (PAM) and ¹⁴C-tracer methods: a comparison. *Marine Ecology Progress Series*. 166: 53-62.
- He, Q., Yang, H., Wu, L. & Hu, C. (2015). Effect of light intensity on physiological changes, carbon allocation and neutral lipid accumulation in oleaginous microalgae. *Bioresource Technology*, 191: 219-228.
- Heasman, M., Diemar, J., O'connor, W., Sushames, T. & Foulkes, L. (2000). Development of extended shelf-life microalgae concentrate diets harvested by centrifugation for bivalve molluscs - a summary. *Aquaculture Research*, 31(8): 637-659.
- Hemaiswarya, S., Raja, R., Kumar, R.R., Ganesan, V., Anbazhagan, C., (2011). Microalgae: A sustainable feed source for aquaculture. *World Journal Microbiology and Biotechnology*, 27: 1737-1746.

- Hu, J., Nagarajan, D., Zhang, Q., Chang, J.S. & Lee, D.J. (2018). Heterotrophic cultivation of microalgae for pigment production: A review. *Biotechnology Advances*, 36, 54-67.
- Huang, J.J., Lin, S., Xu, W. & Cheung, P.C.K. (2017). Occurrence and biosynthesis of carotenoids in phytoplankton. *Biotechnology Advances*, 35(5): 597-618.
- Huerlimann, R., de Nys, R. & Heimann, K. (2010). Growth, lipid content, productivity, and fatty acid composition of tropical microalgae for scale-up production. *Biotechnology and Bioengineering*, 107(2): 245–257.
- Hurtado, M.C., Pilar, M., Moreno, C., Daschner, Á., Riba, R.E., María, R., Pons, G., Fandos, E.G., Arnau, S.G., Gallego, J., Mañes Vinuesa, J., Belloso, O.M., Aránzazu, M., Caballero, M., Martínez Hernández, A., Palop Gómez, A., Lázaro, D.R., Berruezo, G.R., Rubio Armendáriz, C., José, M., Leal, R., Oliag, T., Ángel, J., Buelga, S., Antoni, J., Marí, T., Pascual, V.C., Talens, P., Josep, O. & Marí, A.T. (2017). Section of Food Safety and Nutrition Report approved by the Section of Food Safety and Nutrition of the Scientific Committee in its plenary session.
- Imaizumi, Y., Nagao, N., Yusoff, F.M., Kurosawa, N., Kawasaki, N. & Toda, T. (2016). Lumostatic operation controlled by the optimum light intensity per dry weight for the effective production of *Chlorella zofingiensis* in the high cell density continuous culture. *Algal Research*, 20, 110-117.
- Imaizumi, Y., Nagao, N., Yusoff, F.M., Taguchi, S. & Toda, T. (2014). Estimation of optimum specific light intensity per cell on a high-cell-density continuous culture of *Chlorella zofingiensis* not limited by nutrients or CO₂. *Bioresource Technology*, 162: 53-59.
- Japar, A. S., Takriff, M. S. & Mohd Yasin, N. H. (2021). Microalgae acclimatization in industrial wastewater and its effect on growth and primary metabolite composition. *Algal Research*, 53: 102163.
- Jiang, L., Pei, H., Hu, W., Hou, Q., Han, F. & Nie, C. (2016). Biomass production and nutrient assimilation by a novel microalga, *Monoraphidium* spp. SDEC-17, cultivated in a high-ammonia wastewater. *Energy Conversion and Management*, 123: 423-430.
- Juario, J. V. & Storch, V. (1984). Biological evaluation of phytoplankton (*Chlorella* sp., *Tetraselmis* sp. and *Isochrysis galbana*) as food for milkfish (*Chanos chanos*) fry. *Aquaculture*, 40: 193-198.
- Kaewpintong, K., Shotipruk, A., Powtongsook, S. & Pavasant, P. (2007). Photoautotrophic high-density cultivation of vegetative cells of *Haematococcus pluvialis* in airlift bioreactor. *Bioresource Technology*, 98: 288-295.

- Katayama, T., Nagao, N., Goto, M., Md. Yusoff, F., Sato, M., Takahashi, K. & Furuya, K. (2018). Growth characteristics of shade-acclimated marine *Chlorella vulgaris* under high-cell-density conditions, *Journal of Environmental Biology*, 39: 747-753.
- Katayama, T., Nagao, N., Kasan, N.A., Khatoon, H., Rahman, N.A., Takahashi, K., Furuya, K., Yamada, Y., Wahid, M.E.A. & Jusoh, M. (2020). Bioprospecting of indigenous marine microalgae with ammonium tolerance from aquaculture ponds for microalgae cultivation with ammonium-rich wastewaters. *Journal of Biotechnology*, 323: 113-120.
- Kermanshahi-pour, A., Sommer, T.J., Anastas, P.T. & Zimmerman, J.B. (2014). Enzymatic and acid hydrolysis of *Tetraselmis suecica* for polysaccharide characterization. *Bioresource Technology*, 173: 415-421.
- Khoo, K.H., Culberson, C.H. & Bates, R.G. (1977). Thermodynamics of the dissociation of ammonium ion in seawater from 5 to 40°C. *Journal of Solution Chemistry*, 6: 281-290.
- Kim, G., Bae, J. & Lee, K. (2016). Nitrate repletion strategy for enhancing lipid production from marine microalga *Tetraselmis* sp. *Bioresource Technology*, 205: 274-279.
- Krishna Reddy, Y.V., Adamala, S., Levlin, E. K. & Reddy, K. S. (2017). Enhancing nitrogen removal efficiency of domestic wastewater through increased total efficiency in sewage treatment (ITEST) pilot plant in cold climatic regions of Baltic Sea. *International Journal of Sustainable Built Environment*, 6(2): 351–358.
- Krzemińska, I., Piasecka, A., Nosalewicz, A., Simionato, D. & Wawrzykowski, J. (2015). Alterations of the lipid content and fatty acid profile of *Chlorella protothecoides* under different light intensities. *Bioresource Technology*, 196: 72-77.
- Kuanui, P., Chavanich, S., Viyakarn, V., Omori, M., Fujita, T. & Lin, C. (2020). Effect of light intensity on survival and photosynthetic efficiency of cultured corals of different ages. *Estuarine, Coastal and Shelf Science*, 235: 106515.
- Latowski, D., Kuczyńska, P. & Strzałka, K. (2011). Xanthophyll cycle – a mechanism protecting plants against oxidative stress. *Redox Report*, 16(2): 78-90.
- Lee, S.Y., Kim, S.H., Hyun, S.H., Suh, H.W., Hong, S.J., Cho, B.K., Lee, C.G., Lee, H. & Choi, H.K. (2014). Fatty acids and global metabolites profiling of *Dunaliella tertiolecta* by shifting culture conditions to nitrate deficiency and high light at different growth phases. *Process Biochemistry*, 49(6): 996-1004.

- Lehman, P. W. (1981). Comparison of chlorophyll a and carotenoid pigments as predictors of phytoplankton biomass. *Marine Biology*, 65(3): 237–244.
- Leong, W.H., Lim, J.W., Lam, M.K., Uemura, Y., Ho, C.D. & Ho, Y.C. (2018). Co-cultivation of activated sludge and microalgae for the simultaneous enhancements of nitrogen-rich wastewater bioremediation and lipid production. *Journal of the Taiwan Institute of Chemical Engineers*, 87: 216-224.
- Levasseur, W., Perré, P. & Pozzobon, V. (2020). A review of high value-added molecules production by microalgae in light of the classification. *Biotechnology Advances*, 41: 107545.
- Li, W.K.W. (1980). *Temperature Adaptation in Phytoplankton: Cellular and Photosynthetic Characteristics*. In: Primary Productivity in the Sea. Springer US. pp. 259–279.
- Lim, K.C., Yusoff, F.M., Shariff, M. & Kamarudin, M.S. (2017). Astaxanthin as feed supplement in aquatic animals. *Reviews in Aquaculture*, 10(3): 738-773.
- Liu, S., Daigger, G.T., Kang, J. & Zhang, G. (2019). Effects of light intensity and photoperiod on pigments production and corresponding key gene expression of *Rhodospseudomonas palustris* in a photobioreactor system. *Bioresource Technology*, 294: 122172.
- Liu, J., Sun, Z. & Gerken, H. (2016). *Recent Advances in Microalgal Biotechnology*. OMICS Group eBooks, pp: 11–28.
- Lu, L., Wang, J., Yang, G., Zhu, B. & Pan, K. (2017). Heterotrophic growth and nutrient productivities of *Tetraselmis chuii* using glucose as a carbon source under different C/N ratios. *Journal of Applied Phycology*, 29: 15-21.
- Ma, R., Zhao, X., Xie, Y., Ho, S.-H. & Chen, J. (2019). Enhancing lutein productivity of *Chlamydomonas* sp. via high-intensity light exposure with corresponding carotenogenic genes expression profiles. *Bioresource Technology*, 275: 416-420.
- Ma, X., Chen, K.W. & Lee, Y.K. (1997). Growth of *Chlorella* outdoors in a changing light environment. *Journal of Applied Phycology*, 9: 425-430.
- MacIntyre, H.L., Kana, T.M., Anning, T. & Geider, R.J. (2002). Photoacclimation of photosynthesis irradiance response curves and photosynthetic pigments in microalgae and cyanobacteria. *Journal of Phycology*, 38(1): 17-38.
- Mackey, K.R.M., Paytan, A., Caldeira, K., Grossman, A.R., Moran, D., Mcilvin, M. & Saito, M.A. (2013). Effect of temperature on photosynthesis and growth in marine *Synechococcus* spp. *Plant Physiology*, 163: 815-829.

- Magnotti, C., Lopes, R., Derner, R. & Vinatea, L. (2016). Using residual water from a marine shrimp farming BFT system. Part II: *Artemia franciscana* biomass production fed microalgae grown in reused BFT water. *Aquaculture Research*, 47: 2716-2722.
- Mantecón, L., Moyano, R., Cameán, A. M. & Jos, A. (2019). Safety assessment of a lyophilized biomass of *Tetraselmis chuii* (TetraSOD®) in a 90 day feeding study. *Food and Chemical Toxicology*, 133: 110810.
- Markou, G., Vandamme, D. & Muylaert, K. (2014). Ammonia inhibition on *Arthrospira platensis* in relation to the initial biomass density and pH. *Bioresource Technology*, 166: 259-265.
- Masojídek, J., Torzillo, G., Kopecký, J., Koblížek, M., Nidiaci, L., Komenda, J., Lukavská, A. & Sacchi, A. (2000). Changes in chlorophyll fluorescence quenching and pigment composition in the green alga *Chlorococcum* sp. grown under nitrogen deficiency and salinity stress. *Journal of Applied Phycology*, 12: 417-426.
- Mata, T.M., Martins, A.A. & Caetano, N.S. (2010). Microalgae for biodiesel production and other applications: A review. *Renewable and Sustainable Energy Reviews*, 14(1): 217-232.
- Mathur, S., Agrawal, D. & Jajoo, A. (2014). Photosynthesis: Response to high temperature stress. *Journal of Photochemistry and Photobiology B: Biology*, 137: 116-126.
- Maxwell, K. and Johnson, G.N. (2000). Chlorophyll fluorescence-a practical guide. *Journal of Experimental Botany*, 51: 659-668.
- Mejri, S. C., Tremblay, R., Audet, C., Wills, P. S. & Riche, M. (2021). Essential fatty acid requirements in tropical and cold-water marine fish larvae and juveniles. *Frontiers in Marine Science*, 557.
- Merican, Z. O. & Shim, K. F. (1996). Qualitative requirements of essential fatty acids for juvenile *Penaeus monodon*. *Aquaculture*, 147: 275–291.
- Michels, M.H.A., Camacho-Rodríguez, J., Vermuë, M.H. & Wijffels, R.H. (2014a). Effect of cooling in the night on the productivity and biochemical composition of *Tetraselmis suecica*. *Algal Research*, 6: 145–151.
- Michels, M.H.A., Slegers, P.M., Vermuë, M.H. & Wijffels, R.H., (2014b). Effect of biomass concentration on the productivity of *Tetraselmis suecica* in a pilot-scale tubular photobioreactor using natural sunlight. *Algal Research*, 4: 12–18.
- Minhas, A.K., Hodgson, P., Barrow, C.J. & Adholeya, A. (2016). A review on the assessment of stress conditions for simultaneous production of microalgal lipids and carotenoids. *Frontiers in Microbiology*, 7: 546.

- Mohamed Ramli, N., Giatsis, C., Yusoff, F.M, Verreth, J. & Verdegem, M. (2018). Resistance and resilience of small-scale recirculating aquaculture systems (RAS) with or without algae to pH perturbation. *PLoS One*, 13, e0195862.
- Murata, N., Takahashi, S., Nishiyama, Y. & Allakhverdiev, S.I. (2007). Photoinhibition of photosystem II under environmental stress. *Biochimica et Biophysica Acta- Bioenergetics*, 1767(6): 414-421.
- Nagarajan, D., Lee, D.J., Chen, C.Y. & Chang, J.S. (2020). Resource recovery from wastewaters using microalgae-based approaches: A circular bioeconomy perspective. *Bioresource Technology*, 302: 122817.
- Nagappan, S., Das, P., AbdulQuadir, M., Thaher, M., Khan, S., Mahata, C., Al-Jabri, H., Vatland, A. K. & Kumar, G. (2021). Potential of microalgae as a sustainable feed ingredient for aquaculture. *Journal of Biotechnology*, 341: 1–20.
- National Oceanic and Atmospheric Administration (NOAA). (2020). *Global Climate Report - July 2020*.
- Nayak, M., Suh, W.I., Chang, Y.K. & Lee, B. (2019). Exploration of two-stage cultivation strategies using nitrogen starvation to maximize the lipid productivity in *Chlorella* sp. HS2. *Bioresource Technology*, 276: 110-118.
- Nisar, N., Li, L., Lu, S., Khin, N. C. & Pogson, B. J. (2015). Carotenoid metabolism in plants. *Molecular Plant*, 8(1): 68–82.
- Nixon, P.J., Michoux, F., Yu, J., Boehm, M. & Komenda, J. (2010). Recent advances in understanding the assembly and repair of photosystem II. *Annals of Botany*, 106(1): 1-16.
- Nobel, P.S. & Nobel, P.S. (2009). *Photochemistry of Photosynthesis*. In: *Physicochemical and Environmental Plant Physiology*. Academic Press. pp. 228-275.
- Obata, M., Toda, T. & Taguchi, S. (2009). Using chlorophyll fluorescence to monitor yields of microalgal production. *Journal of Applied Phycology*, 21: 315–319.
- Orefice, I., Chandrasekaran, R., Smerilli, A., Corato, F., Caruso, T., Casillo, A., Corsaro, M.M., Piaz, F.D., Ruban, A. V. & Brunet, C. (2016). Light-induced changes in the photosynthetic physiology and biochemistry in the diatom *Skeletonema marinoi*. *Algal Research*, 17: 1-13.
- Orset, S. & Young, A.J. (1999). Low temperature-induced synthesis of alpha-carotene in the microalga *Dunaliella salina* (Chlorophyta). *Journal of Phycology*, 35(3): 520-527.

- Otero, A. & Fábregas, J. (1997). Changes in the nutrient composition of *Tetraselmis suecica* cultured semicontinuously with different nutrient concentrations and renewal rates. *Aquaculture*, 159: 111-123.
- Ova Ozcan, D. & Ovez, B. (2020). Evaluation of the interaction of temperature and light intensity on the growth of *Phaeodactylum tricornutum*: Kinetic modeling and optimization. *Biochemical Engineering Journal*, 154: 107456.
- Paliwal, C., Mitra, M., Bhayani, K., Bharadwaj, S.V.V., Ghosh, T., Dubey, S. & Mishra, S. (2017). Abiotic stresses as tools for metabolites in microalgae. *Bioresource Technology*, 244: 1216–1226.
- Parkhill, J.P., Maillet, G. & Cullen, J.J. (2001). Fluorescence-based maximal quantum yield for PSII as a diagnostic of nutrient stress. *Journal of Phycology*, 37(4): 517–529.
- Pereira, H., Gangadhar, K.N., Schulze, P.S.C., Santos, T., De Sousa, C.B., Schueler, L.M., Custódio, L., Malcata, F.X., Gouveia, L., Varela, J.C.S. & Barreira, L. (2016). Isolation of a euryhaline microalgal strain, *Tetraselmis* sp. CTP4, as a robust feedstock for biodiesel production. *Science Report*, 6: 1-11.
- Pereira, H., Páramo, J., Silva, J., Marques, A., Barros, A., Maurício, D., Santos, T., Schulze, P., Barros, R., Gouveia, L., Barreira, L. & Varela, J. (2018). Scale-up and large-scale production of *Tetraselmis* sp. CTP4 (Chlorophyta) for CO₂ mitigation: From an agar plate to 100-m³ industrial photobioreactors. *Science Reports*. 8: 5112.
- Perin, G., Bellan, A., Bernardi, A., Bezzo, F. & Morosinotto, T. (2019). The potential of quantitative models to improve microalgae photosynthetic efficiency. *Physiologia Plantarum*. 166(1): 380-391.
- Piorreck, M., Baasch, K. H. & Pohl, P. (1984). Biomass production, total protein, chlorophylls, lipids and fatty acids of freshwater green and blue-green algae under different nitrogen regimes. *Phytochemistry*. 23: 207–216.
- Qiang, H., Guterman, H. & Richmond, A. (1996). Physiological characteristics of *Spirulina platensis* (Cyanobacteria) cultured at high light ultra high cell densities. *Journal of Phycology*, 32: 1066-1073.
- Qu, W., Loke Show, P., Hasunuma, T. & Ho, S. H. (2020). Optimizing real swine wastewater treatment efficiency and carbohydrate productivity of newly microalga *Chlamydomonas* sp. QWY37 used for cell-displayed bioethanol production. *Bioresource Technology*, 305: 123072.
- Quaas, T., Berteotti, S., Ballottari, M., Flieger, K., Bassi, R., Wilhelm, C. & Goss, R. (2015). Non-photochemical quenching and xanthophyll cycle activities in six green algal species suggest mechanistic differences in

the process of excess energy dissipation. *Journal of Plant Physiology*, 172: 92-103.

- Rahman, K.M. (2020). *Food and High Value Products from Microalgae: Market Opportunities and Challenges*. In: *Microalgae Biotechnology for Food, Health and High Value Products*. Springer Singapore. pp. 3–27.
- Ralph, P. J. & Gademann, R. (2005). Rapid light curves: A powerful tool to assess photosynthetic activity. *Aquatic Botany*, 82(3): 222–237.
- Ramanna, L., Guldhe, A., Rawat, I. & Bux, F. (2014). The optimization of biomass and lipid yields of *Chlorella sorokiniana* when using wastewater supplemented with different nitrogen sources. *Bioresource Technology*, 168: 127-135.
- Ramli, N. M., Verdegem, M. C. J., Yusoff, F. M., Zulkifely, M. K. & Verreth, J. A. J. (2017). Removal of ammonium and nitrate in recirculating aquaculture systems by the epiphyte *Stigeoclonium nanum* immobilized in alginate beads. *Aquaculture Environment Interactions*, 9: 213-222.
- Ramos, A., Coesel, S., Marques, A., Rodrigues, M., Baumgartner, A., Noronha, J., Rauter, A., Brenig, B. & Varela, J. (2008). Isolation and characterization of a stress-inducible *Dunaliella salina* Lcy- β gene encoding a functional lycopene β -cyclase. *Applied Microbiology and Biotechnology*, 79, 819-828.
- Ras, M., Steyer, J.P. & Bernard, O. (2013). Temperature effect on microalgae: A crucial factor for outdoor production. *Reviews in Environmental Science and Biotechnology*, 12: 153-164.
- Raven, J.A. & Geider, R.J. (1988). Temperature and algal growth. *New Phytologist*, 110: 441-446.
- Raven, J.A., Wollenweber, B. & Handley, L.L. (1992). A comparison of ammonium and nitrate as nitrogen sources for photolithotrophs. *New Phytologist*. 121(1): 19-32.
- Reddy, K. Y. V., Adamala, S., Levlin, E.K. & Reddy, K.S. (2017). Enhancing nitrogen removal efficiency of domestic wastewater through increased total efficiency in sewage treatment (ITEST) pilot plant in cold climatic regions of Baltic Sea. *International Journal Sustainable Built Environment*, 6: 351-358.
- Renaud, S.M., Thinh, L. Van, Lambrinidis, G. & Parry, D.L. (2002). Effect of temperature on growth, chemical composition and fatty acid composition of tropical Australian microalgae grown in batch cultures. *Aquaculture*, 211: 195-214.
- Roleda, M. Y., Slocombe, S. P., Leakey, R. J. G., Day, J. G., Bell, E. M. & Stanley, M. S. (2013). Effects of temperature and nutrient regimes on

- biomass and lipid production by six oleaginous microalgae in batch culture employing a two-phase cultivation strategy. *Bioresource Technology*, 129: 439–449.
- Rukminasari, N., Omar, S.B.A. & Lukman, M. (2019). Effects of increasing temperature and nitrate concentration on cell abundance, growth rate, biomass and free fatty acid of *Tetraselmis* sp. *IOP Conference Series: Earth Environmental Science*, 370: 012059.
- San Pedro, A., González-López, C. V., Acién, F.G. & Molina-Grima, E. (2013). Marine microalgae selection and culture conditions optimization for biodiesel production. *Bioresource Technology*, 134: 353-361.
- Sansone, C., Galasso, C., Orefice, I., Nuzzo, G., Luongo, E., Cutignano, A., Romano, G., Brunet, C., Fontana, A., Esposito, F. & Ianora, A. (2017). The green microalga *Tetraselmis suecica* reduces oxidative stress and induces repairing mechanisms in human cells. *Scientific Reports*, 7: 1-12.
- Sayegh, F.A.Q. & Montagnes, D.J.S. (2011). Temperature shifts induce intraspecific variation in microalgal production and biochemical composition. *Bioresource Technology*, 102: 3007–3013.
- Schreiber, U., Bilger, W. & Neubauer, C. (1995). Chlorophyll Fluorescence as a Noninvasive Indicator for Rapid Assessment of In Vivo Photosynthesis. In *Ecophysiology of Photosynthesis*. Berlin, Heidelberg: Springer Berlin Heidelberg., pp: 49–70.
- Schreiber, U., Klughammer, C. & Kolbowski, J. (2012). Assessment of wavelength-dependent parameters of photosynthetic electron transport with a new type of multi-color PAM chlorophyll fluorometer. *Photosynthesis Research*, 113: 127–144.
- Schreiber, U., Schliwa, U. & Bilger, W. (1986). Continuous recording of photochemical and non-photochemical chlorophyll fluorescence quenching with a new type of modulation fluorometer. *Photosynthesis Research*. 10: 51-62.
- Schüler, L.M., Santos, T., Pereira, H., Duarte, P., Katkam, N.G., Florindo, C., Schulze, P.S.C., Barreira, L. & Varela, J.C.S. (2020). Improved production of lutein and β -carotene by thermal and light intensity upshifts in the marine microalga *Tetraselmis* sp. CTP4. *Algal Research*, 45: 101732.
- Schulze, P.S.C., Brindley, C., Fernández, J.M., Rautenberger, R., Pereira, H., Wijffels, R.H. & Kiron, V. (2020). Flashing light does not improve photosynthetic performance and growth of green microalgae. *Bioresource Technology Reports*, 9: 100367.

- Schulze, P.S.C., Carvalho, C.F.M., Pereira, H., Gangadhar, K.N., Schüler, L.M., Santos, T.F., Varela, J.C.S. & Barreira, L. (2017). Urban wastewater treatment by *Tetraselmis* sp. CTP4 (Chlorophyta). *Bioresource Technology*, 223: 175-183.
- Seo, S.H., Ha, J.S., Yoo, C., Srivastava, A., Ahn, C.Y., Cho, D.H., La, H.J., Han, M.S. & Oh, H.M. (2017). Light intensity as major factor to maximize biomass and lipid productivity of *Ettlia* sp. in CO₂-controlled photoautotrophic chemostat. *Bioresource Technology*, 244(1): 621-628.
- Serra-Maia, R., Bernard, O., Gonçalves, A., Bensalem, S. & Lopes, F. (2016). Influence of temperature on *Chlorella vulgaris* growth and mortality rates in a photobioreactor. *Algal Research*, 18: 352-359.
- Sforza, E., Simionato, D., Giacometti, G.M., Bertucco, A. & Morosinotto, T. (2012). Adjusted light and dark cycles can optimize photosynthetic efficiency in algae growing in photobioreactors. *PLoS ONE*, 7(6): 1-10.
- Shah, M.M.R., Liang, Y., Cheng, J.J. & Daroch, M. (2016). Astaxanthin-producing green microalga *Haematococcus pluvialis*: From single cell to high value commercial products. *Frontiers in Plant Science*, 7: 531.
- Sheng, J., Kim, H.W., Badalamenti, J.P., Zhou, C., Sridharakrishnan, S., Krajmalnik-Brown, R., Rittmann, B.E. & Vannela, R. (2011). Effects of temperature shifts on growth rate and lipid characteristics of *Synechocystis* sp. PCC6803 in a bench-top photobioreactor. *Bioresource Technology*, 102: 11218-11225.
- Shepherd, C.J. (2013). World of Agriculture for 2012, Aquaculture: are the criticisms justified? Feeding fish to fish. pp: 11-18.
- Shi, T. Q., Wang, L. R., Zhang, Z. X., Sun, X. M. & Huang, H. (2020). Stresses as first-line tools for enhancing lipid and carotenoid production in microalgae. *Frontiers in Bioengineering and Biotechnology*, 8: 610.
- Shi, X.M., Zhang, X.W. & Chen, F. (2000). Heterotrophic production of biomass and lutein by *Chlorella protothecoides* on various nitrogen sources. *Enzyme Microbial Technology*, 27(3), 312–318.
- Sigaud, T.C.S. & Aidar, E. (1993). Salinity and temperature effects on the growth and chlorophyll- α content of some planktonic algae. *Boletim do Instituto Oceanográfico*, 41: 95-103.
- Singh, P., Kumari, S., Guldhe, A., Singh, G. & Bux, F. (2017). ACCase and rbcL gene expression as a function of nutrient and metal stress for enhancing lipid productivity in *Chlorella sorokiniana*. *Energy Conversion and Management*, 148: 809-819.

- Singh, S.P. & Singh, P. (2015). Effect of temperature and light on the growth of algae species: A review. *Renewable, and Sustainable Energy Reviews*, 50: 431-444.
- Solovchenko, A., Solovchenko, O., Khozin-Goldberg, I., Didi-Cohen, S., Pal, D., Cohen, Z. & Boussiba, S. (2013). Probing the effects of high-light stress on pigment and lipid metabolism in nitrogen-starving microalgae by measuring chlorophyll fluorescence transients: Studies with a $\Delta 5$ desaturase mutant of *Parietochloris incisa* (Chlorophyta, Trebouxiophyceae). *Algal Research*, 2(3): 175-182.
- Staehr, P.A. & Birkeland, M.J. (2006). Temperature acclimation of growth, photosynthesis and respiration in two mesophilic phytoplankton species. *Phycologia*, 45: 648-656.
- Stephenson, A.L., Dennis, J.S., Howe, C.J., Scott, S.A. & Smith, A.G. (2010). Influence of nitrogen-limitation regime on the production by *Chlorella vulgaris* of lipids for biodiesel feedstocks. *Biofuels*, 1: 47-58.
- Sun, X.-M., Ren, L.-J., Zhao, Q.-Y., Ji, X.-J. & Huang, H. (2018). Microalgae for the production of lipid and carotenoids: A review with focus on stress regulation and adaptation. *Biotechnology and Biofuels*, 11(1): 1–16.
- Sung, M.G., Lee, B., Kim, C.W., Nam, K. & Chang, Y.K. (2017). Enhancement of lipid productivity by adopting multi-stage continuous cultivation strategy in *Nannochloropsis gaditana*. *Bioresource Technology*, 229: 20-25.
- Sustainable Agriculture and the Environment in the Humid Tropics. (1993). *Sustainable Agriculture and the Environment in the Humid Tropics*. National Academies Press.
- Tam, N.F.Y. & Wong, Y.S. (1996). Effect of ammonia concentrations on growth of *Chlorella vulgaris* and nitrogen removal from media. *Bioresource Technology*, 57: 45-50.
- Tamburic, B., Guruprasad, S., Radford, D.T., Szabó, M., Lilley, R.M., Larkum, A.W.D., Franklin, J.B., Kramer, D.M., Blackburn, S.I., Raven, J.A., Schliep, M. & Ralph, P.J. (2014). The Effect of Diel Temperature and Light Cycles on the Growth of *Nannochloropsis oculata* in a Photobioreactor Matrix. *PLoS ONE*, 9: 1-13.
- Tang, D. Y. Y., Khoo, K. S., Chew, K. W., Tao, Y., Ho, S. H. & Show, P. L. (2020). Potential utilization of bioproducts from microalgae for the quality enhancement of natural products. *Bioresource Technology*, 304: 122997.
- Ting, C.S. & Owens, T.G. (1992). Limitations of the pulse-modulated technique for measuring the fluorescence characteristics of algae. *Plant Physiology*, 100: 367-373.

- Tsai, H.P., Chuang, L. Te & Chen, C.N.N. (2016). Production of long chain omega-3 fatty acids and carotenoids in tropical areas by a new heat-tolerant microalga *Tetraselmis* sp. DS3. *Food Chemistry*, 192: 682-690.
- Valenzuela, J., Carlson, R.P., Gerlach, R., Cooksey, K., Peyton, B.M., Bothner, B. & Fields, M.W. (2013). Nutrient resupplementation arrests bio-oil accumulation in *Phaeodactylum tricornutum*. *Applied Microbiology and Biotechnology*, 97(15): 7049-7059.
- Vaquero, I., Mogedas, B., Ruiz-Domínguez, M. C., Vega, J. M. & Vílchez, C. (2014). Light-mediated lutein enrichment of an acid environment microalga. *Algal Research*, 6: 70–77.
- Varela, J. C., Pereira, H., Vila, M. & León, R. (2015). Production of carotenoids by microalgae: Achievements and challenges. *Photosynthesis Research*, 125(3): 423–436.
- Várkonyi, Z., Masamoto, K., Debreczeny, M., Zsiros, O., Ughy, B., Gombos, Z., Domonkos, I., Farkas, T., Wada, H. & Szalontai, B. (2002). Low-temperature-induced accumulation of xanthophylls and its structural consequences in the photosynthetic membranes of the cyanobacterium *Cylindrospermopsis raciborskii*: An FTIR spectroscopic study. *Proceedings of the National Academy of Sciences of the United States of America*, 99: 2410-2415.
- Veeramani, T. & Santhanam, P. (2015). Effects of temperature and salinity on the growth of microalga *Tetraselmis* sp. and *Tilapia oreochromis* sp. in culture Pond, Tamil Nadu, India. *Journal of Ecology and Environmental Sciences*, 3: 1-9.
- Vonshak, A. & Torzillo, G. (2004). *Environmental stress physiology*. In: Handbook of Microalgal Culture: Biotechnology and Applied Phycology. Blackwell Science, Victoria. pp. 57-82.
- Wang, B. & Jia, J. (2020). Photoprotection mechanisms of *Nannochloropsis oceanica* in response to light stress. *Algal Research*, 46: 101784.
- Wang, C.Y., Fu, C.C. & Liu, Y.C. (2007). Effects of using light-emitting diodes on the cultivation of *Spirulina platensis*. *Biochemical Engineering Journal*, 37(1): 21-25.
- Wang, J., Zhou, W., Chen, H., Zhan, J., He, C. & Wang, Q. (2019). Ammonium nitrogen tolerant *Chlorella* strain screening and its damaging effects on photosynthesis. *Frontiers in Microbiology*, 9: 3250.
- Watanabe, T., Kitajima, C. & Fujita, S. (1983). Nutritional values of live organisms used in Japan for mass propagation of fish: A review. *Aquaculture*, 34(1): 115-143.

- Webb, W.L., Newton, M. & Starr, D. (1974). Carbon dioxide exchange of *Alnus rubra*-A mathematical model. *Oecologia*, 17: 281-291.
- Wei, L., Huang, X. & Huang, Z. (2015). Temperature effects on lipid properties of microalgae *Tetraselmis subcordiformis* and *Nannochloropsis oculata* as biofuel resources. *Chinese Journal of Oceanology and Limnology*, 33: 99-106.
- Weiss, V., Gromet-Elhanan, Z. & Halmann, M. (1985). Batch and continuous culture experiments on nutrient limitations and temperature effects in the marine alga *Tetraselmis suecica*. *Water Research*, 19(2) 185–190.
- World Meteorological Organization (2021). *2020 was one of three warmest years on record - Report on 15 January 2021*
- Xie, J., Chen, S. & Wen, Z. (2021). Effects of light intensity on the production of phycoerythrin and polyunsaturated fatty acid by microalga *Rhodomonas salina*. *Algal Research*, 58: 102397.
- Xie, X., Lu, X., Wang, L., He, L. & Wang, G. (2020). High light intensity increases the concentrations of β -carotene and zeaxanthin in marine red macroalgae. *Algal Research*, 47: 101852.
- Yamori, W., Hikosaka, K. & Way, D.A. (2014). Temperature response of photosynthesis in C₃, C₄, and CAM plants: Temperature acclimation and temperature adaptation. *Photosynthesis Research*, 119: 101-117.
- Yao, C., Ai, J., Cao, X., Xue, S. & Zhang, W. (2012). Enhancing starch production of a marine green microalga *Tetraselmis subcordiformis* through nutrient limitation. *Bioresource Technology*, 118: 438–444.
- Zakar, T., Herman, E., Vajravel, S., Kovacs, L., Knoppová, J., Komenda, J., Domonkos, I., Kis, M., Gombos, Z. & Laczko-Dobos, H. (2017). Lipid and carotenoid cooperation-driven adaptation to light and temperature stress in *Synechocystis* sp. PCC6803. *Biochimica et Biophysica Acta – Bioenergetics*, 1858 (5), 337-350.
- Zhang, X., Endo, M., Sakamoto, T., Fuseya, R., Yoshizaki, G. & Takeuchi, T. (2018). Studies on kuruma shrimp culture in recirculating aquaculture system with artificial ecosystem. *Aquaculture*, 484: 191–196.
- Zhao, X.C., Tan, X.B., Yang, L.B., Liao, J.Y. & Li, X.-Y. (2019). Cultivation of *Chlorella pyrenoidosa* in anaerobic wastewater: The coupled effects of ammonium, temperature and pH conditions on lipids compositions. *Bioresource Technology*, 284: 90-97.
- Zhao, Y., Wang, J., Zhang, H., Yan, C. & Zhang, Y. (2013). Effects of various LED light wavelengths and intensities on microalgae-based simultaneous biogas upgrading and digestate nutrient reduction process. *Bioresource Technology*, 136: 461-468.

Zhu, L., Wang, Z., Takala, J., Hiltunen, E., Qin, L., Xu, Z., Qin, X. & Yuan, Z. (2013). Scale-up potential of cultivating *Chlorella zofingiensis* in piggery wastewater for biodiesel production. *Bioresource Technology*, 137: 318-325.

Zhuang, L.L., Azimi, Y., Yu, D., Wu, Y.H. & Hu, H.Y. (2018). Effects of nitrogen and phosphorus concentrations on the growth of microalgae *Scenedesmus*. LX1 in suspended-solid phase photobioreactors (ssPBR). *Biomass and Bioenergy*, 109: 47-53.

Zhukova, N. V. (2007). Changes in the fatty acid composition of symbiotic dinoflagellates from the hermatypic coral *Echinopora lamellosa* during adaptation to the irradiance level. *Russian Journal of Plant Physiology*, 54, 763-769.

