



**ANTI-PROLIFERATIVE POTENTIAL OF *Eleutherine bulbosa* (Mill.) Urb.
BULB EXTRACTED UNDER OPTIMISED CONDITION ON THREE-
DIMENSIONAL RETINOBLASTOMA CELL CULTURE MODEL (WERI-Rb-1)**

By

'AMMAR AKRAM BIN KAMARUDIN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of
Philosophy**

July 2022

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

A little gift for:

*My beloved parents:
Mr. Kamarudin Napiah & Mrs. Nooryah Hassan*

*My brothers and sisters:
Mrs. Nurhazirah Tasnim, Mr. Ridzuan, Mrs. Durrani Tasnim, Mr. Ammar Asyraf
& Ms. Nurhani Tasnim*



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Doctor of Philosophy

**ANTI-PROLIFERATIVE POTENTIAL OF *Eleutherine bulbosa* (Mill.) Urb.
BULB EXTRACTED UNDER OPTIMISED CONDITION ON THREE-
DIMENSIONAL RETINOBLASTOMA CELL CULTURE MODEL (WERI-Rb-1)**

By

'AMMAR AKRAM BIN KAMARUDIN

July 2022

**Chair : Norhaizan Mohd Esa, PhD
Institute : Bioscience**

According to the Malaysia National Cancer Registry (2007-2011), ocular cancer has been listed among the most prevalent cancers in children after leukaemia, brain and nervous system, lymphoma, as well as bone. A paucity of scientific evidence on the mode of retinoblastoma inhibition remains a great challenge. A folklore medicine locally used among the Dayak community, "Bawang Dayak" or scientifically known as *Eleutherine bulbosa* bulb has been chosen as a potential alternative remedy to inhibit the growth of retinoblastoma. Therefore, the main objective of this study is to scrutinise the new insights on anti-proliferative potential of *E. bulbosa* bulb extract against the mode of retinoblastoma inhibition. The optimisation of the phenolic extraction using response surface methodology was carried out and the results revealed that the temperature of 48 °C; time of 70 min; solid-liquid ratio of 10 g in 146 mL was the best for phenolic extraction. High Performance Liquid Chromatography (HPLC) analysis revealed that eight bioactive compounds were successfully quantified such as gallic acid, chlorogenic acid, rutin, quercetin, epicatechin gallate, eleutheric acid, kaempferol, and myricetin. The results of the cytotoxic study revealed a potent IC₅₀ value of 15.7 ± 2.7 µg/mL compared to cisplatin with 3.6 ± 2.2 µg/mL. The acridine orange/ propidium iodide (AO/PI) dual staining illustrated a significant apoptotic cell death, manifesting apoptotic features such as membrane blebbing, chromatin condensation, and secondary necrosis. In the meantime, Annexin V-FITC portrayed early and late apoptosis as well as cell cycle arrestment in Sub G0/G1 and G0/G1 phases on WERI-Rb-1 cells upon treatment. The apoptosis was further confirmed with qPCR analysis, demonstrating an upregulation of Bax, Bad, p53, Caspase 3, Caspase 8, and Caspase 9. The downregulation of Bcl-2, Bcl-xL, Nrf-2, and HO-1 genes confirmed the apoptotic and antioxidant related pathways were involved in the mode of retinoblastoma inhibition. To further elucidate and compare its anti-proliferative potential, 3D cell culture studies were conducted to investigate the effect of *E. bulbosa* ethanolic bulb extract on apoptotic mechanism and its relation to the antioxidant pathway. The cytotoxic assay was conducted by Resazurin

sodium salt and demonstrated an increased IC₅₀ value of 45.7 ± 1.7 µg/mL and 26.6 ± 6.0 µg/mL for *E. bulbosa* ethanolic bulb extract and cisplatin, respectively. The morphological assessments through 4', 6-diamidino-2-phenylindole (DAPI) and PI double staining as well as scanning electron microscope (SEM) displayed the onset of apoptosis on the 3D retinoblastoma treated cells. The results of gene and protein expressions exhibited that the ratio of pro-survival and pro-apoptotic genes and proteins such as Bcl-2, Bcl-xL, Bax, and Bad were upregulated, suggesting that the extracellular matrix (ECM) hindered the drug penetration resulting in apoptotic resistance. However, the activation of caspase cascades like Caspase 3, 8, and 9 by *E. bulbosa* ethanolic bulb extract confirmed the intrinsic apoptotic mechanism pathway. Of note, the upregulation of antioxidant proteins for instance Nrf-2 and SOD-1 promotes the proliferation of WERI-Rb-1 cells and leads to tumour resistance due the presence of ECM. Surprisingly, the HO-1 protein was downregulated and may be potentially inhibited the growth of retinoblastoma upon treatment by mediating more reactive oxygen species (ROS). Taken together, these findings suggested that the ethanolic bulb extract of *E. bulbosa* may be potential anti-proliferative agent for retinoblastoma cancer as it portrayed selective killing properties in 3D WERI-Rb-1 cells as opposed to cisplatin. Besides, it provides a fundamental understanding on the inhibition of 3D retinoblastoma cancer cells as it mimics more tissue resemblance to in vivo conditions.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**POTENSI ANTI-PROLIFERATIF BEBAWANG *Eleutherine bulbosa* (Mill.)
Urb. DI BAWAH KEADAAN OPTIMUM KE ATAS TIGA-DIMENSI
RETINOBLASTOMA SEL KULTUR MODEL**

Oleh

'AMMAR AKRAM B. KAMARUDIN

Julai 2022

**Pengerusi : Norhaizan Mohd Esa, PhD
Institut : Biosains**

Menurut Pendaftaran Kanser Kebangsaan Malaysia (2007-2011), kanser okular telah disenaraikan antara kanser paling lazim di kalangan kanak-kanak selepas leukemia, otak dan sistem saraf, limfoma, serta tulang. Kekurangan bukti saintifik ke atas cara perencutan retinoblastoma kekal sebagai cabaran besar. Perubatan tradisional yang digunakan secara tempatan dalam kalangan masyarakat Dayak iaitu "Bawang Dayak" atau dikenali secara saintifik sebagai bebwang *Eleutherine bulbosa* telah dipilih sebagai rawatan alternatif yang berpotensi menghalang pertumbuhan retinoblastoma. Oleh itu, objektif utama kajian ini adalah untuk mendalami potensi anti-proliferatif ekstrak bebwang *E. bulbosa* terhadap mod perencutan retinoblastoma. Pengoptimuman ekstrak fenolik menggunakan kaedah permukaan tindak balas (RSM) telah dijalankan dan keputusan menunjukkan bahawa suhu 48°C; masa 70 min; nisbah pepejal-cecair 10 g dalam 146 mL adalah yang terbaik bagi pengekstrakan fenolik. Analisis Kromatografi Cecair Berprestasi Tinggi (HPLC) mendedahkan lapan sebatian bioaktif telah berjaya diidentifikasi seperti asid gallik, asid klorogenik, rutin, kuersetin, epicatechin gallate, eleutherin, kaempferol, dan miricetin. Keputusan potensi sitotoksik yang kuat telah ditunjukkan dengan nilai IC₅₀ sebanyak $15.7 \pm 2.7 \mu\text{g/mL}$ berbanding cisplatin dengan $3.6 \pm 2.2 \mu\text{g/mL}$. Pewarnaan dwi AOPI menggambarkan kematian sel apoptosis yang ketara, menunjukkan ciri apoptosis seperti pengembangan membran, pemeluwapan kromatin dan nekrosis sekunder. Sementara itu, Annexin V-FITC menggambarkan apoptosis awal dan lewat serta penahanan kitaran sel di fasa Sub G0/G1 dan G0/G1 dalam WERI-Rb-1 setelah rawatan. Selanjutnya, apoptosis disahkan melalui analisis qPCR, menunjukkan peningkatan pengawalseliaan Bax, Bad, p53, Caspase 3, Caspase 8, dan Caspase 9. Pengurangan gen Bcl-2, Bcl-xL, Nrf-2, dan HO-1 mengesahkan penglibatan apoptosis dan antioksida. Bagi mendalami dan membandingkan potensi anti-proliferatif dengan lebih lanjut, kajian sel kultur 3D telah dijalankan untuk menyiasat kesan ekstrak etanol bebwang *E. bulbosa* pada mekanisme

apoptosis dan kaitannya dengan antiokksida. Ujian sitotoksik telah dijalankan melalui natrium Resazurin menunjukkan peningkatan nilai IC₅₀ masing-masing sebanyak $45.7 \pm 1.7 \mu\text{g/mL}$ dan $26.6 \pm 6.0 \mu\text{g/mL}$ bagi ekstrak etanol bebewang *E. bulbosa* serta cisplatin. Penilaian morfologi melalui pewarnaan berganda 4', 6-diamidino-2-phenylindole (DAPI) dan PI serta mikroskop pengimbasan elektron (SEM) memaparkan permulaan apoptosis pada sel 3D retinoblastoma yang dirawat. Keputusan gen dan protein menunjukkan nisbah gen dan protein 'pro-survival' dan pro-apoptosis seperti Bcl-2, Bcl-xL, Bax, dan Bad didapati meningkat menunjukkan bahawa ekstrasellular matriks (ESM) menghalang penembusan rawatan dan menyebabkan rintangan apoptosis. Walau bagaimanapun, pengaktifan caspase seperti Caspase 3, 8, dan 9 oleh ekstrak etanol bebewang *E. bulbosa* mengesahkan mekanisme dalaman apoptosis. Selain itu, pengawalseliaan protein antiokksida contohnya Nrf-2 dan SOD-1 meningkat, memberi kesan terhadap rintangan tumor dan menggalakkan proses proliferasi dengan kehadiran ESM. Menariknya, kawal selia protein HO-1 menurun dan mungkin berpotensi menghalang pertumbuhan retinoblastoma setelah rawatan dengan menghasilkan lebih banyak spesies oksigen reaktif (ROS). Secara keseluruhan, ekstrak etanol bebewang *E. bulbosa* boleh dicadangkan sebagai agen anti-proliferatif yang berpotensi bagi kanser retinoblastoma kerana ia menunjukkan sifat membunuh yang terpilih dalam sel 3D WERI-Rb-1 berbanding cisplatin. Kajian ini menyediakan kajian asas pada perencutan sel kanser 3D retinoblastoma kerana ia menggambarkan lebih banyak persamaan tisu terhadap persekitaran *in vivo*.

ACKNOWLEDGEMENTS

“In the name of Allah, the Most Gracious and the Most Merciful”

This thesis is written as a reflection of my bittersweet journey, seeking for His limitless knowledge over the past few years. The odyssey to find knowledge reminded me of those elucidate moments of working relentlessly with my fellow comrades. I owed a great debt of appreciation to countless number of individuals in order to achieve my dream. Alhamdulillah, finally I reached the finishing line.

This thesis would have not been doable without the endless support from my beloved parents and family members. They have endured this long and tiring journey with me since day one. Without their blessings, I would not be able to be where I am now. My appreciation is beyond words.

Then, the utmost gratitude goes to my humble supervisor, Associate Professor Norhaizan Mohd Esa for her kindness, guidance, and unfailing help throughout this entire project. My indebtedness also goes to my co-supervisors, Dr. Norazalina Saad, as well as Dr. Nor Asma Ab. Razak for their continuous assistance and encouragement.

It was such a pleasure to work with Ms. Nor Hafiza Sayuti for her productive assistance, criticism, and valuable discussions throughout this project. Besides that, the inspiration and positivity from my fellow colleagues will always remain in my heart. My sincere gratitude also goes to my helpful staffs of Natural Medicine and Products Research Laboratory, Institute of Bioscience for providing me the complete facilities and making work a pleasant environment.

Lastly, I truly hope that all of these people would forgive me for not mentioning them all by names. Little did they know that, they have a beautiful heart and the reasons I earned this doctorate degree. Thank you!

Sincerely;
Dr. Ammar Akram Kamarudin
Class of 2022

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Norhaizan binti Mohd Esa, PhD

Associate Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Chairman)

Norazalina binti Saad, PhD

Research Officer

Institute of Bioscience

Universiti Putra Malaysia

(Member)

Nor Asma binti Ab. Razak, PhD

Research Officer

Institute of Bioscience

Universiti Putra Malaysia

(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 13 October 2022

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: 'Ammar Akram b. Kamarudin

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee: _____

Associate Prof. Dr. Norhaizan Mohd Esa

Signature: _____

Name of Member
of Supervisory
Committee: _____

Dr. Norazalina Saad

Signature: _____

Name of Member
of Supervisory
Committee: _____

Dr. Nor Asma Ab. Razak

TABLE OF CONTENTS

	Page	
ABSTRACT	i	
ABSTRAK	iii	
ACKNOWLEDGEMENTS	v	
APPROVAL	vi	
DECLARATION	viii	
LIST OF TABLES	xiv	
LIST OF FIGURES	xvi	
LIST OF APPENDICES	xix	
LIST OF ABBREVIATIONS	xxi	
CHAPTER		
1	INTRODUCTION	
1.1	Background	1
1.2	Research problems	3
1.3	Significant of research	4
1.4	Hypothesis	5
1.5	Objectives	5
2	LITERATURE REVIEW	
2.1	Cancer and the risk factors	6
2.2	Hallmarks of cancer	6
2.3	Intraocular tumour	7
2.3.1	Retinoblastoma	8
2.3.1.1	Epidemiology of retinoblastoma in Malaysia	9
2.3.1.2	Clinical presentations of retinoblastoma	11
2.3.1.3	Classification of staging in retinoblastoma	12
2.3.1.4	Treatment options	14
2.4	Apoptosis: A prologue	14
2.4.1	Crosstalk between cancer and apoptosis	14
2.4.2	Morphological and biochemical changes in apoptosis	17
2.4.3	Mechanism of apoptosis	18
2.4.3.1	Intrinsic mitochondria-mediated pathway	18
2.4.3.2	Extrinsic cell surface death receptor pathway	19
2.5	Antioxidant	20
2.5.1	Antioxidant as a double-edged sword in cancer	21
2.5.2	Antioxidant in cancer therapy	23
2.5.3	Future perspectives of chemotherapeutic drugs	24
2.6	Three-dimensional (3D) cell culture model	25
2.6.1	Scaffold-based hydrogel	27

2.6.2	Scaffold requirements for ECM application	27
2.6.3	Collagen	27
2.6.3.1	The structure of collagen	28
2.6.3.2	Sources of collagen	29
2.6.3.3	Extraction methods and sterilisation	29
2.7	Natural product	30
2.8	<i>Eleutherine bulbosa</i> (Mill.) Urb	31
2.8.1	Phytochemistry of <i>Eleutherine bulbosa</i> bulb	31
2.8.2	Pharmacological activity of <i>Eleutherine bulbosa</i> bulb	32
2.9	Design of experiments (DoE): Response surface methodology	32
2.9.1	RSM: Central composite design	33
2.9.2	Evaluation of the fitted model	34
2.9.3	RSM application for optimisation of plant extraction	34
3	THE OPTIMISATION OF THE EXTRACTION CONDITION OF PHENOLIC COMPOUNDS <i>Eleutherine bulbosa</i> BULB USING RESPONSE SURFACE METHODOLOGY AND ITS BIOACTIVE PROFILES	
3.1	Introduction	36
3.2	Materials and methods	37
3.2.1	Chemicals and reagents	37
3.2.2	Raw materials and sample preparation	37
3.2.3	Extraction	37
3.2.4	Experimental design	37
3.2.5	Phytochemical analysis	38
3.2.5.1	Total phenolic content	38
3.2.5.2	Total flavonoid content	38
3.2.6	Antioxidant activities	38
3.2.6.1	DPPH free radical scavenging assay	38
3.2.6.2	ABTS radical cation scavenging assay	39
3.2.7	Validation of the model	39
3.2.8	High performance liquid chromatography (HPLC) analysis	39
3.2.9	Statistical analysis	39
3.3	Results and discussion	40
3.3.1	Fitting the model	40
3.3.2	The effect of experimental parameters on total phenolic content (TPC)	43
3.3.3	The effect of experimental parameters on total flavonoid content (TFC)	46
3.3.4	The effect of experimental parameters on antioxidant activities	46
3.3.5	The optimisation of design parameters and model validation	48
3.3.6	HPLC analysis	49

3.4	Conclusion	52
4	CHEMOTHERAPEUTIC POTENTIAL OF <i>Eleutherine bulbosa</i> ETHANOLIC BULB EXTRACT ON HUMAN RETINOBLASTOMA CANCER CELLS (WERI-Rb-1) UNDER OPTIMISED EXTRACTION CONDITION	
4.1	Introduction	53
4.2	Materials and methods	54
4.2.1	The extraction of <i>E. bulbosa</i> bulb	54
4.2.2	Cell lines and cultures	54
4.2.3	Cytotoxic activity	54
4.2.4	Selectivity index	54
4.2.5	Cell treatments	55
4.2.6	AOPI dual staining	55
4.2.7	Apoptosis assay	55
4.2.8	Cell cycle analysis	55
4.2.9	Quantitative real-time polymerase chain reaction (qPCR)	55
4.2.1	Statistical analysis	56
4.3	Results and discussion	56
4.3.1	The assessment of cytotoxic activity <i>E. bulbosa</i> ethanolic bulb extract on WERI-Rb-1 cells and ARPE-19 cells under optimised extraction condition	59
4.3.2	The cytotoxic activity between optimised and non-optimised extraction condition of <i>E. bulbosa</i> ethanolic bulb extract on WERI-Rb-1 cells	60
4.3.3	The dose-dependent treatments on WERI-Rb-1 cells	60
4.3.4	The morphological assessment of WERI-Rb-1 cells through AOPI staining	60
4.3.5	Apoptosis analysis	63
4.3.6	Cell cycle analysis	65
4.3.7	The effect of the optimised <i>E. bulbosa</i> ethanolic bulb extract on mRNA expression	67
4.4	Conclusion	70
5	CHEMOTHERAPEUTIC POTENTIAL OF <i>Eleutherine bulbosa</i> ETHANOLIC BULB EXTRACT ON THREE-DIMENSIONAL (3D) HUMAN RETINOBLASTOMA CANCER CELLS (WERI-Rb-1) UNDER OPTIMISED EXTRACTION CONDITION	
5.1	Introduction	71
5.2	Materials and methods	72
5.2.1	The extraction of <i>E. bulbosa</i> bulb	72
5.2.2	The preparation of collagen Type I for 3D cell culture	72
5.2.2.1	The extraction of collagen Type I from rat tail tendon	72
5.2.2.2	Sterilisation of collagen Type I	73

5.2.2.3	Neutralisation of sterile collagen Type I	73
5.2.3	3D retinoblastoma cell culture model	73
5.2.4	Cytotoxic assay	73
5.2.5	Cell treatments	74
5.2.6	Morphological assessments of 3D retinoblastoma cancer cells	74
5.2.6.1	DAPI and PI dual staining	74
5.2.6.2	Scanning electron microscopy	74
5.2.7	Gene expression analysis	74
5.2.7.1	Homogenisation and RNA extraction	74
5.2.7.2	RNA precipitation	75
5.2.7.3	RNA wash and solubilisation	75
5.2.7.4	Reverse transcriptase polymerase chain reaction (RT-qPCR)	75
5.2.7.5	Quantitative real-time polymerase chain reaction	75
5.2.8	Protein expression analysis	76
5.2.8.1	Sample preparation and quantification	76
5.2.8.2	Immunoblotting and gel view	76
5.2.9	Statistical analysis	77
5.3	Results and discussion	77
5.3.1	Cytotoxic activity of optimised <i>E. bulbosa</i> ethanolic bulb extract on the developed 3D WERI-Rb-1 cells Imaging and data analysis	77
5.3.2	Cell treatments	80
5.3.3	Surface morphology of 3D WERI-Rb-1 cells	80
5.3.3.1	DAPI and PI dual staining	80
5.3.3.2	Scanning electron microscopy	85
5.3.4	The modulatory effect of the optimised <i>E. bulbosa</i> ethanolic bulb extract on mRNA and protein expressions of 3D WERI-Rb-1 cells	91
5.4	Conclusion	101
6	SUMMARY, GENERAL CONCLUSION, AND FUTURE RECOMMENDATION	
6.1	Summary	102
6.2	General conclusion	103
6.3	Future recommendation	103
BIBLIOGRAPHY		105
APPENDICES		133
BIODATA OF STUDENT		152
LIST OF PUBLICATIONS		153

LIST OF TABLES

Table		Page
2.1	Classification of intraocular retinoblastoma based on IIRC and IRCCB schemes	13
2.2	Comparison between traditional 2D and 3D cell culture model	26
3.1	The central composite design (CCD) for the extraction conditions generated and the corresponding response variables	41
3.2	The regression coefficient (β), coefficient of determination (R^2) and the probability value (p -value) to fit the second order polynomial models for antioxidant study	43
3.3	The optimised extraction conditions for maximum phenolic and flavonoid compounds, as well as antioxidant activities	49
3.4	The predicted versus experimental values of the model responses under optimum extraction conditions	49
3.5	The bioactive compounds detected under optimised extraction conditions	50
4.1	Cytotoxic effects of the tested compounds on retinoblastoma (WERI-Rb-1) and normal cells (ARPE-19)	59
5.1	The cytotoxic effects of the tested compounds on 3D and 2D retinoblastoma (WERI-Rb-1), respectively	78
5.2	The cytotoxic concentrations of the respective tested compounds for the inhibition of 3D retinoblastoma cancer cells	80
5.3	The regulation of mRNA and protein expressions of Bax in 3D WERI-Rb-1 cells upon treatments as compared to the normalised untreated, respectively.	90
5.4	The regulation of mRNA and protein expressions of Bcl-xL in 3D WERI-Rb-1 cells upon treatments as compared to the normalised untreated, respectively.	91
5.5	The mRNA expressions of Bcl-2 and Bad genes in 3D WERI-Rb-1 cells treated with <i>E. bulbosa</i> ethanolic bulb extract and cisplatin as compared to the normalised untreated, respectively.	92

5.6	The regulation of Caspase 3 gene and protein expression in 3D WERI-Rb-1 cells treated with <i>E. bulbosa</i> ethanolic bulb extract and cisplatin as compared to the normalised untreated, respectively.	93
5.7	The regulation of Caspase 9 gene and protein expression in 3D WERI-Rb-1 cells upon treatments as compared to the normalised untreated, respectively.	94
5.8	The regulation of Caspase 8 protein expression in 3D WERI-Rb-1 cells treated with <i>E. bulbosa</i> ethanolic bulb extract and cisplatin as compared to the normalised untreated, respectively.	95
5.9	The regulation of mRNA and protein expression of Nrf-2 in 3D WERI-Rb-1 cells treated with both treatments, respectively.	96
5.10	The regulation of mRNA and protein expression of HO-1 in 3D WERI-Rb-1 cells upon treatments, respectively.	97
5.11	The regulation of mRNA and protein expression of SOD-1 in 3D WERI-Rb-1 cells treated with both treatments, respectively.	98

LIST OF FIGURES

Figure		Page
2.1	The hallmarks of cancer	7
2.2	The percentage of ocular cancer distribution by the anatomical site	8
2.3	The top ten most common cancer among childhood (a)	10
2.4	The common clinical presentations of retinoblastoma	12
2.5	The mechanism of apoptosis in general	16
2.6	The primary changes of morphological characteristics at the onset of apoptosis	17
2.7	The general mechanism of intrinsic and extrinsic pathways in apoptosis	18
2.8	The mechanism of apoptosis through the intrinsic mitochondrial pathway	19
2.9	The mechanism of apoptosis through the extrinsic death receptor pathway	20
2.10	The cumulative effect of ROS in a multistage carcinogenesis process	21
2.11	The antioxidant could act as a double-edged sword in cancer	22
2.12	A comparison of cellular growth in 2D and 3D cell cultures	26
2.13	A classical view of the triple helix structure of the collagen	28
2.14	The morphological feature of <i>E. bulbosa</i> bulb	31
2.15	The chemical structure of eleutherin isolated from <i>E. bulbosa</i> bulb	32
2.16	The design of CCD that comprises of factorial, star, and centre points	34
2.17	The overall procedure of RSM experimental design for the optimisation of plant extraction	35
3.1	The interaction effects between the process parameters and the amount of phenolics produced	45

3.2	The interactive effect between temperature and extraction time (X_{AB}) on the amount of flavonoid obtained	46
3.3	The interactive effect between temperature and solid-liquid ratio on both antioxidant assays, DPPH (a) and ABTS (b), respectively	48
3.4	HPLC profiles of bioactive compounds detected at 280 nm	51
4.1	The cytotoxic activity of the optimised <i>E. bulbosa</i> ethanolic bulb extract (a-b) and cisplatin (c-d) on both retinoblastoma cancer and normal cells, respectively after 72 h	58
4.2	The comparison of the cytotoxic activity between the optimised and non-optimised extraction condition from the bulb of <i>E. bulbosa</i> on WERI-Rb-1 cells	59
4.3	The morphological characteristics of WERI-Rb-1 cells post treatment with <i>E. bulbosa</i> ethanolic bulb extract after 72 h	61
4.4	The morphological characteristics of WERI-Rb-1 cells treated with <i>E. bulbosa</i> ethanolic bulb extract after 72 h	62
4.5	The Annexin V-FITC apoptosis assay of WERI-Rb-1 cells treated with the ethanolic bulb extract of <i>E. bulbosa</i> extract at various concentrations after 72 h	64
4.6	The distribution of WERI-Rb-1 cells treated with the ethanolic bulb extract of <i>E. bulbosa</i> at various concentrations (a)	66
4.7	The mRNA expression of WERI-Rb-1 cells treated with the ethanolic bulb extract of <i>E. bulbosa</i> at 15.7 µg/mL (IC ₅₀)	69
5.1	The cytotoxic activity of the optimised <i>E. bulbosa</i> ethanolic bulb extract (a) and cisplatin (b) on 3D retinoblastoma cancer cells after 72 h, respectively	79
5.2a	The morphological difference between 2D and 3D WERI-Rb-1 cell culture model, respectively (i)	81
5.2b	The morphological assessment of 3D WERI-Rb-1 cells treated with <i>E. bulbosa</i> ethanolic bulb extract at 72 h	82
5.2c	The morphological evaluation of 3D WERI-Rb-1 cells upon treatment with the positive control, cisplatin at 72 h	83
5.3a	The surface morphological observation between 2D and 3D untreated WERI-Rb-1 cells, respectively	85

5.3b	The collagen fibres and the effect of BD ₂₅ treatment on 3D WERI-Rb-1 surface morphology, respectively	86
5.3c	The effect of <i>E. bulbosa</i> ethanolic bulb extract on the surface morphology of 3D WERI-Rb-1 cells	87
5.3d	Varying concentrations of cisplatin-induced apoptotic features on 3D WERI-Rb-1 cells	88
5.3e	The effect of cisplatin (D ₇₅) on the morphological characteristics of 3D WERI-Rb-1 cells	89
5.4	The level of Bax expression on 3D WERI-Rb-1 treated cells.	91
5.5	The level of Bcl-xL protein expression on 3D WERI-Rb-1 treated cells.	92
5.6	The level of Caspase 3 protein expression on 3D WERI-Rb-1 treated cells.	93
5.7	The level of Caspase 9 protein expression on 3D WERI-Rb-1 treated cells.	94
5.8	The level of Caspase 8 protein expression on 3D WERI-Rb-1 treated cells.	95
5.9	The level of Nrf-2 protein expression on 3D WERI-Rb-1 treated cells.	96
5.10	The level of HO-1 protein expression on 3D WERI-Rb-1 treated cells.	97
5.11	The level of SOD-1 protein expression on the 3D WERI-Rb-1 treated cells.	98
6.1	The suggested mechanistic pathway of apoptosis induced by the optimised <i>E. bulbosa</i> ethanolic bulb extract on 3D WERI-Rb-1 cells	102

LIST OF APPENDICES

Appendix		Page
A1	The standard curve for total phenolic (mg GAE) and flavonoid contents (mg RE), respectively	131
A2	The standard curve for total antioxidant activities via DPPH and ABTS radical scavenging assays, respectively	132
B1	The standard curve and chromatographic profile of gallic acid in HPLC analysis, respectively	133
B2	The standard curve and chromatographic profile of epicatechin gallate in HPLC analysis, respectively	134
B3	The standard curve and chromatographic profile of chlorogenic acid in HPLC analysis, respectively	135
B4	The standard curve and chromatographic profile of myricetin in HPLC analysis, respectively	136
B5	The standard curve and chromatographic profile of quercetin in HPLC analysis, respectively	137
B6	The standard curve and chromatographic profile of rutin in HPLC analysis, respectively	138
B7	The standard curve and chromatographic profile of kaempferol in HPLC analysis, respectively	139
B8	The standard curve and chromatographic profile of eleutherin in HPLC analysis, respectively	140
C1	The nucleotide sequences of primers used in qPCR	141
C2	The mRNA expression of apoptotic and antioxidant genes involved on the inhibition of 3D WERI-Rb-1 cells treated with <i>E. bulbosa</i> ethanolic bulb extract and cisplatin.	142
C3	(a) 10X TAE buffer in 1.0 L; (b) 1% Agarose gel	140
D1	The overall process of type I collagen extraction from the rat tail tendon	144
E1	The BCA standard for quantification of protein concentration in immunoblotting	145

- E2 The protein expression of apoptotic and antioxidant genes involved on the inhibition of 3D WERI-Rb-1 cells treated with *E. bulbosa* ethanolic bulb extract and cisplatin. 146
- E3 (a) 10X Tris-Glycine buffer in 2.0 L; (b) 10X Tris-Glycine electrophoresis buffer in 2.0 L; (c) 1X Tris-Glycine transfer buffer in 1.0 L; (d) Resolving buffer in 1.0 L; (e) Stacking buffer in 1.0 L; (f) Polyacrylamide gel; (g) 10X Tris-buffered saline in 1.0 L (h) 1X Tris-buffered saline with Tween 20 (TBST) in 1.0 L; (i) Coomassie blue staining solution in 1.0 L; (j) Destain 1 solution in 1.0 L; (k) Destain 2 solution in 1.0 L 147

LIST OF ABBREVIATIONS

2D	Two-dimensional
3D	Three-dimensional
ABTS	2,2'-azinobis (3-ethylbenzo thiazoline-6-sulphonic acid) disodium salt
AOPI	Acridine orange propidium iodide
ANOVA	Analysis of variance
APAF-1	Apoptotic protease-activating factor-1
ARPE-19	Retinal pigmented epithelium
BCA	Bicinchoninic acid
Bcl-2	B-cell lymphoma 2
BSA	Bovine serum albumin
DAPI	4', 6-diamidino-2-phenylindole
Caspase	cysteine aspartyl-specific proteases
cDNA	Complementary deoxyribonucleic acid
CO ₂	Carbon dioxide
DISC	Death-inducing signalling complex
DMEM	Dulbeccos's modified essential medium
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
DoE	Design of Experiment
DPPH	2,2-diphenyl-1-picrylhydrazyl
ECM	Extracellular matrix
FADD	Fas-associated death domain
FBS	Fetal bovine serum
g	Gram

GAPDH	Glyceraldehyde 3-phosphate dehydrogenase
h	Hour
HPLC	High performance liquid chromatography
HRP	Horseradish peroxidase
HtrA2	Omi/high-temperature requirement protein A
IC	Inhibitory concentration
i.e.	Id est
LOF	Lack of fit
mg	Milligram
min	Minute
mL	Millilitre
mM	Milli Molar
MNCR	Malaysia National Cancer Registry
MTT	(3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide)
NED	National Eye Database
ng	Nanogram
nm	Nanometre
NF- κ B	Nuclear factor kappa-light-chain-enhancer of activated B cells
Nrf-2	Nuclear factor erythroid 2-related factor 2
PBS	Phosphate buffered saline
PCR	Polymerase chain reaction
Ppm	Part per million
PVDF	Polyvinylidene difluoride
qPCR	Quantitative real time polymerase chain reaction
RIPA	Radioimmunoprecipitation
ROCK-1	Rho-associated serine/threonine kinase protein

ROS	Reactive oxygen species
RNA	Ribonucleic acid
RPMI	Roswell Park Memorial Institute
RSM	Response Surface Methodology
RT-qPCR	Reverse transcriptase quantitative polymerase chain reaction
SDS-PAGE	Sodium dodecyl sulphate-polyacrylamide gel
SEM	Scanning electron microscope
SI	Selectivity index
SMAC	Second mitochondria-derived activator of caspase
TBST	Tris buffered saline Tween 20
TNF	Tumour necrosis factor
TRADD	TNF receptor-associated death domain
TRAIL	TNF-related apoptosis-inducing ligand
Trolox	6-hydroxy-2, 5, 7, 8-tetra-methylchroman 2- carboxylic acid
UV	Ultraviolet
µL	Microlitre
WERI-Rb-1	Retinoblastoma
XIAP	X-linked inhibitor of apoptosis protein
%	Percentage

CHAPTER 1

INTRODUCTION

1.1 Background

Retinoblastoma (Rb) is a genetically rare ocular cancer instigated by a bi-allelic mutation of the retinoblastoma gene (RB1), originating from the retinal cells (Dimaras et al., 2015). Paediatrics are more vulnerable than geriatrics, with most cases happening among children with the age of less than 5 years old (Fabian et al., 2018). The annual occurrence of retinoblastoma per country is estimated around 8,000 new cases with an incidence rate between 1 in 16,000 to 18,000 live births (Seregard et al., 2004; Haggerty et al., 2004; Broaddus et al., 2009; Dimaras et al., 2015). The prognosis and the survival rate of Rb patients pivoted on the early diagnosis and treatments received (Ancona-Lezama et al., 2020). In developed countries, the goal of the treatment focuses on the globe salvation and vision preservation (Broaddus et al., 2009; Ancona-Lezama et al., 2020). However, low- and middle-income countries (LMIC) countries have major treatment crises as the majority of the children display late-stages of the advanced tumour at presentation (Kao et al., 2002; Prasad Sah et al., 2013; Gichigo et al., 2015; Goolam et al., 2018). According to Reddy and Anusya (2010), the late presentation of advanced retinoblastoma in Malaysia is due to awareness deprivation among the public. In several cases, parents do not accept the reality that their children were diagnosed with a fatal intraocular tumour and they prefer treatments from the traditional healers, which surprisingly ineffective (Reddy et al., 2010). Besides that, technological and social issues are also perturbing in LMIC regions as compared to developed countries (Chawla et al., 2017). The global burden of Rb encompasses of 43% (3452 of 8099 children) in countries such as India (1486), China (1103), Indonesia (277), Pakistan (260), Bangladesh (184), and the Philippines (142) (Usmanov and Kivelä, 2014). Due to the explosive population growth in the Asia-Pacific region, Dimaras et al. (2012) proposed that Rb has globally triumphed over uveal melanoma as the most prevalent ocular malignancy. With the contemporary trend unveiled, the global burden of Rb is forecasted to escalate by 100 cases annually (Jain et al., 2019).

The management of Rb is still exigent within the remit of ocular oncology. The ultimate goal is to save life, followed by vision salvage (Andersch et al., 2019). Eye preservation is advisable in the case of a localised tumour, which has multiple treatment options such as brachytherapy and local intra-arterial chemotherapy (Andersch et al., 2019). For a massive tumour, systemic chemotherapy would be recommended to reduce the initial tumour size before subsequent treatment options could be prescribed. Like any other cancer, Rb, on the other hand, could disseminate through the optic nerve into the central nervous system and also via sclera, resulting in distant metastatic sites like lymph nodes, bones, and liver (Schaiquevich et al., 2022). In this case, high doses of combinative chemotherapy drugs like vincristine, etoposide, and

carboplatin are less effective as they could impact the quality of life due to the aggressive drug effects (Dunkel et al., 2000; Kremens et al., 2003; Gombos et al., 2007; Maus and June, 2016). Therefore, the urge for alternative and complementary treatment options are warranted.

In search for an alternative treatment, cell-based assays are the essential pillars in drug discovery and development process (Joseph et al., 2018). Mammalian cell cultures provide a solid foundation for deeper exploration of tissue physiology and pathophysiology outside the organisms (Joseph et al., 2018). For over a century, the traditional monolayer 2D cell culture technique has been widely utilised for cellular responses in drug discovery. Up until now, the method is still applicable and has successfully strengthened our understanding on drug mechanisms of action; however, they still have significant drawbacks that require amendments. The primary limitation involves fully grown cells on a culture flask that is made up of stiff platform, rendering them unnatural growth kinetics and cell attachments (Kapałczyńska et al., 2018). As a result, natural microenvironment of the particular cells is not fully represented, which could render misleading data. Most drugs fail during the clinical stage of phases II and III because of the abysmal drug efficacy and safety problems (Arrowsmith and Miller, 2013). It is suggested that the reduced attenuation of drug responses is caused by unfitted preclinical testing i.e. in vitro models that offer insufficient information associated with drug efficacy and safety issues (Kapałczyńska et al., 2018). Hence, new technologies in preclinical testing are rapidly evolving, making improvements for a better in vitro cell culture model that resembles in vivo environment, the three-dimensional (3D) cell culture model (Joseph et al., 2018). The technique is designed to improve the cellular structures and physiology for better mimicry of tissues and organs specific microarchitectures (Ballav et al., 2021).

Cancer remains a perplexing disease for many years, with several strategies conducted to maximise the effectiveness of treatment outcomes (Mitra and Dash, 2018). In view of this, natural products are vigorously explored to fathom the intricate mechanism within cancer. Since immemorial time, natural products have been the mainstay of folklore medicine in treating various ailments. Phytochemicals act directly on the specific molecular targets i.e. genes and proteins, or indirectly by stabilising the conjugates that may affect the metabolic pathways (Saldanha and Tollefsbol, 2012). The shreds of evidence from the past studies displayed that myriad of natural compounds have anti-cancer activities, which could be used as a functional treatment approach through various mechanisms (Li et al., 2013; Zheng et al., 2016; Zhou et al., 2016; Grossi et al., 2017). However, the number of explored natural products for their therapeutic benefits, particularly plants, are still scarce (Ickes et al., 2003; Noor Rain et al., 2007). Notably, polyphenols are the potential candidate for the anti-cancer drug discovery, with a broad range of small molecules to highly polymerised secondary metabolites (Manach et al., 2004). They present abundantly in foods and beverages such as vegetables, fruits, herbs, tea, and wine (Fu et al., 2010; Zhou et al., 2016; Deng et al., 2013).

Thus, extraction plays imperative role in acquiring polyphenols with therapeutic activities. The extraction of polyphenols largely relies on the polarity of the solvent, extraction time, and method as it influences the qualitative and quantitative constitution of the compounds (Rodríguez-Pérez et al., 2015). Organic solvents such as methanol, acetone, hexane, and chloroform are pertinent for extraction as it determines the active component in a particular plant but, significantly toxic for human health (Li et al., 2006). Thus, ethanol is widely chosen for its green solvent quality that could dissolve polar and non-polar compounds efficiently. Besides that, phenolic compounds have different polarities that vary significantly, and the development of a single, optimum extraction is necessary (Garcia-Salas et al., 2010). Conventional optimization is unavailing as it complies one-variable-at-a-time approach, which could potentially miss the interaction effects between parameters and the response of interests (Ibrahim and Elkhidir, 2011). Hence, response surface methodology (RSM), a useful mathematical tool is used to evaluate multiple responses simultaneously, with less labour-intensive management for better extraction reproducibility (Azahar et al., 2017).

Locally known as bawang Dayak or bawang hutan, *Eleutherine bulbosa* Mill. bulb is an herbaceous, perennial flowering plant from the Iridaceae family that is broadly cultivated in Southern America, the African region, and Indonesia (Kusuma et al., 2010; Insanu et al., 2014). In Indonesia, it is widely cultivated across sulphuric land, within 600 to 2000 m above the sea level of Kalimantan Island (Ieyama et al., 2011). The bulb is famous among the local tribe of Kalimantan, the Dayaks, where it is traditionally used to treat diabetes, breast cancer, hypertension, stroke, sexual disorders, as well as to enhance the production of breast-milk (Ieyama et al., 2011). Naphthalene, anthraquinone, and naphthoquinone are the key constituents of *E. bulbosa* bulb which demonstrated various pharmacological responses such as anti-microbial, anti-inflammatory, anti-hypertension, anti-cancer, anti-diabetic, and anti-melanogenesis activity (Kusuma et al., 2010; Insanu et al., 2014). However, studies on the pharmacological activity of *E. bulbosa* bulb are still minimal, primarily on anti-cancer properties.

1.2 Research problems

The prevalence of retinoblastoma cases are arising over the years. It is a genetically rare type of ocular cancer that requires significance attention as their treatments are challenging and involves invasive approaches. Due to the lack of public awareness and parental denial issues, late presentation of advance Rb is a commonplace in Malaysia (Reddy and Anusya, 2010). In view of this, combinative chemotherapies such as etoposide, carboplatin, and vincristine are given to the patients. However, these treatments are less efficient and may aggravate the side effects such as nausea, diarrhoea, fatigue, as well as fever. Invasive procedure such as enucleation is commonly practiced to prevent metastatic secondary cancers however, it does not eliminate tumour recurrence (Yang et al., 2018). Thus, new anti-cancer compound with safe and good efficacy is warranted to treat retinoblastoma.

Eleutherine bulbosa Mill. bulb or locally known as Bawang Dayak is one of the most undervalued plants, traditionally used to treat diabetes, breast cancer, hypertension, stroke, sexual disorders, and production of breast-milk among the Dayak people (Ieyama et al., 2011). Naphthalene, anthraquinone, and naphthoquinone are the key constituents of *E. bulbosa* bulb that are responsible for its anti-microbial, anti-inflammatory, anti-hypertension, anti-cancer, anti-diabetic, and anti-melanogenesis properties (Kusuma et al., 2010; Insanu et al., 2014). However, the studies on the anti-cancer properties are still minimal, which is worth to explore as it exhibits potent anti-cancer activity on breast cancer.

To understand the mechanism of the retinoblastoma inhibition, the development of a cell-based model should be selected appropriately to resembles the real tumour microenvironment. The 2D cell culture model has been used since the early 1900s, however, it promotes uneven growth kinetics on a stiff planar, causing unnatural tumour microenvironment (Cukierman et al., 2001). Using this model, the potential drug efficacy would be affected and the data might be misleading to in vivo and clinical studies. Thus, focus has been shifted to the 3D cell culture system as the technique is designed to improve the cellular structures and physiology for better mimicry of tissues and organs specific microarchitectures (Ballav et al., 2021).

1.3 Significance of research

This study highlights the importance of curative treatment for retinoblastoma through a reliable 3D cell culture model using plant resources, bawang Dayak as an alternative medicine. This study offers a substantial understanding of the native behaviour for cell growth and in vivo tissue resemblance, which could highly minimise animal testing. Since animal testing is the subject of heated debate over these past few years, stringent ethical requirements are necessary, and most of the animal-right extremists and anti-vivisectionists request a total abolishment of animal use in research. Besides that, the efficacy of a potential chemotherapeutic drug such as *E. bulbosa* bulb could be scrutinised optimally using a 3D cell culture approach. Moreover, secondary metabolites derived from the plant usually displays selective cytotoxic capability upon cancer cells, resulting in minimal side effects of chemotherapeutic drugs on healthy cells. Besides, a convenient yet economical method for phenolics extraction of *E. bulbosa* bulb using a green solvent, ethanol could be established. The most significant purpose of using natural resources is due to their cost-effectiveness and relative abundance in nature.

1.4 Hypothesis

It is hypothesised that the optimised extraction of *E. bulbosa* ethanolic bulb extract demonstrates selective cytotoxicity on 2D and 3D model of retinoblastoma cancer cells. It is also postulated that the extract is a potent inhibitor of retinoblastoma cancer cells through the activation of the antioxidant

defence system that could trigger the apoptotic genes and proteins related pathways.

1.5 Objectives

General objective: To investigate the in vitro effect of *E. bulbosa* ethanolic bulb extracted under optimised extraction condition on the retinoblastoma cancer cells via apoptosis and antioxidant signalling pathways.

Specific objectives:

1. To optimise the extraction condition of phenolic compounds from *E. bulbosa* bulb using response surface methodology and obtain its bioactive constituents using high-performance liquid chromatography (HPLC) analysis.
2. To investigate the effects of the optimise *E. bulbosa* ethanolic bulb extract as a chemotherapeutic potential on apoptotic-induced 2D monolayer culture of retinoblastoma cancer cells and its underlying mechanisms.
3. To develop a 3D retinoblastoma cell culture model extracted from the rat tail tendon and observe the effects of *E. bulbosa* ethanolic bulb extract on the morphological changes of the developed 3D retinoblastoma cell culture.
4. To investigate the anti-cancer effects of *E. bulbosa* ethanolic bulb extract on the 3D retinoblastoma cell culture model through regulation of apoptotic and antioxidant related genes and proteins.

BIBLIOGRAPHY

- Abdeen, A. A., Lee, J., and Kilian, K. A. (2016). Capturing extracellular matrix properties in vitro: Microengineering materials to decipher cell and tissue level processes. *Experimental Biology and Medicine*, 241(9), 930-938.
- Abou-Ghali, M., and Stiban, J. (2015). Regulation of ceramide channel formation and disassembly: Insights on the initiation of apoptosis. *Saudi Journal of Biological Sciences*, 22(6), 760-772.
- Adams, C. M., Clark-Garvey, S., Porcu, P., and Eischen, C. M. (2019). Targeting the Bcl-2 family in B cell lymphoma. *Frontiers in Oncology*, 8, 636.
- Agcam, E., Akyıldız, A., and Balasubramaniam, V. M. (2017). Optimization of anthocyanins extraction from black carrot pomace with thermosonication. *FoodnChemistry*, 237, 461-470.
- Agustin, A. R., Faika, S., and Ju, Y. H. (2016). Influence of extracting solvents on its antioxidant properties of bawang Dayak (*Eleutherine palmifolia* L.). *International Journal of Chemical and Petrochemical Technology*, 6(2), 1-10.
- AlAli, A., Kletke, S., Gallie, B., and Lam, W. C. (2018). Retinoblastoma for paediatric ophthalmologists. *The Asia-Pacific Journal of Ophthalmology*, 7(3), 160-168.
- Albert, D. M., and Dryja, T. P. (1988). Recent Studies of the Retinoblastoma Gene: What It Means to the Ophthalmologist. *Archives of Ophthalmology*, 106(2), 181-182.
- Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K., and Walter, P. (2002). Programmed cell death (apoptosis). In *Molecular Biology of the Cell*. 4th edition. Garland Science.
- Almater, A., Alfaleh, A., Alshomar, K., and AlMesfer, S. (2019). Retinoblastoma: Update on current management. *Retinoblastoma—Past, Present and Future*.
- Al-Oqail, M. M. (2021). Anticancer efficacies of *Krameria lappacea* extracts against human breast cancer cell line (MCF-7): Role of oxidative stress and ROS generation. *Saudi Pharmaceutical Journal*, 29(3), 244-251.
- Alvarado, C. J. (1999). Sterilization vs. disinfection vs. clean. *The Nursing clinics of North America*, 34(2), 483-491.
- Ancona-Lezama, D., Dalvin, L. A., and Shields, C. L. (2020). Modern treatment of retinoblastoma: A 2020 review. *Indian Journal of Ophthalmology*, 68(11), 2356.

- Andersch, L., Radke, J., Klaus, A., Schwiebert, S., Winkler, A., Schumann, E., Grunewald, L., Zirngibl, F., Flemmig, C., Jensen, M. C., Rossig, C., Joussen, A., Henssen, A., Eggert, A., Schulte, J. H., Künkele, A. (2019). CD171-and GD2-specific CAR-T cells potently target retinoblastoma cells in preclinical in vitro testing. *BMC Cancer*, 19(1), 895.
- Antoni, D., Burckel, H., Josset, E., and Noel, G. (2015). Three-dimensional cell culture: A breakthrough *in vivo*. *International Journal of Molecular Sciences*, 16(3), 5517-5527.
- Anwar, H., Hussain, G., and Mustafa, I. (2018). Antioxidants from natural sources. *Antioxidants in Foods and its Applications*, 1-27.
- Arbab, I.A., Abdul, A.B., Sukari, M.A., Abdullah, R., Syam, S., Kamalidehghan, B., Ibrahim, M.Y., Taha, M.M.E., Abdelwahab, S.I., Ali, H.M. and Mohan, S. (2013). Dentatin isolated from *Clausena excavata* induces apoptosis in MCF-7 cells through the intrinsic pathway with involvement of NF-κB signalling and G0/G1 cell cycle arrest: A bioassay-guided approach. *Journal of Ethnopharmacology* 145, 343-354.
- Arrowsmith, J., and Miller, P. (2013). Trial watch: phase II and phase III attrition rates 2011-2012. *Nature Reviews. Drug discovery*, 12(8), 569.
- Arseni, L., Lombardi, A., and Orioli, D. (2018). From structure to phenotype: Impact of collagen alterations on human health. *International Journal of Molecular Sciences*, 19(5), 1407.
- Asadi-Samani, M., Farkhad, N. K., Mahmoudian-Sani, M. R., and Shirzad, H. (2019). Antioxidants as a double-edged sword in the treatment of cancer. In *Antioxidants*. IntechOpen.
- Ashkenazi, A. (2002). Targeting death and decoy receptors of the tumour-necrosis factor superfamily. *Nature Reviews Cancer*, 2(6), 420-430.
- Asif, M. (2015). Chemistry and antioxidant activity of plants containing some phenolic compounds. *Chemistry International*, 1(1), 35-52.
- Aybastıer, Ö., İşik, E., Şahin, S., and Demir, C. (2013). Optimization of ultrasonic-assisted extraction of antioxidant compounds from blackberry leaves using response surface methodology. *Industrial Crops and Products*, 44, 558-565.
- Aydar, A. Y. (2018). Utilization of response surface methodology in optimization of extraction of plant materials. *Statistical approaches with emphasis on design of experiments applied to chemical processes*, 157-169.
- Aydar, A. Y., Bağdatlıoğlu, N., and Köseoğlu, O. (2017). Effect of ultrasound on olive oil extraction and optimization of ultrasound-assisted extraction of extra virgin olive oil by response surface methodology (RSM). *Grasas y Aceites*, 68(2), 189.

- Ayodele, B. V., and Abdullah, S. (2018). An overview of response surface methodology approach to optimization of hydrogen and syngas production by catalytic reforming of greenhouse gases (CH₄ and CO₂). *Statistical Approaches with Emphasis on Design of Experiments Applied to Chemical Processes*, 65-78.
- Azahar, N. F., Abd Gani, S. S., and Mokhtar, N. F. M. (2017). Optimization of phenolics and flavonoids extraction conditions of *Curcuma zedoaria* leaves using response surface methodology. *Chemistry Central Journal*, 11(1), 1-10.
- Azzam, E. I., De Toledo, S. M., and Little, J. B. (2004). Stress signalling from irradiated to non-irradiated cells. *Current Cancer Drug Targets*, 4(1), 53-64.
- Baek, J., and Lee, M. G. (2016). Oxidative stress and antioxidant strategies in dermatology. *Redox Report*, 21(4), 164-169.
- Baeza, G., Sarriá, B., Mateos, R., and Bravo, L. (2016). Dihydrocaffeic acid, a major microbial metabolite of chlorogenic acids, shows similar protective effect than a yerba mate phenolic extract against oxidative stress in HepG2 cells. *Food Research International*, 87, 25-33.
- Bahtiar, A. and Annisa, R. (2018). Effects of Dayak onion bulbs (*Eleutherine bulbosa* (Mill.) Urb) on bone development of the hipoestrogen model rat. *Pharmacog Journal*, 10(2).
- Bairati, I., Meyer, F., Jobin, E., Gélinas, M., Fortin, A., Nabid, A., Brochet, F., and Tétu, B. (2006). Antioxidant vitamins supplementation and mortality: A randomized trial in head and neck cancer patients. *International Journal of Cancer*, 119(9), 2221-2224.
- Baker, B. M., and Chen, C. S. (2012). Deconstructing the third dimension—how 3D culture microenvironments alter cellular cues. *Journal of Cell Science*, 125(13), 3015-3024.
- Ballav, S., Deshmukh, A. J., Siddiqui, S., Aich, J., and Basu, S. (2021). In Cell Culture- Advanced Technology and Applications in Medical and Life Sciences (Ed.), *Two-dimensional and three-dimensional cell culture and their applications* (pp. 1-27). IntechOpen.
- Balmer, A., and Munier, F. (2007). Differential diagnosis of leukocoria and strabismus, first presenting signs of retinoblastoma. *Clinical Ophthalmology (Auckland, NZ)*, 1(4), 431.
- Balvan, J., Krizova, A., Gumulec, J., Raudenska, M., Sladek, Z., Sedlackova, M., Babula, P., Sztalmachova, M., Kizek, R., Chmelik, R., and Masarik, M. (2015). Multimodal holographic microscopy: Distinction between apoptosis and oncosis. *PloS one*, 10(3), e0121674.

- Bansal, S., Choudhary, S., Sharma, M., Kumar, S. S., Lohan, S., Bhardwaj, V., Syan, N., and Jyoti, S. (2013). Tea: A native source of antimicrobial agents. *Food research international*, 53(2), 568-584.
- Barabutis, N., Schally, A. V., and Siejka, A. (2018). P53, GHRH, inflammation and cancer. *EBioMedicine*, 37, 557-562.
- Basu, A. K. (2018). DNA damage, mutagenesis and cancer. *International Journal of Molecular Sciences*, 19(4), 970.
- Beesoo, R., Neergheen-Bhujun, V., Bhagooli, R., and Bahorun, T. (2014). Apoptosis inducing lead compounds isolated from marine organisms of potential relevance in cancer treatment. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 768, 84-97.
- Behbahani, M., Moghaddam, M. A., and Arami, M. (2011). Techno-economical evaluation of fluoride removal by electrocoagulation process: Optimization through response surface methodology. *Desalination*, 271(1-3), 209-218.
- Bella, J. (2016). Collagen structure: New tricks from a very old dog. *Biochemical Journal*, 473(8), 1001-1025.
- Benetou, V., Lagiou, A., and Lagiou, P. (2015). Chemoprevention of cancer: Current evidence and future prospects. *F1000Research*, 4(F1000 Faculty Rev).
- Benmeddour, Z., Mehinagic, E., Le Meurlay, D., and Louaileche, H. (2013). Phenolic composition and antioxidant capacities of ten Algerian date (*Phoenix dactylifera* L.) cultivars: A comparative study. *Journal of Functional Foods*, 5(1), 346-354.
- Bezerra, M. A., Santelli, R. E., Oliveira, E. P., Villar, L. S., and Escaleira, L. A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 76(5), 965-977.
- Block, K. I., Koch, A. C., Mead, M. N., Tothy, P. K., Newman, R. A., and Gyllenhaal, C. (2008). Impact of antioxidant supplementation on chemotherapeutic toxicity: A systematic review of the evidence from randomized controlled trials. *International Journal of Cancer*, 123(6), 1227-1239.
- Bonnans, C., Chou, J., and Werb, Z. (2014). Remodelling the extracellular matrix in development and disease. *Nature reviews Molecular Cell Biology*, 15(12), 786-801.
- Bonnier, F., Keating, M. E., Wrobel, T. P., Majzner, K., Baranska, M., Garcia-Munoz, A., Blanco, A., and Byrne, H. J. (2015). Cell viability assessment using the Alamar blue assay: A comparison of 2D and 3D cell culture models. *Toxicology in vitro*, 29(1), 124-131.

- Boubekri, C., Lanez, T., and Djouadi, A. (2015). A comparative study on antioxidant activities and phenolic contents of five algerian eggplant cultivars. *Scientific Study & Research. Chemistry and Chemical Engineering, Biotechnology, Food Industry*, 16(1), 29.
- Brantley, Jr, M. A., and Harbour, J. W. (2001). The molecular biology of retinoblastoma. *Ocular Immunology and Inflammation*, 9(1), 1-8.
- Bray, F., and Møller, B. (2006). Predicting the future burden of cancer. *Nature Reviews Cancer*, 6(1), 63.
- Broaddus, E., Topham, A., and Singh, A. D. (2009). Incidence of retinoblastoma in the USA: 1975–2004. *British Journal of Ophthalmology*, 93(1), 21-23.
- Brougham, C. M., Levingstone, T. J., Shen, N., Cooney, G. M., Jockenhoevel, S., Flanagan, T. C., and O'Brien, F. J. (2017). Freeze-drying as a novel biofabrication method for achieving a controlled microarchitecture within large, complex natural biomaterial scaffolds. *Advanced Healthcare Materials*, 6(21), 1700598.
- Brown, B. N., Valentin, J. E., Stewart-Akers, A. M., McCabe, G. P., and Badylak, S. F. (2009). Macrophage phenotype and remodeling outcomes in response to biologic scaffolds with and without a cellular component. *Biomaterials*, 30(8), 1482-1491.
- Bukhari, S., Aziz-ur-Rehman, B. I., and Qidwai, U. (2011). Presentation pattern of retinoblastoma. *Pakistan Journal of Ophthalmology*, 27(3), 142-5.
- Cancer Research United Kingdom. (2012). Retrieved from <https://www.cancerresearchuk.org/sites/default/files/cstreamnode/incaatomicalsitere.pdf>. Accessed on January 19th, 2019.
- Carneiro, B. A. and El-Deiry, W. S. (2020). Targeting apoptosis in cancer therapy. *Nature reviews Clinical oncology*, 17(7), 395-417.
- Carocho, M., and Ferreira, I. C. (2013). A review on antioxidants, prooxidants and related controversy: Natural and synthetic compounds, screening and analysis methodologies and future perspectives. *Food and Chemical Toxicology*, 51, 15-25.
- Cekanova, M., and Rathore, K. (2014). Animal models and therapeutic molecular targets of cancer: Utility and limitations. *Drug Design, Development and Therapy*, 8, 1911.
- Chaiklahan, R., Chirasuwan, N., Triratana, P., Loha, V., Tia, S., and Bunnag, B. (2013). Polysaccharide extraction from Spirulina sp. and its antioxidant capacity. *International Journal of Biological Macromolecules*, 58, 73-78.
- Chan, L. L. Y., Smith, T., Kumph, K. A., Kuksin, D., Kessel, S., Déry, O., Cribbes, S., Lai, N., Qiu, J. (2016). A high-throughput AO/PI-based cell

- concentration and viability detection method using the Celigo image cytometry. *Cytotechnology*, 68, 2015-2025.
- Chang, A. B., Bush, A., and Bronchiectasis, G. K. (2018). GLOBOCAN 2018: Counting the toll of cancer. *Lancet*, 392(10154), 1196-1196.
- Chantada, G., Fandiño, A., Manzitti, J., Urrutia, L., and Schvartzman, E. (1999). Late diagnosis of retinoblastoma in a developing country. *Archives of Disease in Childhood*, 80(2), 171-174.
- Chattopadhyay, S., and Raines, R. T. (2014). Collagen-based biomaterials for wound healing. *Biopolymers*, 101(8), 821-833.
- Chawla, B., Kumar, K., and Singh, A. D. (2017). Influence of socioeconomic and cultural factors on retinoblastoma management. *Asia-Pacific Journal of Oncology Nursing*, 4(3), 187.
- Chen, D. L., Hu, M. G., Liu, Y. Y., Li, R. T., Yu, M., Xu, X. D., and Ma, G. X. (2018). New naphthalene derivatives from the bulbs of *Eleutherine americana* with their protective effect on the injury of HUVECs. *Molecules*, 23(9), 2111.
- Chen, Y., McMillan-Ward, E., Kong, J., Israels, S. J., and Gibson, S. B. (2008). Oxidative stress induces autophagic cell death independent of apoptosis in transformed and cancer cells. *Cell Death and Differentiation*, 15(1), 171-182.
- Chen, Y., Xie, M. Y., and Gong, X. F. (2007). Microwave-assisted extraction used for the isolation of total triterpenoid saponins from *Ganoderma atrum*. *Journal of Food Engineering*, 81(1), 162-170.
- Chiang, S. K., Chen, S. E., and Chang, L. C. (2021). The role of HO-1 and its crosstalk with oxidative stress in cancer cell survival. *Cells*, 10(9), 2401.
- Chiang, S. K., Chen, S. E., and Chang, L. C. (2018). A dual role of heme oxygenase-1 in cancer cells. *International Journal of Molecular Sciences*, 20(1), 39.
- Chio, I. I. C., and Tuveson, D. A. (2017). ROS in cancer: The burning question. *Trends in Molecular Medicine*, 23(5), 411-429.
- Chi, L. C. M., Ho, T. S., Wong, E. Y. L., and Ooi, V. E. (2006). Ethyl acetate extract of *Patrinia scabiosaeefolia* downregulates anti-apoptotic Bcl-2/Bcl-XL expression, and induces apoptosis in human breast carcinoma MCF-7 cells independent of caspase-9 activation. *Journal of Ethnopharmacology*, 105(1-2), 263-268.
- Clevers, H. (2016). Modelling development and disease with organoids. *Cell*, 165(7), 1586-1597.

- Constantinou, C., Papas, A., and Constantinou, A. I. (2008). Vitamin E and cancer: An insight into the anticancer activities of vitamin E isomers and analog. *International Journal of Cancer*, 123(4), 739-752.
- Copes, F., Pien, N., Van Vlierberghe, S., Boccafoschi, F., and Mantovani, D. (2019). Collagen-based tissue engineering strategies for vascular medicine. *Frontiers in Bioengineering and Biotechnology*, 7, 166.
- Cornell, J. A. (1990). How to apply response surface methodology, Vol. 8. ASQC, Wisconsin, 3.
- Cotter, T. G. (2009). Apoptosis and cancer: The genesis of a research field. *Nature Reviews Cancer*, 9(7), 501-507.
- Couto, C. L., Moraes, D. F., Maria do Socorro, S. C. A., do Amaral, F. M., and Guerra, R. N. (2016). *Eleutherine bulbosa* (Mill.) Urb.: A review study. *Journal of Medicinal Plants Research*, 10(21), 286-297.
- Craig, A. S., Eikenberry, E. F. and Parry, D. A. D. (1987). Ultrastructural organization of skin: Classification on the basis of mechanical role. *Connective Tissue Research*, 16(3), 213-223.
- Ćujić, N., Šavikin, K., Janković, T., Pljevljakušić, D., Zdunić, G., and Ibrić, S. (2016). Optimization of polyphenols extraction from dried chokeberry using maceration as traditional technique. *Food Chemistry*, 194, 135-142.
- Cukierman, E., Pankov, R., Stevens, D. R., and Yamada, K. M. (2001). Taking cell-matrix adhesions to the third dimension. *Science*, 294(5547), 1708-1712.
- D'Arcy, M. S. (2019). Cell death: A review of the major forms of apoptosis, necrosis and autophagy. *Cell Biology International*, 43(6), 582-592.
- Damato, B. (2012). Progress in the management of patients with uveal melanoma. The 2012 Ashton Lecture. *Eye*, 26(9), 1157.
- Daniele, M. A., Adams, A. A., Naciri, J., North, S. H., and Ligler, F. S. (2014). Interpenetrating networks based on gelatin methacrylamide and PEG formed using concurrent thiol click chemistries for hydrogel tissue engineering scaffolds. *Biomaterials*, 35(6), 1845-1856.
- Davinelli, S., Bertoglio, J. C., Zarrelli, A., Pina, R., and Scapagnini, G. (2015). A randomized clinical trial evaluating the efficacy of an anthocyanin–maqui berry extract (Delphinol®) on oxidative stress biomarkers. *Journal of the American College of Nutrition*, 34(sup1), 28-33.
- De Bont, R., and Van Larebeke, N. (2004). Endogenous DNA damage in humans: A review of quantitative data. *Mutagenesis*, 19(3), 169-185.

- de Giffoni de Carvalho, J. T., da Silva Baldivia, D., Leite, D. F., de Araújo, L. C. A., de Toledo Espindola, P. P., Antunes, K. A., Rocha, P. S., de Picoli Souza, K., and dos Santos, E. L. (2019). Medicinal Plants from Brazilian Cerrado: Antioxidant and Anticancer Potential and Protection against Chemotherapy Toxicity. *Oxidative medicine and cellular longevity*, 2019, 16.
- de Oliveira, L. G., de Paiva, A. P., Balestrassi, P. P., Ferreira, J. R., da Costa, S. C., and da Silva Campos, P. H. (2019). Response surface methodology for advanced manufacturing technology optimization: Theoretical fundamentals, practical guidelines, and survey literature review. *The International Journal of Advanced Manufacturing Technology*, 104(5-8), 1785-1837.
- de SS Quintans, J., Soares, B. M., Ferraz, R. P., Oliveira, A. C., da Silva, T. B., Menezes, L. R. A., Sampaio, M. F. C., do N. Prata, A. P., Moraes, M. O., Pessoa, C., Antoniolli, A. R., Costa, E. V., Bezerra, D. P. (2013). Chemical constituents and anticancer effects of the essential oil from leaves of *Xylopia laevigata*. *Planta Medica*, 29, 123-130.
- Delarive, T., Rossier, A., Rossier, S., Ravinet, E., Shaarawy, T., and Mermoud, A. (2003). Aqueous dynamic and histological findings after deep sclerectomy with collagen implant in an animal model. *British Journal of Ophthalmology*, 87(11), 1340-1344.
- Delgado, L. M., Pandit, A., and Zeugolis, D. I. (2014). Influence of sterilisation methods on collagen-based devices stability and properties. *Expert Review of Medical devices*, 11(3), 305-314.
- Deng, C., Zhang, P., Harper, J. W., Elledge, S. J., and Leder, P. (1995). Mice lacking p21CIP1/WAF1 undergo normal development, but are defective in G1 checkpoint control. *Cell*, 82(4), 675-684.
- Deng, G. F., Lin, X., Xu, X. R., Gao, L. L., Xie, J. F., and Li, H. B. (2013). Antioxidant capacities and total phenolic contents of 56 vegetables. *Journal of Functional Foods*, 5(1), 260-266.
- Dickreuter, E., and Cordes, N. (2017). The cancer cell adhesion resistome: Mechanisms, targeting and translational approaches. *Biological Chemistry*, 398(7), 721-735.
- Dimaras, H., Corson, T. W., Cobrinik, D., White, A., Zhao, J., Munier, F. L., Abraham, D. H., Shields, C. L., Chantada, G. L., Njuguna, F., and Gallie, B. L. (2015). Retinoblastoma. *Nature reviews Disease primers*, 1(1), 1-23.
- Dimaras, H., Kimani, K., Dimba, E. A., Grondahl, P., White, A., Chan, H. S., and Gallie, B. L. (2012). Retinoblastoma. *The Lancet*, 379(9824), 1436-1446.

- Dimitorios, B. (2006). Sources of natural phenolics antioxidants. *Trends in Food Sci. and Technol*, 17, 505-512.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S., and Ju, Y. H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatica*. *Journal of Food Drug Analysis*, 22 (3), 296–302.
- Dondey, J. C., Staffieri, S., McKenzie, J., Davie, G., and Elder, J. (2004). Retinoblastoma in Victoria, 1976–2000: Changing management trends and outcomes. *Clinical and Experimental Ophthalmology*, 32(4), 354-359.
- Donehower, L. A., Harvey, M., Slagle, B. L., McArthur, M. J., Montgomery, C. A., Butel, J. S., and Bradley, A. (1992). Mice deficient for p53 are developmentally normal but susceptible to spontaneous tumours. *Nature*, 356(6366), 215-221.
- Dong, C., and Lv, Y. (2016). Application of collagen scaffold in tissue engineering: Recent advances and new perspectives. *Polymers*, 8(2), 42.
- Doyle, A. D., Carvajal, N., Jin, A., Matsumoto, K., and Yamada, K. M. (2015). Local 3D matrix microenvironment regulates cell migration through spatiotemporal dynamics of contractility-dependent adhesions. *Nature Communications*, 6(1), 1-15.
- Dryja, T. P., Cavenee, W., White, R., Rapaport, J. M., Petersen, R., Albert, D. M., and Bruns, G. A. (1984). Homozygosity of chromosome 13 in retinoblastoma. *New England Journal of Medicine*, 310(9), 550-553.
- Dunkel, I. J., Aledo, A., Kernan, N. A., Kushner, B., Bayer, L., Gollamudi, S. V., Finlay, J. L., Abramson, D. H. (2000). Successful treatment of metastatic retinoblastoma. *Cancer*, 89, 2117-2121.
- Duval, K., Grover, H., Han, L. H., Mou, Y., Pegoraro, A. F., Fredberg, J., and Chen, Z. (2017). Modeling physiological events in 2D vs. 3D cell culture. *Physiology*, 32(4), 266-277.
- Eagle Jr, R. C. (2013). The pathology of ocular cancer. *Eye*, 27(2), 128.
- Eilenberger, C., Kratz, S. R. A., Rothbauer, M., Ehmoser, E. K., Ertl, P., and Küpcü, S. (2018). Optimized Alamar Blue assay protocol for drug dose-response determination of 3D tumor spheroids. *MethodsX*, 5, 781-787.
- Elsen, S. R., and Ramesh, T. (2015). Optimization to develop multiple response hardness and compressive strength of zirconia reinforced alumina by using RSM and GRA. *International Journal of Refractory Metals and Hard Materials*, 52, 159-164.

- Eshar, D., Wyre, N. R., and Schoster, J. V. (2011). Use of collagen shields for treatment of chronic bilateral corneal ulcers in a pet rabbit. *Journal of Small Animal Practice*, 52(7), 380-383.
- Ezzati, M., Pearson-Stuttard, J., Bennett, J. E., and Mathers, C. D. (2018). Acting on non-communicable diseases in low- and middle-income tropical countries. *Nature*, 559(7715), 507-516.
- Fabian, I. D., Reddy, A., and Sagoo, M. S. (2018). Classification and staging of retinoblastoma. *Community Eye Health Journal*, 31(101), 11.
- Farghadani, R., and Naidu, R. (2021). The Role of Apoptosis as a Double-Edge Sword in Cancer. In (Ed.) *Regulation and Dysfunction of Apoptosis* (pp. 1-23). IntechOpen.
- Fasihnia, S. H., Peighambardoust, S. H., Peighambardoust, S. J., Oromiehie, A., Soltanzadeh, M., and Peressini, D. (2020). Migration analysis, antioxidant, and mechanical characterization of polypropylene-based active food packaging films loaded with BHA, BHT, and TBHQ. *Journal of Food Science*, 85(8), 2317-2328.
- FDA, U., 2021. Appendix 6: Toxicological Data for Class 3 Solvents. (Accessed 19 February 2021).
- Fitri, Y., and Rosidah, E. S. (2014). Effects of inhibition cell cycle and apoptosis of sabrang onion extract (*Eleutherine bulbosa* (Mill.) Urb.) on breast cancer cells. *International Journal of PharmTech Research*, 6(4), 1392-1396.
- Fitri, Y., Rosidah, Suwarso, E. (2014). Effects of inhibition cell cycle and apoptosis of Sabrang Onion extract (*Eleutherine bulbosa* (Mill) Urb.) on breast cancer cells. *International Journal of PharmaTech Research*, 6(4), 1392-1396.
- Foo, S. C., Yusoff, F. M., Imam, M. U., Foo, J. B., Ismail, N., Azmi, N. H., Tor, Y. S., Khong, N. M. H., Ismail, M. (2019). Increased fucoxanthin in *Chaetoceros calcitrans* extract exacerbates apoptosis in liver cancer cells via multiple targeted cellular pathways. *Biotechnology Report*, 21, e00296.
- Food and Drug Administration, U. S. (2021). Appendix 6: Toxicological Data for Class 3 Solvents. (Accessed on 3rd January 2021). <https://www.fda.gov/regulatory-information/search-fda-guidance/documents/q3cappendix-6>
- Forrester, S. J., Kikuchi, D. S., Hernandes, M. S., Xu, Q., and Griendling, K. K. (2018). Reactive oxygen species in metabolic and inflammatory signaling. *Circulation research*, 122(6), 877-902.
- Friedl, P., Sahai, E., Weiss, S., and Yamada, K. M. (2012). New dimensions in cell migration. *Nature Reviews Molecular Cell Biology*, 13(11), 743-747.

- Friedrich, J., Ebner, R., and Kunz-Schughart, L. A. (2007). Experimental anti-tumor therapy in 3-D: spheroids—old hat or new challenge?. *International Journal of Radiation Biology*, 83(11-12), 849-871.
- Fukai, T. and Ushio-Fukai, M. (2011). Superoxide dismutases: Role in redox signaling, vascular function, and diseases. *Antioxidants and Redox Signaling*, 15(6), 1583- 1606.
- Fu, L., Xu, B. T., Xu, X. R., Qin, X. S., Gan, R. Y., and Li, H. B. (2010). Antioxidant capacities and total phenolic contents of 56 wild fruits from South China. *Molecules*, 15(12), 8602-8617.
- G Murillo, A., and L Fernandez, M. (2017). The relevance of dietary polyphenols in cardiovascular protection. *Current Pharmaceutical Design*, 23(17), 2444-2452.
- Galaris, D., Skiada, V., and Barbouti, A. (2008). Redox signaling and cancer: the role of "labile" iron. *Cancer Letters*, 266(1), 21-29.
- Galingging, R. Y. (2009). Bawang dayak (*Eleutherine palmifolia*) as a multifunctional medicinal plant. *Warta Penelitian dan Pengembangan*, 15(3), 2-4.
- Gallo, F. R., Palazzino, G., Federici, E., Iurilli, R., Galeffi, C., Chifundera, K., and Nicoletti, M. (2010). Polyketides from *Eleutherine bulbosa*. *Natural product Research*, 24(16), 1578-1586.
- Garcia-Salas, P., Morales-Soto, A., Segura-Carretero, A., and Fernández-Gutiérrez, A. (2010). Phenolic compound extraction systems for fruit and vegetable samples. *Molecules*, 15(12), 8813-8826.
- Geismann, C., Arlt, A., Sebens, S., and Schäfer, H. (2014). Cytoprotection "gone astray": Nrf2 and its role in cancer. *Oncotargets and therapy*, 7, 1497.
- Gibbs, D. M., Black, C. R., Dawson, J. I., and Oreffo, R. O. (2016). A review of hydrogel uses in fracture healing and bone regeneration. *Journal of Tissue Engineering and Regenerative Medicine*, 10(3), 187-198.
- Gichigo, E. N., Kariuki-Wanyoike, M. M., Kimani, K., and Nentwich, M. M. (2015). Retinoblastom in Kenya. *Der Ophthalmologe*, 112(3), 255-260.
- Gingras, M., Paradis, I., and Berthod, F. (2003). Nerve regeneration in a collagen-chitosan tissue-engineered skin transplanted on nude mice. *Biomaterials*, 24(9), 1653-1661.
- Gogvadze, V., Orrenius, S., Zhivotovsky, B. 2009, February. Mitochondria as targets for cancer chemotherapy. In *Seminars in cancer biology* (Vol. 19, No. 1, pp. 57-66). Academic Press.

- Goldar, S., Khaniani, M. S., Derakhshan, S. M., and Baradaran, B. (2015). Molecular mechanisms of apoptosis and roles in cancer development and treatment. *Asian Pacific Journal of Cancer Prevention*, 16(6), 2129-2144.
- Gombos, D. S., Hungerford, J., Abramson, D. H., Kingston, J., Chantada, G., Dunkel, I. J., Antoneli, C. B. G., Greenwald, M., Haik, B. G., Leal, C. A., Medina-Sanson, A., Scheffler, A. C., Veerakul, G., Wieland, R., Bornfeld, N., Wilson, M. W., Yu, C. B. O. (2007). Secondary acute myelogenous leukemia in patients with retinoblastoma: is chemotherapy a factor?. *Ophthalmology*, 114(7), 1378-1383.
- Gontijo, V. S., Dos Santos, M. H., and Viegas Jr, C. (2017). Biological and chemical aspects of natural biflavonoids from plants: A brief review. *Mini Reviews in Medicinal Chemistry*, 17(10), 834-862.
- Goolam, S., Kana, H., Welsh, N., Wainwright, L., Poole, J., and Mayet, I. (2018). A 20-year retrospective review of retinoblastoma at two tertiary academic hospitals in Johannesburg, South Africa. *Ocular Oncology and Pathology*, 4(3), 170-175.
- Grantab, R., Sivananthan, S., and Tannock, I. F. (2006). The penetration of anticancer drugs through tumor tissue as a function of cellular adhesion and packing density of tumor cells. *Cancer Research*, 66(2), 1033-1039.
- Green, D. R., and Llambi, F. (2015). Cell death signaling. *Cold Spring Harbor perspectives in biology*, 7(12), a006080.
- Grosso, G., Bella, F., Godos, J., Sciacca, S., Del Rio, D., Ray, S., Galvano, F., and Giovannucci, E. L. (2017). Possible role of diet in cancer: systematic review and multiple meta-analyses of dietary patterns, lifestyle factors, and cancer risk. *Nutrition reviews*, 75(6), 405-419.
- Gu, L., Shan, T., Ma, Y. X., Tay, F. R., and Niu, L. (2019). Novel biomedical applications of crosslinked collagen. *Trends in Biotechnology*, 37(5), 464-491.
- Guarino, V., Cirillo, V., Altobelli, R. and Ambrosio, L. (2015). Polymer-based platforms by electric field-assisted techniques for tissue engineering and cancer therapy. *Expert Review of Medical Devices*, 12(1), 113-129.
- Haggerty, H., Richardson, S., Hrisos, S., Strong, N. P., and Clarke, M. P. (2004). The Newcastle Control Score: a new method of grading the severity of intermittent distance exotropia. *British Journal of Ophthalmology*, 88(2), 233-235.
- Haghshenas, H. F., Khodaii, A., Khedmati, M., and Tapkin, S. (2015). A mathematical model for predicting stripping potential of Hot Mix Asphalt. *Construction and Building Materials*, 75, 488-495.

- Halliwell, B., Cheah, I. K., and Tang, R. M. (2018). Ergothioneine—a diet-derived antioxidant with therapeutic potential. *Febs Letters*, 592(20), 3357-3366.
- Han, A. R., Min, H. Y., Nam, J. W., Lee, N. Y., Wirawan, A., Suprapto, W., Lee, S. K., Lee, K. R., and Seo, E. K. (2008). Identification of a new naphthalene and its derivatives from the bulb of *Eleutherine americana* with inhibitory activity on lipopolysaccharide-induced nitric oxide production. *Chemical and Pharmaceutical Bulletin*, 56(9), 1314-1316.
- Hanahan, D. (2022). Hallmarks of Cancer: New Dimensions. *Cancer Discovery*, 12(1), 31-46.
- Hanahan, D., and Weinberg, R. A. (2011). Hallmarks of cancer: The next generation. *Cell*, 144(5), 646-674.
- Hara, H., Maruyama, N., Yamashita, S., Hayashi, Y., Lee, K. H., Bastow, K. F., Chairul Marumoto, R., Imakura, Y. (1997). Elecanacin, a novel new naphthoquinone from the bulb of *Eleutherine americana*. *Chemical and Pharmaceutical Bulletin*, 45 (10), 1714–1716.
- Harbour, J. W. (2001). Retinoblastoma: Pathogenesis and diagnosis. *Tumors of the eye and orbit*. Philadelphia: BC Decker, 253-65.
- Harbour, J. W. (2006). Eye Cancer: Unique Insights into Oncogenesis: The Cogan Lecture. *Investigative Ophthalmology and visual science*, 47(5), 1737-1745.
- Harrell, C. R., Djonov, V., Fellabaum, C., and Volarevic, V. (2018). Risks of using sterilization by gamma radiation: The other side of the coin. *International Journal of Medical Sciences*, 15(3), 274.
- Hassan, M., Watari, H., AbuAlmaaty, A., Ohba, Y., and Sakuragi, N. (2014). Apoptosis and molecular targeting therapy in cancer. *BioMed research international*, 2014.
- Hekimi, S., Wang, Y., and Noë, A. (2016). Mitochondrial ROS and the effectors of the intrinsic apoptotic pathway in aging cells: The discerning killers!. *Frontiers in Genetics*, 7, 161.
- Hengartner, M. O. (2001). Apoptosis: Corralling the corpses. *Cell*, 104(3), 325-328.
- Hoarau-Véchot, J., Rafii, A., Touboul, C., and Pasquier, J. (2018). Halfway between 2D and animal models: Are 3D cultures the ideal tool to study cancer-microenvironment interactions?. *International Journal of Molecular Sciences*, 19(1), 181.
- Hoffman, A. S. (2012). Hydrogels for biomedical applications. *Advanced Drug Delivery Reviews*, 64, 18-23.

- Holle, A. W., Young, J. L., and Spatz, J. P. (2016). In vitro cancer cell-ECM interactions inform. *Vivo Cancer Treat Adv Drug Delivery Rev*, 97.
- Hollister, S. J. (2009). Scaffold engineering: A bridge to where? *Biofabrication*, 1(1),012001.
- Howes, R. M. (2009). Dangers of antioxidants in cancer patients: a review. *Philica*, 153.<https://www.fda.gov/regulatory-information/search-fdaguidancedocuments/q3appendix-6>
- Huang, J., Zhang, L., Wan, D., Zhou, L., Zheng, S., Lin, S., and Qiao, Y. (2021). Extracellular matrix and its therapeutic potential for cancer treatment. *Signal Transduction and Targeted Therapy*, 6(1), 1-24.
- Huang, Q., Zou, Y., Arno, M. C., Chen, S., Wang, T., Gao, J., Dove, A. P., and Du, J. (2017). Hydrogel scaffolds for differentiation of adipose-derived stem cells. *Chemical Society Reviews*, 46(20), 6255-6275.
- Ibrahim, H. M., and Elkhidir, E. E. (2011). Response surface method as an efficient tool for medium optimisation. *Trends in Applied Sciences Research*, 6(2), 121.
- Ieyama, T., Gunawan-Puteri, M. D., and Kawabata, J. (2011). α -Glucosidase inhibitors from the bulb of *Eleutherine americana*. *Food Chemistry*, 128(2), 308-311.
- Ifesan, B. O. T., Joycharat, N., and Voravuthikunchai, S. P. (2009). The mode of antistaphylococcal action of *Eleutherine americana*. *FEMS Immunology & Medical Microbiology*, 57(2), 193-201.
- Imamura, Y., Mukohara, T., Shimono, Y., Funakoshi, Y., Chayahara, N., Toyoda, M., Kiyota, N., Takao, S., Kono, S., Nakatsura, T., and Minami, H. (2015). Comparison of 2D-and 3D-culture models as drug-testing platforms in breast cancer. *Oncology Reports*, 33(4), 1837-1843.
- Insanu, M., Kusmardiyan, S., and Hartati, R. (2014). Recent Studies on phytochemicals and pharmacological effects of *Eleutherine americana* Merr. *Procedia Chemistry*, 13, 221-228.
- Jagetia, G. C. (2007). Radioprotection and radio sensitization by curcumin. In *The Molecular Targets and Therapeutic Uses of Curcumin in Health and Disease* (pp. 301-320). Springer, Boston, MA.
- Jahangir, M., Abdel-Farid, I. B., Kim, H. K., Choi, Y. H., and Verpoorte, R. (2009). Healthy and unhealthy plants: The effect of stress on the metabolism of Brassicaceae. *Environmental and Experimental Botany*, 67(1), 23-33.
- Jain, M., Rojanaporn, D., Chawla, B., Sundar, G., Gopal, L., and Khetan, V. (2019). Retinoblastoma in Asia. *Eye*, 33(1), 87-96.

- Jänicke, R. U., Ng, P., Sprengart, M. L., and Porter, A. G. (1998). Caspase-3 is required for α -fodrin cleavage but dispensable for cleavage of other death substrates in apoptosis. *Journal of Biological Chemistry*, 273(25), 15540-15545.
- Jensen, C., and Teng, Y. (2020). Is it time to start transitioning from 2D to 3D cell culture?. *Frontiers in Molecular Biosciences*, 7, 33.
- Ježek, J., Cooper, K. F., and Strich, R. (2018). Reactive oxygen species and mitochondrial dynamics: the yin and yang of mitochondrial dysfunction and cancer progression. *Antioxidants*, 7(1), 13.
- Jiang, J. and Xiong, Y. L. (2016). Natural antioxidants as food and feed additives to promote health benefits and quality of meat products: A review. *Meat Science*, 120, 107- 117.
- Joseph, J. S., Malindisa, S. T., and Ntwasa, M. (2018). Two-dimensional (2D) and three-dimensional (3D) cell culturing in drug discovery. *Cell Culture*, 2, 1-22.
- Kaliki, S. (2018). How to do an enucleation for retinoblastoma. *Community Eye Health Journal*, 31(101), 21.
- Kamarudin, A. A., Esa, N. M., Saad, N., Sayuti, N. H., and Razak, N. A. A. (2020). Heat assisted extraction of phenolic compounds from *Eleutherine bulbosa* (Mill.) bulb and its bioactive profiles using response surface methodology. *Industrial Crops and Products*, 144, 112064.
- Kamihara, J., Bourdeaut, F., Foulkes, W. D., Molenaar, J. J., Mossé, Y. P., Nakagawara, A., Parareda, A., Scollon, S. R., Schneider, K. M., Skalet A., States, L. J., Walsh, M. F., Diller, L. R., and Brodeur, G. M. (2017). Retinoblastoma and neuroblastoma predisposition and surveillance. *Clinical Cancer Research*, 23(13), e98-e106.
- Kao, L. Y., Su, W. W., and Lin, Y. W. (2002). Retinoblastoma in Taiwan: Survival and clinical characteristics 1978–2000. *Japanese Journal of Ophthalmology*, 46(5), 577-580.
- Kapałczyńska, M., Kolenda, T., Przybyła, W., Zajączkowska, M., Teresiak, A., Filas, V., Ibbs, M., Blizniak, R., Luczewski, L., and Lamperska, K. (2018). 2D and 3D cell cultures—a comparison of different types of cancer cell cultures. *Archives of Medical Science*, 14(4), 910-919.
- Kapinova, A., Kubatka, P., Golubnitschaja, O., Kello, M., Zubor, P., Solar, P., and Pec, M. (2018). Dietary phytochemicals in breast cancer research: Anticancer effects and potential utility for effective chemoprevention. *Environmental Health and Preventive Medicine*, 23(1), 1-18.
- Kacioglu, Z. A., and Hadjistilianou, D. (2005). Ocular tumors. In *Orbital Tumors* (pp. 257-265). Springer, New York, NY.

- Karimifard, S., and Moghaddam, M. R. A. (2018). Application of response surface methodology in physicochemical removal of dyes from wastewater: a critical review. *Science of the Total Environment*, 640, 772-797.
- Katz, L., and Baltz, R. H. (2016). Natural product discovery: past, present, and future. *Journal of Industrial Microbiology and Biotechnology*, 43(2-3), 155-176.
- Kehrer, J. P., and Klotz, L. O. (2015). Free radicals and related reactive species as mediators of tissue injury and disease: implications for health. *Critical reviews in toxicology*, 45(9), 765-798.
- Khawar, I. A., Kim, J. H., and Kuh, H. J. (2015). Improving drug delivery to solid tumors: Priming the tumor microenvironment. *Journal of Controlled Release*, 201, 78-89.
- Khazaei, S., Ramachandran, V., Esa, N. M., Etemad, A., Moradipoor, S., and Ismail, P. (2017). Flower extract of Allium atroviolaceum triggered apoptosis, activated caspase-3 and down-regulated antiapoptotic Bcl-2 gene in HeLa cancer cell line. *Biomedicine and Pharmacotherapy*, 89, 1216-1226.
- Khedmati, M., Khodaii, A., and Haghshenas, H. F. (2017). A study on moisture susceptibility of stone matrix warm mix asphalt. *Construction and Building Materials*, 144, 42-49.
- Kim, E. K., Jang, M., Song, M. J., Kim, D., Kim, Y., and Jang, H. H. (2019). Redox- mediated mechanism of chemoresistance in cancer cells. *Antioxidants*, 8(10), 471.
- Kim, J. E., Reynolds, D. S., Zaman, M. H., and Mak, M. (2018). Characterization of the mechanical properties of cancer cells in 3D matrices in response to collagen concentration and cytoskeletal inhibitors. *Integrative Biology*, 10(4), 232-241.
- Kim, J. W., and Singh, A. D. (2015). Differential Diagnosis of Leukocoria. In *Clinical Ophthalmic Oncology* (pp. 13-27). Springer, Berlin, Heidelberg.
- Koch, J., Mönch, D., Maaß, A., Gromoll, C., Hehr, T., Leibold, T., Schlitt, H. J., Dahlke, M. H., and Renner, P. (2021). Three-dimensional cultivation increases chemo-and radioresistance of colorectal cancer cell lines. *PLoS One*, 16(1), e0244513.
- Kocyigit, A., Guler, E. M., and Dikilitas, M. (2018). Role of antioxidant phytochemicals in prevention, formation and treatment of cancer. *Reactive Oxygen Species (ROS) in Living Cells*. London: InterchOpen, 21-45.

- Kocyigit, A., Guler, E. M., Haznedaroglu, I. C., and Malkan, U. Y. (2017). Ankaferd hemostat induces DNA damage, apoptosis and cytotoxic activity by generating reactive oxygen species in melanoma and normal cell lines. *International Journal of Clinical and Experimental Medicine*, 10(2), 2116-2126.
- Kocyigit, A., Koyuncu, I., Dikilitas, M., Bahadori, F., and Turkkan, B. (2016). Cytotoxic, genotoxic and apoptotic effects of naringenin-oxime relative to naringenin on normal and cancer cell lines. *Asian Pacific Journal of Tropical Biomedicine*, 6(10), 872-880.
- Komura, H., Mizukawa, K., Minakata, H., Huang, H., Qin, G., and Xu, R. (1983). New anthraquinones from *Eleutherine americana*. *Chemical and Pharmaceutical Bulletin*, 31(11), 4206-4208.
- Kondo, J., and Inoue, M. (2019). Application of cancer organoid model for drug screening and personalized therapy. *Cells*, 8(5), 470.
- König, S. M., Rissler, V., Terkelsen, T., Lambrughi, M., and Papaleo, E. (2019). Alterations of the interactome of Bcl-2 proteins in breast cancer at the transcriptional, mutational and structural level. *PLoS Computational Biology*, 15(12), e1007485.
- Konstantinidis, L., and Damato, B. (2017). Intraocular metastases—A review. *The Asia-Pacific Journal of Ophthalmology*, 6(2), 208-214.
- Kremens, B., Wieland, R., Reinhard, H., Neubert, D., Beck, J. D., Klingebiel, T., Bornfeld, N., Havers, W. (2003). High-dose chemotherapy with autologous stem cell rescue in children with retinoblastoma. *Bone Marrow Transplant*, 31, 281-284.
- Kroemer, G., Petit, P., Zamzami, N., Vayssi  re, J. L., and Mignotte, B. (1995). The biochemistry of programmed cell death. *The FASEB Journal*, 9(13), 1277-1287.
- Kulbacka, J., Saczko, J., Chwilkowska, A., Choroma  ska, A., and Sko  ucka, N. (2012). Apoptosis, free radicals and antioxidant defense in antitumor therapy. *Antioxidant Enzyme*, 265-302.
- Kulesza, J., Paw  owska, M., and Augustin, E. (2021). The Influence of Antitumor Unsymmetrical Bisacridines on 3D Cancer Spheroids Growth and Viability. *Molecules*, 26(20), 6262.
- Kumar, H., Kim, I. S., More, S. V., Kim, B. W., and Choi, D. K. (2014). Natural product-derived pharmacological modulators of Nrf2/ARE pathway for chronic diseases. *Natural Product Reports*, 31(1), 109-139.
- Kumar, S., and Pandey, A. K. (2013). Chemistry and biological activities of flavonoids: An overview. *The Scientific World Journal*, 2013.

- Kurutas, E. B. (2015). The importance of antioxidants which play the role in cellular response against oxidative/nitrosative stress: Current state. *Nutrition journal*, 15(1), 1-22.
- Kusuma, I. W., Arung, E. T., Rosamah, E., Purwatiningsih, S., Kuspradini, H., Astuti, J., Kim, Y. and Shimizu, K. (2010). Antidermatophyte and antimelanogenesis compound from *Eleutherine americana* grown in Indonesia. *Journal of Natural Medicines*, 64(2), 223-226.
- Kutschka, I., Chen, I. Y., Kofidis, T., Arai, T., Von Degenfeld, G., Sheikh, A. Y., Hendry S. L., Pearl, J., Hoyt, G., Sista, R., Yang, P. C., Blau, H. M., Gambhir, S. S., and Robbins, R. C. (2006). Collagen matrices enhance survival of transplanted cardiomyoblasts and contribute to functional improvement of ischemic rat hearts. *Circulation*, 114(1_supplement), I-167.
- Kvansakul, M., Caria, S., and Hinds, M. G. (2017). The Bcl-2 family in host-virus interactions. *Viruses*, 9(10), 290.
- Kyoto Encyclopaedia of Genes and Genomes. (2020). Retrieved from <https://www.kegg.jp/kegg/kegg2.html>. Accessed on July 30th, 2020.
- Lalande, M., Schwob, L., Vizcaino, V., Chirot, F., Dugourd, P., Schlathölter, T., and Pouilly, J. C. (2019). Direct radiation effects on the structure and stability of collagen and other proteins. *ChemBioChem*, 20(24), 2972-2980.
- Lambert, B. J., Mendelson, T. A., and Craven, M. D. (2011). Radiation and ethylene oxide terminal sterilization experiences with drug eluting stent products. *Aaps Pharmscitech*, 12(4), 1116-1126.
- Lancaster, M. A., and Knoblich, J. A. (2014). Organogenesis in a dish: Modeling development and disease using organoid technologies. *Science*, 345(6194).
- Langhans, S. A. (2018). Three-dimensional in vitro cell culture models in drug discovery and drug repositioning. *Frontiers in Pharmacology*, 9, 6.
- Latha, S., Sivarajani, G., and Dhanasekaran, D. (2017). Response surface methodology: A non-conventional statistical tool to maximize the throughput of *Streptomyces* species biomass and their bioactive metabolites. *Critical Reviews in Microbiology*, 43(5), 567-582.
- Lecumberri, E., Dupertuis, Y. M., Miralbell, R., and Pichard, C. (2013). Green teapolyphenol epigallocatechin-3-gallate (EGCG) as adjuvant in cancer therapy. *Clinical Nutrition*, 32(6), 894-903.
- Lee, T. C., and Niederer, P. (Eds.). (2010). *Basic engineering for medics and biologists: An ESEM primer* (Vol. 152). IOS Press.

- L'Espérance, S., Bachvarova, M., Tetu, B., Mes-Masson, A. M., and Bachvarov, D. (2008). Global gene expression analysis of early response to chemotherapy treatment in ovarian cancer spheroids. *BMC Genomics*, 9(1), 1-21.
- Lestari, D., Kartika, R., and Marliana, E. Antioxidant and anticancer activity of *Eleutherine bulbosa* (Mill.) Urb on leukemia cells L1210. In *Journal of Physics: Conference Series*, July 2019, (Vol. 1277, No. 1, p. 012022). IOP Publishing.
- Leu, J. J., Dumont, P., Hafey, M., Murphy, M. E., and George, D. L. (2004). Mitochondrial p53 activates Bak and causes disruption of a Bak–Mcl1 complex. *Nature Cell Biology*, 6(5), 443-450.
- Li, B. B., Smith, B., and Hossain, M. M. (2006). Extraction of phenolics from citrus peels: I. Solvent extraction method. *Separation and Purification Technology*, 48(2), 182-188.
- Li, F., Li, S., Li, H. B., Deng, G. F., Ling, W. H., and Xu, X. R. (2013). Antiproliferative activities of tea and herbal infusions. *Food and Function*, 4(4), 530-538.
- Li, S., Fu, L., Tian, T., Deng, L., Li, H., Xia, W., and Gong, Q. (2018). Disrupting SOD1 activity inhibits cell growth and enhances lipid accumulation in nasopharyngeal carcinoma. *Cell Communication and Signaling*, 16(1), 1-13.
- Li, X., Ohtsuki, T., Koyano, T., Kowithayakorn, T., Ishibashi, M. (2009). New Wnt/β-Catenin signaling inhibitors isolated from *Eleutherine palmifolia*. *Chemistry-An Asian Journal*, 4, 540-547.
- Liang, G., Tang, A., Lin, X., Li, L., Zhang, S., Huang, Z., Tang, H., and Li, Q. Q. (2010). Green tea catechins augment the antitumor activity of doxorubicin in an *in vivo* mouse model for chemoresistant liver cancer. *International Journal of Oncology* 37(1), 111-123.
- Lida, M., Fahimeh, J. A., Fariba, G., Reza, E. P., Masoumeh, M. Z., and Abdol-Mohammad, K. (2020). Evaluation of Different Sterilization Methods for Decellularized Kidney tissue. *Tissue and Cell*, 101396.
- Linn, A. M. (2005). Intraocular retinoblastoma: the case for a new group classification. *Ophthalmology Clinics of North America*, 18(1), 41-53.
- Liu, D., Nikoo, M., Boran, G., Zhou, P., and Regenstein, J. M. (2015). Collagen and gelatin. *Annual Review of Food Science and Technology*, 6, 527-557.
- Liu, H., Su, D., Zhang, J., Ge, S., Li, Y., Wang, F., Gravel, M., Roulston, A., Song, Q., Xu, W., Liang, J. G., Shore, G., Wang, X. and Liang, P. (2017). Improvement of Pharmacokinetic profile of TRAIL via trimer-tag enhances its antitumor activity *in vivo*. *Scientific reports*, 7(1), 8953.

- Liu, Y., Gan, L., Carlsson, D. J., Fagerholm, P., Lagali, N., Watsky, M. A., Munger, R., Hodge, W. G., Priest, D., and Griffith, M. (2006). A simple, cross-linked collagen tissue substitute for corneal implantation. *Investigative Ophthalmology and Visual Science*, 47(5), 1869-1875.
- Lopez, J., and Tait, S. W. G. (2015). Mitochondrial apoptosis: Killing cancer using the enemy within. *British Journal of Cancer*, 112(6), 957.
- Lubis, I.A., Ichwan, M.F., Mustofa, M., Satria, D. Anticancer activity of *Eleutherine bulbosa* (Mill.) Urb. extract on WiDr cell line in vitro. In 2nd Public Health International Conference (PHICo 2017) December 2017, 123-127, Atlantis Press.
- Luo, T., and Kiick, K. L. (2017). Collagen-like peptide bioconjugates. *Bioconjugate Chemistry*, 28(3), 816-827.
- Lyons, F. G., Al-Munajjed, A. A., Kieran, S. M., Toner, M. E., Murphy, C. M., Duffy, G. P., and O'Brien, F. J. (2010). The healing of bony defects by cell-free collagen-based scaffolds compared to stem cell-seeded tissue engineered constructs. *Biomaterials*, 31(35), 9232-9243.
- Machana, S., Weerapreeyakul, N., Barusruks, S., Nonpunya, A., Sripanidkulchai, B., Thitimetharoch, T. (2011). Cytotoxic and apoptotic effects of six herbal plants against the human hepatocarcinoma (HepG2) cell line. *Chinese Medicine*, 6, 1-8.
- Mahabusarakam, W., Hemtasin, C., Chakthong, S., Voravuthikunchai, S. P., and Olawumi, I. B. (2010). Naphthoquinones, anthraquinones and naphthalene derivatives from the bulbs of *Eleutherine americana*. *Planta medica*, 76(04), 345-349.
- Malaysia National Cancer Registry, (2007-2012). Retrieved from <http://nci.moh.gov.my/index.php/ms/pengumuman/340-malaysian-national-cancer-registry-report-2007-2011>. Accessed on January 19th, 2019.
- Manach, C., Scalbert, A., Morand, C., Rémesy, C., and Jiménez, L. (2004). Polyphenols: Food sources and bioavailability. *The American journal of clinical nutrition*, 79(5), 727-747.
- Mansouri, N. (2016). The influence of topography on tissue engineering perspective. *Materials Science and Engineering: C*, 61, 906-921.
- Mantovani, F., Collavin, L., and Del Sal, G. (2019). Mutant p53 as a guardian of the cancer cell. *Cell Death and Differentiation*, 26(2), 199-212.
- Marquardt, J. U., Gomez-Quiroz, L., Camacho, L. O. A., Pinna, F., Lee, Y. H., Kitade, M., Domínguez, M. P., Castven, D., Breuhahn, K., Conner, E. A., Galle, P. R., Andersen, J. B., Factor, V. M., and Thorgeirsson, S. S.

- (2015). Curcumin effectively inhibits oncogenic NF- κ B signaling and restrains stemness features in liver cancer. *Journal of Hepatology*, 63(3), 661-669.
- Marquez, R. T., Tsao, B. W., Faust, N. F., and Xu, L. (2013). Drug resistance and molecular cancer therapy: Apoptosis versus autophagy. *Apoptosis*, 155-177.
- Martins, V. G., Costa, J. A. V., and Prentice-Hernández, C. (2009). Hidrolisado protéico de pescado obtido por vias química e enzimática a partir de corvina (*Micropogonias furnieri*). *Química Nova*, 32(1), 61-66.
- Masfria, M., and Tampubolon, M. S. (2019). The antifungal activity of n-hexane extract of *Eleutherine palmifolia* (L). Merr bulbs against *Candida albicans* and *Trichophyton mentagrophytes*. *Open access Macedonian Journal of Medical Sciences*, 7(22), 3777.
- Maus, M. V., and June, C. H. (2016). Making better chimeric antigen receptors for adoptive T-cell therapy. *Clinical Cancer Research*, 22(8), 1875-1884.
- MedlinePlus. (2020). Retrieved from <https://www.medlineplus.gov/antioxidants.html>. Accessed on July 22nd, 2020.
- Meerloo, J. V., Kaspers, G. J., & Cloos, J. (2011). Cell sensitivity assays: the MTT assay. In *Cancer cell culture* (pp. 237-245). Humana Press.
- Meyer, M. (2019). Processing of collagen-based biomaterials and the resulting materials properties. *Biomedical engineering online*, 18(1), 24.
- Miller, E. (2004). Apoptosis measurement by annexin v staining. In *Cancer cell culture* (pp. 191-202). Humana Press.
- Mitra, S., and Dash, R. (2018). Natural products for the management and prevention of breast cancer. *Evidence-Based Complementary and Alternative Medicine*, 2018.
- Moreira, M. M., da Silva, L. R. R., Mendes, T. A. D., Santiago, S. L., Mazzetto, S. E., Lomonaco, D., and Feitosa, V. P. (2018). Synthesis and characterization of a new methacrylate monomer derived from the cashew nut shell liquid (CNSL) and its effect on dentinal tubular occlusion. *Dental Materials*, 34(8), 1144-1153.
- Morin, P. J. (2003). Drug resistance and the microenvironment: Nature and nurture. *Drug Resistance Updates*, 6(4), 169-172.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. *Journal of Immunological Methods*, 65, 55-63.

- Mouw, J., Ou, G., and Weaver, V. M. (2014). Deconstructing extracellular matrix assembly: A multi-scale road map. *Nature Reviews Cell Molecular Biology*, 15, 771-785.
- Muijnieks, L. D., and Keeley, F. W. (2013). Molecular assembly and mechanical properties of the extracellular matrix: A fibrous protein perspective. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*, 1832(7), 866-875.
- Murphy, C. M., O'Brien, F. J. (2010). Understanding the effect of mean pore size on cell activity in collagen-glycosaminoglycan scaffolds. *Cell Adhesion and Migration*, 4(3), 377-381.
- Murphy, C. M., Haugh, M. G., and O'brien, F. J. (2010). The effect of mean pore size on cell attachment, proliferation and migration in collagen-glycosaminoglycan scaffolds for bone tissue engineering. *Biomaterials*, 31(3), 461-466.
- Murthy, R., Honavar, S. G., Naik, M. N., and Reddy, V. A. (2004). Retinoblastoma. *Modern Ophthalmology*. New Delhi, India, Jaypee Brothers, 849859.
- Murthy, R., Vemuganti, G. K., Honavar, S. G., Naik, M., and Reddy, V. (2009). Extramedullary leukemia in children presenting with proptosis. *Journal of Hematology and Oncology*, 2(1), 1-7.
- Mushtaq, M., Sultana, B., Bhatti, H. N., and Asghar, M. (2015). RSM based optimized enzyme-assisted extraction of antioxidant phenolics from underutilized watermelon (*Citrullus lanatus* Thunb.) rind. *Journal of Food Science and Technology*, 52(8), 5048- 5056.
- Mutiah, R., Choiroh, F., Annisa, R., Listiyana, A. Combinational effect of *Eleutherine palmifolia* (L.) Merr extract and doxorubicin chemotherapy on HeLa cervical cancer cells. In AIP Conference Proceedings, July 2019, (Vol. 2120, No. 1, p. 070001). AIP Publishing LLC.
- Mutiah, R., Listiyana, A., Arief Suryadinata, R. A., Hakim, A., Anggraini, W., and Susilowati, R. (2018). Activity of inhibit the cell cycle and induct apoptosis in HeLa cancer cell with combination of Sabrang onion (*Eleutherine palmifolia* (L.) Merr) and Starfruit Mistletoe (*Macrosolen cochinchinensis* (Lour.) Tiegh.). *Journal of Applied Pharmaceutical Science*, 8(10), 122-128.
- Mut-Salud, N., Álvarez, P. J., Garrido, J. M., Carrasco, E., Aránega, A., and Rodríguez-Serrano, F. (2016). Antioxidant intake and antitumor therapy: Toward nutritional recommendations for optimal results. *Oxidative Medicine and Cellular Longevity*, 2016.
- Naahidi, S., Jafari, M., Logan, M., Wang, Y., Yuan, Y., Bae, H., Dixon , B., and Chen, P. (2017). Biocompatibility of hydrogel-based scaffolds for tissue engineering applications. *Biotechnology Advances*, 35(5), 530-544.

- Nafisa, B. B. (2020). Kajian efek ekstrak umbi bawang dayak (*Eleutherine palmifolia* L. Merr) sebagai anti kanker: Studi literature (Doctoral dissertation, Universitas Islam Negeri Maulana Malik Ibrahim).
- Nagarajan, S., Nagarajan, R., Kumar, J., Salemme, A., Togna, A. R., Saso, L., and Bruno, F. (2020). Antioxidant activity of synthetic polymers of phenolic compounds. *Polymers*, 12(8), 1646.
- Nagata, S. (2018). Apoptosis and clearance of apoptotic cells. *Annual review of immunology*, 36, 489-517.
- Nageotte, J. (1927). *Coagulation fibrillaire in vitro du collagène dissous dans un acide dilué*. Gauthier-Villars.
- National Eye Database. Retrieved from <https://www.acrm.org.my/ned/retinoblastomaRegistry.html>. Accessed on January 22nd, 2021.
- Nawaiseh, I., Al-Hussaini, M., Alhamwi, A., Meyar, M., Sultan, I., Alrawashdeh, K., Jaradat, I. and Yousef, Y. A. (2015). The impact of growth patterns of retinoblastoma (endophytic, exophytic, and mixed patterns)/retinoblastomda büyümeye paternlerinin etkisi (endofitik, ekzofitik ve karışık paternler). *Turkish Journal of Pathology*, 31(1), 45-50.
- Nazari, S. S. (2020). Generation of 3D tumor spheroids with encapsulating basement membranes for invasion studies. *Current Protocols in Cell Biology*, 87(1), e105.
- Netti, P. A., Berk, D. A., Swartz, M. A., Grodzinsky, A. J., and Jain, R. K. (2000). Role of extracellular matrix assembly in interstitial transport in solid tumors. *Cancer Research*, 60(9), 2497-2503.
- Newman, D. J., and Cragg, G. M. (2016). Natural products as sources of new drugs from 1981 to 2014. *Journal of Natural Products*, 79(3), 629-661.
- Niture, S. K., and Jaiswal, A. K. (2012). Nrf2 protein up-regulates antiapoptotic protein Bcl-2 and prevents cellular apoptosis. *Journal of Biological Chemistry*, 287(13), 9873-9886.
- Nowacka, M., Sterzynska, K., Andrzejewska, M., Nowicki, M., and Januchowski, R. (2021). Drug resistance evaluation in novel 3D in vitro model. *Biomedicine & Pharmacotherapy*, 138, 111536.
- O'Brien, F. J., Harley, B. A., Yannas, I. V., and Gibson, L. (2004). Influence of freezing rate on pore structure in freeze-dried collagen-GAG scaffolds. *Biomaterials*, 25(6), 1077-1086.

- O'brien, F. J. (2011). Biomaterials & scaffolds for tissue engineering. *Materials Today*, 14(3), 88-95.
- Oda, E., Ohki, R., Murasawa, H., Nemoto, J., Shibue, T., Yamashita, T., Tokino, T., Taniguchi, T., and Tanaka, N. (2000). Noxa, a BH3-only member of the Bcl-2 family and candidate mediator of p53-induced apoptosis. *Science*, 288(5468), 1053-1058.
- Oetjen, G. W., and Haseley, P. (2004). *Freeze-drying*. John Wiley & Sons.
- Ohan, M. P., and Dunn, M. G. (2003). Glucose stabilizes collagen sterilized with gamma irradiation. *Journal of Biomedical Materials Research Part A: An Official Journal of The Society for Biomaterials, The Japanese Society for Biomaterials, and The Australian Society for Biomaterials and the Korean Society for Biomaterials*, 67(4), 1188-1195.
- Opferman, J. T., and Kothari, A. (2018). Anti-apoptotic BCL-2 family members in development. *Cell Death and Differentiation*, 25(1), 37-45.
- Ozben, T. (2015). Antioxidant supplementation on cancer risk and during cancer therapy: An update. *Current Topics in Medicinal Chemistry*, 15(2), 170-178.
- Paajanen, J., Laaksonen, S., Kettunen, E., Ilonen, I., Vehmas, T., Salo, J., Räsänen, J., Sutinen, E., Ollila, H., Mäyränpää, M. I., Mylläniemi, M., and Wolff, H. (2020). Histopathological features of epithelioid malignant pleural mesotheliomas in patients with extended survival. *Human Pathology*, 98, 110-119.
- Paez-Escamilla, M., Bagheri, N., and Harbour, J. W. (2018). Retinoblastoma with endophytic and exophytic features. *Jama Ophthalmology*, 136(1), e175064-e175064.
- Pandey, A. N. (2014). Retinoblastoma: An overview. *Saudi Journal of Ophthalmology*, 28(4), 310-315.
- Pandey, A., Belwal, T., Sekar, K. C., Bhatt, I. D., and Rawal, R. S. (2018). Optimization of ultrasonic-assisted extraction (UAE) of phenolics and antioxidant compounds from rhizomes of *Rheum moorcroftianum* using response surface methodology (RSM). *Industrial Crops and Products*, 119, 218-225.
- Parenteau-Bareil, R., Gauvin, R., and Berthod, F. (2010). Collagen-based biomaterials for tissue engineering applications. *Materials*, 3(3), 1863-1887.
- Parkin, D. M., Stiller, C. A., Draper, G. J., and Bieber, C. A. (1988). The international incidence of childhood cancer. *International Journal of Cancer*, 42(4), 511-520.

- Persadmehr, A., Torneck, C. D., Cvitkovitch, D. G., Pinto, V., Talior, I., Kazembe, M., Shrestha, S., McCulloch, C. A., and Kishen, A. (2014). Bioactive chitosan nanoparticles and photodynamic therapy inhibit collagen degradation in vitro. *Journal of Endodontics*, 40(5), 703-709.
- Peyman, G. A., and Apple, D. J. (1974). Local excision of choroidal malignant melanoma: Full-thickness eye wall resection. *Archives of Ophthalmology*, 92(3), 216-218.
- Pfeffer, C., and Singh, A. (2018). Apoptosis: A target for anticancer therapy. *International Journal of Molecular Sciences*, 19(2), 448.
- Pimentel, M. F., and Neto, B. B. (1996). Calibração: Uma revisão para químicos analíticos. *Química Nova*, 19(3), 268-277.
- Popgeorgiev, N., Jabbour, L., and Gillet, G. (2018). Subcellular localization and dynamics of the Bcl-2 family of proteins. *Frontiers in Cell and Developmental Biology*, 6, 13.
- Pospíšil, P., Prasad, A., and Rác, M. (2019). Mechanism of the formation of electronically excited species by oxidative metabolic processes: role of reactive oxygen species. *Biomolecules*, 9(7), 258.
- Povea-Cabello, S., Oropesa-Ávila, M., la Cruz-Ojeda, D., Villanueva-Paz, M., De la Mata, M., Suárez-Rivero, J. M., Álvarez-Córdoba, M., Villalón-García, I., Cotán, D., Ybot-González, P., and Sánchez-Alcázar, J. A. (2017). Dynamic reorganization of the cytoskeleton during apoptosis: The two coffins hypothesis. *International journal of molecular sciences*, 18(11), 2393.
- Pozarowski, P., and Darzynkiewicz, Z. (2004). Analysis of cell cycle by flow cytometry. In Checkpoint controls and cancer (pp. 301-311). Humana Press.
- Prasad Sah, K., Saiju, R., Roy, P., and Kafle, S. (2013). Retinoblastoma: Ten years' experience at Kanti Children's Hospital. *Journal of the Nepal Medical Association*, 52(192).
- Prasad, S., Gupta, S. C., and Tyagi, A. K. (2017). Reactive oxygen species (ROS) and cancer: Role of antioxidative nutraceuticals. *Cancer Letters*, 387, 95-105.
- Prayong, P., Barusrux, S., and Weerapreeyakul, N. (2008). Cytotoxic activity screening of some indigenous Thai plants. *Fitoterapia*, 79(7-8), 598-601.
- Prestes, R. C. (2013). Colágeno e seus derivados: características e aplicações em produtos cárneos. *Journal of Health Sciences*, 15(1).
- Purcel, G., Melită, D., Andronescu, E. and Grumezescu, A. M. (2016). Collagen-based nanobiomaterials: challenges in soft tissue engineering.

In *Nanobiomaterials in Soft Tissue Engineering* (pp. 173-200). William Andrew Publishing.

- Puspadewi, R., Adirestuti, P., and Menawati, R. (2013). Efficacy Dayak onion (*Eleutherine palmifolia* (L.) Merr.) bulbs as antimicrobials herbal skin. *Kartika Scientific Journal of Pharmacy*, 1(1), 31-37.
- Qiao, J., Gu, C., Shang, W., Du, J., Yin, W., Zhu, M., Wang, W., Han, M., and Lu, W. (2011). Effect of green tea on pharmacokinetics of 5-fluorouracil in rats and pharmacodynamics in human cell lines in vitro. *Food and Chemical Toxicology*, 49(6), 1410-1415.
- Rahman, K. (2007). Studies on free radicals, antioxidants, and co-factors. *Clinical Interventions in Aging*, 2(2), 219.
- Rai, P., Shah, I. A., Narsani, A. K., Lohana, M. K., Memon, M. K., and Memon, M. A. (2009). Too late presentation of 53 patients with retinoblastoma. *challenge*, 9(2), 227-230.
- Rajan, N., Habermehl, J., Coté, M. F., Doillon, C. J., and Mantovani, D. (2006). Preparation of ready-to-use, storable and reconstituted type I collagen from rat tail tendon for tissue engineering applications. *Nature protocols*, 1(6), 2753-2758.
- Ramshaw, J. A. (2016). Biomedical applications of collagens. *Journal of Biomedical Materials Research Part B: Applied Biomaterials*, 104(4), 665-675.
- Ran, X. G., and Wang, L. Y. (2014). Use of ultrasonic and pepsin treatment in tandem for collagen extraction from meat industry by-products. *Journal of the Science of Food and Agriculture*, 94(3), 585-590.
- Rani, V. S. (2017). In vitro cytotoxic activity and preliminary phytochemical analysis of the crude extracts of *Eleutherine bulbosa* (Miller). *Urban World Journal of Pharmaceutical Research*, 7(4), 1022-1029.
- Raspanti, M., Reguzzoni, M., Protasoni, M. and Basso, P. (2018). Not only tendons: The other architecture of collagen fibrils. *International Journal of Biological Macromolecules*, 107, 1668- 1674.
- Ravi, M., Paramesh, V., Kaviya, S. R., Anuradha, E., and Solomon, F. P. (2015). 3D cell culture systems: Advantages and applications. *Journal of Cellular Physiology*, 230(1), 16-26.
- Re, R., Pellegrini, N., Proteggente, A., Pannala, A., Yang, M., and Rice-Evans, C. (1999). Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical Biology and Medicine*, 26(9-10), 1231-1237.

- Reddy, S. C., and Anusya, S. (2010). Clinical presentation of retinoblastoma in Malaysia: A review of 64 patients. *International Journal of Ophthalmology*, 3(1), 64.
- Reese, A. B. (1976). *Tumors of the Eye* (Vol. 50). Medical Department, Harper & Row.
- Rittié, L. (2017). Type I Collagen Purification from Rat Tail Tendons. In *Fibrosis* (pp. 287-308). Humana Press, New York, NY.
- Rivella, J. (2000). *Plantas da Amazonia: Oportunidades Económicas e sustentaveis*. Manaus: Sebrae-Am/Inpa, 2000. 405p. Sebrae-Am/Inpa.
- Rodriguez-Galindo, C., Orbach, D. B., and VanderVeen, D. (2015). Retinoblastoma. *Paediatric Clinics*, 62(1), 201-223.
- Rodriguez-Galindo, C., Wilson, M. W., Chantada, G., Fu, L., Qaddoumi, I., Antoneli, C., Leal-Leal, C., Sharma, T., Barnoya, M., Epelman, S., Pizzarello, L., Kane, J. R., Barfield, R., Merchant, T. E., Robison, L. L., Murphree, A. L., Chevez-Barrios, P., Dyer, M. A., O'Brien, J., Ribeiro, R. C., Hungerford, J., Helveston, E. M., Haik, B. G., and Wilimas, J. (2008). Retinoblastoma: one world, one vision. *Pediatrics*, 122(3), e763-e770.
- Rodríguez-Pérez, C., Quirantes-Piné, R., Fernández-Gutiérrez, A., and Segura-Carretero, A. (2015). Optimization of extraction method to obtain a phenolic compounds-rich extract from *Moringa oleifera* Lam leaves. *Industrial Crops and Products*, 66, 246-254.
- Roselló-Soto, E., Martí-Quijal, F. J., Cilla, A., Munekata, P. E., Lorenzo, J. M., Remize, F., and Barba, F. J. (2019). Influence of temperature, solvent and pH on the selective extraction of phenolic compounds from tiger nuts by-products: Triple-TOF-LC-MS-MS characterization. *Molecules*, 24(4), 797.
- Rosso, N. D., Daguer, H., Valese, A. C., and Granato, D. (2016). Effects of time and extraction temperature on phenolic composition and functional properties of red rooibos (*Aspalathus linearis*). *Food Research International*, 89, 476-487.
- Rotblat, B., Melino, G., and Knight, R. A. (2012). NRF2 and p53: Januses in cancer? *Oncotarget*, 3(11), 1272.
- Rotblat, B., Southwell, A. L., Ehrnhoefer, D. E., Skotte, N. H., Metzler, M., Franciosi, S., Leprivier, G., Somasekharan, S. P., Barokas, A., Deng, Y., Tang, T., Mathers, J., Cetinbas, N., Daugaard, M., Kwok, B., Li, L., Carnie, C. J., Fink, D., Nitsch, R., Galpin, J. D., Ahern, C. A., Melino, G., Penninger, J. M., Hayden, M. R., and Poulsen, H. Sorensen. (2014). HACE1 reduces oxidative stress and mutant Huntington toxicity by promoting the NRF2 response. *Proceedings of the National Academy of Sciences*, 111(8), 3032-3037.

- Roy, A. S., Tripathy, D. R., Samanta, S., Ghosh, S. K., and Dasgupta, S. (2016). DNA damaging, cell cytotoxicity and serum albumin binding efficacy of the rutin–Cu (II) complex. *Molecular BioSystems*, 12(5), 1687-1701.
- Roy, J., Galano, J. M., Durand, T., Le Guennec, J. Y., and Chung-Yung Lee, J. (2017). Physiological role of reactive oxygen species as promoters of natural defenses. *The FASEB Journal*, 31(9), 3729-3745.
- Sabharwal, S. S., and Schumacker, P. T. (2014). Mitochondrial ROS in cancer: Initiators, amplifiers or an Achilles' heel?. *Nature Reviews Cancer*, 14(11), 709-721.
- Sachlos, E., Reis, N., Ainsley, C., Derby, B., and Czernuszka, J. T. (2003). Novel collagen scaffolds with predefined internal morphology made by solid freeform fabrication. *Biomaterials*, 24(8), 1487-1497.
- Sagoo, M. S., Shields, C. L., Mashayekhi, A., Freire, J., Emrich, J., Reiff, J., Komarnicky, L. and Shields, J. A. (2011). Plaque radiotherapy for juxtapapillary choroidal melanoma: Tumour control in 650 consecutive cases. *Ophthalmology*, 118(2), 402-407.
- Sahin, K., Tuzcu, M., Gencoglu, H., Dogukan, A., Timurkan, M., Sahin, N., Aslan, A., and Kucuk, O. (2010). Epigallocatechin-3-gallate activates Nrf2/HO-1 signaling pathway in cisplatin-induced nephrotoxicity in rats. *Life sciences*, 87(7-8), 240-245.
- Sahu, S., Banavali, S. D., Pai, S. K., Nair, C. N., Kurkure, P. A., Motwani, S. A., and Advani, S. H. (1998). Retinoblastoma: Problems and perspectives from India. *Pediatric hematology and oncology*, 15(6), 501-508.
- Said, K. A. M., and Amin, M. A. M. (2015). Overview on the response surface methodology (RSM) in extraction processes. *Journal of Applied Science & Process Engineering*, 2(1).
- Saldanha, S. N., and Tollefsbol, T. O. (2012). The role of nutraceuticals in chemoprevention and chemotherapy and their clinical outcomes. *Journal of Oncology*, 2012.
- Samimi, H., Sohi, A. N., Irani, S., Arefian, E., Mahdiannasser, M., Fallah, P., and Haghpanah, V. (2021). Alginate-based 3D cell culture technique to evaluate the half-maximal inhibitory concentration: an in vitro model of anticancer drug study for anaplastic thyroid carcinoma. *Thyroid Research*, 14(1), 1-9.
- Santos Araújo, M. D. C., Farias, I. L., Gutierrez, J., Dalmora, S. L., Flores, N., Farias, J., De Cruz, I., Chiesa, J., Morsch, V. M., and Chitolina Schetinger, M. R. (2012). *Uncaria tomentosa*—adjuvant treatment for breast cancer: Clinical trial. *Evidence-Based Complementary and Alternative Medicine*, 2012.

- Santos, J. S., Deolindo, C. T. P., Esmerino, L. A., Genovese, M. I., Fujita, A., Marques, M. B., Rosso, N. D., Daguer, H., Valese, A. C., Granato, D. (2016). Effects of time and extraction temperature on phenolic composition and functional properties of red rooibos (*Aspalathus linearis*). *Food Research International*, 89, 476–487.
- Scarano, A., Chieppa, M., and Santino, A. (2018). Looking at flavonoid biodiversity in horticultural crops: A colored mine with nutritional benefits. *Plants*, 7(4), 98.
- Scelfo, C., Francis, J. H., Khetan, V., Jenkins, T., Marr, B., Abramson, D. H., Shields, C.L., Pe'er, J., Munier, F., Berry, J., Harbour, W., Yarovoy, A., Lucena, E., Murray, T. G., Bhagia, P., Paysse, E., Tuncer, S., Chantada, G. L., Moll, A. C., Ushakova, T., Plager, D. A., Ziyovuddin, I., Leal, C. A., Materin, M. A., Ji, X., Cursino, J. W., Polania, R., Kiratli, H., All-Ericsson, C., Kebudi, R., Honavar, H. G., Vishnevskia-Dai, V., Epelman, S., Daniels, A. B., Ling, J. D., Traore, F. and Ramirez-Ortiz, M. A. (2017). An international survey of classification and treatment choices for group D retinoblastoma. *International Journal of Ophthalmology*, 10(6), 961.
- Schajquevich, P., Francis, J. H., Cancela, M. B., Carcaboso, A. M., Chantada, G. L., & Abramson, D. H. (2022). Treatment of Retinoblastoma: What Is the Latest and What Is the Future. *Frontiers in Oncology*, 12.
- Schieber, M. and Chandel, N. S. (2014). ROS function in redox signalling and oxidative stress. *Current Biology*, 24(10), R453-R462.
- Schmidt, M. M., Dornelles, R. C. P., Mello, R. O., Kubota, E. H., Mazutti, M. A., Kempka, A. P., and Demiate, I. M. (2016). Collagen extraction process. *International Food Research Journal*, 23(3).
- Schoof, H., Apel, J., Heschel, I., and Rau, G. (2001). Control of pore structure and size in freeze-dried collagen sponges. *Journal of Biomedical Materials Research: An Official Journal of The Society for Biomaterials, The Japanese Society for Biomaterials, and The Australian Society for Biomaterials and the Korean Society for Biomaterials*, 58(4), 352- 357.
- Schrieber, R., and Gareis, H. (2007). *Gelatine handbook: theory and industrial practice*. John Wiley & Sons.
- Sebens, S., and Schafer, H. (2012). The tumor stroma as mediator of drug resistance-a potential target to improve cancer therapy?. *Current Pharmaceutical Biotechnology*, 13(11), 2259-2272.
- Selby, M., Delosh, R., Laudeman, J., Ogle, C., Reinhart, R., Silvers, T., Lawrence, S., Kinders, R., Parchment, R., Teicher, B. A., and Evans, D. M. (2017). 3D models of the NCI60 cell lines for screening oncology compounds. *SLAS DISCOVERY: Advancing Life Sciences R&D*, 22(5), 473-483.

- Sengupta, S., Pan, U., and Khetan, V. (2016). Adult-onset retinoblastoma. *Indian Journal of Ophthalmology*, 64(7), 485.
- Seregard, S., Lundell, G., Svedberg, H., and Kivelä, T. (2004). Incidence of retinoblastoma from 1958 to 1998 in Northern Europe: advantages of birth cohort analysis. *Ophthalmology*, 111(6), 1228-1232.
- Shalini, S., Dorstyn, L., Dawar, S., and Kumar, S. (2015). Old, new and emerging functions of caspases. *Cell Death & Differentiation*, 22(4), 526-539.
- Shi, P., Du, W., Wang, Y., Teng, X., Chen, X., and Ye, L. (2019). Total phenolic, flavonoid content, and antioxidant activity of bulbs, leaves, and flowers made from *Eleutherine bulbosa* (Mill.) Urb. *Food Science and Nutrition*, 7(1), 148-154.
- Shields, C. L., Mashayekhi, A., Cater, J., Shelil, A., Meadows, A. T., and Shields, J. A. (2004). Chemoreduction for retinoblastoma. Analysis of tumor control and risks for recurrence in 457 tumors. *American Journal of Ophthalmology*, 138(3), 329-337.
- Shields, C. L., Shields, J. A., and Shah, P. (1991). Retinoblastoma in older children. *Ophthalmology*, 98(3), 395-399.
- Shintani, H. (2017). Ethylene oxide gas sterilization of medical devices. *Biocontrol Science*, 22(1), 1-16.
- Shu, Y. Z. (1998). Recent natural products based drug development: A pharmaceutical industry perspective. *Journal of Natural Products*, 61(8), 1053-1071.
- Sies, H. and Jones, D. P. (2020). Reactive oxygen species (ROS) as pleiotropic physiological signalling agents. *Nature reviews Molecular cell biology*, 21(7), 363-383.
- Simbulan-Rosenthal, C. M., Rosenthal, D. S., Luo, R., and Smulson, M. E. (1999). Poly (ADP-ribosylation) of p53 during apoptosis in human osteosarcoma cells. *Cancer Research*, 59(9), 2190-2194.
- Simon, J. W., and Kaw, P. (2001). Commonly missed diagnoses in the childhood eye examination. *American Family Physician*, 64(4), 623-630.
- Singh, A. V. (2020). Commentary on "Peptide-conjugated nanoparticles as targeted anti-angiogenesis therapeutic and diagnostic in cancer" by Shaker A. Mousa, pharmaceutical research institute, Albany college of pharmacy and health sciences, Rensselaer, NY 12144, United States-peptide-conjugated nanoparticles for multimodal nanomedicine. *Current Medicinal Chemistry*, 27(17), 2927-2928.
- Singh, K., Bhori, M., Kasu, Y. A., Bhat, G., and Marar, T. (2018). Antioxidants as precision weapons in war against cancer chemotherapy induced toxicity—

- Exploring the armoury of obscurity. *Saudi Pharmaceutical Journal*, 26(2), 177-190.
- Singh, R., Letai, A., and Sarosiek, K. (2019). Regulation of apoptosis in health and disease: The balancing act of BCL-2 family proteins. *Nature reviews Molecular cell biology*, 20(3), 175-193.
- Singh, S. B., Adam, A. G., Tripathy, N., Lee, D., and Khang, G. (2018). Reactive oxygen species responsive naturally occurring phenolic-based polymeric prodrug. In *Cutting-Edge Enabling Technologies for Regenerative Medicine* (pp. 291-301). Springer, Singapore.
- Sorushanova, A., Delgado, L. M., Wu, Z., Shologu, N., Kshirsagar, A., Raghunath, R., Mullen, A. M., Bayon, Y., Pandit, A., Raghunath, M., and Zeugolis, D. I. (2019). The collagen suprafamily: From biosynthesis to advanced biomaterial development. *Advanced Materials*, 31(1), 1801651.
- Subramaniam, S., Rahmat, J., Rahman, N. A., Ramasamy, S., Bhoo-Pathy, N., Pin, G. P., and Alagaratnam, J. (2014). Presentation of retinoblastoma patients in Malaysia. *Asian Pacific Journal of Cancer Prevention*, 15(18), 7863-7.
- Suffness, M. (1990). Assays related to cancer drug discovery. Methods in plant biochemistry: Assays for bioactivity. London, Academic Press. 6, 71-133.
- Synopsis of childhood cancer incidence in Malaysia. (2007-2011). Retrieved from <http://nci.moh.gov.my/index.php/ms/pengumuman/340-malaysian-national-cancer-registry-report-2007-2011>. Accessed on January 21th, 2019.
- Sznarkowska, A., Kostecka, A., Meller, K., and Bielawski, K. P. (2017). Inhibition of cancer antioxidant defense by natural compounds. *Oncotarget*, 8(9), 15996.
- Tait, S. W. and Green, D. R. (2013). Mitochondrial regulation of cell death. *Cold Spring Harbor Perspectives in Biology*, 5(9), a008706.
- Tan, B. L., Norhaizan, M. E., and Chan, L. C. (2018). *Manilkara zapota* (L.) P. Royen Leaf Water Extract Induces Apoptosis in Human Hepatocellular Carcinoma (HepG2) Cells via ERK1/2/Akt1/JNK1 Signaling Pathways. *Evidence-Based Complementary and Alternative Medicine*, 2018.
- Thornborrow, E. C., Patel, S., Mastropietro, A. E., Schwartzfarb, E. M., and Manfredi, J. J. (2002). A conserved intronic response element mediates direct p53-dependent transcriptional activation of both the human and murine bax genes. *Oncogene*, 21(7), 990-999.

- Thun, M. J., DeLancey, J. O., Center, M. M., Jemal, A., and Ward, E. M. (2009). The global burden of cancer: priorities for prevention. *Carcinogenesis*, 31(1), 100-110.
- Tor, Y. S., Yazan, L. S., Foo, J. B., Wibowo, A., Ismail, N., Cheah, Y. K., Abdullah, R., Ismail, M., Ismail, I. S., Yeap, S. K. (2015). Induction of apoptosis in MCF-7 cells via oxidative stress generation, mitochondria-dependent and caspase-independent pathway by ethyl acetate extract of *Dillenia suffruticosa* and its chemical profile. *PLoS One*, 10(6), e0127441.
- Trédan, O., Galmarini, C. M., Patel, K., and Tannock, I. F. (2007). Drug resistance and the solid tumor microenvironment. *Journal of the National Cancer Institute*, 99(19), 1441-1454.
- Tsuchiya, A., Kaku, Y., Nakano, T., and Nishizaki, T. (2015). Diarachidonoylphosphoethanolamine induces apoptosis of malignant pleural mesothelioma cells through a Trx/ASK1/p38 MAPK pathway. *Journal of Pharmacological Sciences*, 129(3), 160-168.
- Ullah, R., Khan, M., Shah, S. A., Saeed, K., and Kim, M. O. (2019). Natural antioxidant anthocyanins—A hidden therapeutic candidate in metabolic disorders with major focus in neurodegeneration. *Nutrients*, 11(6), 1195.
- Usmanov, R. H., and Kivelä, T. (2014). Predicted trends in the incidence of retinoblastoma in the Asia-Pacific region. *The Asia-Pacific Journal of Ophthalmology*, 3(3), 151-157.
- Vaseva, A. V., and Moll, U. M. (2009). The mitochondrial p53 pathway. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, 1787(5), 414-420.
- Walczak, H., and Krammer, P. H. (2000). The CD95 (APO-1/Fas) and the TRAIL (APO-2L) apoptosis systems. *Experimental Cell Research*, 256(1), 58-66.
- Wang, F., Xie, C., Ren, N., Bai, S., and Zhao, Y. (2019). Human Freeze-dried Dentin Matrix as a Biologically Active Scaffold for Tooth Tissue Engineering. *Journal of Endodontics*, 45(11), 1321-1331.
- Wang, G., Lei, Z., Zhong, Q., Wu, W., Zhang, H., Min, T., Wu, H., Lai, F. (2017). Enrichment of caffeic acid in peanut sprouts and evaluation of its in vitro effectiveness against oxidative stress-induced erythrocyte haemolysis. *Food Chemistry*, 217, 332–341.
- Wang, Q., Wan, J., Zhang, W., and Hao, S. (2019). MCL-1 or BCL-xL-dependent resistance to the BCL-2 antagonist (ABT-199) can be overcome by specific inhibitor as single agents and in combination with ABT- 199 in acute myeloid leukemia cells. *Leukemia and Lymphoma*, 60(9), 2170-2180.

- Wang, T., Guo, N., Wang, S. X., Kou, P., Zhao, C. J., and Fu, Y. J. (2018). Ultrasound negative pressure cavitation extraction of phenolic compounds from blueberry leaves and evaluation of its DPPH radical scavenging activity. *Food and Bioproducts Processing*, 108, 69-80.
- Wang, X. Y., Wang, Q. H., He, Y., and Wang, H. (2015). Advances in studies on chemical constituents and pharmacological activities of *Eleutherine americana*. *Asia-Pacific Traditional Medicine*, 11, 39-42.
- Weinberg, R. A., and Hanahan, D. (2000). The hallmarks of cancer. *Cell*, 100(1), 57-70.
- WHO Cancer Fact Sheet. Updated September 2018. Retrieved from <https://www.who.int/news-room/fact-sheets/detail/cancer>. Accessed on January 18th, 2019.
- Wiangnon, S., Kamsa-ard, S., Jetsrisuparb, A., Sriplung, H., Sontipong, S., Sumitsawan, Y., and Martin, N. (2003). Childhood cancer in Thailand: 1995-1997. *Asian Pacific Journal of Cancer Prevention*, 4(4), 337-343.
- Wilken, R., Veena, M. S., Wang, M. B., and Srivatsan, E. S. (2011). Curcumin: A review of anti-cancer properties and therapeutic activity in head and neck squamous cell carcinoma. *Molecular cancer*, 10(1), 12.
- Willoughby, C. E., Batterbury, M., and Kaye, S. B. (2002). Collagen corneal shields. *Survey of Ophthalmology*, 47(2), 174-182.
- Witek-Krowiak, A., Chojnacka, K., Podstawczyk, D., Dawiec, A., and Pokomeda, K. (2014). Application of response surface methodology and artificial neural network methods in modelling and optimization of biosorption process. *Bioresource technology*, 160, 150-160.
- World Health Organization. Medicinal Plants in Viet Nam; WHO Regional Office for the Western Pacific: Manila, Philippines, 1990.
- Wu, S., Lu, H., and Bai, Y. (2019). Nrf2 in cancers: A double-edged sword. *Cancer Medicine*, 8(5), 2252-2267.
- Xu, J., Qiu, F., Duan, W., Qu, G., Wang, N., and Yao, X. (2006). New bioactive constituents from *Eleutherine americana*. *Frontiers of Chemistry in China*, 1(3), 320-323.
- Yamada, K. M., and Cukierman, E. (2007). Modeling tissue morphogenesis and cancer in 3D. *Cell*, 130(4), 601-610.
- Yamamoto, S., and Iwakuma, T. (2019). Regulators of oncogenic mutant TP53 gain of function. *Cancers*, 11(1), 4.

- Yang, B., Chen, Y., and Shi, J. (2019). Reactive oxygen species (ROS)-based nanomedicine. *Chemical reviews*, 119(8), 4881-4985.
- Yang, J., Manson, D. K., Marr, B. P., & Carvajal, R. D. (2018). Treatment of uveal melanoma: where are we now?. *Therapeutic Advances in Medical Oncology*, 10,1-17.
- Yang, M., and Wei, W. (2019). Long non-coding RNAs in retinoblastoma. *Pathology-Research and Practice*, 215(8), 152435.
- Yang, X. B., Bhatnagar, R. S., Li, S., and Oreffo, R. O. (2004). Biomimetic collagen scaffolds for human bone cell growth and differentiation. *Tissue engineering*, 10(7-8), 1148-1159.
- Yanık, Ö., Gündüz, K., Yavuz, K., Taçyıldız, N., and Ünal, E. (2015). Chemotherapy in retinoblastoma: Current approaches. *Turkish journal of Ophthalmology*, 45(6), 259.
- Yanumula, A., and Cusick, J. K. (2020). Biochemistry, Extrinsic Pathway of Apoptosis. In *StatPearls [Internet]*. StatPearls Publishing.
- Zaman, S., Wang, R., and Gandhi, V. (2014). Targeting the apoptosis pathway in hematologic malignancies. *Leukemia and lymphoma*, 55(9), 1980-1992.
- Zargar, A., Chang, S., Kothari, A., Snijders, A. M., Mao, J. H., Wang, J., Hernandez, A. C., Keasling, J. D., and Bivona, T. G. (2019). Overcoming the challenges of cancer drug resistance through bacterial-mediated therapy. *Chronic Diseases and Translational medicine*, 5(04), 258-266.
- Zhang, G., Wang, Y., Zhang, Y., Wan, X., Li, J., Liu, K., Liu, Q., Yang, C., Yu, P., Huang, Y., Wang, S., Jiang, P., Qu, Z., Luan, J., Duan, H., Zhang, L., Hou, A., Jin, S., Hsieh, T. C., and Wu, E. (2012). Anti-cancer activities of tea epigallocatechin-3-gallate in breast cancer patients under radiotherapy. *Current Molecular Medicine*, 12(2), 163-176.
- Zhang, H., Birch, J., Pei, J., Mohamed Ahmed, I. A., Yang, H., Dias, G., El-Aty, A. M. A., and Bekhit, A. E. D. (2019). Identification of six phytochemical compounds from *Asparagus officinalis* L. root cultivars from New Zealand and China using UAE-SPE-UPLC-MS/MS: Effects of extracts on H₂O₂-induced oxidative stress. *Nutrients*, 11(1), 107.
- Zhang, L., Zeng, Y., and Cheng, Z. (2016). Removal of heavy metal ions using chitosan and modified chitosan: A review. *Journal of Molecular Liquids*, 214, 175-191.
- Zhang, Y. S., and Khademhosseini, A. (2017). Advances in engineering hydrogels. *Science*, 356(6337), eaaf3627.

- Zhang, Y., Chen, X., Gueydan, C., and Han, J. (2018). Plasma membrane changes during programmed cell deaths. *Cell research*, 28(1), 9.
- Zhang, Y., Duan, S., Jang, A., Mao, L., Liu, X., and Huang, G. (2021). JQ1, a selective inhibitor of BRD4, suppresses retinoblastoma cell growth by inducing cell cycle arrest and apoptosis. *Experimental Eye Research*, 202, 108304.
- Zhao, L., Wang, H., and Du, X. (2021). The therapeutic use of quercetin in ophthalmology: Recent applications. *Biomedicine and Pharmacotherapy*, 137, 111371.
- Zheng, J., Zhou, Y., Li, Y., Xu, D. P., Li, S., and Li, H. B. (2016). Spices for prevention and treatment of cancers. *Nutrients*, 8(8), 495.
- Zheng, T. S., Schlosser, S. F., Dao, T., Hingorani, R., Crispe, I. N., Boyer, J. L., and Flavell, R. A. (1998). Caspase-3 controls both cytoplasmic and nuclear events associated with Fas-mediated apoptosis in vivo. *Proceedings of the National Academy of Sciences*, 95(23), 13618-13623.
- Zhou, H., Zou, P., Chen, Z. C., and You, Y. (2007). A novel vicious cycle cascade in tumor chemotherapy. *Medical Hypotheses*, 69(6), 1230-1233.
- Zhou, Y., Li, Y., Zhou, T., Zheng, J., Li, S., and Li, H. B. (2016). Dietary natural products for prevention and treatment of liver cancer. *Nutrients*, 8(3), 156.
- Zhou, Y., Zheng, J., Li, Y., Xu, D. P., Li, S., Chen, Y. M., and Li, H. B. (2016). Natural polyphenols for prevention and treatment of cancer. *Nutrients*, 8(8), 515.
- Zidan, G., Rupenthal, I. D., Greene, C., and Seyfoddin, A. (2018). Medicated ocular bandages and corneal health: Potential excipients and active pharmaceutical ingredients. *Pharmaceutical Development and Technology*, 23(3), 255-260.
- Zietarska, M., Maugard, C. M., Filali-Mouhim, A., Alam-Fahmy, M., Tonin, P. N., Provencher, D. M., and Mes-Masson, A. M. (2007). Molecular description of a 3D in vitro model for the study of epithelial ovarian cancer (EOC). *Molecular Carcinogenesis*, 46(10), 872–885.
- Zschenker, O., Streichert, T., Hehlsgans, S., and Cordes, N. (2012). Genome-wide gene expression analysis in cancer cells reveals 3D growth to affect ECM and processes associated with cell adhesion but not DNA repair. *PloS one*, 7(4), e34279.