

UNIVERSITI PUTRA MALAYSIA

STUDY ON THE DEVELOPMENT OF EMBRYO AND LARVA OF THE HUMPBACK GROUPER (CROMILEPTES ALTIVELIS VALENCEE) AND ITS FEEDING BEHAVIOUR

USMAN BULANIN

FP 2002 2

STUDY ON THE DEVELOPMENT OF EMBRYO AND LARVA OF THE HUMPBACK GROUPER (CROMILEPTES ALTIVELIS VALENCEE) AND ITS FEEDING BEHAVIOUR

USMAN BULANIN

DOCTOR OF PHILOSOPHY UNIVERSITI PUTRA MALAYSIA

2002



STUDY ON THE DEVELOPMENT OF EMBRYO AND LARVA OF THE HUMPBACK GROUPER (CROMILEPTES ALTIVELIS VALENCEE) AND ITS FEEDING BEHAVIOUR

By

USMAN BULANIN

Thesis Submitted in Fulfilment of the Requirement for the Degree of Doctor of Philosophy in the Faculty of Agriculture Universiti Putra Malaysia

February 2002



DEDICATION

To my parents Bulanin (alm.) and Hj. Mariawan, my wife Misuharti and my child An-nisa Usman who gave me supports and understandings during my study in Malaysia



Abstract of thesis submitted to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

STUDY ON THE DEVELOPMENT OF EMBRYO AND LARVA OF THE HUMPBACK GROUPER (CROMILEPTES ALTIVELIS VALENCEE) AND ITS FEEDING BEHAVIOUR

By

USMAN BULANIN

February 2002

Chairman : Che Roos Saad, Ph.D Faculty : Faculty of Agriculture

A series of experiments were conducted to determine 1) the embryonic and larval development, 2) feeding performance and 3) weaning of humpback grouper, *Cromileptes altivelis*, larva.

The above study was carried out from spawning of broodstocks to obtain the fertilized eggs till they hatched. The results showed that the mean diameter of humpback grouper egg was $828.69 \pm 45.91 \,\mu$ m. The fertilized eggs hatched in 20 hour 10 minutes at the temperature of 27 to 28 °C. The hatched larvae had a mean length of 1.86 mm, and a mean height of 0.460 mm. The yolk sacs and oil droplets were totally utilized after 63 and 65 hour after hatching (HAH), respectively. Within one day, the larvae had an average length and height of 2.253 mm and 0.615 mm, respectively. At 49 DAH, the mean length and height increased to 27.153 mm and 9.190 mm respectively.



The spines on the larvae began to appear at day 7 to day 10. The dorsal spines reached their maximum length at day 28 to day 30, and the spines had a maximum mean length of 5668.30 μ m, while the ventral spines had a maximum mean length of 4415.44 μ m at day 25 – 28. Then the spines began to reduce and change to hard rays as the larvae grew.

The eyes pigments started to be visible at day 3 to day 4 after hatching (DAH). The larvae's mouths were opened for the first time at day 2 or 45 HAH. The relationship of mouth gape and eyes diameter was quadratic with an equation of MG = $-02E-05Ed^2 + 0.3992Ed + 215.05$ and $R^2 = 0.96$. Appearance of black spots at the caudal peduncle, dorsal fin and anal fin occurred between day 25 to day 30. These spots would increase in numbers and would spread towards the head and became perfected at day 40 to day 45, and they would look similar to the juvenile fish.

The teeth of humpback grouper fish larvae began to appear at day 15, but those teeth were still few in numbers and were short. The height and width of larvae teeth increased as the larvae grew. At day 30, the larvae teeth got closer with a mean height and width of 110.18 μ m and 40.13 μ m respectively.

Histologically, no fold occurred in the larvae digestive tract yet before they reach 5 days old. The incipient stomach began to be visible at day 30. The larvae's stomachs were perfected at 45 DAH. The liver began to be visible at day 5, while the pancreas was visible at day 10. The protease enzyme activity increased as the larvae grew. Food was first found in the larvae's digestive tracts at day 3 or 66 HAH. The average number of total food found in the larvae's digestive tracts was 2 ind. larvae⁻¹. Rotifers with a mean size of $55.44 - 96 \mu m$ were first found in the larvae's digestive tracts. The larvae first consumed copepods with a mean size of $104.16 - 116.28 \mu m$ were by at day 6, while *Artemia* nauplii at day 15.

A 1-day old larva had a weight of 0.067 mg and its weight increased to141.114 mg at the end of the experimental with a growth rate of 2.88 mg day⁻¹. The percentage survival of larvae till 49 days was 3.03%.

The larvae that were fed with *Artemia* had a growth rate of 1.91 mg day⁻¹, and were significantly higher (p < 0.01) than larvae that were fed with artificial feed starting from day 35, 40 and 45. The growth rates of the larvae that were fed the artificial diet on day 35, 40 and 45 were 0.09 mg day⁻¹, 0.022 mg day⁻¹ and – 0.02 mg day⁻¹ respectively. The larvae that were given artificial feed at an early stage grew better than larvae fed on artificial feed at a later stage. Larvae fed on artificial feed on day 35 had a higher significant growth rate than the larvae fed with artificial feed on day 40. The mean survival of the larvae that were given *Artemia* was 21.34%, while larvae that were given the artificial feed at day 35, 40 and 45 were 8, 18 and 19.33%, respectively.



Abstrak tesis dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi syarat untuk mendapatkan ijazah Doktor Falsafah

KAJIAN PERKEMBANGAN EMBRIO DAN LARVA TERHADAP IKAN KERAPU TIKUS (*CROMILEPTES ALTIVELIS* VALENCEE) DAN CORAK MAKANNYA

By

USMAN BULANIN

Februari 2002

Pengerusi: Che Roos Saad, Ph.DFakulti: Fakulti Pertanian

Satu siri penyelidikan telah dijalankan untuk mengkaji 1) Perkembangan embrio dan larva, 2) Corak pemakanan 3) Permulaan masa untuk makanan buatan.

Kesemua kajian telah dijalankan mulai dari peneluran oleh induk sehingga penetasan. Keputusan menunjukkan purata diameter telur ikan kerapu tikus ialah 828.69 ± 45.91 µm. Penetasan telur berlaku selepas 20 jam 10 minit pada suhu 27 sehingga 28 °C. Purata panjang larva yang baru menetas ialah 1.86 mm, dan purata tinggi ialah 0.460 mm. Pundi-pundi kuning telur dan titisan minyak masing-masing habis digunakan 63 dan 65 jam selepas menetas. Larva yang berumur 1 hari mempunyai purata panjang 2.253 mm, dan purata tinggi 0.615 mm. Pada umur 49 hari, purata panjang dan tinggi masing-masing meningkat sehingga 27.153 mm dan 9.190 mm.

Spin larva mula kelihatan pada umur 7 sehingga 10 hari. Spin dorsal mencapai panjang maksimum pada umur 28 sehingga 30 hari dengan purata



panjang adalah 5668.30 µm manakala spin ventral telah mencapai purata panjang maksimum ialah 4415.44 µm pada umur 25 sehingga 28 hari. Seterusnya, spin mulai memendek dan berubah menjadi tulang sirip keras.

Pigmen mata mulai kelihatan pada hari ke 3 sehingga hari ke 4 selepas menetas. Mulut larva mulai terbuka pada hari ke 2 atau 45 jam selepas menetas. Hubungan diameter mata dengan bukaan mulut adalah kuadratik dengan persamaan $MG = -0.2E-05Ed^2 + 0.3992Ed + 215.05 dan R^2 = 0.98$. Bintik-bintik hitam telah kelihatan pada pangkal ekor, sirip dorsal dan sirip anal apabila larva mencapai umur 25 hingga 30 hari. Jumlah bintik-bintik ini telah bertambah dan tersebar ke arah kepala dan lengkap pada umur 40 hingga 45 hari. Pada masa itu, ia kelihatan serupa ikan juvenil.

Gigi larva ikan kerapu tikus mulai kelihatan pada umur 15 hari, tetapi jumlah bilangan gigi masih sedikit dan pendek. Tinggi dan lebar gigi meningkat dengan pertambahan umur. Pada umur 30 hari, gigi larva semakin rapat dengan purata tinggi dan lebar masing-masing adalah 110.18 µm dan 40.13 µm.

Dari aspek histologi, tidak terdapat lipatan di dalam saluran penghadaman larva ikan sebelum berumur 5 hari. Pembentukkan perut mulai kelihatan pada umur 30 hari. Perut larva mulai lengkap pada umur 45 hari. Hati larva mulai kelihatan pada umur 5 hari, manakala pankreas mulai kelihatan pada umur 10 hari. Akiviti enzim protease meningkat dengan pertambahan umur.



Makanan mula ditemui di dalam saluran penghadaman pada umur 3 hari atau 66 jam selepas menetas. Purata jumlah makanan yang ditemui di dalam saluran penghadaman adalah 2 ind./larva. Rotifer yang mula-mula dijumpai di dalam saluran penghadaman mempunyai saiz purata 55.44 – 96 μm. Copepoda dengan saiz purata 104.16 – 116.28 μm mula pertama kali dimakan pada umur 6 hari, manakala nauplii *Artemia* pada umur 15 hari.

Larva yang berumur 1 hari mempunyai berat 0.067 mg dan berat akan meningkat sehingga 141.114 mg pada umur 49 hari dengan purata kadar tumbesaran 2.88 mg/hari. Kadar kemandirian larva sehingga 49 hari adalah 3.03%.

Larva yang diberi makan dengan *Artemia* memperolehi kadar tumbesaran 1.91 mg/hari menunjukan perbezaan yang signifikan (p < 0.01) berbanding daripada larva yang diberi makanan buatan mulai pada umur 35, 40 dan 45 hari. Kadar tumbesaran larva yang diberi makanan buatan pada umur 35, 40 dan 45 hari masing-masing adalah 0.09 mg/hari, 0.022 mg/hari and -0.02 mg/hari. Larva yang diberi makanan buatan paling awal mempunyai tumbesaran lebih baik berbanding larva yang lambat diberi makan buatan. Larva yang diberi makanan buatan pada umur 35 hari adalah mempunyai kadar tumbesaran lebih tinggi daripada larva yang diberi makanan buatan pada umur 40 hari. Purata kadar kemandirian larva yang diberi makan Artemia adalah 21.34%, manakala larva yang diberi makanan buatan pada umur 35, 40 dan 45 hari masing-masing adalah 8, 18 dan 19.33%.



ACKNOWLEDGEMENTS

I would like to extend my sincere gratitude to my supervisor Dr. Che Roos Saad for his support and guidance throughout the course of the study, and members of my supervisory committee Dr. Ridwan Affandi, Associate Prof. Dr. Mohd. Salleh Kamarudin, and Associate Prof. Dr. Abdul Razak Alimon for their valuable suggestions.

My sincere gratitude is also extended to: my employer Bung Hatta University in Padang, which sponsored and granted my study leave at Universiti Putra Malaysia.

My sincere gratitude is also due to the Director of Marine Aquaculture Development Centre Hatchery, Lampung, Indonesia and his staff for granting me some facility for my experimental project.

I would also like to thank to my students, Andi Syafinal, Dedi, Yuniarti, Risdawati, Fera Permata Putri, Irmayda, and Nurbaiti for helping me to collect data. I wish also to express my thanks to Dr. Usman Muhamad Tang, who gave some valuable input and always discuss and advice me about my research and Kristiadi, who helped to translate my thesis. Last but not least, I would also like to thank all my Indonesian friends at UPM and those who helped in correcting of my dissertation.



Finally, my deeply appreciation goes to especially my wife, child and my entire family for without their love and constant encouragement, I would have not been able to complete my study.



I certify that an Examination Committee met on 1 February 2002 to conduct the final examination of Usman Bulanin on his Doctor of Philosophy thesis entitled "Study on the Development of Embryo and Larva of the Humpback Grouper (*Cromileptes altivelis* Valencee) and Its Feeding Behaviour" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

Dahlan Bin Ismail, Ph.D. Professor Department of Animal Science Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Che Roos Saad, Ph.D. Department of Agrotechnology Faculty of Agriculture Universiti Putra Malaysia (Member)

Mohd. Salleh Kamarudin, Ph.D. Associate Professor Department of Agrotechnology Faculty of Agriculture Universiti Putra Malaysia (Member)

Abdul Razak Alimon, Ph.D. Associate Professor Department of Animal Science Faculty of Agriculture Universiti Putra Malaysia (Member)

Ridwan Affandi, Ph.D. Department of Aquatic Resources Management Faculty of Fisheries and Marine Science Bogor Agriculture University (Member)

H. Muchtar Ahmad, Ph.D Professor Faculty of Fishery and Marine Science University of Riau (Independent Examiner)

> SHAMSHER MOHAMAD RAMADILI, Ph.D. Professor Deputy Dean, School of Graduate Studies Universiti Putra Malaysia

Date:



TABLE OF CONTENTS

DEDICATION	
ABSTRACT	
ABSTRAK	vi
ACKNOWLEDGEMENTS	ix
APPROVAL SHEETS	xi
DECLARATION FORM	xiii
LIST OF TABLES	xvii
LIST OF FIGURES	xix
LIST OF ABBREVIATION	xxiii

CHAPTER

Ι	GENERAL INTRODUCTION	1
	Background of Study	1
	Survival of Humpback Grouper Larvae	1
	The Statement of Problems	3
	The Objectives of the Study	4
II	LITERATURE REVIEW	6
	Biology and Development of Humpback Grouper Larvae	6
	Eggs and Larval Development	6
	Development of Larval Digestive Tract	9
	Larval Feeding and Other Options	11
	Natural Feeding versus Artificial Feeding	12
	Larval Behavior	13
	The Environmental Factor	14
	Salinity	14
	Temperature	15
	Oxygen	15
	Ammonia Toxicity	16
III	MATERIALS AND METHOD	17
	Location	17
	Eggs and Larvae	17
	Experimental Container	18
	First Study	20
	Embryo and Larval Development	20
	Second Study	21
	Food and Feeding Behavior	21
	Third Study	21
	Application of Artificial Feed	21
	Data Analysis	22



IV	EGGS AND LARVAL DEVELOPMENT OF THE	
	HUMPBACK GROUPER, Cromileptes altivelis,	24
	Introduction	24
	Materials and Methods	26
	Results	30
	Embryonic Development	30
	Larval Development and Absorption of Yolk Sac and	
	Oil Droplet	32
	Post Yolk Sac Development of Larvae	38
	Discussion	47
	Weight, Length and Height Increment	50
	Spine Development	50
	Eye Development	51
	Mouth Development	51
	Spot Development	52
	Survival	52
	Conclusions	52
V	DEVELOPMENT OF DIGESTIVE SYSTEM OF THE	
	HUMPBACK GROUPER, Cromileptes altivelis, LARVAE	54
	Introduction	54
	Materials and Methods	56
	Data Analysis	57
	Results	57
	Anatomical Development	57
	Histological Development	59
	Enzyme Activity	68
	Discussion	69
	Development of Teeth	69
	Anatomical Development of Digestive Tract	70
	Histological Development of Digestive Tract	70
	Development of Liver and Pancreas	72
	Conclusions	73
VI	CHANGES IN FEEDING BEHAVIOR OF THE	
	HUMPBACK GROUPER FISH, Cromileptes altivelis,	
	LARVAE	75
	Introduction	75
	Materials and Methods	/6
	Data Analysis	80
	Results	80
	Food Selectivity Index	87
	Larval Behavior and Feeding Habit	89
	water Quality	93
	Discussion	94
	Benavior	9/
	reeding Habit	100
	Conclusions	101



VII	GROWTH AND PERFORMANCE OF THE HUMPBACK	
	GROUPER, Cromileptes altivelis, LARVAE	103
	Introduction	103
	Materials and Methods	104
	Data Analysis	107
	Results	107
	Environmental Factors	110
	Discussion	110
	Survival	114
	Conclusions	119
VIII	OPTIMAL WEANING PERIOD FOR HUMPBACK	
	CROUPER, Cromileptes altivelis, LARVAE	120
	Introduction	120
	Materials and Methods	121
	Data Analysis	124
	Results	124
	Survival	126
	Discussion	128
	Survival	131
	Conclusions	. 132
IX	GENERAL DISCUSSION AND CONCLUSION	133
	General Discussion	133
	General Conclusion	137
	Recommendation	139
BIBLI	OGRAPHY	140
APPEI	NDICES	153
BIODA	BIODATA OF THE AUTHOR 174	



LIST OF TABLES

Τ	ables		Page
2	.1	Description of eggs of epinepheline seranids.	ິ 7
2	.2	Description of newly hatched larval of epinepheline seranids.	8
4	.1	Time of embryonic development for humpback grouper, Cromileptes altivelis, at 27 – 28 °C.	30
4	.2	Yolk sac and oil droplet absorption of humpback grouper, <i>Cromileptes altivelis</i> , larvae.	33
4	.3	Weight (mg), length and height increment (mm) for humpback grouper, Cromileptes altivelis, larvae.	38
5	.1	Protease enzyme activity value (U/larva) of humpback grouper, <i>Cromileptes altivelis</i> , larval.	68
6.	.1	Percentage of eating larvae and the amount of food in the stomach (ind./fish) from age 62 to 72 hours after hatching.	80
6.	.2	Age, eye size, mouth gape, total food in stomach and the number of individual food in larvae's stomach.	. 81
6.	.3	Water quality parameters of culture media for humpback grouper, <i>Cromileptes altivelis</i> , larval.	93
7.	.1	Physical and chemical parameters of water during culture of humpback grouper, Cromileptes altivelis, larvae.	110
8.	.1	Growth rate (mg day ⁻¹) of humpback grouper, <i>Cromileptes altivel is</i> , larvae fed with artificial feed.	124
8.	.2	Final survival (%) of humpback grouper, Cromileptes altivelis, larvae fed with artificial feed.	126



LIST OF FIGURES

Figures		Page
1.1	Flow chart of problem analysis	5
3.1	Procedure of preparing seawater for research	19
3.2	Types of natural food given to humpback grouper, Cromileptes altivelis, larvae. A) Rotifers, B-D) Copepods, E) Artemia nauplii and F) Mysidopsis sp.	23
4.1	Spawning tank of humpback grouper, Cromileptes altivelis.	27
4.2	Stages of embryonic development of humpback grouper, <i>Cromileptes altivelis</i> , A) fertilized egg, B) 1 cell stage, C) 2 cell, D) 4 cell, E) 8 cell, F) 32 cell, G) 64 cell, H) morula, I) blastula, J) gastrula, K) formation of embryo, L) development of head and tail buds, M) complete embryonic development, N) tail coming out of shell and O) hatch out.	31
4.3	Absorption rate of yolk sac (Ys) and oil droplet (Od) of humpback grouper, <i>Cromileptes altivelis</i> , larvae.	34
4.4	Relationship of (a) age (hour) with yolk sac and (b) oil droplet absorption for humpback grouper, <i>Cromileptes altivelis</i> , larvae	35
4.5	Morphological development of humpback grouper, Cromileptes altivelis, larva: a) newly hatched larva, b) 1 day old, c) 2 days old, d) 3 days old, e) 8 days old, f) 12 days old, g) 18 days old, h) 21 days old i) 24 days old, j) 35 days old, k) 40 days old and l) 45 days old.	36
4.6	Length and height of humpback grouper, Cromileptes altivelis, larva during yolk sac and oil droplet absorption.	37
4.7	Growth rate (mg day ⁻¹) (a) length and weight relationship (b) of humpback grouper, <i>Cromileptes altivelis</i> , larvae.	39
4.8	Total length and height of humpback grouper, Cromileptes altivelis, larvae.	40
4.9	Relationship of length and height of humpback grouper, Cromileptes altivelis, larvae.	41

4.10	Total length of spines for humpback grouper, <i>Cromileptes altivelis</i> , larvae.	42
4.11	Diameter of eye and mouth gape (MG) with age of humpback grouper, <i>Cromileptes altivelis</i> , larvae.	43
4.12	Relationship between eye diameter with mouth gape of humpback grouper, Cromileptes altivelis, larvae.	44
4.13	Spots and spines development of humpback grouper, <i>Cromileptes altivelis</i> , larva at: A) 12 days, B) 15 days, C) 18 days, D) 21 days, E) 24 days, F) 35 days, G) 40 days and H) 45 days old.	45
4.14	Survival (%) of humpback grouper, <i>Cromileptes altivelis,</i> larvae during study period.	46
5.1	Teeth development of humpback grouper, <i>Cromileptes altivelis</i> , larva A) 15 days, B) 20 days, C) 25 days and D) 30 days old.	58
5.2	Digestive tract development of humpback grouper, Cromileptes altivelis, larva: A) day 1, B) day 2, C) day 3 and D) day 4.	59
5.3	Development of alimentary tract in humpback grouper, <i>Cromileptes altivelis</i> , larva at: A) 5 days, B) 10 days, C) 15 days, D) 20 days, E) 25 days, F) 30 days, G) 35 days, H) 40 days and I) 45 days old (stained with HE).1) Esophagus, 2) Stomach, 3) Intestine, 4) Liver, 5) Pancreatic, 6) Anus, 7) Muscle, 8) Food in lumen, 9) Pyloric and 0) Incipient stomach.	60
5.4	Development of intestine in humpback grouper, <i>Cromileptes altivelis</i> , larva at: A) 5 days, B) 10 days, C) 15 days, D) 20 days, E) 25 days, F) 30 days, G) 35 days, H) 40 days and I) 45 days old (stained with HE). 1) Epithelium, 2) Enterocyte, 3) Mucosa epithelium, 4) Sub mucosa, 5) Food in lumen.	62
5.5	Development of stomach in humpback grouper, <i>Cromileptes altivelis</i> , larva at: A) 30 days, B) 35 days, C) 40 days and D) 45 days old (stained with HE). 1) Mucosa, 2) Food in lumen, 3) Sub mucosa, 4) Gastric gland.	64
5.6	Development of liver in humpback grouper, <i>Cromileptes</i> <i>altivelis</i> , larva at: A) 5 days, B) 10 days, C) 15 days, D) 20 days, E) 25 days, F) 30 days, G) 35 days, H) 40 days and I) 45 days old (stained with HE). 1) Hepatic cells, 2) Pericellular	
	ducts.	60



5.7	Development of pancreas in humpback grouper, <i>Cromileptes altivelis</i> , larva at: A) 10 days, B) 15 days, C) 20 days, D) 25 days, E) 30 days, F) 35 days, G) 40 days and H) 45 days old (stained with HE). 1) Exocrine pancreatic 2) Islets of langerhans.	67
5.8	Enzyme activity (U/larva) of humpback grouper, Cromileptes altivelis, larvae at different age.	69
6.1	Equipments used on natural feeding for humpback grouper, <i>Cromileptes altivelis</i> .	77
6.2	(a) Amount of rotifer in stomach day ⁻¹ and (b) its relationship with mouth gape.	82
6.3	(a) Amount of copepod in larvae's stomach day ⁻¹ and (b) its relationship with mouth gape.	84
6.4	(a) Amount of <i>Artemia</i> nauplii in larval stomach day ⁻¹ (b) and ⁻ its relationship with mouth gape.	86
6.5	Food selectivity index of humpback grouper, Cromileptes altivelis, larval by kinds.	88
6.6	Behavior of humpback grouper, <i>Cromileptes altivelis</i> , larvae. A) 1 to 4 days. B) 10 to 12 days C and D) 13 to 25 days E) 30 days and F) 45 days.	90
6.7	Feeding schedule and type of food for humpback grouper, Cromileptes altivelis, larvae until 50 days of culture period	98
7.1	Larva rearing tank of humpback grouper, Cromileptes altivelis,	105
7.2	Mean weight increment (mg day ⁻¹) of humpback grouper, Cromileptes altivelis, larva.	108
7.3	Survival (%) of humpback grouper, <i>Cromileptes altivelis</i> , larvae during the study period.	109
7.4	Other factors affecting mortality of humpback grouper, <i>Cromileptes altivelis</i> , larva.	118
8.1	Experiment tank in artificial feeding for humpback grouper, Cromileptes altivelis, larval	122
8.2	Schedule of weaning from <i>Artemia</i> to artificial feed for humpback grouper, <i>Cromileptes altivelis</i> .	123

8.3	Weight increment (mg) of humpback grouper, <i>Cromileptes altivelis</i> , larvae fed with artificial feed.	125
8.4	Survival (%) of humpback grouper, <i>Cromileptes altivelis</i> , larvae fed with artificial feed.	127



LIST OF ABBREVIATIONS

AF	= After fertilization
ANOVA	= Analysis of variance
С	= Control
cm	= Centimetre
CRD	= Complete randomized design
D	= Day
DAH	= Day after hatching
DO	= Dissolved oxygen
Dsp	= Dorsal spine
E	= Epinephelus
Ed	= Eye diameter
F	= Fish
g	= Gram
GR	= Growth rate
GRH	= Growth rate height
GRL	= Growth rate length
GRW	= Growth rate weight
Н	= Height
h	= Hours
НАН	= Hours after hatching
HE	= Hematoxylin eosin
HUFA	= High unsaturated fatty acid

Ind.	= Individual
U	= Unit
IUC	= Inter university center
L	= Length
L	= Litre
L	= Lutjanus
m	= Metre
MG	= Mouth gape
mg	= Milligram
mm	= Millimetre
ml	= Millilitre
μm	= Micron metre
N	= Nauplii
Od	= Oil droplet
ppm	= Parts per million
ppt.	= Parts per thousand
r	= Radius
SR	= Survival rate
V	= Volume
Vsp.	= Ventral spines
W	= Weight
Ys	= Yolk sac