

LIPID OXIDATION AND PROTEIN CO-OXIDATION IN READY-TO-EAT LOW-MOISTURE SHREDDED (SERUNDING) BEEF AND CHICKEN

By

HAZRATI BINTI WAZIR

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

April 2022

FSTM 2022 14

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

LIPID OXIDATION AND PROTEIN CO-OXIDATION IN READY-TO-EAT LOW-MOISTURE SHREDDED (*SERUNDING*) BEEF AND CHICKEN

By

HAZRATI BINTI WAZIR

April 2022

Chair : Nazamid Saari, PhD Faculty : Food Science and Technology

Ready-to-eat (RTE), low-moisture, high-lipid and high-protein meat products are convenient in emergencies like earthquakes, flash floods, and the global COVID-19 lockdown. Given the long storage times of these products, the lipid and protein components may be more susceptible to oxidation, which causes discolouration, shelf-life reduction, nutritional quality deterioration, lipid rancidity, and protein denaturation. This first objective of the research aimed to determine the effects of meat type, storage time and temperature on lipid oxidation and protein co-oxidation of shredded (serunding) beef and chicken during six months storage. The production of shredded (serunding) beef and chicken was done with desired moisture content (<7.5%) of 6.36% and 7.46% respectively. Immediate post-production analyses of shredded (serunding) beef and chicken include proximate composition, water activity, moisture content, fatty acid composition (FAC), reducing sugar, and scanning electron microscopy (SEM). Upon completion of the analyses, the storage study was instantly conducted to evaluate the physicochemical, lipid oxidation and protein co-oxidation of shredded (serunding) beef and chicken at 25, 40, and 60°C. Due to its higher polyunsaturated fatty acids (PUFA) content in shredded (serunding) chicken (4.12%) than shredded (serunding) beef (1.45%), chicken meat was more susceptible to lipid oxidation than beef, which was evidenced by a higher conjugated dienes (CD) content of shredded (serunding) chicken, 3.0-3.5 mmol CD/mol lipid than CD of shredded (serunding) beef (1.5-2.0 mmol CD/mol lipid) and a rapid accumulation of malondialdehyde (MDA) (week 2 to 10). Chicken serunding also experienced more severe protein co-oxidation than beef serunding, as seen from lower protein solubility (18-24 mg/mL; beef serunding: 28-35 mg/mL), higher protein carbonyl (chicken serunding: 36.63 nmol carbonyl/mg (60°C), 18.59 nmol carbonyl/mg (40°C), 6.80 nmol carbonyl/mg (25°C); beef serunding: 27.40 nmol carbonyl/mg (60°C), 8.68 nmol carbonyl/mg (40°C), 6.23 nmol carbonyl/mg (25°C)), higher tryptophan loss and higher Schiff base formation. The second objective of evaluating the relation of

selected parameters with lipid-protein oxidative markers by principal component analysis (PCA) confirms that meat type, storage time, and storage temperature influence changes in extracted lipid, MDA, and protein co-oxidation reaction (tryptophan loss, Schiff base, and protein carbonyl) over six months. PCA results support the general discussion about colour changes, lipid oxidation, and protein co-oxidation in beef and chicken serunding. Chicken serunding demonstrates lower lipid and protein stability and higher lipid oxidation and protein co-oxidation rates than beef serunding. Subsequently, in the third objective, the butylated hvdroxvanisole (BHA) antioxidant was added (200 ppm based on the lipid content of sample), and different multilayer packaging was used to improve the lipid and protein stability of chicken serunding. The proximate composition, FAC and SEM of chicken serunding with BHA addition (CS-BHA) were measured directly after production, and the results were comparable to control (CS). After the completion of the preceding analyses, the storage study of six months was conducted. During storage, the simultaneous interacting effects of different storage temperatures (25, 40, and 60°C), antioxidant (BHA), and multilayer packaging materials (metallised polyethylene terephthalate (MPET) and aluminum-containing packaging) were evaluated. All lipid and protein markers elevated with increasing temperature $(25 < 40 < 60^{\circ}C)$, indicating that storage of low-moisture meat at high temperature is not feasible. BHA was effective against lipid oxidation, as indicated by the significantly lower extracted lipid content (CS-BHA: 40-44%; CS: 42-46.5%) and delayed formation of MDA, i.e. MDA peaked at week 14 in CS-BHA samples (MPET packaging) while MDA peaked at week 10 in CS samples (MPET packaging). However, BHA is not effective against protein co-oxidation, as shown by the insignificant effect on preventing tryptophan loss, protein carbonyl formation, and Schiff base accumulation. As for the packaging effect, MPET packaging with a superior light and oxygen barrier provided significant protection compared to aluminium. MPET packaging showed lower lipid oxidation, as the CD content in aluminium-packaging peaked earlier (week 2-3). Additionally, MPET packaging consistently had a significantly lower MDA level than aluminium-packaging. In terms of protein oxidation, MPET packaging recorded a tryptophan loss of -32.16% at 25 ° C, -35.98% at 40 ° C and -41.74% at 60 ° C and packaging B a loss of -21.85% at 25 ° C, -22.33% at 40 ° C and -34.64% at 60 ° C, respectively, showing that MPET (packaging A) induced less severe protein co-oxidation than aluminium (packaging B). Finally, PCA reveals the variations in extracted lipid, MDA and protein cooxidation reaction (tryptophan loss, Schiff base and protein carbonyl), monitored over six months of storage, are impacted by storage temperature and packaging material but not by antioxidant. In conclusion, from the quality and economic aspects, 10 to 12 weeks of shelf-life are sufficient to reliably estimate the lipid oxidation onset of shredded (serunding) meat at storage temperature of 25°C and 40°C, therefore three months post-production is the recommended storage term. 25°C, with antioxidant addition in metallised barrier multilayer packaging, is considered as the ideal long-term storage condition to delay simultaneous lipid oxidation and protein co-oxidation. These findings provide insights into simultaneous lipid oxidation and protein co-oxidation in RTE low-moisture, shredded (serunding) meats and might be extrapolated to minimise probable harmful impacts on the quality of low-moisture, high-lipid and high-protein foods.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENGOKSIDAAN LIPID DAN PENGOKSIDAAN BERSAMA PROTEIN DALAM DAGING CARIK (SERUNDING) LEMBU DAN AYAM SEDIA DIMAKAN (RTE) DAN KELEMBAPAN RENDAH

Oleh

HAZRATI BINTI WAZIR

April 2022

Pengerusi : Nazamid Saari, PhD Fakulti : Sains dan Teknologi Makanan

Produk daging sedia untuk dimakan (RTE), lembapan rendah, lemak tinggi dan tinggi protein adalah mudah dalam keadaan kecemasan seperti gempa bumi. banjir kilat dan penutupan global COVID-19. Memandangkan produk ini mempunyai masa penyimpanan yang lama, komponen lipid dan protein mungkin lebih terdedah kepada pengoksidaan, menyebabkan perubahan warna, pengurangan jangka hayat, kemerosotan kualiti pemakanan, ketengikan lipid, dan denaturasi protein. Objektif pertama penyelidikan ini bertujuan untuk menentukan kesan jenis daging, masa penyimpanan dan suhu ke atas pengoksidaan lipid dan pengoksidaan protein daging lembu dan ayam yang dicarik (serunding) selama penyimpanan enam bulan. Pengeluaran daging lembu dan ayam yang dicarik (serunding) dilakukan dengan kandungan lembapan yang dikehendaki (<7.5%) masing-masing sebanyak 6.36% dan 7.46%. Analisis segera pasca pengeluaran serunding daging lembu dan ayam termasuk komposisi proksimat, aktiviti air, kandungan lembapan, komposisi asid lemak (FAC), gula penurun, dan mikroskop elektron pengimbasan (SEM). Setelah selesai analisis, kajian penyimpanan dijalankan serta-merta untuk menilai fizikokimia, pengoksidaan lipid dan pengoksidaan protein daging lembu dan serunding ayam pada suhu 25, 40, dan 60°C. Oleh kerana kandungan asid lemak politaktepu (PUFA) yang lebih tinggi dalam serunding ayam (4.12%) daripada serunding daging lembu (1.45%), daging ayam lebih mudah terdedah kepada pengoksidaan lipid daripada daging lembu, yang dibuktikan oleh kandungan diena terkonjugasi (CD) yang lebih tinggi. serunding ayam, 3.0-3.5 mmol CD/mol lipid daripada CD serunding daging lembu (1.5-2.0 mmol CD/mol lipid) dan pengumpulan pesat malondialdehid (MDA) (minggu 2 hingga 10). Serunding ayam juga mengalami pengoksidaan protein yang lebih teruk daripada serunding daging lembu, seperti yang dilihat daripada keterlarutan protein yang lebih rendah (18-24 mg/mL; serunding daging lembu: 28-35 mg/mL), karbonil protein yang lebih tinggi (serunding ayam: 36.63 nmol karbonil/ mg

(60°C), 18.59 nmol karbonil/mg (40°C), 6.80 nmol karbonil/mg (25°C); serunding daging lembu: 27.40 nmol karbonil/mg (60°C), 8.68 nmol karbonil/mg (40 °C), 6.23 nmol karbonil/mg (25°C)), kehilangan triptofan yang lebih tinggi dan pembentukan asas Schiff yang lebih tinggi. Objektif kedua menilai hubungan parameter terpilih dengan penanda oksidatif lipid-protein oleh analisis komponen utama (PCA) mengesahkan bahawa jenis daging, masa penyimpanan, dan suhu penyimpanan mempengaruhi perubahan dalam tindak balas pengoksidaan lipid (lipid yang diestrak dan MDA), dan pengoksidaan bersama protein (kehilangan triptofan, asas Schiff, dan protein karbonil) selama enam bulan, Keputusan PCA menyokong perbincangan umum tentang perubahan warna, pengoksidaan lipid, dan pengoksidaan bersama protein dalam daging lembu dan serunding ayam. Serunding ayam menunjukkan kestabilan lipid dan protein yang lebih rendah serta kadar pengoksidaan lipid dan pengoksidaan bersama protein yang lebih tinggi daripada serunding daging lembu. Seterusnya, dalam objektif ketiga, antioksidan butilasi hidroksianisole (BHA) telah ditambah (200 ppm berdasarkan kandungan lipid sampel), dan pembungkusan pelbagai lapisan yang berbeza digunakan untuk meningkatkan kestabilan lipid dan protein serunding ayam. Komposisi proksimat, FAC dan SEM serunding ayam dengan penambahan BHA (CS-BHA) diukur secara langsung selepas pengeluaran, dan hasilnya adalah setanding dengan kawalan (CS). Selepas selesai analisis sebelumnya, kajian penyimpanan selama enam bulan telah dijalankan. Semasa penyimpanan, kesan interaksi serentak suhu penyimpanan yang berbeza (25, 40, dan 60°C), antioksidan (BHA), dan bahan pembungkusan berbilang lapisan (polietilena tereftalat berlogam (MPET) dan pembungkusan yang mengandungi aluminium) telah dinilai. Semua penanda lipid dan protein dinaikkan dengan peningkatan suhu (25 < 40 < 60° C), menunjukkan bahawa penyimpanan daging rendah lembapan pada suhu tinggi tidak boleh dilaksanakan. BHA berkesan terhadap pengoksidaan lipid, seperti yang ditunjukkan oleh kandungan lipid yang diekstrak dengan ketara lebih rendah (CS-BHA: 40-44%; CS: 42-46.5%) dan tertunda pembentukan MDA, jaitu MDA memuncak pada minggu ke-14 dalam sampel CS-BHA (pembungkusan MPET) manakala MDA memuncak pada minggu ke-10 dalam sampel CS (pembungkusan MPET). Walau bagaimanapun, BHA tidak berkesan terhadap pengoksidaan bersama protein, seperti yang ditunjukkan oleh kesan yang tidak ketara terhadap mencegah kehilangan triptofan, pembentukan karbonil protein, dan pengumpulan asas Schiff. Bagi kesan pembungkusan, pembungkusan MPET dengan penghalang cahaya dan oksigen yang unggul memberikan perlindungan yang ketara berbanding aluminium. Pembungkusan MPET menunjukkan pengoksidaan lipid yang lebih rendah, kerana kandungan CD dalam pembungkusan aluminium memuncak lebih awal (minggu 2-3). Selain itu, pembungkusan MPET secara konsisten mempunyai tahap MDA yang jauh lebih rendah daripada pembungkusan aluminium. Dari segi pengoksidaan protein, pembungkusan MPET merekodkan kehilangan triptofan sebanyak -32.16% pada 25 ° C, -35.98% pada 40 ° C dan -41.74% pada 60° C dan pembungkusan B kehilangan -21.85% pada -21.85% pada 2.5 . 40 ° C dan -34.64% pada 60 ° C, masing-masing, menunjukkan bahawa MPET (pembungkusan A) menyebabkan pengoksidaan protein yang kurang teruk berbanding aluminium (pembungkusan B). Akhir sekali, PCA mendedahkan variasi dalam tindak balas pengoksidaan lipid, MDA dan protein yang diekstrak (kehilangan triptofan, asas Schiff dan karbonil protein), dipantau selama enam bulan penyimpanan, dipengaruhi oleh suhu penyimpanan dan

bahan pembungkus tetapi bukan oleh antioksidan. Kesimpulannya, dari aspek kualiti dan ekonomi, jangka hayat 10 hingga 12 minggu adalah mencukupi untuk menganggarkan dengan pasti permulaan pengoksidaan lipid serunding daging pada suhu penyimpanan 25°C dan 40°C, oleh itu tiga bulan pasca pengeluaran adalah tempoh penyimpanan yang disyorkan. Suhu penyimpanan pada 25°C dan juga penambahan antioksidan dalam pembungkusan berbilang lapisan penghalang logam, dianggap sebagai keadaan penyimpanan jangka panjang yang ideal untuk menangguhkan pengoksidaan lipid serentak dan pengoksidaan bersama protein. Penemuan ini memberikan gambaran tentang pengoksidaan lipid serentak dan pengoksidaan bersama protein dalam daging RTE yang lembap rendah, dicarik (serunding) dan mungkin diekstrapolasi untuk meminimumkan kemungkinan kesan berbahaya terhadap kualiti makanan rendah lembapan, lemak tinggi dan protein tinggi.

ACKNOWLEDGEMENTS

"It always seems impossible until it's done"

Thank you, Allah SWT, for giving me the strength and courage to complete the thesis. I had considered quitting the programme, but Allah's infinite grace came to my rescue.

I would like to acknowledge my main supervisor, Professor Dr Nazamid Saari, for his warm welcome, tremendous guidance, fantastic advice, unending support and patience. Dr Wan Zunairah Wan Ibadullah and Dr Nor Afizah Mustapha have been equally supportive and helpful. There were scientific viewpoints and constructive criticism on the experiment design and result interpretation. As neither bright nor competent student, I owe gratitude to all supervisors for their presence, patience, trust, and moral support, both mentally and financially.

I would also like to thank Dr Mohamad Zarei and Dr Chay Shyan Yea, postdoctoral researchers, who have given me valuable opinions and guidance during discussions on result interpretation and refining articles writing. Special thanks are dedicated to science officers, assistant science officers, and lab assistants, as well as postgraduate division staff and financial division staff, for their patience and kind assistance when help needed. Sincere appreciations are dedicated to Farah Salina, Hajar, Dhania, Fatema Brishti, Fatimah, 'Atiqah Aqilah, Ariff, Najib, Auwal, Balqis, Nooratikah, Aqilah, Anis, Nusrah, Brisha, Syafiqah, Alia, and Aliah Zannierah for the friendship, discussion and help. You have supported me throughout the countless hours of lab work, data collection and data curation required for my PhD.

Thank you to the Ministry of Higher Education and Universiti Tun Hussein Onn Malaysia for the scholarships awarded and the opportunity to pursue a PhD Degree at Universiti Putra Malaysia (UPM).

Thank you so much to my family. No words can adequately describe my appreciation to my father, Wazir Haron, mother Salabiah Serbaini, spouse Che Maliki Ngah, son Ammar Yasir, daughter Adila Zahra, siblings; Shairah, Anwar Faiz, Akmal Farid. Finally, my close friends; Iman, Fatimah, Afidah, Azni, Ain, Lina, Noorbaiti, and Umi. They helped me financially, physically, and emotionally throughout the study period. Their unwavering love, faith in me, and prayers kept me going till now.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Nazamid bin Saari, PhD

Professor Faculty of Food Science and Technology Universiti Putra Malaysia (Chairman)

Wan Zunairah binti Wan Ibadullah, PhD

Senior Lecturer Faculty of Food Science and Technology Universiti Putra Malaysia (Member)

Nor Afizah binti Mustapha, PhD

Senior Lecturer Faculty of Food Science and Technology Universiti Putra Malaysia (Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and the copyright of the thesis are fullyowned by Universiti Putra Malaysia, as stipulated in the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from the supervisor and the office of the Deputy Vice-Chancellor (Research and innovation) before the thesis is published in any written, printed or electronic form (including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials) as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld in accordance with the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2015-2016) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature:	
Signature.	

Date:

Name and Matric No.: Hazrati binti Wazir

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research and the writing of this thesis were done under our supervision;
- supervisory responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2015-2016) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	
Signature: Name of Member of Supervisory Committee:	
Signature: Name of Member of Supervisory Committee:	
Signature: Name of Member of Supervisory Committee:	

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	vi
APPROVAL	vii
DECLARATION	ix
LIST OF TABLES	xvi
LIST OF FIGURES	xviii
LIST OF APPENDICES	xxiii
LIST OF ABBREVIATIONS	xxiv

CHAPTER

1

INTE	RODUC	TION		1
		round of t	he study	1
		m statem		1
1.3	Object	tives of the	e study	3
LITE	RATU	RE REVIE	W	4
2.1			TE) food products	4
	2.1.1	Definition	n and market of ready-to-eat	4
		(RTE)		
	2.1.2		ready-to-eat (RTE)	6
2.2		and meat		7
	2.2.1		characteristics of meat and	8
		meat pro		
	2.2.2		d protein in meats	11
		2.2.2.1	Lipid in meat and meat	11
			products	
		2.2.2.2	Protein in meat and meat	17
	0.0.0	Dresses	products	10
	2.2.3		ed meat products	19 19
		2.2.3.1	Fresh processed meat products	19
		2222	Cured meat cuts	20
		2.2.3.2		20
		2234	Pre-cooked meat products	20
		2.2.3.5		21
		2.2.0.0	products	21
		2.2.3.6		21
			Frozen meat products	21
		2.2.3.8	·	21
	2.2.4	Ready-to	o-eat (RTE) low-moisture meat-	22
		based pr		
		2.2.4.1		22
2.3	Shred	ded (serui	<i>nding</i>) meat	24
	2.3.1	Preparat	ion	24
	2.3.2	Non-mea	at ingredients	25

		2.3.2.1 Coconut milk	25
		2.3.2.2 Garlic, onions, chillies and spices	25
	2.3.3	•	25
2.4		oxidation and protein co-oxidation (lipid- n interaction) in meat and meat products	26
	2.4.1	General background	33
	2	2.4.1.1 Decomposition of lipid radicals	33
	2.4.2		35
		2.4.2.1 Meat composition	35
		2.4.2.2 Packaging, oxygen	36
		concentration, moisture and	
		presence of light	07
		2.4.2.3 Storage conditions 2.4.2.4 Antioxidants	37 37
	2.4.3	Primary lipid oxidation products:	37
	2.4.5	conjugated dienes (CD)	50
	2.4.4	Secondary oxidation product:	39
		malondialdehydes (MDA)	
2.5		n co-oxidation with oxidised lipid	41
	2.5.1		41
	2.5.2		44
	2.5.3		44 45
	2.5.4	Formation of carbonyl derivatives Formation of Schiff base products	45
		Loss of tryptophan fluorescence	49
2.6		kidant addition: their effect on lipid	50
	oxidat	ion and protein co-oxidation in meat	
	produ		50
	2.6.2	General background Safety aspects of antioxidant changes	50
	2.0.2	in foods	52
	2.6.3		54
		2.6.3.1 2,2-azinobis-(3-	54
		ethylbenzothiazoline-6-	0.
		sulphonic acid (ABTS) assay	
		2.6.3.2 2,2-diphenyl-1-picrylhydrazyl	55
		(DPPH) assay	
		2.6.3.3 Ferric reducing antioxidant	55
27	Dooko	power (FRAP) assay aging material: the effect on lipid oxidation	56
2.7		rotein co-oxidation in meat products	56
	2.7.1	Packaging requirements for meat and	56
		meat products	
		•	
		S AND METHODS / METHODOLOGY	59
3.1	Materi	ials	59

3.2	tempe	rature on ion of sh	at type, storage time and lipid oxidation and protein co- redded (<i>serunding</i>) beef and	60
	3.2.1 3.2.2	Experime	ental design on of shredded (<i>serunding</i>) Lobicken	60 62
	3.2.3	Physicoc	chemical quality assessment on	63
			d (<i>serunding</i>) beef and chicken	62
		3.2.3.1		63
			Reducing sugar content	64 65
		3.2.3.3	Water activity (a _w) Moisture content	
		3.2.3.4		65
		3.2.3.5	Fatty acid composition	65
		3.2.3.6	Colour measurements	67
		3.2.3.7	Scanning electron microscopy (SEM)	67
	3.2.4		dation analysis of shredded ng) beef and chicken	67
		3.2.4.1	Lipid extraction and total lipid	67
		5.2.4.1	content	07
		3.2.4.2	Conjugated dienes (CD)	67
		3.2.4.3	Thiobarbituric acid reactive	68
	3.2.5	Drotoin	substances (TBARS)	60
	3.2.3	Protein	co-oxidation analysis of	69
		3.2.5.1	d (serunding) beef and chicken Protein extraction	69
		3.2.5.1		70
			Soluble protein content	
		3.2.5.3	Amino acid composition	70
		3.2.5.4	Protein carbonyl content	71
		3.2.5.5	Tryptophan loss and Schiff	72
	2.2.0	Otatiatia	base fluorescence	70
~ ~	3.2.6		al data analysis	72
3.3			e temperature, antioxidant and	73
			erials on lipid oxidation and	
	chicke		ation of shredded (serunding)	
			entel decim	70
			ental design	73
	3.3.Z		on of shredded (serunding)	74
		chicken		74
	3.3.3		chemical quality assessment on d (<i>serunding</i>) chicken, without	74
		•	6) or with BHA (CS-BHA)	74
		3.3.3.1		74 74
		3.3.3.2	Fatty acid composition	
		3.3.3.3	Water activity (a _w)	74 75
		3.3.3.4	Colour measurements	75
		3.3.3.5	Scanning electron	75
			microscopy (SEM)	

		idation analysis of shredded	75
		ng) beef and chicken	
	3.3.4.1	Lipid extraction and total lipid content	75
	3.3.4.2	Conjugated dienes (CD)	75
	3.3.4.3	Thiobarbituric acid reactive	75
		substances (TBARS)	
	3.3.4.4	Measurement of total	76
		antioxidant capacity (TAC) in	
		shredded (serunding) chicken	
		on storage stability	
3.3.5	Protein	co-oxidation analysis of	82
0.0.0		d (<i>serunding</i>) chicken	02
	3.3.5.1		82
	3.3.5.2		82
	3.3.5.3		82
	3.3.5.4	Tryptophan loss and Schiff	82
	0.0.0.4	base fluorescence	02
3.3.6	Statistics	al data analysis	82
5.5.0	Statistica		02
RESULTS /		INOISSION	83
		at type, storage time and	83
		lipid oxidation and protein co-	05
		redded (serunding) beef and	
		reduced (Sciuliding) beer and	
c <mark>hicke</mark>	n		02
	n Physicod	chemical quality assessment	83
c <mark>hicke</mark>	n	chemical quality assessment Proximate composition, water	83 83
c <mark>hicke</mark>	n Physicod	chemical quality assessment Proximate composition, water activity (aw), and reducing	
c <mark>hicke</mark>	n Physicoo 4.1.1.1	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar	83
c <mark>hicke</mark>	n Physicoo 4.1.1.1 4.1.2	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC)	83 88
c <mark>hicke</mark>	n Physicoo 4.1.1.1	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and	83
c <mark>hicke</mark>	Physicod 4.1.1.1 4.1.2 4.1.1.3	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement	83 88 90
c <mark>hicke</mark>	n Physicoo 4.1.1.1 4.1.2	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron	83 88
chicke 4.1.1	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.4	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM)	83 88 90 93
c <mark>hicke</mark>	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.4 Lipid oxid	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation	83 88 90 93 94
chicke 4.1.1	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.4 Lipid oxid 4.1.2.1	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation Extracted lipid	83 88 90 93 94 94
chicke 4.1.1	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.4 Lipid oxid	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation Extracted lipid Primary product: conjugated	83 88 90 93 94
chicke 4.1.1	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.4 Lipid oxid 4.1.2.1 4.1.2.2	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation Extracted lipid Primary product: conjugated dienes (CD)	83 88 90 93 94 94 95
chicke 4.1.1	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.4 Lipid oxid 4.1.2.1	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation Extracted lipid Primary product: conjugated dienes (CD) Secondary product:	83 88 90 93 94 94
chicke 4.1.1 4.1.2	Physicod 4.1.1.1 4.1.2 4.1.1.3 4.1.1.4 Lipid oxid 4.1.2.1 4.1.2.2 4.1.2.3	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation Extracted lipid Primary product: conjugated dienes (CD) Secondary product: malondialdehydes (MDA)	 83 88 90 93 94 94 95 97
chicke 4.1.1	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.4 Lipid oxid 4.1.2.1 4.1.2.2 4.1.2.3 Protein oxid	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation Extracted lipid Primary product: conjugated dienes (CD) Secondary product: malondialdehydes (MDA) co-oxidation	83 88 90 93 94 94 95 97 98
chicke 4.1.1 4.1.2	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.4 Lipid oxid 4.1.2.1 4.1.2.2 4.1.2.3 Protein of 4.1.3.1	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation Extracted lipid Primary product: conjugated dienes (CD) Secondary product: malondialdehydes (MDA) co-oxidation Soluble protein content	 83 88 90 93 94 94 95 97 98 98
chicke 4.1.1 4.1.2	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.4 Lipid oxid 4.1.2.1 4.1.2.2 4.1.2.3 Protein of 4.1.3.1 4.1.3.2	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation Extracted lipid Primary product: conjugated dienes (CD) Secondary product: malondialdehydes (MDA) co-oxidation Soluble protein content Amino acid composition	83 88 90 93 94 94 95 97 97 98 98 101
chicke 4.1.1 4.1.2	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.4 Lipid oxid 4.1.2.1 4.1.2.2 4.1.2.3 Protein of 4.1.3.1	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation Extracted lipid Primary product: conjugated dienes (CD) Secondary product: malondialdehydes (MDA) co-oxidation Soluble protein content	83 88 90 93 94 94 95 97 98 98
chicke 4.1.1 4.1.2	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.4 Lipid oxid 4.1.2.1 4.1.2.2 4.1.2.3 Protein of 4.1.3.1 4.1.3.2	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation Extracted lipid Primary product: conjugated dienes (CD) Secondary product: malondialdehydes (MDA) co-oxidation Soluble protein content Amino acid composition Tertiary product: protein	83 88 90 93 94 94 95 97 97 98 98 101
chicke 4.1.1 4.1.2	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.4 Lipid oxid 4.1.2.1 4.1.2.2 4.1.2.3 Protein oxid 4.1.3.1 4.1.3.2 4.1.3.3	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation Extracted lipid Primary product: conjugated dienes (CD) Secondary product: malondialdehydes (MDA) co-oxidation Soluble protein content Amino acid composition Tertiary product: protein carbonyl Tryptophan loss	 83 88 90 93 94 94 95 97 98 98 101 104
chicke 4.1.1 4.1.2	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.4 Lipid oxid 4.1.2.1 4.1.2.2 4.1.2.3 Protein oxid 4.1.3.1 4.1.3.2 4.1.3.3 4.1.3.4	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation Extracted lipid Primary product: conjugated dienes (CD) Secondary product: malondialdehydes (MDA) co-oxidation Soluble protein content Amino acid composition Tertiary product: protein carbonyl	 83 88 90 93 94 94 95 97 98 98 101 104 105
chicke 4.1.1 4.1.2	Physicod 4.1.1.1 4.1.1.2 4.1.1.3 4.1.1.3 4.1.1.4 Lipid oxid 4.1.2.1 4.1.2.2 4.1.2.3 Protein oc 4.1.3.1 4.1.3.2 4.1.3.3 4.1.3.4 4.1.3.5	chemical quality assessment Proximate composition, water activity (aw), and reducing sugar Fatty acid composition (FAC) Visual colour changes and colour measurement Scanning electron microscopy (SEM) dation Extracted lipid Primary product: conjugated dienes (CD) Secondary product: malondialdehydes (MDA) co-oxidation Soluble protein content Amino acid composition Tertiary product: protein carbonyl Tryptophan loss Tertiary product: Schiff base	 83 88 90 93 94 94 95 97 98 98 101 104 105

4.2	Effects of storage temperature, antioxidant and packaging materials on lipid oxidation and protein co-oxidation of shredded (<i>serunding</i>) chicken			
	4.2.1	Physicoc	chemical quality assessment Proximate composition and fatty acid composition (FAC)	110 110
		4.2.1.2	Scanning electron microscopy (SEM)	111
		4.2.1.3	Colour measurement	112
	4.2.2	Lipid oxid	dation	117
		4.2.2.1		117
		4.2.2.2		119
			dienes (CD)	
		4.2.2.3	Secondary product: malondialdehydes (MDA)	121
		4.2.2.4	Total antioxidant capacity (TAC) of shredded	
			(serunding) chicken during	
			storage	
	4.2.3	Protein c	co-oxidation	129
			Soluble protein content	129
		4.2.3.2	Tryptophan loss	131
		4.2.3.3	Tertiary product: protein carbonyl	133
		4.2.3.4	Tertiary product: Schiff base fluorescence	135
	4.2.4	Principal	component analysis (PCA)	137
			ISION AND S FOR FUTURE RESEARCH	139
5.1				139
5.2		al conclus	ion	140
5.3	-		on for future research	141
REFERENCES APPENDICES BIODATA OF S LIST OF PUBLI				143 172 209 210

LIST OF TABLES

Table		Page
2.1	Classification of ready-to-eat (RTE)	6
2.2	Nutritional composition in several raw, cooked meat cuts and typical meat products from different animal sources. All products are standardised to a 100g serving	9
2.3	Fatty acid composition (g/100g of fatty acids) of raw, cooked meat cuts and typical meat products from different animal sources	14
2.4	Amino acid compositions of raw, cooked meat cuts and typical meat products from different animal sources	18
2.5	Review of prior research on lipid and/or protein oxidation in meat and meat products during storage following cooking/manufacturing	28
2.6	Advantages and disadvantages of synthetic and natural antioxidants, respectively	54
3.1	Procedures for protein extraction of shredded (<i>serunding</i>) beef and chicken	69
4.1	Proximate analysis (%), a _w and reducing sugar content (mg glucose/g <i>serunding</i>) for shredded (<i>serunding</i>) beef and chicken obtained from present data and reported data	84
4.2	Effect of storage time and temperature on water activity, aw values for shredded (<i>serunding</i>) beef and chicken	86
4.3	Fatty acid composition (%) of shredded (<i>serunding</i>) beef and chicken	89
4.4	Amino acid composition (mg/g protein) of raw beef and chicken used in the production of shredded (<i>serunding</i>) beef and chicken	101
4.5	Amino acid composition of shredded (<i>serunding</i>) beef and chicken at week 0 and week 24 of storage	102
4.6	Proximate composition (%) and (b) fatty acid composition (%) for shredded (<i>serunding</i>) chicken obtained from present and reported data	110
4.7	Legend description for Figure 4.16–4.21 and 4.25–4.28	113

- 4.8 The total colour difference (ΔE_{0-24}) of the effect of antioxidant 117 addition in the shredded (*serunding*) chicken incubated at 25, 40 and 60°C in 24 weeks of storage
- 4.9 The total colour difference (ΔE_{0-24}) of the different packaging 117 materials used to pack shredded (*serunding*) chicken incubated at -80 and 25°C in 24 weeks of storage



LIST OF FIGURES

	Figure		Page
	2.1	Global ready-to-eat (RTE) food market forecast from 2016 to 2026	5
	2.2	Trends in global demand for meats (in tons) predictions for 2050	7
	2.3	Major lipid groups in meat	12
	2.4	Shredded (serunding) meat	23
	2.5	Shredded cooked meat-based products in different parts of the world	23
	2.6	Alkoxyl and peroxyl radical's decomposition into secondary oxidation products	34
	2.7	Shifting in double bond position caused by the formation of radicals in polyunsaturated fatty acids	38
	2.8	The initial proposed mechanisms of MDA formation	40
	2.9	The effect of lipid radicals with proteins	43
	2.10	Mechanism involved between MDA and lysine in the formation of protein carbonyl	45
	2.11	The enol and enolate forms of MDA	46
	2.12	The potential reaction sites of Micheal-type addition at the carbocations	46
	2.13	The reaction between 2,4-dinitrophenylhydrazine and protein carbonyl groups to form a hydrazone which may be determined spectrophotometrically at 370 nm.	47
	2.14	Example of the protein-lipid covalent bond	48
	2.15	Formation of imines and enamines from Schiff base additions of primary and secondary amines with carbonyls	48
	2.16	Oxidation of tryptophan amino acid	49
	2.17	General reaction of antioxidant addition to free radicals	50
	2.18	Example of synthetic antioxidants	51

2.19	BHA mechanism during oxidation reaction				
3.1	Multipurpose heating machine with a continuous stirrer for shredded (<i>serunding</i>) meat production				
3.2	Experimental design investigating the effect of meat types, storage time and storage temperatures on the physicochemical, lipid oxidation and protein co-oxidation of beef and chicken <i>serunding</i>				
3.3	Storage experiment design	62			
3.4	Product formulation of shredded (<i>serunding</i>) beef and chicken	63			
3.5	Glucose standard calibration curve (DNS assay)	64			
3.6	Lipid extraction method	66			
3.7	Malondialdehyde (MDA) standard calibration curve (TBARS assay)	68			
3.8	Protein standard calibration curve (Bradford assay)	70			
3.9	External standard chromatogram of amino acids	71			
3.10	Experimental design investigating the effect of storage temperatures, antioxidant and packaging materials on the physicochemical, lipid oxidation and protein co-oxidation of shredded (serunding) chicken.	73			
3.11	Trolox standard calibration curve (DPPH assay)	77			
3.12	Trolox standard calibration curve (ABTS assay)	79			
3.13	Trolox standard calibration curve (FRAP assay)	81			
4.1	Visible colour changes in shredded (<i>serunding</i>) beef and chicken at earlier week and towards the end of the 24 weeks storage	91			
4.2	Colour measurements of lightness (L* values) on shredded (<i>serunding</i>) beef and chicken at different storage temperatures during 6 months storage	92			
4.3	Colour measurements of redness (a* values) on shredded (<i>serunding</i>) beef and chicken at different storage temperatures during 6 months storage	92			

G

4.4	Colour measurements of yellowness (b* values) on shredded (<i>serunding</i>) beef and chicken at different storage temperatures during 6 months storage	93
4.5	SEM images of: (A1) shredded (<i>serunding</i>) beef (BS) at magnification ×55; (A2) BS at magnification ×330; (A3) BS at magnification ×600; (B1) shredded (<i>serunding</i>) chicken (CS) at magnification ×55; (B2) CS at magnification ×330; (B3) CS at magnification ×600	94
4.6	Lipid oxidation markers in terms of lipid extractability for shredded (<i>serunding</i>) beef and chicken at different storage temperatures during 6 months storage	95
4.7	Lipid oxidation markers in terms of conjugated dienes (CD) for shredded (<i>serunding</i>) beef and chicken at different storage temperatures during 6 months storage	96
4.8	Lipid oxidation markers in terms of malondialdehydes (MDA) for shredded (<i>serunding</i>) beef and chicken at different storage temperatures during 6 months storage	97
4.9	Procedures of protein extraction for shredded (<i>serunding</i>) beef and chicken	99
4.10	Protein co-oxidation markers in terms of protein solubility for shredded (<i>serunding</i>) beef and chicken at different storage temperatures during 6 months storage	100
4.11	Protein co-oxidation markers in terms of protein carbonyl for shredded (<i>serunding</i>) beef and chicken at different storage temperatures during 6 months storage	104
4.12	Protein co-oxidation markers in terms of tryptophan loss for shredded (<i>serunding</i>) beef and chicken at different storage temperatures during 6 months storage	105
4.13	Protein co-oxidation markers in terms of Schiff base for shredded (<i>serunding</i>) beef and chicken at different storage temperatures during 6 months storage	107
4.14	Principal component analysis on colour, lipid and protein markers shredded (<i>serunding</i>) of beef and chicken stored under different conditions (storage time and temperature), expressed as (a) loading plot and (b) score plot	108
4.15	SEM images of shredded (<i>serunding</i>) chicken (CS) samples: (A1) CS at magnification ×55; (A2) CS at magnification ×330; (B1) CS added with BHA at	112

magnification ×55; (B2) CS added with BHA at magnification ×330

- 4.16 Colour measurements of lightness (L* values) on shredded 114 (*serunding*) chicken in packaging A and B during 6 months storage
- 4.17 Colour measurements of redness (a* values) on shredded 115 (*serunding*) chicken in packaging A and B during 6 months storage
- 4.18 Colour measurements of yellowness on shredded 116 (*serunding*) chicken in packaging A and B during 6 months storage
- 4.19 Changes in lipid oxidation markers, extracted lipid (%) of 118 shredded (*serunding*) chicken at different storage temperatures in packaging A and B during 6 months storage
- 4.20 Changes in lipid oxidation markers, conjugated dienes 120 (CD) of shredded (*serunding*) chicken at different storage temperatures in packaging A and B during 6 months storage
- 4.21 Changes in lipid oxidation markers, Malondialdehydes 122 (MDA) of shredded (*serunding*) chicken at different storage temperatures in packaging A and B during 6 months storage
- 4.22 ABTS assay of shredded (*serunding*) chicken in packaging 125 A and B, with and without BHA addition during 6 months storage
- 4.23 DPPH assay of shredded (*serunding*) chicken in packaging 126 A and B, with and without BHA addition during 6 months storage
- 4.24 FRAP assay of shredded (*serunding*) chicken in packaging 128 A and B, with and without BHA addition during 6 months storage
- 4.25 Changes in protein oxidation markers, protein solubility of 130 shredded (*serunding*) chicken at different storage temperatures during 6 months storage
- 4.26 Changes in protein oxidation markers, tryptophan content 132 (expressed as fluorescence unit, a.u.) of shredded (*serunding*) chicken at different storage temperatures during 6 months storage

- 4.27 Changes in protein oxidation markers, protein carbonyls of 134 shredded (serunding) chicken at different storage temperatures during 6 months storage
- 4.28 Changes in protein oxidation markers, Schiff base 136 (expressed as fluorescence unit, a.u.) of shredded (serunding) chicken at different storage temperatures during 6 months storage
- 4.29 Principal component analysis on colour, lipid and protein markers of shredded (serunding) chicken stored under different conditions (storage temperature, presence of antioxidant, packaging material), expressed as (a) Loading plot and (b) Score plot

LIST OF APPENDICES

Appendix		Page	
A	Effect of storage time and temperature on the moisture content, % for shredded (<i>serunding</i>) beef and chicken		
В	Effect of storage time and temperature on the amino acid 1 composition (AA) of shredded (<i>serunding</i>) beef and chicken		
С	Figure guidelines for effectively presenting control samples (Section 4.2)	185	
D	Statistical data analysis performed using Minitab® statistical software (Section 4.1)	186	

LIST OF ABBREVIATIONS

ΔE_{0-24}	Total colour difference throughout storage
a*	Redness
a _w	Water activity
ABTS	2,2'-Azinobis-(3-ethylbenzothiazoline-6-sulphonic acid)
ALU	Aluminium
AOAC	Association of Official Analytical Chemists
AOCS	American Oil Chemists' Society
b*	Yellowness
BHA	Butylated hydroxyanisole
CD	Conjugated dienes
CS	Chicken serunding
CS-BHA	Chicken serunding with BHA addition
CHCl₃	Chloroform
DNPH	2,4-Dinitrophenyl hydrazine
DPPH	2,2-Diphenyl-1-Picrylhydrazyl
FAO	Food and Agriculture Organization of the United Nations
FRAP	Ferric Reducing Antioxidant Power
L*	Lightness
LLDPE	Linear low-density polyethylene
LO	Alkoxyl radical
LOO	Peroxide radical
LOOH	Hydroperoxide
М	Molar
MCT	Medium chain tryglycerides

	MDA	Malondialdehyde
	MPET	Metallised-polyethylene terephthalate
	MRE	Meals Ready-to-Eat
	MS	Mass spectrometry
	MUFA	Monounsaturated fatty acids
	MΩ	Mili-Q
	Р	Probability (of error)
	Packaging A	PET/PE/MPET (High Barrier)/LLDPE
	Packaging B	PET/PE/ALU/LLDPE
	PBS	Phosphate buffered saline
	PC	Protein carbonyl
	PCA	Principal component analysis
	PE	Polyethylene
	PET	Polyethylene terephthalate
	PITC	Phenyl-isothiocyanate
	PUFA	Polyunsaturated fatty acids
	RTE	Ready-to-eat
	SEM	Scanning electron microscopy
	SFA	Saturated fatty acids
	TAC	Total antioxidant capacity
	TBARS	Thiobarbituric acid reactive substances
\bigcirc	ТСА	Trichloroacetic acid

xxv

CHAPTER 1

INTRODUCTION

1.1 Background of the study

Ready-to-eat (RTE) food products need no additional preparation beyond freezing, cooking, processing before consumption, or little preparation such as boiling or reheating (Howard et al., 2012). Recent trends have observed an increase in consumer demands for convenience in meal preparation, shelf-stable foods as life have gotten more hectic. Faced with increased working hours and lengthy commutes, consumers worldwide seek to maximise their extremely limited free time (Montero et al., 2021). In response to this need, the food industry has expanded its availability of complete pre-prepared ready meal solutions, including ready-to-eat (RTE) options (Diaz, 2021).

RTE food products are a boon for crises like earthquakes, flash floods, and the current global COVID-19 lockdown (Kumar et al., 2016; Telukdarie et al., 2020). With the current pandemic, RTE food products, specifically RTE meat products. become a practical means of providing nutrient-dense sustenance. However, meat-based RTE is lipid and protein-rich, thus susceptible to oxidative deterioration during long-term storage. This study thus assessed the oxidation of such food system using serunding, a locally popular RTE shredded meat product that is cooked under long-hour from beef or chicken meat, known for its high-lipid (6.0–32.3%), high-protein (21.8–40.7%) and low-moisture (4.3–13.6%) (Huda et al., 2012; Knechtges, 2012), as the representative food model. Serunding is a popular specialty dish in Southeast Asia, with distinct flavours derived from a combination of cooked meat, coconut milk, herbs and spices. In general, serunding is acknowledged as shredded meat, meat floss or meat fibre. Serunding is accessible all year in Malaysia, particularly during festive season. Serunding offers convenience to be readily consumed together with staple foods like rice and bread and easy storage up to 6 months without refrigeration.

1.2 **Problem statements**

In the context of food science, *serunding* is categorised as high-lipid, high-protein food system which is highly susceptible to oxidation, a major problem leading to discolouration and shelf-life reduction, sensory and nutritional quality degradation, by developing off-flavour, off-odour, rancidity in lipid molecules and the production of potentially toxic compounds that could risk consumers' safety (Zahid et al., 2020; Gutiérrez-del-Río et al., 2021). The loss of quality in meat and ready-to-eat meat products due to lipid oxidation and protein co-oxidation reactions has impacted the food processing industry. It has resulted in customer

rejection and economic losses. Therefore, controlling oxidative processes is crucial and a challenge for the food industry (Domínguez et al., 2019; Gutiérrezdel-Río et al., 2021; Domínguez et al., 2022).

During lipid oxidation, non-lipid substrates (majorly protein) are spontaneously attacked by highly reactive, protein bound, lipid oxidation-derived aldehyde such as malondialdehyde (MDA) (Requena et al., 1996; Vandemoortele et al., 2019). MDA, an α , β -unsaturated aldehyde, is electrophilic and attacks the nucleophilic groups on proteins, causing unfavourable alteration in the protein structure and stability (Fuentes et al., 2014; Wang et al., 2019), leading to the formation of Schiff base adducts (MDA-modified amino acid residues, predominantly MDA lysine adducts) (Traverso et al., 2004) and a subsequent occurrence of protein co-oxidation, adversely affecting the product functionality and performance. According to Wang et al. (2019), MDA could also promote protein carbonylation and loss of tryptophan fluorescence in the myofibrillar protein while Hellwig (2020) termed the reaction of protein with oxidised lipid product as "lipation", where bulky modifying group on lipid is covalently attached to the nucleophilic site on amino acid. While the effect of protein oxidation/co-oxidation on meat quality is still being investigated in numerous studies, it is generally accepted that it results in the loss of essential amino acids, decreased protein solubility, physicochemical changes, altered protein functionality and formation of protein carbonyl derivatives (Lund et al., 2011). Previous research on the effects of lipid oxidation on protein oxidation has been done in both high-moisture raw and lowmoisture cured meat (Fuentes et al., 2010; Fuentes et al., 2014; Wang et al., 2019). However, these studies concentrated on lipid-protein oxidation because of the processing conditions. Until now, no research has been done on the effect of storage conditions on lipid oxidation and protein co-oxidation in low-moisture, ready-to-eat, cooked meat. To that end, recent work merits special attention since it is a pioneer in examining how a high-lipid, high-protein, low-moisture, RTE shredded meat system analyses the simultaneous progression of lipid oxidation and protein co-oxidation by using two different shredded meats (beef and chicken), three different storage temperatures (25, 40 and 60 °C) within six months storage.

It is hypothesised that the shredded (*serunding*) chicken could have a faster rate of lipid-protein oxidative degradation than shredded (*serunding*) beef throughout long-term storage. The hypothesis is supported by the presence of higher amounts of PUFA in chicken meat than in beef, making shredded (*serunding*) chicken a more favourable substrate to initiate lipid oxidation (Angelo, 2012; Gruffat et al., 2021). Antioxidant's addition could be added to tackle faster lipidprotein oxidative degradation issue in shredded (*serunding*) chicken meat. Currently, synthetic antioxidants are added commercially to various food such as baked goods, fried food, processed meat, dried cereal, processed potatoes, dessert mix and beverages (Movileanu et al., 2013; Shahidi, 2015). Despite the potential side effects of synthetic antioxidant, its application in the food industry is ongoing (Admassu, 2019) to prolong the shelf-life of lipid-rich food products at a low cost. The lipid-soluble (lipophilic) BHA antioxidant was chosen to evaluate the effect of antioxidant addition on lipid oxidation and protein co-oxidation in the shredded (*serunding*) meat because the shredded (*serunding*) meat has a high fat content (3.20–39.00%) (Huda et al., 2012; Sukisman et al., 2014; Sukisman et al., 2018). This facilitates the complete dissolution of BHA during the production of shredded (*serunding*) meat. In addition, the enormous sample size investigated in the present study made it difficult to quantify the effect of various antioxidants; hence a single antioxidant was selected. Aside from antioxidants, packaging with moisture and oxygen barriers are both critical for protecting RTE meat products from lipid-protein oxidative degradation. Considering that oxygen is the most prevalent and essential component for the simultaneous progression of lipid-protein oxidation, packaging that prevents or limits oxygen exposure is an effective technique for preventing and retarding oxidation (Gruffat et al., 2021).

1.3 Objectives of the study

The overall aim of this current study was to determine the physical properties and chemical characteristics of lipid-protein oxidative degradation of ready-toeat (RTE) low-moisture shredded (*serunding*) meats during long-term storage.

Specifically, the objectives were:

- 1. To determine the effect of long-term storage (six months) at different temperatures (25, 40, and 60°C) on simultaneous lipid oxidation and protein co-oxidation in two different meat types of RTE low-moisture shredded (*serunding*) beef and chicken.
- To evaluate the correlation of the different storage times and storage temperatures on simultaneous lipid oxidation and protein co-oxidation in two different meat types of RTE low-moisture shredded (*serunding*) beef and chicken by using principal component analysis (PCA).
- 3. To assess the interacting effect of the different storage temperatures (25, 40, and 60°C), antioxidant butylated hydroxyanisole (BHA) and multilayer packaging materials (metallised polyethene terephthalate (MPET) and aluminium containing packaging) on simultaneous lipid oxidation and protein co-oxidation in RTE low-moisture shredded (*serunding*) chicken during six months of storage.
- 4. To evaluate the interaction of the effect of different storage temperatures, storage time, antioxidant addition and packaging material on simultaneous lipid oxidation and protein co-oxidation in RTE low-moisture shredded (*serunding*) chicken during six months of storage by using PCA.

REFERENCES

- Abeyrathne, E. D. N. S., Nam, K., & Ahn, D. U. (2021). Analytical Methods for Lipid Oxidation and Antioxidant Capacity in Food Systems. *Foods*, 10(10), 1587.
- Adachi, S., Ishiguro, T., & Matsuno, R. (1995). Autoxidation Kinetics for Fatty Acids and Their Esters. *Journal of the American Oil Chemists' Society*, 72(5), 547.
- Admassu, S. (2019). Application of Antioxidants in Food Processing Industry: Options to Improve the Extraction Yields and Market Value of Natural Products. Advances in Food Technology and Nutritional Sciences, 5(2), 38-49.
- Ahmad, R. S., Imran, A., & Hussain, M. B. (2018). Nutritional Composition of Meat. *Meat Science And Nutrition*, *61*(10.5772).
- Ahmad, S., & Augustin, M. A. (1985). Effect of Tertiarybutylhydroquinone on Lipid Oxidation in Fish Crackers. *Journal of the Science of Food Agriculture, 36*(5), 393-401.
- Al-Hijazeen, M., Lee, E. J., Mendonca, A., & Ahn, D. U. (2016). Effects of Tannic Acid on Lipid and Protein Oxidation, Color, and Volatiles of Raw and Cooked Chicken Breast Meat During Storage. *Antioxidants, 5*(2), 19.
- Alfaia, C. M., Alves, S. P., Lopes, A. F., Fernandes, M. J., Costa, A. S., Fontes, C. M., Castro, M. L., Bessa, R. J., & Prates, J. A. (2010). Effect of Cooking Methods on Fatty Acids, Conjugated Isomers of Linoleic Acid and Nutritional Quality of Beef Intramuscular Fat. *Meat Science*, 84(4), 769-777.
- Alojaly, H., & Benyounis, K. Y. (2020). Packaging with Plastics and Polymeric Materials. In *Reference Module in Materials Science and Materials Engineering*: Elsevier.
- Alyaqoubi, S., Abdullah, A., Samudi, M., Abdullah, N., Addai, Z. R., & Musa, K. H. (2015). Study of Antioxidant Activity and Physicochemical Properties of Coconut Milk (*Pati Santan*) in Malaysia. *Journal of Chemical and Pharmaceutical Research*, 7(4), 967-973.
- Amaral, A. B., Silva, M. V., & Lannes, S. C. S. (2018). Lipid Oxidation in Meat: Mechanisms and Protective Factors–a Review. *Food Science and Technology*, 38, 1-15.
- Anderson, G. H., Ashley, D. V. M., & Jones, J. D. (1976). Utilization of L-Methionine Sulfoxide, L-Methionine Sulfone and Cysteic Acid by the Weanling Rat. *The Journal of Nutrition*, *106*(8), 1108-1114.

Angelo, A. J. S. (2012). Warmed-over Flavor of Meat. Elsevier Science.

- AOAC. (2005a). Official Methods of Analysis (18th ed.). Maryland: AOAC International.
- AOAC. (2005b). Official Methods of Analysis of Aoac International (18th Ed.). In The Association of Official Analytical Chemists. Maryland: North Fredick Avenue Gaithersburg: AOAC International.
- AOCS. (2005). Aocs Official Method Ce 1-62. In. Urbana, IL: The American Oil Chemists' Society.
- AOCS. (2011). Aocs Official Method Th 1a-64. In. Urbana, IL: The American Oil Chemists' Society.
- Asian Inspirations. (2019). Beef Floss (*Serunding Daging*). Retrieved from https://asianinspirations.com.au/recipes/beef-floss-serunding-daging
- Aviles, M. V., Naef, E. F., Abalos, R. A., Lound, L. H., Olivera, D. F., & García-Segovia, P. (2020). Effect of Familiarity of Ready-to-Eat Animal-Based Meals on Consumers' Perception and Consumption Motivation. International Journal of Gastronomy and Food Science, 21, 100225.
- Ayala, A., Muñoz, M. F., & Argüelles, S. (2014). Lipid Peroxidation: Production, Metabolism, and Signaling Mechanisms of Malondialdehyde and 4-Hydroxy-2-Nonenal. Oxidative Medicine and Cellular Longevity, 2014, 360438.
- Azima, F. (2016). Chemical Characteristic and Fatty Acid Profile in Rendang of Minangkabau. International Journal on Advanced Science, Engineering and Information Technology, 6(4), 465-468.
- Babji, A. S., & Yusof, S. C. M. (1995). The Nutritional Value of Some Processed Meat Products in Malaysia. *Malaysian Journal of Nutrition*, 1(1), 83-94.
- Baker, P., Machado, P., Santos, T., Sievert, K., Backholer, K., Hadjikakou, M., Russell, C., Huse, O., Bell, C., Scrinis, G., Worsley, A., Friel, S., & Lawrence, M. (2020). Ultra-Processed Foods and the Nutrition Transition: Global, Regional and National Trends, Food Systems Transformations and Political Economy Drivers. *Obesity Reviews*, 21(12), e13126.
- Baldioli, M., Servili, M., Perretti, G., & Montedoro, G. F. (1996). Antioxidant Activity of Tocopherols and Phenolic Compounds of Virgin Olive Oil. *Journal of the American Oil Chemists' Society*, *73*(11), 1589-1593.
- Barden, L., & Decker, E. A. (2016). Lipid Oxidation in Low-Moisture Food: A Review. *Critical Reviews in Food Science and Nutrition*, 56(15), 2467-2482.

- Baskaran, S., Ayob, S. A., Howe, N. C., & Mahadi, N. (2017). Understanding Purchase Intention of Ready-to-Eat Food among Malaysian Urbanites: A Proposed Framework. *International Journal of Academic Research in Business and Social Sciences*, 7, 566-579.
- Bell, R. G. (2012). Meat Packaging: Protection, Preservation, and Presentation. In *Handbook of Meat and Meat Processing* (Second Edition ed.).
- Beltrán, J. A., & Bellés, M. (2019). Effect of Freezing on the Quality of Meat. In P. Ferranti, E. M. Berry, & J. R. Anderson (Eds.), *Encyclopedia of Food Security and Sustainability* (pp. 493-497). Oxford: Elsevier.
- Benzie, I. F., & Strain, J. J. (1999). Ferric Reducing/Antioxidant Power Assay: Direct Measure of Total Antioxidant Activity of Biological Fluids and Modified Version for Simultaneous Measurement of Total Antioxidant Power and Ascorbic Acid Concentration. *Methods Enzymol*, 299, 15-27.
- Benzie, I. F. F., & Choi, S.-W. (2014). Chapter One Antioxidants in Food: Content, Measurement, Significance, Action, Cautions, Caveats, and Research Needs. In J. Henry (Ed.), Advances in Food and Nutrition Research (Vol. 71, pp. 1-53): Academic Press.
- Benzie, I. F. F., & Strain, J. J. (1996). The Ferric Reducing Ability of Plasma (Frap) as a Measure of "Antioxidant Power": The Frap Assay. *Analytical Biochemistry*, 239(1), 70-76.
- Bland, J. M., Grimm, C. C., Bechtel, P. J., Deb, U., & Dey, M. M. (2021). Proximate Composition and Nutritional Attributes of Ready-to-Cook Catfish Products *Foods* 10(11), 2716.

Food Act 1983 (Act 281) & Regulations (as at 25th August 2019), (2019).

- Bodwell, C., & Anderson, B. (1986). Nutritional Composition and Value of Meat and Meat Products. *Muscle as Food*, 321-369.
- Böhm, V. (2000). Measuring of Antioxidative Capacity-Methods and Assessment. In *Ernahrungs-Umschau* (Vol. 47, pp. 372-375).
- Bohrer, B. M. (2017). Review: Nutrient Density and Nutritional Value of Meat Products and Non-Meat Foods High in Protein. *Trends in Food Science* & *Technology*, *65*, 103-112.
- Bohrer, B. M. (2019). An Investigation of the Formulation and Nutritional Composition of Modern Meat Analogue Products. *Food Science and Human Wellness*, 8(4), 320-329.
- Boler, D. D., & Woerner, D. R. (2017). What Is Meat? A Perspective from the American Meat Science Association. *Animal Frontiers*, 7(4), 8-11.

- Botsoglou, E., Govaris, A., Ambrosiadis, I., Fletouris, D., & Papageorgiou, G. (2014). Effect of Olive Leaf (*Olea Europea* L.) Extracts on Protein and Lipid Oxidation in Cooked Pork Meat Patties Enriched with N-3 Fatty Acids. *Journal of the Science of Food Agriculture*, 94(2), 227-234.
- Bower, C., Schilke, K., & Daeschel, M. (2003). Antimicrobial Properties of Raisins in Beef Jerky Preservation. *Journal of Food Science*, 68(4), 1484-1489.
- Bradford, M. M. (1976). A Rapid and Sensitive Method for the Quantitation of Microgram Quantities of Protein Utilizing the Principle of Protein-Dye Binding. *Analytical Biochemistry*, 72(1-2), 248-254.
- Brand-Williams, W., Cuvelier, M. E., & Berset, C. (1995). Use of a Free Radical Method to Evaluate Antioxidant Activity. *LWT - Food Science and Technology*, 28(1), 25-30.
- Butterfield, D. A., & Stadtman, E. R. (1997). Chapter 7: Protein Oxidation Processes in Aging Brain. In P. S. Timiras & E. E. Bittar (Eds.), Advances in Cell Aging and Gerontology (Vol. 2, pp. 161-191): Elsevier.
- Čabarkapa-Pirković, A., Živković, L., Dekanski, D., Topalović, D., & Spremo-Potparević, B. (2021). Chapter 38 - Olive Leaf, DNA Damage and Chelation Therapy. In V. R. Preedy & R. R. Watson (Eds.), Olives and Olive Oil in Health and Disease Prevention (Second Edition) (pp. 457-469). San Diego: Academic Press.
- Caire-Juvera, G., Vázquez-Ortiz, F. A., & Grijalva-Haro, M. I. (2013). Amino Acid Composition, Score and in Vitro Protein Digestibility of Foods Commonly Consumed in Northwest Mexico. *Nutricion Hospitalaria*, 28(2), 365-371.
- Carlsen, C., Møller, J., & Skibsted, L. (2005). Heme-Iron in Lipid Oxidation. Coordination Chemistry Reviews, 249, 485-498.
- Carocho, M., Morales, P., & Ferreira, I. C. F. R. (2018). Antioxidants: Reviewing the Chemistry, Food Applications, Legislation and Role as Preservatives. *Trends in Food Science & Technology, 71*, 107-120.
- Carrascal, J. R., Hidalgo, J., & Morillo, M. (2004, 5–8 September 2). *Fatty Acid Composition of Phospholipid Classes in Beef.* Paper presented at the 3rd Euro Fed Lipid Congress, Edinburgh, Scotland.
- Cartoni Mancinelli, A., Silletti, E., Mattioli, S., Dal Bosco, A., Sebastiani, B., Menchetti, L., Koot, A., van Ruth, S., & Castellini, C. (2021). Fatty Acid Profile, Oxidative Status, and Content of Volatile Organic Compounds in Raw and Cooked Meat of Different Chicken Strains. *Poultry Science*, *100*(2), 1273-1282.

- Carvalho, R. H., Ida, E. I., Madruga, M. S., Martínez, S. L., Shimokomaki, M., & Estévez, M. (2017). Underlying Connections between the Redox System Imbalance, Protein Oxidation and Impaired Quality Traits in Pale, Soft and Exudative (Pse) Poultry Meat. *Food Chemistry*, 215, 129-137.
- Catalán, V., Frühbeck, G., & Gómez-Ambrosi, J. (2018). Chapter 8: Inflammatory and Oxidative Stress Markers in Skeletal Muscle of Obese Subjects. In A. M. Del Moral & C. M. Aguilera García (Eds.), *Obesity* (pp. 163-189): Academic Press.
- Cenci-Goga, B. T., Iulietto, M. F., Sechi, P., Borgogni, E., Karama, M., & Grispoldi, L. (2020). New Trends in Meat Packaging. *Microbiology Research 11*(2), 56-67.
- Chandra, P., Sharma, R. K., & Arora, D. S. (2020). Antioxidant Compounds from Microbial Sources: A Review. *Food Research International, 129*, 108849.
- Chaurasiya, R., Pandey, R., Verma, P., Kek, X. H., Kee, D. M. H., Yeoh, X. Y., Wah, P. J., & Rokiah, R. (2020). Consumer Behavior Towards Readyto-Eat (Rte) Market: A Study of Mtr Foods. *International Journal of Applied Business and International Management*, 5(2), 66-72.
- Cheah, Y. K., Abdul Adzis, A., & Islam, R. (2022). Factors Associated with Household Expenditure on Meat. *Journal Of International Business, Economics And Entrepreneurship, 7*(1), 91.
- Chelh, I., Gatellier, P., & Santé-Lhoutellier, V. (2007). Characterisation of Fluorescent Schiff Bases Formed During Oxidation of Pig Myofibrils. *Meat Science*, *76*(2), 210-215.
- Chen, X., Xu, X., Liu, D., Zhou, G., Han, M., & Wang, P. (2018). Rheological Behavior, Conformational Changes and Interactions of Water-Soluble Myofibrillar Protein During Heating. *Food Hydrocolloids*, 77, 524-533.
- Cheng, H., Erichsen, H., Soerensen, J., Petersen, M. A., & Skibsted, L. H. (2019). Optimising Water Activity for Storage of High Lipid and High Protein Infant Formula Milk Powder Using Multivariate Analysis. *International Dairy Journal*, 93, 92-98.
- Cheng, J. H. (2016). Lipid Oxidation in Meat. *Journal of Nutrition and Food Sciences* 6(3), 1-3.
- Cheng, Z., Moore, J., & Yu, L. (2006). High-Throughput Relative Dpph Radical Scavenging Capacity Assay. *Journal of Agricultural and Food Chemistry*, 54(20), 7429-7436.
- Childs, R. E., & Bardsley, W. G. (1975). The Steady-State Kinetics of Peroxidase with 2,2'-Azino-Di-(3-Ethyl-Benzthiazoline-6-Sulphonic Acid) as Chromogen. *The Biochemical Journal*, *145*(1), 93-103.

- Chio, K., & Tappel, A. L. (1969). Inactivation of Ribonuclease and Other Enzymes by Peroxidizing Lipids and by Malonaldehyde. *Biochemistry*, *8*(7), 2827-2832.
- Choi, M. J., Abduzukhurov, T., Park, D. H., Kim, E. J., & Hong, G. P. (2018). Effects of Deep Freezing Temperature for Long-Term Storage on Quality Characteristics and Freshness of Lamb Meat. *Food Science of Animal Resources*, 38(5), 959-969.
- Christie, W. W. (1978). The Composition, Structure and Function of Lipids in the Tissues of Ruminant Animals. *Progress in Lipid Research, 17*(2), 111-205.
- Coles, R., & Kirwan, M. J. (2011). Food and Beverage Packaging Technology: John Wiley & Sons.
- Comer, F. W. (1979). Functionality of Fillers in Comminuted Meat Products. Canadian Institute of Food Science and Technology Journal, 12(4), 157-165.
- Cropp, M. S. (2022). Can Nitrite-Embedded Packaging Film Induce Curing Reactions in Meat Post-Thermal Processing? (Ph.D.), Iowa State University, Ann Arbor. Retrieved from <u>https://www.proquest.com/dissertations-theses/can-nitrite-embeddedpackaging-film-induce-curing/docview/2681066872/se-2?accountid=27932</u> ProQuest Dissertations & Theses Global database. (28965454)
- Cuq, J. L., Besancon, P., Chartier, L., & Cheftel, C. (1978). Oxidation of Methionine Residues of Food Proteins and Nutritional Availability of Protein-Bound Methionine Sulphoxide. *Food Chemistry*, *3*(2), 85-102.
- Da Silva, S., Marangoni, C., Brum, D., Vendruscolo, R., Silva, M., de Moura, H., Rampelotto, C., Wagner, R., de Menezes, C., & Barin, J. (2018). Effect of Dietary Olive Leaves on the Lipid and Protein Oxidation and Bacterial Safety of Chicken Hamburgers During Frozen Storage. *International Food Research Journal, 25*(1), 383-391.
- Dalle-Donne, I., Carini, M., Orioli, M., Vistoli, G., Regazzoni, L., Colombo, G., Rossi, R., Milzani, A., & Aldini, G. (2009). Protein Carbonylation: 2,4-Dinitrophenylhydrazine Reacts with Both Aldehydes/Ketones and Sulfenic Acids. *Free radical biology & medicine, 46*, 1411-1419.
- Dalziel, C. J., Kliem, K. E., & Givens, D. I. (2015). Fat and Fatty Acid Composition of Cooked Meat from Uk Retail Chickens Labelled as from Organic and Non-Organic Production Systems. *Food Chemistry*, 179, 103-108.
- Damodaran, S., & Parkin, K. L. (2017). Amino Acids, Peptides and Proteins. In *Fennema's Food Chemistry* (pp. 425-439): CRC Press.

- Damodaran, S., Parkin, K. L., & Fennema, O. R. (2007). *Fennema's Food Chemistry*: CRC Press.
- Dave, D., & Ghaly, A. E. (2011). Meat Spoilage Mechanisms and Preservation Techniques: A Critical Review. American Journal of Agricultural Biological Sciences, 6(4), 486-510.
- Davies, K. J., Delsignore, M. E., & Lin, S. W. (1987). Protein Damage and Degradation by Oxygen Radicals. Ii. Modification of Amino Acids. *Journal of Biological Chemistry*, 262(20), 9902-9907.
- Davies, M. J., & Dean, R. T. (2003). *Radical-Mediated Protein Oxidation*. Oxford: Oxford Science Publications.
- De Smet, S., & Vossen, E. (2016). Meat: The Balance between Nutrition and Health. A Review. *Meat Science, 120*, 145-156.
- Decker, E., Elias, R., & McClements, D. J. (2010). Oxidation in Foods and Beverages and Antioxidant Applications: Management in Different Industry Sectors: Elsevier Science.
- Del Rio, D., Stewart, A. J., & Pellegrini, N. (2005). A Review of Recent Studies on Malondialdehyde as Toxic Molecule and Biological Marker of Oxidative Stress. *Nutrition, Metabolism and Cardiovascular Diseases*, 15(4), 316-328.
- Diaz, M. L. M. (2021). Application of Emerging Sensory Methodologies to Characterize Ready-to-Eat-Meals. Washington State University,
- Domínguez, R., Pateiro, M., Gagaoua, M., Barba, F. J., Zhang, W., & Lorenzo, J. M. (2019). A Comprehensive Review on Lipid Oxidation in Meat and Meat Products. *Antioxidants, 8*(10), 429.
- Domínguez, R., Pateiro, M., Munekata, P. E. S., Zhang, W., Garcia-Oliveira, P., Carpena, M., Prieto, M. A., Bohrer, B., & Lorenzo, J. M. (2022). Protein Oxidation in Muscle Foods: A Comprehensive Review. *Antioxidants*, *11*(1), 60.
- Dong, Y. (2011). Protein Modifications in Baked Versus Fried Tortilla Chips. (Master Degree), Rutgers University-Graduate School-New Brunswick,
- Duangmal, K., Hempattarasuwan, P., & Somsong, P. (2015). Current Situation on Food Additives in Thailand: Use and Awareness. In *Food Security and Food Safety for the Twenty-First Century* (pp. 45-60): Springer.
- El Gharbawi, M. I., & Dugan Jr, L. R. (1965). Stability of Nitrogenous Compounds and Lipids During Storage of Freeze - Dried Raw Beef. *Journal of Food Science*, 30(5), 817-822.

- Erejuwa, O. O., Sulaiman, S. A., & Ab Wahab, M. S. (2013). Evidence in Support of Potential Applications of Lipid Peroxidation Products in Cancer Treatment. Oxidative Medicine and Cellular Longevity, 2013, 931251.
- Erickson, M. C. (2002). Lipid Oxidation of Muscle Foods. In C. C. Akoh & D. B. Min (Eds.), *Food Lipids: Chemistry, Nutrition, and Biotechnology* (pp. 384–430). New York, NY, USA: Marcel Dekker, Inc.
- Esteghlal, S., Gahruie, H. H., Niakousari, M., Barba, F. J., Bekhit, A. E.-D., Mallikarjunan, K., & Roohinejad, S. (2019). Bridging the Knowledge Gap for the Impact of Non-Thermal Processing on Proteins and Amino Acids. *Foods*, *8*(7), 262.
- Esterbauer, H., Schaur, R. J., & Zollner, H. (1991). Chemistry and Biochemistry of 4-Hydroxynonenal, Malonaldehyde and Related Aldehydes. *Free Radical Biology and Medicine*, *11*(1), 81-128.
- Estévez, M. (2011). Protein Carbonyls in Meat Systems: A Review. *Meat Science*, *89*(3), 259-279.
- Estévez, M. (2015). Oxidative Damage to Poultry: From Farm to Fork. *Poultry Science*, *94*(6), 1368-1378.
- Estévez, M. (2021). Critical Overview of the Use of Plant Antioxidants in the Meat Industry: Opportunities, Innovative Applications and Future Perspectives. *Meat Science, 181*, 108610.
- Estévez, M., Díaz-Velasco, S., & Martínez, R. (2021). Protein Carbonylation in Food and Nutrition: A Concise Update. *Amino Acids*.
- Estévez, M., & Heinonen, M. (2010). Effect of Phenolic Compounds on the Formation of A-Aminoadipic and Γ-Glutamic Semialdehydes from Myofibrillar Proteins Oxidized by Copper, Iron, and Myoglobin. *Journal* of Agricultural and Food Chemistry, 58(7), 4448-4455.
- Estévez, M., Padilla, P., Carvalho, L., Martín, L., Carrapiso, A., & Delgado, J. (2019a). Malondialdehyde Interferes with the Formation and Detection of Primary Carbonyls in Oxidized Proteins. *Redox Biology, 26*, 101277.
- Estévez, M., Ventanas, S., & Cava, R. (2006). Effect of Natural and Synthetic Antioxidants on Protein Oxidation and Colour and Texture Changes in Refrigerated Stored Porcine Liver Pâté. *Meat science, 74*(2), 396-403.
- Estévez, M., Ventanas, S., & Heinonen, M. (2011). Formation of Strecker Aldehydes between Protein Carbonyls – A-Aminoadipic and Γ-Glutamic Semialdehydes – and Leucine and Isoleucine. *Food Chemistry*, *128*, 1051-1057.

- Estévez, M., & Xiong, Y. (2019b). Intake of Oxidized Proteins and Amino Acids and Causative Oxidative Stress and Disease: Recent Scientific Evidences and Hypotheses. *Journal of Food Science*, *84*(3), 387-396.
- Fagan, J. M., Sleczka, B. G., & Sohar, I. (1999). Quantitation of Oxidative Damage to Tissue Proteins. *The International Journal of Biochemistry & Cell Biology*, 31(7), 751-757.
- Fanani, Z., Hartono, B., & Nugroho, B. A. (2015). The Influence of the Marketing Mix (Product, Price, Promotion, Place, Process, Entrepreneurs and Physical Evidence) to Customer Satisfaction and Loyalty in Buying Shredded Beef in Palu City, Indonesia. *International Journal of Economic Research*, 12(1).
- FAO. (2021, 25 November 2014). Meat and Meat Products: Processing Product Groups. Retrieved from http://www.fao.org/ag/againfo/themes/en/meat/Processing_product.htm l#:~:text=Fresh%20processed%20meat%20products,to%20make%20t he%20products%20palatable.
- FAO, W. H. O. (2015). Code of Hygienic Practice for Low-Moisture Foods. In. Rome, Italy.
- Farouk, M. M., Mills, J., & Bekhit, A. E.-D. A. (2017). Ready-to-Eat Processed Meats. In *Advances in Meat Processing Technology* (pp. 447-486): CRC Press.
- Feng, Y. H., Zhang, S. S., Sun, B. Z., Xie, P., Wen, K. X., & Xu, C. C. (2020). Changes in Physical Meat Traits, Protein Solubility, and the Microstructure of Different Beef Muscles During Post-Mortem Aging. *Foods*, 9(6), 806.
- Fernández, M., Ganan, M., Guerra, C., & Hierro, E. (2014). Protein Oxidation in Processed Cheese Slices Treated with Pulsed Light Technology. *Food Chemistry*, *159*, 388-390.
- Fields, R., & Dixon, H. B. (1971). Micro Method for Determination of Reactive Carbonyl Groups in Proteins and Peptides, Using 2,4-Dinitrophenylhydrazine. *The Biochemical Journal*, *121*(4), 587-589.
- Fillery-Travis, A., Mills, E. N. C., & Wilde, P. (2000). Protein-Lipid Interactions at Interfaces. Grasas Y Aceites, 51(1-2), 50-55.
- Food Act 1983. (2013). Food Act 1983 (Act 281) and Regulations: (as at 1st March 2013). Kuala Lumpur, Malaysia: International Law Book Services.
- Frankel, E. N. (2012). Chapter 7 Stability Methods. In E. N. Frankel (Ed.), *Lipid Oxidation (Second Edition)* (pp. 165-186): Woodhead Publishing.

- Fruehwirth, S., Egger, S., Flecker, T., Ressler, M., Firat, N., & Pignitter, M. (2021). Acetone as Indicator of Lipid Oxidation in Stored Margarine. *Antioxidants, 10*(1), 59.
- Fuentes, V., Estévez, M., Ventanas, J., & Ventanas, S. (2014). Impact of Lipid Content and Composition on Lipid Oxidation and Protein Carbonylation in Experimental Fermented Sausages. *Food Chemistry*, 147, 70-77.
- Fuentes, V., Ventanas, J., Morcuende, D., Estévez, M., & Ventanas, S. (2010). Lipid and Protein Oxidation and Sensory Properties of Vacuum-Packaged Dry-Cured Ham Subjected to High Hydrostatic Pressure. *Meat Science*, 85(3), 506-514.
- Future Market Insights, F. (2016). Ready-to-Eat Food Market by Product Type, Packaging, Distribution Channel & Region for 2016 – 2026 : Analysis and Review. Retrieved from https://www.futuremarketinsights.com/reports/ready-to-eat-food-market
- Gálvez, F., Domínguez, R., Pateiro, M., Carballo, J., Tomasevic, I., & Lorenzo, J. M. (2018). Effect of Gender on Breast and Thigh Turkey Meat Quality. *British Poultry Science*, *59*(4), 408-415.
- Ganhão, R., Morcuende, D., & Estévez, M. (2010). Tryptophan Depletion and Formation of A-Aminoadipic and F-Glutamic Semialdehydes in Porcine Burger Patties with Added Phenolic-Rich Fruit Extracts. *Journal of Agricultural and Food Chemistry*, *58*(6), 3541-3548.
- Gardner, H. W. (1979). Lipid Hydroperoxide Reactivity with Proteins and Amino Acids: A Review. *Journal of Agricultural and Food Chemistry*, 27(2), 220-229.
- Garidel, P. (2013). Protein Solubility from a Biochemical, Physicochemical and Colloidal Perspective. *The Review of American Pharmaceutical Business & Technology*. Retrieved from mericanpharmaceuticalreview.com/Featured-Articles/152568-Protein-Solubility-from-a-Biochemical-Physicochemical-and-Colloidal-Perspective/
- Gatellier, P., Kondjoyan, A., Portanguen, S., & Santé-Lhoutellier, V. (2010). Effect of Cooking on Protein Oxidation in N-3 Polyunsaturated Fatty Acids Enriched Beef. Implication on Nutritional Quality. *Meat Science*, *85*(4), 645-650.
- Geldenhuys, G., Hoffman, L. C., & Muller, N. (2015). The Fatty Acid, Amino Acid, and Mineral Composition of Egyptian Goose Meat as Affected by Season, Gender, and Portion. *Poultry Science*, *94*(5), 1075-1087.
- Ghimire, A., Paudel, N., & Poudel, R. (2022). Effect of Pomegranate Peel Extract on the Storage Stability of Ground Buffalo (*Bubalus Bubalis*) Meat. *LWT-Food Science and Technology, 154*, 112690.

- Ghnimi, S., Budilarto, E., & Kamal Eldin, A. (2017). The New Paradigm for Lipid Oxidation and Insights to Microencapsulation of Omega - 3 Fatty Acids. *Comprehensive Reviews in Food Science and Food Safety, 16*(6), 1206-1218.
- Giera, M., Lingeman, H., & Niessen, W. M. (2012). Recent Advancements in the Lc-and Gc-Based Analysis of Malondialdehyde (Mda): A Brief Overview. *Chromatographia*, 75(9-10), 433-440.
- Goethals, S., Van Hecke, T., Vossen, E., Vanhaecke, L., Van Camp, J., & De Smet, S. (2020). Commercial Luncheon Meat Products and Their in Vitro Gastrointestinal Digests Contain More Protein Carbonyl Compounds but Less Lipid Oxidation Products Compared to Fresh Pork. *Food Research International, 136*, 109585.
- Gruffat, D., Bauchart, D., Thomas, A., Parafita, E., & Durand, D. (2021). Fatty Acid Composition and Oxidation in Beef Muscles as Affected by Ageing Times and Cooking Methods. *Food Chemistry*, *343*, 128476.
- Guo, X., Wang, Y., Lu, S., Wang, J., Fu, H., Gu, B., Lyu, B., & Wang, Q. (2021). Changes in Proteolysis, Protein Oxidation, Flavor, Color and Texture of Dry-Cured Mutton Ham During Storage. *LWT-Food Science and Technology*, 149, 111860.
- Gupta, R. K., & Dudeja, P. (2017). Chapter 45 Ready-to-Eat Meals. In R. K. Gupta, Dudeja, & M. Singh (Eds.), *Food Safety in the 21st Century* (pp. 541-545). San Diego: Academic Press.
- Gutiérrez-del-Río, I., López-Ibáñez, S., Magadán-Corpas, P., Fernández-Calleja, L., Pérez-Valero, Á., Tuñón-Granda, M., Miguélez, E. M., Villar, C. J., & Lombó, F. (2021). Terpenoids and Polyphenols as Natural Antioxidant Agents in Food Preservation. *Antioxidants, 10*(8), 1264.
- Hasan, M. M., Sood, V., Erkinbaev, C., Paliwal, J., Suman, S., & Rodas-Gonzalez, A. (2021). Principal Component Analysis of Lipid and Protein Oxidation Products and Their Impact on Color Stability in Bison Longissimus Lumborum and Psoas Major Muscles. *Meat Science, 178*, 108523.
- Hawkins, C., & Davies, M. (2001). Generation and Propagation of Radical Reactions on Proteins. *Biochimica et Biophysica Acta (BBA) -Bioenergetics, 1504*, 196-219.
- Hellwig, M. (2020). Analysis of Protein Oxidation in Food and Feed Products. Journal of Agricultural and Food Chemistry, 68(46), 12870-12885.
- Hematyar, N., Rustad, T., Sampels, S., & Kastrup Dalsgaard, T. (2019). Relationship between Lipid and Protein Oxidation in Fish. *Aquaculture Research*, *50*(5), 1393-1403.

- Hendley, D. D., Mildvan, A. S., Reporter, M. C., & Strehler, B. L. (1963a). The Properties of Isolated Human Cardiac Age Pigment. I. Preparation and Physical Properties. *Journal of Gerontology*, *18*(2), 144-150.
- Hendley, D. D., Mildvan, A. S., Reporter, M. C., & Strehler, B. L. (1963b). The Properties of Isolated Human Cardiac Age Pigment. Ii. Chemical and Enzymatic Properties. *Journal of Gerontology*, *18*(3), 250-259.
- Hes, M. (2017). Protein-Lipid Interactions in Different Meat Systems in the Presence of Natural Antioxidants: A Review. *Polish Journal of Food and Nutrition Sciences, 67*(1), 5-18.
- Hes, M., Jezewska, M., Szymandera-Buszka, K., & Gramza-Michalowska, A. (2011). Effect of Antioxidant Additives on Nutritive Value of Dried Meat. *Zywnosc-Nauka Technologia Jakosc, 18*(5), 94-106.
- Hes, M., Korczak, J., & Gramza, A. (2007). Changes of Lipid Oxidation Degress and Their Influence on Protein Nutritive Value of Frozen Meat Products. *Polish Journal of Food Nutrition Sciences*, *57*(3), 323-328.
- Howard, S., Adams, J., & White, M. (2012). Nutritional Content of Supermarket Ready Meals and Recipes by Television Chefs in the United Kingdom: Cross Sectional Study. *Bmj* 345, e7607.
- Hu, M. (2016). Chapter 9 Oxidative Stability and Shelf Life of Low-Moisture Foods. In M. Hu & C. Jacobsen (Eds.), Oxidative Stability and Shelf Life of Foods Containing Oils and Fats (pp. 313-371): AOCS Press.
- Huang, S., Dong, X., Zhang, Y., Huang, M., & Zheng, Y. (2022). Effects of Oxidation and Precursors (Lysine, Glyoxal and Schiff Base) on the Formation of Νε-Carboxymethyl-Lysine in Aged, Stored and Thermally Treated Chicken Meat. *Food Science and Human Wellness*, *11*(5), 1252-1258.
- Huang, T. C., & Nip, W. K. (2001). Intermediate-Moisture Meat and Dehydrated Meat. In Y. H. Hui, N. Wai-Kit, R. W. Rogers, & O. A. Young (Eds.), *Meat Science and Applications* (pp. 404-432): CRC Press.
- Huang, X., & Ahn, D. U. (2019). Lipid Oxidation and Its Implications to Meat Quality and Human Health. *Food Science and Biotechnology, 28*(5), 1275-1285.
- Huda, N., Fatma, Y., Fazillah, A., & Adzitey, F. (2012). Chemical Composition, Colour and Sensory Characteristics of Commercial Serunding (Shredded Meat) in Malaysia. *Pakistan Journal of Nutrition*, 11(1), 1-4.
- Huda, N., Shen, Y. H., & Huey, Y. L. (2009). Proximate Composition, Colour, Texture Profile of Malaysian Chicken Balls. *Pakistan Journal of Nutrition*, 8(10), 1555-1558.

- Huff-Lonergan, E., & Lonergan, S. M. (2005). Mechanisms of Water-Holding Capacity of Meat: The Role of Postmortem Biochemical and Structural Changes. *Meat Science*, *71*(1), 194-204.
- Ibadullah, W. Z. W. (2013). Lipid Protein Interactions in Peanut Butter. (Doctoral Degree), Rutgers University-Graduate School-New Brunswick, Retrieved from <u>https://rucore.libraries.rutgers.edu/rutgers-lib/41943/</u>
- Inguglia, E. S., Granato, D., Kerry, J. P., Tiwari, B. K., & Burgess, C. M. (2021). Ultrasound for Meat Processing: Effects of Salt Reduction and Storage on Meat Quality Parameters. *Applied Sciences*, 11(1), 117.
- James, S., & James, C. (2014). Minimal Processing of Ready Meals. *Emerging Technologies for Food Processing*, 599-612.
- Jin, S.-K., Kim, G.-D., & Jeong, J.-Y. (2021). Evaluation of the Effect of Inhibiting Lipid Oxidation of Natural Plant Sources in a Meat Model System. *Journal of Food Quality, 2021*, 6636335.
- Jové, M., Mota-Martorell, N., Pradas, I., Martín-Gari, M., Ayala, V., & Pamplona, R. (2020). The Advanced Lipoxidation End-Product Malondialdehyde-Lysine in Aging and Longevity. *Antioxidants (Basel)*, 9(11).
- Kanner, J. (2007). Dietary Advanced Lipid Oxidation Endproducts Are Risk Factors to Human Health. *Molecular Nutrition & Food Research, 51*(9), 1094-1101.
- Karunasiri, A. N., Gunawardane, M., Senanayake, C. M., Jayathilaka, N., & Seneviratne, K. N. (2020). Antioxidant and Nutritional Properties of Domestic and Commercial Coconut Milk Preparations. *International Journal of Food Science*, *2020*, 3489605.
- Karwowska, M., Wójciak, K. M., & Dolatowski, Z. J. (2014). Comparative Studies on Lipid Oxidation of Organic Model Sausage without Nitrite Produced with the Addition of Native or Autoclaved Mustard Seed and Acid Whey. International Journal of Food Science & Technology, 49(12), 2563-2570.
- Kehm, R., Baldensperger, T., Raupbach, J., & Höhn, A. (2021). Protein Oxidation - Formation Mechanisms, Detection and Relevance as Biomarkers in Human Diseases. *Redox Biology, 42*, 101901.
- Kim, H., Do, H. W., & Chung, H. (2017). A Comparison of the Essential Amino Acid Content and the Retention Rate by Chicken Part According to Different Cooking Methods. *Korean Journal for Food Science of Animal Resources*, 37(5), 626.
- Knechtges, P. L. (2012). *Food Safety : Theory and Practice*. Burlington, MA.: Jones & Bartlett Learning.

- Koleva, I. I., Niederländer, H. A., & van Beek, T. A. (2000). An on-Line Hplc Method for Detection of Radical Scavenging Compounds in Complex Mixtures. *Analytical Chemistry*, 72(10), 2323-2328.
- Koppenol, W. H. (1990). Oxyradical Reactions: From Bond-Dissociation Energies to Reduction Potentials. *FEBS Letters*, *264*(2), 165-167.
- Kumar, P., Verma, A. K., Kumar, D., Umaraw, P., Mehta, N., & Malav, O. (2019). Meat Snacks: A Novel Technological Perspective. In *Innovations in Traditional Foods* (pp. 293-321): Elsevier.
- Kumar, S., Saxena, S., Verma, J., & Gautam, S. (2016). Development of Ambient Storable Meal for Calamity Victims and Other Targets Employing Radiation Processing and Evaluation of Its Nutritional, Organoleptic, and Safety Parameters. LWT-Food Science and Technology, 69, 409-416.
- Kumar, Y., Yadav, D. N., Ahmad, T., & Narsaiah, K. (2015). Recent Trends in the Use of Natural Antioxidants for Meat and Meat Products. *Comprehensive Reviews in Food Science and Food Safety, 14*(6), 796-812.
- Kurutas, E. B. (2016). The Importance of Antioxidants Which Play the Role in Cellular Response against Oxidative/Nitrosative Stress: Current State. *Nutrition Journal, 15*(1), 71-71.
- Levine, R. L., Garland, D., Oliver, C. N., Amici, A., Climent, I., Lenz, A. G., Ahn, B. W., Shaltiel, S., & Stadtman, E. R. (1990). Determination of Carbonyl Content in Oxidatively Modified Proteins. *Methods in Enzymology, 186*, 464-478.
- Levine, R. L., Williams, J. A., Stadtman, E. P., & Shacter, E. (1994). Carbonyl Assays for Determination of Oxidatively Modified Proteins. *Methods in Enzymology*, 233, 346-357.
- Leygonie, C., Britz, T. J., & Hoffman, L. C. (2012). Impact of Freezing and Thawing on the Quality of Meat: Review. *Meat Science*, *91*(2), 93-98.
- Li, K., McKeith, A. G., Shen, C., & McKeith, R. (2017). A Comparison Study of Quality Attributes of Ground Beef and Veal Patties and Thermal Inactivation of Escherichia Coli O157:H7 after Double Pan-Broiling under Dynamic Conditions. *Foods, 7*(1).
- Li, Y. (2013). *Mechanical Denaturation: Forced Unfolding of Proteins.* University of British Columbia,
- Lin, Y., Wang, H., Rao, W., Cui, Y., Dai, Z., & Shen, Q. (2019). Structural Characteristics of Dietary Fiber (*Vigna Radiata* L. Hull) and Its Inhibitory Effect on Phospholipid Digestion as an Additive in Fish Floss. *Food Control, 98*, 74-81.

- Listrat, A., Lebret, B., Louveau, I., Astruc, T., Bonnet, M., Lefaucheur, L., Picard, B., & Bugeon, J. (2016). How Muscle Structure and Composition Influence Meat and Flesh Quality. *The Scientific World Journal, 2016*, 3182746.
- Lonergan, S. M., Topel, D. G., & Marple, D. N. (2019a). Chapter 13 Fresh and Cured Meat Processing and Preservation. In S. M. Lonergan, D. G. Topel, & D. N. Marple (Eds.), *The Science of Animal Growth and Meat Technology* (Vol. Second edition, pp. 205-228): Academic Press.
- Lonergan, S. M., Topel, D. G., & Marple, D. N. (2019b). Chapter 15 Packaging for Meat and Meat Products. In S. M. Lonergan, D. G. Topel, & D. N. Marple (Eds.), *The Science of Animal Growth and Meat Technology* (Second Edition) (pp. 255-269): Academic Press.
- Lorenzo, J. M., Domínguez, R., & Carballo, J. (2017). Chapter 9 Control of Lipid Oxidation in Muscle Food by Active Packaging Technology. In R. Banerjee, A. Verma, & M. Siddiqui (Eds.), *Natural Antioxidants. Applications in Foods of Animal Origin (First Edition)* (pp. 343-382): Apple Academic Press Inc. / CRC Press-Taylor & Francis eBook.
- Lorenzo, J. M., Vargas, F. C., Strozzi, I., Pateiro, M., Furtado, M. M., Sant'Ana, A. S., Rocchetti, G., Barba, F. J., Dominguez, R., Lucini, L., & do Amaral Sobral, P. J. (2018). Influence of Pitanga Leaf Extracts on Lipid and Protein Oxidation of Pork Burger During Shelf-Life. *Food Research International, 114*, 47-54.
- Lu, W., & Tassou, S. A. (2013). Characterization and Experimental Investigation of Phase Change Materials for Chilled Food Refrigerated Cabinet Applications. *Applied Energy*, *112*, 1376-1382.
- Lund, M. N. (2007). *Protein Oxidation in Meat During Chill Storage*. (Doctoral Degree), University of Copenhagen, Copenhagen: Center for Skov, Landskab og Planlægning/Københavns Universitet.
- Lund, M. N., Heinonen, M., Baron, C. P., & Estevez, M. (2011). Protein Oxidation in Muscle Foods: A Review. *Molecular Nutrition & Food Research*, 55(1), 83-95.
- Lund, M. N., & Ray, C. A. (2017). Control of Maillard Reactions in Foods: Strategies and Chemical Mechanisms. *Journal of Agricultural and Food Chemistry*, 65(23), 4537-4552.
- Madane, P., Das, A. K., Pateiro, M., Nanda, P. K., Bandyopadhyay, S., Jagtap,
 P., Barba, F. J., Shewalkar, A., Maity, B., & Lorenzo, J. M. (2019).
 Drumstick (*Moringa Oleifera*) Flower as an Antioxidant Dietary Fibre in Chicken Meat Nuggets. *Foods*, 8(8), 307.
- Małecki, J., Muszyński, S., & Sołowiej, B. G. (2021). Proteins in Food Systems -Bionanomaterials, Conventional and Unconventional Sources,

Functional Properties, and Development Opportunities. *Polymers, 13*(15), 2506.

- Martinaud, A., Mercier, Y., Marinova, P., Tassy, C., Gatellier, P., & Renerre, M. (1997). Comparison of Oxidative Processes on Myofibrillar Proteins from Beef During Maturation and by Different Model Oxidation Systems. *Journal of Agricultural and Food Chemistry*, 45(7), 2481-2487.
- Mathew, S., Abraham, T. E., & Zakaria, Z. A. (2015). Reactivity of Phenolic Compounds Towards Free Radicals under in Vitro Conditions. *Journal* of Food Science and Technology, 52(9), 5790-5798.
- McMillin, K. W., & Belcher, J. N. (2012). 6 Advances in the Packaging of Fresh and Processed Meat Products. In J. P. Kerry (Ed.), *Advances in Meat, Poultry and Seafood Packaging* (pp. 173-204): Woodhead Publishing.

McMurray, J. (2000). Organic Chemistry. Pacific Grove, CA: Brooks/Cole.

- Mercier, Y., Gatellier, P., & Renerre, M. (2004). Lipid and Protein Oxidation in Vitro, and Antioxidant Potential in Meat from Charolais Cows Finished on Pasture or Mixed Diet. *Meat Science*, 66(2), 467-473.
- Miller, N. J., Rice-Evans, C., & Davies, M. J. (1993). A New Method for Measuring Antioxidant Activity. In: Portland Press Ltd.
- Min, B., & Ahn, D. U. (2005). Mechanism of Lipid Peroxidation in Meat and Meat Products: A Review. *Food Science and Biotechnology*, *14*(1), 152-163.
- Min, H., & Jacobsen, C. (2016). Oxidative Stability and Shelf Life of Low-Moisture Foods. London: Elsevier Inc.
- Mohammed, H. H. H., Jin, G., Ma, M., Khalifa, I., Shukat, R., Elkhedir, A. E., Zeng, Q., & Noman, A. E. (2020). Comparative Characterization of Proximate Nutritional Compositions, Microbial Quality and Safety of Camel Meat in Relation to Mutton, Beef, and Chicken. *LWT*, *118*, 108714.
- Mohan, A., Roy, A., Duggirala, K., & Klein, L. (2022). Oxidative Reactions of 4-Oxo-2-Nonenal in Meat and Meat Products. *LWT*, *165*, 113747.
- Moigradean, D., Poiana, M.-A., Gogoasa, I., & Technologies. (2012). Quality Characteristics and Xidative Stability of Coconut Oil During Storage. *Journal of Agroalimentary Processes, 18*(4), 272-276.
- Montero, M. L., Garrido, D., Gallardo, R. K., Tang, J., & Ross, C. F. (2021). Consumer Acceptance of a Ready-to-Eat Meal During Storage as Evaluated with a Home-Use Test. *Foods*, *10*(7), 1623.

- Moon, C. T., Yancey, J. W., S., Apple, J. K., Hollenbeck, J. J., Johnson, T. M., & Winters, A. R. (2016). Quality Characteristics of Fresh and Cooked Ground Beef Can Be Improved by the Incorporation of Lean Finely -Textured Beef (Lftb). *Journal of Food Quality*, *39*(5), 465-475.
- Moreno, M. C. M. M., Olivares, D. M., Lopez, F. J. A., Adelantado, J. V. G., & Reig, F. B. (1999). Analytical Evaluation of Polyunsaturated Fatty Acids Degradation During Thermal Oxidation of Edible Oils by Fourier Transform Infrared Spectroscopy. *Talanta*, *50*(2), 269-275.
- Movileanu, I., Núñez de González, M. T., Hafley, B., Miller, R. K., & Keeton, J. T. (2013). Comparison of Dried Plum Puree, Rosemary Extract, and Bha/Bht as Antioxidants in Irradiated Ground Beef Patties. *International Journal of Food Science*, 2013, 360732.
- Muhamad, I. I., & Karim, N. A. (2015). Trends, Convenience, and Safety Issues of Ready Meals. In *Minimally Processed Foods* (pp. 105-123): Springer.
- Mukhtar, S., Zahoor, T., Randhawa, M., Iqbal, R., Shabbir, A., Liaqat, A., & Ahsan, S. (2018). Synergistic Effect of Chitosan and Clove Oil on Raw Poultry Meat. *Journal of Food Processing and Technology, 09*.
- Muzolf-Panek, M., & Kaczmarek, A. (2021). Chemometric Analysis of Fatty Acid Composition of Raw Chicken, Beef, and Pork Meat with Plant Extract Addition During Refrigerated Storage. *Molecules*, *26*(16), 4952.
- Nadkarni, D. V., & Sayre, L. M. (1995). Structural Definition of Early Lysine and Histidine Adduction Chemistry of 4-Hydroxynonenal. *Chemical research in toxicology*, *8*(2), 284-291.
- Nauman, K., Jaspal, M. H., Asghar, B., Manzoor, A., Akhtar, K. H., Ali, U., Ali, S., Nasir, J., Sohaib, M., & Badar, I. H. (2022). Effect of Different Packaging Atmosphere on Microbiological Shelf Life, Physicochemical Attributes, and Sensory Characteristics of Chilled Poultry Fillets. *Food Science of Animal Resources, 42*(1), 153-174.
- Nawaz, A., Irshad, S., Ali Khan, I., Khalifa, I., Walayat, N., Muhammad Aadil, R., Kumar, M., Wang, M., Chen, F., Cheng, K.-W., & Lorenzo, J. M. (2022).
 Protein Oxidation in Muscle-Based Products: Effects on Physicochemical Properties, Quality Concerns, and Challenges to Food Industry. *Food Research International, 157*, 111322.
- Nissen, L. R., Byrne, D. V., Bertelsen, G., & Skibsted, L. H. (2004). The Antioxidative Activity of Plant Extracts in Cooked Pork Patties as Evaluated by Descriptive Sensory Profiling and Chemical Analysis. *Meat Science, 68*(3), 485-495.
- Nissen, L. R., Månsson, L., Bertelsen, G., Huynh-Ba, T., & Skibsted, L. H. (2000). Protection of Dehydrated Chicken Meat by Natural Antioxidants as

Evaluated by Electron Spin Resonance Spectrometry. *Journal of Agricultural and Food Chemistry, 48*(11), 5548-5556.

- Nollet, L. M. L., & Toldra, F. (2008). Handbook of Processed Meats and Poultry Analysis: CRC Press.
- O'Farrell, M. (2011). 22 Online Quality Assessment of Processed Meats. In J. P. Kerry & J. F. Kerry (Eds.), *Processed Meats* (pp. 546-566): Woodhead Publishing.
- Ockerman, H. W., & Basu, L. (2014). By-Products | Edible, for Human Consumption. In M. Dikeman & C. Devine (Eds.), *Encyclopedia of Meat Sciences (Second Edition)* (pp. 104-111). Oxford: Academic Press.
- Ogunsola, O. O., & Omojola, A. B. (2008). Nutritional Evaluation of a Dehydrated Shredded Meat Product, (*Danbunama*). *Pakistan Journal of Nutrition*, 7(4), 554-556.
- Okkels, S. L., Dybdal, D. R., Beck, A. M., Bügel, S., Klausen, T. W., & Olsen, A. (2019). An Investigation of Main Meal Preferences in Nursing Home Residents. *Journal of Sensory Studies*, *34*(4), e12504.
- Oliver, C. N., Ahn, B. W., Moerman, E. J., Goldstein, S., & Stadtman, E. R. (1987). Age-Related Changes in Oxidized Proteins. *Journal of Biological Chemistry*, 262(12), 5488-5491.
- Orthoefer, F. T., & List, G. R. (2007). 12 Dynamics of Frying. In M. D. Erickson (Ed.), *Deep Frying (Second Edition)* (pp. 253-275): AOCS Press.
- Osório, V. M., & Cardeal, Z. L. (2013). Analytical Methods to Assess Carbonyl Compounds in Foods and Beverages. *Journal of the Brazilian Chemical Society, 24*(11), 1711-1718.
- Paiva, T., Jacinto, T. A., Sarraguça, M. C., & Coutinho, P. (2022). Beef Consumers Behaviour and Preferences—the Case of Portugal. Sustainability, 14(4), 2358.
- Palka, K., & Wesierska, E. (2014). Cooking of Meat | Physics and Chemistry. In
 M. Dikeman & C. Devine (Eds.), *Encyclopedia of Meat Sciences* (Second Edition) (pp. 404-409). Oxford: Academic Press.
- Papastergiadis, A., Mubiru, E., Van Langenhove, H., & De Meulenaer, B. (2012). Malondialdehyde Measurement in Oxidized Foods: Evaluation of the Spectrophotometric Thiobarbituric Acid Reactive Substances (Tbars) Test in Various Foods. *Journal of Agricultural and Food Chemistry*, *60*(38), 9589-9594.

- Park, D., & Xiong, Y. L. (2007). Oxidative Modification of Amino Acids in Porcine Myofibrillar Protein Isolates Exposed to Three Oxidizing Systems. *Food Chemistry*, 103(2), 607-616.
- Park, J.-H., Lee, Y.-J., Lim, J.-G., Jeon, J.-H., & Yoon, K.-S. (2021). Effect of Quinoa (Chenopodium Quinoa Willd.) Starch and Seeds on the Physicochemical and Textural and Sensory Properties of Chicken Meatballs During Frozen Storage. *Foods*, *10*(7), 1601.
- Parr, L. J., & Swoboda, P. A. T. (1976). The Assay of Conjugable Oxidation Products Applied to Lipid Deterioration in Stored Foods. *International Journal of Food Science Technology*, 11(1), 1-12.
- Pateiro, M., Domínguez, R., & Lorenzo, J. M. (2021). Recent Research Advances in Meat Products. *Foods, 10*(6), 1303.
- Peamprasart, T., & Chiewchan, N. (2006). Effect of Fat Content and Preheat Treatment on the Apparent Viscosity of Coconut Milk after Homogenization. *Journal of Food Engineering*, 77(3), 653-658.
- Pereira, A. L. F., & Abreu, V. K. G. (2018). Lipid Peroxidation in Meat and Meat Products. In *Lipid Peroxidation*: IntechOpen.
- Pereira, A. L. F., & Abreu, V. K. G. (2020). Lipid Peroxidation in Meat and Meat Products. In M. A. Mansour (Ed.), *Lipid Peroxidation Research* (pp. 29-42). London, United Kingdon: Intechopen.
- Pereira, P. M. d. C. C., & Vicente, A. F. d. R. B. (2013). Meat Nutritional Composition and Nutritive Role in the Human Diet. *Meat Science*, *93*(3), 586-592.
- Perham, C. C., Gifford, C. L., Woerner, D. R., Engle, T. E., Sellins, K. S., Acheson, R. J., Douglass, L. W., Tatum, J. D., Delmore, R. J., Cifelli, A., McNeill, S. H., & Belk, K. E. (2019). Special-Fed Veal: Separable Components, Proximate Composition, and Nutrient Analysis of Selected Raw and Cooked, Wholesale and Retail Cuts. *Meat Science, 148*, 19-31.
- Pisoschi, A. M., & Pop, A. (2015). The Role of Antioxidants in the Chemistry of Oxidative Stress: A Review. *European Journal of Medicinal Chemistry*, 97, 55-74.
- Pokorný, J. (2001). *Chapter 13: Preparation of Natural Antioxidants*. Cambridge, England: Woodhead Publishing Ltd.
- Pokorny, J., & Davidek, I. (1979). Influence of Interactions of Proteins with Oxidized Lipids on Nutrition and Sensory Value of Food. *Acta Alimentaria Polonica*, *5*(2).

- Pokorný, J., Davidek, J., Chocholata, V., Panek, J., Bulantova, H., Janitz, W., Valentova, H., & Vierecklová, M. (1990). Interactions of Oxidized Lipids with Protein Part Xvi. Interactions of Oxidized Ethyl Linoleate with Collagen. *Die Nahrung*, 34(2), 159-169.
- Pokorny, J., Kołakowska, A., & Bienkiewicz, G. (2010). Chapter 22: Lipid–Protein and Lipid–Saccharide Interactions. In Z. Z. E. Sikorski & A. Kolakowska (Eds.), *Chemical, Biological, and Functional Aspects of Food Lipids* (Second Edition) (pp. 455). Boca Raton, FL: CRC Press.
- Pool-Zobel, B. L., Bub, A., Schröder, N., & Rechkemmer, G. (1999). Anthocyanins Are Potent Antioxidants in Model Systems but Do Not Reduce Endogenous Oxidative DNA Damage in Human Colon Cells. *European Journal of Nutrition*, 38(5), 227-234.
- Popova, T., Marinova, P., Vasileva, V., Gorinov, Y., & Lidji, K. (2009). Oxidative Changes in Lipids and Proteins in Beef During Storage. *Archiva Zootechnica*, *12*(3), 30-38.
- Powell, P. K., Lawler, S., Durham, J., & Cullerton, K. (2021). The Food Choices of Us University Students During Covid-19. *Appetite*, *161*, 105130.
- Pulido, R., Bravo, L., & Saura-Calixto, F. (2000). Antioxidant Activity of Dietary Polyphenols as Determined by a Modified Ferric Reducing/Antioxidant Power Assay. *Journal of Agricultural and Food Chemistry, 48*(8), 3396-3402.
- Puolanne, E., & Halonen, M. (2010). Theoretical Aspects of Water-Holding in Meat. *Meat Science, 86*(1), 151-165.
- Rao, D. N. (1997). Intermediate Moisture Foods Based on Meats—a Review. *Food Reviews International, 13*(4), 519-551.
- RC Machinery. (2022). Mesin Mencarik Daging. Retrieved from http://rcmesin.com/product/mesin-mencarik-daging/
- Refsgaard, H. H., Tsai, L., & Stadtman, E. R. (2000). Modifications of Proteins by Polyunsaturated Fatty Acid Peroxidation Products. *Proceedings of the National Academy of Sciences of the United States of America*, 97(2), 611-616.
- Requena, J. R., Fu, M.-X., Ahmed, M. U., Jenkins, A. J., Lyons, T. J., & Thorpe, S. R. (1996). Lipoxidation Products as Biomarkers of Oxidative Damage to Proteins During Lipid Peroxidation Reactions. *Nephrology Dialysis Transplantation*, 11(supp5), 48-53.
- Reznick, A. Z., & Packer, L. (1994). Oxidative Damage to Proteins: Spectrophotometric Method for Carbonyl Assay. *Methods in Enzymology*, 233, 357-363.

- Rhee, K., Anderson, L., & Sams, A. (1996). Lipid Oxidation Potential of Beef, Chicken, and Pork. *Journal of Food Science*, *61*(1), 8-12.
- Richards, A. T. (2019). Food and Nutritional Analysis | Meat and Meat Products. In P. Worsfold, C. Poole, A. Townshend, & M. Miró (Eds.), *Encyclopedia* of Analytical Science (Third Edition) (pp. 436-450). Oxford: Academic Press.
- Richards, M. P. (2006). Lipid Chemistry and Biochemistry. . In Y. Hui (Ed.), Handbook of Food Science, Technology, and Engineering (Vol. 8, pp. 1-21). Boca Raton, FL: CRC Press, Taylor & Francis.
- Saeed, S., Fawthrop, S. A., & Howell, N. K. (1999). Electron Spin Resonance (Esr) Study on Free Radical Transfer in Fish Lipid–Protein Interaction. *Journal of the Science of Food and Agriculture, 79*(13), 1809-1816.
- Sakanaka, S., Tachibana, Y., Ishihara, N., & Juneja, L. R. (2005). Antioxidant Properties of Casein Calcium Peptides and Their Effects on Lipid Oxidation in Beef Homogenates. *Journal of Agricultural and Food Chemistry*, *53*(2), 464-468.
- Sakomura, N. K., Ekmay, R. D., Mei, S. J., & Coon, C. N. (2015). Lysine, Methionine, Phenylalanine, Arginine, Valine, Isoleucine, Leucine, and Threonine Maintenance Requirements of Broiler Breeders. *Poultry Science*, *94*(11), 2715-2721.
- Sakowski, T., Grodkowski, G., Gołebiewski, M., Slósarz, J., Kostusiak, P., Solarczyk, P., & Puppel, K. (2022). Genetic and Environmental Determinants of Beef Quality-a Review. *Frontiers in veterinary science*, *9*, 819605-819605.
- Salcedo-Sandoval, L., Cofrades, S., Ruiz-Capillas, C., & Jiménez-Colmenero, F. (2014). Effect of Cooking Method on the Fatty Acid Content of Reduced-Fat and Pufa-Enriched Pork Patties Formulated with a Konjac-Based Oil Bulking System. *Meat Science*, 98(4), 795-803.
- Sampaio, G. R., Saldanha, T., Soares, R. A. M., & Torres, E. A. F. S. (2012). Effect of Natural Antioxidant Combinations on Lipid Oxidation in Cooked Chicken Meat During Refrigerated Storage. *Food Chemistry*, *135*(3), 1383-1390.
- Santana, N. D. C. d., Cordeiro, Â. M. T. M., Meireles, B. R. L. A., Araújo, Í. B. S., Estévez, M., Ferreira, V. C. S., & Silva, F. A. P. (2021). Inhibition of Protein and Lipid Oxidation in Ready-to-Eat Chicken Patties by a *Spondias Mombin* L. Bagasse Phenolic-Rich Extract. *Foods, 10*(6), 1338.
- Santeramo, F. G., Carlucci, D., De Devitiis, B., Seccia, A., Stasi, A., Viscecchia, R., & Nardone, G. (2018). Emerging Trends in European Food, Diets and Food Industry. *Food Research International*, 104, 39-47.

- Scerra, M., Foti, F., Caparra, P., Cilione, C., Rao, R., Priolo, A., Natalello, A., Luciano, G., & Chies, L. (2022). Effect of Feeding Pigs with Bergamot by-Product on Fatty Acid Composition and Oxidative Stability of Meat and Salami. *Meat Science*, 183, 108662.
- Schaich. (2008). Co-Oxidations of Oxidizing Lipids: Reactions with Proteins. In A. Kamal-Eldin & D. Min (Eds.), *Lipid Oxidation Pathways* (Vol. 2, pp. 183-274): AOCS Press.
- Schaich, K. M. (2008). Co-Oxidation of Proteins by Oxidizing Lipids. In A. Kamal-Eldin & D. Min (Eds.), (Vol. 2, pp. 183-274): AOCS Press.
- Schaich, K. M. (2013). Challenges in Elucidating Lipid Oxidation Mechanisms: When, Where, and How Do Products Arise? In K. M. Schaich (Ed.), *Lipid Oxidation* (pp. 1-52): Elsevier.
- Schaich, K. M. (2016a). Analysis of Lipid and Protein Oxidation in Fats, Oils, and Foods. In Oxidative Stability and Shelf Life of Foods Containing Oils and Fats (pp. 1-131): Elsevier.
- Schaich, K. M. (2016b). Chapter 1 Analysis of Lipid and Protein Oxidation in Fats, Oils, and Foods. In M. Hu & C. Jacobsen (Eds.), Oxidative Stability and Shelf Life of Foods Containing Oils and Fats (pp. 1-131): AOCS Press.
- Schlesier, K., Harwat, M., Böhm, V., & Bitsch, R. (2002). Assessment of Antioxidant Activity by Using Different in Vitro Methods. *Free Radical Research*, *36*(2), 177-187.
- Seow, C., & Gwee, C. (2003). Coconut Milk: Chemistry and Technology. International Journal of Food Science & Technology, 32, 189-201.
- Serpen, A., Gökmen, V., & Fogliano, V. (2012). Total Antioxidant Capacities of Raw and Cooked Meats. *Meat Science*, *90*(1), 60-65.
- Shahidi, F. (2015). *Handbook of Antioxidants for Food Preservation*: Woodhead Publishing.
- Shahidi, F., & Ambigaipalan, P. (2015). Phenolics and Polyphenolics in Foods, Beverages and Spices: Antioxidant Activity and Health Effects – a Review. *Journal of Functional Foods, 18*, 820-897.
- Shahidi, F., Janitha, P. K., & Wanasundara, P. D. (1992). Phenolic Antioxidants. *Critical Reviews in Food Science and Nutrition, 32*(1), 67-103.
- Shang-gui, D., Zhi-ying, P., Fang, C., Ping, Y., & Tie, W. (2004). Amino Acid Composition and Anti-Anaemia Action of Hydrolyzed Offal Protein from Harengula Zunasi Bleeker. *Food Chemistry*, *87*(1), 97-102.

- Shehab, T. (2016). Effect of Cooking Methods on Amino Acids Composition of Chicken Meat. *Theory Practice of Meat Processing*, *1*(4), 11-18.
- Silberbauer, A., & Schmid, M. (2017). Packaging Concepts for Ready-to-Eat Food: Recent Progress. *Journal of Packaging Technology and Research, 1*(3), 113-126.
- Simuang, J., Chiewchan, N., & Tansakul, A. (2004). Effects of Fat Content and Temperature on the Apparent Viscosity of Coconut Milk. *Journal of Food Engineering*, 64(2), 193-197.
- Skellon, J. H., & Thruston, M. N. (1953). The Oxidation of Monoethenoid Fatty Acids and Esters. Catalytic Oxidation of Elaidic Acid, Methyl Elaidate, and N-Propyl Elaidate. The Oxidation Products. *Journal of the Chemical Society*(0), 138-142.
- Sklan, D., Tenne, Z., & Budowski, P. (1983). The Effect of Dietary Fat and Tocopherol on Lipolysis and Oxidation in Turkey Meat Stored at Different Temperatures. *Poultry Science*, 62(10), 2017-2021.
- Soglia, F., Petracci, M., & Ertbjerg, P. (2016). Novel Dnph-Based Method for Determination of Protein Carbonylation in Muscle and Meat. *Food Chemistry*, 197, 670-675.
- Sohaib, M., Anjum, F. M., Arshad, M. S., Imran, M., Imran, A., & Hussain, S. (2017a). Oxidative Stability and Lipid Oxidation Flavoring Volatiles in Antioxidants Treated Chicken Meat Patties During Storage. *Lipids in Health and Disease*, *16*(1), 27.
- Sohaib, M., Anjum, F. M., Sahar, A., Arshad, M. S., Rahman, U. U., Imran, A., & Hussain, S. (2017b). Antioxidant Proteins and Peptides to Enhance the Oxidative Stability of Meat and Meat Products: A Comprehensive Review. International Journal of Food Properties, 20(11), 2581-2593.
- Soladoye, O., Juárez, M., Aalhus, J., Shand, P., & Estévez, M. (2015). Protein Oxidation in Processed Meat: Mechanisms and Potential Implications on Human Health. *Comprehensive Reviews in Food Science and Food Safety, 14*(2), 106-122.
- Soyer, A., Özalp, B., Dalmış, Ü., & Bilgin, V. (2010). Effects of Freezing Temperature and Duration of Frozen Storage on Lipid and Protein Oxidation in Chicken Meat. *Food Chemistry*, *120*(4), 1025-1030.
- Spencer, K. C. (2005). Modified Atmosphere Packaging of Ready-to-Eat Foods. In *Innovations in Food Packaging* (pp. 185-203): Elsevier.
- Spiegelaar, N., Martin, I. D., & Tsuji, L. J. S. (2019). Indigenous Subarctic Food Systems in Transition: Amino Acid Composition (Including Tryptophan)

in Wild-Harvested and Processed Meats. *International Journal of Food Science*, 2019, 7096416.

- Stadtman, E. R. (1992). Protein Oxidation and Aging. *Science*, 257(5074), 1220-1224.
- Stadtman, E. R., & Levine, R. L. (2000). Protein Oxidation. Annals of the New York Academy of Sciences, 899, 191-208.
- Stadtman, E. R., & Levine, R. L. (2003). Free Radical-Mediated Oxidation of Free Amino Acids and Amino Acid Residues in Proteins. *Amino Acids, 25*(3), 207-218.
- Strube, M., Haenen, G., Van Den Berg, H., & Bast, A. (1997). Pitfalls in a Method for Measurement of Total Antioxidant Capacity. *Free Radical Research*, *26*(512-521), 1237.
- Sukisman, A., Purnomo, H., & Rosyidi, D. (2014). Quality Properties, Antioxidant Capacity and Total Phenolic Content of Traditional Deep Fried Shredded Meat (*Abon*) of Palu, Central Sulawesi. *American Journal of Food Technology 9*, 80-88.
- Sukisman, S., Halid, A., & Rahim, A. (2018). Sifat Fisik, Kimia Dan Aktivitas Antioksidan Abon Daging Ayam Di Kota Palu. *Agroland: Jurnal Ilmu-ilmu Pertanian, 25*(2), 154-163.
- Suman, S. P., & Joseph, P. (2013). Myoglobin Chemistry and Meat Color. Annual Review of Food Science and Technology, 4, 79-99.
- Sun, Q., Senecal, A., Chinachoti, P., & Faustman, C. (2002). Effect of Water Activity on Lipid Oxidation and Protein Solubility in Freeze - Dried Beef During Storage. *Journal of Food Science*, 67(7), 2512-2516.
- Szabo, Z., Marosvölgyi, T., Szabo, E., Koczka, V., Verzar, Z., Figler, M., & Decsi, T. (2022). Effects of Repeated Heating on Fatty Acid Composition of Plant-Based Cooking Oils. *11*(2), 192.
- Tajeddin, B., & Arabkhedri, M. (2020). Chapter 16 Polymers and Food Packaging. In M. A. A. AlMaadeed, D. Ponnamma, & M. A. Carignano (Eds.), *Polymer Science and Innovative Applications* (pp. 525-543): Elsevier.
- Tang, X., Shen, Y., Zhang, Y., Schilling, M. W., & Li, Y. (2021). Parallel Comparison of Functional and Physicochemical Properties of Common Pulse Proteins. *LWT*, 146, 111594.
- Tansakul, A., & Chaisawang, P. (2006). Thermophysical Properties of Coconut Milk. *Journal of Food Engineering*, *73*(3), 276-280.

- Tapia, M. S., Alzamora, S. M., & Chirife, J. (2020). Effects of Water Activity (A_w) on Microbial Stability as a Hurdle in Food Preservation. In G. Barbosa-Cánovas, J. A. J. Fontana, S. J. Schmidt, & T. P. Labuza (Eds.), Water Activity in Foods: Fundamentals and Applications (Second Edition ed., pp. 323-355): John Wiley & Sons, Inc.
- Taşkıran, M., Olum, E., & Candoğan, K. (2020). Changes in Chicken Meat Proteins During Microwave and Electric Oven Cooking. *Journal of Food Processing and Preservation*, 44(2), e14324.
- Telukdarie, A., Munsamy, M., & Mohlala, P. (2020). Analysis of the Impact of Covid-19 on the Food and Beverages Manufacturing Sector. *Sustainability*, *12*(22), 9331.
- Thangaraj, P. (2016). Proximate Composition Analysis. In T. Parimelazhagan (Ed.), *Pharmacological Assays of Plant-Based Natural Products* (pp. 21-31). Cham: Springer International Publishing.
- Tornberg, E. (2005). Effects of Heat on Meat Proteins Implication on Structure and Quality of Meat Products. *Meat Science, 70*, 493-508.
- Traverso, N., Menini, S., Maineri, E. P., Patriarca, S., Odetti, P., Cottalasso, D., Marinari, U. M., & Pronzato, M. A. (2004). Malondialdehyde, a Lipoperoxidation-Derived Aldehyde, Can Bring About Secondary Oxidative Damage to Proteins. *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences*, 59(9), B890-B895.
- Uchida, K., Sakai, K., Itakura, K., Osawa, T., & Toyokuni, S. (1997). Protein Modification by Lipid Peroxidation Products: Formation of Malondialdehyde-Derived N(Epsilon)-(2-Propenol)Lysine in Proteins. *Archives of Biochemistry and Biophysics, 346*(1), 45-52.
- United States Department of Agriculture. (2014). National Nutrient Database for Standard Reference. In *Frankfurter: 07950. Sausage vienna: 07083. Pork cured ham cooked: 10804.*
- Utrera, M., & Estévez, M. (2012). Oxidation of Myofibrillar Proteins and Impaired Functionality: Underlying Mechanisms of the Carbonylation Pathway. *Journal of Agricultural and Food Chemistry, 60*(32), 8002-8011.
- Utrera, M., Morcuende, D., & Estévez, M. (2014a). Fat Content Has a Significant Impact on Protein Oxidation Occurred During Frozen Storage of Beef Patties. *LWT-Food Science and Technology*, *56*(1), 62-68.
- Utrera, M., Morcuende, D., & Estévez, M. (2014b). Temperature of Frozen Storage Affects the Nature and Consequences of Protein Oxidation in Beef Patties. *Meat Science*, *96*(3), 1250-1257.

- Utrera, M., Parra, V., & Estévez, M. (2014c). Protein Oxidation During Frozen Storage and Subsequent Processing of Different Beef Muscles. *Meat Science*, *96*(2), 812-820.
- Valous, N., Zheng, L., Sun, D.-W., & Tan, J. (2016). Quality Evaluation of Meat Cuts. In D.-W. Sun (Ed.), Computer Vision Technology for Food Quality Evaluation (pp. 175-193): Elsevier.
- Vandemoortele, A., & De Meulenaer, B. (2019). Reactivity of Lipid Oxidation Products in Foods – Is Malondialdehyde a Reliable Marker? In L. Melton, F. Shahidi, & P. Varelis (Eds.), *Encyclopedia of Food Chemistry* (pp. 468-477). Oxford: Academic Press.
- Varelis, P., Melton, L., & Shahidi, F. (2018). *Encyclopedia of Food Chemistry*: Elsevier Science.
- Viedma-Poyatos, Á., González-Jiménez, P., Langlois, O., Company-Marín, I., Spickett, C. M., & Pérez-Sala, D. (2021). Protein Lipoxidation: Basic Concepts and Emerging Roles. *Antioxidants (Basel)*, *10*(2).
- Villaverde, A., & Estévez, M. (2013). Carbonylation of Myofibrillar Proteins through the Maillard Pathway: Effect of Reducing Sugars and Reaction Temperature. *Journal of Agricultural and Food Chemistry, 61*(12), 3140-3147.
- Villaverde, A., Ventanas, J., & Estévez, M. (2014). Nitrite Promotes Protein Carbonylation and Strecker Aldehyde Formation in Experimental Fermented Sausages: Are Both Events Connected? *Meat Science*, *98*(4), 665-672.
- Waisundara, V. Y., Perera, C. O., & Barlow, P. J. (2007). Effect of Different Pre-Treatments of Fresh Coconut Kernels on Some of the Quality Attributes of the Coconut Milk Extracted. *Food Chemistry*, 101(2), 771-777.
- Wang, Z., He, Z., Emara, A., Gan, X., & Li, H. (2019). Effects of Malondialdehyde as a Byproduct of Lipid Oxidation on Protein Oxidation in Rabbit Meat. *Food Chemistry*, 288, 405-412.
- Wang, Z., Tu, J., Zhou, H., Lu, A., & Xu, B. (2021). A Comprehensive Insight into the Effects of Microbial Spoilage, Myoglobin Autoxidation, Lipid Oxidation, and Protein Oxidation on the Discoloration of Rabbit Meat During Retail Display. *Meat Science*, *17*2, 108359.
- Wasowicz, E., Gramza, A., Hes, M., Jelen, H., Korczak, J., Malecka, M., Mildner-Szkudlarz, S., Rudzinska, M., Samotyja, U., & Zawirska-Wojtasiak, R. (2004). Oxidation of Lipids in Food. *Polish Journal of Food and Nutrition Sciences, 13*(Special Issue 1).

- Weber, D., Davies, M. J., & Grune, T. (2015). Determination of Protein Carbonyls in Plasma, Cell Extracts, Tissue Homogenates, Isolated Proteins: Focus on Sample Preparation and Derivatization Conditions. *Redox biology*, 5, 367-380.
- Wehr, N. B., & Levine, R. L. (2013). Quantification of Protein Carbonylation. In G. L., V. I., K. O., & K. G. (Eds.), *Cell Senescence. Methods in Molecular Biology (Methods and Protocols)* (pp. 265-281). Totowa, NJ, USA: Humana Press.
- Williams, P. (2007). Nutritional Composition of Red Meat. *Nutrition & Dietetics,* 64, S113-S119.
- Wood, J., Enser, M., Richardson, R., & Whittington, F. (2008). Fatty Acids in Meat and Meat Products. *Fatty Acids in Foods and Their Health Implications*, 3, 87-107.
- Wu, G., Cross, H. R., Gehring, K. B., Savell, J. W., Arnold, A. N., & McNeill, S. H. (2016). Composition of Free and Peptide-Bound Amino Acids in Beef Chuck, Loin, and Round Cuts1,2. *Journal of Animal Science*, 94(6), 2603-2613.
- Wyrwa, J., & Barska, A. (2017). Packaging as a Source of Information About Food Products. *Procedia Engineering, 182*, 770-779.
- Xia, C., Wen, P., Yuan, Y., Yu, X., Chen, Y., Xu, H., Cui, G., & Wang, J. (2021). Effect of Roasting Temperature on Lipid and Protein Oxidation and Amino Acid Residue Side Chain Modification of Beef Patties. *RSC Advances*, *11*(35), 21629-21641.
- Xiong, Q., Zhang, M., Wang, T., Wang, D., Sun, C., Bian, H., Li, P., Zou, Y., & Xu, W. (2020). Lipid Oxidation Induced by Heating in Chicken Meat and the Relationship with Oxidants and Antioxidant Enzymes Activities. *Poultry Science*, *99*(3), 1761-1767.
- Xiong, Y. L. (2000). Food Proteins: Processing Applications. In Nakai S & H. W. Modler (Eds.), *Meat Processing* (pp. 89–146). New York, NY: Wiley-VCH, Inc.
- Xiong, Y. L., & Guo, A. (2021). Animal and Plant Protein Oxidation: Chemical and Functional Property Significance. *Foods, 10*(1), 40.
- Xiong, Y. L., & Mikel, W. B. (2001). Meat and Meat Products. In Y. H. Hui, N. Wai-Kit, R. W. Rogers, & O. A. Young (Eds.), *Meat Science and Applications* (pp. 39-69).
- Yang, T., Liu, R., Yang, L., Yang, W., Li, K., Qin, M., Ge, Q., Yu, H., Wu, M., & Zhou, X. (2022). Improvement Strategies for Quality Defects and Oxidation of Pale, Soft and Exudative (Pse)-Like Chicken Meat: Effects

of Domestic Cooking and Core Temperature. *RSC Advances, 12*(12), 7485-7496.

- Yanishlieva, N. V., & Marinova, E. M. (1992). Inhibited Oxidation of Lipids I: Complex Estimation and Comparison of the Antioxidative Properties of Some Natural and Synthetic Antioxidants. *Lipid/Fett*, 94(10), 374-379.
- Yin, M. C., & Faustman, C. (1993). Influence of Temperature, Ph, and Phospholipid Composition Upon the Stability of Myoglobin and Phospholipid: A Liposome Model. *Journal of Agricultural and Food Chemistry*, 41(6), 853-857.
- Yu, T. Y., Morton, J. D., Clerens, S., & Dyer, J. M. (2017). Cooking Induced Protein Modifications in Meat. *Comprehensive Reviews in Food Science* and Food Safety, 16(1), 141-159.
- Yu, Y., Wang, G., Yin, X., Ge, C., & Liao, G. (2021). Effects of Different Cooking Methods on Free Fatty Acid Profile, Water-Soluble Compounds and Flavor Compounds in Chinese Piao Chicken Meat. Food Research International, 149, 110696.
- Yum, H.-W., Seo, J.-K., Jeong, J.-Y., Kim, G.-D., Rahman, M. S., & Yang, H.-S. (2018). The Quality Improvement of Emulsion-Type Pork Sausages Formulated by Substituting Pork Back Fat with Rice Bran Oil. *Korean journal for food science of animal resources*, 38(1), 123.
- Zahid, M. A., Choi, J. Y., Seo, J.-K., Parvin, R., Ko, J., & Yang, H.-S. (2020). Effects of Clove Extract on Oxidative Stability and Sensory Attributes in Cooked Beef Patties at Refrigerated Storage. *Meat Science, 161*, 107972.
- Zamora, R., & Hidalgo, F. (2011). The Maillard Reaction and Lipid Oxidation. *Lipid Technology*, 23, 59-62.
- Zayas, J. F. (2012). *Functionality of Proteins in Food*: Springer science & business media.
- Zdanowska-Sąsiadek, Ż., Marchewka, J., Horbańczuk, J. O., Wierzbicka, A., Lipińska, P., Jóźwik, A., Atanasov, A. G., Huminiecki, Ł., Sieroń, A., & Sieroń, K. (2018). Nutrients Composition in Fit Snacks Made from Ostrich, Beef and Chicken Dried Meat. *Molecules, 23*(6), 1267.
- Zhang, W., Xiao, S., & Ahn, D. U. (2013). Protein Oxidation: Basic Principles and Implications for Meat Quality. *Critical Reviews in Food Science and Nutrition*, 53(11), 1191-1201.
- Zhao, G. P., Cui, H. X., Liu, R. R., Zheng, M. Q., Chen, J. L., & Wen, J. (2011). Comparison of Breast Muscle Meat Quality in 2 Broiler Breeds. *Poultry Science*, *90*(10), 2355-2359.

Zhao, J., Chen, J., Zhu, H., & Xiong, Y. L. (2012). Mass Spectrometric Evidence of Malonaldehyde and 4-Hydroxynonenal Adductions to Radical-Scavenging Soy Peptides *Journal of Agricultural and Food Chemistry*, *60*(38), 9727-9736.

