

UNIVERSITI PUTRA MALAYSIA

CLINICAL, MICROSCOPIC AND MECHANICAL EVALUATION OF EXPANDED SKIN IN HORSES FOLLOWING SUBCUTANEOUS IMPLANTATION WITH ANISOTROPIC TISSUE EXPANDERS

SADDAM HUSSEIN MOHAMMAED SALEH

FPV 2022 17



CLINICAL, MICROSCOPIC AND MECHANICAL EVALUATION OF EXPANDED SKIN IN HORSES FOLLOWING SUBCUTANEOUS IMPLANTATION WITH ANISOTROPIC TISSUE EXPANDERS



SADDAM HUSSEIN MOHAMMAED SALEH

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

February 2022

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

CLINICAL, MICROSCOPIC AND MECHANICAL EVALUATION OF EXPANDED SKIN IN HORSES FOLLOWING SUBCUTANEOUS IMPLANTATION WITH ANISOTROPIC TISSUE EXPANDERS

By

SADDAM HUSSEIN MOHAMMAED SALEH

February 2022

Chairman : Associate Professor Nurul Hayah binti Khairuddin, PhD Faculty : Veterinary Medicine

Skin expander is a surgical an implant used to stretch cutaneous tissues through a combination of creep and biological stretch processes in other to generate extra cutaneous tissues for potential use in reconstructive skin surgeries. The understanding of the mechanical and histological features of expanded skin in horses is an important step to validate the use of tissue expander for skin extension in skin defect repair in horses. Therefore, this study aims to evaluate the clinical, mechanical, and histological properties of expanded skin following implantation of unidirectional anisotropic subcutaneous tissue expanders at different locations of horse's skin. In addition to that, possible behavioural changes in horses post tissue expander implantations were also investigated. A total of six horses were surgically implanted subcutaneously with unidirectional anisotropic tissue expanders at three different locations: the frontal region of the head, lateral side of the right shoulder, and dorsomedial part of the cannon region of the right forelimb. Each horse was clinically examined, and behaviour patterns were recorded in the stable box during the day and night-time, three days prior to - and six-days post implantation. After 14 days of subcutaneous expansion, skin biopsies of the expanded skin and representative normal skin samples from all the regions were collected. A part of each sample was subjected to mechanical properties study while the remaining was used for histological and immunohistochemical study. All horses tolerated the tissue expander implantation with no effect on the vital parameters as well as the overall horse behaviour. The mechanical properties of expanded skin from the shoulder area showed higher (11.57±1.23 MPa) elastic modulus (EM), maximum stress (MSs) (17.54±3.45 MPa), maximum strain (MSr) (177.70±58.53 %), and maximum force (MF) (150.70±59.89N), than the normal skin when compared to the forehead and lower forelimb. The overall effect of locations on EM and MSs was statistically significant (p < 0.05), however, there was no overall effect of horse factor, treatment factor (normal and expanded skin), and location interaction on the EM, MSS, MF, and MSr. The locations of the expanded or normal skin has effect on the EM and MSs with expanded skin from the frontal and distal limb areas been stiffer (less elastic) and need higher force to failure compared to the expanded skin of the shoulder. Histological evaluation revealed an increase in the thickness of the epidermal, dermal, and total skin layers with up regulation of vascular endothelial growth factor (VEGF) expression in the expanded skins. There was no significant difference in the histo-morphometric data between the three locations. The collagen fibres were more loosely packed in the expanded skin samples. The expression of VEGF was higher in the expanded skin than the normal skin. In conclusion, the implantation of unidirectional anisotropic subcutaneous tissue expanders in horses resulted in successful skin expansion with no physiological and behavioural discomfort. These findings will serve as important information when tissue expansion technique is to be applied in horses for skin reconstructive surgery according to anatomical regions. In general, skin expansion technique is considered a good technique to generate additional cutaneous tissue for equine skin reconstructive surgery.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENILAIAN KLINIKAL, MEKANIKAL DAN MICROSCOPIC KULIT YANG DIPERBAHARUI DALAM MODEL KUDA BERIKUTAN IMPLANTASI SUBKULITAN TISU PENGEMBANG ANISOTROPIK

Oleh

SADDAM HUSSEIN MOHAMMAED SALEH

Februari 2022

Pengerusi : Profesor Madya Nurul Hayah binti Khairuddin, PhD Fakulti : Perubatan Veterinar

Pengembang kulit ialah implan pembedahan yang digunakan untuk meregangkan tisu kutaneus melalui gabungan proses regangan rayapan dan regangan biologi yang lain untuk menjana tisu kutaneus tambahan untuk kegunaan yang berpotensi dalam pembedahan kulit rekonstruktif. Pemahaman tentang ciri mekanikal dan histologi pengembangan kulit pada kuda merupakan langkah penting untuk mengesahkan penggunaan tisu pengembang untuk menyambungkan kulit dan membaiki kecacatan kulit pada kuda. Oleh itu, kajian ini bertujuan untuk menilai sifat klinikal, mekanikal dan histologi kulit yang mengembang berikutan implantasi pengembang tisu subkutaneus anisotropik satu arah di lokasi berbeza pada kulit kuda. Di samping itu, perubahan tingkah laku pasca implantasi tisu pengembang kuda juga telah disiasat. Sebanyak enam ekor kuda telah melalui pembedahan secara subkutan dengan implantasi tisu pengembang anisotropik satu arah di tiga lokasi berbeza: bahagian hadapan kepala, bahagian sisi bahu kanan, dan bahagian dorsomedial kawasan keting pada kaki depan kanan. Setiap kuda telah diperiksa secara klinikal, dan corak tingkah laku direkodkan dalam kandang masing-masing pada waktu siang dan malam, tiga hari sebelum - dan enam hari selepas implantasi. Selepas 14 hari pengembangan subkutaneus, biopsi kulit bagi kulit yang mengembang dan mewakili sampel kulit normal dari semua kawasan telah dikumpulkan. Sebahagian daripada setiap sampel telah tertakluk kepada kajian sifat mekanikal kulit manakala selebihnya digunakan untuk kajian histologi dan imunohistokimia. Semua kuda bertolak ansur dengan prosedur implantasi tisu pengembang tanpa memberi perubahan kepada parameter penting dalam tingkah laku kuda secara keseluruhan. Sifat mekanikal kulit mengembang dari kawasan bahu menunjukkan (11.57±1.23 MPa) modulus elastik (EM), tegasan maksimum (MSs) (17.54±3.45 MPa). ketegangan maksimum (MSr) (177.70±58.53 %), dan maksimum daya (MF) (150.70±59.89N) yang lebih tinggi, daripada kulit biasa jika dibandingkan dengan dahi dan bawah bawah. Kesan

keseluruhan lokasi pada EM dan MS adalah signifikan secara statistik (p < 0.05), walau bagaimanapun, tiada kesan keseluruhan pada faktor kuda, faktor rawatan (kulit normal dan mengembang), dan interaksi lokasi pada EM, MSS, MF, dan MSr. Lokasi kulit yang mengembang atau normal mempunyai kesan pada EM dan MS dengan kulit yang mengembang dari bahagian depan dan bahagian anggota distal adalah lebih kaku (kurang anjal) dan memerlukan daya yang lebih tinggi untuk gagal berbanding dengan kulit bahu yang mengembang. Penilaian histologi mendedahkan peningkatan dalam ketebalan lapisan epidermis, dermis, dan jumlah kulit dengan peningkatan regulasi ekspresi faktor pertumbuhan endothelial vaskular (VEGF) dalam kulit yang mengembang. Tidak terdapat perbezaan yang signifikan dalam data histo-morfometrik antara ketiga-tiga lokasi. Gentian kolagen tersusun lebih longgar dalam sampel kulit yang mengembang. Ekspresi VEGF lebih tinggi pada kulit yang mengembang daripada kulit biasa. Kesimpulannya, implantasi tisu pengembang subkutaneus anisotropik satu arah pada kuda Berjaya menghasilkan pengembangan kulit yang tanpa ketidakselesaan fisiologi dan tingkah laku. Penemuan ini akan menjadi maklumat penting apabila teknik pengembangan tisu akan digunakan pada kuda untuk pembedahan rekonstruktif kulit mengikut kawasan anatomi. Secara umumnya, teknik pengembangan kulit dianggap sebagai teknik yang baik untuk menghasilkan tisu kulit tambahan untuk pembedahan rekonstruktif kulit kuda.

ACKNOWLEDGEMENTS

In the name of Allah, the most gracious, the most merciful.

Firstly, I thank Allah for giving me the patience, persistence, and blessings throughout my learning of study and in completing this project successfully.

Foremost, I would like to express my deepest, sincere gratitude to the chairman of my supervisory committee, Assoc. Prof. Dr. Nurul Hayah Khairuddin for her enthusiasm, support, guidance, immense knowledge sharing, and unique style of teaching. Your guidance and seemingly unending wealth of knowledge has been greatly appreciated throughout the years. Without her guidance and persistent help, this thesis would not have been possible.

To the rest supervisory committee: Dr. Intan Shameha Abdul Razak, Prof. Dr. Zamri Radzi, and Prof. Dr. Mohammad Tariqu Rahman for their support guidance, encouragement and faith in me throughout the completion of this program.

Above all, I would like to thank my father (Hussein) and my mother (Jamilah) for their constant prayers for me were what sustained me this far. Also, to my wife Dr. Ahlam Muhammed Lutf Al-Najhi for her personal support and great patience at all times and stood by me through the good and bad times. To my beloved sunshine, Sultan, Mohammed, and my daughter Dialla, they cheer me up, lift my spirit up and always make my world wonderful every time.

I would also like to extend my appreciation especially to the following:

- Post-graduates' colleagues in the Faculty of Veterinary Medicine, UPM for their kind cooperation rendered at the time of this project were carried out.
- Staffs of the Department of Farm and Exotic Animals Medicine and Surgery, at the University Veterinary Hospital, UPM for their helped throughout this study.
- Ceremonial Mounted Squadron, Malaysian Armed Forces, Malaysia: especially Dr. John Tito Sapalo for his kind help and supported me throughout the practice and experimental surgery.
- Staffs of the histopathology laboratory: Mrs. Jamilah Jahari, Mrs. Latifah Hannan, and Ms. Aisyah. My special thanks are extended to these great persons who have assisted me and making the facilities available during the research work.

- To Dr. Tengku Rinalfi Putra Tengku Azizan for his support and help me throughout the practice and experimental study.
- To Dr. Abubakar Musa Mayaki for his support and helped me throughout the practice at the histopathology lab especially in the Immunohistochemistry work.
- To my sisters and brothers: thank you for being my strongest support system. I am deeply grateful to have all of you around.

A million of thanks to all the friends I met during my study in UPM for making this place a second home away from home. Finally, I thank ALLAH (S.W.T) for HE always directs my path, answer my prayers, accept my supplications and guide me to be a successful and helpful member for the Muslim Ummah.

SADDAM HUSSEIN MOHAMMAED SALEH

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Nurul Hayah binti Khairuddin, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

Intan Shameha Abdul Razak, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

Zamri bin Radzi, DPhil

Professor Faculty of Dentistry University of Malaya (Member)

Mohamad Tariqur Rahman, PhD

Professor Faculty of Dentistry University of Malaya (Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 21 July 2022

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature:

Date:

Name and Matric No: Saddam Hussein Mohammaed Saleh,

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:		
Name of Chairman	Accesiete Drefessor	
of Supervisory	Associate Professor	
Committee:	Dr. Nurul Hayah binti Khairuddin	
Signature:		
Name of Member		
of Supervisory	Associate Professor	
Committee:	Dr. Intan Shameha Abdul Razak	
Signature:		
Name of Member		
of Supervisory	Professor	
Committee:	Dr. Zamri bin Radzi	
001111111001		

Signature: Name of Member of Supervisory Committee:

Professor Dr. Mohamad Tariqur Rahman

TABLE OF CONTENTS

				Page
APPRON DECLAF LIST OF LIST OF LIST OF	A <i>K</i> WLED /AL RATIOI TABL FIGUI	ES		i iii v vii ix xiv xv xvii xviii
CHAPTE	IR			
1	INTR 1.1 1.2 1.3 1.4 1.5 1.6	History Problem Justifica Researc	ch Background of the use of tissue expanders of Statement tion of study ch hypotheses objectives	1 1 2 3 4 5 5 5 5
2	LITE	RATURE	REVIEW	7
-5	2.1 2.2 2.3 2.4	2.1.4 Skin def Reconst	Epidermis Dermis Hypodermis (subcutis) The anatomy of the horse's skin fects and closure tructive surgery expansion History of tissue expansion	7 8 9 9 11 12 13 13 13 14 15 16 17 17 18 19 19

6

	2.5	l issue expansion techniques	21
		2.5.1 Tissue expander2.5.2 Types of tissue expander techniques	22 24
			24
		2.5.3 Nature of the growth of tissue expander: Self-inflating versus non-self-inflating	25
		2.5.4 Directions of tissue expansion	25
		•	27
	2.6		21
	2.0	The use of tissue expanders in experimental animal	20
	07	model	29
	2.7	The use of tissue expanders in veterinary medicine	20
	2.8	and surgery	30 31
	2.0	Microscopic changes of the expanded skin tissue 2.8.1 Roles of growth factors in tissue expansion	31
		2.8.2 Histological changes of the expanded skin	31
		tissue	32
	2.9	Mechanical properties of the expanded skin tissue	33
	2.9	Significance of studies	33
	2.10	Significance of studies	
3	GENE	ERAL MATERIALS AND METHODS	35
5	3.1	Ethical approval	35
	3.2	Experimental design	35
	3.3	Animal and management	37
	3.4	Tissue expanders	37
	3.5	Implantation of Skin Tissue Expanders	38
	0.0	3.5.1 Frontal site	38
		3.5.2 Shoulder site	39
		3.5.3 Forelimb site	40
	3.6	Post-operative care	41
	3.7	Behavioural and Clinical assessment	41
	3.8	Explantation of tissue expanders	43
	3.9	Skin samples collection	44
	3.10	Biomechanical (Tensile) test	45
	3.11	Tissue processing for histo-morphological analysis	
		and evaluation	47
		3.11.1 Haematoxylin and Eosin (H & E) staining	47
		3.11.2 Modified Masson's Trichrome Staining	48
		3.11.3 Immunohistochemistry staining to detect	
		expression of VEGF	48
	3.12	Statistical analysis	49
		,	
4	EFFE	CT OF SUBCUTANEOUS IMPLANTATION OF	
	ANIS	OTROPIC TISSUE EXPANDER ON HORSE	
	CLINI	CAL PARAMETERS AND BEHAVIOUR	50
	4.1	Introduction	50
	4.2	Materials and Methods	51
	4.3	Results	51
		4.3.1 Clinical assessment	51
		4.3.2 Behavioural assessment	51
	4.4	Discussion	56
	4.5	Conclusion	57

xii

5	MECHANICAL PROPERTIES OF EXPANDED SKIN FROM THREE DIFFERENT LOCATIONS IN HORSE FOLLOWING SUBCUTANEOUS IMPLANTATION OF ANISOTROPIC	
	TISSUE EXPANDERS	59
	5.1 Introduction	59
	5.2 Materials and Methods	60
	5.3 Results	60
	5.3.1 Mechanical properties	60
	5.4 Discussion	62
	5.5 Conclusion	65
6	HISTOMORPHOMETRIC AND IMMUNOHISTOCHEMICAL EXPRESSION OF VASCULAR ENDOTHELIAL GROWTH FACTOR (VEGF) IN EXPANDED SKIN OF THE HORSE FOLLOWING SUBCUTANEOUS IMPLANTATION OF	
	ANISOTROPIC TISSUE EXPANDER	66
	6.1 Introduction	66
	6.2 Materials and Methods	67
	6.3 Results	67
	6.3.1 Histo-morphological changes of horse skin	
	due to tissue expansion	67
	6.3.2 Vascular endothelial growth factor (VEGF)	
	expression in horse skin due to tissue	
	expansion	71
	6.4 Discussion	73
	6.5 Conclusion	74
7	GENERAL DISCUSSION, CONCLUSIONS AND	
	RECOMMENDATIONS FOR FUTURE STUDIES	75
	7.1 General Discussion	75
	7.2 General conclusions	78
	7.3 Limitation of study	78
	7.4 Recommendations for future studies	79
RE	FERENCES	80
API	PENDICES	99
BIC	DATA OF STUDENT	108
LIS	T OF PUBLICATIONS	109

LIST OF TABLES

Table		Page	
2.1	Difference between the types of tissue expansion	29	
3.1	Ethogram description of horse behaviours recorded before and after implantation of tissue expanders. The ethogram was based on previous studies	42	
4.1	Proportion of time that the horses spent on each behavioral parameter observed during the daytime pre and post- implantation with anisotropic tissue expanders subcutaneously	53	
4.2	Proportion of time that the horses spent on each behavioral parameter observed during the nighttime pre and post- implantation with anisotropic tissue expanders subcutaneously	53	
4.3	Mean frequency of other miscellaneous behaviors observed during the daytime pre and post-implantation with anisotropic tissue expanders subcutaneously	54	
4.4	Mean frequency of other miscellaneous behaviors observed during the nighttime pre and post-implantation with anisotropic tissue expanders subcutaneously	54	
5.1	Mechanical properties of expanded skin in comparison to normal (non-expanded) skin from three locations in horses following subcutaneous implantation of anisotropic tissue expanders	61	
5.2	Pairwise comparison of the mechanical characteristics of normal skins based on the three locations	61	
5.3	Pairwise comparison of the mechanical characteristics of expanded skins based on the three locations	62	
5.4	The effect of the horse, treatment (normal and expanded skin) and locations on the mechanical characteristics using type III model fit	62	
6.1	The epidermal, dermal, and total skin thickness of expanded and normal skin from three regions in horses	71	

LIST OF FIGURES

Figure		Page
2.1	Vertical section of equine skin from the cranial frontal region. (E) Epidermis, (P) papillary layer, (R) reticular layer, (CL) Cordovan-leather tissue layer (T) accessory cordovan layer and (S) subcutis. Notice the distinct separation of the reticular layer and cordovan-leather tissue layer of the dermis at the level of the arrows. Hematoxylin and Eosin (H&E) stain: x40	10
2.2	Schematic sequence of tissue expander inflation. At biological equilibrium, the skin is in a physiological state of resting tension, unloaded and ungrown. When an expander is implanted and inflated, the skin is stretched, loaded, and ungrown. Mechanical stretch beyond a critical level triggers a series of signaling pathways eventually leading to the creation of new skin to restore the state of resting tension, loader, and grown. Upon expander removal, elastic deformations retract and inelastic deformations remain unloaded and grown	22
3.1	The flow chart of the overall experimental design of this study	36
3.2	Self-inflating anisotropic hydrogel tissue expander devices. A. Each tissue expander is approximately (A) 10 mm in height and (B) 27mm in diameter	38
3.3	Implantation of tissue expander at the frontal region of the horse. Tacking suture was placed to keep the tissue expander in position (yellow arrow) and the incision site was sutured using cross mattress suture pattern (black arrow)	39
3.4	Implantation of tissue expander at the right shoulder region of the horse	40
3.5	Implantation of tissue expander at the dorsomedial part of the cannon region of the right forelimb of the horse	41
3.6	Procedure of explantation of the tissue expanders, A. Raised skin indicating skin expansion, B. vertical longitudinal skin incision made on the expanded skin, C. Removal of the tissue expander, D. Excised skin was sutured with 3-0 Ethilon using cross mattress suture pattern, E. Complete healing of the site after two weeks of explantation	43
3.7	The fully expanded tissue expanders 14 days post implantation. The tissue expander have increased in size: height (A) and diameter (B)	44

 \mathbf{G}

- 3.8 Surgical excision of expanded skin samples (A) of 5 mm wide, 30 mm long, and 3 mm thickness (B) from the three locations
- 3.9 Surgical excision of normal skin samples (A) of 5 mm wide, 30 mm long, and 3 mm thickness (B) from the three locations
- 3.10 Mechanical testing machine model Autograph ag-x (5kn/500N). The speed on the machine was set at a speed of 5 mm/min
- 3.11 Skin sample in between the upper and lower grip of the machine set at 5 mm apart
- 4.1 Rectal temperature (A), Heart rate (B), and Respiratory rate (C) determined pre and six days post-implantation of tissue expander subcutaneously in horses (n=6). *p < 0.05 over the time duration, ns = no significant difference between the time duration
- 4.2 Average percentages of time spent on standing without other activity during the day (A) and night (B) period of the observations. LHL left hind limb, RHL Right hind limb
- 6.1 Photomicrographs of expanded and control skin in horses (H&E, 100x). A, C and E are control skin, and B, D and F are expanded skin from the frontal area of the head, shoulder area and lower forelimb respectively
- 6.2 Photomicrographs of the dermal layers of control and expanded skin shows the arrangement of collagen fibrils (H&E, 400x). The collagen fibres are more compact and dense in expanded skin (B) than the control skin (A)
- 6.3 Photomicrographs of control and expanded skin showing the arrangement of collagen fibrils. A, C and E are control skin, and B, D and F are expanded skin from the frontal area of the head, shoulder area and lower forelimb respectively (Masson's trichrome stain, 100x)
- 6.4 The percentage collagen fibers in the dermis stained with Masson Trichrome due to tissue expansion
- 6.5 Immunohistochemical expressions of vascular endothelial growth factor (VEGF) in expanded and normal skin in horses. A, C and E are control skin, B, D and F are expanded skin from the frontal area of the head, shoulder area and lower forelimb respectively

45

45

46

47

51

68

69

69

70

72

6.6 The percentage expression and staining intensity on VEGF in horse skin due to tissue expansion



6

LIST OF APPENDICES

Appendix		Page
А	Approval of the Institutional Animal Care and Use Committee of Universiti Putra Malaysia (UPM/IACUC/AUP-R066/2018)	99
В	Permission for post graduate research students to be at UPM campus (Serdang)	100
С	Histopathological Procedure (Hematoxylin & Eosin Staining Protocol)	101
D	Masson's Trichrome Staining Protocol for Collagen Fibers	103
Е	Immunohistochemistry protocol	105

 \bigcirc

LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
CBCT	Cone Computed Topography
СМ	Centimetre
Diff	Difference
DM	Dorsomedial
DP	Dorsopalmar
DPX	Dibutylphthalate Polystyrene Xylene
EM	Elastic modulus
НВОТ	Hyperbaric oxygen therapy
H&E	Haematoxylin and Eosin
IACUC	Institutional Animal Care and Use Committee
I.M	Intramuscular
ISO	International Organization for Standardization
IU	International Unit
I.V	Intravenous
Kg	Kilogram
LHL	left hind limb
LO	Lateral Oblique
MF	Maximum force
mm	Millimeter
mmHg	Millimeter high at 0°C
MMA	Methyl Methacrylate
MSs	Maximum stress

MSr	Maximum strain		
OSMED	Hydrogel that uses the osmotic principle to gain volume		
PBS	Phosphate Buffered Saline		
PCNA	Proliferating cell nuclear antigen		
PDGF	Platelet-derived growth factor		
PMM	Polymethyl Methacrylate		
RHL	Right hind limb		
S.C	Subcutaneous		
S.D	Standard Deviation		
SPSS	Statistical Package for Social Science		
STED	Soft tissue expansion device		
UPM	Universiti Putra Malaysia		
VEGF	Vascular endothelial growth factor		
VEGF-R1	Vascular endothelial growth factor receptors 1		
VEGF-R2	Vascular endothelial growth factor receptors 2		
VP	Vinyl Pyrrolidone		

CHAPTER 1

INTRODUCTION

1.1 Research Background

The skin is the largest organ of the horse body. It regulates temperature, comforts the horse with the sense of touch and protects against the immediate environment (Wong *et al.*, 2005, Lim and Nusse, 2013). Other functions of skin include excretion, resorption, and metabolism. The main defensive function of the skin is the homeostasis maintenance that controlled loss of water, ions and serum proteins (Elias and Wakefield, 2010, Darlenski *et al.*, 2011). However, the protective function of the skin from environmental stresses such as dehydration, irradiation, mechanical trauma, and pathogenic infection is through the aid of stratified epithelia lining it. Furthermore, the hair and sebaceous glands help in thermoregulation (Lim and Nusse, 2013).

Nevertheless, horses are prone to traumatic injury because of their fright and flight response and the nature of its environment that exposed horses to injury such as nail, metals or sharp object at the stables or surroundings. The resulting wound can be small or large. Managing small wound is not a problem because it undergoes normal healing process of inflammation, repair and maturation (Baxter, 2004, Knottenbelt, 2013, Theoret and Schumacher, 2017). In the case of large open wound mostly due to accident, burn or dead space following surgical removal of tumour growth, reconstructive surgery is usually required (Zöllner *et al.*, 2012a). Reconstructive surgery is an option to surgically replace the wound tissues with viable tissues. This is done for the purposes of anatomical restoration or cosmetic reasons.

Tissue expansion technique is a type of reconstructive surgery where tissue expander is used to mechanically generate excess soft tissue adjacent to a defect. The significance of this technique is that it allows replacement of loss tissue with analogous tissue with matching texture, colour and other important local features (Austad and Rose, 1982, Argenta et al., 1983b, Radovan, 1984, Pisarski et al., 1998, Agrawal and Agrawal, 2012, Tepole et al., 2012). The process by which the expanded skin tissue keep the same structural and morphological features of the skin collagen and elastin fibers is term mechanical creep (Wilhelmi et al., 1998). The application of tissue expanders was first carried out in a reconstructive surgery as early as 1900 but the clinical trial of the phenomenon was first described in humans, where an inflated rubber balloon was implanted subcutaneously to gain more skin tissue for the reconstruction of an external ear defect (Neumann, 1957). Decades later, the expansion process for breast reconstruction was pursued (Radovan, 1982). The application of tissue expanders in plastic reconstructive surgery has also been exploited by orthopaedic surgeons (Argenta et al., 1983a, Argenta et al.,

1983b, Radovan, 1984). The first attempt to modify the conventional tissue expansion for better result was documented in 1984 (Sasaki and Pang, 1984). Since then, this technique became of immense value and added advantage over the flap surgery technique.

The fact that tissues have the ability to stretch made the application of tissue expanders an accepted practice over the last century in humans particularly in breast reconstruction, paediatric plastic surgery and other skin restorations (Cordeiro and McCarthy, 2006, Yeşilada *et al.*, 2013). Lately, the clinical application of skin tissue expansion was proven useful and it has been performed routinely for repair and reconstruction of skin defects in cosmetic surgery in human (McCauley, 2005, Motamed *et al.*, 2008, Fochtmann *et al.*, 2013, Lei *et al.*, 2015, Wang *et al.*, 2016).

The mechanism of creating additional skin by this protocol, and the physiologic changes occurring in the skin during the process of expansion have been studied in animal models (Sasaki and Pang, 1984). The implantation of silicone device tissue expanders subcutaneously in horses and dog to generate addition skin flap to repair defect have been described (Madison *et al.*, 1989).

However, despite the success of skin expansion with tissue expanders, its application in equine reconstructive surgery is yet to received attention. Most probably, the issues of the traditional type of the tissue expanders need to be addressed if this technique is to be applied clinically in the equine practice.

1.2 History of the use of tissue expanders

Recently, tissue expansion is considered one of the most significant innovative techniques in the reconstructive surgery field. The first reported tissue expansion utilization in reconstructive plastic surgery was in human in 1957 where an artificial subcutaneously implanted air-filled rubber balloon was inflated from outside the host body. In the year 1960 the use of hydrogels was suggested by Wichterle and Lim where it was used in a biomedical application (Wichterle and Lim, 1960). It was noted that hydrogel possessed the ability to swell, but it does not dissolve in aqueous media.

The use of a controlled slow expansion technique that require inflating silicon balloon with a saline solution through the skin filling port has been reported (Austad and Rose, 1982, Radovan, 1982, 1984). The high stretch response of silicon made it an important material for this technique. The mechanism of tissue expansion by controlled slow expansion technique is based on the osmotic gradient created by the saline (i.e. sodium and chloride) on the implanted silicon wall. Therefore, to provide an osmotic driving force the solute

should be non-toxic and hypertonic (Austad and Rose, 1982). It has also been noted that the technique results in instant intraoperative tissue expansion where the skin stretches and widens in surface area (Sasaki, 1984).

Methyl Methacrylate (MMA) and Vinyl-pyrolidone (VP) were another type of osmotic driven tissue expander used for tissue expansion. They have been used extensively in the repair of cleft palate, breast reconstruction and congenital anophthalmia (Cordeiro and McCarthy, 2006, Gundlach *et al.*, 2005, Yeşilada *et al.*, 2013).The MMA and VP possess ability to absorb body fluids, thus increasing the size of the dry gel and causing expansion of the tissue without any practical external intervention (Wiese, 1993). However, the excessive rapid uncontrolled expansion which may lead to tissue necrosis became a limitation to their uses (Chummun *et al.*, 2010).

In order to minimize the possible complications associated with uncontrolled tissue expanders, co-polymer based hydrogel expander which allows considerable self-inflating expansion has been newly developed and its animal pre-clinical trials was tested in pigs (Swan *et al.*, 2012). The use of anisotropic self-inflating tissue expander that has the ability to expand unidirectional gave a positive outcome when tested in a craniofacial plastic reconstructive skin procedure (Swan *et al.*, 2011). Research on mechanical properties of expanded skin from animals such as pig, goat, sheep, and dog have been published (Zeng *et al.*, 2004, Zhang *et al.*, 2006, Manssor *et al.*, 2016). Although, in horses, there have been studies of mechanical behaviours of soft tissues particularly tendon and ligament (Souza *et al.*, 2010, Thorpe *et al.*, 2010), the investigation of skin expansion and its mechanical properties is relatively new in equine clinical research (Al-Majhali *et al.*, 2018, Whittaker *et al.*, 2020).

1.3 Problem Statement

In equine clinical setting, common surgical coverage of large wound at the distal part of horse's legs and face cannot be easily performed due to inadequate skin for closure (Zöllner *et al.*, 2012b). Therefore, large wound in horses has mostly been manage as an open wound, thus the delay in healing period with scar formation resulting in deformation of the injured area. The physiologic changes occurring in the skin during the process of expansion have been studied (Sasaki and Pang, 1984, Maher and Kuebelbeck, 2018). However, considering the high success rate of tissue expander in human plastic surgery, its use in veterinary medicine will ought to have recived some level of attention. Currently, research regarding the application of tissue expander to repair defect in animals is scarce (Madison *et al.*, 1989).

The issue of traditional expanders had been debated in clinical application in equine practice and the use of tissue expanders have been proposed as an alternative technique to skin grafting (Whittaker *et al.*, 2020). It is therefore expected that its application in equine clinical practice will offer an effective alternative to traditional wound grafts.

1.4 Justification of study

To date, an understanding of the mechanical and histological features of expanded skin is required to validate the use of tissue expander for skin extension in skin defect repair in horses. It is therefore our expectation that the investigation into mechanical, histological and immunohistochemical properties of expanded skin using tissue expander is the most common parts of horses susceptible to injury will enrich our understanding of the factors associated with using tissue expansion in managing large wound in horses. Furthermore, there is a need to determine any possible behavioural response to the tissue expander implantation. This is very important to understand if there will be tolerance to the procedure in the horse since the implant will beleft for sometime usually two or more weeks to allow for the expansion of the skin tissue. Afterall, behaviour response is one important factor used to measure animal welfare (Budras et al., 2012, Chung et al., 2018). Moreso, this will be the first study to assess behavioural changes in horses during tissue expansion process. Therefore, any deviation from the normal or usual horse behaviour following the implantation of tissue expander may render the procedure unfavourable for application in the horse. Hence, the results of this study will offer insights to the future developments and application of tissue expander in the field of cosmetic surgery in horse species.

Tissue expander has serve as an excellent strategy to grow skin without changing skin color, texture, hair balance, and thickness of the surrounding healthy skin. More importantly, the cells of expanded tissue have been shown to differentiate normally while keeping its characteristic phenotype on both histological and Immunohistochemical analysis. Furthermore, the use of tissue expander has reduces the incidence of infections associated of skin grafting and speed up the process of healing wounds without scar.

An anisotropic hydrogel tissue expander which is capable of controlled expansion against the uncontrolled expansion associated with an anisotropic expander will be used in this study. Its application in equine reconstructive surgery will help to ameliorate the challenges associated with closure of large wounds, which are most difficult to achieve.

1.5 Research hypotheses

Hypothesis 1

Ho: The mechanical properties of expanded skin due to subcutaneous implantation of anisotropic tissue expander is different from that of normal skin.

Ha: The mechanical properties of expanded skin due to subcutaneous implantation of anisotropic tissue expander is not different from that of normal skin.

Hypothesis 2

Ho: There are no behavioual changes to subcutaneous implantation of anisotropic tissue expander in horses.

Ha: There are behavioual changes to subcutaneous implantation of anisotropic tissue expander in horses.

Hypothesis 3

Ho: There are no difference in the histomorphometric features and expression of vascular endothelial growth factor (VEGF) between expanded and normal skin in the horses.

Ha: There are difference in the histomorphometric features and expression of vascular endothelial growth factor (VEGF) between expanded and normal skin in the horses.

1.6 Aim and objectives

1.6.1 Main aim

The main aim of this study is to investigate the clinical, mechanical, and histological properties of expanded skin at different locations in the horse.

1.6.2 Specific objectives

i. To investigate the possible behavioural changes in horses towards implantation of tissue expander subcutaneously.

- ii. To determine the mechanical properties of expanded skin from different locations in horses following subcutaneous anisotropic self-inflating tissue expander implantation.
- iii. To evaluate histo-morphological changes in expanded skin of horses in comparison to non-expanded skin.
 - 1) To study the histological properties of the cutaneous tissue affected by implantation of self-inflating tissue expanders at different sites of horse skin.
 - 2) To test the occurrence and assess the expression of the vascular endothelial growth factor (VEGF) in tissue expanded at different sites of horse skin.



REFERENCES

- Abrahamsson, P., Isaksson, S., Gordh, M. & Andersson, G. 2009. Periosteal expansion of rabbit mandible with an osmotic self-inflatable expander. *Scandinavian Journal of Plastic and Reconstructive Surgery and Hand Surgery*, 43, 121-125.
- Abramo, F., Pirone, A., Lenzi, C., Vannozzi, I., Della Valle, M. F. & Miragliotta, V. 2016. Establishment of a 2-week canine skin organ culture model and its pharmacological modulation by epidermal growth factor and dexamethasone. *Annals of Anatomy-Anatomischer Anzeiger*, 207, 109-117.
- Agrawal, K. & Agrawal, S. 2012. Tissue regeneration during tissue expansion and choosing an expander. *Indian Journal of Plastic Surgery*, 45, 7-15.
- Ahmad, M. 2010. Tissue expansion. *The Professional Medical Journal*, 17, 611-615.
- Al-Majhali, S. H. 2017. The Implantation of Anisotropic Self-Inflating Tissue Expanders in Various Sites of Horses Skin. Master Thesis, UPM.
- Al-Majhali, S. H., Khairuddin, N. H., Intan-Shameha, A. R., Radzi, Z., Rahman, M. T., Sapalo, J. T., Mayaki, A. M. & Czernuszka, J. T. 2021. Biomechanical Effects of Unidirectional Expansion Using Anisotropic Expanders in Horse Skin Tissue. *Journal of Equine Veterinary Science*, 99, 0737-0806.
- Al-Majhali, S. H., Khairuddin, N. H., Loqman, M. Y., Radzi, Z., Chen, H. C., Lau, S. F. & Rahman, M. T. 2018. Physical measurement of the expansion rate of anisotropic tissue expander in the skin of the horse. *Pertanika Journal of Tropical Agricultural Science*, 41, 1899 -1904.
- Ali, A., Abubakar, A., Kaka, U., Radzi, Z., Khairuddin, N., Yusoff, M. & Loqman, M. 2018. Histological changes of immediate skin expansion of the distal limb of rats. *Veterinary World*, 1706-1711.
- Altamura, G., Strazzullo, M., Corteggio, A., Francioso, R., Roperto, F., D'Esposito, M. & Borzacchiello, G. 2012. O 6-methylguanine-DNA methyltransferase in equine sarcoids: molecular and epigenetic analysis. *BMC Veterinary Research*, 8, 218.
- Annaidh, A. N., Bruyère, K., Destrade, M., Gilchrist, M. D. & Otténio, M. 2012. Characterization of the anisotropic mechanical properties of excised human skin. *Journal of the Mechanical Behavior of Biomedical Materials*, 5, 139-148.

- Antonyshyn, O., Gruss, J., Mackinnon, S. & Zuker, R. 1988. Complications of soft tissue expansion. *British Journal of Plastic Surgery*, 41, 239-250.
- Anwander, T., Schneider, M., Gloger, W., Reich, R. H., Appel, T., Martini, M., Wenghoefer, M., Merkx, M. & Bergé, S. 2007. Investigation of the expansion properties of osmotic expanders with and without silicone shell in animals. *Plastic and Reconstructive Surgery*, 120, 590-595.
- Argenta, L., Watanabe, M., Grabb, W. & Newman, M. 1981. Soft tissue expanders in head and neck surgery: A new method of reconstruction. *Plastic Surgery Forum Australia.*
- Argenta, L. C. 1984. Controlled tissue expansion in reconstructive surgery. British Journal of Plastic Surgery, 37, 520-529.
- Argenta, L. C., Marks, M. W. & Grabb, W. C. 1983a. Selective use of serial expansion in breast reconstruction. *Annals of Plastic Surgery*, 11, 188-195.
- Argenta, L. C., Watanabe, M. J. & Grabb, W. C. 1983b. The use of tissue expansion in head and neck reconstruction. *Annals of Plastic Surgery*, 11, 31-37.
- Arifin, W. N. & Zahiruddin, W. M. 2017. Sample size calculation in animal studies using resource equation approach. *The Malaysian Journal of Medical Sciences*, 24, 101.
- Ashley, F., Waterman-Pearson, A. & Whay, H. 2005. Behavioural assessment of pain in horses and donkeys: application to clinical practice and future studies. *Equine Veterinary Journal*, 37, 565-575.
- Austad, E. A self-inflating tissue expander. Annual Meeting of the American Society of Plastic and Reconstructive Surgeons, 1979 Toronto.
- Austad, E. 1987. Complications in tissue expansion. *Clinics in Plastic Surgery*, 14, 549.
- Austad, E. D., Pasyk, K. A., McClatchey, K. D. & Cherry, G. W. 1982. Histomorphologic evaluation of guinea pig skin and soft tissue after controlled tissue expansion. *Plastic and Reconstructive Surgery*, 70, 704-710.
- Austad, E. D. & Rose, G. L. 1982. A self-inflating tissue expander. *Plastic and Reconstructive Surgery*, 70, 588.
- Austad, E. D., Thomas, S. B. & Pasyk, K. 1986. Tissue expansion: dividend or loan? *Plastic and Reconstructive Surgery*, 78, 63-67.

- Baker, K. & Thomsett, L. 1990. *Canine and Feline Dermatology,* Oxford OX2 0EL, UK, Blackwell Scientific Publications.
- Baker, S. R. 1991. Fundamentals of expanded tissue. *Head & Neck,* 13, 327-333.
- Barker, D. E., Dedrick, D. K., Burney, R. E., Mathes, S. J. & Mackenzie, J. R. 1987. Resistance of rapidly expanded random skin flaps to bacterial invasion. *The Journal of Trauma*, 27, 1061-1065.
- Bates, D. O. & Jones, R. O. P. 2003. The role of vascular endothelial growth factor in wound healing. *Journal of Lower Extremity Wounds*, 2, 107-120.
- Baxter, G. M. 2004. Management of wounds involving synovial structures in horses. *Clinical Techniques in Equine Practice*, 3, 204-214.
- Bell, C. L. & Peppas, N. A. 1996. Water, solute and protein diffusion in physiologically responsive hydrogels of poly (methacrylic acid-g-ethylene glycol). *Biomaterials*, 17, 1203-1218.
- Benjamini, Y. & Hochberg, Y. 1995. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society*, 57, 289-300.
- Bonomo, D. 1986. Subperiosteal tissue expanders for ridge augmentation. CDS review, 79, 34.
- Booth, L. 1991. Equine wound reconstruction using free skin grafting. *Calif Vet*, 45, 13-16.
- Bos, J. D. & Kapsenberg, M. L. 1986. The skin immune system Its cellular constituents and their interactions. *Immunology Today*, 7, 235-240.
- Bowser, J., Elder, S., Pasquali, M., Grady, J., Rashmir-Raven, A., Wills, R. & Swiderski, C. 2014. Tensile properties in collagen-rich tissues of Q uarter H orses with hereditary equine regional dermal asthenia *Equine Veterinary Journal*, 46, 216-222.
- Branco, A., Yoshikawa, F., Pietrobon, A. J., & Sato, M. N. 2018. Role of histamine in modulating the Immune response and inflammation. *Mediators of Inflammation*, 9524075.
- Bristol, D. G. 2005. Skin grafts and skin flaps in the horse. Veterinary Clinics of North America: Equine Practice, 21, 125-144.
- Brown, M. D. & Johnson, T. M. 1993. Complications of tissue expansion. *The Journal of Dermatologic Surgery and Oncology*, 19, 1120.

- Budras, K. D., Sack, W. O., Rock, S., Horowitz, A. & Berg, R. 2012. *Anatomy* of the Horse, London, Schlütersche.
- Calle-Pascual, A. L., Garcia-Torre, N., Moraga, I., Diaz, J. A., Duran, A., Moñux, G., Serrano, F. J., Martín-Alvarez, P. J., Charro, A. & Marañes, J. P. 2001. Epidemiology of nontraumatic lower-extremity amputation in area 7, Madrid, between 1989 and 1999: a population-based study. *Diabetes Care*, 24, 1686-1689.
- Canady, J. W., Squier, C. A., Kelly, K. M. & Bardach, J. 1990. Serial measurement of blood flow in expanded tissue by laser Doppler velocimetry. *Otolaryngology-Head and Neck Surgery*, 103, 986-990.
- Carr, J. A. 2014. Tissue expander-assisted ventral hernia repair for the skingrafted damage control abdomen. *World Journal of Surgery*, 38, 782-787.
- Casanova, D., Bali, D., Bardot, J., Legre, R. & Magalon, G. 2001. Tissue expansion of the lower limb: complications in a cohort of 103 cases. *British Journal of Plastic Surgery*, 54, 310-316.
- Chandawarkar, R. Y., Cervino, A. L. & Pennington, G. A. 2003. Intraoperative acute tissue expansion revisited: a valuable tool for challenging skin defects. *Dermatologic Surgery*, 29, 834-838.
- Chummun, S., Addison, P. & Stewart, K. 2010. The osmotic tissue expander: a 5-year experience. *Journal of Plastic, Reconstructive & Aesthetic Surgery*, 63, 2128-2132.
- Chung, E. L. T., Khairuddin, N. H., Azizan, T. R. P. T. & Adamu, L. 2018. Sleeping patterns of horses in selected local horse stables in Malaysia. *Journal of Veterinary Behavior*, 26, 1-4.
- Collins, S. A. & Swanson, N. A. 1993. Chronic tissue expansion. *The Journal of Dermatologic Surgery and Oncology*, 19, 1090-1098.
- Copcu, E., Sivrioglu, N., Sisman, N., Aktas, A. & Oztan, Y. 2003. Enhancement of tissue expansion by calcium channel blocker: a preliminary study. *World Journal of Surgical Oncology*, 1, 19.
- Cordeiro, P. G. & McCarthy, C. M. 2006. A single surgeon's 12-year experience with tissue expander/implant breast reconstruction: part I. A prospective analysis of early complications. *Plastic and Reconstructive Surgery*, 118, 825-831.
- Darlenski, R., Kazandjieva, J. & Tsankov, N. 2011. Skin barrier function: morphological basis and regulatory mechanisms. *Journal of Clinical Medicine* 4, 36-45.

- De Filippo, R. E. & Atala, A. 2002. Stretch and growth: the molecular and physiologic influences of tissue expansion. *Plastic and Reconstructive Surgery*, 109, 2450-2462.
- De Lorenzi, M., Swan, M., Easter, C. & Chanoit, G. 2018. Outcome of reconstruction of cutaneous limb defects in dogs using hygroscopic "selfinflating" tissue expanders. *Journal of Small Animal Practice*, 59, 98-105.
- De Oliveira MGC, Luna SPL, Nunes TL, et al. 2020. Post-operative pain behaviour associated with surgical castration in donkeys (Equus asinus). Equine Veterinary Journal. 00:1–6.
- De Oliveira MGC, Luna SPL, Nunes TL, et al. 2020. Post-operative pain behaviour associated with surgical castration in donkeys (Equus asinus). Equine Veterinary Journal. 00:1–6.
- Dellman, H. 1993. Textbook of veterinary histology. *Textbook of veterinary histology. 4th edition. Philadelphia, Lea and Febiger.* Philadelphia: Lea and Febiger.
- Dhaval, N. S. 2015. A Histopathological and Immunohistochemical Study of Cutaneous Nodular Lesions and Other Surface Swellings of Kenyan Horses. Masters Degree, University of Nairobi.
- Ding, J., Lei, L., Liu, S., Zhang, Y., Yu, Z., Su, Y. & Ma, X. 2019. Macrophages are necessary for skin regeneration during tissue expansion. *Journal of Translational Medicine*, 17, 1-10.
- Dombi, G. W., Haut, R. C. & Sullivan, W. G. 1993. Correlation of high-speed tensile strength with collagen content in control and lathyritic rat skin. *Journal of Surgical Research*, 54, 21-28.
- Dotan, L., Icekson, M., Yanko-Arzi, R., Ofek, A., Neuman, R. & Margulis, A. 2009. Pediatric tissue expansion: Our experience with 103 expanded flap reconstructive procedures in 41 children. *The Israel Medical Association journal*, 11, 474.
- El-Hawary, M. E. 1998. *Electric power applications of fuzzy systems*, Wiley-IEEE Press.
- Elias, P. M. & Wakefield, J. S. 2010. Skin Barrier Function. In: Krutmann J., Humbert P. (eds). *Nutrition for healthy skin*. Berlin, Heidelberg. https://doi.org/10.1007/978-3-642-12264-4_4: Springer.
- Eming, S. A. & Krieg, T. Molecular mechanisms of VEGF-A action during tissue repair. Journal of Investigative Dermatology Symposium Proceedings, 2006. Elsevier, 79-86.

- Eurell, J. & Frappier, B. 2006. *Text Book of Veterinary Histology. 6th ed.,* USA, Black Well
- Evans, J. W. 2000. *Horses: a guide to selection, care, and enjoyment. 3rd ed. ,* Freeman, New York., Macmillan.
- Fang, L., Zhou, C. & Yang, M. 2013. 'Expansion in-situ'concept as a new technique for expanding skin and soft tissue. *Experimental and Therapeutic Medicine*, 6, 1295-1299.
- Fenton, O. 1987. Expanding possibilities. *British Medical Journal (Clinical Research ed.)*, 295, 684.
- Ferrara, N. 2001. Role of vascular endothelial growth factor in regulation of physiological angiogenesis. *American Journal of Physiology*, 280, C1358-C1366.
- Finlay, M., Yuan, Z., Morgan, I. M., Campo, M. S. & Nasir, L. 2012. Equine sarcoids: Bovine papillomavirus type 1 transformed fibroblasts are sensitive to cisplatin and UVB induced apoptosis and show aberrant expression of p53. *Veterinary Research*, 43, 1-8.
- Fochtmann, A., Keck, M., Mittlbock, M. & Rath, T. 2013. Tissue expansion for correction of scars due to burn and other causes: a retrospective comparative study of various complications. *Burns*, **3**9, 984-989.
- Frank, S., Hübner, G., Breier, G., Longaker, M. T., Greenhalgh, D. G. & Werner, S. 1995. Regulation of vascular endothelial growth factor expression in cultured keratinocytes. Implications for normal and impaired wound healing. *Journal of Biological Chemistry*, 270, 12607-12613.
- Fuchs, C., Kiefner, C., Reese, S., Erhard, M. & Wohr, A. 2016. Narcolepsy: do adult horses really suffer from a neurological disorder or rather from a recumbent sleep deprivation/rapid eye movement (REM)-sleep deficiency? Equine Veterinary Jurnal, 48, 9.
- Gartner, L. P. & Hiatt, J. L. 2006. *Color textbook of histology e-book*, Elsevier Health Sciences.

Geneser, F. 1986. *Textbook of histology*, Denmark, Munksgaard.

- Gibson, T., Kenedi, R. & Craik, J. 1965. The mobile micro-architecture of dermal collagen: A bio-engineering study. *British Journal of Surgery*, 52, 764-770.
- Gibstein, L. A., Abramson, D. L., Bartlett, R. A., Orgill, D. P., Upton, J. & Mulliken, J. B. 1997. Tissue expansion in children: a retrospective study of complications. *Annals of Plastic Surgery*, 38, 358-364.

- Greenbaum, S. S. & Greenbaum, C. H. 1990. Intraoperative tissue expansion using a Foley catheter following excision of a basal cell carcinoma. *The Journal of Dermatologic Surgery and Oncology*, 16, 45-48.
- Greening, L., Shenton, V., Wilcockson, K. & Swanson, J. 2013. Investigating duration of nocturnal ingestive and sleep behaviors of horses bedded on straw versus shavings. *Journal of Veterinary Behavior*, 8, 82-86.
- Gregonis, D. E., Chen, C. M. & Andrade, J. D. 1976. The Chemistry of Some Selected Methacrylate Hydrogels. *Hydrogels for Medical and Related Applications*. ACS Publications.
- Guzey, S., Alhan, D., Şahin, I., Aykan, A., Eski, M. & Nişancı, M. 2015. Our experiences on the reconstruction of lateral scalp burn alopecia with tissue expanders. *Burns*, 41, 631-637.
- Harini, G. & Kaarthikeyan, G. 2014. Periodontal soft tissue expanders-a review. *IOSR Journal of Dental and Medical Sciences*, 13, 31-35.
- Hawary, M. B. 1998. Tissue expansion: King Khalid university experience. Annals of Saudi Medicine, 18, 454-456.
- He, J. F., Visscher, C., van Waas, M. A. & Wismeyer, D. 2010. Expanding soft tissue with Osmed® tissue expanders in the goat maxilla. *Clinical Oral Implants Research*, 22, 121-128.
- Hemmer, K. M., Marsh, J. L. & Picker, S. 1987. Calvarial erosion after scalp expansion. *Annals of Plastic Surgery*, 19, 454-459.
- Hill, D., Moss, N., Pomery, P. & Whittaker, A. 2000. Copolymer hydrogels of 2hydroxyethyl methacrylate with n-butyl methacrylate and cyclohexyl methacrylate: synthesis, characterization and uptake of water. *Polymer*, 41, 1287-1296.
- Hitchon, C. A., Danning, C. L., Illei, G. G., El-Gabalawy, H. S. & Boumpas, D. T. 2002. Gelatinase expression and activity in the synovium and skin of patients with erosive psoriatic arthritis. *The Journal of Rheumatology*, 29, 107-117.
- Hughes, I. 1987. Steroids and Growth. British Medical Journal (Clinical Research ed.), 295, 683.
- Huh, M.-I., An, S. H., Kim, H.-G., Song, Y.-J., Choi, E.-C., An, S.-H., Choi, W.-S., Huh, J. S. & Lim, J. O. 2016. Rapid expansion and auto-grafting efficiency of porcine full skin expanded by a skin bioreactor ex vivo. *Tissue Engineering and Regenerative Medicine*, 13, 31-38.

- Iatridis, J. C., Wu, J., Yandow, J. A. & Langevin, H. M. 2003. Subcutaneous tissue mechanical behavior is linear and viscoelastic under uniaxial tension. *Connective tissue research*, 44, 208-217.
- Ichijo, R., Kobayashi, H., Yoneda, S., Iizuka, Y., Kubo, H., Matsumura, S., Kitano, S., Miyachi, H., Honda, T. & Toyoshima, F. 2017. Tbx3dependent amplifying stem cell progeny drives interfollicular epidermal expansion during pregnancy and regeneration. *Nature Communications*, 8, 1-13.
- Ince, B., Dadaci, M., Oltulu, P., Altuntas, Z. & Bilgen, F. 2015. Effect of dermal thickness on scars in women with type III–IV fitzpatrick skin. *Aesthetic plastic surgery*, 39, 318-324.
- Jacovetti, C., Abderrahmani, A., Parnaud, G., Jonas, J., Peyot, M., Cornu, M., Laybutt, R., Meugnier, E., Rome, S. & Thorens, B. 2012. Micro RNAs contribute to compensatory β cell expansion during pregnancy and obesity. *Journal of Clinical Investigation*, 122, 3541-3551.
- Johnson, K. E. & Wilgus, T. A. 2014. Vascular endothelial growth factor and angiogenesis in the regulation of cutaneous wound repair. *Advances in Wound Care*, 3, 647-661.
- Johnson, P. E., Kernahan, D. A. & Bauer, B. S. 1988. Dermal and epidermal response to soft-tissue expansion in the pig. *Plastic and Reconstructive Surgery*, 81, 390-397.
- Johnson, T. M., Brown, M. D., Sullivan, M. J. & Swanson, N. A. 1990. Immediate intraoperative tissue expansion. *Journal of the American Academy of Dermatology*, 22, 283-287.
- Johnson, T. M., Lowe, L., Brown, M. D., Sullivan, M. J. & Nelson, B. R. 1993. Histology and physiology of tissue expansion. *The Journal of Dermatologic Surgery and Oncology*, 19, 1074-1078.
- Jørgensen, E., Bay, L., Bjarnsholt, T., Bundgaard, L., Sørensen, M. & Jacobsen, S. 2017. The occurrence of biofilm in an equine experimental wound model of healing by secondary intention. *Veterinary Microbiology*, 204, 90-95.
- Jørgensen, E., Lazzarini, G., Pirone, A., Jacobsen, S. & Miragliotta, V. 2018. Normal microscopic anatomy of equine body and limb skin: a morphological and immunohistochemical study. *Annals of Anatomy-Anatomischer Anzeiger*, 218, 205-212.
- Ju, Z., Wei, J., Guan, H., Zhang, J., Liu, Y. & Feng, X. 2012. Effects of hyperbaric oxygen therapy on rapid tissue expansion in rabbits. *Journal* of *Plastic Reconstructive & Aesthetic Surgery*, 65, 1252-1258.

- Kaner, D., Zhao, H., Terheyden, H. & Friedmann, A. 2014. Submucosal implantation of soft tissue expanders does not affect microcirculation. *Clinical Oral Implants Research*, 25, 867-870.
- Kaner, D., Zhao, H., Terheyden, H. & Friedmann, A. 2015. Improvement of microcirculation and wound healing in vertical ridge augmentation after pre-treatment with self-inflating soft tissue expanders-a randomized study in dogs. *Clinical Oral Implants Research*, 26, 720-724.
- Keller, W., Aron, D., Rakich, P., Crowe, D. & Marks, M. 1994. Rapid tissue expansion for the development of rotational skin flaps in the distal portion of the hindlimb of dogs: an experimental study. *Veterinary Surgery*, 23, 31-39.
- Kibblewhite, D., Larrabee, W. F. & Sutton, D. 1992. The effect of relaxin on tissue expansion. Archives of Otolaryngology-Head & Neck Surgery, 118, 153-156.
- Knottenbelt, D. C. 2013. Handbook of Equine Wound Management E-Book, London W1T 4LP, Elsevier Health Sciences.
- Kolarsick, P. A., Kolarsick, M. A. & Goodwin, C. 2011. Anatomy and physiology of the skin. *Journal of the Dermatology Nurses' Association*, 3, 203-213.
- König, H. E., Hans-Georg, H.-G. & Bragulla, H. 2007. Veterinary anatomy of domestic mammals, Germany, Schattauer Verlag.
- Korsmeyer, R. W. & Peppas, N. A. 1981. Effect of the morphology of hydrophilic polymeric matrices on the diffusion and release of water soluble drugs. *Journal of Membrane Science*, 9, 211-227.
- Kotb, M. & Soliman, M. 2007. Guidelines to minimize the complications of tissue expansion. Egypt Journal Plastic Reconstructive Surgery, 31, 79-82.
- Lamalice, L., Le Boeuf, F. & Huot, J. 2007. Endothelial cell migration during angiogenesis. *Circulation Research*, 100, 782-794.
- Lantieri, L. A., Martin-Garcia, N., Wechsler, J., Mitrofanoff, M., Raulo, Y. & Baruch, J. P. 1998. Vascular endothelial growth factor expression in expanded tissue: a possible mechanism of angiogenesis in tissue expansion. *Plastic and Reconstructive Surgery*, 101, 392-397.
- Laurence, V. G., Martin, J. B. & Wirth, G. A. 2012. External tissue expanders as adjunct therapy in closing difficult wounds. *Journal of Plastic, Reconstructive & Aesthetic Surgery*, 65, e297-e299.
- Leach, D. & Caron, J. 1999. *Integumentary system,* Philadelphia, WB Saunders.

- Lee, K., Silva, E. A. & Mooney, D. J. 2011. Growth factor delivery-based tissue engineering: general approaches and a review of recent developments. *Journal of the Royal Society Interface*, 8, 153-170.
- Lei, J., Hou, C., Duan, P., Hao, Z., Zhai, Y. & Meng, Y. 2015. Clinical application of modified skin soft tissue expansion in early repair of devastating wound on the head due to electrical burn. *Chinese Journal of Burns*, 31, 406-409.
- Lentz, A. & Bauer, B. 2014. *Tissue expansion,* Philadelphia, USA: Lippincott Williams & Wilkins.
- Leong, T. Y., Cooper, K. & Leong, A. S. 2010. Immunohistology past, present, and future. *Advances in Anatomic Pathology*, 17, 404-418.
- Lew, D., Clark, R. & Shahbazian, T. 1986. Use of a soft tissue expander in alveolar ridge augmentation: a preliminary report. *Journal of Oral and Maxillofacial Surgery*, 44, 516-519.
- Lim, X. & Nusse, R. 2013. Wnt signaling in skin development, homeostasis, and disease. *Cold Spring Harbor Perspectives in Biology*, 5, a008029.
- Logiudice, J. & Gosain, A. K. 2004. Pediatric tissue expansion: indications and complications. *Plastic Surgical Nursing*, 24, 20-26.
- Lohana, P., Moiemen, N. & Wilson, Y. 2012. The use of OsmedTM tissue expanders in paediatric burns reconstruction. *Annals of Burns and Fire Disasters*, 25, 38.
- Lohsiriwat, V., Peccatori, F. A., Martella, S., Azim Jr, H. A., Sarno, M. A., Galimberti, V., De Lorenzi, F., Intra, M., Sangalli, C. & Rotmensz, N. 2013. Immediate breast reconstruction with expander in pregnant breast cancer patients. *The Breast*, 22, 657-660.
- Machida, B. K., Liu-Shindo, M., Sasaki, G. H., Rice, D. H. & Chandrasoma, P. 1991. Immediate versus chronic tissue expansion. *Annals of Plastic Surgery*, 26, 227-231.
- Madison, J. B., Donawick, W. J., Johnston, D. E. & Orsini, R. A. 1989. The use of skin expansion to repair cosmetic defects in animals. *Veterinary Surgery*, 18, 15-21.
- Magnuson, P. 1913. Lengthening shortened bones of the leg by operation. Ivory screws with removable heads as a means of holding the two bone fragments. *Surg Gynecol Obstet*, 17, 63-71.
- Maher, M. & Kuebelbeck, L. 2018. Nonhealing wounds of the equine limb. *Veterinary Clinics of North America. Equine Practice*, 34, 539-555.

- Manssor, N. A. S., Radzi, Z., Yahya, N. A., Yusof, L. M., Hariri, F., Khairuddin, N. H., Kasim, N. H. A. & Czernuszka, J. T. 2016. Characteristics and Young's modulus of collagen fibrils from expanded skin using anisotropic controlled rate self-inflating tissue expander. *Skin Pharmacology and Physiology*, 29, 55-62.
- Marcus, J., Horan, D. B. & Robinson, J. K. 1990. Tissue expansion: past, present, and future. *Journal of the American Academy of Dermatology*, 23, 813-825.
- Martano, M., Corteggio, A., Restucci, B., De Biase, M. E., Borzacchiello, G. & Maiolino, P. 2016. Extracellular matrix remodeling in equine sarcoid: an immunohistochemical and molecular study. *BMC Veterinary Research*, 12, 24.
- Mason, G. J. & Mench, J. 1997. Using Behaviour to Assess Animal Welfare. Animal Welfare. Hughes: Animal Welfare.
- McCauley, R. L. Tissue expansion reconstruction of the scalp. Seminars in Plastic Surgery, 2005 Seventh Avenue, Newuork. Thieme Medical Publishers, 143-152.
- McGeachie, J. & Tennant, M. 1997. Growth factors and their implications for clinicians: A brief review. *Australian Dental Journal*, 42, 375-380.
- Motamed, S., Niazi, F., Atarian, S. & Motamed, A. 2008. Post-burn head and neck reconstruction using tissue expanders. *Burns*, 34, 878-884.
- Mustoe, T., Bartell, T. & Garner, W. 1989. Physical, biomechanical, histologic, and biochemical effects of rapid versus conventional tissue expansion. *Plastic and Reconstructive Surgery*, 83, 687-691.
- Neumann, C. G. 1957. The expansion of an area of skin by progressive distention of a subcutaneous balloon: use of the method for securing skin for subtotal reconstruction of the ear. *Plastic and Reconstructive Surgery*, 19, 124-130.
- Ninomiya, S., Sato, S., Kusunose, R., Mitumasu, T. & Obara, Y. 2007. A note on a behavioural indicator of satisfaction in stabled horses. *Applied Animal Behaviour Science*, 106, 184-189.
- Noorbala, A. A., Ramezanzadeh, F., Abedinia, N. & Naghizadeh, M. M. 2009. Psychiatric disorders among infertile and fertile women. *Social Psychiatry and Psychiatric Epidemiology*, 44, 587-591.

- Numata, Y., Terui, T., Okuyama, R., Hirasawa, N., Sugiura, Y., Miyoshi, I., Watanabe, T., Kuramasu, A., Tagami, H. & Ohtsu, H. 2006. The accelerating effect of histamine on the cutaneous wound-healing process through the action of basic fibroblast growth factor. *Journal of Investigative Dermatology*, 126 (6): 1403-1409.
- Obdeijn, M. C., Nicolai, J.-P. A. & Werker, P. M. 2009. The osmotic tissue expander: a three-year clinical experience. *Journal of Plastic, Reconstructive & Aesthetic Surgery,* 62, 1219-1222.
- Ottenio, M., Tran, D., Annaidh, A. N., Gilchrist, M. D. & Bruyère, K. 2015. Strain rate and anisotropy effects on the tensile failure characteristics of human skin. *Journal of the Mechanical Behavior of Biomedical Materials*, 41, 241-250.
- Ouhtit, A., Muller, H. K., Davis, D. W., Ullrich, S. E., McConkey, D. & Ananthaswamy, H. N. 2000. Temporal events in skin injury and the early adaptive responses in ultraviolet-irradiated mouse skin. *The American Journal of Pathology*, 156, 201-207.
- Owen, K., Singer, E., Clegg, P., Ireland, J. & Pinchbeck, G. 2012. Identification of risk factors for traumatic injury in the general horse population of north-west England, Midlands and north Wales. *Equine Veterinary Journal*, 44, 143-148.
- Pamplona, D. C., Velloso, R. Q. & Radwanski, H. N. 2014. On skin expansion. Journal of Mechanical Behavior of Biomedical Materials, 29, 655-662.
- Patel, P. A., Elhadi, H. M., Kitzmiller, W. J., Billmire, D. A. & Yakuboff, K. P. 2014. Tissue expander complications in the pediatric burn patient: a 10year follow-up. *Annals of Plastic Surgery*, 72, 150-154.
- Pedersen, G. R., Sondergaard, E. & Ladewig, J. 2004. The influence of bedding on the time horses spend recumbent. *Journal of Equine Veterinary Science*, 24, 153-158.
- Peppas, N. A. & Leobandung, W. 2004. Stimuli-sensitive hydrogels: ideal carriers for chronobiology and chronotherapy. *Journal of Biomaterials Science, Polymer Edition*, 15, 125-144.
- Piérard, G. E., Piérard, S., Delvenne, P. & Piérard-Franchimont, C. 2013. In vivo evaluation of the skin tensile strength by the suction method: pilot study coping with hysteresis and creep extension. *International Scholarly Research Notices*.
- Pisarski, G. P., Mertens, D., Warden, G. D. & Neale, H. W. 1998. Tissue expander complications in the pediatric burn patient. *Plastic and Reconstructive Surgery*, 102, 1008-1012.

- Price, J., Catriona, S., Welsh, E. M. & Waran, N. K. 2003. Preliminary evaluation of a behaviour-based system for assessment of post-operative pain in horses following arthroscopic surgery. *Veterinary Anaesthesia and Analgesia*, 30, 124-137.
- Pritchett LC, Ulibarri C, Roberts MC, Schneider RK. & Sellon DC. 2003. Identification of potential physiological and behavioral indicators of postoperative pain in horses after exploratory celiotomy for colic. *Applied Animal Behaviour Science*, 80:31–43
- Putti, V., 1921. The operative lengthening of the femur. *Clinical Orthopaedics* and *Related Research*[®], 250, pp.4-7.
- Radovan, C. Adjacent flap development using expandable silastic implant. Annual Meeting of the American Society of Plastic and Reconstructive Surgeons, Boston, Mass, 1976.
- Radovan, C. 1982. Breast reconstruction after mastectomy using the temporary expander. *Plastic and Reconstructive Surgery*, 69, 195-208.
- Radovan, C. 1984. Tissue expansion in soft-tissue reconstruction. *Plastic and Reconstructive Surgery*, 74, 482-490.
- Ransom, J. I. & Cade, B. S. 2009. *Quantifying equid behavior a research ethogram for free-roaming feral horses*, U.S. Geological Survey, Reston, Virginia University of Nebraska Lincoln
- Raposio, E., Cella, A., Panarese, P., Caregnato, P., Gualdi, A. & Santi, P. L. 2000. Quantitative benefits provided by acute tissue expansion: a biomechanical study in human cadavers. *British Journal of Plastic Surgery*, 53, 220-224.
- Refojo, M. 1975. Vapor pressure and swelling pressure of hydrogels. *Pascal* and *Francis Bibliographic Databases*, 16, 286-288.
- Remache, D., Caliez, M., Gratton, M. & Dos Santos, S. 2018. The effects of cyclic tensile and stress-relaxation tests on porcine skin. *Journal of the Mechanical Behavior of Biomedical Materials*, 77, 242-249.
- Ricciardelli, C., Horsfall, D.J., Skinner, J.M., Henderson, D.W., Marshall, V.R. and Tilley, W.D., 1989. Development and characterization of primary cultures of smooth muscle cells from the fibromuscular stroma of the guinea pig prostate. *In vitro cellular & developmental biology*, 25(11), pp.1016-1024
- Ronert, M. A., Hofheinz, H., Manassa, E., Asgarouladi, H. & Olbrisch, R. R. 2004. The beginning of a new era in tissue expansion: self-filling osmotic tissue expander four-year clinical experience. *Plastic and Reconstructive Surgery*, 114, 1025-1031.

- Ryu, H. S., Joo, Y. H., Kim, S. O., Park, K. C. & Youn, S. W. 2008. Influence of age and regional differences on skin elasticity as measured by the Cutometer®. Skin Research and Technology, 14, 354-358.
- Saaiq, M. 2013. Tissue expansion: a valuable adjunct to reconstructive surgery. Annals of Pakistan Institute of Medical Sciences, 9, 55-56.
- Salah, M., Usama, K. & Shehata, I. 2007. The evaluation of the tissue expansion technique in the last twenty years. *The Egyptian Journal of Plastic and Reconstructive Surgery*, 31, 227-235.
- Samuelson, D. 2007. Textbook of Veterinary Histology, Saunders Elsevier, St. Louis, missouri.
- Sasaki, G. 1985. Tissue Expansion: Guidelines and Case Analyses. *Dow Corning Wright Publications, USA*.
- Sasaki, G. H. 1984. Plastic surgery important advances in clinical medicine: refinements in tissue expansion. *Western Journal of Medicine*, 141, 234.
- Sasaki, G. H. & Pang, C. Y. 1984. Pathophysiology of skin flaps raised on expanded pig skin. *Plastic and Reconstructive Surgery*, 74, 59-67.
- Sasaki, K., Nozaki, M., Kikutchi, Y., Yamaki, T. & Soejima, K. 1999. Reconstruction of perianal skin defect using a V–Y advancement of bilateral gluteus maximus musculocutaneous flaps: reconstruction considering anal cleft and anal function. *British Journal of Plastic Surgery*, 52, 471-475.
- Scott, D. 1988. Large Animal Dermatology. . Philadelphia. WB Saunders
- Scott, D., Miller, W. & Griffin, C. 2001. Small Animal Dermatology, Philadelphia, WB Saunders
- Scott, D. W. & Miller, W. H. 2011. *Equine dermatology*, Philadelphia, Uruguay, Universidad de la República.
- Shan & Baker 1991. Fundamentals of expanded tissue. *Head & Neck*, 13, 327-333.
- Shergold, O. A., Fleck, N. A. & Radford, D. 2006. The uniaxial stress versus strain response of pig skin and silicone rubber at low and high strain rates. *International Journal of Impact Engineering*, 32, 1384-1402.
- Silver, F. H., Seehra, G. P., Freeman, J. W. & DeVore, D. 2002. Viscoelastic properties of young and old human dermis: a proposed molecular mechanism for elastic energy storage in collagen and elastin. *Journal of Applied Polymer Science*, 86, 1978-1985.

- Silver, F. H., Siperko, L. M. & Seehra, G. P. 2003. Mechanobiology of force transduction in dermal tissue. *Skin Research and Technology*, 9, 3-23.
- Solea, A., Bolwellb, C., Darta, A., Rileyb, C. & Theoretb, C. 2015. Descriptive survey of wounds in horses presented to Australian veterinarians. *Australian Equine Veterinarian*, 34, 68-74.
- Sørensen, M. A., Petersen, L. J., Bundgaard, L., Toft, N. & Jacobsen, S. 2014. Regional disturbances in blood flow and metabolism in equine limb wound healing with formation of exuberant granulation tissue. *Wound Repair and Regeneration*, 22, 647-653.
- Souza, M., van Weeren, P. R., Van Schie, H. & Van De Lest, C. 2010. Regional differences in biochemical, biomechanical and histomorphological characteristics of the equine suspensory ligament. *Equine Veterinary Journal*, 42, 611-620.
- Spodnick, G. J., Pavletic, M. M., Clark, G. N., Schelling, S. H. & Kraus, K. H. 1993. Controlled tissue expansion in the distal extremities of dogs. *Veterinary Surgery*, 22, 436-443.
- Sternberg, S. S. 1992. *Histology for pathologists, New York, Raven Press.*
- Sutton, G.A., Paltiel, O., Soffer, M. & Turner, D. 2013. Validation of two behaviour-based pain scales for horses with acute colic. *Vet. J.* 197, 646–650.
- Sutton, G.A., Paltiel, O., Soffer, M. & Turner, D. 2013. Validation of two behaviour-based pain scales for horses with acute colic. *Veterinary Journal.* 197, 646–650
- Swan, M., Bucknall, D., Goodacre, T. & Czernuszka, J. 2011. Synthesis and properties of a novel anisotropic self-inflating hydrogel tissue expander. *Acta Biomaterialia*, 7, 1126-1132.
- Swan, M. C., Bucknall, D. G., Czernuszka, J. T., Pigott, D. W. & Goodacre, T. E. 2012. Development of a novel anisotropic self-inflating tissue expander: in vivo submucoperiosteal performance in the porcine hard palate. *Plastic Reconstractive Surgery*, 129, 79-88.
- Szotek, S., Bedzinski, R., Kobielarz, M., Gasiorglogowska, M., Komorowska, M., Maksymowicz, K., Hanuza, J. & Hermanowicz, K. 2009. Human skin properties determined by mechanical tests and Raman spectroscopy. *Engineering of Biomaterials*, 89, 208-210.
- Szotek, S., Bedzinski, R., Kobielarz, M., Zywicka, B., Pielka, S. & Kuropka, P. 2008. Investigation of mechanical properties of the skin. *Engineering of Biomaterials*, 11, 81-84.

- Takei, T., Mills, I., Arai, K. & Sumpio, B. E. 1998. Molecular basis for tissue expansion: clinical implications for the surgeon. *Plastic and Reconstructive Surgery*, 102, 247-258.
- Talukdar, A., Calhoun, M. & Stinson, A. 1972. Microscopic anatomy of the skin of the horse. *American Journal of Veterinary Research*, 33, 2365-2390.
- Tepole, A. B., Gosain, A. K. & Kuhl, E. 2012. Stretching skin: The physiological limit and beyond. *International Journal of Non-Linear Mechanics*, 47, 938-949.
- Tercan, M., Çokkeser, Y., Ozyazgan, I., Bekerecioglu, M. & Sar, I. 2001. Facilitated tissue expansion with topical estriol. *Annals of Plastic Surgery*, 46, 617-620.
- Thamm, D., Ehrhart III, E., Charles, J. & Elce, Y. 2008. Cyclooxygenase-2 expression in equine tumors. *Veterinary Pathology*, 45, 825-828.
- Theoret, C., Bolwell, C. & Riley, C. 2016. A cross-sectional survey on wounds in horses in New Zealand. *New Zealand Veterinary Journal*, 64, 90-94.
- Theoret, C. & Schumacher, J. 2016. *Equine wound management*, John Wiley & Sons.
- Theoret, C. & Schumacher, J. 2017. *Equine wound management* [Online]. John Wiley & Sons. Available: <u>http://www.myilibrary.com?id=964112</u>.
- Theoret, C. L. 2004. Wound repair in the horse: Problems and proposed innovative solutions. *Clinical Techniques in Equine Practice*, 3, 134-140.
- Theoret, C. L., Barber, S. M., Moyana, T. N. & Gordon, J. R. 2001. Expression of transforming growth factor β1, β3, and basic fibroblast growth factor in full-thickness skin wounds of equine limbs and thorax. *Veterinary Surgery*, 30, 269-277.
- Thorpe, C., Stark, R., Goodship, A. & Birch, H. 2010. Mechanical properties of the equine superficial digital flexor tendon relate to specific collagen cross-link levels. *Equine Veterinary Journal*, 42, 538-543.
- Timmenga, E. J. & Das, P. K. 1992. Histomorphological observations on dermal repair in expanded rabbit skin: a preliminary report. *British Journal of Plastic Surgery*, 45, 503-507.
- Tonnesen, M. G., Feng, X. & Clark, R. A. Angiogenesis in wound healing. Journal of Investigative Dermatology Symposium Proceedings, 2000. Elsevier, 40-46.

- Topczewska, J. M., Ledwon, J. K., Vaca, E. E. & Gosain, A. K. 2019. Mechanical stretching stimulates growth of the basal layer and rete ridges in the epidermis. *Journal of Tissue Engineering and Regenerative Medicine*, 13, 2121-2125.
- Torcivia, C. and McDonnell, S., 2021. Equine discomfort ethogram. *Animals*, 11(2), p.580. https://doi.org/10.3390/ani11020580.
- Trindade, P.H.E., Taffarel, M.O. & Luna, S.P.L. 2021. Spontaneous Behaviors of Post-Orchiectomy Pain in Horses Regardless of the Effects of Time of Day, Anesthesia, and Analgesia. *Animals*, 11, 1629.
- Tsai, F.-C. 2003. A new method: perforator-based tissue expansion for a preexpanded free cutaneous perforator flap. *Burns*, 29, 845-848.
- Uijlenbroek, H. J., Liu, Y., He, J. F., Visscher, C., van Waas, M. A. & Wismeyer, D. 2011. Expanding soft tissue with Osmed tissue expanders in the goat maxilla. *Clin Oral Implants Res*, 22, 121-128.
- Van Damme, P. A., Freihofer, H. P. M., Maltha, J., Kuijpers-Jagtman, A. M. & Van't Hof, M. A. 1997. Three-dimensional morphometric analysis of the effects of subperiosteal palatal soft-tissue expansion in growing cats. *International journal of oral and maxillofacial surgery*, 26, 61-67.
- Van Damme, P. A., Heidbüchel, K. L., Kuijpers-Jagtman, A.-M., Maltha, J. C. & Freihofer, H. P. M. 1992. Cranio-maxillo-facial tissue expansion, experimentally based or clinically empiric? A review of the literature. *Journal of Cranio-Maxillofacial Surgery*, 20, 61-69.
- Van Rappard, J. 1996. *Histologic changes. In: Nordström REA (ed) Tissue expansion,* Boston, Butterworth-Heinemann, .
- Van Rappard, J. H. 1988. Controlled tissue-expansion in reconstructive surgery. (DIV), University of Groningen.
- Van Rappard, J. H., Jerusalem, C., Sonneveld, G. & Borghouts, J. 1988. Histologic changes in soft tissues due to tissue expansion (in animal studies and humans). *Facial Plastic Surgery*, 5, 280-286.
- Velnar, T., Bailey, T. & Smrkolj, V. 2009. The wound healing process: an overview of the cellular and molecular mechanisms. *Journal of International Medical Research*, 37, 1528-1542.
- Wagh, M. S. & Dixit, V. 2013. Tissue expansion: Concepts, techniques and unfavourable results. *Indian Journal of Plastic Surgery: Official Publication of The Association of Plastic Surgeons of India,* 46, 333.

- Wakuri, H., Mutoh, K., Ichikawa, H. & Liu, B. 1995. Microscopic anatomy of the equine skin with special reference to the dermis. *Okajimas Folia Anatomica Japonica*, 72, 177-183.
- Wang, C., Zhang, J., Yang, S., Hyakusoku, H., Song, P. & Pu, L. L. 2016. The clinical application of preexpanded and prefabricated super-thin skin perforator flap for reconstruction of post-burn neck contracture. *Annals of Plastic Surgery*, 77, S49-S52.
- Werhahn, H., Hessel, E. F., Bachhausen, I. & Van den Weghe, H. F. 2010. Effects of different bedding materials on the behavior of horses housed in single stalls. *Journal of Equine Veterinary Science*, 30, 425-431.
- Whittaker, C. J., Reynolds, B. D., McCarthy, P. M., Taylor, S. F., Major, D., Caruso, K. A. & Smith, J. 2020. Use of a chronic soft tissue expansion device to facilitate blepharoplasty in a horse with lower-lid cicatricial ectropion with a 14-year follow-up. *Veterinary Ophthalmology*, 5, 899-904.
- Wichterle, O. & Lim, D. 1960. Hydrophilic gels for biological use. *Nature*, 185, 117-118.
- Wiese, K. 1993. Osmotically induced tissue expansion with hydrogels: a new dimension in tissue expansion? A preliminary report. *Journal of Cranio-Maxillofacial Surgery*, 21, 309-313.
- Wiese, K., Heinemann, D., Ostermeier, D. & Peters, J. 2001. Biomaterial properties and biocompatibility in cell culture of a novel self-inflating hydrogel tissue expander. *Journal of Biomedical Materials Research: An Official Journal of The Society for Biomaterials and The Japanese Society for Biomaterials*, 54, 179-188.
- Wilhelmi, B. J., Blackwell, S. J., Mancoll, J. S. & Phillips, L. G. 1998. Creep vs. Stretch: A review of the viscoelastic properties of skin. *Annals of Plastic Surgery*, 41, 215-219.
- Wilkinson, H. N. & Hardman, M. J. 2020. Wound healing: Cellular mechanisms and pathological outcomes. *Open biology*, 10, 200-223.
- Wilmink, J., Van den Boom, R., Van Weeren, P. & Barneveld, A. 2006. The modified Meek technique as a novel method for skin grafting in horses: evaluation of acceptance, wound contraction and closure in chronic wounds. *Equine Veterinary Vournal*, 38, 324-329.
- Wilmink, J. M., Stolk, P. T., Van Weeren, P. & Barneveld, A. 1999a. Differences in second-intention wound healing between horses and ponies: macroscopic aspects. *Equine Veterinary Journal*, 31, 53-60.

- Wilmink, J. M., Van Weeren, P., Stolk, P. T., Van Mil, F. & Barneveld, A. 1999b. Differences in second-intention wound healing between horses and ponies: histological aspects. *Equine Veterinary Journal*, 31, 61-67.
- Wittkampf, A. R. 1989. Short-term experience with the subperiosteal tissue expander in reconstruction of the mandibular alveolar ridge. *Journal of Oral and Maxillofacial Surgery*, 47, 469-474.
- Wobeser, B. K. 2015. Skin diseases in horses. Veterinary Clinics: Equine Practice, 31, 359-376.
- Wong, D., Buechner-Maxwell, V. & Manning, T. 2005. Equine skin: structure, immunologic function, and methods of diagnosing disease. *Compendium*, 463-473.
- Yarnell, K., Hall, C. & Billett, E. 2013. An assessment of the aversive nature of an animal management procedure (clipping) using behavioral and physiological measures. *Physiology and Behavior*, 118, 32-39.
- Yeşilada, A. K., Akçal, A., Dağdelen, D., Sucu, D. Ö., Kılınç, L. & Tatlıdede, H. S. 2013. The feasibility of tissue expansion in reconstruction of congenital and aquired deformities of pediatric patients. *International Journal of Burns and Trauma*, 3, 144.
- Yoo, J. M., Ben Amara, H., Kim, M. K., Song, J. D. & Koo, K.-T. 2018. Oral tissue response to soft tissue expanders prior to bone augmentation: in vitro analysis and histological study in dogs. *Journal of Periodontal and Implant Science*, 48, 152-163.
- Zak, M., Kuropka, P., Kobielarz, M., Dudek, A., Kaleta-Kuratewicz, K. & Szotek, S. 2011. Determination of the mechanical properties of the skin of pig foetuses with respect to its structure. *Acta of Bioengineering and Biomechanics*, 13, 37-43.
- Zeng, Y. j., Liu, Y. h., Xu, C. q., Xu, X. h., Xu, H. & Sun, G. c. 2004. Biomechanical properties of skin in vitro for different expansion methods. *Clinical Biomechanics*, 19, 853-857.
- Zhang, E.-p., Liao, D.-h., Liu, A.-z., Wang, X.-b., Li, X.-y., Zeng, Y.-j. & Wang, S.-j. 2006. Biomechanical characteristics investigation on long-term free graft with expanded porcine skin. *Clinical Biomechanics*, 21, 864-869.
- Zöllner, A. M., Tepole, A. B., Gosain, A. K. & Kuhl, E. 2012a. Growing skin: tissue expansion in pediatric forehead reconstruction. *Biomechanics & Modeling in Mechanobiology*, 11, 855-867.
- Zöllner, A. M., Tepole, A. B. & Kuhl, E. 2012b. On the biomechanics and mechanobiology of growing skin. *Journal of Theoretical Biology*, 297, 166-175.