



**UNIVERSITI PUTRA MALAYSIA**

***Piper betle L. AND Persicaria odorata (Lour.) Soják AS ALTERNATIVE  
TO ANTIMICROBIAL GROWTH PROMOTERS IN BROILER CHICKENS***

**ABDUL BASIT MUHAMMAD**

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By

**ABDUL BASIT MUHAMMAD**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
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Philosophy**

**November 2021**

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## DEDICATION

This work is dedicated to the souls of my late father and mother (May Allah rest their soul in the peace and grant them higher ranks in Jannah) My beloved wife, son, and daughter for being my inspiration, their unconditional support made it easier for me to complete the tedious journey of PhD My brother and sisters for their constant encouragement and support My friends and colleagues.

Abdul Basit Muhammad  
November 2021



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

***Piper betle* L. AND *Persicaria odorata* (Lour.) Soják AS ALTERNATIVE TO ANTIMICROBIAL GROWTH PROMOTERS IN BROILER CHICKENS**

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**ABDUL BASIT MUHAMMAD**

**November 2021**

**Chairperson : Assoc. Prof. Arifah binti Abdul Kadir, PhD**  
**Faculty : Veterinary Medicine**

In several countries, except Europe, dietary supplementation of sub-therapeutic antimicrobial growth promoters (AGPs) is commonly practised for sustainable broiler chicken production. However, their indiscriminate use resulted in antibiotic resistance, which leads to a ban against AGPs. Hence, it is essential to find potential and safe alternatives that can improve broiler chickens' production performance. Recently, phytobiotics, especially herbs, have gained attention and have been extensively studied for their possible use as an alternative poultry feed additive. Generally, phytobiotics are assumed to be natural, safe, and residue-free substances; however, scarce literature is available about their optimal dosage and safe use. The present study was carried out to assess the phytochemical screening, quantification of secondary metabolites, acute oral toxicity, appropriate dosage and potential of selected herbs *Piper betle* and *Persicaria odorata* as alternative growth promoters to AGPs in broiler chickens.

*P. betle* and *P. odorata* leaf extracts were evaluated for qualitative and quantitative phytochemical screening, antioxidant and antimicrobial potential. The preliminary phytochemical analysis of *P. betle* and *P. odorata* leaf extracts revealed the presence of flavonoids, phenols, saponins, tannins, glycosides, and volatile oils. The higher total phenolic content and total tannins were quantified from *P. betle* methanolic leaf extract. Additionally, it showed increased antioxidant activity compared to *P. odorata* leaf extracts. The *in vitro* antibacterial potential of both herbs was estimated against *Bacillus subtilis*, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella enterica*, *Pseudomonas aeruginosa*, *Candida albicans*, and *Aspergillus brasiliensis*. The methanolic leaf extract of *Piper betle* showed antibacterial and antifungal activity against selected strains. In conclusion, phytochemical screening of *P. betle* and *P. odorata* leaf extracts showed rich diversity of phytochemicals, antioxidant, and antimicrobial activity.

In the first phase, quantitative phytochemical analyses of methanolic extract of *P. betle* and *P. odorata* was carried out using chromatography. Furthermore, the limit dose acute toxicity of the selected herbs was estimated in broiler chickens according to the guidelines of OECD, 2016. A total of 35 broiler chicks were used in this study. On day 21 of age, the chicks were randomly allocated into 7 treatment groups. The birds in extract-treated groups were orally gavaged with a single dose of methanolic leaf extract of *P. betle* and *P. odorata* at the rate of 500mg/kg, 1000 mg/kg, and 2,000 mg/kg, respectively. At the same time, the control group received 0.5% carboxymethyl cellulose (CMC) as a placebo. The gas chromatography-mass spectrometry (GC-MS) fingerprints showed the presence of various metabolites. Furthermore, the secondary bioactive compounds eugenol and quercetin were successfully quantified from *P. betle* and *P. odorata*, respectively. Acute toxicity study has shown no behavioural or physical changes in the chickens of treated groups. There were no significant differences in growth performance, haematological indices, serum biochemistry, and relative internal organs weights between the control and extract treated groups. The gross and microscopic observations indicated no changes in the liver, heart, lungs, spleen, gizzard, and kidneys of the treated birds. In conclusion, secondary bioactive compounds eugenol and quercetin were successfully quantified from methanolic leaf extract of *P. betle* and *P. odorata*. Based on current study results, the LD<sub>50</sub> value for the tested extracts would be > 2000 mg/kg body weight.

The second phase was conducted to estimate the influence of different dose supplementation of *Piper betle* leaf meal (PBLM) and *Persicaria odorata* leaf meal (POLM) on growth performance, ileal digestibility, gut morphology, haematological indices, serum biochemical attributes, histomorphology of the liver, and internal organs parameters in broiler chickens. A total of 210 day-old broiler chicks were fed either basal diet (BD) or BD + 2, 4, and 8 g/kg of PBLM and POLM. Except for PBLM 8g/kg, graded dose inclusion of PBLM and POLM increased ( $p < 0.05$ ), the body weight gain (BWG) positively modulated the gut architecture and enhanced nutrient digestibility in broiler chickens. Birds fed on PBLM 4g/kg and POLM 8g/kg had significantly higher ( $p < 0.05$ ) BWG with superior ( $p < 0.05$ ) feed efficiency. Haematological indicators like red blood cells and haemoglobin were positively improved in birds fed on diet PBLM 4g/kg and POLM 8g/kg compared to the control group. Additionally, dietary supplementation of PBLM and POLM decreased the activity of AST and ALT with reduced serum concentration of triglyceride and cholesterol, which were observed lowest in PBLM 4g/kg and POLM 8g/kg groups. On the other hand, the serum concentrations of total protein (TP), albumin, and globulin were recorded as increased ( $p < 0.05$ ) in broilers raised on the diets PBLM 4g/kg and POLM 8g/kg. Notably, dietary supplementation of PBLM and POLM did not influence the organ parameters and showed no adverse effects on liver histomorphology. In conclusion, dietary supplementations of phytobiotics (PBLM 4 g/kg and POLM 8 g/kg) positively modulated the intestinal microarchitecture with enhanced nutrient digestibility. Hence, it resulted in maximum body weight gain with improved haematological indicators and serum biochemistry attributes. Furthermore, dietary supplementations of phytobiotics have no deleterious effects

on the internal organs and retained relatively normal hepatic parenchyma, thus enhanced the optimum growth performance of the broiler chickens.

The third phase experiment was designed to estimate the comparative efficacy of PBLM and POLM with AGPs on growth performance, gut morphology, ileal digestibility, caecal microbiota composition, tibial bone morphometry, and meat quality in broilers chickens. Based on the earlier trial results, PBLM 4g/kg and POLM 8g/kg were selected for the comparative efficacy study. A total of 150 day-old broiler chickens were fed basal diet (BD), which served as the negative control (NC) or BD + 0.2g/kg tetracycline as positive control (PC); BD + 0.03g/kg halquinol (HAL), BD + 8g/kg POLM (Po8), and BD + 4g/kg PBLM (Pb4) as treatment groups. Dietary supplementation of phytobiotics Po8 and Pb4 showed improved growth performance, positively modulated the gut architecture, and improved digestibility comparable to AGPs. Additionally, dietary supplementation of phytobiotics (Po8 and Pb4) significantly decreased the *E. coli*, *Salmonella*, and *Staphylococcus aureus* compared to the NC group. However, supplementation of Pb4 resulted in significantly decreased total anaerobic bacteria and Clostridium counts compared to the NC group. Importantly, dietary supplementation of phytobiotics significantly increased the *Lactobacillus* count compared to HAL, PC, and the NC groups. The supplementation of phytobiotics improved meat quality compared to the NC group. Moreover, significantly ( $p < 0.05$ ) higher values of geometric parameters of tibial bones with increased strength were recorded in chickens fed on the phytobiotics, compared to the NC group. Conclusively, phytobiotics showed performance comparable to the selected AGPs. The present study results provided evidence that the dietary supplementation of PBLM 4 g/kg and POLM 8 g/kg are the appropriate and safe doses. PBLM and POLM could be the potential alternatives to AGPs for sustainable broiler chicken production.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

***Piper betle* L. DAN *Persicaria odorata* (Lour.) Soják SEBAGAI ALTERNATIF TERHADAP PENGGALAK PERTUMBUHAN ANTIMIKROB PADA AYAM PEDAGING**

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Di beberapa negara, kecuali Eropah, suplemen makanan sub-terapi penggalak pertumbuhan antimikroba (AGP) biasanya dilakukan untuk pengeluaran ayam pedaging yang lestari. Walau bagaimanapun, penggunaan sembarangan menyebabkan rintangan terhadap antibiotik yang menyebabkan larangan terhadap penggunaan AGP. Oleh itu, adalah mustahak untuk mencari alternatif yang berpotensi dan selamat yang dapat meningkatkan prestasi pengeluaran ayam pedaging. Baru-baru ini, fitobiotik, terutama herba, mendapat perhatian dan telah banyak dikaji untuk kemungkinan penggunaannya sebagai bahan tambahan makanan ternakan. Secara amnya, fitobiotik dianggap sebagai bahan semula jadi, selamat, dan bebas residu; namun, terdapat sedikit maklumat mengenai dos yang optimum dan penggunaan yang selamat. Kajian ini dijalankan untuk menaksir saringan fitokimia, pengkuantitian metabolit sekunder, ketoksikan oral akut, dos dan potensi yang sesuai dari herba terpilih *Piper betle* dan *Persicaria odorata* sebagai penggalak pertumbuhan alternatif terhadap penggalak pertumbuhan antimikrob pada ayam pedaging.

Pada mulanya, ekstrak daun *P. betle* dan *P. odorata* dinilai untuk saringan fitokimia kualitatif dan kuantitatif, potensi antioksidan dan antimikrob. Analisis fitokimia awal ekstrak daun *P. betle* dan *P. odorata* mendedahkan adanya flavonoid, fenol, saponin, tanin, glikosida, dan minyak mudah meruap. Kandungan fenolik total dan jumlah tanin yang lebih tinggi dikuantifikasi dari ekstrak metanol daun *P. betle*. Selain itu, ia menunjukkan peningkatan aktiviti antioksidan berbanding dengan ekstrak daun *P. odorata*. Potensi *in vitro* antibakteria kedua-dua ramuan tersebut dianggarkan terhadap *Bacillus subtilis*, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella enterica*, *Pseudomonas aeruginosa*, *Candida albicans*, dan *Aspergillus brasiliensis*. Ekstrak metanol dari daun *P. betle* menunjukkan aktiviti antibakteria



dan antikulat terhadap jenis-jenis terpilih ini. Kesimpulannya, saringan fitokimia ekstrak daun *P. betle* dan *P. odorata* menunjukkan fitokimia kepelbagaian beraneka, aktiviti antioksidan dan antimikrob.

Pada fasa pertama, analisis kuantitatif fitokimia ekstrak metanol *P. betle* dan *P. odorata* dilakukan dengan menggunakan kromatografi. Selanjutnya, ketoksikan akut dos had ramuan herba terpilih dianggarkan pada ayam pedaging mengikut garis panduan OECD, 2016. Sebanyak 35 ekor ayam pedaging digunakan dalam kajian ini. Pada hari ke-21, anak-anak ayam tersebut diagihkan secara rawak ke dalam 7 kumpulan rawatan. Anak ayam dalam kumpulan yang dirawat ekstrak diberi oral dengan satu dos ekstrak metanol daun *P. betle* dan *P. odorata* pada kadar masing-masing 500mg / kg, 1000 mg / kg, dan 2000 mg / kg. Pada masa yang sama, kumpulan kawalan menerima 0.5% karboksimetil selulosa (CMC) sebagai plasebo. Gambar kromatografi-spektrometri massa (GC-MS) menunjukkan adanya pelbagai metabolit. Selanjutnya, sebatian bioaktif sekunder eugenol dan quercetin berjaya diukur masing-masing dari *P. betle* dan *P. odorata*. Kajian ketoksikan akut tidak menunjukkan perubahan tingkah laku atau fizikal pada kumpulan ayam yang dirawat. Tidak ada perbezaan yang signifikan dalam prestasi pertumbuhan, indeks hematologi, biokimia serum, dan berat organ dalaman relatif antara kumpulan kawalan dan ekstrak yang dirawat. Pemerhatian kasar dan mikroskopik menunjukkan tidak ada perubahan pada hati, jantung, paru-paru, limpa, hempedal, dan ginjal ayam yang dirawat. Kesimpulannya, sebatian bioaktif sekunder eugenol dan quercetin berjaya diukur dari ekstrak daun metanol *P. betle* dan *P. odorata*. Berdasarkan kepada keputusan kajian terkini, nilai LD50 untuk ekstrak *P. betle* dan *P. odorata* teruji ialah > 2000 mg / kg berat badan.

Fasa kedua dijalankan untuk menganggar pengaruh suplemen dos berbeza hidangan daun *P. betle* (PBLM) dan hidangan daun *P. odorata* (POLM) terhadap prestasi pertumbuhan, pencernaan ileal, morfologi usus, indeks hematologi, atribut biokimia serum, histomorfologi hati, dan parameter organ dalaman pada ayam pedaging. Sebanyak 210 ekor anak ayam pedaging diberi makan sama ada diet asas (BD) atau BD + 2, 4, dan 8 g / kg PBLM dan POLM. Namun untuk PBLM 8g / kg, kemasukan dos berperingkat PBLM dan POLM meningkat ( $p < 0.05$ ), kenaikan berat badan (BWG), memodulasi positif arkitektur usus dan meningkatkan pencernaan nutrien pada ayam pedaging. Ayam yang diberi makan PBLM 4g / kg dan POLM 8g / kg secara signifikan lebih tinggi ( $p < 0.05$ ) BWG dengan kecekapan makanan yang unggul ( $p < 0.05$ ). Petunjuk hematologi seperti sel darah merah dan hemoglobin meningkat secara positif pada ayam yang diberi makan PBLM 4g / kg dan POLM 8g / kg berbanding dengan kumpulan kawalan. Selain itu, suplemen makanan PBLM dan POLM menurunkan aktiviti AST dan ALT dengan penurunan kepekatan serum trigliserida dan kolesterol, yang diperhatikan paling rendah dalam kumpulan PBLM 4g / kg dan POLM 8g / kg. Sebaliknya, kepekatan serum protein total (TP), albumin, dan globulin dicatatkan meningkat ( $p < 0.05$ ) pada ayam pedaging yang diberi makan pada diet PBLM 4g / kg dan POLM 8g / kg. Terutamanya, suplemen makanan PBLM dan POLM tidak mempengaruhi parameter organ dan tidak menunjukkan kesan buruk pada histomorfologi hati. Kesimpulannya, makanan tambahan fitobiotik (PBLM 4 g / kg dan POLM 8 g / kg) secara positif memodulasi mikroarkitektur usus

dengan peningkatan pencernaan nutrisi. Oleh itu, ini menghasilkan kenaikan berat badan maksimum dengan penunjuk hematologi yang lebih baik dan atribut biokimia serum. Tambahan pula, makanan tambahan fitobiotik tidak mempunyai kesan buruk pada organ dalaman dan mengekalkan parenkima hepatic yang agak normal, sehingga meningkatkan prestasi pertumbuhan ayam yang optimum.

Eksperimen fasa ketiga dirancang untuk menganggar keberkesanan perbandingan PBLM dan POLM dengan AGP pada prestasi pertumbuhan, morfologi usus, pencernaan ileal, komposisi mikrobiota sekal, morfometri tulang tibial, dan kualiti daging pada ayam pedaging. Berdasarkan hasil percubaan sebelumnya, PBLM 4g / kg dan POLM 8g / kg dipilih untuk kajian perbandingan keberkesanan. Sebanyak 150 ekor ayam pedaging diberi makanan asas (BD), yang berfungsi sebagai kawalan negatif (NC) atau BD + 0.2g / kg tetrasiklin sebagai kawalan positif (PC); BD + 0.03g / kg halquinol (HAL), BD + 8g / kg POLM (Po8), dan BD + 4g / kg PBLM (Pb4) sebagai kumpulan rawatan. Suplemen diet fitobiotik Po8 dan Pb4 menunjukkan peningkatan prestasi pertumbuhan, modulasi positif arkitektur usus, dan peningkatan pencernaan yang setanding dengan AGP. Selain itu, suplemen diet fitobiotik (Po8 dan Pb4) menurunkan *E. coli*, *Salmonella*, dan *Staphylococcus aureus* secara signifikan berbanding kumpulan NC. Walau bagaimanapun, suplementasi Pb4 mengakibatkan penurunan jumlah bakteria anaerob secara signifikan, dan jumlah *Clostridium* dibandingkan dengan kumpulan NC. Yang penting, suplemen diet fitobiotik meningkatkan jumlah *Lactobacillus* secara signifikan berbanding kumpulan HAL, PC, dan NC. Tambahan fitobiotik meningkatkan kualiti daging berbanding dengan kumpulan NC. Selain itu, nilai parameter geometri tulang tibial yang lebih tinggi ( $p < 0.05$ ) lebih tinggi dicatat pada ayam yang diberi makan fitobiotik, berbanding dengan kumpulan NC.

Kesimpulannya, fitobiotik menunjukkan prestasi yang setanding dengan AGP yang dipilih. Hasil kajian ini memberikan bukti bahawa suplemen makanan PBLM 4 g / kg dan POLM 8 g / kg adalah dos yang sesuai dan selamat. PBLM dan POLM boleh menjadi alternatif berpotensi untuk AGP untuk pengeluaran ayam pedaging yang lestari.

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This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

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## LIST OF ABBREVIATIONS

AGPs	Antimicrobial Growth Promoters
PAFs	Phytogetic Feed additives
PBLM	<i>Piper betle</i> leaf meal
POLM	<i>Persicaria odorata</i> leaf meal
TPC	Total Phenolic Content
TFC	Total Flavonoids Content
TTC	Total Tannins Content
HAL	Halquinol
DMSO	Dimethyl Sulfoxide
GC-MS	Gas Chromatography–Mass Spectrometry
HPLC	High-Performance <i>Liquid Chromatography</i>
MIC	Minimum Inhibitory Concentration
MBC	Minimum Bactericidal Concentration
MFC	Minimum Fungicidal Concentration
CMC	Carboxymethyl Cellulose
LD <sub>50</sub>	Lethal Dose Fifty
ICP-MS	Inductively Coupled Plasma Mass Spectrometer
LOD	Limit of Detection
OECD	Organization of Economic Cooperation and Development
BWG	Body Weight Gain
FI	Feed Intake
FCR	Feed Conversion Ration

ALP	Alkaline Phosphatase
ALT	Alanine Aminotransferase
AST	Aspartate Aminotransferase
CFU	Colony Forming Unit
AID	Apparent Ileal Digestibility
TiO <sub>2</sub>	Titanium Dioxide
CP	Crude Protein
DM	Dry Matter
EU	European Union
NRC	National Research Council
Mg	Milligram
mmol/L	Milimol Per Liter
g/kg	Gram Per Kilogram
µm	Micrometer
µL	Microliter
a*	Redness
b*	Yellowness
L*	Lightness
TP	Total protein
%	Percentage
°C	Degree Celsius
SAS	Statistical Analysis System
SEM	Standard Error of Mean

# CHAPTER 1

## INTRODUCTION

### 1.1 Introduction

The rapidly increasing human population resulted in an increased demand for animal-source protein. Broiler chickens` meat offers a considerable and affordable potential to fulfil the human needs of animal source dietary protein (Skarp *et al.*, 2016; Mund *et al.*, 2017; Forte *et al.*, 2018). Intensive broiler chicken farming has been carried out to fulfil the continuously increasing demand for poultry meat. In several countries except for Europe, in-feed supplementation of sub-therapeutic doses of antimicrobial growth promoters (AGPs) has been extensively practised to improve feed efficiency and gut health; hence, to achieve the enhanced growth performance of broiler chickens (Abudabos *et al.*, 2016; Alhidary *et al.*, 2017; Mehdi *et al.*, 2018; Proctor A and Phillips GJ, 2019).

However, this indiscriminate use of AGPs in broilers` diets resulted in the emergence of antimicrobial-resistant microorganisms that may be transferring the resistant genes to the microorganisms pertinent to human health (Boovaragamoorthy *et al.*, 2019; Ma *et al.*, 2020).

Antimicrobial resistance is one of the major global concerns (Hedman *et al.*, 2020). Therefore, European Union (EU) and many other countries have banned the in-feed supplementation of AGPs in food-producing animals ((EC) No 1831/2003; Van Boeckel *et al.*, 2015; Tehseen *et al.*, 2016; Tang *et al.*, 2019).

On the other hand, the removal of in-feed AGPs from the poultry diet has exposed the birds to pathogenic organisms and significantly increased the prevalence of infectious diseases (Kumar *et al.*, 2018). Thus, the removal of AGPs from the poultry diet has a negative effect on the production performance of broiler chickens.

In this scenario, it is vital to find potential alternatives that can replace AGPs as feed additives in food-producing animals (Lilleho *et al.*, 2018). The ban against in-feed inclusion of AGPs has increased momentum to find potential alternatives (Al-Abd *et al.*, 2015; Attia *et al.*, 2018). Thus, extensive research has been carried out to find alternatives with similar antimicrobial and growth-promoting potential in broiler chickens (Yang *et al.*, 2015; Rashid *et al.*, 2020). Numerous alternative feed additives like Prebiotics, probiotics, postbiotics, enzymes, organic acid, and phytogenic feed additives (PFAs) or phytobiotics have been studied as a growth

promoter in broiler chickens. However, natural substances like herbs, plants, plant extracts, and their essential oils have gotten attention as potential feed additives in poultry production (Gadde *et al.*, 2017; Özbudak, 2019). Earlier studies have suggested that phytobiotics might be the potent and viable natural alternative to AGPs (Murugesan *et al.*, 2015; Yang *et al.*, 2015; Wati *et al.*, 2015; Díaz Carrasco *et al.*, 2018).

Among phytobiotics, herbs are of particular significance because of their secondary bioactive metabolites, such as polyphenols, flavonoids, and phenolic compounds. The herbs and their secondary bioactive compounds are potent antioxidants and antimicrobials that can prevent oxidative stress and reduce the risk of chronic diseases (Pietrzyk *et al.*, 2017; Abbas *et al.*, 2017). Additionally, they are anti-inflammatory, immunomodulatory (Grashorn *et al.*, 2010; Muthusamy *et al.*, 2015), detoxifying, and digestion-stimulating substances (Ganguly *et al.*, 2017). Previous studies showed that herbs positively affect the performance and biological health of broiler chickens (Abudabos *et al.*, 2016; Raza *et al.*, 2016). Moreover, herbs can improve haematological indicators and serum biochemical attributes (Gilani *et al.*, 2018; Odetola *et al.*, 2019; Basit *et al.*, 2020b) and have also been reported to regulate kidney and liver functions (Lu *et al.*, 2016; Klaric *et al.*, 2018). Several other beneficial effects of phytobiotics have also been reported in broiler chicken production, including improvement in gut health (Ahsan *et al.*, 2018; Hussein *et al.*, 2020; Basit *et al.*, 2020a) and positive modulation of gut microbiota (Yadav *et al.*, 2019; Basit *et al.*, 2020c). Furthermore, phytobiotics can enhance feed efficiency (Suresh *et al.*, 2018; Tang *et al.*, 2019), thus increasing the growth performance in broilers (Paraskeuas *et al.*, 2017; Mashayekhi *et al.*, 2018; Rahman *et al.*, 2018; Saleh *et al.*, 2018). Additionally, numerous studies have indicated the antibacterial efficacy of phytobiotics against pathogenic bacteria, Like *Clostridium*, *Salmonella*, *Staphylococcus aureus*, and *E. coli* in broiler chickens (Mohiti-Asli *et al.*, 2017; Abudabos *et al.*, 2018; Basit *et al.*, 2020c). On the other hand, one of the primary modes of action of phytobiotics is the balancing of beneficial intestinal microbiota by limiting the pathogenic microorganism (Stevanović *et al.*, 2018; Vase-Khavari *et al.*, 2019).

Natural products like herbs have numerous beneficial and growth-promoting effects in animals and are generally considered safe; however, they may contain some toxic substances (Guldiken *et al.*, 2018; Van *et al.*, 2020). Thus, there are some concerns against the safe use of herbs (Giannenas *et al.*, 2020). The toxicity evaluation of plant-based feed additives can reveal the possible detrimental effects; thus, helping to establish evidence for their safe use (Srinivasa *et al.*, 2018; Williamson *et al.*, 2020). Nevertheless, the data are scarce about the possible detrimental effects of in-feed inclusion of phytobiotics (Karaskova *et al.*, 2015). Hence, it is essential and within scope to investigate the full potential and toxicity evaluation of herbal feed additives, to establish their safe use (Salehi, B *et al.*, 2018; Sharifi-rad *et al.*, 2018).

Among various valuable herbs, one is *Piper betle*. It is an essential herb of the *Piperaceae* family and is widely grown in Southeast Asia and East Africa (Umar *et al.*, 2018). The *P. betle* is a native herb of the central and eastern part of peninsular Malaysia (Pin *et al.*, 2011; Periyannayagam *et al.*, 2012). *Piper betle* is an evergreen climbing shrub producing woody stems 5 - 20 metres long. It is highly abundant and inexpensive, therefore promoting further research and industrialization development, including in the food and pharmaceutical industries. However, its annual yields are estimated at 6 - 10 tonnes per hectare (Pradhan *et al.*, 2013; Nayaka *et al.*, 2021). In Malaysia and Indonesia, it is commonly named as "*daun sirih*." The *P. betle* possesses nutritive and therapeutic properties; thus, widely used in traditional medicinal systems (Faroqui *et al.*, 2016; Biswajit Patra *et al.*, 2016; Taukoorah *et al.*, 2016). This *P. betle* have several biological properties, including antimicrobial (Foo *et al.*, 2015; Chouhan *et al.*, 2017; Singh *et al.*, 2019), antifungal (Basak & Guha, 2017), anti-inflammatory (Biswajit Patra *et al.*, 2016), and antioxidant (Rintu *et al.*, 2015; Kamath & Sabeena, 2018). The reported bioactive compounds of *P. betle* are hydroxychavicol, eugenol, methyl eugenol, and some sterols (Muruganandam *et al.*, 2017). The phenolic compounds of *P. betle* like hydroxychavicol, eugenol, isoeugenol possess potent antioxidant potential (Ali *et al.*, 2018). Previous studies reported the antimicrobial and radical scavenging activities of bioactive compounds (eugenol and isoeugenol) of *P. betle* (Syahidah *et al.*, 2017; Zhang *et al.*, 2017).

Another herb, *Persicaria odorata*/*Polygonum minus* Huds of family *Polygonaceae*, has been widely studied for its therapeutic use. *Persicaria odorata* is perennial herb up to 0.35 m tall. The foliage retention of *P. odorata* is evergreen. is a culinary herb which is indigenous to Tropical south Asia (Sim *et al.*, 2019). This herb has several common names like Vietnamese cilantro and Vietnamese mint. In Malaysia, Indonesia, Singapore, and Brunei, it is called "*daun laksa*" or "*daun kesum*" (Vikram *et al.*, 2014; Dash & Zakaria 2016). Previous literature has shown that *P. odorata* is a strong antioxidant (Christapher *et al.*, 2016; Abdullah *et al.*, 2017) and antimicrobial (Saad *et al.*, 2014; Abubakar *et al.*, 2015). The secondary bioactive compounds like quercetin, myricetin, and gallic acid are important flavonoids of *P. odorata* (Imelda *et al.*, 2014; Christapher *et al.*, 2017; Pawłowska *et al.*, 2020). These compounds are assumed to be responsible for the antioxidant activity of this herb (Ahmad *et al.*, 2014; Christapher *et al.*, 2015). Moreover, previous studies have shown that *P. odorata* was non-toxic in the murine model (Aprianti *et al.*, 2017; Christapher *et al.*, 2017).

## 1.2 Problem Statement

The rapidly rising demand for animal-source protein is soaring worldwide. To meet the increasing demand for animals` protein, antibiotics are extensively used in broiler chicken production to achieve maximum growth. This excessive subtherapeutic use of AGPs has resulted in the emergence of antibiotic resistance. Finding novel alternatives to AGPs as growth promoters is much needed. Numerous studies highlighted the positive effects of phytobiotics like herbs as alternative

growth promoters in broiler chickens (Khan *et al.*, 2017; Paraskeuas *et al.*, 2017; Mashayekhi *et al.*, 2018; Oso *et al.*, 2019). However, literature is scarce about the safe use of herbal plants and their optimal dosage. Additionally, limited studies reported positive effects of herbs containing tannins in broiler chickens production (Starcevic *et al.*, 2015; Bee *et al.*, 2017). This limitation was due to the belief that tannins might have an anti-nutritional effect when supplemented in broiler chickens` diet (Mansoori *et al.*, 2015). In this study, *P. betle* and *P. odorata* leaf meal were evaluated for the first time as the potential alternative to AGPs in broiler chickens.

### 1.3 Objectives

This research was carried out to fulfil the following objectives:

1. To assess the qualitative and quantitative phytochemical screening, antimicrobial and antioxidant potential of *Piper betle* and *Persicaria odorata*.
2. To quantify secondary metabolites and assess the acute oral toxicity of *Piper betle* and *Persicaria odorata* leaf extract in broiler chickens to establish their safe use.
- 3.
4. To determine the effects of graded dose supplementation of *Piper betle* and *Persicaria odorata* leaf meal in broiler chicken feed on growth performance, apparent ileal digestibility, gut morphology, haematological indices, serum biochemical profile, and liver histomorphological changes.
5. To investigate the comparative efficacy of phytobiotics (*Piper betle* and *Persicaria odorata* leaf meal) with halquinol and tetracycline on growth performance, gut morphology, ileal digestibility, meat quality, tibia bone morphometric characteristics, and caecal microbiota composition in broiler chickens.

### 1.4 Hypothesis

It is hypothesised that:

1. The *Piper betle* and *Persicaria odorata* leaf extracts would show antimicrobial and antioxidant potential with successful quantification of secondary bioactive compounds like total phenolic content, total flavonoids, and total tannins.
2. Secondary bioactive compounds eugenol and quercetin would be successfully quantified from the *Piper betle* and *Persicaria odorata* methanolic leaf extract; also, these extracts would show no acute oral toxicity in broiler chickens.

3. Graded dose supplementation of *Piper betle* and *Persicaria odorata* leaf meal would show positive effects on apparent ileal digestibility, improve gut morphology, have no deleterious impact on haematological indices, serum biochemical profile, and liver morphology, thus can enhance growth performance in broiler chickens.
4. Supplementation of *Piper betle* and *Persicaria odorata* leaf meal would show comparable or better performance to halquinol and AGPs as alternative growth promoters in sustainable broiler chickens production.





## REFERENCES

- Abatcha, M. (2017). Salmonella and Listeria monocytogenes: A review of prevalence and antibiotic resistance in chickens and their processing environments. *Advances in Animal Veterinary Sciences*, 5(9), 395-403. [[Google Scholar](#)]
- Abbas, M., Saeed, F., Anjum, F.M., Afzaal, M., Tufail, T., Bashir, M.S., Ishtiaq, A., Hussain, S., & Suleria, H. A. R. (2017). Natural polyphenols: An overview. *International Journal of Food Properties*, 20(8), 1689–1699. <https://doi.org/10.1080/10942912.2016.1220393>
- Abbood, A. A., Kassim, A. Bin, Jawad, H. S. A., Manap, Y. A., & Sazili, A. Q. (2017). Effects of feeding the herb *Borreria latifolia* on the meat quality of village chickens in Malaysia. *Poultry Science*, 96(6), 1767–1782. <https://doi.org/10.3382/ps/pew460>
- Abdulla, N.R., Loh, T.C., Akit, H., Sazili, A.Q., Foo, H.L., Kareem, K.Y., Mohamad, R., & Rahim, R. A. (2017). Effects of dietary oil sources, calcium and phosphorus levels on growth performance, carcass characteristics and bone quality of broiler chickens. *Journal of Applied Animal Research*, 45(1), 423–429. <https://doi.org/10.1080/09712119.2016.1206903>
- Abdullah, M. Z., Mohd Ali, J., Abolmaesoomi, M., Abdul-Rahman, P. S., & Hashim, O. H. (2017). Anti-proliferative, in vitro antioxidant, and cellular antioxidant activities of the leaf extracts from *Polygonum minus* Huds: Effects of solvent polarity. *International Journal of Food Properties*, 20(1), S846–S862. <https://doi.org/10.1080/10942912.2017.1315591>
- Abdurrahman, Z. H., Pramono, Y. B., & Suthama, N. (2016). Feeding effect of inulin derived from dahlia tuber combined with lactobacillus sp. on meat protein mass of crossbred kampong chicken. *Journal of the Indonesian Tropical Animal Agriculture*, 41(1), 37–44. <https://doi.org/10.14710/jitaa.41.1.37-44>
- Abo-EL-Sooud, K. (2018). Ethnoveterinary perspectives and promising future. *International Journal of Veterinary Science and Medicine*, 6(1), 1–7. <https://doi.org/10.1016/j.ijvsm.2018.04.001>
- Abu Hafsa, S. H., & Ibrahim, S. A. (2018). Effect of dietary polyphenol-rich grape seed on growth performance, antioxidant capacity and ileal microflora in broiler chicks. *Journal of Animal Physiology and Animal Nutrition*, 102(1), 268-275. <https://doi.org/10.1111/jpn.12688>
- Abubakar, M. A., Zulkifli, R.M., Hassan, W. N., Shariff, A. H., Malek, N. A.,

- Zakaria, Z., & Ahmad, F. (2015). Antibacterial properties of *Persicaria minor* (Huds.) ethanolic and aqueous-ethanolic leaf extracts. *Journal of Applied Pharmaceutical Science*, 5(Suppl 2), 50–56.  
<https://doi.org/10.7324/JAPS.2015.58.S8>
- Abudabos, A. M., Alyemni, A. H., Dafalla, Y. M., & Khan, R. U. (2016). The effect of phytogetic feed additives to substitute in-feed antibiotics on growth traits and blood biochemical parameters in broiler chicks challenged with *Salmonella typhimurium*. *Environmental Science and Pollution Research*, 23(23), 151–157.  
<https://doi.org/10.1007/s11356-016-7665-2>
- Abudabos, A. M., Alyemni, A. H., Dafalla, Y. M., & Khan, R. U. (2017). Effect of organic acid blend and bacillus subtilis alone or in combination on growth traits, blood biochemical and antioxidant status in broilers exposed to salmonella typhimurium challenge during the starter phase. *Journal of Applied Animal Research*, 45(1), 538–542.  
<https://doi.org/10.1080/09712119.2016.1219665>
- Abudabos, A. M., Alyemni, A. H., Dafalla, Y. M., & Khan, R. U. (2018). The effect of phyto-genics on growth traits, blood biochemical and intestinal histology in broiler chickens exposed to Clostridium perfringens challenge. *Journal of Applied Animal Research*, 46(1), 691–695.  
<https://doi.org/10.1080/09712119.2017.1383258>
- Acamovic, T., & Brooker, J. D. (2005). Biochemistry of plant secondary metabolites and their effects in animals. *Proceedings of the Nutrition Society*, 64(3), 403–412. DOI: <https://doi.org/10.1079/PNS2005449>
- Adegoke, A. V., Abimbola, M. A., Sanwo, K. A., Egbeyale, L. T., Abiona, J. A., Oso, A. O., & Iposu, S. O. (2018). Performance and blood biochemistry profile of broiler chickens fed dietary turmeric (*Curcuma longa*) powder and cayenne pepper (*Capsicum frutescens*) powders as antioxidants. *Veterinary and Animal Science*, 6, 95–102.  
<https://doi.org/10.1016/j.vas.2018.07.005>
- Aguirre, M. E., Owens, C. M., Miller, R. K., & Alvarado, C. Z. (2018). Descriptive sensory and instrumental texture profile analysis of woody breast in marinated chicken. *Poultry Science*, 97(4), 1456–1461.  
<https://doi.org/10.3382/ps/pex428>
- Ahameethunisa, A. R., & Hopper, W. (2010). Antibacterial activity of *Artemisia nilagirica* leaf extracts against clinical and phytopathogenic bacteria. *BMC Complementary and Alternative Medicine*, 10(6), 1–6.  
<https://doi.org/10.1186/1472-6882-10-6>
- Ahmad, R., Baharum, S. N., Bunawan, H., Lee, M., Mohd Noor, N., Rohani, E. R., Ilias, N., & Zin, N. M. (2014). Volatile profiling of aromatic traditional medicinal plant, polygonum minus in different tissues and its biological

activities. *Molecules*, 19(11), 19220–19242.

<https://doi.org/10.3390/molecules191119220>

Ahmad, S., Khalique, A., Pasha, T. N., Mehmood, S., Hussain, K., Ahmad, S., Rasheed, B., Awais, M. M., & Bhatti, S. A. (2018). Influence of feeding *Moringa oleifera* pods as phyto-genic feed additive on performance, blood metabolites, chemical composition and bioactive compounds of breast meat in broiler. *Kafkas Üniversitesi Veteriner Fakültesi Dergisi*, 24(2).  
[[Google Scholar](#)]

Ahmadi, F., Shahbazi, Y., & Karami, N. (2015). Determination of tetracyclines in meat using two phases freezing extraction method and HPLC-DAD. *Food analytical methods*, 8(7), 1883-1891. [[Google Scholar](#)]

Ahsan, U., Kuter, E., Raza, I., Köksal, B. H., Cengiz, Ö., Yıldız, M., Kızanlık, P. K., Kaya, M., Tatlı, O., & Sevim, Ö. (2018). Dietary supplementation of different levels of phyto-genic feed additive in broiler diets: The dynamics of growth performance, caecal microbiota, and intestinal morphometry. *Brazilian Journal of Poultry Science*, 20(4), 737-746.  
<https://doi.org/10.1590/1806-9061-2017-0698>

Akbarian, A., Michiels, J., Degroote, J., Majdeddin, M., Golian, A., & De Smet, S. (2016). Association between heat stress and oxidative stress in poultry; mitochondrial dysfunction and dietary interventions with phytochemicals. *Journal of Animal Science and Biotechnology*, 7(1), 37.  
<https://doi.org/10.1186/s40104-016-0097-5>

Al-Abd, N. M., Mohamed Nor, Z., Mansor, M., Azhar, F., Hasan, M. S., & Kassim, M. (2015). Antioxidant, antibacterial activity, and phytochemical characterization of *Melaleuca cajuputi* extract. *BMC Complementary and Alternative Medicine*, 15(1), 1–13.  
<https://doi.org/10.1186/s12906-015-0914-y>

Alagbe, J. O. (2017). Effect of Dietary Inclusion of *Polyalthia longifolia* Leaf Meal as Phytobiotic Compared with Antibiotics on the Nutrient Retention, Immune Response and Serum Biochemistry of Broiler Chicken. *Greener Journal of Agricultural Sciences*, 7(3), 074–081.  
<https://doi.org/10.15580/gjas.2017.3.052217067>

Al-Dhabi, N. A., Mohammed Ghilan, A. K., Esmail, G. A., Valan Arasu, M., Duraipandiyar, V., & Ponmurugan, K. (2019). Bioactivity assessment of the Saudi Arabian Marine *Streptomyces* sp. Al-Dhabi-90, metabolic profiling and its in vitro inhibitory property against multidrug resistant and extended-spectrum beta-lactamase clinical bacterial pathogens. *Journal of Infection and Public Health*, 12(4), 549–556.  
<https://doi.org/10.1016/j.jiph.2019.01.065>

Alebiosu, C. O., & Yusuf, A. J. (2015). Phytochemical Screening, Thin-layer Chromatographic Studies and UV Analysis of Extracts of *Citrullus*

lanatus. *Journal of Pharmaceutical, Chemical and Biological Sciences*, 3(2), 214–220. [[Google Scholar](#)]

Alhidary, I. A., Abdelrahman, M. M., Uallh Khan, R., & Harron, R. M. (2016). Antioxidant status and immune responses of growing camels supplemented a long-acting multi-trace minerals rumen bolus. *Italian Journal of Animal Science*, 15(2), 343-349. <https://doi.org/10.1080/1828051X.2016.1186502>

Ali Asghar Saki. (2012). Herbal additives and organic acids as antibiotic alternatives in broiler chickens diet for organic production. *African Journal of Biotechnology*, 11(8), 2139–2145. <https://doi.org/10.5897/ajb11.797>

Ali, M., Kenganora, M., & Manjula, S. N. (2016). Health benefits of *Morinda citrifolia* (Noni): A review. *Pharmacognosy Journal*, 8(4). [[Google Scholar](#)]

Ali, A., Chong, C. H., Mah, S. H., Abdullah, L. C., Choong, T. S. Y., & Chua, B. L. (2018). Impact of storage conditions on the stability of predominant phenolic constituents and antioxidant activity of dried piper betle extracts. *Molecules*, 23(2). <https://doi.org/10.3390/molecules23020484>

Ali, A., Lim, X. Y., Chong, C. H., Mah, S. H., & Chua, B. L. (2018). Optimization of ultrasound-assisted extraction of natural antioxidants from Piper betle using response surface methodology. *LWT - Food Science and Technology*, 89(November 2017), 681–688. <https://doi.org/10.1016/j.lwt.2017.11.033>

Aljumaah, M. R., Suliman, G. M., Abdullatif, A. A., & Abudabos, A. M. (2020). Effects of phytobiotic feed additives on growth traits, blood biochemistry, and meat characteristics of broiler chickens exposed to *Salmonella typhimurium*. *Poultry Science*, 99(11), 5744–5751. <https://doi.org/10.1016/j.psj.2020.07.033>

Al-Mnaser, A., & Woodward, M. J. (2017). Oregano extract as an alternative to antibiotics in poultry feed. In: N8 Agrifood International Conference: *Food Production for the Future*. Durham University, Durham, UK.

Alsanad, S. M., Howard, R. L., & Williamson, E. M. (2016). An assessment of the impact of herb-drug combinations used by cancer patients. *BMC Complementary and Alternative Medicine*, 16(1), 393. <https://doi.org/10.1186/s12906-016-1372-x>

Alshelmani, M. I., Loh, T. C., Foo, H. L., Sazili, A. Q., & Lau, W. H. (2016). Effect of feeding different levels of palm kernel cake fermented by *Paenibacillus polymyxa* ATCC 842 on nutrient digestibility, intestinal morphology, and gut microflora in broiler chickens. *Animal Feed Science and Technology*, 216, 216–224. <https://doi.org/10.1016/j.anifeedsci.2016.03.019>

- Alshelmani, M. I., Loh, T. C., Foo, H. L., Sazili, A. Q., & Lau, W. H. (2017). Effect of solid-state fermentation on nutrient content and ileal amino acids digestibility of palm kernel cake in broiler chickens. *Indian Journal of Animal Sciences*, 87(9), 1135-1140. [[Google Scholar](#)]
- Amaral, A. B., Solva, M. V. Da., & Lannes, S. C. D. S. (2018). Lipid oxidation in meat: Mechanisms and protective factors - a review. *Food Science and Technology*, 38(1), 1–15. <https://doi.org/10.1590/fst.32518>
- AMSA. (2012). *AMSA Meat Color Measurement Guidelines* (December 2012 ed.) American Meat Science Association, Champaign, Illinois USA, 61820, 1-135. [[Google Scholar](#)]
- Anand David, A. V., Arulmoli, R., & Parasuraman, S. (2016). Overviews of biological importance of quercetin: A bioactive flavonoid. *Pharmacognosy Reviews*, 10(20), 84–89. <https://doi.org/10.4103/0973-7847.194044>
- Anand, U., Jacobo-Herrera, N., Altemimi, A., & Lakhssassi, N. (2019). A comprehensive review on medicinal plants as antimicrobial therapeutics: Potential avenues of biocompatible drug discovery. *Metabolites*, 9(11), 1–13. <https://doi.org/10.3390/metabo9110258>
- Angel, C. R., Saylor, W., Vieira, S. L., & Ward, N. (2011). Metabolism and nutrition: Effects of a monocomponent protease on performance and protein utilization in 7- to 22-day-old broiler chickens. *Poultry Science*, 90(10), 2281–2286. <https://doi.org/10.3382/ps.2011-01482>
- Angelakis, E. (2017). Weight gain by gut microbiota manipulation in productive animals. *Microbial Pathogenesis*, 106(2017), 162–170. <https://doi.org/10.1016/j.micpath.2016.11.002>
- Annegowda, H. V., Tan, P. Y., Mordi, M. N., Ramanathan, S., Hamdan, M. R., Sulaiman, M. H., & Mansor, S. M. (2013). TLC-Bioautography-Guided Isolation, HPTLC and GC-MS-Assisted Analysis of Bioactives of Piper betle Leaf Extract Obtained from Various Extraction Techniques: In vitro Evaluation of Phenolic Content, Antioxidant and Antimicrobial Activities. *Food Analytical Methods*, 6(3), 715–726. <https://doi.org/10.1007/s12161-012-9470-y>
- Anokwuru, C. P., Anyasor, G. N., Ajibaye, O., Fakoya, O., & Okebugwu, P. (2011). Effect of Extraction Solvents on Phenolic, Flavonoid and Antioxidant activities of Three Nigerian Medicinal Plants. *Nature and Science*, 9(7), 53–61. [[Google Scholar](#)]

- AOAC, Association of Official Analytical Chemist. (1995). Official method of analysis of AOAC International. 16th ed. Arlington: Association of Official Analytical Chemist. [[Google Scholar](#)]
- Aprianti, W., Widiyatno, T. V., & Sudjarwo, S. A. (2017). Effect of Polygonum Minus (Knotweed) Leaves Extract on the Histopathological Changes of Kidney in Mice (*Mus Musculus*) Induced by Mercuric Chloride. *KnE Life Sciences*, 3(6), 753. <https://doi.org/10.18502/cls.v3i6.1206>
- Arasu, M. V., Arokiyaraj, S., Viayaraghavan, P., Kumar, T. S. J., Duraipandiyar, V., Al-Dhabi, N. A., & Kaviyarasu, K. (2019). One step green synthesis of larvicidal, and azo dye degrading antibacterial nanoparticles by response surface methodology. *Journal of Photochemistry and Photobiology B: Biology*, 190(September 2018), 154–162. <https://doi.org/10.1016/j.jphotobiol.2018.11.020>
- Arawwawala, L. D. A. M., Arambewela, L. S. R., & Ratnasooriya, W. D. (2014). Gastroprotective effect of *Piper betle* Linn. leaves grown in Sri Lanka. *Journal of Ayurveda and Integrative Medicine*, 5(1), 38. <https://doi.org/10.4103/0975-9476.128855>
- Archana, P., Samatha, T., Mahitha, B., & Chamundeswari, N. R. (2012). Preliminary phytochemical screening from leaf and seed extracts of *Senna alata* L. Roxb-an ethnomedicinal plant. *International Journal of Pharmaceutical and Biological Research*, 3(3), 82-89. [[Google Scholar](#)]
- Arief Deli, R. N. R., & Adzitey, F. (2017). Prevalence and antibiotic resistance of Salmonella serovars isolated from spent hens and its environmental samples in Penang and Kedah, Malaysia. *Journal of Tropical Agriculture and Food Science*, 45(1), 37-50. [[Google Scholar](#)]
- Aroche, R., Martinez, Y., Ruan, Z., Guan, G., Waititu, S., Nyachoti, C. M., Más, D., & Lan, S. (2018). Dietary inclusion of a mixed powder of medicinal plant leaves enhances the feed efficiency and immune function in broiler chickens. *Journal of Chemistry*, 2018(1), 1–6. <https://doi.org/10.1155/2018/4073068>
- Arokiyaraj, S., Saravanan, M., & Vijayakumar, B. (2015). Green synthesis of Silver nanoparticles using aqueous extract of *Taraxacum officinale* and its antimicrobial activity Manuscript details Abstract. *South Indian Journal of Biological Sciences*, 1(2), 1. [[Google Scholar](#)]
- Arsi, K., Donoghue, A. M., Woo-Ming, A., Blore, P. J., & Donoghue, D. J. (2015). The efficacy of selected probiotic and prebiotic combinations in reducing *Campylobacter* colonization in broiler chickens. *Journal of Applied Poultry Research*, 24(3), 327–334. <https://doi.org/10.3382/japr/pfv032>
- Asgari Lajayer, B., Ghorbanpour, M., & Nikabadi, S. (2017). Heavy metals in

contaminated environment: Destiny of secondary metabolite biosynthesis, oxidative status and phytoextraction in medicinal plants. *Ecotoxicology and Environmental Safety*, 145(April), 377–390.  
<https://doi.org/10.1016/j.ecoenv.2017.07.035>

Atanasov, A. G., Waltenberger, B., Pferschy-Wenzig, E. M., Linder, T., Wawrosch, C., Uhrin, P., Temml, V., Wang, L., Schwaiger, S., Heiss, E. H., & Stuppner, H. (2015). Discovery and resupply of pharmacologically active plant-derived natural products: A review. *Biotechnology Advances*, 33(8), 1582–1614.  
<https://doi.org/10.1016/j.biotechadv.2015.08.001>

Atiya, A., Sinha, B. N., & Ranjan Lal, U. (2018). New chemical constituents from the Piper betle Linn. (Piperaceae). *Natural product research*, 32(9), 1080–1087.  
<https://doi.org/10.1080/14786419.2017.1380018>

Attia, Y. A., Bakhashwain, A. A., & Bertu, N. K. (2018). Utilisation of thyme powder (*Thyme vulgaris* L.) as a growth promoter alternative to antibiotics for broiler chickens raised in a hot climate. *European Poultry Science*, 82.  
<https://doi.org/10.1399/eps.2018.238>

Aumeeruddy-Elalfi, Z., Gurib-Fakim, A., & Mahomoodally, F. (2015). Antimicrobial, antibiotic potentiating activity and phytochemical profile of essential oils from exotic and endemic medicinal plants of Mauritius. *Industrial Crops and Products*, 71(2015), 197–204.  
<https://doi.org/10.1016/j.indcrop.2015.03.058>

Ayodele, S. O., Oloruntola, O. D., & Agbede, J. O. (2016). Effect of alchornea cordifolia leaf meal inclusion and enzyme supplementation on performance and digestibility of rabbits. *World Rabbit Science*, 24(3), 201–206.  
<https://doi.org/10.4995/wrs.2016.3933>

Aziz, M., & Karboune, S. (2018). Natural antimicrobial/antioxidant agents in meat and poultry products as well as fruits and vegetables: A review. *Critical Reviews in Food Science and Nutrition*, 58(3), 486–511.  
<https://doi.org/10.1080/10408398.2016.1194256>

Bahri, S. I. S., Ariffin, A. S., & Mohtar, S. (2019). Critical Review on Food Security in Malaysia for Broiler Industry. *International Journal of Academic Research in Business and Social Sciences*, 9(7), 869–876.  
<https://doi.org/10.6007/ijarbss/v9-i7/6186>

Banerjee, S., Mukhopadhyay, S. K., Haldar, S., Ganguly, S., Pradhan, S., Patra, N. C., Niyogi, D., & Isore, D. (2013). Effect of phyto-genic growth promoter on broiler birds. *Indian Journal of Veterinary Pathology*, 37(1), 34–37.  
[\[Google Scholar\]](#)

- Banso, A., & Adeyemo, S. O. (2007). Evaluation of antibacterial properties of tannins isolated from *Dichrostachys cinerea*. *African Journal of Biotechnology*, 6(15), 1785–1787.  
<https://doi.org/10.5897/AJB2007.000-2262>
- Basak, S., & Guha, P. (2017). Use of predictive model to describe sporocidal and cell viability efficacy of betel leaf (*Piper betle* L.) essential oil on *Aspergillus flavus* and *Penicillium expansum* and its antifungal activity in raw apple juice. *LWT - Food Science and Technology*, 80, 510–516.  
<https://doi.org/10.1016/j.lwt.2017.03.024>
- Basit, M. A., Arifah, A. K., Loh, T. C., Saleha, A. A., Salleh, A., Kaka, U., & Idris, S. B. (2020a). Effects of graded dose dietary supplementation of Piper betle leaf meal and *Persicaria odorata* leaf meal on growth performance, apparent ileal digestibility, and gut morphology in broilers. *Saudi Journal of Biological Sciences*, 27(6), 1503–1513.  
<https://doi.org/10.1016/j.sjbs.2020.04.017>
- Basit, M. A., Kadir, A. A., Loh, T. C., Aziz, S. A., Salleh, A., Kaka, U., & Idris, S. B. (2020b). Effects of inclusion of different doses of *Persicaria odorata* leaf meal (POLM) in broiler chicken feed on biochemical and haematological blood indicators and liver histomorphological changes. *Animals*, 10(7), 1–18. <https://doi.org/10.3390/ani10071209>
- Basit, M. A., Kadir, A. A., Loh, T. C., Abdul Aziz, S., Salleh, A., Zakaria, Z. A., & Banke Idris, S. (2020c). Comparative Efficacy of Selected Phytobiotics with Halquinol and Tetracycline on Gut Morphology, Ileal Digestibility, Cecal Microbiota Composition and Growth Performance in Broiler Chickens. *Animals*, 10(11), 2150. <https://doi.org/10.3390/ani10112150>
- Baskaran, S. A., Kollanoor-Johny, A., Nair, M. S., & Venkitanarayanan, K. (2016). Efficacy of plant-derived antimicrobials in controlling enterohemorrhagic *Escherichia coli* virulence in vitro. *Journal of Food Protection*, 79(11), 1965–1970.  
<https://doi.org/10.4315/0362-028X.JFP-16-104>
- Batiha, G. E. S., Beshbishy, A. M., Mulla, Z. S., Ikram, M., El-Hack, M. E. A., Taha, A. E., Algammal, A. M., & Elewa, Y. H. A. (2020). The pharmacological activity, biochemical properties, and pharmacokinetics of the major natural polyphenolic flavonoid: quercetin. *Foods*, 9(3), 374.  
<https://doi.org/10.3390/foods9030374>
- Bauer, A. W., Kirby, W. M. M., Sherris, J. C., & Turck, M. (1966). Antibiotic Susceptibility Testing by a Standardized Single Disk Method. *American Journal of Clinical Pathology*, 45(4\_ts), 493–496.  
[https://doi.org/10.1093/ajcp/45.4\\_ts.493](https://doi.org/10.1093/ajcp/45.4_ts.493)
- Baynes, R. E., Dedonder, K., Kissell, L., Mzyk, D., Marmulak, T., Smith, G., Tell, L., Gehring, R., Davis, J., & Riviere, J. E. (2016). Health concerns and



management of select veterinary drug residues. *Food and Chemical Toxicology*, 88, 112-122.

<https://doi.org/10.1016/j.fct.2015.12.020>

Bee, G., Silacci, P., Ampuero-Kragten, S., Čandek-Potokar, M., Wealleans, A. L., Litten-Brown, J., Salminen, J. P., & Mueller-Harvey, I. (2017). Hydrolysable tannin-based diet rich in gallotannins has a minimal impact on pig performance but significantly reduces salivary and bulbourethral gland size. *Animal*, 11(9), 1617-1625.

<https://doi.org/10.1017/S1751731116002597>

Begam K. M. F., Ravichandra, P., and Manimekalai, V. (2018). Phytochemical Analysis of Some Selected Varieties of Piper Betle L. *International Journal of Current Pharmaceutical Research*, 10(2), 89.

<https://doi.org/10.22159/ijcpr.2018v10i2.25884>

Bengtsson-Palme, J., Kristiansson, E., & Larsson, D. J. (2018). Environmental factors influencing the development and spread of antibiotic resistance. *FEMS microbiology reviews*, 42(1), fux053.

<https://doi.org/10.1093/femsre/fux053>

Bertelli, D., Brighenti, V., Marchetti, L., Reik, A., & Pellati, F. (2018). Nuclear magnetic resonance and high-performance liquid chromatography techniques for the characterization of bioactive compounds from *Humulus lupulus* L. (hop). *Analytical and Bioanalytical Chemistry*, 410(15), 3521–3531.

<https://doi.org/10.1007/s00216-018-0851-y>

Bindhu R, K., & K, S. (2018). in Vitro Study on Antioxidant Activity of Methanolic Leaf Extract of Piper Betle Linn. *Journal of Evolution of Medical and Dental Sciences*, 7(24), 2865–2869.

<https://doi.org/10.14260/jemds/2018/646>

Biswajit Patra, Das, M. T., Dey, S. K., & Das, T. (2016). A review on Piper betle L. *J Med Plants Stud*, 4(6), 185-192. [\[Google Scholar\]](#)

Boovaragamoorthy, G. M., Anbazhagan, M., Piruthiviraj, P., Pugazhendhi, A., Kumar, S. S., Al-Dhabi, N. A., Ghilan, A. K. M., Arasu, M.V., & Kaliannan, T. (2019). Clinically important microbial diversity and its antibiotic resistance pattern towards various drugs. *Journal of Infection and Public Health*, 12(6), 783–788.

<https://doi.org/10.1016/j.jiph.2019.08.008>

Braykov, N. P., Eisenberg, J. N., Grossman, M., Zhang, L., Vasco, K., Cevallos, W., Muñoz, D., Acevedo, A., Moser, K. A., Marrs, C. F., & Foxman, B. (2016). Antibiotic resistance in animal and environmental samples associated with small-scale poultry farming in northwestern Ecuador. *Mosphere*, 1(1).

[DOI: 10.1128/mSphere.00021-15](https://doi.org/10.1128/mSphere.00021-15)

- Brenes, A., & Roura, E. (2010). Essential oils in poultry nutrition: Main effects and modes of action. *Animal Feed Science and Technology*, 158(1–2), 1–14.  
<https://doi.org/10.1016/j.anifeedsci.2010.03.007>
- Brenes, A., Viveros, A., Chamorro, S., & Arija, I. (2016). Use of polyphenol-rich grape by-products in monogastric nutrition. A review. *Animal Feed Science and Technology*, 211(2016), 1–17.  
<https://doi.org/10.1016/j.anifeedsci.2015.09.016>
- Brodersen, D. E., Clemons Jr, W. M., Carter, A. P., Morgan-Warren, R. J., Wimberly, B. T., & Ramakrishnan, V. (2000). The structural basis for the action of the antibiotics tetracycline, pactamycin, and hygromycin B on the 30S ribosomal subunit. *Cell*, 103(7), 1143-1154.  
[https://doi.org/10.1016/S0092-8674\(00\)00216-6](https://doi.org/10.1016/S0092-8674(00)00216-6)
- Brown, K., Uwiera, R. R., Kalmokoff, M. L., Brooks, S. P., & Inglis, G. D. (2017). Antimicrobial growth promoter use in livestock: a requirement to understand their modes of action to develop effective alternatives. *International journal of Antimicrobial Agents*, 49(1), 12-24.  
<https://doi.org/10.1016/j.ijantimicag.2016.08.006>
- Brus, M., Dolinšek, J., Cencič, A., & Škorjanc, D. (2013). Effect of chestnut (*Castanea sativa* Mill.) wood tannins and organic acids on growth performance and faecal microbiota of pigs from 23 to 127 days of age. *Bulgarian Journal of Agricultural Science*, 19(4), 841-847.  
[[Google Scholar](#)]
- Brussow, H. (2015). Growth promotion and gut microbiota: Insights from antibiotic use. *Environmental Microbiology*, 17(7), 2216–2227.  
<https://doi.org/10.1111/1462-2920.12786>
- Butkhup, L., Chowtivanakul, S., Gaensakoo, R., Prathepha, P., & Samappito, S. (2010). Study of the phenolic composition of Shiraz red grape cultivar (*Vitis vinifera* L.) cultivated in Northeastern Thailand and its antioxidant and antimicrobial activity. *South African Journal of Enology and Viticulture*, 31(2), 89–98.  
<https://doi.org/10.21548/31-2-1405>
- Cai, Y., Zhang, J., Chen, N. G., Shi, Z., Qiu, J., He, C., & Chen, M. (2017). Recent Advances in Anticancer Activities and Drug Delivery Systems of Tannins. *Medicinal Research Reviews*, 37(4), 665–701.  
<https://doi.org/10.1002/med>
- Calo, J. R., Crandall, P. G., O'Bryan, C. A., & Ricke, S. C. (2015). Essential oils as antimicrobials in food systems—A review. *Food Control*, 54, 111-119.  
[doi.org/10.1016/j.foodcont.2014.12.040](https://doi.org/10.1016/j.foodcont.2014.12.040)
- Carrasco, J. D., Cabral, C., Redondo, L. M., Viso, N. P., Farber, M. D., & Miyakawa, M. E. F. (2016, December). Impact of dietary tannins on rumen

microbiota of bovines. In *2nd International Symposium on Alternative to Antibiotics (ATA) Challenges and Solution in Animal Production*, OIE Headquarter, Paris, France (pp. 12-15).

[\[Google Scholar\]](#)

Carvalho, I. T., & Santos, L. (2016). Antibiotics in the aquatic environments: a review of the European scenario. *Environment International*, 94, 736-757. [doi.org/10.1016/j.envint.2016.06.025](https://doi.org/10.1016/j.envint.2016.06.025)

Chamorro, S., Romero, C., Brenes, A., Sánchez-Patán, F., Bartolomé, B., Viveros, A., & Arija, I. (2019). Impact of a sustained consumption of grape extract on digestion, gut microbial metabolism and intestinal barrier in broiler chickens. *Food and Function*, 10(3), 1444–1454.

<https://doi.org/10.1039/c8fo02465k>

Chan, E. W. C., Tan, Y. P., Chin, S. J., Gan, L. Y., Kang, K. X., Fong, C. H., Chang, H. Q., & Chern How, Y. (2014). Antioxidant properties of selected fresh and processed herbs and vegetables. *Free Radicals and Antioxidants*, 4(1), 39–46. <https://doi.org/10.5530/fra.2014.1.7>

Chang, C. L. T., Chung, C. Y., Kuo, C. H., Kuo, T. F., Yang, C. W., & Yang, W. C. (2016). Beneficial effect of *Bidens pilosa* on body weight gain, food conversion ratio, gut bacteria and coccidiosis in chickens. *PLoS ONE*, 11(1), 1–15. <https://doi.org/10.1371/journal.pone.0146141>

Chansiw, N., Chotinantakul, K., & Srichairatanakool, S. (2019). Anti-inflammatory and Antioxidant Activities of the Extracts from Leaves and Stems of *Polygonum odoratum* Lour. *Anti-Inflammatory & Anti-Allergy Agents in Medicinal Chemistry*, 18(1), 45–54.

<https://doi.org/10.2174/1871523017666181109144548>

Chitnis, K. S. (2017). Quantitation of eugenol in betel leaf varieties by HPTLC. *International Journal of Pharmaceutical Sciences and Research*, 8(11), 4858-4862. [\[Google Scholar\]](#)

Chopra, I., & Roberts, M. (2001). Tetracycline antibiotics: mode of action, applications, molecular biology, and epidemiology of bacterial resistance. *Microbiology and Molecular Biology Reviews*, 65(2), 232-260. [doi: 10.1128/MMBR.65.2.232-260.2001](https://doi.org/10.1128/MMBR.65.2.232-260.2001)

Chouhan, S., Sharma, K., & Guleria, S. (2017). Antimicrobial Activity of Some Essential Oils Present Status and Future Perspectives. *Medicines*, 4(3), 58. <https://doi.org/10.3390/medicines4030058>

Chowdhury, P. R., McKinnon, J., Wyrsh, E., Hammond, J. M., Charles, I. G., & Djordjevic, S. P. (2014). Genomic interplay in bacterial communities: Implications for growth promoting practices in animal husbandry. *Frontiers in Microbiology*, 5(AUG), 1–12.

<https://doi.org/10.3389/fmicb.2014.00394>

- Chowdhury, S., Mandal, G. P., Patra, A. K., Kumar, P., Samanta, I., Pradhan, S., & Samanta, A. K. (2018). Different essential oils in diets of broiler chickens: 2. Gut microbes and morphology, immune response, and some blood profile and antioxidant enzymes. *Animal Feed Science and Technology*, 236, 39-47.  
[doi.org/10.1016/j.anifeedsci.2017.12.003](https://doi.org/10.1016/j.anifeedsci.2017.12.003)
- Christopher, P., Parasuraman, S., Christina, J. A., Vikneswaran, M., & Asmawi, M. Z. (2015). Review on Polygonum minus. Huds, a commonly used food additive in Southeast Asia. *Pharmacognosy Research*, 7(1),  
[doi:10.4103/0974-8490.147125](https://doi.org/10.4103/0974-8490.147125)
- Christopher, P. V., Parasuraman, S., Vasanth Raj, P., Saghir, S. A. M., Asmawi, M. Z., & Vikneswaran, M. (2016). Influence of extracting solvent on pharmacological activity and cytotoxicity of Polygonum minus, a commonly consumed herb in Southeast Asia. *Pharmacognosy Magazine*, 12(47), S424–S430.  
<https://doi.org/10.4103/0973-1296.191451>
- Christopher, P. V., Parasuraman, S., Asmawi, M. Z., & Murugaiyah, V. (2017). Acute and subchronic toxicity studies of methanol extract of Polygonum minus leaves in Sprague Dawley rats. *Regulatory Toxicology and Pharmacology*, 86, 33–41.  
<https://doi.org/10.1016/j.yrtph.2017.02.005>
- Chua, A. Q., Verma, M., Hsu, L. Y., & Legido-Quigley, H. (2021). An analysis of national action plans on antimicrobial resistance in Southeast Asia using a governance framework approach. *The Lancet Regional Health-Western Pacific*, 7, 100084. <https://doi.org/10.1016/j.lanwpc.2020.100084>
- Chumngoen, W., & Tan, F. J. (2015). Relationships between descriptive sensory attributes and physicochemical analysis of broiler and Taiwan native chicken breast meat. *Asian-Australasian Journal of Animal Sciences*, 28(7), 1028–1037. <https://doi.org/10.5713/ajas.14.0275>
- Clavijo, V., & Flórez, M. J. V. (2018). The gastrointestinal microbiome and its association with the control of pathogens in broiler chicken production: a review. *Poultry science*, 97(3), 1006-1021.  
<https://doi.org/10.3382/ps/pex359>
- Cogliani, C., Goossens, H., & Greko, C. (2011). Restricting antimicrobial use in food animals: lessons from Europe. *Microbe*, 6(6), 274. [[Google Scholar](#)]
- Cosby, D. E., Cox, N. A., Harrison, M. A., Wilson, J. L., Buhr, R. J., & Fedorka-Cray, P. J. (2015). Salmonella and antimicrobial resistance in broilers: A review. *Journal of Applied Poultry Research*, 24(3), 408-426.  
<https://doi.org/10.3382/japr/pfv038>
- Costa, M. C., Bessegatto, J. A., Alfieri, A. A., Weese, J. S., Filho, J. A. B., & Oba,

- A. (2017). Different antibiotic growth promoters induce specific changes in the cecal microbiota membership of broiler chicken. *PLoS ONE*, 12(2), 1–13.  
<https://doi.org/10.1371/journal.pone.0171642>
- Das, B., Mandal, D., Dash, S. K., Chattopadhyay, S., Tripathy, S., Dolai, D. P., Dey, S.K., & Roy, S. (2016). Eugenol provokes ROS-mediated membrane damage-associated antibacterial activity against clinically isolated multidrug-resistant *Staphylococcus aureus* strains. *Infectious Diseases: Research and Treatment*, 9, IDRT-S31741.  
[DOI: 10.4137/idrt.s31741](https://doi.org/10.4137/idrt.s31741)
- Das, S., Parida, R., Sriram Sandeep, I., Nayak, S., & Mohanty, S. (2016). Biotechnological intervention in betelvine (*Piper betle* L.): A review on recent advances and future prospects. *Asian Pacific Journal of Tropical Medicine*, 9(10), 938–946.  
<https://doi.org/10.1016/j.apjtm.2016.07.029>
- Das, Q., Islam, M., Lepp, D., Tang, J., Yin, X., Mats, L., Liu, H., Ross, K., Kennes, Y. M., Yacini, H. & Diarra, M. S. (2020). Gut Microbiota, Blood Metabolites and Spleen Immunity in Broiler Chickens fed Berry Pomaces and Phenolic-Enriched Extractives. *Frontiers in Veterinary Science*, 7, 150.  
[DOI: 10.3389/fvets.2020.00150](https://doi.org/10.3389/fvets.2020.00150)
- Dash, G. K., & Zakaria, Z. B. (2016). Pharmacognostic studies on *Persicaria odorata* (Lour.) Sojak. *Journal of Pharmacy Research*, 10(6), 377–380.  
[\[Google Scholar\]](#)
- Davies, J. (2007). Microbes have the last word: A drastic re-evaluation of antimicrobial treatment is needed to overcome the threat of antibiotic-resistant bacteria. *EMBO reports*, 8(7), 616–621.  
<https://doi.org/10.1038/sj.embor.7401022>
- Debnath, B. C., Choudhary, K. B. D., Ravikanth, K., Thakur, A., Maini, S., & Nagar, R. K. (2014). Comparative efficacy of natural growth promoter (Av/Agp/10) with antibiotic growth promoter on overall growth performance and intestinal morphometry in broiler birds. *International Journal of Pharmaceutical Science and Health Care*, 2(4), 155–168.  
[\[Google Scholar\]](#)
- Deepak, P., Balamuralikrishnan, B., Park, S., Sowmiya, R., Balasubramani, G., Aiswarya, D., Amutha, V., & Perumal, P. (2019). Phytochemical profiling of marine red alga, *Halymenia palmata* and its bio-control effects against Dengue Vector, *Aedes aegypti*. *South African Journal of Botany*, 121, 257–266.  
<https://doi.org/10.1016/j.sajb.2018.11.011>

- Department of Standards Malaysia. (2009). *Halal Food—Production, Preparation, Handling and Storage—General Guideline*; Department of Standards Malaysia: Cyberjaya, Malaysia, pp. 1–26. Available online: [www.jsm.gov.my](http://www.jsm.gov.my) (accessed on 26 September 2020)
- Department of Veterinary Services DVS. (2019). *Livestock Statistics, Putrajaya*: Department of Veterinary Services (DVS), Ministry of Agriculture and Agro-Based Industry Malaysia. [https://www.dosm.gov.my/v1/index.php?r=column/cthemByCat&cat=164&bul\\_id=OTM1TDMzS11vYm5mU1JiU1Fwekt3UT09&menu\\_id=Z0VTZGU1UHBTU1VJMF1paXRRR0xpdz09](https://www.dosm.gov.my/v1/index.php?r=column/cthemByCat&cat=164&bul_id=OTM1TDMzS11vYm5mU1JiU1Fwekt3UT09&menu_id=Z0VTZGU1UHBTU1VJMF1paXRRR0xpdz09) (accessed on 20 November 2020)
- Di Sotto, A., Di Giacomo, S., Amatore, D., Locatelli, M., Vitalone, A., Toniolo, C., Rotino, G. L., Lo Scalzo, R., Palamara, A. T., Maccocci, M. E., & Nencioni, L. (2018). A polyphenol rich extract from *Solanum melongena* L. DR2 peel exhibits antioxidant properties and anti-herpes simplex virus type 1 activity in vitro. *Molecules*, 23(8), 2066. <https://doi.org/10.3390/molecules23082066>
- Diaz Carrasco, J. M., Redondo, E. A., Pin Viso, N. D., Redondo, L. M., Farber, M. D., & Fernandez Miyakawa, M. E. (2018). Tannins and bacitracin differentially modulate gut microbiota of broiler chickens. *BioMed Research International*, 1–11. <https://doi.org/10.1155/2018/1879168>
- Diaz-Sanchez, S., D'Souza, D., Biswas, D., & Hanning, I. (2015). Botanical alternatives to antibiotics for use in organic poultry production. *Poultry Science*, 94(6), 1419–1430. <https://doi.org/10.3382/ps/pev014>
- Dida, M. F. (2016). Review paper on enzyme supplementation in poultry ration. *International Journal of Bioorganic Chemistry*, 1(1), 1-7. [[Google Scholar](#)]
- Dos Santos, C. M., Campos, J. F., Dos Santos, H. F., Balestieri, J. B. P., Silva, D. B., De Picoli Souza, K., Carlos Alexandre Carollo, C. A., Estevinho, L. M., & Dos Santos, E. L. (2017). Chemical Composition and Pharmacological Effects of Geopropolis Produced by *Melipona quadrifasciata anthidioides*. *Oxidative Medicine and Cellular Longevity*, 2017. <https://doi.org/10.1155/2017/8320804>
- Dubey, K., Anand, B. G., Shekhawat, D. S., & Kar, K. (2017). Eugenol prevents amyloid formation of proteins and inhibits amyloid-induced hemolysis. *Scientific Reports*, 7, 40744. <https://doi.org/10.1038/srep40744>
- El-Faras, A. A., & Elsayaf, A. L. (2017). Hepatoprotective activity of quercetin against paracetamol-induced liver toxicity in rats. *Tanta Medical Journal*, 45(2), 92. [[Google Scholar](#)]

- El-Tarabily, K. A., M. T. El-Saadony, M. Alagawany, M. Arif, G. E. Batiha, A. F. Khafaga, H. A. Elwan, S. S. Elnesr, and M. E. Abd El-Hack. 2021. Using essential oils to overcome bacterial biofilm formation and their antimicrobial resistance. *Saudi Journal of Biological Sciences*. 28:5145–5156. [doi:10.1016/j.sjbs.2021.05.033](https://doi.org/10.1016/j.sjbs.2021.05.033).
- Elshikh, M., Ahmed, S., Mcgaw, M., Marchant, R., Funston, S., Dunlop, P., & Banat, I. M. (2016). Resazurin-based 96-well plate microdilution method for the determination of minimum inhibitory concentration of biosurfactants. *Biotechnology Letters*, 38(6), 1015–1019. <https://doi.org/10.1007/s10529-016-2079-2>
- El-Toumy, S. A., Salib, J. Y., El-Kashak, W. A., Marty, C., Bedoux, G., & Bourgougnon, N. (2018). Antiviral effect of polyphenol rich plant extracts on herpes simplex virus type 1. *Food Science and Human Wellness*, 7(1), 91–101. <https://doi.org/10.1016/j.fshw.2018.01.001>
- Espinosa-Diez, C., Miguel, V., Mennerich, D., Kietzmann, T., Sánchez-Pérez, P., Cadenas, S., & Lamas, S. (2015). Antioxidant responses and cellular adjustments to oxidative stress. *Redox biology*, 6, 183-197. <https://doi.org/10.1016/j.redox.2015.07.008>
- Falcinelli, S., Picchiatti, S., Rodiles, A., Cossignani, L., Merrifield, D. L., Taddei, A. R., Maradonna, F., Olivotto, I., Gioacchini, G. & Carnevali, O. (2015). Lactobacillus rhamnosus lowers zebrafish lipid content by changing gut microbiota and host transcription of genes involved in lipid metabolism. *Scientific Reports*, 5(9336), 8–10. <https://doi.org/10.1038/srep09336>
- FAO. (2020). *Meat Market Review, Overview of global meat market developments in 2019*, April 2020. Rome. <http://www.fao.org/3/ca8819en/CA8819EN.pdf> (accessed on 01-09-2020)
- Farag, S. A., & El-Rayes, T. (2016). Research article effect of bee-pollen supplementation on performance, carcass traits and blood parameters of broiler chickens. *Asian Journal of Animal and Veterinary Advances*, 11(3), 168-77. DOI: [10.3923/ajava.2016.168.177](https://doi.org/10.3923/ajava.2016.168.177)
- Farahat, M. H., Abdallah, F. M., Ali, H. A., & Hernandez-Santana, A. (2017). Effect of dietary supplementation of grape seed extract on the growth performance, lipid profile, antioxidant status & immune response of broiler chickens. *Animal*, 11(5), 771–777. <https://doi.org/10.1017/S1751731116002251>
- Farooqui, M., Hassali, M. A., Shatar, A. K. A., Farooqui, M. A., Saleem, F., Haq, N. ul., & Othman, C. N. (2016). Use of complementary and alternative medicines among Malaysian cancer patients: A descriptive study. *Journal of Traditional and Complementary Medicine*, 6(4), 321–326. <https://doi.org/10.1016/j.jtcme.2014.12.008>

- Fasuyi, A. O., Dairo, F. A. S., & Adeniji, A. O. (2008). Tropical vegetable (*Amaranthus cruentus*) leaf meal as alternative protein supplement in broiler starter diets: Bionutritional evaluation. *Journal of Central European Agriculture*, 9(1), 23–33. [[Google Scholar](#)]
- Ferdous, M. F., Arefin, M. S., Rahman, M. M., Ripon, M. M. R., Rashid, M. H., Sultana, M. R., Hossain, M. T., Ahammad, M. U., & Rafiq, K. (2019). Beneficial effects of probiotic and phytobiotic as growth promoter alternative to antibiotic for safe broiler production. *Journal of Advanced Veterinary and Animal Research*, 6(3), 409. [doi: 10.5455/javar.2019.f361](https://doi.org/10.5455/javar.2019.f361)
- Finley, R. L., Collignon, P., Larsson, D. J., McEwen, S. A., Li, X. Z., Gaze, W. H., Reid-Smith, R., Timinouni, M., Graham, D. W., & Topp, E. (2013). The scourge of antibiotic resistance: the important role of the environment. *Clinical Infectious Diseases*, 57(5), 704-710. <https://doi.org/10.1093/cid/cit355>
- Filazi, A., Şireli, U. T., Pehlivanlar Önen, S., Çadırcı, Ö., & Aksoy, A. (2013). Comparative pharmacokinetics of gentamicin in laying hens. *Kafkas Üniversitesi Veteriner Fakültesi Dergisi* .19(3), 495-498 [DOI:10.9775/kvfd.2012.8138](https://doi.org/10.9775/kvfd.2012.8138).
- Fomentini, M., Haese, D., Kill, J. L., Sobreiro, R. P., Puppo, D. D., Haddade, I. R., Lima, A. L., & Saraiva, A. (2016). Prebiotic and antimicrobials on performance, carcass characteristics, and antibody production in broilers. *Ciência Rural*, 46(6), 1070-1075. <http://dx.doi.org/10.1590/0103-8478cr20150133>
- Foo, S. C., Yusoff, F. M., Ismail, M., Basri, M., Khong, N. M. H., Chan, K. W., & Yau, S. K. (2015). Efficient solvent extraction of antioxidant-rich extract from a tropical diatom, *Chaetoceros calcitrans* (Paulsen) Takano 1968. *Asian Pacific Journal of Tropical Biomedicine*, 5(10), 834–840. <https://doi.org/10.1016/j.apjtb.2015.06.003>
- Food and Drug Administration (FDA). (2012). The judicious use of medically important antimicrobial drugs in food-producing animals. *Guidance for Industry* #209, 1–26, <http://www.fda.gov/downloads/AnimalVeterinary/GuidanceComplianceEnforcement/GuidanceforIndustry/UCM216936.pdf> (Access on august 2020)
- Forte, C., Branciarri, R., Pacetti, D., Miraglia, D., Ranucci, D., Acuti, G., Balzano, M., Frega, N.G., & Trabalza-Marinucci, M. (2018). Dietary oregano (*Origanum vulgare* L.) aqueous extract improves oxidative stability and consumer acceptance of meat enriched with CLA and n-3 PUFA in broilers. *Poultry Science*, 97(5), 1774-1785. <https://doi.org/10.3382/ps/pex452>



- Founou, L. L., Founou, R. C., & Essack, S. Y. (2016). Antibiotic resistance in the food chain: a developing country-perspective. *Frontiers in Microbiology*, 7, 1881.  
<https://doi.org/10.3389/fmicb.2016.01881>
- Fritz, J. W., & Zuo, Y. (2007). Simultaneous determination of tetracycline, oxytetracycline, and 4-epitetracycline in milk by high-performance liquid chromatography. *Food Chemistry*, 105(3), 1297-1301.  
<https://doi.org/10.1016/j.foodchem.2007.03.047>
- Fujita K. I., Chavasiri, W., & Kubo, I. (2015). Anti-Salmonella Activity of Volatile Compounds of Vietnam Coriander. *Phytotherapy Research*, 29(7), 1081–1087. <https://doi.org/10.1002/ptr.5351>
- Gadde, U., Kim, W. H., Oh, S. T., & Lillehoj, H. S. (2017). Alternatives to antibiotics for maximizing growth performance and feed efficiency in poultry: A review. *Animal Health Research Reviews*, 18(1), 26–45.  
<https://doi.org/10.1017/S1466252316000207>
- Galvao, S. S., A. S. Monteiro, E. P. Siqueira, M. R. Bomfim, M. V. Dias-Souza, G. F. Ferreira, A. M. Denadai, A. R. Santos, V. Lucia Dos Santos, E. M. de Souza-Fagundes, E. S., Fernandes, E. S., & Monteiro-Neto, V. (2016). Annona glabra flavonoids act as antimicrobials by binding to Pseudomonas aeruginosa cell walls. *Frontiers in Microbiology*, 7(DEC), 1–9.  
<https://doi.org/10.3389/fmicb.2016.02053>
- Ganguly, S. (2015). A comprehensive review on physiological and nutritional properties of prebiotics as poultry feed supplement. *Octa Journal of Biosciences*, 3(1), 5–6. <https://doi.org/10.1017/1>
- Ganguly, S. (2017). *Herbal Antioxidant Agents and its Pharmacological and Medicinal Properties*. Lulu Press, Inc. Research maGma Group: Solapur, India, 2017, 15-21. ISBN 978-1-365-90767-8.
- Garcia-Martinez, O., De Luna-Bertos, E., Ramos-Torrecillas, J., Ruiz, C., Milia, E., Lorenzo, M.L., Jimenez, B., Sánchez-Ortiz, A., & Rivas, A. (2016). Phenolic compounds in extra virgin olive oil stimulate human osteoblastic cell proliferation. *PLoS ONE*, 11(3), 1–15.  
<https://doi.org/10.1371/journal.pone.0150045>
- Gaucher, M. L., Quessy, S., Letellier, A., Arsenault, J., & Boulianne, M. (2015). Impact of a drug-free program on broiler chicken growth performances, gut health, Clostridium perfringens and Campylobacter jejuni occurrences at the farm level. *Poultry science*, 94(8), 1791-1801.  
<https://doi.org/10.3382/ps/pev142>

- Gedikli, S., Ozkanlar, S., Gur, C., Sengul, E., & Gelen, V. (2017). Preventive effects of quercetin on liver damages in high-fat diet-induced obesity. *Journal of Histology & Histopathology*, 4, 7.  
<https://doi.org/10.7243/2055-091x-4-7>
- Gentile, D., Fornai, M., Colucci, R., Pellegrini, C., Tirotta, E., Benvenuti, L., Segnani, C., Ippolito, C., Duranti, E., Virdis, A., & Carpi, S. (2018). The flavonoid compound apigenin prevents colonic inflammation and motor dysfunctions associated with high fat diet-induced obesity. *PLoS One*, 13(4), 1–19.  
<https://doi.org/10.1371/journal.pone.0195502>
- George, O. S., Kaegon, S. G., & Igbokwe, A. A. (2015). Feed additive effects of graded levels of ginger (*Zingiber Officinale*) on serum metabolites of broilers. *IOSR Journal of Agriculture and Veterinary Science*, 8, 59-62.  
<https://doi.org/10.9790/2380-08325962>
- Ghasemi, H. A., Kasani, N., & Taherpour, K. (2014). Effects of black cumin seed (*Nigella sativa* L.), a probiotic, a prebiotic and a synbiotic on growth performance, immune response and blood characteristics of male broilers. *Livestock Science*, 164, 128-134.  
<https://doi.org/10.1016/j.livsci.2014.03.014>
- Ghasemzadeh, A., Jaafar, H. Z. E., Rahmat, A., & Ashkani, S. (2015). Secondary metabolites constituents and antioxidant, anticancer and antibacterial activities of *Etlingera elatior* (Jack) R.M.Sm grown in different locations of Malaysia. *BMC Complementary and Alternative Medicine*, 15(1), 1–10. <https://doi.org/10.1186/s12906-015-0838-6>
- Gholami-Ahangaran, M., Ahmadi-Dastgerdi, A., & Karimi-Dehkordi, M. (2020). Thymol and carvacrol; as antibiotic alternative in green healthy poultry production. *Plant Biotechnology Persa*, 2(1), 22-25. [[Google Scholar](#)]
- Giannenas, I., Bonos, E., Filliouis, G., Stylianaki, I., Kumar, P., Lazari, D., Christaki, E., & Florou-Paneri, P. (2019). Effect of a polyherbal or an arsenic-containing feed additive on growth performance of broiler chickens, intestinal microbiota, intestinal morphology, and lipid oxidation of breast and thigh meat. *Journal of Applied Poultry Research*, 28(1), 164-175.  
<https://doi.org/10.3382/japr/pfy059>
- Giannenas, I., Sidiropoulou, E., Bonos, E., Christaki, E., & Florou-Paneri, P. (2020). *The history of herbs, medicinal and aromatic plants, and their extracts. Feed Additives*. Elsevier Inc.  
<https://doi.org/10.1016/b978-0-12-814700-9.00001-7>
- Gibson, G. R., & Roberfroid, M. B. (1995). Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. *The Journal of nutrition*, 125(6), 1401-1412.

<https://doi.org/10.1093/jn/125.6.1401>

- Gilani, S. M. H., Zehra, S., Galani, S., & Ashraf, A. (2018). Effect of natural growth promoters on immunity, and biochemical and haematological parameters of broiler chickens. *Tropical Journal of Pharmaceutical Research*, 17(4), 627-633. DOI: [10.4314/tjpr.v17i4.9](https://doi.org/10.4314/tjpr.v17i4.9)
- Gibson, G. R., Hutkins, R., Sanders, M. E., Prescott, S. L., Reimer, R. A., Salminen, S. J., Scott, K., Stanton, C., Swanson, K.S., Cani, P.D., Verbeke Kristin & Reid, G. (2017). Expert consensus document: The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nature reviews Gastroenterology & hepatology*, 14(8), 491-502. <https://doi.org/10.1038/nrgastro.2017.75>
- Gole, M., Manwar, S. J., Chaudhary, S. P., Kawitkar, S. V., & Khose, K. K. (2020). The impact of feeding clove essential oils and organic acids on immunity , gut health and economics of broiler production. *Journal of Pharmacognosy and Phytochemistry*, 9(3), 1417–1422. [[Google Scholar](#)]
- Gonzalez Ronquillo, M., & Angeles Hernandez, J. C. (2017). Antibiotic and synthetic growth promoters in animal diets: Review of impact and analytical methods. *Food Control*, 72, 255–267. <https://doi.org/10.1016/j.foodcont.2016.03.001>
- Gopi, M., Karthik, K., Manjunathachar, H.V., Tamilmahan, P., Kesavan, M., Dashprakash, M., Balaraju, B. L., & Purushothaman, M. R. (2014). Essential oils a Feed Additive in Poultry Nutrition. *Advances in Animal and Veterinary Sciences*, 2(1), 2307–8316. [[Google Scholar](#)]
- Gor, M. C., Ismail, I., Mustapha, W. A. W., Zainal, Z., Noor, N. M., Othman, R., & Hussein, Z. A. M. (2011). Identification of cDNAs for jasmonic acid-responsive genes in *Polygonum minus* roots by suppression subtractive hybridization. *Acta physiologiae plantarum*, 33(2), 283-294. [[Google Scholar](#)]
- Gorniak, I., Bartoszewski, R., & Króliczewski, J. (2019). Comprehensive review of antimicrobial activities of plant flavonoids. *Phytochemistry Reviews*, 18(1), 241-272. <https://doi.org/10.1007/s11101-018-9591-z>
- Grashorn, M. A. (2010). Use of phytobiotics in broiler nutrition—an alternative to infeed antibiotics. *Journal of Animal and Feed Sciences*, 19(3), 338-347. DOI: <https://doi.org/10.22358/jafs/66297/2010>
- Grossman, T. H. (2016). Tetracycline antibiotics and resistance. *Cold Spring Harbor perspectives in medicine*, 6(4), a025387. [doi:10.1101/cshperspect.a025387](https://doi.org/10.1101/cshperspect.a025387)

- Guldiken, B., Ozkan, G., Catalkaya, G., Ceylan, F. D., Ekin Yalcinkaya, I., & Capanoglu, E. (2018). Phytochemicals of herbs and spices: Health versus toxicological effects. *Food and Chemical Toxicology*, 119(May), 37–49. <https://doi.org/10.1016/j.fct.2018.05.050>
- Guran, M., Şanlıtürk, G., Kerküklü, N. R., Altundağ, E. M., & Yalçın, A. S. (2019). Combined effects of quercetin and curcumin on anti-inflammatory and antimicrobial parameters in vitro. *European journal of pharmacology*, 859, 172486. <https://doi.org/10.1016/j.ejphar.2019.172486>
- Habib, M. A., Haque, M. A., Islam, M. S., & Liton, M. R. (2019). Effect of dietary Halquinol supplementation on the productive performances, carcass traits and blood profile of Sonali chicken. *Asian Journal of Medical and Biological Research*, 5(4), 316–323. <https://doi.org/10.3329/ajmbr.v5i4.45270>
- Hafeez, A., Mader, A., Boroogeni, F. G., Ruhnke, I., Röhe, I., Männer, K., & Zentek, J. (2014). Impact of thermal and organic acid treatment of feed on apparent ileal mineral absorption, tibial and liver mineral concentration, and tibia quality in broilers. *Poultry Science*, 93(7), 1754–1763. <https://doi.org/10.3382/ps.2013-03750>
- Hascik, P., Trembecká, L., Bobko, M., Kacániová, M., Bucko, O., Tkáčová, J., & Kunová, S. (2015). Effect of different dietary supplements on selected quality indicators of chicken meat. *Potravinárstvo*, 9(1), 427–434. <https://doi.org/10.5219/517>
- Hashemi, S. R., Zulkifli, I., Bejo, M. H., Farida, A., & Somchit, M. N. (2008). Acute toxicity study and phytochemical screening of selected herbal aqueous extract in broiler chickens. *International Journal of Pharmacology*, 4(5), 352–360. <https://doi.org/10.3923/ijp.2008.352.360>
- Hashmi, M. A., Khan, A., Farooq, U., & Khan, S. (2017). Alkaloids as Cyclooxygenase Inhibitors in Anticancer Drug Discovery. *Current Protein & Peptide Science*, 19(3), 292–301. <https://doi.org/10.2174/1389203718666170106103031>
- Haslan, H., Suhaimi, F. H., Thent, Z. C., & Das, S. (2015). The underlying mechanism of action for various medicinal properties of Piper betle (betel). *Clinical Therapeutics*, 166(5), 208–214. [doi: 10.7417/CT.2015.1880](https://doi.org/10.7417/CT.2015.1880)
- Hassali, M. A., Yann, H. R., Verma, A. K., Hussain, R., & Sivaraman, S. (2018). Antibiotic Use in Food Animals : Malaysia Overview. *Discipline of Social & Administrative Pharmacy. School of Pharmaceutical Sciences. Universiti Sains Malaysia*. Retrieved from [https://www.reactgroup.org/wpcontent/uploads/2018/11/Antibiotic\\_Use\\_in\\_Food\\_Animals\\_Malaysia\\_Overview\\_2018web](https://www.reactgroup.org/wpcontent/uploads/2018/11/Antibiotic_Use_in_Food_Animals_Malaysia_Overview_2018web).

- Hassan, H. M. A., Youssef, A. W., Ali, H. M., & Mohamed, M. A. (2015). Adding phytogetic material and/or organic acids to broiler diets: Effect on performance, nutrient digestibility and net profit. *Asian Journal of Poultry Science*, 9(2), 97–105. <https://doi.org/10.3923/ajpsaj.2015.97.105>
- Hassan, S. M., Khalaf, M. M., Sadek, S. A., & Abo-Youssef, A. M. (2017). Protective effects of apigenin and myricetin against cisplatin-induced nephrotoxicity in mice. *Pharmaceutical Biology*, 55(1), 766–774. <https://doi.org/10.1080/13880209.2016.1275704>
- Hassan, H. M. A., Samy, A., Youssef, A. W., & Mohamed, M. A. (2018). Using different feed additives as alternative to antibiotic growth promoter to improve growth performance and carcass traits of broilers. *International Journal of Poultry Science*, 17(6), 255–261. <https://doi.org/10.3923/ijps.2018.255.261>
- Hassim, N., Markom, M., Anuar, N., Dewi, K. H., Baharum, S. N., & Noor, N. M. (2015). Antioxidant and Antibacterial Assays on Polygonum minus Extracts: Different Extraction Methods. *International Journal of Chemical Engineering*, 2015, 1–10. <https://doi.org/10.1155/2015/826709>
- Hatab, M. H., Elsayed, M. A., & Ibrahim, N. S. (2016). Effect of some biological supplementation on productive performance, physiological and immunological response of layer chicks. *Journal of Radiation Research and Applied Sciences*, 9(2), 185–192. <https://doi.org/10.1016/j.jrras.2015.12.008>
- Hayashi, K., Fukushima, A., Hayashi-Nishino, M., & Nishino, K. (2014). Effect of methylglyoxal on multidrug-resistant *Pseudomonas aeruginosa*. *Frontiers in Microbiology*, 5(APR), 1–6. <https://doi.org/10.3389/fmicb.2014.00180>
- He, J., Dong, L., Xu, W., Bai, K., Lu, C., Wu, Y., Huang, Q., Zhang, L., & Wang, T. (2015). Dietary tributyrin supplementation attenuates insulin resistance and abnormal lipid metabolism in suckling piglets with intrauterine growth retardation. *PLoS ONE*, 10(8), 1–14. <https://doi.org/10.1371/journal.pone.0136848>
- Hedman, H. D., Vasco, K. A., & Zhang, L. (2020). A review of antimicrobial resistance in poultry farming within low-resource settings. *Animals*, 10(8), 1–39. <https://doi.org/10.3390/ani10081264>
- Henchion, M., Hayes, M., Mullen, A. M., Fenelon, M., & Tiwari, B. (2017). Future protein supply and demand: strategies and factors influencing a sustainable equilibrium. *Foods*, 6(7), 53. <https://doi.org/10.3390/foods6070053>
- Hidanah, S., Sabdongrum, E. K., Wahjuni, R. S., & Chusniati, S. (2018). Effects of meniran (*Phyllanthus niruri* L.) administration on leukocyte profile of broiler chickens infected with *Mycoplasma gallisepticum*. *Veterinary*

- World*, 11(6), 834–839. <https://doi.org/10.14202/vetworld.2018.834-839>
- Honikel, K. O. (1998). Reference methods for the assessment of physical characteristics of meat. *Meat Science*, 49(4), 447–457. [https://doi.org/10.1016/S0309-1740\(98\)00034-5](https://doi.org/10.1016/S0309-1740(98)00034-5)
- Hossain, M. F., Khairunnesa, M., & Das, S. C. (2015). Use of non-antibiotic growth promoter Grow Power in commercial broiler diet. *Bangladesh Journal of Animal Science*, 44(1), 33-39. DOI: <https://doi.org/10.3329/bjas.v44i1.23139>
- Hossain, M. A., & Awad, E. A. (2018). The Efficacy of Plant based Diets on Growth Potential, Energy Utilization, Nutrient Digestibility, Leg Bone Development and Litter Quality of Meat Chickens. *Iranian Journal of Applied Animal Science*, 8(1), 1-7. [[Google Scholar](#)]
- Hu, Y., Wang, Y., Li, A., Wang, Z., Zhang, X., Yun, T., Qiu, L., & Yin, Y. (2016). Effects of fermented rapeseed meal on antioxidant functions, serum biochemical parameters and intestinal morphology in broilers. *Food and Agricultural Immunology*, 27(2), 182–193. <https://doi.org/10.1080/09540105.2015.1079592>
- Hu, Q., Zhou, M., & Wei, S. (2018). Progress on the antimicrobial activity research of clove oil and eugenol in the food antiseptis field. *Journal of food science*, 83(6), 1476-1483. <https://doi.org/10.1111/1750-3841.14180>
- Huang, X., Liu, Y., Lu, Y., & Ma, C. (2015). Anti-inflammatory effects of eugenol on lipopolysaccharide-induced inflammatory reaction in acute lung injury via regulating inflammation and redox status. *International immune pharmacology*, 26(1), 265-271. <https://doi.org/10.1016/j.intimp.2015.03.026>
- Huang, J., Chen, L., Xue, B., Liu, Q., Ou, S., Wang, Y., & Peng, X. (2016). Different flavonoids can shape unique gut microbiota profile in vitro. *Journal of food science*, 81(9), H2273-H2279. doi: [10.1111/1750-3841.13411](https://doi.org/10.1111/1750-3841.13411)
- Huang, Q., Liu, X., Zhao, G., Hu, T., & Wang, Y. (2018). Potential and challenges of tannins as an alternative to in-feed antibiotics for farm animal production. *Animal Nutrition*, 4(2), 137–150. <https://doi.org/10.1016/j.aninu.2017.09.004>
- Humam, A. M., Loh, T. C., Foo, H. L., Samsudin, A. A., Mustapha, N. M., Zulkifli, I., & Izuddin, W. I. (2019). Effects of feeding different postbiotics produced by *Lactobacillus plantarum* on growth performance, carcass yield, intestinal morphology, gut microbiota composition, immune status, and growth gene expression in broilers under heat stress. *Animals*, 9(9), 644. <https://doi.org/10.3390/ani9090644>

- Hussein, E. O., Ahmed, S. H., Abudabos, A. M., Aljumaah, M. R., Alkhulaili, M. M., Nassan, M. A., Suliman, G. M., Naiel, M. A., & Swelum, A. A. (2020). Effect of Antibiotic, Phytobiotic and Probiotic Supplementation on Growth, Blood Indices and Intestine Health in Broiler Chicks Challenged with *Clostridium perfringens*. *Animals*, *10*(3), 507. <https://doi.org/10.3390/ani10030507>
- Imelda, F., Faridah, D. N., & Kusumaningrum, H. D. (2014). Bacterial inhibition and cell leakage by extract of *Polygonum minus* Huds. leaves. *International Food Research Journal*, *21*(2). [[Google Scholar](#)]
- Interagency Coordination Group on Antimicrobial Resistance. Antimicrobial resistance: national action plans. 2017. <https://www.un.org/sg/en/content/sg/personnel-appointments/2017-03-17/interagency-coordination-group-antimicrobial-resistance>  
Accessed on (28-01-2022)
- Jaganathan, S. K., & Supriyanto, E. (2012). Antiproliferative and molecular mechanism of eugenol-induced apoptosis in cancer cells. *Molecules*, *17*(6), 6290-6304. <https://doi.org/10.3390/molecules17066290>
- Jaisinghani, R. N. (2017). Antibacterial properties of quercetin. *Microbiology Research*, *8*;6877(1), 13–14. <https://doi.org/10.4081/mr.2017.6877>
- Jaiswal, S. G., Patel, M., Saxena, D. K., & Naik, S. N. (2014). Antioxidant Properties of Piper Betel ( *L* ) Leaf Extracts from Six Different Geographical Domain of India. *Journal of Bioresource Engineering and Technology*, *2*(2), 12–20. [[Google Scholar](#)]
- Jamroz, D., Orda, J., Kamel, C., Wiliczkiwicz, A., Wertelecki, T., & Skorupińska, J. (2003). The influence of phytogetic extracts on performance, nutrient digestibility, carcass characteristics, and gut microbial status in broiler chickens. *Journal of Animal and Feed Sciences*, *12*(3), 583–596. <https://doi.org/10.22358/jafs/67752/2003>
- Janabi, A. H. W., Kamboh, A. A., Saeed, M., Xiaoyu, L., BiBi, J., Majeed, F., Naveed, M., Mughal, M. J., Korejo, N. A., Kamboh, R., & Lv, H. (2020). Flavonoid-rich foods (FRF): A promising nutraceutical approach against lifespan-shortening diseases. *Iranian Journal of Basic Medical Sciences*, *23*(2), 140–153. <https://doi.org/10.22038/IJBMS.2019.35125.8353>
- Jarriyawattanachaikul, W., Chaveerach, P., & Chokesajjawatee, N. (2016). Antimicrobial activity of Thai-herbal plants against food-borne pathogens *E. coli*, *S. aureus* and *C. jejuni*. *Agriculture and Agricultural Science Procedia*, *11*, 20-24. <https://doi.org/10.1016/j.aaspro.2016.12.004>

- Jiang, S., Jiang, Z., Zhou, G., Lin, Y., & Zheng, C. (2014). Effects of Dietary Isoflavone Supplementation on Meat Quality and Oxidative Stability During Storage in Lingnan Yellow Broilers. *Journal of Integrative Agriculture*, 13(2), 387–393.  
[https://doi.org/10.1016/S2095-3119\(13\)60386-X](https://doi.org/10.1016/S2095-3119(13)60386-X)
- Józefiak, D., Kaczmarek, S., & Rutkowski, A. (2008). A note on the effects of selected prebiotics on the performance and ileal microbiota of broiler chickens. *Journal of Animal and Feed Sciences*, 17(3), 392-397.  
[\[Google Scholar\]](#)
- Judice, J. P. M., Bertechini, A. G., Muniz, J. Á., Rodrigues, P. B., & Fassani, J. E. (2002). Cation-anionic balance of rations and food management for layers of second cycle. *Agrotechnical Science*, 26(3), 598-609.
- Julian, R. J. (2005). Production and growth related disorders and other metabolic diseases of poultry - A review. *Veterinary Journal*, 169(3), 350–369.  
<https://doi.org/10.1016/j.tvjl.2004.04.015>
- Kadir, A., Sher, S., Siddiqui, R. A., & Mirza, T. (2020). Nephroprotective role of eugenol against cisplatin-induced acute kidney injury in mice. *Pakistan Journal of Pharmaceutical Sciences*, 33(3), 1281–1287.  
<https://doi.org/doi.org/10.36721/PJPS.2020.33.3.SUP.1281-1287.1>
- Kalantari, H., Foruozandeh, H., Khodayar, M. J., Siahpoosh, A., Saki, N., & Kheradmand, P. (2018). Antioxidant and hepatoprotective effects of Capparis spinosa L. fractions and Quercetin on tert-butyl hydroperoxide-induced acute liver damage in mice. *Journal of traditional and complementary medicine*, 8(1), 120-127. <https://doi.org/10.1016/j.jtcme.2017.04.010>
- Kamath, B. R., & Sabeena, K. (2018). In vitro study on antioxidant activity of methanolic leaf extract of piper betle linn. *Journal of Evolution of Medical and Dental Sciences*, 7(24), 2865–2870.  
<https://doi.org/10.14260/jemds/2018/646>
- Kamboh, A. A., Arain, M. A., Mughal, M. J., Zaman, A., Arain, Z. M., & Soomro, A. H. (2015). Flavonoids: health promoting phytochemicals for animal production-a review. *Journal of Animal Health and Production*, 3(1), 6-13.  
[\[Google Scholar\]](#)
- Kamboh, A. A., Hang, S. Q., Khan, M. A., & Zhu, W. Y. (2016). In vivo immunomodulatory effects of plant flavonoids in lipopolysaccharide-challenged broilers. *Animal*, 10(10), 1619-1625.  
[DOI:10.1017/S1751731116000562](https://doi.org/10.1017/S1751731116000562)
- Kamboh, A. A., Khan, M. A., Kaka, U., Awad, E. A., Memon, A. M., Saeed, M., Korejo, N. A., Bakhethgul, M., & Kumar, C. (2018). Effect of dietary



- supplementation of phytochemicals on immunity and haematology of growing broiler chickens. *Italian Journal of Animal Science*, 17(4), 1038–1043. <https://doi.org/10.1080/1828051X.2018.1438854>
- Kandepu, N., Kodaganur, S. C., Mantri, A. P., & Saha, S. (2012). RP-HPLC method for quantitative estimation of Halquinol in pharmaceutical dosage forms. *Eurasian Journal of Analytical Chemistry*, 7(1), 7–12. [\[Google Scholar\]](#)
- Karak, S., Das, S., Biswas, M., Choudhury, A., Dutta, M., Chaudhury, K., & De, B. (2019). Phytochemical composition,  $\beta$ -glucuronidase inhibition, and antioxidant properties of two fractions of Piper betle leaf aqueous extract. *Journal of Food Biochemistry*, 43(12), 1–12. <https://doi.org/10.1111/jfbc.13048>
- Karami, M., Karimi, A., Sadeghi, A., Zentek, J., & Goodarzi Borojeni, F. (2020). Evaluation of interactive effects of phytase and benzoic acid supplementation on performance, nutrients digestibility, tibia mineralisation, gut morphology and serum traits in male broiler chickens. *Italian Journal of Animal Science*, 19(1), 1428–1438. <https://doi.org/10.1080/1828051X.2020.1846468>
- Karangiya, V. K., Savsani, H. H., Patil, S. S., Garg, D. D., Murthy, K. S., Ribadiya, N. K., & Vekariya, S. J. (2016). Effect of dietary supplementation of garlic, ginger and their combination on feed intake, growth performance and economics in commercial broilers. *Veterinary World*, 9(3), 245. [doi: 10.14202/vetworld.2016.245-250](https://doi.org/10.14202/vetworld.2016.245-250)
- Karaskova, K., Suchy, P., & Straková, E. (2015). Current use of phytogetic feed additives in animal nutrition: A review. *Czech Journal of Animal Science*, 60(12), 521–530. <https://doi.org/10.17221/8594-CJAS>
- Karthivashan, G., Arulselvan, P., Alimon, A. R., Safinar Ismail, I., & Fakurazi, S. (2015). Competing role of bioactive constituents in Moringa oleifera extract and conventional nutrition feed on the performance of cobb 500 broilers. *BioMed Research International*, 2015, 1–11. <https://doi.org/10.1155/2015/970398>
- Kaschubek, T., Mayer, E., Rzesnik, S., Grenier, B., Bachinger, D., Schieder, C., König, J., & Teichmann, K. (2018). Effects of phytogetic feed additives on cellular oxidative stress and inflammatory reactions in intestinal porcine epithelial cells. *Journal of Animal Science*, 96(9), 3657–3669. <https://doi.org/10.1093/jas/sky263>
- Kasote, D. M., Katyare, S. S., Hegde, M. V., & Bae, H. (2015). Significance of antioxidant potential of plants and its relevance to therapeutic applications. *International Journal of Biological Sciences*, 11(8), 982–991. <https://doi.org/10.7150/ijbs.12096>

- Kearney, J. (2010). Food consumption trends and drivers. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 365(1554), 2793-2807. <https://doi.org/10.1098/rstb.2010.0149>
- Khalaji, S., Zaghari, M., Hatami, K. H., Hedari-Dastjerdi, S., Lotfi, L., & Nazarian, H. (2011). Black cumin seeds, Artemisia leaves (*Artemisia sieberi*), and Camellia L. plant extract as phytogetic products in broiler diets and their effects on performance, blood constituents, immunity, and cecal microbial population. *Poultry Science*, 90(11), 2500-2510. <https://doi.org/10.3382/ps.2011-01393>
- Khalil, A. A., Rahman, U. ur, Khan, M. R., Sahar, A., Mehmood, T., & Khan, M. (2017). Essential oil eugenol: sources, extraction techniques and nutraceutical perspectives. *RSC Advances*, 7(52), 32669-32681. [DOI: 10.1039/C7RA04803C](https://doi.org/10.1039/C7RA04803C)
- Khan, R. U., Naz, S., Javdani, M., Nikousefat, Z., Selvaggi, M., Tufarelli, V., & Laudadio, V. (2012). The use of Turmeric (*Curcuma longa*) in poultry feed. *World's Poultry Science Journal*, 68(1), 97-103. <https://doi.org/10.1017/S0043933912000104>
- Khan, R. U., Naz, S., Nikousefat, Z., Tufarelli, V., Javdani, M., Qureshi, M. S., & Laudadio, V. (2012). Potential applications of ginger (*Zingiber officinale*) in poultry diets. *World's Poultry Science Journal*, 68(2), 245-252. <https://doi.org/10.1017/S004393391200030X>
- Khan, N., Choi, J. Y., Nho, E. Y., Jamila, N., Habte, G., Hong, J. H., Hwang, I. M., & Kim, K. S. (2014). Determination of minor and trace elements in aromatic spices by micro-wave assisted digestion and inductively coupled plasma-mass spectrometry. *Food Chemistry*, 158, 200-206. <https://doi.org/10.1016/j.foodchem.2014.02.103>
- Khan, I., Zaneb, H., Masood, S., Yousaf, M. S., Rehman, H. F., & Rehman, H. (2017). Effect of Moringa oleifera leaf powder supplementation on growth performance and intestinal morphology in broiler chickens. *Journal of Animal Physiology and Animal Nutrition*, 101, 114-121. <https://doi.org/10.1111/jpn.12634>
- Khattak, F., Ronchi, A., Castelli, P., & Sparks, N. (2014). Effects of natural blend of essential oil on growth performance, blood biochemistry, cecal morphology, and carcass quality of broiler chickens. *Poultry Science*, 93(1), 132-137. <https://doi.org/10.3382/ps.2013-03387>
- Khatun, J., Loh, T. C., Akit, H., Foo, H. L., & Mohamad, R. (2018). Influence of different sources of oil on performance, meat quality, gut morphology, ileal digestibility and serum lipid profile in broilers. *Journal of Applied Animal Research*, 46(1), 479-485. <https://doi.org/10.1080/09712119.2017.1337580>

- Kim, D. K., Lillehoj, H. S., Lee, S. H., Jang, S. I., Park, M. S., Min, W., Lillehoj, E. P., & Bravo, D. (2013). Immune effects of dietary anethole on *Eimeria acervulina* infection. *Poultry Science*, 92(10), 2625-2634.  
<https://doi.org/10.3382/ps.2013-03092>
- Kim, D., Hong, E., Kim, J., Bang, H., Choi, J., & Ji, S. (2015). Effects of dietary quercetin on growth performance, blood biochemical parameter, immunoglobulin and blood antioxidant activity in broiler chickens. *Korean Journal of Poultry Sciences*, 42(1), 33–40. [[Google Scholar](#)]
- Kirubakaran, A., Moorthy, M., Chitra, R., & Prabakar, G. (2016). Influence of combinations of fenugreek, garlic, and black pepper powder on production traits of the broilers. *Veterinary World*, 9(5), 470.  
[doi: 10.14202/vetworld.2016.470-474](https://doi.org/10.14202/vetworld.2016.470-474)
- Klaric, I., Pavic, M., Miskulin, I., Blazicevic, V., Domic, A., & Miskulin, M. (2018). Influence of dietary supplementation of propolis and bee pollen on liver pathology in broiler chickens. *Animals*, 8(4), 54.  
<https://doi.org/10.3390/ani8040054>
- Kleczek, K., Majewska, K., & Makowski, W. (2012). The effect of diet supplementation with propolis and bee pollen on the physicochemical properties and strength of tibial bones in broiler chickens. *Archiv Tierzucht*, 55(1), 97–103. <https://doi.org/10.5194/aab-55-97-2012>
- Krauze, M., Abramowicz, K., & Ognik, K. (2020). The effect of addition of probiotic bacteria (*Bacillus subtilis* or *Enterococcus faecium*) or phytobiotic containing cinnamon oil to drinking water on the health and performance of broiler chickens. *Annals of Animal Science*, 20(1), 191-205.  
[DOI:10.2478/aoas-2019-0059](https://doi.org/10.2478/aoas-2019-0059)
- Krauze, M. (2021). Phytobiotics, a Natural Growth Promoter for Poultry. In *Advanced Studies in the 21st Century Animal Nutrition*. *IntechOpen*.  
[DOI: http://dx.doi.org/10.5772/intechopen.99030](https://doi.org/10.5772/intechopen.99030)
- Kroliczewska, B., Miśta, D., Króliczewski, J., Zawadzki, W., Kubaszewski, R., Wincewicz, E., Żuk, M. & Szopa, J. (2017). A new genotype of flax (*Linum usitatissimum* L.) with decreased susceptibility to fat oxidation: consequences to hematological and profiles of blood indices. *Journal of the Science of Food and Agriculture*, 97(1), 165–171.  
<https://doi.org/10.1002/jsfa.7705>
- Kudva, A. K., Rao, S., Rao, P., Periera, R., Bhandari, G., Mathew, J. M., Ashwini, K., Pais, M. L., Swamy, M. K., & Baliga, M. S. (2018). Piper betle Linn. in Cancer: Past, Present, and Future. In *Anticancer Plants: Properties and Application* (pp. 327-347). Springer, Singapore.  
[https://doi.org/10.1007/978-981-10-8548-2\\_14](https://doi.org/10.1007/978-981-10-8548-2_14)
- Kulshreshtha, G., Rathgeber, B., Stratton, G., Thomas, N., Evans, F., Critchley, A.,

- Hafting, J., & Prithiviraj, B. (2014). Feed supplementation with red seaweeds, *Chondrus crispus* and *Sarcodiotheca gaudichaudii*, affects performance, egg quality, and gut microbiota of layer hens. *Poultry Science*, 93(12), 2991–3001. <https://doi.org/10.3382/ps.2014-04200>
- Kumar, M., Kumar, V., Roy, D., Kushwaha, R., & Vaswani, S. (2014). Application of Herbal Feed Additives in Animal Nutrition - A Review. *International Journal of Livestock Research*, 4(9), 1–8. <https://doi.org/10.5455/ijlr.20141205105218>
- Kumar, N., & Chaiyasut, C. (2017). Health Promotion Potential of Vegetables Cultivated in Northern Thailand: A Preliminary Screening of Tannin and Flavonoid Contents, 5 $\alpha$ -Reductase Inhibition, Astringent Activity, and Antioxidant Activities. *Journal of Evidence-Based Complementary and Alternative Medicine*, 22(4), 573–579. <https://doi.org/10.1177/2156587216686689>
- Kumar, S., Chen, C., Indugu, N., Werlang, G. O., Singh, M., Kim, W. K., & Thippareddi, H. (2018). Effect of antibiotic withdrawal in feed on chicken gut microbial dynamics, immunity, growth performance and prevalence of foodborne pathogens. *PLoS One*, 13(2), e0192450. <https://doi.org/10.1371/journal.pone.0192450>
- Kwiatkowska, K., Kwiecień, M., & Winiarska-Mieczan, A. (2017). Fast-growing chickens fed with lucerne protein-xanthophyll concentrate: Growth performance, slaughter yield and bone quality. *Journal of Animal and Feed Sciences*, 26(2), 131–140. <https://doi.org/10.22358/jafs/70194/2017>
- Kyuchukova, R. (2020). Antibiotic residues and human health hazard-review. *Bulgarian Journal of Agricultural Science*, 26(3), 664-668. [\[Google scholar\]](#)
- Laptev, G. Y., Filippova, V. A., Kochish, I. I., Yildirim, E. A., Ilina, L. A., Dubrovin, A. V., Brazhnik, E. A., Novikova, N. I., Novikova, O. B., Dmitrieva, M. E., & Romanov, M. N. (2019). Examination of the expression of immunity genes and bacterial profiles in the caecum of growing chickens infected with *Salmonella* Enteritidis and fed a phytobiotic. *Animals*, 9(9), 1–25. <https://doi.org/10.3390/ani9090615>
- Laxminarayan, R., Duse, A., Wattal, C., Zaidi, A. K., Wertheim, H. F., Sumpradit, N., Vlieghe, E., Hara, G. L., Gould, I. M., Goossens, H., & Cars, O. (2013). Antibiotic resistance the need for global solutions. *The Lancet infectious diseases*, 13(12), 1057-1098. [https://doi.org/10.1016/S1473-3099\(13\)70318-9](https://doi.org/10.1016/S1473-3099(13)70318-9)
- Lee, D. N., Lyu, S. R., Wang, R. C., Weng, C. F., & Chen, B. J. (2011). Exhibit differential functions of various antibiotic growth promoters in broiler

growth, immune response and gastrointestinal physiology. *International Journal of Poultry Science*, 10(3), 216-220. [[Google Scholar](#)]

- Lei, Y., Wang, K., Deng, L., Chen, Y., Nice, E. C., & Huang, C. (2015). Redox Regulation of Inflammation: Old Elements, a New Story. *Medicinal Research Reviews*, 35(2), 306–340. <https://doi.org/10.1002/med>
- Li, H. L., Zhao, P. Y., Lei, Y., Hossain, M. M., & Kim, I. H. (2015). Phytoncide, phytogetic feed additive as an alternative to conventional antibiotics, improved growth performance and decreased excreta gas emission without adverse effect on meat quality in broiler chickens. *Livestock Science*, 181, 1–6. <https://doi.org/10.1016/j.livsci.2015.10.001>
- Li, Y., Xu, Q., Huang, Z., Lv, L., Liu, X., Yin, C., Yan, H., & Yuan, J. (2016). Effect of *Bacillus subtilis* CGMCC 1.1086 on the growth performance and intestinal microbiota of broilers. *Journal of Applied Microbiology*, 120(1), 195–204. <https://doi.org/10.1111/jam.12972>
- Liao, X., Wen, Q., Zhang, L., LU, L., Zhang, L., & Luo, X. (2018). Effect of dietary supplementation with flavonoid from *Scutellaria baicalensis* Georgi on growth performance, meat quality and antioxidative ability of broilers. *Journal of Integrative Agriculture*, 17(5), 1165–1170. [https://doi.org/10.1016/S2095-3119\(17\)61803-3](https://doi.org/10.1016/S2095-3119(17)61803-3)
- Liaquat, S., Mahmood, S., Ahmad, S., Kamran, Z., & Koutoulis, K. C. (2016). Replacement of canola meal with *Moringa oleifera* leaf powder affects performance and immune response in broilers. *Journal of Applied Poultry Research*, 25(3), 352–358. <https://doi.org/10.3382/japr/pfw018>
- Lillehoj, H., Liu, Y., Calsamiglia, S., Fernandez-Miyakawa, M.E., Chi, F., Cravens, R.L., Oh, S., & Gay, C. G. (2018). Phytochemicals as antibiotic alternatives to promote growth and enhance host health. *Veterinary Research*, 49(1), 1–18. <https://doi.org/10.1186/s13567-018-0562-6>
- Lin, S. Y., Wang, Y. Y., Chen, W. Y., Chuang, Y. H., Pan, P. H., & Chen, C. J. (2014). Beneficial effect of quercetin on cholestatic liver injury. *The Journal of Nutritional Biochemistry*, 25(11), 1183-1195. <https://doi.org/10.1016/j.jnutbio.2014.06.003>
- Linden, J. (2013). Global Poultry trends 2013: Asia consumes 40 Percent of world's chicken. The Poultry site. (accessed on 25 October 2020) <https://www.thepoultrysite.com/articles/global-poultry-trends-2013-asia-consumes-40-per-cent-of-worlds-chicken>
- Lister, I. N. E., Ginting, C. N., Girsang, E., Amansyah, A., Chiuman, L., Yanti, N. L. NL, Rizal R, and Widowati, W. (2019). Hepatoprotective effect of Eugenol on Acetaminophen-Induced Hepatotoxicity in HepG2 cells. *Journal of Physics: Conference Series*, 1374(1), 1–7.

<https://doi.org/10.1088/1742-6596/1374/1/012009>

- Liu, X. L., Hao, Y. Q., Jin, L., Xu, Z. J., McAllister, T. A., & Wang, Y. (2013). Anti-Escherichia coli O157:H7 properties of purple prairie clover and sainfoin condensed tannins. *Molecules*, *18*(2), 2183–2199.  
<https://doi.org/10.3390/molecules18022183>
- Liu, Q. W., Feng, J. H., Chao, Z., Chen, Y., Wei, L. M., Wang, F., Sun, R. P., & Zhang, M. H. (2016). The influences of ambient temperature and crude protein levels on performance and serum biochemical parameters in broilers. *Journal of Animal Physiology and Animal Nutrition*, *100*(2), 301–308.  
<https://doi.org/10.1111/jpn.12368>
- Liu, J., & Mao, Y. (2019). Eugenol attenuates concanavalin A-induced hepatitis through modulation of cytokine levels and inhibition of mitochondrial oxidative stress. *Archives of Biological Sciences*, *71*(2), 339–346.  
<https://doi.org/10.2298/ABS190121016L>
- Lokaewmanee, K., & Promdee, P. (2018). Mao pomace on carcass and meat quality of broiler. *International Journal of Poultry Science*, *17*(5), 221–228.  
<https://doi.org/10.3923/ijps.2018.221.228>
- Lomiwes, D., Farouk, M. M., Wu, G., & Young, O. A. (2014). The development of meat tenderness is likely to be compartmentalised by ultimate pH. *Meat Science*, *96*(1), 646–651. <https://doi.org/10.1016/j.meatsci.2013.08.022>
- Lu, W., Wang, J., Zhang, H. J., Wu, S. G., & Qi, G. H. (2016). Evaluation of *Moringa oleifera* leaf in laying hens: Effects on laying performance, egg quality, plasma biochemistry and organ histopathological indices. *Italian Journal of Animal Science*, *15*(4), 658–665.  
<https://doi.org/10.1080/1828051X.2016.1249967>
- Lu, H., Wu, L., Liu, L., Ruan, Q., Zhang, X., Hong, W., Wu, S., Jin, G., & Bai, Y. (2018). Quercetin ameliorates kidney injury and fibrosis by modulating M1/M2 macrophage polarization. *Biochemical Pharmacology*, *154*(March), 203–212. <https://doi.org/10.1016/j.bcp.2018.05.007>
- Ly, Z., Xing, K., Li, G., Liu, D., & Guo, Y. (2018). Dietary genistein alleviates lipid metabolism disorder and inflammatory response in laying hens with fatty liver syndrome. *Frontiers in Physiology*, *9*, 1493.  
<https://doi.org/10.3389/fphys.2018.01493>
- Ma, J. Q., Li, Z., Xie, W. R., Liu, C. M., & Liu, S. S. (2015). Quercetin protects mouse liver against CCl4-induced inflammation by the TLR2/4 and MAPK/NF- $\kappa$ B pathway. *International Immunopharmacology*, *28*(1), 531–539.  
<https://doi.org/10.1016/j.intimp.2015.06.036>

- Ma, J. L., Zhao, L. H., Sun, D. D., Zhang, J., Guo, Y. P., Zhang, Z. Q., Ma, Q. G., Ji, C., & Zhao, L. H. (2020). Effects of Dietary Supplementation of Recombinant Plectasin on Growth Performance, Intestinal Health and Innate Immunity Response in Broilers. *Probiotics and Antimicrobial Proteins*, 12(1), 214–223. <https://doi.org/10.1007/s12602-019-9515-2>
- Madhumita, M., Guha, P., & Nag, A. (2020). Bio-actives of betel leaf (Piper betle L.): A comprehensive review on extraction, isolation, characterization, and biological activity. *Phytotherapy Research*, 34(10), 2609–2627. <https://doi.org/10.1002/ptr.6715>
- Mahayothee, B., Koomyart, I., Khuwijitjaru, P., Siriwongwilaichat, P., Nagle, M., & Müller, J. (2016). Phenolic Compounds, Antioxidant Activity, and Medium Chain Fatty Acids Profiles of Coconut Water and Meat at Different Maturity Stages. *International Journal of Food Properties*, 19(9), 2041–2051. <https://doi.org/10.1080/10942912.2015.1099042>
- Mahboubi, A., Asgarpanah, J., Sadaghiyani, P. N., & Faizi, M. (2015). Total phenolic and flavonoid content and antibacterial activity of *Punica granatum* L. var. *pleniflora* flowers (Golnar) against bacterial strains causing foodborne diseases. *BMC Complementary and Alternative Medicine*, 15(1), 1–7. <https://doi.org/10.1186/s12906-015-0887-x>
- Mainente, F., Menin, A., Alberton, A., Zoccatelli, G., & Rizzi, C. (2019). Evaluation of the sensory and physical properties of meat and fish derivatives containing grape pomace powders. *International Journal of Food Science and Technology*, 54(4), 952–958. <https://doi.org/10.1111/ijfs.13850>
- Mak, K. K., Kamal, M., Ayuba, S., Sakirolla, R., Kang, Y. B., Mohandas, K., Balijepalli, M., Ahmad, S., & Pichika, M. (2019). A comprehensive review on eugenol's antimicrobial properties and industry applications: A transformation from ethnomedicine to industry. *Pharmacognosy Reviews*, 13(25), 1-9. DOI: [10.4103/phrev.phrev\\_46\\_18](https://doi.org/10.4103/phrev.phrev_46_18)
- Makwana, S. J., and Jadeja, B.A. (2016). Exploration of bioactive compounds in *Operculina Turpethum* L. *Journal of Pharmacognosy and Phytochemistry*, 5(5), 209. [[Google Scholar](#)]
- Malahubban, M., & Ab Aziz, Z. F. (2016). Serum biochemical properties and liver morphology of broiler chicken as affected by feeding Misai kucing (*Orthosiphon stamineus*) as a supplementary diet. *Research in Biotechnology*, 7. [[Google Scholar](#)]
- Manafi, M. (2015). Comparison study of a natural non-antibiotic growth promoter and a commercial probiotic on growth performance, immune response and biochemical parameters of broiler chicks. *The Journal of Poultry Science*, 52(4), 274-281. <https://doi.org/10.2141/jpsa.0150027>

- Mancabelli, L., Ferrario, C., Milani, C., Mangifesta, M., Turrone, F., Duranti, S., Lugli, G. A., Viappiani, A., Ossiprandi, M. C., Van Sinderen, D. & Ventura, M. (2016). Insights into the biodiversity of the gut microbiota of broiler chickens. *Environmental Microbiology*, 18(12), 4727–4738. <https://doi.org/10.1111/1462-2920.13363>
- Manjaniq, A., Wihandoyo, W., & Dono, N. D. (2017). The Effect of Dietary Violet Roselle Flower and Moringa Leaves Meal Supplementation on Blood Profile of Broiler Chickens. In *International Seminar on Tropical Animal Production (ISTAP)* (pp. 251-255).
- Mansoori, B., Rogiewicz, A., & Slominski, B. A. (2015). The effect of canola meal tannins on the intestinal absorption capacity of broilers using a D-xylose test. *Journal of Animal Physiology and Animal Nutrition*, 99(6), 1084–1093. <https://doi.org/10.1111/jpn.12320>
- Marchese, A., Barbieri, R., Coppo, E., Orhan, I. E., Daglia, M., Nabavi, S. F., Izadi, M., Abdollahi, M., Nabavi, S. M., & Ajami, M. (2017). Antimicrobial activity of eugenol and essential oils containing eugenol: A mechanistic viewpoint. *Critical Reviews in Microbiology*, 43(6), 668-689. <https://doi.org/10.1080/1040841X.2017.1295225>
- Marchini, C. F. P., Silva, P. L., Nascimento, M. R. B. M., Beletti, M. E., Silva, N. M., & Guimarães, E. C. (2011). Body weight, intestinal morphometry and cell proliferation of broiler chickens submitted to cyclic heat stress. *International Journal of Poultry Science*, 10(6), 455-460. [[Google Scholar](#)]
- Maron, D. F., Smith, T. J., & Nachman, K. E. (2013). Restrictions on antimicrobial use in food animal production: an international regulatory and economic survey. *Globalization and Health*, 9(1), 48. <https://doi.org/10.1186/1744-8603-9-48>
- Mashayekhi, H., Mazhari, M., & Esmaeilipour, O. (2018). Eucalyptus leaves powder, antibiotic and probiotic addition to broiler diets: Effect on growth performance, immune response, blood components and carcass traits. *Animal*, 12(10), 2049–2055. <https://doi.org/10.1017/S1751731117003731>
- Massele, A., Tiroyakgosi, C., Matome, M., Desta, A., Muller, A., Paramadhas, B. D. A., Malone, B., Kurusa, G., Didimalang, T., Moyo, M., & Godman, B. (2017). Research activities to improve the utilization of antibiotics in Africa. *Expert review of pharmacoeconomics & outcomes research*, 17(1), 1-4. [DOI: 10.1586/14737167.2016.1164040](https://doi.org/10.1586/14737167.2016.1164040)
- Mcallister, T. A., Martinez, T., Hee, D. B., Muir, A. D., Yanke, L. J., & Jones, G. A. (2005). Characterization of condensed tannins purified from legume forages: Chromophore production, protein precipitation, and inhibitory effects on cellulose digestion. *Journal of Chemical Ecology*, 31(9), 2049–2068. <https://doi.org/10.1007/s10886-005-6077-4>



- Mc Dermott, P. F., Walker, R. D., & White, D. G. (2003). Antimicrobials: modes of action and mechanisms of resistance. *International journal of toxicology*, 22(2), 135-143. DOI: [10.1080/10915810305089](https://doi.org/10.1080/10915810305089)
- McIvor, I. R., Douglas, G. B., Hurst, S. E., Hussain, Z., & Foote, A. G. (2008). Structural root growth of young Veronese poplars on erodible slopes in the southern North Island, New Zealand. *Agroforestry Systems*, 72(1), 75-86. DOI: [10.1007/s10457-007-9090-5](https://doi.org/10.1007/s10457-007-9090-5)
- Mehaisen, G. M., Eshak, M. G., Elkaiaty, A. M., Atta, A. R. M., Mashaly, M. M., & Abass, A. O. (2017). Comprehensive growth performance, immune function, plasma biochemistry, gene expressions and cell death morphology responses to a daily corticosterone injection course in broiler chickens. *PLoS One*, 12(2), e0172684. <https://doi.org/10.1371/journal.pone.0172684>
- Mehdi, Y., Létourneau-Montminy, M. P., Gaucher, M. L., Chorfi, Y., Suresh, G., Rouissi, T., Brar, S. K., Côté, C., Ramirez, A. A., & Godbout, S. (2018). Use of antibiotics in broiler production: Global impacts and alternatives. *Animal Nutrition*, 4(2), 170–178. <https://doi.org/10.1016/j.aninu.2018.03.002>
- Mehdipour, Z., Afsharmanesh, M., & Sami, M. (2013). Effects of dietary synbiotic and cinnamon (*Cinnamomum verum*) supplementation on growth performance and meat quality in Japanese quail. *Livestock Science*, 154(1–3), 152–157. <https://doi.org/10.1016/j.livsci.2013.03.014>
- Melguizo-Rodríguez, Manzano-Moreno, Illescas-Montes, Ramos-Torrecillas, Luna-Bertos, Ruiz, & García-Martínez. (2019). Bone Protective Effect of Extra-Virgin Olive Oil Phenolic Compounds by Modulating Osteoblast Gene Expression. *Nutrients*, 11(8), 1722. <https://doi.org/10.3390/nu11081722>
- Mellata, M. (2013). Human and avian extraintestinal pathogenic *Escherichia coli*: infections, zoonotic risks, and antibiotic resistance trends. *Foodborne Pathogens and Disease*, 10(11), 916-932. <https://doi.org/10.1089/fpd.2013.1533>
- Mihailovic, V., Mišić, D., Matić, S., Mihailović, M., Stanić, S., Vrvić, M. M., Katanić, J., Mladenović, M., Stanković, N., Boroja, T., and Stanković, M. S. (2015). Comparative phytochemical analysis of *Gentiana cruciata* L. roots and aerial parts, and their biological activities. *Industrial Crops and Products*, 73, 49–62. <https://doi.org/10.1016/j.indcrop.2015.04.013>
- Miltonprabu, S., Tomczyk, M., Skalicka-Woźniak, K., Rastrelli, L., Daglia, M., Nabavi, S. F., Alavian, S. M., & Nabavi, S. M. (2017). Hepatoprotective effect of quercetin: From chemistry to medicine. *Food and Chemical Toxicology*, 108, 365-374. <https://doi.org/10.1016/j.fct.2016.08.034>

- Mir, M. A., Parihar, K., Tabasum, U., Kumari, E., Mir, A., & Amin Mir, M. (2016). Estimation of alkaloid, saponin and flavonoid, content in various extracts of *Crocus sativa*. *Journal of Medicinal Plants Studies*, 4(5), 171–174. Retrieved from <https://www.plantsjournal.com/archives/2016/vol4issue5/PartC/4-5-10-775.pdf>
- Mir, N. A., Rafiq, A., Kumar, F., Singh, V., & Shukla, V. (2017). Determinants of broiler chicken meat quality and factors affecting them: a review. *Journal of Food Science and Technology*, 54(10), 2997–3009. <https://doi.org/10.1007/s13197-017-2789-z>
- Mishra, B., & Jha, R. (2019). Oxidative stress in the poultry gut: Potential challenges and interventions. *Frontiers in Veterinary Science*, 6(MAR), 1–5. <https://doi.org/10.3389/fvets.2019.00060>
- Mohammadagheri, N., Najafi, R., & Najafi, G. (2016). Effects of dietary supplementation of organic acids and phytase on performance and intestinal histomorphology of broilers. *Veterinary Research Forum : An International Quarterly Journal*, 7(3), 189–195. [Google Scholar]
- Mohammadi Gheisar, M., Hosseindoust, A., & Kim, I. H. (2016). Effects of dietary *Enterococcus faecium* on growth performance, carcass characteristics, faecal microbiota, and blood profile in broilers. *Veterinarni Medicina*, 61(1), 28–34. <https://doi.org/10.17221/8680-VETMED>
- Mohammadi Gheisar, M., & Kim, I. H. (2018). Phytobiotics in poultry and swine nutrition—a review. *Italian Journal of Animal Science*, 17(1), 92–99. <https://doi.org/10.1080/1828051X.2017.1350120>
- Mohiti-Asli, M., & Ghanaatparast-Rashti, M. (2017). Comparing the effects of a combined phyto-genic feed additive with an individual essential oil of oregano on intestinal morphology and microflora in broilers. *Journal of Applied Animal Research*, 46(1), 1–6. <https://doi.org/10.1080/09712119.2017.1284074>
- Ministry of Health Malaysia [MOH] (2018). *Malaysian Action Plan on Antimicrobial Resistance (MyAP-AMR) 2017-2021*. Available online at: [https://www.moh.gov.my/moh/resources/Penerbitan/Garis%20Panduan/Garis%20panduan%20Umum%20\(Awam\)/National\\_Action\\_Plan\\_-\\_FINAL\\_29\\_june.pdf](https://www.moh.gov.my/moh/resources/Penerbitan/Garis%20Panduan/Garis%20panduan%20Umum%20(Awam)/National_Action_Plan_-_FINAL_29_june.pdf) (accessed 25-01- 2022).
- Moorthy, M., Khoo, J. J., & Palanisamy, U. D. (2019). Acute oral toxicity of the ellagitannin geraniin and a geraniin-enriched extract from *Nephelium lappaceum* L rind in Sprague Dawley rats. *Heliyon*, 5(8), e02333. <https://doi.org/10.1016/j.heliyon.2019.e02333>

- Movahhedkhah, S., Rasouli, B., Seidavi, A., Mazzei, D., Laudadio, V., & Tufarelli, V. (2019). Summer savory (*Satureja hortensis* L.) extract as natural feed additive in broilers: Effects on growth, plasma constituents, immune response, and ileal microflora. *Animals*, 9(3), 87. <https://doi.org/10.3390/ani9030087>
- Mpofu, D. A., Marume, U., Mlambo, V., & Hugo, A. (2016). The effects of *Lippia javanica* dietary inclusion on growth performance, carcass characteristics and fatty acid profiles of broiler chickens. *Animal Nutrition*, 2(3), 160-167. <https://doi.org/10.1016/j.aninu.2016.05.003>
- Muloi, D., Ward, M. J., Pedersen, A. B., Fevre, E. M., Woolhouse, M. E., & Van Bunnik, B. A. (2018). Are food animals responsible for transfer of antimicrobial-resistant *Escherichia coli* or their resistance determinants to human populations? A systematic review. *Foodborne Pathogens and Disease*, 15(8), 467-474. <https://doi.org/10.1089/fpd.2017.2411>
- Mund, M. D., Khan, U. H., Tahir, U., Mustafa, B. E., & Fayyaz, A. (2017). Antimicrobial drug residues in poultry products and implications on public health: A review. *International Journal of Food Properties*, 20(7), 1433-1446. [doi.org/10.1080/10942912.2016.1212874](https://doi.org/10.1080/10942912.2016.1212874)
- Muntean, E., Michalski, R., Muntean, N., & Duda, M. (2016). Chemical risk due to heavy metal contamination of medicinal plants. *Hop and Medicinal Plants*, 24(1-2), 71-78. [Google Scholar]
- Murate, L. S., Paião, F. G., de Almeida, A. M., Berchieri, A., & Shimokomaki, M. (2015). Efficacy of prebiotics, probiotics, and synbiotics on laying hens and broilers challenged with *Salmonella enteritidis*. *Journal of Poultry Science*, 52(1), 53–56. <https://doi.org/10.2141/jpsa.0130211>
- Muruganandam, L., Krishna, A., Reddy, J., & Nirmala, G. S. (2017). Optimization studies on extraction of phytochemicals from betel leaves. *Resource-Efficient Technologies*, 3(4), 385–393. <https://doi.org/10.1016/j.reffit.2017.02.007>
- Murugesan, G. R., Gabler, N. K., & Persia, M. E. (2014). Effects of direct-fed microbial supplementation on broiler performance, intestinal nutrient transport and integrity under experimental conditions with increased microbial challenge. *British Poultry Science*, 55(1), 89-97. <https://doi.org/10.1080/00071668.2013.865834>
- Murugesan, G. R., Syed, B., Haldar, S., & Pender, C. (2015). Phytochemical feed additives as an alternative to antibiotic growth promoters in broiler chickens. *Frontiers in Veterinary Science*, 2(AUG), 1–6. <https://doi.org/10.3389/fvets.2015.00021>
- Mushtaq, M. M. H., & Pasha, T. N. (2013). Electrolytes, dietary electrolyte balance and salts in broilers: An updated review on acid-base balance, blood and

- carcass characteristics. *World's Poultry Science Journal*, 69(4), 833–852. <https://doi.org/10.1017/S0043933913000846>
- Mustafa, M. A. G. (2019). Effect of eucalyptus leaves and its supplementation with diet on broiler performance, microbial and physiological statues to alleviate cold stress. *Iraqi Journal of Agricultural Sciences*, 50(1), 359–368. [[Google Scholar](#)]
- Muthusamy, N., Sankar, V., & Sheep, M. (2015). Phytogetic compounds used as a feed additive in poultry production. *International Journal of Science, Environment and Technology*, 4(1), 167-171. [[Google Scholar](#)]
- Narasimhulu, G., & Mohamed, J. (2014). Medicinal phytochemical and pharmacological properties of kesum (*Polygonum minus* Linn.): A mini review. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(4), 682-688. [[Google Scholar](#)]
- Nayaka, N. M. D. M. W., Sasadara, M. M. V., Sanjaya, D. A., Yuda, P. E. S. K., Dewi, N. L. K. A. A., Cahyaningsih, E., & Hartati, R. (2021). Piper betle (L): Recent Review of Antibacterial and Antifungal Properties, Safety Profiles, and Commercial Applications. *Molecules*, 26(8), 2321. <https://doi.org/10.3390/molecules26082321>
- Nazzaro, F., Fratianni, F., De Martino, L., Coppola, R., & De Feo, V. (2013). Effect of essential oils on pathogenic bacteria. *Pharmaceuticals*, 6(12), 1451-1474. <https://doi.org/10.3390/ph6121451>
- Nhung, N. T., Cuong, N. V., Thwaites, G., & Carrique-Mas, J. (2016). Antimicrobial usage and antimicrobial resistance in animal production in Southeast Asia: a review. *Antibiotics*, 5(4), 37. <https://doi.org/10.3390/antibiotics5040037>
- Nhung, N. T., Chansiripornchai, N., & Carrique-Mas, J. J. (2017). Antimicrobial resistance in bacterial poultry pathogens: a review. *Frontiers in veterinary science*, 4, 126. <https://doi.org/10.3389/fvets.2017.00126>
- Nidaullah, H., Abirami, N., Shamila-Syuhada, A. K., Chuah, L. O., Nurul, H., Tan, T. P., Abidin, F. W. Z., & Rusul, G. (2017). Prevalence of Salmonella in poultry processing environments in wet markets in Penang and Perlis, Malaysia. *Veterinary World*, 10(3), 286. [doi: 10.14202/vetworld.2017.286-292](https://doi.org/10.14202/vetworld.2017.286-292)
- Niewold, T. A. (2007). The nonantibiotic anti-inflammatory effect of antimicrobial growth promoters, the real mode of action? A hypothesis. *Poultry Science*, 86(4), 605-609. <https://doi.org/10.1093/ps/86.4.605>
- Niu, Y., Wan, X. L., Zhang, L. L., Wang, C., He, J. T., Bai, K. W., Zhang, X. H., Zhao, L. G., & Wang, T. (2019). Effect of different doses of fermented

Ginkgo biloba leaves on serum biochemistry, antioxidant capacity hepatic gene expression in broilers. *Animal Feed Science and Technology*, 248, 132-140.

[doi.org/10.1016/j.anifeedsci.2019.01.003](https://doi.org/10.1016/j.anifeedsci.2019.01.003)

Niu, Y., Zhang, J. F., Wan, X. L., Huang, Q., He, J. T., Zhang, X. H., Zhao, L. G., Zhang, L. L., & Wang, T. (2019). Effect of fermented Ginkgo biloba leaves on nutrient utilisation, intestinal digestive function and antioxidant capacity in broilers. *British Poultry Science*, 60(1), 47-55.

<https://doi.org/10.1080/00071668.2018.1535166>

NRC (1994). *Nutrient Requirements of Poultry. 9<sup>th</sup> Revised Edition*. The National Academies Press, Washington, D. C, USA.

Odetola, O. M., Adejinmi, O. O., Owosibo, O. A., Banjo, O. T., & Awodola-Peters, O. O. (2019). Growth response, serum biochemistry and organ histopathology of broilers fed diets supplemented with graded levels of Petiveria alliacea root meal. *International Journal of Poultry Science*, 18, 45-50. DOI: 10.3923/ijps.2019.45.50

OECD. (2010). *Test No. 223: Avian Acute Oral Toxicity Test*. OECD Publishing. <https://doi.org/10.1787/9789264090897-en>

Ofongo-Abule, R. T. S., Etebu, E., & Ohimain, E. I. (2016). Performance and Molecular Identification of Bacteria Isolated from the Gut of Broiler Birds After Antibiotic Administration and Enzyme Supplementation. *Journal of Microbiology, Biotechnology and Food Sciences*, 6(3), 924–929. <https://doi.org/10.15414/jmbfs.2016/17.6.3.924-929>

Oghenebrorhie, O., & Oghenesuvwe, O. (2016). Performance and haematological characteristics of broiler finisher fed Moringa oleifera leaf meal diets. *Journal of Northeast Agricultural University (English Edition)*, 23(1), 28-34.

[https://doi.org/10.1016/S1006-8104\(16\)30029-0](https://doi.org/10.1016/S1006-8104(16)30029-0)

Ohadoma, S. C., Nnatuanya, I., Amazu, L. U., & Okolo, C. E. (2014). Antimicrobial activity of the leaf extract and fractions of Lupinus arboreus. *Journal of Medicinal Plants Research*, 8(8), 386–391.

<https://doi.org/10.5897/jmpr12.1263>

Oloruntola, O. D., Ayodele, S. O., Agbede, J. O., & Oloruntola, D. A. (2016). Effect of feeding broiler chickens with diets containing Alchornea cordifolia leaf meal and enzyme supplementation. *Archivos de Zootecnia*, 65(252), 489-498. [Google Scholar]

Oloruntola, O. D. (2019). Effect of pawpaw leaf and seed meal composite mix dietary supplementation on haematological indices, carcass traits and histological studies of broiler chicken. *Bulletin of the National Research Centre*, 43(1), 129. <https://doi.org/10.1186/s42269-019-0172-0>

- Oloruntola, O. D., Agbede, J. O., Ayodele, S. O., & Oloruntola, D. A. (2019). Neem, pawpaw and bamboo leaf meal dietary supplementation in broiler chickens: Effect on performance and health status. *Journal of Food Biochemistry*, 43(2), e12723. <https://doi.org/10.1111/jfbc.12723>
- Orlowski, S., Flees, J., Greene, E. S., Ashley, D., Lee, S. O., Yang, F. L., Owens, C. M., Kidd, M., Anthony, N., & Dridi, S. (2018). Effects of phytogetic additives on meat quality traits in broiler chickens. *Journal of Animal Science*, 96(9), 3757–3767. <https://doi.org/10.1093/jas/sky238>
- Oso, A. O., Suganthi, R. U., Reddy, G. M., Malik, P. K., Thirumalaisamy, G., Awachat, V. B., Selvaraju, S., Arangasamy, A., & Bhatta, R. (2019). Effect of dietary supplementation with phytogetic blend on growth performance, apparent ileal digestibility of nutrients, intestinal morphology, and cecal microflora of broiler chickens. *Poultry Science*, 98(10), 4755–4766. <https://doi.org/10.3382/ps/pez191>
- Ouyang, K., Xu, M., Jiang, Y., & Wang, W. (2016). Effects of alfalfa flavonoids on broiler performance, meat quality, and gene expression. *Canadian Journal of Animal Science*, 96(3), 332–341. <https://doi.org/10.1139/cjas-2015-0132>
- Oz, H. S. (2017). Nutrients, infectious and inflammatory diseases. *Nutrients*, 9(10), 1–9. <https://doi.org/10.3390/nu9101085>
- Ozbudak, S. (2019). Phytoiotics and Their Roles in Broiler Nutrition. *Journal of Poultry Research*. *Journal of Poultry Research*, 16(1), 23-29. <https://doi.org/10.34233/jpr.465575>
- Padhi, L., & Panda, S. K. (2015). Antibacterial activity of *Eleutherine bulbosa* against multidrug-resistant bacteria. *Journal of Acute Medicine*, 5(3), 53–61. <https://doi.org/10.1016/j.jacme.2015.05.004>
- Palaniappan, K., & Holley, R. A. (2010). Use of natural antimicrobials to increase antibiotic susceptibility of drug resistant bacteria. *International Journal of Food Microbiology*, 140(2-3), 164-168. <https://doi.org/10.1016/j.ijfoodmicro.2010.04.001>
- Pan, D., & Yu, Z. (2014). Intestinal microbiome of poultry and its interaction with host and diet. *Gut Microbes*, 5(1), 108-119. [[Google Scholar](#)]
- Panaite, T. D., Saracila, M., Papuc, C. P., Predescu, C. N., & Soica, C. (2020). Influence of dietary supplementation of salix alba bark on performance, oxidative stress parameters in liver and gut microflora of broilers. *Animals*, 10(6), 958. <https://doi.org/10.3390/ani10060958>
- Panda, S., Sikdar, M., Biswas, S., Sharma, R., & Kar, A. (2019). Allylpyrocatechol, isolated from betel leaf ameliorates thyrotoxicosis in rats by altering thyroid peroxidase and thyrotropin receptors. *Scientific Reports*, 9(1), 1–12. <https://doi.org/10.1038/s41598-019-48653-9>

- Pandey, A. K., Kumar, P., & Saxena, M. J. (2019). Feed Additives in Animal Health. In *Nutraceuticals in Veterinary Medicine* (pp. 345-362). Springer, Cham. [https://doi.org/10.1007/978-3-030-04624-8\\_23](https://doi.org/10.1007/978-3-030-04624-8_23)
- Paraskeuas, V., Fegeros, K., Palamidi, I., Hunger, C., & Mountzouris, K. C. (2017). Growth performance , nutrient digestibility , antioxidant capacity , blood biochemical biomarkers and cytokines expression in broiler chickens fed different phytogetic levels. *Animal Nutrition*, 3(2), 114–120. <https://doi.org/10.1016/j.aninu.2017.01.005>
- Park, M., Cho, H., Jung, H., Lee, H., & Hwang, K. T. (2014). Antioxidant and anti-inflammatory activities of tannin fraction of the extract from black raspberry seeds compared to grape seeds. *Journal of Food Biochemistry*, 38(3), 259–270. <https://doi.org/10.1111/jfbc.12044>
- Parmar, A., Patel, V., Patel, J., Usadadia, S., Rathwa, S., Prajapati, D., & Rathva, A. (2019). Quercetin, a health promising phytoadditive for poultry production: Trends & advances. *The Pharma Innovation Journal*, 8(9), 68-74. [\[Google Scholar\]](#)
- Parsaie, S., Shariatmadari, F., Zamiri, M. J., & Khajeh, K. (2007). Influence of wheat-based diets supplemented with xylanase, bile acid and antibiotics on performance, digestive tract measurements and gut morphology of broilers compared with a maize-based diet. *British Poultry Science*, 48(5), 594–600. <https://doi.org/10.1080/00071660701615788>
- Patil, S. D., Sharma, R., Srivastava, S., Navani, N. K., & Pathania, R. (2013). Downregulation of yidC in Escherichia coli by antisense RNA expression results in sensitization to antibacterial essential oils eugenol and carvacrol. *PLoS One*, 8(3), e57370. <https://doi.org/10.1371/journal.pone.0057370>
- Paul, S., Islam, M., Tanvir, E. M., Ahmed, R., Das, S., Rumpa, N. E., Hossen, M., Parvez, M., Gan, S. H., & Khalil, M. (2016). Satkara (*Citrus macroptera*) fruit protects against acetaminophen-induced hepatorenal toxicity in rats. *Evidence-Based Complementary and Alternative Medicine*, 2016. <https://doi.org/10.1155/2016/9470954>
- Paul, T. K., Hasan, M. M., Haque, M. A., Talukder, S., Sarker, Y. A., Sikder, M. H., Khan, M., Sakib, M. N., & Kumar, A. (2020). Dietary supplementation of Neem (*Azadirachta indica*) leaf extracts improved growth performance and reduced production cost in broilers, *Veterinary World*, 13(6): 1050-1055. [doi: 10.14202/vetworld.2020.1050-1055](https://doi.org/10.14202/vetworld.2020.1050-1055)
- Pawar, S., Kalyankar, V., Dhamangaonkar, B., Dagade, S., Waghmode, S., & Cukkemane, A. (2017). Biochemical profiling of antifungal activity of betel leaf(*Piper betle* L.) extract and its significance in traditional

- medicine. *Journal of Advanced Research in Biotechnology*, 2(1), 1–4.  
<https://doi.org/10.15226/2475-4714/2/1/00116>
- Pawłowska, K. A., Strawa, J., Tomczyk, M., & Granica, S. (2020). Changes in the phenolic contents and composition of *Persicaria odorata* fresh and dried leaves. *Journal of Food Composition and Analysis*, 91(August 2019).  
<https://doi.org/10.1016/j.jfca.2020.103507>
- Peng, Q. Y., Li, J. D., Li, Z., Duan, Z. Y., & Wu, Y. P. (2016). Effects of dietary supplementation with oregano essential oil on growth performance, carcass traits and jejunal morphology in broiler chickens. *Animal Feed Science and Technology*, 214, 148–153.  
<https://doi.org/10.1016/j.anifeedsci.2016.02.010>
- Perera, H. D. S. M., Samarasekera, J. K. R. R., Handunnetti, S. M., & Weerasena, O. V. D. S. J. (2016). In vitro anti-inflammatory and antioxidant activities of Sri Lankan medicinal plants. *Industrial Crops and Products*, 94, 610–620. <https://doi.org/10.1016/j.indcrop.2016.09.009>
- Perez-Gregorio, M. R., Mateus, N., & de Freitas, V. (2014). Rapid Screening and Identification of New Soluble Tannin–Salivary Protein Aggregates in Saliva by Mass Spectrometry (MALDI-TOF-TOF and FIA-ESI-MS). *Langmuir*, 30(28), 8528–8537. <https://doi.org/10.1021/la502184f>
- Periyanyagam, K., Jagadeesan, M., Kavimani, S., & Vetrivelvan, T. (2012). Pharmacognostical and Phyto-physicochemical profile of the leaves of Piper betle L. var Pachaikodi (Piperaceae) - Valuable assessment of its quality. *Asian Pacific Journal of Tropical Biomedicine*, 2(2 SUPPL.), S506–S510. [https://doi.org/10.1016/S2221-1691\(12\)60262-7](https://doi.org/10.1016/S2221-1691(12)60262-7)
- Pietrzyk, L. (2017). Food properties and dietary habits in colorectal cancer prevention and development. *International Journal of Food Properties*, 20(10), 2323–2343. <https://doi.org/10.1080/10942912.2016.1236813>
- Polycarpo, G. V., Andretta, I., Kipper, M., Cruz-Polycarpo, V. C., Dadalt, J. C., Rodrigues, P. H. M., & Albuquerque, R. (2017). Meta-analytic study of organic acids as an alternative performance-enhancing feed additive to antibiotics for broiler chickens. *Poultry science*, 96(10), 3645–3653.  
<https://doi.org/10.3382/ps/pex178>
- Pin, K. Y., Ilicali, C., Luqman Chuah, A., Choong, T. S. Y., Muhammad Shahrul, M. N., Rasadah, M. A., & Law, C. L. (2011). Modelling of freezing kinetics of extract of betel leaves (*Piper betle* L.). *International Journal of Food Engineering*, 7(1), 3758.  
<https://doi.org/10.2202/1556-3758.1692>
- Pingili, R. B., Challa, S. R., Pawar, A. K., Toleti, V., Kodali, T., & Koppula, S. (2020). A systematic review on hepatoprotective activity of quercetin against various drugs and toxic agents: Evidence from preclinical studies. *Phytotherapy Research*, 34(1), 5–32.



<https://doi.org/10.1002/ptr.6503>

Pirgozliev, V., Bravo, D., Mirza, M. W., & Rose, S. P. (2015). Growth performance and endogenous losses of broilers fed wheat-based diets with and without essential oils and xylanase supplementation. *Poultry Science*, 94(6), 1227–1232. <https://doi.org/10.3382/ps/peu017>

Popova, T. (2017). Effect of probiotics in poultry for improving meat quality. *Current Opinion in Food Science*, 14, 72–77. <https://doi.org/10.1016/j.cofs.2017.01.008>

Porras, D., Nistal, E., Martínez-Flórez, S., Pisonero-Vaquero, S., Olcoz, J. L., Jover, R., González-Gallego, J., García-Mediavilla, M. V., & Sánchez-Campos, S. (2017). Protective effect of quercetin on high-fat diet-induced non-alcoholic fatty liver disease in mice is mediated by modulating intestinal microbiota imbalance and related gut-liver axis activation. *Free Radical Biology and Medicine*, 102, 188–202. <https://doi.org/10.1016/j.freeradbiomed.2016.11.037>

Pourabedin, M., & Zhao, X. (2015). Prebiotics and gut microbiota in chickens. *FEMS Microbiology Letters*, 362(15), 1–8. <https://doi.org/10.1093/femsle/fnv122>

Prabakar, G., Gopi, M., Karthik, K., Shanmuganathan, S., Kirubakaran, A., & Pavulraj, S. (2016). Review Article Phytobiotics: Could the Greens Inflation the Poultry Production. *Asian Journal of Animal and Veterinary Advances*, 11(7), 383–392. <https://doi.org/10.3923/ajava.2016.383.392>

Pradhan, D., Suri, K. A., Pradhan, D. K., & Biswasroy, P. (2013). Golden Heart of the Nature: Piper betle L. *Journal of Pharmacognosy and Phytochemistry*, 1(6). [[Google Scholar](#)]

Pradhan, D., Biswasroy, P., & Suri, K. (2014). Various factors influencing the percentage content of hydroxychavicol in different extracts of Piper betle L. by altering the extraction parameters. *International Journal of Advanced Scientific and Technical Research*, 2, 517–530. [[Google Scholar](#)]

Prakatur, I., Miskulin, M., Pavic, M., Marjanovic, K., Blazicevic, V., Miskulin, I., & Domacinovic, M. (2019). Intestinal morphology in broiler chickens supplemented with propolis and bee pollen. *Animals*, 9(6), 301. <https://doi.org/10.3390/ani9060301>

Proctor, A., & Phillips, G. J. (2019). Differential effects of bacitracin methylene disalicylate (BMD) on the distal colon and cecal microbiota of young broiler chickens. *Frontiers in Veterinary Science*, 6(MAR), 1–9. <https://doi.org/10.3389/fvets.2019.00114>

- Punuri, J. B., Sharma, P., Sibyala, S., Tamuli, R., & Bora, U. (2012). Piper betle-mediated green synthesis of biocompatible gold nanoparticles, *International Nano Letters*, 2(1), 18.  
<https://doi.org/10.1186/2228-5326-2-18>
- Qazi, M. A., & Molvi K. I. (2016). Herbal Medicine: A Comprehensive Review. *International Journal of Pharmaceutical Research*, 8(2), 8-12
- Qureshi, S., Bandy, M. T., Shakeel, I., Adil, S., Mir, M. S., Beigh, Y. A., & Amin, U. (2016). Histomorphological studies of broiler chicken fed diets supplemented with either raw or enzyme treated dandelion leaves and fenugreek seeds. *Veterinary World*, 9(3), 269.  
[doi: 10.14202/vetworld.2016.269-275](https://doi.org/10.14202/vetworld.2016.269-275)
- Rahman, M. M., & Yang, D. K. (2018). Effects of Ananas comosus leaf powder on broiler performance, haematology, biochemistry, and gut microbial population. *Revista Brasileira de Zootecnia*, 47(e20170064), 1–6.  
<https://doi.org/10.1590/rbz4720170064>
- Ramirez-Acosta, M., Jiménez-Plascencia, C., Juárez-Woo, C., Rendón-Guizar, J., Ángeles-Espino, A., & Sánchez-Chiprés, D. (2018). Inclusion of the Moringa oleifera leaf on immunological constants in broiler chickens. *Abanico veterinario*, 8(3), 68-74.  
<https://doi.org/10.21929/abavet2018.83.4>
- Raphael, K. J. (2017). Growth performance and serum biochemical profile of broiler chickens fed on diets supplemented with Afrostyrax lepidophyllus fruit and bark as alternative to antibiotic growth promoters. *Journal of Veterinary Medicine and Research*. [[Google Scholar](#)]
- Rashid, Z., Mirani, Z. A., Zehra, S., Gilani, S. M. H., Ashraf, A., Azhar, A., Al-Ghanim, K. A., Al-Misned, F., Al-Mulahim, N., Mahboob, S., & Galani, S. (2020). Enhanced modulation of gut microbial dynamics affecting body weight in birds triggered by natural growth promoters administered in conventional feed. *Saudi Journal of Biological Sciences*, 27(10), 2747-2755. <https://doi.org/10.1016/j.sjbs.2020.06.027>
- Ravindran, V. (2013). Feed enzymes: The science, practice, and metabolic realities. *Journal of Applied Poultry Research*, 22(3), 628-636.  
<https://doi.org/10.3382/japr.2013-00739>
- Raza, A., Muhammad, F., Bashir, S., Aslam, B., Anwar, M. I., & Naseer, M. U. (2016). In-vitro and in-vivo anthelmintic potential of different medicinal plants against Ascaridia galli infection in poultry birds. *World's Poultry Science Journal*, 72(1), 115-124. [DOI:10.1017/S0043933915002615](https://doi.org/10.1017/S0043933915002615)
- Re, R., Pellegrini, N., Proteggente, A., Pannala, A., Yang, M., & Rice-Evans, C. (1999). Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical Biology and Medicine*, 26(9–10),

1231–1237. [https://doi.org/10.1016/S0891-5849\(98\)00315-3](https://doi.org/10.1016/S0891-5849(98)00315-3)

- Redondo, L. M., Chacana, P. A., Dominguez, J. E., & Fernandez Miyakawa, M. E. D. (2014). Perspectives in the use of tannins as alternative to antimicrobial growth promoter factors in poultry. *Frontiers in Microbiology*, 5, 118. <https://doi.org/10.3389/fmicb.2014.00118>
- Rehman, H. F., Zaneb, H., Masood, S., Yousaf, M. S., Ashraf, S., Khan, I., Shah, M., Khilji, M. S., & Rehman, H. (2018). Effect of moringa oleifera leaf powder supplementation on pectoral muscle quality and morphometric characteristics of tibia bone in broiler chickens. *Revista Brasileira de Ciencia Avicola*, 20(4), 817–824. <https://doi.org/10.1590/1806-9061-2017-0609>
- Rehman, Z. U., Che, L., Ren, S., Liao, Y., Qiu, X., Yu, S., Sun, Y., Tan, L., Song, C., Liu, W., & Ding, Z. (2018). Supplementation of vitamin e protects chickens from Newcastle disease virus-mediated exacerbation of intestinal oxidative stress and tissue damage. *Cellular Physiology and Biochemistry*, 47(4), 1655-1666. [[Google Scholar](#)]
- Reis, J. H., Gebert, R. R., Barreta, M., Baldissera, M. D., Dos Santos, I. D., Wagner, R., Campigotto, G., Jaguezeski, A. M., Gris, A., de Lima, J. L., & Da Silva, A. S. (2018). Effects of phytogetic feed additive based on thymol, carvacrol and cinnamic aldehyde on body weight, blood parameters and environmental bacteria in broilers chickens. *Microbial Pathogenesis*, 125, 168-176. <https://doi.org/10.1016/j.micpath.2018.09.015>
- Rekha, V. P. B., Kollipara, M., Gupta, B. R. S. S., Bharath, Y., & Pulicherla, K. K. (2014). A review on Piper betle L.: nature's promising medicinal reservoir. *American Journal of Ethnomedicine*, 1(5), 276-289. [[Google Scholar](#)]
- Renu, S., Jakhar, K. K., Deepika, L., Vikas, N., & Adya, P. (2017). Clinico-pathological studies of thiacloprid toxicity in broiler chickens. *Haryana Veterinarian*, 56(2), 204-206. [[Google Scholar](#)]
- Requena, R., Vargas, M., & Chiralt, A. (2019). Eugenol and carvacrol migration from PHBV films and antibacterial action in different food matrices. *Food Chemistry*, 277(June 2018), 38–45. <https://doi.org/10.1016/j.foodchem.2018.10.093>
- Reygaert, W. C. (2018). An overview of the antimicrobial resistance mechanisms of bacteria. *AIMS microbiology*, 4(3), 482. [doi: 10.3934/microbiol.2018.3.482](https://doi.org/10.3934/microbiol.2018.3.482)
- Rhiouani, H., El-Hilaly, J., Israili, Z. H., & Lyoussi, B. (2008). Acute and sub-chronic toxicity of an aqueous extract of the leaves of *Herniaria glabra* in rodents. *Journal of Ethnopharmacology*, 118(3), 378–386.

<https://doi.org/10.1016/j.jep.2008.05.009>

- Ridzuan, P. M., Hamzah, H. A., Shah, A., Hassan, N. M., & Roesnita, B. (2017). Synergistic effects of *Persicaria odorata* (Daun Kesom) leaf extracts with standard antibiotics on pathogenic bacteria. *IJUM Medical Journal Malaysia*, 16(2), 27–32. <https://doi.org/10.31436/imjm.v16i2.321>
- Rinttila, T., & Apajalahti, J. (2013). Intestinal microbiota and metabolites—Implications for broiler chicken health and performance. *Journal of Applied Poultry Research*, 22(3), 647–658. <https://doi.org/10.3382/japr.2013-00742>
- Rintu, D., Shinjini, M., & Pramathadhip P, K. M. (2015). Anti-Oxidant and Anti-Inflammatory Activities of Different Varieties of Piper Leaf Extracts (*Piper Betle* L.). *Journal of Nutrition & Food Sciences*, 05(05), 1–15. <https://doi.org/10.4172/2155-9600.1000415>
- Rios, H. V., Vieira, S. L., Stefanello, C., Kindlein, L., Soster, P., dos Santos, P. I., & Toscan, A. B. (2017). Energy and nutrient utilisation of maize-soy diet supplemented with a xylanase- $\beta$ -glucanase complex from *Talaromyces versatilis*. *Animal Feed Science and Technology*, 232, 80–90. <https://doi.org/10.1016/j.anifeedsci.2017.08.009>
- Ripon, M. M. R., Rashid, M. H., Rahman, M. M., Ferdous, M. F., Arefin, M. S., Sani, A. A., Hossain, M. T., Ahammad, M. U., & Rafiq, K. (2019). Dose-dependent response to phytobiotic supplementation in feed on growth, hematology, intestinal pH, and gut bacterial load in broiler chicken. *Journal of Advanced Veterinary and Animal Research*, 6(2), 253–259. <https://doi.org/10.5455/javar.2019.f341>
- Rodriguez-Garcia, A., Hosseini, S., Martinez-Chapa, S. O., & Cordell, G. A. (2017). Multi-target Activities of Selected Alkaloids and Terpenoids. *Mini-Reviews in Organic Chemistry*, 14(4). <https://doi.org/10.2174/1570193X14666170518151027>
- Roman, Y., Bomsel-Demontoy, M. C., Levrier, J., Ordonneau, D., Chaste-Duvernoy, D., & Saint Jalme, M. (2009). Influence of molt on plasma protein electrophoretic patterns in bar-headed geese (*Anser indicus*). *Journal of Wildlife Diseases*, 45(3), 661-671. <https://doi.org/10.7589/0090-3558-45.3.661>
- Ronquillo, M. G., & Hernandez, J. C. A. (2017). Antibiotic and synthetic growth promoters in animal diets: review of impact and analytical methods. *Food Control*, 72, 255-267. <https://doi.org/10.1016/j.foodcont.2016.03.001>
- Ross, K. A., Ehret, D., Godfrey, D., Fukumoto, L., & Diarra, M. (2017). Characterization of pilot scale processed canadian organic cranberry (*Vaccinium macrocarpon*) and blueberry (*Vaccinium angustifolium*) juice pressing residues and phenolic-enriched extractives. *International Journal of Fruit Science*, 17(2), 202-232. [[Google Scholar](#)]

- Rossi, B., Toschi, A., Piva, A., & Grilli, E. (2020). Single components of botanicals and nature-identical compounds as a non-antibiotic strategy to ameliorate health status and improve performance in poultry and pigs. *Nutrition Research Reviews*, 1-17. DOI: [10.1017/S0954422420000013](https://doi.org/10.1017/S0954422420000013)
- Rubio, M. S., Laurentiz, A. C., Sobrane, F., Mello, E. S., Filardi, R. S., Silva, M. L. A., & Laurentiz, R. S. (2019). Performance and Serum Biochemical Profile of Broiler Chickens Supplemented with Piper Cubeba Ethanolic Extract. *Brazilian Journal of Poultry Science*, 21(1). <http://dx.doi.org/10.1590/1806-9061-2018-0789>
- Saad, R., Khan, J., Krishnanmurthi, V., Asmani, F., & Yusuf, E. (2014). Effect of Different Extraction Techniques of *Persicaria odorata* Extracts Utilizing Anti-bacterial Bioassay. *British Journal of Pharmaceutical Research*, 4(18), 2146–2154. <https://doi.org/10.9734/bjpr/2014/12232>
- Saeed, M., Naveed, M., Arain, M. A., Arif, M., Abd El-Hack, M. E., Alagawany, M., Siyal, F. A., Soomro, R. N., & Sun, C. (2017). Quercetin: Nutritional and beneficial effects in poultry. *World's Poultry Science Journal*, 73(2), 355-364. [[Google Scholar](#)]
- Saeid, J. M., Mohamed, A. B., & Baddy, M. A. (2013). Effect of garlic powder (*Allium sativum*) and black seed (*Nigella sativa*) on broiler growth performance and intestinal morphology. *Iranian Journal of Applied Animal Science*, 3(1), 185-188. [[Google Scholar](#)]
- Saifullah, N., Siddiqui, R. A., Memon, Z., Shahid, M. A., & Mirza, T. (2019). Eugenol ameliorates rhabdomyolysis-induced acute kidney in Mice. *Pakistan Journal of Medicine and Dentistry*, 8(03), 25–29. [[Google Scholar](#)]
- Salaheen, S., Kim, S. W., Haley, B. J., Van Kessel, J. A. S., & Biswas, D. (2017). Alternative growth promoters modulate broiler gut microbiome and enhance body weight gain. *Frontiers in Microbiology*, 8, 2088. <https://doi.org/10.3389/fmicb.2017.02088>
- Salami, S. A., Majoka, M. A., Saha, S., Garber, A., & Gabarrou, J. F. (2015). Efficacy of dietary antioxidants on broiler oxidative stress, performance and meat quality: science and market. *Avian Biology Research*, 8(2), 65-78. DOI: [10.3184/175815515X14291701859483](https://doi.org/10.3184/175815515X14291701859483)
- Saleh, A. A., Ragab, M. M., Ahmed, E. A. M., Abudabos, A. M., & Ebeid, T. A. (2018). Effect of dietary zinc-methionine supplementation on growth performance, nutrient utilization, antioxidative properties and immune response in broiler chickens under high ambient temperature. *Journal of Applied Animal Research*, 46(1), 820–827. <https://doi.org/10.1080/09712119.2017.1407768>

- Salehi, B., Mishra, A. P., Shukla, I., Sharifi-Rad, M., Contreras, M. D. M., Segura-Carretero, A., Fathi, H., Nasrabadi, N. N., Kobarfard, F., & Sharifi-Rad, J. (2018). Thymol, thyme, and other plant sources: Health and potential uses. *Phytotherapy Research*, 32(9), 1688–1706.  
<https://doi.org/10.1002/ptr.6109>
- Sampaio, T. P. D., Cartaxo-Furtado, N. A. O., De Medeiros, A. C. D., Alves, H. S., Rosalen, P. L., & Pereira, J. V. (2017). Antimicrobial potential of plant extracts and chemical fractions of sideroxylon obtusifolium (roem. & schult.) t.d. penn on oral microorganisms. *Journal of Contemporary Dental Practice*, 18(5), 392–398.  
<https://doi.org/10.5005/jp-journals-10024-2053>
- San Chin, P., Yu, C. Y., Ang, G. Y., Yin, W. F., & Chan, K. G. (2017). Draft genome sequence of multidrug-resistant *Salmonella enterica* serovar Brancaster strain PS01 isolated from chicken meat, Malaysia. *Journal of Global Antimicrobial Resistance*, 9, 41–42.  
<https://doi.org/10.1016/j.jgar.2016.12.017>
- Sanghavi, N., Bhosale, S. D., & Malode, Y. (2014). RP-HPLC method development and validation of Quercetin isolated from the plant *Tridax procumbens* L. *Journal of Scientific and Innovative Research*, 3(6), 594–597.  
[[Google Scholar](#)]
- Sanver, D., Murray, B. S., Sadeghpour, A., Rappolt, M., & Nelson, A. L. (2016). Experimental Modeling of Flavonoid-Biomembrane Interactions. *Langmuir*, 32(49), 13234–13243.  
<https://doi.org/10.1021/acs.langmuir.6b02219>
- Sarkozy, G. (2001). Quinolones: a class of antimicrobial agents. *Veterinarni Medicina-Praha-*, 46(9/10), 257–274. [[Google Scholar](#)]
- Sarma, C., Rasane, P., Kaur, S., Singh, J., Singh, J., Gat, Y., Garba, U., Kaur, D., & Dhawan, K. (2018). Antioxidant and antimicrobial potential of selected varieties of *Piper betle* L. (Betel leaf). *Anais da Academia Brasileira de Ciências*, 90(4), 3871–3878.  
<https://doi.org/10.1590/0001-3765201820180285>
- SAS (2012). *User's Guide, 9.4 ed.*; SAS Institute, Inc.: Cary, NC, USA.
- Savon, L., Scull, I., Orta, M., & Martinez, M. (2006). Integral foliage meal for poultry feeding. I Chemical composition, physical properties, and phytochemical screening. *Revista Cubana de Ciencia Avícola*. 41:359–369. [[Google Scholar](#)]
- Savsani, H., Srivastava, A., Gupta, S., & Patel, K. (2020). Strengthening antioxidant defense & cardio protection by *Piper betle*: An in-vitro study. *Heliyon*, 6(1), e03041. <https://doi.org/10.1016/j.heliyon.2019.e03041>

- Sazili, A. Q., Parr, T., Sensky, P. L., Jones, S. W., Bardsley, R. G., & Buttery, P. J. (2005). The relationship between slow and fast myosin heavy chain content, calpastatin and meat tenderness in different ovine skeletal muscles. *Meat Science*, 69(1), 17–25.  
<https://doi.org/10.1016/j.meatsci.2004.06.021>
- Scanes, C. G. (2014). Protein *Metabolism*. *Sturkie's Avian Physiology, 6th ed.*; Academic Press, Elsevier Inc.: Waltham, MA, USA, 2014; pp. 455–468.  
[[Google Scholar](#)]
- Schar, D., Sommanustweechai, A., Laxminarayan, R., & Tangcharoensathien, V. (2018). Surveillance of antimicrobial consumption in animal production sectors of low-and middle-income countries: Optimizing use and addressing antimicrobial resistance. *PLoS medicine*, 15(3), e1002521.  
<https://doi.org/10.1371/journal.pmed.1002521>
- Scherer, R., Junior, S. B., de Albuquerque, R., & Godoy, H. T. (2014). Microencapsulated eucalyptol and eugenol as growth promoters in broilers. *Brazilian Journal of Food Research.*, 5, 26-32. [[Google Scholar](#)]
- Sebola, N. A., Mlambo, V., & Mokoboki, H. K. (2019). Chemical characterisation of *Moringa oleifera* (MO) leaves and the apparent digestibility of MO leaf meal-based diets offered to three chicken strains. *Agroforestry Systems*, 93(1), 149–160. <https://doi.org/10.1007/s10457-017-0074-9>
- Sgavioli, S., de Faria Domingues, C. H., Castiblanco, D. M. C., Praes, M. F. F. M., Andrade-Garcia, G. M., Santos, E. T., Baraldi-Artoni, S. M., Garcia, R. G., & Junqueira, O. M. (2016). Silicon in broiler drinking water promotes bone development in broiler chickens. *British Poultry Science*, 57(5), 693–698. <https://doi.org/10.1080/00071668.2016.1206190>
- Shah, S., Garg, G., Jhade, D., & Patel, N. (2016). Piper Betle: Phytochemical, Pharmacological and Nutritional Value in Health Management T Review. *International Journal of Pharmaceutical Sciences Review and Research*, 38(34), 181–189.  
<https://pdfs.semanticscholar.org/9d62/4399323d94ff6800221ef9b4d00b99b08ac9.pdf>
- Shahid, M., Dumat, C., Khalid, S., Schreck, E., Xiong, T., & Niazi, N. K. (2017). Foliar heavy metal uptake, toxicity and detoxification in plants: A comparison of foliar and root metal uptake. *Journal of Hazardous Materials*, 325, 36–58.  
<https://doi.org/10.1016/j.jhazmat.2016.11.063>
- Shallcross, L. J., & Davies, S. C. (2014). The World Health Assembly resolution on antimicrobial resistance. *Journal of Antimicrobial Chemotherapy*, 69(11), 2883–2885. <https://doi.org/10.1093/jac/dku346>
- Shang, Y., Regassa, A., Kim, J. H., & Kim, W. K. (2015). The effect of dietary

fructooligosaccharide supplementation on growth performance, intestinal morphology, and immune responses in broiler chickens challenged with *Salmonella* Enteritidis lipopolysaccharides. *Poultry Science*, 94(12), 2887–2897. <https://doi.org/10.3382/ps/pev275>

- Sharifi-Rad, M., Fokou, P. V. T., Sharopov, F., Martorell, M., Ademiluyi, A. O., Rajkovic, J., Salehi, B., Martins, N., Iriti, M., & Sharifi-Rad, J. (2018). Antiulcer agents: From plant extracts to phytochemicals in healing promotion. *Molecules*, 23(7), 1751. <https://doi.org/10.3390/molecules23071751>
- Sharma, O. P., & Bhat, T. K. (2009). DPPH antioxidant assay revisited. *Food chemistry*, 113(4), 1202-1205. <https://doi.org/10.1016/j.foodchem.2008.08.008>
- Sharma, G., Raturi, K., Dang, S., Gupta, S., & Gabrani, R. (2014). Combinatorial antimicrobial effect of curcumin with selected phytochemicals on *Staphylococcus epidermidis*. *Journal of Asian Natural Products Research*, 16(5), 535–541. <https://doi.org/10.1080/10286020.2014.911289>
- Sharma, U. K., Sharma, A. K., & Pandey, A. K. (2016). Medicinal attributes of major phenylpropanoids present in cinnamon. *BMC Complementary and Alternative Medicine*, 16(1), 1–11. <https://doi.org/10.1186/s12906-016-1147-4>
- Sharma, U. K., Kumar, R., Gupta, A., Ganguly, R., Singh, A. K., Ojha, A. K., & Pandey, A. K. (2019). Ameliorating efficacy of eugenol against metanil yellow induced toxicity in albino Wistar rats. *Food and Chemical Toxicology*, 126(October 2018), 34–40. <https://doi.org/10.1016/j.fct.2019.01.032>
- Shawle, K., Urge, M., & Animut, G. (2016). Effect of different levels of *Lepidium sativum* L. on growth performance, carcass characteristics, haematology and serum biochemical parameters of broilers. *SpringerPlus*, 5(1), 1-15. <https://doi.org/10.1186/s40064-016-3118-0>
- Sheoran, N., Kumar, R., Kumar, A., Batra, K., Sihag, S., Maan, S., & Maan, N. S. (2017). Nutrigenomic evaluation of garlic (*Allium sativum*) and holy basil (*Ocimum sanctum*) leaf powder supplementation on growth performance and immune characteristics in broilers. *Veterinary World*, 10(1), 121–129. <https://doi.org/10.14202/vetworld.2017.121-129>
- Shewita, R. S., & Taha, A. E. (2018). Influence of dietary supplementation of ginger powder at different levels on growth performance, haematological profiles, slaughter traits and gut morphometry of broiler chickens. *South African Journal of Animal Science*, 48(6). DOI: [10.4314/sajas.v48i6.1](https://doi.org/10.4314/sajas.v48i6.1)



- Shi, D., Bai, L., Qu, Q., Zhou, S., Yang, M., Guo, S., Li, Q., & Liu, C. (2019). Impact of gut microbiota structure in heat-stressed broilers. *Poultry Science*, 98(6), 2405–2413. <https://doi.org/10.3382/ps/pez026>
- Short, F. J., Gorton, P., Wiseman, J., & Boorman, K. N. (1996). Determination of titanium dioxide added as an inert marker in chicken digestibility studies. *Animal Feed Science and Technology*, 59(4), 215–221. [https://doi.org/10.1016/0377-8401\(95\)00916-7](https://doi.org/10.1016/0377-8401(95)00916-7)
- Sikder, K., Das, N., Kesh, S. B., & Dey, S. (2014). Quercetin and  $\beta$ -sitosterol prevent high fat diet induced dyslipidemia and hepatotoxicity in Swiss albino mice. *Indian Journal of experimental Biology*, 52, 60-66. [\[Google Scholar\]](#)
- Sim, O. P., Rasid, R. A., Hardy, N., Daud, A., David, D., Haya, B. A., Saibeh, K., Silip, J. J., Milan, A. R., & Alimon, A. R. (2019). Preliminary investigation on the chemical composition of local medicinal herbs (*Curcuma longa* L., *Persicaria odorata* L. and *Eleutherine palmifolia* L.) as potential layer feed additives for the production of healthy eggs. *Transactions on Science and Technology*, 6, 221-227. [\[Google Scholar\]](#)
- Singh, P., Jayaramaiah, R. H., Agawane, S. B., Vannuruswamy, G., Korwar, A. M., Anand, A., Dhaygude, V. S., Shaikh, M. L., Joshi, R. S., Boppana, R., & Kulkarni, M. J. (2016). Potential dual role of eugenol in inhibiting advanced glycation end products in diabetes: proteomic and mechanistic insights. *Scientific Reports*, 6(1), 1-13. <https://doi.org/10.1038/srep18798>
- Singh, H., Bhushan, S., Arora, R., Singh Buttar, H., Arora, S., & Singh, B. (2017). Alternative treatment strategies for neuropathic pain: Role of Indian medicinal plants and compounds of plant origin-A review. *Biomedicine and Pharmacotherapy*, 92, 634–650. <https://doi.org/10.1016/j.biopha.2017.05.079>
- Singh, D., Narayanamoorthy, S., Gamre, S., Majumdar, A. G., Goswami, M., Gami, U., Cherian, S., & Subramanian, M. (2018). Hydroxychavicol, a key ingredient of Piper betle induces bacterial cell death by DNA damage and inhibition of cell division. *Free Radical Biology and Medicine*, 120(March), 62–71. <https://doi.org/10.1016/j.freeradbiomed.2018.03.021>
- Singh, A., Dwivedy, A. K., Singh, V. K., Upadhyay, N., Chaudhari, A. K., Das, S., & Dubey, N. K. (2019). Essential oils based formulations as safe preservatives for stored plant masticatories against fungal and mycotoxin contamination: A review. *Biocatalysis and Agricultural Biotechnology*, 17(October 2018), 313–317. <https://doi.org/10.1016/j.bcab.2018.12.003>
- Sinurat, A. P., Wina, E., Rakhmani, S. I. W., Wardhani, T., Haryati, T., & Purwadaria, T. (2018). Bioactive substances of some herbals and their

effectiveness as antioxidant, antibacteria and antifungi. *Jurnal Ilmu Ternak Dan Veteriner*, 23(1), 18.  
<https://doi.org/10.14334/jitv.v23i1.1660>

- Sivareddy, B., Reginald, B., Sireesha, D., Samatha, M., Reddy, K. H., & Subrahmanyam, G. (2019). Antifungal activity of solvent extracts of Piper betle and Ocimum sanctum Linn on Candida albicans: An in vitro comparative study. *Journal of Oral and Maxillofacial Pathology*, 23(3), 333. [https://doi.org/10.4103/jomfp.JOMFP\\_167\\_19](https://doi.org/10.4103/jomfp.JOMFP_167_19)
- Skarp, C. P. A., Hänninen, M. L., & Rautelin, H. I. K. (2016). Campylobacteriosis: The role of poultry meat. *Clinical Microbiology and Infection*, 22(2), 103–109. <https://doi.org/10.1016/j.cmi.2015.11.019>
- Sohaib, M., Butt, M. S., Shabbir, M. A., & Shahid, M. (2015). Lipid stability, antioxidant potential and fatty acid composition of broilers breast meat as influenced by quercetin in combination with  $\alpha$ -tocopherol enriched diets. *Lipids in Health and Disease*, 14(1), 1–15. <https://doi.org/10.1186/s12944-015-0058-6>
- Soliman, N. (2019). Efficiency of Phytogenic Feed Additives in Improving Broiler Performance, Intestinal Bacteria and Ileal Histomorphology. *Egyptian Journal of Nutrition and Feeds*, 22(3), 625–635. <https://doi.org/10.21608/ejnf.2019.79498>
- Somporn, N., Saenthaweek, S., Naowaboot, J., Thaeomor, A., & Kukongviriyapan, V. (2018). Effect of lemongrass water extract supplementation on atherogenic index and antioxidant status in rats. *Acta Pharmaceutica*, 68(2), 185–197. <https://doi.org/10.2478/acph-2018-0015>
- Specialty Feed Additives. (2016). Market by type (flavors & sweeteners, minerals, binders, vitamins, acidifiers, antioxidants), livestock (swine, ruminants, poultry, aquatic animals), function, form, and region global forecast to 2022. *Research and Markets Report*, 173, ID: 3897708
- Srinivasa Rao, B., Chandrasekaran, C. V., Srikanth, H. S., Sasikumar, M., Edwin Jothie, R., Haseena, B., Bharathi, B., Selvam, R., & Prashanth, D. S. (2018). Mutagenicity and acute oral toxicity test for herbal poultry feed supplements. *Journal of Toxicology*, 2018. <https://doi.org/10.1155/2018/9412167>
- Stanley, D., Hughes, R. J., & Moore, R. J. (2014). Microbiota of the chicken gastrointestinal tract: influence on health, productivity and disease. *Applied Microbiology and Biotechnology*, 98(10), 4301–4310. <https://doi.org/10.1007/s00253-014-5646-2>
- Starcevic, K., Krstulović, L., Brozić, D., Maurić, M., Stojević, Z., Mikulec, Ž., Bajić, M., & Mašek, T. (2015). Production performance, meat composition and

oxidative susceptibility in broiler chicken fed with different phenolic compounds. *Journal of the Science of Food and Agriculture*, 95(6), 1172–1178. <https://doi.org/10.1002/jsfa.6805>

Stevanovic, Z. D., Bošnjak-Neumüller, J., Pajić-Lijaković, I., Raj, J., & Vasiljević, M. (2018). Essential oils as feed additives future perspectives. *Molecules*, 23(7), 1717. <https://doi.org/10.3390/molecules23071717>.

Sundang, M., Nasir, S. N. S., Sipaut, C. S., & Othman, H. (2012). Antioxidant Activity, Phenolic, Flavonoid and Tannin Content of Piper Betle and Leucosyke Capitella Murni. *Malaysian Journal of Fundamental & Applied Sciences*, 8(1), 1–6. [[Google Scholar](#)]

Suresh, G., Das, R. K., Kaur Brar, S., Rouissi, T., Avalos Ramirez, A., Chorfi, Y., & Godbout, S. (2018). Alternatives to antibiotics in poultry feed: molecular perspectives. *Critical Reviews in Microbiology*, 44(3), 318–335. <https://doi.org/10.1080/1040841X.2017.1373062>

Syahidah, A., Saad, C. R., Hassan, M. D., Rukayadi, Y., Norazian, M. H., & Kamarudin, M. S. (2017). Phytochemical Analysis , Identification and Quantification of Antibacterial Active Compounds in Betel Leaves , Piper betle Methanolic Extract. *Pakistan Journal of Biological Sciences*, 20(2), 70–81. <https://doi.org/10.3923/pjbs.2017.70.81>

Tahir, M., Chuzaemi, S., & Widodo, E. (2019). The Performance of Broilers given Eugenol of Clove Leaf Essential Oil as a Feed Additive. *Russian Journal of Agricultural and Socio-economic Sciences*, 95(11). [[Google Scholar](#)]

Talaty, P. N., Katanbaf, M. N., & Hester, P. Y. (2010). Bone mineralization in male commercial broilers and its relationship to gait score. *Poultry Science*, 89(2), 342–348. <https://doi.org/10.3382/ps.2009-00382>

Tang, K. L., Caffrey, N. P., Nóbrega, D. B., Cork, S. C., Ronksley, P. E., Barkema, H. W., Polachek, A. J., Ganshorn, H., Sharma, N., Kellner, J. D., & Checkley, S. L. (2019). Examination of unintended consequences of antibiotic use restrictions in food-producing animals: Sub-analysis of a systematic review. *One Health*, 7(March), 100095. <https://doi.org/10.1016/j.onehlt.2019.100095>

Taukoorah, U., Lall, N., & Mahomoodally, F. (2016). Piper betle L. (betel quid) shows bacteriostatic, additive, and synergistic antimicrobial action when combined with conventional antibiotics. *South African Journal of Botany*, 105, 133–140. <https://doi.org/10.1016/j.sajb.2016.01.006>

Teff, K. L., & Kim, S. F. (2011). Atypical antipsychotics and the neural regulation of food intake and peripheral metabolism. *Physiology and Behavior*, 104(4), 590–598. <https://doi.org/10.1016/j.physbeh.2011.05.033>

- Tehseen, M., Tahir, M., Khan, R. U., Jabbar, A., Ahmad, B., Ahsan, T., Khan, M. S., Khan, S., & Abudabos, A. M. (2016). Additive effect of *Nigella sativa* and *Zingiber officinale* herbal mixture on performance and cholesterol profile in Broilers. *Philippine Agricultural Scientist*, 99(4), 408–413. [[Google Scholar](#)]
- Thakuria, P., Nath, R., Sarma, S., Kalita, D., Dutta, D., Borah, P., Sharma, R., Barman, C., & Hussain, J. (2018). Phytochemical screening of medicinal plants occurring in local area of Assam. *Journal of Pharmacognosy and Phytochemistry JPP*, 186(73), 186–188. [[Google Scholar](#)]
- Thanan, R., Oikawa, S., Hiraku, Y., Ohnishi, S., Ma, N., Pinlaor, S., Yongvanit, P., Kawanishi, S., & Murata, M. (2014). Oxidative stress and its significant roles in neurodegenerative diseases and cancer. *International Journal of Molecular Sciences*, 16(1), 193–217. <https://doi.org/10.3390/ijms16010193>
- Thanganadar Appapalam, S., & Panchamoorthy, R. (2017). Aerva lanata mediated phytofabrication of silver nanoparticles and evaluation of their antibacterial activity against wound associated bacteria. *Journal of the Taiwan Institute of Chemical Engineers*, 78, 539–551. <https://doi.org/10.1016/j.jtice.2017.06.035>
- Thrall, M. A., Weiser, G., Allison, R., & Campbell, T. (2012). *Veterinary Haematology and Clinical Chemistry*, 2<sup>nd</sup> ed; John Wiley & Sons: Oxford, UK, 2012; pp. 582–598
- Toghyani, M., Toghyani, M., Gheisari, A., Ghalamkari, G., & Eghbalsaid, S. (2011). Evaluation of cinnamon and garlic as antibiotic growth promoter substitutions on performance, immune responses, serum biochemical and haematological parameters in broiler chicks. *Livestock Science*, 138(1–3), 167–173. <https://doi.org/10.1016/j.livsci.2010.12.018>
- Tonda, R. M., Rubach, J. K., Lumpkins, B. S., Mathis, G. F., & Poss, M. J. (2018). Effects of tannic acid extract on performance and intestinal health of broiler chickens following coccidiosis vaccination and/or a mixed-species *Eimeria* challenge. *Poultry Science*, 97(9), 3031–3042. <https://doi.org/10.3382/ps/pey158>
- Tothova, C., Nagy, O., & Kovac, G. (2016). Serum proteins and their diagnostic utility in veterinary medicine: A review. *Veterinari Medicina*, 61(9), 475–496. <https://doi.org/10.17221/19/2016-VETMED>
- Touchette, K. J. J. A., Carroll, G. L., Allee, R. L., Matteri, C. J., Dyer, L. A., Beausang, & M. E. Z. (2002). Effect of spray-dried plasma and lipopolysaccharide exposure on weaned pigs: II. Effects on the hypothalamic-pituitary-adrenal axis of weaned pigs. *Journal of Animal*

*Science*, 80(2), 502–509. <https://doi.org/10.2527/2002.802502x>

- Tuo, K., Béourou, S., Touré, A. O., Ouattara, K., Meité, S., Ako, A. A. Yao SS, Koffi D, Coulibay B, Coulibaly A, & Djaman, A. J. (2015). Antioxidant activities and estimation of the phenols and flavonoids content in the extracts of medicinal plants used to treat malaria in Ivory Coast. *International Journal of Current Microbiology and Applied Sciences*, 4(1), 862-874. [[Google Scholar](#)]
- Turcu, R. P., Panaite, T. D., Untea, A. E., Şoica, C., Iuga, M., & Mironeasa, S. (2020). Effects of Supplementing Grape Pomace to Broilers Fed Polyunsaturated Fatty Acids Enriched Diets on Meat Quality. *Animals*, 10(6), 947. <https://doi.org/10.3390/ani10060947>
- Umar R. A. N., Adani Sanusi, M. A. K., & Rohin, S. I. (2018). Chemical Composition and The Potential Biological Activities of Piper Betel-A Review. *Malaysian Journal of Applied Sciences*, 3(1), 1–8. [[Google Scholar](#)]
- Umatiya, R. V., Srivastava, A. K., Pawar, M. M., Chauhan, H. D., & Jain, A. K. (2018). Efficacy of garlic ( *Allium sativum* ) and ginger ( *Zingiber officinale* ) powder as phytogetic feed additive in diet of broiler chickens Efficacy of ginger ( *Zingiber officinale* ) and garlic ( *Allium sativum* ) powder as phytogetic feed additives in diet of. *Journal of Pharmacognosy and Phytochemistry*, 7(3), 1136–1140. [[Google Scholar](#)]
- Union, O., of, B. B.-P. L. in O. (2003), undefined. *Regulation (EC) No. 1831/2003 of European Parliament and the Council of 22 September 2003 on Additives for Use in Animal Nutrition*.
- Upadhyay, A., Upadhyaya, I., Karumathil, D. P., Yin, H. B., Nair, M. S., Bhattaram, V., Chen, C. H., Flock, G., Mooyottu, S., & Venkitanarayanan, K. (2015). Control of *Listeria monocytogenes* on skinless frankfurters by coating with phytochemicals. *LWT-Food Science and Technology*, 63(1), 37-42. <https://doi.org/10.1016/j.lwt.2015.03.100>
- Upadhyay, A., Arsi, K., Wagle, B. R., Upadhyaya, I., Shrestha, S., Donoghue, A. M., & Donoghue, D. J. (2017). Trans-cinnamaldehyde, carvacrol, and eugenol reduce *Campylobacter jejuni* colonization factors and expression of virulence genes in vitro. *Frontiers in Microbiology*, 8, 713. <https://doi.org/10.3389/fmicb.2017.00713>
- Upadhaya, S. D., & Kim, I. H. (2017). Efficacy of phytogetic feed additive on performance, production and health status of monogastric animals - A review. *Annals of Animal Science*, 17(4), 929–948. <https://doi.org/10.1515/aoas-2016-0079>
- Valenzuela-Grijalva, N. V., Pinelli-Saavedra, A., Muhlia-Almazan, A., Domínguez-Díaz, D., & González-Ríos, H. (2017). Dietary inclusion effects of

- phytochemicals as growth promoters in animal production. *Journal of Animal Science and Technology*, 59(1), 1–17.  
<https://doi.org/10.1186/s40781-017-0133-9>
- Valle, D. L., Puzon, J. J. M., Cabrera, E. C., & Rivera, W. L. (2016). Thin Layer Chromatography-Bioautography and Gas Chromatography-Mass Spectrometry of Antimicrobial Leaf Extracts from Philippine Piper betle L. against Multidrug-Resistant Bacteria. *Evidence-Based Complementary and Alternative Medicine*, 2016. <https://doi.org/10.1155/2016/4976791>
- Valsalam, S., Agastian, P., Arasu, M. V., Al-Dhabi, N. A., Ghilan, A. K. M., Kaviyarasu, K., Ravindran, B., Chang, S.W., & Arokiyaraj, S. (2019). Rapid biosynthesis and characterization of silver nanoparticles from the leaf extract of *Tropaeolum majus* L. and its enhanced in-vitro antibacterial, antifungal, antioxidant and anticancer properties. *Journal of Photochemistry and Photobiology B: Biology*, 191, 65–74.  
<https://doi.org/10.1016/j.jphotobiol.2018.12.010>
- Van Boeckel, T. P., Brower, C., Gilbert, M., Grenfell, B. T., Levin, S. A., Robinson, T. P., Teillant, A., Laxminarayan, R. (2015). Global trends in antimicrobial use in food animals. *Proceedings of the National Academy of Sciences*, 112(18), 5649-5654.  
<https://doi.org/10.1073/pnas.1503141112>
- Van Boeckel, T. P., Pires, J., Silvester, R., Zhao, C., Song, J., Criscuolo, N. G., Gilbert, M., Bonhoeffer, S., & Laxminarayan, R. (2019). Global trends in antimicrobial resistance in animals in low-and middle-income countries. *Science*, 365 (6459). DOI: 10.1126/science.aaw1944
- Van Wyk, A. S., & Prinsloo, G. (2020). Health, safety and quality concerns of plant-based traditional medicines and herbal remedies. *South African Journal of Botany*, 133, 54–62. <https://doi.org/10.1016/j.sajb.2020.06.031>
- Vase-Khavari, K., Mortezaei, S. H., Rasouli, B., Khusro, A., Salem, A. Z. M., & Seidavi, A. (2019). The effect of three tropical medicinal plants and superzist probiotic on growth performance, carcass characteristics, blood constituents, immune response, and gut microflora of broiler. *Tropical Animal Health and Production*, 51(1), 33–42.  
<https://doi.org/10.1007/s11250-018-1656-x>
- Venkadeswaran, K., Muralidharan, A. R., Annadurai, T., Ruban, V. V., Sundararajan, M., Anandhi, R., Thomas, P. A., & Geraldine, P. (2014). Antihypercholesterolemic and antioxidative potential of an extract of the plant, Piper betle, and its active constituent, eugenol, in triton WR-1339-induced hypercholesterolemia in experimental rats. *Evidence-Based Complementary and Alternative Medicine*, 2014.  
<https://doi.org/10.1155/2014/478973>

- Verlinden, M., Pasmans, F., Mahu, M., Maele, L. V., De Pauw, N., Yang, Z., Haesebrouck, F., & Martel, A. (2013). In vitro sensitivity of poultry *Brachyspira intermedia* isolates to essential oil components and in vivo reduction of *Brachyspira intermedia* in rearing pullets with cinnamaldehyde feed supplementation. *Poultry Science*, 92(5), 1202-1207.  
<https://doi.org/10.3382/ps.2012-02690>
- Vijayanchali, S. S. (2018). Nutrient Composition, Phytonutrient Constituents and Antioxidant Activity of the Dried Betel Leaves (Piper Betle). *International Journal for Research and Development*, 5(5).  
<https://ssrn.com/abstract=3345532>
- Vikram, P., Chiruvella, K. K., Ripain, I. H. A., & Arifullah, M. (2014). A recent review on phytochemical constituents and medicinal properties of kesum (*Polygonum minus* Huds.). *Asian Pacific Journal of Tropical Biomedicine*, 4(6), 430–435.  
<https://doi.org/10.12980/APJTB.4.2014C1255>
- Vitalini, S., Madeo, M., Tava, A., Iriti, M., Vallone, L., Avato, P., Cocuzza, C.E., Simonetti, P. and Argentieri, M. P. (2016). Chemical profile, antioxidant and antibacterial activities of *Achillea moschata* Wulfen, an endemic species from the Alps. *Molecules*, 21(7), 830.  
<https://doi.org/10.3390/molecules21070830>
- Vispute, M. M., Sharma, D., Mandal, A. B., Rokade, J. J., Tyagi, P. K., & Yadav, A. S. (2019). Effect of dietary supplementation of hemp (*Cannabis sativa*) and dill seed (*Anethum graveolens*) on performance, serum biochemicals and gut health of broiler chickens. *Journal of Animal Physiology and Animal Nutrition*, 103(2), 525–533.  
<https://doi.org/10.1111/jpn.13052>
- Viveros, A., Chamorro, S., Pizarro, M., Arijia, I., Centeno, C., & Brenes, A. (2011). Effects of dietary polyphenol-rich grape products on intestinal microflora and gut morphology in broiler chicks. *Poultry Science*, 90(3), 566-578.  
<https://doi.org/10.3382/ps.2010-00889>
- Vuong, C. N., Chou, W. K., Hargis, B. M., Berghman, L. R., & Bielke, L. R. (2016). Role of probiotics on immune function and their relationship to antibiotic growth promoters in poultry, a brief review. *International Journal of Probiotics & Prebiotics*, 11(1), 1-7.[[Google Scholar](#)]
- Wagle, B. R., Arsi, K., Shrestha, S., Upadhyay, A., Upadhyaya, I., Bhargava, K., Donoghue, A., & Donoghue, D. J. (2019). Eugenol as an antimicrobial wash treatment reduces *Campylobacter jejuni* in postharvest poultry. *Journal of Food Safety*, 39(6), e12704.  
<https://doi.org/10.1111/jfs.12704>

- Walsh, T. R. (2018). A one-health approach to antimicrobial resistance. *Nature microbiology*, 3(8), 854-855. <https://doi.org/10.1038/s41564-018-0208-5>
- Wan-Ibrahim, W. I., Sidik, K., & Kuppasamy, U. R. (2010). A high antioxidant level in edible plants is associated with genotoxic properties. *Food Chemistry*, 122(4), 1139–1144. <https://doi.org/10.1016/j.foodchem.2010.03.101>
- Wang, S., Yao, J., Zhou, B., Yang, J., Chaudry, M. T., Wang, M., Xiao, F., Li, Y., & Yin, W. (2018). Bacteriostatic effect of quercetin as an antibiotic alternative in vivo and its antibacterial mechanism in vitro. *Journal of Food Protection*, 81(1), 68–78. <https://doi.org/10.4315/0362-028X.JFP-17-214>
- Wang, Y., Yin, C., Wang, D., Huang, J., Ho, C. T., Zhou, Y., & Wan, X. (2018). Supplemental summer-autumn tea leaf (*Camellia sinensis*) improve the immune status of broilers. *Journal of Applied Animal Research*, 46(1), 1260-1267. [[Google Scholar](#)]
- Wang, M., Xiao, F. L., Mao, Y. J., Ying, L. L., Zhou, B., & Li, Y. (2019). Quercetin decreases the triglyceride content through the PPAR signalling pathway in primary hepatocytes of broiler chickens. *Biotechnology & Biotechnological Equipment*, 33(1), 1000-1010. <https://doi.org/10.1080/13102818.2019.1635528>
- Wang, H., Liang, S., Li, X., Yang, X., Long, F., & Yang, X. (2019). Effects of encapsulated essential oils and organic acids on laying performance, egg quality, intestinal morphology, barrier function, and microflora count of hens during the early laying period. *Poultry science*, 98(12), 6751-6760. <https://doi.org/10.3382/ps/pez391>
- Wati, T., Ghosh, T. K., Syed, B., & Haldar, S. (2015). Comparative efficacy of a phyto-genic feed additive and an antibiotic growth promoter on production performance, caecal microbial population and humoral immune response of broiler chickens inoculated with enteric pathogens. *Animal Nutrition*, 1(3), 213–219. <https://doi.org/10.1016/j.aninu.2015.08.003>
- Williamson, E. M., Liu, X., & Izzo, A. A. (2020). Trends in use, pharmacology, and clinical applications of emerging herbal nutraceuticals. *British Journal of Pharmacology*, 177(6), 1227–1240. <https://doi.org/10.1111/bph.14943>
- Wink, M. (2015). Modes of Action of Herbal Medicines and Plant Secondary Metabolites. *Medicines*, 2(3), 251–286. <https://doi.org/10.3390/medicines2030251>
- Wojcik, A., Ska, J. S. Ń., Witkowska, D., Y, Ł. C. A. Ż., & Piotrowska, J. (2011). Wp ł yw obrotu przedubojowego kurcz ą t brojlerów na jako ś ć technologiczn ą mi ę sa. *Inż. Ap. Chem.*, 2(3), 85–86. [[Google Scholar](#)]



- WHO. (2012). Evaluation of certain veterinary drug residues in food. *World Health Organization Technical Report Series*, (969).
- WHO. (2015). Antimicrobial resistance–SEA/RC68/R3 (No. SEA/RC68/R3) World Health Organization. [[Google Scholar](#)] Accessed on (28-01-2022)
- WHO. (2017). WHO guidelines on use of medically important antimicrobials in food-producing animals: web annex A: evidence base (No. WHO/NMH/FOS/ FZD/17.2). World Health Organization. [[Google Scholar](#)]
- WHO; FAO, ( 2018). Evaluation of Certain Veterinary Drug Residues in Food (Eighty-Fifth Report of the Joint FAO/WHO Expert Committee on Food Additives) WHO Technical Report Series, No 1008; WHO: Geneva, Switzerland, 2018; Available online: <https://apps.who.int/iris/bitstream/handle/10665/259895/9789241210171-eng.pdf;sessionid=E271946E46D0616CAC0C94420CA4E03B?sequence=1> (accessed on 16 August 2020).
- Xia, Y., Kong, J., Zhang, G., Zhang, X., Seviour, R., & Kong, Y. (2019). Effects of dietary inulin supplementation on the composition and dynamics of cecal microbiota and growth-related parameters in broiler chickens. *Poultry Science*, 98(12), 6942-6953. <https://doi.org/10.3382/ps/pez483>
- Xiong, Y., Tang, X., Meng, Q., & Zhang, H. (2016). Differential expression analysis of the broiler tracheal proteins responsible for the immune response and muscle contraction induced by high concentration of ammonia using iTRAQ-coupled 2D LC-MS/MS. *Science China Life Sciences*, 59(11), 1166-1176 <https://doi.org/10.1007/s11427-016-0202-8>
- Xu, D., Hu, M. J., Wang, Y. Q., & Cui, Y. L. (2019). Antioxidant activities of quercetin and its complexes for medicinal application. *Molecules*, 24(6), 1123. <https://doi.org/10.3390/molecules24061123>
- Yadav, S., & Jha, R. (2019). Strategies to modulate the intestinal microbiota and their effects on nutrient utilization, performance, and health of poultry. *Journal of Animal Science and Biotechnology*, 10(1), 1–11. <https://doi.org/10.1186/s40104-018-0310-9>
- Yang, Y., Iji, P. A., & Choct, M. (2009). Dietary Modulation Of Gut Microflora In Broiler Chickens: A Review Of The Role Of Six Kinds Of Alternatives To In-Feed Antibiotics. *World's Poultry Science Journal*, 65(01), 97. <https://doi.org/10.1017/S0043933909000008>
- Yang, C., Chowdhury, M. A. K., Hou, Y., & Gong, J. (2015). Phytochemicals as alternatives to in-feed antibiotics: Potentials and challenges in application. *Pathogens*, 4(1), 137–156. <https://doi.org/10.3390/pathogens4010137>

- Yang, J. Y., Zhang, H. J., Wang, J., Wu, S. G., Yue, H. Y., Jiang, X. R., & Qi, G. H. (2017). Effects of dietary grape proanthocyanidins on the growth performance, jejunum morphology and plasma biochemical indices of broiler chicks. *Animal*, *11*(5), 762-770.  
<https://doi.org/10.1017/S1751731116002056>
- Yang, H., Song, Y., Liang, Y. N., & Li, R. (2018). Quercetin treatment improves renal function and protects the kidney in a rat model of adenine-induced chronic kidney disease. *Medical Science Monitor*, *24*, 4760–4766.  
<https://doi.org/10.12659/MSM.909259>
- Yang, Y. F., Zhao, L. L., Shao, Y. X., Liao, X. D., Zhang, L. Y., Lin, L. U., & Luo, X. G. (2019). Effects of dietary graded levels of cinnamon essential oil and its combination with bamboo leaf flavonoid on immune function, antioxidative ability and intestinal microbiota of broilers. *Journal of Integrative Agriculture*, *18*(9), 2123-2132.  
[https://doi.org/10.1016/S2095-3119\(19\)62566-9](https://doi.org/10.1016/S2095-3119(19)62566-9)
- Yilmaz, B., & Li, H. (2018). Gut microbiota and iron: The crucial actors in health and disease. *Pharmaceuticals*, *11*(4), 1–20.  
<https://doi.org/10.3390/ph11040098>
- Yitbarek, melkamu B. (2015). Phytochemicals As Feed Additives In Poultry Production : A Review. *International Journal of Extensive Research*, *3*(1), 49–60.  
[\[Google Scholar\]](#)
- Yousefdoost, S., Samadi, F., Jafari, S. M., Ramezanzpour, S. S., Hassani, S., & Ganji, F. (2019). Application of nanoencapsulated silymarin to improve its antioxidant and hepatoprotective activities against carbon tetrachloride-induced oxidative stress in broiler chickens. *Livestock Science*, *228*, 177-186.  
<https://doi.org/10.1016/j.livsci.2019.08.015>
- Zagmutt, S., Guzmán, L., Orrego, R., Wehinger, S., & Leiva, E. (2016). Phenolic Compound Identification and Antioxidant Capacity of Alperujo Extracts from Region del Maule, Chile. *International Journal of Food Properties*, *19*(9), 2016–2025. <https://doi.org/10.1080/10942912.2015.1092162>
- Zaidan, M. R., Noor Rain, A., Badrul, A. R., Adlin, A., Norazah, A., & Zakiah, I. (2005). In vitro screening of five local medicinal plants for antibacterial activity using disc diffusion method. *Tropical Biomedicine*, *22*(2), 165–170. [\[Google Scholar\]](#)
- Zahira A., and Tamilmani, K (2016). Evaluation of Bioactive Compound Present in Piper Betle Linn. By Elution Chromatography Coupling Technique. *World Journal of Pharmacy and Pharmaceuticals Sciences*, *5*(5), 1405–1413. <https://doi.org/10.20959/wjpps20165-6786>

- Zakeri, B., & Wright, G. D. (2008). Chemical biology of tetracycline antibiotics. *Biochemistry and Cell Biology*, 86(2), 124-136.  
<https://doi.org/10.1139/O08-002>
- Zellweger, R. M., Carrique-Mas, J., Limmathurotsakul, D., Day, N. P. J., Thwaites, G. E., Baker, S., & Southeast Asia Antimicrobial Resistance Network. (2017). A current perspective on antimicrobial resistance in Southeast Asia. *Journal of Antimicrobial Chemotherapy*, 72(11), 2963-2972.  
<https://doi.org/10.1093/jac/dkx260>
- Zeng, Z., Zhang, S., Wang, H., & Piao, X. (2015). Essential oil and aromatic plants as feed additives in non-ruminant nutrition: A review. *Journal of Animal Science and Biotechnology*, 6(7), 40104.  
<https://doi.org/10.1186/s40104-015-0004-5>
- Zhang, L., Li, J., Yun, T. T., Qi, W. T., Liang, X. X., Wang, Y. W., & Li, A. K. (2015). Effects of pre-encapsulated and pro-encapsulated *Enterococcus faecalis* on growth performance, blood characteristics, and cecal microflora in broiler chickens. *Poultry Science*, 94(11), 2821-2830.  
<https://doi.org/10.3382/ps/pev262>
- Zhang, L. L., Zhang, L. F., Xu, J. G., & Hu, Q. P. (2017). Comparison study on antioxidant, DNA damage protective and antibacterial activities of eugenol and isoeugenol against several foodborne pathogens. *Food and Nutrition Research*, 61(1).  
<https://doi.org/10.1080/16546628.2017.1353356>
- Zhang, Y., Chen, S., Wei, C., Chen, J., & Ye, X. (2017). Proanthocyanidins from Chinese bayberry (*Myrica rubra* Sieb. et Zucc.) leaves regulate lipid metabolism and glucose consumption by activating AMPK pathway in HepG2 cells. *Journal of Functional Foods*, 29, 217-225.  
<https://doi.org/10.1016/j.jff.2016.12.030>
- Zhao, X., Guo, Y., Guo, S., & Tan, J. (2013). Effects of *Clostridium butyricum* and *Enterococcus faecium* on growth performance, lipid metabolism, and cecal microbiota of broiler chickens. *Applied Microbiology and Biotechnology*, 97(14), 6477-6488. <https://doi.org/10.1007/s00253-013-4970-2>
- Zhao, Y., Kumar, D., Prasad, D. N., Singh, R. K., & Ma, Y. (2015). Morphoanatomic, physicochemical, and phytochemical standardization with HPTLC fingerprinting of aerial parts of *Aerva lanata* (Linn) Juss ex Schult. *Journal of Traditional Chinese Medical Sciences*, 2(1), 39-44.  
<https://doi.org/10.1016/j.jtcms.2014.12.002>
- Zhao, L. L., Liao, X. D., Zhang, L. Y., Luo, X. G., & Lu, L. (2017). Bacteriostatic effects of plant extracts and their compounds on chicken pathogenic bacteria in vitro. *Chinese Journal of Animal Nutrition*, 29, 3277-3286.  
[\[Google Scholar\]](#)

- Zhou, H., Wang, C., Ye, J., Chen, H., & Tao, R. (2015). Effects of dietary supplementation of fermented *Ginkgo biloba* L. residues on growth performance, nutrient digestibility, serum biochemical parameters and immune function in weaned piglets. *Animal Science Journal*, 86(8), 790-799. <https://doi.org/10.1111/asj.12361>
- Zhou, Y., Mao, S., & Zhou, M. (2019). Effect of the flavonoid baicalein as a feed additive on the growth performance, immunity, and antioxidant capacity of broiler chickens. *Poultry Science*, 98(7), 2790–2799. <https://doi.org/10.3382/ps/pez071>
- Ziaie, H., Bashtani, M., & Torshizi, M. A. K. (2011). Effect of Antibiotic and its Alternatives on Morphometric Characteristics , Mineral Content and Bone Strength of Tibia in Ross Broiler Chickens. *Global Veterinaria*, 7(4), 315–322. [[Google Scholar](#)]
- Zou, W., Liu, W., Yang, B., Wu, L., Yang, J., Zou, T., Liu, F., Xia, L., & Zhang, D. (2015). Quercetin protects against perfluorooctanoic acid-induced liver injury by attenuating oxidative stress and inflammatory response in mice. *International Immunopharmacology*, 28(1), 129-135. <https://doi.org/10.1016/j.intimp.2015.05.043>
- Zywica, R., Charzynska, D. G., & Banach, J. K. (2011). Wpływ procesu oszłamiania elektrycznego kurcząt za pomocą urządzenia własnej konstrukcji na barwę mięsa. *Żywność Nauka Technologia Jakość*, 18(1), 52-67. [[Google Scholar](#)]