



UNIVERSITI PUTRA MALAYSIA

***DEVELOPMENT OF CHLOROPLAST GENOME LIBRARY FOR DNA
TRACKING AND CONSERVATION OF ENDANGERED AQUILARIA
SPECIES NATIVE TO MALAYSIA***

MUHAMMAD SYAHMI BIN HISHAMUDDIN

FPAS 2022 17



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MUHAMMAD SYAHMI BIN HISHAMUDDIN

Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Philosophy

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DEDICATION

In the name of ALLAH the Beneficent and the Merciful

Special dedication to:

Supervisory Committee

PROF. DR. ROZI MOHAMED

DR. SHAIRUL IZAN BINTI RAMLEE

DR. DHILIA UDIE LAMASUDIN

Parents

Kol. (B) JAMALUDDIN BIN AHAMAD & CHE ROHANA BINTI AHMAD

To my late father

Mej. (B) HISHAMUDDIN BIN HAJI KADIR

My wife and daughter

**NURSUHAILI BINTI ZAHARI & NURSAKINAH MADEENA BINTI
MUHAMMAD SYAHMI**

and

My siblings and friends

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

DEVELOPMENT OF CHLOROPLAST GENOME LIBRARY FOR DNA TRACKING AND CONSERVATION OF ENDANGERED AQUILARIA SPECIES NATIVE TO MALAYSIA

By

MUHAMMAD SYAHMI BIN HISHAMUDDIN

January 2022

Chair : Professor Rozi Mohamed, PhD
Faculty : Forestry and Environment

Aquilaria Lam., species [*Aquilaria beccariana* Tiegh., (*gaharu buaya*), *Aquilaria crassna* Pierre ex Lecomte, (*gaharu*), *Aquilaria hirta* Ridl., (*chandan bulu*), *Aquilaria malaccensis* Lam., (*ching karas*), *Aquilaria microcarpa* Baill., (*gaharu putih*), *Aquilaria rostrata* Ridl., (*candan gunung*), *Aquilaria sinensis* (*gaharu*) and *Aquilaria subintegra* Ding Hou, (*gaharu*)] is a naturally distributed in the Indomalesian region and are protected against over-exploitation. *Aquilaria* is highly prized for its unique scented resin, agarwood, which is often the subject of unlawful trade activities. Survival of the tree is heavily threatened by destructive harvesting and agarwood poaching, leading to its protection under the Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES). Ambiguous species delimitation and limited genetic information within *Aquilaria* are among the impediments to conservation efforts. In this study, a comparative analysis was performed on eight *Aquilaria* species complete chloroplast (cp) genomes, of which seven were newly sequenced using Illumina HiSeq X Ten platform followed by de novo assembly. All *Aquilaria* species utilised in this study were collected during fieldwork and the *Aquilaria* germplasm of Forest Research Institute of Malaysia (FRIM), and grown in the greenhouse of the Faculty of Forestry and Environment, Universiti Putra Malaysia. All *Aquilaria* species have been authenticated by experts. To conduct comparative cp genome analysis, the sequences of all eight *Aquilaria* cp genomes were aligned using MAFFT v7 and then imported into DnaSP v5.10.1 to determine nucleotide diversity in the total genome, LSC, SSC, and IR regions. The boundaries between the IR and SC regions were manually examined to determine the differences in length variation between *Aquilaria* cp genomes. *Aquilaria* cp genomes possess a typical quadripartite structure including gene order and genomic structure. The length of each of the cp genome is about 174 kbp and encoded between 89 and 92 proteins, 38 tRNAs, and 8 rRNAs, with 27 duplicated in the IR (inverted repeat) region. Besides, 832 repeats (forward, reverse, palindrome and complement repeats) and nine highly variable regions

(HVR) were also identified. The phylogenetic analysis performed using Maximum likelihood (ML) and Bayesian inference (BI) suggests that the topology structure of *Aquilaria* cp genomes were well presented with strong support values based on the cp genomes data set and matches their geographic distribution pattern. Five of the nine HVR (*matK-rps16*, *ndhF-rpl32*, *psbJ-petA*, *trnD*, and *trnT-trnL*) were selected based on a cut off value of >0.01. These regions were further analyzed using the neighbor-joining (NJ) method to assess their ability at discriminating the eight species. Coupled with in silico primer design, two potential barcoding regions, *psbJ-petA* and *trnT-trnL*, were identified. Their strengths in species delimitation were evaluated individually and in combination, via DNA barcoding analysis. The results showed that the combined dataset, *psbJ-petA+trnT-trnL*, effectively resolved members of the genus *Aquilaria* by clustering all species into their respective clades. In addition, this study show great potential for agarwood identification that the newly proposed DNA barcode was capable at identifying the species of origin of six commercial agarwood samples that were included as unknown samples. Such achievement offers a new technical advancement, useful in the combat against illicit agarwood trades and in assisting the conservation of these valuable species in natural populations. The identification of simple sequence repeats (SSR) across complete cp genome sequences from eight *Aquilaria* species has never been reported before. Perhaps because developing these genetic are regarded as laborious and expensive. To overcome this issue, the current work also aims to find SSRs in *Aquilaria* cp genomes using silico methods. For identification of the simple sequence repeats (SSRs), MISA PERL script which had a repeat length of 12 for mononucleotides (mono-), 6 for dinucleotides (di-), 4 for trinucleotides (tri-), 3 for tetranucleotides (tetra-), pentanucleotides (penta-), and hexanucleotides (hexa-), respectively, along with frequency were utilized. From a total of 312 SSRs that were discovered, merely 50 (16%) were found localized within the coding region while the majority (84%) were within the intergenic regions, with an average of one SSR per 4.5 kb. The mean length of the SSRs were 11.63 bp. Mono- repeats were the predominant motifs (29.2%), followed by tetra- (28.8%), di- (20.5%), tri- (19.9%), and penta- (1.6%). Whereas the most recurring motifs were A/T (97.8%) for mono-, AT/AT (87.5%) for di-, AAT/ATT (48.4%) for tri-, and AAAT/ATT (45.6%) for tetra-. GO analysis using the REVIGO software identified four molecular functions, six biological processes and three cellular components. In conclusion, findings of this SSR offer a scientific foundation for future phylogenetics, evolutionary genetics, diversity studies and breeding programs on *Aquilaria* species.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PEMBANGUNAN PERPUSTAKAAN DATA GENOM KLOROPLAS BAGI
MENJEJAKI DNA DAN PEMULIHARAAN SPESIS *AQUILARIA*
(TERANCAM) DI MALAYSIA**

Oleh

MUHAMMAD SYAHMI BIN HISHAMUDDIN

Januari 2022

Pengerusi : Profesor Rozi Mohamed, PhD
Fakulti : Perhutanan dan Alam Sekitar

Aquilaria spesis [*Aquilaria beccariana* Tiegh., (*gaharu buaya*), *Aquilaria crassna* Pierre ex Lecomte, (*gaharu*), *Aquilaria hirta* Ridl., (*chandan bulu*), *Aquilaria malaccensis* Lam., (*ching karas*), *Aquilaria microcarpa* Baill., (*gaharu putih*), *Aquilaria rostrata* Ridl., (*candan gunung*), *Aquilaria sinensis* (*gaharu*) and *Aquilaria subintegra* Ding Hou, (*gaharu*)] tersebar secara semula jadi di wilayah Indomalesia dan dilindungi daripada eksplorasi berlebihan. *Aquilaria* sangat berharga kerana resinnya yang unik dan berbau wangi, gaharu, yang sering terlibat dalam aktiviti perdagangan yang tidak sah. Kelangsungan hidup pokok ini sangat terancam dengan penuaian yang merosakkan dan perburuan gaharu secara besar-besaran, menyebabkan ianya dilindungi dan disenaraikan dibawah Konvensyen Perdagangan Antarabangsa Spesies Fauna dan Flora Liar yang Terancam (CITES). Persempadan spesies yang kurang jelas dan maklumat genetik yang terhad dalam *Aquilaria* adalah antara halangan usaha pemuliharaan. Dalam kajian ini, analisis perbandingan dilakukan terhadap lapan urutan genom kloroplas lengkap (cp) spesies *Aquilaria*, di mana tujuh daripadanya baru disusun mengikut urutan menggunakan platform Illumina HiSeq X Ten diikuti dengan pemasangan de novo. Semua spesies *Aquilaria* yang digunakan dalam kajian ini telah diperolehi semasa kerja lapangan dan juga daripada germplasma *Aquilaria* Institut Penyelidikan Perhutanan Malaysia (FRIM), dan ditanam di rumah hijau Fakulti Perhutanan dan Alam Sekitar, Universiti Putra Malaysia. Semua spesies *Aquilaria* telah disahkan oleh pakar akan ketulenan spesisnya. Untuk menjalankan analisis perbandingan genom kloroplas, jujukan kesemua lapan genom kloroplas *Aquilaria* telah diselaraskan menggunakan MAFFT v7 dan kemudian diimport ke dalam DnaSP v5.10.1 untuk menentukan kepelbagaiannya nukleotida dalam keseluruhan genom, LSC, SSC dan wilayah IR. Sempadan antara kawasan IR dan SC telah diperiksa secara manual untuk menentukan perbezaan variasi panjang antara genom kloroplas *Aquilaria*. Genom kloroplas *Aquilaria* mempunyai struktur kuadripartit termasuk susunan gen dan struktur genom. Panjang setiap genom kloroplas adalah kira-

kira 174 kbp dan dikodkan antara 89 dan 92 protein, 38 tRNA, dan 8 rRNA, dengan 27 pengulangan di wilayah IR (ulangan terbalik). Selain itu, 832 pengulangan (ke depan, bertentangan, palindrom dan pengulangan pelengkap) dan sembilan kawasan yang sangat berubah (HVR) juga dikenal-pasti. Analisis filogenetik yang dilakukan menggunakan kaedah kemungkinan maksimum (ML) dan Inferens Bayesian (BI) menunjukkan bahawa struktur topologi genom kloroplas *Aquilaria* dibentangkan dengan baik dengan nilai sokongan yang kuat berdasarkan set data genom kloroplas dan sepadan dengan corak taburan geografi mereka. Lima daripada sembilan HVR (*matK-rps16*, *ndhF-rp32*, *psbj-petA*, *trnD*, and *trnT-trnL*) dipilih berdasarkan nilai pemotongan > 0.01 . Kawasan-kawasan ini selanjutnya dianalisa menggunakan kaedah hubung-kait jiran (NJ) untuk menilai kemampuan mereka dalam mendiskriminasi lapan spesies. Ditambah dengan reka bentuk primer yang dihasilkan secara *in-silico*, dua wilayah barkod yang berpotensi, *psbj-petA* dan *trnT-trnL*, dikenal-pasti. Kekuatan mereka dalam penghadan sempadan spesies dinilai secara individu dan gabungan, melalui analisis barkoding DNA. Hasil kajian menunjukkan bahawa kumpulan data gabungan antara, *psbj-petA* + *trnT-trnL*, berjaya menyelesaikan anggota genus *Aquilaria* dengan mengelaskan semua spesies ke dalam kelompok masing-masing. Di samping itu, kajian ini menunjukkan bahawa barkod DNA yang dicadangkan mampu mengenal-pasti spesies asal enam sampel gaharu komersial yang dimasukkan sebagai sampel yang tidak diketahui. Pencapaian ini memberikan kemajuan teknikal baru, berguna dalam memerangi perdagangan gaharu secara haram dan dapat membantu pemuliharaan spesies berharga ini dalam populasi semula jadi. Pengenalpastian ulangan jujukan mudah (SSR) merentasi jujukan genom kloroplas lengkap daripada lapan spesies *Aquilaria* tidak pernah dilaporkan sebelum ini. Mungkin kerana membangunkan genetik ini dianggap rumit dan melibatkan kos yang mahal. Untuk mengatasi isu ini, kajian ini dilakukan bertujuan untuk mencari SSR dalam genom kloroplas *Aquilaria* menggunakan kaedah silico. Untuk mengenal pasti ulangan jujukan mudah (SSR), skrip MISA PERL yang mempunyai panjang ulangan 12 untuk mononukleotida (mono-), 6 untuk dinukleotida (di-), 4 untuk trinukleotida (tri-), 3 untuk tetranukleotida (tetra-), pentanukleotida (penta-), dan heksanukleotida (hexa-), telah digunakan. Daripada sejumlah 312 SSR yang ditemui, hanya 50 (16%) ditemui dalam kawasan pengekodan manakala majoriti (84%) berada dalam kawasan intergenik, dengan purata satu SSR setiap 4.5 kb. Purata panjang SSR ialah 11.63 bp. Mono- ulangan adalah motif utama (29.2%), diikuti oleh tetra- (28.8%), di- (20.5%), tri- (19.9%), dan penta- (1.6%). Manakala motif yang paling berulang ialah A/T (97.8%) untuk mono-, AT/AT (87.5%) untuk di-, AAT/ATT (48.4%) untuk tri-, dan AAAT/ATTT (45.6%) untuk tetra-. Analisis GO menggunakan perisian REVIGO mengenal pasti empat fungsi molekul, enam proses biologi dan tiga komponen selular. Kesimpulannya, penemuan SSR ini menawarkan asas saintifik untuk filogenetik, genetik evolusi, kajian kepelbagai dan program pembiakan spesies *Aquilaria*.

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**SPECIALLY DEDICATED TO MY LATE FATHER,
Mej. (B) Haji Hishamuddin Bin Haji Kadir**

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LIST OF ABBREVIATIONS

UPM	Universiti Putra Malaysia
°C	degree Celsius
+ F	base frequencies
+ G4	Gamma model with default 4 rate categories
%	Percentage
s	second
min	minutes
Ta	Annealing Temperature
Tm	Melting Temperature
ng	Nanogram
µL	Microliter
µmol/L	Micromole per litre
w/v	Weight per Volume
AFLP	Amplified Fragment-Length Polymorphism
AS	Agarwood Stick
AT	Agarwood Tea
AWB	Agarwood Wood Block
AWC	Agarwood Wood Chips
Bar-HRM	Barcode-High-Resolution Melting
BI	Bayesian inference
bp	Base Pair
C	Complement
CBOL	The Consortium for the Barcode of Life-Plant Working Group
CDS	Coding Sequence

cemA	chloroplast envelope membrane protein
CE	Capillary Electrophoresis
CO1	cytochrome c oxidase 1
cp	Chloroplast
cpDNA	Chloroplast DNA
cpSSRs	Chloroplast SSRs
CITES	Convention on International Trade in Endangered Species of Wild Fauna and Flora
CTAB	cetyltrimethylammonium bromide
di-	Dinucleotide
DNA	Deoxyribonucleic Acid
DNASP	DNA Sequence Polymorphism
F	Forward
FASTA	Fast Alignment
FDR	False Discovery Rate
FRIM	Forest Research Institute of Malaysia
Gb	Gigabyte
GC content	guanine-cytosine content
GO	Gene Ontology
h	Hour
HVRs	Highly Variable Regions
IGS	Intergenic Spacer
IUCN	International Union for Conservation of Nature and Natural Resources
ITS	Internal Transcribed Spacer
ITS1	Internal Transcribed Spacer 1
ITS2	Internal Transcribed Spacer 2

IR	Inverted Repeat
ISSR	Inter-Simple Sequence Repeat
K2P	Kimura Two-Parameter
kb	Kilobases
kbp	Kilobase pairs
LSC	Large Single Copy
MAFFT	Multiple Sequence Alignment
matK	maturaseK
Mb	Million Base
MCMC	Markov chain Monte Carlo
MISA	Microsatellite Finder
mg	Milligram
mL	Millilitre
ML	Maximum likelihood
mono-	Mononucleotide
mtDNA	Mitochondrial DNA
mtSSRs	Mitochondrial SSRs
nSSRs	Nuclear SSRs
NCBI	National Center for Biotechnology Information
NJ	Neighbor Joining
NGS	Next Generation Sequencing
OGDRAW	OrganellarGenomeDRAW
ORFs	Open Reading Frames
P	Palindrome
PCR	Polymerase Chain Reaction
Pi	Nucleotide Variability

PIS	Parsimony Informative Sites
PIS2V	Parsimony Informative Sites with Two Variants
PIS3V	Parsimony Informative Sites with Three Variants
PVP	Polyvinylpyrrolidone
R	Reverse
rRNA	Ribosomal RNA
RAPD	Random Ampified Polymorphic DNA
RNA	Ribonucleic Acid
ROS	Reactive Oxygen Species
SH-aLRT	SH-like Approximate Likelihood-Ratio Test
SNP	Single Nucleotide Polymorphism
spp	Species
SRAP	Sequence-Related Amplified Polymorphism
SSC	Small Single Copy
SSR	Simple Sequence Repeat
SVS	Singleton Variable Sites
SV2V	Singleton Variable with Two Variants
SV3V	Singleton Variable with Three Variants
tri-	Trinucleotide
tRNA	Transfer Ribonucleic Acid
TVM	Transversion Model
UFboot	Ultrafast Bootstrapping Algorithm

CHAPTER 1

INTRODUCTION

1.1 General

Aquilaria Lam. (Thymelaeaceae) is an evergreen tropical forest tree endemic to the Indomalesia region (Whitmore, 1972). This tree is well-known for producing aromatic, rare and precious wood, commonly known as agarwood (*gaharu*) that has significant industrial values. The woodchip of agarwood is processed into incense powder, sticks, cubes, cosmetic accessories like bracelets and pendants, and hygienic goods, such as shampoo and soap and scented balms, and perfumes essential oil (Snelder and Lasco, 2008). There are 21 accepted species in the genus *Aquilaria*. Five of them, namely *Aquilaria beccariana* Tiegh., *Aquilaria hirta* Ridl., *Aquilaria malaccensis* Lam., *Aquilaria microcarpa* Baill., and *Aquilaria rostrata* Ridl., are reported to be native to Malaysia (Lee and Mohamed, 2016a). *Aquilaria rostrata* Ridl., an endemic species to Peninsular Malaysia, was previously thought to be extinct until it was rediscovered in 2015, nearly 100 years after its first record in 1911 (Lee and Mohamed, 2016b).

Currently, all the species in this genus are listed in Appendix II of CITES and are also classified as vulnerable to extinction on the Red List of the International Union for Conservation of Nature (UNEP-WCMC, 2019). Consequently, agarwood trade is strictly controlled by international legislation, as these species are listed in Appendix II. Commercial activities require specific permits, and traders frequently attempt to circumvent these regulations through different means, including illegal ways. Similar to other plant species, the identification of *Aquilaria* species is mostly based on its physical and morphometric traits, specifically on the floral reproductive structures (Lee and Mohamed, 2016a). However, due to its appearance and phenotypic plasticity, agarwood is relatively easy to be separated from wood from other genera by hand lens or microscopy, but difficult or impossible for species identification using wood anatomy (Yin et al., 2016). In general, the identification of agarwood species using the naked eye is a nearly impossible and difficult task.

The DNA sequence of an organism provides an effective and efficient tool for species identification (Hollingsworth et al., 2011). As a result, several researchers have attempted to develop a library of useful gene sequences for *Aquilaria* species delineation. Previous studies showed that the *trnL-trnF* region is useful in examining the phylogenetic relationships between different *Aquilaria* species (Eurlings and Gravendeel, 2005). Thus, this genomic region is potentially useful as biomarker in distinguishing degraded agarwood products by using a real-time polymerase chain reaction (PCR) approach (Mohamed et al., 2012). It was also reported that the DNA nuclear ribosomal internal transcribed spacer 1 (ITS1) region and the intergenic spacer region *psbC-trnS* could differentiate between different *Aquilaria* species (Ito and Honda, 2005). In

addition to the identification techniques based on gene markers, the advent of the DNA barcoding technique shed light on *Aquilaria* species identification and bolstering traditional wood identification. However, this initiative is not always attainable at the species level. The feasibility of DNA barcoding on *Aquilaria* wood samples was tested by confirming the limitation of extracting useful genomic information through the amplification process performed on heat-treated samples (Jiao et al., 2014). Furthermore, the combination loci of *trnL-trnF+ITS* and *trnL-trnF+ITS2* were reported as potential DNA barcoding loci for *Aquilaria* species discrimination and identification of agarwood-related products (Lee et al., 2016a). Despite the potential use of single gene markers, there are still limitations in identifying very closely related species, necessitating further investigation by using alternative DNA resources, such as the chloroplast (cp) genome.

The chloroplast genomes exist in a semi-autonomous state relative to the nuclear genome, having their own genome and gene expression system. Chloroplast genes are derived from subsets of endosymbiont genes, the products of which regulate their own gene transcription. These genes remain confined to a single membrane-bound compartment, together with their protein products (Allen, 2015). In addition, the cp genomes also comprise a separate transcription and transport mechanism encoded by ribosomal proteins related to photosynthesis (Bungard, 2004). Although the cp genome is significantly smaller than the nuclear genome, each cell has around 400 to 1600 copies. These numbers are enough to provide an important foundation for the research of species relationships, evolutionary and genetic polymorphisms. The comparative study of the cp genome is crucial for the evolution of chloroplasts and the discovery of their basic variation (Bennett, 1987). To date, most of the study on the cp genome were one-sided and focused only on individual area or were based on specific species. The comparative researches on the cp genome levels between related species are not widely studied and available.

Previous researcher had reported the challenges in using universal DNA barcoding genes to distinguish and identify highly evolutionarily related *Aquilaria* species (Lee et al., 2016a). Therefore, this study is proposing to sequence the cp genome as this genome is smaller than the whole genome, therefore the sequencing data are more manageable. In addition, the genome is potentially valuable in giving numerous highly polymorphic loci for discriminating between various *Aquilaria* species. In other words, this study aims to develop a cp genome library based on five native *Aquilaria* species in Malaysia. The suitable regions from each species will be identified to develop DNA barcodes, and a DNA tracking system will be developed. The identification of potential DNA barcode sites is essential in developing conservation plans for each species. Furthermore, the library will provide a solid platform for future researches, such as by identifying *Aquilaria* evolutionary history and ecophysiology.

1.2 Problem Statement

The demand for natural agarwood on the international market has continued to rise over the years (Azren et al., 2019). The high prices range from US\$100/kg for low grade to US\$100,000/kg for good grade, and the lucrative profits from the sales of wild agarwood have resulted in massive illegal hunting and harvesting activities of *Aquilaria* trees (Ismail et al., 2015). These actions further lowered the number of wild *Aquilaria* that can reproduce naturally and, as a consequence, gradually reduced their population. Due to the rapid reduction in wild *Aquilaria* species populations, nearly all the *Aquilaria* species are now being placed on the Appendix II (CITES), including five Malaysian *Aquilaria* species, namely *A. beccariana*, *A. hirta*, *A. malaccensis* Lam., *A. microcarpa*, and *A. rostrata* Ridl.. Three of these five *Aquilaria* species have been listed as endangered (*A. microcarpa*) and critically endangered (*A. malaccensis* and *A. rostrata*). Even though most nations have stringent cross-border monitoring, there are still reports of illegal international agarwood commencing and cross-border smuggling. One main reason contributing to this is that different forms of agarwood, ranging from powders to hardwood beams, are easily blended with non-protected species, thus foiling the detection. The development of a complete cp genome of *Aquilaria* sequence database, especially from five *Aquilaria* species native to Malaysia, can aid in enhancing species-specific molecular techniques for differentiating between various *Aquilaria* species. This initiative is critical for the agarwood sector to ensure consumers right, authenticate market products, and effectively combat agarwood trade fraud.

1.3 Justification

The decreased population of natural *Aquilaria* and the listing of *Aquilaria* as an endangered species have prompted the researchers, naturalists, and conservationists to enhance species gene pool resources to conserve and protect this genus from extinction. To successfully establish programmed reproduction and characterize the genetic information from the natural *Aquilaria* population, the developments of a complete cp genome sequences database and the research on data utilization from this database are critical for conservation efforts. With the availability of a complete cp genome, the researchers can increase the efficacy of traditional methods for identifying *Aquilaria* species that are based on a partial genome sequence. The complete cp genome contains a huge quantity of genetic information, thus is ideal for studying genetical and structural varieties of natural populations. DNA-based studies on *Aquilaria* species began in 2005 after TRAFFIC emphasized the importance of conserving the natural *Aquilaria* resources in the wild, while the studies based on the cp genome of *Aquilaria* were started only in 2016.

The study of the complete cp genome can assist in the understanding of tree life, evolution, taxonomy, and genetic diversity. Cp genome is widely used in phylogenetic studies, as it is maternally inherited and is recombination-free. The highly conserved structure of cp genome facilitates sequencing and identification

of potential barcodes useful for plant species identification. In this study, genetic information of these endangered tree species are sequenced as a necessary step towards conserving their natural habitat and genetic diversity. Adequate genetic information and an understanding of species identification techniques will aid in the conservation efforts, specifically by assisting in the establishment of appropriate breeding programs and timber trade controls.

1.4 Research Objectives

The general objective of this study is to develop a complete sequences database of chloroplast genomes retrieved from five *Aquilaria* species that are native to Malaysia

Specific objectives:

1. To conduct a comparative analysis between the chloroplast genomes of eight *Aquilaria* species, including the characterization of chloroplast genomes, the identification of interspecies sequence variations and highly polymorphic regions in the library, followed by the determination of their molecular placements in phylogenetic trees;
2. To generate DNA barcodes from complete chloroplast genomes of *Aquilaria* species and demonstrate their utility for agarwood species tracking;
3. To identify the cross-transferability of simple sequence repeats (SSRs) in the chloroplast genomes between eight *Aquilaria* species.

REFERENCES

- Akkaya, M. S., Bhagwat, A. A., & Cregan, P. B. (1992). Length polymorphisms of simple sequence repeat DNA in soybean. *Genetics*, 132(4), 1131-1139. Akkaya, M.S., Bhagwat, A.A. and Cregan, P.B., 1992. Length polymorphisms of simple sequence repeat DNA in soybean. *Genetics*, 132(4), 1131-1139.
- Allen, G. C., Flores-Vergara, M. A., Krasynanski, S., Kumar, S., & Thompson, W. F. (2006). A modified protocol for rapid DNA isolation from plant tissues using cetyltrimethylammonium bromide. *Nature protocols*, 1(5), 2320-2325. Allen, G.C., Flores-Vergara, M.A., Krasynanski, S., Kumar, S. and Thompson, W.F., 2006. A modified protocol for rapid DNA isolation from plant tissues using cetyltrimethylammonium bromide. *Nature Protocols*, 1(5), 2320-2325.
- Allen, J. F. (2015). Why chloroplasts and mitochondria retain their own genomes and genetic systems: colocation for redox regulation of gene expression. *Proceedings of the National Academy of Sciences*, 112(33), 10231-10238.
- Alves, C., Pereira, R., Prieto, L., Aler, M., Amaral, C. R., Arévalo, C., Berardi, G., Di Rocco, F., Caputo, M., Carmona, C.H. & Pereira, F. (2017). Species identification in forensic samples using the SPInDel approach: a GHEP-ISFG inter-laboratory collaborative exercise. *Forensic Science International: Genetics*, 28, 219-224.
- Anderson, J.A.R., (1980). A checklist of the trees of Sarawak. Kuching: Forest Department, Sarawak.
- Antonopoulou, M., Compton, J. A. M. E. S., Perry, L. S., & Al-Mubarak, R. A. Z. A. N. (2010). *The trade and use of agarwood (Oudh) in the United Arab Emirates*. Selangor: TRAFFIC Southeast Asia.
- Arenas, M., Pereira, F., Oliveira, M., Pinto, N., Lopes, A. M., Gomes, V., Carracedo, A. & Amorim, A. (2017). Forensic genetics and genomics: much more than just a human affair. *PLoS Genetics*, 13(9), e1006960.
- Azhari, H., Mohamad, A. and Othman, R., 2015, September. Molecular identification of *Aquilaria* spp. by using inter-simple sequence repeat (ISSR). In AIP Conference Proceedings (Vol. 1678, No. 1, p. 030030). AIP Publishing.
- Azhari, H., Mohamad, A., & Othman, R. (2015). In AIP Conference Proceedings (Vol. 1678, No. 1, p. 030030): Molecular identification of *Aquilaria* spp. by using inter-simple sequence repeat (ISSR). AIP Publishing LLC.
- Azren, P. D., Lee, S. Y., Emang, D., & Mohamed, R. (2019). History and perspectives of induction technology for agarwood production from

cultivated *Aquilaria* in Asia: a review. *Journal of Forestry Research*, 30(1), 1-11.

Barden, A., Anak, N. A., Mulliken, T., & Song, M. (2000). Heart of the matter: agarwood use and trade and CITES implementation for *Aquilaria malaccensis*. *TRAFFIC International, Cambridge, UK*.

Beaumont, A. J., Edwards, T. J., Manning, J., Maurin, O., Rautenbach, M., Motsi, M. C., Fay, M.F., Chase, M.W. & Van Der Bank, M. (2009). Gnidia (Thymelaeaceae) is not monophyletic: taxonomic implications for Thymelaeoideae and a partial new generic taxonomy for Gnidia. *Botanical Journal of the Linnean Society*, 160(4), 402-417.

Bell, K. L., Burgess, K. S., Okamoto, K. C., Aranda, R., & Brosi, B. J. (2016). Review and future prospects for DNA barcoding methods in forensic palynology. *Forensic Science International: Genetics*, 21, 110-116.

Bennett, M. D. (1987). Variation in genomic form in plants and its ecological implications. *New Phytologist*, 106, 177-200.

Bi, Y., Zhang, M. F., Xue, J., Dong, R., Du, Y. P., & Zhang, X. H. (2018). Chloroplast genomic resources for phylogeny and DNA barcoding: a case study on *Fritillaria*. *Scientific Reports*, 8(1), 1-12.

Bock, R., & Knoop, V. (Eds.). (2012). *Genomics of Chloroplasts and Mitochondria* (Vol. 35). Springer Science & Business Media.

Bosmali, I., Ganopoulos, I., Madesis, P. and Tsafaris, A., (2012). Microsatellite and DNA-barcode regions typing combined with High Resolution Melting (HRM) analysis for food forensic uses: A case study on lentils (*Lens culinaris*). *Food Research International*, 46(1), 141-147.

Browne, F. G. (1955). Forest trees of Sarawak and Brunei and their products. *Forest trees of Sarawak and Brunei and their products*.

Bruni, I., Galimberti, A., Caridi, L., Scaccabarozzi, D., De Mattia, F., Casiraghi, M., & Labra, M. (2015). A DNA barcoding approach to identify plant species in multiflower honey. *Food Chemistry*, 170, 308-315.

Bungard, R. A. (2004). Photosynthetic evolution in parasitic plants: insight from the chloroplast genome. *BioEssays*, 26(3), 235-247.

Buschiazzo, E., & Gemmell, N. J. (2006). The rise, fall and renaissance of microsatellites in eukaryotic genomes. *Bioessays*, 28(10), 1040-1050.

Cavagnaro, P. F., Senalik, D. A., Yang, L., Simon, P. W., Harkins, T. T., Kodira, C. D., Huang, S. & Weng, Y. (2010). Genome-wide characterization of simple sequence repeats in cucumber (*Cucumis sativus* L.). *BMC Genomics*, 11(1), 1-18.

- Chan, C. X., & Ragan, M. A. (2013). Next-generation phylogenomics. *Biology Direct*, 8(1), 1-6.
- Chen, C. H., Kuo, T. C. Y., Yang, M. H., Chien, T. Y., Chu, M. J., Huang, L. C., Chen, C.Y., Lo, H.F., Jeng, S.T. & Chen, L. F. O. (2014). Identification of cucurbitacins and assembly of a draft genome for *Aquilaria agallocha*. *BMC Genomics*, 15(1), 1-11.
- Chen, H., Wang, L., Wang, S., Liu, C., Blair, M. W., & Cheng, X. (2015a). Transcriptome sequencing of mung bean (*Vigna radiata* L.) genes and the identification of EST-SSR markers. *PloS One*, 10(4), e0120273.
- Chen, J., Hao, Z., Xu, H., Yang, L., Liu, G., Sheng, Y., Zheng, C., Zheng, W., Cheng, T. & Shi, J. (2015b). The complete chloroplast genome sequence of the relict woody plant *Metasequoia glyptostroboides* Hu et Cheng. *Frontiers in Plant Science*, 6, 447.
- Chen, J., Zhao, J., Erickson, D. L., Xia, N., & Kress, W. J. (2015c). Testing DNA barcodes in closely related species of *C urcumoides* (Zingiberaceae) from Myanmar and *C hina*. *Molecular Ecology Resources*, 15(2), 337-348.
- Cheng, J., Zhao, Z., Li, B., Qin, C., Wu, Z., Trejo-Saavedra, D. L., Luo, X., Cui, J., Rivera-Bustamante, R.F., Li, S. & Hu, K. (2016). A comprehensive characterization of simple sequence repeats in pepper genomes provides valuable resources for marker development in *Capsicum*. *Scientific Reports*, 6(1), 1-12.
- Cho, W. B., Han, E. K., Choi, G., & Lee, J. H. (2018). The complete chloroplast genome of *Daphne kiusiana*, an evergreen broad-leaved shrub on Jeju Island. *Conservation Genetics Resources*, 10(1), 103-106. Species *Aquilaria malaccensis* (Thymelaeaceae) in Peninsular Malaysia.
- Chua L, Leong L, Hoo L, Zakaria N, Hong T, Ting L, Hong N, Siong K. (2016). Conservation Action Plan for the Threatened Agarwood Species *Aquilaria malaccensis* (Thymelaeaceae) in Peninsular Malaysia. Kepong (MY): Forest Research Institute.
- Chua, L. S. L. (2008). Agarwood (*Aquilaria malaccensis*) in Malaysia. NDF Werkshop Case Studies, Forest Research Institute Malaysia.
- Coissac, E., Hollingsworth, P.M., Lavergne, S. & Taberlet, P., (2016). From barcodes to genomes: extending the concept of DNA barcoding, 25(7), 1423-1428.
- Compton, J., & Ishihara, A. (2004). The use and trade of agarwood in Japan. A TRAFFIC report to the CITES Secretariat, 6, 1-21.
- Conesa, A., Götz, S., García-Gómez, J. M., Terol, J., Talón, M., & Robles, M. (2005). Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics*, 21(18), 3674-3676.

- Cullis, C. A., Vorster, B. J., Van Der Vyver, C., & Kunert, K. J. (2009). Transfer of genetic material between the chloroplast and nucleus: how is it related to stress in plants?. *Annals of Botany*, 103(4), 625-633.
- Dahham, S. S., Tabana, Y. M., Ahmed Hassan, L. E., Khadeer Ahamed, M. B., Abdul Majid, A. S., & Abdul Majid, A. M. S. (2016). In vitro antimetastatic activity of Agarwood (*Aquilaria crassna*) essential oils against pancreatic cancer cells. *Alexandria Journal of Medicine*, 52(2), 141-150.
- Deng, X., Jiang, Z., Jiang, Q., Guo, W., Li, Y., & Zhang, X. (2020). Characterization of the complete chloroplast genome of *Aquilaria sinensis*, an endangered agarwood-producing tree. *Mitochondrial DNA Part B*, 5(1), 422-423.
- Dieffenbach, C. W., Lowe, T. M., & Dveksler, G. S. (1993). General concepts for PCR primer design. *PCR Methods and Applications*, 3(3), S30-S37.
- Dierckxsens, N., Mardulyn, P., & Smits, G. (2017). NOVOPlasty: de novo assembly of organelle genomes from whole genome data. *Nucleic Acids Research*, 45(4), e18-e18.
- Domke, W. (1934). Untersuchungen über die systematische und geographische-Gliederung der Thymelaeaceen nebst einer Neubeschreibung ihrer Gattung. *Bibliotheca Botanische*, 27(1), 1-151.
- Don, G. (1831). A General History of the Dichlamydeous Plants, Comprising Complete Descriptions of the Different Orders. Volume 1, *Thalamiflorae*. Rivington.
- Dong, W. L., Wang, R. N., Zhang, N. Y., Fan, W. B., Fang, M. F., & Li, Z. H. (2018). Molecular evolution of chloroplast genomes of orchid species: insights into phylogenetic relationship and adaptive evolution. *International Journal of Molecular Sciences*, 19(3), 716.
- Dormontt, E. E., Boner, M., Braun, B., Breulmann, G., Degen, B., Espinoza, E., Gardner, S., Guillery, P., Hermanson, J.C., Koch, G. & Lowe, A. J. (2015). Forensic timber identification: It's time to integrate disciplines to combat illegal logging. *Biological Conservation*, 191, 790-798.
- Downie, S. R., & Jansen, R. K. (2015). A comparative analysis of whole plastid genomes from the Apiales: expansion and contraction of the inverted repeat, mitochondrial to plastid transfer of DNA, and identification of highly divergent noncoding regions. *Systematic Botany*, 40(1), 336-351.
- Downie, S. R., & Palmer, J. D. (1992). Use of chloroplast DNA rearrangements in reconstructing plant phylogeny. In *Molecular Systematics of Plants* (pp. 14-35). Springer, Boston, MA.
- Echt, C. S., DeVerno, L. L., Anzidei, M., & Vendramin, G. G. (1998). Chloroplast microsatellites reveal population genetic diversity in red pine, *Pinus resinosa* Ait. *Molecular Ecology*, Vol. 7: 307-316.

- Ellegren, H. (2004). Microsatellites: simple sequences with complex evolution. *Nature Reviews Genetics*, 5(6), 435-445.
- Eurlings, M. C. M., & Gravendeel, B. (2005). *trn L-trn F* sequence data imply paraphyly of Aquilaria and Gyrinops (Thymelaeaceae) and provide new perspectives for agarwood identification. *Plant Systematics and Evolution*, 254(1), 1-12.
- Eurlings, M. C., van Beek, H. H., & Gravendeel, B. (2010). Polymorphic microsatellites for forensic identification of agarwood (*Aquilaria crassna*). *Forensic Science International*, 197(1-3), 30-34.
- Fagundes, B. S., da Silva, L. F., Giacomin, R. M., Secco, D., Díaz-Cruz, J. A., & Da-Silva, P. R. (2016). Transferability of microsatellite markers among Myrtaceae species and their use to obtain population genetics data to help the conservation of the Brazilian Atlantic Forest. *Tropical Conservation Science*, 9(1), 408-422.
- Farah, A. H., Lee, S. Y., Gao, Z., Yao, T. L., Madon, M., & Mohamed, R. (2018). Genome size, molecular phylogeny, and evolutionary history of the tribe Aquilarieae (Thymelaeaceae), the natural source of agarwood. *Frontiers in Plant Science*, 9, 712.
- Faridah-Hanum, I., Mustapa, M. Z., Lepun, P., Tuan Marina, T. I., Nazre, M., Alan, R. I. B. K. A., & Mohamed, R. (2009). Notes on the distribution and ecology of *Aquilaria* Lam.(Thymelaeaceae) in Malaysia. *Malaysian Forester*, 72(2), 247-259.
- Feng, T., Li, Q., Wang, Y., Qiu, S., He, M., Zhang, W., Dong, J. & Zhu, S. (2019). Phylogenetic analysis of *Aquilaria* Lam.(Thymelaeaceae) based on DNA barcoding. *Holzforschung*, 73(6), 517-523.
- Finkemeier, I. and Leister, D., (2010). Plant chloroplasts and other plastids. eLS.
- Ganopoulos, I., Argiriou, A., and Tsafaris, A. (2011). Microsatellite high resolution melting (SSR-HRM) analysis for authenticity testing of protected designation of origin (PDO) sweet cherry products. *Food Control*, 22(3-4), 532-541.
- Gasson, P. (2011). How precise can wood identification be? Wood anatomy's role in support of the legal timber trade, especially CITES. *IAWA Journal*, 32(2), 137-154.
- Gilg, E. (1894). Thymelaeaceae. 216–245. A. Engler and K. Prantl. *Die Natürlichen Pflanzenfamilien*, 3. Gilg, E., 1894. Thymelaeaceae. 216–245. A. Engler and K. Prantl. *Die natürlichen Pflanzenfamilien*, 3.
- Gonçalves, J., Marks, C. A., Obendorf, D., Amorim, A., & Pereira, F. (2015). A multiplex PCR assay for identification of the red fox (*Vulpes vulpes*) using the mitochondrial ribosomal RNA genes. *Conservation Genetics Resources*, 7(1), 45-48.

- Gregory, T. R. (2005). DNA barcoding does not compete with taxonomy. *Nature*, 434(7037), 1067-1067. Gregory, T. R., 2005. DNA barcoding does not compete with taxonomy. *Nature*, 434(7037), 1067-1067.
- Greiner, S., Lehwerk, P., & Bock, R. (2019). OrganellarGenomeDRAW (OGDRAW) version 1.3. 1: expanded toolkit for the graphical visualization of organellar genomes. *Nucleic Acids Research*, 47(W1), W59-W64.
- Greppin, J. A. (1988). The various aloës in ancient times. *The Journal of Indo-European Studies*, 16(1-2), 33-48.
- Group, C. P. W., Hollingsworth, P. M., Forrest, L. L., Spouge, J. L., Hajibabaei, M., Ratnasingham, S., van der Bank, M., Chase, M.W., Cowan, R.S., Erickson, D.L. & Little, D. P. (2009). A DNA barcode for land plants. *Proceedings of the National Academy of Sciences*, 106(31), 12794-12797.
- Guindon, S., Dufayard, J. F., Lefort, V., Anisimova, M., Hordijk, W., & Gascuel, O. (2010). New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. *Systematic Biology*, 59(3), 307-321.
- Hamilton, F. (1836). Chapter 11. A commentary on the second book of the herbarium amboinense. *Memoirs of the Wernerian Natural History Society*, 6, 268-333.
- Hebert, P. D., Cywinska, A., Ball, S. L., & DeWaard, J. R., (2003). Biological identifications through DNA barcodes. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 270(1512), 313-321.
- Herber, B.E., (2002). Pollen morphology of the Thymelaeaceae in relation to its taxonomy. *Plant Systematics and Evolution*, 232(1), 107-121.
- Hishamuddin, M. S., Lee, S. Y., Isa, N. M., Lamasudin, D. U., Abidin, S. A. Z., & Mohamed, R. (2019). Time-based LC-MS/MS analysis provides insights into early responses to mechanical wounding, a major trigger to agarwood formation in *Aquilaria malaccensis* Lam. *RSC Advances*, 9(32), 18383-18393.
- Hishamuddin, M. S., Lee, S. Y., Ng, W. L., Ramlee, S. I., Lamasudin, D. U., & Mohamed, R. (2020). Comparison of eight complete chloroplast genomes of the endangered *Aquilaria* tree species (Thymelaeaceae) and their phylogenetic relationships. *Scientific Reports*, 10(1), 1-13.
- Hoang, D. T., Chernomor, O., Von Haeseler, A., Minh, B. Q., & Vinh, L. S. (2018). UFBoot2: improving the ultrafast bootstrap approximation. *Molecular Biology and Evolution*, 35(2), 518-522.
- Hollingsworth, P. M., Graham, S. W., & Little, D. P., (2011). Choosing and using a plant DNA barcode. *PloS One*, 6(5), e19254.

- Hou, D., (1964). Notes on some Asiatic species of *Aquilaria* (Thymelaceae). *Blumea*, 12(2). 285-8.
- Huang H, Shi C, Liu Y, Mao S-Y, Gao L-Z. (2014). Thirteen *Camellia* chloroplast genome sequences determined by high-throughput sequencing: genome structure and phylogenetic relationships. *BMC Evolutionary Biology*, 14:151.
- Huelsenbeck, J.P. and Ronquist, F., (2001). MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics*, 17(8), 754-755.
- Hughes, K. (2007). *The Incense Bible: Plant Scents that Transcend World Culture, Medicine, and Spirituality*. New York: Haworth Press.
- Hung, K. H., Lin, C. H., & Ju, L. P. (2017). Tracking the geographical origin of timber by DNA fingerprinting: a study of the endangered species *Cinnamomum kanehirae* in Taiwan. *Holzforschung*, 71(11), 853-862.
- Hurst, G. D., & Jiggins, F. M., (2005). Problems with mitochondrial DNA as a marker in population, phylogeographic and phylogenetic studies: the effects of inherited symbionts. *Proceedings of the Royal Society B: Biological Sciences*, 272(1572), 1525-1534.
- Ismail, N., Rahiman, M. H. F., Taib, M. N., Ibrahim, M., Zareen, S., & Tajuddin, S. N. (2015). A review on agarwood and its quality determination. In *2015 IEEE 6th Control and System Graduate Research Colloquium (ICSGRC)* (pp. 103-108). IEEE.
- Ito, M. & Honda, G., (2005). Taxonomical identification of agarwood-producing species. *Natural medicines*, 59(3), 104-112
- Ito, T., Kakino, M., Tazawa, S., Oyama, M., Maruyama, H., Araki, Y., Hara, H. & Iinuma, M. (2012). Identification of phenolic compounds in *Aquilaria crassna* leaves via liquid chromatography-electrospray ionization mass spectroscopy. *Food Science and Technology Research*, 18(2), 259-262.
- Jiang, S., Jiang, Y., Guan, Y. F., Tu, P. F., Wang, K. Y., & Chen, J. M. (2011). Effects of 95% ethanol extract of *Aquilaria sinensis* leaves on hyperglycemia in diabetic db/db mice. *Journal of Chinese Pharmaceutical Sciences*, 20(6), 609.
- Jiao, L., Lu, Y., He, T., Li, J. & Yin, Y., (2019). A strategy for developing high-resolution DNA barcodes for species discrimination of wood specimens using the complete chloroplast genome of three *Pterocarpus* species. *Planta*, 250(1), 95-104.
- Jiao, L., Lu, Y., He, T., Guo, J., & Yin, Y. (2020). DNA barcoding for wood identification: Global review of the last decade and future perspective. *IAWA Journal*, 41(4), 620-643.

- Jiao, L., Yin, Y., Cheng, Y., & Jiang, X. (2014). DNA barcoding for identification of the endangered species *Aquilaria sinensis*: comparison of data from heated or aged wood samples. *Holzforschung*, 68(4), 487-494.
- Kale, S.M., Pardeshi, V.C., Kadoo, N.Y., Ghorpade, P.B., Jana, M.M. & Gupta, V.S., (2012). Development of genomic simple sequence repeat markers for linseed using next-generation sequencing technology. *Molecular Breeding*, 30(1), 597-606.
- Kalia, R.K., Rai, M.K., Kalia, S., Singh, R. & Dhawan, A.K., (2011). Microsatellite markers: an overview of the recent progress in plants. *Euphytica*, 177(3), 309-334.
- Kalyaanamoorthy, S., Minh, B.Q., Wong, T.K., Von Haeseler, A. & Jermiin, L.S., (2017). ModelFinder: fast model selection for accurate phylogenetic estimates. *Nature methods*, 14(6), 587-589.
- Kato, S., Imai, A., Rie, N. & Mukai, Y., (2013). Population genetic structure in a threatened tree, *Pyrus calleryana* var. *dimorphophylla* revealed by chloroplast DNA and nuclear SSR locus polymorphisms. *Conservation Genetics*, 14(5), 983-996.
- Katoh, K. & Standley, D.M., (2013). MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Molecular Biology and Evolution*, 30(4), 772-780.
- Katoh, K., Rozewicki, J., & Yamada, K. D. (2019). MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Briefing in Bioinformatics*, 20(4), 1160-1166.
- Kearse, M., Moir, R., Wilson, A., Stones-Havas, S., Cheung, M., Sturrock, S., Buxton, S., Cooper, A., Markowitz, S., Duran, C. and Thierer, T., (2012). Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics*, 28(12), 1647-1649.
- Knight, C. (1840). *The Penny Magazine of the Society for the Diffusion of Useful Knowledge* (Vol. 9). Charles Knight. Society for the Diffusion of Useful Knowledge, 1838. The Penny Cyclopaedia of the Society for the Diffusion of Useful Knowledge, vol. 9. London: Charles Knight and Company
- Ku, C., Nelson-Sathi, S., Roettger, M., Sousa, F.L., Lockhart, P.J., Bryant, D., Hazkani-Covo, E., McInerney, J.O., Landan, G. & Martin, W.F., (2015). Endosymbiotic origin and differential loss of eukaryotic genes. *Nature*, 524(7566), 427-432.
- Kumar, S., Stecher, G., Li, M., Knyaz, C. & Tamura, K., (2018). MEGA X: molecular evolutionary genetics analysis across computing platforms. *Molecular Biology and Evolution*, 35(6), 1547.

- Kumphune, S., Prompunt, E., Phaebuaw, K., Sriudwong, P., Pankla, R., & Thongyoo, P. (2011). Anti-inflammatory effects of the ethyl acetate extract of *Aquilaria crassna* inhibits LPS-induced tumour necrosis factor-alpha production by attenuating P38 MAPK activation. *International Journal of Green Pharmacy*, 5(1), 43.
- Kurtz, S., Choudhuri, J.V., Ohlebusch, E., Schleiermacher, C., Stoye, J. & Giegerich, R., (2001). REPuter: the manifold applications of repeat analysis on a genomic scale. *Nucleic Acids Research*, 29(22), 4633-4642.
- Laiou, A., Mandolini, L. A., Piredda, R., Bellarosa, R., & Simeone, M. C., (2013). DNA barcoding as a complementary tool for conservation and valorisation of forest resources. *ZooKeys*, (365), 197.
- Lee, S. Y., & Mohamed, R. (2016a). The origin and domestication of *Aquilaria*, an important agarwood-producing genus. In *Agarwood* (pp. 1-20). Springer, Singapore.
- Lee, S. Y., & Mohamed, R. (2016b). Rediscovery of *Aquilaria rostrata* (Thymelaeaceae), a species thought to be extinct, and notes on *Aquilaria* conservation in Peninsular Malaysia. *Blumea-Biodiversity, Evolution and Biogeography of Plants*, 61(1), 13-19.
- Lee, S. Y., Ng, W. L., Mahat, M. N., Nazre, M., & Mohamed, R. (2016a). DNA barcoding of the endangered *Aquilaria* (Thymelaeaceae) and its application in species authentication of agarwood products traded in the market. *PloS One*, 11(4), e0154631.
- Lee, S. Y., Weber, J., & Mohamed, R., (2011). Genetic variation and molecular authentication of selected *Aquilaria* species from natural populations in Malaysia using RAPD and SCAR markers. *Asian Journal of Plant Sciences*, 10(3), 202.
- Lee, S.Y., Ng, W.L. & Mohamed, R., (2016b). Rapid species identification of highly degraded agarwood products from *Aquilaria* using real-time PCR. *Conservation Genetics Resources*, 8(4), 581-585.
- Lee, S.Y., Ng, W.L., Hishamuddin, M.S. & Mohamed, R., (2019). The complete chloroplast genome sequence of Chengal (*Neobalanocarpus heimii*, Dipterocarpaceae), a durable tropical hardwood. *Mitochondrial DNA Part B*, 4(1), 19-20.
- Lee, S.Y., Ng, W.L., Mohamed, R. & Terhem, R., (2018). The complete chloroplast genome of *Aquilaria malaccensis* Lam. (Thymelaeaceae), an important and threatened agarwood-producing tree species. *Mitochondrial DNA Part B*, 3(2), 1120-1121.
- Li R, Ma PF, Wen J, Yi TS. (2013). Complete Sequencing of Five Araliaceae Chloroplast Genomes and the Phylogenetic Implications. *PLoS One* 8, 1–15.

- Li, G.D., Rao, P.Y., Guo, J.L. & Zhang, Y.H., (2019). The complete chloroplast genome of a critically endangered agarwood tree, *Aquilaria crassna* (Thymelaeaceae). *Mitochondrial DNA Part B*, 4(1), 1810-1811.
- Li, H., Xiao, W., Tong, T., Li, Y., Zhang, M., Lin, X., Zou, X., Wu, Q. & Guo, X., (2021). The specific DNA barcodes based on chloroplast genes for species identification of Orchidaceae plants. *Scientific Reports*, 11(1), 1-15.
- Li, Q., Yan, H., Lin, D., Wang, Y., He, M., Zhang, W., Gao, X. & Zhu, S., (2018). Molecular identification of three *Aquilaria* (Thymelaeaceae) species through DNA barcoding. *Biological and Pharmaceutical Bulletin*, 41(6), 967-971.
- Li, X., Yang, Y., Henry, R. J., Rossetto, M., Wang, Y., & Chen, S., (2015). Plant DNA barcoding: from gene to genome. *Biological Reviews*, 90(1), 157-166.
- Librado, P. & Rozas, J., (2009). DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. *Bioinformatics*, 25(11), 1451-1452.
- Lin, C.P., Hsiao, Y.T., Hsiao, Y.J.A., Chou, S.J., Chen, A.P., Kuo, C.I. & Chen, L.F.O., (2019). The complete chloroplast genome of agarwood producing species, *Aquilaria sinensis* (Lour.) Gilg: a species on IUCN red list. *Mitochondrial DNA Part B*, 4(2), 2992-2993.
- Linacre, A., and& Tobe, S. S., (2011). An overview to the investigative approach to species testing in wildlife forensic science. *Investigative Genetics*, 2(1), 1-9.
- Liu, G., Xie, Y., Zhang, D. & Chen, H., (2018). Analysis of SSR loci and development of SSR primers in *Eucalyptus*. *Journal of Forestry Research*, 29(2), 273-282.
- Liu, J. I. E., Moeller, M., Gao, L. M., Zhang, D. Q., & Li, D. Z., (2011). DNA barcoding for the discrimination of Eurasian yews (*Taxus* L., Taxaceae) and the discovery of cryptic species. *Molecular Ecology Resources*, 11(1), 89-100.
- Liu, S.R., Li, W.Y., Long, D., Hu, C.G. & Zhang, J.Z., (2013). Development and characterization of genomic and expressed SSRs in citrus by genome-wide analysis. *PloS One*, 8(10), p.e75149.
- Liu, Y., Chen, H., Yang, Y., Zhang, Z., Wei, J., Meng, H., Chen, W., Feng, J., Gan, B., Chen, X. & Gao, Z., (2013). Whole-tree agarwood-inducing technique: an efficient novel technique for producing high-quality agarwood in cultivated *Aquilaria sinensis* trees. *Molecules*, 18(3), 3086-3106.
- Liu, Y.Y., Wei, J.H., Gao, Z.H., Zhang, Z. & Lyu, J.C., (2017). A review of quality assessment and grading for agarwood. *Chinese Herbal Medicines*, 9(1), 22-30.

- Long, G. (1839). *The penny cyclopaedia of the Society for the Diffusion of Useful Knowledge* (Vol. 15). C. Knight.
- López-Sampson, A. & Page, T., (2018). Elliptical Fourier descriptors of leaf outlines: a tool to discriminate among *Aquilaria* species (Thymelaeaceae). *Silvae Genetica*, 67(1), 89-92.
- Lynch, M., Koskella, B., & Schaack, S., (2006). Mutation pressure and the evolution of organelle genomic architecture. *Science*, 311(5768), 1727-1730.
- Mabberley, D.J., (2008). Mabberley's Plant Book: A Portable Dictionary of Plants, their Classification and Uses 3rd Edn., Cambridge University Press, Cambridge, ISBN-13: 9780521820714,59.
- Martin, W., Stoebe, B., Goremykin, V., Hansmann, S., Hasegawa, M. & Kowallik, K.V., (1998). Gene transfer to the nucleus and the evolution of chloroplasts. *Nature*, 393(6681), 162-165.
- Mathews, R. H. (1931). *A Chinese-English dictionary: compiled for the China Inland Mission: Enghish index*. China inland mission and presbyterian mission Press.
- McCauley, D.E., (1995). The use of chloroplast DNA polymorphism in studies of gene flow in plants. *Trends in Ecology and Evolution*, 10(5), 198-202.
- McCormack, J. E., Hird, S. M., Zellmer, A. J., Carstens, B. C., & Brumfield, R. T., (2013). Applications of next-generation sequencing to phylogeography and phylogenetics. *Molecular Phylogenetics and Evolution*, 66(2), 526-538.
- Mehmood, F., Shahzadi, I., Waseem, S., Mirza, B., Ahmed, I., & Waheed, M. T., (2020). Chloroplast genome of *Hibiscus rosa-sinensis* (Malvaceae): comparative analyses and identification of mutational hotspots. *Genomics*, 112(1), 581-591.
- Meier, R., Shiyang, K., Vaidya, G. & Ng, P.K., (2006). DNA barcoding and taxonomy in Diptera: a tale of high intraspecific variability and low identification success. *Systematic Biology*, 55(5), 715-728.
- Meng, J., Li, X., Li, H., Yang, J., Wang, H. & He, J., (2018). Comparative analysis of the complete chloroplast genomes of four *Aconitum* medicinal species. *Molecules*, 23(5), 1015.
- Miller, J.I. (1969). The spice trade of the Roman Empire. Oxford, UK: Oxford University Press.
- Min, L., Gao, X., Zhang, J. Y., Deng, H. N., & Xu, B. (2021). Comparative chloroplast genomics of *Sophora* species: Evolution and phylogenetic relationships in the early-diverging legume subfamily Papilioideae (Fabaceae). *Frontiers in Plant Science*, 2785.

- Minh, B.Q., Nguyen, M.A.T. & von Haeseler, A., (2013). Ultrafast approximation for phylogenetic bootstrap. *Molecular Biology and Evolution*, 30(5), 1188-1195.
- Moe, K.T., Chung, J.W., Cho, Y.I., Moon, J.K., Ku, J.H., Jung, J.K., Lee, J. & Park, Y.J., (2011). Sequence information on simple sequence repeats and single nucleotide polymorphisms through transcriptome analysis of mungbean. *Journal of Integrative Plant Biology*, 53(1), 63-73.
- Mohamed, R. & Lee, S.Y., (2016). Keeping up appearances: Agarwood grades and quality. In *Agarwood* (pp. 149-167). Springer, Singapore.
- Mohamed, R. ed., 2016. Agarwood: Science behind the fragrance. Springer, Singapore.
- Mohamed, R., Jong, P.L. & Zali, M.S., (2010). Fungal diversity in wounded stems of *Aquilaria malaccensis*. *Fungal Diversity*, 43(1), 67-74.
- Mohamed, R., Tan, H. Y., & Siah, C. H. (2012). A real-time PCR method for the detection of *trnL-trnF* sequence in agarwood and products from *Aquilaria* (Thymelaeaceae). *Conservation Genetic Resources*, 4(3), 803-806.
- Moore, M. J., Soltis, P. S., Bell, C. D., Burleigh, J. G., & Soltis, D. E. (2010). Phylogenetic analysis of 83 plastid genes further resolves the early diversification of eudicots. *Proceedings of the National Academy of Sciences*, 107(10), 4623-4628.
- Moreira, F., Carneiro, J., & Pereira, F., (2017). A proposal for standardization of transgenic reference sequences used in food forensics. *Forensic Science International: Genetics*, 29, e26-e28.
- Moreira, P. A., & Oliveira, D. A. (2011). Leaf age affects the quality of DNA extracted from *Dimorphandra mollis* (Fabaceae), a tropical tree species from the Cerrado region of Brazil. *Genetics and Molecular Research*, 10(1), 353-358.
- Morgante, M. & Olivieri, A.M., (1993). PCR-amplified microsatellites as markers in plant genetics. *The Plant Journal*, 3(1), 175-182.
- Mullet JE. (1988). Chloroplast Development and Gene Expression. 1967:475–502.
- Neuhaus, H.E. & Emes, M.J., 2000. Nonphotosynthetic metabolism in plastids. *Annual review of plant biology*, 51(1), 111-140.
- Ng, L.T., (1997). A review on agar (gaharu) producing *Aquilaria* species. *Journal of Tropical Forest Product*, 2, 272-285.
- Nguyen, L.T., Schmidt, H.A., Von Haeseler, A. & Minh, B.Q., (2015). IQ-TREE: a fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Molecular Biology and Evolution*, 32(1), 268-274.

- Niu, X.L., Ji, K.P. & Lu, G.Q., (2010). Preliminary studies on identification of *Aquilaria sinensis* (L.) Gilg by the PCR product of rDNA ITS sequencing. *Guangdong Agricultural Science*, 37, 167-169.
- Ogden, R., & Linacre, A., (2015). Wildlife forensic science: a review of genetic geographic origin assignment. *Forensic Science International: Genetics*, 18, 152-159.
- Ozyigit, I.I., Dogan, I. & Filiz, E., (2015). In silico analysis of simple sequence repeats (SSRs) in chloroplast genomes of 'Glycine' species. *Plant Omics*, 8(1), 24-29.
- Paiva, J.A., Prat, E., Vautrin, S., Santos, M.D., San-Clemente, H., Brommonschenkel, S., Fonseca, P.G., Grattapaglia, D., Song, X., Ammiraju, J.S. & Kudrna, D., (2011). Advancing *Eucalyptus* genomics: identification and sequencing of lignin biosynthesis genes from deep-coverage BAC libraries. *BMC Genomics*, 12(1), 1-13.
- Patel, R.K. & Jain, M., (2012). NGS QC Toolkit: a toolkit for quality control of next generation sequencing data. *PloS One*, 7(2), p.e30619.
- Perdereau, A.C., Kelleher, C.T., Douglas, G.C. & Hodgkinson, T.R., (2014). High levels of gene flow and genetic diversity in Irish populations of *Salix caprea* L. inferred from chloroplast and nuclear SSR markers. *BMC Plant Biology*, 14(1), 1-12.
- Pereira, F., Carneiro, J., & Amorim, A., (2008). Identification of species with DNA-based technology: current progress and challenges. *Recent Patents on DNA & Gene Sequences (Discontinued)*, 2(3), 187-200.
- Pern, Y. C., Lee, S. Y., Ng, W. L., & Mohamed, R., (2020). Cross-amplification of microsatellite markers across agarwood-producing species of the Aquilarieae tribe (Thymelaeaceae). *3 Biotech*, 10(3), 1-9.
- Phumichai, C., Phumichai, T. & Wongkaew, A., (2015). Novel chloroplast microsatellite (cpSSR) markers for genetic diversity assessment of cultivated and wild Hevea rubber. *Plant Molecular Biology Reporter*, 33(5), 1486-1498.
- Powell, W., Morgante, M., Andre, C., Hanafey, M., Vogel, J., Tingey, S. & Rafalski, A., (1996). The comparison of RFLP, RAPD, AFLP and SSR (microsatellite) markers for germplasm analysis. *Molecular Breeding*, 2(3), 225-238.
- POWO. (2019). Plants of the world online. Facilitated by the Royal Botanic Gardens, Kew.
- Pranakhon, R., Aromdee, C., & Pannangpatch, P. (2015). Effects of iriflophenone 3-C- β -glucoside on fasting blood glucose level and glucose uptake. *Pharmacognosy Magazine*, 11(41), 82.

- Provan, J., Powell, W. & Hollingsworth, P.M., (2001). Chloroplast microsatellites: new tools for studies in plant ecology and evolution. *Trends in Ecology & Evolution*, 16(3), 142-147.
- Qian, J., Song, J., Gao, H., Zhu, Y., Xu, J., Pang, X., Yao, H., Sun, C., Li, X.E., Li, C. & Liu, J., (2013). The complete chloroplast genome sequence of the medicinal plant *Salvia miltiorrhiza*. *PLoS One*, 8(2), p.e57607.
- Qin, M., Zhu, C. J., Yang, J. B., Vatanparast, M., Schley, R., Lai, Q., Zhang, D.Y., Tu, T.Y., Klitgård, B.B., Li, S.J. & Zhang, D. X. (2020). Comparative analysis of complete plastid genome reveals powerful barcode regions for identifying wood of *Dalbergia odorifera* and *D. tonkinensis* (Leguminosae). *Journal of Systematics and Evolution*.
- Rachmayanti, Y., Leinemann, L., Gailing, O. & Finkeldey, R., (2009). DNA from processed and unprocessed wood: factors influencing the isolation success. *Forensic Science International: Genetics*, 3(3), 185-192.
- Rajendrakumar, P., Biswal, A.K., Balachandran, S.M., Srinivasarao, K. and& Sundaram, R.M., (2007). Simple sequence repeats in organellar genomes of rice: frequency and distribution in genic and intergenic regions. *Bioinformatics*, 23(1), 1-4.
- Rambaut, A. (2019) FigTree v1.4.4. Available online: <http://tree.bio.ed.ac.uk/software/figtree/> (accessed on 31 October 2019).
- Rautenbach M. & Gnidia L., (2008). (Thymelaeaceae) is not monophyletic: taxonomic implications for Gnidia and its relatives in Thymelaeoideae, Doctoral dissertation, University of Johannesburg.
- Ridley, H.N., (1901). Garu and chanietnamdan. Journal of the Straits Branch of the Royal Asiatic Society, (35), 73-82.
- Rogalski, M., Ruf, S. & Bock, R., (2006). Tobacco plastid ribosomal protein S18 is essential for cell survival. *Nucleic Acids Research*, 34(16), 4537-4545.
- Royle, J.F., (1839). Illustrations of the botany and other branches of the natural history of the Himalayan Mountains and of the flora of Cashmere (Vol. 2). Allen.
- Ruan, X. S., Lin, X. C., Lou, Y. F., Guo, X. Q., Fang, W., & Chen, C. J. (2008). Genetic diversity of *Phyllostachys heterocycla* var. *pubescens* provenances by AFLP and ISSR. *Journal of Zhejiang Forestry Science and Technology*, 28, 29-33.
- S.S. Dahham, S. S., Ahamed, M. B. K., Saghir, S. M., Alsuede, F. S., Iqbal, M. A., & Majid, A. M. S. A. (2014). Bioactive essential oils from *Aquilaria crassna* for cancer prevention and treatment. *Global Journal on Advances Pure and Applied Sciences*, 4.

- Santos, C., & Pereira, F., (2017). Design and evaluation of PCR primers for amplification of four chloroplast DNA regions in plants. *Conservation Genetics Resources*, 9(1), 9-12.
- Särkinen, T., & George, M., (2013). Predicting plastid marker variation: can complete plastid genomes from closely related species help? *PLoS One*, 8(11), e82266.
- Seberg, O., & Petersen, G., (2009). How many loci does it take to DNA barcode a crocus? *PloS One*, 4(2), e4598.
- Shanker, A., (2013). *Identification of microsatellites in chloroplast genome of Anthoceros formosae*. Univ.-Bibliothek.
- Shanker, A., (2014). *Computationally mined microsatellites in chloroplast genome of Pellia endiviifolia*. Univ.-Bibliothek.
- Sharma, P., Jha, A. B., Dubey, R. S., & Pessarakli, M. (2012). Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. *Journal of botany*, 2012.
- Shen, Y., Yan, P., Zhao, X., Pang, Q., & Zhao, S., (2008). Application of ISSR marker and ITS sequence to investigation of genetic variation of *Aquilaria sinensis*. *Journal of South China University of Technology (Natural Science Edition)* 36: 128-132
- Shinozaki, K., Ohme, M., Tanaka, M., Wakasugi, T., Hayashida, N., Matsubayashi, T., Zaita, N., Chunwongse, J., Obokata, J., Yamaguchi-Shinozaki, K. & Ohto, C., (1986). The complete nucleotide sequence of the tobacco chloroplast genome: its gene organization and expression. *The EMBO Journal*, 5(9), 2043-2049.
- Shukla, N., Kuntal, H., Shanker, A. & Sharma, S.N., (2018). Mining and analysis of simple sequence repeats in the chloroplast genomes of genus Vigna. *Biotechnology Research and Innovation*, 2(1), 9-18.
- Singh P, Sharma H, Nag A, Bhau BS, & Sharma RK., (2015). Development and characterization of polymorphic microsatellites markers in endangered *Aquilaria malaccensis*. *Conservation Genetics Resources*, 7(1), 61-63.
- Singh, H. K., Parveen, I., Raghuvanshi, S., & Babbar, S. B., (2012). The loci recommended as universal barcodes for plants on the basis of floristic studies may not work with congeneric species as exemplified by DNA barcoding of *Dendrobium* species. *BMC Research Notes*, 5(1), 1-11.
- Singh, V.K., Singh, A.K., Singh, S. & Singh, B.D., (2015). Next-generation sequencing (NGS) tools and impact in plant breeding. In Advances in Plant Breeding Strategies: *Breeding, Biotechnology and Molecular Tools* (pp. 563-612). Springer, Cham.

- Snelder, D. J., & Lasco, R. D. (Eds.). (2008). *Smallholder tree growing for rural development and environmental services: Lessons from Asia* (Vol. 5). Springer Science & Business Media
- Soehartono, T. & Newton, A.C., (2001). Reproductive ecology of *Aquilaria* spp. in Indonesia. *Forest Ecology and Management*, 152(1-3), 59-71.
- Sonah, H., Deshmukh, R. K., Sharma, A., Singh, V. P., Gupta, D. K., Gacche, R. N., Rana, J.C., Singh, N.K. & Sharma, T. R. (2011). Genome-wide distribution and organization of microsatellites in plants: an insight into marker development in Brachypodium. *Plos One*, 6(6), e21298.
- Song, Y., Chen, Y., Lv, J., Xu, J., Zhu, S., Li, M., & Chen, N., (2017). Development of chloroplast genomic resources for Oryza species discrimination. *Frontiers in Plant Science*, 8, 1854.
- Stegemann, S., Keuthe, M., Greiner, S. & Bock, R., (2012). Horizontal transfer of chloroplast genomes between plant species. *Proceedings of the National Academy of Sciences*, 109(7), 2434-2438.
- Stothard, P., (2000). The sequence manipulation suite: JavaScript programs for analyzing and formatting protein and DNA sequences. *Biotechniques*, 28(6), 1102-1104.
- Subasinghe, S.M.C.U.P., (2013). Proceedings of International Forestry and Environment Symposium (Vol. 18): Agarwood Production in *Gyrinops walla* (Walla patta) Myths and Reality. In *Proceedings of International Forestry and Environment Symposium* (Vol. 18).
- Sugiura, M. (1992). The chloroplast genome. *10 Years Plant Molecular Biology*, 149-168.Sugiura, M., 1992. The chloroplast genome. *10 Years plant molecular biology*, 149-168.
- Sun, Y. X., Moore, M. J., Meng, A. P., Soltis, P. S., Soltis, D. E., Li, J. Q., & Wang, H. C. (2013). Complete plastid genome sequencing of Trochodendraceae reveals a significant expansion of the inverted repeat and suggests a Paleogene divergence between the two extant species. *PLoS One*, 8(4), e60429.
- Takemoto, H., Ito, M., Shiraki, T., Yagura, T., & Honda, G. (2008). Sedative effects of vapor inhalation of agarwood oil and spikenard extract and identification of their active components. *Journal of Natural Medicines*, 62(1), 41-46.
- Tambarussi, E. V., Melotto-Passarin, D. M., Gonzalez, S. G., Brigati, J. B., de Jesus, F. A., Barbosa, A. L., Dressano, K. & Carrer, H. (2009). In silico analysis of Simple Sequence Repeats from chloroplast genomes of Solanaceae species. *Crop Breeding & Applied Biotechnology*, 9(4)

- Tanaka, S. & Ito, M., (2020). DNA barcoding for identification of agarwood source species using *trnL-trnF* and mat K DNA sequences. *Journal of Natural Medicines*, 74(1), 42-50.
- Tangmitcharoen, S., Yongrattana, P., Luangvirivasaeng, V., Tasen, W., Chantbep, P. & Saengtubtim, S., (2008). Floral biology of *Aquilaria crassna* Pierre ex Lecomte. *Thailand Journal of Forestry*, 27(2), 1-13.
- Tautz, D., (1989). Hypervariability of simple sequences as a general source for polymorphic DNA markers. *Nucleic Acids Research*, 17(16), 6463-6471.
- Tawan, C. S. (2004). Thymelaeaceae. In: Soepadmo, Saw, L.G. Saw and Chung, R.C.K. (Eds.). Tree Flora of Sabah and Sarawak. Vol. 5. Sabah Forestry Department, FRIM and Sarawak Forestry Department. Kuala Lumpur Malaysia. 133-484.
- Techen, N., Parveen, I., Pan, Z., & Khan, I. A., (2014). DNA barcoding of medicinal plant material for identification. *Current Opinion in Biotechnology*, 25, 103-110.
- The Angiosperm Phylogeny Group, (1998). An ordinal classification for the families of flowering plants. *Annals of the Missouri Botanical Garden*, 531-553.
- The IUCN Red List of Threatened Species. Version 2017-2. www.iucnredlist.org. (Accessed November 06, 2019).
- The Plant List. Version 1.1. (2013). Available: <http://www.theplantlist.org/>. Accessed 28 March 2021
- Thitikornpong, W., Palanuvej, C. & Ruangrungsi, N., 2018. DNA barcoding for authentication of the endangered plants in genus *Aquilaria*. *Thai Journal of Pharmaceutical Sciences (TJPS)*, 42(4).
- Thuy, D. T. T., Tuyen, T. T., Thuy, T. T. T., Minh, P. T. H., Tran, Q. T., Long, P. Q., Nguyen, D.C., Bach, L.G. & Chien, N. Q. (2019). Isolation process and compound identification of agarwood essential oils from *Aquilaria crassna* cultivated at three different locations in vietnam. *Processes*, 7(7), 432.
- Tillich, M., Lehwerk, P., Pellizzer, T., Ulbricht-Jones, E. S., Fischer, A., Bock, R., & Greiner, S. (2017). GeSeq—versatile and accurate annotation of organelle genomes. *Nucleic Acids Research*, 45(W1), W6-W11.
- Tnah, L. H., Lee, C. T., Lee, S. L., Ng, K. K. S., Ng, C. H., Nurul-Farhanah, Z., Lau, K.H. & Chua, L. S. L. (2012). Isolation and characterization of microsatellite markers for an important tropical tree, *Aquilaria malaccensis* (Thymelaeaceae). *American Journal of Botany*, 99(11), e431-e433.
- Tropicos. Missouri Botanical Garden. (2016). [<http://www.tropicos.org>]. Access date 27 October 2019

- Turker, A. U., & Camper, N. D. (2002). Biological activity of common mullein, a medicinal plant. *Journal of Ethnopharmacology*, 82(2-3), 117-125.
- Ueno, S., Moriguchi, Y., Uchiyama, K., Ujino-Ihara, T., Futamura, N., Sakurai, T., Shinohara, K. & Tsumura, Y., (2012). A second-generation framework for the analysis of microsatellites in expressed sequence tags and the development of EST-SSR markers for a conifer, *Cryptomeria japonica*. *BMC Genomics*, 13(1), 1-16.
- UNEP-WCMC. (2019). Te checklist of CITES species website. Compiled by UNEP-WCMC, Cambridge, UK. CITES Secretariat, Geneva. <http://checklist.cites.org>. Accessed 2019 June 28.
- Van De Wiel, C. C. M., Van Der Schoot, J., Van Valkenburg, J. L. C. H., Duistermaat, H., & Smulders, M. J. M., (2009). DNA barcoding discriminates the noxious invasive plant species, floating pennywort (*Hydrocotyle ranunculoides* Lf), from non-invasive relatives. *Molecular Ecology Resources*, 9(4), 1086-1091.
- Van der Bank, M., Fay, M. F., & Chase, M. W. (2002). Molecular phylogenetics of Thymelaeaceae with particular reference to African and Australian genera. *Taxon*, 51(2), 329-339.
- Varshney, R.K., Nayak, S.N., May, G.D. & Jackson, S.A., (2009). Next-generation sequencing technologies and their implications for crop genetics and breeding. *Trends in Biotechnology*, 27(9), 522-530.
- Varshney, R.K., Thiel, T., Stein, N., Langridge, P. & Graner, A., (2002). In silico analysis on frequency and distribution of microsatellites in ESTs of some cereal species. *Cellular and Molecular Biology Letters*, 7(2A), 537-546.
- Vaughn, J. N., Chaluvadi, S. R., Rangan, L., & Bennetzen, J. L. (2014). Whole plastome sequences from five ginger species facilitate marker development and define limits to barcode methodology. *PLoS One*, 9(10), e108581.
- Vieira, M.L.C., Santini, L., Diniz, A.L. & Munhoz, C.D.F., 2016. Microsatellite markers: what they mean and why they are so useful. *Genetics and Molecular Biology*, 39, 312-328.
- Wang, Y., Zhan, D. F., Jia, X., Mei, W. L., Dai, H. F., Chen, X. T., & Peng, S. Q. (2016). Complete chloroplast genome sequence of *Aquilaria sinensis* (Lour.) Gilg and evolution analysis within the Malvales order. *Frontiers in Plant Science*, 7, 280.
- Wang, Z.F., Cao, H.L., Cai, C.X., Guo, Y. & Wang, Z.M., (2018). Microsatellites development and cross-amplification for *Aquilaria sinensis*, an endangered agarwood-producing tree. *Journal of Genetics*, 97(1), 139-145.

- Watt G., (2014). A dictionary of the economic products of India, vol. 1. Cambridge: Cambridge University Press.
- Wen-jie, J.I.A., (2008). Extraction of Genomic DNA and Optimization of ISSR Reaction Condition for *Aquilaria sinensis* (Lour.) Gilg [J]. *Journal of Anhui Agricultural Sciences*, 24(1).
- Whitmore, T. C. (1972). Thymelaeaceae. Tree Flora of Malaya, vol. 2, 383–391.
- Wicke, S., Schneeweiss, G. M., Depamphilis, C. W., Müller, K. F., & Quandt, D. (2011). The evolution of the plastid chromosome in land plants: gene content, gene order, gene function. *Plant Molecular Biology*, 76(3), 273-297.
- Williams, A. V., Miller, J. T., Small, I., Nevill, P. G., & Boykin, L. M. (2016). Integration of complete chloroplast genome sequences with small amplicon datasets improves phylogenetic resolution in *Acacia*. *Molecular Phylogenetics and Evolution*, 96, 1-8.
- Wolfe, K. H., Li, W. H., & Sharp, P. M., (1987). Rates of nucleotide substitution vary greatly among plant mitochondrial, chloroplast, and nuclear DNAs. *Proceedings of the National Academy of Sciences*, 84(24), 9054-9058.
- Woodson, J.D. & Chory, J., (2008). Coordination of gene expression between organellar and nuclear genomes. *Nature Reviews Genetics*, 9(5), 383-395.
- Wu, F. H., Chan, M. T., Liao, D. C., Hsu, C. T., Lee, Y. W., Daniell, H., Duvall, M.R. & Lin, C. S. (2010). Complete chloroplast genome of *Oncidium Gower Ramsey* and evaluation of molecular markers for identification and breeding in Oncidiinae. *BMC Plant Biology*, 10(1), 1-12.
- Wyn, L.T. & Anak, N.A., (2010). Wood for the trees: a review of the agarwood (gaharu) trade in. *Kuala Lumpur: TRAFFIC Southeast Asia*.
- Wynen, L.P., Goldsworthy, S.D., Insley, S.J., Adams, M., Bickham, J.W., Francis, J., Gallo, J.P., Hoelzel, A.R., Majluf, P., White, R.W & Slade, R., (2001). Phylogenetic relationships within the eared seals (Otariidae: Carnivora): implications for the historical biogeography of the family. *Molecular Phylogenetics and Evolution*, 21(2), 270-284.
- Yan, F., Wang, Q.L., Zhang, Y.J., Zhang, C.M. & Chen, Y., (2019). The complete chloroplast genome sequence of medicinal plant, *Daphne tangutica* Maxim. (Thymelaeaceae). *Mitochondrial DNA Part B*, 4(1), 1776-1777.
- Yang, J. B., Li, D. Z., & Li, H. T. (2014). Highly effective sequencing whole chloroplast genomes of angiosperms by nine novel universal primer pairs. *Molecular Ecology Resources*, 14(5), 1024-1031.
- Yang, J. B., Tang, M., Li, H. T., Zhang, Z. R., & Li, D. Z., (2013). Complete chloroplast genome of the genus *Cymbidium*: lights into the species

- identification, phylogenetic implications and population genetic analyses. *BMC Evolutionary Biology*, 13(1), 1-12.
- Yang, J., Vázquez, L., Chen, X., Li, H., Zhang, H., Liu, Z., & Zhao, G., (2017). Development of chloroplast and nuclear DNA markers for Chinese oaks (*Quercus* subgenus *Quercus*) and assessment of their utility as DNA barcodes. *Frontiers in Plant Science*, 8, 816.
- Yin, Y., Jiao, L., Dong, M., Jiang, X., & Zhang, S. (2016). Wood resources, identification, and utilization of agarwood in China. In *agarwood* (pp. 21-38). Springer, Singapore.
- Yu, X. Q., Drew, B. T., Yang, J. B., Gao, L. M., & Li, D. Z. (2017). Comparative chloroplast genomes of eleven *Schima* (Theaceae) species: Insights into DNA barcoding and phylogeny. *PLoS One*, 12(6), e0178026.
- Yuan, C., Zhong, W., Mou, F., Gong, Y., Pu, D., Ji, P., Huang, H., Yang, Z. & Zhang, C. (2017). The complete chloroplast genome sequence and phylogenetic analysis of Chuanminshen (*Chuanminshenviolaceum* Sheh et Shan). *Physiology and Molecular Biology of Plants*, 23(1), 35-41.
- Yuan, C.Y., Wang, P., Chen, P.P., Xiao, W.J., Zhang, C., Hu, S., Zhou, P., Chang, H.P., He, Z., Hu, R. & Lu, X.T., (2015). Genetic diversity revealed by morphological traits and ISSR markers in 48 Okras (*Abelmoschus esculentus* L.). *Physiology and Molecular Biology of Plants*, 21(3), 359-364.
- Yun, N., Park, J. & Oh, S.H., (2019). The complete chloroplast genome of the traditional medicinal plant *Stellera chamaejasme* L. (Thymelaeaceae). *Mitochondrial DNA Part B*, 4(1), 1796-1797.
- Zaya, D. N., & Ashley, M. V. (2012). Plant genetics for forensic applications. *Plant DNA Fingerprinting and Barcoding*, 35-52.
- Zhang, D.Q., Tian, H., Xie, Y.J., Tan, X.F., Huang, Q.Y., Gu, Z.J., Cao, J.G. & ZENG, Y.L., (2009). Genetic Diversity of *Eucalyptus tereticornis* by ISSR [J]. *Journal of Central South University of Forestry & Technology*, 5.
- Zhang, Y. J., Ma, P. F., & Li, D. Z. (2011). High-throughput sequencing of six bamboo chloroplast genomes: phylogenetic implications for temperate woody bamboos (Poaceae: Bambusoideae). *PloS One*, 6(5), e20596.
- Zhang, Y. T., Wang, Z. F., Cao, H. L., Li, X. Y., Wu, L. F., Zhuo, S. B., & Huang, X. F. (2010). Isolation and characterization of polymorphic microsatellite loci in *Aquilaria sinensis* (Lour.) Gilg. *Conservation Genetics Resources*, 2(1), 5-6.
- Zhang, Y., Du, L., Liu, A., Chen, J., Wu, L., Hu, W., Zhang, W., Kim, K., Lee, S.C., Yang, T.J. & Wang, Y. (2016). The complete chloroplast genome sequences of five *Epimedium* species: lights into phylogenetic and taxonomic analyses. *Frontiers in Plant Science*, 7, 306.

- Zhang, Y.H., Huang, Y., Li, Z.M. & Zhang, S.D., (2019). Characterization of the complete chloroplast genome of the vulnerable agarwood tree, *Aquilaria yunnanensis* (Thymelaeaceae). *Conservation Genetics Resources*, 11(2), 161-164.
- Zong, D., Gan, P., Zhou, A., Li, J., Xie, Z., Duan, A., & He, C. (2019). Comparative analysis of the complete chloroplast genomes of seven *Populus* species: Insights into alternative female parents of *Populus tomentosa*. *PLoS One*, 14(6), e0218455.
- Zou, C., Lu, C., Zhang, Y., & Song, G. (2012). Distribution and characterization of simple sequence repeats in *Gossypium raimondii* genome. *Bioinformation*, 8(17), 801.
- Zou, M., Xia, Z., Lu, C., Wang, H., Ji, J., & Wang, W. (2012). Genetic diversity and differentiation of *Aquilaria sinensis* (Lour.) Gilg revealed by ISSR and SRAP markers. *Crop Science*, 52(5), 2304-2313.