

UNIVERSITI PUTRA MALAYSIA

ELUCIDATING FELINE HEPATIC LIPIDOSIS THROUGH SERUM BIOCHEMISTRY, HISTOPATHOLOGY AND GENE EXPRESSION IN THE KLANG VALLEY STRAY CATS

MUHAMMAD FADZLY BIN SALLEH

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MUHAMMAD FADZLY BIN SALLEH

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

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DEDICATION

This thesis is dedicated to my father, mother, brothers and sister; to my supervisor, supervisory committee, fellow postgraduates, and friends.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

ELUCIDATING FELINE HEPATIC LIPIDOSIS THROUGH SERUM BIOCHEMISTRY, HISTOPATHOLOGY AND GENE EXPRESSION IN THE KLANG VALLEY STRAY CATS

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November 2020

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Feline hepatic lipidosis (FHL) is a common metabolic dysfunction related to lipid metabolism in cats. In conditions of hepatic lipidosis, steatosis or excessive lipid deposition occurs in the liver due to the cat's inability to efficiently breakdown and utilise the excess lipids. As such, lipid vacuolation in the hepatocytes and associated clinical signs, may arise if the condition worsens. The main cause of hepatic lipidosis remains unclear. This study aims to determine the lipid composition of liver samples in Klang Valley stray cats and to relate their serum and liver biochemistry with their histopathological changes, alongside identifying expression of hepatic genes associated with energy regulation through lipid metabolism associated with FHL in stray cats.

In total, 18 stray cats that were emaciated, dehydrated and lethargic were collected from pounds located in the Klang Valley area. Alanine aminotransferase (ALT), a common biomarker to determine liver injury in small animals, was measured in cat blood serum, as well as creatinine and urea in determining kidney function. Triglycerides (TAG) and cholesterol concentrations were measured in serum through serum analysis and in liver samples through Bligh & Dyer lipid extraction to relate to their steatosis levels. Histological analysis of liver samples were carried out to determine the severity of fatty change of the liver (FCL).

Through this study, cats with increasing severity of FCL in liver histology was observed to have increased levels of serum ALT, creatinine and urea, signifying liver injury, renal challenges and dehydration. However, TAG levels in serum seemed to decrease with increasing severity of lipid vacuolation, with an increase of TAG concentration in liver sections extracted for total lipids. Peripheral fats break down into free fatty acids and enter the liver through the blood stream, hence, the lower levels of serum TAG despite increasing severity of liver steatosis. Serum and liver cholesterol levels exhibit the same trend in TAG movement: decreasing serum cholesterol levels against increasing severity of FCL in contrast to increased liver cholesterol concentrations with FCL severity.

Expression of PPAR- γ was observed, indicating macrophage activity which may be involved in hepatic recovery as well as fatty acid oxidation. PPAR- δ expression was observed as well, a signifier of increased lipogenesis and consequently induction of inflammatory reactions.

Blood biomarkers (ALT, TAG, urea and creatinine) and liver TAG concentrations reflect on the histopathological changes in FHL cats in correlation to their FCL severity, coupled with hepatic gene expression of PPAR- δ and PPAR- γ in promoting hepatic recovery from FHL. These blood biomarkers in addition to further studies into blood-gene expression of the PPAR constituents could lead to earlier and less invasive diagnosis of FHL in cats.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

MENJELASKAN LIPIDOSIS HEPATIK FELIN MELALUI BIOKIMIA SERUM, HISTOPATOLOGI DAN UNGKAPAN GEN PADA KUCING-KUCING TERBIAR DI LEMBAH KLANG

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Lipidosis hepatik felin merupakan disfungsi proses metabolisme yang berkait dengan metabolisme lipid di dalam kucing. Kucing yang menghidap lipidosis hepatik akan mengalami pengumpulan lipid secara berlebihan di dalam hati disebabkan ketidakupayaan kucing mencerna secara cekap lipid-lipid tersebut serta menggunakannya. Oleh itu, pengumpulan lipid di dalam hepatosit, selain petanda-petanda klinikal yang lain bakal menjadi sekiranya penyakit ini berlarutan. Punca utama lipidosis hepatik masih lagi belum terurai. Pengajian ini bertujuan untuk menentukan kandungan lipid di dalam sampel-sampel kucing terbiar di Lembah Klang dan mengaitkannya kepada penganalisisan biokimia darah dan hati mereka, perubahan-perubahan histopatologinya, serta memastikan penyataan genetik hepatik yang berkait dengan pengawalan penggunaan tenaga melalui metabolisme lipid.

Secara kerseluruhannya, 18 kucing terbiar yang kurus kering, dinyahhidratkan, dan lesu telah dikumpul dari paun-paun di dalam kawasan Lembah Klang. *Alanine aminotransferase* (ALT), biopenanda yang lazim digunakan bagi memastikan kecedaraan hati pada haiwan-haiwan kecil, telah diukur daripada serum darah, serta kreatinina dan urea bagi memastikan kebolehfungsian buah pinggang. Paras trigliserida (TAG) dan kolesterol telah diukur di dalam serum melalui penganalisisan serum dan di dalam hati melalui pengekstrakan lipid *Bligh* dan *Dyer* bagi pengatian tahap steatosis. Analisis histologi keratan hati telah dilaksana bagi menentukan tahap keparahan perubahan lemak di dalam hati (PLH).

Pemerhatian sampel-sampel bertahap tinggi PLH pada histologinya menunjukkan jumlah ALT, kretinina dan urea menaik, menunjukkan kejadian keradangan hati, masalah renal dan penyahhidratan. Walaubagaimanapun, jumlah TAG di dalam darah didapati semakin menurun berdasarkan kenaikan keparahan PLH, tetapi terdapat kenaikan jumlah TAG di dalam keratan-keratan hati berdasarkan pengekstrakan lipid keseluruhan. Pemecahan lemak periferi menjadi asid lemak bebas akan memasuki hati melalui aliran darah, justeru, menjelaskan penurunan jumlah TAG serum dan menguatkan lagi keparahan steatosis hati. Jumlah kolesterol serum dan hati mempamerkan trend yang menyamai trend TAG: kolestrol serum semakin menurun apabila dibandingkan dengan peningkatan keparahan PLH sedangkan kolestrol hati semakin menaik selari dengan naiknya keparahan PLH.

Terdapat penyataan PPAR-γ yang menunjukkan kewujudan aktiviti makrofaj yang mungkin terlibat dalam penyembuhan hepatik selain pengoksidaan asid lemak. Penyataan PPAR-δ juga dicerap, suatu petanda ternaiknya kadar *lipogenesis* dan seterusnya penggalakan reaksi-reaksi keradangan.

Petanda-petanda darah (ALT, TAG, urea dan kreatinina) dan TAG hepatik mencerminkan perubahan-perubahan histopatologi dalam lipidosis hepatik felin yang berkait dengan keparahan PLH pada kucing, bersamaan ungkapan gen hepatik *PPAR-* δ and *PPAR-* γ yang menggalakkan kesembuhan hepatic daripada kesan PLH. Oleh itu, pengesanan awal lipidosis hepatik felin yang tak invasif mampu dilaksanakan menggunakan petanda-petanda tersebut di samping kajian lanjut pada ungkapan gen PPAR dalam darah.

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This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

α-KG	α-ketoglutaric
AA	Amino acids
ADP	Adenosine diphosphate
ALT	Alanine aminotransferase
ATP	Adenosine triphosphate
2-ME	2-mercaptoethanol
CN	Carnitine
CV	Central vein
DGAT	Diacylglycerol acyltransferase
DM	Diabetes mellitus
DNL	De novo lipogenesis
ERR-α	Estrogen related receptor- α
FA	Fatty acids
FATP1	Fatty acid transport protein-1
FCL	Fatty change of liver
FOX01	Fibroblast growth factor 21
FHL	Feline hepatic lipidosis
GH	Growth hormone
GI	Gastrointestinal
GSH	Glutathione
HE	Hepatic encephalopathy
HE	Hematoxylin and Eosin
HL	Hepatic lipidosis

IBD	Inflammatory bowel disease	
IL	Interleukin	
LCPUFA	Long-chain polyunsaturated fatty acid	
LD	Lipid droplet	
LPL	Lipoprotein lipase	
LPS	Lipopolysaccharides	
NAC	N-acetylcysteine	
NRF	Nuclear respiratory factor	
PCR	Polymerase chain reaction	
PEPCK	Phosphoenolpyruvate carboxykinase	
PLH	Perubahan lemak di dalam hati	
PPAR	Peroxisome proliferator activated receptor	
PT	Portal triad	
RT	Reverse transcription	
UC	Ulcerative colitis	
SAMe	S-adenosylmethionine	
SU	Sulphonylureas	
TNF	Tumor necrosis factor	
TAG	Triglycerides	
TPP	Thiamine pyrophosphate	
UV	Ultraviolet	
VAT	Visceral adipose tissue	
VLDL	Very low density lipoprotein	

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CHAPTER 1

INTRODUCTION

1.1 General Background of Study

The common cat, *Felis catus*, can be found in abundance in Malaysia. Many homeowners raise them for a variety of reasons including companionship, competition or even welfare. However, some domestic cats end up outside of their homes, abandoned, becoming strays on the streets. Among the common issues faced by these predatorial animals include feline hepatic lipidosis (FHL) where fatty deposition occurs in the liver of these cats. Prolonged anorexia in these stray cats force the use of all remaining fat stores, leading to the irregular build-up of the fats in the liver for storage. This study will be looking at abnormal lipid metabolism in stray cats, more specifically in FHL.

Feline hepatic lipidosis (FHL) is characterised by the excessive deposition of TAGs into the hepatocytes (greater than 80%), increasing the liver weight to more than 50% of its original weight. Secondary impairment of liver functions can occur as well as intrahepatic cholestasis (Valtolina & Favier, 2017a).

Unique dietary requirements of cats also bring concern to proper metabolic function, in particular to protein deficiency. A key feature of FHL is observed in anorectic cats where they can quickly suffer from protein malnutrition. This is due to their inability to adapt to diminishing protein resources in the body and continuously utilising the proteins, in particular, in urea cycle enzymes, and failing to conserve nitrogen (Center, 2005a). Ammonia detoxification into urea could be compromised, inflicting hyperammonemia in malnourished cats (Verbrugghe & Bakovic, 2013).

Lipidosis in cats is usually linked to anorexia, where, in healthy cats, prolonged periods of anorexia coupled with an underlying disease may lead to the development of lipidosis. This condition is coined "secondary HL". However, healthy cats can also develop lipidosis when they undergo rapid weight loss through food deprivation, or even unintentional food deprivation from their owner, food changes that the cat may not favour, unfavourable lifestyle changes or stress. Lipidosis due to such reasons is coined "primary HL". Evidence has shown overweight cats to exhibit FHL within two weeks in an experimental model, however, veterinarians share their experiences whereby FHL can develop faster in cases of percent losses in caloric intake against degree of adiposity, in about one week (Armstrong & Blanchard, 2009a).

In FHL, TAGs are deposited excessively in hepatocytes due to lipolysis from peripheral fat stores. The liver plays a vital role in regulating the body's energy homeostasis through metabolising glucose and fatty acids. Adenosine triphosphate (ATP), the body's main unit of energy, usually comes from the catabolism of glucose. In excess, glucose turns into glycogen for storage, which converts into fatty acids through lipogenesis. Through lipogenesis, TAGs are stored in white adipose tissue, where it can be stored virtually forever. However, in conditions of excessive food intake or impairment of fatty acid metabolism, the excessive lipids can be stored in large amounts in the liver through steatosis. In conditions where the body is low in glucose and insulin, hepatic glycogen stores are depleted and fatty acid production is reduced. Following this, adipose tissues are hydrolysed to catabolise TAGs into free fatty acids which enter the body's blood plasma. Upon reaching the liver, these free fatty acids are oxidised into ketone bodies to be used by tissues outside of the liver (Reddy & Sambasiva Rao, 2006).

1.2 Objective of Study

The welfare of stray cats in the Klang Valley is unknown, moreover if they suffer from the common metabolic disease that is FHL. Although there is much research into FHL, there is still a lack of the correlating factors between the blood biochemistry of cats with FHL, their histopathology and genetic expression related to the disease. Understanding these parameters can propose new insights into the workings of FHL.

This study was conducted to identify the biochemical parameters for cats undergoing lipidosis. The parameters of concern include blood serum and liver homogenate where several analyses were conducted respectively. Blood serum parameters reveal a broader physiological condition of the cat with liver function of concern. To go in hand with this data, liver samples were studied to identify presence and severity of lipidosis with further investigation into gene expression for lipid metabolism. Securing these data points allowed for a more wholesome investigation into lipidosis in the feline model. The study was conducted in line with these three objectives;

- 1. To determine serum biochemistry and lipid composition of serum and liver samples in cats in relation with FHL.
- 2. To assess histopathological changes associated with FHL in cats.
- 3. To detect targeted genes responsible for energy regulation through lipid metabolism associated with FHL from cats.

REFERENCES

- Armstrong, P. J., & Blanchard, G. (2009a). Hepatic Lipidosis in Cats. Veterinary Clinics of North America - Small Animal Practice, 39(3), 599–616. https://doi.org/10.1016/j.cvsm.2009.03.003
- Armstrong, P. J., & Blanchard, G. (2009b). Hepatic Lipidosis in Cats. Veterinary Clinics of North America - Small Animal Practice. https://doi.org/10.1016/j.cvsm.2009.03.003
- Armstrong, P. J., & Blanchard, G. (2009c). Hepatic Lipidosis in Cats. Veterinary Clinics of North America - Small Animal Practice. https://doi.org/10.1016/j.cvsm.2009.03.003
- Armstrong, P. J., & Williams, D. A. (2012). Pancreatitis in Cats. *Topics in Companion Animal Medicine*, 27(3), 140–147. https://doi.org/10.1053/j.tcam.2012.09.001
- Aroch, I., Shechter-Polak, M., & Segev, G. (2012). A retrospective study of serum β-hydroxybutyric acid in 215 ill cats: Clinical signs, laboratory findings and diagnoses. *Veterinary Journal*, 191(2), 240–245. https://doi.org/10.1016/j.tvjl.2011.01.010
- Aulbach, A. D., & Amuzie, C. J. (2017). Biomarkers in Nonclinical Drug Development. A Comprehensive Guide to Toxicology in Nonclinical Drug Development (Second Edi). Elsevier Inc. https://doi.org/10.1016/b978-0-12-803620-4.00017-7
- Barmore, W., Azad, F., & Stone, W. L. (2020). *Physiology , Urea Cycle*. NCBI Bookshelf.
- Baum, N., Dichoso, C. C., & Carlton, C. E. (1975). Blood urea nitrogen and serum creatinine. Physiology and interpretations. *Urology*, *5*(5), 583– 588. https://doi.org/10.1016/0090-4295(75)90105-3
- Bazelle, J., & Watson, P. (2014). Pancreatitis in cats: Is it acute, is it chronic, is it significant? *Journal of Feline Medicine and Surgery*, *16*(5), 395–406. https://doi.org/10.1177/1098612X14523186
- Boag, F., Weerakoon, J., Ginsburg, J., Havard, C. W., & Dandona, P. (1985). Diminished creatinine clearance in anorexia nervosa: Reversal with weight gain. *Journal of Clinical Pathology*, 38, 60–63. https://doi.org/10.1136/jcp.38.1.60
- Bolsoni-Lopes, A., & Alonso-Vale, M. I. C. (2015). Lipolysis and lipases in white adipose tissue An update. *Archives of Endocrinology and Metabolism*, *59*(4), 335–342. https://doi.org/10.1590/2359-399700000067

- Cárdenas, A., & Ginès, P. (2009). A Patient With Cirrhosis and Increasing Creatinine Level: What Is It and What to Do? *Clinical Gastroenterology and Hepatology*, *7*(12), 1287–1291. https://doi.org/10.1016/j.cgh.2009.08.010
- Center, S. A. (2005a). Feline hepatic lipidosis. Veterinary Clinics of North America: Small Animal Practice, 35(1), 225–269. https://doi.org/10.1016/j.cvsm.2004.10.002
- Center, S. A. (2005b). Feline Hepatic Lipidosis. Veterinary Clinics of North America - Small Animal Practice, 47(3), 225–269. https://doi.org/10.1016/j.cvsm.2004.10.002
- Center, S. A. (2007). Interpretation of Liver Enzymes. Veterinary Clinics of North America - Small Animal Practice. https://doi.org/10.1016/j.cvsm.2006.11.009
- Center, S. A., Crawford, M. A., Guida, L., Erb, H. N., & King, J. (1993a). A Retrospective Study of 77 Cats With Severe Hepatic Lipidosis: 1975-1990. Journal of Veterinary Internal Medicine, 7(6), 349–359. https://doi.org/10.1111/j.1939-1676.1993.tb01030.x
- Center, S. A., Crawford, M. A., Guida, L., Erb, H. N., & King, J. (1993b). A Retrospective Study of 77 Cats With Severe Hepatic Lipidosis: 1975– 1990. Journal of Veterinary Internal Medicine, 7(6), 349–359. https://doi.org/10.1111/j.1939-1676.1993.tb01030.x
- Center, S., Warner, K., Randolph, J., Sunvold, G., & Vickers, J. (2012). Influence of dietary supplementation with L-carnitine on metabolic rate, fatty acid oxidation, body condition, and weight loss in overweight cats. American Veterinary Medical Association.

Committee, W. G. N. (2013). Body Condition Score 1. WSAVA.

- Cristo, T. G., Biezus, G., Noronha, L. F., Gaspar, T., Dal Pont, T. P., Withoeft, J. A., ... Casagrande, R. A. (2019). Feline Leukaemia Virus Associated with Leukaemia in Cats in Santa Catarina, Brazil. *Journal of Comparative Pathology*, 170, 10–21. https://doi.org/10.1016/j.jcpa.2019.05.002
- Cullen, J. M., van den Ingh, T. S. G. A. M., Van Winkle, T., Charles, J. A., & Desmet, V. J. (2006). Morphological classification of parenchymal disorders of the canine and feline liver: 1. Normal histology, reversible hepatocytic injury and hepatic amyloidosis. In WSAVA Standards for *Clinical and Histological Diagnosis of Canine and Feline Liver Diseases* (pp. 77–83). Elsevier. https://doi.org/10.1016/B978-0-7020-2791-8.50010-1

- de la Rosa Rodriguez, M. A., & Kersten, S. (2017). Regulation of lipid dropletassociated proteins by peroxisome proliferator-activated receptors. *Biochimica et Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids*, (July), 0–1. https://doi.org/10.1016/j.bbalip.2017.07.007
- Delanaye, P., Cavalier, E., & Pottel, H. (2017). Serum Creatinine: Not so Simple! Nephron, 136, 302–308. https://doi.org/10.1159/000469669
- El-Gayar, S., Thüring-Nahler, H., Pfeilschifter, J., Röllinghoff, M., & Bogdan, C. (2003). Translational Control of Inducible Nitric Oxide Synthase by IL-13 and Arginine Availability in Inflammatory Macrophages. *The Journal* of Immunology, 171(9), 4561–4568. https://doi.org/10.4049/jimmunol.171.9.4561
- Glavind, E., Aagaard, N. K., Grønbæk, H., Møller, H. J., Orntoft, N. W., Vilstrup, H., & Thomsen, K. L. (2016). Alcoholic Hepatitis Markedly Decreases the Capacity for Urea Synthesis. *PLOS ONE*, *11*(7), e0158388. https://doi.org/10.1371/journal.pone.0158388
- Hall, J. A., Barstad, L. A., & Connor, W. E. (1997). Lipid Composition of Hepatic and Adipose Tissues From Normal Cats and From Cats With Idiopathic Hepatic Lipidosis. *Journal of Veterinary Internal Medicine*, 11(4), 238– 242. https://doi.org/10.1111/j.1939-1676.1997.tb00097.x
- Hamper, B., Bartges, J., Kirk, C., Witzel, A. L., Murphy, M., & Raditic, D. (2012). *The Unique Nutritional Requirements of the Cat: A Strict Carnivore. The Cat.* Elsevier Inc. https://doi.org/10.1016/B978-1-4377-0660-4.00015-6
- Higgins, C. (2016). Urea and the clinical value of measuring blood urea concentration. *Acutecaretesting.Org*, (August), 1–6. Retrieved from https://acutecaretesting.org/-/media/acutecaretesting/files/pdf/urea-and-the-clinical-value-of-measuring-blood-ans-approved.pdf%0Ahttps://acutecaretesting.org/~/media/acutecaretesting.g/files/pdf/urea-and-the-clinical-value-of-measuring-blood-ans-approved.pdf%0Ahttps://acutecaretesting.org/~/media/acutecaretesting.plood-ans-approved.pdf/urea-and-the-clinical-value-of-measuring-blood-ans-approved.pdf/urea-and-the-clinical-value-of
- Hinz, B., Phan, S. H., Thannickal, V. J., Galli, A., Bochaton-Piallat, M. L., & Gabbiani, G. (2007). The myofibroblast: One function, multiple origins. *American Journal of Pathology*, 170(6), 1807–1816. https://doi.org/10.2353/ajpath.2007.070112
- Hoenig, M. (2006). The cat as a model for human nutrition and disease, 584–588.
- Jegatheesan, P., & De Bandt, J. P. (2017). Fructose and NAFLD: The multifaceted aspects of fructose metabolism. *Nutrients*, *9*(3), 1–13. https://doi.org/10.3390/nu9030230

- Kalaitzakis, E., Roubies, N., Panousis, N., Pourliotis, K., Kaldrymidou, E., & Karatzias, H. (2007). Clinicopathologic evaluation of hepatic lipidosis in periparturient dairy cattle. *Journal of Veterinary Internal Medicine / American College of Veterinary Internal Medicine*, 21, 835–845. https://doi.org/10.1892/0891-6640(2007)21[835:CEOHLI]2.0.CO;2
- Kang, K., Reilly, S. M., Karabacak, V., Gangl, M. R., Hatano, B., & Lee, C. (2009). Adipocyte-dreved Th2 cytokines and myeloid PPAR delta. *Cell Metabolism*, 7(6), 485–495. https://doi.org/10.1016/j.cmet.2008.04.002.Adipocyte-derived
- Kiapidou, S., Liava, C., Kalogirou, M., Akriviadis, E., & Sinakos, E. (2020). Chronic kidney disease in patients with non-alcoholic fatty liver disease: What the Hepatologist should know? *Annals of Hepatology*, *19*(2), 134– 144. https://doi.org/10.1016/j.aohep.2019.07.013
- Kim, W. R., Flamm, S. L., Di Bisceglie, A. M., & Bodenheimer, H. C. (2008). Serum activity of alanine aminotransferase (ALT) as an indicator of health and disease. *Hepatology*, 47(4), 1363–1370. https://doi.org/10.1002/hep.22109
- Klaassen, J. K. (1999). Reference values in veterinary medicine. *Laboratory Medicine*, *30*(3), 194–197. https://doi.org/10.1093/labmed/30.3.194
- Krishna, M. (2013). Role of special stains in diagnostic liver pathology. *Clinical Liver Disease*, 2(S1), S8–S10. https://doi.org/10.1002/cld.148
- Kuzi, S., Segev, G., Kedar, S., Yas, E., & Aroch, I. (2017). Prognostic markers in feline hepatic lipidosis: A retrospective study of 71 cats. Veterinary Record, 181(19), 512. https://doi.org/10.1136/vr.104252
- Lawler, D. F., Chase, K., Teckenbrock, R., & Lark, K. G. (2006). Heritable components of feline hematology, clinical chemistry, and acid-base profiles. *Journal of Heredity*, *97*(6), 549–554. https://doi.org/10.1093/jhered/esl041
- Ley, K. (2017). M1 Means Kill; M2 Means Heal. *The Journal of Immunology*, *199*(7), 2191–2193. https://doi.org/10.4049/jimmunol.1701135
- Liss, K. H. H., & Finck, B. N. (2017). Biochimie PPARs and nonalcoholic fatty liver disease. *Biochimie*, 136, 65–74. https://doi.org/10.1016/j.biochi.2016.11.009
- Liu, J., Han, L., Zhu, L., & Yu, Y. (2016). Free fatty acids, not triglycerides, are associated with non-alcoholic liver injury progression in high fat diet induced obese rats. *Lipids in Health and Disease*, *15*(1). https://doi.org/10.1186/s12944-016-0194-7

- Lonsdale, D. (2006). A review of the biochemistry, metabolism and clinical benefits of thiamin(e) and its derivatives. *Evidence-Based Complementary and Alternative Medicine*, *3*(1), 49–59. https://doi.org/10.1093/ecam/nek009
- Mezey, E. (1982). Liver Disease and Protein Needs. *Annual Review of Nutrition*, 2(1), 21–50. https://doi.org/10.1146/annurev.nu.02.070182.000321
- Minamoto, T., Walzem, R. L., Hamilton, A. J., Hill, S. L., Payne, H. R., Lidbury, J. A., ... Steiner, J. M. (2018). Altered lipoprotein profiles in cats with hepatic lipidosis. *Journal of Feline Medicine and Surgery*. https://doi.org/10.1177/1098612X18780060
- Musso, G., Gambino, R., Tabibian, J. H., Ekstedt, M., Kechagias, S., Hamaguchi, M., ... Cassader, M. (2014). Association of Non-alcoholic Fatty Liver Disease with Chronic Kidney Disease: A Systematic Review and Meta-analysis. *PLoS Medicine*, *11*(7). https://doi.org/10.1371/journal.pmed.1001680
- Nakamura, M. T., Yudell, B. E., & Loor, J. J. (2014a). Regulation of energy metabolism by long-chain fatty acids. *Progress in Lipid Research*, *53*(1), 124–144. https://doi.org/10.1016/j.plipres.2013.12.001
- Nakamura, M. T., Yudell, B. E., & Loor, J. J. (2014b). Regulation of energy metabolism by long-chain fatty acids. *Progress in Lipid Research*. Elsevier Ltd. https://doi.org/10.1016/j.plipres.2013.12.001
- Orecchioni, M., Ghosheh, Y., Pramod, A. B., & Ley, K. (2019). Macrophage polarization: Different gene signatures in M1(Lps+) vs. Classically and M2(LPS-) vs. Alternatively activated macrophages. *Frontiers in Immunology*, 10(MAY), 1–14. https://doi.org/10.3389/fimmu.2019.01084
- Ostermann, M., Kashani, K., & Forni, L. G. (2016). The two sides of creatinine: both as bad as each other? *Journal of Thoracic Disease*, *8*(7), E628– E630. https://doi.org/10.21037/jtd.2016.05.36
- Perrone, R. D., Madias, N. E., & Levey, A. S. (1992). Serum creatinine as an index of renal function: New insights into old concepts. *Clinical Chemistry*, 38(10), 1933–1953.
- Porta, C., Riboldi, E., Ippolito, A., & Sica, A. (2015). Molecular and epigenetic basis of macrophage polarized activation. *Seminars in Immunology*, 27(4), 237–248. https://doi.org/10.1016/j.smim.2015.10.003
- Reddy, J. K., & Sambasiva Rao, M. (2006). Lipid Metabolism and Liver Inflammation. II. Fatty liver disease and fatty acid oxidation. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 290(5), G852–G858. https://doi.org/10.1152/ajpgi.00521.2005

- Shangraw, R. E., & Jahoor, F. (1999). Effect of liver disease and transplantation on urea synthesis in humans: relationship to acid-base status. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 276(5), G1145–G1152. https://doi.org/10.1152/ajpgi.1999.276.5.G1145
- Simões, I. C. M., Fontes, A., Pinton, P., Zischka, H., & Wieckowski, M. R. (2018). Mitochondria in non-alcoholic fatty liver disease. *International Journal of Biochemistry and Cell Biology*, 95(October 2017), 93–99. https://doi.org/10.1016/j.biocel.2017.12.019
- Simpson, K. W. (2015). Pancreatitis and triaditis in cats: causes and treatment, 56(January). https://doi.org/10.1111/jsap.12313
- Slack, A., Yeoman, A., & Wendon, J. (2010). Renal dysfunction in chronic liver disease. *Critical Care*, *14*(2), 214. https://doi.org/10.1186/cc8855
- Softic, S., Cohen, D. E., & Kahn, C. R. (2016). Role of Dietary Fructose and Hepatic de novo Lipogenesis in Fatty Liver Disease. *Digestive and Liver Disease*, 61(5), 1282–1293. https://doi.org/10.1007/s10620-016-4054-0
- Takabatake, T., Ohta, H., Ishida, Y., Hara, H., & Ushiogi, Y. (1989). Low serum creatinine levels in severe hepatic disease. *Archives of Internal Medicine*, 149(6), 1313–1315. https://doi.org/10.1001/archinte.1989.00390060173051
- Tan, N. S., Vázquez-Carrera, M., Montagner, A., Sng, M. K., Guillou, H., & Wahli, W. (2016). Transcriptional control of physiological and pathological processes by the nuclear receptor PPARβ/δ. *Progress in Lipid Research*, 64, 98–122. https://doi.org/10.1016/j.plipres.2016.09.001
- Thongprayoon, C., Cheungpasitporn, W., & Kashani, K. (2016). Serum creatinine level, a surrogate of muscle mass, predicts mortality in critically ill patients. *Journal of Thoracic Disease*, *8*(5), E305–E311. https://doi.org/10.21037/jtd.2016.03.62
- Valtolina, C., & Favier, R. P. (2017a). Feline Hepatic Lipidosis. Veterinary Clinics of North America - Small Animal Practice, 47(3), 683–702. https://doi.org/10.1016/j.cvsm.2016.11.014
- Valtolina, C., & Favier, R. P. (2017b). Feline Hepatic Lipidosis. Veterinary Clinics of North America - Small Animal Practice, 47(3), 683–702. https://doi.org/10.1016/j.cvsm.2016.11.014
- Valtolina, C., & Favier, R. P. (2017c). Feline Hepatic Lipidosis. Veterinary Clinics of North America - Small Animal Practice, 47(3), 683–702. https://doi.org/10.1016/j.cvsm.2016.11.014

- Valtolina, C., & Favier, R. P. (2017d). Feline Hepatic Lipidosis. Veterinary Clinics of North America - Small Animal Practice, 47(3), 683–702. https://doi.org/10.1016/j.cvsm.2016.11.014
- Verbrugghe, A., & Bakovic, M. (2013). Peculiarities of one-carbon metabolism in the strict carnivorous cat and the role in feline hepatic lipidosis. *Nutrients*, *5*(7), 2811–2835. https://doi.org/10.3390/nu5072811
- Walther, T. C., & Farese Jr., R. V. (2012). Lipid Droplets And Cellular Lipid Metabolism. *Annual Review of Biochemistry*, *81*, 687–714. https://doi.org/10.1146/annurev-biochem-061009-102430.Lipid
- Walther, T. C., & Farese, R. V. (2009). The life of lipid droplets. *Biochimica et Biophysica Acta Molecular and Cell Biology of Lipids*, 1791(6), 459–466. https://doi.org/10.1016/j.bbalip.2008.10.009
- Washabau, R. J., & Day, M. J. (2013). Canine and Feline Gastroenterology. (R. J. Washabau & M. J. Day, Eds.), Elsevier. Elsevier Inc. https://doi.org/https://doi.org/10.1016/C2009-0-34969-7
- Washabau, R. J., Meyer, H. P., Rothuizen, J., Lidbury, J. A., Steiner, J. M., Richter, K., ... Watson, P. (2013). Liver. In *Canine and Feline Gastroenterology* (pp. 849–957). Elsevier. https://doi.org/10.1016/B978-1-4160-3661-6.00061-4
- Willard, M. D., & Twedt, D. C. (2003). Gastrointestinal, pancreatic, and hepatic disorders. Small Animal Clinical Diagnosis by Laboratory Methods (4th ed.). Elsevier Inc. https://doi.org/10.1016/B0-72-168903-5/50014-8
- Witzel, A. L., Bartges, J., Kirk, C., & Hamper, B. (2012). Nutrition for the Normal Cat, 243–247. https://doi.org/10.1016/B978-1-4377-0660-4.00016-8
- YERIAN, L. (2011). Histopathological evaluation of fatty and alcoholic liver diseases. *Journal of Digestive Diseases*, *12*(1), 17–24. https://doi.org/10.1111/j.1751-2980.2010.00472.x
- Zawie, D. A., & Garvey, M. S. (1984). Feline hepatic disease. *The Veterinary Clinics of North America. Small Animal Practice*, 14(6), 1201–1230. https://doi.org/10.1016/S0195-5616(84)50154-5
- Zini, E., Linscheid, P., Franchini, M., Kaufmann, K., Monnais, E., Kutter, A. P., ... Reusch, C. E. (2009). Partial sequencing and expression of genes involved in glucose metabolism in adipose tissues and skeletal muscle of healthy cats. *Veterinary Journal*, 180(1), 66–70. https://doi.org/10.1016/j.tvjl.2007.10.022



activated receptor-5. DNA ladder *300bp, **400bp. (c) Gene expression of peroxisome proliferator activated receptor-y. Gene expression of Interleukin-6 (IL6). DNA ladder *300bp, **400bp. (b) Gene expression of peroxisome proliferator DNA ladder *400bp, **500bp. Numbers represent cat sample number from study

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Appendix B

Sample ID	Concentration (ng/µl)	260/280 Ratio
1	444.88	2.11
3	682.16	2.11
4	505.04	2.1
9	1095.52	2.12
15	368.88	2.13
16	391.76	2.11
18	289.04	2.12
29	324.8	2.11
30	339.76	2.12
31	224.16	2.1
32	440.88	2.12
35	291.36	2.1
40	350.56	2.13
42	289.52	2.12
46	113.04	1.97
55	377.44	2.13
57	356	2.13
63	193.6	2.11
65	184.24	2.11
67	379.28	2.11
69	89.52	2.15
71	306	2.12
73	542.4	2.11

Sample ID of samples and respective concentration of extracted RNA and 260/280 ratio

G

Appendix C

Protocol:	Supplier
Anaesthetisation	
Item:	Virbac, Hamilton, New Zealand
Zoletil	
Euthanisation	
Item: Dolethal	Vetoquinol, Northamptonshire, United Kingdom
Blood serum separation	
Item: Eppendorf centrifuge 5702 R	Merck, Darmstadt, Germany
Liver sample storage	
Item:	
Formalin	Sigma-Aldrich, Missouri, United States of America
RNAlater Stabilization Solution	Invitrogen by Thermo Fisher Scientific, Vilnius,
Liver lipid extraction	Lithuania
Item:	and a second a second as a
Methanol	DCU sharen Denskek Theiland
Chloroform	KUI Lapscan, Bangkok, Inaliand
Pierce BCA Protein Assay	Thermo Fisher, Illinois, United States of America
Liver processing for histology	
Item:	
TP1020 Semi-enclosed Benchtop Tissue Processor Hematoxylin & Eosin	Leica Biosystems, Illinois, United States of America
Alcohol (70-100%)	Veterinary Histopathology Lab, Faculty of Veterinary Medicine UPM
Paraffin	
Xvlene	
RM2045 Manual Microtome	Leica Biosystems, Illinois, United States of America
Serum biochemistry analysis	
Item:	
Automated chemistry analyer TRX 7010	Rierov, Mannheim Germany
Biolis 24i	blorex, Marinneini Gernary
PCR	
Item:	
RNeasy Mini Kit	Qiagen, Maryland, United States of America
Access RT-PCR System	
RNAsin [™] Ribonuclease Inhibitors	Promeda Corporation Wisconsin United States of
Agarose powder	America
Loading dye (Blue/Orange 6×)	Amonou
DNA ladder, 25bp, 100bp	
PCR primers	Apical Scientific, Selangor, Malaysia
Sensoquest Labcycler®	Sensoquest GmbH, Göttingen, Germany
Thermocycler	-
2-mercaptoethanol	
TAE buffer	Biorad, California, United States of America
Gel Doc XR + Image Lab Software	

List of materials used and respective suppliers.

BIODATA OF STUDENT

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Citizenship	: Malaysian
Age	: 27 (2020
Marital Status	: Single

Born in 1993, Fadzly is the eldest among three younger siblings. Primary education was a mix between local studies in Selangor, Malaysia, up to Primary 3, then migrated to Wyoming, United States of America, for his 5-6th grade. His father was studying for his doctorate, allowing for the opportunity for the family to experience education in a foreign land. Upon completion of the doctorate, the family moved back to Malaysia where Fadzly continued his secondary education in Sekolah Menengah Kebangsaan Jalan Reko, Kajang, Selangor, Malaysia. There, he was introduced to the field of sciences, with teachers whom showed passion for their syllabus, sparking a greater interest in him to pursue the field in Kolej Matrikulasi Pulau Pinang, Penang, Malaysia. A year of the matriculation program allowed Fadzly to continue in the field of biology where he pursued a Bachelor's Degree in cellular and molecular biology in Universiti Putra Malaysia, Serdang, Malaysia. Challenges were faced as he delved deeper into science, culminating hard work and perseverance which allowed his graduation from the Bachelor's program with a second class honour's. Trudging on, he continued with a Master's program in the pursuit of knowledge and even greater selfdiscovery.

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