

UNIVERSITI PUTRA MALAYSIA

PREVALENCE OF PREGNANCY KETOSIS IN GOAT FARMS, BLOOD PROFILING AND HISTOPATHOLOGICAL CHANGES IN LATE GESTATION AND POST-PARTUM GOATS WITH KETOSIS

MUHAMAD AFFAN BIN AB AZID

FPV 2022 3



PREVALENCE OF PREGNANCY KETOSIS IN GOAT FARMS, BLOOD PROFILING AND HISTOPATHOLOGICAL CHANGES IN LATE GESTATION AND POST-PARTUM GOATS WITH KETOSIS



By

MUHAMAD AFFAN BIN AB AZID

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Science

June 2021

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia

 \mathbf{C}



DEDICATION

This thesis is wholeheartedly dedicated to my beloved parents, Ab Azid bin Mat Jusoh and Nazrini binti Mat Zain who have been my source of inspiration that gave me strength when I thought of giving up and who continually provide their moral, spiritual and financial supports to me throughout this journey.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

PREVALENCE OF PREGNANCY KETOSIS IN GOAT FARMS, BLOOD PROFILING AND HISTOPATHOLOGICAL CHANGES IN LATE GESTATION AND POST-PARTUM GOATS WITH KETOSIS

By

MUHAMAD AFFAN BIN AB AZID

June 2021

Chairman : Hasliza Abu Hassim, PhD Faculty : Veterinary Medicine

Pregnancy ketosis is one of the common metabolic diseases that affect goats' meat and milk production, resulting in low productivity and hence, high economic losses. Thus, this study aimed to evaluate the prevalence of natural ketosis in selected dairy goat farms and its associated risk factors, and also to study the serum biochemical profiles and the histopathological changes in late gestation and post-partum goats with experimental ketosis. A total of 255 pregnant goats from ten farms from Terengganu (n=5) and Selangor (n=5) were screened for ketosis, which comprised of 20 to 30 random pregnant goats for each farm. Chi square analysis was done to determine the risk factors that associated with the prevalence of ketosis in the farms. For experimental trial, twelve (n=12) pregnant goats were divided into control (n=3) and treatment (n=9) groups. The control group was fed diet with adequate energy, while the treatment group was exposed to 50% reduction in the energy intake to induce ketosis. Blood was collected at weekly intervals for biochemical analysis, which included glucose, Beta-hydroxybutyrate (BHBA), free fatty acid (FFA), calcium, sodium, potassium, chloride, cortisol and insulin. On days 20 (PK20) and 40 (PK40) post-induction, 3 induced and 1 control goats were slaughtered while the remaining 3 ketosis-induced goats were provided with normal balanced diet to allow for recovery (PKRD). At slaughter, the liver, heart and brain were collected for histopathological study. Following ketosis screening, Selangor farms showed high prevalence (47.33%) of ketosis as compared to Terengganu farms (20.97%), which was characterized by high BHBA level ($\geq 0.08 \text{ mmol/L}$) and presence of clinical signs. The survey revealed that all the nutritional risk factors, namely type, composition, nutritive value and quality of feed were significantly associated (p<0.05) with the prevalence of ketosis. Moreover, the experimental induction resulted in acute ketosis after 20 days with clinical signs including weakness, low body condition score and recumbency. Chronic ketosis was observed at 40 days, showing incoordination and abortion. Following re-introduction of adequate energy supply, the recovered PKRD goats showed no clinical sign. There were significant (p<0.05) decrease in the glucose,

insulin, calcium and potassium levels among the induced goats while the concentrations of BHBA, FFA and cortisol were significantly (p<0.05) higher. Histopathological examinations revealed that the liver of goats with ketosis showed fatty degeneration and congestion with polymorphonuclear leukocytes and mononuclear cells infiltrations. The heart of ketotic does were congested while the brain had congestion and cerebellar spongiosis. Indeed, pregnancy ketosis has been associated with insufficient diet during the late stage of pregnancy which the severity of the disease caused changes in serum biochemical and hormonal profiles during the late gestation and post-partum period of the goats as well as the histopathological changes of various organs such as liver, heart and brain.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PREVALENS KETOSIS KEBUNTINGAN DI LADANG-LADANG KAMBING, PROFIL DARAH DAN HISTOPATOLOGI PADA KAMBING KETOSIS SEMASA PERINGKAT AKHIR KEBUNTINGAN DAN POSTPARTUM

Oleh

MUHAMAD AFFAN BIN AB AZID

Jun 2021

Pengerusi : Hasliza Abu Hassim, PhD Fakulti`: Perubatan Veterinar

Ketosis kebuntingan merupakan salah satu penyakit metabolik yang sering kali mempengaruhi pengeluaran daging dan susu kambing jaitu boleh menyebabkan pengeluaran yang rendah dan kerugian yang tinggi pada ekonomi. Oleh itu, kajian ini bertujuan untuk menilai prevalens ketosis secara semula jadi di ladang-ladang kambing tenusu terpilih dan faktor-faktor risiko yang berkait dan juga untuk mengkaji perubahanperubahan pada profil serum biokimia dan histopatologi semasa tempoh akhir kebuntingan dan postpartum pada kambing-kambing melalui eksperimen ketosis. Sebanyak 255 ekor kambing bunting dari sepuluh ladang dari Terengganu (n=5) dan Selangor (n=5) telah disaring untuk ketosis kebuntingan yang mana telah melibatkan 20 ke 30 ekor kambing bunting yang rawak bagi setiap ladang. Analisis Khi kuasa dua telah dijalankan bagi mengenalpasti faktor-faktor risiko yang berkait rapat dengan kadar prevalens ketosis di ladang-ladang tersebut. Untuk ujikaji percubaan, dua belas (n=12) ekor kambing bunting telah dibahagikan kepada kumpulan kawalan (n=3) dan rawatan (n=9). Kumpulan kawalan telah diberi makan dengan tenaga yang mencukupi manakala kumpulan rawatan telah dihadkan pengambilan tenaga sehingga 50% daripada keperluan harian untuk mendorong ketosis kebuntingan. Sampel-sampel darah telah dikumpul setiap minggu untuk analisis biokimia yang melibatkan glukosa, Beta-hidroksibutirat (BHBA), asid lemak bebas (FFA), kalsium, elektrolit (natrium, kalium, klorida), kortisol dan insulin. Pada hari ke 20 (PK20) dan 40 (PK40) pasca induksi, tiga (n=3) kambing rawatan dan satu (n=1) kambing kawalan telah disembelih manakala tiga (n=3) kambing rawatan yang selebihnya telah diberi diet normal yang seimbang untuk pemulihan (PKRD). Semasa penyembelihan, organ hati, jantung dan otak telah diambil untuk kajian histopatologi. Dalam saringan ketosis, ladang di Selangor menunjukkan prevalens ketosis yang tinggi (47.33%) berbanding dengan ladang di Terengganu (20.97%) yang mana bercirikan dengan kepekatan BHBA yang tinggi (≥0.8 mmol/L) dan kewujudan tanda-tanda klinikal ketosis. Tinjauan menunjukkan yang semua faktor-faktor risiko iaitu jenis, komposisi, kadar nutrisi dan kualiti makanan adalah berkait (p<0.05) dengan prevalens ketosis. Seterusnya, induksi ujikaji telah mengakibatkan ketosis kebuntingan akut selepas 20 hari dengan tanda-tanda klinikal termasuk kelemahan, kemerosotan keadaan badan dan sentiasa terbaring. Ketosis yang kronik telah dikesan pada hari ke-40, iaitu menunjukkan tanda-tanda seperti pergerakan tidak seimbang dan juga keguguran. Hasil daripada pemberian semula bekalan tenaga yang mencukupi, kambing pemulihan PKRD tidak menunjukkan sebarang tanda klinikal. Terdapat penurunan yang signifikan (p<0.05) pada kadar glukosa, insulin, kalsium dan kalium dekat kambing yang telah didorong dengan ketosis manakala terdapat pertambahan yang signifikan (p<0.05) bagi kepekatan BHBA, FFA dan kortisol. Hasil daripada pemerhatian histopatologi menunjukkan yang tisu hati bagi kambing ketosis telah dikenalpasti dengan kewujudan hati berlemak, pengumpulan darah, leukosit polimorfonuklear dan penyusupan sel mononuklear. Tisu hati pada kambing ketosis telah dikesan dengan lesi pengumpulan darah manakala tisu otak juga telah dikesan dengan lesi pengumpulan darah dan serebelum spongiosis. Sesungguhnya, ketosis kebuntingan telah dikenalpasti sebagai penyakit yang berkait dengan kekurangan diet semasa peringkat akhir kebuntingan yang mana penyakit itu telah menyebabkan perubahan pada profil serum biokimia dan hormon semasa tempoh akhir kebuntingan dan postpartum pada kambing serta menyebabkan perubahan pada pelbagai organ seperti hati, jantung dan otak.

ACKNOWLEDGEMENT

In the name of God, the Most Gracious, the Most Merciful...

All praises to Allah S.W.T for His blessings for completion of this Master thesis. I am thankful to Him for giving me the strength, knowledge, ability and opportunity to undertake this research study and to persevere and complete it satisfactorily. Without His blessings, this achievement would not have been possible.

First and foremost, I would like to show my greatest appreciation to my supervisory committee. To my supervisor, Assoc. Prof. Dr. Hasliza Abu Hassim, thank you for your guidance and encouragement throughout this project. Your patience, understanding and coolness make me feel comfortable whenever I have lost my path in this project. Not to forget to my co-supervisors, Prof Zamri Saad and Dr Annas Salleh, for their guidance and opinions regarding my project. I thank you all for being a great supervisory committee ever.

Next, I want to express my thankfulness to the fellow graduate students in particular, Megat Nadzrin, Dr Afifi, Amirul Faiz, Dr Taqiuddin, Azri, Hashikin, Fadzlin, Zaid, Qayyum, Adha, Fariz, Zakiah and Mursyidah for helping me during the sampling, experimental trial, lab works and during my difficult times. I also would like to take this opportunity to thank UPM staffs, especially Puan Farah (Nutrition lab), Puan Latifah and Puan Jamilah (Histopathology lab), Encik Raziman (ARC), Encik Nizam (Slaughter house), Encik Eddy and Encik Meng (ITAFoS) for the technical help and cooperation to make sure I can use all facilities for my project. Without you guys, this project would not be successful.

Another group of people that I will not forget is my beloved parents, Ab Azid Mat Jusoh (Ayah) and Nazrini Mat Zain (Ma) as well as my siblings, Alif, Asraf and Alya for their du'a, moral and financial support throughout my master journey. Without them, I would not be able to come until the end of this journey.

Besides, I would like to thank Institute of Agriculture and Tropical of Food Security (ITAFoS) for their cooperation for giving me permission to do my experimental trial and usage of all facilities provided in their place. Last but not least, I would like to thank Universiti Putra Malaysia (UPM) for funding this study through Matching-Geran Putra/2018/9300439. Without the fund support, this project would not be started.

I certify that a Thesis Examination Committee has met on 29th June 2021 to conduct the final examination of Muhamad Affan bin Ab Azid on his thesis entitled "Prevalence of Pregnancy Ketosis in Goat Farms, Blood Profiling and Histopathological Changes in Late Gestation and Post-partum Goats with Ketosis" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Hafandi bin Ahmad, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

Siti Zubaidah binti Ramanoon, PhD

Senior Lecturer Faculty of Veterinary Medicine Universiti Putra Malaysia (Internal Examiner)

Jasni bin Sabri, PhD Professor Faculty of Veterinary Medicine

Universiti Malaysia Kelantan (External Examiner)

> Prof. Ts. Dr. Zuriati Ahmad Zukarnain Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Hasliza binti Abu Hassim, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

Mohd Zamri bin Saad, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

Annas bin Salleh, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 14 April 2022

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	
Signature.	_

Date:

Name and Matric No.: Muhamad Affan bin Ab Azid

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of	
Supervisory	
Committee:	Assoc. Prof. Dr. Hasliza Abu Hassim
Signature:	
Name of Member of	
Supervisory	
Committee:	Prof. Dr. Mohd Zamri Saad
Signature:	
Name of Member of	
Supervisory	
Committee:	Assoc. Prof. Dr. Annas Salleh

TABLE OF CONTENTS

		Page
ABSTRACT		i
ABSTRAK		iii
ACKNOWLED	GEMENT	v
APPROVAL		vi
DECLARATIO	N	viii
LIST OF TABL	ES	xiii
LIST OF FIGU	RES	XV
LIST OF APPE	NDICES	xvii
LIST OF ABBR	EVIATIONS	xviii
CHAPTER		
1	INTRODUCTION	1
	1.1 Background of study	1
	1.2 Problem statement	2
	1.3 Objectives	2
	1.4 Hypotheses	2
		2
2	LITERATURE REVIEW	3
	2.1 Factors affecting livestock production in Malaysia	3
	2.2 Pregnancy ketosis in small runnants	4
	2.5 Prevalence and fisk factors of pregnancy ketosis	3
	2.4 Diagnosis of pregnancy ketosis	7
	2.4.1 Subclinical signs	7
	2.4.2 Chinical signs	/ Q
	2.5 1 Dietary changes and negative energy	8
	balance during pregnancy ketosis	8
	2.5.2 Metabolic changes during pregnancy	8
	ketosis	0
	2.5.2.1 Lipolysis in adipocyte	8
	2.5.2.2 Oxidation of fatty acid and	10
	ketogenesis in liver	10
	2.6 Biochemical and histopathological changes during	13
	pregnancy ketosis in goats	
	2.6.1 Dlood mediling shanges during programmy	12
	2.6.1 Blood profiling changes during pregnancy	15
	2.6.2 Histopathological changes in internal	14
	organs during pregnancy ketosis	14
	2.7 Challenges in diagnosing pregnancy ketosis	15
	2.7 Containing of pregnancy Recosts 2.8 Potential control and prevention of pregnancy	16
	ketosis	10
	PREVALENCE OF DDECNANCY KETOSIS	18
5	DURING I ATE GESTATION IN SEI FOTED COAT	10
	FARMS IN TERENGCANII AND SELECTED GOAT	
	3.1 Introduction	18
		10

3.2 Materials and methods 19

	3.2.1	Study design	19
	3.2.2	Farms selection	19
	3.2.3	Sample size	20
	3.2.4	Animal selection	20
	3.2.5	Survey analysis	20
		32.5.1 Questionnaire pre-testing	21
		3252 Questionnaire form outlines	21
	276	Fand analysis	21
	5.2.0	2.2.6.1 Determination of dry matter and	$1 \qquad 22$
		5.2.0.1 Determination of dry matter and	. 22
		moisture	22
		3.2.6.2 Determination of crude protein	22
		3.2.6.3 Determination of crude fiber	23
		3.2.6.4 Determination of crude fat	24
		3.2.6.5 Determination of metabolizable energy	25
	3.2.7	Measurement of body condition score	25
	3.2.8	Presence of clinical signs	25
	3.2.9	Serum biochemical profiles	26
		3.2.9.1 Plasma collection	26
		3292 Serum biochemical and hormon	al 26
		profiles	.11 20
	3 2 10	Statistical analysis	27
33	Doculto	Statistical analysis	27
5.5	221	Cumror atu da	20
	2.2.1	Survey study	20
	3.3.2	Body condition score and chinical signs	20
	3.3.3	Feed analysis	32
	3.3.4	Serum biochemical profiles	34
	3.3.5	Prevalence of pregnancy ketosis in goa	ts 35
		from Terengganu and Selangor farms	
	3.3.6	Association between prevalence of	f 36
		pregnancy ketosis and its potential risk	
		factors in Terengganu and Selangor farms	
3.4	Discus	sion	38
3.5	Conclu	sion	40
SER	UM	BIOCHEMICAL PROFILES O	F 41
SUB	CLINIC	CAL AND CLINICAL KETOSIS I	N
GOA	TS DU	RING LATE GESTATION AND POST]-
PAR	TUM P	ERIOD	
4.1	Introdu	iction	41
4.2	Materia	als and methods	42
	421	Experimental design	42
	422	Measurement of body condition score	43
	1.2.2	Ketone strip test and clinical signs	/3
	12.5	Blood collection	43
	4.2.4 125	Serum biochemical and hormonal profiles	45 /2
	4.2.J 1 2 6	Statistical analysis	43
4.2	4.∠.0	Staustical analysis	43
4.3	Kesults		44
	4.3.1	Clinical signs of pregnancy ketosis	44
	4.3.2	Measurement of body condition score	44
	4.3.3	Ketone strip test	44

4

 \bigcirc

		4.3.4 Serum biochemical and hormonal profiles	47
	4.4	Discussion	51
	4.5	Conclusion	54
5	5 GROSS AND HISTOPATHOLOGICAL CHANGES OF SUBCLINICAL AND CLINICAL KETOSIS IN GOATS DURING LATE GESTATION AND POST- PARTUM PERIOD		55
	5.1	Introduction	55
	5.2	Materials and methods	56
		5.2.1 Experimental design	56
		5.2.2 Gross pathology and histopathology	56
		5.2.3 Histopathological examination	56
		5.2.3.1 Fixation and staining	56
		5.2.3.2 Microscopic study	56
		5.2.4 Statistical analysis	57
	5.3	Results	58
		5.3.1 Gross examination	58
	5.4	5.5.2 Histopathological changes	01 76
	5.4	Conclusion	70
5.5 Conclusion		//	
6	GEN	IERAL DISCUSSION	78
7	GEN	IERAL CONLCUSION AND	80
	REC	OMMENDATION	
	7. <mark>1</mark>	Conclusion	80
	7. <mark>2</mark>	Recommendation for future research	80
REFEREN	CES		82
APPENDIC	CES		91
BIODATA OF STUDENT 9			96
LIST OF PUBLICATIONS 97			97

xii

6

LIST OF TABLES

Table		Page
2.1	Serum biochemical profiles of pregnancy ketosis in goat	13
3.1	Determination of body condition score in goats	25
3.2	Clinical signs of pregnancy ketosis	26
3.3	History of selected pregnant goats in Terengganu and Selangor farms	28
3.4	Distribution of number of foetus and type of breed of selected pregnant goats in Terengganu and Selangor farms	29
3.5	Distribution of natural vegetation and climate changes and awareness of the farmers among respondents in Terengganu and Selangor farms	29
3.6	Body condition score of selected pregnant goats in Terengganu and Selangor farms	30
3.7	Clinical signs of pregnancy ketosis in selected goat farms in Terengganu and Selangor	31
3.8	Type and composition of feeds in Terengganu and Selangor farms	32
3.9	Nutritional composition of feeds in Terengganu and Selangor farms	33
3.10	Nutritional composition of feeds in total mixed rations in Terengganu and Selangor farms	34
3.11	Serum biochemical profiles of pregnant goats in Terengganu vs Selangor farms	34
3.12	Prevalence rate of pregnancy ketosis in Terengganu and Selangor farms	35
3.13	Association between BHBA concentration and risk factors (nutritional and non-nutritional) and prevalence of pregnancy ketosis in Terengganu and Selangor farms	37
4.1	Nutritional composition of feeds (Napier grass and goat concentrate)	42

4.2	Characteristics features of ketosis induced pregnant goats	45
4.3	Serum biochemical and hormonal profiles in treatment vs control group	48
5.1	Lesion scoring system of organs according to percentage of lesion distribution	57
5.2	Histopathological lesions scoring of liver in control and treatment groups (PK20, PK40 and PKRD)	67
5.3	Histopathological lesion scorings for the heart of control and treatment groups (PK20, PK40 and PKRD)	70
5.4	Histopathological lesions scoring of brain in control and treatment groups (PK20, PK40 and PKRD)	75

 \bigcirc

LIST OF FIGURES

Figure	3	Page
2.1	Metabolism of adipocyte	9
2.2	Lipolysis of adipocyte tissue	10
2.3	Metabolism linkage between the adipose tissue and liver during ketosis state	11
2.4	Pathway of ketogenesis during non-availability of oxaloacetate	12
3.1	Map of Peninsular Malaysia	19
4.1	Ketone strip results	46
4.2	Concentration of cortisol hormone in healthy pregnant and pregnancy ketotic does (PK20, PK40 and PKRD)	49
4.3	Concentration of insulin hormone in healthy pregnant and pregnancy ketotic does (PK20, PK40 and PKRD)	50
5.1	Comparison between the gross appearance of liver of control group (healthy pregnant does) and treatment group (pregnancy ketosis does)	58
5.2	Comparison between the hearts of control group (healthy pregnant does) and treatment group (pregnancy ketosis does)	59
5.3	Brain of healthy control pregnant goat vs treatment group with pregnancy ketosis	60
5.4	Hepatocytes of a healthy control doe showing absence of lipid droplets in liver tissue	62
5.5	Presence of clear fat droplets (arrows) in the cytoplasm of many hepatocytes of a doe in PK20 group, suggestive of hepatic lipidosis	63
5.6	Presence of intracytoplasmic lipid droplets (arrows) in almost all hepatocytes suggestive of severe hepatic lipidosis in a PK40 doe	64
5.7	Presence of polymorphonuclear cells	65
5.8	Infiltration of mononuclear cells	66

 \bigcirc

- 5.9 Normal histopathological appearance with absence of 68 lesions in the heart of a control does (healthy pregnant does)
- 5.10 Presence of cardiac congestion
- 5.11 Normal histopathological appearance with absence of lesion in the brain of a control doe (healthy pregnant doe)
- 5.12 Diffused, severe congestion (arrows) of the ependymal lining the brain ventricular system in a ketosis does (PK40)
- 5.13 Congestion (arrows) in the brain was prominent, especially in the submeningeal blood vessels of a ketosis doe (PK40)
- 5.14 Presence of scattering circular spongiosis (arrow) in the cerebellum of a ketosis does (PK40).

73

69

71

72

74

LIST OF APPENDICES

Appendix		Page
1	Example of questionnaires form for the survey	91
2	Healthy pregnant goat with body condition score of 3.5	93
3	Pregnancy ketotic does with body condition score of 2	93
4	Microplate reader for ELISA analysis	94
5	Chemistry analyser machine	94
6	Harris' Haematoxylin and Eosin (H&E) staining	95

(C)

LIST OF ABBREVIATIONS

AcAc	Acetoacetic acid
ANOVA	Analysis of variance
ATP	Adenosine triphosphate
BHBA	Beta-hydroxybutyrate
cAMP	Cyclic adenosine monophosphate
СоА	Coenzyme A
DM	Dry matter
DPX	Distyrene Plasticiser Xylene
EDTA	Ethylenediaminetetraacetic acid
ELISA	Enzyme-linked immunosorbent assay
FFA	Free fatty acid
GTP	Guanosine triphosphate
HSL	Hormone sensitive lipase
H&E	Haematoxylin and eosin
kJ	Kilo joule
ME	Metabolizable energy
mmol	Millimole
МЈ	Megajoule
PMN	Polymorphonuclear leukocytes
РК	Pregnancy ketosis
РКС	Palm kernel cake
PKRD	Pregnancy ketosis reversed diet
SEM	Standard error mean
SPSS	Statistical package for the social sciences

6

WMetabolic body weightμmMicrometreχ2Chi square

(C)



CHAPTER 1

INTRODUCTION

1.1 Background of study

Goat rearing plays a vital role for the economics of farming community. Goats are reared for meat, milk and hide. However, morbidity and mortality among goats, including pregnant goats have caused bad economic impact on the livelihood of marginal farmers. Pregnancy ketosis, also known as pregnancy toxaemia, is one of the common metabolic diseases in pregnant ruminant around the world that affect livestock's meat and milk productions (Bani Ismail et al., 2008). The disease is commonly occurring in ruminants such as goat, sheep and cattle during the late stage of pregnancy, which generally has a low morbidity rate (2-6%) but a high mortality rate (80%). It is caused by abnormal metabolism of carbohydrates and fats, as a result of negative energy balance that occurs during the late stage of gestation. It is characterized by relatively high concentrations of the ketone bodies specifically acetoacetate, Beta-hydroxybutyrate (BHBA) and acetone as well as low concentration of glucose in blood circulation (Brozos et al., 2011). According to Schlumbohm and Harmeyer (2014), obese ewes or does carrying multiple foetuses are at higher risk of developing the disease due to the high demand of feed intake.

In Malaysia, pregnancy ketosis has been observed but not been thoroughly studied such as the information regarding the detailed serum biochemical changes and disease development, risk factors and impact of ketosis on dairy goats. Information regarding the predisposing risk factors such as the nutritional and non-nutritional aspects that contributed to the outbreak of pregnancy ketosis in goat farms would help goat farmers as an early measure to prevent the occurrence of the disease. Diagnosis of clinical pregnancy ketosis is based on history, clinical signs (e.g.: teeth grinding, weakness, recumbency) and the serum biochemical profiles (Lima et al., 2012). In addition, the use of these serum biochemical profiles as biomarkers in developmental stage of pregnancy ketosis offers a promising opportunity to develop rapid and accurate test kit that could be used by dairy farmers or veterinarians to screen and diagnose herds for pregnancy ketosis.

Histopathological reports related to any disease are usually referred to data obtained from internal organ biopsy or during post-mortem examination. Many researchers claimed that during ketosis changes occur in some tissues, particularly the cerebral and cerebellar neuronal necrosis and vacuolation, early structural immaturity of placenta, and liver steatosis (Andrews, 2017; Jeffrey and Higgins, 2012). Thus, it is believed that understanding the pathogenesis of ketosis could highlight critical points that can be used to prevent the development of the disease from becoming more severe which will affect the dairy goats that may further affect various internal organs such as liver, heart and brain which can be observed through development of lesions.

In the present study, the prevalence of pregnancy ketosis from selected goat farms in Malaysia was determined. Apart from that, the serum biochemical profiles and histopathological changes also were analyzed during the late gestation and post-partum period of experimental pregnancy ketosis does.

1.2 Problem statement

Although pregnancy ketosis is frequently observed among sheep and goats, the prevalence of the disease in goat farms in Malaysia has not been studied thus far. Hence, the predisposing risk factors in terms of nutritional and non-nutritional aspects that may cause the pregnancy ketosis in animal farms remained unknown. Furthermore, clinical ketosis could be classified as acute or chronic. Therefore, the serum biochemical profiles and histopathological changes in organs could be used to classify the stage of ketosis in pregnant animals. In facts, the observation of serum biochemical profiles can be used as an early prevention for subsequent attempt to treat or control pregnancy ketosis in dairy goat farms while the histopathological changes in various organs of affected animals can be used as a guideline or reference in pregnancy ketosis study. Therefore, understanding the risk factors on the prevalence of pregnancy ketosis in dairy goat, the serum biochemical profiles and the histopathological changes in experimental trial warrant further investigation.

1.3 Objectives

- a) To evaluate the prevalence of subclinical and clinical ketosis among dairy goats in Terengganu and Selangor farms.
- b) To evaluate the association between the risk factors and prevalence of subclinical and clinical ketosis among dairy goats in Terengganu and Selangor farms.
- c) To evaluate the serum biochemical profiles among pregnant goats with subclinical and clinical ketosis at late gestation and post-partum period.
- d) To observe the gross and histopathological changes in selected organs of pregnant goats with experimental ketosis at late gestation and post-partum period.

1.4 Hypotheses

High prevalence of pregnancy ketosis in Selangor farms as compared to Terengganu farms are associated with the risk factors (nutritional and non-nutritional) that contributed to occurrence of the disease in each state. The development of pregnancy ketosis in affected animals without proper treatment will cause severe changes in serum biochemical profiles and histopathological in various organs (e.g.: liver, heart, brain) during late gestation as well as postpartum period.

REFERENCES

- Abd-Elghany, H., Seham, Y., & Saad, S. (2010). Some immunohormonal changes in experimentally pregnant toxaemic goats. *Veterinary Medicine International*, 8: 5-11.
- Abd-Elghany, H., Seham, Y., & Saad, S. (2011). Hematobiochemical profile of pregnant and experimentally pregnancy toxemic goats. *Journal of Basic and Applied Chemistry*, 1(8): 65-69.
- Abdolmaleky, M. (2012). Predictions of small-farmers' empowerment to success in farm operations in Lorestan province, Iran. *World Applied Sciences Journal*, 20(10): 1416-1422.
- Abdullah, F. F. J., Rofie, A. M. B., Tijjani, A., Lim, E., Chung, T., Mohammed, K., & Abba, Y. (2015). Survey of goat farmers' compliance on proper herd health program practices. *International Journal of Livestock Research*, 5(11): 8–14.
- Abitbol, M. M., Driscoll, S. G., & Ober, W. B. (2011). Placental lesions in experimental toxaemia in the rabbit. *American Journal of Obstetrics and Gynaecology*, 125(7): 942–948.
- Agata, B., Mateusz, O., & Magdalena, H. D. (2018). Pregnancy ketonemia and development of the fetal central nervous system. *International Journal of Endocrinology*, 20(4): 1-7.
- Al-Qudah, K. M. (2011). Oxidant and antioxidant profile of hyperketonemic ewes affected by pregnancy toxaemia. *Veterinary Clinical Pathology*, 40(1): 60-65.
- Ali, O., Gholamali, M., & Mitra, B. (2013). Diagnosis, treatment and prevention of pregnancy toxaemia in ewes. *International Journal of Advanced Biological* and Biomedical Research, 1(11): 1452-1456.
- Anoushepour, A., Mottaghian, P., & Mehdi, S. (2014). The comparison of some biochemical parameters in hyperketonemic and normal ewes. *European Journal of Experimental Biology*, 4(3): 83-87.

Andrews, A. H. (2017). Pregnancy toxaemia in the ewe. In Practice, 19: 306-314.

- AOAC. (2007). Official methods of analysis. In: *Association of Official Analytical Chemists* (pp. 55-72). USA: Arlington Press.
- Arner, P. (2005). Human fat cell lipolysis: biochemistry, regulation and clinical role. Best Practice Research Clinical Endocrinology and Metabolism, 19(4): 471-482.
- Aziz, A. J. (2007). Proceedings from 19th Veterinary Association Malaysia Congress: Wealth creation through livestock production.

- Azmi, F. M. A, Ghani, A. A. A., Ajat, M., Lai, K. S., Zamri-Saad., Zuki, A. B., & Hassim, H. A. (2016). Histological changes of liver tissue and serobiochemical relation in does with pregnancy ketosis. *Jurnal Ilmu Ternak dan Veteriner*, 21: 96.
- Balikci, E., Yildiz, A., & Gurdogan, F. (2009). Investigation on some biochemical and clinical parameters for pregnancy toxaemia in Akkaraman ewes. *Journal of Animal and Veterinary Advances*, 8(7): 1268-1273.
- Bani Ismail, Z. A., Al-Rawashdeh, O., Al-Majali, A. M., & Amireh, F. (2015). Prevalence and risk factors for pregnancy toxemia of goats in Jordan. Animal Biology and Animal Husbandry, 4(2): 121-134.
- Bani Ismail, Z. A., Al-Majali, A. M., & Amireh, F. (2008). Metabolic profiles in goat does in late pregnancy with and without subclinical pregnancy toxaemia. *Veterinary Clinical Pathology*, 37(4): 434-437.
- Barakat, S. E. M., Al-Bhanasawi, G. E., Elazhari, N. M., & Bakhiet, A. O. (2007). Clinical and serobiochemical studies on naturally occurring pregnancy toxaemia in Shamia goats. *Journal of Animal and Veterinary Advances*, 6(6): 768-772.
- Benoit, M., & Laignel, G. (2010). Energy consumption in mixed crop-sheep farming systems: What factors of variation and how to decrease? *Journal of Animal and Veterinary Sciences*, 4(9): 1597-1605.
- Behera, A. K., Hildebrand, E., Bronson, R. T., Perides, G., Uematsu, S., Akira, S., & Hu, L. T. (2016). MyD88 deficiency results in tissue-specific changes in cytokine induction and inflammation in interleukin-18-independent mice infected with *Borrelia burgdorferi*. *Infection and Immunity*, 74: 1462-1470.
- Beitz, D. C. (2014). Carbohydrate Metabolism. In Reese, W.O. Dukes (12th Ed.), *Physiology of Domestic Animals* (pp. 501-515). Cornell University Press.
- Berk, P. D. (2008). Regulatable fatty acid transport mechanisms are central to the pathophysiology of obesity, fatty liver, and metabolic syndrome. *Hepatology* (*Baltimore, Md*), 48: 1362–1376.
- Block, S. S., Butler, W. R., Ehrhardt, R. A., Bell, A. W., Van Amburgh, M. E., & Boisclair, Y. R. (2011). Decreased concentration of plasma leptin in periparturient dairy cows is caused by negative energy balance. *Journal of Endocrinology*, 171: 339-348.
- Bobe, G., Young, J. W., & Beitz, D. C. (2014). Invited review: pathology, etiology, prevention, and treatment of fatty liver in dairy cows. *Journal of Dairy Science*, 87(10): 3105–3124.
- Bradford, P. (2006). *Large Animal Internal Medicine*. University of California: Mosby Press.

- Brockmanr, P., & Laarveld, B. (2010). Effects of insulin on net hepatic metabolism of acetate and 8-hydroxybutyrate in sheep (*Ovis aries*). Comp. Biochem. Physiol., 16(5): 255-257.
- Brown, C. R., Blaho, V. A., & Loiacono, C. M. (2013). Susceptibility to experimental Lyme arthritis correlates with KC and monocyte chemoattractant protein-1 production in joints and requires neutrophil recruitment via CXCR2. *Journal* of Immunology, 171: 893-901.
- Brozos, C., Mavrogianni, V. S., & Fthenakis, G. C. (2011). Treatment and control of peri parturient metabolic diseases: pregnancy toxaemia, hypocalcaemia, hypomagnesemia. *Veterinary Clinical North America: Food Animal Practice*, 27: 105-113.
- Burswell, J, Hadd, P., & Bywater, J. (2009). Treatment of pregnancy toxemia in sheep using a concentrated oral rehydration solution. *Veterinary Record*, 118: 208-209.
- Callan, R. J., & Van-Metre, D. C. (2014). Viral diseases of the ruminant nervous system. *Veterinary Clinical North America: Food Animal Practice*, 20: 327.
- Cal-Pereyra, L., Borteiro, C., Benech, A., Rodas, E., Abreu, M. N., Cruz, J. R., & Montana, G. (2009). Histological changes of the liver and metabolic correlates in ewes with pregnancy toxemia. *Arquivo Brasileiro de Medicina Veterianaria Zootecnia*, 61(2): 306-312.
- Cal-Pereyra, L., Benech, A, González-Montaña, Acosta-Dibarrat, J., Da Silva, S., & Martín, A. (2015). Changes in the metabolic profile of pregnant ewes to an acute feed restriction in late gestation. *New Zealand Veterinary Journal*, 63(3): 141-146.
- Chandrawathani, P., Chang, K. W., Nurulaini, R., Waller, P. J., Adnan, M., Zaini., C. M., Jamnah, O., Khadijah, S., & Vincent, N. (2006). Daily feeding of fresh Neem leaves (*Azadirachta indica*) for worm control in sheep. *Tropical Biomedicine*, 23(1): 23-30.
- Collard, B. L., Boettcher, P. J., Dekkers, J. C. M., Petitclerc, D., & Schaeffer, L. R. (2010). Relationships between energy balance and health traits of dairy cattle in early lactation. *Journal of Dairy Science*, 83: 2683-2690.
- Cotter, D. G., Avignon, D. A., Wentz, A. E., Weber, M. L., & Crawford, P. A. (2011). Obligate role for ketone body oxidation in neonatal metabolic homeostasis. *Journal of Biological Chemistry*, 286(9): 6902–6910.
- Crnkic, C., & Hodzic, A. (2012). Nutrition and Health of Dairy Animals. *Overview of Animal Nutrition, Management and Health* (pp. 1-64). New York: Wordy Publisher.
- Dalrymple, E. F. (2004). Pregnancy toxaemia in a ferret. *The Canadian Veterinary Journal*, 45: 150-152.

- Darzi, M. M. (2012). Pathology of *Taenia hydatigena* cysticercosis in a naturally infected Corrie dale lamb. *Veterinary Parasitology*, 16(2): 173–174.
- Department of Mineral and Geological Malaysia (JMG). (2020, November 24). Geological Map of Peninsular Malaysia. Scale 1:1 000 000. Retrieved from <u>https://jmg.gov.my/add_on/mt/smnjg/tiles/</u>
- Department of Statistics Malaysia. (2020, June 20). Demographic statistics first quarter 2020, Malaysia. Retrieved from https://www.dosm.gov.my/v1/index.php?r=column/cthemeByCat&cat=430.
- Department of Veterinary Services. (2019, August 13). Livestock statistics. Malaysia: Livestock population, 2015-2016. Retrieved from http://www.dvs.gov.my/dvs/resources/user 1/DVS%20pdf/Perangkaan%202.
- Delarue, J., & Magnan, C. (2007). Free fatty acids and insulin resistance. *Current Opinion in Clinical Nutrition and Metabolic Care*, 10: 142–148.
- Devendra, C. (2007). Enhancing animal protein supplies in Malaysia: Opportunities and challenges. *ASM Science Journal*, 1(1): 63-73.
- Dore, V., Dubuc, J., Belanger, A. M., & Buczinski, S. (2013). Short communication: Evaluation of the accuracy of an electronic on-farm test to quantify blood β hydroxybutyrate concentration in dairy goats. *Journal of Dairy Science*, 96: 4505-4507.
- Dubuc, J., Duffield, T. F., Leslie, K. E., Walton, K. E., & LeBlanc, S. J. (2012). Risk factors and effects of postpartum anovulation in dairy cows. *Journal of Dairy Science*, 95: 1845–1854.
- Duffield, T. F., Kelton, D. F., Leslie, K. E., Lissemore, K. D., & Lumsden, J. H. (2009). Use of test day milk fat and milk protein to detect subclinical ketosis in dairy cattle in Ontario. *Canadian Veterinary Journal*, 38: 713–718.
- El-Dee, W. M. (2012). Novel biomarkers for pregnancy toxaemia in ewes: acute phase proteins and pro-inflammatory cytokines. *Open Access Scientific Report*, 1(4): 242-243.
- Ermilio, E. M., & Smith, M. C. (2011) Treatment of emergency conditions in sheep and goats. Veterinary Clinical North America: Food Animal Practice, 27: 105-106.
- Ford, E. J. H., Evans, J., & Robinson, I. (2010). Cortisol in pregnancy toxaemia of sheep. *British Veterinary Journal*, 146: 539-542.
- Ganong, W. F. (2007). Review of Medical Physiology. United States: McGraw Hill.
- Gatenby, M. (2006). *Sheep production in the tropics and sub-tropics*. New York: Longman Inc.

- Gordan, E, D. (2012). Proceedings from 18th Annual Dairy Sheep Association of North America Symposium: *Ewe and Flock Health Overview*.
- Harwood, D. (2014). Diseases of dairy goats. In Practice, 26:248-259.
- Hefnawy, A. E., Shousha, S., & Youssef, S. (2011). Haematobiochemical profile of pregnant and experimentally pregnancy toxaemic goats. *Journal of Basic Applied Chemistry*, 1(8): 65-69.
- Henze, P., Bickhardt, K., Furhmann, H., & Sallmann, H. P. (2018). Spontaneous pregnancy toxaemia (ketosis) in sheep and the role of insulin. *Journal of Veterinary Sciences*, 45(5): 255-266.
- Henze, P., Bickhardt, K., & Furhmann, H. (2017). Human growth hormone and cortisol response to insulin. *Journal of Dairy Science*, 101: 61-65.
- Herdt, T. H. (2000). Ruminant adaptation to negative energy balance. *Veterinary Clinical North America: Food Animal Practice*, 16(2): 215-230.
- Houten, S. M., & Wanders, R. J. A. (2010). A general introduction to the biochemistry of mitochondrial fatty acid β-oxidation. *Journal of Inherited Metabolic Disease*, 33(5): 469–477.
- Jaeschke, H., Gores, G. J., & Cederbaum, A. I. (2012). Mechanisms of Hepatotoxicity. *Toxicological and Sciences*, 65(2): 166-176.
- Jeffrey, M., & Higgins, R. J. (2012). Brain lesions of naturally occurring pregnancy toxaemia of sheep. *Veterinary Pathology*, 29: 301-307.
- Jeremy, M. B., John, L. T., & Lubert, S. (2012). *Biochemistry*. New York: W.H. Freeman and Company.
- Kabakci, N., Yarim, G., Yarim, M., Duru, O., Yagci, B. B., & Kisa, U. (2003). Pathological, clinical and biochemical investigation of naturally occurring pregnancy toxemia of sheep. *Acta Veterinaria (Beograd)*, 53(3): 161–169.
- Kahn, C. (2005). *The Merck Veterinarian Manual*. New Jersey, USA: Merck & Company Incorporated.
- Kaneko, J. J., Harvey, J. W., & Bruss, M. L. (2008). Clinical biochemistry of domestic animals. 33-36. New York: Academic Press.
- Kara, M., & Doganay, A. (2005). Investigation of antigenic specificity against *Cysticercus tenuicollis* cyst fluid antigen in dogs experimentally infected with *Taenia hydatigena*. *Turkish Journal of Veterinary and Animal Sciences*, 29: 835–840.
- Kim, H. I., & Suh, G. K. (2013). Effect of the amount of body condition loss from the dry to near calving periods on the subsequent body condition change, occurrence of postpartum diseases, metabolic parameters and reproductive performance in Holstein dairy cows. *Theriogenology Journal*, 60: 1445-1456.

- King, J. C. (2010). Physiology of pregnancy and nutrient metabolism. *American Journal of Clinical Nutrition*, 71(5): 1218-1225.
- Koyuncu, M. (2013). Importance of body condition score in dairy goats. *Macedonian Journal of Animal Science*, 3: 167–173.
- Kristina, B. F., Orjan, A. L., & Bernt, V. J. (2010). Low cortisol levels in blood from dairy cows with ketosis: a field study. Acta Veterinaria Scandinavic, 52: 31.
- Lars, P., Bechmann, R. A., Hannivoort, G. G., Gokhan, S., Hotamisligil, M. T., & Ali, C. (2012). The interaction of hepatic lipid and glucose metabolism in liver diseases. *Journal of Hepatology*, 56(4): 952-964.
- Lehninger, A. L., Nelson, D. L., & Cox, M. M. (2005). Lehninger principles of biochemistry. New York: W.H. Freeman.
- Lima, L. S., Alcalde, C. R., Freitas, H. S., Molina, B. S. D., Macedo, F. D. F., & Horst, J. A. (2012). Performance of dairy goats fed diets with dry yeast from sugar cane as protein source. *Revista Brasileira de Zootecnia*, 41(1): 232-236.
- Lucas, E. S., Finn, S. L., Cox, A., Lock, F. R., & Watkins, A. J. (2009). The impact of maternal high fat nutrition on the next generation. *Journal of Physiology*, 587(14): 34–39.
- McArt, J. A. A., Nydam, D. V., Ospina, P. A., & Oetzel, G. R. (2011). A field trial on the effect of propylene glycol on milk yield and resolution of ketosis in fresh cows diagnosed with subclinical ketosis. *Journal of Dairy Science*, 94: 6011-6020.
- Menzies, P. I. (2011). Pregnancy Toxemia in Ewes: Hepatic Lipidosis. In Merck, *Veterinary Manual* (pp. 13-19). USA: Merial Press.
- Mirnawati, M., Gita, C., & Ferawati, B. H. (2019). Improving the quality and nutrient content of palm kernel cake through fermentation with *Bacillus subtilis*. *Livestock Research for Rural Development*, 31(7): 14-22.
- Mitchell, J. H., Wildenthal, K., & Johnson, R. L. (2011). The effects of acid-base disturbances on cardiovascular and pulmonary function. *Kidney International*, 1(5): 375-389.
- Morais, M. J., Sevillaa, C. C., Dizonb, J. T., Manulatc, G. L., Abesc, E. E. C., & Angeles, A. A. (2018). Growth Performance and Ruminal Metabolic Variables of Goats Fed Rain Tree (*Samanea saman*) Pods. *Tropical Animal Science Journal*, 41(1): 22-28.
- Motshakeri, M., Ebrahimi, M., Goh, Y. M., Othman, H. H., Hair-Bejo, M., & Mohamed, S. (2014). Effects of brown seaweed (*Sargassum polycystum*) extracts on kidney, liver, and pancreas of type 2 diabetic rat model. *Evidence Based Complementary and Alternative Medicine*, 1-11.

- Mursyidah, A. K., Khadijah, S., & Rita, N. (2017). Fasciola and Paramphistomum infections in small ruminants (sheep and goat) in Terengganu. Malaysian Journal of Veterinary Research, 8(2): 8-12.
- National Research Council (NRC). (2007). Nutrient Requirement of Small Ruminants: Sheep, Goats, Corvids and New World Camelids. Washington, D.C: National Academic Press.
- Nguyen, P., Leray, V., Diez, M., Serisier, S., Le Bloc'h., J., & Siliart, B. (2008). Liver lipid metabolism. *Journal of Animal Physiology and Animal Nutrition*, 92: 272–283.
- Oetzel, G. R. (2014). Monitoring and testing dairy herds for metabolic disease. *Veterinary Clinical North American Food Animal Practice*, 20: 651–674.
- Ondieki, G., & Renita, M. (2012). Nutritional strategies to mitigate the effects of negative energy balance on reproductive performance of early postpartum. *American Journal of Animal and Veterinary Sciences*, 7(4): 194-197.
- Ospina, P. A., Nydam, D. V., Stokol, T., & Overton, T. R. (2010). Associations of elevated non-esterified fatty acids and β -hydroxybutyrate concentrations with early lactation reproductive performance and milk production in transition dairy cattle in the north eastern United States. *Journal of Dairy Science*, 93: 1596–1603.
- Pethick, D. W., Harper, G. S., & Dunshea, F. R. (2005). Fat metabolism and turnover. In J. Dijkstra (Ed.), *Quantitative Aspects of Ruminants Digestion and Metabolism* (pp. 345-371). Cambridge, USA: CABI Publishing.
- Pichler, M., Damberger, A., Amholdt, T., Schwendenwein, I., & Gasteiner, J. (2014). Evaluation of two electronic handheld devices for diagnosis of ketonemia and glycemia in dairy goats. *Journal of Dairy Science*, 97: 7538-7546.
- Pough, D. G. (2002). Diseases of the gastrointestinal system. In J.M Saunders (1st Ed.), Sheep and Goat Medicine (pp. 69-105). Philadelphia, Pennsylvania: An imprint of Elsivier.
- Prasannkumar, R. V., Jani, R. G., Goswami, H. V., Rathwa, S. D., & Tandel, F. B. (2016). Studies on clinical signs and biochemical alteration in pregnancy toxemic goats. *Veterinary World*, 9(8): 869-874.
- Radostits, M., Gay, C., Blood, C., & Kenneth, W. (2006). Veterinary Medicine. A *Textbook of The Disease of Cattle, Horses, Sheep, Pigs and Goats* (pp. 1668-1671). London: Baillire Tindall publisher.
- Rajala-Schultz, P. J., Gröhn, Y. T., & McCulloch, C. E. (2009). Effects of milk fever, ketosis, and lameness on milk yield in dairy cows. *Journal of Dairy Science*, 82(2): 288–294.
- Rahman, A. R., (2015). Proceedings from 32nd MSAP Annual Conference: *Issues and challenges in commercializing new livestock technologies*.

- Ramin, A. G., Asri, S., & Majdani, R. (2005). Correlations among serum glucose, beta hydroxybutyrate and urea concentrations in non-pregnant ewes. *Small Ruminant Research*, 57(3): 265-269.
- Roach, J. O., Bemyon, S., Horton-Szar, D., & Dominiczak, N. (2014). Lipid Metabolism and Transport. *Metabolism and Nutrition* (pp. 55-82). Philadelphi: Elsevier Ltd.
- Rook, J. S. (2010). Pregnancy toxemia of ewes, does, and beef cows. *Veterinary Clinical North America Food Animal Practice*, 16: 293–317.
- Sadjadian, R., Seifi, H. A., Mohri, M., Naseian, A. A., & Farzaneh, N. (2013). Variations of energy biochemical metabolites in periparturient dairy Saanen goats. *Comparative Clinical Pathology*, 22: 449–456.
- Sakha, M., Ameri, M., Sharifi, H., & Taheri, I. (2007). Bovine subclinical ketosis in dairy herds in Iran. *Veterinary Research Communications*, 31: 673-679.
- Sargison, N. D., Scott, P. R., Penny, C. D., Pirie, R. S., & Kelly, J. M. (2007). Plasma enzymes and metabolites as potential prognostic indices of ovine pregnancy toxaemia a preliminary study. *British Veterinary Journal*, 150: 271.
- Sato, S., Kohno, M., & Ono, H. (2015). Relation between blood β-hydroxybutyric acid and glucose, non-esterified fatty acid and aspartate aminotransferase in dairy cows with subclinical ketosis. *Japanese Journal of Veterinary Clinics*, 28: 7-13.
- Schlumbohm, C., Sporleder, H. P., Gortler, H., & Harmeyer, J. (2007). Insulin and glucogon secretion in goats. *Veterinary Clinical Pathology*, 104: 359-365.
- Schlumbohm, C., & Harmeyer, J. (2013). Hypocalcaemia reduces endogenous glucose production in hyperketonemic sheep. *Journal of Dairy Science*, 68: 1953–1962.
- Schlumbohm, C., & Harmeyer, J. (2014). Hyperketonemia impairs glucose metabolism in pregnant ewes. *Journal of Dairy Science*, 87: 350-358.
- Silk, L. (2013). Metabolic diseases in sheep: developments and treatment. *Veterinary Times*, 43: 16-19.
- Singh, J., Bal, M., Aradhana, S., & Gumber, S. (2004). Efficacy of different flukicides against fascioliasis in sheep and goats. *Journal of Research Punjab Agricultural University*, 41(2): 287-289.
- Sithambaram, S., & Hassan, Q. N. (2014). *Country report Malaysia*. Kuala Lumpur: Asian Australasian Dairy Goat Network.
- Smith, B. P. (2005). Large Animal Internal Medicine. *The C.V. Mosby Company*, 16(2): 14-19.

- Smith, M. C., & Sheramn, D. M. (2009). *Goat Medicine*. New Jersey: Wiley Blackwell.
- Sordillo, L. M., & Raphael, W. (2013). Significance of metabolic stress, lipid mobilization and inflammation on transition cow disorders. *Veterinary Clinics: Food Animal Practice*, 29: 267–278.
- Stockhaus, C., Van Den Ingh, T., & Rothuizen, J. (2004). A multistep approach in the cytologic evaluation of liver biopsy samples of dogs with hepatic diseases. *Veterinary Pathology*, 41: 461-470.
- Suriyasathaporn, W., Heuer, C. E. N., Noordhuizen-Stassen, M., & Schukken, Y. H. (2010). Hyperketonemia and udder defense: a review. *Veterinary Research*, 31: 397–412.
- Tadesse, D. T. (2012). Management practices, productive performances and farm management in Shewa, Ethiopia. Agricultural Research Outputs, 19(2): 21-26.
- Van Saun, R. J. (2010). Pregnancy toxemia in a flock of sheep. Journal of the American Veterinary Medical Association, 217: 1536-1539.
- Vernon, R. G. (2005). Metabolic Regulation. In E.J. Dijkstra (Ed.), Quantitative Aspects of Ruminant Digestion and Metabolism (pp. 443-468). Cambridge, US: CABI Publishing.
- Zambom, M. A., Alcalde, C. R., & Silva, K. T. (2005). Ingestion, diets digestibility and milk yield in Saanen goats submitted to different roughage ratios. *Brazilian Journal of Animal Science*, 34(6): 2505-2514.