

UNIVERSITI PUTRA MALAYSIA

IDENTIFICATION AND CHARACTERIZATION OF DROUGHT-INDUCED GENES IN MALAYSIA RICE CULTIVARS, MR220 AND MR211 USING RNA-SEQUENCING PLATFORM

TAJUL ARIFFIEN BIN OTHMAN

FBSB 2019 33



IDENTIFICATION AND CHARACTERIZATION OF DROUGHT-INDUCED GENES IN MALAYSIA RICE CULTIVARS, MR220 AND MR211 USING RNA-SEQUENCING PLATFORM

By

TAJUL ARIFFIEN BIN OTHMAN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

June 2019

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia

 \mathbf{O}



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

IDENTIFICATION AND CHARACTERIZATION OF DROUGHT-INDUCED GENES IN MALAYSIA RICE CULTIVARS, MR220 AND MR211 USING RNA-SEQUENCING PLATFORM

By

TAJUL ARIFFIEN BIN OTHMAN June 2019 Chair : Assoc. Prof. Noor Azmi Shaharuddin, PhD Faculty : Biotechnology and Biomolecular Sciences

Drought is one of the abiotic stresses on plants, causing significant detrimental impacts, especially to lowland rice (Oryza sativa L.) ecosystems. In order to obtain new insights on osmotic stress in rice, a comparative study using a Next-Generation Sequencing platform was conducted to elucidate osmotic-responsive genes from two local Malaysian rice cultivars, namely the commercially available drought-tolerant MR220 and the drought-sensitive MR211. In the study, 21-day-old seedlings of MR220 and MR211 were exposed to 6% PEG 6000 for 24 hours, which produced osmotic stress that mimicked the drought condition. The samples were collected and total RNA were extracted. Two transcriptomic libraries were constructed from both rice cultivars using the Illumina HiSeq 2000 platform. A total of 77,964,138 and 92,699,454 raw sequence reads were generated from these libraries. From the expressed genes from both libraries, around 44, 902 genes have been found overlapping each other. Then, 8,095 and 2,081 gen have been found uniquely in MR220 and MR211, respectively. Based on the gene annotation of O. sativa, a total of 106 genes were identified as differentially and significantly expressed in drought-tolerant and drought-susceptible cultivars, and a total of 29 genes were categorized as unknown genes. From the 106 differentially expressed genes (DEGs), 14 genes were up-regulated, while another 92 were downregulated in MR220. Gene Ontology (GO) and KEGG analysis were conducted to obtain the functional and biological role of the differentially expressed genes. Six drought related genes were selected for validating the RNA-Seq analysis using semiquantitative PCR. The validation result showed four out of six DEGs followed the RNA-Seq analysis. Then, two genes were selected form six semi-quantitative analysis, HP and Thia genes were validated using real-time qRT-PCR. The result showed same expression with semi-quantitative PCR. Overall, this study gives further insight on rice defense mechanisms during osmotic stress at early stage.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

MENGENAL PASTI DAN MENGKAJI CIRI-CIRI GEN YANG TERLIBAT DENGAN KEMARAU PADA PADI MALAYSIA, MR220 AND MR211 DENGAN MENGUNAKAN PLATFORM PENJUJUKAN RNA

Oleh

TAJUL ARIFFIEN OTHMAN

Jun 2019

Pengerusi : Prof. Madya Noor Azmi Shaharuddin, PhD Fakulti : Bioteknologi dan Sains Biomolekul

Kemarau merupakan salah satu juzuk abiotic terhadap tumbuh-tumbuhan, yang memberi kesan yang sangat buruk terutamanya ekosistem padi (Oryza sativa L.) tanah rendah. Untuk mendapat gambaran sebenar kesan kemarau terhadap padi, kajian perbandingan mengunakan platform Penjujukan Generasi Seterusnya telah dijalankan dalam mencari gen yang terlibat kepada kemarau, daripada dua jenis padi tempatan, padi komersal tahan kemarau, MR220 dan padi sensitif terhadap kemarau, MR211. Dalam kajian ini, anak padi MR220 dan MR211 yang berusia 21 hari dikenakan 6% daripada PEG 6000 selama 24 jam, menghasilkan tekanan osmotik yang meniru keadaan kemarau. Sampel padi dikumpul dan total RNA akan diekstrak dari sampel pokok. Dua perpustakaan transkriptomik telah dibina dari kedua-dua kultivar padi dengan mengunakan HiSeq Platform 2000. Sebanyak 77, 964,138 dan 92,699,454 'sequence read' telah dijana dari data tersebut. Daripada gen yang terzahir daripada kedua-dua perpustakaan, sebanyak 44,902 gen telah dijumpai terzahir. Sementara itu, sebanyak 8,095 dan 2,081 gen yang unik dijumpai dalam MR220 dan MR211. Berdasarkan anotasi gen O. Sativa, sebanyak 106 gen telah dikenal pasti sebagai yang sangat ketara dalam kultivar tahan kemarau dan tidak tahan kemarau, dan sejumlah 29 gen adalah gen baru yang tidak dikenal pasti. Daripada 106 gen yang dinyatakan sebagai (DEG), 14 gen telah menunjukkan peningkatan dan 92 gen menunjukkan penurunan dalam MR220. Gen Ontologi (GO) dan analisis KEGG telah dijalankan untuk memperolehi fungsi dan proses biologi untuk setiap gen tersebut. Enam gen telah dipilih untuk mengesahkan analisis RNA-Seq dengan mengunakan RT-PCR separakuantitatif. Keputusan pengesahan menunjukkan daripada empat gen menyerupai analisis RNA-Seq tersebut. Kemudian, dua gen dipilih, HP dan Thia gen dalam mengesahkan keputusan RNA-Seq, dengan mengunakan qRT-PCR kuantitatif. Hasil kajian ini menunjukkan penzahiran yang sama dengan PCR separa kuantitatif. Secara keseluruhan, kajian ini memberikan satu pandangan berkenaan dengan mekanisma pertahanan pokok padi semasa tekanan osmosis pada peringkat awal.

ACKNOWLEDGEMENTS

In the of Allah, Most Gracious, Most Merciful, I have completed this thesis

I would like to express my appreciation to Universiti Putra Malaysia (UPM) for giving me a place to pursue my master study in Plant Biotechnology Program and finally accomplish my study. I also would like to thank to the Dean of Faculty of Biotechnology and Biomolecular Science, Prof. Dr. Arbakariya Ariff, for his support and help towards postgraduate student affairs.

Special appreciation goes to my supervisor, Assoc. Prof. Dr. Noor Azmi Shaharuddin, for his supervision and constant support, fantastic idea and guidance to increase the value of my project as well as taught me the great value of how to be a good scientist. In addition, heartfelt appreciation conveys to all my co-supervisors, Dr. Norliza Binti Abu Bakar, Puan Rabiatual Adawiyah Binti Zainal Abidin, Allahyarhamah Prof. Dr. Maziah Mahmood and Dr. Nor Baity Binti Saidi who have contributed to the success of this research.

Also thanks goes to Assoc. Prof. Dr. Awang Yahya as his guidance and contribution on field experiments. Special thanks for laboratory assistants of Faculty of Biotechnology and Biomolecular Sciences for giving me help, support, and guidance during my lab work.

Sincere thanks also to all my friends and lab mates for their kindness and moral support during my study. Finally yet importantly, my deepest gratitude goes to my beloved parents, (Othman bin Mamat and Alimah binti Amirusin), my siblings and my wife, Sharifah Nur Nusrah binti Syed Yassin for their endless love, prayers and encouragement. To those who indirectly contributed in this research, your kindness means a lot to me.

Thank you very much.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Noor Azmi bin Shaharuddin, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

Nor Baity binti Saidi, PhD

Senior Lecturer Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

Maziah binti Mahmood, PhD

Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 17 October 2019

Declaration by Graduate Student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustrations, and citations have been referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from thesis and copyright of thesis are fully owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books journals, modules, proceedings, popular writings, seminar papers, manuscript, posters, reports, lecture notes, learning modules, or any materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/ fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiasrism detection software.

Signature:	Date:
Name and Matric No.:	

Declaration by Members of Supervisory Committee

This is to confirm that:

G

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory	
Committee:	
U	
Signature:	
Name of Member of Supervisory	
Committee:	
Signature:	
Name of Membe <mark>r of</mark>	
Supervisory	
Committee:	

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	ii
ACKNOWLEDGEMENTS	iii
APPROVAL	iv
DECLARATION	vi
LIST OF TABLES	xi
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xvi

CHAPTER

1	INT	RODUC"	ΓΙΟΝ	1
	1.1	Gene	eral Introduction	1
	1.2	Rese	arch Objectives	2
2	LIT		RE REVIEW	3
	2.1	Ory <mark>za s</mark>	ativa L.	3
		2.1.1	Origin and Taxonomy	3
		2.1.2	1	3
			Rice as Monocot Model Plant	4
	2.2	Rice Pr	oduction in Malaysia	6
		2.2.1	Malaysia Rice Varieties	9
		2	.2.1.1 MR220	9
		2	.2.1.2 MR211	9
	2.3	Stress		9
		2.3.1	Abiotic Stress	10
		2.3.2	Drought Stress and Adaption in	10
			Plant	
	2.4	Biotech	nology Approach for Crop	13
		Improv		
	2.5	Next-G	eneration Sequencing (NGS)	14
		Techno	logy	
		2.5.1	RNA-Sequencing (RNA-Seq)	15
		2.5.2	RNA-Sequencing Platform	16
		2.5.3	RNA-Seq Application in Plants	17
3	MΔ	FFRIAT	S AND METHODS /	19
5		THODO		17
	3.1		reatment and Total RNA Preparation	19
	5.1	3.1.1	Source of <i>Oryza sativa</i>	19
		3.1.2		19
		3.1.2	Total RNA Extraction of Plant	21
		5.1.5	Samples	21
		3.1.4	DNAse Treatment in Total RNA	22

		Samples	
	3.1.5	RNA Gel Electrophoresis	22
	3.1.6	Quantification of Total RNA	22
	3.1.7	RNA Integrity Number (RIN)	23
3.2		uction of RNA-Sequencing Libraries	23
0.1	3.2.1	mRNA Construction and Illumina	23
	3.2.1	sequencing	25
	3.2.2	Pre-Processing Sequence Reads	23
	3.2.3	Mapping the Sequence Reads	23
	5.2.5	Against Reference Genome	23
	224	Tuxedo Protocol	24
	3.2.4		
	3.2.5	MapMan	24
	3.2.6	Gene Ontology (GO) and Kyoto	24
		Encyclopedia Gene and Genome	
		(KEGG)	
3.3		tion of DEGs from RNA-Seq Analysis	26
	-	RT-PCR	
	3.3.1	Primer Design	26
	3.3.2	Plant Samples Preparation	26
	3.3.3	Total RNA Extraction and cDNA	27
		Synthesis	
	3.3.4	PCR Cycle and Annealing	27
		Temperature Optimization	
	3.3.5	DNA Purification	29
	3.3.6	DNA-Sequencing	29
	3.3.7		29
	3.3.8		29
	3.3.9	Expression Profiles of Genes	30
3.4		tion of DEGs from RNA-Seq Analysis	30
		RT-PCR	
	3.4.1	Primer Design	30
	3.4.2	Total RNA Extraction and cDNA	30
	3.4.2	Synthesis	50
	3.4.3	Gene Expression Analysis of Thia	30
	5.4.5	and HP genes using qRT-PCR	50
		and HP genes using qR1-PCR	
DEG		ND DISCUSSION	22
		AND DISCUSSION Freatment and Osmotic Stress	32
4.1			32
4.0	• •	om Evaluation	24
4.2	Total I		34
	4.2.1	Total RNA Extraction	34
	4.2.2	RIN and Quantification of total	34
		RNA	
4.3	RNA-S	Sequencing	36
	4.3.1	Construction of Transcriptome	36
		Libraries	
	4.3.2	Data Analysis and Expression Study	40
	4.3.3	Gene Ontology (GO) Classification	47
	4.3.4	Pathway Enrichment Analysis	51
4.4		tion of RNA-Sequencing Analysis	52
	4.4.1	Purity and Quantification of total	52

4

(1

 \bigcirc

 4.4.2 Optimization of PCR Cycles and Annealing Temperature 4.4.3 Identity Confirmation of PCR 4.4.3 Identity Confirmation of PCR 5 CONCLUSION AND RECOMMENDATIONS 4.4.2 Optimization of PCR (PCR) 4.4.5 Optimization of PCR (PCR) 4.4.5 Optimization of PCR (PCR) 4.4.5.1 Standard Curve and PCR 4.4.5.2 Melt Curve Analysis 60 4.4.5.3 Gene Expression Analysis of Thia and HP in Drought Condition using qRT-PCR 			RNA i	in Validating RNA-Seq	
 4.4.3 Identity Confirmation of PCR Products via BLAST 4.4.4 Semi-Quantitative Reverse 56 Transcription-PCR (RT-PCR) 4.4.5 Quantitative Reverse transcription-PCR (qRT-PCR) 4.4.5.1 Standard Curve and PCR 9 efficiency of Designed Primers 4.4.5.2 Melt Curve Analysis 60 4.4.5.3 Gene Expression Analysis of 60 Thia and HP in Drought Condition using qRT-PCR 5 CONCLUSION AND RECOMMENDATIONS 64 		4.4.2			54
Products via BLAST4.4.4Semi-Quantitative Reverse56Transcription-PCR (RT-PCR)594.4.5Quantitative Reverse transcription- PCR (qRT-PCR)594.4.5.1Standard Curve and PCR efficiency of Designed Primers594.4.5.2Melt Curve Analysis604.4.5.3Gene Expression Analysis of Condition using qRT-PCR605CONCLUSION AND RECOMMENDATIONS64			Annea	aling Temperature	
 4.4.4 Semi-Quantitative Reverse Transcription-PCR (RT-PCR) 4.4.5 Quantitative Reverse transcription-PCR (qRT-PCR) 4.4.5.1 Standard Curve and PCR 59 efficiency of Designed Primers 4.4.5.2 Melt Curve Analysis 60 4.4.5.3 Gene Expression Analysis of 60 Thia and HP in Drought Condition using qRT-PCR 5 CONCLUSION AND RECOMMENDATIONS 64 		4.4.3			56
Transcription-PCR (RT-PCR)4.4.5Quantitative Reverse transcription- PCR (qRT-PCR)59 PCR (qRT-PCR)4.4.5.1Standard Curve and PCR efficiency of Designed Primers59 efficiency of Designed Primers4.4.5.2Melt Curve Analysis Gene Expression Analysis of Thia and HP in Drought Condition using qRT-PCR605CONCLUSION AND RECOMMENDATIONS64			Produ	cts via BLAST	
4.4.5 Quantitative Reverse transcription- PCR (qRT-PCR) 59 4.4.5.1 Standard Curve and PCR efficiency of Designed Primers 59 4.4.5.2 Melt Curve Analysis 60 4.4.5.3 Gene Expression Analysis of Condition using qRT-PCR 60 5 CONCLUSION AND RECOMMENDATIONS 64		4.4.4	Semi-	Quantitative Reverse	56
PCR (qRT-PCR) 4.4.5.1 Standard Curve and PCR 59 efficiency of Designed Primers 4.4.5.2 Melt Curve Analysis 60 4.4.5.3 Gene Expression Analysis of 60 Thia and HP in Drought Condition using qRT-PCR 5 CONCLUSION AND RECOMMENDATIONS 64					
 4.4.5.1 Standard Curve and PCR 59 efficiency of Designed Primers 4.4.5.2 Melt Curve Analysis 60 4.4.5.3 Gene Expression Analysis of 60 Thia and HP in Drought Condition using qRT-PCR 5 CONCLUSION AND RECOMMENDATIONS 64 		4.4.5			59
 efficiency of Designed Primers 4.4.5.2 Melt Curve Analysis 4.4.5.3 Gene Expression Analysis of 4.4.5.3 Gene Expression Analysis of <i>Thia</i> and <i>HP</i> in Drought Condition using qRT-PCR 5 CONCLUSION AND RECOMMENDATIONS 64 					
Primers4.4.5.2Melt Curve Analysis604.4.5.3Gene Expression Analysis of60 <i>Thia</i> and <i>HP</i> in Drought Condition using qRT-PCR64			4.4.5.1		59
4.4.5.2Melt Curve Analysis604.4.5.3Gene Expression Analysis of Thia and HP in Drought Condition using qRT-PCR605CONCLUSION AND RECOMMENDATIONS64					
4.4.5.3Gene Expression Analysis of Thia and HP in Drought Condition using qRT-PCR605CONCLUSION AND RECOMMENDATIONS64					
Thia and HP in Drought Condition using qRT-PCR5CONCLUSION AND RECOMMENDATIONS64					
Condition using qRT-PCR5CONCLUSION AND RECOMMENDATIONS64			4.4.5.3	· ·	60
5 CONCLUSION AND RECOMMENDATIONS 64					
				Condition using qRT-PCR	
	-	CONCLUS			<i></i>
FOR FUTURE RESEARCH	5				64
		FORFUTU	RE RES	SEARCH	
DEPEDENCES	DEFEDEN	CEC			
REFERENCES66APPENDICES83					
BIODATA OF STUDENT 112			т		
LIST OF PUBLICATIONS 113					
	LISTOFT	UDLICATIO			115

 (\mathbf{C})

LIST OF TABLES

Table		Page
2.1	Area harvested, yield and rice production from 2015 to 2017 Asian and World	4
2.2	Area harvested, yield and rice production from 2010 to 2017 in Malaysia	6
3.1	Primer sequences were used in RNA-Seq validation using RT-PCR	28
3.2	General view of plant treatment for the validation of selected RNA-Sequencing data	29
3.3	The components used for cDNA synthesis as to make 20 uL of cDNA templates	29
3.4	List of PCR components used in RT-PCR	30
3.5	Optimized PCR protocol which used in semi quantitative PCR	30
3.6	Primer sequences were used in RNA-Seq validation using qRT-PCR	32
3.7	List of PCR components used in qPCR	33
3.8	The qPCR program which were used for quantifying the genes expressions	33
4.1	The total read before and after trimming in Malaysia rice cultivars, MR220 and MR211 under osmotic stress	38
4.2	The DEGs identified in Malaysia rice cultivars, MR220 vs MR211	43
4.3	The unknown DEGs annotated from Rice Annotation Project Database (RAP-DB), blast using NCBI	44
4.4	The DEGs selected from the RNA-Seq data	55
4.5	The purity and concentration of total RNA from 21-old day of the rice cultivars, MR220 and MR211	57
4.6	Primer sequences, R ² and PCR amplification efficiency of genes used in the qRT-PCR analysis	63
5.1	List of DEGs from the RNA-Seq analysis of Malaysia	91

rice cultivars

6

5.2	Band intensity value in optimization of cycles of Ubq5	96
5.3	Band intensity value in optimization of cycles of HP	97
5.4	Band intensity value in optimization of cycle of Thia	97
5.5	Band intensity value in optimization of cycle of AAi	97
5.6	Band intensity value in optimization of cycle of P450	98
5.7	Band intensity value in optimization of cycle of <i>Cys</i>	98
5.8	Band intensity value in optimization of cycle of <i>Def</i>	98
5.9	Band intensity value in optimization annealing temperature of <i>Ubq5</i>	100
5.10	Band intensity value in optimization annealing temperature of <i>HP</i>	101
5.11	Band intensity value in optimization annealing temperature of <i>Thia</i>	101
5.12	Band intensity value in optimization annealing temperature of <i>AAi</i>	101
5.13	Band intensity value in optimization annealing temperature of <i>P450</i>	102
5.14	Band intensity value in optimization annealing temperature of <i>Cys</i>	102
5.15	Band intensity value in optimization annealing temperature of <i>Def</i>	102
5.16	The relative density of drought induced genes form RNA-Sequencing analysis between S"MR220" and S"MR211"	108
5.17	The relative density of drought induced genes form RNA-Sequencing analysis between C"MR220" and S"MR220"	109
5.18	The relative density of drought induced genes form RNA-Sequencing analysis between C"MR211" and S"MR211"	110

LIST OF FIGURES

Figure		Page
2.1	Trend of import and local rice which supplied to local market in Malaysia, from 2000 to 2016	7
2.2	Map of paddy growing area in Peninsular Malaysia	8
2.3	The template generation <i>via</i> bridge amplification	17
3.1	Rice plant which planted in glass house in Universiti Putra Malaysia	21
3.2	The rice seeds were placed and submerged into distilled water under a box for three days	21
3.3	The rice plant were planted using hydroponic system (Yoshida nutrient solution) in controlled parameters	22
3.4	The 21-day old rice plant were treated with PEG 6000 under control parameters and harvested after 24 hours treatment	22
3.5	The RNA-sequencing data workflow	27
4.1	The example of 21 days of seedling stage of Malaysia rice cultivar before being introduced to osmotic stress	35
4.2	Different response to drought condition between MR220 and MR211	35
4.3	Total RNA of MR220 and MR211 after DNAse treatment	36
4.4	The results of total RNA for MR220 and MR211	37
4.5	Comparison of genes between the two transcriptome libraries of different drought perception cultivars, MR220 and MR211	39
4.6	Snapshot of known genes (genes before proceed CuffDif) which generated using MapMan in main metabolic pathways and gene regulation	41
4.7	The distribution of DEGs genes annotation from IRGSP database were classified	43
4.8	The amount of fold change for each DEGs (FC>1)	47
4.9	Cluster of significant differential expression genes in	48

MR211 and MR220 presented in volcano

4.10	Cluster of significant differential expression genes in MR211 and MR220 based on log ratio FPKM data	49
4.11	Functional annotation of differentially expressed genes (DEGs) between MR220 and MR211 within the Gene Ontology (GO) (level2)	52
4.12	Gel documentation of total RNA of rice treatment for replicate 1, replicate 2, and replicate 3	56
4.13	The validation analysis of RNA-Sequencing for genes <i>AAi, HP, P450,</i> and <i>Thia</i> using semi-quantitative PCR	60
4.14	The validation analysis of RNA-Sequencing for genes <i>Cys</i> and <i>Def</i> using semi-quantitative PCR	60
4.15	Relative intensity (RI) value of each DEGs which normalized With <i>Ubq5</i>	61
4.16	The expression profile of HP and <i>Thia</i> in MR220 and MR211	64
4.17	The translated protein of <i>HP</i> genes	66
4.18	The features suggested form the gene which generated using SMART Domain	66
5.1 5.2	QC result of MR220 RNA integrity for RNA-Seq library Graph generated from total RNA of MR220	86 87
5.3 5.4	QC result of MR211 RNA integrity for RNA-Seq library Graph generated from total RNA of MR211	88 89
5.5	DNA and RNA ladders used for mass estimation size of DNA dan RNA	90
5.6	The comparison of gene ontology between MR220 and MR211	94
5.7	The genes were successfully identified in MR220 rice samples which were <i>Ubq5</i> , <i>AAi</i> , <i>HP</i> , <i>Cys</i> , and <i>P450</i>	95
5.8	The genes were successfully identified in MR220 rice samples which were <i>Thia</i> and <i>Def</i>	95
5.9	Six genes have been suscesfully optimized for PCR cycles for <i>Ubq5</i> , <i>AAi</i> , <i>HP</i> , <i>Thia</i> , <i>P450</i> , <i>Cys</i> and <i>Def</i> gene	96
5.10	Relationship of PCR cycles with relative intensity of	99

6

	bands were simplified into graph for Ubq5, AAi, Thia, Def, HP, Cys, and P450 genes	
5.11	Six genes have been successfully optimized for annealing temperature for <i>Ubq5</i> , <i>AAi</i> , <i>HP</i> , <i>Thia</i> , <i>P450</i> , <i>Cys</i> and <i>Def</i> genes	100
5.12	Relationship of annealing temperature with relative intensity of bands were simplified into graph for <i>Ubq5</i> , <i>AAi</i> , <i>Thia</i> , <i>Def</i> , <i>HP</i> , <i>Cys</i> , and <i>P450</i> genes	103
5.13	Blast hit result of housekeeping gene, Ubq5	104
5.14	Blast hit result of Thiamine biosynthesis gene, Thia	104
5.15	Blast hit result of hypothethical protein, <i>HP</i>	105
5.16	Blast hit result of cytochorome, P450	105
5.17	Blast hit result of alpha amylase inhibitor, <i>AAi</i>	106
5.18	Blast hit result of drought induced gene, Cys	106
5.19	Blast hit of defense hypothetical gene, <i>Def</i>	107
5.20	Standard curve of housekeeping gene, HNR	111
5.21	qPCR melting curve of HNR gene	111
5.22	Standard curve of housekeeping gene, EP	112
5.23	qPCR melting curve of EP gene	112
5.24	Standard curve of target gene, Thia	113
5.25	qPCR melting curve of Thia gene	113
5.26	Standard curve of target gene, HP	114
5.27	qPCR melting curve of HP gene	114
G		

xv

LIST OF ABBREVIATIONS

bp	BasePair
cDNA	Complementary Deoxyribonucleotide acid
°C	Celsius
DEGs	Differential Expressed Genes
DNA	Deoxyribonucleic Acid
ds	Double Stranded
FPKM	Fragment Per Kilobase of transcript per Million mapped reads
GO	Gene Ontology
KEGG	Kyoto Encyclopedia of Genes and Genomes
MARDI	Malaysia Agricultural Research and Development Institute
min	Minute
μg	Microgram
mM	MiliMolar
μL	Microlitre
NGS	Next Generation Sequencing
NCBI	National Centre for Biotechnology Information
nt	nucleotide
PCR	Polymerase chain reaction
PEG	Polyethylene glycol
qPCR	Quantitative polymerse chain reaction
RIN	RNA integrity number
RNA	Ribonuclease acid
RNA-Seq	RNA Sequencing

C,

- Rpm revolutions per minute
- RT Reverse-transcription
- RT-PCR Reverse-transcription polymerase chain reaction
- ROS Reactive Oxygen Species
- RPKM Read Per Kilobase of transcript per Million mapped reads
- s Second
- Ss Single Stranded
- UPM Universiti Putra Malaysia
- UK United Kingdom
- US United State Of America

CHAPTER 1

INTRODUCTION

1.1 General Introduction

Rice, (*Oryza sativa L.*) is one of the most important crops in the world as over half of the world's populations consume rice as their staple food (Hadiarto and Tran, 2011). In 2017, over 167.25 million hectares were planted with rice all over the world with 87.02% of the rice harvested from Asia. Total production yield from all over the world is around 769.65 million tonnes, with 89.98% of the rice production came from Asia (FAO, 2018). In Malaysia, rice is grown on 689,268 hectares of land which produced 2.9 million tons of paddy grain annually (FAO, 2018). In the midst of meeting with the increasing world demand, rice production is still facing limitations, mostly due to abiotic and biotic stresses, as well as drought stress and the latter is a major issue as reported in numerous scientific reports (Najmuddin, *et al.*, 2018; Todaka *et al.*, 2015; Mostajeran and Rahimi-Eichi, 2009).

Drought is one of the environmental factors that affect the rice production. It is severely influencing many regions in the world especially in lowland rice ecosystems (Zhang *et al.*, 2018; Shukla *et al.*, 2012; Passioura, 2007). Statistical analysis showed that the land affected by drought have increased more than twice in percentage from 1970s to early 2000 (Isendahl and Schmidt, 2006). The situation becomes more serious due to global climate change in agricultural areas and increasing world population (Lesk *et al.*, 2016; Hongbo *et al.*, 2005).

Rice is a crop that is susceptible to drought due to its shallow root system, rapid stomatal closure and leaf senescence in mild water stress (Obidiegwu *et al.*, 2015; Hirasawa, 1999). The drought stress in different stage of rice growth gives different effects. At vegetative stage, the height of rice plant is significantly reduced and lower grain yield is produced (Sarvestani *et al.*, 2008). Therefore, it is highly important to study the molecular mechanism of drought tolerance for future production of drought tolerant rice.

Rice genome has been fully sequenced and generated and 2859 unique genes were discovered and need to be characterized (Sasaki, 2005). Due to the lack of information about their functional characterization, validation needs to be carried out. RNA-Seq is a high-throughput sequencing-based method, which has become an important technique used in transcriptome study. Much information can be obtained and studied from the RNA-Seq data. This includes identification of transcription sites and new splicing variants, monitoring of allele expression, cataloguing species transcripts, quantifying expression level of each transcript in different conditions and quantification of exon expression and splicing site (Nagalakshmi *et al.*, 2010; Tsuchihara *et al.*, 2009; Wang

et al., 2009). As the RNA-Seq is one of the most used methods in most transcriptome study, this technology can be used to identify drought related tolerant genes.

MR220 has been identified as Malaysian drought tolerant commercial rice cultivar (Zulkarnain *et al.*, 2013) while MR211 has been identified as susceptible cultivar to drought stress (Abdul Rahim *et al.*, 2012). Moreover, MR220 gives high yield (Zain *et al.*, 2014) and has been planted widely in Malaysia for the past few years. Hence, a study was designed to characterize and profile the expression of drought-induced genes in both these Malaysia cultivars, MR220 and MR211 *via* high-throughput sequencing technology. These results can reveal pathways, alternative splicing sites for genes and help to identify novel genes that could play major roles during plant tolerance to drought stress.

1.2 Research Objectives

Therefore, the objectives of this study were:

- 1. To induce drought treatment to Malaysian rice cultivars, MR220 and MR211 using PEG 6000 and generate transcriptome libraries of drought-induced MR220 and MR211 *via* RNA-Seq technology
- 2. To identify Differential Expressed Genes (DEGs) from the transcriptome libraries
- 3. To validate the expression of the DEGs using reverse transcription-polymerase Chain Reaction (RT-PCR) and real-time quantitative reverse-transcription Polymerase Chain Reaction (qRT-PCR)

REFERENCES

- Abdul-Rahim, H., Shafika, Z.K., Bhuiyan, M.A.R., Narimah, M.K., Wickneswari, R., Abdullah, M.Z., Anna, L.P.K., Sobri, H., Rusli. I. and Khairuddin, A.R. (2012). Characterization of advanced rice mutant line of rice (Oryza Sativa), MR219-4 and MR219-9 under drought condition. *International Atomic Energy Agency Evaluation* 44(43): 033.
- Abdul-Rahim, F.H., Hawari, N.N. and Abidin, N.Z. (2017). Supply & Demand of Rice in Malaysia: A System Dynamics A pproach. *International Journal of Supply Chain Management* 6: 4.
- Abdullah, A.A. (2009). Genetic studies on leaf rolling and some root traits under drought conditions in rice (Oryza sativa L.). *African Journal of Biotechnology* 8(22): 6241-6248.
- Abdullah, S.N.A., Panjaitan, S.B., Aziz, M.A., Sariah, M. and Omar, O. (2009). Somatic embryogenesis from Scutellar embryo of oryza sativa L. var. MR219. *Pertanika Journal of Tropical Agriculture Science* 32(2): 185-194.
- Agarwal, P.K., Shukla, P.S., Gupta, K. and Jha, B. (2013). Bioengineering for salinity tolerance in plants: State of the art. *Molecular Biotechnology* 54: 102-123.
- Agarwal, P., Arora, R., Ray, S., Singh, A.K., Singh, V.P., Takatsuji, H., Kapoor, S. and Tyagi, A.K (2007). Genome-wide identification of C2H2 zinc-finger gene family in rice and their phylogeny and expression analysis. *Plant Molecular Biology* 65: 467-485
- Amirbakhtiar, N., Ismaili, A., Ghaffari, M.R., Nazarian, F.F. and Shobbar. Z.S. (2019). Transcriptome response of roots to salt stress in a salinity-tolerant bread wheat cultivar. *PLoS ONE* 14(3): e0213305.
- Anuar, N.H.S., Mazlan, N., Engku-Ariff, E.A.K., Juraimi, A.S. and Yusop, M.R. (2014). A comparative study of vegetative and reproductive growth of local weedy and Clearfield® rice varieties in Malaysia. *Journal of International Society for Southeast Asian Agricultural Science* 20(1): 41-51.
- Auer, H., Liyanarachchi, S., Newsom, D., Klisovic, M.I., Marcucci, G. and Kornacker, K. (2003). Chipping away at the chip bias: RNA degradation in microarray analysis. *Nature Genetics* 35: 292-293.
- Arora, K., Panda, K.K., Mittal, S., Mallikarjuna, M.G., Rao, A.R., Dash, P.K. and Thirunavukkarasu, N. (2017). RNA-seq revealed the important gene pathways controlling adaptive mechanisms under waterlogged stress in maize. *Scientific Reports* 7: 10950.
- Azizi, P., Rafii, M.Y., Mahmood, M., Akmar Abdullah, S.N., Hanafi, M.M., Latif, M.A., Sahebi, M., and Ashkani, S. (2017). Evaluation of RNA extraction methods in rice and their application in expression analysis of resistance genes

against Magnaporthe oryzae. *Biotechnology and Biotechnological Equipment* 31(1): 75-84.

- Baier, M., Kandlbinder, A., Golldack, D. and Dietz, K.J. (2005). Oxidative stress and ozone: Perception, signalling and response. *Plant, Cell and Environment* 28: 1012-1020.
- Baldoni, E., Genga, E. and Cominelli, E. (2015). Plant MYB transcription factors: Their role in drought response mechanisms. *International Journal of Molecular Science* 16: 15811-15851.
- Bass, H.W., Krawetz, J.E., OBrian, G.R., Zinselmeier, C., Habben, J.E. and Boston, R. (2004). Maize ribosome-inactivating proteins (RIPs) with distinct expression patterns have similar requirements for proenzyme activation. *Journal of Experimental Botany* 55(406): 2219-2233.
- Basu, S. and Roychoudhury, A. (2014). Expression profiling of abiotic stress-inducible genes in response to multiple stresses in rice (Oryza sativa L.) varieties with contrasting level of stress tolerance. *BioMed Research International* 12: 706890.
- Bota, J., Medrano, H. and Flexas, J. (2004). Is photosynthesis limited by decreased Rubisco activity and RuBP content under progressive water stress?. *New Phytologist Journal* 163(3): 671-681.
- Bowman, M.J., Park, W., Bauer, P.J., Udall, J.A., Page, J.T. and Raney, J. (2013). RNA-Seq transcriptome profiling of upland cotton (Gossypium hirsutum L.) root tissue under water-deficit stress. *PLoS One* 8(12): e82634.
- Breton, G., Danyluk, J., Charron, J.B.F. and Sarhan, F. (2003). Expression profiling and bioinformatic analyses of a novel stress-regulated multispanning transmembrane protein family from cereals and Arabidopsis. *Plant physiology* 132(1): 64-74.
- Buermans, H.P.J. and Dunnen, J.T. (2014). Next generation sequencing technology: Advances and applications. *Biochimica et Biophysica Acta* 1842: 1932-1941.
- Burla, B., Pfrunder, S., Nagy, R., Francisco, R.M., Lee, Y. and Martinoia. (2013). Vacuolar transport of abscisic acid glucosyl ester is mediated by ATP-binding cassette and proton-antiport mechanisms in Arabidopsis. *Plant Physiology* 163:1446-1458.
- Champagne, E.T., Wood, D.F., Juliano, B.O. and Bechtel, D.B. (2004). The rice grain and its gross composition. In: Champagne, E.T. (Ed.), Rice Chemistry and Technology. American Association of Cereal Chemists, pp. 93e96
- Chatterjee, S. (1997). The Rise of Birds. Baltimore, Maryland: John Hopkins University Press.

- Chen, T. H. and Murata, N. (2002). Enhancement of tolerance of abiotic stress by metabolic engineering of betaines and other compatible solutes. *Current Opinion in Plant Biology* 5(3): 250-257.
- Chuong, C.M. (1993). The making of a feather: Homeoproteins, retinoids and adhesion molecules. *BioEssays* 15: 513-521.
- Chutia, J. and Borah, S.P. (2012). Water stress effects on leaf growth and chlorophyll content but not the grain yield in traditional rice (Oryza sativa Linn.) Genotypes of Assam, India II. Protein and proline status in seedlings under peg induced water stress. *American Journal of Plant Sciences* 3(7): 20679.
- Claeys, H. and Inzé, D. (2013). The agony of choice: How plants balance growth and survival underwater-limiting conditions. *Plant Physiology* 162:1768-1779.
- Conesa, A., Madrigal, P., Tarazona, S., Cabrero, D.G., Cervera, A., McPherson, A., Szcześniak, M.W., Gaffney, D.J., Elo, L.L., Zhang, X. and Mortazavi, A. (2016). A survey of best practices for RNA-seq data analysis. *Genome Biology* 17:13.
- Cloonan, N. and Grimmond, S. (2008). Transcriptome content and dynamics at singlenucleotide resolution. *Genome Biology* 9: 234.
- Cutler, S.R., Rodriguez, P.L., Finkelstein, R.R. and Abrams, S.R. (2010). Abscisic acid: Emergence of a core signaling network. *Annual Review of Plant Biology* 61: 651-679.
- Datta, S.K., Malabuyoc, J.A. and Aragon, E.L. (1988). A field screening technique for evaluating rice germplasm for drought tolerance during vegetative stage. *Field Crops Research* 19: 123-124.
- David, L., Huber, W., Granovskaia, M., Toedling, J., Palm, C.J., Bofkin, L., Jones, T., Davis, R.W. and Steinmetz, L.M. (2006). A high-resolution map of transcription in the yeast genome. *Proceeding of the National Academic Science* 103: 5320-5325.
- Degenkolbe, T., Do, P.T., Zuther, E., Repsilber, D., Walther, D., Hincha, D.K. and Köhl, K.I. (2009). Expression profiling of rice cultivars differing in their tolerance to long-term drought stress. *Plant Molecular Biology* 69(1-2): 133-153.
- Dipti, S.S., Bergman, C., Indrasari, S.D., Herath, T., Hall, R. and Lee, H. (2012). The potential of rice to offer solutions for malnutrition and chronic diseases. *Rice* 5: 1-18.
- Dong, Y., Fan, G., Zhao, Z., and Deng, M. (2014). Transcriptome expression profiling in response to drought stress in *Paulownia australis*. *International Journal of Molecular Science* 15(3): 4583-4607.

- Du, H., Wang, Z., Yu, W. and Huang, B. (2012). Metabolic responses of hybrid Bermudagrass to short-term and long-term drought stress. *Journal of American Society for Horticultural Science* 137(6): 411-420.
- Delauney, A.J. and Verma, D.P.S. (1993). Proline biosynthesis and osmoregulation in plants. *Plant Journal* 4: 215-223
- Estrada, A.D., Freese, N.H., Blakley, I.C. and Loraine, A.E. (2015). Analysis of pollenspecific alternative splicing in *Arabidopsis thaliana via* semi-quantitative PCR. *PeerJ* 3: e919.
- Fang, C., Dou, L.L., Liu, Y.R., Yu, J.S. and Tu, J. (2018). Heat stress-responsive transcriptome analysis in heat susceptible and tolerant rice by high-throughput sequencing. *Ecological Genetics and Genomics* 6: 33-40
- FAO. Retrieved 6 December 2018 from http://www.faostat.fao.org
- Farooq, M.A., Wahid, N., Kobayashi, D., Fujita, S. and Basra, M.A. (2009). Plant drought stress: Effects, 320 mechanisms and management. Mechanisms and management. Agronomy for Sustainable Development 29: 185-212.
- Feduccia, A. (1999). The Origin and Evolution of Birds 2nd, Ed. New Haven, Connecticut: Yale University Press.
- Feng, Q., Zhang, Y., Hao, P., Wang, S., Fu, G., Huang, Y., Li, Y., Zhu, J., Liu, Y., Hu, X., Jia, P., Zhang, Y., Zhao, Q., Ying, K., Yu, Y., Tang, Y., Weng, Q., Zhang, L., Lu, Y., Mu, J., Lu, Y., Zhang, L.S., Yu, Z., Fan, D., Liu, X., Lu, T., Li, C., Wu, Y., Sun, T., Lei, H., Li, T., Hu, H., Guan, J., Wu, M., Zhang, R., Zhou, B., Chen, Z., Chen, L., Jin, Z., Wang, R., Yin, H., Cai, Z., Ren, S., Lv, S., Gu, W., Zhu, G., Tu, Y., Jia, J., Zhang, Y., Chen, J., Kang, H., Chen, X., Shao, C., Sun, Y., Hu, Q., Zhang, Z., Zhang, W., Wang, L., Ding, C., Sheng, H., Gu, J., Chen, S., Ni, L., Zhu, F., Chen, W., Lan, L., Lai, Y., Cheng, Z., Gu, M., Jiang, J., Li, J., Hong, G., Xue, Y., and Han, B.(2002). Sequence and analysis of rice chromosome 4. *Nature* 420: 316-320.
- Ferre, F. (1992). Quantitative or semi-quantitative PCR: Reality versus myth. Carlsbad, California: Cold Spring Harbor Laboratory Press 2:1-99.
- Flexas, J., Ribas-Carbó, M., Bota, J., Galmés, J., Henkle, M., Martínez-Cañellas, S. and Medrano, H. (2006). Decreased Rubisco activity during water stress is not induced by decreased relative water content but related to conditions of low stomatal conductance and chloroplast CO2 concentration. *New Phytologist* 172(1): 73-82.
- Franca, M., Prados, L., Lemos-Filho, J., Ranieri, B. and Vale, F. (2012). Morphophysiological differences in leaves of Lavoisiera campos-portoana (Melastomataceae) enhance higher drought tolerance in water shortage events. *Journal of Plant Research* 125: 85-92.

- Fujii, H., Verslues, P.E. and Zhu, J.K. (2011). Arabidopsis decuple mutant reveals the importance of SnRK2 kinases in osmotic stress responses in vivo. *Proceeding* of the National Academy Science of United State of America 108: 1717-1722.
- Gasulla, F., Vom, D.K., Dombrink, I., Zähringer, U., Gisch, N. and Dörmann, P. (2013). The role of lipid metabolism in the acquisition of desiccation tolerance in Craterostigmaplantagineum: A comparative approach. *The Plant Journal* 75: 726-741.
- Gautier, L., Cope, L., Bolstad, B.M. and Irizarry, R.A. (2004). Affy-analysis of Affymetrix Gene Chip data at the pro be level. *Bioinformatics* 20: 307-315.
- Gechev, T.S., Dinakar, C., Benina, M., Toneva, V. and Bartels, D. (2012). Molecular mechanisms of desiccation tolerance in resurrection plants. *Celllar and Molecular Life Science* 69: 3175-3186.
- Gill, P.K., Sharma, A.D., Singh, P. and Bhullar, S.S. (2001). Effect of various abiotic stresses on the growth soluble sugars and water relations of sorghum seedlings grown in light and darkness. *Bulgarian Journal of Plant Physiology* 27: 72-84.
- Golldack, D., Li, C., Mohan, H. and Probst, N. (2014). Tolerance to drought and salt stress in plants: unraveling the signaling networks. *Frontiers in Plant Science* 5(151): 1.
- Golldack, D., Lüking, I. and Yang, O. (2011). Plant tolerance to drought and salinity: Stress regulating transcription factors and their functional significance in the cellular transcriptional network. *Plant Cell Reports* 30: 1383-1391.
- Gonzalez, M.P., Kurbatova, N., Griebel, T., Ferreira, P.G., Barann, M., Wieland, T., Greger, L., Iterson, M., Almlof, J., Ribeca, P., Pulyakhina, I., Esser, D., Giger, T., Tikhonov, A., Sultan, M., Bertier, G., MacArthur, D.G., Lek, M., Lizano, E., Buermans, H.P.J., Padioleau, I., Schwarzmayr, T., Karlberg, O., Ongen, H., Kilpinen, H., Beltran, S., Gut, M., Kahlem, K., Amstislavskiy, V., Stegle, O., Pirinen, M., Montgomery, S.B., Donnelly, P., McCarthy, M.I., Flicek, P., Strom, T.M., Consortium, T.G., Lehrach, H., Schreiber, S., Sudbrak, R., Carracedo, A., Antonarakis, S.E., Hasler, R., Syvanen, G.-J., van Ommen, A. C., Brazma, A., Meitinger, T., Rosenstiel, P., Guigo, R., Gut, I.G., Estivill, X. and Dermitzakis, E.T. (2013). Transcriptome and genome sequencing uncovers functional variation in humans. *Nature* 501: 506-511.
- Guo, L., Yang, H., Zhang, X. and Yang, S. (2013). Lipid transfer protein 3 as a target of MYB96 mediates freezing and drought stress in *Arabidopsis*. *Journal of Experimental Botany* 64: 1755-1767.
- Gupta, A.K and Kaur, N. (2005). Sugar signaling and gene expression in relation to carbohydrate metabolism under abiotic stresses in plants. *Journal of Bioscience* 30: 761-76.
- Gupta, R., Lee, S.E., Agrawal, G.K., Rakwal, R., Park, S., Wang, Y. and Kim, S.T. (2015). Understanding the plant pathogen interactions in the context of

proteomics-generated apoplastic proteins inventory. *Frontiers in Plant Science* 6: 352.

- Habib, H. and Fazili K.M. (2007). Protease inhibitor: A defense strategy in plant. *Biotechnology and Molecular Biology Review* 2(3): 068-085.
- Hadiarto, T. and Tran, L.S.P. (2011). Progress studies of drought-responsive genes in rice. *Plant Cell Reports* 30(3): 297-310.
- Halford, N.G. and Hey, S.J. (2009). Snf1-related protein kinases (SnRKs) act within an intricate network that links metabolic and stress signaling in plants. *Biochemical Journal* 419: 247-259.
- Harb, A., Krishnan, A., Madana, M.R., Ambavaram, and Pereira, A. (2010). Molecular and physiological analysis of drought stress in Arabidopsis reveals early responses leading to acclimation in plant growth. *Plant Physiology* 154: 1254-1271.
- Harrak, H., Azelmat, S., Baker, E.N. and Tabaeizadeh, Z. (2001). Isolation and characterization of a gene encoding a drought-induced cysteine protease in tomato (Lycopersicon esculentum). *Genome* 44: 368-374.
- Hey, S.J., Byrne, E. and Halford, N.G. (2010). The interface between metabolic and stress signalling. *Annals of Botany* 105: 197-203.
- Hilarion, S.G., Paulet, D., Lee, K.T., Hon, C.C., Lechat, P., Mogensen, E., Moyrand, F., Proux, C., Barboux, R., Bussotti, G., Hwang, J., Coppée, J.Y., Bahn, Y, Sun. and Janbon, G. (2016). Intron retention-dependent gene regulation in Cryptococcus neoformans. *Scientific Reports* 6: 32252.
- Hirasawa, T. (1999). Physiological characterization of rice plant for tolerance of water deficit, in genetic improvement of rice for water-limited environments. Ed. Ito O, O'Toole JC, Hardy B., pp. 89–98. Philippines: *International Rice Research Institute*, Los Baños.
- Hochberg, U., Degu, A., Toubiana, D., Gendler, T., Nikoloski, Z., Rachmilevitch, S. and Fait, A. (2013). Metabolite profiling and network analysis reveal coordinated changes in grapevine water stress response. *BMC Plant Biology* 13: 184.
- Hoekstra, F.A., Golovina, E.A. and Buitink, J. (2001). Mechanisms of plant desiccation tolerance, Trends. *Plant Science* 6: 431-438.
- Hoen, P.A.T., Ariyurek, Y., Thygesen, H.H., Vreugdenhil, E.R., Vossen, H., Menezes, R.X., Boer, J.M., Ommen, G.J.B. and Dunnen, J.T. (2008). Deep sequencingbased expression analysis shows major advances in robustness, resolution and inter-lab portability over five microarray platforms. *Nucleic Acids Research* 36: 141.

- Hongbo, S., Zongsuo, L. and Mingan, S. (2005). Changes of anti-oxidative enzymes and MDA content under soil water deficits among 10 wheat (Triticum aestivum L.) genotypes at maturation stage. Colloids and Surface B: *Biointerfaces* 45: 7-13.
- Hossain, Z., Nouri, M.Z. and Komatsu, S. (2012). Plant cell organelle proteomics in response to abiotic stress. *Journal of Proteome Research* 11(1): 37-48.
- Huang, G.T., Ma SL, Bai, L.P., Zhang, L., Ma, H. and Jia, P. (2012). Signal transduction during cold, salt, and drought stresses in plants. *Molecular Biology Reports* 2: 969-987.
- Huang, H., Møller, I.M. and Song, S. (2012). Proteomics of desiccation tolerance during development and germination of maize embryos. *Journal of Proteomics* 75: 1247-1262.
- Hussain, B. (2015). Modernization in plant breeding approaches for improving biotic stress resistance in crop plants. *Turkish Journal of Agriculture* 39.
- Ichimura, K., Mizoguchi, T., Yoshida, R., Yuasa, T. and Shinozaki, K. (2000). Various abiotic stresses rapidly activate Arabidopsis MAP kinases ATMPK4 and ATMPK6. *The Plant Journal* 24: 655-65.
- International Barley Genome Sequencing Consortium. (2012). A physical, genetic and functional sequence assembly of the barley genome. *Nature* 491: 711-716.
- International Wheat Genome Sequencing Consortium. (2014). A chromosomebased draft sequence of the hexaploid bread wheat (*Triticum aestivum*) genome. Science 345: 1251788.
- IRRI, Rice. Research Institute. (1988). Annual report for 1987. P.O. Box 933, Manila, Philippines.
- Isendahl, N. and Schmidt, G. (2006). Drought in the Mediterranean-WWF Policy Proposals. A WWF Report, Madrid.
- Ives, C.L., Johanson, A. and Lewis, J. (2001). Agricultural Biotechnology: A Review of Contemporary Issues. United States Agency for International Development. http://pdf.dec.org/pdf_docs/PNACN153.pdf
- Jain, M., Nijhawan, A., Tyagi., A.K. and Khurana, J.P. (2006). Validation of housekeeping genes as internal control for studying gene expression in rice by quantitative real-time PCR. *Biochemical and Biophysical Research Communication* 345: 646-651.
- Jarzyniak, K.M. and Jasi'nski, M. (2014). Membrane transporters and drought resistance–a complex issue. *Frontiers in Plant Science* 5: 687.
- Jaspers, P. and Kangasjärvi, J. (2010). Reactive oxygen species in abiotic stress signaling. Physiologia Plantarum 138: 405-413.

- Jiang, M. and Zhang, J. (2002). Water stress-induced abscisic acid accumulation triggers the increased generation of reactive oxygen species and up-regulates the activities of antioxidant enzymes in maize leaves. *Journal of Experimental Botany* 53(379): 2401-2410.
- JiangA S.S., Liang, X.N., Li, X., Wang, S.L., Lv, D.W., Ma, C.Y., Li, X.H., Ma, W.J. and Yan, Y.M. (2012). Wheat Drought-Responsive Grain Proteome Analysis by Linear and Nonlinear 2-DE and MALDI-TOF Mass Spectrometry. *Journal* of Molecular Science 13(12): 16065-16083.
- JiangB S.Y., Bhalla, R., Ramamoorthy, R., Luan, H.F., Venkatesh, P.N., Cai, M. and Ramachandran, S. (2012). Over-expression of OSRIP18 increases drought and salt tolerance in transgenic rice plants. *Transgenic Research* 21: 785-795.
- Jones, H.G., Flowers, T.J. and Jones, M.B. (1989). Plants under Stress. Cambridge: Cambridge University Press.
- Kang, G.Z., Wang, Z.X. and Sun, G.C. (2003). Participation of H2O2 in enhancement of cold chilling by salicylic acid in banana seedlings. *Acta Botanica Sinica* 45: 567-573.
- Kinoshita, T., Nishimura, M. and Shimazaki, K.I. (1995). Cytosolic concentration of Ca2+ regulates the plasma membrane H+-ATPase in guard cells of fava bean. *Plant Cell* 7: 1333-1342.
- Kolker, E., Makarova, K.S., Shabalina, S., Picone, A.F., Purvine, S., Holzman, T., Cherny, T., Armbruster, D., Jr, R.S.M., Kolesov, G., Frishman, D. and Galperin, M.Y. (2004). Identification and functional analysis of `hypothetical' genes expressed in Haemophilus influenzae. *Nucleic Acids Research* 32(8): 2353±2361.
- Kong, X., Pan, J., Zhang, M., Xing, X., Zhou, Y., Liu, Y. and Li, D. (2011). ZmMKK4, A novel group C mitogen-activated protein kinase kinase in maize (Zea mays), confers salt and cold tolerance in transgenic Arabidopsis. *Plant, Cell and Environment* 34: 1291-1303.
- Krasensky, J. and Jonak, C. (2012). Drought, salt, and temperature stress-induced metabolic rearrangements and regulatory networks. *Journal of Experimental Botany* 1-16.
- Krochko, J.E., Abrams, G.D., Loewen, M.K., Abrams, S.R. and Cutler, A.J. (1998). (+)-Abscisic acid 8'-hydroxylase is a cytochrome P450 monooxygenase. *Plant Physiology* 118(3): 849-60.
- Kuromori, T., Miyaji, T., Yabuuchi, H., Shimizu, H., Sugimoto, E. and Kamiya, A. (2010). ABC transporter AtABCG25 is involved in abscisic acid transport and responses. *Proceedings of the National Academy of Sciences* 107: 2361-2366.
- Lawson, T. and Blatt, M.R. (2014). Stomatal size, speed, and responsiveness impact on photosynthesis and water use efficiency. *Plant Physiology* 164: 1556-1570.

- Lesk, C., Rowhani, P. and Ramankutty, N. (2016). Influence of extreme weather disasters on global crop production. *Nature* 529: 84–87.
- Li, K.Q., Xu, X.Y. and Huang, X.S. (2016). Identification of differentially expressed genes related to dehydration resistance in a highly drought-tolerant pear, Pyrus betulaefolia, as through RNA-Seq. *PLoS ONE* 11(2): e0149352.
- Lian, H.L., Yu, X., Ye, Q., Ding, X.S., Kitagawa, Y., Kwak, S.S., Su, W.I. and Tang, Z.C. (2004). The Role of Aquaporin RWC3 in Drought Avoidance in Rice. *Plant Cell Physiology* 45(4): 481-489.
- Liu, S.C., Jin, J.Q., Ma, J.Q., Yao, M.Z., Ma, C.L. and Li, C.F. (2016). Transcriptomic analysis of tea plant responding to drought stress and recovery. *PLoS ONE* 11(1): e0147306.
- Liu, X., Du, F., Li, N., Chang, Y. and Yao, D. (2016). Gene expression profile in the long-living lotus: insights into the heat stress response mechanism. *PLoS ONE* 11(3): e0152540.
- Liu, T., Zhu, S., Tang, Q., Yu, Y. and Tang, S. (2013). Identification of drought stressresponsive transcription factors in ramie (Boehmeria nivea L. Gaud). *BMC Plant Biology* 13: 130.
- Liu, Y., Zhou, J. and White, K.P. (2014). RNA-seq differential expression studies: more sequence or more replication?. *Bioinformatics* 30(3): 301–304.
- Liu, Y., Jiang, Y., Lan, J., Zou, Y. and Gao, J. (2014). Comparative transcriptomic analysis of the response to cold acclimation in Eucalyptus dunnii. *PLoS ONE* 9(11): e113091
- Livak, K.J. and Schmittgen, T.D. (2001). Analysis of relative gene expression data us¬ing real-time quantitative PCR and the 2(-Del¬ta Delta C(T)) method. *Methods* 25: 402-408.
- Loresto, G.C. and Chang, T.T. (1981). Decimal scoring system for drought reactions and recovery ability in screening murseries of rice. *International Rice Research Newsletter* 6(2): 9-10.
- Lucas, A.M. and Stettenheim, P.R. (1972). Avian Anatomy-Integument. Agricultural Handbook 362: Agricultural Research Services Eds, US Department of Agriculture, Washington DC.
- Ma, J., Li, R., Wang, H., Li, D., Wang, X., Zhang, Y., Zhen, W., Duan, H., Yan, G. and Li, Y. (2017). Transcriptomics analyses reveal wheat responses to drought stress during reproductive stages under field conditions. *Frontiers of Plant Science* 8: 592.
- Ma, N.N., Zuo, Y.Q., Liang, X.Q., Yin, B., Wang, G.D. and Meng, Q.W. (2013). The multiple stress-responsive transcription factor SINAC1 improves the chilling tolerance of tomato. *Physiologia Plantarum Physiologia Plantarum* 149: 474– 486.

- Mahajan, S. and Tuteja, N. (2005). Cold, salinity and drought stresses: An overview. *Archives of Biochemistry and Biophysics* 444: 139-158.
- Marguerat, S., Wilhelm, B. and Bähler, J. (2008). Next-generation sequencing: Applications beyond genomes. *Biochemical Society Transactions* 36: 1091-1096.
- Marioni, J., Mason, C., Mane, S., Stephens, M. and Gilad, Y. (2008). RNA-Seq: An assessment of technical reproducibility and comparison with gene expression arrays. *Genome Research* 18: 1509-1517.
- Martinez, M., Cambra, I., Carrillo, L., Diaz-Mendoza, M. and Diaz, I. (2009). Characterization of the entire cystatin gene family in barley and their target cathepsin l-like cysteine-proteases, partners in the hordein mobilization during seed germination. *Plant Physiology* 151(3): 1531-1545.
- Matsumoto, T., Wu, J., Itoh, T., Numa, H. and Antonio, B.T.S. (2016). The Nipponbare genome and the next generation of rice genomics research in Japan. *Rice* 9:33.
- McPherson, M.J., Hames, B.D. and Taylor, G.R. (1995). PCR: A Practical Approach. Oxford: IRL Press.
- Menichelli, C., Gascuel, O. and Bréhélin, L. (2018). Improving pairwise comparison of protein sequences with domain co-occurrence. *PLOS Computational Biology* 14(1):e1005889.
- Miller, H.C., Biggs, P.J., Voelckel, C. and Nelson, N.J. (2012). De novo sequence assembly and characterisation of a partial transcriptome for an evolutionarily distinct reptile, the tuatara (Sphenodon punctatus). *BMC Genomics* 13: 439.
- Mitra, R.D. and Church, G.M. (1999). In situ localized amplification and contact replication of many individual DNA molecules. *Nucleic Acids Research* 27: e34-e39.
- Mittler, R. (2002). Oxidative stress, antioxidants and stress tolerance. *Trends Plant Science* 7: 405-410.
- Marone, M., Mozzetti, S., De Ritis, D., Pierelli, L. and Scambia, G. (2001). Semiquantitative RT-PCR analysis to assess the expression levels of multiple transcripts from the same sample. *Biology Proceeding Online* 3(1): 19-25.
- Mohapatra, D. and Bal, S. (2006). Cooking quality and instrumental textural attributes of cooked rice for different milling fractions. *Journal of Food Engineering* 73(3): 253-259.
- Mostajeran, A. and Rahimi-Eichi, V. (2009). Effects of drought stress on growth and yield of rice (Oryza sativa L.) cultivars and accumulation of proline and soluble sugars in sheath and blades of their different ages leaves. *American-Eurasian Journal of Agricultural and Environmental Science* 5(2): 264-272.

- Najmuddin, O., Rasul, G., Hussain, A., Molden, D., Wahid, S. and Debnath, B. (2018). Low Water Productivity for Rice in Bihar, India: A Critical Analysis, *Water* 10: 1082.
- Nagalakshmi, U., Waern, K. and Snyder, M. (2010). RNA-Seq: A method for comprehensive transcriptome analysis. *Current Protocols in Molecular Biology* 4(11): 1-4.
- Negi, J., Matsuda, O., Nagasawa, T., Oba, Y., Takahashi, H. and Kawai-Yamada, M. (2008). CO2 regulator SLAC1 and its homologues are essential for anion homeostasis in plant cells. *Nature* 452: 483-486.
- Obidiegwu, J.E., Bryan, G.J., Jones, H.G. and Prashar, P. (2015). Coping with drought: stress and adaptive responses in potato and perspectives for improvement. *Frontiers in Plant Science* 6: 542.
- Osakabe, Y., Arinaga. N., Umezawa, T., Katsura, S., Nagamachi, K. and Tanaka, H. (2013). Osmotic stress responses and plant growth controlled by potassium transporters in *Arabidopsis*. *Plant Cell* 25: 609-624.
- Osakabe, Y., Maruyama, K., Seki, M., Satou, M., Shinozaki, K. and Yamaguchi-Shinozaki, K. (2005). Leucine-rich repeat receptor-like kinasel is a key membrane-bound regulator of abscisic acid early signaling in arabidopsis. *Plant Cell* 17(4): 1105-1119.
- Osakabe, Y., Osakabe, K., Shinozak, K. and Tran, L.S.P. (2014). Mini review article: Response of plants to water stress. *Frontiers in Plant Science* 5(86): 1.
- Othman, T.A., Norliza, A.B., Rabiatul, A.Z.A., Mahmood, M., Saidi N.B.B. and Shaharuddin, N.A. (2014). Potential of plant's Bowman-Birk protease inhibitor in combating abiotic stresses: A mini review. *Bioremediation Science* & *Technology Research* 2(2): 53-61.
- Passioura, J.B. (2007). The drought environment: Physical, biological and agricultural perspectives. *Journal of Experimental Botany* 58: 113-117.
- Paterson, A., Bowers, J., Bruggmann, R., Dubchak, I., Grimwood, J. and Gundlach, H. (2009). The Sorghum bicolor genome and the diversification of grasses. *Nature* 457: 551-556.
- Price, A., Garhyan, J. and Gibas, C. (2017). The impact of RNA secondary structure on read start locations on the Illumina sequencing platform. *PLoS One* 12(2): e0173023.
- Qureshi, A.A., Mo, H., Packer, L. and Peterson, D.M. (2000). Isolation and identification of novel tocotrienols from rice bran with hypocholesterolemic, antioxidant, and antitumor properties. *Journal of Agricultural and Food Chemistry* 48: 3130-3140.
- Rapala-Kozik, M., Wolak, N., Kujda, M. and Banas, A.K. (2012). The upregulation of thiamine (vitamin B1) biosynthesis in Arabidopsis thaliana seedlings under

salt and osmotic stress conditions is mediated by abscisic acid at the early stages of this stress response. *BMC Plant Biology* 12: 2.

- Rajamoorthy, Y., Rahim, K.B.A. and Munusamy, S. (2015). Rice industry in Malaysia: challenges, policies and implications. *Procedia Economics and Finance* 31: 861-867.
- Rashid B., Tariq M., Khalid A., Shams F., Ali Q., Ashraf F., Ghaffar I., Khan M.I., Rehman R., and Husnain T (2017). Crop improvement: new approaches and modern techniques. *Plant Gene and Trait* 8(3): 18-30.
- Rice Chromosome 10 Sequencing Consortium. (2003). In-depth view of structure, activity, and evolution of rice chromosome 10. *Science* 300: 1566-1569.
- Romero, I.G., Pail, A.A., Tung, J. and Gilad, Y. (2014). RNA-Seq: impact of RNA degradation on transcript quantification. *BMC Biology* 12: 42.
- Ryan, C.A. (1990). Proteinase inhibitors in plants: genes for improving defenses against insects and pathogens. *Annual Review of Phytopathology* 28: 425-449.
- Saglam, A., Terzi, R. and Demiralay, M. (2014). Effect of polyethylene glycol induced drought stress on photosynthesis in two chickpea genotypes with different drought tolerance. *Acta Biologica Hungarica* 65(2): 6.
- Sakata, K., Antonio, B.A., Mukai, Y., Nagasaki, Y., Sakai, Y., Makino, K., and Sasakia, T. (2000). A rice genome database with an integrated map view. *Nucleic Acids Research* 28(1): 97-101.
- Sánchez-Hernández, C., Martínez-Gallardo, N., Guerrero-Rangel, A., Valdés-Rodríguez, S. and Délano-Frier, J. (2004). Trypsin and α-amylase inhibitors are differentially induced in leaves of amaranth (*Amaranthus hypochondriacus*) in response to biotic and abiotic stress. *Physiologia Plantarum* 122(2): 254-264.
- Sanger, F. (1977). DNA sequencing with chain-terminating inhibitors. *Proceeding of the National Academy Science of United State of America* 74: 5463-5467.
- Sanghera, G.S., Wani, S.H., Hussain, W. and Singh, N.B. (2011). Engineering cold stress tolerance in crop plants. *Current Genomics* 12: 30-43.
- Sarunyaporn, M., Kanyaratt, S. and Gopalan, S. (2013). High-quality reference genes for quantifying the transcriptional responses of Oryza sativa L. (ssp. indica and japonica) to abiotic stress conditions. *Chinese Science Bulletin* 58: 1919-1930.
- Sarvestani, Z.T., Pirdashti, H., Sanavy, S.A.M.M. and Balouchi, H. (2008). Study of water stress in effects in the different growth stages on yield and yield component of different rice Oryza sativa L. cultivar. Pakistan Journal of Biological Sciences 11(10): 1303-1309.

Sasaki, T. (2005). The map-based sequence of the rice genome. Nature 436: 783-800.

- Sasaki, T. *et al.* (2002). The genome sequence and structure of rice chromosome 1. Nature 420: 312-316.
- Satoh, K., Doi, K., Nagata, T., Kishimoto, N., Suzuki, K., Otomo, Y., Kawai, J., Nakamura, M., Hirozane-Kishikawa, T., Kanagawa, s., Arakawa, T., Takahashi-Iida, J., Murata, M., Ninomiya, M., Sasaki, d., Fukuda, S., Tagami, M., Yamagata, M., Kurita, K., Kamiya, K.,Yamamoto, M., Kikuta, A., Bito, T., Fujitsuka, N., Ito, I., Kanamori, H., Choi, R., Nagamura, Y., Matsumoto, T., Murakami, K., Matsubara, K.I., Carninci, P., Hayashizaki, Y. and Kikuchi, S. (2007). Gene organization in rice revealed by full-length cDNA mapping and gene expression analysis through microarray. *PLoSONE* 2(11): e1235.
- Schmutz, J., Cannon, S.B., Jessica S.J., Ma, J., Mitros, T., Nelson, W. and Hyten, D.L. (2010). Genome sequence of the palaeopolyploid soybean. *Nature* 463:178-183.
- Schnable, P., Ware, D., Fulton, R.S., Joshua, C., Stein, J.C., Fusheng, W.F. and Shiran, P.S. (2009). The B73 maize genome: Complexity, diversity, and dynamics. *Science* 326: 1112-1115.
- Schroeder, A., Mueller, O., Stocker, S., Salowsky, R., Leiber, M., Gassmann, M., Lightfoot, S., Menzel, W., Granzow, M. and Ragg, T. (2006). The RIN: An RNA integrity number for assigning integrity values to RNA measurements. BMC Molecular Biology 7: 3.
- Shi, H., Wang, X., Ye, T., Chen, F., Deng, J., Yang, P., Zhang, Y. and Chan, Z (2014). The Cysteine2/Histidine2-Type Transcription Factor Zinc Finger Of Arabidopsis Thaliana Modulates Biotic and Abiotic Stress Responses by Activating Salicylic Acid-Related Genes and C-Repeat-Binding Factor Genes in Arabidopsis. *Plant Physiology* 165:1367-1379
- Shinozaki, K. and Yamaguchi-Shinozaki, K. (2007). Gene networks involved in drought stress response and tolerance. *Journal of Experimental Botany* 58(2): 221-227.
- Shukla, N., Awasthi, R.P., Rawat, L. and Kumar, J. (2012). Biochemical and physiological responses of rice (Oryza sativa L.) as influenced by Trichoderma harzianum under drought stress. *Plant Physiology and Biochemistry* 54: 78-88.
- Silveira, R.D.D., Abreu, F.R.M., Mamidi, S., McClean, P.E., Vianello, R.P., Lanna, A.C., Carneiro, N.P. and Brondani, C. (2015). Expression of drought tolerance genes in tropical upland rice cultivars (*Oryza sativa*). *Genetic Molecular Research* 14(3): 8181-8200.
- Singh, D., Singh, C.K., Taunk, J., Tomar, R.S.S., Chaturvedi, A.K., Gaikwad K. and Pal, M. (2017). Transcriptome analysis of lentil (*Lens Culinaris Medikus*) in response to seedling drought stress. *BMC Genomics* 18: 206.

- Sinha, A.K., Jaggi, M., Raghuram, B. and Tuteja, N. (2011). Mitogen-activated protein kinase signaling in plants under abiotic stress. *Plant Signaling & Behavior* 6(2):196-203.
- Sotowa, M., Ootsuka, K., Kobayashi, Y., Hao, Y., Tanaka, K., Ichitani, K., Flowers, J.M., Purugganan, M.D., Nakamura, I., Sato, Y.I., Sato, T., Crayn, D., Simon, B., Waters, D.L.E., Henry, R.J. and Ishikawa, R. (2013). Molecular relationships between Australian annual wild rice, *Oryza meridionalis*, and two related perennial forms. *Rice* 6(1): 26.
- Sreenivasulu, N., Sopory, S.K. and Kishor, P.B.K. (2007). Deciphering the regulatory mechanisms of abiotic stress tolerance in plants by genomic approaches. *Gene* 388: 1-13.
- Subburaj, S., Zhu, D., Li, X., Hu, Y. and Yan, Y. (2017). Molecular characterization and expression profiling of *Brachypodium distachyon L*. Cystatin genes reveal high evolutionary conservation and functional divergence in response to abiotic stress. *Frontiers of Plant Science* 8: 743.
- Tamiru, M., Undan, J.R., Takagi, H., Abe, A., Yoshida, K., Undan, J.Q., Natsume, S., Uemura, A., Saitoh, H., Matsumura, H., Urasaki, N., Yokota, T. and Terauchi, R. (2015). A cytochrome P450, OsDSS1, is involved in growth and drought stress responses in rice (Oryza sativa L.). *Plant Molecular Biology* 88(1-2): 85-99.
- Tan, Y., Wang, S., Liang, D., Li, M. and Ma, F. (2014). Genome-wide identification and expression profiling of the cystatin gene family in apple (Malus × domestica Borkh.). *Plant Physiology Biochemistry* 79:88-97.
- Tarkow, H., Feist, W.C. and Southerland, C.F. (1996). Interaction of wood and polymeric materials penetration versus molecular size. *Forest Products Journal* 16: 61-65.
- Teige, M., Scheik, E., Eulgem, T., Doczi, R., Ichimura, K., Shinozaki, K., Dang, J.L. and Hirt, H. (2004). The MKK2 pathway mediates cold and salt stress signaling in Arabidopsis. *Molecular Cells* 15: 141-152.
- Tester, M. and Langridge, P. (2010). Breeding technologies to increase crop production in a changing world. *Science* 327: 818-822.
- Todaka, D., K. Shinozaki and K. Yamaguchi-Shinozaki, 2015. Recent advances in the dissection of drought-stress regulatory networks and strategies for development of drought-tolerant transgenic rice plants. *Frontiers in Plant Science* 6: 84
- Torres-Franklin, M.L., Gigon, A., Melo, D.F., Zuily-Fodil, Y. and Pham-Thi, A.T. (2007). Drought stress and rehydration affect the balance between MGDG and DGDG synthesis in cowpea leaves. *Physiologia Plantarum* 131: 201-210.
- Trapnell, C., Roberts, A., Goff, L., Pertea, G., Kim, D., Kelley, D.R., Pimentel, H., Salzberg, S.L., Rinn, J.L. and Pachter, L. (2012). Differential gene and

transcript expression analysis of RNA-Seq experiments with TopHat and Cufflinks. *Nature Protocols* 7(3): 562.

- Tsuchihara, K., Suzuki, Y., Wakaguri, H., Irie, T., Tanimoto, K., Hashimoto, S.I., Matsushima, K., Sugano, J.M., Yamashita, R., Nakai, K., Bentley, D., Esumi, H. and Sugano, S. (2009). Massive transcriptional start site analysis of human genes in hypoxia cells. *Nucleic Acid Research* 37: 2249-2263.
- Tsukagoshi, H., Suzuki, T., Nishikawa, K., Agarie, S., Ishiguro, S. and Higashiyama, T. (2015). RNA-Seq Analysis of the response of the halophyte, Mesembryanthemum crystallinum (Ice Plant) to high salinity. *PLoS ONE* 10(2): e0118339.
- Tuberosa, R. (2012). Phenotyping for drought tolerance of crops in the genomics era. *Frontier in Physiology* 3: 347.
- Vahisalu, T., Kollist, H., Wang, Y.F., Nishimura, N., Chan, W.Y. and Valerio, G. (2008). SLAC1 is required for plant guard cell S-type anion channel function in stomatal signalling. *Nature* 452: 487-491.
- Vanderauwera, S., Vandenbroucke, K., Inzé, A., VandeCotte, B., Mühlenbock, P. and Rycke, R. (2012). At WRKY15 perturbation abolishes the mitochondrial stress response that steers osmotic stress tolerance in Arabidopsis. Proceedings of the National Academy of Sciences of the United States of American 109: 20113-20118.
- Wang, M., Jiang, B., Liu, W., Lin, Y., Liang, Z., He, X. and Peng, Q. (2019). Transcriptome analyses provide novel insights into heat stress responses in Chieh-Qua (Benincasa hispida Cogn. var. Chieh-Qua How). International Journal of Molecular Science 20(4): 883.
- Wang, D., Yang, C., Dong, L., Zhu, J., Wang, J. and Zhang, S. (2015). Comparative transcriptome analyses of drought-resistant and - susceptible Brassica Napus L. and development of EST-SSR markers by RNA-Seq. *Journal of Plant Biology* 58: 259-269.
- Wang, Y.C., Jiang, J., Zhao, X., Liu, G.F., Yang, C.P. and Zhan, L.P. (2006). A novel LEA gene from *Tamarix Androssowii* confers drought tolerance in transgenic tobacco. *Plant Science*. 171(6): 655-62.
- Wang, P., Yang, C., Chen, H., Song, C., Zhang, X. and Wang, D. (2017). Transcriptomic basis for drought-resistance in *Brassica napus* L. *Scientific Reports* 7: 40532.
- Wang, W., Vinocur, B. and Altman, A. (2003). Plant responses to drought, salinity and extreme temperatures: Towardss genetic engineering for stress tolerance. *Planta* 218: 1-14.
- Wang, Z., Gerstein, M. and Snyder, M. (2009). RNA-Seq: A revolutionary tool for transcriptomics. *Nature Reviews Genetics* 10: 57-63.

- Wei, S., Hu, W., Deng, X., Zhang, Y., Liu, X., Zhao, X., Luo, X., Jin, Z., Li, Y., Zhou, S., Sun, T., Wang, L., Yang, G. and He, G. (2014). A rice calcium-dependent protein kinase OsCPK9 positively regulates drought stress tolerance and spikelet fertility. *BMC Plant Biology* 14: 133.
- Wei, S., Du, Z., Gao, F., Ke, X., Li, J. and Liu, J. (2015). Global transcriptome profiles of 'meyer' zoysiagrass in response to cold stress. *PLoS ONE* 10(6): e0131153.
- Wei, X. and Huang, X. (2019). Origin, taxonomy, and phylogenetics of rice. Published by Elsevier Inc. in cooperation with AACC International.
- Weiner, J.J., Peterson, F.C., Volkman, B.F. and Cutler, S.R. (2010). Structural and functional in sights in to core ABA signaling. *Current Opinion in Plant Biology* 13: 495-502.
- Weingartner, M., Subert, C. and Sauer, N (2011). LATE, a C2H2 zinc-finger protein that acts as floral repressor. *Plant Journal* 68:681-692.
- Welch, R.M. and Graham, R.D. (2004). Breeding for micronutrients in staple food crops from a human nutrition perspective. *Journal of Experimental Botany* 55: 353-364.
- Wickramasinghe, S., Cánovas, A., Rincón, G. and Medrano, N.F. (2014). RNA-Sequencing: A tool to explore new frontiers in animal genetics. *Livestock Science* 166: 206-216.
- Xu, Z., Hua, N. and Godber, J. S. (2001). Antioxidant activity of tocopherols, tocotrienols and γ -oryzanol components from rice bran against cholesterol oxidation accelerated by 2,2-azobis (2-methylpropionamidine) dihydrochloride. *Journal of Agricultural and Food Chemistry* 49: 2077-2081.
- Xue, G.P., McIntyre, C.L., Glassop, D. and Shorter, R. (2008). Use of expression analysis to dissect alterations in carbohydrate metabolism in wheat leaves during drought stress. *Plant Molecular Biology* 67: 197-214.
- Yamamoto, T., Nagasaki, H., Yonemaru, J., Ebana, K., Nakajima, M., Shibaya, T. and Yano, M. (2010). Fine definition of the pedigree haplotypes of closely related rice cultivars by means of genome-wide discovery of single-nucleotide polymorphisms. *BMC Genomics* 11: 267.
- Yee, W.S., Syamimi Aziz, S.D.A. and Yusof, Z.N.B (2016). Osmotic stress upregulates the transcription of thiamine (vitamin B1) biosynthesis genes (THIC and THI4) in oil palm (Elaies guineensis). *African Journal of Biotechnology* 15(29); 1566-1574.
- Yoshida, S., Forno, D.A., Cock, J.H. and Gomez, K.A. (1976). Laboratory Manual for Physiological Studies of Rice. 83. IRRI, Las Banos, Laguna.
- Zain, N.A.M., Ismail, M.Z., Puteh, A., Mahmood, M. and Islam, M.R. (2014). Impact of cyclic water stress on growth, physiological responses and yield of rice

(Oryza sativa L.) grown in tropical environment. Ciência Rural, Santa Maria 44(12): 2136-2141.

- Zainuddin, H., Sariam, O., Chan, C.S., Azmi, M., Saad, A., Alias, I. and Marzukhi, H. (2014). Performance of selected aerobic rice varieties cultivated under local condition. *Journal of Tropical Agriculture and Food Science* 42(2): 1Z7a5i n– u d1i8n2.
- Zang, O.W., Wang, C.X., Li, X.Y., Guo, Z.A., Jing, R.L., Zhao, J. and Chang, X.P. (2010). Isolation and characterization of a gene encoding a polyethylene glycol-induced cysteine protease in common wheat. *Journal of Biosciences* 35: 379-388.
- Zhang, Q. (2007). Strategies for developing Green Super Rice. Proceedings of the National Academy of Sciences of the United States of America 104:16402-16409.
- Zhang, X., Liu, S. and Takano, T (2008). Two cysteine proteinase inhibitors from Arabidopsis thaliana, AtCYSa and AtCYSb, increasing the salt, drought, oxidation and cold tolerance. *Plant Molecular Biology* 68:131.
- Zhang, H., Liu, Y., Wen, F., Yao, D., Wang, L., Guo, J., Ni, L., Zhang, A., Tan, M. and Jiang, M. (2014). A novel rice C2H2-type zinc finger protein, ZFP36, is a key player involved in abscisic acid-induced antioxidant defense and oxidative stress tolerance in rice. *Journal of Experimental Botany* 65(20): 5795-5809.
- Zhang, Y., Kong, X., Dai, J., Luo, Z., Li, Z., and Lu, H. (2017). Global gene expression in cotton (*Gossypium hirsutum L*.) leaves to waterlogging stress. *PLoS ONE* 12(9): e0185075.
- Zhang, J., Zhang, S., Cheng, M., Jiang, H., Zhang, X., Peng, C., Lu, X., Zhang, X. and Jin, J. (2018). Effect of Drought on Agronomic Traits of Rice and Wheat: A Meta-Analysis. *International Journal of Environment Research Public Health* 15: 839.
- Zhao, X., Schmidl, C., Suzuki, T., Ntini, E., Arner, E., Valen, E., Li, K., Schwarzfischer, L. and Zhu, J.K. (2002). Salt and drought stress signal transduction in plants. *Annual Review of Plant Physiology and Plant Molecular Biology* 53: 247-273.
- Zhu, J., Lee, B.H., Dellinger, M., Cui, X., Zhang, C. and Wu, S. (2010). A cellulose synthase-like protein is required for osmotic stress to 1-erancein *Arabidopsis*. *The Plant Journal* 63: 128-140.
- Zhu, Y.N., Shi, D.Q., Ruan, M.B., Zhang, L.L., and Meng, Z.H. (2013). Transcriptome Analysis Reveals Crosstalk of Responsive Genes to Multiple Abiotic Stresses in Cotton (*Gossypium hirsutum L.*). *PLoS ONE* 8(11): e80218.
- Zulkarnain, W.M., Ismail, M.Z., Saud, H.M., Othman, R., Habib, S.H. and Kausar, H. (2013). Growth and yield response to water availability at different growth stages of rice. *Journal of Food, Agriculture and Environment* 2: 540-544.