



**UNIVERSITI PUTRA MALAYSIA**

***CLONING AND EXPRESSION ANALYSIS OF  
MANGANESE PEROXIDASE AND LACCASE TRANSCRIPTS FROM  
Ganoderma boninense PER71 IN RESPONSE TO DIFFERENT  
NITROGEN SOURCES, PHYTOHORMONES AND HYDROGEN  
PEROXIDE***

**HO PEI YIN**

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By

**HO PEI YIN**

**Thesis Submitted to the School of Graduate Studies,  
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**December 2019**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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**December 2019**

**Chairman : Prof. Ho Chai Ling, PhD**  
**Faculty : Biotechnology and Biomolecular Sciences**

Basal Stem Rot (BSR) is a serious disease caused by *Ganoderma* species. *Ganoderma boninense* produces lignin degrading enzymes (LDEs) that are able to degrade the lignin component of plant cell wall causing oil palms to rot and eventually collapse. The transcripts and expressions of LDEs including manganese peroxidase (MnP) and laccase (Lac) in *G. boninense* PER71 during oil palm-*Ganoderma* interaction have not been reported. Likewise, the effect of nitrogen sources in fertilizers, phytohormones and hydrogen peroxide on the growth and gene expression of *G. boninense* are unknown. Therefore, the objectives of this study were to clone the transcripts encoding these LDEs; to measure their gene expression in *G. boninense* PER71 treated with different nitrogen sources (ammonium sulphate, ammonium nitrate, sodium nitrate and potassium nitrate), phytohormones (jasmonic acid, JA and salicylic acid, SA) and hydrogen peroxide; and to evaluate the effect of different nitrogen sources on the *in vitro* growth of *G. boninense* and oil palm seedlings inoculated with *G. boninense*. The full-length cDNA of four MnPs and three Lacs were cloned from *G. boninense* by Rapid Amplification of cDNA Ends (RACE)-PCR and confirmed by sequence analysis. Real-time reverse transcription-PCR (qRT-PCR) analysis showed that only Unigene 6011 (MnP) from *G. boninense* was up-regulated by all nitrogen sources and hydrogen peroxide but down-regulated in JA treatment. Unigene 87 (MnP) showed up-regulation in *G. boninense* treated with JA. Unigene 35959 (MnP) of *G. boninense* was up-regulated by ammonium sulphate treatment, down-regulated by hydrogen peroxide and suppressed by sodium nitrate and SA. Meanwhile, Unigene 30636 (Lac) was up-regulated by SA; down-regulated by hydrogen peroxide and suppressed by ammonium sulphate, potassium nitrate and JA. Unigene 36023 (Lac) was up-regulated by JA and hydrogen peroxide while Unigene 90667 (Lac) was up-regulated by ammonium nitrate, JA, SA and hydrogen peroxide. The growth of *G. boninense* cultured on ammonium nitrate-containing Czapek-Dox agar was the fastest while the growth on sodium nitrate was the slowest based on the

measurement of radial mycelial diameter. The optical mycelial density of *G. boninense* cultured on ammonium nitrate was also denser than that of *G. boninense* cultured on sodium nitrate. However, the highest optical mycelial density was observed for *G. boninense* cultured on ammonium sulphate. On the other hand, *G. boninense*-infected oil palm seedlings treated with ammonium nitrate were the least infected; white mycelia were not observed at the basal region and root surface as compared to oil palm seedlings in other nitrogen treatments. Inoculated oil palm seedlings without additional nitrogen; treated with ammonium sulphate, sodium nitrate and potassium nitrate showed increased disease symptoms. The most serious disease symptoms were observed in oil palm seedlings without nitrogen supplement, followed by sodium nitrate and potassium nitrate, ammonium sulphate then ammonium nitrate. The results showed that ammonium nitrate is a preferable source of nitrogen for growth of *G. boninense* and could slow down BSR development. In conclusion, the study contributes to the basic understanding of the effects of different nitrogen sources, phytohormones and hydrogen peroxide on *G. boninense* and the expression of MnP and Lac, as well as the disease development of BSR in oil palm seedlings.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**PENGLONAN DAN ANALISIS PENGEKSPRESAN MANGANAN  
PEROKSIDASE DAN LAKASE DARIPADA *Ganoderma boninense* PER71  
YANG BERTINDAK BALAS TERHADAP SUMBER NITROGEN YANG  
BERBEZA, FITOHORMON DAN HIDROGEN PEROKSIDA**

Oleh

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Reput pangkal batang (BSR) merupakan sejenis penyakit yang membimbangkan disebabkan oleh spesies *Ganoderma*. *Ganoderma boninense* menghasilkan enzim pereput lignin (LDE) yang boleh mereputkan komponen lignin pada dinding sel kelapa sawit menyebabkan pereputan dan seterusnya keruntuhan. Transkrip dan pengekspresan LDE termasuk manganan peroksidase (MnP) dan lakase (Lac) daripada *G. boninense* PER71 dalam interaksi kelapa sawit-*Ganoderma* belum pernah dilaporkan. Begitu juga dengan kesan sumber nitrogen dalam baja, fitohormon dan hidrogen peroksida terhadap pertumbuhan dan ekspresi gen *G. boninense* yang tidak diketahui. Oleh itu, objektif kajian ini adalah untuk mengklonkan transkrip LDE; mengukur gen ekspresi mereka dalam *G. boninense* PER71 yang telah dirawat dengan sumber nitrogen yang berbeza (ammonium sulfat, ammonium nitrat, natrium nitrat dan kalium nitrat), fitohormone tumbuhan (asid jasmonik, JA dan asid salisilik, SA) dan hidrogen peroksida; serta menilaikan kesan sumber nitrogen yang berbeza ke atas pertumbuhan *G. boninense* secara *in vitro* dan anak benih kelapa sawit yang telah diinokulasi dengan *G. boninense*. cDNA lengkap empat MnP dan tiga Lac telah diklonkan daripada *G. boninense* dengan menggunakan *Rapid Amplification of cDNA Ends (RACE)-PCR* dan dikenalpasti dengan analisis jujukan. Analisis tindakbalas berantai polimerase-masa nyata (qRT-PCR) menunjukkan bahawa pengekspresan Unigene 6011 (MnP) telah dipertingkatkan oleh semua sumber nitrogen dan hidrogen peroksida tetapi diperturunkan dalam rawatan JA. Unigene 87 (MnP) hanya menunjukkan peningkatan pengekspresan gen dalam *G. boninense* yang dirawat JA. Gen pengekspresan Unigene 35959 (MnP) dalam *G. boninense* dipertingkatkan oleh ammonium sulfat, diturunkan oleh hidrogen peroksida dan ditindas oleh natrium nitrate dan SA. Sementara itu, pengekspresan Unigene 30636 (Lac) dipertingkatkan oleh SA; diturunkan oleh hidrogen peroksida dan ditindas oleh ammonium sulfat, kalium nitrat dan JA. Ekspresi Unigene 36023 (Lac) dipertingkatkan oleh JA dan hidrogen peroksida manakala ekspresi Unigene 90667 (Lac) dipertingkatkan dalam ammonium nitrate, JA, SA dan hidrogen peroksida. Berdasarkan pengukuran diameter jejari, pertumbuhan *G. boninense* adalah paling cepat di atas agar Czapek-Dox

yang mengandung ammonium nitrat manakala pertumbuhan atas natrium nitrat adalah paling lambat. Kepadatan optikal mycelium *G. boninense* di atas media ditambah ammonium nitrat adalah lebih tinggi daripada *G. boninense* yang ditumbuh atas media ditambah natrium nitrat. Walau bagaimanapun, kepadatan optikal mycelium *G. boninense* adalah paling tinggi untuk *G. boninense* yang ditumbuh atas media ditambah ammonium sulfat. Selain itu, anak benih kelapa sawit yang dijangkiti dengan *G. boninense* dan dirawat dengan ammonium nitrat menunjukkan jangkitan yang paling minima; micelium tidak didapati pada kawasan pangkal dan permukaan akar berbanding dengan anak benih kelapa sawit yang dirawat dengan sumber nitrogen yang lain. Anak benih kelapa sawit yang diinokulasi tanpa penambahan sumber nitrogen; yang dirawat dengan ammonium sulfat, natrium nitrat dan kalium nitrat menunjukkan peningkatan simptom penyakit. Simptom penyakit yang paling ketara didapati pada anak benih kelapa sawit yang tidak dibekalkan sumber nitrogen tambahan, diikuti oleh anak benih kelapa sawit yang dirawat natrium nitrat, kalium nitrat, ammonium sulfat dan ammonium nitrat. Penemuan kajian menunjukkan ammonium nitrat merupakan sumber nitrogen yang sesuai untuk pertumbuhan *G. boninense* dan boleh melambatkan perkembangan BSR. Kesimpulannya, penemuan kajian ini telah menyumbang terhadap pemahaman asas tentang kesan sumber nitrogen yang berbeza, fitohormon dan hidrogen peroksida terhadap *G. boninense* dan pengekspresan MnP dan Lac, serta perkembangan simptom penyakit BSR pada anak benih kelapa sawit.

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## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiii
<b>LIST OF FIGURES</b>	xv
<b>LIST OF ABBREVIATIONS</b>	xx
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	1
<b>2 LITERATURE REVIEW</b>	3
2.1 Oil palm	3
2.1.1 Oil palm disease and basal stem rot (BSR)	3
2.1.2 Management of BSR	4
2.2 White rot fungi and <i>Ganoderma boninense</i>	5
2.2.1 The role of <i>Ganoderma</i> spp. in disease development	6
2.3 Lignin	7
2.4 Lignin degrading enzymes (LDEs)	8
2.4.1 Lignin peroxidase (LiP)	9
2.4.2 Manganese peroxidase (MnP)	10
2.4.3 Laccase (Lac)	12
2.4.4 Applications of LDEs	13
2.5 Effect of nitrogen and carbon sources on LDEs	14
2.6 Role of phytohormones and hydrogen peroxide during interactions with fungal pathogens	15
2.7 Effects of phytohormones and hydrogen peroxide on LDEs	16
<b>3 CLONING AND SEQUENCE ANALYSIS OF TRANSCRIPTS ENCODING LIGNIN DEGRADING ENZYMES FROM <i>G. boninense</i> PER71</b>	18
3.1 Introduction	18
3.2 Materials and methods	18
3.2.1 Sequence analysis of partial transcript sequences encoding LDEs	18
3.2.2 Primer design	19
3.2.3 Preparation of culture media	19
3.2.4 Preparation of <i>G. boninense</i> PER71 culture	19
3.2.5 Total RNA extraction	19
3.2.6 DNase treatment	21
3.2.7 cDNA synthesis for SMART™ Rapid Amplification of cDNA Ends (RACE)	21

3.2.8	3'- and 5'-RACE-PCR	22
3.2.9	Gel purification and direct purification of PCR products	22
3.2.10	Cloning of purified PCR products	23
3.2.10.1	Ligation	23
3.2.10.2	Preparation of competent cells	23
3.2.10.3	Transformation	23
3.2.10.4	Colony PCR	24
3.2.11	Plasmid DNA extraction	24
3.2.12	Restriction enzyme digestion	24
3.2.13	Isolation of full-length transcript	25
3.2.14	Addition of dATP to PCR product	25
3.2.15	Sequence analysis	25
3.3	Results and discussion	26
3.3.1	Molecular cloning and sequence analysis of transcripts encoding MnP	27
3.3.1.1	Unigene 87	29
3.3.1.2	Unigene 6011	33
3.3.1.3	Unigene 35959	37
3.3.1.4	Unigene 67998	41
3.3.2	Molecular cloning and sequence analysis of transcripts encoding Lac	46
3.3.2.1	Unigene 30636	48
3.3.2.2	Unigene 36023 and Unigene 46830	52
3.3.2.3	Unigene 90667	60
3.3.2.4	Isolation of full-length sequence for Unigene 36023	63
3.3.3	Sequence analysis of cloned Unigenes	64
3.4	Conclusions	71
<b>4</b>	<b>EFFECTS OF NITROGEN SOURCES, PHYTOHORMONES AND HYDROGEN PEROXIDE ON THE GROWTH AND GENE EXPRESSION OF LIGNIN DEGRADING ENZYMES OF <i>Ganoderma boninense</i> PER71</b>	<b>72</b>
4.1	Introduction	72
4.2	Materials and methods	73
4.2.1	Preparation of culture media	73
4.2.2	Inoculation of fungus on Czapek-Dox agar and broth with different nitrogen sources	73
4.2.3	Inoculation of fungus in PDB with phytohormones and hydrogen peroxide	74
4.2.4	Quantitative reverse transcription PCR (qRT-PCR)	74
4.2.4.1	Primer design	74
4.2.4.2	RNA extraction, DNase treatment and first strand cDNA synthesis	74
4.2.4.3	Melt curve and standard curve analysis	75
4.2.4.4	Gene expression profiling of LDEs	76
4.2.5	Inoculation of oil palms with <i>G. boninense</i>	76

	in the presence of different nitrogen sources	
4.3	Results and discussion	77
4.3.1	Growth of fungus on Czapek-Dox Agar with different nitrogen sources	77
4.3.2	Gene expression analysis of transcripts encoding LDEs	81
4.3.2.1	Expression profiles of LDEs in <i>G. boninense</i> treated with different nitrogen sources, phytohormones and hydrogen peroxide	81
4.3.3	Effects of different nitrogen sources on BSR disease symptoms of oil palms treated with <i>G. boninense</i>	84
4.4	Conclusions	93
<b>5</b>	<b>SUMMARY, GENERAL CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH</b>	<b>94</b>
5.1	Recommendations for future research	95
<b>REFERENCES</b>		<b>97</b>
<b>APPENDICES</b>		<b>116</b>
<b>BIODATA OF STUDENT</b>		<b>124</b>

## LIST OF TABLES

Table		Page
2.1	LDEs produced by different species of white rot fungi.	9
2.2	Lignin peroxidase sequences retrieved from NCBI database.	10
2.3	MnP sequences retrieved from NCBI database.	12
2.4	White rot fungi Lac sequences retrieved from NCBI database.	13
2.5	Types of carbon and nitrogen source preferred for the highest LDE activities.	15
2.6	Preferred source of carbon and nitrogen for maximum mycelial growth in white rot fungi.	15
3.1	Primers for amplification of LDE transcripts by 3'- and 5'-RACE.	20
3.2	Universal sequencing primers used for sequencing.	25
3.3	Forward and reverse primers for the amplification of full-length sequence.	25
3.4	Concentration and quality of total RNA and DNase-treated RNA.	27
3.5	List of Unigenes subjected to blastn searched against NCBI database.	28
3.6	The putative identity of the Unigenes subjected to blastn searched against NCBI database.	47
3.7	Summary of the sequences obtained from RACE-PCR for all selected Unigenes.	65
3.8	Identities (%) among nucleotide sequences of Unigenes of MnP.	65
3.9	Identities (%) among protein sequences of Unigenes of MnP.	65
3.10	Identities (%) among nucleotide sequences of Unigenes of Lac.	65

3.11	Identities (%) among protein sequences of Unigenes of Lac.	66
4.1	Primers (forward and reverse) for quantitative real-time PCR.	75
4.2	Disease severity index (DSI) based on external signs and symptoms of oil palm seedlings treated with <i>G. boninense</i> (Abdullah, 2003).	77
4.3	One-way ANOVA of effect of different nitrogen sources on the radial mycelial diameter (cm) of <i>G. boninense</i> on Czapek-Dox agar over 7 dpi.	80





## LIST OF FIGURES

Figure		Page
2.1	Components of lignocellulose.	7
2.2	Basic units of lignin.	7
2.3	Oxidation of organic acid by MnP.	11
3.1	Agarose gel electrophoresis of total RNA extracted from <i>G. boninense</i> PER71 stained with ethidium bromide.	26
3.2	Sequence alignment of partial sequence of Unigene 87 and <i>G. applanatum</i> (Accession number: AB035734.1).	29
3.3	Agarose gel electrophoresis of the 3'-RACE-PCR amplicon for Unigene 87 encoding MnP amplified from <i>G. boninense</i> PER 71.	30
3.4	Gel-purified 3'-RACE-PCR product of Unigene 87.	30
3.5	Colony PCR for 3'-RACE-PCR product of Unigene 87.	31
3.6	Purified plasmid DNA and restriction enzyme analysis for 3'-RACE-PCR product of Unigene 87.	31
3.7	Nucleotide and amino acid sequence of Unigene 87 encoding for MnP obtained through 3'-RACE-PCR.	32
3.8	Multiple alignment of 3'-RACE-PCR of Unigene 87 with other Unigenes.	33
3.9	Agarose gel electrophoresis of 5'-RACE-PCR amplicon for Unigene 6011 encoding MnP amplified from <i>G. boninense</i> PER 71.	34
3.10	Gel-purified 5'-RACE-PCR product of Unigene 6011.	34
3.11	Colony PCR for 5'-RACE-PCR product of Unigene 6011.	35
3.12	Purified plasmid DNA and restriction enzyme analysis for 5'-RACE-PCR product of Unigene 6011.	35
3.13	Nucleotide and amino acid sequence of Unigene 6011 encoding for MnP.	36

3.14	Agarose gel electrophoresis of 3'- and 5'-RACE-PCR amplicons for Unigene 35959 encoding MnP amplified from <i>G. boninense</i> PER 71.	37
3.15	Gel-purified 3'- and 5'-RACE-PCR products of Unigene 35959.	38
3.16	Colony PCR for 3'- and 5'-RACE-PCR products of Unigene 35959.	38
3.17	Purified plasmid DNA and restriction enzyme analysis by using <i>Hind</i> III for 3'- and 5'-RACE-PCR products of Unigene 35959.	39
3.18	Consensus nucleotide and amino acid sequence of Unigene 35959 encoding for MnP obtained through 3'-RACE-PCR and 5'-RACE-PCR.	40
3.19	Agarose gel electrophoresis of 3'- and 5'-RACE-PCR amplicon for Unigene 67998 encoding MnP amplified from <i>G. boninense</i> PER 71.	41
3.20	Gel-purified 3'- and 5'-RACE-PCR products of Unigene 67998.	42
3.21	Colony PCR for 3'- and 5'-RACE-PCR products of Unigene 67998.	42
3.22	Purified plasmid DNA and restriction enzyme analysis by using <i>Hind</i> III for 3'- and 5'-RACE-PCR product of Unigene 67998.	43
3.23	Consensus nucleotide and amino acid sequence of Unigene 67998 encoding for MnP obtained through 3'- and 5'-RACE-PCR.	44
3.24	The similarity of nucleotide sequence of 5'-RACE-PCR product of Unigene 6011 and consensus sequence of 3'- and 5'-RACE-PCR product of Unigene 67998 is 96.1%.	45
3.25	The similarity of amino acid sequence (ORF) of 5'-RACE-PCR product of Unigene 6011 and consensus sequence of 3'- and 5'-RACE-PCR products of Unigene 67998 is 99.7%.	46
3.26	Pairwise alignment of partial sequence of Unigene 30636 and <i>G. lucidum</i> (Accession number: DQ914872.1).	48
3.27	Agarose gel electrophoresis of 3'-RACE-PCR amplicon for Unigene 30636 encoding Lac amplified from <i>G. boninense</i> PER 71.	49

3.28	Gel-purified 3'-RACE PCR product of unigene 30636.	49
3.29	Colony PCR for 3-RACE-PCR product of Unigene 30363.	50
3.30	Purified plasmid DNA and restriction enzyme analysis for 3'-RACE-PCR product of Unigene 30636.	50
3.31	Nucleotide and amino acid sequence of Unigene 30636 encoding for Lac obtained through 3'-RACE-PCR.	51
3.32	Multiple alignment of 3'-RACE-PCR sequence of Unigene 30636 with other Unigene sequences.	52
3.33	Agarose gel electrophoresis of 3'-RACE-PCR amplicons for Unigene 36023 and Unigene 46830 encoding Lac amplified from <i>G. boninense</i> PER71.	53
3.34	Direct purified 3'-RACE-PCR product of Unigene 36023 and Unigene 46830.	53
3.35	Colony PCR for 3'-RACE-PCR products of Unigene 36023 and Unigene 46830.	54
3.36	Purified plasmid DNA and restriction enzyme analysis for 3'-RACE-PCR products of Unigene 36023 and Unigene 46830.	54
3.37	Comparison of the sequences of Unigene 36023, Unigene 46830, 3'-RACE-PCR transcripts for Unigene 36023 and Unigene 46830.	55
3.38	Agarose gel electrophoresis of 5'-RACE-PCR amplicon for Unigene 36023 encoding Lac amplified from <i>G. boninense</i> PER 71.	57
3.39	Gel-purified 5'-RACE-PCR product for Unigene 36023.	57
3.40	Colony PCR for 5'-RACE-PCR product of Unigene 36023.	58
3.41	Purified plasmid DNA and restriction enzyme analysis for 5'-RACE-PCR product of Unigene 36023.	58
3.42	Consensus nucleotide and amino acid sequence of Unigene 36023 encoding for Lac obtained through RACE-PCR.	59
3.43	Agarose gel electrophoresis of 5'-RACE-PCR amplicon for Unigene 90667 encoding Lac amplified from <i>G. boninense</i> PER 71.	60
3.44	Gel purified 5'-RACE-PCR products for Unigene 90667.	60
3.45	Colony PCR for 5'-RACE-PCR product of Unigene 90667.	61

3.46	Purified plasmid DNA and restriction enzyme analysis for 5'-RACE-PCR product of Unigene 90667.	61
3.47	Nucleotide and amino acid sequence of Unigene 90667 encoding for Lac obtained through 5'-RACE-PCR.	62
3.48	Agarose gel electrophoresis of full-length PCR product for Unigene 36023 amplified from cDNA of <i>G. boninense</i> PER71.	63
3.49	Purified PCR products for Unigene 36023.	63
3.50	Colony PCR for randomly selected positive clones.	64
3.51	(a) Purified plasmid DNA and (b) Restriction enzyme analysis for Unigene 36023.	64
3.52	Multiple alignment of MnP protein sequences from <i>G. boninense</i> , <i>G. lucidum</i> (Accession number: AMK79470.1) and <i>G. applanatum</i> (Accession number: BAA88392.1).	67
3.53	The phylogenetic tree of deduced amino acid sequences of MnP from <i>G. boninense</i> PER 71 and other fungal MnP from white-rot fungi.	68
3.54	Multiple alignment of Lac protein sequences from <i>G. boninense</i> , <i>G. lucidum</i> (Accession number: AHA83584.1) and <i>G. weberianum</i> (Accession number: ANA53145.1).	69
3.55	The phylogenetic tree of deduced amino acid sequences of Lac from <i>G. boninense</i> PER 71 and other fungal Lac from white-rot fungi.	70
4.1	Effect of different nitrogen sources on <i>in vitro</i> mycelial density and hyphal extension of <i>G. boninense</i> on Czapek-Dox Agar. <i>G. boninense</i> had the highest optical mycelial density on medium with ammonium sulphate, and the highest radial mycelial diameter at 7 dpi on medium with ammonium nitrate. Scale bar represents 1 cm.	78
4.2	Changes of radial mycelial diameter over 7 dpi of <i>G. boninense</i> on Czapek-Dox agar with different inorganic nitrogen sources. <i>G. boninense</i> had the fastest mycelial growth on medium with ammonium nitrate.	79
4.3	Relative expression of selected Unigenes of LDEs in <i>G. boninense</i> in response to different nitrogen sources (upper panel), phytohormones (lower panel) and hydrogen peroxide (lower panel).	82

4.4	Comparison of <i>Ganoderma</i> -inoculated and uninoculated oil palm seedlings at 22 wpi.	86
4.5	Root mass of oil palm seedlings inoculated with <i>G. boninense</i> PER71 and treated with 50 mM of different inorganic nitrogen sources (lower panel) compared to control oil palm seedlings under the same treatments (upper panel) at 24 wpi.	87
4.6	Dissected root of oil palm seedlings inoculated with <i>G. boninense</i> PER71 and treated with 50 mM of different inorganic nitrogen sources (lower panel) compared to control oil palm seedlings under the same treatments (upper panel) at 24 wpi.	88
4.7	Formation of white mycelia on the basal region of oil palm seedlings inoculated with <i>G. boninense</i> under treatments of different inorganic nitrogen sources except ammonium nitrate.	90
4.8	Root condition of oil palm seedlings treated with different nitrogen source and inoculated with <i>G. boninense</i> at 24 wpi. White mycelia is not observed on the <i>Ganoderma</i> -inoculated roots under ammonium nitrate treatment.	91

## LIST OF ABBREVIATIONS

AAO	Aryl alcohol oxidase
ANOVA	Analysis of variance
Avr	Avirulence
°C	Degree Celsius
BLAST	Basic local alignment search tool
bp	Base pair
BSA	Bovine serum albumin
BSR	Basal stem rot
CAZys	Carbohydrate active enzymes
cDNA	Complementary deoxyribonucleic acid
CDS	Coding sequence
cm	Centimeter
CWDEs	Cell wall degrading enzymes
dATP	Deoxyadenosine triphosphate
DBQH <sub>2</sub>	Hydroquinone
DEPC	Diethyl pyrocarbonate
DMRT	Duncan's Multiple Range Test
DNA	Deoxyribonucleic acid
DNase	Deoxyribonuclease
dNTP	Deoxyribonucleic triphosphate
dpi	Days post inoculation
DSI	Disease severity index
DTT	Dithiothreitol
EDTA	Ethylenediaminetetraacetic acid

ELISA	Enzyme-linked immunosorbent assay
EPR	Electron paramagnetic resonance
EtBr	Ethidium bromide
G	Guaiacyl
g	Gram
GSPs	Gene specific primers
G-type	Guaiacyl type
HCL	Hydrochloric acid
H <sub>2</sub> O <sub>2</sub>	Hydrogen peroxide
HR	Hypersensitive response
IPTG	Isopropyl β-D-1-thiogalactopyranoside
JA	Jasmonic acid
L	Liter
Lac	Laccase
LB	Luria-Bertani broth
LDEs	Lignin degrading enzymes
LiP	Lignin peroxidase
M	Molarity
MEA	Malt extract agar
min	Minutes
mg	Milligram
ml	Milliliter
mm	Millimeter
mM	Millimolar
Mn <sup>2+</sup>	Manganese
MnP	Manganese peroxidase

MPOB	Malaysian Palm Oil Board
NaCl	Sodium chloride
NCBI	National centre for biotechnology information
ng	Nanogram
NPK	Nitrogen, phosphorus and potassium
NUP	Nested universal primer A
OD	Optical density
ORF	Open reading frame
PCR	Polymerase chain reaction
PCI	Phenol:Chloroform:Isoamyl Alcohol (25:24:1, v/v)
PDA	Potato dextrose agar
PDB	Potato dextrose broth
POPW	Paddy and oil palm wood
qRT-PCR	Quantitative real-time polymerase chain reaction
R	Resistance
R <sup>2</sup>	Correlation coefficient squared
RACE	Rapid amplification of cDNA ends
RNA	Ribonucleic acid
ROS	Reactive oxygen species
Rpm	Revolutions per minute
RT	Reverse transcription
S	Syringyl
s	Seconds
SA	Salicylic acid
SDS	Sodium dodecyl sulphate
SPSS	Statistical package for the social sciences



S-type	Syringyl type
TAE	Tris-Acetate-EDTA
Tm	Melting temperature
U	Unit
Mg	Microgram
μl	Microliter
UPM	Universal primer A mix
UPM	University Putra Malaysia
USR	Upper stem rot
UTR	Untranslated region
UV	Ultraviolet
Vol	Volume
VP	Versatile peroxidase
wpi	Week post-inoculation
w/v	Weight per volume
X-gal	5-bromo-4-chloro-3-indolyl-β-D-galactopyranoside

## CHAPTER 1

### INTRODUCTION

Agriculture is an important sector to the sustenance of daily life and economic system of a country. Being an efficient oil crop, oil palm (*Elaeis guineensis* Jacq.), a tropical perennial tree has been planted in many countries including Malaysia. Oil palm was first brought to Malaya by British in 1870s as an ornamental plant which then turned into a commercial plant in the agriculture sector. Palm oil has gained the fastest growing demand globally because it serves as an ingredient in many products including food, cosmetic and bioenergy (Ferdous Alam *et al.*, 2015). In Malaysia, the latest oil palm land coverage has reached 5.84 million hectares in 2018 giving rise to 17.16 tonnes/hectare of fresh fruit bunch (FFB) and 3.42 tonnes/hectare of palm oil (Malaysian Palm Oil Board, 2019a; Malaysian Palm Oil Board, 2019b; Malaysian Palm Oil Board, 2019c). Palm oil production is threatened by stem rot caused by *Ganoderma boninense*, leading to major yield loss (Tisné *et al.*, 2017).

*Ganoderma boninense* is a deadly white-rot basidiomycete which inflicts death to oil palm trees. White-rot basidiomycetes fully digest lignin while leaving behind white cellulose (Paterson, 2007). The major disease caused by *G. boninense* is Basal Stem Rot (BSR), a common recurrent disease in oil palm plantations (Ariffin *et al.*, 2000) when oil palm trees are replanted. The disease progresses slowly without obvious symptoms during initial stage. Fruiting body appeared at a later stage of infection whereby the transportation of water and nutrient supply are greatly destructed (Ahmadi *et al.*, 2017). Successful penetration of pathogenic fungi through cuticle and cell wall into host cells is the key to pathogenicity (An *et al.*, 2018). Cell wall degrading enzymes (CWDEs) are needed by plant pathogenic fungi to penetrate plant cell wall (Kubicek *et al.*, 2014).

Lignin degrading enzymes (LDEs) are a subset of CWDEs specialize in depolymerisation of lignin. LDEs include LiP, MnP and Lac. Ho *et al.* (2016) found that LDE genes for MnPs and Lacs were up-regulated when oil palm was infected. There are many factors that can affect LDE secretion such as ratio of carbon to nitrogen, pH, temperature and mediators (Asgher *et al.*, 2016b). The production of LDEs from white-rot basidiomycetes was found to be suppressed upon application of nitrogen fertilizer (Magill and Aber, 1998).

Soil fertility is important in ensuring plant biomass production. The macronutrients of soil are nitrogen, phosphorus and potassium (Emamgholizadeh *et al.*, 2017). Over time, soil fertility declines with continuous cropping. This often resulted in increased usage of fertilizers to restore soil fertility and supply nutrients for plant growth. Insufficient nitrogen in plants increases susceptibility of plants to pathogens (Snoeijers *et al.*, 2000). In addition, the forms of nitrogen (ammonium or nitrate) also affect pathogenesis (Huber and Watson, 1974). In the case of tobacco, nitrate induces resistance and ammonium compromises resistance host plant against hemibiotrophic pathogen (Gupta *et al.*, 2013).

Other than nutrient availability, phytohormones also affect pathogenicity (Ma and Ma, 2016). Salicylic acid (SA) is a phenolic compound produced by plants, and its biosynthesis and signaling pathway have been well characterized, demonstrating its important role as a signal involved in the plant defense against biotrophic pathogens (De Coninck *et al.*, 2015). Jasmonic acid (JA) is a signalling molecule in plant defense against necrotrophic pathogens (De Coninck *et al.*, 2015). *Ganoderma boninense* is a hemibiotroph, having both biotrophic and necrotrophic lifestyles. Thus, SA and JA may affect the pathogenicity of *G. boninense*. It was found that SA affects the secretion and enzyme activities of LDEs of *G. boninense* (Surendran *et al.*, 2018). Surendran *et al.* (2018) also stated that phenolic compounds can be inhibitors for LDEs. However, literatures on the effect of SA and JA on gene expression for LDEs in *G. boninense* are not available.

To date, there is no efficient ways for BSR management despite many approaches have been taken (Ahmadi *et al.*, 2017). Studies on the effect of different nitrogen sources and phytohormones on the gene expression of LDEs might provide insights into the relationship between oil palm and *G. boninense* during infection. It is hoped that this information can help to lessen the severity of the oil palm disease in the future. This could be achieved by applying suitable nitrogen sources for growth of oil palm while limiting the oil palm disease.

The specific objectives of this study were:

1. To clone transcripts encoding fungal lignin degrading enzymes (LDEs) from *G. boninense* PER71;
2. To measure fungal growth and expression of genes for LDEs of *G. boninense* treated with different nitrogen sources, phytohormones and hydrogen peroxide;
3. To measure plant growth and disease symptoms of oil palm seedlings infected with *G. boninense* under treatment of different nitrogen sources.

## REFERENCES

- Abadulla, E., Tzanov, T., Costa, S., Robra, K., Cavaco-Paulo, A. and Gubitz, G.M. 2000. Decolorization and Detoxification of Textile Dyes with a Laccase from *Trametes hirsuta*. *Applied and Environmental Microbiology*, 66: 3357-3362.
- Abdullah, F., Ilias, G.N.M., Nelson, M., Mohd-Zainudin, N.A.I., Umi Kalsom, Y. 2003. Disease assessment and the efficacy of *Trichoderma* as a biocontrol agent of basal stem rot of oil palm. *Science Putra Research Bulletin*, 11: 31-33.
- Achyuthan, K.E., Achyuthan, A.M., Adams, P.D., Dirk, S.M., Harper, J.C., Simmons, B.A. and Singh, A.K. 2010. Supramolecular self-assembled chaos: Polyphenolic lignin's barrier to cost-effective lignocellulosic biofuels. *Molecules*, 15: 8641-8688.
- Ahmadi, P., Muharam, F.M., Ahmad, K., Mansor, S. and Abu Seman, I. 2017. Early detection of *Ganoderma* basal stem rot of oil palms using artificial neural network spectral analysis. *Plant Disease*, 101: 1009-1016.
- Akyilmaz, E., Yorganci, E. and Asav, E. 2010. Do copper ions activate tyrosine enzyme? Abiosensor model for the solution. *Bioelectrochemistry*, 78: 155-160.
- Alexander, A., Sipaut, C.T., Dayou, J. and Chong, K.P. 2017. Oil palm roots colonisation by *Ganoderma boninense*: An insight study using scanning electron microscopy. *Journal of Oil Palm Research*, 29: 262-266.
- Amara, S., Perrot, T., Navarro, D., Deroy, A. Benkhelfallah, A., Chalak, A., Daou, M., Chevret, D., Faulds, C.B., Berrin, J-G., Morel-Rouhier, M., Gelhaye, E. and Record, E. 2018. Enzyme activities of two recombinant heme-containing peroxidases, TvDyP1 and TvVP2, identified from the secretome of *Trametes versicolor*. *Applied and Environmental Microbiology*, 84: e02826-17.
- An, B., Wang, W., Guo, Y., Wang, Q., Luo, H. and He, C. 2018. BAS2 is required for conidiation and pathogenicity of *Colletotrichum gloeosporioides* from *Hevea brasiliensis*. *International Journal of Molecular Sciences*, 19: 1860.
- Ander, P. and Marzullo, L. 1997. Sugar oxidoreductases and veratryl alcohol oxidase as related to lignin degradation. *Journal of Biotechnology*, 53: 115-131.
- Ansari, Z., Karimi, A., Sedghi, S. and Razzaghi, M. 2017. Glucose oxidase effect on treatment of textile effluent containing reactive azo dyes by *Phanerochaete chrysosporium*. *Journal of Chemical Technology and Biotechnology*, 92: 1721-1726.
- Argyropoulos, D.S. and Menachem, S.B., 1997. Lignin. In Eriksson, K.L. (Ed.), *Advances in Biochemical Engineering Biotechnology* (pp. 127-158). Germany, Springer.

- Ariffin, D., Idris, A.S. and Singh, G. 2000. Status of *Ganoderma* in oil palm. In Flood, J., Bridge, P.D. and Holderness, M. (Eds.), *Ganoderma Disease of Perennial Crops* (pp.49-68). London: CABI Publishing.
- Armenteros, J.J.A., Tsirigos, K.D., Sonderby, C.K., Petersen, T.N., Winther, O., Brunak, S., Heijne, G.V. and Nielsen, H. 2019. SignalP 5.0 improves signal peptide predictions using deep neural networks. *Nature Biotechnology*, 37: 420-423.
- Arora, D.S. and Sharma, R.K. 2010. Ligninolytic fungal laccases and their biotechnological applications. *Applied Biochemistry and Biotechnology*, 160: 1760-1788.
- Asgher, M., Shah, S.A.H. and Iqbal, H.M.N. 2016a. Statistical correlation between ligninolytic enzymes secretion and remazol brilliant yellow-3GL dye degradation potential of *Trametes versicolor* IBL-04. *Water Environment Research*, 88: 338–345.
- Asgher, M., Wahab, A., Bilal, M. and Iqbal, H.M.N. 2016b. Lignocellulose degradation and production of lignin modifying enzymes by *Schizophyllum commune* IBL-06 in solid-state fermentation. *Biocatalysis and Agricultural Biotechnology*, 6: 195-201.
- Auclair, S.M., Bhanu, M.K. and Kendall, D.A. 2012. Signal peptidase 1: Cleaving the way to mature proteins. *Protein Science*, 21: 13-25.
- Bajpai, P. 1999. Application of enzymes in the pulp and paper industry. *Biotechnology Progress*, 15: 147-157.
- Barba-Espín, G., Diaz-Vivancos, P., Job, D., Belghazi, M., Job, C. and Hernández, J.A. 2011. Understanding the role of H<sub>2</sub>O<sub>2</sub> during pea seed germination: A combined proteomic and hormone profiling approach. *Plant Cell Environment*, 34: 1907-1919.
- Barcelos, E., Rios, S.A., Cunha, R.N.V., Lopes, R., Motoike, S.Y., Babychuk, E., Skirycz, A. and Kushnir, S. 2015. Oil palm natural diversity and the potential for yield improvement. *Frontiers in Plant Science*, 6: 190.
- Bellettini, M.B., Fiorda, F.A., Maievas, H.A., Teixeira, G.L., Avila, S., Hornung, P.S., Junior, A.M. and Ribani, R.H. 2016. Factors affecting mushroom *Pleurotus* spp. *Saudi Journal of Biological Sciences*, 2016: 1-14.
- Bellincampi, D., Cervone, F. and Lionetti, V. 2014. Plant cell wall dynamics and wall-related susceptibility in plant-pathogen interactions. *Frontiers in Plant Science*, 5: 228.
- Berg, B. and McLaugherty, C. 2014. Decomposer organisms. In *Plant litter: decomposition, humus formation, carbon sequestration* (pp.35–52). Berlin, Heidelberg: Springer Berlin Heidelberg.

- Bivi, M.S.H.R., Paiko, A.S., Khairulmazmi, A., Akhtar, M.S. and Idris, A.S. 2016. Control of basal stem rot disease in oil palm by supplementation of calcium, copper, and salicylic acid. *The Plant Pathology Journal*, 32: 396-406.
- Blackman, L.M., Cullerne, D.P., Torreña, P., Taylor, J. and Hardham, A.R. 2015. RNA-seq analysis of the expression of genes encoding cell wall degrading enzymes during infection of Lupin (*Lupinus angustifolius*) by *Phytophthora parasitica*. *PLoS One*, 10.
- Blodig, W., Doyle, W.A., Smith, A.T., Winterhalter, K., Choinowski, T. and Piontek, K. 1998. Autocatalytic formation of a hydroxy group at C beta of trp171 in lignin peroxidase. *Biochemistry*, 37: 8832-8838.
- Bower, N.I. and Johnston, I.A. 2010. Targeted rapid amplification of cDNA ends (TRACE)-an improved RACE reaction through degradation of non-target sequences. *Nucleic Acid Research*, 38: e194.
- Bradshaw, R.E., Guo, Y., Sim, A.D., Kabir, M.S., Chettri, P., Ozturk, I.K., Hunziker, L., Ganley, R.J. and Cox, M.P. 2016. Genome-wide gene expression dynamics of the fungal pathogen *Dothistroma septosporum* throughout its infection cycle of the gymnosperm host *Pinus radiata*. *Molecular Plant Pathology*, 17: 210-224.
- Buswell, J.A., Cai, Y. and Chang, S.T. 1995. Effect of nutrient nitrogen and manganese on manganese peroxidase and laccase production by *Lentinula* (*Lentinus*) *edodes*. *FEMS Microbiology Letters*, 128: 81-88.
- Caarls, L., Pieterse, C.M.J. and Wees, S.C.M.V. 2015. How salicylic acid takes transcriptional control over jasmonic signalling. *Frontier Plant Science*, 6: 170.
- Cairney, J.W.G. 2005. Basidiomycete mycelia in forest soils: Dimensions, dynamics and roles in nutrient distribution. *Mycological Research*, 109: 7-20.
- Champagne, P.P., Nesheim, M.E. and Ramsay, J.A. 2013. A mechanism for NaCl inhibition of reactive blue 19 decolorization and ABTS oxidation by laccase. *Applied Microbiology and Biotechnology*, 97: 6263-6269.
- Chen, B., Xu, W.Q., Pan, X.R. and Lu, L. 2015. A novel non-blue laccase from *Bacillus amyloliquefaciens*: Secretory expression and characterization. *International Journal of Biological Macromolecules*, 76: 39-44.
- Chong, K.P., Atong, M. and Rossall, S. 2012. The role of syringic acid in the interaction between oil palm, and *Ganoderma boninense*, the causal agent of basal stem rot. *Plant Pathology*, 61: 953-963.
- Chong, K.P., Dayou, J. and Alexander, A. 2017. *Detection and Control of Ganoderma boninense in Oil Palm Crop*. Cham, Switzerland: Springer Nature.
- Chung, G.F. 2011. Management of *Ganoderma* disease in oil palm plantations. *Planter*, 87: 325-339.

- Coelho-Moreira, J. da S., Brugnari, T., Sá-Nakanishi, A.B., Castoldi, R., de Souza, C.G.M., Bracht, A. and Peralta, R.M. 2018. Evaluation of diuron tolerance and biotransformation by white white-rot fungus *Ganoderma lucidum*. *Fungal Biology*, 122: 471-478.
- Coleman, J., Inukai, M. and Inouye, M. 1985. Dual functions of the signal peptide in protein transfer across the membrane. *Cell Press*, 43: 351-360.
- Corley, R.H.V. 2009. How much palm oil do we need?. *Environmental Science and Policy*, 12: 134-139.
- Corley, R.H.V. and Tinker, P.B. 2003. *The Oil Palm* (4<sup>th</sup> ed.). New Jersey, USA: Wiley.
- Corley, R.H.V. and Tinker, P.B. 2016. *The Oil Palm* (5<sup>th</sup> ed.). New Jersey, USA: Wiley.
- Dai, X.D., Zhan, Y.G., Zhang, J.C., Zhang, P.Q., Han, Z.H., Ma, Q.F., Kong, X.H., Liu, J.N. and Ma, Y.P. 2015. Regulatory effect of salicylic acid and methyl jasmonate supplementation on ergosterol production in *Hericium erinaceus* mycelia. *Journal of Forestry Research*, 26: 71-77.
- De Coninck, B., Timmermans, P., Vos, C., Cammue, B.P.A. and Kazan, K. 2015. What lies beneath: Belowground defense strategies in plants. *Trends in Plant Science*, 20: 91-101.
- Deeba, F., Pruthi, V. and Negi, Y.S. 2017. Effect of emerging contaminants from paper mill industry into the environment and their control. In Gupta, T., Agarwal, A., Agarwal, R. and Labhsetwar, N. (Eds.), *Environmental Contaminants, Energy, Environment, and Sustainability* (pp.391-408). Singapore: Springer.
- Di, X.T., Takken, F.L.W and Tintor, N. 2016. How phytohormones shape interactions between plants and the soil-borne fungus *Fusarium oxysporum*. *Frontiers in Plant Science*, 7: 170.
- Dietrich R., Ploss K. and Heil, M. 2004. Constitutive and induced resistance to pathogens in *Arabidopsis thaliana* depends on nitrogen supply. *Plant, Cell and Environment*, 27: 896-906.
- D'Souza, T. M., Merritt, C. S. and Reddy, C. A. 1999. Lignin modifying enzymes of the white rot basidiomycete *Ganoderma lucidum*. *Applied and Environmental Microbiology*, 65: 5307-5313.
- Eggert, C., Temp, U. and Eriksson, K.E. 1996. The ligninolytic system of the white rot fungus *Pycnoporus cinnabarinus*: Purification and characterization of the laccase. *Applied and Environmental Microbiology*, 62: 1151-1158.
- Elisashvili, V., Kachlishvili, E. and Penninckx, M. 2008. Effect of growth substrate, method of fermentation, and nitrogen source on lignocellulose-degrading enzymes production by white-rot basidiomycetes. *Journal of Industrial Microbiology and Biotechnology*, 35: 1531-1538.

- Emamgholizadeh, S., Shahsavani, S. and Eslami, M.A. 2017. Comparison of artificial neural networks, geographically weighted regression and Cokriging methods for predicting the spatial distribution of soil macronutrients (N, P and K). *Chinese Geographical Science*, 27: 747-759.
- Erden, E., Ucar, M.C., Gezer, T. and Pazarlioglu, N.K. 2009. Screening for ligninolytic enzymes from autochthonous fungi and applications for decolorization of remazole marine blue. *Brazilian Journal of Microbiology*, 40: 346-353.
- Falade, A.O., Nwodo, U.U., Iweriebor, B.C., Green, E., Mabinya, L.V. and Okoh, A.I. 2016. Lignin peroxidase functionalities and prospective applications. *Microbiology Open*, 6: e00394.
- Ferdous Alam, A.S.A., Er, A.C. and Begum, H. 2015. Malaysia oil palm industry: Prospect and problem. *Journal of Food, Agriculture and Environment*, 13: 143-148.
- Fernandez, J., Marroquin-Guzman, M. and Wilson, R.A. 2014. Mechanisms of nutrient acquisition and utilization during fungal infections of leaves. *Annual Review of Phytopathology*, 52: 155-174.
- Fleige, S. and Pfaffl, M.W. 2006. RNA integrity and the effect on the real-time qRT-PCR performance. *Molecular Aspects of Medicine*, 27: 126-139.
- Flor, H.H. 1942. Inheritance of pathogenicity in *Melampsora lini*. *Phytopathology*, 32: 653 - 669.
- Francoz, E., Ranocha, P., Nguyen-Kim, H., Jamet, E., Burlat, V. and Dunand, C. 2015. Roles of cell wall peroxidases in plant development. *Phytochemistry*, 112: 15-21.
- Freedman, Z. and Zak, D.R. 2014. Atmospheric N deposition increases bacterial laccase-like multicopper oxidases: Implications for organic matter decay. *Applied and Environmental Microbiology*, 8: 4460-4468.
- Fujii, K., Uemura, M., Hayakawa, C., Funakawa, S. and Kosaki, T. 2012. Environmental control of lignin peroxidase, manganese peroxidase, and laccase activities in forest floor layers in humid Asia. *Soil Biology and Biochemistry*, 57: 109-115.
- Gallagher, S. 1998. Quantitation of nucleic acids with absorption spectroscopy. *Current Protocols in Protein Science*, 13: A.4K.1-A.4K.3.
- Giardina, P., Faraco, V., Pezzella, C., Piscitelli, A., Vanhulle, S. and Sannia, G. 2010. Laccases: A never ending story. *Cellular and Molecular Life Sciences*, 67: 369-385.
- Glazebrook, J. 2005. Contrasting mechanisms of defense against biotrophic and necrotrophic pathogens. *Annual Review of Phytopathology*, 43: 205-227.



- Goh, K.M., Ganeson, M. and Supramaniam, C.V. 2014. Infection potential of vegetative incompatible *Ganoderma boninense* isolates with known ligninolytic enzyme production. *African Journal of Biotechnology*, 13: 1056-1066.
- Goh, Y.K., Marzuki, N.F., Tan, S.Y., Tan, S.S., Tung, H.J., Goh, Y.K. and Goh, K.J. 2016. Experimental mixture design as a tool to optimize the growth of various *Ganoderma* species cultivated on media with different sugars. *Mycology*, 7: 36-44.
- Gold, M.H. and Alic, M. 1993. Molecular biology of the lignin-degrading basidiomycete *Phanerochaete chrysosporium*. *Microbiology and Molecular Biology Reviews*, 57: 605-622.
- Gómez-Toribio, V., Garcia-Martin, A.B., Martinez, M.J., Martinez, A.T. and Guillen, F. 2009. Enhancing the production of hydroxyl radicals by *Pleurotus eryngii* via quinone redox cycling for pollutant removal. *Application and Environmental Microbiology*, 75: 3954-3962.
- Gupta, K.J., Brotman, Y., Segu, S., Zeier, T., Zeier, J., Persijn, S.T., Cristescu, S.M., Harren, F.J.M., Bauwe, H., Fernie, A.R., Kaiser, W.M. and Mur, L.A.J. 2013. The form of nitrogen nutrition affects resistance against *Pseudomonas syringae* pv. *phaseolicola* in tobacco. *Journal of Experimental Botany*, 64: 553-568.
- Hall, T.A. 1999. BioEdit: A user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Oxford University Press*, 41: 95-98.
- Hatfield, R. and Vermerris, W. 2001. Lignin formation in plants: The dilemma of linkage specificity. *Plant Physiology*, 126: 1351-1357.
- Hatti-Kaul, R. and Ibrahim, V. 2013. Lignin degrading enzymes: An overview. In Yang, S.T., El-Enshasy, H.A. and Thongchul, N. (Eds.), *Bioprocessing Technologies in Biorefinery for Sustainable Production of Fuels, Chemicals, and Polymers* (pp.167-192). New Jersey, USA: Wiley AIChE.
- Hibi, M., Hatahira, S., Nakatani, M., Yokozeki, K., Shimizu, S. and Ogawa, J. 2012. Extracellular oxidases of *Cerrena* sp complementarily functioning in artificial dye decolorization including laccase, manganese peroxidase, and novel versatile peroxidases. *Biocatalyst and Agricultural Biotechnology*, 1: 220-225.
- Ho, C.L., Tan, Y.C., Yeoh, K.A., Ghazali, A.K., Yee, W.Y., and Hoh, C.C. 2016. *De novo* transcriptome analyses of host-fungal interactions in oil palm (*Elaeis guineensis* Jacq.). *BMC Genomics*, 17: 66.
- Ho, Y.W. and Nawawi, A. 1985. *Ganoderma boninense* Pat. From basal stem rot of oil palm (*Elaeis guineensis*) in Peninsular Malaysia. *Pertanika*, 8: 425-428.
- Hoa, H.T. and Wang, C.L. 2014. The effects of temperature and nutritional conditions on mycelium growth of two oyster mushrooms (*Pleurotus ostreatus* and *Pleurotus cystidiosus*). *Mycobiology*, 43: 14-23.

- Holzbaun, E. and Tien, M. 1988. Structure and regulation of a lignin peroxidase gene from *Phanerochaete chrysosporium*. *Biochemical and Biophysical Research Communications*, 155: 626–633.
- Hosseinzadeh, P., Mirts, E.N., Pfister, T.D., Gao, Y.G., Mayne, C., Robinson, H., Tajkhorshid, E. and Lu, Y. 2016. Enhancing Mn(II)-binding and manganese peroxidase activity in a designed cytochrome C peroxidase through fine-tuning secondary-sphere interactions. *Biochemistry*, 55: 1494-1502.
- Huang, S.T., Tzean, S.S., Tsai, B.Y. and Hsieh, H.J. 2009. Cloning and heterologous expression of a novel ligninolytic peroxidase gene from poroid brown-rot fungus *Antrodia cinnamomea*. *Microbiology*, 155: 424-433.
- Huber, D.M. and Watson, R.D. 1974. Nitrogen form and plant disease. *Annual Review of Phytopathology*, 12:139-165.
- Hushiarian, R., Yusof, N.A. and Dutse, S.W. 2013. Detection and control of *Ganoderma boninense*: Strategies and perspectives. *SpingerPlus*, 2: 555.
- Hussin, M. H., Rahim, A.A., Ibrahim, M.N.M., Perrin, D. and Brosse, N. 2015. Enhanced properties of oil palm fronds (OPF) lignin fractions produced via tangential ultrafiltration technique. *Industrial Crops and Products*, 66: 1-10.
- Idris, A., Kushairi, A., Ismail, S. and Ariffin, D. 2004. Selection for partial resistance in oil palm progenies to *Ganoderma* basal stem rot. *Journal of Oil Palm Research*, 16: 12-18.
- Isaac, I. L. Walter, A.W.C.Y. Abu Bakar, M.F., Idris, A.S., Abu Bakar, F.D. Bharudin, I. and Abdul Murad, A.M. 2018. Transcriptome datasets of oil palm pathogen *Ganoderma boninense*. *Data in Brief*, 17: 1108-1111.
- Isaac, S. 1991. *Fungal-plant interactions*. United Kingdom: Chapman & Hall.
- Janda, M. and Ruelland, E. 2014. Magical mystery tour: Salicylic acid signaling. *Environmental and Experimental Botany*, 114: 117-128.
- Janusz, G., Kucharzyk, K.H., Pawlik, A., Staszczak, M. and Paszczynski, A.J. 2013. Fungal laccase, manganese peroxidase and lignin peroxidase: Gene expression and regulation. *Enzyme Microbiology Technology*, 52: 1-12.
- Janusz, G., Pawlik, A., Sulej, J., Swiderska-Burek, U., Jarosz-Wilkolazka, A. and Paszczynski, A. 2017. Lignin degradation: Microorganisms, enzymes involved, genomes analysis and evolution. *FEMS Microbiology Reviews*, 41: 941-962.
- Järvinen, J., Taskila, S., Isomäki, R. and Ojamo, H. 2012. Screening of white-rot fungi manganese peroxidases: A comparison between the specific activities of the enzyme from different native producers. *AMB Express*, 2: 62.

- Jayasinghe, C., Imtiaj, A., Lee, G.W., Im, K.H., Hur, H., Lee, M.W., Yang, H.S. and Lee, T.S. 2008. Degradation of three aromatic dyes by white rot fungi and the production of ligninolytic enzymes. *Mycobiology*, 36: 114-120.
- Jones, L.H. and Hughes, W.A. 1989. Oil palm (*Elaeis guineensis* Jacq.). *Biotechnology in Agriculture and Forestry*, 5: 176-202.
- Joo, S.S., Ryu, I.W., Park, J.K., Yoo, Y.M., Lee, D.H., Hwang, K.W., Choi, H.T., Lim, C.J., Lee, D.I. and Kim, K. 2008. Molecular cloning and expression of a laccase from *Ganoderma lucidum*. *Molecules and Cells*, 25: 112-118.
- Kaal, E.E.J., Jong, E.D. and Field, J.A. 1993. Stimulation of ligninolytic peroxidase activity by nitrogen nutrients in the white rot fungus *Bjerkandera* sp. strain BOS55. *Applied and Environmental Microbiology*, 59: 4031-4036.
- Karimi, A., Mahdizadeh, F., Salari, D. and Niaei, A. 2011. Bio-deoxygenation of water using glucose oxidase immobilized in mesoporous MnO<sub>2</sub>. *Desalination*, 275: 148-153.
- Kastelein, P., Slobbe, W.G.V. and Leeuw, G.T.N.D. 1990. Symptomatological and histopathological observations on oil palms from Brazil and Ecuador affected by fatal yellowing. *Netherlands Journal of Plant Pathology*, 96: 113-117.
- Kazan, K. and Lyons, R. 2014. Intervention of phytohormone pathways by pathogen effectors. *The Plant Cell*, 26: 2285-2309.
- Kirk, T.K. and Farrell, R.L. 1987. Enzymatic “combustion”: The microbial degradation of lignin. *Annual Review of Microbiology*, 41: 465-505.
- Kersten, P. and Cullen, D. 2007. Extracellular oxidative systems of the lignin-degrading basidiomycete *Phanerochaete chrysosporium*. *Fungal Genetics and Biology*, 44: 77-87.
- Kong, W., Chen, H., Lyu, S., Ma, F.Y., Yu, H.B. and Zhang, X. Y. 2016. Characterization of a novel manganese peroxidase from white-rot fungus *Echinodontium taxodii* 2538, and its use for the degradation of lignin-related compounds. *Process Biochemistry*, 51: 1776-1783.
- Koyani, R.D., Sanghvi, G.V., Sharma, K.R. and Rajput, K.S. 2013. Contribution of lignin degrading enzymes in decolourisation of reactive textile dyes. *International Biodeterioration & Biodegradation*, 77: 1-9.
- Kubicek, C.P. 2013. Fungi and lignocellulosic biomass. New York, USA: Wiley.
- Kubicek, C.P., Starr, T.L. and Glass, N.L. 2014. Plant cell wall-degrading enzymes and their secretion in plant-pathogenic fungi. *Annual Review of Phytopathology*, 52: 427-451.

- Kües, U., Nelson, D.R., Liu, C., Yu, G.J., Zhang, J.H., Li, J.Q., Wang, X.C. and Sun, H. 2015. Genome analysis of medicinal *Ganoderma* spp. with plant-pathogenic and saprotrophic life-styles. *Phytochemistry*, 114: 18-37.
- Kunjadia, P.D., Sanghvi, G.V., Kunjadia, A.P., Mukhopadhyay, P.N. and Dave, G.S. 2016. Role of ligninolytic enzymes of white rot fungi (*Pleurotus* spp.) grown with azo dyes. *SpringerPlus*, 5: 1487.
- Lee, M.W., Qi, M. and Yang, Y.N. 2001. A novel jasmonic acid-inducible rice *myb* gene associates with fungal infection and host cell death. *The American Phytopathological Society*, 14: 527-535.
- Leonowicz, A., Matuszewska, A., Luterek, J., Ziegenhagen, D., Wojtaś-Wasilewska, M., Cho, N.S., Hofrichter, M. and Rogalski, J. 1999. Biodegradation of lignin by white rot fungi. *Fungal Genetics and Biology*, 27: 175-185.
- Lim, F.H., Nor Fakhrana, I., Abdul Rasid, O., Idris, A.S., Ahmad Parveez, G.K., Ho, C.L. and Shaharuddin, N.A. 2014. Isolation and selection of reference genes for *Ganoderma boninense* gene expression study using quantitative real-time PCR (qPCR). *Journal of Oil Palm Research*, 26: 170-181.
- Lombard, V., Ramulu, H.G., Drula, E., Coutinho, P.M. and Henrissat, B. 2014. The carbohydrate-active enzymes database (CAZy) in 2013. *Nucleic Acids Research*, 42: D490-D495.
- Long, D.H., Lee F.N. and TeBeest D.O. 2000. Effect of nitrogen fertilization on disease progress of rice blast on susceptible and resistant cultivars. *Plant Disease*, 84: 403-409.
- López-Berges, M.S., Rispaíl, N., Prados-Rosales, R.C. and Pietro, A.D. 2010. A nitrogen response pathway regulates virulence functions in *Fusarium oxysporum* via the protein kinase TOR and the bZIP protein MeaB. *The Plant Cell*, 22: 2459-2475.
- Lundell, T., Mäkelä, M. R. and Hildén, K. 2010. Lignin-modifying enzymes in filamentous basidiomycetes – ecological, functional and phylogenetic review. *Journal of Basic Microbiology*, 50: 5-20.
- Maciel, M.J., Silva, A.C. and Ribeiro, H.C. 2010. Industrial and biotechnological applications of ligninolytic enzymes of the basidiomycota: A review. *Electronic Journal of Biotechnology*, 13.
- Ma, K.W. and Ma, W.B. 2016. Phytohormone pathways as targets of pathogens to facilitate infection. *Plant Molecular Biology*, 91: 713-725.
- Magill, A.H. and Aber, J.D. 1998. Long-term effects of experimental nitrogen additions on foliar litter decay and humus formation in forest ecosystems. *Plant Soil*, 203: 301-311.
- Mäkelä, M.R., Lundell, T., Hatakka, A. and Hildén, K. 2013. Effect of copper, nutrient nitrogen, and wood-supplement on the production of lignin-modifying enzymes by the white-rot fungus *Phlebia radiata*. *Fungal Biology*, 117: 62-70.

- Malaysian Palm Oil Board. 2019a. *Oil palm planted area by state at December 2018 (hectares)*. Retrieved 17 January 2019 from [http://bepi.mpob.gov.my/images/area/2018/Area\\_summary.pdf](http://bepi.mpob.gov.my/images/area/2018/Area_summary.pdf)
- Malaysian Palm Oil Board. 2019b. *Monthly FFB yield (tonnes/hectare)*. Retrieved 21 January 2019 from [http://bepi.mpob.gov.my/images/Yield/Yield-2018/FFB\\_Yield\\_July-December\\_2018.pdf](http://bepi.mpob.gov.my/images/Yield/Yield-2018/FFB_Yield_July-December_2018.pdf)
- Malaysian Palm Oil Board. 2019c. *Monthly crude palm oil yield (tonnes/hectares)*. Retrieved 21 January 2019 from [http://bepi.mpob.gov.my/images/Yield/Yield-2018/Oil\\_Yield\\_July-December\\_2018.pdf](http://bepi.mpob.gov.my/images/Yield/Yield-2018/Oil_Yield_July-December_2018.pdf)
- Malinovsky, F.G., Fangel, J.U. and Willats, W.G.T. 2014. The role of the cell wall in plant immunity. *Frontiers in Plant Science*, 5: 178.
- Manavalan, T., Manavalan, A. and Heese, K. 2014. Characterization of Lignocellulolytic Enzymes from White-Rot Fungi. *Current Microbiology*, 70, 485–498.
- Manavalan, T., Manavalan, A., Thangavelu, K.P. and Heese, K. 2013. Characterization of optimized production, purification and application of laccase from *Ganoderma lucidum*. *Biochemical Engineering Journal*, 70: 106-114.
- Marschnert, H., Kirkby, E.A. and Engels, C. 1997. Importance of cycling and recycling of mineral nutrients within plants for growth and development. *Botanica Acta*, 110: 265-273.
- Martínez, A.T., Speranza, M., Ruiz-Dueñas, F.J., Ferreira, P., Camarero, S., Guillén, F., Martínez, M.J., Gutiérrez, A. and del Río, J.C. 2005. Biodegradation of lignocellulosics: Microbial, chemical, and enzymatic aspects of the fungal attack of lignin. *International Microbiology*, 8: 195-204.
- Martinez, D., Larrondo, L.F., Putnam, N., Gelpke, M.D.S., Huang, K., Chapman, J., Helfenbein, K.G., Ramaiya, P., Detter, J.C., Larimer, F., Coutinho, P.M., Henrissat, B., Berka, R., Cullen, D. and Rokhsar, D. 2004. Genome sequence of the lignocellulose degrading fungus *Phanerochaete chrysosporium* strain RP78. *Nature Biotechnology*, 22: 695-700.
- Marzluf, G.A. 1997. Genetic regulation of nitrogen metabolism in the fungi. *Microbiology and Molecular Biology Reviews*, 61: 17–32.
- Mayer, A.M. 2006. Pathogenesis by fungi and by parasitic plants: Similarities and differences. *Phytoparasitica*, 34: 3-16.
- Mayer, A.M. and Staples, R.C. 2002. Laccase: New functions for an old enzyme. *Phytochemistry*, 60: 551-565.
- McMullan, G., Meehan, C., Conneely, A., Robinson, N.K., Nigam, P., Banat, I.M., Marchant, R. and Symth, W.F. 2001. Microbial decolorisation and degradation of textile dyes. *Applied Microbiology and Biotechnology*, 56: 81-87.

- Mepsted, R., Flood, J. and Cooper, R.M. 1995. Fusarium wilt of oil palm II. Stunting as a mechanism to reduce water stress. *Physiological and Molecular Plant Pathology*, 46: 373-387.
- Mercière, M., Boulord, R., Carasco-Lacombe, C., Klopp, C., Lee, Y.P., Tan, J.S., Alwee, S.S.R.S., Zaremski, A., De Franqueville, H., Breton, F. and Camus-Kulandaivelu, L. 2017. About *Ganoderma boninense* in oil palm plantations of Sumatra and peninsular Malaysia: Ancient population expansion, extensive gene flow and large scale dispersion ability. *Fungal Biology*, 121: 529–540.
- Mester, T. and Field, J.A. 1998. Characterization of a novel manganese peroxidase-lignin peroxide hybrid isozyme produced by *Bjerkandera* species strain BOS55 in the absence of manganese. *The Journal of Biological Chemistry*, 273: 15412-15417.
- Miles, P.G. and Chang, S.T. 1997. *Mushroom biology concise basics and current developments*. World Scientific, Singapore.
- Mitchell, C.E., Reich, P.B., Tilman, D. and Groth, J.V. 2003. Effects of elevated CO<sub>2</sub>, nitrogen deposition, and decreased species diversity on foliar, fungal plant disease. *Global Change Biology*, 9: 438-451.
- Mohammed, C.L., Rimbawanto, A. and Page, D.E. 2014. Management of basidiomycete root- and stem-rot diseases in oil palm, rubber and tropical hardwood plantation crops. *Forest Pathology*, 44: 428-446.
- Mohammed, M.A.A., Salmiaton, A., Wan Azlina, W.A.K.G., Mohammad Amran, M.S., Fakhru'l-Razi, A. and Taufiq-Yap, Y.H. 2011. Hydrogen rich gas from oil palm biomass as a potential source of renewable energy in Malaysia. *Renewable and Sustainable Energy Reviews*, 15: 1258-1270.
- Mohan, D. 2018. *Effects of herbicides, hydrogen peroxide, and phytohormones on Ganoderma infection in oil palm (Elaeis guineensis Jacq.) roots* (Unpublished master's thesis). Universiti Putra Malaysia, Selangor, Malaysia.
- Mohd-Zainudin, N.A.I. and Faridah, A. 2008. Disease suppression in *Ganoderma*-infected oil palm seedlings treated with *Trichoderma harzianum*. *Plant Protection Science*, 44: 101-107.
- Monteiro, M.C. and De Carvalho, M.E.A. 1998. Pulp bleaching using laccase from *Trametes versicolor* under high temperature and alkaline conditions. *Applied Biochemistry and Biotechnology*, 70–72: 983-993.
- Moreno-Chacón, A.L., Camperos-Reyes, J.E., Diazgranados, R.A.A. and Romero, H.M. 2013. Biochemical and physiological responses of oil palm to bud rot caused by *Phytophthora palmivora*. *Plant Physiology and Biochemistry*, 70: 246-251.
- Morozova, O.V., Shumakovich, G.P., Gorbacheva, M.A., Shleev, S.V. and Yaropolov, A.I. 2007. “Blue” laccases. *Biochemistry*, 72: 1136–1150.

- Nawawi, A. and Ho, Y.W. 1990. Effect of temperature and pH on growth pattern of *Ganoderma boninense* from oil palm in Peninsular Malaysia. *Pertanika*, 13: 303-307.
- Niladevi, K.N. 2009. Ligninolytic enzymes. In Nigam, P.S.N. and Pandey, A. (Eds.), *Biotechnology for Agro-Industrial Residues Utilisation* (pp.397-414). Berlin, Germany: Springer.
- Nousiainen, P., Kontro, J., Manner, H., Hatakka, A. and Sipilä, J. 2014. Phenolic mediators enhance the manganese peroxidase catalyzed oxidation of recalcitrant lignin model compounds and synthetic lignin. *Fungal Genetics and Biology*, 72: 137-149.
- Nunes, C.S. and Kunamneni, A. 2018. Laccases: Properties and applications. *Enzymes in Human and Animal Nutrition*, 2018: 133-161.
- Olsson, S. 1995. Mycelial density profiles of fungi on heterogeneous media and their interpretation in terms of nutrient reallocation patterns. *Mycological Research*, 99: 143-153.
- Ong, E., Pollock, W.B. and Smith, M. 1997. Cloning and sequence analysis of two laccase complementary DNAs from the ligninolytic basidiomycete *Trametes versicolor*. *Gene*, 196: 113-119.
- Osma, J.F., Saravia, V., Herrera, J.L.T. and Couto, S.R. 2007. Mandarin peelings: The best carbon source to produce laccase by static cultures of *Trametes pubescens*. *Chemosphere*, 67: 1677-1680.
- Palma, C., Moreira, M.T., Feijoo, G. and Lema, J.M. 1997. Enhanced catalytic properties of MnP by exogenous addition of manganese and hydrogen peroxide. *Biotechnology Letters*, 19: 263-267.
- Papaspyridi, L. M., Katapodis, P., Gonou-Zagou, Z., Kapsanaki-Gotsi, E. and Christakopoulos, P. 2011. Growth and biomass production with enhanced beta-glucan and dietary fibre contents of *Ganoderma australe* ATHUM 4345 in a batch-stirred tank bioreactor. *Engineering in Life Sciences*, 11: 65-74.
- Paterson, R.R.M. 2007. *Ganoderma* disease of oil palm-A white rot perspective necessary for integrated control. *Crop Protection*, 26: 1369-1376.
- Paterson, R.R.M. 2019a. *Ganoderma boninense* disease deduced from simulation modelling with large data sets of future Malaysian oil palm climate. *Phytoparasitica*, 47: 255-262.
- Paterson, R.R.M. 2019b. *Ganoderma boninense* disease of oil palm to significantly reduce production after 2050 in Sumatra if projected climate change occurs. *Microorganisms*, 7: 24.
- Paterson, R.R.M. and Lima, N. 2018. Climate change affecting oil palm agronomy, and oil palm cultivation increasing climate change, require amelioration. *Ecology and Evolution*, 8: 452-461.

- Paterson, R.R.M., Moen, S. and Lima, N. 2009. The feasibility of producing oil palm with altered lignin content to control *Ganoderma* disease. *Journal of Phytopathology*, 157: 649-656.
- Patrick, F., Mtui, G., Mshandete, A.M. and Kivaisi, A. 2011. Optimization of laccase and manganese peroxidase production in submerged culture of *Pleurotus sajorajju*. *African Journal of Biotechnology*, 10: 10166-10177.
- Pattathil, S., Hahn, M.G., Dale, B.E. and Chundawat, S.P.S. 2015. Insights into plant cell wall structure, architecture, and integrity using glycome profiling of native and AFECTM-pre-treated biomass. *Journal of Experimental Botany Advance Access*, 66: 4279-4294.
- Peralta, R.M., Silva, B.P., Corrêa, R.C.G., Kato, C.G., Seixas, F.A.V. and Bracht, A. 2017. Enzymes from basidiomycetes: Peculiar and efficient tools for Biotechnology. In Brahmachari, G., Demain, A.L. and Adrio, J.L. (Eds.). *Biotechnology of Microbial Enzymes: Production, biocatalysis and industrial applications* (pp.119-149). London: Academic Press.
- Pieterse, C.M.J., Zamioudis, C., Berendsen, R.L., Weller, D.M., Van Wees, S.C. and Bakker, P.A. 2014. Induced systemic resistance by beneficial microbes. *Annual Review of Phytopathology*, 52: 347-375.
- Pieterse, C.M.J., Van der Does, D., Zamioudis, C., Leon-Reyes, A. and Van Wees, S.C.M. 2012. Hormonal modulation of plant immunity. *Annual Review of Cell and Developmental Biology*. 28: 489-521.
- Piscitelli, A., Giardina, P., Lettera, V., Pezzella, C., Sannia, G. and Faraco, V. 2011. Induction and transcriptional regulation of laccases in fungi. *Current Genomics*, 12: 104-112.
- Pollegioni, L., Tonin, F. and Rosini, E. 2015. Lignin degrading enzymes. *The FEBS Journal*, 282, 1190-1213.
- Poulos, T.L. and Kraut, J. 1980. The stereochemistry of peroxidase catalysis. *The Journal of Biological Chemistry*, 255: 8199-8205.
- Prasher, I.B. and Chauhan, R. 2015. Effect of carbon and nitrogen sources on the growth, reproduction and ligninolytic enzymes activity of *Dictyoarthrinium Synnematum* Somrith. *Advances in Zoology and Botany*, 3: 24-30.
- Pribnow, D., Mayfield, M.B., Nipper, V.J., Brown, J.A. and Gold, M.H. 1989. Characterization of a cDNA encoding a manganese peroxidase, from lignin-degrading basidiomycete *Phanerochaete chrysosporium*. *The Journal of Biological Chemistry* 264: 5036-5040.
- Pryor, R.J. and Wittwer, C.T. Real-time polymerase chain reaction and melting curve analysis. *Methods in Molecular Biology*, 336: 19-32.



- Ramakers, C., Ruijter, J.M., Deprez, R.H.L. and Moorman, A.F.M. 2003. Assumption-free analysis of quantitative real-time polymerase chain reaction (PCR) data. *Neuroscience Letters*, 339: 62-66.
- Reddy, C.A. 1995. The potential for white rot fungi in the treatment of pollutants. *Current Opinion in Biotechnology*, 6: 320-328.
- Rees, R.W., Flood, J., Hasan, Y. and Cooper, R.M. 2007. Effects of inoculum potential, shading and soil temperature on root infection of oil palm seedlings by the basal stem rot pathogen *Ganoderma boninense*. *Molecular Plant Pathology*, 56: 862-870.
- Rees, R.W., Flood, J., Hasan, Y., Potter, U. and Cooper, R.M. 2009. Basal stem rot of oil palm (*Elaeis guineensis*): Mode of root infection and lower stem invasion by *Ganoderma boninense*. *Plant Pathology*, 58: 982-989.
- Rees, R.W., Flood, J., Hasan, Y., Wills, M.A. and Cooper, R.M. 2011. *Ganoderma boninense* basidiospores in oil palm plantations: Evaluation of their possible role in stem rots of *Elaeis guineensis*. *Plant Pathology*, 61: 567-578.
- Revankar, M.S. and Lele, S.S. 2006. Synthetic dye decolorization by white rot fungus, *Ganoderma* sp. WR-1. *Bioresource Technology*, 98: 775-780.
- Rinkes, Z.L., Bertrand, I., Amin, B.A.Z., Grandy, A.S., Wickings, K. and Weintraub, M.N. 2016. Nitrogen alters microbial enzyme dynamics but not lignin chemistry during maize decomposition. *Biogeochemistry*, 128: 171-186.
- Ritch, T.G., Nipper, V.J., Akileswaran, L., Smith, A.J., Pribnow, D.G. and Gold, M.H. 1991. Lignin peroxidase from the basidiomycete *Phanerochaete chrysosporium* is synthesized as a preproenzyme. *Gene*. 107: 119-126.
- Rodríguez-Rincón, F., Suarez, A., Lucas, M., Larrondo, L.F., de la Rubia, T, Polaina, J. and Martínez, J. 2010. Molecular and structural modelling of the *Phanerochaete flavidoalba* extracellular laccase reveals its ferroxidase structure. *Archives of Microbiology*, 192: 883-892.
- Roslan, A. and Idris, A.S. 2012. Economic impact of *Ganoderma* incidence on Malaysian Oil Palm Plantation – A case study in Johor. *Oil Palm Industry Economic Journal*, 13: 24-30.
- Ruiz-Deñás, F.J. and Martínez, A.T. 2009. Microbial degradation of lignin: How a bulky recalcitrant polymer is efficiently recycled in nature and how we take advantage of this. *Microbial Biotechnology*, 2: 164-177.
- Sahebi, M., Hanafi, M.M., Mohidin, H., Rafii, M.Y., Azizi, P., Idris, A.S., Fariz, A., Abiri, R., Taheri, S. and Moradpoor, M. 2018. Antioxidant enzyme activities and secondary metabolite profiling of oil palm seedlings treated with combination of NP fertilizers infected with *Ganoderma boninense*. *BioMed Research International*, 2018: 1-18.

- Saloheimo, M., Barajas, V., Niku-Paavola, M-L. and Knowles, J.K.C. 1989. A lignin peroxidase-encoding cDNA from the white-rot fungus *Phlebia radiata*: Characterization and expression in *Trichoderma reesei*. *Gene*, 85: 343-351.
- Sambrook, J., Fritsch, E.F. and Maniatis, T. 1989. *Molecular cloning: A laboratory manual* (2<sup>nd</sup> ed.). New York, USA: Cold Spring Harbor Laboratory Press.
- Sambrook, J. and Russell, D.W. 2006. Preparation and transformation of competent *E. coli* using calcium chloride. USA: Cold Spring Harbor Protocols.
- Sandhu, D.K. and Arora, D.S. 1985. Laccase production by *Polyporus sanguineus* under different nutritional and environmental conditions. *Experientia*, 41: 355–356.
- Sandrine, I., Graudens, E., Boulanger, V., Barlet, X., Zaborski, P., Eveno, E., Mueller, O., Schroeder, A. and Auffray, C. 2005. Towards standardization of RNA quality assessment using user-independent classifiers of microcapillary electrophoresis traces. *Nucleic Acids Research*, 33: e56.
- Santhanam, N, Vivanco, J.M., Decker, S.R. and Reardon, K.F. 2011. Expression of industrially relevant laccases: Prokaryotic style. *Trends in Biotechnology*, 29: 480–489.
- Sattler, S.E. and Funnell-Harris, D.L. 2013. Modifying lignin to improve bioenergy feedstocks: Strengthening the barrier against pathogens?. *Frontiers in Plant Science*, 4: 70.
- Schmidt, J.H. and Weidema, B.P. 2008. Shift in the marginal supply vegetable oil. *The International Journal of the Life Cycle Assessment*, 13: 235-239.
- Schneider, W.D.H., Fontana, R.C., Mendonça, S., Siqueira, F.G.D., Dillon, A.J.P. and Camassola, M. 2018. High level production of laccases and peroxidases from the newly isolated white-rot basidiomycete *Marasmiellus palmivorus* VE111 in a stirred-tank bioreactor in response to different carbon and nitrogen sources. *Process Biochemistry*, 69: 1-11.
- Sheikhi, F., Ardakani, M.R., Enayatizamir, N. and Rodriguez-Couto, S. 2012. The determination of assay for laccase of *Bacillus subtilis* WPI with two classes of chemical compounds as substrates. *Indian Journal of Microbiology*, 52: 701-707.
- Shetty, N.P., Mehrabi, R., Lütken, H., Haldrup, A., Kema, G.H., Collinge, D.B. and Jørgensen, H.J. 2007. Role of hydrogen peroxide during the interaction between the hemibiotrophic fungal pathogen *Septoria tritici* and wheat. *New Phytologist*, 174: 637-647.
- Shraddha, Shekher, R., Sehgal, S., Kamthania, M. and Kumar, A. 2011. Laccase: Microbial sources, production, purification, and potential biotechnological applications. *Enzyme Research*, 2011: 1-11.

- Sigoillot, J-C., Berrin, J-G., Bey, M., Lesage-Meessen, L., Levasseur, A., Lomascolo, A., Record, E. and Uzan-Boukhris, E. 2012. Fungal strategies for lignin degradation. In Jousnin, L. and Lapiere, C. (Eds.), *Advances in Botanical Research* (pp.263–308). London, UK: Academic Press, Elsevier.
- Sjöström, E. 1993. Wood chemistry, fundamentals and applications (2<sup>nd</sup> ed). New York, USA: Academic Press.
- Skyba, O., Douglas, C.J. and Mansfield, S.D. 2013. Syringyl-rich lignin renders poplars more resistant to degradation by wood decay fungi. *Applied and Environmental Microbiology*. 79: 2560–2571.
- Snoeijsers, S.S., Pérez-García, A., Joosten, M.H.A.J. and De Wit, P.J.G.M. 2000. The effect of nitrogen on disease development and gene expression in bacterial and fungal plant pathogens. *European Journal of Plant Pathology*, 106: 493–506.
- Sokolovsky, V., Kaldenhoff, R., Ricci, M. and Russo, V.E.A. 1990. Fast and reliable mini-prep RNA extraction from *Neurospora crassa*. *Fungal Genetics Reports*, 37: 27.
- Srivastava, P., Andersen, P.C., Marois, J.J., Wright, D.L., Srivastava, M. and Harmon, P.F. 2013. Effect of phenolic compounds on growth and ligninolytic enzyme production in *Botryosphaeria* isolates. *Crop Protection*, 43: 146-156.
- Stokland, J.N. 2012. Evolution of saproxylic organisms. In Stokland, J.N., Siitonen, J. and Jonsson, B.G. (Eds.), *Biodiversity in dead wood* (pp.218-247). England: Cambridge University Press.
- Suderman, R.J., Dittmer, N.T., Kanost, M.R. and Kramer, K.J. 2006. Model reactions for insect cuticle sclerotization: Cross-linking of recombinant cuticular proteins upon their laccase-catalyzed oxidative conjugation with catechols. *Insect Biochemistry and Molecular Biology*, 36: 353–365.
- Sundaramoorthy, M., Kishi, K., Gold, M.H. and Poulos, T.L. 1994. The crystal structure of manganese peroxidase from *Phanerochaete chrysosporium* at 2.06-Å resolution. *The Journal of Biological Chemistry*, 269: 32759-32767.
- Surendran, A., Siddiqui, Y., Saud, H.M., Ali, N.S. and Manickam, S. 2018. Inhibition and kinetic studies of lignin degrading enzymes of *Ganoderma boninense* by natural occurring phenolic compounds. *Journal of Applied Microbiology*, 125: 876-887.
- Susanto. A. 2009. Basal stem rot in Indonesia: Biology, economic importance, epidemiology, detection and control. In: *Proceedings of the International Workshop on Awareness, Detection and Control of Oil Palm Devastating Diseases, November 2009, Kuala Lumpur Convention Centre, Malaysia*. Serdang, Malaysia: Universiti Putra Malaysia Press.

- Susanto, A., Sudharto, P.S. and Purba, R.Y. 2005. Enhancing biological control of basal stem rot disease (*Ganoderma boninense*) in oil palm plantations. *Mycopathologia*, 159: 153-157.
- Sutherland, G.R.J., Zapanta, L.S., Tien, M. and Aust, S.D. 1997. Role of calcium in maintaining the heme environment of manganese peroxidase. *Biochemistry*, 36: 3654-3662.
- Suwannarach, N., Sujarit, K., Kumla, J. and Bussaban, B. 2013. First report of leaf spot disease on oil palm caused by *Pestalotiopsis theae* in Thailand. *Journal of General Plant Pathology*, 79: 277-279.
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M. and Kumar, S. 2011. MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Molecular Biology and Evolution*, 28: 2731-2739.
- Tan, Y.C., Yeoh, K.A., Wong, M.Y. and Ho, C.L. 2013. Expression profiles of putative defence-related proteins in oil palm (*Elaeis guineensis*) colonized by *Ganoderma boninense*. *Journal of Plant Physiology*, 170: 1455-1460.
- Thurston, C.F. 1994. The structure and function of fungal laccases. *Microbiology Society Journals*, 140: 19-26.
- Tien, M. and Kirk, T.K. 1988. Lignin peroxidase of *Phanerochaete chrysosporium*. In Wood, W.A. and Kellogg, S.T. (Eds.), *Methods in Enzymology: Biomass (Part B), Lignin, Pectin and Chitin* (pp.238-249). San Diego: Academic Press Inc.
- Tisné, S., Pomiès, V., Riou, V., Syahputra, I., Cochard, B. and Denis, M. 2017. Identification of *Ganoderma* disease resistance loci using natural field infection of an oil palm multiparental population. *Genes, Genomes, Genetics*, 7: 1683-1692.
- Tuomela, M., Vikman, M., Hatakka, A. and Itävaara, M. 2000. Biodegradation of lignin in a compost environment: A review. *Bioresource Technology*, 72: 169-183.
- Turner, P.D. 1981. Oil palm diseases and disorders. Kuala Lumpur: Oxford University Press.
- Turner, P.D. 1965. The incidence of *Ganoderma* disease of oil palm in Malaysia and its relation to previous crop. *Annals of Applied Biology*, 55: 417-423.
- Urzúa, U., Kersten, P.J. and Vicuña, R. 1998. Manganese peroxidase-dependent oxidation of glyoxylic and oxalic acids synthesized by *Ceriporiopsis subvermisporea* produces extracellular hydrogen peroxide. *Applied and Environmental Microbiology*, 64: 68-73.
- Usha, K.Y., Praveen, K. and Reddy, B.R. 2014. Enhanced production of ligninolytic enzymes by a mushroom *Stereum ostrea*. *Biotechnology Research International*, 2014: 1-9.

- Utomo, C., Werner, S., Niepold, F. and Deising, H.B. 2005. Identification of *Ganoderma*, the causal agent of basal stem rot disease in oil palm using a molecular method. *Mycopathologia*, 159: 159-170.
- Vanisa, D.V., Mustafaev, O.N., Moiseenko, K.V., Sadovskaya, N.S., Glazunova, O.A., Tyurin, A.A., Fedorova, T.V., Pavlov, A.R., Tyazhelova, T.V., Goldenkova-Pavlova, I.V. And Koroleva, O.V. 2015. The *Trametes hirsuta* 072 laccase multigene family: Genes identification and transcriptional analysis under copper ions induction. *Biochimie* 116: 154-164.
- Vares, T., Kalsi, M. and Hatakka, A. 1995. Lignin peroxidases, manganese peroxidases, and other ligninolytic enzymes produced by *Phlebia radiata* during solid-state fermentation of wheat straw. *Applied and Environmental Microbiology*, 61: 3515-3520.
- Vartiamäki, H., Majjala, P., Uotila, A. and Hantula, J. 2008. Characterization of growth and enzyme production of *Chondrostereum purpureum* isolates and correlation of these characteristics with their capability to prevent sprouting of birch in field. *Biological Control*, 47: 46-54.
- Vasiliadou, I.A., Molina, R., Pariente, M.I., Christoforidis, K.C., Martinez, F. and Melero, J.A. 2019. Understanding the role of mediators in the efficiency of advanced oxidation processes using white-rot fungi. *Chemical Engineering Journal*, 359: 1427-1435.
- Veresoglou, S.D., Barto, E.K., Menexes, G. and Rillig, M.C. 2013. Fertilization affects severity of disease caused by fungal plant pathogens. *Plant Pathology*, 62: 961-969.
- Vicuña, R. 2000. Ligninolysis. *Molecular Biotechnology*, 14: 173-176.
- Vijay, V., Pimm, S.L., Jenkins, C.N. and Smith, S.J. 2016. The impacts of oil palm on recent deforestation and biodiversity loss. *PLoS ONE*, 11: e0159668.
- Vos, A.M., Jurak, E., Pelkmans, J.F., Herman, K., Pels, G., Baars, J.J., Hendrix, E., Kabel, M.A., Lugones, L.G. and Wösten, H.A.B. 2017. H<sub>2</sub>O<sub>2</sub> as a candidate bottleneck for MnP activity during cultivation of *Agaricus bisporus* in compost. *AMB Express*, 7: 124.
- Walker, G. M. and White, N.A. 2017. Introduction to fungal physiology. In Kavanagh, K. (Ed.), *Fungi: Biology and applications* (pp.1-34). New Jersey, USA: Wiley Blackwell.
- Wang, Y.S., Thorup-Kristensen, K., Jensen, L.S. and Magid, J. 2016. Vigorous root growth is a better indicator of early nutrient uptake than root hair traits in spring wheat grown under low fertility. *Frontier in Plant Science*, 7: 865.
- Welinder, K.G., Mauro, J.M. and Norskov-Lauritsen, L. 1992. Structure of plant and fungal peroxidases. *Biochemistry Society Transactions*, 20: 337– 340.

- Wilson, R.A., Fernandez, J., Quispe, C.F., Gradnigo, J., Seng, A., Moriyama, E. and Wright, J.D. 2012. Towards defining nutrient conditions encountered by the rice blast fungus during host infection. *PLoS ONE*, 7: e47392.
- Wong, C.L., Bong, J.F.C. and Idris, A.S. 2012. *Ganoderma* species associated with basal stem rot disease of oil palm. *American Journal of Applied Sciences*, 9: 879-885.
- Wong, D.W. 2009. Structure and action mechanism of ligninolytic enzymes. *Applied Biochemistry and Biotechnology*, 157: 174-209.
- Wong, Y.X. and Yu, J. 1999. Laccase-catalyzed decolorization of synthetic dyes. *Water Research*, 33: 3512-3520.
- Xu, P., Ding, Z.Y., Qian, Z., Zhao, C.X. and Zhang, K.C. 2008. Improved production of mycelial biomass and ganoderic acid by submerged culture of *Ganoderma lucidum* SB97 using complex media. *Enzyme and Microbial Technology*, 42: 325-331.
- Yadav, S. and Chandra, R. 2015. Syntrophic co-culture of *Bacillus subtilis* and *Klebsiella pneumonia* for degradation of kraft lignin discharged from rayon grade pulp industry. *Journal of Environmental Sciences*, 33: 229-328.
- Yaver, D.S. and Golightly, E.J. 1996. Cloning and characterization of three laccase genes from white-rot basidiomycete *Trametes villosa*: Genomic organization of the laccase gene family. *Gene*, 181: 95-102.
- Yesilada, O., Birhanli, E. and Geckil, H. 2018. Bioremediation and decolorization of textile dyes by white rot fungi and laccase enzymes. In Prasad, R. (Ed.), *Mycoremediation and Environmental Sustainability* (pp.121-153). Cham, Switzerland: Springer.
- You, L.F., Liu, Z.M., Lin, J.F., Guo, L.Q., Huang, X.L. and Yang, H.X. 2014. Molecular cloning of a laccase gene from *Ganoderma lucidum* and heterologous expression in *Pichia pastoris*. *Journal of Basic Microbiology*, 54: 134-141.
- Youn, H.D., Hah, Y.C. and Kang, S.O. 1995. Role of laccase in lignin degradation by white-rot fungi. *FEMS Microbiology Letters*, 132: 183-188.
- Zamocky, M., Gasselhuber, B., Furtmuller, P.G. and Obinger, C. 2014 Turning points in the evolution of peroxidase-catalase superfamily: Molecular phylogeny of hybrid heme peroxidases. *Cellular Molecular Life Sciences*, 71: 4681-4696.
- Zhuo, R., Ma, L., Fan, F., Gong, Y., Wan, X., Jiang, M., Zhang, X. and Yang, T. 2011. Decolorization of different dyes by a newly isolated white-rot fungi strain *Ganoderma* sp.EN3 and cloning and functional analysis of its laccase gene. *Journal of Hazardous Materials*, 192: 855-873.