



UNIVERSITI PUTRA MALAYSIA

***PHYTOCHEMICAL CONTENT IN *Polyalthia bullata* King AND THE
EFFECT OF AUXINS, ELICITORS AND PRECURSORS ON TOTAL
ALKALOID CONTENT IN CALLUS***

MUNIRAH ADIBAH BINTI KAMARUL ZAMAN

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By

MUNIRAH ADIBAH BINTI KAMARUL ZAMAN

Thesis Submitted to School of Graduate Studies, Universiti Putra Malaysia, in
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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of
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Polyalthia bullata or locally known as Tongkat Ali Hitam is one of the species belongs to genus of *Polyalthia*. The plant has been reported to possess an ability to treat many diseases and enhance human health and life quality, which might be contributed by the presence of bioactive compounds. However, due to limited reports on phytochemical compounds present in *P. bullata*, the phytochemical profiling can help in clarifying the types of phytocompounds, therefore, choosing the right extraction solvent is important in order to get optimum yield. One of the factors that might affect the extraction yield is the polarity of extraction solvent. Apart from that, overcollection of *P. bullata* from wild habitat has become serious problem that may lead to species extinction. The establishment of *P. bullata* callus culture and addition of elicitors and precursors can help in enhancing the production of phytochemical compounds, therefore reducing the extinction of *P. bullata* from native habitat. Hence, the aims of this study were to determine the total alkaloid content (TAC), total phenolic content (TPC), total flavonoid content (TFC), and total terpenoid content (TTC) as well as antioxidant activity of hexanic, ethanolic, methanolic, and distilled water extracts of *P. bullata* root, leaf and stem, to profile biochemical compounds using gas chromatography- mass spectrometry (GC-MS), to induce callus from *P. bullata* using different explants, basal media, and plant growth regulators (PGRs), and to determine the effectiveness of auxins, precursors, and elicitors in enhancing alkaloid production in callus. For callus induction, the sterilized leaf and midrib explants were used and cultured on Murashige and Skoog (MS) and Woody Plant Medium (WPM) basal media supplemented with B5 vitamin, and different concentrations and types of auxins (2,4-dichlorophenoxyacetic acid (2,4-D), α -naphthaleneacetic acid (NAA), picloram and dicamba). The MS and WPM basal media supplemented with different types and concentrations (10, 20, 30, 40 50 μ M) of auxins (2,4-D, NAA, picloram, dicamba, indole-3-acetic acid (IAA), indole-3-butyric acid (IBA)) were used to determine the best multiplication medium and alkaloid content after six weeks of culture. The elicitors (methyl jasmonate (MeJA), salicylic acid (SA), and chitosan) and precursors (L-phenylalanine, L-tyrosine, and L-tryptophan) at concentration of 50, 100 and 150 μ M was respectively added into the best alkaloid

production medium to enhance the alkaloid production. The results from the studies revealed that the methanolic extract of *P. bullata* leaf exhibited the highest TPC, TFC, TTC and total antioxidant activity at 1042.52 ± 1.97 mg GAE/g DW, 80.88 ± 0.24 mg QE/g DW, 0.19 ± 0.00 mg LE/g DW and $85.19 \pm 1.16\%$, respectively. Meanwhile, the methanolic extract of *P. bullata* stem showed the highest TAC at 7.71 ± 0.00 mg AE/g DW. The fatty acids, phenolics, and carboxylic acid were found in methanolic stem extract; carbohydrates, alkaloids, and fatty acids were found in methanolic root extract; and terpenoids, phenolics, and alcohol were found in methanolic leaf extract of *P. bullata* using GC-MS. Among the media tested for *in vitro* callus induction, WPM basal medium supplemented with $16.56 \mu\text{M}$ picloram exhibited the highest callus induction percentage with $53.33 \pm 22.06\%$. As for the callus multiplication, the callus cultured on MS + $30 \mu\text{M}$ dicamba was found to significantly produce the highest fresh weight (1180.00 ± 159.43 mg FW) and dry weight (58.00 ± 6.66 mg DW) of callus after three weeks of culture. The addition of auxins into culture medium managed to enhance the alkaloid production in callus with the highest alkaloid content was observed in callus cultured on MS medium supplemented with $30 \mu\text{M}$ 2,4-D (31.07 ± 0.05 $\mu\text{g/g}$ DW). Among auxins, elicitors, and precursors tested, the MS + $30 \mu\text{M}$ 2,4-D and MS + $30 \mu\text{M}$ 2,4-D + $50 \mu\text{M}$ chitosan were found to be the best media for alkaloid production with the amount of 31.07 ± 0.05 and 31.30 ± 0.23 $\mu\text{g/mg}$ DW after six weeks of culture, respectively. As a conclusion, methanol was found to be the best extraction solvent to extract phytochemical compounds from *P. bullata*. The incorporation of auxins like 2,4-D into the culture medium is the best strategy to enhance alkaloid production in *P. bullata* callus. Therefore, the data obtained from this study can be used to further investigate biological activities of phytochemical compounds present in *P. bullata*, and therefore can reduce overcollection of this plant from the forest.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai
memenuhi keperluan untuk ijazah Ijazah Master Sains

**KANDUNGAN FITOKIMIA DALAM *Polyalthia bullata* King DAN KESAN
AUKSIN, PENGELISIT DAN PREKURSOR TERHADAP JUMLAH
KANDUNGAN ALKALOID DALAM KALUS**

Oleh

MUNIRAH ADIBAH BINTI KAMARUL ZAMAN

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Polyalthia bullata atau dikenali sebagai Tongkat Ali Hitam adalah salah satu spesies tergolong dalam genus *Polyalthia*. Tumbuhan ini dilaporkan mempunyai keupayaan untuk merawat pelbagai penyakit dan meningkatkan kesihatan dan kualiti hidup manusia, di mana ia mungkin disumbangkan oleh kehadiran sebatian bioaktif. Walau bagaimanapun, disebabkan oleh laporan yang terhad mengenai sebatian fitokimia yang terdapat dalam *P. bullata*, pemprofilan fitokimia dapat membantu dalam menjelaskan jenis-jenis fitosebatian, oleh itu, pemilihan pelarut pengekstrakan yang tepat adalah penting untuk mendapatkan hasil yang optimum. Salah satu faktor yang boleh menjelaskan hasil pengekstrakan ialah polariti pelarut pengekstrakan. Selain itu, pengambilan berlebihan *P. bullata* dari habitat liar telah menjadi masalah serius di mana boleh mengakibatkan kepupusan spesies. Penubuhan kultur kalus *P. bullata* dan penambahan pengelisit dan prekursor boleh membantu dalam meningkatkan pengeluaran sebatian fitokimia, seterusnya mengurangkan kepupusan *P. bullata* dari habitat asal. Oleh itu, matlamat kajian ini adalah untuk menentukan jumlah kandungan alkaloid (TAC), jumlah kandungan fenolik (TPC), jumlah kandungan flavonoid (TFC), dan jumlah kandungan terpenoid (TTC) serta aktiviti antioksidan ekstrak heksana, etanol, metanol, dan air suling dalam akar, daun, dan batang *P. bullata*, untuk memprofil sebatian biokimia menggunakan kromatografi gas-spektroskopi jisim (GC-MS), untuk menghasilkan kalus dari *P. bullata* menggunakan eksplan, media asas, dan pengawal atur pertumbuhan (PGRs) yang berbeza, dan untuk menentukan keberkesanan auksin, precursor, dan pengelisit dalam meningkatkan penghasilan alkaloid dalam kalus. Bagi induksi kalus, eksplan daun dan tulang daun yang telah disterilkan digunakan dan dikultur di atas media asas Murashige dan Skoog (MS) dan Woody Plant Medium (WPM) yang ditambah dengan vitamin B5, dan kepekatan dan jenis auksin yang berbeza (asid 2,4-diklorofenoksiasetik (2,4-D), asid α -naftalenaasetik (NAA), pikloram dan dikamba). Media asas MS dan WPM ditambah dengan jenis dan kepekatan (10, 20, 30, 40 50 μ M) auksin yang berbeza (2,4-D, NAA, pikloram, dikamba, asid indol-3-asetik (IAA), asid indol-3-butirik (IBA)) telah digunakan untuk menentukan media

penggandaan kalus dan kandungan alkaloid yang terbaik selepas enam minggu kultur. Pengelisit (metil jasmonat (MeJA), asid salisilik (SA), dan kitosan) dan prekursor (L-fenilalanina, L-tirosina, dan L-triptofan) masing-masing pada kepekatan 50, 100, 150 μM ditambah ke dalam media penghasilan alkaloid yang terbaik untuk meningkatkan pengeluaran alkaloid. Keputusan dari kajian ini menunjukkan bahawa ekstrak metanol daun *P. bullata* menunjukkan jumlah TPC, TFC, TTC dan jumlah aktiviti antioksidan tertinggi masing-masing pada $1042.52 \pm 1.97 \text{ mg GAE / g DW}$, $80.88 \pm 0.24 \text{ mg QE / g DW}$, $0.19 \pm 0.00 \text{ mg LE / g DW}$ dan $85.19 \pm 1.16\%$. Sementara itu, ekstrak metanol batang *P. bullata* menunjukkan TAC tertinggi pada $7.71 \pm 0.00 \text{ mg AE / g DW}$. Asid lemak, fenolik dan asid karboksilit telah dijumpai dalam ekstrak metanol batang; karbohidrat, alkaloid dan asid lemak telah dijumpai dalam ekstrak metanol akar; dan terpenoid, fenolik dan alkohol dijumpai dalam ekstrak metanol daun *P. bullata* menggunakan GC-MS. Antara media yang diuji untuk induksi kalus *in vitro*, media asas WPM yang ditambah dengan $16.56 \mu\text{M}$ pikloram menunjukkan peratusan induksi kalus tertinggi dengan $53.33 \pm 22.06\%$. Manakala bagi penggandaan kalus, kalus yang dikultur di atas media MS + 30 μM dikamba didapati dengan ketara menghasilkan kalus dengan berat segar ($1180.00 \pm 159.43 \text{ mg FW}$) dan berat kering ($58.00 \pm 6.66 \text{ mg DW}$) tertinggi pada minggu ketiga. Penambahan auksin dalam medium kultur berjaya meningkatkan penghasilan alkaloid dalam kalus dengan kandungan alkaloid tertinggi dihasilkan dalam media MS ditambah dengan 30 μM 2,4-D ($31.07 \pm 0.05 \mu\text{g / g DW}$). Antara auksin, pengelisit, dan prekursor yang diuji, MS + 30 μM 2,4-D dan MS + 30 μM 2,4-D + 50 μM kitosan merupakan media yang terbaik untuk penghasilan alkaloid masing-masing dengan jumlah 31.07 ± 0.05 and $31.30 \pm 0.23 \mu\text{g/mg DW}$ selepas enam minggu kultur. Kesimpulannya, metanol telah didapati menjadi pelarut pengekstrakan yang terbaik untuk mengekstrak sebatian fitokimia dalam *P. bullata*. Penggabungan auksin seperti 2,4-D ke dalam medium kultur adalah strategi terbaik untuk meningkatkan penghasilan alkaloid dalam kalus *P. bullata*. Oleh itu, data yang diperolehi daripada kajian ini boleh digunakan untuk mengetahui dengan lebih lanjut aktiviti biologi sebatian biokimia yang terdapat dalam *P. bullata* dan dengan itu dapat mengurangkan pengambilan tumbuhan ini dari hutan.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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4.13	Fresh weight (a) and Dry weight (b) of <i>P. bullata</i> callus grown on WPM medium at different concentrations of dicamba. Data represent as means \pm SE. The asterisk (*) represents a significant difference of treated callus with 1-week old callus grown on WPM 0 at $p \leq 0.05$ Tukey's range test.	74
4.14	Total alkaloid content of <i>P. bullata</i> callus grown on MS medium at different concentrations of (a) NAA, (b) IBA, (c) IAA, (d) 2,4-D, (e) picloram and (f) dicamba. Data represent as means \pm SE. The asterisk (*) represent a significant difference of treated callus with 1-week old callus grown on MSO at $p \leq 0.05$ Tukey's range test.	77
4.15	Total alkaloid content of <i>P. bullata</i> callus grown on WPM medium at different concentrations of (a) NAA, (b) IBA, (c) IAA, (d) 2,4-D, (e) picloram and (f) dicamba. Data represented as means \pm SE. The asterisk (*) represented as significant difference of treated callus with 1-week old callus grown on WPM 0 at $p \leq 0.05$ Tukey's range test.	82
4.16	Total alkaloid content of <i>P. bullata</i> callus grown on MSO, MS + 30 μ M 2,4-D and MS + 30 μ M 2,4-D supplemented with 50, 100, and 150 μ M of elicitors (MeJA, SA and chitosan) after six weeks of incubation. Treatment with MSO and 30 μ M 2,4-D served as controls. Data represent as means \pm SE. Different letters represent as significant difference at $p \leq 0.05$ Tukey's range test.	88
4.17	Total alkaloid content of <i>P. bullata</i> callus grown on MSO, MS + 30 μ M 2,4-D and MS + 30 μ M 2,4-D supplemented with 50, 100, and 150 μ M of precursors (phenylalanine, tyrosine and tryptophan) after six weeks of incubation. Data represent as means \pm SE. Different letters represent as significant difference at $p \leq 0.05$ Tukey's range test.	90

LIST OF ABBREVIATIONS

2,4-D	2,4-dichlorophenoxyacetic acid
AE	Atropine equivalent
Bi(NO ₃) ₃ .5H ₂ O	Bismuth nitrate pentahydrate
DPPH	2,2-Diphenyl-1-picrylhydrazyl
DR	Dragendorff's reagent
DW	Dry weight
FW	Fresh weight
GAE	Gallic acid equivalent
GC-MS	Gas chromatography – mass spectrometry
HCl	Hydrochloric acid
HNO ₃	Nitric acid
IAA	Indole-3-acetic acid
IBA	Indole-3-butyric acid
LE	Linalool equivalent
MeJA	Methyl jasmonate
MS	Murashige and Skoog
Na ₂ S	Disodium sulfide
NAA	α -Naphthaleneacetic acid
NaOH	Sodium hydroxide
PGRs	Plant growth regulators
QE	Quercetin equivalent
rpm	Revolutions per minute
SA	Salicylic acid
SE	Standard error
TAC	Total alkaloid content
TFC	Total flavonoid content
TPC	Total phenolic content
TTC	Total terpenoid content
UV-VIS	Ultraviolet - Visible
WPM	Woody plant medium

CHAPTER 1

INTRODUCTION

1.1 Introduction

Plant bioactive compounds can be defined as compounds that have biological activities and give positive and negative effects towards humans and animals (Saponjac *et al.*, 2016). These metabolites are produced when plants responding to biotic and abiotic stresses, and therefore, they are involved in the plant adaptation process (Ncube and Van, 2015; Ahmed *et al.*, 2017). Plant bioactive compounds can be classified into three major categories which are terpenes, alkaloids, and phenolics (Azmir *et al.*, 2013). Many studies have reported that the plant-derived antioxidant compounds are mainly from phenolic group like flavonoids, polyphenols and tannins (Cartea *et al.*, 2010; Nagavani *et al.*, 2010). They have capacity to combat the free radicals with minimal side effects on human (Patel *et al.*, 2012). According to Gangwar *et al.* (2014), the consumption of natural antioxidant can reduce the risk of cancer and many chronic diseases. Commonly, the solvent extraction is a method used for separation and isolation of plant bioactive compounds (Barchan *et al.*, 2014). The use of suitable solvent extraction is a key factor to extract maximum yield of the phytochemical and antioxidant compounds because the compounds have different characteristics and polarities that make them soluble in the solvent used (Fatiha *et al.*, 2012; Pham *et al.*, 2015).

Polyalthia bullata or locally known as Tongkat Ali Hitam is a medicinal plant belongs to genus *Polyalthia* and Annonaceae family. The plant is a shrub that can grow up to two- to three-meter height mainly in lowland of primary or secondary forest located in Peninsular Malaysia and Sabah. The genus *Polyalthia* has been reported to contain numerous types of bioactive compounds that are mostly derived from alkaloids, flavonoids, acetogenin and triterpenoids groups (Paarakh and Khosa, 2009). The *P. bullata* flower, root, and leaf extracts are reported to have capabilities in treating high blood pressure, diabetes and liver diseases, while its root is involved in boosting men sexual desires (Virmala, 2013). These factors, therefore, lead to the exploitation of *P. bullata* plant from the wild habitat, which may become a reason of species extinction in the near future. Apart from that, the accumulation of the plant secondary metabolites is naturally time consuming, and most of the bioactive compounds in *P. bullata* are remain unknown.

Among the bioactive compounds, alkaloids play a crucial role in pharmaceutical industries as important drugs for controlling numerous diseases (Dias *et al.*, 2012; Perviz *et al.*, 2016). These compounds have been used as a source of remedies to treat diverse range of human diseases (Amirkia and Heinrich, 2014). However, the alkaloid content in the intact plant is low, and the accumulation of this compound is influenced by environmental factors (Bienaimé *et al.*, 2015; Gupta *et al.*, 2015). Thus, the *in vitro* technique can be utilized to mass produce bioactive compounds without harvesting the entire plant (Efferth, 2019).

Plant tissue culture technique has been widely used due to its ability to produce disease free plant and pharmaceutically important secondary metabolites such as phenolics, alkaloids, and terpenoids (Adhikari and Pant, 2013; Upadhyay and Koche, 2015; Shahzad *et al.*, 2017). Callus culture is one of the techniques that can be used to facilitate the mass production of these compounds. The presence of auxins is significant in callus formation and mass production of bioactive compounds (Ahmad *et al.*, 2013; Ikeuchi *et al.*, 2013). The application of exogenous auxins may affect the biosynthesis of bioactive compounds including alkaloids in callus culture (Raj *et al.*, 2015).

The yield of bioactive compounds especially alkaloid in callus culture can be improved by adding the elicitors and precursors into the callus culture media. Elicitors are the chemical compounds that can promote stress responses in plants and lead to the mass production of secondary metabolites (Naik and Al-Khayri, 2016). As for precursor feeding, the addition of precursors that is involved in the metabolic biosynthetic pathway has stimulates the production of bioactive compounds (Zuldin *et al.*, 2013).

1.2 Hypothesis

The hypotheses of this study were:

- 1) Polar extraction solvent is the best solvent to extract high amount of bioactive compounds from *P. bullata* leaf, stem, and root
- 2) The different concentrations of auxins applied in callus induction media can induce the formation of *P. bullata* callus grown under dark condition
- 3) Application of different types of auxins, elicitors, and precursors can enhance the production of total alkaloids in callus

1.3 Objectives

The objectives of this study are:

- 1) To determine the efficiency of different extraction solvents in extracting phytochemical content and their antioxidative capacity in leaf, stem, and root of *P. bullata*
- 2) To profile phytochemical compounds in leaf, stem, and root of *P. bullata* using GC-MS
- 3) To induce callus from *P. bullata* using different explants, culture media, and auxins
- 4) To determine the effectiveness of auxins, elicitors, and precursors in enhancing alkaloid production in callus of *P. bullata*

REFERENCES

- Abbas, M. S., El-Shabrawi, H. M., Soliman, A. S. and Selim, M. A. (2018). Optimization of germination, callus induction, and cell suspension culture of African locust beans *Parkia biglobosa* (Jacq.) Benth. *Journal of Genetic Engineering and Biotechnology*, 16: 191-201.
- Abraham, J. and Thomas, T. D. (2015). Plant regeneration from organogenic callus and assessment of clonal fidelity in *Elephantopus scaber* Linn., an ethnomedicinal herb. *Physiology and Molecular Biology of Plants*, 21: 269-277.
- Addla, D., Sridhar, B., Devi, A. and Kantevari, S. (2012). Design, synthesis and antimicrobial evaluation of novel 1-benzyl 2-butyl-4-chloroimidazole embodied 4-azafluorenones via molecular hybridization approach. *Bioorganic & Medicinal Chemistry Letters*, 22: 7475–7480.
- Adhikari, S. R. and Pant, B. (2013). Induction and proliferation of *in vitro* mass of callus of *Withania somnifera* (L.) Dunal. *Research in Plant Sciences*, 1: 58-61.
- Adil, M., Ren, X. and Jeong, B. R. (2019). Light elicited growth, antioxidant enzymes activities and production of medicinal compounds in callus culture of *Cnidium officinale* Makino. *Journal of Photochemistry and Photobiology B: Biology*, 196: 111509-111516.
- Adnan, M., Chy, N. U., Mostafa Kamal, A. T. M., Azad, M. O. K., Paul, A., Uddin, S. B., Barlow, J. W., Faruque, M. O., Park, C. H. and Cho, D. H. (2019). Investigation of the biological activities and characterization of bioactive constituents of *Ophiorrhiza rugosa* var. *prostrata* (D. Don) & Mondal leaves through *in vivo*, *in vitro*, and *in silico* approaches. *Molecules*, 24: 1367-1390.
- Aghaali, Z., Hoshino, Y., Monfared, S. R. and Moeini, A. (2019). Regulation of dedifferentiation and differentiation in different explants of *Papaver rheas* L. by one-step culture. *Scientia Horticulturae*, 246: 366-370.
- Agrawal, R., Sethiya, N. K. and Mishra, S. H. (2013). Antidiabetic activity of alkaloids of *Aerva lanata* roots on streptozotocin-nicotinamide induced type-II diabetes in rats. *Pharmaceutical Biology*, 51: 635-642.
- Ahmad, S., Garg, M., Tamboli, E. T., Abdin, M. Z. and Ansari, S. H. (2013). *In vitro* production of alkaloids: factors, approaches, challenges and prospects. *Pharmacognosy Reviews*, 7: 27-33.
- Ahmed, E., Arshad, M., Zakriyya Khan, M., Shoaib Amjad, H., Mehreen Sadaf, H., Riaz, I., Sabir, S., Ahmad, N. and Sabaoon, M. A. (2017). Secondary metabolites and their multidimensional prospective in plant life. *Journal of Pharmacognosy and Phytochemistry*, 6: 205-214.

- Ahmed, S. A. and Baig, M. M. V. (2014). Biotic elicitor enhanced production of psoralen in suspension cultures of *Psoralea corylifolia* L. *Saudi Journal of Biological Sciences*, 21: 499-504.
- Ahsan, T., Chen, J., Zhao, X., Irfan, M. and Wu, Y. (2017). Extraction and identification of bioactive compounds (eicosane and dibutyl phthalate) produced by *Streptomyces* strain KX852460 for the biological control of *Rhizoctonia solani* AG-3 strain KX852461 to control target spot disease in tobacco leaf. *AMB Express*, 7: 54-62.
- Ain, Q. U., Khan, H., Mubarak, M. S. and Pervaiz, A. (2016). Plant alkaloids as antiplatelet agent: drugs of the future in the light of recent developments. *Frontiers in Pharmacology*, 7: 292-300.
- Al Hashmi, L. S., Hossain, M. A., Weli, A. M., Al-Riyami, Q. and Al-Sabahi, J. N. (2013). Gas chromatography-mass spectrometry analysis of different organic crude extracts from the local medicinal plant of *Thymus vulgaris* L. *Asian Pacific Journal of Tropical Biomedicine*, 3: 69-73.
- Ali, M., Boonerjee, S., Islam, M. N., Saha, M. L., Hoque, M. I. and Sarker, R. H. (2018). Endogenous bacterial contamination of plant tissue culture materials: identification and control strategy. *Plant Tissue Culture and Biotechnology*, 28: 99-108.
- Almeida, É. S., de Oliveira, D. and Hotza, D. (2019). Properties and applications of *Morinda citrifolia* (Noni): a review. *Comprehensive Reviews in Food Science and Food Safety*, 18: 883-909.
- Altemimi, A., Lakhssassi, N., Baharlouei, A., Watson, D. and Lightfoot, D. (2017). Phytochemicals: Extraction, isolation, and identification of bioactive compounds from plant extracts. *Plants*, 6: 42-65.
- Amirkia, V. and Heinrich, M. (2014). Alkaloids as drug leads—a predictive structural and biodiversity-based analysis. *Phytochemistry Letters*, 10: 48-53.
- Anand, D., Yadav, P. K., Patel, O. P. S., Parmar, N., Maurya, R. K., Vishwakarma, P., Raju, K. S. R., Taneja, I., Wahajuddin, M., Kar, S. and Yadav, P. P. (2017). Antileishmanial activity of pyrazolopyridine derivatives and their potential as an adjunct therapy with miltefosine. *Journal of Medicinal Chemistry*, 60: 1041–1059.
- Anandan, R., Deenathayalan, T., Kumar, N. S. and Deepak. K. V. (2018). An alternative *in vitro* plant regeneration system in papaya (*Carica papaya* L.) through callus derived nodular cultures. *Meta Gene*, 17: 147-152.
- Andi, S. A., Gholami, M., Ford, C. M. and Maskani, F. (2019). The effect of light, phenylalanine and methyl jasmonate, alone or in combination, on growth and secondary metabolism in cell suspension cultures of *Vitis vinifera*. *Journal of Photochemistry & Photobiology, B: Biology*, 199: 111625-111634.

- Ang, H. H. and Lee, K. L. (2006). Analysis of lead in Tongkat Ali Hitam herbal preparations in Malaysia. *Toxicological and Environmental Chemistry*, 87: 521-528.
- Anjusha, S and Gangaprasad, A. (2017). Callus culture and *in vitro* production of anthraquinone in *Gynochthodes umbellata* (L.) Razafim. and B. Bremer (Rubiaceae). *Industrial Crops and Products*, 95: 608-614.
- Anyasor, G. N., Funmilayo, O., Odutola, O., Olugbenga, A. and Oboutor, E. M. (2014). Chemical constituents in n-butanol fractions of *Castus afer* ker Gawl leaf and stem. *Journal of Intercultural Ethnopharmacology*, 3: 78-84.
- Asiah, O., Nurhanan, M. Y. and Ilham, A. M. (2007). Determination of bioactive peptide (4.3 kDa) as an aphrodisiac marker in six Malaysian plants. *Journal of Tropical Forest Science*, 19: 61-63.
- Asuk, A. A., Agiang, M. A., Dasofunjo, K. and Willie, A. J. (2015). The biomedical significance of the phytochemical, proximate and mineral compositions of the leaf, stem bark and root of *Jatropha curcas*. *Asian Pacific Journal of Tropical Biomedicine*, 5: 650-657.
- Awad, V., Kuvallek, A. and Harsulkar, A. (2014). Microbial elicitation in root cultures of *Taverniera cuneifolia* (Roth) Arn. for elevated glycyrrhizic acid production. *Industrial Crops and Products*, 54: 13-16.
- Azliza, M. A., Ong, H. C., Vikineswary, S., Noorlidah, A. and Haron, N. W. (2012). Ethno-medicinal resources used by the Temuan in Ulu Kuang Village. *Studies on Ethno-Medicine*, 6: 17-22.
- Azmir, J., Zaidul, I. S. M., Rahman, M. M., Sharif, K. M., Mohamed, A., Sahena, F., Jahurul, M. H. A., Ghafoor, K., Norulaini, N. A. N. and Omar, A. K. M. (2013). Techniques for extraction of bioactive compounds from plant materials: A review. *Journal of Food Engineering*, 117: 426-436.
- Baek, S., Ho, T. T., Lee, H., Jung, G., Kim, Y. E., Jeong, C. S. and Park, S. Y. (2020). Enhanced biosynthesis of triterpenoids in *Centella asiatica* hairy root culture by precursor feeding and elicitation. *Plant Biotechnology Reports*, 14: 45-53.
- Bañas, N., García-Viguera, C. and Moreno, D. (2014). Elicitation: a tool for enriching the bioactive composition of foods. *Molecules*, 19: 13541-13563.
- Banakar, P. and Jayaraj, M. (2018). GC-MS analysis of bioactive compounds from ethanolic leaf extract of *Waltheria indica* Linn. and their pharmacological activities. *International Journal of Pharmaceutical Sciences and Research*, 9: 2005-2010.
- Barchan, A., Bakkali, M., Arakrak, A., Pagan, R. and Laglaoui, A. (2014). The effects of solvents polarity on the phenolic contents and antioxidant activity of three *Mentha* species extracts. *International Journal of Current Microbiology and Applied Sciences*, 3: 399-412.

- Bartwal, A., Mall, R., Lohani, P., Guru, S. K. and Arora, S. (2013). Role of secondary metabolites and brassinosteroids in plant defense against environmental stresses. *Journal of Plant Growth Regulation*, 32: 216-232.
- Behbahani, M., Shanehsazzadeh, M. and Hessami, M. J. (2011). Optimization of callus and cell suspension cultures of *Barringtonia racemosa* (Lecythidaceae family) for lycopene production. *Scientia Agricola*, 68: 69-76.
- Behrens, M. R., Mutlu, N., Chakraborty, S., Dumitru, R., Jiang, W. Z., LaVallee, B. J., Herman, P. L., Clemente, T. E. and Weeks, D. P. (2007). Dicamba resistance: enlarging and preserving biotechnology-based weed management strategies. *Science*, 316: 1185-1188.
- Bekheet, S. A., El-Bahr, M. K., Ali, S. A. and Hamed, M. A. (2014). Callus production of Globe artichoke and Milk thistle: in vitro hypolipidemic and antioxidant activities. *World Journal of Pharmaceutical Research*, 3: 1-17.
- Benderradjji, L., Brini, F., Kellou, K., Ykhlef, N., Djekoun, A., Masmoudi, K. and Bouzerzour, H. (2011). Callus induction, proliferation, and plantlets regeneration of two bread wheat (*Triticum aestivum* L.) genotypes under saline and heat stress conditions. *ISRN Agronomy*, 2012: 1-8.
- Bendiabdellah, A., Dib, M. E. A., Meliani, N., Djabou, N., Allali, H. and Tabti, B. (2012). Preliminary phytochemical screening and antioxidant activities of solvent extracts from *Daucus crinitus* Desf., from Algeria. *Journal of Applied Pharmaceutical Science*, 2: 92-95.
- Berini, J. L., Brockman, S. A., Hegeman, A. D., Reich, P. B., Muthukrishnan, R., Montgomery, R. A. and Forester, J. D. (2018). Combinations of abiotic factors differentially alter production of plant secondary metabolites in five woody plant species in the boreal-temperate transition zone. *Frontiers in Plant Science*, 9: 1257-1264.
- Besher, S., Al-Ammouri, Y. and Murshed, R. (2014). Production of tropane alkaloids in the *in vitro* and callus cultures of *Hyoscyamus aureus* and their genetic stability assessment using ISSR markers. *Physiology and Molecular Biology of Plants*, 20: 343-349.
- Bhat, P. and Bhat, A. (2016). Silver nanoparticles for enhancement of accumulation of capsaicin in suspension culture of *Capsicum* sp. *Journal of Experimental Sciences*, 7: 1-6.
- Bhojwani, S. S. and Dantu, P. K. (2013). Tissue and cell culture. In Bhojwani, S. S. and Dantu, P. K. (Ed.), *Plant Tissue Culture: An Introductory Text* (pp. 39-50). India: Springer.
- Bienaimé, C., Melin, A., Bensaddek, L., Attoumbré, J., Nava-Saucedo, E. and Baltora-Rosset, S. (2015). Effects of plant growth regulators on cell growth and alkaloids production by cell cultures of *Lycopodiella inundata*. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 123: 523-533.

- Bimakr, M., Rahman, R. A., Taip, F. S., Ganjloo, A., Salleh, L. M., Selamat, J., Hamid, A. and Zaidul, I. S. M. (2011). Comparison of different extraction methods for the extraction of major bioactive flavonoid compounds from spearmint (*Mentha spicata* L.) leaves. *Food and Bioproducts Processing*, 89: 67-72.
- Binte Mostafiz, S. and Wagiran, A. (2018). Efficient callus induction and regeneration in selected Indica rice. *Agronomy*, 8: 77-96.
- Bitis, L., Sen, A. Ozsoy., N., Birteksoz-Tan, S., Kultur, S. and Melikoglu, G. (2017). Flavonoids and biological activities of various extracts from *Rosa sempervirens* leaves. *Biotechnology and Biotechnological Equipment*, 31: 299-303.
- Block, A. K., Vaughan, M. M., Schmelz, E. A. and Christensen, S. A. (2019). Biosynthesis and function of terpenoid defense compounds in maize (*Zea mays*). *Planta*, 249: 21-30.
- Boamponsem, G. A. and Leung, D. W. (2017). Use of compact and friable callus cultures to study adaptive morphological and biochemical responses of potato (*Solanum tuberosum*) to iron supply. *Scientia Horticulturae*, 219: 161-172.
- Boeing, J. S., Barizao, E. O., e Silva, B. C., Montanher, P. F., de Cinque Almeida, V. and Visentainer, J. V. (2014). Evaluation of solvent effect on the extraction of phenolic compounds and antioxidant capacities from the berries: application of principal component analysis. *Chemistry Central Journal*, 8: 1-9.
- Brahmi, F., Mechri, B., Dabbou, S., Dhibi, M. and Hammami, M. (2012). The efficacy of phenolics compounds with different polarities as antioxidants from olive leaves depending on seasonal variations. *Industrial Crops and Products*, 38: 146-152.
- Brahmkshatriya, P. P. and Brahmkshatriya, P. S. (2013). Terpenes: Chemistry, biological role, and therapeutic applications. In Ramawat, K. and Mérillon, J. M. (Ed.), *Natural Products* (pp. 2665-2691). Berlin, Heidelberg: Springer.
- Bunchom, N., Padhungkir, M., Saijuntha, W., Thanonkeo, P. and Thanonkeo, S. (2014). Production of resveratrol from callus cultures of *Artocarpus lacucha* buch.-ham. *KKU Research Journal*, 19: 262-267.
- Cadar, E., Tomescu, A., Erimia, C. L., Mustafa, A. and Sîrbu, R. (2015). The impact of alkaloids structures from natural compounds on public health. *European Journal of Social Science Education and Research*, 2: 34-39.
- Can, E., Çeliktaş, N. and Hatipoğlu, R. (2008). Effect of auxin type and concentrations in different media on the callus induction and shoot formation of crested wheatgrass (*Agropyron cristatum* (L.) Gaertn). *Biotechnology and Biotechnological Equipment*, 22: 782-786.
- Carocho, M. and Ferreira, I. C. (2013). A review on antioxidants, prooxidants and related controversy: natural and synthetic compounds, screening and analysis

- methodologies and future perspectives. *Food and Chemical Toxicology*, 51: 15-25.
- Cartea, M. E., Francisco, M., Lema, M., Soengas, P. and Velasco, P. (2010). Resistance of cabbage (*Brassica oleracea capitata* group) crops to *Mamestra brassicae*. *Journal of Economic Entomology*, 103: 1866-1874.
- Castro, A. H. F., Braga, K. D. Q., Sousa, F. M. D., Coimbra, M. C. and Chagas, R. C. R. (2016). Callus induction and bioactive phenolic compounds production from *Brysonima verbascifolia* (L.) DC. (Malpighiaceae). *Revista Ciéncia Agronómica*, 47: 143-151.
- Casuga, F. P., Castillo, A. L. and Corpuz, M. J. A. T. (2016). GC-MS analysis of bioactive compounds present in different extracts of an endemic plant *Broussonetia luzonica* (Blanco) (Moraceae) leaves. *Asian Pacific Journal of Tropical Biomedicine*, 6: 957-961.
- Chakraborty, N., Banerjee, D., Ghosh, M., Pradhan, P., Gupta, N. S., Acharya, K. and Banerjee, M. (2013). Influence of plant growth regulators on callus mediated regeneration and secondary metabolites synthesis in *Withania somnifera* (L.) Dunal. *Physiology and Molecular Biology of Plants*, 19: 117-125.
- Chandrasekara, A. and Shahidi, F. (2018). Herbal beverages: bioactive compounds and their role in disease risk reduction-a review. *Journal of Traditional and Complementary Medicine*, 8: 451-458.
- Chen, Y. M., Huang, J. Z., Hou, T. W. and Pan, I. C. (2019). Effects of light intensity and plant growth regulators on callus proliferation and shoot regeneration in the ornamental succulent Haworthia. *Botanical Studies*, 60: 10-18.
- Chen, M., Zhang, J., Zhou, Y., Li, S., Fan, X., Yang, L., Guan, Y. and Zhang, Y. (2019). Transcriptome analysis of *Lilium Oriental* × *Trumpet* hybrid roots reveals auxin-related genes and stress-related genes involved in picloram-induced somatic embryogenesis induction. *Journal of Horticultural Science and Biotechnology*, 94: 317-330.
- Cho, K. S., Lim, Y. R., Lee, K., Lee, J., Lee, J. H. and Lee, I. S. (2017). Terpenes from forests and human health. *Toxicological Research*, 33: 97-106.
- Chodisetti, B., Rao, K., Gandi, S. and Giri, A. (2015). Gymnemic acid enhancement in the suspension cultures of *Gymnema sylvestre* by using the signaling molecules—methyl jasmonate and salicylic acid. *In Vitro Cellular and Developmental Biology-Plant*, 51: 88-92.
- Christiana, O. A., Johnbull, O. E., Raphael, C. M., Joseph, O. O., Paul, M. O. and Emmanuel, G. J. (2019). Gas chromatographic study of bio-active compounds in methanolic extract of leaf of *Crateva adansonii* DC. *Journal of Physics: Conference Series*, 1299: 012014-012022.

- Chu, C. C., Wang, C. C., Sun, C. S., Chen, H., Yin, K. C. and Chu, C. Y. (1975). Establishment of an efficient medium for anther culture of rice through comparative experiments on the nitrogen sources. *Scientia Sinica*, 18: 659-668.
- Cioć, M., Szewczyk, A., Źupnik, M., Kalisz, A. and Pawłowska, B. (2018). LED lighting affects plant growth, morphogenesis and phytochemical contents of *Myrtus communis* L. *in vitro*. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 132: 433-447.
- Connolly, J. D., Haque, M. E. and Kadir, A. A. (1996). Two 7, 7'-bisdehydroaporphine alkaloids from *Polyalthia bullata*. *Phytochemistry*, 43: 295-297.
- Cushnie, T. T., Cushnie, B. and Lamb, A. J. (2014). Alkaloids: an overview of their antibacterial, antibiotic-enhancing and antivirulence activities. *International Journal of Antimicrobial Agents*, 44: 377-386.
- Dai, J. and Mumper, R. J. (2010). Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*, 15: 7313-7352.
- Das, J., Mao, A. A. and Handique, P. J. (2013). Callus-mediated organogenesis and effect of growth regulators on production of different valepotriates in Indian valerian (*Valeriana jatamansi* Jones.). *Acta Physiologiae Plantarum*, 35: 55-63.
- de Aguiar Lage, D., da Silva Tirado, M., Vanicore, S. R., de Carvalho Sabino, K. C. and Albarello, N. (2015). Production of betalains from callus and cell suspension cultures of *Pereskia aculeata* Miller, an unconventional leafy vegetable. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 122: 341-350.
- de Moraes, J., de Oliveira, R. N., Costa, J. P., Junior, A. L., de Sousa, D. P., Freitas, R. M., Allegretti, S. M. and Pinto, P. L. (2014). Phytol, a diterpene alcohol from chlorophyll, as a drug against neglected tropical disease *Schistosomiasis mansoni*. *PLoS Neglected Tropical Diseases*, 8: e2617-e2628.
- de Souza, R. R., de Oliveira Paiva, P. D., da Silva, R. R., da Silva, D. P. C., dos Reis, M. V. and Paiva, R. (2019). Morphogenetic potential of different sources of explants for efficient *in vitro* regeneration of *Genipa* sp. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 136: 153-160.
- Dian-Nashiela, F., Abdullah, N., Hashim, N. and Abdul Hamid, A. (2015). Antioxidant activity of herbal tea prepared from *Cosmos caudatus* leaves at different maturity stages. *International Food Research Journal*, 22: 1189-1194.
- Dias, D. A., Urban, S. and Roessner, U. (2012). A historical overview of natural products in drug discovery. *Metabolites*, 2: 303-336.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S. and Ju, Y. H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatica*. *Journal of Food and Drug Analysis*, 22: 296-302.

- Dou, H., Niu, G., Gu, M. and Masabni, J. (2017). Effects of light quality on growth and phytonutrient accumulation of herbs under controlled environments. *Horticulturae*, 3: 36-47.
- Dube, P., Meyer, S. and Marnewick, J. L. (2017). Antimicrobial and antioxidant activities of different solvent extracts from fermented and green honeybush (*Cyclopia intermedia*) plant material. *South African Journal of Botany*, 110: 184-193.
- Efferth, T. (2019). Biotechnology applications of plant callus cultures. *Engineering*, 5: 50-59.
- El-Nabarawy, M. A., El-Kafafi, S. H., Hamza, M. A. and Omar, M. A. (2015). The effect of some factors on stimulating the growth and production of active substances in *Zingiber officinale* callus cultures. *Annals of Agricultural Sciences*, 60: 1-9.
- Elshorbagy, M. I., Ibrahim, S. M., El-Seoud, K. A. A. and Abd El-Maksoud, A. I. (2018). Tropane alkaloids production from callus culture of *Atropa belladonna* L. as affected by elicitors and precursor feeding. *International Research Journal of Pharmacy*, 9: 116-125.
- Fan, G., Nie, T., Fan, J. and Zhan, Y. (2017). Exogenous feeding of fructose and phenylalanine further improves betulin production in suspended *Betula platyphylla* cells under nitric oxide treatment. *Molecules*, 22: 1035-1046.
- Farhadi, N., Panahandeh, J., Azar, A. M. and Salte, S. A. (2017). Effects of explant type, growth regulators and light intensity on callus induction and plant regeneration in four ecotypes of Persian shallot (*Allium hirtifolium*). *Scientia Horticulturae*, 218: 80-86.
- Fatiha, B., Khodir, M., Farid, D., Tiziri, R., Karima, B., Sonia, O. and Chibane, M. (2012). Optimisation of solvent extraction of antioxidants (phenolic compounds) from Algerian mint (*Mentha spicata* L.). *Pharmacognosy Communications*, 2: 72-86.
- Fazal, H., Abbasi, B. H., Ahmad, N., Ali, S. S., Akbar, F. and Kanwal, F. (2016). Correlation of different spectral lights with biomass accumulation and production of antioxidant secondary metabolites in callus cultures of medicinally important *Prunella vulgaris* L. *Journal of Photochemistry and Photobiology B: Biology*, 159: 1-7.
- Feduraev, P., Chupakhina, G., Maslennikov, P., Tacenko, N. and Skrypnik, L. (2019). Variation in phenolic compounds content and antioxidant activity of different plant organs from *Rumex crispus* L. and *Rumex obtusifolius* L. at different growth stages. *Antioxidants*, 8: 237-242.
- Fehér, A. (2019). Callus, dedifferentiation, totipotency, somatic embryogenesis: What these terms mean in the era of molecular plant biology?. *Frontiers in Plant Science*, 10: 536-547.

- Ferdausi, A., Chang, X., Hall, A. and Jones, M. (2020). Galanthamine production in tissue culture and metabolomic study on Amaryllidaceae alkaloids in *Narcissus pseudonarcissus* cv. Carlton. *Industrial Crops and Products*, 144: 112058-112070.
- Filova, A. (2014). Production of secondary metabolites in plant tissue culture. *Research Journal of Agricultural Science*, 46: 236-245.
- Forni, C., Facchiano, F., Bartoli, M., Pieretti, S., Facchiano, A., D'Arcangelo, D., Norelli, S., Valle, G., Nisini, R., Beninati, S., Jadeja, R. N. and Tabolacci, C. (2019) Beneficial role of phytochemicals on oxidative stress and age-related diseases. *BioMed Research International*, 2019: 1-16.
- Fu, Z. F., Tu, Z. C., Zhang, L., Wang, H., Wen, Q. H. and Huang, T. (2016). Antioxidant activities and polyphenols of sweet potato (*Ipomoea batatas* L.) leaves extracted with solvents of various polarities. *Food Bioscience*, 15: 11-18.
- Gadzovska, S., Maury, S., Delaunay, A., Spasenoski, M., Hagège, D., Courtois, D. and Joseph, C. (2013). The influence of salicylic acid elicitation of shoots, callus, and cell suspension cultures on production of naphtodianthrone and phenylpropanoids in *Hypericum perforatum* L. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 113: 25-39.
- Gaikwad, A. V., Singh, S. K. and Gilhotra, R. (2017). Plant tissue culture – a review. *Journal of Pharmaceutical Research and Education*, 2: 217-220.
- Galanakis, C. M., Goulas, V., Tsakona, S., Manganaris, G. A. and Gekas, V. (2013). A knowledge base for the recovery of natural phenols with different solvents. *International Journal of Food Properties*, 16: 382-396.
- Galáz-Ávalos, R. M., Aguilar-Díaz, S., Xool-González, P. A., Huchín-May, S. M. and Loyola-Vargas, V. M. (2018). Callus, suspension culture, and hairy roots. Induction, maintenance and characterization. In Loyola-Vargas V. and Ochoa-Alejo N. (Ed.), *Plant Cell Culture Protocols. Methods in Molecular Biology (Methods and Protocols)* (pp. 29-40). Totowa, NJ: Humana Press.
- Gamborg, O. L., Miller R. A. and Ojima K. (1968). Nutrient requirements of suspension cultures of soybean root cells. *Experimental Cell Research*, 50: 151-158.
- Gangwar, M., Gautam, M. K., Sharma, A. K., Tripathi, Y. B., Goel, R. K. and Nath, G. (2014). Antioxidant capacity and radical scavenging effect of polyphenol rich *Mallotus philippensis* fruit extract on human erythrocytes: an in vitro study. *The Scientific World Journal*, 2014: 1-12.
- Gaosheng, H. and Jingming, J. (2012). Production of useful secondary metabolites through regulation of biosynthetic pathway in cell and tissue suspension culture of medicinal plants. In Leva, A. and Rinaldi, L. (Ed.), *Recent Advances in Plant In Vitro Culture*, (pp. 197-210). United Kingdom: IntechOpen.

- Geetha, D. H., Rajeswari, M. and Jayashree, I. (2013). Chemical profiling of *Elaeocarpus serratus* L. by GC-MS. *Asian Pacific Journal of Tropical Biomedicine*, 3: 985-987.
- Geetha, N., Subha, D. and Chandralega, N. (2015). Phytochemical screening of *Tanacetum Parthenium* L. (Feverfew) leaves: an important medicinal plant. *International Journal of Pharmacy and Pharmaceutical Research*. 2: 98-126.
- Gensicka-Kowalewska, M., Cholewiński, G. and Dzierzbicka, K. (2017). Recent developments in the synthesis and biological activity of acridine/acridone analogues. *RSC Advances*, 7: 15776-15804.
- George, E. F., Hall, M. A. and Klerk, G. J. D. (2008). Plant tissue culture procedure - Background. In George, E. F., Hall, M. A. and Klerk, G. J. D. (Ed.), *Plant Propagation by Tissue Culture* (pp. 1-28). Dordrecht: Springer.
- Ghosh, N., Ghosh, R., Mandal, V. and Mandal, S. C. (2011). Recent advances in herbal medicine for treatment of liver diseases. *Pharmaceutical Biology*, 49: 970-988.
- Global Information Hub on Integrated Medicine. N.d. Retrieved 30 August 2019 from <http://www.globinmed.com/>.
- Godara, P., Dulara, B. K., Barwer, N. and Chaudhary, N. S. (2019). Comparative GC-MS analysis of bioactive phytochemicals from different plant parts and callus of *Leptadenia reticulata* Wight and Arn. *Pharmacognosy Journal*, 11: 129-140.
- Grąbkowska, R., Matkowski, A., Grzegorczyk-Karolak, I. and Wysokińska, H. (2016). Callus cultures of *Harpagophytum procumbens* (Burch.) DC. ex Meisn.; production of secondary metabolites and antioxidant activity. *South African Journal of Botany*, 103: 41-48.
- Grossmann, K. (2010). Auxin herbicides: current status of mechanism and mode of action. *Pest Management Science: Formerly Pesticide Science*, 66: 113-120.
- Guerriero, G., Berni, R., Muñoz-Sánchez, J., Apone, F., Abdel-Salam, E., Qahtan, A., Alatar, A. A., Cantini, C., Cai, G., Hausman, J., Hernandez-Sotomayor, S. M. T., Faisal, M. and Siddiqui, K. (2018). Production of plant secondary metabolites: examples, tips and suggestions for biotechnologists. *Genes*, 9: 309-331.
- Gupta, P., Sharma, S. and Saxena, S. (2015). Biomass yield and steviol glycoside production in callus and suspension culture of *Stevia rebaudiana* treated with proline and polyethylene glycol. *Applied Biochemistry and Biotechnology*, 176: 863-874.
- Habibah, N. A., Moeljopawiro, S., Dewi, K. and Indrianto, A. (2018). Callus induction and flavonoid production on the immature seed of *Stelechocarpus burahol*. *Journal of Physics: Conference Series*, 983: 12186-12193.

- Hadi, M. A. M., Zhang, F. J., Wu, F. F., Zhou, C. H. and Tao, J. (2013). Advances in fruit aroma volatile research. *Molecules*, 18: 8200–8229.
- Hajam, M. A., Hassan, G. I., Bhat, T. A., Bhat, I. A., Rather, A. M., Parry, E. A., Wani, M. A. and Khan, I. F. (2017). Understanding plant growth regulators, their interplay: For nursery establishment in fruits. *International Journal of Chemical Studies*, 5: 905-910.
- Halliday, K. J., Martínez-García, J. F. and Josse, E. M. (2009). Integration of light and auxin signaling. *Cold Spring Harbor Perspectives In Biology*, 1: 1586-1597.
- Halliwell, B. (2007). Biochemistry of oxidative stress. *Biochemical Society Transactions*, 35: 1147-1150.
- Hari, G., Vadlapudi, K., Vijendra, P. D., Rajashekhar, J., Sannabommaji, T. and Basappa, G. (2018). A combination of elicitor and precursor enhances psoralen production in *Psoralea corylifolia* Linn. suspension cultures. *Industrial Crops and Products*, 124: 685-691.
- Hazuba-Przybył, T. (2019). Propagation of *Juniper* species by plant tissue culture: a mini-review. *Forests*, 10: 1028-1045.
- Hesami, M. and Daneshvar, M. H. (2018). Indirect organogenesis through seedling-derived leaf segments of *Ficus religiosa*-a multipurpose woody medicinal plant. *Journal of Crop Science and Biotechnology*, 21: 129-136.
- Hosseini, N. S., Hagh, Z. G. and Khoshghalb, H. (2020). Morphological, antioxidant enzyme activity and secondary metabolites accumulation in response of polyethylene glycol-induced osmotic stress in embryo-derived plantlets and callus cultures of *Salvia leuviifolia*. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 140: 143-155.
- Hussain, A. M., Mansoor, S. S., Aswin, K., and Sudhan, S. P. N. (2014). Pentafluorophenylammonium triflate: An effective and reusable organocatalyst for the one-pot preparation of 2,4-diaryl-5H-indeno [1, 2-b] pyridin-5-one derivatives. *Journal of King Saud University-Science*, 26: 213-221.
- Hussain, G., Rasul, A., Azwar, H., Aziz, N., Razzaq, A., Wei, W., Ali, M., Li, J. and Li, X. (2018). Role of plant derived alkaloids and their mechanism in neurodegenerative disorders. *International Journal of Biological Sciences*, 14: 341-357.
- Hussain, M. S., Fareed, S., Saba Ansari, M., Rahman, A., Ahmad, I. Z. and Saeed, M. (2012). Current approaches toward production of secondary plant metabolites. *Journal of Pharmacy and Bioallied Sciences*, 4: 10-20.
- Ikeuchi, M., Sugimoto, K. and Iwase, A. (2013). Plant callus: mechanisms of induction and repression. *The Plant Cell*, 25: 3159-3173.

- Iloki-Assanga, S. B., Lewis-Lujan, L. M., Lara-Espinoza, C. L., Gil-Salido, A. A., Fernandez-Angulo, D., Rubio-Pino, J. L. and Haines, D. D. (2015). Solvent effects on phytochemical constituent profiles and antioxidant activities, using four different extraction formulations for analysis of *Bucida buceras* L. and *Phoradendron californicum*. *BMC Research Notes*, 8: 396-410.
- Iqbal, J., Abbasi, B. A., Mahmood, T., Kanwal, S., Ali, B., Shah, S. A. and Khalil, A. T. (2017). Plant-derived anticancer agents: a green anticancer approach. *Asian Pacific Journal of Tropical Biomedicine*, 7: 1129-1150.
- Irene, W. M., Alumiro, H. L., Asava, K. K., Agwanda, C. O. and Anami, S. E. (2019). Effects of genotype and plant growth regulators on callus induction in leaf cultures of *Coffea arabica* L.F1 hybrid. *Journal of Plant Biochemistry & Physiology*, 7: 236-248.
- Iriti, M. and Faoro, F. (2009). Chitosan as a MAMP, searching for a PRR. *Plant Signaling & Behavior* 4: 66-68.
- Isah, T. (2016). Anticancer alkaloids from trees: development into drugs. *Pharmacognosy Reviews*, 10: 90-99.
- Isah, T., Umar, S., Mujib, A., Sharma, M. P., Rajasekharan, P. E., Zafar, N. and Frukha, A. (2018). Secondary metabolism of pharmaceuticals in the plant *in vitro* cultures: strategies, approaches, and limitations to achieving higher yield. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 132: 239-265.
- Jadid, N., Hidayati, D., Hartanti, S. R., Arraniry, B. A., Rachman, R. Y. and Wikanta, W. (2017). Antioxidant activities of different solvent extracts of *Piper retrofractum* Vahl. using DPPH assay. *AIP Conference Proceedings*, 1854: 020019-020025.
- Jahanban-Esfahlan, A., Ostadrahimi, A., Tabibazar, M. and Amarowicz, R. (2019). A comparative review on the extraction, antioxidant content and antioxidant potential of different parts of walnut (*Juglans regia* L.) fruit and tree. *Molecules*, 24: 2133-2173.
- Jawahar, G., Punita, D. L., Rajasheker, G., Manoharachary, C., Venkatachalam, P. and Kishor, P. K. (2018). Feeding elicitors and precursors enhance colchicine accumulation in morphogenic cultures of *Gloriosa superba* L. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 135: 235-245.
- Jayaraman, S., Daud, N. H., Halis, R. and Mohamed, R. (2014). Effects of plant growth regulators, carbon sources and pH values on callus induction in *Aquilaria malaccensis* leaf explants and characteristics of the resultant calli. *Journal of Forestry Research*, 24: 535-540.
- Jayanthi, M., Susanthi, B., Mohan, N. M. and Mandal, P. K. (2015). In vitro somatic embryogenesis and plantlet regeneration from immature male inflorescence of adult dura and tenera palms of *Elaeis guineensis* (Jacq.). *SpringerPlus*, 4: 1-7.

- Jeet, A., Singh, Y., Singh, P., Nimoriya, R., Bilung, C. J., Kanojiya, S., Tripathi, V. and Mishra, D. K. (2020). Strategies for indole alkaloids enrichment through callus culture from *Alstonia scholaris* (L.) R. Br. *Plant Growth Regulation*, 1-10.
- Jiang, Z., Kempinski, C. and Chappell, J. (2016). Extraction and analysis of terpenes/terpenoids. *Current Protocols in Plant Biology*, 1: 345-358.
- Jin, X. L., Zhang, R. Q., Zhang, D. L., He, P. and Cao, F. X. (2009). In vitro plant regeneration of *Zelkova schneideriana*, an endangered woody species in China, from leaf explants. *The Journal of Horticultural Science and Biotechnology*, 84: 415-420.
- John, B. I. J. U., Sulaiman, C. T., George, S. and Reddy, V. R. K. (2014). Spectrophotometric estimation of total alkaloids in selected *Justicia* species. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6: 647-648.
- Kabera, J. N., Semana, E., Mussa, A. R. and He, X. (2014). Plant secondary metabolites: biosynthesis, classification, function and pharmacological properties. *Journal of Pharmacy and Pharmacology*, 2: 377-392.
- Kamarul Zaman, M. A. and Azzeme, A. M. (2019). Plant toxins: alkaloids and their toxicities. *GSC Biological and Pharmaceutical Sciences*, 6: 21-29.
- Kapoor, S., Sharma, A., Bhardwaj, P., Sood, H., Saxena, S. and Chaurasia, O. P. (2019). Enhanced production of phenolic compounds in compact callus aggregate suspension cultures of *Rhodiola imbricata* Edgew. *Applied Biochemistry and Biotechnology*, 187: 817-837.
- Katkar, K. V., Suthar, A. C. and Chauhan, V. S. (2010). The chemistry, pharmacologic, and therapeutic applications of *Polyalthia longifolia*. *Pharmacognosy Reviews*, 4: 62-68.
- Kaur, J., Singh, A., Pathak, T. and Kumar, K. (2017). Role of PGRs in anticancer alkaloids (vincristine and vinblastine) production. In Naeem, M., Aftab, T. and Khan, M. (Ed.), *Catharanthus roseus* (pp. 309-319). Cham: Springer.
- Kaur, S. and Mondal, P. (2014). Study of total phenolic and flavonoid content, antioxidant activity and antimicrobial properties of medicinal plants, *Journal of Microbiology and Experimentation*, 1: 1-6.
- Khan, T., Abbasi, B. H. and Khan, M. A. (2018). The interplay between light, plant growth regulators and elicitors on growth and secondary metabolism in cell cultures of *Fagonia indica*. *Journal of Photochemistry and Photobiology B: Biology*, 185: 153-160.
- Khan, T., Abbasi, B. H., Zeb, A. and Ali, G. S. (2018). Carbohydrate-induced biomass accumulation and elicitation of secondary metabolites in callus cultures of *Fagonia indica*. *Industrial Crops and Products*, 126: 168-176.

- Khatiwora, E., Adsul, V. B., Kulkarni, M., Deshpande, N. R. and Kashalkar, R. V. (2012). Antibacterial activity of dibutyl phthalate: a secondary metabolite isolated from *Ipomoea carnea* stem. *Journal of Pharmacy Research*, 5: 150-152.
- Kittakoop, P., Mahidol, C. and Ruchirawat, S. (2014). Alkaloids as important scaffolds in therapeutic drugs for the treatments of cancer, tuberculosis, and smoking cessation. *Current Topics in Medicinal Chemistry*, 14: 239-252.
- Kotta, S., Ansari, S. H. and Ali, J. (2013). Exploring scientifically proven herbal aphrodisiacs. *Pharmacognosy Reviews*, 7: 1-10.
- Krishnaiah, D., Sarbatly, R. and Nithyanandam, R. (2011). A review of the antioxidant potential of medicinal plant species. *Food and Bioproducts Processing*, 89: 217-233.
- Krishnamoorthy, K. and Subramaniam, P. (2014). Phytochemical profiling of leaf, stem, and tuber parts of *Solena amplexicaulis* (Lam.) Gandhi using GC-MS. *International Scholarly Research Notices*, 2014: 1-13.
- Krishnan, J. J., Gangaprasad, A. and Satheeshkumar, K. (2018). Biosynthesis of camptothecin from callus and cell suspension cultures of *Ophiorrhiza mungos* L. var. *angustifolia* (Thw.) Hook. f. *Proceedings of the National Academy of Sciences, India Section B: Biological Sciences*, 89: 893-902.
- Kukowska, M. (2017). Amino acid or peptide conjugates of acridine/acridone and quinoline/quinolone-containing drugs. A critical examination of their clinical effectiveness within a twenty-year timeframe in antitumor chemotherapy and treatment of infectious diseases. *European Journal of Pharmaceutical Sciences*, 109: 587-615.
- Kumar, R., Mamrutha, H. M., Kaur, A., Venkatesh, K., Grownal, A., Kumar, R. and Tiwari, V. (2017). Development of an efficient and reproducible regeneration system in wheat (*Triticum aestivum* L.). *Physiology and Molecular Biology of Plants*, 23: 945-954.
- Kumar, S. and Pandey, A. K. (2013). Chemistry and biological activities of flavonoids: an overview. *The Scientific World Journal*, 2013: 1-16.
- Kumar, S. S., Aruna, M. C. and Giridhar, P. (2016). Improvement of green leafy vegetables: the role of plant tissue culture and biotechnology. In Anis, M. and Ahmad, N. (eds), *Plant Tissue Culture: Propagation, Conservation and Crop Improvement* (pp. 547-582). Singapore: Springer.
- Kumar, S., Sharma, U. K., Sharma, A. K. and Pandey, A. K. (2012). Protective efficacy of *Solanum xanthocarpum* root extracts against free radical damage: phytochemical analysis and antioxidant effect. *Cellular and Molecular Biology*, 58: 171-178.

- Kumlay, A. M. and Ercisli S. (2015). Callus induction, shoot proliferation and root regeneration of potato (*Solanum tuberosum* L.) stem node and leaf explants under long-day conditions. *Biotechnology and Biotechnological Equipment*, 29: 1075-1084.
- Kurmukov, A. G. (2013). Phytochemistry of medicinal plants. In Eisenman S. W. and Zaurov, D. E. (Ed.), *Medicinal Plants of Central Asia: Uzbekistan and Kyrgyzstan* (pp. 13-14). New York, United States: Springer.
- Kurniasari, D., Yulianti, E., Sugiyarto, L. and Mercuriani, I. S. (2019). Increasing *Rhynchosystylis retusa*'s callus formation by immersing explant in ascorbic acid and planting on activated charcoal contained medium. *Journal of Physics: Conference Series*, 1241: 012060.
- Labarrere, B., Prinzing, A., Dorey, T., Chesneau, E. and Hennion, F. (2019). Variations of secondary metabolites among natural populations of sub-antarctic *Ranunculus* species suggest functional redundancy and versatility. *Plants*, 8: 234-257.
- Lamaoui, M., Chakhchar, A., Benlaouane, R., El Kharrassi, Y., Farissi, M., Wahbi, S. and El Modafar, C. (2019). Uprising the antioxidant power of *Argania spinosa* L. callus through abiotic elicitation. *Comptes Rendus Biologies*, 342: 7-17.
- Lee, K. W., Chinzorig, O., Choi, G. J., Kim, K. Y., Ji, H. C., Park, H. S., Kim, W. H. and Lee, S. H. (2012). Factors influencing callus induction and plant regeneration of dahurian wildrye grass (*Elymus dahuricus* L.). *African Journal of Biotechnology*, 11: 815-820.
- Lemos, V. C., Reimer, J. J. and Wormit, A. (2019). Color for life: Biosynthesis and distribution of phenolic compounds in pepper (*Capsicum annuum*). *Agriculture*, 9: 1-30.
- Li, X. and Tian, T. (2018). Phytochemical characterization of *Mentha spicata* L. under differential dried-conditions and associated nephrotoxicity screening of main compound with organ-on-a-chip. *Frontiers in Pharmacology*, 9: 1067-1076.
- Linsmaier, E. M. and Skoog, F. (1965). Organic growth factor requirements of tobacco tissue cultures. *Physiologia Plantarum*, 18: 100-127.
- Liu, Y., Liang, Z. and Zhang, Y. (2010b). Induction and *in vitro* alkaloid yield of calluses and protocorm-like bodies (PLBs) from *Pinellia ternata*. *In Vitro Cellular and Developmental Biology-Plant*, 46: 239-245.
- Liu, Z., Ren, Z., Zhang, J., Chuang, C. C., Kandaswamy, E., Zhou, T. and Zuo, L. (2018). Role of ROS and nutritional antioxidants in human diseases. *Frontiers in Physiology*, 9: 477-490.
- Lloyd, G. and McCown, B. (1980). Commercially-feasible micropropagation of mountain laurel, *Kalmia latifolia*, by use of shoot-tip culture. *International Plant Propagators' Society*, 30: 421-427.

- Lulai, E. C., Suttle, J. C., Olson, L. L., Neubauer, J. D., Campbell, L.G. and Campbell, M. A. (2016). Wounding induces changes in cytokinin and auxin content in potato tuber, but does not induce formation of gibberellins. *Journal of Plant Physiology*, 191: 22-28.
- Machado, M. W., Stern Neto, C., Salgado, J., Zaffari, G., Barison, A., Campos, F. R., de Corilo, Y. E., Eberlin, M. N. and Biavatti, M. W. (2010). Search for alkaloids on callus culture of *Passiflora alata*. *Brazilian Archives of Biology and Technology*, 53: 901-910.
- Mahmood, M., Normi, R. and Subramaniam, S. (2010). Optimization of suitable auxin application in a recalcitrant woody forest plant of *Eurycoma longifolia* (Tongkat Ali) for callus induction. *African Journal of Biotechnology*, 9: 8417-8428.
- Majda, M. and Robert, S. (2018). The role of auxin in cell wall expansion. *International Journal of Molecular Sciences*, 19: 951-971.
- Malik, K., Birla, D., Yadav, H., Sainger, M., Chaudhary, D. and Jaiwal, P. K. (2017). Evaluation of carbon sources, gelling agents, growth hormones and additives for efficient callus induction and plant regeneration in Indian wheat (*Triticum aestivum* L.) genotypes using mature embryos. *Journal of Crop Science and Biotechnology*, 20: 185-192.
- Mamphisiwana, N. D., Mashela, P. W. and Mdee, L. K. (2010). Distribution of total phenolics and antioxidant activity in fruit, leaf, stem and root of *Monsonia burkeana*. *African Journal of Agriculture Research*, 5: 2570-2575.
- McCown, B. H. and Lloyd, G. (1981). Woody plant medium (WPM) – a mineral nutrient formulation for microculture of woody plant species. *HortScience*, 16: 453-453.
- Mediani, A., Abas, F., Tan, C. P. and Khatib, A. (2014). Effects of different drying methods and storage time on free radical scavenging activity and total phenolic content of *Cosmos caudatus*. *Antioxidants*, 3: 358-370.
- Meenakshi, V. K., Gomathy, S., Senthamarai, S., Paripooranaselvi, M. and Chamundeswari, K. P. (2012). GC-MS Determination of the bioactive components of *Microcosmus exasperatus* Heller, 1878. *Journal of Current Chemical and Pharmaceutical Sciences*, 2: 271-276.
- Memar, M. Y., Raei, P., Alizadeh, N., Aghdam, M. A. and Kafil, H. S. (2017). Carvacrol and thymol: strong antimicrobial agents against resistant isolates. *Reviews in Medical Microbiology*, 28: 63-68.
- Mendonça, E. G., Paiva, L. V., Stein, V. C., Pires, M. F., Santos, B. R. and Pereira, F. J. (2012). Growth curve and development of the internal calli structure of *Eucalyptus camaldulensis* Dehn. *Brazilian Archives of Biology and Technology*, 55: 887-896.
- Michael, J. P. (2007). Quinoline, quinazoline and acridone alkaloids. *Natural Product Reports*, 24: 223-246.

- Mikulic- Petkovsek, M., Schmitzer, V., Stampar, F., Veberic, R. and Koron, D. (2014). Changes in phenolic content induced by infection with *Didymella applanata* and *Leptosphaeria coniothyrium*, the causal agents of raspberry spur and cane blight. *Plant Pathology*, 63: 185-192.
- Mirmazloum, I., Kiss, A., Ladányi, M. and György, Z. (2019). Production of cinnamyl alcohol glycosides by biotransformation in roseroot callus cells. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 139: 1-9.
- Mishra, M.R., Srivastava, R.K. and Akhtar, N. (2018). Enhanced alkaloid production from cell culture system of *Catharanthus roseus* in combined effect of nutrient salts, sucrose and plant growth regulators. *Journal of Biotechnology and Biomedical Science*, 4: 14-34.
- Mohamad, M., Mohsin, H. F. and Singh, G. K. S. (2017). A study on the effect of *Eurycoma longifolia* and *Polyalthia bullata* on reproductive organs and androgens of male sprague-dawley rats. *Journal of Engineering and Applied Sciences*, 12: 6994-6999.
- Mohammed, A., Chiruvella, K. K., Rao, Y. K., Geethangili, M., Raghavan, S. C. and Ghanta, R. G. (2015). *In vitro* production of echiodinin, 7-o-methywogonin from callus cultures of *Andrographis lineata* and their cytotoxicity on cancer cells. *PLoS One*, 10: e0141154-e0141168.
- Moreira, R., Pereira, D. M., Valentão, P. and Andrade, P. B. (2018). Pyrrolizidine alkaloids: chemistry, pharmacology, toxicology and food safety. *International Journal of Molecular Sciences*, 19: 1668-1690.
- Moura, L. C. D., Xavier, A., Cruz, A. C. F. D., Gallo, R., Gatti, K. C., Miranda, N. A., and Otoni, W. C. (2017). Effects of explant type, culture media and picloram and dicamba growth regulators on induction and proliferation of somatic embryos in *Eucalyptus grandis* x *E. urophylla*. *Revista Árvore*, 41: e410502-e410512.
- Muhamad, S. N. S., Ling, A. P. K. and Wong, C. L. (2018). Effect of plant growth regulators on direct regeneration and callus induction from *Sargassum polycystum* C. Agardh. *Journal of Applied Phycology*, 30: 3299-3310.
- Mujeeb, F., Bajpai, P. and Pathak, N. (2014). Phytochemical evaluation, antimicrobial activity, and determination of bioactive components from leaves of *Aegle marmelos*. *BioMed Research International*, 2014: 1-11.
- Murashige, T. and Skoog, F. (1962). A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiologia Plantarum*, 15: 473-497.
- Nagavani, V., Madhavi, Y., Rao, D. B., Rao, P. K. and Rao, T. R. (2010). Free radical scavenging activity and qualitative analysis of polyphenols by RP-HPLC in the flowers of *Couroupita guianensis* Abul. *Electronic Journal of Environmental, Agricultural and Food Chemistry*, 9: 1471-1478.

- Naidu, J. R., Ismail, R. B., Yeng, C., Sasidharan, S. and Kumar, P. (2012). Chemical composition and antioxidant activity of the crude methanolic extracts of *Mentha spicata*. *Journal of Phytological Research*, 4: 13-18.
- Naik, P. M., and Al-Khayri, J. M. (2016). Impact of abiotic elicitors on *in vitro* production of plant secondary metabolites: a review. *Journal of Advanced Research in Biotechnology*, 1: 1-7.
- Namdeo, A. G. (2007). Plant cell elicitation for production of secondary metabolites: a review. *Pharmacognosy Review*, 1: 69-79.
- Nanda, A. K. and Melnyk, C. W. (2018). The role of plant hormones during grafting. *Journal of Plant Research*, 131: 49-58.
- Nantapap, S. S., Punyanitya, N., Nuntasaen, W., Pompimon, W. and Meepowpan, P. (2017). Flavones from aerial parts of *Polyalthia bullata* and cytotoxicity against cancer cell lines. *Chemistry of Natural Compounds*, 53: 762-763.
- Narayani, M., Chadha, A. and Srivastava, S. (2017). Callus and cell suspension culture of *Viola odorata* as *in vitro* production platforms of known and novel cyclotides. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 130: 289-299.
- Nasution, N. H. and Nasution, I. W. (2019). The effect of plant growth regulators on callus induction of mangosteen (*Garcinia mangostana* L.). *IOP Conference Series: Earth and Environmental Science*, 305: 012049-012056.
- Naz, S. and Khatoon, K. (2014). The effect of auxins on callus induction in *Achyranthes aspera*. *Pakistan Journal of Botany*, 46: 2203-2207.
- Ncube, B. and Van, J. S. (2015). Tilting plant metabolism for improved metabolite biosynthesis and enhanced human benefit. *Molecules*, 20: 12698-12731.
- Nevzorova, Y. A., Grossmann, J. and Trautwein, C. (2017). Anti-tumorigenic and anti-angiogenic effects of natural conifer *Abies sibirica* terpenoids *in vivo* and *in vitro*. *Biomedicine and Pharmacotherapy*, 89: 386-395.
- Ng, Y. P., Or, T. C. T. and Ip, N. Y. (2015). Plant alkaloids as drug leads for Alzheimer's disease. *Neurochemistry International*, 89: 260-270.
- Nimse, S. B. and Pal, D. (2015). Free radicals, natural antioxidants, and their reaction mechanisms. *Rsc Advances*, 5: 27986-28006.
- Nishiumi, S., Miyamoto, S., Kawabata, K., Ohnishi, K., Mukai, R., Murakami, A., Ashida, H. and Terao, J. (2011). Dietary flavonoids as cancer-preventive and therapeutic biofactors. *Frontiers in Bioscience*, 3: 1332-1362.
- Osman, N. I., Sidik, N. J. and Awal, A. (2016). Effects of variations in culture media and hormonal treatments upon callus induction potential in endosperm explant of

Barringtonia racemosa L. *Asian Pacific Journal of Tropical Biomedicine*, 6: 143-147.

- Osuntokun, O. T. and Cristina, G. M. (2019). Bio-guided isolation, chemical purification, identification, antimicrobial and synergistic efficacy of extracted essential oils from stem bark extract of *Spondias mombin* (Linn). *International Journal of Molecular Biology: Open Access*, 4: 135-143.
- Paarakh, P. M. and Khosa, R. L. (2009). Phytoconstituents from the genus Polyalthia- a review. *Journal of Pharmacy Research*, 2: 594-605.
- Panche, A. N., Diwan, A. D. and Chandra, S. R. (2016). Flavonoids: an overview. *Journal of Nutritional Science*, 5: 1-15.
- Parthasarathy, A., Cross, P. J., Dobson, R., Adams, L. E., Savka, M. A. and Hudson, A. O. (2018). A three-ring circus: metabolism of the three proteogenic aromatic amino acids and their role in the health of plants and animals. *Frontiers in Molecular Biosciences*, 5: 29-40.
- Parthipan, B., Suky, M. G. T. and Mohan, V. R. (2015). GC-MS analysis of phytocomponents in *Pleiospermum alatum* (Wall. ex Wight & Arn.) Swingle,(Rutaceae). *Journal of Pharmacognosy and Phytochemistry*, 4: 216-222.
- Patel, D. K., Prasad, S. K., Kumar, R. and Hemalatha, S. (2012). An overview on antidiabetic medicinal plants having insulin mimetic property. *Asian Pacific Journal of Tropical Biomedicine*, 2: 320-330.
- Pavarini, D. P., Pavarini, S. P., Niehues, M. and Lopes, N. P. (2012). Exogenous influences on plant secondary metabolite levels. *Animal Feed Science and Technology*, 176: 5-16.
- Perveen, S. and Al-Taweel, A. (2018). Introductory Chapter: Terpenes and Terpenoids. In Perveen, S. (Ed.), *Terpenes and Terpenoids* (pp. 1-12). United Kingdom: IntechOpen.
- Perviz, S., Khan, H. and Pervaiz, A (2016). Plant alkaloids as an emerging therapeutic alternative for the treatment of depression. *Frontiers in Pharmacology*, 7: 1-7.
- Pham, H. N. T., Nguyen, V. T., Vuong, Q., V., Bowyer, M., C. and Scarlett, C. J. (2015). Effect of extraction solvents and drying methods on the physicochemical and antioxidant properties of *Helicteres hirsuta* Lour. leaves. *Technologies*, 3: 285-301.
- Pillai, S. K. and Siril, E. A. (2019). Enhanced production of berberine through callus culture of *Tinospora cordifolia* (Willd.) Miers ex Hook F. and Thoms. *Proceedings of the National Academy of Sciences, India Section B: Biological Sciences*, 1-9.

Pliankong, P., Suksa-Ard, P. and Wannakrairoj, S. (2018). Chitosan elicitation for enhancing of vincristine and vinblastine accumulation in cell culture of *Catharanthus roseus* (L.) G. Don. *Journal of Agricultural Sciences*, 10: 287–293.

Polyalthia bullata King in GBIF Backbone Taxonomy. N.d. Retrieved on 30 September 2019 from <https://doi.org/10.15468/39omei>

Ptak, A., Simlat, M., Morańska, E., Skrzypek, E., Warchoł, M., Tarakemeh, A. and Laurain-Mattar, D. (2019). Exogenous melatonin stimulated Amaryllidaceae alkaloid biosynthesis in *in vitro* cultures of *Leucojum aestivum* L. *Industrial Crops and Products*, 138: 111458-111466.

Ptak, A., Tahchy, A., Skrzypek, E., Wójtowicz, T. and Laurain-Mattar, D. (2013). Influence of auxins on somatic embryogenesis and alkaloid accumulation in *Leucojum aestivum* callus. *Open Life Sciences*, 8: 591-599.

Radušienė, J., Karpavičienė, B. and Stanius, Ž. (2012). Effect of external and internal factors on secondary metabolites accumulation in St. John's worth. *Botanica Lithuanica*, 18: 101-108.

Rafieian-Kopaei, M. (2012) Medicinal plants and the human needs. *Journal of Herbmed Pharmacology*, 1: 1-2.

Rahayu, S., Roostika, I. and Bermawie, N. (2016). The effect of types and concentrations of auxins on callus induction of *Centella asiatica*. *Nusantara Bioscience*, 8: 283-287.

Raj, D., Kokotkiewicz, A., Drys, A., and Luczkiewicz, M. (2015). Effect of plant growth regulators on the accumulation of indolizidine alkaloids in *Securinega suffruticosa* callus cultures. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 123: 39-45.

Ramakrishna, A. and Ravishankar, G. A. (2011). Influence of abiotic stress signals on secondary metabolites in plants. *Plant Signaling Behaviour*, 6: 1720-1731.

Ramirez-Estrada, K., Vidal-Limon, H., Hidalgo, D., Moyano, E., Golenioswki, M., Cusidó, R. and Palazon, J. (2016). Elicitation, an effective strategy for the biotechnological production of bioactive high-added value compounds in plant cell factories. *Molecules*, 21: 182-206.

Rao, S., Usha, K., and Arjun. (2015). Production of secondary metabolites from callus cultures of *Centella asiatica* (L.) Urban. *Annals of Phytomedicine-An International Journal*, 4: 74-78.

Ratnadewi, D. (2017). Alkaloids in plant cell cultures. In Georgiev, V. and Pavlov, A. (Ed.), *Alkaloids-Alternatives in Synthesis, Modification and Application* (pp. 1-23). United Kingdom: IntechOpen.

- Rezaei, M. and Ghasemi Pirbalouti, A. (2019). Phytochemical, antioxidant and antibacterial properties of extracts from two spice herbs under different extraction solvents. *Journal of Food Measurement and Characterization*, 13: 2470-2480.
- Ribeiro, I. G., Gayer, C. R. M., de Castro, T. C., Coelho, M. G. P. and Albarello, N. (2015). Compact callus cultures and evaluation of the antioxidant activity of *Hovenia dulcis* Thunb. (Rhamnaceae) under *in vivo* and *in vitro* culture conditions. *Journal of Medicinal Plant Research*, 9: 8-15.
- Saeed, N., Khan, M. R. and Shabbir, M. (2012). Antioxidant activity, total phenolic and total flavonoid contents of whole plant extracts *Torilis leptophylla* L. *BMC Complementary and Alternative Medicine*, 12: 221-233.
- Sahin, G., Verma, S. K. and Gurel, E. (2013). Calcium and magnesium elimination enhances accumulation of cardenolides in callus cultures of endemic *Digitalis* species of Turkey. *Plant Physiology and Biochemistry*, 73: 139-143.
- Salehi, M., Moieni, A. and Safaie, N. (2017). A novel medium for enhancing callus growth of hazel (*Corylus avellana* L.). *Scientific Reports*, 7: 15598-15606.
- Santos, M. R. A. D., Souza, C. A. D. and Paz, E. S. (2017). Growth pattern of friable calluses from leaves of *Capsicum annuum* var *annuum* cv Iberaba Jalapeño. *Revista Ciencia Agronomica*, 48: 523-530.
- Šaponjac, V.T., Čanadanović-Brunet, J., Ćetković, G. and Djilas, S. (2016). Detection of bioactive compounds in plants and food products. In Nedović, V., Raspor, P., Lević, J., Tumbas Šaponjac, V. and Barbosa-Cánovas, G. (Ed.), *Emerging and Traditional Technologies for Safe, Healthy and Quality Food. Food Engineering Series* (pp 81-109), Cham: Springer.
- Satyal, P., Murray, B. L., McFeeters, R. L. and Setzer, W. N. (2016). Essential oil characterization of *Thymus vulgaris* from various geographical locations. *Foods*, 5: 70-82.
- Sayhan, H., Beyaz, S. G. and Çeliktaş, A. (2017). The local anesthetic and pain relief activity of alkaloids. In Georgiv, V. (Ed.), *Alkaloids: Alternatives in Synthesis, Modification and Application* (pp. 57-84). United Kingdom: IntechOpen.
- Schäfer, H. and Wink, M. (2009). Medicinally important secondary metabolites in recombinant microorganisms or plants: progress in alkaloid biosynthesis. *Biotechnology Journal: Healthcare Nutrition Technology*, 4: 1684-1703.
- Schaller, G. E., Bishopp, A. and Kieber, J. J. (2015). The yin-yang of hormones: cytokinin and auxin interactions in plant development. *The Plant Cell*, 27: 44-63.
- Schenk, R. U. and Hildebrandt, A. C. (1972). Medium and techniques for induction and growth of monocotyledonous and dicotyledonous plant cell cultures. *Canadian Journal of Botany*, 50: 199-204.

- Scherer, R., Lemos, M. F., Martinelli, G. C., Martins, J. D. L. and da Silva, A. G. (2013). Antioxidant and antibacterial activities and composition of Brazilian spearmint (*Mentha spicata* L.). *Industrial Crops and Products*, 50: 408-413.
- Semwal, P., Painuli, S., Badoni, H. and Bacheti, R. K. (2018). Screening of phytoconstituents and antibacterial activity of leaves and bark of *Quercus leucotrichophora* A. Camus from Uttarakhand Himalaya. *Clinical Phytoscience*, 4: 30-36.
- Seo, J., Lee, S., Elam, M. L., Johnson, S. A., Kang, J. and Arjmandi, B. H. (2014). Study to find the best extraction solvent for use with guava leaves (*Psidium guajava* L.) for high antioxidant efficacy. *Food Science and Nutrition*, 2: 174-180.
- Shaheed, K. A., AlGaraawi, N. I., Alsultany, A. K., Abbas, Z. H., Khshayyish, I. K. and Al Khazali, M. T. (2019). Analysis of bioactive phytochemical compound of (*Cyperus iria* L.) by using gas chromatography–mass spectrometry. *IOP Conference Series: Earth and Environmental Science*, 388: 012064-012076.
- Shahzad, A., Sharma, S., Parveen, S., Saeed, T., Shaheen, A., Akhtar, R., Yadav, V., Upadhyay, A. and Ahmad, Z. (2017). Historical perspective and basic principles of plant tissue culture. In Abdin, M., Kiran, U. and Kamaluddin, A. A. (Ed.), *Plant Biotechnology: Principles and Applications* (pp. 1-36). Singapore ; Springer.
- Shaikh, R., Pund, M., Dawane, A. and Iliyas, S. (2014). Evaluation of anticancer, antioxidant, and possible anti-inflammatory properties of selected medicinal plants used in Indian traditional medication. *Journal of Traditional and Complementary Medicine*, 4: 253–257.
- Sharma, A. and Zheng, B. (2019). Molecular responses during plant grafting and its regulation by auxins, cytokinins, and gibberellins. *Biomolecules* 9: 397-416.
- Sharma, A., Mathur, A. K., Ganpathy, J., Joshi, B. and Patel, P. (2019). Effect of abiotic elicitation and pathway precursors feeding over terpenoid indole alkaloids production in multiple shoot and callus cultures of *Catharanthus roseus*. *Biologia*, 74: 543-553.
- Sharma, P., Patil, A. and Patil, D. (2017). Effect of culture media and growth hormones on callus induction in *Crataeva tapia* L. *International Journal of Pharmaceutical Research*, 9: 70-76.
- Sharmila, M., Rajeswari, M., Jayashree, I. and Geetha, D. H. (2016). GC-MS analysis of bioactive compounds of *Amaranthus polygonoides* Linn. (Amaranthaceae). *International Journal of Applied and Advanced Scientific Research (IJAASR)*, 1: 174-180.
- Shi, Q. I. U., Hui, S. U. N., Zhang, A. H., Hong-Ying, X. U., Guang-Li, Y. A. N., Ying, H. A. N. and Xi-Jun, W. A. N. G. (2014). Natural alkaloids: basic aspects,

- biological roles, and future perspectives. *Chinese Journal of Natural Medicines*, 12: 401-406.
- Shinde, A. N., Malpathak, N. and Fulzele, D. P. (2009). Enhanced production of phytoestrogenic isoflavones from hairy root cultures of *Psoralea corylifolia* L. using elicitation and precursor feeding. *Biotechnology and Bioprocess Engineering*, 14: 288-294.
- Shobi, T. M. and Viswanathan, M. B. G. (2018). Antibacterial activity of di-butyl phthalate isolated from *Begonia malabarica*. *Journal of Applied Biotechnology and Bioengineering*, 5: 97-100.
- Shon, M. S., Lee, Y., Song, J. H., Park, T., Lee, J. K., Kim, M., Park, E. and Kim, G. N. (2014). Anti-aging potential of extracts prepared from fruits and medicinal herbs cultivated in the Gyeongnam area of Korea. *Preventive Nutrition and Food Science*, 19: 178–186.
- Singh, B., Virk, G. S. and Nagpal, A. K. (2011). An efficient plant regeneration protocol from callus cultures of *Citrus jambhiri* Lush. *Physiology and Molecular Biology of Plants*, 17: 161-169.
- Singh, R. and Chaturvedi, P. (2019). Phytochemical characterization of rhizome, fruit, leaf and callus of *Rheum emodi* Wall. using GC-MS. *Pharmacognosy Journal*, 11: 617-623.
- Singh, R., Rai, M. K. and Kumari, N. (2015). Somatic embryogenesis and plant regeneration in *Sapindus mukorossi* Gaertn. from leaf-derived callus induced with 6-benzylaminopurine. *Applied Biochemistry and Biotechnology*, 177: 498-510.
- Singh, T., Yadav, R. and Agrawal, V. (2020). Effective protocol for isolation and marked enhancement of psoralen, daidzein and genistein in the cotyledon callus cultures of *Cullen corylifolium* (L.) Medik. *Industrial Crops and Products*, 143: 111905-111915.
- Sivanandhan, G., Selvaraj, N., Ganapathi, A. and Manickavasagam, M. (2014). Enhanced biosynthesis of withanolides by elicitation and precursor feeding in cell suspension culture of *Withania somnifera* (L.) Dunal in shake-flask culture and bioreactor. *PLoS One*, 9: e104005-e104016.
- Skrzypczak-Pietraszek, E., Słota, J. and Pietraszek, J. (2014). The influence of L-phenylalanine, methyl jasmonate and sucrose concentration on the accumulation of phenolic acids in *Exacum affine* Balf. f. ex Regel shoot culture. *Acta Biochimica Polonica*, 61: 47-53.
- Small, C. C. and Degenhardt, D. (2018). Plant growth regulators for enhancing revegetation success in reclamation: a review. *Ecological Engineering*, 118: 43-51.

- Smetanska, I. (2008). Production of secondary metabolites using plant cell cultures. *Food Biotechnology*, 187-228.
- Song, Y. (2014). Insight into the mode of action of 2, 4- dichlorophenoxyacetic acid (2, 4- D) as an herbicide. *Journal of Integrative Plant Biology*, 56: 106-113.
- Souza, J. M., Berkov, S. and Santos, A. S. (2014). Improvement of friable callus production of Boerhaavia paniculata Rich and the investigation of its lipid profile by GC/MS. *Anais da Academia Brasileira de Ciências*, 86: 1015-1027.
- Sree, S. J., Vijayakumar, N. and Gomathi, K. S. (2016). Effect of plant growth regulators on callus induction in *Atalantia monophylla* and *Pamburus missionis*. *International Journal of Current Research in Biosciences and Plant Biology*, 3: 21-25.
- Srinivasan, K. (2014). Antioxidant potential of spices and their active constituents. *Critical Reviews in Food Science and Nutrition*, 54: 352-372.
- Stevenson, P. C., Nicolson, S. W. and Wright, G. A. (2017). Plant secondary metabolites in nectar: impacts on pollinators and ecological functions. *Functional Ecology*, 31: 65-75.
- Stohs, S. J. and Bagchi, D. (2015). Antioxidant, anti-inflammatory, and chemoprotective properties of *Acacia catechu* heartwood extracts. *Phytotherapy Research*, 29: 818-824.
- Subha, J. S. and Divakar, K. M. (2016). GC-MS analysis of the phytoconstituents of methanolic fruit extracts of *Terminalia chebula* Retz. *International Journal of Innovative Pharmaceutical Sciences and Research*, 4: 53-61.
- Sudha, G. and Ravishankar, G. A. (2003). Elicitation of anthocyanin production in callus cultures of *Daucus carota* and involvement of calcium channel modulators. *Current Science*, 84: 775-779.
- Sudhan, P. N., Ghashang, M. and Mansoor, S. S. (2016). Tribromo melamine-catalyzed one-pot synthesis of a series of 4-aryl-4, 5-dihydro-1 H-indeno [1,2-b] pyridine derivatives. *Journal of Taibah University for Science*, 10: 709-717.
- Sumazian, Y., Syahida, A., Hakiman, M., and Maziah, M. (2010). Antioxidant activities, flavonoids, ascorbic acid and phenolic contents of Malaysian vegetables. *Journal of Medicinal Plants Research*, 4: 881-890.
- Summart, J., Panichajakul, S., Prathepha, P. and Thanonkeo, P. (2008). Callus induction and influence of culture condition and culture medium on growth of Thai aromatic rice, Khao Dawk Mali 105, cell culture. *World Applied Sciences Journal*, 5: 246-251.

- Sun, C., Li, X., Xu, C., Zhang, S., Chen, K., Chen, Q. and Liu, C. (2007). Determination of 9 (10 H)- Acridone by HPLC with fluorescence detection. *Journal of Liquid Chromatography & Related Technologies*, 30: 245-254.
- Suwanseree, V., Phansiri, S. and Yapwattanaphun, C. (2019). A comparison of callus induction in 4 *Garcinia* species. *Electronic Journal of Biotechnology*, 40: 45-51.
- Swamy, M. K., Arumugam, G., Kaur, R., Ghasemzadeh, A., Yusoff, M. M. and Sinniah, U. R. (2017). GC-MS based metabolite profiling, antioxidant and antimicrobial properties of different solvent extracts of Malaysian *Plectranthus amboinicus* leaves. *Evidence-Based Complementary and Alternative Medicine*, 1-10.
- Szopa, A. and Ekiert, H. (2014). Production of biologically active phenolic acids in *Aronia melanocarpa* (Michx.) Elliott *in vitro* cultures cultivated on different variants of the Murashige and Skoog medium. *Plant Growth Regulation*, 72: 51-58.
- Takaidza, S., Mtunzi, F. and Pillay, M. (2018). Analysis of the phytochemical contents and antioxidant activities of crude extracts from *Tulbaghia* species. *Journal of Traditional Chinese Medicine*, 38: 272-279.
- Thavamoney, N., Sivanadian, L., Tee, L. H., Khoo, H. E., Prasad, K. N. and Kong, K. W. (2018). Extraction and recovery of phytochemical components and antioxidative properties in fruit parts of *Dacryodes rostrata* influenced by different solvents. *Journal of Food Science and Technology*, 55: 2523-2532.
- Tholl, D. (2015). Biosynthesis and biological functions of terpenoids in plants. In: Schrader J., Bohlmann J. (Ed.), *Biotechnology of Isoprenoids. Advances in Biochemical Engineering/Biotechnology* (pp. 63-106). Cham: Springer.
- Tian, W., Zhi, H., Yang, C., Wang, L., Long, J., Xiao, L., Liang, J., Huang, Y., Zheng, X., Zhao, S., Zhang, K. and Zheng, J. (2018). Data on chemical composition of alkaloids of *Plumula nelumbinis* and antioxidant activity from thirteen habitats in China. *Data in Brief*, 21:1591-1597.
- Tomsone, L., Kruma, Z. and Galoburda, R. (2012). Comparison of different solvents and extraction methods for isolation of phenolic compounds from horseradish roots (*Armoracia rusticana*). *World Academy of Science, Engineering and Technology*, 64: 903-908.
- Truong, D. H., Nguyen, D. H., Ta, N. T. A., Bui, A. V., Do, T. H. and Nguyen, H. C. (2019). Evaluation of the use of different solvents for phytochemical constituents, antioxidants, and *in vitro* anti-inflammatory activities of *Severinia buxifolia*. *Journal of Food Quality*, 2019: 1-9.
- Turner, I. M. and Utteridge, T. M. A. (2016). Whither Polyalthia (Annonaceae) in Peninsular Malaysia? Synopses of Huberantha, Maasia, Monoon and Polyalthia s.s. *European Journal of Taxonomy*, 183: 1-26.

- Tyagi, T. and Agarwal, M. (2017). Phytochemical screening and GC-MS analysis of bioactive constituents in the ethanolic extract of *Pistia stratiotes* L. and *Eichhornia crassipes* (Mart.) solms. *Journal of Pharmacognosy and Phytochemistry*, 6: 195-206.
- Upadhyay, S. and Koche, V. (2015). Comparison of different medium and establishment of an efficient micropropagation technique of *Clerodendrum serratum* L. An endangered medicinal plant. *IOSR Journal of Environmental Science, Toxicology and Food Technology (IOSR-JESTFT)*, 2: 37-45.
- Vagiri, M., Johansson, E. and Rumpunen, K. (2017). Phenolic compounds in black currant leaves—an interaction between the plant and foliar diseases? *Journal of Plant Interactions*, 12: 193-199.
- Vauzour, D., Rodriguez-Mateos, A., Corona, G., Oruna-Concha, M. J. and Spencer, J. P. (2010). Polyphenols and human health: prevention of disease and mechanisms of action. *Nutrients*, 2: 1106-1131.
- Veraplakorn, V. (2016). Micropropagation and callus induction of *Lantana camara* L.—a medicinal plant. *Agriculture and Natural Resources*, 50: 338-344.
- Vergara-Jimenez, M., Almatrafi, M. and Fernandez, M. (2017). Bioactive components in *Moringa oleifera* leaves protect against chronic disease. *Antioxidants*, 6: 91-103.
- Verma, A. K., Singh, R. R. and Singh, S. (2011). Improved alkaloid content in callus culture of *Catharanthus roseus*. *Botanica Serbica*, 36: 123-130.
- Verma, R. S., Padalia, R. C., Chanotiya, C. S. and Chauhan, A. (2010). Chemical investigation of the essential oil of *Thymus linearis* (Benth. ex Benth) from western Himalaya, India. *Natural Product Research*, 24: 1890-1896.
- Verma, S. K., Das, A. K., Cingoz, G. S., Uslu, E. and Gurel, E. (2016). Influence of nutrient media on callus induction, somatic embryogenesis and plant regeneration in selected Turkish crocus species. *Biotechnology Reports*, 10: 66-74.
- Virmala, S. (2013). Malaysian herbal heritage. pp 60-61, Forest Reserach Institute Malaysia (FRIM): Kuala Lumpur.
- Wadood, A., Ghufran, M., Jamal, S. B., Naeem, M., Khan, A. and Ghaffar, R. (2013). Phytochemical analysis of medicinal plants occurring in local area of Mardan. *Biochemistry and Analytical Biochemistry*, 2: 1-4.
- Wang, C., Wan, J., Mei, Z. and Yang, X. (2014). Acridone alkaloids with cytotoxic and antimalarial activities from *Zanthoxylum simulans* Hance. *Pharmacognosy Magazine*, 10: 73-76.
- White, P. R. (1964). The cultivation of animal and plant cells. *Soil Science*, 97: 74-75.

- Widyawati, P. S., Budianta, T. D. W., Kusuma, F. A. and Wijaya, E. L. (2014). Difference of solvent polarity to phytochemical content and antioxidant activity of *Pluchea indica* less leaves extracts. *Journal of Pharmacognosy and Phytochemistry*, 6: 850-855.
- Wink, M. (2015). Modes of action of herbal medicines and plant secondary metabolites. *Medicines*, 2: 251-286.
- Xin, Z., Yu, Z., Erb, M., Turlings, T. C., Wang, B., Qi, J., Liu, S. and Lou, Y. (2012). The broad- leaf herbicide 2, 4- dichlorophenoxyacetic acid turns rice into a living trap for a major insect pest and a parasitic wasp. *New Phytologist*, 194: 498-510.
- Xue, B., Su, Y. C. F., Thomas, D. C. and Saunders, R. M. K. (2012). Pruning the polyphyletic genus *Polyalthia* (Annonaceae) and resurrecting the genus monoon. *Taxon*, 61: 1021-1039.
- Yadav, M., Chatterji, S., Gupta, S. K. and Watal, G. (2014). Preliminary phytochemical screening of six medicinal plants used in traditional medicine. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6: 2-14.
- Yamuna, P., Abirami, P., Vijayashalini, P. and Sharmila, M. (2017). GC-MS analysis of bioactive compounds in the entire plant parts of ethanolic extract of *Gomphrena decumbens* Jacq. *Journal of Medicinal Plants Studies*, 5: 31-37.
- Yang, L., Wen, K. S., Ruan, X., Zhao, Y. X., Wei, F. and Wang, Q. (2018). Response of plant secondary metabolites to environmental factors. *Molecules*, 23: 762-788.
- Yao, L. J., Jalil, J., Attiq, A., Hui, C. C. and Zakaria, N. A. (2018). The medicinal uses, toxicities and anti-inflammatory activity of *Polyalthia* species (Annonaceae). *Journal of Ethnopharmacology*, 229: 303-325.
- Zahid, M., Arif, M., Rahman, M. A., Singh, K. and Mujahid, M. (2018). Solvent extraction and gas chromatography-mass spectrometry analysis of *Annona squamosa* L. seeds for determination of bioactives, fatty acid/fatty oil composition, and antioxidant activity. *Journal of Dietary Supplements*, 15: 613-623.
- Zang, Q., Zhou, L., Zhuge, F., Yang, H., Wang, X. and Lin, X. (2016). Callus induction and regeneration via shoot tips of *Dendrocalamus hamiltonii*. *SpringerPlus*, 5: 1799-1805.
- Zhang, X., Jin, Y., Wu, Y., Zhang, C., Jin, D., Zheng, Q. and Li, Y. (2018). Anti-hyperglycemic and anti-hyperlipidemia effects of the alkaloid-rich extract from barks of *Litsea glutinosa* in ob/ob mice. *Scientific Reports*, 8: 12646-12656.
- Zhu, J., Wang, M., Wen, W. and Yu, R. (2015). Biosynthesis and regulation of terpenoid indole alkaloids in *Catharanthus roseus*. *Pharmacognosy Reviews*, 9: 24-28.

Zlotek, U., Mikulska, S., Nagajek, M. and Świeca, M. (2016). The effect of different solvents and number of extraction steps on the polyphenol content and antioxidant capacity of basil leaves (*Ocimum basilicum* L.) extracts. *Saudi Journal of Biological Sciences*, 23: 628-633.

Zuldin, M., Nahazima, N., Said, I. M., Mohd Noor, N., Zainal, Z., Jin Kiat, C. and Ismail, I. (2013). Induction and analysis of the alkaloid mitragynine content of a *Mitragyna speciosa* suspension culture system upon elicitation and precursor feeding. *The Scientific World Journal*, 2013: 1-11.