



UNIVERSITI PUTRA MALAYSIA

***DEVELOPMENT OF A BIO-NANOGATE-BASED ELECTROCHEMICAL
IMMUNOSENSING STRATEGY FOR THE DETECTION OF
ANTI-HEPATITIS B SURFACE ANTIGEN ANTIBODY***

NOOR SYAMILA BINTI OTHMAN

FBSB 2022 12



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By

NOOR SYAMILA BINTI OTHMAN

**Thesis Submitted to School of Graduate Studies, Universiti Putra
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of Philosophy**

March 2022

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

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NOOR SYAMILA BINTI OTHMAN

March 2022

Chair: Asilah binti Ahmad Tajudin, PhD
Faculty: Biotechnology and Biomolecular Sciences

An area requiring real-time analysis is the diagnosis of infectious disease and monitoring of vaccination efficiency against the disease to determine immunity level particularly among high risk group including immunocompromised patients against infectious diseases in screening for immunization program including hepatitis B virus (HBV). However, the conventional methods require laborious work and tedious fabrication of sensing platforms which limit the efficiency in upscaling the screening tests. These further impede the analysis of large cohort of clinical samples. Therefore, an effort is needed in order to improve the outcome of this lab-based technology. Generally, a bio-nanogate system involves the use of synthetic or natural molecules as a 'gate' towards bioreceptors and ideally, the gating mechanism should respond only upon the presence of external stimuli i.e. targeted analytes in a nanoscale dimension. The versatility of polyamidoamine (PAMAM) dendrimers to form conjugates with proteins can be utilized to form a bio-nanogate. PAMAM interaction with protein bioreceptor and the ability of a bio-nanogate-based immunosensing strategy in detecting an antibody i.e. anti-hepatitis B surface antigen (anti-HBsAg) antibody electrochemically were of interest in this study.

An antigenic determinant (aD) region of HBV fused with maltose binding protein (MBP-aD) was synthesized to form a specific bioreceptor for anti-HBsAg antibody in the bio-nanogate system. The bio-nanogate interaction was further analysed for its binding affinity, thermal stability, and thermodynamic analysis. Following that, a proof of concept utilizing displacement immunosensing strategy was conducted electrochemically, where the MBP-aD was immobilized on the screen-printed carbon electrode (SPCE) platform, and further sandwiched with electroconductive PAMAM encapsulated gold nanoparticles (PAMAM-Au), forming the 'gate'.

PAMAM-Au here also functions as a monitoring agent capable of generating a signal response upon a displacement event in the presence of anti-HBsAg antibody in differential pulse voltammetry (DPV) analysis. Finally, the PAMAM-Au displacement efficiency was further improved via implementation of acoustic mixing on modified SPCE platform coupled with piezoelectric transducer.

The synthesized MBP-aD was confirmed with western blotting technique. The interaction study revealed that the interaction of MBP-aD with anti-HBsAg antibody has a higher thermal stability and binding affinity ($K_A = 1.6 \times 10^5 \text{ Lmol}^{-1}$) as compared to its interaction with PAMAM ($K_A = 2.9 \times 10^6 \text{ Lmol}^{-1}$). Thermodynamic parameters also demonstrated that the bio-nanogate components interact through van der Waals and hydrogen bonding. Based on the interaction study, it was hypothesized that the interactions among the bio-nanogate components may be able to be manipulated at the nanoscale level for the detection of anti-HBsAg antibody. Under optimal conditions, the hypothesis was proven that the high specificity of anti-HBsAg antibody towards MBP-aD displaced PAMAM-Au, in a range of 1 mIU/mL to 1000 mIU/mL with a detection limit (LOD) of 2.5 mIU/mL. The results also showed high specificity and selectivity of the immunosensor platform in detecting anti-HBsAg antibody both in spiked buffer and human serum samples. Furthermore, the incubation/reaction time for detecting anti-HBsAg antibody has been reduced from an initial incubation time of 20 min to 8 min via the improvement of PAMAM-Au displacement efficiency under acoustic streaming effect. The newly developed immunosensor platform utilizing the manipulation of lower interaction between PAMAM-Au (gate) and the candidate bioreceptor (anchor) would ultimately eliminate the need of having specifically designed and labelled analogues which has been commonly used in displacement-based immunoassays.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PEMBANGUNAN STRATEGI PENDERIAAN IMUNO ELEKTROKIMIA
BERDASARKAN GET NANO BIO UNTUK PENGESANAN ANTIBODI
BAGI ANTIGEN PERMUKAAN VIRUS HEPATITIS B**

Oleh

NOOR SYAMILA BINTI OTHMAN

Mac 2022

Pengerusi: Asilah binti Ahmad Tajudin, PhD
Fakulti: Bioteknologi dan Sains Biomolekul

Bidang yang memerlukan analisis waktu nyata adalah diagnosis penyakit berjangkit dan pemantauan kecekapan vaksinasi terhadap penyakit untuk menentukan kadar imuniti terutamanya golongan berisiko tinggi dan pesakit terimunokompromi terhadap penyakit berjangkit di dalam saringan program imunisasi termasuklah terhadap virus hepatitis B (HBV). Namun, kaedah konvensional yang memerlukan gerak kerja makmal yang rumit dan pembuatan platform pengesanan yang memakan masa mengurangi kecekapan dalam meningkatkan ujian saringan. Ini seterusnya membatasi analisis kohort sampel klinikal yang besar. Oleh itu, usaha diperlukan untuk meningkatkan hasil teknologi berdasarkan makmal ini. Secara amnya, sistem get nano bio melibatkan molekul yang disintesis atau molekul semula jadi sebagai 'pintu pagar' terhadap reseptor bio dan secara idealnya, mekanisme pintu pagar harus bertindak balas hanya apabila terdapat rangsangan luaran, iaitu analit yang disasarkan dalam dimensi skala nano. Keserbagunaan dendrimer poliamidoamina (PAMAM) untuk membentuk konjugat dengan protein dapat digunakan dalam membentuk get nano bio. Interaksi PAMAM dengan reseptor protein dan kemampuan strategi penderiaan imuno berasaskan get nano bio dalam mengesan antibodi iaitu antibodi anti-HBsAg, secara elektrokimia menarik minat kajian ini.

Bahagian penentuan antigen (aD) yang dilakurkan dengan protein penambat maltosa (MBP-aD) telah disintesis untuk membentuk reseptor bio khusus kepada antibodi anti-HBsAg dalam sistem get nano bio. Interaksi get nano bio dianalisis lebih lanjut untuk afiniti pengikat, kestabilan terma, dan analisis termodinamik. Berikutan itu, bukti konsep yang menggunakan strategi pengalihan dalam penderiaan imuno dilakukan secara elektrokimia, di mana MBP-aD dipegunkan di atas

platform skrin bercetak elektrod karbon (SPCE), dan selanjutnya dilapisi dengan nanopartikel emas berkapsul PAMAM (PAMAM-Au) yang elektrokonduktif untuk membentuk 'pintu pagar'. PAMAM-Au di sini juga berfungsi sebagai agen pemantauan yang mampu menghasilkan isyarat tindak balas apabila berlaku kejadian peralihan dengan kehadiran antibodi anti-HBsAg menerusi analisis voltammetri nadi pembezaan (DPV). Akhirnya, kecekapan peralihan PAMAM-Au ditingkatkan lagi melalui pelaksanaan pencampuran akustik pada platform SPCE yang digunakan bersama dengan transduser piezoelektrik.

MBP-aD yang disintesis disahkan dengan kaedah 'western blotting'. Kajian interaksi menunjukkan bahawa interaksi MBP-aD dengan antibodi anti-HBsAg mempunyai kestabilan terma dan afiniti pengikatan yang lebih tinggi ($K_A = 1.6 \times 10^{-5} \text{ Lmol}^{-1}$) berbanding interaksinya dengan PAMAM ($K_A = 2.9 \times 10^{-6} \text{ Lmol}^{-1}$). Parameter termodinamik menunjukkan bahawa komponen get nano bio berinteraksi melalui ikatan van der Waals dan hidrogen. Berdasarkan kajian interaksi, dihipotesiskan bahawa interaksi antara komponen get nano bio dapat dimanipulasi pada skala nano untuk pengesanan antibodi anti-HBsAg. Dalam keadaan optimum, hipotesis terbukti bahawa kekhususan tinggi antibodi anti-HBsAg terhadap MBP-aD dapat mengalihkan PAMAM-Au, dalam julat 1mIU / mL hingga 1000 mIU / mL dengan had pengesanan (LOD) 2.5 mIU / mL. Hasilnya juga menunjukkan kekhususan dan selektiviti tinggi platform immunosensor dalam mengesan antibodi anti-HBsAg dalam sampel larutan penimbal dan sampel serum manusia. Masa inkubasi / reaksi untuk mengesan antibodi anti-HBsAg juga dapat dikurangkan dari masa inkubasi awal 20 min menjadi 8 min melalui peningkatan kecekapan peralihan PAMAM-Au di bawah kesan penstriman akustik. Platform immunosensor yang baru dibangunkan menggunakan manipulasi interaksi rendah antara PAMAM-Au (pintu pagar) dan calon reseptor bio (jangkar) ini akan menyingkirkan keperluan dalam menggunakan analog yang direka dan dilabel secara khusus yang biasa digunakan dalam imunoasai berdasarkan peralihan.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Asilah binti Ahmad Tajudin, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Tan Wen Siang, PhD

Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Amir Syahir bin Amir Hamzah, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Haslina binti Ahmad, PhD

Associate Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

Yusran bin Sulaiman, PhD

Associate Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

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Signature: _____ Date: _____

Name and Matric No.: Noor Syamila binti Othman

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electrochemical immunosensor

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LIST OF ABBREVIATIONS

%	Percentage
[a]	Concentration of analyte
°C	Degree celsius
μ	Viscosity
μA	Microampere
μL	Microliter
μm	Micrometer
μM	micromolar
3D	Three-dimensional
$A_{600\text{nm}}$	Optical density at wavelength 600 nanometer
aa	amino acid
aD	antigenic determinant
AFM	Atomic force microscopy
Ag	silver
Anti-HBcAg	Anti-hepatitis B core antigen antibody
Anti-HBsAg	Anti-hepatitis B surface antigen antibody
AP	Alkaline phosphatase
APS	Ammonium persulfate
Au	Gold
AuCl_4^-	Tetrachloroaurate ion
<i>B</i>	Signal intensity
<i>b</i>	y-intercept
BCIP-NBT	5-Bromo-4-chloro-3-indolyl phosphate – nitro blue tetrazolium

BSA	Bovine serum albumin
C	Carbon
C=O	Carbonyl group
CaCl ₂	Calcium chloride
CIMA	Chemiluminescence assay
C—N	Carbon-nitrogen bond
CV	Cyclic voltammetry
dH ₂ O	Deionized water
DNA	Deoxyribonucleic acid
dNTP	Deoxynucleoside triphosphate
DPV	Differential pulse voltammetry
DTGS	Deuterated Triglycine sulfate
E	potential
<i>E. coli</i>	<i>Escherichia coli</i>
EI	Electrochemical immunosensor
EIS	Electrochemical impedance spectroscopy
<i>f</i>	Frequency
FESEM-EDX	field emission scanning electron microscopy with Energy Dispersive X-ray spectroscopy
F_{max}	Maximal fluorescence
F_{min}	Minimal fluorescence
FTIR	Fourier-transform infrared spectroscopy
g	Gram
H ⁺	Hydrogen ion
H ₂ PO ₄	Dihydrogen phosphate

HBsAg	Hepatitis B surface antigen
hr	Hour
Hz	Hertz
I	current
IgG	Immunoglobulin
IPTG	Isopropyl β -D-1-thiogalactopyranoside
J	Joule
K_A	Binding adsorption constant
K	Kelvin
$K_3Fe(CN)_6$	potassium hexacyanoferrate (III)
$K_4[Fe(CN)_6] \cdot 3H_2O$	potassium hexacyanoferrate (II) trihydrate
KBr / Ge	Potassium bromide / germanium
kDa	kilodaltons
kHz	kilo-Hertz
KPL	Milk diluent / blocking solution
L	Litre
LB	Luria-Bertani
LOD	Limit of detection
LSV	Linear sweep voltammetry
M	Molar
m	Slope
mA	Milliampere
MBP	Maltose binding protein
MBP-aD	Maltose binding protein fused with antigenic determinant

MCS	Multiple cloning sites
MgCl ₂	Magnesium chloride
MHz	Mega-Hertz
min	Minute
mIU	milli-international unit
MOPS	3-(N-morpholino)propanesulfonic acid
<i>N</i>	Number of available sites
Na ₂ HPO ₄	disodium hydrogen phosphate
NaCl	Sodium chloride
NaH ₂ PO ₄	sodium dihydrogen phosphate
Nano-DSF	Nano-differential scanning fluorometry
ng	Nanogram
N—H	Nitrogen-nitrogen bond
NH ₃ ⁺	Ammoniumyl
nm	Nanometer
NPs	Nanoparticles
O	Oxygen
<i>P</i>	Fluorescence intensity
PAMAM	Polyamidoamine dendrimer
PBS	Phosphate buffered solution
R	Resistance
RbCl	Rubidium chloride
R _{ct}	Resistant of charge transfer
RMS	Root mean square
rpm	Revolutions per minute

RT	Room temperature
SDS-PAGE	Sodium dodecyl sulphate-polyacrylamide gel electrophoresis
sec	Second
SEM	Scanning electron microscope
SPCE	Screen-printed carbon electrode
TBS	Tris-buffered saline
TBST	Tris-buffered saline with 0.1% TWEEN® 20 Detergent
TFBI	Transformation buffer I
TFBII	Transformation buffer II
T_i	Inflection temperature
Tris-HCl	Tris hydrochloride
Trp	Tryptophan
UV-vis	Ultraviolet-visible spectroscopy
V	Volt
V _{pp}	Voltage peak-to-peak
W	Watt
Y	Fluorescence ratio
β	Beta
ΔR_{ct}	Changes of resistant of charge transfer
$\Delta_A G^\circ$	standard Gibbs free energy of associated protein-polymer
$\Delta_A H^\circ$	enthalpy
$\Delta_A S^\circ$	Entropy
ζ -potential	Zeta potential

ρ density
 σ Viscous boundary thickness



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CHAPTER 1

INTRODUCTION

The emergence and re-emergence of viral infectious diseases including hepatitis B virus, (HBV), E-bola virus, immunodeficiency virus (HIV), and the recent severe acute respiratory syndrome (SARS)-associated coronavirus 2 (SARS-CoV-2) that caused COVID-19 have caused major public health concerns worldwide. The general transmission factor involving human as a host risks in viral infection attacking vital organs and this can cause acute or chronic diseases particularly in immunocompromised individuals. The protection against viral infection can be obtained through natural or acquired immunity such as vaccination, where it can trigger the immune response to recognize and fight against the viral-causing diseases.

Vaccination efficiency is typically assessed by determining the level of immunity developed in the body particularly among high risk group such as healthcare and foreign workers, immunocompromised patients and antenatal mothers during their visit in primary healthcare centres (Tam et al., 2017; Roupa et al., 2019). Monitoring antibodies against viral antigens is critical for maintaining immunity, especially in vaccinated individuals and immunocompromised patients (Grzegorzewska, 2012; Tan & Ho, 2014; Yong et al., 2015). Antibody screening tests frequently involve the detection of virus-specific antibodies through the use of conventional methods such as enzyme-linked immunosorbent assay (ELISA) and western blot analysis. Immunodetection assays identify and quantify specific targeted antibodies. The results of this detection are required in diagnostic and treatment procedures for viral infections. However, these conventional methods require laborious work and are time consuming. These limit the efficiency of upscaling the screening tests and further impede the analysis of large cohorts of clinical samples (Poiteau et al., 2017; Tam et al., 2017). Rapid diagnostic tests (RDTs) on the other hand, were developed to assist clinicians in preliminary screening and predicting the risk of reinfection in the infected individuals, particularly in remote areas. This adds benefit in disease surveillance and epidemiologic research. The most well-known RDT to date is probably the lateral flow tests, however they are limited in sensitivity, expensive, require multistep fabrication, and have reproducible issues due to the limitation of fade dyes in colorimetric-based sensor, which may lead to false negative results (Sher et al., 2017). Henceforth, more research is required to overcome the current limitations and develop reliable antibody detection systems in the future.

There are various advantages of using electrochemical-immunosensors for infectious disease markers. Among the benefits are ease of handling, robustness, portability, shorter detection times and relatively inexpensive analytical systems, all of which contribute to the development of point-of-care tests (POCTs) (Kaushik et al., 2018; Layqah & Eissa, 2019; Fabiani et al., 2021; Eissa & Zourob, 2021). Among them is the displacement-based electrochemical immunosensor, which uses the competitive-heterogenous-like immunoassay concept to measure signals from the immunocomplex formation via current or impedance change (Tang et al., 2018; Safarian et al., 2021). The displacement of non-specific binding of analyte analogue from bound bioreceptors in the presence of free targeted analyte offers a simple fabrication such as reagentless and washing-free steps. However, the high steric effect of analogue makes displacement difficult and occurred only after a longer incubation time (Khor et al., 2013; Lu et al., 2016; Avella-Oliver et al., 2018).

Hepatitis B virus (HBV) is a DNA virus that infects the human liver and causes both acute and chronic liver diseases. It affects approximately two billion people worldwide, of which around 257 million people suffering from chronic HBV infections (Papastergiou et al., 2015; Huzair & Sturdy, 2017; World Health Organization, 2019). A common serological marker of HBV, the anti-hepatitis B surface antigen antibody (anti-HBsAg antibody) indicates the immune status and the viral clearance in infected individuals. Such antibody is developed upon natural exposure towards HBV or through immunization with HBsAg (Pondé, 2011). To date, the majority of anti-HBsAg antibody detection is done using the conventional immunoassay, which has sensitivity issues when detecting low antibody titres and requires longer incubation time, limiting the turnaround analysis in time (Grzegorzewska, 2012; Tan & Ho, 2014; Yong et al., 2015). Meanwhile, RDT demonstrated low sensitivity in detecting low antibody titres and was only reliable in the presence of positive test (Bottero et al., 2013; Chevaliez & Pawlowsky, 2018). As a result, in this study, anti-HBsAg antibody was chosen as a model analyte marker for the formation of the current antibody detection system.

Recently, the fabrication of nanogate/nanoswitch has attracted the interest of researchers, particularly in developing biosensors and drug carriers, as the 'gating' mechanism. This 'gating' mechanism which can be in the form of synthesized polymers or biomolecules may possess the designated recognition sites that can respond selectively to external stimuli and is able to regulate signal release from a solid support (Tam et al., 2013; Bilalis et al., 2016; Huang & Szeleifer, 2017; Luo et al., 2019; Yang et al., 2020). This mechanism allows small molecules to be detected in a sensitive manner (Lee et al., 2016; Vivero-Escoto et al., 2017). Several studies have shown that the use of nanogate systems in conjunction with the use of porous materials can transiently hold the signalling molecules that are engulfed within the sensor surface (Wang et al., 2015; Huang & Szeleifer, 2017). Apart from that, the ability of DNA and aptamer-based molecules to form a 'gate' within a bio-nanogate system that relies on low interactions of

electrostatic, hydrophobic and hydrogen bonding also demonstrated selectivity towards targeting biological components (Wang et al., 2015). However, the aptamer-based bio-nanogate system's time-consuming fabrication and the requirement of multi-reagent steps limit the analysis turnaround time. The manipulation of 'gating' mechanism could be beneficial in improving the displacement-based electrochemical immunosensing strategy particularly for antibody detection. An appropriate component to function as the 'gate' in a bio-nanogate system is crucial for this purpose to ensure good control over its operation.

The sensitivity of the biosensors can also be improved by monitoring the small detectable current changes that may have occurred on the electrode. The current changes can be obtained when displacement-based immunosensor applies the non-specific binding components to react with the bioreceptors, and it will then be displaced with the presence of a specific targeted analyte (Zhang & Ding, 2016; Nadhakumar et al., 2018; Roy & Sharan, 2019). The displacement occurrences can be elevated by incorporating the acoustic streaming forces, which can manipulate and enhance the mass transport of surface-attached biomolecules by causing streaming-induced drag forces and swirling motion in the interacted media. Acoustic streaming has been reported to aid in many applications, including acceleration of DNA-hybridization assay, particle concentration, manipulation of micro/nanoparticles and fluid mixing within microfluidic channels (Liu et al., 2003; Daniels et al., 2011; Collins et al., 2016; Cui et al., 2016; Collins et al., 2017).

Therefore, this study was carried out to develop an immunosensor which incorporates a bio-nanogate mechanism to detect anti-HBsAg antibody on electrode surface, while utilizing electrochemical signalling, which in turn is crucial for the development of a POCT tool. The interaction of bio-nanogate key component biomolecules of synthesized maltose binding protein (MBP) harboring antigenic determinant (aD) region of HBsAg (MBP-aD) as a bioreceptor, PAMAM (the gate) and anti-HBsAg antibody was investigated. This is followed by the 'proof-of-concept' of the bio-nanogate-based immunosensing strategy employing the concept of displacement immunosensing via nonspecific-binding of PAMAM-Au to form weak interactions, which can act as a 'gate' towards its 'anchor' of potential bioreceptors. The presence of PAMAM-Au is also useful as a signal label for the monitoring of the oxidation peak changes upon displacement of the 'gate' from the 'anchor' in detecting the analyte using differential pulse voltammetry (DPV). The displacement efficiency of PAMAM-Au can be improved via acoustic streaming forces and reduces the standing incubation/reaction time of the developed immunosensor in targeting anti-HBsAg antibody.

1.1 Hypotheses

1. Bio-nanogate-based immunosensor utilizing displacement immunosensing strategy may improve rapidity, simplicity and sensitivity in targeting anti-HBsAg antibody, electrochemically.
2. The presence of the aD region as bioreceptor can enhance the specificity and accuracy of the sensing platform.
3. PAMAM will have lower binding interaction with the MBP-aD to form bio-nanogate.
4. The presence of anti-HBsAg antibody which has a higher affinity towards MBP-aD will displace PAMAM in the anti-HBsAg antibody detection system.
5. The synthesized PAMAM-Au can play a major role as a monitoring agent to monitor the displacement occurrences upon detection.
6. The incubation time for detecting anti-HBsAg antibody will be reduced via improvement of PAMAM-Au displacement efficiency under acoustic streaming effect.

1.2 General Objective

To develop an immunosensor which manipulates the interaction among bio-nanogate components at nanoscale level to improve simplicity, rapidity and sensitivity in targeting anti-HBsAg utilizing displacement immunosensing strategy, electrochemically

1.3 Specific Objectives

1. To synthesize the bioreceptor of maltose binding protein harbouring the antigenic determinant (MBP-aD) targeting the anti-HBsAg antibody in *Escherichia coli* expression system.
2. To analyze the interaction of bio-nanogate key components of MBP-aD, PAMAM and anti-HBsAg antibody using binding, thermal stability and thermodynamic analyses.
3. To synthesize and characterize the PAMAM encapsulated gold nanoparticles (PAMAM-Au)
4. To develop the bio-nanogate-based electrochemical immunosensor via non-specific binding of PAMAM-Au and MBP-aD for detection of anti-HBsAg antibody.
5. To improve the PAMAM-Au displacement efficiency via acoustic streaming forces for detecting anti-HBsAg antibody.

REFERENCES

- Abara, W. E., Qaseem, A., Schlie, S., McMahon, B. J. & Harris, A. M. (2017). Hepatitis B Vaccination, Screening, and Linkage to Care: Best Practice Advice From the American College of Physicians and the Centers for Disease Control and Prevention. *Ann. Intern. Med.*, 167: 794-804.
- Abd Muain, M. F., Cheo, K. H., Omar, M. N., Amir Hamzah, A. S., Lim, H. N., Salleh, A. B., Tan, W. S. & Ahmad Tajudin, A. (2018). Gold nanoparticle-decorated reduced-graphene oxide targeting anti hepatitis B virus core antigen. *Bioelectrochemistry*, 122: 199-205
- An, Y., Jiang, X., Bi, W., Chen, H., Jin, L., Zhang, S., Wang, C. & Zhang, W. (2012). Sensitive electrochemical immunosensor for α -synuclein based on dual signal amplification using PAMAM dendrimer-encapsulated Au and enhanced gold nanoparticle labels. *Biosens. Bioelectron.*, 32: 224-230.
- An, Y., Zhu, G., Bi, W., Lu, L., Feng, C., Xu, Z. & Zhang, W. (2017). Highly sensitive electrochemical immunoassay integrated with polymeric nanocomposites and enhanced SiO₂@Au core-shell nanobioprobes for SirT1 determination. *Anal. Chim. Acta*, 966: 54-61
- Assari, P., Rafati, A. A., Feizollahi, A. & Joghani, R. A. (2020). Fabrication of a sensitive label free electrochemical immunosensor for detection of prostate specific antigen using functionalized multi-walled carbon nanotubes/polyaniline/AuNPs. *Mater. Sci. Eng. C*, 115: 111066.
- ATLAS Medical (2015). HBsAb One Step Hepatitis B Surface Antibody Test Strip (Serum/Plasma), Cambridge, UK
- Avella-Oliver, M., Ferrando, V., Monsoriu, J. A., Puchades, R. & Maquieira, A. (2018). A label-free diffraction-based sensing displacement immunosensor to quantify low molecular weight organic compounds. *Anal. Chim. Acta*. 1033: 173-179
- Avila-Salas, F., Gonzales, R. I., Rios, P.L., Araya-Duran, I. & Camarada, M.B. (2020) Effect of the Generation of PAMAM Dendrimers on the Stabilization of Gold Nanoparticles. *J Chem Inf Model*, 60 (6): 2966-2976.
- Aydin, E. B., Aydin, M. & Sezgintürk, M. K. (2018). Electrochemical immunosensor based on chitosan/conductive carbon black composite modified disposable ITO electrode: An analytical platform for p53 detection. *Biosens. Bioelectron.*, 121: 80-89.

- Bahadir, E. B. & Sezgintürk, M. K. (2016). Poly(amidoamine) PAMAM: an emerging material for electrochemical bio(sensing) applications. *Talanta*, 148, 427-438
- Bakshi, S., Mehta, S., Kumeria, T. Shiddiky, M. J. A., Ponat, A., Choudhury, S., Bose, S. & Nayak, R. (2021). Rapid fabrication of homogenously distributed hyper-branched gold nanostructured electrode based electrochemical immunosensor for detection of protein biomarkers. *Sens. Actuators B Chem.*, 326: 128803
- Ballew, J. T., Murray, J. A., Collin, P., Mäki, M., Kagnoff, M. F., Kaukinen, K. & Daugherty, P. S. (2013). Antibody biomarker discovery through in vitro directed evolution of consensus recognition epitopes. *Proc. Natl. Acad. Sci. USA*, 110: 19330–19335.
- Bekale, L., Chanpai, P., Sanyakamdhom, S., Agudelo D. & Tajmir-Riahi, H. A. (2014). Microscopic and thermodynamic analysis of PEG- β -lactoglobulin interaction. *RSC Adv.*, 4: 31084-31093.
- Blilalis, P., Tziveleka, L. A., Varlas, S. & Latrou, H. (2016). pH-Sensitive nanogates based on poly(L-histidine) for controlled drug release from mesoporous silica nanoparticles. *Polym. Chem.*, 7: 1475-1485.
- Boonkaew, S., Teengam, P., Jampasa, S., Rengpipat, S., Siangproh, W. & Chailapakul, O. (2020). Cost-effective paper-based electrochemical immunosensor using a label-free assay for sensitive detection of ferritin. *Analyst.*, 145: 5019-5026.
- Bottero, J., Boyd, A., Gozlan, J., Lemoine, M., Carrat, F., Collignon, A., Boo, N., Dhotte, P., Varsat, B., Muller, G., Cha, O., Picard, O., Nau, J., Campa, P., Silbermann, B., Bary, M., Girard, P. M. & Lacombe, K. (2013). Performance of rapid tests for detection of HBsAg and anti-HBsAb in a large cohort, France. *J. Hepatol.*, 58: 473-478.
- Boufermel, A., Joly, N., Lotton, P., Amari, M. & Gusev, V. (2011). Velocity of Mass Transport to Model Acoustic Streaming: Numerical Application to Annular Resonators. *Acta Acust. united Acust.*, 97 (2): 219-227
- Bradford, M. M. (1976). A Rapid and Sensitive Method for the Quantitation Microgram Quantities of Protein Utilizing the Principle of Protein-Dye Binding. *Anal. Biochem.*, 254: 248–254.
- Brunetto, M. R. (2010) A new role for an old marker, HBsAg. *J. Hepatol.*, 52: 475–477.

- Burcu Bahadır, E. & Kemal Sezgintürk, M. (2015). Applications of electrochemical immunosensors for early clinical diagnostics. *Talanta*, 132: 162–174.
- Buynak, E. B., Roehm, R. R., Tytell, A. A., Bertland, A. U., II, Lampson, G. P. & Hilleman, M. R. (1976). Vaccine against human hepatitis B. *J. Am. Med. Assoc.*, 235: 2832–2834.
- Cabral-Miranda, G., Cardoso, A. R., Ferreira, L. C. S., Sales, M. G. F. & Bachmann, M. F. (2018). Biosensor-based selective detection of Zika virus specific antibodies in infected individuals. *Biosens Bioelectron*, 15: 113-107
- Cao, L., Xiao, H., Fang, C., Zhao, F. & Chen, Z. (2020). Electrochemical immunosensor based on binary nanoparticles decorated rGO-TEPA as magnetic capture and Au@PtNPs as probe for CEA detection. *Microchim. Acta*, 187 (584).
- Cardoso, A. R., Cabral-Miranda, G., Reyes-Sandoval, A., Bachmann, M. F. & Sales, M. G. F. (2017). Detecting circulating antibodies by controlled surface modification with specific target proteins: Application to malaria. *Biosens. Bioelectron.*, 91: 833-841
- Catarino, S. O., Silva, L. R., Mendes, P. M., Miranda, J. M., Lanceros-Mendez, S. & Minas, G. (2014). Piezoelectric actuators for acoustic mixing in microfluidic devices-Numerical prediction and experimental validation of heat and mass transport. *Sens. Actuators B Chem.*, 205: 206-214
- Chanpai, P., & Tajmir-Riahi, H. A. (2016). Thermodynamic analysis of biogenic and synthetic polyamines conjugation with PAMAM-G4 nanoparticles. *J. Photochem. Photobiol. B, Biol.*, 155: 13-19.
- Chanpai, P., Froehlich, E., Mandeville, J. S., & Tajmir-Riahi, H. A. (2017). Protein conjugation with PAMAM nanoparticles: Microscopic and thermodynamic analysis. *Colloids Surf. B*, 15: 168-174.
- Chen, D., Wang, J., Xu, Y. (2013). Highly Sensitive Lateral Field Excited Piezoelectric Film Acoustic Enzyme Biosensor. *IEEE Sens. J.*, 13 (6): 2217-2222
- Chen, W., Tomalia, D. A. & Thomas, J. L. (2000). Unusual pH-dependent polarity changes in PAMAM dendrimers: evidence for pH-responsive conformational changes, *Macromolecules*, 3: 9169–9172.
- Chen, Y., Yuan, P. X., Wang, A. J. Luo, X., Xue, Y. Zhang, L. & Feng, J. J. (2019). A novel electrochemical immunosensor for highly sensitive detection of prostate-specific antigen using 3D open-structured

PtCu nanoframes for signal amplification. *Biosens. Bioelectron.*, 126: 187-192

- Chevaliez, S. & Pawlotsky, J. M. (2018). New virological tools for screening, diagnosis and monitoring of hepatitis B and C in resource-limited settings. *J. Hepatol.*, 69 (4): 916-926.
- Ciolkowski, M., Rozanek, M., Bryszewska, M. & Klajnert, B. (2013). The influence of PAMAM dendrimers surface groups on their interaction with porcine pepsin. *Biochim Biophys Acta Proteins Proteom*, 1834(10): 1982-1987.
- Ciucci, F. (2019). Modeling electrochemical impedance spectroscopy. *Curr Opin Electrochem*, 13: 132-139.
- Collins, D. J., Ma, Z. & Ai, Y. (2016). Highly localized acoustic streaming and size-selective submicrometer particle concentration using high frequency microscale focused acoustic fields. *Anal. Chem.*, 88 (10): 5513-5522
- Collins, D. J., Ma, Z., Han, J. & Ai, Y. (2017). Continuous micro-vortex-based nanoparticle manipulation via focused surface acoustic waves. *Lab Chip*, 17(1): 91-103
- Cui, W., Zhang, H., Zhang, H., Yang, Y., He, M., Qu, H., Pang, W., Zhang, D. & Duan, X. (2016). Localized ultrahigh frequency acoustic fields induced micro-vortices for submilliseconds microfluidic mixing. *Appl. Phys. Lett.*, 109 (25): 253503.
- Dai, Y. & Liu, C. C. (2019). Recent Advances on Electrochemical Biosensing Strategies toward Universal Point-of-Care Systems. *Angew. Chem. Int. Ed.*, 58: 12355-12368
- Dane, D. S., Cameron, C. H. & Briggs, M. (1970). Virus-like particles in serum of patients with Australia-antigen associated hepatitis. *Lancet*, 1: 695-698.
- Daniels, T. M., Pogfai, T., Rodaree, K., Chaotheing, S., Jomphoak, A., Wisitsoraat, A., Suwannakitti, N., Wongsombat, C., Jaruwongrunsee, K., Shaw, P., Kamchonwongpaisan, S. & Tuantranont, A. (2011). Enhancement of DNA hybridization under acoustic streaming with three piezoelectric-transducer system. *Lab on a Chip* 12(1): 133-8
- Delsing, P., Clelang, A. N., Schuetz, M. J. A., Knörzer, J., Giedke, G., Cirac, J. I., Srinivasan, K., Wu, M. & Balram, K. C. (2019). The 2019 surface acoustic roadmap. *J. Phys. D: Appl. Phys.* 52: 353001.

- Désiré, N., Ngo, Y., Franetich, J. F., Dembele, L., Mazier, D., Vaillant, J. C., Poynard, T. & Thibault, V. (2015). Definition of an HBsAg to DNA international unit conversion factor by enrichment of circulating hepatitis B forms. *J. Viral Hepat.*, 22: 718–726.
- Devandran, C. & Gralinski, I. (2014). Separation of particles using acoustic streaming and radiation forces in an open microfluidic channel. *Microfluid Nanofluidics*, 17: 879-890.
- Di Guana, G., Lib, P., Riggsa, P. D. & Inouyeb, H. (1988). Vectors that facilitate the expression and purification of foreign peptides in *Escherichia coli* by fusion to maltose-binding protein. *Genes* 67 (1): 21-30
- Diagnostic Automation/ Cortez Diagnostics, Inc. (2017). OneStep HBV Combo RapiCard™ InstaTest (Serum/Plasma), California, USA
- DIAsource Immunoassays (2011). HBeAg/Anti-HBe Elisa, Louvain-la-Neuve-Belgium.
- Dip Gandarilla, A. M., Regiart, M., Bertotti, M., Glória, J. C., Mariuba, L. A. M. & Brito, W. R. (2020). One-step enzyme-free dual electrochemical immunosensor for histidine-rich protein 2 determination. *RSC Adv*, 11: 408-415.
- Dreesman, G. R., Hollinger, F. B., Suriano, J. R., Fujioka, R. S., Brunshwig, J. P. & Melnick, J. L. (1972). Biophysical and biochemical heterogeneity of purified hepatitis B antigen. *J. Virol.*, 10: 469–476.
- Eble, B. E., Lingappa, V. R. & Ganem, D. (1986). Hepatitis B surface antigen: An unusual secreted protein initially synthesized as a transmembrane polypeptide. *Mol. Cell. Biol.*, 6: 1454–1463.
- Eble, B. E., MacRae, D. R., Lingappa, V. R. & Ganem, D. (1987). Multiple topogenic sequences determine the transmembrane orientation of hepatitis B surface antigen. *Mol. Cell. Biol.*, 7: 3591–3601
- Ebrahimi, M., Norouzi, P., Safarnejad, M. R., Tabaei, O. & Haji-Hashemi, H. (2019). Fabrication of a label-free electrochemical immunosensor for direct detection of *Candidatus Phytoplasma Aurantifolia*. *J. Electroanal. Chem.*, 851: 113451
- Eissa, S. & Zourob, M. (2021). Development of a Low-Cost Cotton-Tipped Electrochemical Immunosensor for the detection of SARS-CoV-2, *Anal. Chem.*, 93: 1826-1833
- Elanchezian, M. & Senthilkumar, S. (2019). Covalent immobilization and enhanced electrical wiring of haemoglobin using gold nanoparticles encapsulated PAMAM dendrimer for

electrochemical sensing of hydrogen peroxide. *Appl. Surf. Sci.*, 495: 143540.

- Elanchezian, M. & Senthilkumar, S. (2021). Redox-active gold nanoparticle-encapsulated poly(amidoamine) dendrimer for electrochemical sensing of 4-aminophenol. *J.Mol. Liq.*, 325: 115131.
- Elgrishi, N., Rountree, K. J., McCarthy, B. D., Rountree, E. S., Eisenhart, T. T. & Dempsey, J. L. (2017). A Practical Beginner's Guide to Cyclic Voltammetry. *J. Chem. Educ.*, 95: 197-206.
- Fabiani, L., Saroglia, M., Galatá, g., Santis, R. D., Fillo, S., Luca, V., Faggioni, G., D'Amore, N., Regalbuto, E., Salvatori, P., Terova, G., Moscone, D., Lista, F. & Arduini, F. (2021). Magnetic beads combined with carbon black-based screen-printed electrodes for COVID-19: A reliable and miniaturized electrochemical immunosensor for SARS-CoV-2 detection in *Saliva*, *Biosensors and Bioelectronics*, 171: 112686.
- Ferguson, M., Yu, M. W. & Heath, A. (2010). Calibration of the second international standard for hepatitis B immunoglobulin in an international collaborative study. *Vox Sang* 99: 77–84
- Gelin, P., Sukas, Özlem, S. S., Hellemans, K., Maes, D. & Malsche, W. (2019). Study on the mixing and migration behaviour of micron-size particles in acoustofluidics. *Chem. Eng. J.* 369: 370-375
- Gogola, J. L., Martins, G., Caetano, F. R., Ricciardi-Jorge, T., Duarte dos Santos, C. N., Marcolino-Junior, L. H. & Bergamini, M. F. (2019). Label-free electrochemical immunosensor for quick detection of anti-hantavirus antibody. *J. Electroanal. Chem.*, 842: 140-145.
- Gray, D., Gray, M. & Barr, T. (2007). Innate responses of B cells. *Eur J Immunol* 37(12): 3304–3310
- Greub, G., Zysset, F., Genton, B., Spertini, F. & Frei, P. C. (2001). Absence of anti-hepatitis B surface antibody after vaccination does not necessarily mean absence of immune response. *Med Microbiol Immunol* 189(3): 165–168
- Grzegorzewska, A. E. (2012). Hepatitis B vaccination in Chronic Kidney Disease: Review of Evidence in Non-Dialyzed Patients. *Hepat. Mon.*, 12 (11): E7359.
- Guo, X. & Hu, N. (2009). Increment of Density of Au Nanoparticles Deposited in Situ within Layer-by-Layer Films and Its Enhancement on the Electrochemisry of Ferrocenecarboxylic Acid and Bioeletrocatalysis. *J. Phys. Chem. C*, 113: 9831-9837.

- Guo, X. & Wang, J. (2019). Comparison of linearization methods for modelling the Langmuir adsorption isotherm. *J. Mol. Liq.*, 296: 111850.
- Habib, S. & Shaikh, O. (2007). Hepatitis B immune globulin. *Drugs Today (Barc)*. 43(6): 379–394
- Haji-Hashemi, H., Norouzi, P., Safarnejad, M. R. & Ganjali, M. R. (2017). Label-free electrochemical immunosensor for direct detection of *Citrus tristeza virus* using modified gold electrode. *Sens. Actuators B Chem.*, 244: 211-216
- Hammarström, B. (2014). *Acoustic Trapping in Biomedical Research* (Unpublished doctoral dissertation). Lund University, Lund, Sweden
- Han, J. L., Hu, H., Huang, Q. Y. & Lei, Y. L. (2021). Particle separation by standing surface acoustic waves inside a sessile droplet. *Sens. Actuator, A*, 326: 112731.
- Hasmoni, S. H., Mau, G. K., Karsani, S. A., Cass, A. A. & Shahir, S. (2016). *Strep-tag II* Mutant Maltose-binding Protein for Reagentless Fluorescence Sensing. *Trop Life Sci Res.*, 27(1): 63-75
- Heo, N. S., Zheng, S., Yang, M. H., Lee, S. J., Lee, S. Y., Kim, H. J., Park, J. Y., Lee, C. S. & Park, T. J. (2012). Label-free Electrochemical Diagnosis of Viral Antigens with Genetically Engineered Fusion Protein. *Sensors*, 12 (8): 10097-10108
- Hirano, A., Tanaka, T. Kataura, H., & Kameda, T. (2014). Arginine Side Chains as a Dispersant for Individual Single-Wall Carbon Nanotubes. *Chem. Eur. J.*, 20 (17): 4922-4930
- Holmes, M., Parker, N. & Povey, M. (2011). Temperature dependence of bulk viscosity in water using acoustic spectroscopy, *J. Phys. Conf. Ser.*, 012011.
- Hong, C. C., Lin, C. C., Hong, C. L., Lin, Z. X., Chung, M. H. & Hsieh, P. W. (2016). Handheld analyzer with on-chip molecularly-imprinted biosensors for electrical detection of propofol in plasma samples. *Biosens. Bioelectron.*, 86: 623-629.
- Hong, Z., Zhou, Z. M., Chen, X. Y. & Ying, F. C. (1991). Sound Propagation in Glass-Ceramic. In: Leroy O., Breazeale M. A. (eds) *Physical Acoustics*. Springer, Boston, MA.
- Honorato de Castro, A. C., Alves, L. M., Siquieroli, A. C. S., Madurro, J. M. & Brito-Madurrio, A. G. (2020). Label-free electrochemical

- immunosensor for detection of oncomarker CA125 in serum. *Microchem. J.*, 155: 104746
- Hou, J., Ren, J., Song, L., Zhao, F. & Liang, P. (2018). Analytical performance of three diagnostic reagents for HBsAg on an automatic ELISA analyser. *J. Clin. Lab. Anal.*, 32: e22159
- Howard, C. R. & Allison, L. M. C. (1995). Hepatitis B Surface Antigen Variation and Protective Immunity. *Intervirology*, 38: 35-40.
- Hu, L., Dong, T., Zhao, K., Deng, A. & Li, J. (2017). Ultrasensitive electrochemiluminiscent brombuterol immunoassay by applying a multiple signal amplification strategy based on a PAMAM-gold nanoparticle conjugate as the bioprobe an Ag@Au core shell nanoparticles as a substrate. *Microchim. Acta* 184 (9): 3415-3423.
- Huang, K. & Szeifer, I. (2017). , Design of Multifunctional Nanogate in Response to Multiple External Stimuli Using Amphiphilic Diblock Copolymer. *J. Am. Chem. Soc.*, 139: 6422-6430
- Huang, Q. Y., Hu, H., Lei, Y. L., Han, J. L., Zhang, P. & Dong, J. (2020). Simulation and experimental investigation of surface acoustic wave streaming velocity. *Jpn. J. Appl. Phys.* 59; 064001
- Huzair, F. & Sturdy, S. (2017). Biotechnology and the transformation of vaccine innovation: The case of the hepatitis B vaccines 1968–2000. *Stud Hist Philos Bio Biomed Sci*, 64: 11–21.
- Huzly, D., Schenk, T., Jilg, W. & Neumann-Haefelin, D. (2008). Comparison of nine commercially available assays for quantification of antibody response to hepatitis B virus surface antigen. *J. Clin. Microbiol.*, 46: 1298–1306
- Hyun, C. S., Lee, S. & Ventura, W. R. (2019). The prevalence and significance of isolated hepatitis B core antibody (anti-HBc) in endemic population. *BMC Res. Notes*, 12: 251.
- Iglesias-Mayor, A., Amor-Gutiérrez, O., Costa-García, A. & Escosura-Muñiz, A. (2019). Nanoparticles as Emerging Labels in Electrochemical Immunosensors. *Sensors*, 19: 5137
- Jain, A., & Cheng, K. (2017). The principles and applications of avidin-based nanoparticles in drug delivery and diagnosis. *J. Control. Release*, 245: 27–40.
- Jung, M. C. & Pape, G. R. (2002). Immunology of hepatitis B infection. *Lancet* 2(1): 43–50

- Kang, J. Li, Z. & Wang, G. (2021). A novel signal amplification strategy electrochemical immunosensor for ultra-sensitive determination of p53 protein. *Bioelectrochemistry*, 137: 107647
- Kao, J. H. (2008). Diagnosis of hepatitis B virus infection through serological infection through serological and virological markers. *Expert Rev. Gastroenterol. Hepatol.*, 2(4): 553-562
- Kapust, R. B. & Waugh, D. S. (1999). Escherchia coli maltose-binding protein is uncommonly effective at promoting the solubility of polypeptides to which it is fused. *Protein Sci.* 8: 1668-74.
- Kaushik, A., Yndart, A., Kumar, S., Jayant, R. D., Vashist, A., Brown, A. N., Li, C. Z. & Nair, M. (2018). A sensitive electrochemical immunosensor for label-free detection of Zika-virus protein, *Sci. Rep.*, 8: 9700
- Kavosi, B., Hallaj, R., Teymourian, H. & Salimi, A. (2014). Au nanoparticles/PAMAM dendrimer functionalized wired ethyleneamine-viologen as highly efficient interface for ultra-sensitive α -fetoprotein electrochemical immunosensor. *Biosens. Bioelectron.* 59: 389-396.
- Kawahara, N., Yarin, A. L. & Brenn, G. (2000). Effect of acoustic streaming on the mass transfer from a sublimating sphere. *Phys. Fluids.*, 12: 912-923
- Kessler, D. A. & Jimenez, A. (2019). Transfusion Medicine and Hemostasis Clinical and Laboratory Aspects (Third Edition), *Chapter 13-Hepatitis B Virus Screening* (pp. 73-76). Elsevier Science
- Khoo, M. M., Ng, K. L., Alias, Y. & Khor, S. M. (2017). Impedimetric biotin-Immunosensor with excellent analytical performance for real sample analysis. *J. Electroanal. Chem.*, 799: 111-121
- Khor, S. M., Thordason, P. & Gooding, J. J (2013). The impact of antibody/epitope affinity strength on the sensitivity of electrochemical immunosensors for detecting small molecules. *Anal. Bioanal. Chem.* 405: 3889-3898
- Kim, A. R., Park, T. J., Kim, M. S., Kim, I. H., Kim, K. S., Chung, K. H. & Ko, S. (2017). Functional fusion proteins and prevention of electrode fouling for a sensitive electrochemical immunosensor. *Anal. Chim. Acta.*, 967: 70-77
- Kinn, S., Akhavan, S., Agut, H. & Thibault, V. (2011). Performance of the DiaSorin LIAISON® anti-HBs II for the detection of hepatitis B surface antibodies: Comparison with the Abbott Architect anti-HBs assay. *J. Clin. Virol.*, 50: 297-302

- Kirtland, J. D. (2010). *Interfacial Mass Transfer In Microfluidic Systems: Existence And Persistence Of The Modified Graetz Behavior* [Doctoral dissertation, Cornell University]. Cornell University Research Repository. <https://hdl.handle.net/1813/17771>
- Kozlowski, L. P. (2016). IPC - Isoelectric Point Calculator. *Biol. Direct*, 11: 55.
- Kyriakidou, E.A., Khivantsev, K., Gostanian, T. M., Alexxev, O.S. & Amiridis, M. D. (2015). Silica-supported gold/dendrimer nanocomposites with controlled sizes of gold particles. *APPL CATAL A-GEN*, 504: 482-492.
- Lau, P. Y., Ng, K. L., Yusof, N. A., Liu, G., Alias, Y. & Khor, S. M. (2019). A sample pre-treatment free electrochemical immunosensor with negative electro-pulsion for quantitative detection of acrylamide in coffee, cocoa and prune juice. *Anal. Methods*, 11: 4299-4313
- Layqah, L. A. & Eissa, S. (2019). An electrochemical immunosensor for the corona virus associated with the Middle East respiratory syndrome using an array of gold nanoparticle-modified carbon electrodes, *Microchimica Acta*, 186: 224.
- Leathers, J. S., Pisano, M. B., Re, V., van Oord, G., Sultan, A., Boonstra, A. & Debes, J. D. (2019). Validation of a point-of-care rapid diagnostic test for hepatitis C for use in resource-limited settings. *Int. Health* 11 (4): 314-315.
- Lee, B. Y., Li, Z., Clemens, D. L., Dillon, B. J., Hwang, A. A., Zink, J. I., & Horwitz, M. A. (2016). Redox-triggered release of moxifloxacin from mesoporous silica nanoparticles functionalized with disulfide snap-tops enhances efficacy against pneumonic tularemia in mice. *Small*, 12: 3690–3702.
- Lei, Y. M., Zhou, J., Chai, Y. Q., Zhuo, Y., Yuan, R. (2018). SnS₂ quantum dots as new emitters with strong electrochemiluminescence for ultrasensitive antibody detection. *Anal. Chem.*, 90: 12270-12277
- Li, Q., Yu, C., Gao, R., Xia, C., Yuan, G., Li, Y., Zhao, Y., Chen, Q. & He, J. (2016). A novel DNA biosensor integrated with Polypyrrole/streptavidin and Au-PAMAM-CP bionanocomposite probes to detect the rs4839469 locus of the *vangl1* gene for dysontogenesis prediction. *Biosens. Bioelectron*, 80: 674-681.
- Li, Y., Liu, L., Liu, X., Ren, Y., Xu, K., Zhang, N., Sun, X., Yang, X., Ren, X. & Wei, Q. (2020). A dual-mode PCT electrochemical immunosensor with CuCo₂S₄ bimetallic sulphides as enhancer. *Biosens. Bioelectroni*, 163: 112280.

- Liao, X., Ma, C., Zhao, C., Li, W., Song, L. Y., Hong, C. & Qiao, X. (2021). An immunosensor detects carcinoembryonic antigen by dual catalytic signal enhancer-hydrogen peroxide based on in-situ reduction of silver nanoparticles with dopamine and graphene high-load cobalt tetroxide. *Microchem. J.*, 160: 105602.
- Liu, B., Li, M., Zhao, Y., Pan, M., Gu, Y., Sheng, W., Fang, G. & Wang, S. (2018). A sensitive electrochemical immunosensor based on PAMAM dendrimer-encapsulated Au for detection of norfloxacin in animal-derived foods. *Sensors*, 18: 1946
- Liu, F., Guo, Z. & Dong, C. (2017). Influences of obesity on the immunogenicity of Hepatitis B vaccine. *Hum Vaccin Immunother* 4 13(5):1014–1017.
- Liu, G. & Lin, Y. (2007). Nanomaterial Labels in Electrochemical Immunosensors and Immunoassays. *Talanta*, 74: 308–317
- Liu, J., Li, S., & Bhethanabotla, V. R. (2018). Integrating Metal-Enhanced Fluorescence and Surface Acoustic Waves for Sensitive and Rapid Quantification of Cancer Biomarkers from Real Matrices. *ACS Sensors*, 3(1): 222–229.
- Liu, R. H., Lenigk, R., Druyor-Sanchez, R. L., Yang, J. & Grodzinski, P. (2003). Hybridization enhancement using cavitation microstreaming. *Anal Chem* 75(8): 1911-7
- Lu, C. Y., Ni, Y. H., Chiang, B. L., Chen, P. J., Chang, M. H. & Chang, L.Y. (2008). Humoral and cellular immune responses to a hepatitis B vaccine booster 15–18 years after neonatal immunization. *J. Infect. Dis.* 197: 1419–1426
- Lu, Y., Peterson, J. R., Luais, E., Gooding, J. J. & Lee, N. A. (2016). Effects of Surface Epitope Coverage on the Sensitivity of Displacement Assays that Employ Modified Nanoparticles: Using Bisphenol A as a Model Analyte. *Biosensors*, 6: 43.
- Luanwathi, S. Krittayavathananon, A., Srimuk, P. & Sawangphruk, M. (2015). *In situ* synthesis of permselective zeolitic imidazolate framework-8/graphene oxide composites: rotating disk electrode and Langmuir adsorption isotherm. *RSC. Adv.*, 5: 46617-46623.
- Luo, W., Xu, X., Zhou, B., He, P., Li, Y. & Liu, C. (2019). Formation of enzymatic/redox-switching nanogates on mesoporous silica nanoparticles for anticancer drug delivery. *Mater. Sci. Engin. C*, 100: 855-861.
- Luong, T. D., Phan, V. N. & Nguyen, N. T. (2010). High-throughput micromixers based on acoustic streaming induced by surface acoustic wave. *Microfluid Nanofluidics*, 10(3): 619–625.

- Luong, T. D., Phan, V. N. & Nguyen, N. T. (2011). High-throughput micromixers based on acoustic streaming induced by surface acoustic wave. *Microfluid Nanofluid*, 10: 619-625
- Mahnashi, M. H., Mahmoud, A. M., Alhazzani, K., Alanazi, A. Z., Alaseem, A. M., Algahtani, M. M. & El-Wekil, M. M. (2021). Ultrasensitive and selective molecularly imprinted electrochemical oxaliplatin sensor based on a novel nitrogen-doped carbon nanotubes/Ag@cu MOF as a signal enhancer and reported nanohybrid. *Microchim. Acta*, 188 (124).
- Mahshid, S. S., Mahshis, S., Vallée-Bélisle, A. & Kelley, S. O. (2019). Peptide-Mediated Electrochemical Steric Hindrance Assay for One-Step Detection of HIV Antibodies. *Anal. Chem*, 91: 4943-4947
- Maina, C. V., Riggs, P. D., Granda III, A. G., Slatko, B. E., Moran, L. S., Tagliamonte, J. A., McReynolds, L. A. & di Guan, C. (1988). An *Escherichia coli* vector to express and purify foreign proteins by fusion to and separation from maltose-binding protein. *Gene*, 74: 365-373
- Mangold, C. M. T., Unckell, F., Werr, M. & Streeck, R. E. (1995). Secretion and antigenicity of hepatitis B virus small envelope proteins lacking cysteines in the major antigenic region. *Virology*, 211: 535-543
- Mangold, C. M. T., Unckell, F., Werr, M. & Streeck, R. E. (1997). Analysis of intermolecular disulfide bonds and free sulfhydryl groups in hepatitis B surface antigen particles. *Arch. Virol.*, 142: 2257-2267
- Maupas, P., Goudeau, A., Coursaget, P., Drucker, J. & Bagros, P. (1976). Immunisation against hepatitis B in man. *Lancet*, 307: 1367-1370.
- Medetalibeyoglu, H., Kotan, G., Atar, N. & Yola, M. L. (2020). A novel and ultrasensitive sandwich-type electrochemical immunosensor based on delaminated MXene@AuNPs as signal amplification for prostate specific antigen (PSA) detection and immunosensor validation. *Talanta*, 220: 121403
- Mehmood, S., Corradi, V., Choudhury, H. G., Hussain, R., Becker, P., Axford, D., Zirah, S., Rebuffat, S., Tieleman, D. P., Robinson, C. V. & Beis, K. (2016). Structural and Functional Basis for Lipid Synergy on the Activity of the Antibacterial Peptide ABC Transporter McjD. *J. Biol. Chem.*, 291: 21656-21668.
- Milich, D. R. & Leroux-Roels. G. G. (2003). Immunogenetics of the response to HBsAg vaccination. *Autoimmun. Rev* 2(5): 248-257

Ministry of Health Malaysia (2019). National Strategic Plan for Hepatitis B and C 2019 – 2023.

Miodek, A., Mejri, N., Gomgnimbou, M., Sola, C. & Korri-Youssofi, H. (2015). E-DNA Sensor of *Mycobacterium tuberculosis* Based on Electrochemical Assembly of Nanomaterials (MWCNTs/PPy/PAMAM). *Anal. Chem.* 87: 9257-9264

Miodek, A., Mejri-Omrani, N., Khoder, R. & Korri-Youssofi, H. (2016). Electrochemical functionalization of polypyrrole through amine oxidation of poly(amidoamine) dendrimers: Application to DNA biosensor. *Talanta*, 154: 446-454

Mitome, H. (1998). The mechanism of generation of acoustic microstreaming. *Electron Commun Jpn.*, 81 (10): 1-8.

Mondrago'n, L., Mas, N., Ferragud, V., de la Torre, C., Agostini, A., Mart'inez-Ma'n'ez, R., Sanceno'n, F., Amoro's, P., Perez-Paya', E., & Orza'ez, M. (2014). Enzyme-responsive intracellular-controlled release using silica mesoporous nanoparticles capped with ϵ -poly-L-lysine. *Chem. A Eur. J.*, 20: 5271–5281.

Mössner, B. K., Staugaard, B., Jensen, J., Lillevang, S. T., Christensen, P. B. & Holm, D. K. (2016). Dried blood spots, valid screening for viral hepatitis and human immunodeficiency virus in real-life. *World J. Gastroenterol.*, 22(33): 7604-7612.

Muniswamy, V., Pattnaik, P. K. & Krishnaswamy, N. (2019). Modeling and Analysis of SOI Gratings-Based Opto-Fluidic Biosensor for Lab-on-a-chip Applications. *Photonics*, 6 (2).

Nadhakumar, P., Haque, A. J., Lee, N. S., Yoon, Y. H. & Yang, H. (2018). Washing-free displacement immunosensor for cortisol in human serum containing numerous interfering species. *Anal. Chem.*, 90 (18): 10982-10989.

Nasir, S., Ali, M. & Ensinger, W. (2012). Thermally controlled permeation of ionic molecules through synthetic nanopores functionalized with amine-terminated polymer brushes. *Nanotechnology*, 23 (22): 5502.

Nestorova, G., Adapa, B. S., Kopparchy, L. & Guilbeau, E. J. (2016). Lab-on-a-chip thermoelectric DNA biosensor for label-free detection of nucleic acid sequences. *Sens. Actuators B Chem.*, 225: 174-180.

Nguyen, T., Andreasen, S. Z., Wolff, A. & Bang, D. D. (2018). From Lab on a Chip to Point of Care Devices: The Role of Open Source Microcontrollers. *Micromachines*, 9 (8): 403

- Nowacka, O., Milowska, K. & Bryszewska (2015). Interaction of PAMAM dendrimers with bovine insulin depends on nanoparticle end-groups. *J. Lumin.*, 162: 87-91
- Ortho Clinical Diagnostics (2017). Hepatitis B Virus Core Antigen (Recombinant) ORTHO® HBc ELISA Test System, USA
- Ozcelik, A., Ahmed, D., Xie, Y., Nama, N., Qu, Z., Nawaz, A. A. & Huang, T. J. (2014). An Acoustofluidic Micromixer via Bubble Inception and Cavitation from Microchannel Sidewalls. *Anal. Chem.* 86: 5083-5088
- Pan, G., Zhao, G., Wei, M., Wang, Y. & Zhao, B. (2019). Design of nanogold electrochemical immunosensor for detection of four phenolic estrogens. *Chem. Phys. Lett.*, 732: 136657.
- Pan, S., Zhang, H., Liu, W., Wang, Y., Pang, W. & Duan, X. (2017). Biofouling Removal and Protein Detection Using a Hypersonic Resonator. *ACS Sens.*, 2(8): 1175–1183
- Papastergious, V., Lombardi, R. & MacDonald, D. (2015). Global Epidemiology of Hepatitis B Virus (HBV). *Curr Hepatology Rep*, 14: 171-178.
- Park, H. S., Park, T. J., Huh, Y. S., Choi, B. G., Ko, S., Lee, S. Y. & Hong, W. H. (2010). Immobilization of genetically engineered fusion proteins on gold-decorated carbon nanotube hybrid films for the fabrication of biosensor platforms. *J. Colloid Interface Sci.*, 350 (2): 453-458
- Park, J. W., Lee, S. J., Ren, S., Lee, S., Kim, S. & Laurell, T. (2016). Acousto-microfluidics for screening of ssDNA aptamer. *Sci. Rep.*, 6: 27121
- Pashchenko, O., Shelby, T., Banerjee, T. & Santra, S. (2018). A comparison of optical, electrochemical, magnetic, and colorimetric point-of-care biosensors for infectious disease diagnosis. *ACS Infect Dis.*, 4 (8): 1162-1178
- Paxton, W. F., O'Hara, M. J., Peper, S. M., Petersen, S. L. & Grate, J. W. (2008). Accelerated Analyte Uptake on Single Beads in Microliter-Scale Batch Separations Using Acoustic Streaming: Plutonium Uptake by Anion Exchange for Analysis by Mass Spectrometry. *Anal. Chem.*, 80 (11): 4070-4077
- Penna, A., Artini, M., Cavalli, A., Levrero, M., Bertoletti, A., Pilli, M., Chisari, F. V., Rehermann, B., Del Prete, G., Fiaccadori, F. & Ferrari, C. (1996). Long-lasting memory T cell responses following self-limited acute hepatitis B. *J Clin Invest* 98(5): 1185–1194

- Pérez-Fernández, B., Mercader, J V., Abad-Fuentes, A., Checa-Orrego, B. I., Costa-García, A. & Escosura-Muñiz, A. (2020). Direct competitive immunosensor for Imidacloprid pesticide detection on gold nanoparticle-modified electrodes. *Talanta*, 209: 120465
- Poiteau, L., Soulier, A., Roudot-Thoraval, F., Hezode, C., Challine, D., Pawlowsky, J. M. & Chevaliez, S. (2017). Performance of rapid diagnostic tests for the detection of anti-HBs in various patient populations. *J. Clin. Virol.*, 96: 64-66.
- Pondé, R. A. A. (2011). The underlying mechanisms for the “simultaneous HBsAg and anti-HBs serological profile”. *Eur J Clin Microbiol Infect Dis*, 30: 1325-1340
- Porchetta, A., Ippodrino, R., Marini, B., Caruso, A., Caccuri, F. & Ricci, F. (2018). Programmable Nucleic Acid Nanoswitches for the Rapid, Single-Step Detection of Antibodies in Bodily Fluids. *J. Am. Chem. Soc.*, 140: 947-953
- Qi, Q. & Brereton, G. J. (1995). Mechanisms of removal of micron-sized particles by high-frequency ultrasonic waves. *IEEE Trans Ultrason Ferroelectr Freq Control*, 42(4): 619-629.
- Qiu, Z., Tang, D., Shu, J., Chen, G. & Tang, D. (2016). Enzyme-triggered formation of enzyme-tyramine concatamers on nanogold-functionalized dendrimer for impedimetric detection of Hg(II) with sensitivity enhancement. *Biosens Bioelectrons*. 75: 108-115
- Ranallo, S., Rossetti, M., Plaxco, K. W., Vallée-Bélisle, A. & Ricci, F. A. (2015). DNA-Based Beacon for Single-Step Fluorescence Detection of Antibodies and Other Proteins. *Angew. Chem. Int. Ed.*, 54: 13214-13218.
- Rao, S., Lu, S., Guo, Z., Li, Y., Chen, D., & Xiang, Y. (2014). A light-powered bio-capacitor with nanochannel modulation. *Adv. Mater.*, 26 (33): 5846-5850.
- Rashid, S., Nawaz, M. H., Rehman, I. U, Hayat, A. & Marty, J. L. (2021). Dopamine/mucin-1 functionalized electro-active carbon nanotubes as a probe for direct competitive electrochemical immunosensing of breast cancer biomarker. *Sens. Actuators B Chem.*, 330: 129351
- Raven, S., Hautvast, J., Steenbergen, J.V., Akkermans, R., Weykamp, C., Smits, F., Hoebe, C. & Vossen, A. (2017). Diagnostic performance of serological assays for anti-HBs testing: results from a quality assessment program. *J Clin Virol* 87: 17–22
- Ribeiro, L. F., Amarelle, V., Ribeiro, L. F. C. & Guazzaroni, M. E. (2019). Converting a Periplasmic Binding Protein into a Synthetic

Biosensing Switch through Domain Insertion. *BioMed Res. Int.*, 4798793

- Riggs, P. (2000). *Expression and Purification of Recombinant Proteins by Fusion to Maltose-Binding Protein*. Beverly MA: New England Biolabs
- Rodella, A., Galli, C., Terlenghi, L., Perandin, F., Bonfanti, C. & Manca, N. (2006). Quantitative analysis of HBsAg, IgM anti-HBc and anti-HBc avidity in acute and chronic hepatitis B. *J. Clin. Virol.*, 37: 206-212
- Roupa, Z., Noula, M., Farazi, E., Stylianides, A. & Papanephytous, C. (2019). Vaccination Coverage and Awareness of Hepatitis B Virus Among Healthcare Students at a University in Cyprus. *Mater Sociomed*, 31(3): 190-196.
- Roy, S. K. & Sharan, P. (2019). Design of ultra-high sensitive biosensor to detect *E. Coli* in water. *Int. J. Inf. Technol.*, 12: 775-780.
- Ruehle, B., Clemens, D.L., Lee, B. Y., Horwitz, M.A., & Zink, J. I. (2017). A pathogen-specific cargo delivery platform based on mesoporous silica nanoparticles. *J. Am. Chem. Soc.*, 137: 6663–6668.
- Safarian, S. M., Kusov, P. A., Kosolobov, S. S., Borzenkova, O. V., Khakimov, A. V., Kotelevtsev, Y. V. & Drachev, V. P. (2021). Surface-specific washing-free immunosensor for time-resolved cortisol monitoring. *Talanta*, 225: 122070
- Salama, I., Sami, S., Salama, S., Rabah, T., El Etreby, L., Abdel Hamid, A., Elmosalami, D., El Hariri, H. & Said, Z. N. (2016). Immune response to second vaccination series of hepatitis B virus among booster dose non-responders. *Vaccine* 34(16): 1904–1908
- Schuster, B. (2018). S-Layer Protein-Based Biosensors. *Biosensors*, 8 (2), 40
- Scott, A. M. & Wolchok, J. D. (2012). Old, L.J. Antibody therapy of cancer. *Nat. Rev. Cancer*, 12: 278.
- Sekli Belaidi, F., Tsopela, A., Salvagnac, L., Ventalon, V., Bedel-Pereira, E., Bardinal, V., Séguy, I., Temple-Boyer, P., Juneau, P., Izquierdo, R. & Launay, J. (2016). Lab-on-chip with microalgal based biosensor for water assessment. *2016 IEEE Nanotechnology Materials and Devices Conference (NMDC)*: 1-2
- Sekowski, S., Buczkowski, A., Palecz, B. & Gabryelak, T. (2011). Interaction of polyamidoamine (PAMAM) succinamic acid dendrimers generation 4 with human serum albumin. *Spectrochim. Acta A Mol. Biomol. Spectrosc.*, 81 (1): 706-710.

- Selby, C. (1999) Interference in immunoassay. *Ann Clin Biochem* 36: 704–721
- Shepard, C. W., Simard, E.P., Finelli, L., Fiore, A. E. & Bell, B. P. (2006). Hepatitis B virus infection: epidemiology and vaccination. *Epidemiol Rev* 28: 112–125
- Sher, M., Zhuang, R., Demirci, U., & Asghar, W. (2017). Paper-based analytical devices for clinical diagnosis: recent advances in the fabrication techniques and sensing mechanisms. *Expert Rev Mol Diagn.* 17 (4): 351-366
- Society for Maternal-Fetal Medicine, Dionne-Odom, J., Tita, A. T. N. & Silverman, N. S. (2016). Society for Maternal-Fetal Medicine (SMFM) Consult Series: Hepatitis B in pregnancy screening, treatment, and prevention of vertical transmission. *Am. J. Obstet. Gynecol.*: 6-14.
- Stephenne, J. (1988). Recombinant versus plasma-derived hepatitis B vaccines: Issues of safety, immunogenicity and cost-effectiveness. *Vaccine*, 6, 299–303
- Stramer, S. L., Glynn, S. A., Kleinman, S. H., Strong, D. M., Caglioti, S., Wright, D. J., Dodd, R. Y. & Busch, M. P. (2004). Detection of HIV-1 and HCV infections among antibody-negative blood donors by nucleic acid - Amplification testing. *N. Engl. J. Med.*, 351: 760–768.
- Su, S., Sun, Q., Wan, L., Gu, X., Zhu, D., Zhou, Y., Chao, J. & Wang, L. (2019). Ultrasensitive analysis of carcinoembryonic antigen based on MOS₂-based electrochemical immunosensor with triple signal amplification. *Biosens. Bioelectron.*, 140: 111353
- Suffner, S., Gerstenberg, N., Patra, M., Ruibal, P., Orabi, A., Schindler, M. and Bruss, V. (2018). Domains of the hepatitis B virus small surface protein S mediating oligomerization. *J. Virol.*, 92: e02232-17
- Sun, C., Liao, X., Huang, P., Shan, G., Ma, X., Fu, L., Zhou, L. & Kong, W. (2020). A self-assembled electrochemical immunosensor for ultrasensitive detection of ochratoxin A in medicinal and edible malt. *Food Chem.*, 315: 126289.
- Suresh, L., Bondili, J. S. & Brahman, P. K. (2020). Development of proof of concept for prostate cancer detection: an electrochemical immunosensor based on fullerene-C₆₀ and copper nanoparticles composite film as diagnostic tool. *Mater.Today Chem.*, 16: 100257.

- Taleghani, A. S., Ebrahimnejad, P., Heydarinasab, A. & Akbarzadeh, A. (2020). Adsorption and controlled release of iron-chelating drug from the amino-terminated PAMAM/ordered mesoporous silica hybrid materials. *J Drug Deliv Sci Technol*, 56: 101579
- Tam, D., Xue, M. & Zink, J. I. (2013). pH-Responsive Dual Cargo Delivery from Mesoporous Silica Nanoparticles with a Metal-Latched Nanogate. *Inorg. Chem.*, 52: 2044-2049
- Tam, Y. J., Zeenathul, N. A., Rezaei, M. A., Mustafa, N. H., Azmi, M. L. M., Bahaman, A. R., Lo, S. C., Tan, J. S., Hani, H. and Rasedee, A. (2017). Wide dynamic range of surface-plasmon-resonance-based assay for hepatitis B surface antigen antibody optimal detection in comparison with ELISA. *Biotechnol. Appl. Biochem.* 64: 735-744.
- Tan, G. H., Yusoff, K., Seow, H. F. & Tan, W. S. (2005). Antigenicity and immunogenicity of the immunodominant region of hepatitis B surface antigen displayed on bacteriophage T7. *J Med Virol.* 77 (4): 475-480.
- Tan, W. S. & Ho, K. L. (2014). Phage display creates innovative applications to combat hepatitis B virus. *World J Gastroenterol.* 20 (33): 11650-11670.
- Tang, Y., Tang, D., Zhang, J. & Tang, D. (2018). Novel quartz crystal microbalance immunodetection of aflatoxin B₁ coupling cargo-encapsulated liposome with indicator-triggered displacement assay. *Anal. Chim. Acta*, 1031: 161-168.
- Tenda, K., Van Gerven, B., Arts, R., Hiruta, Y., Merckx, M. & Citterio, D. (2018). Paper-Based Antibody Detection Devices using Bioluminescent BRET-Switching Sensor Proteins. *Angew. Chem.*, 130: 15595-15599
- Torigoe, K., Suzuki, A. & Esumi, K. (2001). Au(III)-PAMAM Interaction and Formation of Au-PAMAM Nanocomposites in Ethyl Acetate. *J. Colloid Interface Sci.*, 241 (2): 346-356.
- Tsopela, A., Laborde, A., Salvagnac, L., Ventalon, V., Bedel-Pereira, E., Séguy, I., Temple-Boyer, P., Juneau, P., Izquierdo, R. & Launay, J. (2016). Development of a lab-on-chip electrochemical biosensor for water quality analysis based on microalgal photosynthesis. *Biosens. Bioelectron.*, 79: 568-573
- Tu Ho, J. K., Jeevan-Raj, B. & Jürgen, H. (2020). Hepatitis B Virus (HBV) Subviral Particles as Protective Vaccine and Vaccine Platforms. *Viruses*, 12 (2): 126

- Valats, J., Tuailleon, E., Funakoshi, N., Hoa, D., Brabet, M. C., Bolloré, K., Ducos, J., Vendrell, J. P. & Blanc, P. (2010). Investigation of memory B cell responses to hepatitis B surface antigen in health care workers considered as non-responders to vaccination. *Vaccine* 28(39): 6411–6416
- Van Assche, T. R. C., Baron, G. V. & Denayer, J. F. M. (2018). An explicit multicomponent adsorption isotherm model: accounting for the size-effect for components with Langmuir adsorption behaviour. *Adsorption*, 24: 517-530.
- Van Oss, C. J. (2000). Nature of Specific Ligand-Receptor Bonds, in Particular the Antigen-Antibody Bond. *Journal of Immunoassay*, 21 (2-3): 109-142.
- Vivero-Escoto, J. L., Jeffords, L., Dreau, D., Alvarez-Berrios, M. & Mukherjee, P. (2017). Mucin1 antibody-conjugated dye-doped mesoporous silica nanoparticles for breast cancer detection in vivo. *Proc. SPIE 10078, Colloidal Nanoparticles for Biomedical Applications XII*, 100780B1–100780B8.
- Vusa, C. S. R., Manju, V. Berchmans, S. & Arumugam, P. (2016). Electrochemical amination of graphene using nanosized PAMAM dendrimers for sensing applications. *RSC Adv.*, 6: 33409-33418
- Wang, B., Sun, Y., Davis, T. P., Ke, P. C., Wu, Y. & Ding, F. (2018). Understanding Effects of PAMAM Dendrimer Size and Surface Chemistry on Serum Protein Binding with Discrete Molecular Dynamic Simulations. *ACS Sustainable Chem. Eng.*, 6(9): 11704-11715.
- Wang, R., Xu, L. & Li, Y. (2015). Bio-nanogate controlled enzymatic reaction for virus sensing. *Biosens. Bioelectron.*, 67: 400-407
- Wang, X., Cai, X., Hu, J., Shao, N., Wang, F., Zhang, Q., Xiao, J. & Cheng, Y. (2013). Glutathione-Triggered “Off-On” Release of Anticancer Drugs from Dendrimer-Encapsulated Gold Nanoparticles. *J. AM. Chem. Soc.*, 135 (26): 9805-9810.
- Weihrauch, M. R., von Bergwelt-Baildon, M., Kandic, M., Weskott, M., Klamp, W. & Rosler, J. (2008). T cell responses to hepatitis B surface antigen are detectable in non-vaccinated individuals. *World J Gastroenterol.*, 14: 2529–2533
- White, H. B., & Hughes, A. R. (1981). Biotin-Binding Proteins in Chicken Eggs and the Biotin Requirements of Chicken Embryos. *Poult. Sci. J.*, 60(7): 1454–1457
- World Health Organization. (2019). Hepatitis B. Key Facts; World Health Organization: Geneva, Switzerland.

- Wright, T.A., Stewart, J. M., Page, R. C. & Konkolewicz, D. (2017). Extraction of Thermodynamic Parameters of Protein Unfolding Using Parallelized Differential Scanning Fluorimetry. *J. Phys. Chem. Lett.*, 8: 553-558.
- Wu, M., Ozcelik, A., Rufo, J., Wang, Z., Fang, R. & Huang, T. J. (2019). Acoustofluidic separation of cells and particles. *Microsyst. Nanoeng.*, 5 (32)
- Xiao, H., Wei, S., Gu, M., Chen, Z. & Cao, L. (2021). A sandwich-type electrochemical immunosensor using rGO-TEPA-Thi-Au as sensitive platform and CMK-3@AuPtNPs as signal probe for AFP detection. *Microchem. J.*, 170: 106641.
- Xie, Y., Chindam, C., Nama, N., Yang, S., Lu, M., Zhao, Y., Mai, J. D., Costanzo, F. & Huang, T. J. (2015). Exploring bubble oscillation and mass transfer enhancement in acoustic-assisted liquid-liquid extraction with a microfluidic device. *Sci. Rep.*, 5: 12572
- Xu, X. X., Zhou, C. L., Zeng, B. R. Xia, H. P., Lan, W. G. & He, X. M. Structure and properties of polyamidoamine/polyacrylonitrile composite nanofiltration membrane prepared by interfacial polymerization. *Sep. Purif. Technol.*, 96: 229-236
- Yan, H., Yang, C. J., Tang, N., Zou, Y., Chakravarty, S., Roth, A. & Chen, R. T. (2017). Specific Detection of Antibiotics by Silicon-on-Chip Photonic Crystal Biosensor Arrays. *IEEE Sens. J.*, 17 (18).
- Yang, G., Lai, Y., Xiao, Z., Tang, C. & Deng, Y. (2018). Ultrasensitive electrochemical immunosensor of carcinoembryonic antigen based on gold-label silver-stain signal amplification. *Chin. Chem. Lett.*, 29(12): 1857-1860
- Yang, H., Nishitani, S. & Sakata, T. (2018). Potentiometric Langmuri Isotherm Analysis of Histamine-Selective Molecularly Imprinted Polymer-Based Field-Effect Transistor. *ECS J Solid State Sci Technol*, 7(7): 3079-3082.
- Yang, J., Dai, D., Lou, X., Ma, L., Wang, B. & Wang, Y. W. (2020). Supramolecular nanomaterials based on hollow mesoporous drug carriers and macrocycle-capped CuS nanogates for synergistic chemo-photothermal therapy. *Theranostics*, 10: 615-629
- Yang, Z. H., Zhuo, Y., Yuan, R. & Chai, Y. Q. (2015). An amplified electrochemical immunosensor based on in situ-produced 1-naphthol as electroactive substance and graphene oxide Pt nanoparticles functionalized CeO₂ nanocomposites as signal enhancer. *Biosens. Bioelectron.*, 69: 321-327.

- Yin, H., Zhou, Y., Ai, S., Chen, Q., Zhu, X., Liu, X. & Zhu, L. (2010). Sensitivity and selectivity determination of BPA in real water samples using PAMAM dendrimer and CoTe quantum dots modified glassy carbon electrode. *J. Hazard. Mater.* 174: 236-243
- Yin, H., Cui, L., Chen, Q., Shi, W., Ai, S., Zhu, L. & Lu, L. (2011). Amperometric determination of bisphenol A in milk using PAMAM- Fe_3O_4 modified glassy carbon electrode. *Food Chem.* 125: 1097-1103
- Yong, C. Y., Yeap, S. K., Goh, Z. H., Ho, K. L., Omar, A. R. & Tan, W. S. (2015). Induction of Humoral and Cell-Mediated Immune Responses by Hepatitis B Virus Epitope Displayed on the Virus-Like Particles of Prawn Nodavirus. *Appl. Environ. Microbiol.*, 81: 882-889.
- Younger, A. K. D, Su, P. Y., Shepard, A. J., Udani, S. V., Cybulski, T. R., Tyo, K. E. J. & Leonard, J. N. (2018). Development of novel metabolite-responsive transcription factors via transposon-mediated protein fusion. *Protein Eng. Des. Sel.*, 31(2): 55–63
- Zaffina, S., Marcellini, V., Santoro, A. P., Scarsella, M., Camisa, V., Vinci, M. R., Musolino, A. M. & Nicolosi, L. (2014). Repeated vaccinations do not improve specific immune defenses against hepatitis B in non-responder health care workers. *Vaccine* 32(51): 6902–6910
- Zeng, Y., Bao, J., Zhao, Y., Huo, D., Chen, M., Yang, M., Fa, H. & Huo, C. (2018). A sensitive label-free electrochemical immunosensor for detection of cytokeratin 19 fragment antigen 21-1 based on 3D graphene with gold nanoparticle modified electrode. *Talanta*, 178: 122-128
- Zhang, B. & Ding, C. (2016). Displacement-type amperometric immunosensing platform for sensitive determination of tumour markers. *Biosens. Bioelectron.* 82: 112-118.
- Zhang, C., Kaluvan, S., Zhang, H., Wang, G. & Zuo, L. (2018). A study on the Langmuir adsorption for quartz crystal resonator based low pressure CO₂ gas sensor. *Measurement*, 124: 286-290.
- Zhang, H. M., Lou, K., Cao, J. & Wang, Y. Q. (2014). Interaction of a Hydrophobic-Functionalized PAMAM Dendrimer with Bovine Serum Albumin: Thermodynamic and Structural Changes. *Langmuir*, 30(19): 5536-5544.
- Zhang, Q., Zhang, Z., Zhou, H., Xie, Z., Wen, L., Zhaoyue, L., Zhai, J. & Zhai, J. (2017). Redox switch of ionic transport in conductive polypyrrole-engineered unipolar nanofluidic diodes. *Nano Res.* 10 (11): 3715-3725.

- Zhang, X., Wang, J., Chen, X., Yu, M., Yu, S., Sun, Y., Duan, J., Sun, H. & Yuan, P. (2018). Short-term immunogenicity of standard and accelerated hepatitis B virus vaccination schedules in healthy adults: a comparative field study in China. *Biosci Rep.* 38(5): BSR20180846.
- Zhao, M., Du, L., Qi, L., Li, Y., Li, Y. & Li, X. (2018). Numerical simulations and electrochemical experiments of the mass transfer of microvias electroforming under ultrasonic agitation. *Ultrason Sonochem*, 48: 424-431
- Zhong, H. & Hu, N. (2007). Conductive Effect of Gold Nanoparticles Encapsulated Inside Polyamidoamine (PAMAM) Dendrimers on Electrochemistry of Myoglobin (Mb) in {PAMAM-Au/Mb}_n Layer-by-Layer Films. *J. Phys. Chem. B*, 111 (35): 10583-10590.
- Zhou, M., Gao, D., Yang, Z., Zhou, C., Tan, Y., Wang, W. & Jiang, Y. (2021). Streaming-enhanced, chip-based biosensor with acoustically active, biomarker-functionalized micropillars: A case study of thrombin detection. *Talanta*, 222: 121480
- Zhu, J., Ye, Z., Fan, X., Wang, H., Wang, Z. & Chen, B. (2019). A highly sensitive biosensor based on Au NPs/rGO-PAMAM-Fc nanomaterials for detection of cholesterol. *Int J Nanomedicine*, 14: 835-849.