

# **UNIVERSITI PUTRA MALAYSIA**

IN VITRO INVESTIGATION OF ANTIDIABETIC AND ANTIGLYCATION ACTIVITIES OF METHANOLIC EXTRACT OF Ficus deltoidea Jack VARIETIES

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By

NUR SUMIRAH MOHD DOM

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

July 2021

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

#### IN VITRO INVESTIGATION OF ANTIDIABETIC AND ANTIGLYCATION ACTIVITIES OF METHANOLIC EXTRACT OF *Ficus deltoidea* Jack VARIETIES

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Diabetes mellitus has been listed as one of the leading death factors in the world which associated with glucose uptake activity in the human body. Nowadays, the usage of medicinal plants for the management of diabetes mellitus has gained interest even though many antidiabetic drugs are available in the market. This could possibly be due to the limitations of these drugs such as adverse effects and poor clinical efficacy. Therefore, searching for new effective antidiabetic agents should be continued. *Ficus deltoidea (F. deltoidea)* or also known as Mas Cotek is a local medicinal plant in Malaysia that has been used as a supplement to promote health and traditionally claimed to possess antidiabetic effect. However, the scientific studies to confirm its efficacy and its possible mode of actions are still inadequate.

This study was done to authenticate the antidiabetic property of *in vitro* antihyperglycemic mechanisms evaluation of *F. deltoidea* varieties, to elucidate the potential of the plant to stimulate insulin secretion from  $\beta$ -pancreatic cells, to enhance glucose uptake by adipocytes and muscle cells, to screen glucose uptake inhibition activity in adipocytes and muscle cells in the presence of LY294002 (PI3-Kinase inhibitor) and ALLN (CAP inhibitor), to assess the insulin sensitizing activity in adipocytes cells and to observe the expression of PPAR-gamma, PI3 Kinase, GLUT4 and CAP gene. The inhibition of advanced glycation end products formation, protein oxidation, total phenolic contents and antioxidative effects of the plant were also monitored. The viability of cells that were used in the *in vitro* evaluation of antihyperglycemic mechanisms in the presence of *F. deltoidea* extracts was determined using MTT assay.

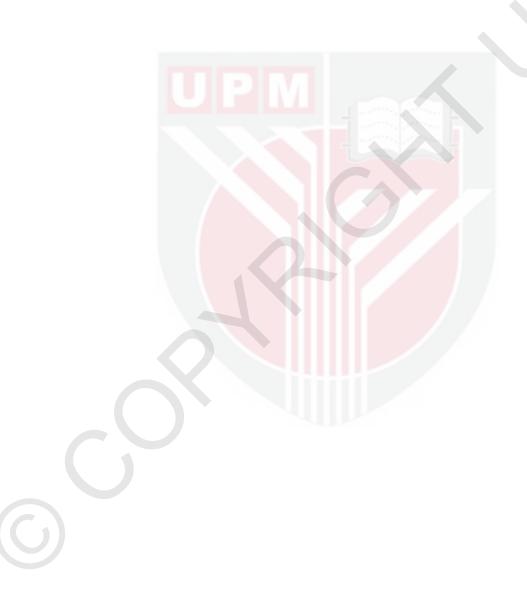
The viability study showed that methanolic extract of *F. deltoidea* varieties did not possess any cytotoxic effect at concentration of 100  $\mu$ g/ ml and this concentration was used to measure the antidiabetic mechanism of *F. deltoidea* varieties extract in BRIN BD11 pancreatic, 3T3FF42A adipocytes and L6 myotubes cells. For insulin secretion study, *F. deltoidea* variety *intermedia* stimulated the highest insulin secretion followed by *F. deltoidea* variety *kunstleri* and *F. deltoidea* variety *trengganuensis.* The result indicated that the insulin secretory action of the extracts involved KATP channel-dependent and KATP channel-independent pathways. *F. deltoidea* variety *trengganuensis, F. deltoidea* variety *kunstleri* and *F. deltoidea* variety *intermedia* also significantly enhanced basal and insulin-mediated glucose uptake into adipocytes and muscles cells. The extracts showed either insulin-mimetic or insulin-sensitizing activity or combination of both activities during enhancing glucose uptake into these cells.

Meanwhile, the result of glucose uptake inhibition assay of the extracts demonstrated that insulin stimulated glucose uptake followed phosphatidylinositiol-3-kinase-independent (PI3K) mechanism in L6 muscle and 3T3F442A adjpocytes cells. In L6 myotubes cells, the highest glucose uptake activity was observed when treated with F. deltoidea variety trengganuensis and F. deltoidea variety intermedia meanwhile in adipocytes, the highest activity was found when treated with F. deltoidea variety trengganuensis. Additionally, in gene expression study, the most upregulation of PPAR-gamma gene was expressed significantly at 24 hours with the highest in F. deltoidea variety kunstleri followed by F. deltoidea variety intermedia and F. deltoidea variety trengganuensis.

The novelty of this study is about the investigation of antiglycation properties of *F*. *deltoidea* varieties which significantly inhibited the formation of advancedglycation end-products (AGEs). In conjunction with a reduction of fructosamine level, the plant extracts also increased the thiol groups level and inhibited the formation of protein carbonyl. No previous report published about the potential of *F. deltoidea* plant to inhibit the formation of AGEs. The highest antiglycation activity was observed in *F. deltoidea* variety *intermedia* which also depicted the highest phenolic content and reducing power activities. The correlation study between total phenolic content and antiglycation activity, DPPH radical scavenging activity and reducing power activity showed  $(-1.0 \ge r \le 1)$ ,  $(-1.0 \ge r \le 1)$  and  $(-0.69 \ge r \le 1.0)$  respectively.

In conclusion, this study had shown that *F. deltoidea* able to reduce hyperglycemia with mechanisms such as stimulation of insulin secretion from pancreatic  $\beta$ -cells, enhancement of glucose uptake into adipocytes and muscle cells and effects of insulin stimulated glucose uptake is through independent type of PI3K pathway. Insulin-sensitizing activities exhibited by *F. deltoidea* varieties indicated that this plant has the ability to alleviate insulin resistance and may potentially be beneficial for the treatment of type 2 diabetes mellitus with insulin-resistance condition. Finally, the *F. deltoidea* plant extract also contained phenolic compounds, possessed antioxidant and antiglycation effects that may offer remarkable prospects for the preventive treatment of AGE-mediated diabetic complications.

Hence, *F. deltoidea* varieties may offer good therapeutic potential to reduce the complications of diabetes mellitus.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

## PENYIASATAN SECARA IN VITRO AKTIVITI ANTIDIABETIK DAN ANTIGLIKASI EKSTRAK METANOLIK PELBAGAI VARIASI Ficus deltoidea Jack

#### Oleh

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Diabetes mellitus telah disenaraikan sebagai salah satu faktor kematian di dunia yang dikaitkan dengan aktiviti pengambilan glukosa dalam badan manusia. Pada masa kini, penggunaan tumbuhan ubatan dalam mengawal diabetes mellitus telah mendapat perhatian walaupun terdapat banyak ubat antidiabetik berada di pasaran. Ini boleh disebabkan oleh limitasi ubat-ubat yang berkaitan di mana ianya mempunyai kesan buruk dan keberkesanan klinikal yang lemah. Sehubungan itu, pencarian kepada agen antidiabetik yang berkesan perlu diteruskan. *F. deltoidea* atau dikenali juga sebagai Mas Cotek merupakan tumbuhan ubatan tempatan di Malaysia yang diambil sebagai makanan tambahan untuk kesihatan dan secara tradisional didakwa mempunyai sifat antidiabetik. Namun begitu, kajian-kajian saintifik untuk mengesahkan keberkesanan dan mod tindakan yang mungkin bagi tumbuhan tersebut masih berkurangan.

Kajian ini dilaksanakan untuk mengesahkan ciri antidiabetik bagi pelbagai variasi *F. deltoidea* melalui penentuan mekanisme antihiperglisemik secara *in-vitro*. Kajian ini untuk menjelaskan potensi tumbuhan tersebut bagi meransang rembesan insulin daripada sel beta-pankreatik, untuk meningkatkan pengambilan glukosa oleh sel adipos dan sel otot, untuk melihat aktiviti pengambilan glukosa dengan kehadiran LY294002 (perencat PI3-Kinase) dan ALLN (perencat CAP) dalam sel adipos dan sel otot, untuk menilai aktiviti pemekaan insulin dalam sel adipos dan untuk melihat pengungkapan gen PPAR-gamma, PI3 Kinase, GLUT4 dan CAP. Perencatan pembentukan produk akhir glikasi lanjutan, pengoksidaan protein dan kesan antioksida ekstrak tumbuhan juga dipantau. Keupayaan untuk hidup sel-sel yang digunakan dalam kajian mekanisme antihiperglisemik secara *in-vitro* dengan kehadiran *F. deltoidea* ditentukan menggunakan ujian MTT.

Kajian keupayaan sel untuk hidup menunjukkan bahawa ekstrak pelbagai variasi *F. deltoidea* tidak mempunyai kesan sitotoksik pada kepekatan 100 µg/ ml dan kepekatan ini digunakan untk mengukur mekanisme antidiabetik ke atas sel pankreatik BRIN BD11, sel adipos 3T3F442A dan sel otot L6 myotube. Kajian tindakan perembesan insulin menunjukkan ekstrak *F. deltoidea* variasi *intermedia* merangsang perembesan insulin tertinggi diikuti oleh *F. deltoidea* variasi *kunstleri* dan *F. deltoidea* variasi *trengganuensis*. Ini menunjukkan tindakan perembesan insulin tersebut melibatkan laluan kebergantungan-KATP dan ketidakbergantungan-KATP. *F. deltoidea* variasi *trengganuensis, F. deltoidea* variasi *kunstleri* dan *F. deltoidea* variasi *intermedia* juga dengan ketara meningkatkan pengambilan glukosa pada tahap basal dan dengan perantaraan insulin dalam sel adipos dan sel otot. Ekstrak tersebut menunjukkan sama ada aktiviti peniru insulin atau pemekaan insulin atau gabungan kedua-dua aktiviti semasa meningkatkan pengambilan glukosa ke dalam sel-sel terlibat.

Sementara itu, hasil ujian perencatan pengambilan glukosa oleh F. deltoidea variasi trengganuensis, F. deltoidea variasi kunstleri and F. deltoidea variasi intermedia menunjukkan bahawa pengambilan glukosa yang dirangsang oleh insulin adalah sebahagiannya mengikuti mekanisma PI3-kinaseketidakbergantungan pada sel otot L6 dan sel adipos 3T3F442A. Pada L6 myotubes, aktiviti penambahan glukosa tertinggi boleh dilihat apabila sel dirawat dengan F. deltoidea variasi trengganuensis dan F. deltoidea variasi intermedia manakala pada sel adipos, aktiviti tertinggi apabila sel dirawat dengan ekstrak *F. deltoidea* variasi *trengganuensis*. Sebagai tambahan, kajian pengungkapan gen menunjukkan bahawa peningkatan aktiviti pengambilan glukosa melalui pengungkapan gen PPAR-gamma adalah signifikan selepas 24 jam dengan pengungkapan tertinggi pada ekstrak F. deltoidea variasi kunstleri diikuti dengan F. deltoidea variasi intermedia dan F. deltoidea variasi trengganuensis.

Kebaharuan bagi kajian ini adalah mengenai sifat antiglikasi bagi pelbagai variasi *F. deltoidea* dalam merencat pembentukan produk akhir glikasi lanjutan. Kajian juga menunjukkan ekstrak tumbuhan menurunkan tahap fruktosamin, meningkatkan tahap kumpulan *thiol* dan menghalang pembentukan protein karbonil. Sehingga kini, tiada laporan direkodkan mengenai potensi tumbuhan *F. deltoidea* untuk merencatkan pembentukan produk akhir glikasi lanjutan. Aktiviti antiglikasi tertinggi dilihat pada *F. deltoidea* variasi *intermedia* yang mana ia juga menunjukan kandungan fenolik tertinggi dan aktiviti daya pengurangan. Kajian korelasi antara keseluruhan kandungan fenolik dan aktiviti antiglikasi, aktiviti pengambilan radikal DPPH dan aktiviti daya pengurangan menunjukkan (-1.0 ≥ r ≤ 1), (-1.0 ≥ r ≤ 1) dan (-0.69 ≥ r ≤ 1.0) masing-masing.

Sebagai kesimpulan, kajian ini telah menunjukkan bahawa F. deltoidea kemampuan untuk mengurangkan hiperglisemia mempunyai dengan mekanisme seperti merangsang perembesan insulin dari sel-sel pankreas, peningkatan pengambilan glukosa ke dalam sel adipos dan sel otot serta pengambilan glukosa yang dirangsang oleh insulin mengikut laluan PI3-kinaseketidakbergantungan. Kegiatan pemekaan insulin oleh F. deltoidea menunjukkan bahawa tumbuhan ini mempunyai keupayaan untuk

mengurangkan ketahanan insulin dan berpotensi memberi manfaat untuk rawatan diabetes mellitus jenis 2 dengan keadaan ketahanan insulin. Akhirnya, ekstrak tumbuhan *F. deltoidea* juga mengandungi sebatian fenolik, mempunyai kesan antioksida dan antiglikasi yang menawarkan potensi yang bagus untuk rawatan pencegahan komplikasi diabetes yang disebabkan oleh produk akhir glikasi lanjutan. Oleh itu, pelbagai variasi *F. deltoidea* boleh menawarkan potensi terapi yang baik untuk mengurangkan komplikasi diabetes mellitus.



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Signature: Name of Member of Supervisory Committee:	Dr. Zainah Adam

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### LIST OF ABBREVIATIONS

ADP Adenosine diphosphate AGEs Advanced Glycation Endproducts AKT Protein Kinase B ALLN N-acetyl-Leu-Leu-Norleu-al AMPK Adenosine 5' Monophosphate-activated Protein Kinase Analysis of Variance ANOVA ATP Adenosine Triphosphate B40 Bottom 40% households Cyclic Adenosine Monophosphate cAMP CAP Catabolite Activator Protein DAG Diacylglycerol DMEM **Dulbecco's Modified Eagle Medium** DNA Deoxyribonucleic Acid DPP-4 **Dipeptidyl Peptidase-4** DPPH 2,2-diphenyl-1-picrylhydrazyl ELISA Enzyme-Linked Immunosorbent Assay GDM **Gestational Diabetes Mellitus** GDP Guanosine diphosphate GLP-1 Glucagon-like Peptide 1 GTP **Guanosine Triphosphate** HbA1c Glycated haemoglobin IFN-γ Interferon-gamma IL-6 Interleukin 6

- *IL-1β* Interleukin 1 beta
- IRS-1 Insulin receptor Substrate 1
- JNK c-Jun NH2-terminal kinase
- K<sub>ATP</sub> ATP-sensitive potassium
- KRB Kreb's Ringer Buffer
- MAPK Mitogen-Activated Protein Kinase
- MODY Maturity onset of diabetes in youth
- mRNA Messenger Ribonucleic Acid
- mTOR Mechanistic Target of Rapamycin
- MTT 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
- NF-kB Nuclear Factor kappa B
- PBS Phosphate Buffer Saline
- PI3K Phosphatidylinositol 3-kinase
- PPAR Peroxisome proliferator-activated receptor
- RAGE Receptor Advanced Glycation Endproducts
- ROS Reactive Oxygen Species
- RPMI 1640 Roswell Park Memorial Institute 1640
- SDS Sodium Dodecyl Sulfate
- SGLT-2 Sodium-glucose co-transporter-2
- SUs Sulfonylureas
- TNF-α Tumor Necrosis Factor Alpha
- TZDs Thiazolidinediones

#### CHAPTER 1

#### INTRODUCTION

### 1.1 Background of Study

Diabetes mellitus is a chronic metabolic disease characterized by hyperglycemia due to the deficiency in either insulin secretion or insulin action or by combination of both factors. Diabetes mellitus is the seventh leading death factor in the world and has caused 1 million mortalities in 2000 and increased to 1.6 million in 2016 (WHO, 2018). The same report discovered that 422 million people around the world suffered from diabetes, mainly in middle- and poor-income countries. It is also stated that over the years, more patients experienced with diabetes and become more predominance. In India, around 41 million people were diagnosed with diabetic and expected to escalate to 70 million in 2025 (Sicree *et al.*, 2006). Skyler *et al.*, (2017) reported that more diabetic adults originated from East Asia, South Asia and Australia compared to other regions with a total of 153 million patients.

Institute for Public Health (2020) has conducted a National Health and Morbidity Survey (NHMS) among Malaysian and discovered a prevalence rate of diabetes in adults was 13.4% in 2015 and increased to 18.3% in 2019. The highest prevalence of diabetes was recorded among Indians (31.4%), followed by Malays (21.6%), Chinese (15.1%), and Bumiputera Sarawak (12.3%). The report also classified the disease was more common among widow(er)/ divorcee (33.2%), patient with no formal education (28.7%), retirees (45.8%) and people with low socio-economic (B40) group (18.5%). The predominance of diabetes in the urban area was depicted at 9.7%, not much different to rural area at 8.2%. However, when compared between states, the highest prevalence was found in Perak (15.2%) followed by Melaka (13.7%), Perlis (13.5%), Sabah (4.1%) and Sarawak (7.7%) respectively with no record mentioned on other states in Malaysia (Institute for Public Health, 2020). In conjunction, it was stated that the development of T2DM is associated with low socio-economic conditions such as occupational rank, educational status and income level (Agardh *et al.*, 2011).

The NHMS 2019 survey also found that around 3.9 million Malaysians are suffering from diabetes. The figure depicted as the highest rate in Asia and resembled among the highest in the world (Institute for Public Health, 2020). Meanwhile, around 8.1% of the adult population in Malaysia, or similar to 1.7 million people, possessed high possibilities to suffer from diabetes, hypertension, and high cholesterol diseases. They also projected that by 2025, 7 million Malaysian adults are likely to suffer from diabetes which encounter for 31.3% diabetes prevalence for adults aged 18 years and above. The same report mentioned a ratio of 1:5 adults aged 18 years and above are suffering from

diabetes and the trend of prevalence has increased from 7.2% (2011) followed by 8.3% (2015) and steadily increased to 9.4% (2019) which have been diagnosed with diabetes meanwhile, 4.0% (2011), 5.1% (2015) and 8.9% (2019) of adults claimed that they did not realize they have diabetes (Institute for Public Health, 2020).

Environmental factors such as obesity, in-active lifestyles, aging, daily intake containing processed food, hereditary factors, and epigenetic modifications responsible to the increasing of diabetes cases (Ma *et al.*, 2014). Other than focused on the lifestyle interventions, intake of oral medications, injection of insulin and other treatment based on pharmacogenomic, proteomic, and metabolomic approaches could help in fight against diabetes mellitus (Hu and Jia, 2018). From the data of Adult Diabetes Control and Management (ADCM) of Malaysia, the usage of insulin injection has recorded 12.9% and 85.6% were prescribed with anti-hyperglycemic agents such as biguanides (83.2%) and sulfonylureas (69.9%). Other than that, the diabetic patients were prescribed with angiotensin-converting enzyme inhibitors (63.9%) and calcium channel blockers (<40%), meanwhile, about 58.2% were on antihypertensive agent treatment (Mastura *et al.*, 2011).

### 1.2 Statement of Problem

The ultimate mission for diabetes treatment is to reduce and maintain glycated hemoglobin (HbA1c) level below 7%. Higher HbA1c levels (7.0% and above) can progress to macrovascular and microvascular complications. The macrovascular complications are known as cardiovascular, cerebrovascular, and peripheral vascular disease meanwhile, microvascular risks are identified as nephropathy, neuropathy, and retinopathy complications (Stein *et al.*, 2013; He *et al.*, 2015). Therefore, to maintain normal blood glucose levels and minimize the complications of diabetes, the usage of oral antidiabetic drugs has been approved by Drug Administration or European Medicines Agency (He *et al.*, 2015).

Currently, several oral synthetic antidiabetic drugs have been used to treat the disease but have displayed undesirable side effects. Metformin is the first choice of oral medication to diabetic's patients followed by second-line prescription of sodium-glucose co-transporter-2 (SGLT-2) inhibitors, thiazolidinediones (TZDs), sulfonylureas (SUs), dipeptidyl peptidase 4 (DPP-4) inhibitors, alpha-glucosidase inhibitors (AGIs), and meglitinides (Qian *et al.*, 2018). The adverse effects of metformin are known as risk of getting neuropathy in elderly, may cause anemia, mild weight loss, and nausea/ vomiting or diarrhea; meanwhile for DPP-4 inhibitors (bone fractures, genital mycosis and may increase low-density lipoprotein cholesterol); sulfonylureas (weight gain and increase cardiovascular disease risk); and TZDs (weight gain, cardiac failure, risk of liver disease, anemia risk, swelling of legs or ankles) (Chaudhury *et al.*, 2017). Other than that, meglitinides and alpha-glucosidase inhibitors are not frequently

prescribed as both drugs require multiple divided dosage and develop inefficient treatment (Hussein *et al.*, 2015). In addition, referring to NHMS 2019 report, about 25.7% of diabetes patients claimed that they were on insulin therapy, 85.6% were prescribed with oral antidiabetic drugs within the past 2 weeks, meanwhile 88.0% had a diabetes diet advice from healthcare staffs. Another 75.4% were advised to lose some weight and 23.0% had shifted to traditional and complementary medicines (Institute for Public Health, 2020).

Therefore, searching for alternative and advanced solution for diabetes mellitus are favorable and urgently needed. In recent years, many group of researchers had focused on research using plant-based products to treat various diseases including diabetes mellitus (Abbas *et al.*, 2019). The safety aspects in the use of oral antidiabetic drugs should be considered and limitations of the antidiabetic drugs such as the undesirable side effects, lack of clinical efficacy and inconsistent effects of the antidiabetics drugs on each patient need to be further studied.

## 1.3 Justification of the Study

Even though no specific cure has been found for diabetes mellitus yet, there are many ways to overcome the disease. Aside from oral antidiabetic drugs (conventional medicine), there are complementary and alternative medicine (CAM) therapies suggested for diabetes which refers to other method of healing treatment. In the meantime, National Center for Complementary and Integrative Health (NCCIH), National Institute of Health, defined complementary medicine as other practice that paired together with conventional medicine meanwhile alternative medicine as other routine that used in place of conventional medicine (NCCIH, 2018). In addition, WHO has estimated for the total world population, around 80% are using complementary and alternative medicine for their basic wellness (Yang et al., 2015). The report also outlined the complementary and alternative medicine therapies such as dietary supplement from natural products (herbs), mind and body practices (yoga, chiropractic, acupuncture), Ayurvedic medicine, traditional Chinese medicine, homeopathy and naturopathy. Additionally, another recent term introduced is integrative health care which refers to combination of conventional and complementary approaches together in a coordinated way (NCCIH, 2018).

In traditional practices, various medicinal plants have been utilized to forbid long term complications in diabetes because of low cost, easy to find and less adverse effects (Deepa *et al.*, 2018). It was recorded that around 200 bioactive compounds consist of phenolic acids, flavonoids, triterpenoids, alkaloids and carbohydrates have been isolated from therapeutic plants and possessed antidiabetic effects (Misbah *et. al.*, 2013). A review report by Birdee *et al.*, (2010) listed some plants that have been used to treat diabetes for instance *Allium sativum*, Aloe vera and *Gymnema sylvestre*, which have insulin secretagogue effects. In addition, *Coccinia indica* has insulin mimetic effect, *Momordica charantia* possessed insulin mimetic effect and decreased hepatic glucose

production, meanwhile *Trigonella foenum graecum* acts as insulin secretagogue and decreased carbohydrate absorption. Apart from that, a plant called *Opuntia streptacantha* decreased carbohydrate absorption and lastly *Panex ginseng*, *P. quiquefolius* works as insulin mimetic and alters the hepatic glucose metabolism.

Other than that, Ficus deltoidea (F. deltoidea) or local folks addressed it as Mas Cotek is one of the medicinal plants which possessed antidiabetic properties but not extensively been studied yet. F. deltoidea is a shrub tree from Moraceae family which is primarily cultivated and can be found in Peninsular Malaysia, Java, Thailand, Borneo, Sumatra and Moluccas (Berg et al., 2005). In Malaysia, this plant has been used as an alternative treatment and has been reported for antidiabetic, antinociceptive, antiulcer, antioxidant, anti-inflammatory as well as anti-melanogenic properties (Misbah et al., 2013). Meanwhile, in vivo study showed that oral intake of ethanolic extract of F. deltoidea leaves significantly reduced the blood glucose level in the diabetic rats (Mohammad Noor et al., 2016). Previous research by Adam et al. (2012) reported on in vivo and in vitro antidiabetic study of F. deltoidea extract but not specific for each variety and the mechanism involved. Nevertheless, extensive study on antidiabetics mechanisms and efficiency of F. deltoidea plant in inhibiting diabetes risk factor is still incomplete and need further research.

### 1.4 Objectives of the Study

The main objective of this study is to elucidate the antidiabetic mechanisms of three varieties of methanolic *F. deltoidea* extracts using *in vitro* models. The specific objectives of the study are:

- 1. To evaluate insulin secreting activity of  $\beta$ -pancreatic cells treated with methanolic extracts of *F*. *deltoidea* varieties and the molecular mechanisms underlies such activity.
- 2. To determine the potential of methanolic extracts of *F. deltoidea* varieties to enhance glucose uptake into insulin-targeting cells and molecular mechanisms underlies such activity.
- 3. To evaluate insulin sensitizing activity of adipocytes cells treated with methanolic extracts of *F. deltoidea* varieties.
- 4. To determine the potential of methanolic extracts of *F. deltoidea* varieties towards inhibition of advanced glycation end products formation, total phenolic content and antioxidative properties.

#### REFERENCES

- Abdul-Ghani, M. A., Norton, L., & Defronzo, R. A. 2011. Role of sodium-glucose cotransporter 2 (SGLT 2) inhibitors in the treatment of type 2 diabetes. *Endocrine Reviews* 32 (4): 515–531.
- Abbas, G., Al Harrasi, A., Hussain, H., Hamaed, A. and Supuran, C.T. 2019. The management of diabetes mellitus-imperative role of natural products against dipeptidyl peptidase-4, α-glucosidase and sodium-dependent glucose co-transporter 2 (SGLT2). *Bioorganic Chemistry* 86: 305-315.
- Abdulla, M.A., Abdul-Aziz Ahmed, K., Abu-Luhoom, F.M. and Muhanid, M. 2010. Role of *Ficus deltoidea* extract in the enhancement of wound healing in experimental rats. *Biomedical Research* 21 (3): 241–245.
- Abdullah, Z., Hussain, K., Zhari, I., Rasadah, M.A., Mazura, P., Jamaludin, F. and Sahdan, R. 2009. Evaluation of extracts of leaf of three *Ficus deltoidea* varieties for antioxidant activities and secondary metabolites. *Pharmacognosy Research* **1** (4): 216–223.
- Abrahim, N.N., Abdul-Rahman, P.S. and Aminudin, N. 2018. The antioxidant activities, cytotoxic properties, and identification of water-soluble compounds of *Ficus deltoidea* leaves. *PeerJ* 6: e5694; DOI 10.7717/peerj.5694.
- Aćimović, J.M., Stanimirović, B.D., Todorović, N., Jovanović, V.B., and Mandić, L.M. 2010. Influence of the microenvironment of thiol groups in low molecular mass thiols and serum albumin on the reaction with methylglyoxal. *Chemico-Biological Interactions* 188 (1): 21–30.
- Adam, Z., Khamis, S. Ismail, A. and Hamid. 2012. *Ficus deltoidea*: A potential alternative medicine for diabetes mellitus. *Evidence-Based Complementary and Alternative Medicine*. Article ID 632763. https://doi.org/10.1155/2012/632763.
- Adisakwattana, S., Thilavech, T., and Chusak, C. 2014. Mesona Chinensis Benth extract prevents AGE formation and protein oxidation against fructose-induced protein glycation in vitro. *BMC Complementary and Alternative Medicine.* 14: 130. https://doi.org/10.1186/1472-6882-14-130
- Agardh E, Allebeck P, Hallqvist J, Moradi T, Sidorchuk A. 2011. Type 2 diabetes incidence and socio-economic position: a systematic review and metaanalysis. *International Journal of Epidemiology* 40: 804–818.
- Aguilar-Bryan, L. and Bryan, J. 1999. Molecular biology of adenosine triphosphate-sensitive potassium channels. *Endocrine Reviews* 20 (2): 101–135,1999.

- Ahmed, M.U., Thorpe, S.R. and Baynes, J.W. 1986. Identification of Νεcarboxymethyllysine as a degradation product of fructoselysine in glycated protein. *The Journal of Biological Chemistry* 261 (11): 4889-4894.
- Ahmed, N. 2005. Advanced glycation endproducts-role in pathology of diabetic complications. *Diabetes Research and Clinical Practice* 67: 3 21.
- Aiyegoro, O.A., and Okoh, A.I. 2010. Preliminary phytochemical screening and in vitro antioxidant activities of the aqueous extract of Helichrysum longifolium DC. *BMC Complementary and Alternative Medicine*. https://doi.org/10.1186/1472-6882-10-21
- Akash, M., Rehman, K., and Liaqat, A. 2018. Tumor Necrosis Factor-Alpha: Role in Development of Insulin Resistance and Pathogenesis of Type 2 Diabetes Mellitus. *Journal of Cellular Biochemistry*. 119 (1): 105–110.
- Al-Zuady, M.H., Ismail, A., Mohamed, S., Abdul Razis, A. F., Mumtaz, M.W. and Azizah, A.H. 2018. Antioxidant effect, glucose uptake activity in cell lines and cytotoxic potential of Melicope *lunu-ankenda leaf* extract. *Journal of Herbal Medicine* 14: 55-60.
- American Diabetes Association. 2009. Diagnosis and classification of diabetes mellitus. *Diabetes Care 32* (Supplement 1): S62–S67.
- American Diabetes Association. 2014. Diagnosis and classification of diabetes mellitus. *Diabetes Care.* 37 (Supplement 1): S81-S90.
- Amiera, Z.U.R., Nihayah, M., Wahida, I.F. and Rajab, N.F. 2014. Phytochemical characteristic and uterotonic effect of aqueous extract of *Ficus deltoidea* leaves in rats uterus. *Pakistan Journal of Biological Sciences* 17: 1046–1051.
- Amori, R.E., Lau, J., and Pittas, A.G. 2007. Efficacy and safety of incretin therapy in type 2 diabetes: systematic review and meta-analysis. *The Journal of the American Medical Association (JAMA)* 298 (2): 194–206.
- Anandharajan, R., Pathmanathana, K., Shankernarayananc, N.P., Vishwakarmab, R.A. and Balakrishnana, A. 2005. Upregulation of Glut-4 and PPARγ by an isoflavone from *Pterocarpus marsupium* on L6 myotubes: A possible mechanism of action. *Journal of Ethnopharmacology* 97: 253-260.
- Ardestani, A., and Yazdanparast, R. 2007. Cyperus rotundus suppresses AGE formation and protein oxidation in a model of fructose-mediated protein glycoxidation. *International Journal of Biological Macromolecules*. 41 (5): 572–578.
- Ashcroft F. M. (2005). ATP-sensitive potassium channelopathies: focus on insulin secretion. *The Journal of Clinical Investigation*, 115(8), 2047–2058.

- Ashraf, J.M., Shahabb, U., Tabrezc, S., Leea, E.J., Choi, I. and Ahmad, S. 2015. Quercetin as a finer substitute to aminoguanidine in the inhibition of glycation products. *International Journal of Biological Macromolecules* 77: 188-192.
- Aslantürk, O.S. 2017. In Vitro Cytotoxicity and Cell Viability Assays: Principles, Advantages, and Disadvantages, Genotoxicity - A Predictable Risk to Our Actual World. *IntechOpen*, DOI: 10.5772/intechopen.71923.
- Aslantürk, Ö.S., Çelik, T., Karabey, B., and Karabey, F. 2017. Active phytochemical detecting, antioxidant, cytotoxic, apoptotic activities of ethyl acetate and methanol extracts of Galium aparine L. *British Journal of Pharmaceutical Research* 15 (6):1-16.
- Asokan, A., and Thangavel, D. 2014. Invitro Cytotoxic Studies of crude methanolic extract of Saraca indica bark extract. *IOSR Journal of Pharmacy and Biological Sciences* 9: 26-30.
- Averna, M., De Tullio, R., Salamino, F., Minafra, R., Pontremoli, S. and Melloni, E. 2001. Age-dependent degradation of calpastatin in kidney of hypertensive rats. *The Journal of Biological Chemistry* 276 (42): 38426– 38432.
- Awang, N.A., S.M.Z. Hasan and M.S.B. Shafie, 2013. Morphological study of *Ficus deltoidea* jack in Malaysia. *Journal of Agricultural Science and Technology* 3 (2): 144-150.
- Balu, M., Sangeetha, P., Murali, G., and Panneerselvam, C. 2005. Age-related oxidative protein damages in central nervous system of rats: modulatory role of grape seed extract. *International Journal of Developmental Neuroscience* 23 (6): 501–507.
- Barros, L., Baptista, P., and Ferreira, I.C. 2007. Effect of Lactarius piperatus fruiting body maturity stage on antioxidant activity measured by several biochemical assays. *Food and Chemical Toxicology* 45 (9): 1731–1737.
- Baumann, C. A., Ribon, V., Kanzaki, M., Thurmond, D. C., Mora, S., Shigematsu, S., Bickel, P. E., Pessin, J. E., and Saltiel, A. R. 2000. CAP defines a second signalling pathway required for insulin-stimulated glucose transport. *Nature* 407 (6801): 202–207.
- Bellamy, L., Casas, J. P., Hingorani, A. D., and Williams, D. 2009. Type 2 diabetes mellitus after gestational diabetes: a systematic review and meta-analysis. *Lancet* 373 (9677): 1773–1779.
- Berg, C.C. 2003. Flora Malesiana precursor for the treatment of Moraceae 3: Ficus subgenus Ficus. *Blumea Journal* of Plant Taxonomy and Plant Geography 48 (3): 529–550.

- Berg, C.C., Corner, E.J.H. and Nooteboom, H.P. 2005. Flora Malesiana Series I (Seed Plants). In: *Moraceae (Ficus)* 17 (2): pp.730. National Herbarium of the Netherlands, Leiden. **ISBN** 9071236617.
- Birdee, G.S. and Yeh. G. 2010. Complementary and Alternative Medicine Therapies for Diabetes: A Clinical Review. *Clinical Diabetes* 28 (4): 147-155.
- Bielski, B.H.J., Shiue, G.G., Bajuk, S. 1980. Reduction of nitro blue terazolium by CO-2 and O-2 radicals. *The Journal of Physical Chemistry*. 84: 830– 833.
- Borst, S.E., and Bagby, G.J. 2004. Adipose tumor necrosis factor-α is reduced during onset of insulin resistance in Sprague-Dawley rats. *Cytokine* 26 (5): 217-222.
- Boughton, C.K., Munro, N., Whyte, M. 2017. Targeting beta-cell preservation in the management of type 2 diabetes. *The British Journal of Diabetes* 17 (4): 134-144.
- Brachmann, S.M., Yballe, C.M., Innocenti, M., Deane, J.A., Fruman, D.A., Thomas, S.M., and Cantley, L.C. 2005. Role of phosphoinositide 3kinase regulatory isoforms in development and actin rearrangement. *Molecular and Cellular Biology* 25 (7): 2593–2606.
- Brennan, L., Shine, A., Hewage, C., Malthouse, J.P., Brindle, K.M., McClenaghan, N., Flatt, P.R., and Newsholme, P. 2002. A nuclear magnetic resonance-based demonstration of substantial oxidative Lalanine metabolism and L-alanine-enhanced glucose metabolism in a clonal pancreatic beta-cell line: metabolism of L-alanine is important to the regulation of insulin secretion. *Diabetes* 51(6), 1714–1721.
- Brown, A, Guess, N, Dornhorst, A, Taheri, S, Frost, G. 2017. Insulin-associated weight gain in obese type 2 diabetes mellitus patients: What can be done? *Diabetes Obesity and Metabolism*; 19: 1655 1668.
- Bunawan, H., Amin, N.M., Bunawan, S.N., Baharum, S.N., and Mohd Noor, N. 2014. Ficus deltoidea Jack: A Review on Its Phytochemical and Pharmacological Importance. *Evidence-based Complementary and Alternative Medicine: eCAM* 902734. https://doi.org/10.1155/2014/902734
- Butler, A.E., Janson, J., Bonner-Weir, S., Ritzel, R., Rizza, R.A. and Butler, P.C. 2003. Beta-cell deficit and increased beta-cell apoptosis in humans with type 2 diabetes. *Diabetes* 52 (1): 102–110.
- Camaforte, N., Saldanha, L.L., Vareda, P., Rezende-Neto, J.M., Senger, M.R., Delgado, A.Q., Morgan, H., Violato, N.M., Pieroni, L.G., Dokkedal, A.L., Silva-Júnior, F.P., and Bosqueiro, J.R. 2019. Hypoglycaemic activity of

Bauhinia holophylla through GSK3-β inhibition and glycogenesis activation. *Pharmaceutical Biology* 57(1): 269–279.

- Carmichael, J., DeGraff, W.G., Gadzar, A., Minna, J.D. and Mitchell, J.B. 1987. Evaluation of tetrazolium-based colorimetric assay: assessment of chemosensitivity testing. *Cancer Research* 47: 936-942.
- Caron, A.Z., He, X., Mottawea, W., Seifert, E.L., Jardine, K., Dewar-Darch, D., Cron, G.O., Harper, M.E., Stintzi, A. and McBurney, M.W. 2014, The SIRT1 deacetylase protects mice against the symptoms of metabolic syndrome. *The FASEB Journal* 28: 1306-1316.
- Carragher, N.O. 2006. Calpain inhibition: a therapeutic strategy targeting multiple disease states. *Current Pharmaceutical Design* 12 (5): 615–638.
- Ceddia, R.B. 2005. Direct metabolic regulation in skeletal muscle and fat tissue by leptin: implications for glucose and fatty acids homeostasis. *International Journal of Obesity* 29 (10): 1175–1183.
- Chadt, A. and Al-Hasani, H. 2020. Glucose transporters in adipose tissue, liver, and skeletal muscle in metabolic health and disease. *Pflugers Arch - Eur J Physiol* 472, 1273–1298.
- Chaudhury, A., Duvoor, C., Reddy Dendi, V.S., Kraleti, S., Chada, A., Ravilla, R., Marco, A., Shekhawat, N.S., Montales, M.T., Kuriakose, K., Sasapu, A., Beebe, A., Patil, N., Musham, C.K., Lohani, G.P., and Mirza, W. 2017. Clinical Review of Antidiabetic Drugs: Implications for Type 2 Diabetes Mellitus Management. *Frontier in Endocrinology* 8 (6). https://doi: 10.3389/fendo.2017.00006.
- Chen, C.C., Hsiang, C.Y., Chiang, A.N., Lo, H.Y., and Li, C.I. 2008. Peroxisome proliferator-activated receptor gamma transactivation-mediated potentiation of glucose uptake by Bai-Hu-Tang. *Journal of Ethnopharmacology*, 118(1), 46–50.
- Chen, Y.F., Roan, H.Y., Lii, C.K., Huang, Y.C., Wang, T.S. 2011. Relationship between antioxidant and antiglycation ability of saponins, polyphenols, and polysaccharides in Chinese herbal medicines used to treat diabetes. *Journal of Medicinal Plants Research*. 5: 2322-2331.
- Chen, J. H., Lin, X., Bu, C., and Zhang, X. 2018. Role of advanced glycation end products in mobility and considerations in possible dietary and nutritional intervention strategies. *Nutrition & Metabolism* 15: 72. https://doi.org/10.1186/s12986-018-0306-7.
- Cherney, D.Z., Perkins, B.A., Soleymanlou, N., Maione, M., Lai, V., Lee, A., Fagan, N.M., Woerle, H.J., Johansen, O.E., Broedl, U.C., and von Eynatten, M. 2014. Renal hemodynamic effect of sodium-glucose cotransporter 2 inhibition in patients with type 1 diabetes mellitus. *Circulation* 129 (5): 587–597.

- Cheung, N., Mitchell, P., and Wong, T.Y. 2010. Diabetic retinopathy. *Lancet* 376 (9735): 124–136.
- Chiang, S.H., Baumann, C.A., Kanzaki, M., Thurmond, D.C., Watson, R.T., Neudauer, C.L., Macara, I.G., Pessin, J.E., and Saltiel, A.R. 2001. Insulin-stimulated GLUT4 translocation requires the CAP-dependent activation of TC10. *Nature* 410 (6831): 944–948.
- Chompoo, J., Upadhyay, A., Kishimoto, W., Makise, T., and Tawata, S. 2011. Advanced glycation end products inhibitors from Alpinia zerumbet rhizomes. *Food chemistry* 129 (3): 709–715.
- Choo, C.Y., Sulong, N.Y., Man, F. and Wong, T.W. 2012. Vitexin and isovitexin from the Leaves of *Ficus deltoidea* with in-vivo α-glucosidase inhibition. *Journal of Ethnopharmacology* 142: 776–781.
- Chua, M.T., Tung, Y.T., and Chang, S.T. 2008. Antioxidant activities of ethanolic extracts from the twigs of Cinnamomum osmophloeum. *Bioresource Technology*. 99 (6): 1918-1925
- Ciaraldi, T. P., Carter, L., Mudaliar, S., & Henry, R. R. (2010). GSK-3beta and control of glucose metabolism and insulin action in human skeletal muscle. *Molecular and cellular endocrinology*, 315(1-2), 153–158.
- Coelho, S., Carrilho, P. and Inchaustegui, L. 2013. Management of hyperglycaemia in type 2 diabetic patients with chronic kidney disease. *Portuguese Journal of Nephrology & Hypertension* 27 (2): 91–100.
- Corner, E.J.H. 1969. The Complex of Ficus deltoidea; A Recent Invasion of the Sunda Shelf. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, 256 (808): 281-317. Retrieved August 27, 2020, from http://www.jstor.org/stable/2416885
- Cox S. L. 2004. Rosiglitazone maleate/metformin hydrochloride: a new formulation therapy for type 2 diabetes. *Drugs of Today* (Barcelona, Spain: 1998), 40(7), 633–643.
- Dagogo-Jack, S., and Santiago, J.V. 1997. Pathophysiology of type 2 diabetes and modes of action of therapeutic interventions. *Archives of Internal Medicine* 157 (16): 1802–1817.
- Dali-Youcef, N., Mecili, M., Ricci, R., & Andrès, E. (2013). Metabolic inflammation: connecting obesity and insulin resistance. *Annals of medicine* 45 (3): 242–253.
- Dalle-Donne, I., Rossi, R., Giustarini, D., Milzani, A., and Colombo, R. 2003. Protein carbonyl groups as biomarkers of oxidative stress. *Clinica Chimica Acta; International Journal of Clinical Chemistry* 329 (1-2): 23– 38.

- Deepa, P., Sowndhararajan, K., Kim, S., and Park, S.J. 2018. A role of Ficus species in the management of diabetes mellitus: A review. *Journal of Ethnopharmacology* 215: 210–232.
- DeFronzo, R.A. and Tripathy, D. 2009. Skeletal muscle insulin resistance is the primary defect in type 2 diabetes. Diabetes Care 32 (Supplement 2): S157–S163.
- Derosa, G., and Maffioli, P. 2012. α-Glucosidase inhibitors and their use in clinical practice. *Archives of Medical Science* 8 (5): 899–906.
- Doan, H.V., Riyajan, S., Iyara, R., and Chudapongse, N. 2018. Antidiabetic activity, glucose uptake stimulation and α-glucosidase inhibitory effect of Chrysophyllum cainito L. stem bark extract. *BMC complementary and Alternative Medicine* 18 (1): 267. https://doi.org/10.1186/s12906-018-2328-0
- Ducluzeau, P.H., Fletcher, L.M., Vidal, H., Laville, M., and Tavaré, J. M. 2002. Molecular mechanisms of insulin-stimulated glucose uptake in adipocytes. *Diabetes & Metabolism* 28 (2): 85–92.
- Duraisamy, Y., Gaffney, J., Slevin, M., Smith, C.A., Williamson, K., and Ahmed, N. 2003. Aminosalicylic acid reduces the antiproliferative effect of hyperglycaemia, advanced glycation endproducts and glycated basic fibroblast growth factor in cultured bovine aortic endothelial cells: comparison with aminoguanidine. *Molecular and Cellular Biochemistry* 246 (1-2): 143–153.
- Durruty, P., Sanzana, M., and Sanhueza, L. 2019. Pathogenesis of Type 2 Diabetes Mellitus, Type 2 Diabetes - From Pathophysiology to Modern Management. IntechOpen, DOI: 10.5772/intechopen.83692.
- Eizirik, D.L., Colli, M.L., and Ortis, F. 2009. The role of inflammation in insulitis and beta-cell loss in type 1 diabetes. *Nature Reviews Endocrinology* 5 (4): 219–226.
- Eldor, R., DeFronzo, R.A., and Abdul-Ghani, M. 2013. In vivo actions of peroxisome proliferator-activated receptors: glycemic control, insulin sensitivity, and insulin secretion. *Diabetes care* 36 (Supplement 2): S162–S174.
- Ellman, G.L. 1959. Tissue sulfhydryl groups. Archives of Biochemistry and Biophysics 82 (1): 70–77.
- Elmore, E., Siddiqui, S., Desai, N., Moyer, M.P., Steele, V.E. and Redpath, J.L. 2002. The human epithelial cell cytotoxicity assay for determining tissue specific toxicity: method modifications. *Methods in Cell Science* 24 (4): 145-153.
- Elosta, A., Ghous, T., and Ahmed, N. 2012. Natural products as anti-glycation agents: possible therapeutic potential for diabetic complications. *Current Diabetes Reviews*, 8 (2): 92–108.

- Engelman, J.A., Berg, A.H., Lewis, R.Y., Lisanti, M.P. and Scherer, P.E. 2000. Tumor Necrosis Factor α-Mediated Insulin Resistance, but Not Dedifferentiation, Is Abrogated by MEK1/2 Inhibitors in 3T3-L1 Adipocytes. *Molecular Endocrinology* 14 (10): 1557–1569.
- Fatihah, H.N.N., Nashriyah, M., Zaimah, A.R.N., Khairil, M., Ghani, A.Y., and Ali, A.M. 2014. Leaf morphology and anatomy of 7 varieties of *Ficus deltoidea* (Moraceae). *Turkish Journal of Botany* 38: 677-685.
- Fetita, L.S., Sobngwi, E., Serradas, P., Calvo, F., and Gautier, J.F. 2006. Consequences of fetal exposure to maternal diabetes in offspring. *The Journal of Clinical Endocrinology and Metabolism* 91(10): 3718–3724.
- Francis, G.A., Annicotte, J.S., Auwerx, J., 2003. PPAR agonists in the treatment of atherosclerosis. Current Opinion in Pharmacology 3 (2): 186–191.
- Furusyo, N., and Hayashi, J. 2013. Glycated albumin and diabetes mellitus. *Biochimica et Biophysica Acta* 1830 (12): 5509–5514.
- Gastaldelli, A., Ferrannini, E., Miyazaki, Y., Matsuda, M., Mari, A., and DeFronzo, R.A. 2007. Thiazolidinediones improve beta-cell function in type 2 diabetic patients. *American Journal of Physiology. Endocrinology and Metabolism* 292 (3): E871–E883.
- Geering, B., Pedro, R. Cutillas, G.N., Severine, I. and Gharbi, B.V. 2007. Class IA phosphoinositide 3-kinases are obligate p85-p110 heterodimers. *Proceedings of the National Academy of Sciences* 104 (19): 7809-7814.
- George, P. and McCrimmon, R. 2012. Diazoxide. *Practical Diabetes* 29 (1): 36-37.
- Goldin, A., Beckman, J.A., Schmidt, A.M., and Creager, M.A. 2006. Advanced glycation end products: sparking the development of diabetic vascular injury. *Circulation*. 114 (6): 597–605.
- Gray, A.M., and Flatt, P.R. 1999. Insulin-releasing and insulin-like activity of the traditional anti-diabetic plant Coriandrum sativum (coriander). *The British Journal of Nutrition* 81 (3): 203–209.
- Gray, A.M., Abdel-Wahab, Y.H. and Flatt, P.R. 2000. The traditional plant treatment, Sambucus nigra (elder), exhibits insulin-like and insulin-releasing actions *in vitro*. *Journal of Nutrition* 130: 15-20.
- Guilherme, A., Virbasius, J.V., Puri, V. and Czech, M.P. 2008. Adipocyte dysfunctions linking obesity to insulin resistance and type 2 diabetes. *Nature Reviews Molecular Cell Biology* 9 (5): 367–377.
- Hajer, G.R., van Haeften, T.W., Visseren, F.L. 2008. Adipose tissue dysfunction in obesity, diabetes, and vascular diseases. *European Heart Journal* 29 (4): 2959–2971.

- Hakiman, M. and Marziah, M. 2009. Non enzymatic and enzymatic antioxidant activities in aqueous extract of different Ficus deltoidea accessions. *Journal of Medicinal Plants Research* 3 (3): 120-131.
- Hanafi, M., Afzan, A., Yaakob, H., Aziz, R., Sarmidi, M.R., Wolfender, J.L., and Prieto, J.M. 2017. *In Vitro* Pro-apoptotic and Anti-Migratory Effects of *Ficus deltoidea* L. Plant Extracts on the Human Prostate Cancer Cell Lines PC3. *Frontiers in Pharmacology* 8: 895. https://doi.org/10.3389/fphar.2017.00895.
- Hannan, J.M.A., Marenah, L., Ali, L., Rokeya, B., Flatt, P.R., and Abdel-Wahab, Y.H. 2006. Ocimum sanctum leaf extracts stimulate insulin secretion from perfused pancreas, isolated islets and clonal pancreatic betacells. *The Journal of endocrinology* 189 (1): 127–136.
- Hannan, J.M.A., Marenah, L., Ali, L., Rokeya, B., Flatt, P.R., and Abdel-Wahab, Y.H. 2007. Insulin secretory actions of extracts of Asparagus racemosus root in perfused pancreas, isolated islets and clonal pancreatic betacells. *The Journal of endocrinology* 192 (1): 159–168.
- Hauguel-de Mouzon, S., Lepercq, J., and Catalano, P. 2006. The known and unknown of leptin in pregnancy. *American Journal of Obstetrics and Gynecology*. 194 (6): 1537–1545.
- He, L., Sabet, A., Djedjos, S., Miller, R., Sun, X., Hussain, M. A., Radovick, S., and Wondisford, F.E. 2009. Metformin and insulin suppress hepatic gluconeogenesis through phosphorylation of CREB binding protein. *Cell* 137 (4): 635–646.
- He, Z.X., Zhou, Z.W., Yang, Y., Yang, T., Pan, S.Y., Qiu, J.X., and Zhou, S.F. 2015. Overview of clinically approved oral antidiabetic agents for the treatment of type 2 diabetes mellitus. *Clinical and Experimental Pharmacology & Physiology* 42(2): 125–138.
- Henquin, J.C. (2000). Triggering and amplifying pathways of regulation of insulin secretion by glucose. *Diabetes* 49 (11): 1751–1760.
- Herman, M.A., and Kahn, B.B. 2006. Glucose transport and sensing in the maintenance of glucose homeostasis and metabolic harmony. *The Journal of Clinical Investigation* 116 (7): 1767–1775.
- Hober, D. and Alidjinou, E.K. 2013. Enteroviral pathogenesis of type 1 diabetes: queries and answers. *Current Opinion in Infectious Diseases* 26: 263– 269.
- Horikawa, Y., Oda, N., Cox, N.J., Li, X., Orho-Melander, M., Hara, M., Hinokio, Y., Lindner, T.H., Mashima, H., Schwarz, P.E., del Bosque-Plata, L., Horikawa, Y., Oda, Y., Yoshiuchi, I., Colilla, S., Polonsky, K.S., Wei, S., Concannon, P., Iwasaki, N., Schulze, J., ... Bell, G. I. 2000. Genetic variation in the gene encoding calpain-10 is associated with type 2 diabetes mellitus. *Nature Genetics* 26 (2): 163–175.

- Hotamisligil, G.S., Peraldi, P., Budavari, A., Ellis, R., White, M. F., and Spiegelman, B.M. 1996. IRS-1-mediated inhibition of insulin receptor tyrosine kinase activity in TNF-alpha- and obesity-induced insulin resistance. *Science* 271(5249): 665–668.
- Hu, C. and Jia, W. 2018. Diabetes in China: Epidemiology and Genetic Risk Factors and Their Clinical Utility in Personalized Medication. *Diabetes* 67 (1): 3-11.
- Huang, S., and Czech, M.P. 2007. The GLUT4 glucose transporter. *Cell Metabolism* 5 (4): 237–252.
- Huang, T.H., Peng, G., Kota, B.P., Li, G.Q., Yamahara, J., Roufogalis, B.D., and Li, Y. 2005. Anti-diabetic action of Punica granatum flower extract: activation of PPAR-gamma and identification of an active component. *Toxicology and Applied Pharmacology* 207 (2):160–169.
- Hussein, Z., Taher, S.W., Gilcharan Singh, H.K., and Chee Siew Swee, W. 2015. Diabetes Care in Malaysia: Problems, New Models, and Solutions. *Annals of Global Health* 81 (6): 851–862.
- Imam, M.U., Ismail, M., Ithnin, H., Tubesha, Z., and Omar, A.R. 2013. Effects of germinated brown rice and its bioactive compounds on the expression of the peroxisome proliferator-activated receptor gamma gene. *Nutrients* 5 (2): 468–477.
- Institute for Public Health. 2020. National Health and Morbidity Survey (NHMS) 2019: Non-communicable diseases, healthcare demand, and health literacy - Key Findings.http://www.iku.gov.my/images/IKU/Document/REPORT/NHM S2019/Report\_NHMS2019-NCD\_v2.pdf. Accessed on 7 September 2020.
- Jaldin-Fincati, J.R., Pavarotti, M., Frendo-Cumbo, S., Bilan, P.J., and Klip, A. 2017. Update on GLUT4 Vesicle Traffic: A Cornerstone of Insulin Action. *Trends in Endocrinology and Metabolismn* 28 (8): 597–611.
- Janus, A., Szahidewicz-Krupska, E., Mazur, G., and Doroszko, A. 2016. Insulin Resistance and Endothelial Dysfunction Constitute a Common Therapeutic Target in Cardiometabolic Disorders. *Mediators of inflammation* 3634948. https://doi.org/10.1155/2016/3634948.
- Jariyapamornkoon, N., Yibchok-anun, S., and Adisakwattana, S. 2013. Inhibition of advanced glycation end products by red grape skin extract and its antioxidant activity. *BMC Complementary and Alternative Medicine* 13: 171. https://doi.org/10.1186/1472-6882-13-171
- Joseph, J.J., Echouffo-Tcheugui, J.B., Golden, S.H., Chen, H., Jenny, N.S., Carnethon, M.R., Jacobs, D., Jr, Burke, G.L., Vaidya, D., Ouyang, P., and Bertoni, A.G. 2016. Physical activity, sedentary behaviors and the incidence of type 2 diabetes mellitus: The Multi-Ethnic Study of

Atherosclerosis (MESA). *BMJ Open Diabetes Research & Care* 4 (1): e000185. https://doi.org/10.1136/bmjdrc-2015-000185

- Jurgens, C.A., Toukatly, M.N., Fligner, C.L., Udayasankar, J., Subramanian, S.L., Zraika, S., Aston-Mourney, K., Carr, D.B., Westermark, P., Westermark, G.T., Kahn, S.E., and Hull, R.L. 2011. β-cell loss and β-cell apoptosis in human type 2 diabetes are related to islet amyloid deposition. *The American Journal of Pathology* 178 (6): 2632–2640.
- Kaku, K. 2010. Pathophysiology of Pathophysiology of Type 2 Diabetes and Its Treatment Policy. *Japan Medical Association Journal* 53 (1): 41–46.
- Kalra, S. 2014. Sodium Glucose Co-Transporter-2 (SGLT2) Inhibitors: A Review of Their Basic and Clinical Pharmacology. *Diabetes Therapy: Research, Treatment and Education of Diabetes and Related Disorders* 5 (2): 355– 366.
- Kalwat, M. A., and Cobb, M. H. (2017). Mechanisms of the amplifying pathway of insulin secretion in the β cell. *Pharmacology & Therapeutics*, 179, 17–30.
- Kamarudin, A.M. and Latiff, M.S. 2002. Tumbuhan Ubatan Malaysia. Percetakan Watan Sdn. Bhd., Malaysia (in Bahasa Melayu).
- Kamuhabwa, A., Nshimo, C. and De Witte, P. 2000. Cytotoxicity of some medicinal plant extracts used in Tanzanian traditional medicine. *Journal of Ethnopharmacology* 70: 143-149.
- Kanzaki M. 2006. Insulin receptor signals regulating GLUT4 translocation and actin dynamics. *Endocrine Journal* 53 (3): 267–293.
- Kanzaki, M., and Pessin, J. E. 2001. Insulin-stimulated GLUT4 translocation in adipocytes is dependent upon cortical actin remodeling. *The Journal of Biological Chemistry* 276 (45): 42436–42444.
- Kanzaki, M. and Pessin, J.E. 2003. Insulin signaling: GLUT4 vesicles exit via the exocyst. *Current Biology* 13: R574–R576.
- Karlsson, M. G., and Ludvigsson, J. 2000. The ABBOS-peptide from bovine serum albumin causes an IFN-gamma and IL-4 mRNA response in lymphocytes from children with recent onset of type 1 diabetes. *Diabetes Research and Clinical Practice*, 47(3), 199–207.
- Karpe, F., Dickmann, J. R., and Frayn, K. N. 2011. Fatty acids, obesity, and insulin resistance: time for a reevaluation. *Diabetes* 60 (10): 2441–2449.
- Kawasaki, Y., Fujii, J., Miyazawa, N., Hoshi, A., Okado, A., Tano, Y., and Naoyuki, T. 1998. Specific detections of the early process of the glycation re- action by fructose and glucose in diabetic rat lens. *FEBS Letters* 441: 116-120.

- Kellerer, M., Lammers, R., and Haring, H.U. 1999. Insulin signal transduction; possible mechanism for insulin resistance. *Experimental and Clinical Endocrinology and Diabetes* 107: 97–106.
- Knip, M., Virtanen, S. M., Becker, D., Dupré, J., Krischer, J. P., Åkerblom, H. K., and TRIGR Study Group. 2011. Early feeding and risk of type 1 diabetes: experiences from the Trial to Reduce Insulin-dependent diabetes mellitus in the Genetically at Risk (TRIGR). *The American Journal of Clinical Nutrition*, 94(6 Suppl), 1814S–1820S. https://doi.org/10.3945/ajcn.110.000711
- Kochummen, K.M. 1978. *Moraceae* In: Ng FSP (ed) Tree flora of Malaya (A manual of foresters) volume three, *Longman Malaysia Sdn. Bhd*.
- Körner, A., Wabitsch, M., Seidel, B., Fischer-Posovszky, P., Berthold, A., Stumvoll, M., Blüher, M., Kratzsch, J., and Kiess, W. 2005. Adiponectin expression in humans is dependent on differentiation of adipocytes and down-regulated by humoral serum components of high molecular weight. *Biochemical and Biophysical Research Communications* 337 (2): 540–550.
- Koster, J.C., Permutt, M.A., and Nichols, C.G. 2005. Diabetes and insulin secretion: the ATP-sensitive K+ channel (KATP) connection. *Diabetes* 54 (11): 3065–3072.
- Kousar, S., Sheikh, M.A., Asghar, M. and Rashid, R. 2008. Effect of onion (*Allium* cepa L.) extract on Maillard reaction under in vitro conditions. *Pakistan Journal of Agricultural Sciences* 45 (1):103-106.
- Kovacs, L., and Su, Y. 2014. The Critical Role of Calpain in Cell Proliferation. Journal of Biomolecular Research & Therapeutics, 3(3), 1000112. https://doi.org/10.4172/2167-7956.1000112
- Krogvold, L., Skog, O., Sundström, G., Edwin, B., Buanes, T., Hanssen, K.F., Ludvigsson, J., Grabherr, M., Korsgren, O., and Dahl-Jørgensen, K.
  2015. Function of Isolated Pancreatic Islets from Patients at Onset of Type 1 Diabetes: Insulin Secretion Can Be Restored After Some Days in a Nondiabetogenic Environment in Vitro: Results from the DiViD Study. *Diabetes* 64 (7), 2506–2512.
- Kumar, S., Narwal, S., Kumar, V., and Prakash, O. 2011. α-glucosidase inhibitors from plants: A natural approach to treat diabetes. *Pharmacognosy Reviews*, 5(9), 19–29.
- Kumar, P.M., Venkataranganna, M.V., Manjunath, K., Viswanatha, G.L., and Ashok, G. 2014. Methanolic extract of Momordica cymbalaria enhances glucose uptake in L6 myotubes in vitro by up-regulating PPAR-γ and GLUT-4. *Chinese Journal of Natural Medicines* 12 (12): 895-900.
- Kusirisin, W., Srichairatanakool, S., Lerttrakarnnon, P., Lailerd, N., Suttajit, M., Jaikang, C., and Chaiyasut, C. 2009. Antioxidative activity, polyphenolic

content and anti-glycation effect of some Thai medicinal plants traditionally used in diabetic patients. *Medicinal Chemistry* 5 (2): 139–147.

- Leahy, J.L. 2005. Pathogenesis of type 2 diabetes mellitus. *Archives of Medical Research* 36 (3): 197–209.
- Lehrke, M., and Lazar, M.A. 2005. The many faces of PPAR-gamma. *Cell* 123 (6): 993–999.
- Leney, S.E. and Tavare, J.M. 2009. The molecular basis of insulin-stimulated glucose uptake: signalling, trafficking and potential drug targets. *Journal of Endocrinology* 203: 1-18.
- Levine, R.L., Garland, D., Oliver, C.N., Amici, A., Climent, I., Lenz, A.G., Ahn, B.W., Shaltiel, S., and Stadtman, E.R. 1990. Determination of carbonyl content in oxidatively modified proteins. *Methods in Enzymology*. 186: 464–478.
- Li, M., Han, Z., Bei, W., Rong, X., Guo, J., and Hu, X. 2015. Oleanolic acid attenuates insulin resistance via NF-ÎoB to regulate the IRS1-GLUT4 pathway in HepG2Cells. *Evidence-based Complementary and Alternative Medicine* 643102. https://doi.org/10.1155/2015/643102
- Li, N., Brun, T., Cnop, M., Cunha, D. A., Eizirik, D. L., and Maechler, P. (2009). Transient oxidative stress damages mitochondrial machinery inducing persistent beta-cell dysfunction. *The Journal of Biological Chemistry* 284 (35): 23602–23612.
- Lin, S.H., Cheng, P.C., Tu, S.T., Hsu, S.R., Cheng, Y.C., and Liu, Y.H. 2018. Effect of metformin monotherapy on serum lipid profile in statin-naïve individuals with newly diagnosed type 2 diabetes mellitus: a cohort study. *PeerJ* 6 e4578. https://doi.org/10.7717/peerj.4578
- Lip, J.M., Hisham, D.N., Zaidi, J.A., Musa, Y., Ahmad, A.W., Normah, A., Sharizan, A. 2009. Isolation and identification of moretenol from *Ficus deltoidea* leaves. *Journal of Tropical Agriculture and Food Science* 37: 195–201.
- Liu, R.H., Mizuta, M., Kurose, T. and Matsukura, S. 2002. Early Events Involved in the Development of Insulin Resistance in Zuker Fatty Rats. *International Journal of Obesity* 26: 318-326.
- Lobo, V., Patil, A., Phatak, A. and Chandra, N. 2010. Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacognosy Reviews.* 4 (8): 118-26.
- Logie, L.J., Brown, A.E., Yeaman, S.J., and Walker, M. 2005. Calpain inhibition and insulin action in cultured human muscle cells. Molecular Genetics and Metabolism. 85 (1): 54–60.

- Lotito, S.B., and Frei, B. 2006. Consumption of flavonoid-rich foods and increased plasma antioxidant capacity in humans: cause, consequence, or epiphenomenon?. *Free Radical Biology & Medicine*, 41 (12): 1727–1746.
- Ma, H., Fukiage, C., Kim, Y. H., Duncan, M. K., Reed, N. A., Shih, M., Azuma, M., & Shearer, T. R. (2001). Characterization and expression of calpain 10. A novel ubiquitous calpain with nuclear localization. *The Journal of Biological Chemistry* 276 (30): 28525–28531.
- Ma, R.C., Lin, X., and Jia, W. 2014. Causes of type 2 diabetes in China. *Lancet Diabetes Endocrinology* 2: 980–991
- Ma, X., Iwanaka, N., Masuda, S., Karaike, K., Egawa, T., Hamada, T., Toyoda, T., Miyamoto, L., Nakao, K., and Hayashi, T. 2009. Morus alba leaf extract stimulates 5'-AMP-activated protein kinase in isolated rat skeletal muscle. *Journal of Ethnopharmacology* 122 (1): 54–59.
- Maarof Alam, M., Ahmad, I., Naseem, I. 2015. Inhibitory effect of quercetin in the formation of advance glycation end products of human serum albumin: An in vitro and molecular interaction study. *International Journal of Biological Macromolecules* 79: 336-343.
- Mahomoodally, F.M., Subratty, A.H., Gurib-Fakim, A. and Choudhary, M.I. 2012. Antioxidant, antiglycation and cytotoxicity evaluation of selected medicinal plants of the Mascarene Islands. *BMC Complementary and Alternative Medicine* 12 (165) https://doi.org/10.1186/1472-6882-12-165.
- Manurung, H., Kustiawan, W., Kusuma, I.W., Marjenah. 2017. Total Flavonoid Content and Antioxidant Activity in Leaves and Stems Extract of Cultivated and Wild Tabat Barito (*Ficus deltoidea* Jack.) *AIP Conference Proceedings* 183 (020007) doi: 10.1063/1.4975945.
- Marín-Peñalver, J.J., Martín-Timón, I., Sevillano-Collantes, C., del Cañizo-Gómez, F.J. 2016. Update on the treatment of type 2 diabetes mellitus. World Journal of Diabetes 7 (17): 354-395.
- Martineau, L.C., Couture, A., Spoor, D., Benhaddou-Andaloussi, A., Harris, C., Meddah, B., Leduc, C., Burt, A., Vuong, T., Mai Le, P., Prentki, M., Bennett, S.A., Arnason, J.T., and Haddad, P.S. 2006. Anti-diabetic properties of the Canadian lowbush blueberry Vaccinium angustifolium Ait. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology* 13 (9-10): 612–623.
- Mastura, I., Chew, B.H., Lee, P.Y., Cheong, A., Ghazali, S., and Abdul, S. 2011. Control and treatment profiles of 70,889 adult type 2 diabetes mellitus patients in Malaysia - a cross sectional survey in 2009. *International Journal of Collaborative Research on Internal Medicine and Public Health 3*: 98-113.

- Mat Akhir, N.A., Chua, L.S., Majid, F.A.A. and Sarmidi, M.R. 2011. Cytotoxicity of Aqueous and Ethanolic Extracts of *Ficus deltoidea* on Human Ovarian Carcinoma Cell Line. *British Journal of Medicine and Medical Research* 1 (4): 397-409
- Mat, N., N.A. Rosni, N.Z. Ab Rashid, N. Haron and Z.M. Nor, Hasan Nudin, N.F., Yunus, A.G. and Ali, A.M. 2012. Leaf morphological variations and heterophylly in Ficus deltoidea Jack (Moraceae). *Sains Malaysiana* 41: 527-538.
- Mat-Salleh, K. and Latif, A., 2002. Dikotiledon: Subkelas Hamamelidae. *Ficus deltoidea* Jack. In: *Tumbuhan Ubatan Malaysia*. Pusat Pengurusan Penyelidikan, Universiti Kebangsaan Malaysia., pp: 184-185. ISBN 983-2446-39-2.
- Mathews, J.N., Flatt, P.R. and Abdel-Wahab, Y.H. 2006. Asparagus adscendens (Shweta musali) stimulates insulin secretion, insulin action and inhibits starch digestion. *British Journal of Nutrition*. 95(3):576–581.
- Matsuzawa, Y., Funahashi, T., Kihara, S., and Shimomura, I. 2004. Adiponectin and metabolic syndrome. *Arteriosclerosis, Thrombosis, and Vascular Biology* 24 (1): 29–33.
- McArdle, M.A., Finucane, O.M., Connaughton, R.M., McMorrow, A.M., and Roche, H.M. 2013. Mechanisms of obesity-induced inflammation and insulin resistance: insights into the emerging role of nutritional strategies. *Frontiers in Endocrinology* 4 (52). https://doi.org/10.3389/fendo.2013.00052.
- McIntyre, H.D., Catalano, P., Zhang, C., Desoye, G., Mathiesen, E.R., and Damm, P. 2019. Gestational diabetes mellitus. *Nature Reviews. Disease Primers* 5 (1) 47. https://doi.org/10.1038/s41572-019-0098-8
- Meigs, J.B., Cupples, L.A., and Wilson, P.W. 2000. Parental transmission of type 2 diabetes: The Framingham Offspring Study. *Diabetes* 49 (12): 2201–2207.
- Menser, M.A., Forrest, J.M., and Bransby, R.D. 1978. Rubella infection and diabetes mellitus. *Lancet* 1 (8055): 57–60.
- Misbah, H., Aziz, A.A., and Aminudin, N. 2013. Antidiabetic and antioxidant properties of Ficus deltoidea fruit extracts and fractions. *BMC Complementary and Alternative Medicine* 13: 118. https://doi.org/10.1186/1472-6882-13-118
- Modi, P. 2007. Diabetes beyond insulin: review of new drugs for treatment of diabetes mellitus. *Current Drug Discovery Technologies* 4 (1): 39–47.
- Mohammad Noor, H.S., Ismail, N.H. Kasim, N., Mohd Zohdi, R. and Ali, A.M. 2016. Hypoglycemic and glucose tolerance activity of standardized

extracts Ficus deltoidea varieties in normal rats. Journal of Medicinal Plants Studies 4 (5): 275-279.

- Mohammad Noor, H.S., Ismail, N.H., Kasim, N., Mediani, A., Mohd Zohdi, R., Ali, A.M., Mat, N., and Al-Mekhlafi, N.A. 2020. Urinary Metabolomics and Biochemical Analysis of Antihyperglycemic Effect of Ficus deltoidea Jack Varieties in Streptozotocin-Nicotinamide-Induced Diabetic Rats. *Applied Biochemistry and Biotechnology* 192 (1): 1–21.
- Mohd, K.S., Rosli, A.S., Azemin, A., Mat, N., Zakaria, A.J. 2016. Comprehensive Biological Activities Evaluation and Quantification of Marker Compounds of *Ficus deltoiea Jack* Varieties. *International Journal of Pharmacognosy and Phytochemical Research*. 8 (10): 1698-1708.
- Moini, J. 2019. Type 2 Diabetes. Epidemiology of Diabetes. Elsevier 91-114. ISBN: 9780128168646.
- Moller D.E. 2000. Potential role of TNF-alpha in the pathogenesis of insulin resistance and type 2 diabetes. *Trends in Endocrinology and Metabolism* 11 (6): 212–217.
- Mosihuzzman, M., Naheed, S., Hareem, S., Talib, S., Abbas, G., Khan, S.N., Choudhary, M.I., Sener, B., Tareen, R.B., and Israr, M. 2013. Studies on α-glucosidase inhibition and anti-glycation potential of Iris loczyi and Iris unguicularis. *Life Sciences* 92 (3): 187–192.
- Mosmann T. 1983. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of Immunological Methods* 65 (1-2): 55–63.
- Mukhtar, H.M., Singh, A., Soni, V., Singh, A., and Kaur, H. 2019. Development and validation of RP-HPLC method for the determination of stigmasterol in the botanical extract of *Ficus deltoidea*. *Natural Product Research* 33 (19): 2868–2872.
- Musa, Y. 2005. Variability in morphology and agronomy of emas cotek accessions found in Kelantan and Terengganu. Buletin Teknologi Tanaman 2: 35–48.
- Musapha, Z. and Harun, H. 2015. Phytochemical Constituents in leaves and callus of *Ficus deltoidea Jack* var. *Kunstleri* (King) corner. *Walailak Journal of Science and Technology* 12: 431–439.
- Nagai, R., Ikeda, K., Higashi, T., Sano, H., Jinnouchi, Y., Araki, T., and Horiuchi, S. 1997. Hydroxyl radical mediates N epsilon-(carboxymethyl)lysine formation from Amadori product. *Biochemical and Biophysical Research Communications* 234 (1): 167–172.
- Nakatani, S., Kakehashi, A., Ishimura, E., Yamano, S., Mori, K., Wei, M., Inaba, M., and Wanibuchi, H. 2011. Targeted proteomics of isolated glomeruli

from the kidneys of diabetic rats: sorbin and SH3 domain containing 2 is a novel protein associated with diabetic nephropathy. *Experimental Diabetes Research* 979354. https://doi.org/10.1155/2011/979354

- Nasma, A., Aishath, N. Azilah, A. and Sulaiman, A.Z. 2018. Optimization of vitexin and isovitexin compounds extracted from dried Mas Cotek leaves using one-factor-at-a-time (OFAT) approach in aqueous extraction. *International Food Research Journal* 25 (6): 2560-2571.
- National Center for Complementary and Integrative Health, National Institutes of Health. 2018. Complementary, Alternative, or Integrative Health: What's in a Name? [article online] Available from https://www.nccih.nih.gov/health/complementary-alternative-orintegrative-health-whats-in-a-name. Accessed on 11 September 2020.
- Nawrocki, A.R., Rajala, M.W., Tomas, E., Pajvani, U.B., Saha, A.K., Trumbauer, M.E., Pang, Z., Chen, A.S., Ruderman, N.B., Chen, H. Rossetti, L., and Scherer, P.E. 2006. Mice lacking adiponectin show decreased hepatic insulin sensitivity and reduced responsiveness to peroxisome proliferator-activated receptor gamma agonists. *The Journal of Biological Chemistry* 281 (5): 2654–2660.
- Nikzamir, A., Palangi, A., Kheirollaha, A., Tabar, H., Malakaskar, A., Shahbazian, H., and Fathi, M. 2014. Expression of Glucose Transporter 4 (GLUT4) is Increased by Cinnamaldehyde in C2C12 Mouse Muscle Cells. *Iranian Red Crescent Medical Journal* 16 (2): e13426. https://doi.org/10.5812/ircmj.13426
- Norris, J.M., Barriga, K., Klingensmith, G., Hoffman, M., Eisenbarth, G.S., Erlich, H.A. and Rewers, M. 2003. Timing of initial cereal exposure in infancy and risk of islet autoimmunity. *The Journal of the American Medical Association* 290 (13): 1713–1720.
- Norris, J.M., Yin, X., Lamb, M.M., Barriga, K., Seifert, J., Hoffman, M., Orton, H.D., Barón, A.E., Clare-Salzler, M. and Chase, H.P., Szabo, N.J., Erlich, H., Eisenbarth, G.S., and Rewers, M. 2007. Omega-3 polyunsaturated fatty acid intake and islet autoimmunity in children at increased risk for type 1 diabetes. *JAMA*. 298 (12): 1420-1428.
- Ntambi, J.M., and Young Cheul, K. 2000. Adipocyte differentiation and gene expression. *The Journal of Nutrition* 130 (12): 3122S-3126S.
- Ojo O.O. and Ojo C.C. 2015. Insulinotropic actions of *Moringa oleifera* involves the induction of membrane depolarization and enhancement of intracellular calcium concentration. *Journal of Experimental and Integrative Medicine* 5 (1): 36 – 41.
- Olefsky, J.M. 2000. Treatment of insulin resistance with peroxisome proliferator– activated receptor  $\gamma$  agonists. *The Journal of Clinical Investigation* 106: 467–72.

- Omar, M.H., Mullen, W. and Crozier, A. 2011. Identification of proanthocyanidin dimers and trimers, flavone C-glycosides, and antioxidants in *ficus deltoidea*, a Malaysian herbal tea. *Journal of Agricultural and Food Chemistry* 59 (4): 1363–1369.
- Ormazabal, V., Nair, S., Elfeky, O., Aguayo, C., Salomon, C., and Zuñiga, F.A. 2018. Association between insulin resistance and the development of cardiovascular disease. *Cardiovascular Diabetology* 17 (1): 122. https://doi.org/10.1186/s12933-018-0762-4
- Otani, K., Han, D.H., Ford, E.L., Garcia-Roves, P.M., Ye, H., Horikawa, Y., Bell, G.I., Holloszy, J.O., and Polonsky, K.S. 2004. Calpain system regulates muscle mass and glucose transporter GLUT4 turnover. *The Journal of Biological Chemistry*, 279 (20): 20915–20920.
- Ott, C., Jacobs, K., Haucke, E., Navarrete Santos, A., Grune, T., and Simm, A. 2014. Role of advanced glycation end products in cellular signaling. *Redox Biology* 2: 411–429.
- Ottum, M. S., and Mistry, A. M. 2015. Advanced glycation end-products: modifiable environmental factors profoundly mediate insulin resistance. *Journal of Clinical Biochemistry and Nutrition* 57 (1): 1–12.
- Owen, M. R., Doran, E., & Halestrap, A. P. 2000. Evidence that metformin exerts its anti-diabetic effects through inhibition of complex 1 of the mitochondrial respiratory chain. *The Biochemical Journal*, 348 Pt 3(Pt 3), 607–614.
- Oyaizu, M. 1986. Studies on product of browning reaction prepared from glucose amine. *The Japanese Journal of Ntrition and Diatetics*. 44: 307-315.
- Pajvani, U.B., Du, X., Combs, T.P., Berg, A.H., Rajala, M.W., Schulthess, T., Engel, J., Brownlee, M., and Scherer, P.E. 2003. Structure-function studies of the adipocyte-secreted hormone Acrp30/adiponectin: Implications for metabolic regulation and bioactivity. *Journal of Biological Chemistry* 278 (11): 9073-9085.
- Pal, J., Ganguly, S., Tahsin, K.S., and Acharya, K. 2010. In vitro free radical scavenging activity of wild edible mushroom *Pleurotus squarrosulus* (Mont.) Singer. *Indian Journal of Experimental Biology*. 47: 1210-1218.
- Park, Y.J. and Woo, M. 2019. Pancreatic β cells: Gatekeepers of type 2 diabetes. *Journal of Cell Biology* 218 (4): 1094–1095.
- Paschou, S.A., Papadopoulou-Marketou, N., Chrousos, G.P. and Kanaka-Gantenbein, C. 2018. On type 1 diabetes mellitus pathogenesis. *Endocrine Connections.* 7: R38–R46. https://doi.org/10.1530/EC-17-0347.

- Paul, D.S., Harmon, A.W., Winston, C.P., and Patel, Y.M. 2003. Calpain facilitates GLUT4 vesicle translocation during insulin-stimulated glucose uptake in adipocytes. *The Biochemical Journal* 376 (3): 625–632.
- Paul, S., Ali, A., and Katare, R. 2020. Molecular complexities underlying the vascular complications of diabetes mellitus - A comprehensive review. *Journal of Diabetes and Its Complications* 34 (8): 107613. DOI: 10.1016/j.jdiacomp.2020.107613.
- Peppa, M., and Vlassara, H. 2005. Advanced glycation end products and diabetic complications: a general overview. *Hormones* 4 (1): 28–37.
- Petersen, K.F., Krssak, M., Inzucchi, S., Cline, G.W., Dufour, S., and Shulman, G.I. 2000. Mechanism of troglitazone action in type 2 diabetes. *Diabetes*, 49 (5): 827–831.
- Phielix, E., Meex, R., Moonen-Kornips, E., Hesselink, M.K., and Schrauwen, P. 2010. Exercise training increases mitochondrial content and ex vivo mitochondrial function similarly in patients with type 2 diabetes and in control individuals. *Diabetologia*, 53 (8): 1714–1721.
- Popa, S. and Mota, M. 2013. Beta-Cell Function and Failure in Type 2 Diabetes, Type 2 Diabetes. IntechOpen, DOI: 10.5772/56467.
- Povichit, N., Phrutivorapongkul, A., Suttajit, M., Chaiyasut, C.C., and Leelapornpisid, P. 2010. Phenolic content and in vitro inhibitory effects on oxidation and protein glycation of some Thai medicinal plants. *Pakistan Journal of Pharmaceutical Sciences.* 23 (4): 403-408.
- Pratt, E.P.S., Harvey, K.E., Salyer, A.E. and Hockerman, G.H. 2019. Regulation of cAMP accumulation and activity by distinct phosphodiesterase subtypes in INS-1 cells and human pancreatic β-cells. *PLOS ONE* 14(8): e0215188. https://doi.org/10.1371/journal.pone.0215188
- Proks, P., Reimann, F., Green, N., Gribble, F., and Ashcroft, F. 2002. Sulfonylurea stimulation of insulin secretion. *Diabetes* 51 (Supplement 3): S368–S376.
- Qian, D., Zhang, T., Zheng, P., Liang, Z., Wang, S., Xie, J., Zhao, L., Zahng, Y., and Situ, B. 2018. Comparison of Oral Antidiabetic Drugs as Add-On Treatments in Patients with Type 2 Diabetes Uncontrolled on Metformin: A Network Meta-Analysis. *Diabetes Therapy* 9: 1945–1958.
- Qiu, C., Williams, M.A., Vadachkoria, S., Frederick, I.O., and Luthy, D.A. 2004. Increased maternal plasma leptin in early pregnancy and risk of gestational diabetes mellitus. *Obstetrics and Gynecology.* 103 (3): 519– 525.
- Quintanilla Rodriguez, B.S., and Correa, R. Rosiglitazone. 2021 [Updated 2021 Jul 18]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing. https://www.ncbi.nlm.nih.gov/books/NBK544230/

- Raab, J., Giannopoulou, E.Z., Schneider, S., Warncke, K., Krasmann, M., Winkler, C. and Ziegler, A.G. 2014. Prevalence of vitamin D deficiency in pre-type 1 diabetes and its association with disease progression. *Diabetologia* 57 (5): 902–908.
- Rabbani, N., and Thornalley, P.J. 2012. Methylglyoxal, glyoxalase 1 and the dicarbonyl proteome. *Amino acids* 42 (4): 1133–1142.
- Rahbar, S., and Figarola, J.L. 2003. Novel inhibitors of advanced glycation endproducts. *Archives of Biochemistry and Biophysics* 419 (1): 63–79.
- Rahimi, R., Nikfar, S., Larijani, B. and Abdollahi, M. 2005. A review on the role of antioxidants in the management of diabetes and its complications. *Biomedicine & Pharmacotherapy* 59: 365–373.
- Ramachandran, A. 2014. Know the signs and symptoms of diabetes. *The Indian Journal of Medical Research* 140 (5):579-81.
- Ramkissoon, J.S., Mahomoodally, M.F., Ahmed, N., Subratty, A.H. 2013. Antioxidant and anti-glycation activities correlates with phenolic composition of tropical medicinal herbs. *Asian Pacific Journal of Tropical Medicine*. 561-569.
- Ramkissoon, J.S., Mahomoodally, M.F., Subratty, A.H. and Ahmed, N. 2016. Inhibition of glucose- and fructose-mediated protein glycation by infusions and ethanolic extracts of ten culinary herbs and spices. *Asian Pacific Journal of Tropical Biomedicine* 6 (6): 492–500.
- Razani, B., Chakravarthy, M.V. and Semenkovich, C.F. 2008. Insulin resistance and atherosclerosis. *Endocrinology and Metabolism Clinics of North America* 37: 603-621.
- Redondo, M.J., Rewers, M., Yu, L., Garg, S., Pilcher, C.C., Elliott, R.B. and Eisenbarth, G.S. 1999. Genetic determination of islet cell autoimmunity in monozygotic twin, dizygotic twin, and non-twin siblings of patients with type 1 diabetes: prospective twin study. *British Medical Journal* 318 (7185): 698–702.
- Reilly, M.P., Iqbal, N., Schutta, M., Wolfe, M.L., Scally, M., Localio, A.R., Rader, D.J., and Kimmel, S.E. 2004. Plasma leptin levels are associated with coronary atherosclerosis in type 2 diabetes. *The Journal of Clinical Endocrinology and Metabolism*, 89 (8): 3872–3878.
- Rena, G., Hardie, D.G., and Pearson, E.R. 2017. The mechanisms of action of metformin. *Diabetologia* 60 (9): 1577–1585.
- Ribon, V., Johnson, J.H., Camp, H.S., Saltiel, A.R. 1998. Thiazilidinediones and insulin resistance: peroxisome proliferator activated receptor gamma activation stimulates expression of the CAP gene. *Proceedings of the National Academy of Sciences* 25: 14751-14756.

- Röhling, M., Herder, C., Stemper, T., and Müssig, K. 2016. Influence of Acute and Chronic Exercise on Glucose Uptake. *Journal of Diabetes Research* 2868652. https://doi.org/10.1155/2016/2868652.
- Rose, A.J., and Richter, E.A. 2005. Skeletal muscle glucose uptake during exercise: how is it regulated? *Physiology* 20: 260–270.
- Ruiz-Ojeda, F.J., Rupérez, A.I., Gomez-Llorente, C., Gil, A., and Aguilera, C.M. 2016. Cell Models and Their Application for Studying Adipogenic Differentiation in Relation to Obesity: A Review. *International journal of molecular sciences*, 17(7), 1040. https://doi.org/10.3390/ijms17071040
- Sami, W., Ansari, T., Butt, N. S., and Hamid, M. 2017. Effect of diet on type 2 diabetes mellitus: A review. International journal of health sciences, 11(2), 65–71.
- Satoh, T. 2014. Molecular Mechanisms for the Regulation of Insulin-Stimulated Glucose Uptake by Small Guanosine Triphosphatases in Skeletal Muscle and Adipocytes. *International Journal of Molecular Sciences* 15, 18677-18692.
- Schalkwijk, C.G., Stehouwer, C.D., and van Hinsbergh, V.W. 2004. Fructosemediated non-enzymatic glycation: sweet coupling or bad modification. *Diabetes/ Metabolism Research and Reviews* 20 (5): 369– 382.
- Scott, P.H., Brunn, G.J., Kohn, A.D., Roth, R.A., and Lawrence, J.C., Jr 1998. Evidence of insulin-stimulated phosphorylation and activation of the mammalian target of rapamycin mediated by a protein kinase B signaling pathway. *Proceedings of the National Academy of Sciences of the United States of America* 95 (13): 7772–7777.
- Sharma, S.D., Pandey, B.N., Mishra, K.P., and Sivakami, S. 2002. Amadori product and age formation during nonenzymatic glycosylation of bovine serum albumin in vitro. *Journal of Biochemistry, Molecular Biology, and Biophysics*. 6 (4): 233–242.
- Sharma, Y., Saxena, S., Mishra, A., Saxena, A., and Natu, S. M. 2013. Advanced glycation end products and diabetic retinopathy. Journal of ocular biology, diseases, and informatics, 5(3-4), 63–69.
- Sharp, G.W. 1979 The adenylate cyclase-cyclicAMP system in islets of Langerhans and its role in the control of insulin release. *Diabetologia* 16 (5): 287–297.
- Shepherd, P.R., and Kahn, B.B. 1999. Glucose transporters and insulin action-implications for insulin resistance and diabetes mellitus. *The New England Journal of Medicine* 341 (4): 248–257.
- Shimada, K., Fujikawa, K., Yahara, K., Nakamura, T. 1992. Antioxidative Properties of Xanthan on the Autoxidation of Soybean Oil in Cyclodextrin Emulsion. *Journal of Agricultural and Food Chemistry*. 40 (6): 945-948.

- Shivaprasad, H.N., Mohan, S., Kharya, M.D., Shiradkar, R.M. and Lakshman, K. 2005. Invitro models for antioxidant activity evaluation: A review. Pharmainfo Net 3 (4): 1-11
- Sicree, R., Shaw, J., Zimmet, P. 2006. Prevalence and projections. In: Gan D (ed.). Diabetes Atlas International Diabetes Federation, 3rd edition. *International Diabetes Federation* 17–71.
- Sierra-Honigmann, M.R., Nath, A.K., Murakami, C., García-Cardeña, G., Papapetropoulos, A., Sessa, W.C., Madge, L.A., Schechner, J.S., Schwabb, M.B., Polverini, P.J., and Flores-Riveros, J.R. 1998. Biological action of leptin as an angiogenic factor. *Science* 281 (5383): 1683– 1686.
- Singh, A. 2014. Dipeptidyl peptidase-4 inhibitors: novel mechanism of actions. Indian Journal of Endocrinology and Metabolism 18 (6): 753-759.
- Singh, V.P., Bali, A., Singh, N. and Jaggi, A.S. 2014. Advanced Glycation end products and diabetic complications. *The Korean Journal of Physiology* & *Pharmacology* 18 (1): 1-14. https://doi.org/10.4196/kjpp.2014.18.1.1
- Singleton, V.L. and Rossi, J.A. 1965. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American Journal of Enology and Viticulture*. 16: 144-158.
- Siti Fatimah Zahra, M.A., Mahmood, A.A., Hapipah, M.A., Suzita, M.N. and Salmah, I. 2009. Anti-ulcerogenic Activity of Aqueous Extract of Ficus deltoidea Against Ethanol-induced Gastric Mucosal Injury in Rats. *Research Journal of Medical Sciences* 3: 42-46.
- Skyler, J.S., Bakris, G.L., Bonifacio, E., Darsow, T., Eckel, R.H., Groop, L., Groop, P.H., Handelsman, Y., Insel, R.A., Mathieu, C., McElvaine, A.T., Palmer, J.P., Pugliese, A., Schatz, D.A., Sosenko, J.M., Wilding, J.P., and Ratner, R.E. 2017. Differentiation of Diabetes by Pathophysiology, Natural History, and Prognosis. *Diabetes* 66 (2): 241–255.
- Smith, P.R., and Thornalley, P.J. 1992. Mechanism of the degradation of nonenzymatically glycated proteins under physiological conditions. Studies with the model fructosamine, N epsilon-(1-deoxy-D-fructos-1-yl) hippuryl-lysine. *European Journal of Biochemistry* 210 (3): 729–739.
- Snijder, M.B., Heine, R.J., Seidell, J.C., Bouter, L.M., Stehouwer, C.D., Nijpels, G., Funahashi, T., Matsuzawa, Y., Shimomura, I. and Dekker, J.M. 2006. Associations of adiponectin levels with incident impaired glucose metabolism and type 2 diabetes in older men and women: *The Hoorn study. Diabetes Care* 29: 2498-2503

- Sola, D., Rossi, L., Schianca, G. P., Maffioli, P., Bigliocca, M., Mella, R., Corlianò, F., Fra, G. P., Bartoli, E., & Derosa, G. 2015. Sulfonylureas and their use in clinical practice. Archives of medical science: AMS, 11(4), 840–848.
- Sompong, W., Meeprom, A., Cheng, H., and Adisakwattana, S. 2013. A comparative study of ferulic acid on different monosaccharide-mediated protein glycation and oxidative damage in bovine serum albumin. *Molecules* 18 (11): 13886–13903.
- Spiteller, G. 2006. Peroxyl radicals: inductors of neurodegenerative and other inflammatory diseases. Their origin and how they transform cholesterol, phospholipids, plasmalogens, polyunsaturated fatty acids, sugars, and proteins into deleterious products. *Free Radical Biology & Medicine*, 41 (3): 362–387.
- Spranger, J., Kroke, A., Möhlig, M., Bergmann, M.M., Ristow, M., Boeing, H., and Pfeiffer, A.F. 2003. Adiponectin and protection against type 2 diabetes mellitus. *Lancet* 361 (9353): 226–228.
- Sreenan, S.K., Zhou, Y.P., Otani, K., Hansen, P.A., Currie, K.P., Pan, C.Y., Lee, J.P., Ostrega, D.M., Pugh, W., Horikawa, Y., Cox, N.J., Hanis, C.L., Burant, C.F., Fox, A.P., Bell, G.I., and Polonsky, K.S. 2001. Calpains play a role in insulin secretion and action. *Diabetes* 50 (9): 2013–2020.
- Starr, F., Starr, K., and Loope, L. 2003. *Ficus deltoidea.* Plants of Hawai'i reports. United States Geological Survey - Biological Resources Division, Haleakala Field Station, Maui, Hawai'i.
- Stein, S.A., Lamos, E.M. and Davis, S.N. 2013. A review of the efficacy and safety of oral antidiabetic drugs. *Expert Opinion on Drug Safety* 12 (2): 153-175.
- Stipanuk, M.H. and Caudill, M.A. 2019. Biochemical, Physiological, and Molecular Aspects of Human Nutrition Fourth Edition. Chapter 12: Metabolism of Carbohydrate. *Elsevier Inc.* ISBN978-0-323-44181-0.
- Størling, J., and Pociot, F. 2017. Type 1 Diabetes Candidate Genes Linked to Pancreatic Islet Cell Inflammation and Beta-Cell Apoptosis. *Genes* 8 (2): 72. https://doi.org/10.3390/genes8020072.
- Suárez, G., Rajaram, R., Oronsky, A.L., and Gawinowicz, M.A. 1989. Nonenzymatic glycation of bovine serum albumin by fructose (fructation). Comparison with the Maillard reaction initiated by glucose. *The Journal of Biological Chemistry*. 264 (7): 3674–3679.
- Seino, S., Takahashi, H., Takahashi, T. and Shibasaki, T. 2012. Treating diabetes today: a matter of selectivity of sulphonylureas. *Diabetes, Obesity and Metabolism*, 14: 9-13.
- Sulaiman, M.R., Hussain, M.K., Zakaria, Z.A., Somchit, M.N., Moin, S., Mohamad, A.S., and Israf, D.A. 2008. Evaluation of the antinociceptive

activity of Ficus deltoidea aqueous extract. *Fitoterapia* 79 (7-8): 557–561.

- Suryati, S., Nurdin, H., Dachriyanus, D. and Lajis, M.N. 2011. Structure Elucidation Of Antibacterial Compound From *Ficus Deltoidea* Jack Leaves. *Indonesian Journal of Chemistry* 11: 67–70.
- Syed Aris, S.R., Mustafa, S., Ahmat, N., Mohd Jaafar, F. and Ahmad, R. 2009. Phenolic content and antioxidant activity of fruits of *Ficus deltoidea* var angustifolia sp. *The Malaysian Journal of Analytical Sciences* 13(2): 146 – 150.
- Szeto, V., Chen, N., Sun, H. and Feng, Z. 2018. The role of KATP channels in cerebral ischemic stroke and diabetes. *Acta Pharmacologica Sinica* 39: 683–694.
- Taylor M. Triolo, Alexandra Fouts, Laura Pyle, Liping Yu, Peter A. Gottlieb, Andrea K. Steck, the Type 1 Diabetes TrialNet Study Group. 2019. Identical and Nonidentical Twins: Risk and Factors Involved in Development of Islet Autoimmunity and Type 1 Diabetes. *Diabetes Care Feb*, 42 (2) 192-199
- Taniguchi, C.M., Emanuelli, B., Kahn, C.R. 2006. Critical nodes in signalling pathways: insights into insulin action. *Nature Reviews. Molecular Cell Biology*. 7 (2): 85-96.
- Toledo, L.M., Lydon, N.B., and Elbaum, D. 1999. The structure-based design of ATP-site directed protein kinase inhibitors. *Current Medicinal Chemistry* 6 (9): 775–805.
- Tontonoz, P., and Spiegelman, B.M. 2008. Fat and beyond: the diverse biology of PPAR-gamma. *Annual Review of Biochemistry* 77: 289–312.
- Tremblay, F., Lavigne, C., Jacques, H. and Marette, A. 2001. Defective Insulin-Induced GLUT4 Translocation in Skeletal Muscle of High Fat–Fed Rats Is Associated with Alterations in Both Akt/Protein Kinase B and Atypical Protein Kinase C ( $\zeta/\lambda$ ) Activities. *Diabetes Aug*, 50 (8) 1901-1910.
- Tsakiridis, T., McDowell, H.E., Walker, T., Downes, C.P., Hundal, H.S., Vranic, M., and Klip, A. 1995. Multiple roles of phosphatidylinositol 3-kinase in regulation of glucose transport, amino acid transport, and glucose transporters in L6 skeletal muscle cells. *Endocrinology* 136 (10): 4315– 4322
- Tseng, M.Y., Huang, Y.F., Liang, J, Wang, J.S. Yang, C.T., Wug, C.C., Cheng, H.S., Cheni, C.H., Lin, Y.E., Wang, W.S. and Shyu, Y.I.L. 2019. Diabetic neuropathies influence recovery from hip-fracture surgery in older persons with diabetes. *Experimental Gerontology* 119:168–173.
- Tsukaya H. 2005. Leaf shape: genetic controls and environmental factors. *The International Journal of Developmental Biology* 49 (5-6): 547–555.

- Turban, S., Stretton, C., Drouin, O., Green, C. J., Watson, M. L., Gray, A., Ross, F., Lantier, L., Viollet, B., Hardie, D. G., Marette, A., and Hundal, H. S. 2012. Defining the contribution of AMP-activated protein kinase (AMPK) and protein kinase C (PKC) in regulation of glucose uptake by metformin in skeletal muscle cells. *The Journal of Biological Chemistry*, 287(24), 20088–20099.
- Tyagi, S., Gupta, P., Saini, A.S., Kaushal, C., and Sharma, S. 2011. The peroxisome proliferator-activated receptor: A family of nuclear receptors role in various diseases. *Journal of Advanced Pharmaceutical Technology & Research* 2 (4): 236–240.
- Unoki, H., Takahashi, A., Kawaguchi, T., Hara, K., Horikoshi, M., Andersen, G., Ng, D.P., Holmkvist, J., Borch-Johnsen, K., Jørgensen, T., Sandbaek, A., Lauritzen, T., Hansen, T., Nurbaya, S., Tsunoda, T., Kubo, M., Babazono, T., Hirose, H., Hayashi, M., Iwamoto, Y., Kashiwagi, A., Kaku, K., Kawamori, R., Tai, S., Pederson, O., Kamatani, N., Kadowaki, T. Kikkawa, R., Nakamura, Y. and Maeda, S. 2008. SNPs in KCNQ1 are associated with susceptibility to type 2 diabetes in East Asian and European populations. *Nature Genetics* 40 (9): 1098–1102.
- USDA ARS, National genetic resources program, Germplasm resources information network–(GRIN) database (National germplasm resources laboratory, Beltsville, Maryland) https://npgsweb.arsgrin.gov/gringlobal/taxonomydetail.aspx?16826 (accessed on Apr 20, 2020).
- van Dam, E.M., Govers, R. and James, D.E. 2005. Akt activation is required at a late stage of insulin-induced GLUT4 translocation to the plasma membrane. *Molecular Endocrinology* 19 (4): 1067-1077.
- van der Zijl, N.J., Goossens, G.H., Moors, C.C., van Raalte, D.H., Muskiet, M.H., Pouwels, P.J., Blaak, E.E., and Diamant, M. 2011. Ectopic fat storage in the pancreas, liver, and abdominal fat depots: impact on β-cell function in individuals with impaired glucose metabolism. *The Journal of Clinical Endocrinology and Metabolism* 96 (2): 459–467.
- Varadi, A., Grant, A., McCormack, M., Nicolson, T., Magistri, M., Mitchell, K.J., Halestrap, A.P., Yuan, H., Schwappach, B., and Rutter, G.A. 2006. Intracellular ATP-sensitive K+ channels in mouse pancreatic beta cells: against a role in organelle cation homeostasis. *Diabetologia* 49 (7): 1567–1577.
- Vasanth Rao, V.R., Tan, S.H., Candasamy, M., and Bhattamisra, S.K. 2019. Diabetic nephropathy: An update on pathogenesis and drug development. *Diabetes & Metabolic Syndrome* 13 (1): 754–762.
- Vieira, R., Souto, S.B., Sánchez-López, E., Machado, A.L., Severino, P., Jose, S., Santini, A., Fortuna, A., García, M.L., Silva, A.M., and Souto, E.B. 2019. Sugar-Lowering Drugs for Type 2 Diabetes Mellitus and Metabolic

Syndrome-Review of Classical and New Compounds: Part-I. *Pharmaceuticals* 12 (4): 152. https://doi.org/10.3390/ph12040152.

- Vlavcheski, F., Naimi, M., Murphy, B., Hudlicky, T., and Tsiani, E. 2017. Rosmarinic Acid, a Rosemary Extract Polyphenol, Increases Skeletal Muscle Cell Glucose Uptake and Activates AMPK. *Molecules* 22 (10): 1669. https://doi.org/10.3390/molecules22101669.
- Wahid, A., Manek, N., Nichols, M., Kelly, P., Foster, C., Webster, P., Kaur, A., Friedemann Smith, C., Wilkins, E., Rayner, M., Roberts, N., and Scarborough, P. 2016. Quantifying the Association Between Physical Activity and Cardiovascular Disease and Diabetes: A Systematic Review and Meta-Analysis. *Journal of the American Heart Association* 5 (9): e002495. https://doi.org/10.1161/JAHA.115.002495.
- Wahli, W., and Michalik, L. 2012. PPARs at the crossroads of lipid signaling and inflammation. *Trends in Endocrinology and Metabolism* 23 (7): 351–363.
- Walker, E.H., Pacold, M.E., Perisic, O., Stephens, L., Hawkins, P.T., Wymann, M.P., and Williams, R.L. 2000. Structural determinants of phosphoinositide 3-kinase inhibition by wortmannin, LY294002, quercetin, myricetin, and staurosporine. *Molecular Cell* 6 (4): 909–919.
- Wang, Y., Liu, Q., Kang, S-G., Huang, K., Tong, T. 2021. Dietary Bioactive Ingredients Modulating the cAMP Signaling in Diabetes Treatment. *Nutrients*; 13(9):3038. https://doi.org/10.3390/nu13093038
- Wheeler, M.L., Dunbar, S.A., Jaacks, L.M., Karmally, W., Mayer-Davis, E.J., Wylie-Rosett, J., and Yancy, W.S., Jr. 2012. Macronutrients, food groups, and eating patterns in the management of diabetes: a systematic review of the literature, 2010. *Diabetes Care* 35 (2): 434– 445.
- Woon, S.M., Seng, Y.W., Ling, A.P., Chye, S.M, and Koh, R.Y. 2014. Antiadipogenic effects of extracts of *Ficus deltoidea* var. *deltoidea* and var. *angustifolia* on 3T3-L1 adipocytes. *Journal of Zhejiang University*. *Science. B* 15 (3): 295–302.
- World Health Organization (WHO). 2020. Diabetes. Accessed on 29 August 2021 at https://www.who.int/news-room/fact-sheets/detail/diabetes.

World Health Organization (WHO) 2018. Diabetes. https://www.who.int/newsroom/fact-sheets/detail/diabetes. Accessed on 14 September 2020.

- Wu, G., and Meininger, C.J. 2009. Nitric oxide and vascular insulin resistance. *BioFactors (Oxford, England)* 35 (1): 21–27.
- Wu, C.H., Huang, S.M., Lin, J.A., and Yen, G.C. 2011. Inhibition of advanced glycation endproduct formation by foodstuffs. *Food & Function* 2 (5): 224–234.

- Yamanouchi, T. 2010. Concomitant therapy with pioglitazone and insulin for the treatment of type 2 diabetes. *Vascular Health and Risk Management* 6: 189–197.
- Yang, X., Pratley, R.E., Baier, L.J., Horikawa, Y., Bell, G.I., Bogardus, C., and Permana, P.A. 2001. Reduced skeletal muscle calpain-10 transcript level is due to a cumulative decrease in major isoforms. *Molecular Genetics and Metabolism*. 73 (1): 111–113.
- Yang, H. and Li, X. 2012. The role of fatty acid metabolism and lipotoxicity in pancreatic beta-cell injury: identification of potential therapeutic targets. *Acta Pharmaceutica Sinica B* 2 (4): 396–402.
- Yang, W.C., Chang, C.L., Li, C.R., Nammi, S., and Cho, W.C. 2015. Complementary and alternative medicine for diabetes 2014. *Evidence-Based Complementary and Alternative Medicine*: eCAM, 685248. https://doi.org/10.1155/2015/685248.
- Yap, A., Nishiumi, S., Yoshida, K., and Ashida, H. 2007. Rat L6 myotubes as an in vitro model system to study GLUT4-dependent glucose uptake stimulated by inositol derivatives. *Cytotechnology*, 55(2-3), 103–108.
- Yen, W., Chang, L., and Duh, P. 2005. Antioxidant Activity of Peanut Seed Testa and Its Antioxidative Component, Ethyl Protocatechuate. LWT-Food Science and Technology. 38 (3): 193-200.
- Yonamine, C.Y., Pinheiro-Machado, E., Michalani, M.L., Alves-Wagner, A.B., Esteves, J.V., Freitas, H.S., and Machado, U.F. 2017. Resveratrol Improves Glycemic Control in Type 2 Diabetic Obese Mice by Regulating Glucose Transporter Expression in Skeletal Muscle and Liver. *Molecules* 22 (7): 1180. https://doi.org/10.3390/molecules22071180
- Young, I.S., and Woodside, J.V. 2001. Antioxidants in health and disease. *Journal of Clinical Pathology* 54 (3), 176–186.
- Yunusa, A.K., Rashid, Z.M., Mat, N., Bakar, C.A.A., and Ali, A.M. 2018. Chemicals and Bioactivity Discrimination of Syconia of Seven Varieties of *Ficus deltoidea* Jack Via ATR-IR Spectroscopic-Based Metabolomics. *Pharmacognosy Journal* 10 (Supplement 6): s147-s151.
- Zakaria, Z.A., Hussain, M.K., Mohamad, A.S., Abdullah, F.C., and Sulaiman, M.R. 2012. Anti-inflammatory activity of the aqueous extract of ficus deltoidea. *Biological Research for Nursing* 14 (1): 90–97.
- Zand, H., Morshedzadeh, N., and Naghashian, F. 2017. Signaling pathways linking inflammation to insulin resistance. *Diabetes & Metabolic Syndrome* 11 (Supplement 1): S307–S309.