



UNIVERSITI PUTRA MALAYSIA

**DEVELOPMENT OF GRAFTED SALT-TOLERANT ROCKMELON
(*Cucumis melo L.*) USING VARIOUS SALINITY SOURCES TO
IMPROVE PLANT PERFORMANCE**

MUHAMAD HAFIZ BIN MUHAMAD HASSAN

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MUHAMAD HAFIZ BIN MUHAMAD HASSAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillments of the Requirements for the Degree of
Master of Science**

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Abstracts of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**DEVELOPMENT OF GRAFTED SALT-TOLERANT ROCKMELON
(*Cucumis melo* L.) USING VARIOUS SALINITY SOURCES TO
IMPROVE PLANT PERFORMANCE**

By

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November 2021

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Nowadays, there is an increase in demand for high fruit quality of rockmelon for local and export market. To improve fruit quality, salt addition in nutrient solution is a viable approach that can be implemented. However, excess and continuous supply deleteriously affects the rockmelon's growth and yield. One of the potential strategies to utilize varying sources of salinity for fruit quality improvement without causing growth and yield reduction is by growing grafted rockmelon using salt-tolerant *Cucurbitaceae* rootstock. Therefore, the objectives of this research are to determine a suitable rockmelon scion age to be grafted on cucurbit rootstocks, to identify the salinity tolerance levels of grafted rockmelon and to evaluate the growth, physiological process, yield and fruit quality of salt-tolerant grafted rockmelon under varying salinity sources. Three scion ages (8, 13 and 18 days after sowing; DAS) of rockmelon (cv. Glamour) were grafted onto two types of *Cucurbitaceae* viz. bottle gourd (*Lagenaria siceraria*) and bitter gourd (*Momordica charantia*). These factorial combinations were arranged in a Randomized Complete Block Design (RCBD) with three replications. Rockmelon scion ages at 13 and 18 DAS were found to be suitable for both cucurbit rootstocks, as they exhibited higher scion height, leaf number, leaf and stem dry weight compared to 8 DAS. Thus, 13 DAS scion age of rockmelon which served as the fastest scion produced was selected to be used for grafting in the subsequent study. The following experiment involved grafting of 13 DAS rockmelon scion age onto three cucurbit rootstocks (self-grafted, bottle gourd, bitter gourd) and subjected to four levels of NaCl salinity (0, 25, 50, 75 mM). These factorial combinations were arranged in a RCBD with three replications. Among graft combinations, rockmelon/ bottle gourd was classified as the least salt-sensitive due to its capability to reduce

toxic ions (Na^+ and Cl^-) accumulation in the leaf, acquired moderate plant water status among others while having the least significant relationships in growth reductions under saline conditions. Furthermore, NaCl levels at 75 mM were found to be detrimental on most of the growth parameters, leaf gas exchanges and relative water content, which further causes ionic stress that negatively affected nutrient availability in rockmelon. Therefore, graft combination of rockmelon/ bottle gourd and salinity level at 50 mM were selected to be tested in the last experiment. In the last experiment, rockmelon/bottle gourd was subjected to four types of salinity sources (Basic nutrient solution; BNS, NaCl (50 mM) + BNS, KNO_3 (50 mM) + BNS, high strength nutrient solution; NS). These treatments were arranged in a RCBD with four replications. It is revealed that, salinity induced by NaCl (50 mM) salt and high strength NS is capable to increase fruit quality components without interfering the physiological process. When treated with KNO_3 (50 mM) salt, most of the growth and fruit quality parameters were sustained. However, the physiological process such as photosynthetic pigments was significantly impaired. Yield component such as fruit weight were decreased regardless of salinity sources by NaCl (28.8%), KNO_3 (28.26%) and high strength NS (27.72%) applications as compared to BNS treatment. Overall, 13 DAS scion age is recommended to be grafted with bottle gourd as a salt-tolerant rootstock for rockmelon. Taking into account the cost saving factor, incorporation of NaCl (50 mM) in nutrient solution is a feasible approach on rockmelon/ bottle gourd due to high fruit quality results whilst sustaining all the physiological process.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

**PENGHASILAN CANTUMAN ROCKMELON (*Cucumis melo L.*) YANG
TAHAN KEMASINAN BAGI PENGGUNAAN KEMASINAN YANG
PELBAGAI UNTUK MENINGKATKAN PRESTASI TANAMAN**

Oleh

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Pada masa ini, terdapatnya peningkatan dalam permintaan terhadap rockmelon yang berkualiti tinggi bagi tujuan pasaran tempatan dan eksport. Untuk meningkatkan kualiti buah, penambahan garam ke dalam larutan nutrien merupakan pendekatan yang relevan untuk diperlakukan. Bagaimanapun, pemberian yang berlebihan dan berterusan mengakibatkan kesan yang buruk terhadap pertumbuhan dan hasil rockmelon. Salah satu strategi yang berpotensi untuk menggunakan sumber kemasinan bagi peningkatan kualiti buah tanpa menyebabkan penurunan pertumbuhan dan hasil adalah melalui penanaman rockmelon yang dicantum menggunakan penanti *Cucurbitaceae* yang toleran kepada kemasinan. Maka itu, objektif kajian ini dijalankan adalah untuk menentukan umur scion rockmelon yang sesuai untuk dicantumkan ke atas penanti cucurbit, untuk mengenalpasti tahap ketahanan cantuman rockmelon terhadap kemasinan dan untuk menilai pertumbuhan, proses fisiologi, hasil dan kualiti buah cantuman rockmelon yang toleran terhadap kemasinan pada sumber kemasinan yang pelbagai. Tiga umur scion (8, 13 dan 18 hari lepas penyemaian; DAS) rockmelon (varieti Glamour) dicantumkan ke atas dua jenis *Cucurbitaceae* iaitu labu botol (*Lagenaria siceraria*) dan peria katak (*Momordica charantia*). Gabungan faktorial ini disusun di dalam rekabentuk rawak blok lengkap (RCBD) dengan tiga replikasi. Umur scion rockmelon pada 13 dan 18 DAS dilihat bersesuaian untuk kedua-dua penanti cucurbit, di mana ianya memperkenalkan tinggi scion, bilangan daun, berat kering daun dan batang yang lebih tinggi dibandingkan kepada 8 DAS. Maka, 13 DAS umur scion rockmelon yang mana berupaya menjadi scion yang terpantas untuk dihasilkan adalah dipilih untuk digunakan bagi cantuman pada kajian yang berikutnya. Eksperimen yang seterusnya melibatkan cantuman scion rockmelon yang berusia 13 DAS yang dilakukan ke atas tiga penanti cucurbit

(rockmelon, labu botol, peria katak) dan dirawat menggunakan empat tahap kemasinan NaCl (0, 25, 50, 75 mM). Gabungan faktorial ini disusun di dalam RCBD dengan tiga replikasi. Antara kombinasi cantuman, rockmelon/ labu botol diklasifikasikan sebagai paling kurang sensitif kepada kemasinan disebabkan oleh kemampuannya untuk mengurangkan pengumpulan ion-ion toksik (Na^+ and Cl^-) di daun, beroleh status air pokok yang sederhana antara semua, di samping memiliki pertalian ketara yang paling kurang dalam pengurangan pertumbuhan di bawah keadaan yang masin. Selain itu, tahap NaCl pada 75 mM dilihat menjelaskan pada hampir kesemua parameter pertumbuhan, pertukaran gas daun dan kandungan air relatif, di mana seterusnya menyebabkan tegasan ion yang secara negatifnya mempengaruhi kedapatan nutrien di dalam rockmelon. Maka itu, kombinasi cantuman rockmelon/ labu botol dan kadar kemasinan pada 50 mM adalah dipilih untuk diuji pada eksperimen yang terakhir. Di dalam eksperimen terakhir ini, rockmelon/ labu botol dirawat menggunakan empat jenis sumber kemasinan (Larutan nutrien asas; BNS, NaCl (50 mM) + BNS, KNO_3 (50 mM) + BNS, larutan nutrien asas berkepekatan tinggi). Kajian membuktikan bahawa, kemasinan yang dirawat menggunakan garam NaCl (50 mM) dan larutan nutrien asas berkepekatan tinggi berupaya untuk meningkatkan komponen kualiti buah tanpa menjelaskan proses fisiologi. Apabila diaruh menggunakan garam KNO_3 (50 mM), hampir keseluruhan parameter pertumbuhan dan kualiti buah adalah tidak terjejas. Walaubagaimanapun, proses fisiologi seperti pigmen fotosintesis adalah terjejas dengan ketara. Komponen hasil seperti berat buah adalah berkurangan tanpa mengira sumber kemasinan pada aplikasi NaCl (28.8%), KNO_3 (28.26%) dan larutan nutrien asas berkepekatan tinggi (27.72%) apabila dibandingkan dengan rawatan larutan nutrien asas. Secara keseluruhannya, 13 DAS umur scion adalah dicadangkan untuk dicantumkan dengan labu botol sebagai penanti yang tahan kemasinan kepada rockmelon. Mengambil kira faktor kos, penggunaan NaCl (50 mM) di dalam larutan nutrien adalah pendekatan yang terbaik pada rockmelon/ labu botol disebabkan penghasilan kualiti buah yang tinggi di samping tidak menjelaskan keseluruhan proses fisiologinya.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

mM	Milimolar
mmol/L	Milimolar per litre
MPa	Megapascal
RWC	Relative water content
μmol	Micromole
nmol	Nanomole
DW	Dry weight
FW	Fresh weight
TW	Turgid weight
EC	Electrical conductivity
MDA	Malondialdehyde
ROS	Reactive oxygen species
dS/m	Decisiemens per metre
TW	Turgid weight
K/Na	Potassium to sodium ratio
N	Newton
$^{\circ}\text{C}$	Degree celsius

CHAPTER 1

INTRODUCTION

Rockmelon which is also known as muskmelon (*Cucumis melo* L.) is a short-term horticultural crop that belongs to the *Cucurbitaceae* family. It is contained with source of antioxidants such as carotenes and flavonoids and being enriched with vitamin B, vitamin C, folate and pantothenic acid. It is also high in fiber, low in fat and cholesterol free, as well as source of many essential nutrients including potassium and magnesium (Lester, 1997). In Malaysia, rockmelon is commercially grown to fulfill the demand for local and export markets in the form of fresh fruit and processed products. Over the years, rockmelon productions have increased drastically to 45.56% since 2012-2018 with total productions recorded at 5,845.71 mt in 2018 (Department of Agriculture, 2018). The increase in production areas is due to high demand from consumers for a high fruit quality of rockmelon. According to Lester (2006), fruit quality characteristics including sweetness, taste, texture and flavour are the most important reason that highly preferred by the consumers in melon productions.

In order to increase fruit quality, salinity application by supplementation of salt into nutrient solution is a feasible approach and fairly easy to be adapted. Accumulation of salt may increase the root zone solute which further caused low soil water potential. Thus, the plant had reduced their water absorption capacity that increased dry-matter components. Increased in dry matter components ultimately enhanced fruit quality attributes including total soluble solid, total titratable acidity and sugar acid ratio. This is accordance with Costa *et al.*, (2013) that the fruit quality enhancement was accompanied by the increase of dry-matter concentration which is associated with water uptake reduction. It also has been broadly reported that, salinity could enhance fruit quality attributes and this has been demonstrated for a number of crops, such as tomato and watermelon (Machado and Serralheiro, 2017). Unfortunately, there is no current research regarding this approach has been conducted on rockmelon to this point in time.

While, excess and continuous supply of salt could lead to salinity development and deleteriously affect the rockmelon's growth, physiological process and yield. Reduction of growth and physiological process under saline environment simultaneously resulted to lower yield productions. Lower fruit yield in salinized rockmelon is not marketable as the fruit is perceived to be of a lower grade. Since, rockmelon has been classified as moderately sensitive to salt stress among cucurbit species (Pessarakli, 2016), supplementation using different sources of salinity can be done to improve fruit quality without causing growth and yield reduction (Zhang *et al.*, 2016).

Therefore, rockmelon variety that is tolerant to salinity is needed for enabling the use of various sources of salinity. Developing such salt tolerance variety through the selection and conventional breeding program is time-consuming with low commercial success due to the complexity of the traits, as salt resistance characteristics contain dynamic genetic and physiological interactions (Flowers, 2004).

Hence, grafting among potential salt-tolerant cucurbit rootstocks has a higher potential to increase the salt tolerance level in rockmelon. By using salt-tolerant grafted rockmelon from grafting procedure established, the salinity tolerance level could be enhanced and the various sources of salinity can be utilized for the growth and yield improvement without interfering in the physiological process. This research may contribute to the future development of growing rockmelon with salt addition, which will improve fruit quality without causing growth and yield reduction. Knowledge produced in this study may be useful in improving rockmelon development practices in Malaysia as well as exploiting new research pathways for rockmelon in the future. Considering these factors, this research is thus necessary to be done with the objectives:

1. to determine the suitable age of rockmelon scion to be grafted on potential salt-tolerant cucurbit rootstocks
2. to identify the salinity tolerance levels of rockmelon grafted on potential salt-tolerant cucurbit rootstocks
3. to evaluate the growth, physiological process, yield, and fruit quality of rockmelon grafted on salt-tolerant rootstock under varying salinity sources.

REFERENCES

- Abdelgawad, F. K., El-Mogy, M., Mohamed, M., Garchery, C., and Stevens, R. (2019). Increasing ascorbic acid content and salinity tolerance of cherry tomato plants by suppressed expression of the ascorbate oxidase gene. *Agronomy*, 9(2): 51.
- Adabnejad, H., Kavousi, H. R., Hamidi, H., and Tavassolian, I. (2015). Assessment of the vacuolar Na^+/H^+ antiporter (NHX1) transcriptional changes in *Leptochloa fusca L.* in response to salt and cadmium stresses. *Molecular Biology Research Communications*, 4(3): 133.
- Agarwal, S., and Shaheen, R. (2007). Stimulation of antioxidant system and lipid peroxidation by abiotic stresses in leaves of *Momordica charantia*. *Brazilian Journal of Plant Physiology*, 19: 149-161.
- Ahmad, H., Hayat, S., Ali, M., Ghani, M. I., and Zhihui, C. (2017). Regulation of growth and physiological traits of cucumber (*Cucumis sativus L.*) through various levels of 28-homobrassinolide under salt stress conditions. *Canadian Journal of Plant Science*, 98(1): 132-140.
- Akrami, M., and Arzani, A. (2018). Physiological alterations due to field salinity stress in melon (*Cucumis melo L.*). *Acta Physiologiae Plantarum*, 40(5): 1-14.
- Albacete, A., Martinez-Andujar, Ghanem, M. E., Acosta, M., and Perez-Alfocea, F. (2009). Rootstock-mediated changes in xylem ionic and hormonal status are correlated with delayed leaf senescence, and increased leaf area and crop productivity in salinized tomato. *Plant, Cell and Environment*, 32(7): 928-938.
- Al-Hamzawi, M. K. A. (2010). Growth and storability of plastic houses cucumber (*Cucumis sativus L.* cv. Al-Hytham). *American Journal of Plant Physiology*, 5(5): 278-290.
- Al-Harbi, A. R., Wahb-Allah, M. A., and Al-Omran, A. M. (2008). Effects of salinity and irrigation management on growth and yield of tomato grown under greenhouse conditions. In *International Symposium on Strategies Towards Sustainability of Protected Cultivation in Mild Winter Climate*, 807: 201-206.
- Ali Q, Ashraf M, Athar H. (2007). Exogenously applied proline at different growth stages enhances growth of two maize cultivars grown under water deficit conditions. *Pak. J. Bot.*, 39: 1133–1144.

- Al-Ismaily, S. S., Al-Yahyai, R. A., and Al-Rawahy, S. A. (2014). Mixed fertilizer can improve fruit yield and quality of field-grown tomatoes irrigated with saline water. *Journal of Plant Nutrition*, 37(12): 1981-1996.
- Aloni, B., Karni, L., Deventurero, G., Levin, Z., Cohen, R., and Kapulnik, Y. (2008). Physiological and biochemical changes at the rootstock-scion interface in graft combinations between *Cucurbita* rootstocks and a melon scion. *The Journal of Horticultural Science and Biotechnology*, 83(6): 777-783.
- Al-Saady, N. A., Khan, A. J., Rajesh, L., and Esechie, H. A. (2012). Effect of salt stress on germination, proline metabolism and chlorophyll content of fenugreek (*Trigonella gracium* L.). *Journal of plant Sciences*, 7(5): 176.
- Apse, M. P., Aharon, G. S., Snedden, W. A., and Blumwald, E. (1999). Salt tolerance conferred by overexpression of a vacuolar Na^+/H^+ antiport in *Arabidopsis*. *Science*, 285(5431): 1256-1258.
- Arefin, S. M., Zeba, N., Solaiman, A. H., Naznin, M. T., Azad, M. O. K., Tabassum, M., and Park, C. H. (2019). Evaluation of compatibility, growth characteristics, and yield of tomato grafted on potato ('Pomato'). *Acta Horticulturae*, 5(2): 37.
- Arzani, A. (2008). Improving salinity tolerance in crop plants: a biotechnological view. *In Vitro Cellular and Developmental Biology-Plant*, 44(5): 373-383.
- Ashraf, M. Y., and Bhatti, A. S. (2000). Effect of salinity on growth and chlorophyll content in rice. *Biological Sciences-PJSIR*, 43(2): 130-131.
- Awang, Y. B., Atherton, J. G., and Taylor, A. J. (1993). Salinity effects on strawberry plants grown in rockwool. II. Fruit quality. *Journal of Horticultural Science*, 68(5): 791-795.
- Ayres, R. S., and Westcot, D. W. (1985). Water Quality for Agriculture. *Journal of Irrigation and Drainage*, 29: 103-107.
- Azarmi, R., Taleshmikail, R. D., and Gikloo, A. (2010). Effects of salinity on morphological and physiological changes and yield of tomato in hydroponics system. *Journal of Food, Agriculture and Environment*, 8(2): 573-576.
- Bahrami, M., Heidari, M., and Ghorbani, H. (2016). Variation in antioxidant enzyme activities, growth and some physiological parameters of bitter melon (*Momordica charantia*) under salinity and chromium stress. *Journal of Environmental Biology*, 37(4): 529.

- Baligar, V. C., Fageria, N. K., and He, Z. L. (2001). Nutrient use efficiency in plants. *Journal of Soil Science and Plant Analysis*, 32(7-8): 921-950.
- Balkaya, A. (2016). Effects of salt stress on vegetative growth parameters and ion accumulations in cucurbit rootstock genotypes. *Ekin Journal of Crop Breeding and Genetics*, 2(2): 11-24.
- Bates, L., Waldren, R. and Teare, I. (1973). Rapid Determination of Free Proline for Water-Stress Studies. *Plant and Soil*, 39(1): 205-207.
- Bayuelo-Jimenez, J. S., Debouck, D. G., and Lynch, J. P. (2003). Growth, gas exchange, water relations, and ion composition of Phaseolus species grown under saline conditions. *Field Crops Research*, 80(3): 207-222.
- Bielorai, H., S. Darsberg, Y. and Brum, M. (1988). The effect of saline irrigation water on Shamuti orange production. *Proc. Int. Citrus Congress.*, 6: 707-715.
- Botia, P., Carvajal, M., Cerdá, A., and Martínez, V. (1998). Response of eight *Cucumis melo* cultivars to salinity during germination and early vegetative growth. *Agronomie*, 18(8-9): 503-513.
- Carillo, P., Annunziata, M. G., Pontecorvo, G., Fuggi, A., and Woodrow, P. (2011). Salinity stress and salt tolerance. *Abiotic stress in plants mechanisms and adaptations*, 1: 21-38.
- Cavins, T., Whipker, B., Fonteno, W., Harden, B., and Gibson, J. (2000). Monitoring and managing pH and EC using the PourThru extraction method. *Horticulture Information Leaflet*, 590: 1-17.
- Chaves, M. M., Flexas, J., and Pinheiro, C. (2009). Photosynthesis under drought and salt stress: regulation mechanisms from whole plant to cell. *Annals of Botany*, 103(4): 551-560.
- Chen, B. M., Wang, Z. H., Li, S. X., Wang, G. X., Song, H. X., and Wang, X. N. (2004). Effects of nitrate supply on plant growth, nitrate accumulation, metabolic nitrate concentration and nitrate reductase activity in three leafy vegetables. *Plant Science*, 167(3): 635-643.
- Chen, B., Liu, E., Tian, Q., Yan, C., and Zhang, Y. (2014). Soil nitrogen dynamics and crop residues. A review. *Agronomy for Sustainable Development*, 34(2): 429-442.
- Chen, G. and Wang, R. (2008). Effects of salinity on growth and concentrations of sodium, potassium, and calcium in grafted cucumber seedlings. *Acta Horticulturae*, 771: 217-224.

- Chen, S., and Polle, A. (2010). Salinity tolerance of *Populus*. *Journal of Plant Biology*, 12(2): 317-333.
- Chipojola, F. M., Mwase, W. F., Kwapata, M. B., Njoloma, J. P., Bokosi, J. M., and Maliro, M. F. (2013). Effect of tree age, scion source and grafting period on the grafting success of cashew nut (*Anacardium occidentale L.*). *African Journal of Agricultural Research*, 8(46): 5785-5790.
- Chookhampaeng, S. (2011). The effect of salt stress on growth, chlorophyll content proline content and antioxidative enzymes of pepper (*Capsicum annuum L.*) seedling. *European Journal of Scientific Research*, 49(1): 103-109.
- Colla, G., Roupahel, Y., Leonardi, C., and Bie, Z., (2010). Role of grafting in vegetable crops grown under saline conditions. *Scientia Horticulturae*, 127(2): 147-155.
- Colla, G., Roupahel, Y., Cardarelli, M., and Rea, E. (2006). Effect of salinity on yield, fruit quality, leaf gas exchange, and mineral composition of grafted watermelon plants. *Journal of HortScience*, 41(3): 622-627.
- Colla, G., Roupahel, Y., Cardarelli, M., Svecova, E., Rea, E., and Lucini, L. (2013). Effects of saline stress on mineral composition, phenolic acids and flavonoids in leaves of artichoke and cardoon genotypes grown in floating system. *Journal of the Science of Food and Agriculture*, 93(5): 1119-1127.
- Colla, G., Roupahel, Y., Leonardi, C., and Bie, Z. (2010). Role of grafting in vegetable crops grown under saline conditions. *Scientia Horticulturae*, 127(2): 147-155.
- Costa, A. R., and Medeiros, J. F. D. (2018). Nitrogen, phosphorus and potassium accumulation in watermelon cultivars irrigated with saline water. *Engenharia Agricola*, 38: 343-350.
- Costa, L. D. F., Casartelli, M. R. D. O., & Wallner-Kersanach, M. (2013). Labile copper and zinc fractions under different salinity conditions in a shipyard area in the patos lagoon estuary, south of Brazil. *Química Nova*, 36: 1089-1095.
- Davis, A. R., Perkins-Veazie, P., Sakata, Y., Lopez-Galarza, S., Maroto, J. V., and Lee, J. M. (2008). Cucurbit grafting. *Critical Reviews in Plant Sciences*, 27(1): 50-74.
- Debouba, M., Gouia, H., and Ghorbel, M. H. (2006). NaCl effects growth, ions and water status of Tomato (*Lycopersicon esculentum*) seedlings. *Acta Botanica Gallica*, 153(3): 297-307.

- Del-Amor, F. M., Martinez, V., and Cerda, A. (1999). Salinity duration and concentration affect fruit yield and quality, and growth and mineral composition of melon plants grown in perlite. *HortScience*, 34(7): 1234-1237.
- Delauney, A. J., and Verma, D. P. S. (1990). A soybean gene encoding 1-pyrroline-5-carboxylate reductase was isolated by functional complementation in *Escherichia coli* and is found to be osmoregulated. *Molecular and General Genetics MGG*, 221(3): 299-305.
- Department of Agriculture, Malaysia (2018). Fruit Crop Statistics, 11-21pp.
- Dias, N. D. S., Palacio, V. S., Moura, K. D. F., and Sousa Neto, O. N. (2015). Growth of melon plants in coconut saline nutrient solution, *Journal of Irrigation*, 20(1): 1-12.
- Dias, N., Morais, P. L., Abrantes, J. D., Nogueira de Sousa Neto, O., Palacio, V. S., and Freitas, J. J. R. (2018). Nutrient solution salinity effect of greenhouse melon (*Cucumis melon* L. cv. Nectar). *Acta Agronomica*, 67(4): 517-524.
- Dobrota, C. (2006). Energy dependent plant stress acclimation. *Reviews in Environmental Science and Bio/Technology*, 5(2-3): 243-251.
- Duarte, B., Santos, D., Marques, J. C., and Casador, I. (2013). Ecophysiological adaptations of two halophytes to salt stress: photosynthesis, PS II photochemistry and anti-oxidant feedback-implications for resilience in climate change. *Plant Physiology and Biochemistry*, 67: 178-188.
- Duarte, H., and Souza, E. R. (2016). Soil water potentials and *Capsicum annuum* L. under salinity. *Revista Brasileira de Ciencia*, 40pp.
- Edelstein, M. (2004). Grafting vegetable-crop plants: pros and cons. In *VII International Symposium on Protected Cultivation in Mild Winter Climates: Production, Pest Management and Global Competition* 659: 235-238.
- Edelstein, M., Plaut, Z., and Ben-Hur, M. (2010). Water salinity and sodicity effects on soil structure and hydraulic properties – A review. *Adv. Hort. Sci.*, 24:154-160.
- EI-Eslamboly, A. A., and Abdel-Wahab, M. A. S. (2014). Grafting salinity tolerant rootstocks and magnetic iron treatments for cantaloupe production under conditions of high salinity soil and irrigation water. *Middle East J. Agric. Res.*, 3(3): 677-693.

- El-Mogy, M., Garchery, C., and Stevens, R. (2018). Irrigation with salt water affects growth, yield, fruit quality, storability and marker-gene expression in cherry tomato. *Acta Agriculturae Scandinavica, Section B-Soil and Plant Science*, 68(8): 727-737.
- Elsheery, N. I., and Cao, K. F. (2008). Gas exchange, chlorophyll fluorescence, and osmotic adjustment in two mango cultivars under drought stress. *Acta Physiologiae Plantarum*, 30(6): 769-777.
- El-Shraiy, A. M., Mostafa, M. A., Zaghloul, S. A., and Shehata, S. A. M. (2011). Physiological aspect of NaCl-salt stress tolerant among cucurbitaceous cultivars. *Australian Journal of Basic and Applied Sciences*, 5(11): 62-71.
- Estan, M. T., Martinez-Rodriguez, M. M., Perez-Alfocea, F., Flowers, T. J., and Bolarin, M. C. (2005). Grafting raises the salt tolerance of tomato through limiting the transport of sodium and chloride to the shoot. *Journal of Experimental Botany*, 56(412): 703-712.
- FAO and ITPS. (2015). Status of the World's Soil Resources (SWSR) – Main Report. Food and Agriculture Organization of the United Nations and Intergovernmental Technical Panel on Soils, Rome, Italy.
- Farhoudi, R. (2011). Effect of salt stress on physiological and morphological parameters of rapeseed cultivars. *Advances in Environmental Biology*, 2501-2509pp.
- Fisarakis, I., Nikolaou, N., Tsikalas, P., Therios, I., and Stavrakas, D. (2005). Effect of salinity and rootstock on concentration of potassium, calcium, magnesium, phosphorus, and nitrate in Thompson seedless grapevine. *Journal of Plant Nutrition*, 27(12): 2117-2134.
- Flowers, T.J. (2004). Improving crop salt tolerance. *Journal of Experimental Botany* 55(396): 307–319.
- Freitas L.D.A., Figueiredo V.B., Porto-Filho F.Q., Costa J.C.D., and Cunha E.M.D. (2014). Growth and production of melon grown under different levels of salinity and nitrogen. *Rev. Bras. Eng. Agric. Amb.*, 18: 20-26.
- Gabrijel, O., Davor, R., Zed, R., Marija, R., and Monika, Z. (2009). Cadmium accumulation by muskmelon under salt stress in contaminated organic soil. *Science of the Total Environment*, 407(7): 2175-2182.
- Garcia-Sanchez, F., Carvajal, M., Porras, I., Botia, P., and Martinez, V. (2003). Effects of salinity and rate of irrigation on yield, fruit quality and mineral composition of 'Fino 49' lemon. *European Journal of Agronomy*, 19(3): 427-437.

- Ghanem, M. E., van Elteren, J., Albacete, A., Quinet, M., Martinez-Andujar, C., Kinet, J. and Lutts, S. (2009). Impact of salinity on early reproductive physiology of tomato (*Solanum lycopersicum*) in relation to a heterogeneous distribution of toxic ions in flower organs. *Functional Plant Biology*, 36(2): 125-136.
- Ghani, M. N. O., Awang, Y., and Ismail, M. F. (2018). Effects of NaCl salinity on leaf water status, proline and mineral ion content of four *Cucurbitaceae* species. *Australian Journal of Crop Science*, 12(9): 1434-1439.
- Gloria, I. A., and Lilia, A. (2010). Salinity effects on protein content, lipid peroxidation, pigments and proline in *Paulownia imperialis* (Siebold and Zuccarini) and *Paulownia fortunei* (Seemann and Hemsley) grown in vitro. *Electronic Journal of Biotechnology*, 1051-1062pp.
- Goldschmidt, E. E. (2014). Plant grafting: new mechanisms, evolutionary implications. *Frontiers in Plant Science*, 5: 727.
- Gomes, M. A., Suzuki, M. S., Cunha, M. D., and Tullii, C. F. (2011). Effect of salt stress on nutrient concentration, photosynthetic pigments, proline and foliar morphology of *Salvinia auriculata*. *Acta Limnologica Brasiliensis*, 23: 164-176.
- Gong, B., Wen, D., Langenberg, K., Wei, M., Yang, F., Shi, Q., and Wang, X. (2013). Comparative effects of NaCl stress on photosynthetic parameters, nutrient metabolism, and the antioxidant system in tomato leaves. *Scientia Horticulturae*, 157: 1-12.
- Grattan, S. R., and Grieve, C. M. (1992). Mineral element acquisition and growth response of plants grown in saline environments. *Agriculture, ecosystems and environment*, 38(4): 275-300.
- Grattan, S. R., and Grieve, C. M. (1998). Salinity-mineral nutrient relations in horticultural crops. *Scientia Horticulturae*, 78(1-4): 127-157.
- Grisso R.B., Mark A.W.G., Holshouser D., and Thomason W. (2007). Precision Farming Tools: Soil electrical conductivity. *Virginia cooperative Extension*, 442-508pp.
- Haddad, M., Bani-Hani, N. M., Al-Tabbal, J. A., and Al-Fraihat, A. H. (2016). Effect of different potassium nitrate levels on yield and quality of potato tubers. *Journal of Food, Agriculture and Environment*, 14(1): 101-107.
- Hansen, E. H., and Munns, D. N. (1988). Effects of CaSO_4^{2-} and NaCl on growth and nitrogen fixation of *Leucaena leucocephala*. *Plant and soil*, 107(1): 95-99.

- Hartmann, H. T., Kester, D. E., Davies, F. T., and Geneve, R. L. (1997). *Plant propagation: principles and practices* (No. Ed. 6). Prentice-Hall Inc.
- He, Y., Zhu, Z., Yang, J., Ni, X., and Zhu, B. (2009). Grafting increases the salt tolerance of tomato by improvement of photosynthesis and enhancement of antioxidant enzymes activity. *Environmental and Experimental Botany*, 66(2): 270-278.
- Helaly, M. N., El-Hoseiny, H., El-Sheery, N. I., Rastogi, A., and Kalaji, H. M. (2017). Regulation and physiological role of silicon in alleviating drought stress of mango. *Plant physiology and biochemistry*, 118: 31-44.
- Hernandez, J. A., and Almansa, M. S. (2002). Short-term effects of salt stress on antioxidant systems and leaf water relations of pea leaves. *Physiologia Plantarum*, 115(2): 251-257.
- Heuvelink, E. P., Bakker, M., and Stanghellini, C. (2003). Salinity effects on fruit yield in vegetable crops: a simulation study. *International Symposium on Managing Greenhouse Crops in Saline Environment*, 609: 133-140.
- Hiscox, J. D., and Israelstam, G. F. (1979). A method for the extraction of chlorophyll from leaf tissue without maceration. *Canadian journal of botany*, 57(12): 1332-1334.
- Huang, Y., Tang, R., Cao, Q., and Bie, Z. (2009). Improving the fruit yield and quality of cucumber by grafting onto the salt tolerant rootstock under NaCl stress. *Scientia Horticulturae*, 122(1): 26-31.
- Jaarsma, R., de Vries, R. S., and de Boer, A. H. (2013). Effect of salt stress on growth, Na⁺ accumulation and proline metabolism in potato (*Solanum tuberosum*) cultivars. *PloS one*, 8(3): 60183.
- Jamil, A., Riaz, S., Ashraf, M., and Foolad, M. R. (2011). Gene expression profiling of plants under salt stress. *Critical Reviews in Plant Sciences*, 30(5): 435-458.
- Jawandha, S. K., Gill, P. P. S., Harminder, S., and Thakur, A. (2017). Effect of potassium nitrate on fruit yield, quality and leaf nutrients content of plum. *Vegetos*, 30(2): 325-328.
- Kader, F., Nicolas, J. P., and Metche, M. (1999). Degradation of pelargonidin 3-glucoside in the presence of chlorogenic acid and blueberry polyphenol oxidase. *Journal of the Science of Food and Agriculture*, 79(4): 517-522.

- Kawaguchi, M., Taji, A., Backhouse, D., and Oda, M. (2008). Anatomy and physiology of graft incompatibility in solanaceous plants. *Journal of Horticultural Science and Biotechnology*, 83(5): 581-588.
- Kaya, C., Tuna, A. L., Ashraf, M., and Altunlu, H. (2007). Improved salt tolerance of melon (*Cucumis melo L.*) by the addition of proline and potassium nitrate. *Environmental and Experimental Botany*, 60(3): 397-403.
- Khoshbakht, D., Ghorbani, A., Baninasab, B., Naseri, L. A., and Mirzaei, M. (2014). Effects of supplementary potassium nitrate on growth and gas-exchange characteristics of salt-stressed citrus seedlings. *Photosynthetica*, 52(4): 589-596.
- Kusvuran, S. (2012). Effects of drought and salt stresses on growth, stomatal conductance, leaf water and osmotic potentials of melon genotypes (*Cucumis melo L.*). *African Journal of Agricultural Research*, 7(5): 775-781.
- Kusvuran, S. (2021). Microalgae (*Chlorella vulgaris*) alleviates drought stress of broccoli plants by improving nutrient uptake, secondary metabolites, and antioxidative defense system. *Journal of Horticultural Plant*, 7(3): 221-231.
- Taylor L. (2002). Technical Data Report for Bitter melon (*Momordica charantia*): Herbal Secrets of the Rainforest, Sage Press Inc.
- Lee, J. M., and Oda, M. (2003). *Grafting of herbaceous vegetable and ornamental crops*. *Horticultural Reviews*, 28: 61-124.
- Lee, J. M., Kubota, C., Tsao, S. J., Bie, Z., Echevarria, P. H., Morra, L., and Oda, M. (2010). Current status of vegetable grafting: Diffusion, grafting techniques, automation. *Scientia Horticulturae*, 127(2): 93-105.
- Lester, G. (1997). Melon (*Cucumis melo L.*) fruit nutritional quality and health functionality. *Hort Technology*, 7(3), 222-227.
- Lester, G. (2006). Consumer preference quality attributes of melon fruits. In *IV International Conference on Managing Quality in Chains-The Integrated View on Fruits and Vegetables Quality*, 712: 175-182.
- Liu, T., Dai, W., Sun, F., Yang, X., Xiong, A., and Hou, X. (2014). Cloning and characterization of the nitrate transporter gene BraNRT2 in non-heading Chinese cabbage. *Acta physiologae plantarum*, 36(4): 815-823.

- Machado, R. M. A., and Serralheiro, R. P. (2017). Soil salinity: Effect on vegetable crop growth. Management practices to prevent and mitigate soil salinization. *Acta Horticulturae*, 3(2): 30.
- Maiti, R. K., Vidyasagar, P., Umashanker, P., Gupta, A., Rajkumar, D., and Rodriguez, H. G. (2010). Comparative levels of salinity tolerance of different vegetable crops. *Int J Bio-Resources Stress Management*, 2: 105-109.
- Maksimovic, I., and Ilin, Z. (2012). Effects of salinity on vegetable growth and nutrients uptake. *Irrigation Systems and Practices in Challenging Environments*, 9-12pp.
- Martinez-Ballesta, M. C., Alcaraz-Lopez, C., Muries, B., Mota-Cadenas, C., and Carvajal, M. (2010). Physiological aspects of rootstock–scion interactions. *Scientia Horticulturae*, 127(2): 112-118.
- Martinez-Rodriguez, M. M., Estan, M. T., Moyano, E., Garcia-Abellán, J. O., Flores, F. B., and Bolarín, M. C. (2008). The effectiveness of grafting to improve salt tolerance in tomato when an 'excluder'genotype is used as scion. *Environmental and Experimental Botany*, 63(1-3): 392-401.
- Melnyk, C. W. (2017). Plant grafting: Insights into tissue regeneration. *Regeneration*, 4(1): 3-14.
- Meng, X., Zhou, J., and Sui, N. (2018). Mechanisms of salt tolerance in halophytes: Current understanding and recent advances. *Open Life Sciences*, 13(1): 149-154.
- Mngersquo, S. A., Sileshi, G. W., Jamnadass, R., Akinnifesi, F. K., and Mhango, J. (2012). Scion and stock diameter size effect on growth and fruit production of *Sclerocarya birrea* (Marula) trees. *Journal of Horticulture and Forestry*, 4(9): 153-160.
- Morais, P. L., Dias, N., Oliveira, A. M., Sousa, O. N., Sarmento, J. D., and Gonzaga, M. I. (2018). Effects of nutrient solution salinity on the physiological performance of melon cultivated in coconut fiber. *Revista Caatinga*, 31: 713-718.
- Mudge, K., Janick, J., Scofield, S., and Goldschmidt, E. E. (2009). A history of grafting. *Horticulture Reviews*, 35(9): 437-493.
- Munns, R. (2002). Comparative physiology of salt and water stress. *Plant, cell and environment*, 25(2): 239-250.
- Munns, R. (2005). Genes and salt tolerance: bringing them together. *New phytologist*, 167(3): 645-663.

- Munns, R. and Tester, M. (2008). Mechanisms of salinity tolerance. *Annual Review of Plant Biology*, 59: 651-681.
- Munns, R., James, R.A. and Lauchli, A. (2006). Approaches to increasing the salt tolerance of wheat and other cereals. *Journal of Experimental Botany* 57:1025-1043.
- Muradoglu, F., Gundogdu, M., Ercisli, S., Encu, T., Balta, F., Jaafar, H. Z., and Zia-Ul-Haq, M. (2015). Cadmium toxicity affects chlorophyll a and b content, antioxidant enzyme activities and mineral nutrient accumulation in strawberry. *Biological research*, 48: 1-7.
- Musyimi, D. M., Netondo, G. W., and Ouma, G. (2007). Effects of salinity on growth and photosynthesis of avocado seedlings. *International Journal of Botany*, 3(1): 78-84.
- Navarro, A., Banon, S., Olmos, E., and Sanchez-Blanco, M. D. J. (2007). Effects of sodium chloride on water potential components, hydraulic conductivity, gas exchange and leaf ultrastructure of *Arbutus unedo* plants. *Plant Science*, 172(3): 473-480.
- Navarro, J. M., Botella, M. A., Cerdá, A., and Martínez, V. (2001). Phosphorus uptake and translocation in salt-stressed melon plants. *Journal of plant physiology*, 158(3): 375-381.
- Nazarbeygi, E., Yazdi, H. L., Naseri, R., and Soleimani, R. (2011). The effects of different levels of salinity on proline and A-B chlorophylls in canola. *American-Eurasian Journal of Agricultural and Environmental Sciences*, 10(1): 70-74.
- Neocleous, D., Ntatsi, G., and Savvas, D. (2017). Physiological, nutritional and growth responses of melon (*Cucumis melo L.*) to a gradual salinity built-up in recirculating nutrient solution. *Journal of Plant Nutrition*, 40(15): 2168-2180.
- Nguyen, V. H., and Yen, C. R. (2018). Rootstock age and grafting season affect graft success and plant growth of papaya (*Carica papaya L.*) in greenhouse. *Chilean Journal of Agricultural Research*, 78(1): 59-67.
- Noor, R. S., Wang, Z., Umair, M., Yaseen, M., Ameen, M., and Sun, Y. (2019). Interactive effects of grafting techniques and scion-rootstocks combinations on vegetative growth, yield and quality of cucumber (*Cucumis sativus L.*). *Agronomy*, 9(6): 288.
- Noreen, Z., and Ashraf, M. (2009). Assessment of variation in antioxidative defense system in salt-treated pea (*Pisum sativum*) cultivars and its putative use as salinity tolerance markers. *Journal of Plant Physiology*, 166(16): 1764-1774.

- Oda, M., Maruyama, M., and Mori, G. (2005). Water transfer at graft union of tomato plants grafted onto *Solanum* rootstocks. *Journal of the Japanese Society for Horticultural Science*, 74(6): 458-463.
- Olfati, J. A., Saadatian, M., and Moqbeli, E. (2015). Optimisation of nitrogen and potassium for *Aloe vera* (L.) Burm. f. in a soilless culture system. *South African Journal of Plant and Soil*, 32(4): 249-252.
- Oosthuyse, S. A. (1996). Effect of KNO_3 sprays to flowering mango trees on fruit retention, fruit size, tree yield, and fruit quality. In *V International Mango Symposium* 455: 359-366.
- Padder, B. M., Agarwal, R. M., Kaloo, Z. A., and Singh, S. (2012). Nitrate reductase activity decreases due to salinity in mungbean (*Vigna radiata* L.). *Int. J. Curr. Res. Rev*, 4: 117-123.
- Pardossi, A., Malorgio, F., Incrocci, L., Campiotti, C. A., and Tognoni, F. (2002). A comparison between two methods to control nutrient delivery to greenhouse melons grown in recirculating nutrient solution culture. *Scientia Horticulturae*, 92(2): 89-95.
- Parida, A. K., and Das, A. B. (2005). Salt tolerance and salinity effects on plants: A review. *Ecotoxicology and environmental safety*, 60(3): 324-349.
- Patel, R. K., Babu, K. D., Singh, A., Yadav, D. S., and De, L. C. (2007). Soft Wood Grafting in Mandarin (*C. reticulata Blanco*) A Novel Vegetative Propagation Technique. *International Journal of Fruit Science*, 7(2): 31-41.
- Pereira, F. A. D. L., Medeiros, J. F. D., Gheyi, H. R., Dias, N. D. S., Preston, W., and Vasconcelos, C. B. (2017). Tolerance of melon cultivars to irrigation water salinity. *Revista Brasileira de Engenharia Agrícola e Ambiental*, 21: 846-851.
- Pessarakli, M. (2016). Handbook of Cucurbits, Growth, Cultural Practices, and Physiology. Florida: CRC Press, Taylor and Francis Publishing Group, 574pp.
- Peterson, D. G., Pearson, W. R., and Stack, S. M. (1998). Characterization of the tomato (*Lycopersicon esculentum*) genome using in vitro and in situ DNA reassociation. *Plant Genome*, 41(3): 346-356.
- Pisoschi, A. M., Cheregi, M. C., and Danet, A. F. (2009). Total antioxidant capacity of some commercial fruit juices: electrochemical and spectrophotometrical approaches. *Molecules*, 14(1): 480-493.

- Plaut, J. A., Wadsworth, W. D., Pangle, R., Yepez, E. A., Dowell, N. G., and Pockman, W. T. (2013). Reduced transpiration response to precipitation pulses precedes mortality in a piñon-juniper woodland subject to prolonged drought. *New Phytologist*, 200(2): 375-387.
- Pooyeh, F., Peyvast, G., and Olfati, J. A. (2012). Growing media including palm waste in soilless culture of cucumber. *International journal of vegetable science*, 18(1): 20-28.
- Prajapati, R. P., Kalariya, M., Parmar, S. K., and Sheth, N. R. (2010). Phytochemical and pharmacological review of *Lagenaria siceraria*. *Journal of Ayurveda and Integrative Medicine*, 1(4): 266.
- Raman, A., and Lau, C. (1996). Anti-diabetic properties and phytochemistry of *Momordica charantia* L. (Cucurbitaceae). *Journal of Phytomedicine*, 2(4): 349-362.
- Razeek, N., Sittampalam, T., and Kapilan, R. (2015). Optimization of salinity and temperature for the efficient seed germination of *Momocardia charantia*. *Int. J. Adv. Res. Biol. Sci*, 2(12): 73-80.
- Reddy, K. S., and Menary, R. C. (1990). Nitrate reductase and nitrate accumulation in relation to nitrate toxicity in *Boronia megastigma*. *Physiologia Plantarum*, 78(3): 430-434.
- Richards, L.A. (1954). Diagnosis and improvement of saline and alkaline soils. *Journal of Soil Science* 78(2): 154.
- Romero, L., Belakbir, A., Ragala, L., and Ruiz, J. M. (1997). Response of plant yield and leaf pigments to saline conditions: effectiveness of different rootstocks in melon plants (*Cucumis melo* L.). *Soil Science and Plant Nutrition*, 43(4): 855-862.
- Rouphael, Y., Cardarelli, M., Rea, E., and Colla, G. (2012). Improving melon and cucumber photosynthetic activity, mineral composition, and growth performance under salinity stress by grafting onto Cucurbita hybrid rootstocks. *Photosynthetica*, 50(2): 180-188.
- Rouphael, Y., Cardarelli, M., Rea, E., Battistelli, A., and Colla, G. (2006). Comparison of the subirrigation and drip-irrigation systems for greenhouse zucchini squash production using saline and non-saline nutrient solutions. *Agricultural water management*, 82(1-2): 99-117.
- Rouphael, Y., Schwarz, D., Krumbein, A., and Colla, G. (2010). Impact of grafting on product quality of fruit vegetables. *Scientia Horticulturae*, 127(2): 172-179.

- Ruiz, J. M., Belakbir, A., Lopez-Cantarero, A., and Romero, L. (1997). Leaf macronutrient content and yield in grafted melon plants: A model to evaluate the influence of rootstock genotype. *Scientia Horticulturae*, 71: 227–234.
- Rutkowski, K. P., Michalczuk, B., and Konopacki, P. (2008). Nondestructive determination of ‘Golden Delicious’ apple quality and harvest maturity. *Journal of Fruit Ornamental Plant Research*, 16(1): 39-52.
- Saha, S., Sadle, J., Van Der Heiden, C., and Sternberg, L. (2015). Salinity, groundwater, and water uptake depth of plants in coastal uplands of Everglades National Park (Florida, USA). *Journal of Ecohydrology*, 8(1): 128-136.
- Saligedar, F., Sedaqathour, S., and Olfati, J. A. (2014). Nutrient solution on Aloin content and other quality characteristics of Aloe vera. *Journal of Medicinal Plant and Byproducts*, 1: 59-62.
- Sanchez-Castillo, C. P., and James, W. P. (2013). *Salt: Epidemiology*. Encyclopedia of Human Nutrition (Third Edition), Academic Press.
- Santa-Cruz, A., Martinez-Rodriguez, M. M., Perez-Alfocea, F., Romero-Aranda, R., and Bolarin, M. C. (2002). The rootstock effect on the tomato salinity response depends on the shoot genotype. *Plant Science*, 162(5): 825-831.
- Sarabi, B., Bolandnazar, S., Ghaderi, N., and Ghashghaie, J. (2017). Genotypic differences in physiological and biochemical responses to salinity stress in melon (*Cucumis melo L.*) plants: prospects for selection of salt tolerant landraces. *Plant Physiology and Biochemistry*, 119: 294-311.
- Shabala, S. (2013). Learning from halophytes: physiological basis and strategies to improve abiotic stress tolerance in crops. *Annals of botany*, 112(7): 1209-1221.
- Shafieizargar, A., Awang, Y., Ajamgard, F., Juraimi, A. S., Othman, R., and Ahmadi, A. K. (2015). Assessing five citrus rootstocks for NaCl salinity tolerance using mineral concentrations, proline and relative water contents as indicators. *Asian Journal of Plant Sciences*, 14(1): 20-26.
- Shahid M., Salim J., Mohd Noor M.R., Ab Hamid A.H., Abd Manas M., and Ahmad S. A. (2009). Manual Teknologi Fertigasi Penanaman Cili, Rockmelon dan Tomato. Publisher MARDI, 42-56pp.
- Shannon, M. C., and Grieve, C. M. (1998). Tolerance of vegetable crops to salinity. *Scientia Horticulturae*, 78(1-4): 5-38.

- Sheoran, I. S., and Saini, H. S. (1996). Drought-induced male sterility in rice: changes in carbohydrate levels and enzyme activities associated with the inhibition of starch accumulation in pollen. *Sexual Plant Reproduction*, 9(3): 161-169.
- Shrivastava, P., and Kumar, R. (2015). Soil salinity: A serious environmental issue and plant growth promoting bacteria as one of the tools for its alleviation. *Saudi Journal of Biological Sciences*, 22(2): 123-131.
- Shu, S., Yuan, L. Y., Guo, S. R., Sun, J., and Liu, C. J. (2012). Effects of exogenous spermidine on photosynthesis, xanthophyll cycle and endogenous polyamines in cucumber seedlings exposed to salinity. *African Journal of Biotechnology*, 11(22): 6064-6074.
- Silva Oliveira, W., Stamford, N. P., Vila Nova da Silva, E., de Rosalia Silva Santos, C. E., Santiago de Freitas, A. D., Stamford Arnaud, T. M., and Sarmento, B. F. (2014). Biofertilizer produced by interactive microbial processes affects melon yield and nutrients availability in a Brazilian semiarid soil. *Australian Journal of Crop Science*, 8(7): 1124-1130.
- Singh, A. L., Hariprassana, K., and Solanki, R. M. (2008). Screening and selection of groundnut genotypes for tolerance of soil salinity. *Australian Journal of Crop Science*, 1(3): 69-77.
- Sivritepe, N., Sivritepe, H. O., and Eris, A. (2003). The effects of NaCl priming on salt tolerance in melon seedlings grown under saline conditions. 97(3-4): 229-237.
- Slama, I., Abdelly, C., Bouchereau, A., Flowers, T., and Savoure, A. (2015). Diversity, distribution and roles of osmoprotective compounds accumulated in halophytes under abiotic stress. *Annals of Botany*, 115(3): 433-447.
- Soomro, A., Soomro, K. B., Akhtar, J., Soomro, S., and Ali, A. (2015). Impact of salinity on growth, yield and water use efficiency of *Momordica charantia* L. under raisedbed irrigation. *Int. J. Biol. Biotechnology*, 12: 485-491.
- Soumelidou, K., Li, H., Barnett, J. R., John, P., and Battey, N. H. (1994). The effect of auxin and calcium antagonists on tracheary element differentiation in *Phaseolus vulgaris*. *Journal of Plant Physiology*, 143(6): 717-721.
- Sousa, A. B., Duarte, S. N., Sousa, O. N. D., Souza, A., Sampaio, P. R., and Dias, C. T. (2016). Production and quality of mini watermelon cv. Smile irrigated with saline water. *Revista Brasileira de Engenharia Agricola Ambiental*, 20: 897-902.

- Sousa, V. F., Costa, C. C., Diniz, G. L., Santos, J. B., and Bomfim, M. P. (2018). Physiological behavior of melon cultivars submitted to soil salinity. *Pesquisa Agropecuaria Tropical*, 48: 271-279.
- Sparks, D. L. (2003). *Environmental soil chemistry*. Elsevier.
- Stepien, P., and Kłbus, G. (2006). Water relations and photosynthesis in *Cucumis sativus L.* leaves under salt stress. *Biologia Plantarum*, 50(4): 610-616.
- Tandonnet, J. P., Cookson, S. J., Vivin, P., and Ollat, N. (2010). Scion genotype controls biomass allocation and root development in grafted grapevine. *Australian Journal of Grape and Wine Research*, 16(2): 290-300.
- Tang, W., Yueh, S. H., Hayashi, A., Fore, A. G., Jones, W. L., Santos-Garcia, A., and Jacob, M. M. (2015). Rain-induced near surface salinity stratification and rain roughness correction for Aquarius SSS retrieval. *IEEE Journal of Selected Topics in Applied Earth Observations and Remote Sensing*, 8(12): 5474-5484.
- Tanuja, P., and Thippesha, D. (2017). Effect of different age groups of scions on success rate of softwood grafting in sapota. *Journal of Pharmacognosy and Phytochemistry*, 6(5): 1886-1889.
- Teakle, N. L., and S. D. Tyerman. (2010). Mechanisms of Cl⁻ transport contributing to salt tolerance. *Plant, Cell and Environment*, 33: 566–589.
- Tester, M., and Davenport, R. (2003). Na⁺ tolerance and Na⁺ transport in higher plants. *Annals of Botany*, 91(5): 503-527.
- Toumi, M., Nedjimi, B., Halitim, A., and Garcia, M. (2016). Effects of K-Mg ratio on growth and cation nutrition of grafted *Vitis vinifera L.* cv. "Dattier de Beiruth". *Journal of Plant Nutrition*, 39(7): 904-911.
- Traka-Mavrona, E., Koutsika-Sotiriou, M., and Pritsa, T. (2000). Response of squash (*Cucurbita* spp.) as rootstock for melon (*Cucumis melo L.*). *Scientia Horticulturae*, 83(3-4): 353-362.
- Tavakkoli, E., Rengasamy, P. and Mc Donald, K. (2010). High concentrations of Na⁺ and Cl⁻ ions in soil solution have simultaneous detrimental effects on growth of faba bean under salinity stress. *Journal of Experimental Botany*, 61(15): 4449-4459.
- Tuttle, M. A., and Gotlieb, A. R. (1985). Graft union histology and distribution of tomato ringspot virus in infected McIntosh/ Malling Merton 106 apple trees. *Journal of Phytopathology*, 75(3): 347-351.

- Ulas, F., Aydin, A., Ulas, A., and Yetisir, H. (2019). Grafting for sustainable growth performance of melon (*Cucumis melo L.*) under salt stressed hydroponic condition. *European Journal of Sustainable Development*, 8(1): 201-205.
- Umar, S. (2006). Alleviating adverse effects of water stress on yield of sorghum, mustard and groundnut by potassium application. *Pakistan J. Bot.* 38:1373–1380.
- US Salinity Laboratory Staff. (1954). Reclamation and improvement of saline and sodic soils. USDA Handbook 60. Riverside, California.
- Usherwood, N. R. (1985). The role of potassium in crop quality. *Potassium in Agriculture*, 489-513pp.
- Verbruggen, N., and Hermans, C. (2013). Physiological and molecular responses to magnesium nutritional imbalance in plants. *Journal of Plant and Soil*, 368(1): 87-99.
- Wang, H. M., Xiao, X. R., Yang, M. Y., Gao, Z. L., Zang, J., Fu, X. M., and Chen, Y. H. (2014). Effects of salt stress on antioxidant defense system in the root of *Kandelia candel*. *Journal of Botany*, 55(1): 1-7.
- Weatherly, P. (1950). Studies in the water relations of the cotton plant. *New Phytol.* 49: 81-97.
- Wellburn, A. R. (1994). The spectral determination of chlorophylls a and b, as well as total carotenoids, using various solvents with spectrophotometers of different resolution. *Journal of plant physiology*, 144(3): 307-313.
- Xiong, L., Schumaker, K. S., and Zhu, J. K. (2002). Cell signaling during cold, drought, and salt stress. *The Plant Cell*, 14: 165-183.
- Yang, Y., Wang, L., Tian, J., Li, J., Sun, J. and He, L. (2012). Proteomic study participating the enhancement of growth and salt tolerance of bottle gourd rootstock-grafted watermelon seedlings. *Plant Physiology and Biochemistry*, 58:54–65.
- Yanyan, Y. A. N., Shuoshuo, W. A. N. G., Min, W. E. I., Biao, G. O. N. G., and Qinghua, S. H. I. (2018). Effect of different rootstocks on the salt stress tolerance in watermelon seedlings. *Horticultural Plant Journal*, 4(6): 239-249.
- Yarsi, G., Sivaci, A., Dasgan, H. Y., Altuntas, O., Binzet, R., and Akhoundnejad, Y. (2017). Effects of salinity stress on chlorophyll and carotenoid contents and stomata size of grafted and ungrafted galia C8 melon cultivar. *Pak. J. Bot.* 49(2): 421-426.

- Yetisir, H., and Sari, N. (2003). Effect of different rootstock on plant growth, yield and quality of watermelon. *Australian Journal of Experimental Agriculture*, 43(10): 1269-1274.
- Yetisir, H., and Uygur, V. (2010). Responses of grafted watermelon onto different gourd species to salinity stress. *Journal of Plant Nutrition*, 33(3): 315-327.
- Yetişir, H., Denli, N., Ata, A., Ulaş, A., Pınar, H., Kirnak, H., and Yamac, M. (2016). Determination of salt tolerance potential of Turkish bottle gourd (*Lagenaria siceraria*) germplasm. *Meeting on Cucurbit Genetics and Breeding*, 278-282pp.
- Zayan, M., Zeerban, S., El-Samak, A., and Ezzat, A. (2011). Evaluation study for the effect of four mango rootstocks on behaviour of Keit mango variety. *International Journal of Horticultural Science*, 17(4-5): 49-53.
- Zhang, L., Ma, G., Kato, M., Yamawaki, K., Takagi, T., Kiriwa, Y., and Nesumi, H. (2012). Regulation of carotenoid accumulation and the expression of carotenoid metabolic genes in citrus juice sacs in vitro. *Journal of experimental botany*, 63(2), 871-886.
- Zhang, P., Senge, M., and Dai, Y. (2016). Effects of salinity stress on growth, yield, fruit quality and water use efficiency of tomato under hydroponics system. *Reviews in Agricultural Science*, 4: 46-55.
- Zhu, J. K. (2001). Plant salt tolerance. *Trends in Plant Science*, 6(2): 66-71.
- Zhu, J., Bie, Z. and Li, Y. (2008). Physiological and growth responses of two different salt-sensitive cucumber cultivars to NaCl stress, *Soil Science and Plant Nutrition* 54(3): 400-407.
- Zulkarami, B., Ashrafuzzaman, M., and Razi, I. M. (2010). Morphophysiological growth, yield and fruit quality of rock melon as affected by growing media and electrical conductivity. *Journal of Food Agriculture and Environment*, 8(1): 249-252.