

UNIVERSITI PUTRA MALAYSIA

EFFECT OF AQUEOUS NEEM LEAVES EXTRACT IN CONTROLLING Fusarium WILT BANANA (Musa spp.)

UNG YI

FP 2021 65



EFFECT OF AQUEOUS NEEM LEAVES EXTRACT IN CONTROLLING Fusarium WILT BANANA (Musa spp.)



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

Jun 2020

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

EFFECT OF AQUEOUS NEEM LEAVES EXTRACT IN CONTROLLING Fusarium WILT OF BANANA (Musa spp.)

By

UNG YI

Jun 2020

Chair Faculty : Assoc. Prof. Siti Zaharah Sakimin, PhD : Agriculture

Neem leaf extracts (NLE) have frequently been applied to inhibit plant diseases and for the development of bio-fertilizer that leading to the commercial exploitation of this tree in agriculture. However, previous studies had indicated contradictory outcomes when it was applied as antifungal disease treatment and as bio-fertilizer when applied through the soil for several crops, including banana. Therefore, the present investigation was undertaken (i) to assess the growth of Fusarium oxysporum f. sp. cubense (Foc) fungus using a suitable media for control of pH in-vitro condition and (ii) to examine the occurrence and severity of diseases caused by Foc on banana "Cavendish" treated with NLE at the optimum soil acidity in the glasshouse condition. In 1A in-vitro experiment, different NLE rate from 0 to 10 ml was added into 250 ml of potato dextrose agar (PDA) media to grow the Foc fungus the while in the 1B experiment, the Foc were growth in different pH level (5, 5.5 as control, 6 and 7) of PDA media under complete randomized design (CRD) experimental design. In 1A experiment, colony radial extension decreased while the percentage of inhibition of radial growth (PIRG, %) increased significantly with increasing NLE rate for 7 days after incubation (DAI). However, no significant difference of all parameters was recorded for both 8 ml (8:250) and 10 ml (10:250) of NLE. Besides that, both pH 5 and 7 of PDA media also had the same effective significant effect in all parameters than under any other pH of the PDA media in the 1B experiment. Due to the planting condition of banana plants and according to significant differences, in Experiment 2, the best level of 8 ml aqueous NLE per 250 ml media (or at the ratio of 8:250 or 4:125) from Experiment 1A was applied to Fusarium-inoculated Cavendish banana (Musa spp. AAA group cv. 'Grand Nain') under the best soil media pH condition (optimum pH 7) from Experiment 1B. The 40 ml Fusarium fungus solution with the population of 2.50 x 10⁸ spores ml⁻¹ was poured onto the soil. The solution of NLE treatment was applied after one month of Fusarium application. The treatments in Experiment 2 were arranged by randomized complete block design (RCBD). Banana plants associated fungus were detrimental significantly and showed wilting symptoms. Foc resulting in high

disease severity (leaves and rhizome) and incident of *Fusarium* wilt (%). Moreover, it was observed that the application of extract improves significantly in plant height, pseudo-stem diameter, root size and distribution, plant biomass production as well as soil physiochemical properties and tend to resist *Fusarium* wilt diseases. Thus, the major finding of this study shows that the application of NLE solution promotes better growth of cavendish banana plants and tend to withstand *Fusarium* wilt infection under optimum pH 7 media. Based on the results, aqueous NLE (at the ratio of 4:125) was recommended to be applied in the field as a treatment to improve Cavendish banana growth and delay *Fusarium* diseases damage to plants under pH7 soil condition.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

POTENSI EXTRAK AIR NEEM SEBAGAI PENGAWALAN PENYAKIT Fusarium PADA TANAMAN PISANG (Musa spp.)

Oleh

UNG YI

Jun 2020

Pengerusi : Prof. Madya Siti Zaharah Sakimin, PhD Fakulti : Pertanian

Daun neem extract (NLE) sering digunakan oleh petani untuk mengatasi pelbagai masalah penyakit tanaman dan mempunyai potensi yang tinggi untuk pembangunan komersial kerana permintaan baja organic semasa telah meningkat secara drastic. Walau bagaimanapun, kajian dan penyelidikan yang sedia masih kurang bukti untuk mengenal pasti keberkesanan penggunaan daun neem extract sebagai rawatan penyakit dan sebagai baja organic terhadap pertumbuhan pelbagai penanaman termasuk pisang. Sehubung dengan itu, berikut adalah objektif penyelidikan ini bagi menjawab permasalahan tersebut iaitu (i) mengkaji pertumbuhan Fusarium oxysporum f.sp. cubense (Foc) fungi dalam keadaan in-vitro menggunakan media yang sesuai di bawah keadaan pH yang beza dan (ii) menentukan kesan NLE terhadap pertumbuhan pokok pisang 'Cavendish' yang diinokulasi dengan Foc dan pengurangan penyakit layu Fusarium atas keasidan tanah yang optimum dalam rumah kaca. Dalam Kajian 1A, NLE dengan kepekatan yang berbeza dari 0 hingga 10 ml telah ditambah dalam potato dextrose agar (PDA) untuk tumbuh Foc fungi manakala dalam Kajian 1B, Foc tumbuh pada PDA media dengan kadar pH yang beza (5, 5.5control, 6 dan 7) di bawah reka bentuk eksperimen rawak lengkap (CRD). Peningkatan NLE dari 0 ml hingga 10 ml dalam Kajian 1A telah meningkatkan secara ketara terhadap peratusan perencatan pertumbuhan jejari (PIRG, %) Foc tetapi kecikan koloni jejari Foc dari hari pertama hingga hari ketujuh selepas inkubasi (DAI). Walau bagaimanapun, aplikasi antara 8 ml (8:250) dan 10 ml (10:250) NLE tidak menunjukan significant yang beza untuk semua parameter. Selain daripada itu, keputusan Kajian 1B menunjukkan bahawa pH 5 dan 7 memberi kesan signifikan yang sama terhadap koloni jejari, kadar pertumbuhan dan PIRG Foc. Menurut keadaan penanaman pokok pisang termasuk kesan signifikan, dalam Kajian 2 kepekatan terbaik 8 ml air NLE per 250 ml media (kadar 8:250 atau 4:125) dari Kajian 1A telah digunakan atas Fusarium inokulasi pokok pisang Cavendish (Musa spp. AAA group cv. 'Grand Nain') dengan pH media terbaik (optimum pH 7) dari Kajian 1B. Media tanah diinokulasi dengan kadar 40 ml Fusarium pada kepekatan 2.50 x 10⁸ spora ml⁻¹ serta diaplikasi dengan rawatan NLE sebulan selepas inokulasi Fusarium fungus. Rawatan dalam Eksperimen 2 disusun dengan reka bentuk blok lengkap secara rawak (RCBD). Pokok pisang diinokulasikan dengan fungus menunjuk tanda-tanda layu. Keputusan mendapati bahawa keparahan (severity) penyakit (daun dan sulur) dan insiden layu Fusarium (%) meningkat dengan inokulasi Foc. Selain daripada itu, aplikasi NLE menunjukkan peningkatkan yang ketara bagi parameter ketinggian pokok, diameter batang, ukuran dan pengedaran akar (luas permukaan akar, diameter akar dan isipadu akar), berat pokok (berat segar akar, berat kering akar dan nisbah pucuk akar) serta sifat fizikokimia tanah (kapasiti pertukaran kation, nitrogen, fosforus, kalium, kalsium dan magnesium) dan mungkin berkesan dalam melawan penyakit layu Fusarium. Oleh itu, penemuan utama daripada hasil kajian ini membuktikan bahawa NLE memberi faedah atas penumbuhan pokok dan berpotensi meningkatkan kerintangan pokok pisang cavendish terhadap serangan penyakit layu Fusarium atas optimum pH7 media. Secara keseluruhannya, air NLE (4:125) disarankan untuk diaplikasi di ladang sebagai rawatan untuk melambatkan penyakit Fusarium yang mengakibantkan kerosakan teruk pada pokok dan memperbaiki pertumbuhan pokok pisang Cavendish di bawah keadaan tanah pH7.

ACKNOWLEDGEMENTS

First of all, I would like to express the deepest appreciation to my supervisor, Assoc. Prof. Siti Zaharah Sakimin for being my tremendous supervisor, she continually and convincingly conveyed a spirit of adventure in regard to research and an excitement in regard to teaching. I am extremely grateful and indebted to her for her expert, sincere and valuable guidance and encouragement extended to me. Without her guidance and persistent help, this research dissertation would not have been done.

Meanwhile, I am also grateful to all my committee supervisor Prof. Dr. Mohamed Hanafi Musa and Dr. Siti Izera binti Ismail. I am extremely thankful and indebted to them for sharing expertise, and sincere and valuable guidance and encouragement extended to me.

I would also like to extend my gratitude to Puchong field, UPM for supplying Cavendish banana seedlings, neem leaves as well as providing assistance in preparing the experimental material.

Special thanks for all the staffs from Department of Crop Science, Department of Plant Protection and Department of Soil Science Management for providing all the necessary facilities, help, suggestion, cooperation and encouragement to complete my research.

Moreover, I would also like to extend my thanks to my family members, Ung Toon Sun, my father, Chin Thui Kiew, my mother while also my sister Ung Tian and Ung Wen who give moral support and caring to finish my research. The comment and supportive from them make me has more strength, power and spirit to continue my task. In addition, I also thank my friends for the encouragement, support and attention. Last but not least, it is a genuine pleasure to express my deep sense of thanks and gratitude to all of those who involved directly or indirectly for their kind help and co-operation throughout my study period. This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Siti Zaharah binti Sakimin, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Siti Izera binti Ismail, PhD

Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Member)

Mohamed Hanafi bin Musa, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 9 December 2021

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of	
Supervisory Committee:	Assoc. Prof. Dr. Siti Zaharah Sakimin
Signature:	
Name of Member of	E serve E
Supervisory Committee:	Dr. Siti Izera Ismail
Signature:	
Name of Member of	
Supervisory Committee	Prof. Dr. Mohamed Hanafi Musa

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiv
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	xviii

CHAP ⁻	TER
-------------------	-----

СНА	PTER				
1	INTE	RODUC			1
2	LITE 2.1 2.2	ERATUI Banar 2.1.1 2.1.2 2.1.3 2.1.4 2.1.5 Diseas 2.2.1 2.2.2	RE REVIE Botany, Propaga Growth r World ar Cultivations Ses and por Common Fusariur 2.2.2.1 2.2.2.2 2.2.2.3 2.2.2.4	W morphology and varieties tion requirement and condition nd Malaysia production of banana on in Malaysia ests of banana crops n disease in banana n wilt or panama diseases Fusarium wilt diseases Fusarium wilt diseases effects on agriculture crops Morphology and characteristic of Fusarium fungus The global history and distribution of Fusarium wilt diseases Fusarium wilt diseases Fusarium wilt diseases	3 3 5 6 7 8 8 9 9 10 11
	2.3	Contro 2.3.1	b) of <i>Fusar</i> Neem 2.3.1.1 2.3.1.2 2.3.1.3 Disease 2.3.2.1 2.3.2.2	symptom and spreading factors rium diseases Neem products and their agricultural application Characteristic and properties of neem leave The effect of neem leaves product as biological control for suppression of <i>Fusarium</i> fungus under in-vitro and in- vivo condition s suppressive soils Soil condition (pH of media) The effect of media pH as biological control for suppression of Fusarium	15 17 17 19 20 23 24 24 24

fungus under in-vitro and in-vivo condition

3	THE AQU pH L	EFFE	CTS OF DIFFERENT CONCENTRATION OF NEEM LEAVES EXTRACT AND DIFFERENT ON THE <i>Fusarium</i> spp. GROWTH UNDER <i>in</i> -	29
	3.1	Introdu	uction	29
	3.2	Materi	als and methods	30
		3.2.1	Experimental site	30
		3.2.2	Preparation of PDA media as growing materials	30
		3.2.3	Preparation and culture of <i>Fusarium oxysporum</i> fungus	30
		3.2.4	Experiment 1A: Treatment, method of application and experimental design	31
			3.2.4.1 Preparation of aqueous neem leaves extract at a different rate	31
			3.2.4.2 Method of application and	31
			experimental design	
		3.2.5	Experiment 1B: Treatment, method of application and experimental design	32
			3.2.5.1 Preparation of PDA media at a different pH level	32
			3.2.5.2 Method of application and experimental design	32
	3.3	Data C	Collection	32
		3.3.1	Colony radial extension of Foc	32
		3 <mark>.3.2</mark>	Growth rate of Foc	32
		3 <mark>.3.3</mark>	Percentage of Inhibition of Radial Growth of Foc	33
	3.4	Statist	ical Analysis	33
	3.5	Result	ts	33
		3.5.1	Experiment 1A: The effects of different aqueous	33
			neem leaves extract on the Fusarium growth	
			under the <i>in-vitro</i> condition	
			3.5.1.1 Colony radial extension of Foc	33
			3.5.1.2 Growth rate of Foc	34
			3.5.1.3 Percentage of Inhibition of Radius Growth of Foc	34
		3.5.2	Experiment 1B: The effects of different pH level	37
			of growth media on the <i>Fusarium</i> growth under	
			the <i>in-vitro</i> condition	07
			3.5.2.1 Colony radial extension of Foc	37
			3.5.2.2 Growth rate of Foc	37
	_		3.5.2.3 Percentage of Inhibition of Radius Growth of Foc	37
	3.6	Discus	ssions	40
	3.7	Conclu	usion	44

(C)

4 EFF CON	ECTS (ITROLI	DF AQUEOUS NEEM LEAVES EXTRACT IN LING <i>Fusarium</i> WILT OF <i>Musa</i> spp. GROWN	45
UND	DER OP	TIMIZED pH MEDIA	
4.1	Introd	uction	45
4.2	Materi	als and methods	46
	4.2.1	Experimental site	46
	4.2.2	Preparation of soil at selected pH level	46
	4.2.3	Preparation and culture of Fusarium oxysporum	46
		fungus	
	4.2.4	Preparation of aqueous neem leaves extract as	47
	4.2.5	Preparation of Cavendish banana as planting	47
		materials	
	4.2.6	Treatment and method of application	47
	4.2.7	Experimental design	48
4.3	Data c	ollections	48
	4.3.1	Nutrient contents of neem leaves	48
	4.3.2	Plant growth and morphology	48
		4.3.2.1 Plant height	49
		4.3.2.2 Pseudo-stem diameter	49
		4.3.2.3 Total leaf number	49
		4.3.2.4 Leaf dimension parameters (area,	49
		length, width)	
		4.3.2.5 Nutrient contents of banana leaf	49
		4.3.2.6 Root size and distribution	50
		4.3.2.7 Plant biomass production	50
	4.3.3	Physiological changes	50
		4.3.3.1 Photosynthesis rate, stomatal	50
		conductance, transpiration rate and	
		water vapor deficit	
		4.3.3.2 Chlorophyll content	51
	4.3.4	Disease assessments	51
		4.3.4.1 Leaf symptom index and rhizome	51
		4342 Leaf disease severity index rhizome	54
		diseases severity index and	54
		effectiveness of neem leaves extract	
		treatment	
		1313 Lest damage percentage and rhizome	55
		wilting percentage	55
		4.2.4.4 Poduco in Eusprium wilt	55
		4.3.4.4 Reduce in Fusanum with	55
	125	A.S.4.5 Soli microbial population	50
	4.3.3		50
		4.3.5.1 SUII μ	20
		4.3.5.2 Callon exchange Capacity	20 57
		4.5.5.5 Available prosphorus	5/ F7
	01-1-1	4.3.5.4 Soll exchangeable bases	5/
4.4	Statis	tical analysis	57
4.5	Result	is and discussion	57
	4.5.1	Nutrient contents of neem leaves	57
	4.5.2	Plant growth and morphology	58

			4.5.2.1	Plant height	58
			4.5.2.2	Pseudo-stem diameter	58
			4.5.2.3	Total leaf number	59
			4.5.2.4	Leaf area	61
			4.5.2.5	Nutrient contents of banana leaf	61
			4.5.2.6	Root size and distribution	62
			4.5.2.7	Plant biomass production	63
		4.5.3	Physiolo	gical changes	64
			4.5.3.1	Photosynthesis rate, stomata	64
				conductance, transpiration rate and	
				vapour pressure deficit	
			4.5.3.2	Chlorophyll content	65
		4.5.4	Disease	assessments	66
			4.5.4.1	Leaf wilting symptom	66
			4.5.4.2	Rhizome discoloration symptom	71
			4.5.4.3	Soil microbial population	73
		4.5.5	Soil phys	siochemical properties	73
			4.5.5.1	Soil pH	74
			4.5.5.2	Cation exchange capacity	/4
			4.5.5.3	Soil exchangeable bases and	74
	46	Discus	sion	available prospriorus	77
	4.0	Conclu	ision		86
	7.7	Conoic	101011		00
5	GEN	IERAL	DISC	USSION, CONCLUSION AND	87
	REC	O <mark>MME</mark>	NDATION	FOR FUTURE RESEARCH	
	_				
REFE	REN	CES			89
APPE	APPENDICES 109				109
BIOD			TIONS		153
LIST (UBLICA	TIONS		154

LIST OF TABLES

Table		Page
2.1	Total world and Malaysia production of banana during 2013 to 2017.	7
2.2	Previous studies that highlighted the symptom of agriculture crops attack by <i>Fusarium</i> fungi.	13
2.3	Previous studies that highlighted the effect of biological control products on <i>Fusarium</i> diseases.	15
2.4	Previous studies that highlighted the effect of neem products on different agricultural diseases and growth of various agriculture crops.	18
2.5	Previous studies that highlighted the effect of neem leaves extract on different pathogen and <i>Fusarium</i> fungus under <i>in-vitro</i> and <i>in-vivo</i> condition.	20
2.6	Previous studies that highlighted the effect of neem leaves extract on growth of various agriculture crops.	21
2.7	Previous studies that highlighted the effect of neem leaves on physiochemical properties of the soil.	22
2.8	Previous studies that highlighted the effect of media pH on <i>Fusarium</i> fungus under <i>in-vitro</i> and <i>in-vivo</i> condition.	24
2.9	Previous studies that highlighted the effect of media pH on growth of various agriculture crops and physiochemical properties of the soil.	26
3.1	Effect of different aqueous NLE rate per 250 ml PDA media on colony radial extension of Foc from 1st to 7th DAI.	33
3.2	Effect of different aqueous NLE rate per 250 ml PDA media on growth rate of Foc from 1st to 7th DAI.	34
3.3	Effect of different aqueous NLE rate per 250 ml PDA media on PIRG of Foc from 1st to 7th DAI.	35
3.4	Effect of different pH level of the PDA media on colony radial extension of Foc from 1st to 7th DAI.	37
3.5	Effect of different pH level of the PDA media on growth rate of Foc from 1st to 7th DAI.	38

6

3.6 Effect of different pH level of the PDA media on PIRG 39 of Foc from 1st to 7th DAI. 4.1 The leaf symptom index of Cavendish banana crops 52 4.2 The rhizome discoloration index of Cavendish 53 banana crops. 4.3 The leaf disease severity index, rhizome diseases 54 severity index and effectiveness of neem leaves extract treatment evaluation on banana plant. 4.4 Comparison of nutrient content of neem leaf. 46 4.5 Effect of aqueous NLE on Nutrient contents of 59 banana leaf of Foc uninoculated (Control +ve) and inoculated (Control -ve) Cavendish banana plants under optimum soil pH 7 condition. 4.6 Effect of aqueous NLE on root length, root area, root 60 diameter and root volume of Foc uninoculated (Control +ve) and inoculated (Control -ve) -Cavendish banana plants under optimum soil pH 7 condition. 4.7 61 Effect of aqueous NLE on shoot fresh weight, shoot dry weight, root fresh weight, root dry weight and root shoot ratio of Foc uninoculated (Control +ve) and inoculated (Control -ve)-Cavendish banana plants under optimum soil pH 7 condition. 4.8 Effect of aqueous NLE on photosynthesis rate, 62 stomata conductance, transpiration rate and vapour pressure deficit of Foc uninoculated (Control +ve) and inoculated (Control -ve) Cavendish banana plants under optimum soil pH 7 condition. 4.9 Effect of aqueous NLE on total chlorophyll content of 63 Foc uninoculated (Control +ve) and inoculated (Control -ve)-Cavendish banana plants under optimum soil pH 7 condition. 4.10 Effect of aqueous NLE on leaf symptom index of Foc 65 uninoculated (Control +ve) and inoculated (Control ve) Cavendish banana plants throughout 10 weeks after planting under optimum soil pH 7 condition. 4.11 Effect of aqueous NLE on leaf disease severity 66 index, effectiveness of neem leaves extract treatment on leaf of Foc uninoculated (Control +ve)

and inoculated (Control –ve)-Cavendish banana plants throughout 10 weeks after planting under optimum soil pH 7 condition.

- 4.12 Effect of aqueous NLE on leaf damage percentage of Foc uninoculated (Control +ve) and inoculated (Control –ve)-Cavendish banana plants throughout 10 weeks after planting under optimum soil pH 7 condition.
- 4.13 Effect of aqueous NLE on rhizome discoloration index, rhizome diseases severity index, effectiveness of neem leaves extract treatment on rhizome, rhizome wilting percentage and reduce in *Fusarium* wilt of Foc uninoculated (Control +ve) and inoculated (Control –ve)-Cavendish banana plants on week 10 after planting under optimum soil pH 7 condition.
- 4.14 Effect of aqueous NLE on microbial population of Foc uninoculated (Control +ve) and inoculated (Control –ve) Cavendish banana plants under optimum soil pH 7 condition.
- 4.15 Effect of aqueous NLE on soil pH and cation exchange capacity of Foc uninoculated (Control +ve) and inoculated (Control –ve) -Cavendish banana plants under optimum soil pH 7 condition.
- 4.16 Effect of aqueous NLE on soil exchangeable bases, available phosphorus (P) of Foc uninoculated (Control +ve) and inoculated (Control -ve) -Cavendish banana plants under optimum soil pH 7 condition.

69

67

71

70

72

LIST OF FIGURES

Figure		Page
2.1	Morphology of <i>Fusarium</i> fungus with white or purple cottony aerial mycelium surface.	10
2.2	Banana-shaped, multicellular macroconidia together with kidney-shaped, uni-or two-celled microconidia of <i>Fusarium</i> fungus.	11
2.3	The processes of Fusarium infection.	12
2.4	Disease triangle component	22
4.1	Effect of aqueous NLE on plant height (A), pseudo- stem diameter (B), total leaf number (C) and leaf area (D) of Foc uninoculated (Control +ve) and inoculated (Control –ve) Cavendish banana plants throughout 10 weeks after planting under optimum soil pH 7 condition. Mean values with the same letter (NLE treatment) are not significantly difference using LSD at P>0.05.	58
4.2	Effect of aqueous NLE on leaf symptom index of Foc uninoculated (Control +ve) and inoculated (Control –ve) Cavendish banana plants on week 10 after planting under optimum soil pH 7 condition. Note: T1 = Without NL:Foc uninoculated (Control +ve), T2 = With NL:Foc uninoculated (Control +ve), T3 = Without NL:Foc inoculated (Control -ve) and T4 = With NL:Foc inoculated (Control +ve).	64
4.3	Effect of aqueous NLE on stem (horizontal cut), stem (longitudinal cut) and rhizome visually symptom of Foc uninoculated (Control +ve) and inoculated (Control –ve) Cavendish banana plants on week 10 after planting under optimum soil pH 7 condition. Note: T1 = Without NL:Foc uninoculated (Control +ve), T2 = With NL:Foc uninoculated (Control +ve), T3 = Without NL:Foc inoculated (Control -ve) and T4 = With NL:Foc inoculated (Control +ve).	68

LIST OF ABBREVIATIONS

	\$	Dollar sign
	%	Percentage
	±	Plus-minus sign
	0.1N	0.1 Normality
	Α.	Azadirachta
	ANOVA	Analysis of Variance
	AA	Auto Analyzer
	AAS	Atomic Absorption Spectrometer
	ABA	Abscisic Acid
	A. Juss	Adrien-Henri de Jussieu (author abbreviation)
	BBMV	Banana Bract Mosaic Virus
	BBTV	Banana Bunchy Top Virus
	BIO	Bioorganic
	BSV	Banana Streak Disease
	CV.	Cultivar
	Са	Calcium
	CEC	Cation Exchange Capacity
	CFU	Colony Forming Unit
	cm	Centimetre
	cmol	Centimole
	CMV	Cucumber Mosaic Virus
	CRD	Complete Randomized Design
	Cu	Copper
	CZA	Czapek-Dox Agar

°C	Degree celcius
DAI	Days After Incubation
DF	Degree of Freedom
DOA	Department of Agriculture
DSI	Diseases Severity Index
EC	Electrical conductivity
EID	Emerging infectious disease
et al.	And friends
F.	Fusarium
FAO	Food and Agriculture Organization
FAOSTAT	Food and Agriculture Organization Statistical Database
Fe	Iron
Foc	Fusarium oxysporum f.sp. cubense
f. sp.	Forma specialis
g	Gram
GA3	Gibberellic Acid
GLM	General Linear Models
GML	Ground Magnesium Limestone
H+	Hydrogen Ion.
ha	Hectare
HCI	Hydrocholoric Acid
H2O2	Hydrogen Peroxide
HPLC	High Performance Liquid Chromatographic
HSD H2SO4	High Significant Different Sulfuric Acid
IAA	Indole-3-acetic acid

xix

	Im	Immune
	IU	International unit
	kg	Kilogram
	К	Potassium
	kPa	Kilopascals
	K2SO4	Potassium Sulphate
	L	Litre
	LSD	Least Significant Different
	LSI	Leaf Symptom Index
	М	Molarity
	m	Meter
	mm	Millimeter
	Mardi	Malaysian Agricultural Research and Development Institute
	Mg	Magnesium
	mg	Milligram
	ml	Millilitre
	MMEA	Maize Meal Extract Agar
	Mn	Manganese
	Mr	Moderately resistance
	Ms	Moderately susceptible
	μ	Micro
	Ν	Nitrogen
	nm ns	Nanometre Not significant
	NaOCI	Sodium Hypochlorite
	NaOH	Sodium Hydroxide

NAP	National Agriculture Policy
NH4F	Ammonium Fluoride
NH4OAc	Ammonium Acetate
NL	Neem Leaves
NLE	Neem Leaves Extract
NaOCI	Sodium Hypochlorite
Р	Phosphorus
PDA	Potato Dextrose Agar
PDB	Potato Dextrose Broth
рН	Measurement of Acidity / Alkalinity
PIRG	Percentage of Inhibition of Radial Growth
RDI	Rhizome Discoloration Index
RCBD	Randomized complete block design
RM	Ringgit Malaysia
Rs	Resistance
R/S	Root/shoot ratio
s	Second
SAS	Statistical analysis system
Sc	Susceptible
Si	Silicon
sp.	Species (single)
spp. SSR	Species (plural) Self-sufficiency ratio
st	-first
TR4	Tropical Race 4
th	Suffixes (other than -first, -second and -third)
	NAP NH4F NH4OAc NL NLE NaOCI P PDA PDB PDB PH PDB PIRG RDI RCBD RM RS R/S SAS SAS SAS SAS SSR SAS SSR Si sp. SSR sp. SSR st TR4

xxi

- UPM Universiti Putra Malaysia
- USD United states dollar
- Vs Very susceptible
- v/v Volume/Volume
- Zn Zinc



CHAPTER 1

INTRODUCTION

Banana (*Musa spp.*) also known as Pisang in Malaysia is one of the most wellknown fruits in the world. However, this world's most popular fruit might soon go extinct due to outbreaks of catastrophic wilt diseases induced by *Fusarium oxysporum* f. sp. *cubense* (Foc). *Fusarium* wilt disease symptoms come in various forms such as yellowing of the leaf, wilting of banana trees, discoloration of the vascular system. The lethal strain of Foc led to huge banana production losses all over the world especially Cavendish plantations in China, Indonesia, Philippines including Malaysia and causing heavy economic losses of over 75 million USD (around RM 300 million) mark. According to Masdek *et al.* (2003) and Nasdir (2003) this destructive fungus destroying more than 5,000 ha of banana annually while losses in banana plantations of both Malaysia and Indonesia country exceed 8 million of plants. Disease's control is thus urgent to eliminating this pathogenic-fungus and prevent the spread of disease.

The suppressive Fusarium fungus is not effective by chemical (Stover, 1962, Lakshmanan et al., 1987; Herbert and Marx, 1990) and biological control strategies. Preventive means with disease-resistant cultivars and disease-free propagation material is the best strategy (Saravanan et al. 2003; Cao et al. 2005; Nel et al. 2007; Lian et al. 2009). However, efforts to develop effective, reliable and environmentally friendly biocontrol strategies should not be neglected as it remains an important organic strategy against crop diseases. Numerous biological control technique such as inoculating beneficial microbial organisms (Alabouvette, 1986; Larkin et al., 1996; Larkin and Fravel, 1998; Ploetz et al., 2003), bioorganic product application (Runia, 2014; Xue et al., 2015) and soil amendment (Nasir et al., 2003) have been reported to shows slight suppression in controlling Fusarium diseases on different agriculture crops. At present, there are still very few studies that have focused on the antifungal activities of both Malaysia neem extract and Malaysia land condition against the Fusarium pathogen, especially on local Cavendish banana crops.

The researcher of Malaysia has given high attention and priority to the neem tree (*Azadirachta indica* A.Juss.) powerful products. Rich nutrient content in neem leaves extracts (NLE) improving fertility of soil and plants' nutrients uptake. Meanwhile, this extract also helps in controlling several parasitic fungi due to their active ingredient, azadirachtin which responsible for stopping the growth cycle of microorganism without any side effect on crops. However, under favorable environmental conditions, fungal spores still germinate and spread easily in the agriculture field even in the present of the fungicide.

Host, pathogen, and environment in the "disease triangle", are the three components that determined the potential risk of an agriculture crop in Emerging Infectious Disease (EID). Thus, changing the media pH level can suppress the

pathogen directly or indirectly through impact on this soil pathogen activity and on host colonization by converting the disease-conducive media to diseasesuppressive media (Kloepper *et al.*, 1980). Thus, the pH level that optimum for the agriculture crops but not for the disease's pathogens should be chosen as it could make the crops strong enough to resist the pathogens. However, while the general principles are theorized, there is a lack of detailed scientific knowledge and prove especially on Cavendish plants in Malaysia under both laboratory and field performance.

Currently, the rapid spread of *Fusarium* diseases in Malaysia had caused serious economic losses problem in local banana agriculture (Husain and William, 2011). Besides that, acidic topsoil (pH 4.2 to pH 4.8) in most of the land areas in Malaysia, may even result in more serious *Fusarium* diseases incident (Gordon *et al.*, 2019; Orr and Nelson, 2018). Acidic soils also cause significant losses in banana production as it does not suitable for crops growth and were low in disease resistance. Thus, in view of the above problems, the main objective of this study was to systematically study the effects of different aqueous neem leaves extract rate (NLE) and different pH level of media in controlling *Fusarium* (Foc) growth, soil physicochemical properties, growth and physiological changes of Cavendish banana. The specific objectives of the present study were:

- i. To study the effects of different aqueous NLE rate applications on the *Fusarium* growth under *in-vitro* condition.
- ii. To determine the effects of different pH levels of growth media on the *Fusarium* growth under *in-vitro* condition.
- iii. To determine the effect of aqueous NLE on controlling *Fusarium* wilt, soil physicochemical properties, growth performance and physiology changes of Cavendish banana under optimized soil pH condition.

REFERENCES

- Abdullah Shukor, A.R., Jamaluddin, S.H., Nik Masdek, N.H., Zabedah, M., Abdullah, H., 2000. Malaysian banana industry-Prospects and challengers. In: Proceedings of the international workshop on the banana *Fusarium* wilt disease (Molina A.B., Nik Masdek, N. H. and Liew, K. W., eds.). Genting Highlands, Malaysia, 18-20 October 1999. INIBAP and MARDI. 32-38.
- Abubakar, A., Suberu, H.A., Bello, I.M., Abdulkadir, R., Daudu, O.A., Lateef, A.A., 2013. Effect of pH on mycelial growth and sporulation of *Aspergillus para siticus*. Journal of Plant Sciences (1)4: 64-67.
- Agbenin, N.O., Emechebe, A. M and Marley, P.S., 2004. Evaluation of neem seed powder for *Fusarium* wilt and Meloidogyne control on tomato. Archives of Phytopathology and Plant Protection 37(4): 319-326.
- Agrios, G.N., 2006. Plant Pathology. Academic Press, San Diego, USA.
- Ahlem, H., Mohammed, E., Badoc, A. and Ahmed, L., 2012. Effect of pH, temperature and water activity on the inhibition of *Botrytis cinerea* by *Bacillus amyloliquefaciens* isolates. African Journal of Biotechnology 11(9): 2210-2217.
- Aini, Z., 2006. Soil-enhancing technologies for improving crop productivity in malaysia and considerations for their use [online]. Available from: https://www.fftc.org.tw/ [Accessed on 27 August 2020]
- Aini, Z., Sivapragasam, A., Vimala, P. and Mohamad Roff, M.N., 2005. Organic vegetable cultivationin Malaysia. Kuala Lumpur: Malaysian Agricultural Research and Development Institute (MARDI) pp 6-130.
- Ahmad, M.S., Mukhtar, T, Ahmad, R, 2004. Some studies on the control of citrus nematode (*Tylenchulus semipenetrans*) by leaf extracts of three plants and their effects on plant growth variables. Asian Journal of Plant Sciences 3(5): 544-548.
- Akila, R., Rajendran, L., Harish, S., Saveetha, K., Raguchander, T., and Samiyappan, R., 2011. Combined application of botanical formulations and biocontrol agents for the management of *Fusarium oxysporum* f. sp. *cubense* (Foc) causing *Fusarium* wilt in banana. Biological control 57: 175-183.
- Akhtar, M., 1999a. Biological control of plant-parasitic nematodes in pigeon pea filed crops using neem-based products and manurial treatments. Applied Soil Ecology 12: 191-195.
- Akhtar, M., 1999b. Plant growth and nematode dynamics in response to soil amendments with neem products, urea and compost. Bioresource Technology 69: 181–183.

- Alabouvette, C., 1986. *Fusarium*-wilt suppressive soils from the Châteaurenard region: review of 10-years study. Agronomie 6: 273–284.
- Alabouvette, C., Lemanceau, P. and Steinberg, C., 1993. Recent advances in the biological control of *Fusarium* wilts. Pesticide Science 37: 363–373.
- Al-Hazmi, R.H.M. 2013. Effect of neem (*Azadirachta indica*) leaves and seeds extract on the growth of six of the plant diseases causing fungi. Global Advanced Research Journal of Microbiology. 2: 89–98.
- Alhussaen, K.M., 2012. Effect of soil acidity on diseases caused by *Pythium ultimum* and *Fusarium oxysporum* on tomato plants. Journal of Biological Sciences 12 (7): 416-420.
- Ali, A.S.M.Y., Solaiman, A.H.M., and Saha, K.C., 2016. Influence of organic nutrient sources and neem (*Azadirachta*) products on growth and yield of carrot. International Journal of Crop Science and Technology 2(1): 19-25.
- Amadioha, A.C. and Obi, V.I., 1998. Fungitoxic activity of extracts from *Azadirachta indica* and *Xylopia aethiopica* on *Colletotrichum lindemuthianum* in cowpea. Journal of Herbs, Spices and Medicinal Plants 6: 33-40.
- Amadioha, A.C., 2000. Controlling rice blast in vitro and in vivo with extracts of Azadirachta indica. Crop Protection 19 (5): 287-290.
- Ansari, J., Sohail, H.K., Ulhaq, A. and Yousaf, M., 2012. Effects of the level of *Azadirachta indica* dried leaf meal as phytogenic feed additive on the growth performance and haemato-biochemical parameters in broiler chicks. Journal of Applied Animal Research 40(4): 336-345.
- Arumugam, P.A., Mohamad, I., Salim, R. and Mohamed, Z., 2015. Antifungal effect of Malaysian neem leaf extract on selected fungal species causing otomycosis in *in-vitro* culture medium. Malaysian Journal of Medicine and Health Sciences (ISSN 1675-8544) 11(2): 69-84.
- Baker, N.R., Hardwick, K., 1973. Biochemical and physiological aspects of leaf development in cocoa (*Theobroma cacao*). I. Development of chlorophyll and photosynthetic activity. New Phytologist 72: 1315-1324.
- Beckman, C.H. and Roberts, E.M., 1995. On the nature and genetic basis for resistance and tolerance of fungal wilt diseases. Advances in Botanical Research 21: 35-77.
- Bhatnagar, D., 1988. The inhibitory effect of neem (*Azadirachta indica*) leaf extracts on aflatoxin synthesis in *Aspergillus parasiticus*. Journal of the American Oil Chemists' Society 65(7):1166-1168.

- Bhattarai A, Bhattarai B, Pandey S, 2015. Variation of soil microbial population in different soil horizons. Journal of Microbiology and Experimentation 2(2):75-78.
- Bhende, S.S. and Kurien, S., 2015. Sucker production in banana. Journal of Tropical Agriculture 53(2): 97-106.
- Bhowmik, D., Chiranjib, Yadav., J., Tripathi., K.K. and Kumar, K.P.S., 2010. Herbal remedies of *Azadirachta indica* and its medicinal application. Journal of Chemical and Pharmaceutical Research 2(1): 62-72.
- Bishop, C.D. and Cooper, R.M., 1983a. An ultrastructural study of root invasion of three vascular wilt diseases. Physiological Molecular Plant Pathology 22: 15-27.
- Black, G.R. and Hartge, K.H., 1986. Bulk density. In: A. Klute (Ed.). Methods of Soil Analysis. Part I. Physical and Mineralogical Methods. 2nd. Ed., Agronomy No. 9 (part I). ASA-SSSA. Madison, Wisconsin, USA, 363-375.
- Bohra, B., Vyas, B. N. and Mistry K. B., 2006. Eco-friendly management of damping-off in winter vegetables and tobacco using microbial agents and neem for mulations. Journal of Mycology and Plant Pathology 36: 178-181.
- Brahmachari G., 2004. Neem-An Omnipotent Plant: A Retrospection. Chem Bio Chem (Wiley-VCH) 5: 408-421.
- Bray, R.H. and Kurtz, L.T., 1945. Determination of total, organic, and available forms of phosphorus is soils. Soil Science 59:39-45.
- Butler, D., 2013. Fungus threatens top banana. Nature 504: 195-196.
- Cachinero, J.M., Hervas, A., Jimenez-Diaz, R.M., Tena, M., 2002. Plant defence reactions against *Fusarium* wilt in chickpea induced by incompatible race 0 of *Fusarium oxysporum* f.sp. *ciceris* and non-host isolates of *Fusarium oxysporum*. Plant Pathology 51: 765-776.
- Cao, L.X., Qiu, Z.Q., You, J.L., Tan, H.M. and Zhou, S.N., 2005. Isolation and characterization of endophytic streptomycete antagonists of *Fusarium* wilt pathogen from surface-sterilized banana roots. FEMS Microbiology Letters 247: 147-152.
- Caracuel, Z., Roncero, M.I., Espeso, E.A., González-Verdejo, C.I., García-Maceira, F.I. and Di Pietro, A., 2003. The pH signalling transcription factor PacC controls virulence in the plant pathogen *Fusarium oxysporum*. Molecular Microbiology 48(3): 765 – 779.
- Casey, N.G., 2015. A survey of cultivated bananas in Perak. UTAR Agriculture Science Journal 1(1): 13-21.

- Cha, S.D., Jeon, Y.J., Ahn, G.R., Han, J.I., Han, K.H and Kim, S.H., 2007. Characterization of *Fusarium oxysporum* isolated from Paprika in Korea. Mycobiology 35(2): 91-96.
- Chen, L.H., Huang, X.Q., Yang, X.M. and Shen, Q.R., 2013a. Modeling the effects of environmental factors on the population of *Fusarium oxysporum* in cucumber continuously cropped soil. Communications in Soil Science and Plant Analysis 44 (15): 2219-2232.
- Chen, Y.F., Chen, W., Huang, X., Hu, X., Zhao, J.T., Gong, Q., Li, X.J. and Huang, X.L., 2013b. *Fusarium* wilt-resistant lines of Brazil banana (*Musa* spp. AAA) obtained by EMS-induced mutation in a micro-crosssection cultural system. Plant Pathology 62: 112-119.
- CITYFARM, 2020. Green Eagle Alterdew F11 (Bio-Fungicide) [online]. Available from: https://cityfarm.my/ [27 August 2020].
- Clark, R.B. and Zeto, S.K., 1996. Mineral acquisition by mycorrhizal maize grown on acid and alkaline soil. Soil Biology and Biochemistry (28)10-11: 1495-1503.
- Cook, R.J. and Baker, K.F., 1983. The Nature and Practice of Biological Control of Plant Pathogens. APS Press, St. Paul, MN.
- Coombs, J., Hind, G., Leegood, R.C., Tieszen, L.L. and Vonshak, A., 1985. Analytical Techniques. In: Techniques in Bioproductivity and photosynthesis 2nd edition. (Eds) J. Coombs, D.O. Hall, S.P. Long and J.M.O. Scurlock. pp. 219-220, Pergamon Press.
- Daniells, J. 1995. Illustrated guide to the identification of banana varieties in the South Pacific. Australian Centre for International Agricultural Research (ACIAR). Australia, Canberra: CPN Publications.
- Deacon, J.W., 1984. Panama disease of banana in South Africa. Horticultural Science 1: 29–31.
- Depieril, R.A., Martinez, S.S. and Menezes Jr.A.O., 2005. Compatibility of the fungus *Beauveria Bassiana* (Bals.) Vuill. (Deuteromycetes) with extracts of neem seeds and leaves and the emulsible oil. Neotropical Entomology 34(4): 1519-1566.
- Dita, M.A., Waalwijk, C., Paiva, L.V., Souza Jr., M.T. and Kema, G. H.J., 2011. A greenhouse bioassay for the *Fusarium oxysporum* f. sp. *cubense* x 'Grand Nain' (*Musa*, AAA, Cavendish Subgroup) Interaction. Acta horticulturae: 897(897).
- Dugdale, L.J., Mortimer, A.M., Isaac, S. and Collin, H.A., 2000. Disease response of carrot and carrot somaclones to *Alternaria dauci*. Plant Pathology 49: 57-67.

- Egunjobi, O.A. and Afolami, S.O., 1976. Effects of neem (*Azadirachta indica*) leaf extracts on populations of *Pratylenchus Brachyurus* and on the growth ad yield of maize. Nematologica 22:125-132.
- Ekdahl, I., 1957. The growth of root hairs and roots in nutrient media and distilled water and the effects of oxylate. Lantbrukshoegsk Ann 23: 497-518.
- El-Khallal, S.M., 2007. Induction and modulation of resistance in tomato plants against *Fusarium* wilt disease by bioagent fungi (*Arbuscular mycorrhiza*) and/or hormonal elicitors (jasmonic acid & salicylic acid): 2-changes in the antioxidant enzymes, phenolic compounds and pathogen related-proteins. Australian Journal of Basic and Applied Science 1 (4): 717–732.
- Embayeab, K. Weiha, M., Ledinc, S. and Christersson, L., 2005. Biomass and nutrient distribution in a highland bamboo forest in southwest Ethiopia: implications for management. Forest Ecology and Management 204 (2-3): 159-169.
- Emmanuel I.M.J., 2012. Comparative evaluation of modified neem leaf, neem leaf and woodash extracts on soil fertility improvement, growth and yields of maize (*Zea mays* L.) and watermelon (*Citrullus lanatus*) (sole and intercrop). Agricultural Sciences 3(1): 90-97.
- Emmanuel, I.M.J., 2014. Effects of water extracts of neem (*Azadirachta indica* L.) leaf, wood ash and their mixture on soil chemical composition and growth and yield of plantain (*Musa sapientum* L.). American Journal of Experimental Agriculture 4(7): 836-848.
- Epp, M.D., 1987. Somaclonal variation in bananas: a case study with *Fusarium* wilt. Banana and Plantain Strategies. ACIAR Proceedings 21: 140-150.
- Ezeonu, C.S., Imo, C. and Agwaranze, D., Iruka, A. and Joseph, A., 2018. Antifungal effect of aqueous and ethanolic extracts of neem leaves, stem bark and seeds on fungal rot diseases of yam and cocoyam. Chemical and Biological Technologies in Agriculture 5(18): 1-9.
- Fageria, N.K. and Baligar V.C., 2005. Nutrient availability. Encyclopedia of Soils in the Environment (D. Hillel, Ed.). Elsevier, San Diego, CA, 63-71.
- Fageria, N.K., Baligar, V.C. and Clark, R.B., 2002. Micronutrients in crop production. Advances in Agronomy. 77: 185–268.
- FAMA, n.d.. Laporan Warta Barangan [online]. Available from: https://sdvi.fama.gov.my/ [Accessed on 12 May 2019].
- FAO, 2003. The world banana economy 1985-2002. Rome: Economic and Social Department, Food and Agriculture Organization.

- FAO, 2017. Food and agriculture organization of the united nation, Banana facts and figures [online]. Available from: http://www.fao.org [Accessed on 13 Jun 2017].
- FAO, 2019. World banana forum, *Fusarium* TR4 [online]. Available from: http://www.fao.org [Accessed on 24 August 2019].
- FAOSTAT, 2019. FAOSTAT. 2019 [online]. Available from: http://faostat.fao.org/ [Accessed on 12 May 2019].
- Foy, C.D., 1984. Physiological effects of hydrogen, aluminium and manganese toxicities in acid soil. In: Pearson RW, Adams F, eds. Soil acidity and liming, 2nd edn. Wisconsin: American Society of Agronomy pp 57-97.
- Farag Hanaa, R.M., Abdou, Z.A., Salama, D.A., Ibrahim, M.A.R., Sror, H.A.M., 2011. Effect of neem and willow aqueous extracts on *Fusarium* wilt disease in tomato seedlings: Induction of antioxidant defensive enzymes. Annals of Agricultural Sciences (56)1: 1-7.
- Francis S.P.N., 2015. A brief history of bananas. UTAR Agriculture Science Journal 1(1): 3-9.
- Frison, E.A. and Sharrock, S.L., 1999. The economic, nutritional and social importance of bananas in the world, Bananas and Food Security, PICQ C., et al. Eds. Montpellier: INIBAP.
- Fuchs, J.G., Moënne-Loccoz, Y. and Défago, G., 1997. Nonpathogenic *Fusarium oxysporum* strain Fo47 induces resistance to *Fusarium* wilt in tomato. Plant Disease 81(5): 492-496.
- Garcia, F.A., Ordonez, N., Konkol, J., Al Qasem, M., Naser, Z., Abdelwali, M., Salem, N.M., Waalwijk, C., Ploetz, R.C. and Kema, G., 2014. First report of *Fusarium oxysporum* f. sp. *cubense* tropical race 4 associated with Panama disease of banana outside Southeast Asia. Plant Disease 98:694.
- Garrigues, E., Doussan, C. and Pierret, A., 2006. Water uptake by plant roots: I–Formation and propagation of a water extraction front in mature root systems as evidenced by 2D light transmission imaging. Plant and Soil. 283(1-2): 83.
- Ghini, R., Fortes, N.L.P., Navas-Cortés, J.A., Silva, C.A. and Bettiol, W., 2016. Combined effects of soil biotic and abiotic factors, influenced by sewage sludge incorporation, on the incidence of corn stalk rot. PloS one 11(5): e0155536.
- Girish, K., Shankara Bhat, S., 2008. Neem A Green Treasure. Electronic Journal of Biology 4(3): 102-111.

- Goh, Y.K., Choon, K.L., Cheng, C.R., Tan, S.Y., Cheah, L.W., Ah Tung, P.G., Goh, K.Y. and Goh, K.J., 2017. Effects of chemical properties of different soils on *Ganoderma* disease in oil palm (*Elaeis guineensis*). Oil Palm Bulletin 75: 17-26.
- Goicoechea, N., Aguirreolea, J., Cenoz, S. and Garcia, M.J.M., 2001. Gas exchange and flowering in Verticillium-wilted pepper plants. Journal of Phytopathology 149(5): 281-286.
- Gonçalves, A.L. and Kernaghan, J.R., 2014. Banana production methods. A comparative study p1-40 [online]. Available from: https://www.naturskyddsforeningen.se [Accessed on 5 May 2019]
- Gordon, T.R., Stueven, M., Pastrana, A.M., Henry, P.M., Dennehy, C.M., Kirkpatrick, S.C. and Daugovish, O., 2019. The effect of pH on spore germination, growth, and infection of strawberry roots by *Fusarium oxysporum* f. sp. *fragariae*, cause of *Fusarium* wilt of strawberry. The American Phytopathological Society 103(4): 697-704.
- Haas, D. and Défago, G., 2005. Biological control of soil-borne pathogens by fluorescent pseudomonads. Nature Reviews Microbiology 3(4):307-319.
- Hadian, S., Rahnama, K., Jamali, S. and Eskandari, A., 2011. Comparing Neem extract with chemical control on *Fusarium oxysporum* and *Meloidogyne incognita* complex of tomato. Advances in Environmental Biology 5(8): 2052-2057.
- Harris, R.W., 1992. Root-shoot ratios. Journal of Arboriculture 18(1):39-42.
- Hassan, N.M.M., 2004. Banana R & D in Malaysia: update and highlights. In: Advancing Banana and Plantain R & D in Asia and the Pacific Vol. 12, edited by Molina A.B., Eusebio J.E., Roa V.N., Van Den Bergh I. & Maghuyop M.A.G. Jakarta: INIBAP.
- Hayman, X.V. and Tavares, M., 1985. Plant growth responses to vesicular *Arbuscular mycorrhiza* xv. influence of soil pH on the symbiotic efficiency of different endophytes. New Phytologist 100: 367-377.
- Herbert, J.A. and Marx, D., 1990. Short-term control of Panama disease in South Africa. Phytophylactica 22: 339–340.
- Heslop-Harisson, J.S. and Schawarzacher, T., 2007. Domestication, genomics and the future of banana. Annals of Botany 100: 1073-1084.
- Hoper, H., Steinberg, G. and Alabouvette, C., 1995. Involvement of clay type and pH in mechanisms of soil suppressiveness *Fusarium* wilt of flax. Soil Biology and Biochemistry 27(7): 955-967.

- Hossain, M.A., Al-Toubi, W.A.S., Weli, A.M., Al-Riyami, Q.A., Al-Sabahi, J.N., 2013. Identification and characterization of chemical compounds in different crude extracts from leaves of Omani neem. Journal of Taibah University for Science. 7: 181–188.
- Howard, D.D. and Adams, F., 1965. Calcium Requirement for Penetration of Subsoils by Primary Cotton Roots. Soil Science Society of America 29(5): 558-562.
- Huang, Y.H., Wang, R.C., Li, C.H., Zuo, C.W., Wei, Y.R., Zhang, L. and Yi, G.J., 2012. Control of *Fusarium* wilt in banana with Chinese leek. European Journal of Plant Pathology 134(1): 87-95.
- Husain M. and William R., 2011. Status of banana cultivation and disease incidences in Malaysia. Crop Protection and Plant Quarantine Division Department of Agriculture [online]. Available from: http://www.itfnet.org [Accessed on 13 Jun 2017].
- Islam, A.K.M.S., Edwards, D.G. and Asher, C.J., 1980. pH optima for crop growth: Results of a flowing nutrient culture experiment with six species. Plant Soil 54:339–357.
- Jacobson M., 1986. Pharmacological and toxicological effects of neem and chinaberry on warm-blooded animals. Neem newsl 3:39-43.
- Jaizme-Vega, M. C., Sosa Hernandez, B. and Hernandez Hernandez, J. M., 1998. Interaction of *Arbuscular mycorrhizal* fungi and the soil pathogen *Fusarium oxysporum* f. sp. *cubense* on the first stages of micropropagated Grande Naine banana. Acta Horticulturae 490: 285-295.
- Jonathan, H.C. and Carlos, F.B., 2016. banana growing in the florida home landscape. florida cooperative extensionservice, Electronic Digital International Source (EDIS), HS10. [online]. Available from: https://hos.ifas.ufl.edu [Accessed on 12 May 2019]
- Jones, L.R., 1924. The relation of environment to disease in plants. American Journal of Botany11: 601-609.
- Joseph, B., Dar, M.A. and Kumar, V., 2008. Bioefficacy of plant extracts to control *Fusarium solani* f. sp. *Melongenae incitant* of brinjal wilt. Global Journal of Biotechnology and Biochemistry 3 (2): 56-59.
- Jung, B., Lee, S., Ha, J., Park, J. C., Han, S. S., Hwang, I., Lee, Y. W. and Lee, J., 2013. Development of a selective medium for the fungal pathogen *Fusarium graminearum* using toxoflavin produced by the bacterial pathogen *Burkholderia glumae*. Plant Pathology 29(4): 446-450.

- Kaiser, C., van der Merwe, R., Bekker, T.F. and Labuschagne, N., 2005. *In-vitro* inhibition of mycelial growth of several phytopathogenic fungi, including *Phytophthora cinnamomi* by soluble silicon. South African Avocado Growers' Association Yearbook 28.
- Kapadiya, I.B., Undhad, S.V., Talaviya, J.R. and Siddhapara, M.R., 2014. Evaluation of phytoextracts against *Fusarium solani* causing root rot of okra. Journal of Biopesticides 7 (Supp.): 7-9.
- Karim Rezaul, S.M, Noorjanna Rahmatullah, Mariam Firdaus Mad Nordin and Ataul Karim Rajin, S.M., 2018. Effect of stage of maturity and frying time on the quality of banana springs. Pertanika Journal of Tropical Agricultural Science 41 (3): 1097-1110.
- Kartika, K., Lakitan, B., Sanjaya, N., Wijaya, A., Kadir, S., Kurnianingsih, A., Widuri, L.I., Siaga, E. and Meihana, M., 2018. Internal versus edge row comparison in jajar legowo 4: 1 rice planting pattern at different frequency of fertilizer applications. AGRIVITA, Journal of Agricultural Science 40(2): 222-232.
- Kasai, M. and Takahashi, W., 2013. interaction of photosynthetic source-sink balance and activities of membrane H+ pumps in soybean. A comprehensive survey of international soybean research - Genetics, Physiology, Agronomy and Nitrogen relationships. James E. Board. Rijeka, Croatia: IntechOpen.
- Kerridge, P.C., 1969. Aluminum toxicity in wheat. Ph. D. thesis. Oregon State University, Corvallis, OR. 170 p.
- Khazanah Research Institute, 2019. Banana: The world's most popular fruit. Kuala Lumpur: Khazanah Research Institute. License: Creative Commons Attribution CC BY 3.0.
- Kidd, P.S. and Proctor, J., 2001. Why plants grow poorly on very acid soils: are ecologists missing the obvious? Journal of Experimental Botany 52 (357): 791-799.
- Kloepper, J.W., Leong, J., Teintze, M. and Schroth, M.N., 1980. *Pseudomonas* siderophores: a mechanism explaining disease-suppressive soils. Current Microbiology 4: 317-320.
- Kocić-Tanackov, S., Dimić, G., Lević, J., Tanackov, I., and Tuco, D., 2011. Antifungal activities of basil (*Ocimum basilicum* L.) extract on *Fusarium* species. African Journal of Biotechnology 10(50): 10188-10195.
- Kumar D. and Pandey V., 2010. Relationship of pseudostem cross-sectional area with bunch weight, fruit quality and nutrient status in banana cv. *Rasthali* (Pathkapoora-AAB). Indian Journal of Horticulture 67(1): 26-29.

- Kumar, A., Zacharia, S., Maurya, A. K. and John, V., 2019. Effect of fungicides and neem oil on the rhizoctonia root rot of soybean (*Glycine max* L.). International Journal of Current Microbiology and Applied Sciences 8(1): 368-372.
- Lakshmanan, P., Selvaraj, P. and Mohan, S., 1987. Efficiency of different methods for the control of Panama disease. Tropical Pest Management 33:373–376.
- Larkin, R.P. and Fravel, D.R., 1998. Efficacy of various fungal and bacterial biocontrol organisms for the control of *Fusarium* wilt of tomato. Plant Disease 82: 1022–1118.
- Larkin, R.P., Hopkins, D.L. and Martin, F.N., 1996. Suppression of *Fusarium* wilt of watermelon by non-pathogenic *Fusarium oxysporum* and other microorganisms recovered from a disease-suppressive soil. Phytopathology 86: 812–819.
- Lemanceau, P. and Alabouvette, C., 1991. Biological control of *Fusarium* diseases by fluorescent Pseudomonas and non-pathogenic *Fusarium*. Crop Protection 10: 279-286.
- Li, T.T., Wu, Q.X., Wang, Y., John, A., Qu, H.X., Gong, L., Duan, X.W., Zhu, H., Yun, Z. and Jiang, Y.M., 2017. Application of proteomics for the investigation of the effect of initial ph on pathogenic mechanisms of *Fusarium proliferatum* on banana fruit. Frontiers in Microbiology 8: 23-27.
- Lian, J., Wang, Z. F., Cao, L.X., Tan, H.M., Inderbitzin, P., Jiang, Z.D. and Shining, Z., 2009. Artificial inoculation of banana tissue culture plantlets with indigenous endophytes originally derived from native banana plants. Biological Control 51: 427-434.
- Lokanadhan, S., Muthukrishnan, P. and Jeyaraman, S., 2012. Neem products and their agricultural applications. Journal of Biopesticides 5: 72–76.
- Loke, W.H., Heng, C.K., Azman Rejab and Norlaili Basirun, 1990. Studies on neem (*Azadirachta indica* A. Juss) in Malaysia [online]. Available from: http://agris.upm.edu.my [Accessed on 27 August 2020]
- Lopez, A. and Espinosa, J. 1998. Banana response to potassium. Better Response International 12(1): 1-5.
- Mak, C., Mohamed, A.A., Liew, K.W., Ho, Y.W., 2004. Early screening technique for *Fusarium* wilt resistance in banana micropropagated plants. Banana Improvement 18: 219-227.
- Mane, P.N., Sakhare, S. B., Gaikwad, M., Ghatol, P.U., Fatak, S. and Wankhade, N. J., 2018. Comparative effect of Pseudomonas fluorescens, neem seed karnel extract and fungicide against powdery mildew of sunflower. Journal of Plant Disease Sciences 13(2): 103-106.

- Marin, S., Sanchis, V. and Magan, G., 1995. Water activity, temperature, and pH effects on growth of *Fusarium moniliforme* and *Fusarium proliferatum* isolates from maize. Canadian Journal of Microbiology 41: 1063-1070.
- Marriott, J. and Palmer, J.K., 2009. Bananas physiology and biochemistry of storage and ripening for optimum quality. C R C Critical Reviews in Food Science and Nutrition 13(1): 41-88.
- Marschner, H., 1995. Mineral Nutrition of Higher Plants, 2nd edn. Academic Press, London.
- Masdek, N., Mahmood, M., Molina, A., Hwang, S.C., Dimyati, A., Tangaveli, R. and Omar, I., 2003. Global significance of *Fusarium* wilt: Asia. Abstracts of Papers 2nd International Symposium on *Fusarium* wilt on banana. PROMUSAINIBAP/EMBRAPA. Salvador de Bahía: Brazil 22 - 26 Sept.
- McCauley, A., Jones, C. and Jacobsen, J., 2009. Soil pH and organic matter. Nutrient management module 8: 1-12.
- McLean, E.O., 1982. Soil pH and lime requirement. In Page, A. L., R. H. Miller and D. R. Keeney (eds.) Methods of soil analysis. Part 2 - Chemical and microbiological properties. (2nd Ed.). Agronomy 9:199-223.
- Medina, E., Cuevas, E., Figueroa, J. and Lugo, A.E., 1994. Mineral content of leaves from trees growing on serpentine soils under contrasting rainfall regimes in Puerto Rico. Plant and Soil 158(1): 13-21.
- Merhej, J., Boutigny, A., Pinson-Gadais, L., Richard-Forget F. and Barreau, C., 2010. Acidic pH as a determinant of TRI gene expression and trichothecene B biosynthesis in *Fusarium graminearum*. Food additives and contaminants. Part A, Chemistry, analysis, control, exposure and risk assessment 27: 710-717.
- MacHardy, W.E. and Beckman, C.H., 1981. Vascular wilt Fusaria: Infections and Pathogenesis. In *Fusarium*: Diseases, Biology and Taxonomy, Nelson, P.E., Toussoun, T.A. and Cook, R.J. edition: 365-390. The Pennysylvania State University. Press, University Park and London.
- Michielse, C.B. and Rep, M., 2009. Pathogen profile update: *Fusarium* oxysporum. Molecular Plant Pathology 10: 311–324.
- Milsum, J.N., 2015. Malayan banana varieties in 1919. UTAR Agriculture Science Journal 1(1): 10-12.
- Minotta, G. and Pinzauti, S., 1996. Effects of light and soil fertility on growth, leaf chlorophyll content and nutrient use efficiency of beech (*Fagus sylvatica* L.) seedlings. Forest Ecology and Management 86(1-3): 61-71.
- Mohamad Roff, M.N., Tengku Abdul Malik T.M. and Sharif, H., 2012. Challenges to banana production in Malaysia: A threat to food security. The Planter 88: 13-21.

- Mohammed, A.A., Mak, C., Liew, K.W. and Ho, Y.W, 1999. Early evaluation of banana plants at nursery stage for *Fusarium* wilt tolerance. 174-186. In Banana *Fusarium* wilt management: toward sustainable cultivarion. Proceedings of the International workshop on banana *Fusarium* wilt disease, Malaysia 18 - 20 October 1999.
- Mondal, S.N., and Timmer, L.W. 2003. Effect of urea, CaCO₃, and dolomite on pseudothecial development and ascospore production of *Mycosphaerella citri*. Plant Disease 87:478-483.
- Mondali, N.K., Mojumdar, A., Chatterje, S.K., Banerjee, A., Datta, J.K. and Gupta, S., 2009. Antifungal activities and chemical characterization of Neem leaf extracts on the growth of some selected fungal species in vitro culture medium. Journal of Applied Science and Environment Management 13(1): 49-53.
- Moore, N.Y., 1994. *Fusarium* wilt of banana: pathogen variability and host pathogen interaction. Ph.D. Thesis, The University of Queensland, Brisbane, Australia.
- Moore, N.Y., Pegg, K.G., Allen, R.N. and Irwin, J.A.G., 1993. Vegetative compatibility and distribution of *Fusarium oxysporum* f. sp. *cubense* in Australia. Australian Journal of Experimental Agriculture 33: 797-802.
- Moreira, R.S., 1999. Banana: teoria e prática de cultivo (2a ed., CD ROM). Campinas, SP: Fundação Cargill.
- Morgan, E.D. 2009. *Azadirachtin*, a scientific gold mine. Bioorganic & Medicinal Chemistry 17: 4096-4105.
- Moslem M.A. and El-Kholie E.M., 2009. Effect of Neem (*Azardirachta indica* A. Juss) seeds and leaves extract on some plant pathogenic fungi. Pakistan Journal of Biological Sciences12 (14): 1045-1048.
- Mostert, D., Molina, A.B., Daniells, J., Fourie, G., Hermanto, C., Chao, C.P., Fabregar, E., Sinohin, V.G., Masdek, N., Thangavelu, R., Li, C., Yi, G., Mostert, L. and Viljoen, A., 2017. The distribution and host range of the banana *Fusarium* wilt fungus, *Fusarium oxysporum* f.sp. *cubense*, in Asia. PLoS One 12 (7).
- Moyin-Jesu, E.I., 2012. Comparative evaluation of modified neem leaf, neem leaf and woodash extracts on soil fertility improvement, growth and yields of maize (*Zea mays* L.) and watermelon (*Citrullus lanatus*) (sole and intercrop). Agricultural Sciences 3(1): 90-97.
- Moyin-Jesu, E.I., 2014. Effects of water extracts of Neem (*Azadirachta indica* L.) leaf, wood ash and their mixture on soil chemical composition and growth and yield of plantain (*Musa sapientum* L.). American Journal of Experimental Agriculture 4(7): 836-848.

- Murakami, H., Tsushima, S., Kuroyanagi, Y. and Shishido, Y., 2002. Reduction of resting spore density of *Plasmodiophora brassicae*. Soil Science and Plant Nutrition 48 (5): 685-691.
- Musabyimana, T. and Saxena, R.C., 1999. Efficacy of Neem seed derivatives against nematodes affecting banana. Phytoparasitica 27(1): 43-49.
- Mustaffa, M.M and Kumar, V., 2012. Banana production and productivity enhancement through spatial, water and nutrient management. European Journal for Horticultural Science 7(1): 1-28.
- Nahak, G. and Sahu, R. K., 2010. Antioxidant activity in bark and roots of neem (*Azadirachta indica*) and Mahaneem (*Melia azedarach*). Journal of Pharmaceutical Sciences 4: 28-34.
- Nahak, G. and Sahu, R.K., 2014. Bioefficacy of leaf extract of neem (*Azadirachta indica* A. Juss) on growth parameters, wilt and leafspot diseases of brinjal. Research Journal of Medicinal Plants 8 (6): 269-276.
- Nasdir, N., 2003. *Fusarium* wilt race 4 in Indonesia. Research Institute for Fruits west. Sumatra, Indonesia. Abstracts of Papers 2nd. International Symposium on *Fusarium* wilt on banana. PROMUSA-INIBAP/EMBRAPA. Salvador de Bahía: Brazil 22 26 Sept.
- Nasir, N., Pittaway, P.A. and Pegg, K.G., 2003. Effect of organic amendments and solarisation on *Fusarium* wilt in susceptible banana plantlets, transplanted into naturally infested soil. Australian Journal of Agricultural Research 54: 251-257.
- Nel, B., Steinberg, C., Labuschagne, N. and Viljoen, A., 2006. Isolation and characterization of nonpathogenic *Fusarium oxysporum* isolates from the rhizosphere of healthy banana plants. Plant pathology 55(2): 207-216.
- Nel, B., Steinberg, C., Labuschagne, N. and Viljoen, A., 2007. Evaluation of fungicides and sterilants for potential application in the management of *Fusarium* wilt of banana. Crop Protection 26: 697-705.
- Nelson, C.S., Ploetz, R. and Kepler, A.K., 2006. Musa species (banana and plantain). Species profiles for pacific island agro forestry [online]. Available from: https://traditionaltree.org [Accessed on 12 May 2019]
- Ngamsaeng, A., Wanapat, M. and Khampa, S., 2006. Evaluation of local tropical plants by in vitro rumen fermentation and their effects on fermentation end-products. Pakistan Journal of Nutrition 5 (5): 414-418.
- Niranjan, P.S., Udeybir, Singh, J. and Verma, D.N., 2008. Mineral and antinutritional factors of common tree leaves. Indian Veterinary Journal 85: 1067-1069.

- Njukwe, E., Tenkouano, A., Amah, D., Sadik, K., Perez, M., Nyine, M. and Dubois, T., 2007. Macro-propagation of banana and plantain: training manual. Entebbe: ASARECA.
- Noor, U.N.M., Memon, K.S., Anwar, R., Ahmad, S. and Nafees, M., 2010. Status and response to improved NPK fertilization practices in banana. Pakistan Journal of Botany 42(4): 2369-2381.
- Nor Aslan, Nurul Aqidah and Marimuthu, Murugaiyah, 2010. Neem (*Azadirachta indica*) oil as an anthelmintic in goats. Serdang, Selangor: Seminar on Veterinary Sciences p. 78.
- Ntamwira, J., Sivirihauma, C., Ocimati, W., Bumba, M., Vutseme, L., Kamira, M. and Blomme, G., 2017. Macropropagation of banana/plantain using selected local materials: A cost-effective way of mass propagation of planting materials for resource-poor households. European Journal for Horticultural Science 82: 38-53.
- Nyanjage, M.O., Wainwright, H., Bishop, C.F.H. and Cullum, F.J., 2001. A comparative study on the ripening and mineral content of organically and conventionally grown cavendish bananas. Biological Agriculture and Horticulture 18(3): 221-234.
- Okungbowa, F. I. and Shittu, H., 2013. *Fusarium* Wilts: An Overview. Environmental Research Journal 6(2):83-102.
- Ordonez, N., Garcia, F., Laghari, H., Akkary, M., Harfouche, E., Al Awar, B. and Kema, G., 2015. First report of *Fusarium oxysporum* f. sp. *cubense* tropical race 4 causing Panama disease in Cavendish bananas in Pakistan and Lebanon. Plant Disease: In press.
- Oritsejafor, J.J., 1986. Influence of moisture and pH on growth and survival of *Fusarium oxysporum* f.sp. *elaeidis* in soil. Transactions of the British Mycological Society 87 (4): 511-517.
- Orr, R. and Nelson, P.N., 2018. Impacts of soil abiotic attributes on *Fusarium* wilt, focusing on bananas. Applied Soil Ecology 132: 20-33.
- Parkinson, D., Gray, T.R.G. and Williams, S.T., 1971. Methods for studying the ecology of soil microorganisms. IBP Handbook No. 19. Blackwell Scientific Publications, Oxford.
- Paul, P.K. and Sharma, P.D., 2002. Azadirachta indica leaf extract induces resistance in barley against leaf stripe disease. Physiological and Molecular Plant Pathology 61: 3-13.
- Pauziah, M., Suhana, O., Rozeita, L. and Maimun, T., 2016. Status of *Fusarium* wilt diseases in Malaysia. Malaysian Agricultural Research and Development Institute MARDI [online]. Available from: http://banananetworks.org [Accessed on 28 Oct 2017].

- Ploetz, R.C. and Pegg, K.G., 2000. Fungal diseases of root, corm andpseudostem. In: Jones, D.R. (Ed.), Diseases of Banana, Abaca´and Enset. CAB International, Wallingford, UK, pp. 143-172.
- Ploetz, R.C., 2006a. *Fusarium* wilt of banana is caused by several pathogens referred to as *Fusarium oxysporum* f. sp. *cubense*. Phytopathology 96:653-656.
- Ploetz, R.C., 2006b. Panama disease, an old nemesis rears its ugly head: Part 2, the Cavendish era and beyond. Plant Health Progress. March 2006.
- Ploetz, R.C., Kema, G.H.J. and Ma, L.J., 2015. Impact of diseases on export and smallholder production of banana. Annual Review of Phytopathology 53: 269-288.
- Ploetz, R.C., Kepler, A.K., Daniells J., and Nelson S.C., 2007. Banana and plantain-an overview with emphasis on Pacific island cultivars. Musaceae (banana family). Species Profiles for Pacific Island Agroforestry. Permanent Agriculture Resources (PAR). In: Elevitch, C.R. (ed.).
- Ploetz, R.C., Thomas, J.E. and Slabaugh, W.R., 2003. Diseases of banana and plantain. In: Ploetz RC, ed. Diseases of Tropical Fruit Crops. Wallingford, UK: CAB International Publishing, 109-112.
- Ploetz, R.C. and Pegg, K.G., 2000. Fungal diseases of root, corm andpseudostem. In: Jones, D.R. (Ed.), Diseases of Banana, Abaca[´]and Enset. CAB International, Wallingford, UK, pp. 143–172.
- Ploetz, R.C., 2005. *Fusarium* wilt of banana is caused by several pathogens referred to as *Fusarium* oxysporum f. sp. cubense. The American Phytopathological Society 96(6): 653-656.

Ploetz, R.C., 1990. Fusarium Wilt of Banana. APS Press, St Paul.

- Poly-Mbah, C.P., Onuoha, R.E. and Uzowuru, E.I., 2010. Effect of soil nursery mixtures on the growth of pepper seedlings in Owerri, South Eastern Nigeria. International Journal of Agriculture and Rural Development 13: 2.
- Prasad, R., 2007. Strategy for increasing fertilizer use efficiency. Indian Journal of Fertilizers 3(1): 53-62.
- Prasanthrajan, M., Pandiyan, M. and Shalini, S., 2017. Reducing the electrical conductivity of bore well water using natural bioadsorbents and augmenting azolla growth by neem bark powder-clay sorbent. Water Science and Technology Water Supply 17(5): 1298–1305.

- Puvan Arul, A., Irfan M., Rosdan S., Zeehaida M., 2015. Antifungal effect of Malaysian neem leaf extract on selected fungal species causing otomycosis in *in-vitro* culture medium. Malaysian Journal of Medicine and Health Sciences 11(2): 69-84.
- Qasem, J.R. and Abu-Blan, H.A., 1966. Fungicidal activity of some common weed extracts against different plant pathogenic fungi. Journal of Phytopathology 144: 157-161.
- Raghupathi, H.B., Srinivas, K., Reddy, B.M.C. and Padma, P., 2000. Concentration and distribution of primary nutrient in banana under fertigation. Indian Journal of Horticulture 57(3).
- Raheem, A.A. and Olatunde Ibiwoye, E., 2018. A study of neem seed husk ash as partial replacement for cement in concrete. International Journal of Sustainable Construction Engineering and Technology 9(2): 55-65.
- Rahman, M.A., Begum, M.F. and Alam, M.F., 2009. Screening of *Trichoderma* isolates as a biological control agent against *Ceratocystis paradoxa* causing pineapple disease of sugarcane. Mycobiology 37(4): 277–285.
- Rathore, S.S., Saxena, S.N., Sharma, Y.K. and Mishra, B.K. and Singh, B., 2015. Effect of pH and salt levels on growth of *Fusarium oxysporum* f. sp. *cumini* isolate from cumin. International Journal of Seed Spices 5: 100-101.
- Reddy, M.S., Kumar, S. and Khosla, B., 2002. Biosolubilization of poorly soluble rock phosphates by *Aspergillus tubingensis* and *Aspergillus niger*. Bioresource Technology 84: 187-189.
- Riaz, T., Khan, S.N. and Javaid, A., 2010. Management of *Fusarium* corm rot of gladiolus (*Gladiolus grandiflorus* sect. Blandus cv. Aarti) by using leaves of allelopathic plants. African Journal of Biotechnology 9(30): 4681-4686.
- Richards, L.A., 1947. Pressure-membrane apparatus, construction and use. Agriculture Engineering 28: 451-454.
- Ribeiro, L.R., Amorim, E.P., Cordeiro, Z.J.M., de Oliveira e Silva, S. and Dita M.A., 2011. Discrimination of banana genotypes for *Fusarium* wilt resistance in the greenhouse. Acta horticulturae 897: 381-386.
- Richards, L.A., 1947. Pressure-membrane apparatus-construction and use. Agricultural Engineering, 28: 451-454.
- Ros, M., Hernandez, M.T., Garcia, C., Bernal, A. and Pascual, J.A., 2005. Biopesticide effect of green compost against *Fusarium* wilt on melon plants. Journal of Applied Microbiology 98(4): 845-854.

- Rossner, J., Zebitz, C.P.W., 1986. Effect of neem products on nematodes on tomato (*Lycopersicon esculentum*) plants. In: Proceedings of the 3rd International Neem Conference, Nairobi, Kenya. pp 10-15.
- Rousk, J., Brookes, P.C. and Bååth, E., 2009. Contrasting soil pH effects on fungal and bacterial growth suggest functional redundancy in carbon mineralization. Applied and Environmental Microbiology 75 (6): 1589– 1596.
- Rowe, R.N., Farr, D.J. and Richards, B.A.J., 1994. Effects of foliar and root applications of methanol or ethanol on the growth of tomato plants (*Lycopersicon esculentum* Mill). New Zealand Journal of Crop and Horticultural Science 22: 335-337.
- Runia, W.T., 2014. Management of *Fusarium oxysporum* f.sp *cubense* (Foc-TR4) from banana by anaerobic soil disinfestation (ASD). Lelystad: PPO AGV.
- Rupe, J.C., 1989. Frequency and pathogenicity of *Fusarium solani* recovered from soybeans with sudden death syndrome. Plant Diseases 73:581-584.
- Samuel, T.A., Hussaini, A.M., Titilayo, A. and Ibrahim K., 2008. Effects of *Fusarium* verticilloides, its metabolites and neem leaf extract on germination and vigour indices of maize (*Zea mays* L.). African Journal of Biotechnology 7(14): 2402-2406.
- Sana, N., Shoaib, A., Javaid, A. and Farooq, N., 2015. Effect of neem leaves as soil amendment on southern blight disease, growth and physiology of chili. Pakistan Journal of Phytopathology 27(2): 115-120.
- Sánchez-Rangel, D., Hernández-Domínguez, E., Pérez-Torres, C., Ortiz-Castro, R., Villafán, E., Rodríguez-Haas, B., Alonso-Sánchez, A., López-Buenfil, A., Carrillo-Ortiz, N., Hernández-Ramos, L. and Ibarra-Laclette E., 2018. Environmental pH modulates transcriptomic responses in the fungus *Fusarium* sp. associated with KSHB *Euwallacea* sp. near fornicates. BMC Genomics 19:721-735.
- Santhi, S.R. and Palaniappan, Sp., 1986. Effect of Neem Leaf (*Azadirachta indica* L.) on growth and yield of low land rice. Journal of Agronomy and Crop Science Volume 157 (2): 114 117.
- Saravanan, T., Muthusamy, M. and Marimuthu, T., 2003. Development of integrated approach to manage the fusarial wilts of banana. Crop Protection 22: 1117-1123.
- Sarawaneeyaruk, S., Krajangsang, S. and Pringsulaka, O., 2015. The effects of neem extract and azadirachtin on soil microorganisms. Journal of soil science and plant nutrition 15 (4): 1071-1083.

- SAS Institute Inc., 1999. Statistical Analysis Systems, SAS Producers Guide, Version 8. Cary, Newcastle.
- Scher, F.M. and Baker, R., 1980. Mechanism of biological control in a *Fusarium*suppressive soil. Phytopathology 70: 412-417.
- Shin, C.H. and Wen H.K., 2004. Cavendish banana cultivars resistant to *Fusarium* wilt acquired through soma clonal variation in Taiwan. The American Phytopathological Society 88 (6): 580-588.
- Simmonds, N.W., 1962. The evolution of the bananas. London: Longman. p 124.
- Singh, P., Singh, M.V.K.A. and Vyas, D., 2013. *Arbuscular Mycorrhizal* Fungi: biocontrol against *Fusarium* wilt of chickpea. International Journal of Scientific and Research Publications 3 (1): 2250-3153.
- Singh, P.P., Shin, Y.C., Park, C.S. and Chung, Y.R., 1999. Biological control of *Fusarium* wilt of cucumber by *Chitinolytic* Bacteria. Phytopathology 89(1): 92-99.
- Singh, U., Singh, H. and Singh, R., 1980. The fungicidal effect of neem (*Azadirachta indica*) extracts on some soil-borne pathogens of gram (*Cicer arietinum*). Mycologia 72:1077-1093.
- Sirsat, S.B., Sobita, S. and Abhilasha A.L., 2017. Comparative effect of *Pseudomonas* sp., Neem leaf extract and fungicides against leaf blight *Alternaria alternata* disease of chickpea. Journal of Pharmacognosy and Phytochemistry 6(4): 1992-1994.
- Skidmore, A.M., Dickinson, C. H., 1976. Colony interactions and hyphal interference between *Septoria Nodorum* and phylloplane fungi. Transactions of the British Mycological Society 66: 57–64.
- Smiley, R.W., Collins, H.P. and Rasmussen, P.E., 1996. Diseases of wheat in long-term agronomic experiments at Pendleton, Oregon. Plant Disease 80: 813–820.
- Staver, C. and Lescot, T., 2015. Propagating quality planting material to improve plant health and crop performance. Key practices for dessert banana, plantain and cooking banana: Illustrated guide. Rome: Bioversity International. p56.
- Steijl, H., Niemann, G.J. and Boon, J.J., 1999. Changes in chemical composition related to fungal infection and induced resistance in carnation and radish investigated by pyrolysis mass spectrometry. Physiological and Molecular Plant Pathology 55(5): 297-311.
- Stover, R.H., 1953a. The effect of soil moisture on *Fusarium* species. Canadian Journal of Botany 31: 693-697.

- Stover, R.H., 1953b. The effect of soil moisture on the growth and survival of *Fusarium oxysporum* f. *cubense* in the laboratory. Phytopathology 43: 499-504.
- Stover, R.H., 1962. Fusarial wilt (Panama Disease) of bananas and other Musa species. Phytopathol. Surrey: Commonw. Mycol. Inst., Kew.
- Subapriya, R. and Nagini, S., 2005. Medicinal properties of neem leaves: a review. Current Medicinal Chemistry Anti-Cancer Agents 5(2):149-156.
- Sullivan, P., 2001. Sustainable management of soil-borne plant diseases. ATTRA, USDA's Rural Business Cooperative Service. [online]. Available from: https://www.attra.org [Accessed on 19 Jun 2019].
- Summerell, B.A., Salleh, B. and Leslie, J.F., 2003. A utilitarian approach to *Fusarium* identification. The American Phytopathological Society 87(2): 117-128.
- Swennen, R., Wilson, G.F. and De Langhe. E. 1984. Preliminary investigation of the effects of gibberellic acid (GA3) on sucker development in plantain (*Musa* cv. AAB) under field conditions. Tropical Agriculture (Trin.) 61:253–256.
- Tan, M.T., 1974. Studies on the antifeedant properties of extracts from the neem tree (*Azadirachta indica* A. Juss) and their effectiveness against the diamondback moth, *Plutella xylostella* (L.). Bachelor of Science Project, University of Malaya, Kuala Lumpur.
- Tariq, V.N. and Magee A.C., 1990. Effect of volatiles from garlic bulb extract on *Fusarium oxysporum* f. sp. *lycopersici*. Mycological Research 94(5): 617-620.
- Teixeira, L.A.J., Nomura, E.S., Damatto Jr., E.R. and Fuzitani, E.J., 2014. Banana. In Aguiar, A.T.E., Gonçalves, C., Paterniani, M.E.A.G., Tucci, M.G.S. and Castro, CE. F. (Eds.), Instruções agrícolas para as principais culturas econômicas (p.46-51, Boletim Técnico, 200). Campinas, SP: Instituto Agronômico.
- Tomlinson, P, 1969. Anatomy of the monocotyledons. III. Commelinales– Zingiberales. Oxford: Clarendon Press.
- Trinci, A.P.J., 1971. Influence of the width of the peripheral growth zone on the radial growth rate of fungal colonies on solid media. Journal of General Microbiology 67:325–344.
- Tyagi, S. and Paudel, R., 2014. Effect of different pH on the growth and sporulation of *Fusarium oxysporum*: The causal organism of wilt disease of tomato. International Journal of Basic and Applied Biology 2 (1): 103 – 106.

- USDA, 2017. United States Department of Agriculture, Agricultural Research Service. National Nutrient Database [online]. Available from: https://ndb.nal.usda.gov [Accessed on 13 Jun 201].
- Vassilev, N., Medina, A., Azcon, R. and Vassileva, M., 2006. Microbial solubilization of rock phosphate on media containing agro-industrial wastes and effect of the resulting products on plant growth and P uptake. Developments in Plant and Soil Sciences 287: 77-84.
- Viljoen, A., 2002. The status of *Fusarium* wilt (Panama disease) of banana in South Africa. South African Journal of Science 98: 341–344.
- Weller, D.M., Raaijmakers, J.M., McSpadden Gardener, B.B. and Thomashow, L.S., 2002. Microbial populations responsible for specific soil suppressiveness to plant pathogens. Annual Review of Phytopathology 40: 309 – 348.
- Wheeler, K.A., Hurdman, BF. and Pitt J. I., 1990. Influence of pH on the growth of some toxigenic species of *Aspergillus, Penicillium* and *Fusarium*. International Journal of Food Microblology 12: 141-150.
- Wills, R., Wimalasiri, P. and Greenfield, H., 1984. Dehydroascorbic acid levels in fresh fruit and vegetables in relation to total vitamin C activity. Journal of Agricultural and Food Chemistry 32: 836–838.
- Wöjcik, D., 2004. Convergence in corporate governance: Empirical evidence from Europe 2000–2003. Working Paper, School of Geography, University of Oxford.
- Wright, I.J., Reich, P.B. and Westoby, M., 2001. Strategy shifts in leaf physiology, structure and nutrient content between species of high- and low-rainfall and high- and low-nutrient habitats. Functional Ecology 15 (4): 423 434.
- Xue, C., Penton, C., Shen, Z., Zhang, R., Huang, Q., Li, R., Ruan, Y., Shen, Q., 2015. Manipulating the banana rhizosphere microbiome for biological control of Panama disease. Scientific Reports 5: 11124.
- Yao, H.Y. and Wu, F.Z., 2010. Soil microbial community structure in cucumber rhizosphere of different resistance cultivars to *Fusarium* wilt. FEMS Microbiology Ecology (72): 456–463.
- Yusnita and Sudarsono, 2004. Metode inokulasi dan reaksi ketahanan 30 genotipe kacang tanah terhadap penyakit busuk batang Sclerotium. Hayati 11: 53-58.