



UNIVERSITI PUTRA MALAYSIA

***EFFECTS OF Brachiaria decumbens STAPF IN SHORT- AND LONG-TERM FEEDING ON In Vitro RUMEN FERMENTATION AND GROWTH PERFORMANCE IN SHEEP***

MIMI SYAZWANI BINTI JAAPAR

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PERFORMANCE IN SHEEP**

By

**MIMI SYAZWANI BINTI JAAPAR**

Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of Master of  
Science

**December 2021**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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**December 2021**

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*Brachiaria* species have been planted on more than 80% of improved farming pastures with *Brachiaria decumbens* as the most favoured species. It is important because of its high productivity under intensive use, and its tolerance of low fertility soil. However, the presence of steroid saponins in *B. decumbens* which had become a limiting factor led to the outbreaks of photosensitivity in ruminants. This current study focused on the effects of *B. decumbens* in short and long-term feeding on *In Vitro* gas production, rumen fermentation, nutrient digestibility and growth performance in sheep. A total of 30 (6-month-old) male Dorper cross sheep were divided randomly into 3 treatment groups with 10 sheep per treatment. This study was conducted in 2 phases which is the short-term (7 days) and long-term (90 days) stages excluding the 2 weeks adaptation period. Treatment 1 (control) sheep were fed with *Pennisetum purpureum* and concentrates as the basal diet, whereas Treatment 2 and 3 sheep were fed with low (10%) and high (60%) levels of *B. decumbens* respectively according to 3% kg/feed per body weight. The digestibility trial was done at day 7 during the short-term and at day 90 during the long-term. The amount of feed offered and refusals were recorded daily while body weight gain and body measurement were recorded weekly to determine the feed efficiency. The *In Vitro* gas production showed no significant changes in net gas production and gas production kinetics during the 48 hours on incubation. However, the gas production significantly decreased as the level of *B. decumbens* increases, with T3 diet demonstrated poorest gas production. The concentration of ammonia and pH showed no significant difference among treatments but decreasing as *B. decumbens* level increases. The same goes with all individual VFA except for acetic acid and total VFA ( $p<0.05$ ) The sheep digestibility of organic matter, neutral detergent fiber, and crude protein except dry matter showed no significant differences ( $p>0.05$ ) during the short-term, while DM, CP, NDF and acid detergent fiber decreased significantly ( $p<0.05$ ) during the long-term. For the growth performance and feed intake, there were also no significant differences during the short-term.

Meanwhile, there were significant differences in total weight gained each week, average daily gain (ADG), total feed intake and daily feed intake recorded daily during the long-term. T3 sheep fed with the highest concentration of *B. decumbens* exhibited the worst growth performance than other treatments. Significant differences ( $p<0.05$ ) were detected on day 7, 30, 60, and 90 among treatment sheep. T1 showed the highest growth hormones compare to other treatments and T3 sheep revealed the highest reduction in growth hormone concentration. Despite the ADG and weight gain result for long-term and growth hormones, there were no significant differences on the body measurement during both terms including the body indexes. Overall, feeding sheep with the high level of *B. decumbens* showed inferior in the gas production and rumen fermentation, growth performance and digestibility as compared to the control group fed with *P. purpureum* with 0% of *B. decumbens*. The effects of both low and high saponin levels of *B. decumbens* on *In Vitro* study and growth performance on sheep from this study may contribute to future research of *B. decumbens*.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk Ijazah Master Sains

**KESAN JANGKA PENDEK DAN JANGKA PANJANG PEMBERIAN MAKAN  
*Brachiaria decumbens* STAPF PADA FERMENTASI RUMEN *In Vitro* DAN  
PRESTASI PERTUMBUHAN TERHADAP BIRI – BIRI**

Oleh

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Spesies *Brachiaria* telah ditanam lebih daripada 80% padang rumput pertanian yang telah diperbaik pulih dengan *Brachiaria decumbens* sebagai spesies yang paling digemari. Hal ini penting kerana produktivitinya yang tinggi dalam penggunaan intensif, dan toleransi yang tinggi terhadap masalah kesuburan tanah. Walau bagaimanapun, saponin steroid dalam *B. decumbens* menjadi faktor pembatas dalam menyebabkan masalah fotosensitisasi pada ruminan. Kajian semasa ini fokus pada kesan jangka pendek dan jangka panjang pemberian makan *Brachiaria decumbens* pada fermentasi rumen, produksi gas *In Vitro*, kebolehcernaan nutrien dan prestasi pertumbuhan tahap rendah terhadap biri – biri. Sebanyak 30 ekor bebiri (Dorper cross) jantan berusia 6 bulan dibahagikan secara rawak kepada 3 kumpulan dengan 10 ekor bebiri setiap rawatan. Kajian ini dilakukan pada 2 fasa: jangka pendek (7 hari) dan jangka panjang (90 hari) tidak termasuk tempoh penyesuaian 2 minggu. Bebiri kumpulan 1 (kawalan) diberi makan *Pennisetum purpureum* dan pellet sebagai diet basal, manakala bebiri kumpulan 2 dan 3 diberi makan dengan tahap *B. decumbens* yang rendah (10%) dan tinggi (60%) menurut 3% kg/makanan per berat badan. Percubaan pencernaan dilakukan pada hari ke-7 semasa jangka pendek dan pada hari ke-90 semasa jangka panjang. Jumlah makanan yang ditawarkan dan baki direkod secara harian manakala kenaikan berat badan dan pengukuran badan dicatat secara mingguan untuk menentukan kecekapan makanan. Pengeluaran gas *In Vitro* tidak menunjukkan perubahan ketara dalam pengeluaran gas bersih dan pengeluaran gas kinetik selama 48 jam semasa inkubasi. Walaubagaimanapun, pengeluaran gas menurun dengan ketara apabila tahap *B. decumbens* meningkat, dengan diet kumpulan 3 menunjukkan pengeluaran gas paling rendah. Konsentrasi amonia dan pH tidak menunjukkan perbezaan yang signifikan antara kumpulan bebiri tetapi menurun apabila tahap *B. decumbens* meningkat. Hal yang sama berlaku untuk semua individu VFA kecuali asid asetik dan jumlah keseluruhan VFA ( $p<0.05$ ). Pencernaan nutrisi

bebiri: bahan organik, fiber detergen neutral, dan protein mentah kecuali bahan kering tidak menunjukkan perbezaan yang signifikan ( $p > 0,05$ ) semasa jangka pendek, sementara DM, CP, NDF dan fiber detergen asid menurun dengan ketara ( $p < 0,05$ ) semasa jangka panjang. Bagi prestasi pertumbuhan dan pengambilan makanan, tidak ada perbezaan yang signifikan semasa jangka pendek. Sementara itu, terdapat perbezaan yang signifikan dalam jumlah kenaikan berat badan, kenaikan harian purata (ADG), jumlah pengambilan makanan dan pengambilan makanan harian semasa jangka panjang. Bebiri kumpulan 3 yang diberi konsentrasi *B. decumbens* tertinggi menunjukkan prestasi pertumbuhan terburuk daripada kumpulan lain. Perbezaan yang ketara ( $p < 0,05$ ) dikesan pada hari ke-7, 30, 60, dan 90 di antara bebiri. Kumpulan menunjukkan hormon pertumbuhan tertinggi dibandingkan dengan kumpulan lain manakala bebiri kumpulan 3 menunjukkan penurunan konsentrasi hormon pertumbuhan tertinggi. Walaupun terdapat signifikan pada ADG dan kenaikan berat badan untuk jangka panjang dan penurunan hormon pertumbuhan, tiada perbezaan yang signifikan didapati pada pengukuran badan selama kedua - dua jangka termasuk indeks badan. Secara keseluruhan, pemberian *B. decumbens* di konsentrasi tertinggi sebagai makanan kepada bebiri menunjukkan penurunan dalam prestasi pertumbuhan, pencernaan dan pengeluaran gas dan fermentasi rumen berbanding kumpulan kawalan yang diberi makan *P. purpureum* dengan 0% kandungan *B. decumbens*. Kesan tahap konsentrasi saponin rendah dan tinggi dalam *B. decumbens* bagi kajian *In Vitro* dan prestasi pertumbuhan pada bebiri dari kajian ini boleh menyumbang kepada penyelidikan *B. decumbens* di masa hadapan.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

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## LIST OF ABBREVIATIONS

AMT	<i>Astragalus membranaceus</i> root
APS	<i>Astragalus polysaccharide</i>
<i>B. brizantha</i>	<i>Brachiaria brizantha</i>
<i>B. decumbens</i>	<i>Brachiaria decumbens</i>
<i>B. humidicola</i>	<i>Brachiaria humidicola</i>
<i>B. mutica</i>	<i>Brachiaria mutica</i>
<i>B. ruziziensis</i>	<i>Brachiaria ruziziensis</i>
<i>M. uniflorum</i>	<i>Macrotyloma uniflorum</i>
<i>P. chartarum</i>	<i>Pithomyces chartarum</i>
<i>P. maximum</i>	<i>Panicum maximum</i>
<i>P. purpureum</i>	<i>Pennisetum purpureum</i>
<i>Q. Saponaria</i>	<i>Quillaja Saponaria</i>
<i>T. terrestris</i>	<i>Tribulus terrestris</i>
<i>Y. schidigera</i>	<i>Yucca schidigera</i>
%	Percent
°C	Celsius
µL	Microlitre
ADF	Acid detergent fibre
ADG	Average daily gain
BL	Body length
BLI	Body length index
cm	Centimetre
CO <sub>2</sub>	Carbon dioxide
CP	Crude protein

DM	Dry matter
DMI	Dry matter intake
EE	Ether extract
FE	Feed efficiency
g	Gram
GH	Growth hormone
GRF	Growth hormone - releasing factor
h	Hour
HG	Heart girth
HGI	Heart girth index
IGF-1	Insulin – like growth factor
IVDMD	In vitro dry matter digestibility
kg	Kilogram
L	Litre
mg	Milligram
min	Minute
mL	Millilitre
mM	Millimolar
NDF	Neutral detergent fibre
NH <sub>3</sub> -N	Ammonia nitrogen
nm	Nanometre
OM	Organic matter
OMD	Organic matter digestibility
ppm	Parts per million
rpm	Revolution per minute

SAS	Statistical analysis system
SI	Somatic index
SRIH	Somatostatin
ST	Somatotropin
v/v	Volume/volume
v/w	Volume/weight
VFA	Volatile fatty acid
w/w	Weight/weight
WH	Whither height

## CHAPTER 1

### INTRODUCTION

#### 1.1 *Brachiaria decumbens* intoxication

The ruminant sector is one of the main livestock industries in Malaysia with feed being the main factor influencing both large and small ruminant production (Yusoff et al., 2016). One of the most important grasses for ruminant production is *Brachiaria* spp. Owing to the hot and humid climatic condition in Malaysia, *B. decumbens* is considered to have the potential as an excellent pasture species for the ruminant industries (Assumaidae and Mustapha 2012). Other species of *Brachiaria* spp. namely *B. brizantha*, *B. humidicola*, *B. mutica*, and *B. ruziziensis* are also widely used as ruminant feed. Livestock production on this pasture system has been variable even when the quality and quantity of the pasture are high (Low, 2015). *B. decumbens* originally came from the highlands of Central and Eastern Africa. It is now widespread in the tropics and sub-tropics. Initially, *Pithomyces chartarum* was suspected as the primary cause of the outbreak of photosensitivity due to the presence of spores. However, several authors have shown that none of the *P. chartarum* isolates produce sporodesmin that cause hepatotoxicity (Graydon et al., 1991; Salam Abdullah et al., 1992). Further study has revealed that the crystal-associated cholangiohepatopathy lesions in the liver tissue of sheep are similar to that found in *Panicum* spp. and *Tribulus terrestris* poisoning. These forages contain lithogenic saponins known to cause photosensitization (Bridges et al., 1987; Miles et al., 1993). The toxicity symptoms were reproduced and confirmed in sheep through oral dosing of crude saponins extracted from the plant *T. terrestris* (Zygophyllaceae) (Lascano and Euclides, 1996).

Similarly, Lemos et al. (1998) suggested that *Brachiaria* spp poisoning was also due to lithogenic saponins contained in the plant. Cruz et al. (2001) have induced cholangiopathy in lambs by administering *B. decumbens* extracts showing that the poisoning was caused by lithogenic saponins contained in the grass. Furthermore, Salam Abdullah et al. (1992) claimed that steroid saponins were found in the rumen contents of poisoned sheep fed with *B. decumbens*. Supporting this, a study has confirmed the presence of plant saponins in the rumen of sheep suffering from *B. decumbens* toxicity (Lajis et al., 1993). The presence of steroid saponins in *B. decumbens* is the main cause of photosensitivity in goats, sheep, llama, buffaloes, deer, and cattle (Low, 2015).

Although feeding the animals with *B. decumbens* has improved their overall performance, the risk of toxicity has become a limiting factor (Muniandy et al., 2020). Even though most of the outbreaks related to hepatogenous photosensitization are caused by *B. decumbens*, other *Brachiaria* species such as *B. brizantha*, *B. humidicola*, and *B. ruziziensis* could also cause poisoning in cattle, sheep, goats, and buffalo (Riet-Correa et al., 2011). The toxicity of *B.*

*decumbens* is due to the presence of steroid saponins and intoxication occurs when the grass is fed as the main source of feed for grazing animals (Assumaidae and Mustapha, 2012). Clinical symptoms such as liver damage in histopathology, changes in haematological results, hepatic jaundice, and photosensitization on the skin especially around the eyelid and muzzle have been observed in sheep fed with *B. decumbens*, indicating hepatotoxicity due to toxic damage (Abas et al., 1983). Souza et al. (2010) stated that *B. decumbens* poisoning in sheep can occur at any time of year and any maturity stage of the plant.

According to Riet-Correa et al. (2011), sheep are more susceptible to *B. decumbens* poisoning than goats and other ruminants. Wina et al. (2006) mentioned that goats are categorized as browsers, so they tend to confront more secondary metabolite plants compared to sheep. Hence, a higher proportion of entodiniomorphid protozoa, which aid the digestion process was found in the rumen of a goat than in sheep which led to the rumen micro-organisms of goats being more adaptable to these substances than those of sheep. This was supported by studies conducted by Abas et al. (1983) and Low (1993) who recorded the absence of photosensitization in goats grazing together with sheep suggesting that goats are more resistant than sheep. Furthermore, lamb and naïve sheep are more susceptible to toxicity poisoning as compared to experienced flocks (Faccin et al., 2014). In short, this study contributes to the distribution of *B. decumbens* during emergency for a while or taking in livestock that is previously exposed of *B. decumbens*.

## 1.2 Problem Statements

1. Many studies had only reported the clinico-pathology effects of *B. decumbens* in both grazing and experimental sheep.
2. Limited data is available on the effect of feeding *B. decumbens* on the nutrient composition, *In Vitro* gas production, and rumen fermentation characteristic of sheep.
3. No study has been conducted to justify the impact of feeding different levels of *B. decumbens* diets on the growth performance and digestibility of sheep during the short and long-terms feeding.

### **1.3      Hypotheses**

1. It was hypothesized that there will be responses in both *In Vitro* and *In Vivo* studies using different levels of *B. decumbens* diets and at different time phases.
2. The *In Vitro* gas production and rumen fermentation characteristic will be decreased significantly at a higher level of *B. decumbens* diet.
3. Sheep fed with a high level of *B. decumbens* will have the lowest growth performance and digestibility.
4. Sheep fed with a high level of *B. decumbens* will be severely affected physically and morphologically during the long-term feeding.

### **1.4.     Objectives**

1. The general aim of this study was to investigate the effect of different levels of *B. decumbens* diets on the *In Vitro* rumen fermentation and performances of sheep at different time phases. The specific objectives were:
2. To evaluate the effects of different *B. decumbens* level on the nutrient composition, *In Vitro* gas production, and rumen fermentation characteristic of sheep.
3. To study the effects of different *B. decumbens* levels on the feed intake and nutrient digestibility of sheep during the short and long-terms.
4. To determine the effects of different levels of *B. decumbens* on the growth performance, growth hormone concentration, and body measurement of sheep during the short and long-terms.

## REFERENCES

- Aazami, M.H., Tahmasbi, A.M., Ghaffari, M.H., Naserian, A.A., Valizadeh, R., and Ghaffari, A.H. (2013). Effects of saponins on rumen fermentation, nutrients digestibility, performance, and plasma metabolites in sheep and goat kids. *Annual Research and Review in Biology*, 596-607.
- Abas, M.O., Khusahry, M.Y., and Sheikh Omar, A.R. (1983). Jaundice and photosensitization in indigenous sheep of Malaysia grazing on *Brachiaria decumbens*. *Malaysian journal of Veterinary Research*, 7, 254-263.
- Abdel-Raheem, S. M., and Hassan, E. H. (2021). Effects of dietary inclusion of *Moringa oleifera* leaf meal on nutrient digestibility, rumen fermentation, ruminal enzyme activities and growth performance of buffalo calves. *Saudi Journal of Biological Sciences. Agricultural and Food Chemistry*, 41, 914-917.
- AOAC, (1995). Association of Official Analytical Chemists, Official Methods of Analysis. 16 th Edn. Arlington, Virginia, USA.
- Aregheore, E.M. (2001). Nutritive value and utilization of three grass species by crossbred Anglo-Nubian goats in Samoa. *Asian-Australasian Journal of Animal Science*, 14, 1389–343 1393.
- Arroyave, C., Tolrà, R., Thuy, T., Barceló, J., and Poschenrieder, C. (2013). Differential aluminum resistance in *Brachiaria* species. *Environmental and Experimental Botany*, 89, 11-18.
- Assumaidae, A. A. M., and Mustapha, N. M. (2012). Toxicity of signal grass (*Brachiaria decumbens*): a review article. *Journal of Advanced Medical Research*, 2(2012), 18-39.
- Babayemi, O. J., Hamzat, R. A., Bamikole, M. A., Anurudu, N. F., and Olomola, O. O. (2006). Preliminary studies on spent tea leaf: in vitro gas production as affected by chemical composition and secondary metabolites. *Pakistan Journal of Nutrition*, 5(5), 497-500.
- Baile, C. A., and Pfander, W. H. (1966). A possible chemosensitive regulatory mechanism of ovine feed intake. *American Journal of Physiology-Legacy Content*, 210(6), 1243-1250.
- Bauman, D. E., and Currie, W. B. (1980). Partitioning of nutrients during pregnancy and lactation: a review of mechanisms involving homeostasis and homeorhesis. *Journal of dairy science*, 63(9), 1514-1529.
- Bauman, D.E., Eppard, P.J., DeGeeter, M.J., and Lanza, G.M. (1985). Responses of high-producing dairy cows to long-term treatment with pituitary somatotropin and recombinantsomatotropin. *Journal of Dairy Science*, 68, 1352.

- Bergman, E. N. (1990). Energy contributions of volatile fatty acids from the gastrointestinal tract in various species. *Physiological reviews*, 70(2), 567-590.
- Blümmel, M., and Becker, K. (1997). The degradability characteristics of fifty-four roughages and roughage neutral-detergent fibres as described by in vitro gas production and their relationship to voluntary feed intake. *British Journal of Nutrition*, 77(5), 757-768.
- Blümmel, M., Cone, J. W., Van Gelder, A. H., Nshalai, I., Umunna, N. N., Makkar, H. P. S., and Becker, K. (2005). Prediction of forage intake using in vitro gas production methods: Comparison of multiphase fermentation kinetics measured in an automated gas test, and combined gas volume and substrate degradability measurements in a manual syringe system. *Animal feed science and technology*, 123, 517-526.
- Bosler, D.A., Blummel, M., Bullerdieck, P., Makkar, H.P.S. (1997). Influence of a saponin-containing feed additive on mass development and carcass evaluation of growing lambs. *Proceedings of the Society of Nutrition Physiology*, 6, 46.
- Bridges, C.H., Camp, B.J., Livingston, C.W., and Bailey, E.M. (1987). Kleingrass (*Panicum coloratum* L.) poisoning in sheep. *Veterinary Pathology*, 24, 525-531.
- Brum, K. B., Haraguchi, M., Lemos, R. A., Riet-Correa, F., & Fioravanti, M. C. S. (2007). Crystal-associated cholangiopathy in sheep grazing Brachiaria decumbens containing the saponin protodioscin. *Pesquisa Veterinária Brasileira*, 27, 39-42.
- Bulo, D., Blair, G.J., Till, A.R., and Stür, W. (1994). Yield and digestibility of forages in East Indonesia II. grasses. *Asian-Australasian Journal of Animal Science*, 7, 335-342.
- Cardona-Álvarez, J., Vargas-Vilória, M., & Paredes-Herbach, E. (2016). Clinical and histopathological study of the phototoxic dermatitis in Zebu calves in grazing of Brachiaria decumbens. *Revista MVZ Córdoba*, 21(2), 5366-5380.
- Cardozo, P.W., Calsamiglia, S., Ferret, A., and Kamel, C. (2005). Screening for the effects of natural plant extracts at different pH on in vitro rumen microbial fermentation of a high-concentrate diet for beef cattle. *Journal of Animal Science*, 83, 2572-2579.
- Castro, M.B., Santos, Jr, H.L., Mustafa, V.S., Gracindo, C.V., Moscardini, A.C.R., Louvandini, H., Paludo, G.R., Borges, J.R.J., Haraguchi, Ferreira, M.B., and Riet-Correa, F. (2009). *Brachiaria* spp poisoning in sheep in Brazil: Experimental and epidemiological findings. 366 8th International Symposium on Poisonous Plants, Brazil p.12. (Abstract).

- Cavalcanti Filho, L.F.M., Santos, M.V.F.D., Ferreira, M.D.A., Lira, M.D.A., Farias, I., Ferreira, R.L.C., and Lucena, J.E.C. (2004). Heifers forage intake and performance on *Brachiaria decumbens* after supplementation. *Pesquisa Agropecuaria Brasileira*, 39,1247-1252.
- Cheeke, P. R. (1995). Biological effects of feed and forage saponins and their impacts on animal production. *Saponins used in Food and Agriculture*, 377-385.
- Cheok, C. Y., Salman, H. A. K., and Sulaiman, R. (2014). Extraction and quantification of saponins: A review. *Food Research International*, 59, 16-40.
- Chung, E.L.T., Predith, M., Nobilly, F., Samsudin, A.A., Jesse F.F.A., Loh T.C. (2018). Can treatment of *Brachiaria decumbens* (signal grass) improve its utilisation in the diet in small ruminants? —a review. *Tropical Animal Health and Production*, 50, 1727-1732.
- Corrêa, T. G., Ferreira, J. M., Riet-Correa, G., Ruas, J. L., Schild, A. L., Riet-Correa, F., ... & Felippe-Bauer, M. L. (2007). Seasonal allergic dermatitis in sheep in southern Brazil caused by Culicoides insignis (Diptera: Ceratopogonidae). *Veterinary parasitology*, 145(1-2), 181-185.
- Crowder, L.V., Chaverra, H., and Lotero, J. (1970). Productive improved grasses in Colombia. In Proceedings of the 11th International Grassland Congress, Queensland, Australia, 13– 23 April 1970; Wageningen Academic Publishers: Wageningen, The Netherlands pp. 147–149.
- Cruz, C., Driemeier, D., Pires, V.S., Colodel, E.M., Taketa, A.T., Schenkel, E.P. (2000). Isolation of steroid saponins implicated in experimentally induced cholangiopathy of sheep grazing *Brachiaria decumbens* in Brazil. *Veterinary and Human Toxicology*, 42, 142–145.
- Cruz, C., Driemeier, D., Pires, V.S., Schenkel, E.P. (2001). Experimentally induced cholangiopathy by dosing sheep with fractionated extracts from *Brachiaria decumbens*. *Journal of Veterinary Diagnostic Investigation*, 13, 170-172.
- da Costa, M. C. M., Ítavo, L. C. V., Ítavo, C. C. B. F., Dias, A. M., dos Santos Difante, G., Buschinelli de Goes, R. H. D. T., ... and de Souza Arco, T. F. F. (2021). Natural intoxication caused by protodioscin in lambs kept in *Brachiaria* pastures. *Tropical Animal Health and Production*, 53(3), 1-9.
- Das, T. K., Banerjee, D., Chakraborty, D., Pakhira, M. C., Shrivastava, B., and Kuhad, R. C. (2012). Saponin: Role in animal system. *Veterinary World*, 5(4), 248.
- De Oliveira, C. D., Barbosa, J. D., Oliveira, C. M. C., Bastianetto, E., Melo, M. M., Haraguchi, M., ... & Leite, R. C. (2013). Hepatic photosensitization in

- buffaloes intoxicated by *Brachiaria decumbens* in Minas Gerais state, Brazil. *Toxicon*, 73, 121-129.
- de Paula Barbosa, A. (2014). Saponins as immunoadjuvant agent: a review. *African journal of pharmacy and pharmacology*, 8(41), 1049-1057.
- Debus, N., Breen, K. M., Barrell, G. K., Billings, H. J., Brown, M., Young, E. A., & Karsch, F. J. (2002). Does cortisol mediate endotoxin-induced inhibition of pulsatile luteinizing hormone and gonadotropin-releasing hormone secretion? *Endocrinology*, 143(10), 3748-3758.
- Driver, P. M., and Forbes, J. M. (1981). Episodic growth hormone secretion in sheep in relation to time of feeding, spontaneous meals and short-term fasting. *The Journal of physiology*, 317(1), 413-424.
- Erasmus, L. J., Botha, P. M., and Kistner, A. (1992). Effect of yeast culture supplement on production, rumen fermentation, and duodenal nitrogen flow in dairy cows. *Journal of Dairy Science*, 75(11), 3056-3065.
- Eugene, M., Archimede, H., and Sauvant, D. (2004). Quantitative meta-analysis on the effects of defaunation of the rumen on growth, intake and digestion in ruminants. *Livestock Production Science*, 85, 81-97.
- Faccin, T.C., Riet-Correa, F., Rodrigues, F.S., Santos, A.C., Melo, G.K., Silva, J.A., Ferreira, R., Itavo, C.C., and Lemos, R.A. (2014). Poisoning by *Brachiaria brizantha* in flocks of naïve and experienced sheep. *Toxicon*, 82, 1-8.
- FAO. (2016). Grassland Index. A searchable catalogue of grass and forage legumes. FAO, Rome, Italy.
- Filípek, J., and Dvořák, R. (2009). Determination of the volatile fatty acid content in the rumen liquid: comparison of gas chromatography and capillary isotachophoresis. *Acta Veterinaria Brno*, 78(4), 627-633.
- Finlay, B.J., Esteban, G., Clarke, K.J., Williams, A.G., Embley, T.M., Hirt, R.P. (1994). Some rumen ciliates have endosymbiotic methanogens. *FEMS Microbiology Letters*, 117, 157-161.
- Flåøyen, A., and Wilkins, A. L. (1997). Metabolism of saponins from *Narthecium ossifragum*-a plant implicated in the aetiology of alveld, a hepatogenous photosensitization of sheep. *Veterinary research communications*, 21(5), 335-345.
- Francis, G., Kerem, Z., Makkar, H. P., and Becker, K. (2002). The biological action of saponins in animal systems: a review. *British journal of Nutrition*, 88(6), 587-605.
- Galgal, K.K., Shelton, H.M., Mullen, B., and Komolong, M. (2000). Growth of cattle grazing Leucaena genotypes in the humid lowland tropics of Papua New Guinea. *Asian- Australasian Journal of Animal Science*, 13, 242–245.

- Gee, J. M., Wortley, G. M., Johnson, I. T., Price, K. R., Rutten, A. A. J. J. L., Houben, G. F., and Penninks, A. H. (1996). Effects of saponins and glycoalkaloids on the permeability and viability of mammalian intestinal cells and on the integrity of tissue preparations in vitro. *Toxicology In Vitro*, 10(2), 117-128.
- Getachew, G., Blümmel, M., Makkar, H.P.S., and Becker, K. (1998). *In Vitro* gas measuring techniques for assessment of nutritional quality of feeds: a review. *Animal Feed Science and Technology*, 72, 261-281.
- Goel, G., Makkar, H. P. S., & Becker, K. (2008). Changes in microbial community structure, methanogenesis and rumen fermentation in response to saponin-rich fractions from different plant materials. *Journal of applied microbiology*, 105(3), 770-777.
- Goetsch, A.L., and Owens, F.N. (1985). Effects of sarsaponin on digestion and passage rates in cattle fed medium to low concentrates. *Journal of Dairy Science*, 68, 2377–2384.
- Goldman, A., Genizi, A., Yulzari, A., and Seligman, N.G. (1987). Improving the reliability of the two-stage in vitro assay for ruminant feed digestibility by calibration against in vivo data from a wide range of sources. *Animal Feed Science and Technology*, 18, 233-245.
- Gomar, M. S., Driemeier, D., Colodel, E. M., & Gimeno, E. J. (2005). Lectin histochemistry of foam cells in tissues of cattle grazing Brachiaria spp. *Journal of Veterinary Medicine Series A*, 52(1), 18-21.
- Gracindo, C.V., Louvandini, H., Riet-Correa, F., Barbosa-Ferreira, M., and Castro, M.B. (2014). Performance of sheep grazing in pastures of *Brachiaria decumbens*, *Brachiaria brizantha*, *Panicum maximum* and *Andropogon gayanus* with different protodioscin concentrations. *Tropical Animal Health and Production*, 46, 733–737.
- Granzotto, F., Branco, A. F., dos Santos, A. L., Barreto, J. C., Teixeira, S., Serrano, R. C., ... and Coneglian, S. M. (2011). Proteic supplements with and without sulfur sources on intake behavior of steers fed with low quality hay. *Semina: Ciências Agrárias*, 32(3), 1151-1162.
- Graydon, R.I., Hamid, H., and Zahari, P. (1991). Photosensitization and crystal associated cholangiohepatopathy in sheep grazing *Brachiaria decumbens*. *Australian Veterinary Journal*, 68, 234-236.
- Guenni, O., Marín, D., and Baruch, Z. (2002). Responses to drought of five *Brachiaria* species. I. Biomass production, leaf growth, root distribution, water use and forage quality. *Plant Soil*, 243, 229–241.
- Guil-Guerrero, J. L., Ramos, L., Moreno, C., Zúñiga-Paredes, J. C., Carlosama-Yépez, M., and Ruales, P. (2016). Plant-food by-products to improve farm-animal health. *Animal Feed Science and Technology*, 220, 121-135.

- Gunun, P., Gunun, N., Cherdthong, A., Wanapat, M., Polyorach, S., Sirilaophaisan, S., ... and Kang, S. (2018). In vitro rumen fermentation and methane production as affected by rambutan peel powder. *Journal of Applied Animal Research*, 46(1), 626-631.
- Harding, W.A.T. and Grof, B. (1978). Effect of fertilizer nitrogen on yield, nitrogen content and animal productivity of *Brachiaria decumbens* cv. *Basilisk* on the wet tropical coast of north Queensland. *Queensland Journal of Agriculture and Animal Science*, 34, 11–21.
- Hare, M.D.P., Tatsapong, S., and Phengphet. (2009). Herbage yield and quality of *Brachiaria* cultivars, *Paspalum atratum* and *Panicum maximum* in north-east Thailand. *Tropical grasslands – Forrajes Tropicales*, 43, 65-72.
- Hart, I. C., Bines, J. A., Morant, S. V., and Ridley, J. L. (1978). Endocrine control of energy metabolism in the cow: comparison of the levels of hormones (prolactin, growth hormone, insulin and thyroxine) and metabolites in the plasma of high-and low-yielding cattle at various stages of lactation. *Journal of Endocrinology*, 77(3), 333-345.
- Hart, I. C., Chadwick, P. M. E., James, S., and Simmonds, A. D. (1985). Effect of intravenous bovine growth hormone or human pancreatic growth hormone-releasing factor on milk production and plasma hormones and metabolites in sheep. *Journal of Endocrinology*, 105(2), 189-196.
- Hart, K. J., Yanez-Ruiz, D. R., Duval, S. M., McEwan, N. R., & Newbold, C. J. (2008). Plant extracts to manipulate rumen fermentation. *Animal feed science and technology*, 147(1-3), 8-35.
- Hart, K. J., Yáñez-Ruiz, D. R., Duval, S. M., McEwan, N. R., and Newbold, C. J. (2008). Plant extracts to manipulate rumen fermentation. *Animal Feed Science and Technology*, 147(1-3), 8-35.
- Hasiah, H., Elsheikh, H.A., Abdullah, A.S., Khairi, H.M., and Rajion, M.A. (2000). Effect of phenobarbitone treatment against signal grass (*Brachiaria decumbens*) toxicity in sheep.
- Hess, H.D., Beuret, R.A., Lotscher, M., and Hindrichsen, I.K. (2004). Ruminal fermentation, methanogenesis and nitrogen utilization of sheep receiving tropical grass hay-concentrate diets offered with *Sapindus saponaria* fruits and *Cratylia argentea* foliage. *Animal Science*, 79, 177–189.
- Hess, H.D., Kreuzer, Díaz, T., Lascano, C., Carulla, J., Soliva, R., Machmüller, A. (2003). Saponin rich tropical fruits affect fermentation and methanogenesis in faunated and defaunated rumen fluid. *Animal Feed Science and Technology*, 109, 79-94.
- Hess, H.D., Monsalve, L.M., Lascano, C.E., Carulla, J., and Kreuzer, M. (2003). Supplementation of a tropical grass diet with forage legumes and *Sapindus*

- saponaria* fruits: effects on in vitro ruminal nitrogen turnover and methanogenesis. *Australian Journal of Agricultural Research*, 54, 703–713.
- Hiai, S., Oura, H., and Nakajima, T. (1976). Color reaction of some saponins and saponins with vanillin and sulfuric acid. *Planta Medica*, 29(02), 116-122.
- Holtshausen, L., Chaves, A. V., Beauchemin, K. A., McGinn, S. M., McAllister, T. A., Odongo, N. E., ... and Benchaar, C. (2009). Feeding saponin-containing *Yucca schidigera* and *Quillaja saponaria* to decrease enteric methane production in dairy cows. *Journal of Dairy Science*, 92(6), 2809-2821.
- Hu, W. L., Liu, J. X., Ye, J. A., Wu, Y. M., and Guo, Y. Q. (2005). Effect of tea saponin on rumen fermentation in vitro. *Animal Feed Science and Technology*, 120(3-4), 333-339.
- Hundal, J. S., Wadhwa, M., and Bakshi, M. P. S. (2019). Herbal feed additives containing essential oil: 1. Impact on the nutritional worth of complete feed in vitro. *Tropical animal health and production*, 51(7), 1909-1917.
- Istiqomah, L., Hardian, H., Febrisantosa, A., and Putra, D. (2011). Waru leaf (*Hibiscus tiliaceus*) as saponin source on in vitro ruminal fermentation characteristic. *Journal of the Indonesian Tropical Animal Agriculture*, 36(1), 43-49.
- Izuddin, W. I., Loh, T. C., Foo, H. L., Samsudin, A. A., and Humam, A. M. (2019). Postbiotic *L. plantarum* RG14 improves ruminal epithelium growth, immune status and upregulates the intestinal barrier function in post-weaning lambs. *Scientific reports*, 9(1), 1-10.
- Izuddin, W. I., Loh, T. C., Samsudin, A. A., and Foo, H. L. (2018). In vitro study of postbiotics from *Lactobacillus plantarum* RG14 on rumen fermentation and microbial population. *Revista Brasileira de Zootecnia*, 47.
- Jasmin, B. H., Boston, R. C., Modesto, R. B., and Schaer, T. P. (2011). Perioperative ruminal pH changes in domestic sheep (*Ovis aries*) housed in a biomedical research setting. *Journal of the American Association for Laboratory Animal Science*, 50(1), 27-32.
- Klita, P.T., Mathison, G.W., Fenton, T.W., and Hardin, R.T. (1996). Effect of alfalfa root saponins on digestive function in sheep. *Journal of Animal Science*, 74, 1144–1156.
- Kung Jr, L., Huber, J. T., Bergen, W. G., and Petitclerc, D. (1984). Amino acids in plasma and duodenal digesta and plasma growth hormone in cows fed varying amounts of protein of differing degradability. *Journal of dairy science*, 67(11), 2519-2524.
- Lajis, N.H., Abdullah, A.S., Salim, S.J., Bremner, J.B., Khan, M.N. (1993). Epi-sarsasapogenin and epi-smilagenin: Two saponins isolated from the

- rumen content of sheep intoxicated by *Brachiaria decumbens*. *Steroids*, 58, 387–389.
- Lascano, C.E., and Euclides, V.P.B. (1996). Nutritional quality and animal production of *Brachiaria* pastures. In *Brachiaria: Biology, Agronomy and Improvement*. Eds. Miles JW, Maass B, do Valle CB pp. 106–123.
- Law, G. J., Ray, K. P., and Wallis, M. (1984). Effects of growth hormone-releasing factor, somatostatin and dopamine on growth hormone and prolactin secretion from cultured ovine pituitary cells. *FEBS letters*, 166(1), 189-193.
- Leal, E. S., Itavo, L. C. V., do Valle, C. B., Itavo, C. C. B. F., Dias, A. M., dos Santos Difante, G., ... and Gurgel, A. L. C. (2020). Influence of protodioscin content on digestibility and in vitro degradation kinetics in *Urochloa brizantha* cultivars. *Crop and Pasture Science*, 71(3), 278-284.
- Lelis, D. L., Rennó, L. N., Chizzotti, M. L., Pereira, C. E. R., Silva, J. C. P., Moreira, L. G. T., ... and Chizzotti, F. H. M. (2018). Photosensitization in naïve sheep grazing signal grass (*Brachiaria decumbens*) under full sunlight or a silvopastoral system. *Small Ruminant Research*, 169, 24-28.
- Lemos, R.A.A., Ferreira, L.C.L., Silva, S.M., Nakazato, L., and Salvador, S.C. (1996b). Photosensitization and Cholangioathy associated with crystal in sheep grazing. *Ciencia Rural*, 26, 109-113.
- Lemos, R.A.A., Nakazato, L., Herrero Jr, G.O., Silveira, A.C., and Porfírio, L.C. (1998). Photosensitization and crystal-associated cholangiopathy in goats kept under *Brachiaria decumbens* pastures in Mato Grosso do Sul. *Ciencia Rural*, 28, 507-510.
- Lila, Z. A., Mohammed, N., Kanda, S., Kurihara, M., and Itabashi, H. (2005). Sarsaponin effects on ruminal fermentation and microbes, methane production, digestibility and blood metabolites in steers. *Asian-australasian Journal of Animal Sciences*, 18(12), 1746-1751.
- Lila, Z., Mohammed, N., Kanda, S., Kamada, T., and Itabashi, H. (2003). Effect of sarsaponin on ruminal fermentation with particular reference to methane production in vitro. *Journal of Dairy Science*, 86, 3330-3336.
- Liu, C., Qu, Y. H., Guo, P. T., Xu, C. C., Ma, Y., and Luo, H. L. (2018). Effects of dietary supplementation with alfalfa (*Medicago sativa* L.) saponins on lamb growth performance, nutrient digestibility, and plasma parameters. *Animal Feed Science and Technology*, 236, 98-106.
- Lovett, D. K., Stack, L., Lovell, S., Callan, J., Flynn, B., Hawkins, M., and O'Mara, F. P. (2006). Effect of feeding *Yucca schidigera* extract on performance of lactating dairy cows and ruminal fermentation parameters in steers. *Livestock Science*, 102(1-2), 23-32.

- Low, S. G. (2000). *Problems Associated with Signal Grass (*Brachiaria decumbens*) Pastures in the Tropical Lowlands of Papua New Guinea.* University of New England.
- Low, S.G. (1993). Photosensitization of cattle grazing Signal grass (*Brachiaria decumbens*) in Papua New Guinea. *New Zealand Veterinary Journal*, 41, 220–221.
- Low, S.G. (2015). Signal grass (*Brachiaria decumbens*) toxicity in grazing ruminants. *Agriculture*, 5, 971–990.
- Lu, C.D., and Jorgensen, N.A. (1987). Alfalfa saponins affect site and extent of nutrient digestion in ruminants. *The Journal of Nutrition*, 117, 919–927.
- Mabjeesh, S. J., Cohen, M., and Arieli, A. (2000). In vitro methods for measuring the dry matter digestibility of ruminant feedstuffs: comparison of methods and inoculum source. *Journal of dairy science*, 83(10), 2289-2294.
- Mader, T.L., and Brumm, M.C. (1987). Effect of feeding sarsaponin in cattle and swine diets. *Journal of Animal Science*, 65, 9–15.
- Makkar, H. P. S. (2003). Effects and fate of tannins in ruminant animals, adaptation to tannins, and strategies to overcome detrimental effects of feeding tannin-rich feeds. *Small ruminant research*, 49(3), 241-256.
- Makkar, H. P. S. (2003). Effects and fate of tannins in ruminant animals, adaptation to tannins, and strategies to overcome detrimental effects of feeding tannin-rich feeds. *Small ruminant research*, 49(3), 241-256.
- Makkar, H. P. S., and Becker, K. (1997). Degradation of quillaja saponins by mixed culture of rumen microbes. *Letters in Applied Microbiology*, 25(4), 243-245.
- Makkar, H. P. S., and Becker, K. (2000). Beneficial effects of saponins on animal production. In *Saponins in food, feedstuffs and medicinal plants* (pp. 281-286). Springer, Dordrecht.
- Makkar, H.P.S., Sen, S., Blummel, M., and Becker, K. (1998). Effects of fractions containing saponins from *Yucca schidigera*, *Quillaja saponaria* and *Acacia auriculiformis* on rumen fermentation. *Journal of Agricultural and Food Chemistry*, 46, 4324–4328.
- Mavule, B. S., Muchenje, V., Bezuidenhout, C. C., and Kunene, N. W. (2013). Morphological structure of Zulu sheep based on principal component analysis of body measurements. *Small Ruminant Research*, 111(1-3), 23-30.
- Meagher, L. P., Smith, B. L., and Wilkins, A. L. (2001). Metabolism of diosgenin-derived saponins: implications for hepatogenous photosensitization diseases in ruminants. *Animal Feed Science and Technology*, 91(3-4), 157-170.

- Meale, S.J., Chaves, A.V., Baah, J., and McAllister, T.A. (2012). Methane production of different forages in in vitro ruminal fermentation. *Asian-Australasian Journal of Animal Science*, 25, 86–91.
- Menke, K. H. (1988). Estimation of the energetic feed value obtained from chemical analysis and in vitro gas production using rumen fluid. *Animal research and development*, 28, 7-55.
- Menke, K.H., Raab, L., Salewski, A., Steingass, H., Fritz, D., and Schneider, W. (1979). The estimation of the digestibility and metabolizable energy content of ruminant feedstuffs from the gas production when they are incubated with rumen liquor in vitro. *Journal of Agricultural Science (Camb)*, 92, 217-222.
- Miles, C.O., Wilkins, A.L., Munday, S.C., Flayen, A., Holland, P.T., and Smith, B.L. (1993). Identification of insoluble salts of the beta-D-glucuronides of episarsasapogenin and epismilagenin in the bile of lambs with alveld and examination of *Narthecium ossifragum*, *Tribulus terrestris*, and *Panicum miliaceum* for sapogenins. *Journal of*
- Mirza, I.H., Khan, A.G., Azim, A., and Mirza, M.A. (2002). Effect of supplementing grazing cattle calves with urea-molasses blocks, with and without *Yucca schidigera* extract, on performance and carcass traits. *Asian-Australasian Journal of Animal Science*, 15, 1300–1306.
- Muniandy, K.V., Chung, E.L.T., Jaapar, M.S., Hamdan, M.H.M., Salleh, A., Jesse, F.F.A. (2020). Filling the gap of *Brachiaria decumbens* (signal grass) research on clinico-pathology and haemato-biochemistry in small ruminants: a review. *Toxicon*, 174, 26-31.
- Nasri, S., Salem, H. B., Vasta, V., Abidi, S., Makkar, H. P. S., and Priolo, A. (2011). Effect of increasing levels of *Quillaja saponaria* on digestion, growth and meat quality of Barbarine lamb. *Animal Feed Science and Technology*, 164(1-2), 71-78.
- Ogah, D. M., Yakubu, A., Momoh, M. O., and Dim, N. I. (2011). Relationship between some body measurements and live weight in adult Muscovy ducks using path analysis. *Trakia Journal of Sciences*, 9(1), 58-61.
- Ørskov, E. R., and McDonald, I. (1979). The estimation of protein degradability in the rumen from incubation measurements weighted according to rate of passage. *The Journal of Agricultural Science*, 92(2), 499-503.
- Patra, A. K., & Saxena, J. (2009). Dietary phytochemicals as rumen modifiers: a review of the effects on microbial populations. *Antonie van Leeuwenhoek*, 96(4), 363-375.
- Patra, A.K., Kamra, D.N., and Agarwal, N. (2006). Effect of plant extracts on in vitro methanogenesis, enzyme activities and fermentation of feed in rumen liquor of buffalo. *Animal Feed Science and Technology*, 128, 276–291.

- Pell, J. M., and Bates, P. C. (1990). The nutritional regulation of growth hormone action. *Nutrition research reviews*, 3(1), 163-192.
- Pen, B., Sar, C., Mwenya, B., Kuwaki, K., Morikawa, R., and Takahash, J. (2006). Effects of *Yucca schidigera* and *Quillaja saponaria* extracts on in vitro ruminal fermentation and methane emission. *Animal Feed Science and Technology*, 129, 175–186.
- Pen, B., Takaura, K., Yamaguchi, S., Asa, R., and Takahashi, J. (2007). Effects of *Yucca schidigera* and *Quillaja saponaria* with or without  $\beta$  1–4 galacto-oligosaccharides on ruminal fermentation, methane production and nitrogen utilization in sheep. *Animal Feed Science and Technology*, 138(1), 75-88.
- Potter, S.M., Jimenez-Flores, R., Pollack, J., Lone, T.A., and Berber-Jimenez, M.D. (1993). Protein saponin interaction and its influence on blood lipids. *Journal of Agricultural and Food Chemistry*, 41, 1287-1291.
- Quinn, J. C., Kessell, A., and Weston, L. A. (2014). Secondary plant products causing photosensitization in grazing herbivores: Their structure, activity and regulation. *International Journal of Molecular Sciences*, 15(1), 1441-1465.
- Ramírez-Restrepo, C. A., Tan, C., López-Villalobos, N., Padmanabha, J., Wang, J., and McSweeney, C. S. (2016). Methane production, fermentation characteristics, and microbial profiles in the rumen of tropical cattle fed tea seed saponin supplementation. *Animal Feed Science and Technology*, 216, 58-67.
- Reh, C.S., and Geffner, M.E. (2010). Somatotropin in the treatment of growth hormone deficiency and Turner syndrome in pediatric patients: a review. *The Journal of Clinical Pharmacology*, 2, 111–122.
- Riet-Correa, B., Castro, M., Lemos, R., Riet-Correa, G., Mustafa, V., and Riet-Correa, F. (2011). Brachiaria spp. poisoning of ruminants in Brazil. *Pesquisa Veterinaria Brasileira*, 31, 183-192.
- Rueda, B.L., Blake, R.W., Nicholson, C.F., Fox, D.G., Tedeschi, L.O., Pell, A.N., Fernandes, E.C.M., Valentim, J.F., and Carneiro, J.C. (2003). Production and economic potentials of cattle in pasture-based system in western Amazon region of Brazil. *Journal of Animal Science*, 81, 2923-2937.
- Russell, J. B., and Rychlik, J. L. (2001). Factors that alter rumen microbial ecology. *Science*, 292(5519), 1119-1122.
- Salako, A. E. (2006). Application of morphological indices in the assessment of type and function in sheep. *Int. J. Morphol*, 24(1), 13-18.
- Salam-Abdullah, A., Lajis, N.H., Bremner, J.B., Davies, N.W., Mustapha, W., and Rajion, M.A. (1992). Hepatotoxic constituents in the rumen of *Brachiaria*

- decumbens* intoxicated sheep. *Veterinary and Human Toxicology*, 34, 154-155.
- Santoso, B., & Hariadi, B. T. (2007). Effect of supplementation of *Acacia mangium* Willd in *Pennisetum purpureum* on fermentation characteristic and methane production in vitro. *Med. Pet.*, 30(2), 106-113.
- Santoso, B., Kilmaskossu, A., and Sambodo, P. (2007). Effects of saponins from *Biophytum petersianum* Klotzsch on ruminal fermentation, microbial protein synthesis and nitrogen utilization in goats. *Animal Feed Science and Technology*, 137, 58-68.
- Santoso, B., Mwenya, B., Sar, C., Gamo, Y., Kobayashi, T., Morikawa, R., ... and Takahashi, J. (2004). Effects of supplementing galacto-oligosaccharides, *Yucca schidigera* or nisin on rumen methanogenesis, nitrogen and energy metabolism in sheep. *Livestock Production Science*, 91(3), 209-217.
- Souza, M. A., Detmann, E., Paulino, M. F., Sampaio, C. B., Lazzarini, I., and Valadares Filho, S. C. (2010). Intake, digestibility and rumen dynamics of neutral detergent fibre in cattle fed low-quality tropical forage and supplemented with nitrogen and/or starch. *Tropical Animal Health and Production*, 42(6), 1299-1310.
- Suhaimi, D., Sharif, S., Normah, M.A., Norain Nadia, M., Wan Syahidah, H. (2017). Estimating relative feed value of local *Brachiaria decumbens*. *Malaysian Journal of Veterinary Research*, 8, 78-87.
- Suharti, S., Astuti, D. A., Wina, E., & Toharmat, T. (2011). Rumen microbial population in the in vitro fermentation of different ratios of forage and concentrate in the presence of whole lerak (*Sapindus rarak*) fruit extract. *Asian-Australasian Journal of Animal Sciences*, 24(8), 1086-1091.
- Taye, M., Abebe, G., Gizaw, S., Lemma, S., Mekoya, A., & Tibbo, M. (2010). Growth performances of Washera sheep under smallholder management systems in Yilmanadensa and Quarit districts, Ethiopia. *Tropical animal health and production*, 42(4), 659-667.
- Thalib, A., Widiawati, Y., Hamid, H., Suherman, D., and Sabrani, M. (1996). The effects of saponin from *Sapindus rarak* fruit on rumen microbes and performance of sheep. *Jurnal*
- Tilley, J.M.A., and Terry, R.A.A. (1963). Two-Stage Technique for in Vitro Digestion of Forages Crops. *Journal of the British Grassland Society*, 18, 104-111.
- Tuggle, C.K., and Trenkle, A. (1996). Control of growth hormone synthesis. *Domestic Animal Endocrinology*, 13, 133.
- Unnawong, N., Cherdthong, A., and So, S. (2021). Crude saponin extract from *Sesbania grandiflora* (L.) Pers pod meal could modulate ruminal

- fermentation, and protein utilization, as well as mitigate methane production. *Tropical Animal Health and Production*, 53(2), 1-9.
- Valdez, F.R., Bush, L.J., Goetsch, A.L., and Owens, F.N. (1986). Effect of steroid saponins on ruminal fermentation and on production of lactating dairy cows. *Journal of Dairy Science*, 69, 1568-1575.
- Van Soest, P. J. (1994). *Nutritional ecology of the ruminant*. Cornell university press.
- Van Soest, P. V., Robertson, J. B., and Lewis, B. (1991). Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. *Journal of dairy science*, 74(10), 3583-3597.
- Vendramini, J.M.B., Sollenberger, L.E., Soares, A.B., Silva, W.L., Sanchez, J.M.D., Valente, A.L., Aguiar, A.D., and Mullenix, M.K. 2014. Harvest frequency affects herbage accumulation and nutritive value of brachiaria grass hybrids in Florida. *Tropical Grassland-Forrajes Tropicales*, 2, 197-206.
- Villalba, J. J., Provenza, F. D., K Clemensen, A., Larsen, R., and Juhnke, J. (2011). Preference for diverse pastures by sheep in response to intraruminal administrations of tannins, saponins and alkaloids. *Grass and Forage Science*, 66(2), 224-236.
- Wanapat, M., Kongmun, P., Poungchompu, O., Cherdthong, A., Khejornsart, P., Pilajun, R., and Kaenpakdee, S. (2012). Effects of plants containing secondary compounds and plant oils on rumen fermentation and ecology. *Tropical Animal Health and Production*, 44(3), 399-405.
- Wang, C.J., Wang, S.P., and Zhou, H. (2009). Influences of flavomycin, ropadiar, and saponin on nutrient digestibility, rumen fermentation, and methane emission from sheep. *Animal Feed Science and Technology*, 148, 157–166.
- Wang, Y., McAllister, T. A., Yanke, L. J., and Cheeke, P. R. (2000). Effect of steroid saponin from *Yucca schidigera* extract on ruminal microbes. *Journal of Applied Microbiology*, 88(5), 887-896.
- Wang, Y., Mcallister, T.A., Yanke, L.J., Xu, Z.J., Cheeke, P.R., and Cheng, K. (2000). In vitro effects of steroid saponins from *Yucca schidigera* extract on rumen microbial protein synthesis and ruminal fermentation. *Journal of the Science of Food and Agriculture*, 80, 2114-2122.
- Whiteman, P.C., Halim, N.R., Norton, B.W., Hales, and J.W. (1985). Beef production from three tropical grasses in south-eastern Queensland. *Australian Journal of Experimental Agriculture*, 25, 481–488.
- Wina, E., Muetzel, S., & Becker, K. (2006). The dynamics of major fibrolytic microbes and enzyme activity in the rumen in response to short-and long-

- term feeding of *Sapindus rarak* saponins. *Journal of Applied Microbiology*, 100(1), 114-122.
- Wina, E., Muetzel, S., and Becker, K. (2005). The impact of saponins or saponin-containing plant materials on ruminant production: A review. *Journal of Agricultural and Food Chemistry*, 53, 8093–8105.
- Yuliana, P., Laconi, E. B., Wina, E., and Jayanegara, A. (2014). Extraction of tannins and saponins from plant sources and their effects on in vitro methanogenesis and rumen fermentation. *Journal of the Indonesian Tropical Animal Agriculture*, 39(2), 91-97.
- Yusof, A. M., and Isa, M. L. M. (2016). Prevalence of gastrointestinal nematodiasis and coccidiosis in goats from three selected farms in Terengganu, Malaysia. *Asian Pacific journal of tropical biomedicine*, 6(9), 735-739.
- Zheng, C., Zhou, J., Zeng, Y., and Liu, T. (2021). Effects of mannan oligosaccharides on growth performance, nutrient digestibility, ruminal fermentation and hematological parameters in sheep. *PeerJ*, 9, e11631.
- Zhong, R. Z., Yu, M., Liu, H. W., Sun, H. X., Cao, Y., and Zhou, D. W. (2012). Effects of dietary *Astragalus* polysaccharide and *Astragalus membranaceus* root supplementation on growth performance, rumen fermentation, immune responses, and antioxidant status of lambs. *Animal feed science and technology*, 174(1-2), 60-67.
- Zhou, C. S., Xiao, W. J., Tan, Z. L., Salem, A. Z. M., Geng, M.M., Tang, S. X., ... and Kang, j. h. (2012). Effects of dietary supplementation of tea saponins (*Ilex kudingcha* CJ Tseng) on ruminal fermentation, digestibility and plasma antioxidant parameters in goats. *Animal feed science and technology*, 16(1-4), 163-169.