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OPTIMIZATION OF EXTRACTION METHOD AND IMPROVEMENT OF GROWTH, PHYSIOLOGY, YIELD AND ANTIOXIDANTS PROPERTIES AS AFFECTED BY CHITOSAN IN Ocimum basilicum L.

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QAZIZADAH AHMAD ZUBAIR

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

March 2022

DEDICATION

To my beloved mother who always prays for me day and night to achieve my goal

To my family members:



and

To all my friends who supported me all these years

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

OPTIMIZATION OF EXTRACTION METHOD AND IMPROVEMENT OF GROWTH, PHYSIOLOGY, YIELD AND ANTIOXIDANTS PROPERTIES AS AFFECTED BY CHITOSAN IN *Ocimum basilicum* L.

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March 2022

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Sweet basil (Ocimum basilicum L.) is one of aromatic herbs belonging to Lamiaceae family and is widely used in pharmaceutical, culinary and traditional medicine industries. Sweet basil holds high potential and demand in mentioned industries, but biomass production and phytochemical content are still not sufficient to cater the demand. The quantity of the extraction of phytochemical content is depends on the extraction technique. It is therefore, important to increase biomass production and the content of phytochemicals at field level and establish suitable extraction technique, in order to achieve high amount of extraction. Two experiments were carried out with the objectives of (i) to establish a suitable combination of ethanol concentration and temperature in order to obtain high amount of selected antioxidant constituents from sweet basil leaves, (ii) to identify the suitable concentration of chitosan and time of application for improvement of the growth, herbal yield and antioxidant contents in sweet basil at field. In the first experiment, three different concentrations of ethanol (60, 75 and 90% v/v) were combined with three different temperatures (40, 60 and 80°C) for the extraction of sweet basil leaves. The experiment was arranged in 3 X 3 factorial Complete Randomized Design (CRD) with four replications and three samples per replicate. The result showed that 90% ethanol at 80°C had the highest percentage of extraction yield (11.56%) compared to other treatments. However, extracting dried sweet basil leaves with 60% ethanol at 80°C temperature recorded the highest extraction of total phenolic content (67.02 mg of GAE/g of DE), total phenolic yield (693.5 mg of GAE/100 g of DW), total flavonoid content (44.7 mg of QUE/g of DE), total flavonoid yield (462.52 mg of QUE/100 g of DW) and antioxidant activity (66.8%). Results from the correlation analysis showed a positive relationship between phenolic, flavonoid

and antioxidant activity of the extract. Therefore, it is recommended to extract sweet basil leaves using 60% ethanol under 80°C. This combination was applied in the second experiment. In the second experiment, chitosan at four different concentrations (0, 2, 4 and 6 ml/L) were applied at three different application times of (20, 40 and 20 + 40 days after transplanting, DAT) at the field. The experiment was organized in 4 X 3 factorial Randomized Complete Block Design (RCBD) with four replications and five plants per replicate. The results showed an interaction between chitosan concentrations and times of applying on growth, herbal yield and secondary metabolites performance of sweet basil plants. From the results, application of chitosan at concentration of 4 ml/L on 20 DAT gave the highest biomass yield components such as leaf fresh weight and leaf dry weight by 43.45 and 59.71% as well as total content of flavonoid and antioxidant activity by 40.21 and 22.81%, respectively, when compared with the control group. In addition, the same treatment resulted in the highest value of plant height, stem diameter, dry weight of stem per plant, number of leaves per plant, total leaf area per plant, average chlorophyll-a, chlorophyll-b, total chlorophyll, actual chlorophyll and dry weight of root per plant by 40.73, 37.47, 40.38, 25.71, 37.19, 9.90, 37.36, 18.75, 18.86 and 143.97%, respectively compare to the Besides, correlation analysis showed positive relationship control groups. among variables as well as dry weight of leaves with total leaf area (r = 0.96) and total chlorophyll (r = 0.76), the AA with TPC (r = 0.80) and TFC (r = 0.57). Most of the growth and physiology variables positively correlated with biomass yield components. In conclusion, it is recommended that the application of 60% ethanol at 80°C extraction temperature showed greater strength in extracting phenolic, flavonoid and antioxidant activity from sweet basil's leaf. Meanwhile, application of 90% ethanol at 80°C temperature showed greater percentage of extraction yield in comparison to other treatments. Besides, the performance of sweet basil plants at field level could be improved by the application of 4 ml/L chitosan at 20 DAT.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PENGOPTIMUMAN KAEDAH PENGEKSTRAKAN DAN PENAMBAHBAIKAN PERTUMBUHAN, FISIOLOGI, HASIL DAN BAHAN ANTIOKSIDA TERPILIH YANG DIPENGARUHI OLEH KITOSAN KE ATAS Ocimum basilicum L.

Oleh

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Selasih (Ocimum basilicum L.) adalah salah satu herba aromatik di bawah keluarga Lamiaceae dan digunakan secara meluas di dalam industri farmaseutikal, masakan dan perubatan tradisional. Selasih mempunyai potensi dan permintaan yang tinggi di dalam industri, namun, pengeluaran biojisim serta kandungan fitokimianya masih rendah berbanding permintaan. Kuantiti pengekstrakan kandungan fitokimia terletak kepada teknik pengekstrakan. Oleh itu, adalah penting untuk meningkatkan pengeluaran biojisim serta kandungan fitokimia semasa di peringkat ladang dan juga teknik pengekstrakan yang sesuai, bagi mencapai jumlah pengekstrakan yang tinggi. Dua eksperimen telah dijalankan dengan objektif, (i) untuk mengukur kepekatan etanol dan suhu yang diperlukan bagi mendapatkan kuantiti bahan antioksida t erpiih yang tinggi daripada daun selasih, (ii) untuk mengenal pasti kepekatan kitosan dan masa penggunaan yang sesuai untuk meningkatkan pertumbuhan, hasil herba dan antioksida tanaman selasih di ladang. Di dalam eksperimen pertama, tiga kepekatan etanol yang berbeza (60, 75 dan 90% v/v) telah digabungkan dengan tiga suhu berbeza (40, 60 dan 80°C) untuk pengekstrakan daun selasih. Eksperimen telah disusun dalam bentuk 3 X 3 faktorial Reka Bentuk Rawak Lengkap (CRD). Keputusan menunjukkan bahawa kepekatan etanol pada 90% dan pada suhu 80°C menghasilkan peratusan hasil pengekstrakan yang paling tinggi (11.56%) berbanding rawatan lain. Walau bagaimanapun, pengekstrakan daun selasih dengan 60% etanol pada suhu 80°C mencatatkan pengekstrakan tertinggi bagi jumlah kandungan fenolik (67.02 mg of GAE/g of DE), jumlah hasil fenolik (693.5 mg of GAE/100 g of DW), jumlah kandungan flavonoid (44.7). mg QUE/g DE), jumlah hasil flavonoid (462.52 mg QUE/100 g DW) dan aktiviti

antioksida (66.8%). Keputusan daripada analisis korelasi menunjukkan terdapatnya hubungan positive di antara fenolik, flavonoid dan aktiviti antioksida di dalam ekstrak tersebut. Oleh itu, adalah disyorkan untuk mengekstrak daun selasih dengan menggunakan etanol pada 60% dengan suhu 80°C. Keputusan ini seterusnya digunakan di dalam eksperimen kedua. Di dalam eksperimen kedua, empat kepekatan kitosan yang berbeza (0, 2, 4 dan 6 ml/L) telah digunakan pada tiga masa penggunaan yang berbeza (20, 40 dan 20 + 40 hari selepas pemindahan anak pokok, DAT) di lapangan. Eksperimen ini disusun dalam Reka Bentuk Rawak Blok Lengkap (RCBD) 4 X 3 faktorial. Keputusan menunjukkan terdapatnya interaksi di antara kepekatan kitosan dan masa penggunaannya terhadap hasil herba dan prestasi metabolit sekunder tumbuhan selasih. Daripada keputusan, data dengan hasil tertinggi seperti berat segar daun (54.26 g/pokok), berat kering daun (8.8 g/pokok) serta kandungan flavonoid (33.23 mg QUE/g DE) dan aktiviti antioksidan (92.34%) adalah diperolehi daripada penggunaan 4 ml/L kitosan pada 20 DAT, jika dibandingkan dengan kawalan (37.84 g/pokok, 5.51 g/pokok, 23.70 mg QUE/g DE dan 75.18%), masing-masing. Selain itu, rawatan yang sama ini telah menghasilkan ketinggian tumbuhan (55.08 cm), diameter batang (11.08 mm), bilangan daun (296.57), jumlah luas daun (2234.31 cm²), purata klorofil-a (4.33). mg/cm²), klorofil-b (2.5 mg/cm²), jumlah klorofil (6.84 mg/cm²), klorofil sebenar (4.79 mg/cm²), berat kering batang (11.09 g/pokok) dan berat kering akar (2.83 g/pokok). Selain itu, analisis korelasi juga turut menunjukkan hubungan positif antara pembolehubah serta berat kering daun dengan jumlah luas daun (r = 0.96) dan jumlah klorofil (r = 0.76), AA dengan TPC (r = 0.80) dan TFC (r = 0.57). Kesimpulannya, adalah disyorkan bahawa penggunaan 60% etanol pada suhu pengekstrakan 80°C mampu mengekstrak fenolik, flavonoid dan aktiviti antioksidan yang paling tinggi daripada daun selasih. Sementara itu, penggunaan 90% etanol pada suhu 80°C menunjukkan peratusan hasil pengekstrakan yang lebih besar berbanding rawatan lain. Di samping itu, prestasi tumbuhan selasih di peringkat ladang boleh dipertingkatkan dengan penggunaan 4 ml/L kitosan pada 20 DAT.

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TA₁=Time of Application at 20 DAT, TA₂=Time of Application at 40 DAT and TA₃=Time of Application at 20 + 40 DAT, and C₀=Chitosan 0 ml /L, C₁=Chitosan 2 ml/L, C₂=Chitosan 4 ml/L and C₃=Chitosan 6 ml/L, the red line separates the underground and above ground parts of the stems.

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LIST OF ABBREVIATIONS

ROS	Reactive oxygen species
EY	Extraction yield (%)
TPC	Total phenolic content
mg of GAE/g of DE	Milligram gallic acid equivalent per gram of dry extract
TPY	Total phenolic yield (mg of GAE/100 g of DW)
mg GAE/100 g DW	Milligram gallic acid equivalent per 100 gram of dry weight
TFC	Total flavonoid content (mg of QUE/g of WE)
mg of QUE/g of WE	Milligram of quercetin equivalent per gram of dry extract
TFY	Total flavonoid yield (mg of QUE/100 g of DW)
mg of QUE/100 g of DW	Milligram of quercetin equivalent per 100 gram of dry weight
AA	Antioxidant activity (%)
LSD	Least significant difference
СНІ	Chitosan
ТА	Times of application
DAT	Days after transplanting

CHAPTER 1

INTRODUCTION

1.1 Introduction

Sweet basil (*Ocimum basilicum* L.) is a common herb which is extensively consumed in pharmaceutical, cosmetic, nutraceutical and beverage industries. Because of its strong aroma and a wide range of beneficial properties, sweet basil is also known as "King of Aromatic Herbs" (Filip, 2017). Sweet basil consists of more than 200 secondary metabolite bioactive compounds (Ghasemzadeh et al., 2016). However, phenolics and flavonoids are the most important groups of antioxidant compounds, involving health beneficial properties in sweet basil (Filip, 2017; Ghasemzadeh et al., 2016; Koca and Karaman, 2015).

Although sweet basil contains various types of phytochemicals, however, good extraction method is the key to extracting high phytochemicals content from the leaves. According to Dorta et al. (2012) the solvent concentration and temperature used during the extraction are two essential factors in achieving high extraction yield. Khoddami et al. (2013) stated that phenolic compounds could be easily extracted using organic solvents. Among the organic solvents, it was said that ethanol is the most inexpensive solvent to be used in extraction (Chiaramonti et al., 2012), nontoxic to human body (Casagrande et al., 2018) and effective in extracting the phytochemicals from plant materials (Truong et al., 2019; Chigayo et al., 2016; Dent et al., 2013). It was reported that ethanol at 62.7% concentration was able to extract high phenolic compounds (7.21 g of GAE/g) with 80% of antioxidant activity at 49.7°C in Feronia limonia fruit (Ilaiyaraja et al., 2015). This is supported by Hassan et al. (2021) where low concentration of ethanol (60%) recorded to extract the highest phenolic compounds (807.20 mg of GAE/g) from Padina australis. However, another researcher proved that high concentration of ethanol (90%) yielded better result in extracting those antioxidants in Toona sinensis (Maulana et al., 2019). Due to high variations in concentration that should be used, it is then important to test the concentration on a specific plant.

Apart from this, earlier report by Su et al. (2017) and Dent et al. (2013) revealed that temperature plays a vital role during the extraction of phytochemical. The use of higher temperature during the extraction can help in enhancing the solvent to enter the cells and stimulate the solubility of bioactive compounds which lead to the increase in extraction efficiency (Najafabadi et al., 2020; Su et al., 2017; Setford et al., 2017). However, the optimum extraction temperature is not same for all plants. This is proven when Juntachote et al. (2006) tested different extraction temperatures on different crops. They found that suitable temperature for extracting phenolic compounds from *Cymbopogon citratus* was 25 °C, while from *Alpinia galanga* plants was 75 °C. Similar finding was also obtained by

Oreopoulou et al, (2019) and Milevskaya et al. (2019) where different crops required different temperatures to extract the same compound. Thus, it is important to test different temperatures on different plant species. Besides, in order to increase the efficiency in the extract, Ilaiyaraja et al. (2015) suggested to combine the temperature with different percentage of ethanol. This is agreed by Dorta et al. (2012) who stated that the efficiency of extraction process depends on solvent concentration and level of extraction temperature.

Furthermore, as sweet basil is rich in phytochemical compounds, there is an increase in the number of products that use sweet basil as part of the ingredients. There were several attempts from previous researchers on methods to increase the number of leaves in plants other than sweet basil, which were by using auxin, fertilization and managing the light intensity (Jebapriya and Somasundaram, 2021; Uzun, 2006; Dieleman and Heuvelink, 1992). However, these methods are costly, time consuming and hard to maintain.

Alternatively, one of the growth regulators that is gaining more attention in agriculture is chitosan. Chitosan is one of the plant regulators that is safe, easily available and low-cost (Khan, et al., 2018; Emami-Bistgani, et al., 2017). It is said that chitosan increases growth performances in several medicinal herbs such as *Carum copticum* L. (Razavizadeh et al., 2020), *Pisum sativum* (Khan et al., 2018) and *Salvia officinalis* L. (Vosoughi et al., 2018). However, the effectiveness of chitosan was reported to be dependent on its concentration (Irawati et al., 2019; EI-Miniawy et al., 2013; Abdel-Mawgoud, 2010) and growing stage of the plants (Rasheed et al., 2020; Pichyangkura and Chadchawan, 2015). Lei et al. (2011) stated that 100 mg/L of chitosan produced higher biosynthesis of phytochemicals in *Artemisia annua* herbs, while maintaining plant growth and development. However, Singh (2016) stated that low concentration at 10 mg/L was sufficient to obtain the highest accumulation of phytochemicals in *Spinacia oleracea*.

In terms of plant's growing stage, Ali et al. (1997) suggested that it is recommended to apply chitosan at later growing stage which is approximately 42 days after planting compared to early growing stage (28 days after planting) in *Glycine max* Merr. Nevertheless, this is disagreed by Cuibu and Shiayama (2001) who recommended to applying chitosan at the early growing stage of *Glycine max*, *Solanum lycopersicum* and *Oryza sativa* where the plant height, leaf surface area, chlorophyll contents and dry plant materials are higher. This is also supported by Mondal et al. (2016) who stated applying chitosan at 25 days after transplanting was more suitable compared to 35 DAT in *Solanum lycopersicum*. Besides, Ohta et al. (2004) stated that higher accumulation of phytochemicals was found when applying chitosan at early growing stage in *Torenia fournieri, Calceolaria herbeohybrida* and *Campanula fragilis* L. plants. Other than affecting the plant's growth and development, chitosan has also been described to increase the biosynthesis of phytochemicals in several plants (Silva et al., 2020; Jiao et al., 2018 and Talón et al., 2017).

1.2 **Problem statement**

Today providing antioxidants as well as in natural form is very challenging for the researchers (Mairapetyan and Mamikonyan, 2016). Medicinal herbs are the most important source of natural antioxidant compounds. Due to this, the demand for this crops in global market will be increased from \$6.2 billion to \$5 trillion till 2050 (Govindaraju and Arulselvi, 2018; Rao et al., 2017; Khanna, 2015; Kumar and Janagam, 2011). Particularly, sweet basil as a common mostly consumed herb is highly contributing to this demand (Rahman et al., 2021; Dou et al., 2018; Liaros et al., 2016). This demand is including both biomass (as spice) and phytochemicals as well as antioxidant compounds. As king of medicinal herbs, it is needed to increase leaf yield and phytochemicals in sweet basil in order to cater the demand. In literature, attempts were introduced to increase leaf yield, however, they were not economic, easy to maintain and ecofriendly.

On the other hand, the quantity of phytochemicals lies on the efficiency extraction technique. In extraction process, concentration of solvent and level of temperature are key factors and needs to be optimized since they are very from crop to crop.

Therefore, the problem is how to improve biomass and phytochemical production of sweet basil at field, and establish suitable extraction technique in order to obtain optimum phytochemicals with higher antioxidant property from sweet basil leaves.

1.3 Research objectives

The main objective of this study was to improve herbal yield and phytochemical production of sweet basil at field, and optimize extraction of its phytochemicals. Hence, the specific objectives of this study were as follows:

- 1) To establish a suitable combination of ethanol concentration and temperature in order to obtain high amount of selected antioxidant constituents from sweet basil leaves.
- To identify the suitable concentration of chitosan and time of application for improvement of the growth, herbal yield and antioxidant contents in sweet basil at field.

1.4 Significance of the study

This study provides new insights in to the easy, economic and ecofriendly attempt to improve production of biomass and antioxidant phytochemicals of

sweet basil at filed. Besides of that, as a result of this study suitable extraction technique will be established. While the first attempt will benefit farmers who are seeking for easy way to increase quality and quantity of sweet basil's production as well as in Malaysia. The second attempt will contribute to medicinal and food industries who extract and process antioxidant phytochemicals of sweet basil in all over the world. As a result, practical application of findings from this study will be a great contribution to meet the increasing demand for antioxidant resources. Moreover, the analysis that presented in this study will convey valuable information for future research that will explore various phytochemicals from sweet basil.



REFERENCES

- Aadesariya, M. K., Ram, V. R., and Dave, P. N. (2017). Evaluation of antioxidant activities by use of various extracts from *Abutilon pannosum* and *Grewia tenax* in the Kachchh Region. *MOJ Food Process and Technology*, 5 (1), 00116.
- Abas, F., Lajis, N. H., Israf, D. A., Khozirah, S., and Kalsom, Y. U. (2006). Antioxidant and Nitric Oxide Inhibition Activities of Selected Malay Traditional Vegetables. *Food Chemistry*, 95 (4), 566-573.
- Abdel-aziz, H. M. M. (2019). Effect of Priming with Chitosan Nanoparticles on Germination, Seedling Growth and Antioxidant Enzymes of Broad Beans. *Egypcian Society for Environmental Science Effect* 18 (1), 81–86.
- Abdel-Mawgoud, A. M. R., Tantawy, A. S., El-Nemr, M. A., and Sassine, Y. N. (2010). Growth and Yield Responses of Strawberry Plants to Chitosan Application. *European Journal of Scientific Research*, 39 (1), 170-177.
- Abdel-Wahed, G. A., and Shaker, A. M. (2020). Management Powdery Mildew of Marigold (*Calendula officinalis*) Using Some Nanoparticles and Chitosan. *Journal of Plant Protection and Pathology*, 11 (9), 435-440.
- Abu-Bakar, F. I., Abu Bakar, M. F., Abdullah, N., Endrini, S., and Fatmawati, S. (2020). Optimization of Extraction Conditions of Phytochemical Compounds and Anti-gout Activity of *Euphorbia hirta* L. (Ara Tanah) Using Response Surface Methodology and Liquid Chromatography-mass Spectrometry (LC-MS) Analysis. *Evidence-Based Complementary and Alternative Medicine*, 2020.
- Acemi, A., Bayrak, B., Çakır, M., Demiryürek, E., Gün, E., El Gueddari, N. E., and Özen, F. (2018). Comparative Analysis of the Effects of Chitosan and Common Plant Growth Regulators on in Vitro Propagation of *Ipomoea purpurea* (L.) Roth from nodal explants. *In Vitro Cellular & Developmental Biology-Plant*, 54 (5), 537-544.
- Achat, S., Rakotomanomana, N., Madani, K., and Dangles, O. (2016). Antioxidant Activity of Olive Phenols and Other Dietary Phenols in Model Gastric Conditions: Scavenging of the Free Radical DPPH and Inhibition of the Haem induced Peroxidation of Linoleic Acid. *Food chemistry*, 213, 135-142.
- Adiaha, M. S., and Agba, O. A. (2016). Influence of Different Methods of Fertilizer Application on the Growth of Maize (*Zea mays* L.). for Increase Production in South Nigeria. *World Scientific News*, 54, 73-86.
- Agbodjato, N. A., Noumavo, P. A., Adjanohoun, A., Agbessi, L., and Baba-Moussa, L. (2016). Synergistic Effects of Plant Growth Promoting

Rhizobacteria and Chitosan on in Vitro Seeds Germination, Greenhouse Growth, and Nutrient Uptake of Maize (*Zea mays* L.). *Biotechnology Research International.*

- Agriculture Forestry and Fisheries, Republic of South Africa. (2012). Basil Production. Directorate Communication Services – *Department of Agriculture Forestry and Fisheries, South Africa*. P: 1-13.
- Ahamed, T. E. S. (2019). Bioprospecting Elicitation with Gamma Irradiation Combine with Chitosan to Enhance, Yield Production, Bioactive Secondary Metabolites and Antioxidant Activity for Saffron. *Journal of Plant Sciences*, 7 (6), 137.
- Ahmad, B., Khan, M. M. A., Jaleel, H., Sadiq, Y., Shabbir, A., and Uddin, M. (2017). Exogenously Sourced γ-irradiated Chitosan-mediated Regulation of Growth, Physiology, Quality Attributes and Yield in *Mentha piperita* L. *Turkish Journal of Biology*, 41 (2), 388-401.
- Ahmed, K. B. M., Khan, M. M. A., Siddiqui, H., and Jahan, A. (2020). Chitosan and its Oligosaccharides, a Promising Option for Sustainable Crop Production a Review. *Carbohydrate Polymers*, 227, 115331.
- Akbari, G. A., Soltani, E., Binesh, S., and Amini, F. (2018). Cold Tolerance, Productivity and Phytochemical Diversity in Sweet Basil (Ocimum basilicum L.) Accessions. Industrial Crops and Products, 124, 677-684.
- Al Harthy, Khalid Mubarak Saleh (2019) *Growth and Physiological Responses* of Banana (Musa spp.) under Different Levels of Salinity and Organic Fertilizer in the Northern Sultanate of Oman. Doctoral thesis, Universiti Putra Malaysia.
- Alara, O. R., Abdurahman, N. H., Mudalip, S. K. A., and Olalere, O. A. (2018). Characterization and Effect of Extraction Solvents on the Yield and Total Phenolic Content from Vernonia amygdalina Leaves. Journal of Food Measurement and Characterization, 12 (1), 311-316.
- Alberti, A., Zielinski, A. A. F., Zardo, D. M., Demiate, I. M., Nogueira, A., and Mafra, L. I. (2014). Optimisation of the Extraction of Phenolic Compounds from Apples Using Response Surface Methodology. *Food Chemistry*, 149, 151-158.
- Ali, M., Horiuchi, T., and Miyagawa, S. (1997). Nodulation, Nitrogen Fixation and Growth of Soybean Plants (*Clycine max* Merr.) in Soil Supplemented with Chitin or Chitosan. *Japanese Journal of Crop Science*, 66 (1), 100-107.
- Almeida, L., da Silva, E. M., Magalhaes, P., Karam, D., dos Reis, C. O., Gomes Júnior, C. C., and Marques, D. (2020). Root System of Maize Plants Cultivated under Water Deficit Conditions with Application of Chitosan. *Embrapa Milho e Sorgo-Artigo em periódico indexado (ALICE)*.

- Al-Tawaha, A. R., Turk, M. A., Al-Tawaha, A. R. M., Alu'datt, M. H., Wedyan, M., Al-Ramamneh, E. A. D. M., and Hoang, A. T. (2018). Using Chitosan to Improve Growth of Maize Cultivars under Salinity Conditions. *Bulg J Agric Sci*, 24 (3), 437-442.
- Alvarez, M. A. (2014). Plant Secondary Metabolism. In *Plant Biotechnology for Healt*, Springer, Cham: pp. 15-31.
- Amine, R., Abla, E. H., Mohammed, B. I., and Khadija, O. (2020). The Amendment with Chitin and/or Chitosan Improves the Germination and Growth of Lycopersicon esculentum L., Capsicum annuum L. and Solanum melongena L. Indian Journal of Agricultural Research, 54 (4).
- Anusuya, S., and Sathiyabama, M. (2016). Effect of Chitosan on Growth, Yield and Curcumin Content in Turmeric under Field Condition. *Biocatalysis and agricultural biotechnology*, 6, 102-106.
- Arceusz, A., Wesolowski, M., and Konieczynski, P. (2013). Methods for Extraction and Determination of Phenolic Acids in Medicinal Plants: A Review. *Natural Product Communications*, Vol. 8 No. (12) 1821 – 1829.
- Aryal, S., Baniya, M. K., Danekhu, K., Kunwar, P., Gurung, R., and Koirala, N. (2019). Total Phenolic Content, Flavonoid Content and Antioxidant Potential of Wild Vegetables from Western Nepal. *Plants*, 8 (4), 96.
- Aryal, S., Baniya, M. K., Danekhu, K., Kunwar, P., Gurung, R., and Koirala, N. (2019). Total Phenolic Content, Flavonoid Content and Antioxidant Potential of Wild Vegetables from Western Nepal. *Plants*, 8 (4), 96.
- Ashraf, I., Zubair, M., Rizwan, K., Rasool, N., Jamil, M., Khan, S. A., ... and Jaafar, H. Z. (2018). Chemical Composition, Antioxidant and Antimicrobial Potential of Essential Oils from Different Parts of *Daphne mucronata* Royle. *Chemistry Central Journal*, 12 (1), 1-8.
- Ashraf, M. A., Ashraf, M., and Ali, Q. (2010). Response of Two Genetically Diverse Wheat Cultivars to Salt Stress at Different Growth Stages: Leaf Lipid Peroxidation and Phenolic Contents. *Pak J Bot*, 42 (1), 559-565.
- Atait, M., and Qureshi, U. S. (2020). Efficacy of Different Primers on Growth and Yield of Tulip (*Tulipa gesneriana* L.). *World Journal of Biology and Biotechnology*, 5 (2), 31-35.
- Aung, K. K., and Htun, K. T. (2020) Preparations and Characterizations of Chitosan, ZnO Nanoparticle and Chitosan-ZnO Nanocomposite. *Journal of the Myanmar Academy of Arts and Science*. Vol. XVIII. No.1A

- Avestan, S., Naseri, L., and Barker, A. V. (2017). Evaluation of Nano-silicon Dioxide and Chitosan on Tissue Culture of Apple under Agar-induced Osmotic stress. *Journal of Plant Nutrition*, 40 (20), 2797-2807.
- Baddeley, J. A., and Watson, C. A. (2005). Influences of Root Diameter, Tree Age, Soil Depth and Season on Fine Root Survivorship in *Prunus avium. Plant and Soil*, 276 (1-2), 15-22.
- Barber, S. A. (1979). Growth Requirements for Nutrients in Relation to Demand at the Root Surface. In *The soil–root Interface*, Academic Press: pp. 5-20.
- Barros, N. A., Rocha, R. R., de Assis, A. V. R., and Mendes, M. F. (2013).
 Extraction of Basil Oil (*Ocimum basilicum* L.) Using Supercritical Fluid. In III Ibero-american Conference on Supercritical Fluids. *Cartagena de Indi as, Colombia*.
- Barry, C. S. (2009). The Stay-green Revolution: Recent Progress in Deciphering the Mechanisms of Chlorophyll Degradation in Higher Plants. *Plant Science*, 176 (3), 325-333.
- Basit, A., Alam, M., Ahmad, I., Ullah, I., Alam, N., Ullah, I., ... and Ul Ain, N. (2020). Efficacy of Chitosan on Performance of Tomato (*Lycopersicon esculentum* L.) Plant under Water Stress condition. *Pakistan Journal of Agricultural Research*, 33 (1), 27.
- Becker, T., Schlaak, M., and Strasdeit, H. (2000). Adsorption of Nickel (II), Zinc (II) and Cadmium (II) by New Chitosan Derivatives. *Reactive and Functional Polymers*, 44 (3), 289-298.
- Bekhradi, F., Luna, M. C., Delshad, M., Jordan, M. J., Sotomayor, J. A., Martínez-Conesa, C., and Gil, M. I. (2015). Effect of Deficit Irrigation on the Postharvest Quality of Different Genotypes of Basil Including Purple and Green Iranian Cultivars and a Genovese variety. *Postharvest Biology and Technology*, 100, 127-135.
- Benosman, A., Slimane, S. K., and Roger, P. (2021). Adsorption of Anionic Dye by Cross-linked Chitosan–polyaniline Composites. *Journal of Water Chemistry and Technology*, 43 (1), 14-21.
- Berliana, A. I., Kuswandari, C. D., Retmana, B. P., Putrika, A., and Purbaningsih, S. (2020). Analysis of the Potential Application of Chitosan to Improve Vegetative Growth and Reduce Transpiration Rate in *Amaranthus Hybridus*. *E&ES*, 481 (1), 012021.
- Biro-Janka, B., Nyaradi, I. I., Varban, D. I., Molnar, K., and Duda, M. M. (2019). Comparing the Plant Biomass and the Volatile Oil Content of Sweet Basil (*Ocimum basilicum* L.) Cultivars Grown in Târgu Mureş. *Hop and Medicinal Plants*, 27 (1-2), 96-103.

- Biswas, T. (2018). Estimation of Rosmarinic Acid Content of Genetically Stable Clones of Ocimum basilicum L. (Sweet Basil) Regenerated through in Vitro Shoot Bud Multiplication and Root Culture. Open Access Journal of Medicinal and Aromatic Plants, 9 (2), 34.
- Boonlertnirun, S., Boonraung, C., and Suvanasara, R. (2008). Application of Chitosan in Rice Production. *Journal of Metals, Materials and Minerals*, 18 (2).
- Bouma, T. J., Nielsen, K. L., and Koutstaal, B. A. S. (2000). Sample Preparation and Scanning Protocol for Computerised Analysis of Root Length and Diameter. *Plant and Soil*, 218 (1-2), 185-196.
- Boyer, J., and Liu, R. H. (2004). Apple Phytochemicals and Their Health Benefits. *Nutrition Journal*, 3 (1), 1-15.
- Brian, P. W. (1958). Gibberellic Acid: A New Plant Hormone Controlling Growth and Flowering. *Journal of the Royal Society of Arts*, 106 (5022), 425-441.
- Bridgers, E. N., Chinn, M. S., and Truong, V. D. (2010). Extraction of Anthocyanins from Industrial Purple-fleshed Sweet Potatoes and Enzymatic Hydrolysis of Residues for Fermentable Sugars. *Industrial Crops and Products*, 32 (3), 613-620.
- Bubalo, M. C., Ćurko, N., Tomašević, M., Ganić, K. K., and Redovniković, I. R. (2016). Green Extraction of Grape Skin Phenolics by Using Deep Eutectic Solvents. *Food Chemistry*, 200, 159-166.
- Bucktowar, K., Bucktowar, M., and Bholoa, L. D. (2016). A Review on Sweet Basil Seeds: *Ocimum basilicum* (L.). *World Journal of Pharmacy and Pharmaceutical Sciences*, 5 (12), 554-567.
- Burducea, M., Zheljazkov, V. D., Dincheva, I., Lobiuc, A., Teliban, G. C., Stoleru, V., and Zamfirache, M. M. (2018). Fertilization Modifies the Essential Oil and Physiology of Basil Varieties. *Industrial Crops and Products*, 121, 282-293.
- Cabrera, J. C., Messiaen, J., Cambier, P., and Van Cutsem, P. (2006). Size, Acetylation and Concentration of Chito-oligosaccharide Elicitors Determine the Switch from Defence Involving PAL Activation to Cell Death and Water Peroxide Production in *Arabidopsis* Cell Suspensions. *Physiologia Plantarum*, 127, 44–56.
- Cacace, J. E., and Mazza, G. (2003). Optimization of Extraction of Anthocyanins from Black Currants with Aqueous Ethanol. *Journal of Food Science*, 68 (1), 240-248.

- Cacace, J. E., and Mazza, G. (2003). Optimization of Extraction of Anthocyanins from Black Currants with Aqueous Ethanol. *Journal of Food Science*, 68 (1), 240-248.
- Casagrande, M., Zanela, J., Júnior, A. W., Busso, C., Wouk, J., Iurckevicz, G., ... and Malfatti, C. R. M. (2018). Influence of Time, Temperature and Solvent on the Extraction of Bioactive Compounds of *Baccharis dracunculifolia*: In Vitro Antioxidant Activity, Antimicrobial Potential, and Phenolic Compound Quantification. *Industrial Crops and Products*, 125, 207-219.
- Ch, M. A., Naz, S. B., Sharif, A., Akram, M., and Saeed, M. A. (2015). Biological and Pharmacological Properties of the Sweet Basil (*Ocimum basilicum* L.). *Journal of Pharmaceutical Research International*, 330-339.
- Chakraborty, M., Karun, A., and Mitra, A. (2009). Accumulation of Phenylpropanoid Derivatives in Chitosan-induced Cell Suspension Culture of (*Cocos nucifera*). *Journal of Plant Physiology*, 166 (1), 63-71.
- Chamnanmanoontham, N., Pongprayoon, W., Pichayangkura, R., Roytrakul, S., and Chadchawan, S. (2015). Chitosan Enhances Rice Seedling Growth via Gene Expression Network Between Nucleus and Chloroplast. *Plant Growth Regulation*, 75 (1), 101-114.
- Chandrkrachang S (2002) The Applications of Chitin in Agriculture in Thailand. *Advances in Chitin Science* 5, 458-462.
- Chang, X., Alderson, P., and Wright, C. (2005). Effect of Temperature Integration on the Growth and Volatile Oil Content of Basil (*Ocimum basilicum* L.). *The Journal of Horticultural Science and Biotechnology*, 80 (5), 593-598.
- Chen, Q. M. (2021). Nrf2 for Cardiac Protection: Pharmacological Options Against Oxidative Stress. *Trends in Pharmacological Sciences*.
- Chew, K. K., Khoo, M. Z., Ng, S. Y., Thoo, Y. Y., Aida, W. W., and Ho, C. W. (2011). Effect of Ethanol Concentration, Extraction Time and Extraction Temperature on the Recovery of Phenolic Compounds and Antioxidant Capacity of *Orthosiphon stamineus* Extracts. *International Food Research Journal*, 18 (4), 1427.
- Chiaramonti, D., Prussi, M., Ferrero, S., Oriani, L., Ottonello, P., Torre, P., and Cherchi, F. (2012). Review of Pretreatment Processes for Lignocellulosic Ethanol Production, and Development of an Innovative Method. *Biomass and Bioenergy*, 46, 25-35.
- Chibu, H. and H. Shibayama, (2001). Effects of Chitosan Applications on the Growth of Several Crops, in: T. Uragami, K. Kurita, T. Fukamizo (Eds.), *Chitin and Chitosan in life science, Yamaguchi*, pp. 235-239.

- Chigayo, K., Mojapelo, P. E. L., Mnyakeni-Moleele, S., and Misihairabgwi, J. M. (2016). Phytochemical and Antioxidant Properties of Different Solvent Extracts of *Kirkia wilmsii* Tubers. *Asian Pacific Journal of Tropical Biomedicine*, 6 (12), 1037-1043.
- Chonan, N. (1965). Studies on the Photosynthetic Tissues in the Leaves of Cereal Crops: I. The Mesophyll Structure of Wheat Leaves Inserted at Different Levels of the Shoot. *Japanese Journal of Crop Science*, 33 (4), 388-393.
- Choudhary, R. C., Kumaraswamy, R. V., Kumari, S., Sharma, S. S., Pal, A., Raliya, R., ... and Saharan, V. (2017). Cu-chitosan Nanoparticle Boost Defense Responses and Plant Growth in Maize (*Zea mays* L.). *Scientific Reports*, 7 (1), 1-11.
- Chowdhury, M. A., and Yahya, K. (2014) Sustainable Seafood Production: Malaysian Status and Comparison with the World. *Conference Paper*.
- Clark, L. J., Price, A. H., Steele, K. A., and Whalley, W. R. (2008). Evidence from Near-isogenic lines that root penetration increases with root diameter and bending stiffness in Rice. *Functional Plant Biology*, 35 (11), 1163-1171.
- Claussen, M., Lüthe, H., Blatt, M., and Böttger, M. (1997). Auxin-induced Growth and its Linkage to Potassium Channels. Planta, 201 (2), 227-234.
- Coombs, J., Hall, P.O., Long, S.P. and Schlock, J.M.O. (1987). Techniques in Bio-productivity and Photosynthesis. *Pergamon Press, Oxford*, 159.
- Costa, D. C., Costa, H. S., Albuquerque, T. G., Ramos, F., Castilho, M. C., and Sanches-Silva, A. (2015). Advances in Phenolic Compounds Analysis of Aromatic Plants and Their Potential Applications. *Trends in Food Science* & *Technology*, 45 (2), 336-354.
- Croft, H., Chen, J. M., Luo, X., Bartlett, P., Chen, B., and Staebler, R. M. (2017). Leaf Chlorophyll Content as a Proxy for Leaf Photosynthetic Capacity. *Global Change Biology*, 23 (9), 3513-3524.
- Dahmoune, F., Nayak, B., Moussi, K., Remini, H., and Madani, K. (2015). Optimization of Microwave-assisted Extraction of Polyphenols from *Myrtus communis* L. Leaves. *Food Chemistry*, 166, 585-595.
- Dash, J., Curran, P. J., Tallis, M. J., Llewellyn, G. M., Taylor, G., and Snoeij, P. (2010). Validating the MERIS Terrestrial Chlorophyll Index (MTCI) with Ground Chlorophyll Content Data at MERIS Spatial Resolution. *International Journal of Remote Sensing*, 31 (20), 5513-5532.
- Davis, A. S., and Jacobs, D. F. (2005). Quantifying Root System Quality of Nursery Seedlings and Relationship to Out Planting Performance. *New Forests*, 30 (2-3), 295-311.

- Delgado-Pelayo, R., Gallardo-Guerrero, L., and Hornero-Méndez, D. (2014). Chlorophyll and Carotenoid Pigments in the Peel and Flesh of Commercial Apple Fruit Varieties. *Food Research International*, 65, 272-281.
- Deng, F., Wang, L., Yao, X., Wang, J. J., Ren, W. J., and Yang, W. Y. (2009). Effects of Different Growing Stage Shading on Rice Grain-filling and Yield. *Journal of Sichuan Agricultural University*, 27 (3), 265-269.
- Dent, M., Dragović-Uzelac, V., Penić, M., Bosiljkov, T., and Levaj, B. (2013). The Effect of Extraction Solvents, Temperature and Time on the Composition and Mass Fraction of Polyphenols in Dalmatian Wild Sage (*Salvia officinalis* L.) Extracts. *Food Technology and Biotechnology*, 51 (1), 84-91.
- Dieleman, J. A., and Heuvelink, E. (1992). Factors Affecting the Number of Leaves Preceding the First Inflorescence in the Tomato. *Journal of Horticultural Science*, 67 (1), 1-10.
- Divya, K., and Jisha, M. S. (2018). Chitosan Nanoparticles Preparation and Applications. *Environmental Chemistry Letters*, 16 (1), 101-112.
- Divya, K., Vijayan, S., Nair, S. J., and Jisha, M. S. (2019). Optimization of Chitosan Nanoparticle Synthesis and Its Potential Application as Germination Elicitor of *Oryza sativa* L. *International Journal of Biological Macromolecules*, 124, 1053-1059.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S., and Ju, Y. H. (2014). Effect of Extraction Solvent on Total Phenol Content, Total Flavonoid Content, and Antioxidant Activity of *Limnophila aromatica. Journal of Food and Drug Analysis*, 22 (3), 296-302.
- Dong, C., Chen, W., and Liu, C. (2014). Flocculation of Algal Cells by Amphoteric Chitosan Based Flocculant. *Bioresource Technology*, 170, 239-247.
- Dorta, E., Lobo, M. G., and Gonzalez, M. (2012). Reutilization of Mango Byproducts: Study of the Effect of Extraction Solvent and Temperature on Their Antioxidant Properties. *Journal of Food Science*, 77 (1), C80-C88.
- Dou, H., Niu, G., Gu, M., and Masabni, J. G. (2018). Responses of Sweet Basil to Different Daily Light Integrals in Photosynthesis, Morphology, Yield, and Nutritional Quality. *HortScience*, 53 (4), 496-503.
- Dwyer, P. J., Bannister, P., and Jameson, P. E. (1995). Effects of Three Plant Growth Regulators on Growth, Morphology, Water Relations, and Frost Resistance in Lemonwood (*Pittosporum eugenioides* A. Cunn). *New Zealand Journal of Botany*, 33 (3), 415-424.
- Dzung, N. A., Khanh, V. T. P., and Dzung, T. T. (2011). Research on Impact of Chitosan Oligomers on Biophysical Characteristics, Growth, Development

and Drought Resistance of Coffee. *Carbohydrate Polymers*, 84 (2), 751-755.

- Efthymiopoulos, I., Hellier, P., Ladommatos, N., Russo-Profili, A., Eveleigh, A., Aliev, A., ... and Mills-Lamptey, B. (2018). Influence of Solvent Selection and Extraction Temperature on Yield and Composition of Lipids Extracted from Spent Coffee Grounds. *Industrial Crops and Products*, 119, 49-56.
- Eggink, L. L., Park, H., and Hoober, J. K. (2001). The Role of Chlorophyll b in Photosynthesis: Hypothesis. *BMC Plant Biology*, 1 (1), 2.
- El Knidri, H., Belaabed, R., Addaou, A., Laajeb, A., and Lahsini, A. (2018). Extraction, Chemical Modification and Characterization of Chitin and Chitosan. *International Journal of Biological Macromolecules*, 120, 1181-1189.
- Elboughdiri, N. (2018). Effect of Time, Solvent-solid Ratio, Ethanol Concentration and Temperature on Extraction Yield of Phenolic Compounds from Olive Leaves. *Eng. Technol. Appl. Sci. Res*, 8 (2), 2805-2808.
- El-Khateeb, M. A., Nasr, A. A. M., and Hassan, N. A. A. (2018). Growth and Quality Improvement of *Dracaena surculosa* Lindl. by the Foliar Application of Some Bio-stimulants. *Int. J. Environ*, 7 (2), 53-64.
- El-Miniawy, S. M., Ragab, M. E., Youssef, S. M., and Metwally, A. A. (2013). Response of Strawberry Plants to Foliar Spraying of Chitosan. *Res. J. Agric. Biol. Sci*, 9 (6), 366-372.
- El-Tantawy, E. M. (2009). Behavior of Tomato Plants as Affected by Spraying with Chitosan and Aminofort as Natural Stimulator Substances under Application of Soil Organic Amendments. *Pakistan Journal of Biological Sciences: PJBS*, 12 (17), 1164-1173.
- Emami-Bistgani, Z., Siadat, S. A., Bakhshandeh, A., Pirbalouti, A. G., and Hashemi, M. (2017). Morpho-physiological and Phytochemical Traits of (*Thymus daenensis* Celak.) in Response to Deficit Irrigation and Chitosan Application. *Acta Physiologiae Plantarum*, 39 (10), 1-13.
- Emami-Bistgani, Z., Siadat, S. A., Bakhshandeh, A., Pirbalouti, A. G., and Hashemi, M. (2017). Interactive Effects of Drought Stress and Chitosan Application on Physiological Characteristics and Essential Oil Yield of *Thymus daenensis* Celak. *The Crop Journal*, 5 (5), 407-415.
- Esclapez, M. D., García-Pérez, J. V., Mulet, A., and Cárcel, J. A. (2011). Ultrasound-assisted Extraction of Natural Products. *Food Engineering Reviews* 3: 108–120.

- Espada-Bellido, E., Ferreiro-González, M., Carrera, C., Palma, M., Barroso, C. G., and Barbero, G. F. (2017). Optimization of the Ultrasound-assisted Extraction of Anthocyanins and Total Phenolic Compounds in Mulberry (*Morus nigra*) Pulp. *Food Chemistry*, 219, 23-32.
- Evans, S. D., and Barber, S. A. (1964). The Effect of Rubidium-86 Diffusion on the Uptake of Rubidium-86 by Corn. Soil Science Society of America Journal, 28 (1), 56-57.
- Fahmy, A. A., and Nosir, W. S. (2021). Influence of Chitosan and Micronutrients (FE+ ZN) Concentrations on Growth, Yield Components and Volatile oil of Lavender Plant. Scientific Journal of Flowers and Ornamental Plants, 8 (1), 87-100.
- Falcón-Rodríguez, A. B., Costales, D., Gónzalez-Peña, D., Morales, D., Mederos, Y., Jerez, E., Cabrera, J. C. (2017). Chitosans of Different Molecular Weight Enhance Potato (*Solanum tuberosum* L.) Yield in a Field Trial. Spanish Journal of Agricultural Research, Volume 15, Issue 1, e0902.
- Farouk, S., and Amany, A. R. (2012). Improving Growth and Yield of Cowpea by Foliar Application of Chitosan under Water Stress. *Egyptian Journal of Biology*, 14, 14-16.
- Farouk, S., and Omar, M. M. (2020). Sweet Basil Growth, Physiological and Ultrastructural Modification, and Oxidative Defense System under Water Deficit and Silicon Forms Treatment. *Journal of Plant Growth Regulation*, 1-25.
- Farouk, S., Mosa, A. A., Taha, A. A., and El-Gahmery, A. M. (2011). Protective Effect of Humic Acid and Chitosan on Radish (*Raphanus sativus* L. var. sativus) Plants Subjected to Cadmium Stress. *Journal of Stress Physiology* & *Biochemistry*, 7 (2).
- Filip, S. (2017). Basil (*Ocimum basilicum L.*) A Source of Valuable Phytonutrients. *International Journal of Clinical Nutrition and Dietetics*, Volume 3. 118.
- Gallo, M., Formato, A., Ciaravolo, M., Formato, G., and Naviglio, D. (2020). Study of the Kinetics of Extraction Process for the Production of Hemp Inflorescences Extracts by Means of Conventional Maceration (CM) and Rapid Solid-liquid Dynamic Extraction (RSLDE). *Separations*, 7 (2), 20.
- Gao, X., Chen, X., Zhang, J., Guo, W., Jin, F., and Yan, N. (2016). Transformation of Chitin and Waste Shrimp Shells into Acetic Acid and Pyrrole. ACS Sustainable Chemistry and Engineering, 4 (7), 3912-3920.
- Gaska, J. M., and Oplinger, E. S. (1988). Use of Ethephon as a Plant Growth Regulator in Corn Production. *Crop science*, 28 (6), 981-986.

- Gechev, T., Gadjev, I., Van Breusegem, F., Inzé, D., Dukiandjiev, S., Toneva, V., and Minkov, I. (2002). Hydrogen Peroxide Protects Tobacco from Oxidative Stress by Inducing a Set of Antioxidant Enzymes. *Cellular and Molecular Life Sciences*, 59, 708–714. 708–714.
- Genc, Y., Huang, C. Y., and Langridge, P. (2007). A Study of the Role of Root Morphological Traits in Growth of Barley in Zinc Deficient Soil. *Journal of Experimental Botany*, 58 (11), 2775-2784.
- Genwali, G. R., Acharya, P. P., and Rajbhandari, M. (2013). Isolation of Gallic Acid and Estimation of Total Phenolic Content in Some Medicinal Plants and Their Antioxidant Activity. *Nepal Journal of Science and Technology*, 14 (1), 95-102.
- Gharibreza, M., Raj, J. K., Yusoff, I., Othman, Z., Tahir, W. Z. W. M., and Ashraf, M. A. (2013). Land Use Changes and Soil Redistribution Estimation Using 137Cs in the Tropical Bera Lake Catchment, Malaysia. *Soil and Tillage Research*, 131, 1-10.
- Ghasemzadeh, A., Ashkani, S., Baghdadi, A., Pazoki, A., Jaafar, H. Z., and Rahmat, A. (2016). Improvement in Flavonoids and Phenolic Acids Production and Pharmaceutical Quality of Sweet Basil (*Ocimum basilicum* L.) by Ultraviolet-B Irradiation. *Molecules*, 21 (9), 1203.
- Ghitescu, R. E., Volf, I., Carausu, C., Bühlmann, A. M., Gilca, I. A., and Popa, V.
 I. (2015). Optimization of Ultrasound Assisted Extraction of Polyphenols from Spruce Wood Bark. *Ultrasonics Sonochemistry*, 22, 535-541.
- Giannakoula, A. E., Ilias, I. F., Maksimović, J. J. D., Maksimović, V. M., and Živanović, B. D. (2012). The Effects of Plant Growth Regulators on Growth, Yield, and Phenolic Profile of lentil Plants. *Journal of Food Composition and Analysis*, 28 (1), 46-53.
- Gibaud, M., Bourmaud, A., and Baley, C. (2015). Understanding the Lodging Stability of Green Flax Stems; The Importance of Morphology and Fibre Stiffness. *Biosystems Engineering*, 137, 9-21.
- Gironi, F., and Piemonte, V. (2011). Temperature and Solvent Effects on Polyphenol Extraction Process from Chestnut Tree Wood. *Chemical Engineering Research and Design*, 89 (7), 857-862.
- Gitelson, A. A., Gritz, Y., and Merzlyak, M. N. (2003). Relationships Between Leaf Chlorophyll Content and Spectral Reflectance and Algorithms for Nondestructive Chlorophyll Assessment in Higher Plant Leaves. *Journal of Plant Physiology*, 160 (3), 271-282.
- Golkar, P., Taghizadeh, M., and Yousefian, Z. (2019). The Effects of Chitosan and Salicylic Acid on Elicitation of Secondary Metabolites and Antioxidant

Activity of Safflower under in Vitro Salinity Stress. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 137 (3), 575-585.

- Gomes, S. V., Portugal, L. A., dos Anjos, J. P., de Jesus, O. N., de Oliveira, E. J., David, J. P., and David, J. M. (2017). Accelerated Solvent Extraction of Phenolic Compounds Exploiting a Box-Behnken Design and Quantification of Five Flavonoids by HPLC-DAD in *Passiflora* species. *Microchemical Journal*, 132, 28-35.
- González-Montelongo, R., Lobo, M. G., and González, M. (2010). Antioxidant Activity in Banana Peel Extracts: Testing Extraction Conditions and Related Bioactive Compounds. *Food Chemistry*, 119 (3), 1030-1039.
- González-Salvatierra, C., Badano, E. I., Flores, J., and Rodas, J. P. (2013). Shade Shelters Increase Survival and Photosynthetic Performance of Oak Transplants at Abandoned Fields in Semi-arid Climates. *Journal of Forestry Research*, 24 (1), 23-28.
- Govindaraju, S., and Arulselvi, P. I. (2018). Effect of Cytokinin Combined Elicitors (I-phenylalanine, Salicylic Acid and Chitosan) on in Vitro Propagation, Secondary Metabolites and Molecular Characterization of Medicinal Herb *Coleus Aromaticus* Benth (L). *Journal of the Saudi Society of Agricultural Sciences*, 17 (4), 435-444.
- Gruber, B. D., Giehl, R. F., Friedel, S., Wirén, N. (2013) Plasticity of the Arabidopsis Root System under Nutrient Deficiencies. *Plant Physiology*, 163: 161–179.
- Guan, Y. J., Hu, J., Wang, X. J., & Shao, C. X. (2009). Seed Priming with Chitosan Improves Maize Germination and Seedling Growth in Relation to Physiological Changes under Low Temperature Stress. *Journal of Zhejiang University Science B*, 10 (6), 427-433.
- Guerrero-Lagunes, L. A., Ruiz-Posadas, L. D. M., Rodríguez-Mendoza, M. D. L.
 N., and Soto-Hernández, M. (2020). Quality and Yield of Basil (*Ocimum basilicum* L.) Essential Oil under Hydroponic Cultivation. *AGROProductividad*, 13 (9), 89-94.
- Guttridge, C. G., and Thompson, P. A. (1959). Effect of Gibberellic Acid on Length and Number of Epidermal Cells in Petioles of Strawberry. *Nature*, 183 (4655), 197-198.
- Hafez, Y., Attia, K., Alamery, S., Ghazy, A., Al-Doss, A., Ibrahim, E., ... and Abdelaal, K. (2020). Beneficial Effects of Biochar and Chitosan on Antioxidative Capacity, Osmolytes Accumulation, and Anatomical Characters of Water-stressed Barley plants. *Agronomy*, 10 (5), 630.
- Hajji, S., Younes, I., Ghorbel-Bellaaj, O., Hajji, R., Rinaudo, M., Nasri, M., and Jellouli, K. (2014). Structural Differences Between Chitin and Chitosan

Extracted from Three Different Marine Sources. *International Journal of Biological Macromolecules*, 65, 298-306.

- Halim, R. (2020). Industrial Extraction of Micro-algal Pigments. In Pigments from Microalgae Handbook. *Springer, Cham* pp. 265-308.
- Hanudin, E., Wismarini, H., Hertiani, T., and Hendro Sunarminto, B. (2012). Effect of Shading, Nitrogen and Magnesium Fertilizer on Phyllanthin and Total Flavonoid Yield of *Phyllanthus Niruri* in Indonesia Soil. *Journal of Medicinal Plants Research*, 6 (30), 4586-4592.
- Harmayani, E., Anal, A. K., Wichienchot, S., Bhat, R., Gardjito, M., Santoso, U.,
 ... and Payyappallimana, U. (2019). Healthy Food Traditions of Asia: Exploratory Case Studies from Indonesia, Thailand, Malaysia, and Nepal. *Journal of Ethnic Foods*, 6 (1), 1-18.
- Hassan, I. H., Pham, H. N. T., and Nguyen, T. H. (2021). Optimization of Ultrasound-assisted Extraction Conditions for Phenolics, Antioxidant, and Tyrosinase Inhibitory Activities of Vietnamese Brown Seaweed (*Padina australis*). *Journal of Food Processing and Preservation*, e15386.
- Hassnain, M., Alam, I., Ahmad, A., Basit, I., Ullah, N., Alam, I., ... and Shair, M. M. (2020). Efficacy of Chitosan on Performance of Tomato (*Lycopersicon esculentum* L.) Plant under Water Stress Condition. Pak. J. Agric. Res, 33, 27-41.
- Heidari, J., Amooaghaie, R., and Kiani, S. (2020). Impact of Chitosan on Nickel Bioavailability in Soil, the Accumulation and Tolerance of Nickel in *Calendula tripterocarpa. International Journal of Phytoremediation*, 1-10.

Hernández-Jiménez, A., Kennedy, J. A., Bautista-Ortín, A. B., and Gómez-Plaza, E. (2012). Effect of Ethanol on Grape Seed Proanthocyanidin Extraction. *American Journal of Enology and Viticulture*, 63 (1), 57-61.

- Hesami, M., Naderi, R., Tohidfar, M., and Yoosefzadeh-Najafabadi, M. (2020). Development of Support Vector Machine-based Model and Comparative Analysis with Artificial Neural Network for Modeling the Plant Tissue Culture Procedures: Effect of Plant Growth Regulators on Somatic Embryogenesis of Chrysanthemum, as a Case Study. *Plant Methods*, 16 (1), 1-15.
- Hidangmayum, A., Dwivedi, P., Katiyar, D., and Hemantaranjan, A. (2019). Application of Chitosan on Plant Responses with Special Reference to Abiotic Stress. *Physiology and Molecular Biology of Plants*, 25 (2), 313-326.
- Holguín-Peña, R. J., Medina-Hernández, D., Vázquez-Islas, G., Nieto-Navarro, F., and Puente, E. O. (2021). Anti-infective Properties of Medicinal Plants from the Baja California peninsula, Mexico for the treatment of *Fusarium*

oxysporum f. sp. Basilici in Organic Sweet basil (*Ocimum basilicum*). *Revista de la Facultad de Ciencias Agrarias UNCuyo*, 53 (1), 234-244.

- Hollman, P. C. H., Hertog, M. G. L., and Katan, M. B. (1996). Role of Dietary Flavonoids in Protection Against Cancer and Coronary Heart Disease. *Biochemical Society Transactions*, 24 (3), 785-789.
- Hu, Y., Ye, X., Shi, L., Duan, H., and Xu, F. (2010). Genotypic Differences in Root Morphology and Phosphorus Uptake Kinetics in *Brassica napus* under Low Phosphorus Supply. *Journal of Plant Nutrition*, 33 (6), 889-901.
- Hussain, I., Ahmad, S., Ullah, I., Basit, A., Ahmad, I., Sajid, M., Alam, M., Khan, S., and Ayaz, S. (2019). Foliar Application of Chitosan Modulates the Morphological and Biochemical Characteristics of Tomato. *Asian J. Agric. Biol.* 7 (3): 365-372.
- Hutchings, M. J., and John, E. A. (2003). Distribution of Roots in Soil, and Root Foraging Activity. In Root Ecology. *Springer*, pp. 33-60.
- Ibrahem-nasr, S. I., and Soliman, G. M. (2020). Application of Methyl Jasmonate and Chitosan on Behaviour of "Anna" Apple Seedlings Grown under Water and Heat Stress Conditions. *Arab Universities Journal of Agricultural Sciences*.
- Iglesias, M. J., Colman, S. L., Terrile, M. C., Paris, R., Martín-Saldaña, S., Chevalier, A. A., ... and Casalongué, C. A. (2019). Enhanced Properties of Chitosan Micro particles over Bulk Chitosan on the Modulation of the Auxin Signalling Pathway with Beneficial Impacts on Root Architecture in Plants. *Journal of Agricultural and Food Chemistry*, 67 (25), 6911-6920.
- Ilaiyaraja, N., Likhith, K. R., Babu, G. S., and Khanum, F. (2015). Optimisation of Extraction of Bioactive Compounds from *Feronia limonia* (wood Apple) Fruit Using Response Surface Methodology (RSM). *Food Chemistry*, 173, 348-354.
- Irawati, E. B., Sasmita, E. R., and Suryawati, A. (2019). Application of Chitosan for Vegetative Growth of *Kemiri sunan* Plant in Marginal Land. In *IOP Conference Series: Earth and Environmental Science* (Vol. 250, No. 1, p. 012089). IOP Publishing.
- Izadiyan, P., and Hemmateenejad, B. (2016). Multi-response Optimization of Factors Affecting Ultrasonic Assisted Extraction from Iranian Basil Using Central Composite Design. *Food Chemistry*, 190, 864-870.
- Janmohammadi, M., Mostafavi, H., Kazemi, H., Mahdavinia, G. R., and Sabaghnia, N. (2014). Effect of Chitosan Application on the Performance of Lentil Genotypes under Rainfed Conditions. *Acta Technologica Agriculturae*, 17 (4), 86-90.

- Jannat, R., Shaha, M., Rubayet, M. T., and Sultana, S. (2018). Role of Chitosan in Induction of Defence Response Against *Phomopsis vexans* and Augmentation of Growth and Yield of Eggplant. *Global Journal of Science Frontier Research: C Biological Science*, 18, 6-13.
- Jebapriya, G. R., and Somasundaram, R. (2021). Responses of Auxin Derivatives on Rooting and Sprouting Behaviour of *Excoecaria agallocha* L. Stem Cuttings. *Journal of Stress Physiology and Biochemistry*, 17 (1).
- Jeong, S. M., Kim, S. Y., Kim, D. R., Jo, S. C., Nam, K. C., Ahn, D. U., and Lee, S. C. (2004). Effect of Heat Treatment on the Antioxidant Activity of Extracts from Citrus Peels. *Journal of Agricultural and Food Chemistry*, 52 (11), 3389-3393.
- Jiao, J., Gai, Q. Y., Wang, X., Qin, Q. P., Wang, Z. Y., Liu, J., and Fu, Y. J. (2018). Chitosan Elicitation of *Isatis tinctoria* L. Hairy Root Cultures for Enhancing Flavonoid Productivity and Gene Expression and Related Antioxidant Activity. *Industrial Crops and Products*, 124, 28-35.
- Jovanović, A. A., Đorđević, V. B., Zdunić, G. M., Pljevljakušić, D. S., Šavikin, K. P., Gođevac, D. M., and Bugarski, B. M. (2017). Optimization of the Extraction Process of Polyphenols from *Thymus serpyllum* L. Herb Using Maceration, Heat-and Ultrasound-assisted Techniques. *Separation and Purification Technology*, 179, 369-380.
- Jung, W. S., Chung, I. M., Kim, S. H., Kim, M. Y., Ahmad, A., and Praveen, N. (2011). In Vitro Antioxidant Activity, Total Phenolics and Flavonoids from Celery (*Apium graveolens*) Leaves. *Journal of Medicinal Plants Research*, 5 (32), 7022-7030.
- Juntachote, T., Berghofer, E., Bauer, F., and Siebenhandl, S. (2006). The Application of Response Surface Methodology to the Production of Phenolic Extracts of Lemon Grass, Galangal, Holy Basil and Rosemary. *International Journal of Food Science and Technology*, 41 (2), 121-133.
- Kahromi, S., and Khara, J. (2020). Chitosan Stimulate Secondary Metabolites Production and Nutrient Uptake in Medicinal Plant *Dracocephalum kotschyi. Journal of the Science of Food and Agriculture*.
- Kahromi, S., and Khara, J. (2021). Chitosan Stimulates Secondary Metabolite Production and Nutrient Uptake in Medicinal Plant *Dracocephalum kotschyi. Journal of the Science of Food and Agriculture*, 101 (9), 3898-3907.
- Kalia, K., Sharma, K., Singh, H. P., and Singh, B. (2008). Effects of Extraction Methods on Phenolic Contents and Antioxidant Activity in Aerial Parts of *Potentilla atrosanguinea* Lodd. and Quantification of its Phenolic Constituents by RP-HPLC. *Journal of Agricultural and Food Chemistry*, 56 (21), 10129-10134.

- Kamalipourazad, M., Sharifi, M., Maivan, H. Z., Behmanesh, M., and Chashmi, N. A. (2016). Induction of Aromatic Amino Acids and Phenylpropanoid Compounds in *Scrophularia striata* Boiss. Cell Culture in Response to Chitosan-induced Oxidative stress. *Plant Physiology and Biochemistry*, 107, 374-384.
- Kapasakalidis, P. G., Rastall, R. A., and Gordon, M. H. (2006). Extraction of Polyphenols from Processed Black Currant (*Ribes nigrum* L.) Residues. *Journal of Agricultural and Food Chemistry*, 54 (11), 4016-4021.
- Karadeniz, A., Kaya, B., Savaş, B., and Topcuoğlu, Ş. F. (2011). Effects of Two Plant Growth Regulators, Indole-3-acetic Acid and β-naphthoxyacetic Acid, on Genotoxicity in *Drosophila* SMART Assay and on Proliferation and Viability of HEK293 Cells from the Perspective of Carcinogenesis. *Toxicology and Industrial Health*, 27 (9), 840-848.
- Kareem, S. A. A. (2011). Physiological Studies for Different Concentration from Biochikol 020 PC (Chitosan) on Bean Plant. *Journal of Asian Scientific Research*, 1 (2), 73.
- Katiyar, D., Hemantaranjan, A., and Singh, B. (2015). Chitosan as a Promising Natural Compound to Enhance Potential Physiological Responses in Plant: A Review. *Indian Journal of Plant Physiology*, 20 (1), 1-9.
- Katiyar, D., Hemantaranjan, A., Singh, B., and Bhanu, A. N. (2014). A Future Perspective in Crop Protection: Chitosan and its Oligosaccharides. *Advances in Plants and Agriculture Research*, 1 (1), 1-8.
- Khan, R., Manzoor, N., Zia, A., Ahmad, I., Ullah, A., Shah, S. M., Naeem, M., Ali, S., Khan, H. I., Zia, D., and Malik, S. (2018). Exogenous Application of Chitosan and Humic Acid Effects on Plant Growth and Yield of Pea (*Pisum sativum*). *International Journal of Biosciences*, Vol. 12, No. 5, p. 43-50.
- Khan, T., Khan, T., Hano, C., and Abbasi, B. H. (2019). Effects of Chitosan and Salicylic Acid on the Production of Pharmacologically Attractive Secondary Metabolites in Callus Cultures of *Fagonia indica*. *Industrial Crops and Products*, 129, 525-535.
- Khan, W., Costa, C., Souleimanov, A., Prithiviraj, B., and Smith, D. L. (2011). Response of *Arabidopsis thaliana* Roots to Lipo-chito-oligosaccharide from *Bradyrhizobium japonicum* and Other Chitin-like Compounds. *Plant Growth Regulation*, 63 (3), 243-249.
- Khanna, R. (2015). Customer Perception Towards Brand: A Study on 'Patanjali'. *Global Journal of Management and Business Research*, 15 (9), 8-12.

Khattak, M. K., Mamoon-ur-Rashid, S. A. S., and Islam, H. T. (2006).

Comparative Effect of Neem (*Azadirachta indica A. Juss*) Oil, Neem Seed Water Extract and Baythroid TM Against Whitefly, Jassids and Trips on Cotton. *Pak. Entomol.* Vol. 28, No.1.

- Khoddami, A., Wilkes, M. A., and Roberts, T. H. (2013). Techniques for Analysis of Plant Phenolic Compounds. *Molecules*, 18 (2), 2328-2375.
- King, A. M. Y., and Young, G. (1999). Characteristics and Occurrence of Phenolic Phytochemicals. *Journal of the American Dietetic Association*, 99 (2), 213-218.
- Knekt, P., Jarvinen, R., Reunanen, A., and Maatela, J. (1996). Flavonoid Intake and Coronary Mortality in Finland: A Cohort Study. *BMJ*, 312 (7029), 478-481.
- Ko, H. M., Eom, T. K., Kim, K. C., Kim, C. J., Lee, J. G., and Kim, J. S. (2018). Antioxidant Effects and Tyrosinase and Elastase Inhibitory Activities of Mountain Ginseng Adventitious Roots Extracts at Different Ethanol Concentrations. *Korean Journal of Agricultural Science*, 45 (3), 499-508.
- Koca, N., and Karaman, Ş. (2015). The Effects of Plant Growth Regulators and L-phenylalanine on Phenolic Compounds of Sweet Basil. *Food Chemistry*, 166, 515-521.
- Kopsell, D. A., Kopsell, D. E., Lefsrud, M. G., Curran-Celentano, J., and Dukach, L. E. (2004). Variation in Lutein, β-carotene, and Chlorophyll Concentrations Among *Brassica oleracea* Cultigens and Seasons. *HortScience*, 39 (2), 361-364.
- Koroch, A. R., Simon, J. E., and Juliani, H. R. (2017). Essential Oil Composition of Purple Basils, Their Reverted Green Varieties (*Ocimum basilicum*) and Their Associated Biological Activity. *Industrial Crops and Products*, 107, 526-530.
- Kőszeghi, S., Bereczki, C., Balog, A., and Benedek, K. (2014). Comparing the Effects of Benzyladenine and Meta-Topolin on Sweet Basil (*Ocimum basilicum*) Micropropagation. *Notulae Scientia Biologicae*, 6 (4), 422-427.
- Krell, V., Unger, S., Jakobs-Schoenwandt, D., and Patel, A. V. (2018). Importance of Phosphorus Supply through Endophytic *Metarhizium brunneum* for Root: Shoot Allocation and Root Architecture in Potato Plants. *Plant and Soil*, 430 (1-2), 87-97.
- Krupa-Małkiewicz, M., and Fornal, N. (2018). Application of Chitosan in Vitro to Minimize the Adverse Effects of Salinity in *Petuniax atkinsiana* D. don. *Journal of Ecological Engineering*, 19 (1), 143-149.
- Kumar, B. (2012). Prediction of Germination Potential in Seeds of Indian Basil (Ocimum basilicum L.). Journal of Crop Improvement, 26 (4), 532-539.

- Kumar, M. R., and Janagam, D. (2011). Export and Import Pattern of Medicinal Plants in India. *Indian Journal of Science and Technology*, 4 (3), 245-248.
- Kushwaha, R., Kumar, V., Vyas, G., and Kaur, J. (2018). Optimization of Different Variable for Eco-friendly Extraction of Betalains and Phytochemicals from Beetroot Pomace. *Waste and Biomass Valorization*, 9 (9), 1485-1494.
- Lai, Q. X., Bao, Z. Y., Zhu, Z. J., Qian, Q. Q., and Mao, B. Z. (2007). Effects of Osmotic Stress on Antioxidant Enzymes Activities in Leaf Discs of PSAG12-IPT Modified Gerbera. *Journal of Zhejiang University Science* B, 8, 458–464.
- Landi, L., Feliziani, E., and Romanazzi, G. (2014). Expression of Defense Genes in Strawberry Fruits Treated with Different Resistance Inducers. *Journal of Agricultural and Food Chemistry*, 62 (14), 3047-3056.
- Lanfer-Marquez, U. M., Barros, R. M., and Sinnecker, P. (2005). Antioxidant Activity of Chlorophylls and Their Derivatives. *Food Research International*, 38 (8-9), 885-891.
- Lawton, M. A., and Lamb, C. J. (1987). Transcriptional Activation of Plant Defense Genes by Fungal Elicitor, Wounding, and Infection. *Molecular and Cellular Biology*, 7, 335–341. 335–341.
- Lee, W. C. (2020). Marketing Margins of Aquaculture Shrimp Production in Kedah. *Borneo Journal of Marine Science and Aquaculture (BJoMSA)*, 4 (1), 20-23.
- Lee, Y. S., Kim, Y. H., and Kim, S. B. (2005). Changes in the Respiration, Growth, and Vitamin C Content of Soybean Sprouts in Response to Chitosan of Different Molecular Weights. *HortScience*, 40 (5), 1333-1335.
- Lei, C., Ma, D., Pu, G., Qiu, X., Du, Z., Wang, H., ... and Liu, B. (2011). Foliar Application of Chitosan Activates Artemisinin Biosynthesis in (*Artemisia annua* L.). *Industrial Crops and Products*, 33 (1), 176-182.
- Li, B. B., Smith, B., and Hossain, M. M. (2006a). Extraction of Phenolics from Citrus Peels: I. Solvent Extraction Method. *Separation and Purification Technology*, 48 (2), 182-188.
- Li, B. B., Smith, B., and Hossain, M. M. (2006b). Extraction of Phenolics from Citrus Peels: II. Enzyme-assisted Extraction Method. *Separation and Purification Technology*, 48 (2), 189-196.
- Li, H., Ge, Y., Luo, Z., Zhou, Y., Zhang, X., Zhang, J., and Fu, Q. (2017). Evaluation of the Chemical Composition, Antioxidant and Antiinflammatory Activities of Distillate and Residue Fractions of Sweet Basil Essential Oil. *Journal of Food Science and Technology*, 54 (7), 1882-1890.

- Li, R., He, J., Xie, H., Wang, W., Bose, S. K., Sun, Y., ... and Yin, H. (2019a). Effects of Chitosan Nanoparticles on Seed Germination and Seedling Growth of Wheat (*Triticum aestivum* L.). *International Journal of Biological Macromolecules*, 126, 91-100.
- Li, Y., Cui, Y., Hu, X., Liao, X., and Zhang, Y. (2019b). Chlorophyll Supplementation in Early Life Prevents Diet-Induced Obesity and Modulates Gut Microbiota in Mice. *Molecular nutrition & food research*, 63 (21), 1801219.
- Liaros, S., K. Botsis, and G. Xydis. (2016). Techno-economic Evaluation of Urban Plant Factories: The Case of Basil (*Ocimum basilicum*). *Sci. Total Environ.* 554: 218–227.
- Liazid, A., Palma, M., Brigui, J., and Barroso, C. G. (2007). Investigation on Phenolic Compounds Stability During Microwave-assisted Extraction. *Journal of Chromatography A*, 1140 (1-2), 29-34.
- Linkohr, B. I., Williamson, L. C., Fitter, A. H., and Leyser, H. (2002). Nitrate and Phosphate Availability and Distribution Have Different Effects on Root System Architecture of Arabidopsis. *Plant J*, 29: 751–760.
- Liu, W., Yu, Y., Yang, R., Wan, C., Xu, B., and Cao, S. (2010). Optimization of Total Flavonoid Compound Extraction from *Gynura medica* Leaf Using Response Surface Methodology and Chemical Composition Analysis. *International Journal of Molecular Sciences*, 11 (11), 4750-4763.
- Liu, Z., Liu, T., Liang, L., Li, Z., Hassan, M. J., Peng, Y., and Wang, D. (2020). Enhanced Photosynthesis, Carbohydrates, and Energy Metabolism Associated with Chitosan-induced Drought Tolerance in Creeping Bentgrass. *Crop Science*.
- Ljung, K., Bhalerao, R. P., and Sandberg, G. (2001). Sites and Homeostatic Control of Auxin Biosynthesis in Arabidopsis during Vegetative Growth. *The plant journal*, 28 (4), 465-474.
- Lopez-Moya, F., Suarez-Fernandez, M., and Lopez-Llorca, L. V. (2019). Molecular Mechanisms of Chitosan Interactions with Fungi and Plants. *International Journal of Molecular Sciences*, 20 (2), 332.
- Mahdavi, B., and Rahimi, A. (2013). Seed Priming with Chitosan Improves the Germination and Growth Performance of Ajowan (*Carum copticum*) under Salt Stress. *EurAsian Journal of BioSciences*, 7 (1), 69-76.
- Mahdi-Pour, B., Jothy, S. L., Latha, L. Y., Chen, Y., and Sasidharan, S. (2012). Antioxidant Activity of Methanol Extracts of Different Parts of *Lantana camara. Asian Pacific Journal of Tropical Biomedicine*, 2 (12), 960-965.

- Mahmoody, M. and Noori, M. (2014). Effect of Gibberellic Acid on Growth and Development Plants and its Relationship with Abiotic Stress. *International Journal of Farming and Allied Sciences*, 3 (6): 717-721.
- Mairapetyan, S., Alexanyan, J., Tovmasyan, A., Daryadar, M., Stepanian, B., and Mamikonyan, V. (2016). Productivity, Biochemical Indices and Antioxidant Activity of Peppermint (*Mentha piperita* L.) and Basil (*Ocimum basilicum* L.) in Conditions of Hydroponics. Journal of Science, Technology and Environamental Informatics. 03 (02): 191-194.
- Majidian, P., Gerami, M., Ghorbanpour, A., and Alipour, Z. (2020). Study of Some Morphological Responses of Stevia (*Stevia rebaudiana* Bertoni) to Chitosan Elicitor under Salt Stress. *Journal of Crop Breeding*, Volume 12, Number 33; Page (s) 150 To 161.
- Malekpoor, F., Salimi, A., and Ghasemi Pirbalouti, A. (2015). Effects of Jasmonic acid on essential oil yield and chemical compositions of two Iranian landraces of basil (*Ocimum basilicum*) under reduced irrigation. *Journal of Medicinal Herbs*, 6 (1), 13-21.
- Malerba, M., and Cerana, R. (2016). Chitosan Effects on Plant Systems. International Journal of Molecular Sciences, 17 (7), 996.
- Malerba, M., and Cerana, R. (2018). Recent Advances of Chitosan Applications in Plants. *Polymers*, 10 (2), 118.
- Malerba, M., and Cerana, R. (2019). Recent Applications of Chitin-and Chitosanbased Polymers in Plants. *Polymers*, 11 (5), 839.
- Malerba, M., and Cerana, R. (2020). Chitin-and Chitosan-based Derivatives in Plant Protection Against Biotic and Abiotic Stresses and in Recovery of Contaminated Soil and Water. *Polysaccharides*, 1 (1), 21-30.
- Marschener, H. (1998). Role of Root Growth, *Arbuscular mycorrhiza*, and Root Exudates for the Efficiency in Nutrient Acquisition. *Field Crops Research*, 56 (1-2), 203-207.
- Maulana, T. I., Falah, S., and Andrianto, D. (2019). Total Phenolic Content, Total Flavonoid Content, and Antioxidant Activity of Water and Ethanol Extract from Surian (*Toona sinensis*) Leaves. In *IOP Conference Series: Earth and Environmental Science* (Vol. 299, No. 1, p. 012021). IOP Publishing.
- Maulana, T. I., Falah, S., and Andrianto, D. (2019, July). Total Phenolic Content, Total Flavonoid Content, and Antioxidant Activity of Water and Ethanol Extract from Surian (*Toona sinensis*) Leaves. In IOP Conference Series: *Earth and Environmental Science*, Vol. 299, No. 1, p. 012021.
- Mc-Cullough, M. L., Peterson, J. J., Patel, R., Jacques, P. F., Shah, R., and Dwyer, J. T. (2012). Flavonoid Intake and Cardiovascular Disease Mortality

in a Prospective Cohort of US Adults. *The American Journal of Clinical Nutrition*, 95 (2), 454-464.

- Meng, S., Jia, Q., Zhou, G., Zhou, H., Liu, Q., and Yu, J. (2018). Fine Root Biomass and its Relationship with Aboveground Traits of *Larix gmelinii* Trees in Northeastern China. *Forests*, 9 (1), 35.
- Milevskaya, V. V., Prasad, S., and Temerdashev, Z. A. (2019). Extraction and Chromatographic Determination of Phenolic Compounds from Medicinal Herbs in the Lamiaceae and Hypericaceae Families: A Review. *Microchemical Journal*, 145, 1036-1049.
- Minke, R. A. M., and Blackwell, J. (1978). The Structure of α-chitin. *Journal of Molecular Biology*, 120 (2), 167-181.
- Mirzajani, Z., Hadavi, E., and Kashi, A. (2015). Changes in the Essential Oil Content and Selected Traits of Sweet Basil (*Ocimum basilicum* L.) as Induced by Foliar Sprays of Citric Acid and Salicylic acid. *Industrial Crops and Products*, 76, 269-274.
- Mith, H., Yayi-Ladékan, E., Sika Kpoviessi, S. D., Yaou Bokossa, I., Moudachirou, M., Daube, G., and Clinquart, A. (2016). Chemical Composition and Antimicrobial Activity of Essential Oils of *Ocimum basilicum*, *Ocimum* canum and *Ocimum* gratissimum in Function of Harvesting Time. Journal of Essential Oil Bearing Plants, 19 (6), 1413-1425.
- Mohamad, M., Ali, M. W., and Ahmad, A. (2010). Modelling for Extraction of Major Phytochemical Components from (*Eurycoma longifolia*). *Journal of Applied Sciences*, 10 (21), 2572-2577.
- Mohammadi, H., Dizaj, L. A., Aghaee, A., and Ghorbanpour, M. (2020). Chitosan-Mediated Changes in Dry Matter, Total Phenol Content and Essential Oil Constituents of Two *Origanum* Species under Water Deficit Stress. *Gesunde Pflanzen*, 1-11.
- Mokrani, A., and Madani, K. (2016). Effect of Solvent, Time and Temperature on the Extraction of Phenolic Compounds and Antioxidant Capacity of Peach (*Prunus persica* L.) Fruit. *Separation and Purification Technology*, 162, 68-76.
- Mondal, M. M. A., Malek, M. A., Puteh, A. B., and Ismail, M. R. (2013). Foliar Application of Chitosan on Growth and Yield Attributes of Mungbean (*Vigna radiata* (L.) Wilczek). *Bangladesh Journal of Botany*, 42 (1), 179-183.
- Mondal, M. M. A., Malek, M. A., Puteh, A. B., Ismail, M. R., Ashrafuzzaman, M., and Naher, L. (2012). Effect of Foliar Application of Chitosan on Growth and Yield in Okra. *Australian Journal of Crop Science*, 6 (5), 918.

- Mondal, M., Puteh, A. B., and Dafader, N. C. (2016). Foliar Application of Chitosan Improved Morphophysiological Attributes and Yield in Summer Tomato (*Solanum lycopersicum*). *Pakistan Journal of Agricultural Sciences*, 53 (2).
- Morganti, P., Danti, S., and Coltelli, M. B. (2018). Chitin and Lignin to Produce Biocompatible Tissues. *Res Clin Dermatol*, 1, 5-11.
- Mosadegh, H., Trivellini, A., Ferrante, A., Lucchesini, M., Vernieri, P., and Mensuali, A. (2018). Applications of UV-B Lighting to Enhance Phenolic Accumulation of Sweet Basil. *Scientia Horticulturae*, 229, 107-116.
- Mujtaba, M., Khawar, K. M., Camara, M. C., Carvalho, L. B., Fraceto, L. F., Morsi, R. E., ... and Wang, D. (2020). Chitosan-based Delivery Systems for Plants: A Brief Overview of Recent Advances and Future Directions. *International Journal of Biological Macromolecules*, 154, 683-697.
- Mukta, J. A., Rahman, M., Sabir, A. A., Gupta, D. R., Surovy, M. Z., Rahman, M., and Islam, M. T. (2017). Chitosan and Plant Probiotics Application Enhance Growth and Yield of Strawberry. *Biocatalysis and Agricultural Biotechnology*, 11, 9-18.
- Mukta, J. A., Rahman, M., Sabir, A. A., Gupta, D. R., Surovy, M. Z., Rahman, M., and Islam, M. T. (2017). Chitosan and Plant Probiotics Application Enhance Growth and Yield of Strawberry. *Biocatalysis and Agricultural Biotechnology*, 11, 9-18.
- Muley, A. B., Shingote, P. R., Patil, A. P., Dalvi, S. G., and Suprasanna, P. (2019). Gamma Radiation Degradation of Chitosan for Application in Growth Promotion and Induction of Stress Tolerance in Potato (*Solanum tuberosum* L.). *Carbohydrate Polymers*, 210, 289-301.
- Murali, M., and Prabakaran, G. (2018). Effect of Different Solvents System on Antioxidant Activity and Phytochemical Screening in Various Habitats of Ocimum basilicum L. (Sweet basil) Leaves. International Journal of Zoology and Applied Biosciences, 3, 375-381.
- Murugan, P. (2021). Antioxidants Effect on Herbs. *International Journal of Current Research in Life Sciences*, 10 (01), 3399-3402.
- Nagar, C. K., and Gayatri, A. B., Sinha, S. K., Venkatesh, K., Mandal, P. K.
 (2018). Nitrogen Stress Induced Changes in Root System Architecture (RSA) in Diverse Wheat (*T. aestivum* L.) Genotypes at Seedling Stage. *Wheat and Barley Research*, 10 (2), 93-101.
- Najafabadi, N. S., Sahari, M. A., Barzegar, M., and Esfahani, Z. H. (2020). Role of Extraction Conditions in the Recovery of Some Phytochemical Compounds of the Jujube Fruit. *Journal of Agricultural Science and Technology*, 22, 439-451.

- Ngo, T. V., Scarlett, C. J., Bowyer, M. C., Ngo, P. D., and Vuong, Q. V. (2017). Impact of Different Extraction Solvents on Bioactive Compounds and Antioxidant Capacity from the Root of *Salacia chinensis* L. *Journal of Food Quality*, Volume 2017, 8 pages.
- Ngome, M. T., Alves, J. G. L. F., de Oliveira, A. C. F., da Silva Machado, P., Mondragón-Bernal, O. L., and Piccoli, R. H. (2018). Linalool, Citral, Eugenol and Thymol: Control of Planktonic and Sessile Cells of *Shigella flexneri*. *AMB Express*, 8 (1), 105.
- Niklas, K. J., and Cobb, E. D. (2008). Evidence for "Diminishing Returns" from the Scaling of Stem Diameter and Specific Leaf Area. *American Journal of Botany*, 95 (5), 549-557.
- Nordin, S. M., Ariffin, Z., Jajuli, R., Abdullah, W. D. W., and Denis, M. G. (2007). Country Report on the State of Plant Genetic Resources for Food and Agriculture in Malaysia (1997-2007). *FAO, Rome, Italy*.
- Nour, V., Trandafir, I., and Cosmulescu, S. (2016). Optimization of Ultrasoundassisted Hydroalcoholic Extraction of Phenolic Compounds from Walnut Leaves Using Response Surface Methodology. *Pharmaceutical Biology*, 54 (10), 2176-2187.
- Nyamai, D. W., Arika, W., Ogola, P. E., Njagi, E. N. M., and Ngugi, M. P. (2016). Medicinally Important Phytochemicals: An Untapped Research Avenue. *Journal of Pharmacognosy and phytochemistry*, 4 (4), 2321-6182.
- Ohta, K., Morishita, S., Suda, K., Kobayashi, N., and Hosoki, T. (2004). Effects of Chitosan Soil Mixture Treatment in the Seedling Stage on the Growth and Flowering of Several Ornamental Plants. *Journal of the Japanese Society for Horticultural Science*, 73 (1), 66-68.
- Onofrei, V., Benchennouf, A., Jancheva, M., Loupassaki, S., Ouaret, W., Burducea, M., ... and Robu, T. (2018). Ecological Foliar Fertilization Effects on Essential Oil Composition of Sweet Basil (*Ocimum basilicum* L.) Cultivated in a Field System. *Scientia Horticulturae*, 239, 104-113.
- Oosterhuis, D. M. (1990). Growth and Development of a Cotton Plant. *Nitrogen nutrition of Cotton: Practical Issues*, 1-24.
- Oreopoulou, A., Tsimogiannis, D., and Oreopoulou, V. (2019). Extraction of Polyphenols from Aromatic and Medicinal Plants: An Overview of the Methods and the Effect of Extraction Parameters. *Polyphenols in Plants*, 243-259.
- Ouyang, S., and Langlai, X. (2003). Effect of Chitosan on Nutrient Quality and Some Agronomic Characteristic of Non-heading Chinese Cabbage. *Plant Physiology Communication* 39 (1): 21-24.

- Patriani, P., Hellyward, J., Hafid, H., and Apsari, N. L. (2021). Application of Sweet Basil (*Ocimum basilicum*) on Physical and Organoleptic Qualities of Chicken Meatballs. In *IOP Conference Series: Earth and Environmental Science* (Vol. 782, No. 2, p. 022083). IOP Publishing.
- Patriani, P., Hellyward, J., Hafid, H., and Apsari, N. L. (2021). Application of Sweet Basil (*Ocimum basilicum*) on Physical and Organoleptic Qualities of Chicken Meatballs. In *IOP Conference Series: Earth and Environmental Science* (Vol. 782, No. 2, p. 022083). IOP Publishing.
- Pemmaraju, D., Appidi, T., Minhas, G., Singh, S. P., Khan, N., Pal, M., ... and Rengan, A. K. (2018). Chlorophyll Rich Biomolecular Fraction of A. Cadamba Loaded into Polymeric Nanosystem Coupled with Photothermal Therapy: A Synergistic Approach for Cancer Theranostics. *International Journal of Biological Macromolecules*, 110, 383-391.
- Philibert, T., Lee, B. H., and Fabien, N. (2017). Current Status and New Perspectives on Chitin and Chitosan as Functional Biopolymers. *Applied Biochemistry and Biotechnology*, 181 (4), 1314-1337.
- Pichyangkura, R., and Chadchawan, S. (2015). Biostimulant Activity of Chitosan in Horticulture. *Scientia Horticulturae*, 196, 49-65.
- Prakash, V. (2019). Leafy spices. CRC press.
- Price, G. J., and Smith, P. F. (1993). Ultrasonic Degradation of Polymer Solutions—III. The Effect of Changing Solvent and Solution Concentration. *European Polymer Journal*, 29 (2-3), 419-424.
- Proestos, C., Boziaris, I. S., Nychas, G. J., and Komaitis, M. (2006). Analysis of Flavonoids and Phenolic Acids in Greek Aromatic Plants: Investigation of Their Antioxidant Capacity and Antimicrobial Activity. *Food Chemistry*, 95 (4), 664-671.
- Pushpangadan, P., and George, V. (2012). Basil. In *Handbook of Herbs and Spices* (pp. 55-72). Woodhead Publishing.
- Putievsky, E., and Galambosi, B. (1999). Production Systems of Sweet Basil. Basil: The Genus Ocimum, 10, 39-65.
- Rademacher, W. (2015). Plant Growth Regulators: Backgrounds and Uses in Plant Production. *Journal of Plant Growth Regulation*, 34 (4), 845-872.
- Rafińska, K., Pomastowski, P., Rudnicka, J., Krakowska, A., Maruśka, A., Narkute, M., and Buszewski, B. (2019). Effect of Solvent and Extraction Technique on Composition and Biological Activity of *Lepidium sativum* Extracts. *Food Chemistry*, 289, 16-25.

Rahimi-Shokooh, A., Mehrafarin, A., and Naghdi-Badi, H. (2013). The Effects of

Bio-fertilizers and Bio-stimulators on Morphophisiological Properties of (*Ocimum basilicum* L.). 2nd *National Congress on Medicinal Plants*, Tehran- Iran.

- Rahman, M. M., Vasiliev, M., and Alameh, K. (2021). LED Illumination Spectrum Manipulation for Increasing the Yield of Sweet Basil (*Ocimum basilicum* L.). *Plants*, 10 (2), 344.
- Rahman, M., Mukta, J. A., Sabir, A. A., Gupta, D. R., Mohi-Ud-Din, M., Hasanuzzaman, M., ... and Islam, M. T. (2018). Chitosan Biopolymer Promotes Yield and Stimulates Accumulation of Antioxidants in Strawberry Fruit. *PloS One*, 13 (9), e0203769.
- Raj, G. B., and Dash, K. K. (2020). Ultrasound-assisted Extraction of Phytocompounds from Dragon Fruit Peel: Optimization, Kinetics and Thermodynamic Studies. *Ultrasonics Sonochemistry*, 68, 105180.
- Raji, M. N. A., Ab Karim, S., Ishak, F. A. C., and Arshad, M. M. (2017). Past and Present Practices of the Malay Food Heritage and Culture in Malaysia. *Journal of Ethnic Foods*, 4 (4), 221-231.
- Ramakrishna, A., and Ravishankar, G. A. (2011). Influence of Abiotic Stress Signals on Secondary Metabolites in Plants. *Plant Signaling and Behavior*, 6 (11), 1720–1731.
- Rao, N. S., Geetha, K. A., and Maiti, S. (2017). DPLMAP: A Digital Photo Library Reference System for Medicinal and Aromatic Plants.
- Rasheed, R., Ashraf, M. A., Arshad, A., Iqbal, M., and Hussain, I. (2020). Interactive Effects of Chitosan and Cadmium on Growth, Secondary Metabolism, Oxidative Defense, and Element Uptake in Pea (*Pisum sativum* L.). *Arabian Journal of Geosciences*, 13 (17), 1-14.
- Rastogi, A., Siddiqui, A., Mishra, B. K., Srivastava, M., Pandey, R., Misra, P., ... and Shukla, S. (2013). Effect of Auxin and Gibberellic Acid on Growth and Yield Components of Linseed (*Linum usitatissimum* L.). *Crop Breeding and Applied Biotechnology*, 13 (2), 136-143.
- Razavizadeh, R., Adabavazeh, F., and Komatsu, S. (2020). Chitosan Effects on the Elevation of Essential Oils and Antioxidant Activity of *Carum copticum* L. Seedlings and Callus Cultures under in Vitro Salt Stress. *Journal of Plant Biochemistry and Biotechnology*, 29 (3), 473-483.
- Reis, C. O., Magalhães, P. C., Avila, R. G., Almeida, L. G., Rabelo, V. M., Carvalho, D. T., ... and de Souza, T. C. (2019). Action of N-Succinyl and N, O-Dicarboxymethyl Chitosan Derivatives on Chlorophyll Photosynthesis and Fluorescence in Drought-sensitive Maize. *Journal of Plant Growth Regulation*, 38 (2), 619-630.

- Richardson, A. D., Duigan, S. P., and Berlyn, G. P. (2002). An Evaluation of Noninvasive Methods to Estimate Foliar Chlorophyll Content. *New Phytologist*, 153 (1), 185-194.
- Rinaudo, M. (2006). Chitin and Chitosan: Properties and Applications. *Progress in Polymer Science*, 31 (7), 603-632.

Rizky F. (2013). Miracle of Vegetables. Jakarta: Agromedia.

- Rocha, R. P., and Melo, E. C. (2011). Influence of Drying Process on the Quality of Medicinal Plants: A Review. *Journal of Medicinal Plants Research*, 5 (33), 7076-7084.
- Rodríguez, A. F., Costales, D., Peña, D. G., Morales, D., Mederos, Y., Jerez, E., and Pino, J. C. (2017). Chitosans of Different Molecular Weight Enhance Potato (*Solanum tuberosum* L.) Yield in a Field Trial. *Spanish Journal of Agricultural Research*, 15 (1), 25.
- Roesti, D., Gaur, R., Johri, B. N., Imfeld, G., Sharma, S., Kawaljeet, K., and Aragno, M. (2006). Plant Growth Stage, Fertiliser Management and Bioinoculation of *Arbuscular mycorrhizal* Fungi and Plant Growth Promoting Rhizobacteria Affect the Rhizobacterial Community Structure in Rain-fed Wheat Fields. *Soil Biology and Biochemistry*, 38 (5), 1111-1120.
- Ruan, Z. P., Zhang, L. L., and Lin, Y. M. (2008). Evaluation of the Antioxidant Activity of *Syzygium cumini* Leaves. *Molecules*, 13 (10), 2545-2556.
- Sae-Lee, N., Kerdchoechuen, O., Laohakunjit, N., Thumthanaruk, B., Sarkar, D., and Shetty, K. (2017). Improvement of Phenolic Antioxidant-linked Cancer Cell Cytotoxicity of Grape Cell Culture Elicited by Chitosan and Chemical Treatments. *HortScience*, 52 (11), 1577-1584.
- Safahani, A. (2020). Alleviatory Activities of Salicylic Acid and Chitosan in Burdock Plant (*Arctium lappa* L.) under Drought Stress.
- Saharan, V., and Pal, A. (2016). Chitosan Based Nanomaterials in Plant Growth and Protection. *Springer*, 33-41.
- Saharan, V., Kumaraswamy, R. V., Choudhary, R. C., Kumari, S., Pal, A., Raliya, R., and Biswas, P. (2016). Cu-chitosan Nanoparticle Mediated Sustainable Approach to Enhance Seedling Growth in Maize by Mobilizing Reserved Food. *Journal of Agricultural and Food Chemistry*, 64 (31), 6148-6155.
- Saharan, V., Sharma, G., Yadav, M., Choudhary, M. K., Sharma, S. S., Pal, A., ... and Biswas, P. (2015). Synthesis and in Vitro Antifungal Efficacy of Cu– chitosan Nanoparticles Against Pathogenic Fungi of Tomato. *International Journal of Biological Macromolecules*, 75, 346-353.

- Said, M. B. (2009). Traditional Malaysian Salads (Ulam) as a Source of Antioxidants. In *Prosiding Seminar Kimia Bersama UKM-ITB* VIII (Vol. 9, p. 11).
- Saikia, S., Mahnot, N. K., and Mahanta, C. L. (2015). Optimisation of Phenolic Extraction from Averrhoa carambola Pomace by Response Surface Methodology and its Microencapsulation by Spray and Freeze Drying. Food Chemistry, 171, 144-152.
- Saint-Germain, A., Ligerot, Y., Dun E. A., Pillot, JP., Ross, J. J., Beveridge, C. A., Rameau, C. (2013) Strigolactones Stimulate Internode Elongation Independently of Gibberellins. *Plant Physiology*, 163 (2):1012–1025.
- Salachna, P., Wilas, J., and Zawadzińska, P. (2014). The Effect of Chitosan Coating of Bulbs on the Growth and Flowering of Ornithogalum saundersiae Baker. In Proceedings of the 29th International Horticultural Congress: Sustaining Lives, Livelihoods and Landscapes (Aug. 17-22), Brisbane, Australia.
- Salehi, S., and Rezayatmand, Z. (2017). The Effect of Foliar Application of Chitosan on Yield and Essential Oil of Savory (*Satureja isophylla* L.) under Salt Stress. *Journal of Herbal Drugs (An International Journal on Medicinal Herbs*), 8 (2), 101-108.
- Santos, V. P., Marques, N. S., Maia, P. C., Lima, M. A. B. D., Franco, L. D. O., and Campos-Takaki, G. M. D. (2020). Seafood Waste as Attractive Source of Chitin and Chitosan Production and Their Applications. *International journal of molecular sciences*, 21 (12), 4290.
- Santos Zea, L., Gutiérrez-Uribe, J. A., and Benedito, J. (2019). Effect of Ultrasound Intensification on the Supercritical Fluid Extraction of Phytochemicals from *Agave salmiana* Bagasse. *The Journal of Supercritical Fluids*, 144, 98-107.
- Sari, S. G., Selvia, E., Nisa, C., and Junaidi, A. B. (2020) The Effect of Liquid Smoke and Chitosan Composition on the Growth of Red Cayenne Pepper *Capsicum Frutescens* Linn. *Biotropika Journal of Tropical Biology*. Vol. 8 | No. 1: 8-11.
- Sarker, U., Oba, S., and Daramy, M. A. (2020). Nutrients, Minerals, Antioxidant Pigments and Phytochemicals, and Antioxidant Capacity of the Leaves of Stem Amaranth. *Scientific Reports*, 10 (1), 1-9.
- Sathiyabama, M., and Parthasarathy, R. (2016). Biological Preparation of Chitosan Nanoparticles and its in Vitro Antifungal Efficacy Against some Phytopathogenic Fungi. *Carbohydrate Polymers*, 151, 321-325.
- Scapin, G., Schmidt, M. M., Prestes, R. C., and Rosa, C. S. (2016). Phenolics Compounds, Flavonoids and Antioxidant Activity of Chia Seed Extracts

(*Salvia hispanica*) Obtained by Different Extraction Conditions. *International Food Research Journal*, 23 (6).

- Sen, S., De, B., Devanna, N., and Chakraborty, R. (2013). Total Phenolic, Total Flavonoid Content, and Antioxidant Capacity of the Leaves of *Meyna spinosa* Roxb., an Indian Medicinal Plant. *Chinese Journal of Natural Medicines*, 11 (2), 149-157.
- Setford, P. C., Jeffery, D. W., Grbin, P. R., and Muhlack, R. A. (2017). Factors Affecting Extraction and Evolution of Phenolic Compounds During Red Wine Maceration and the Role of Process Modelling. *Trends in Food Science and Technology*, 69, 106-117.
- Setyaningsih, W., Saputro, I. E., Carrera, C. A., and Palma, M. (2019). Optimisation of an Ultrasound-assisted Extraction Method for the Simultaneous Determination of Phenolics in Rice Grains. *Food Chemistry*, 288, 221-227.
- Shah, M., Jan, H., Drouet, S., Tungmunnithum, D., Shirazi, J. H., Hano, C., and Abbasi, B. H. (2021). Chitosan Elicitation Impacts Flavonolignan Biosynthesis in *Silybum marianum* (L.) Gaertn Cell Suspension and Enhances Antioxidant and Anti-Inflammatory Activities of Cell Extracts. *Molecules*, 26 (4), 791.
- Shahrajabian, M. H., Sun, W., and Cheng, Q. (2020). Chemical components and pharmacological benefits of Basil (*Ocimum basilicum*): a review. *International Journal of Food Properties*, 23 (1), 1961-1970.
- Sharafzadeh, S., Esmaeili, M., and Mohammadi, A. H. (2011). Interaction Effects of Nitrogen, Phosphorus and Potassium on Growth, Essential Oil and Total Phenolic Content of Sweet Basil. *Adv. Environ. Biol*, 5 (6), 1285-1289.
- Sharif, R., Mujtaba, M., Ur Rahman, M., Shalmani, A., Ahmad, H., Anwar, T., ... and Wang, X. (2018). The Multifunctional Role of Chitosan in Horticultural Crops; a Review. *Molecules*, 23 (4), 872.
- Sharma, G., Kumar, A., Devi, K. A., Prajapati, D., Bhagat, D., Pal, A., ... and Saharan, V. (2020). Chitosan Nanofertilizer to Foster Source Activity in Maize. *International Journal of Biological Macromolecules*, 145, 226-234.
- Sharma, N., and Tomar, R. S. (2021). Association of Nonenzymatic Antioxidants in Plant Holobiont. In Antioxidants in Plant-Microbe Interaction, Singapore. *Springer*, pp. 59-73.
- Shehata, M. G., Abd El Aziz, N. M., Youssef, M. M., and El-Sohaimy, S. A. (2021). Optimization Conditions of Ultrasound-assisted Extraction of Phenolic Compounds from Orange Peels Using Response Surface Methodology. *Journal of Food Processing and Preservation*, 45 (10).

- Shehata, S. A., Fawzy, Z. F., and El-Ramady, H. R. (2012). Response of Cucumber Plants to Foliar Application of Chitosan and Yeast under Greenhouse Conditions. *Australian Journal of Basic and Applied Sciences*, 6 (4), 63-71.
- Sheikha, S. A., and Al-Malki, F. M. (2011). Growth and Chlorophyll Responses of Bean Plants to the Chitosan Applications. *European Journal of Scientific Research*, 50 (1), 124-134.
- Sheng, Z. L., Wan, P. F., Dong, C. L., and Li, Y. H. (2013). Optimization of Total Flavonoids Content Extracted from *Flos Populi* Using Response Surface Methodology. *Industrial Crops and Products*, 43, 778-786.
- Shenglei, G. U. O., Zhang, S., Liwei, J. I. A., Mingyuan, X. U., and Zhenyue, W. A. N. G. (2020). Root Growth of Eleuthero (*Eleutherococcus senticosus* [Rupr. and Maxim.] Maxim.) Seedlings Cultured with Chitosan Oligosaccharide Addition under Different Light Spectra. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 48 (2), 626-635.
- Silva, E. M., Rogez, H., and Larondelle, Y. (2007). Optimization of Extraction of Phenolics from *Inga edulis* Leaves Using Response Surface Methodology. *Sep Purif Technol*, 55:381–38.
- Silva, V., Singh, R. K., Gomes, N., Soares, B. G., Silva, A., Falco, V., ... and Poeta, P. (2020). Comparative Insight Upon Chitosan Solution and Chitosan Nanoparticles Application on the Phenolic Content, Antioxidant and Antimicrobial Activities of Individual Grape Components of Sousão Variety. *Antioxidants*, 9 (2), 178.
- Silva-Antonio, A., Aguiar, A. T. C., Santos, G. R. C., Pereira, H. M. G., Veiga-Junior, V. F., and Wiedemann, L. S. M. (2020). Phytochemistry by Design: A Case Study of the chemical composition of *Ocotea guianensis* optimized extracts focused on Untargeted Metabolomics Analysis. *RSC Advances*, 10 (6), 3459-3471.
- Silva-Ferreira, V., and Sant'Anna, C. (2017). Impact of Culture Conditions on the Chlorophyll Content of Microalgae for Biotechnological Applications. *World Journal of Microbiology and Biotechnology*, 33 (1), 20.
- Singh, S. (2016). Enhancing Phytochemical Levels, Enzymatic and Antioxidant Activity of Spinach Leaves by Chitosan Treatment and an Insight into the Metabolic Pathway Using DART-MS Technique. *Food Chemistry*, 199, 176-184.
- Singh, S., Lal, R. K., Maurya, R., and Chanotiya, C. S. (2018). Genetic Diversity and Chemotype Selection in Genus Ocimum. *Journal of Applied Research on Medicinal and Aromatic Plants*, 9, 19-25.

- Skrypnik, L., Novikova, A., and Tokupova, E. (2019). Improvement of Phenolic Compounds, Essential Oil Content and Antioxidant Properties of Sweet Basil (*Ocimum basilicum* L.) Depending on Type and Concentration of Selenium Application. *Plants*, 8 (11), 458.
- Slougui, N., Tlili, A., Hammoudi, R., Bentayeb, H., and Mahammed, M. H. (2018). Composition of Essential Oil of Ocimum basilicum L., Minimum and Variability in Antioxidant Activity of Essential Oil of Leaves and Flowering Tops of Ocimum basilicum L. Genovese Following Seasons of Culture under Arid Climate (Southeast of Algeria). International Journal of Biosciences, Vol. 12, No. 4, p. 370-382.
- Sopee, M. S. M., Azlan, A., and Khoo, H. E. (2019). Comparison of Antioxidants Content and Activity of *Nephelium mutabile* Rind Extracted Using Ethanol and Water. *Journal of Food Measurement and Characterization*, 13 (3), 1958-1963.
- Souri, M. K., Naiji, M., and Kianmehr, M. H. (2019). Nitrogen Release Dynamics of a Slow Release Urea Pellet and its Effect on Growth, Yield, and Nutrient Uptake of Sweet Basil (*Ocimum basilicum* L.). *Journal of Plant Nutrition*, 42 (6), 604-614.
- Spigno, G., and De-Faveri, D. M. (2007). Antioxidants from Grape Stalks and Marc: Influence of Extraction Procedure on Yield, Purity and Antioxidant Power of the Extracts. *Journal of Food Engineering*, 78:793–801.
- Spigno, G., Tramelli, L., and De Faveri, D. M. (2007). Effects of Extraction Time, Temperature and Solvent on Concentration and Antioxidant Activity of Grape Marc Phenolics. *Journal of Food Engineering*, 81 (1), 200-208.
- Sprangers, K., Thys, S., van Dusschoten, D., and Beemster, G. T. (2020). Gibberellin Enhances the Anisotropy of Cell Expansion in the Growth Zone of the Maize Leaf. *Frontiers in Plant Science*, 11, 1163.
- Srivastava, R. K., Kumar, S., and Sharma, R. S. (2018). Ocimum as a promising commercial crop. In *The Ocimum Genome. Springer*, Cham. Pp: 1-7.
- Stützel, H., and Hanafy, M. S. (2020). Impact of Chitosan on Shoot Regeneration from Faba Bean Embryo Axes through its Effect on Phenolic Compounds and Endogenous Hormones. *Plant Archives*, 20 (1), 2269-2279.
- Su, C. H., Lai, M. N., and Ng, L. T. (2017). Effects of Different Extraction Temperatures on the Physicochemical Properties of Bioactive Polysaccharides from *Grifola frondosa*. *Food Chemistry*, 220, 400-405.
- Subramoniam, A., Asha, V. V., Nair, S. A., Sasidharan, S. P., Sureshkumar, P. K., Rajendran, K. N., ... and Ramalingam, K. (2012). Chlorophyll Revisited: Anti-inflammatory Activities of Chlorophyll a and Inhibition of Expression of TNF-α Gene by the Same. *Inflammation*, 35 (3), 959-966.

- Sumere, B. R., de Souza, M. C., Dos Santos, M. P., Bezerra, R. M. N., da Cunha, D. T., Martinez, J., and Rostagno, M. A. (2018). Combining Pressurized Liquids with Ultrasound to Improve the Extraction of Phenolic Compounds from Pomegranate Peel (*Punica granatum* L.). Ultrasonics Sonochemistry, 48, 151-162.
- Sun, J., Wang, M., Lyu, M., Niklas, K. J., Zhong, Q., Li, M., and Cheng, D. (2019). Stem Diameter (and not Length) Limits Twig Leaf Biomass. *Frontiers in Plant Science*, 10, 185.
- Swoish, M., and Steinke, K. (2017). Plant Growth Regulator and Nitrogen Applications for Improving Wheat Production in Michigan. *Crop, Forage and Turfgrass Management*, 3 (1), 1-7.
- Tagliavini, M., Veto, L. J., and Looney, N. E. (1993). Measuring Root Surface Area and Mean Root Diameter of Peach Seedlings by Digital Image Analysis. *HortScience*, 28 (11), 1129-1130.
- Taha, R. S., Alharby, H. F., Bamagoos, A. A., Medani, R. A., and Rady, M. M. (2020). Elevating Tolerance of Drought Stress in *Ocimum basilicum* Using Pollen Grains Extract; A Natural Bio-stimulant by Regulation of Plant Performance and Antioxidant Defense System. *South African Journal of Botany*, 128, 42-53.
- Talón, E., Trifkovic, K. T., Nedovic, V. A., Bugarski, B. M., Vargas, M., Chiralt, A., and González-Martínez, C. (2017). Antioxidant Edible Films Based on Chitosan and Starch Containing Polyphenols from Thyme Extracts. *Carbohydrate Polymers*, 157, 1153-1161.
- Tanimoto, E. (2005). Regulation of Root Growth by Plant Hormones—roles for Auxin and Gibberellin. *Critical Reviews in Plant Sciences*, 24 (4), 249-265.
- Tayo, P. M. T., Ewane, C. A., Effa, P. O., and Boudjeko, T. (2017). Effects of Chitosan and Snail Shell Powder on Cocoa (*Theobroma cacao* L.) Growth and Resistance Against Black Pod Disease Caused by *Phytophthora megakarya*. *African Journal of Plant Science*, 11 (8), 331-340.
- Teale, W. D., Paponov, I. A., and Palme, K. (2006). Auxin in Action: Signalling, Transport and the Control of Plant Growth and Development. *Nature Reviews Molecular Cell Biology*, 7 (11), 847-859.
- Tidemann, B. D., O'Donovan, J. T., Izydorczyk, M., Turkington, T. K., Oatway, L., Beres, B., ... and de Gooijer, H. (2020). Effects of Plant Growth Regulator Applications on Malting Barley in Western Canada. *Canadian Journal of Plant Science*, 100 (6), 653-665.
- Toh, P. Y., Leong, F. S., Chang, S. K., Khoo, H. E., and Yim, H. S. (2016). Optimization of Extraction Parameters on the Antioxidant Properties of

Banana Waste. *Acta Scientiarum Polonorum Technologia Alimentaria*, 15 (1), 65-78.

- Truong, D. H., Nguyen, D. H., Ta, N. T. A., Bui, A. V., Do, T. H., and Nguyen, H. C. (2019). Evaluation of the Use of Different Solvents for Phytochemical Constituents, Antioxidants, and in Vitro Anti-inflammatory Activities of Severinia buxifolia. Journal of Food Quality, Volume 2019, ID 8178294, 9 pages.
- Turan V. 2019. Confident Performance of Chitosan and Pistachio Shell Biochar on Reducing Ni Bioavailability in Soil and Plant Plus Improved the Soil Enzymatic Activities, Antioxidant Defense System and Nutritional Quality of Lettuce. *Eco Environ Saf.* 183: 109594.
- Turrini, F., Donno, D., Beccaro, G. L., Zunin, P., Pittaluga, A., and Boggia, R. (2019). Pulsed Ultrasound-assisted Extraction as an Alternative Method to Conventional Maceration for the Extraction of the Polyphenolic Fraction of *Ribes nigrum* Buds: A New Category of Food Supplements Proposed by the Finnover Project. *Foods*, 8 (10), 466.
- Ullah, N., Basit, A., Ahmad, I., Ullah, I., Shah, S. T., Mohamed, H. I., and Javed, S. (2020). Mitigation the Adverse Effect of Salinity Stress on the Performance of the Tomato Crop by Exogenous Application of Chitosan. *Bulletin of the National Research Centre*, 44 (1), 1-11.
- Urquiza-Martínez, M. V., and Navarro, B. F. (2016). Antioxidant Capacity of Food. *Free Radicals and Antioxidants*, 6 (1), 01-12.
- Uthairatanakij, A., Teixeira da Silva, J. A., and Obsuwan, K. (2007). Chitosan for Improving Orchid production and Quality. Orchid Science and Biotechnology, 1 (1), 1-5.
- Uzun, S. (2006). The Quantitative Effects of Temperature and Light on the Number of Leaves Preceding the First Fruiting Inflorescence on the Stem of Tomato (*Lycopersicon esculentum*, Mill.) and Aubergine (*Solanum melongena* L.). *Scientia Horticulturae*, 109 (2), 142-146.
- Van, S. N., Minh, H. D., and Anh, D. N. (2013). Study on Chitosan Nanoparticles on Biophysical Characteristics and Growth of *Robusta coffee* in Green House. *Biocatalysis and Agricultural Biotechnology*, 2 (4), 289-294.
- Vasyukova, N. I., Zinov'Eva, S. V., II'Inskaya, L. I., Perekhod, E. A., Chalenko, G. I., Gerasimova, N. G., ... and Ozeretskovskaya, O. L. (2001). Modulation of Plant Resistance to Diseases by Water-soluble Chitosan. *Applied Biochemistry and Microbiology*, 37 (1), 103-109.
- Volf, I., Ignat, I., Neamtu, M., and Popa, V. I. (2014). Thermal Stability, Antioxidant Activity, and Photo-oxidation of Natural Polyphenols. *Chemical Papers*, 68 (1), 121-129.

- Vongsak, B., Sithisarn, P., Mangmool, S., Thongpraditchote, S., Wongkrajang, Y., and Gritsanapan, W. (2013). Maximizing Total Phenolics, Total Flavonoids Contents and Antioxidant Activity of *Moringa oleifera* Leaf Extract by the Appropriate Extraction Method. *Industrial Crops and Products*, 44, 566-571.
- Vosoughi, N., Gomarian, M., Pirbalouti, A. G., Khaghani, S., and Malekpoor, F. (2018). Essential Oil Composition and Total Phenolic, Flavonoid Contents, and Antioxidant Activity of Sage (*Salvia officinalis* L.) Extract under Chitosan Application and Irrigation Frequencies. *Industrial Crops and Products*, 117, 366-374.
- Wakeel, A., Jan, S. A., Ullah, I., Shinwari, Z. K., and Xu, M. (2019). Solvent Polarity Mediates Phytochemical Yield and Antioxidant Capacity of *Isatis Tinctoria. PeerJ*, 7, e7857.
- Wang, C., He, J., Zhao, T. H., Cao, Y., Wang, G., Sun, B., ... and Li, M. H. (2019b). The Smaller the Leaf is, the Faster the Leaf Water Loses in a Temperate forest. *Frontiers in Plant Science*, 10, 58.
- Wang, C., Shi, L., Fan, L., Ding, Y., Zhao, S., Liu, Y., and Ma, C. (2013). Optimization of Extraction and Enrichment of Phenolics from Pomegranate (*Punica granatum* L.) Leaves. *Industrial Crops and Products*, 42, 587-594.
- Wang, E., and Wink, M. (2016). Chlorophyll Enhances Oxidative Stress Tolerance in *Caenorhabditis elegans* and Extends its Lifespan. *PeerJ*, 4, e1879.
- Wang, M., Nie, H., Han, D., Qiao, X., Yan, H., and Shen, S. (2019a). Cauliflowerlike Resin Microspheres with Tuneable Surface Roughness as Solid-phase Extraction Adsorbent for Efficient Extraction and Determination of Plant Growth Regulators in Cucumbers. *Food Chemistry*, 295, 259-266.
- Wang, W., Li, S., Zhao, X., Du, Y., and Lin, B. (2008). Oligo-chitosan Induces Cell Death and Hydrogen Peroxide Accumulation in Tobacco Suspension Cells. Pesticide Biochemistry and Physiology, 90, 106–113.
- Wang, Y., Zhao, J., Lu, W., and Deng, D. (2017). Gibberellin in Plant Height Control: Old Player, New Story. *Plant Cell Reports*, 36 (3), 391-398.
- Went, F. W. (1935). Auxin, the Plant growth-hormone. *The Botanical Review*, 1 (5), 162-182.
- Wittkopp, A., and Schreiner, P. R. (2003). Metal-free, Noncovalent Catalysis of Diels–alder Reactions by Neutral Hydrogen Bond Donors in Organic Solvents and in Water. *Chemistry–A European Journal*, 9 (2), 407-414.
- Wongsen, W., Bodhipadma, K., Noichinda, S., and Leung, D. W. M. (2015). Influence of Different 2, 4-D Concentrations on Antioxidant Contents and

Activities in Sweet Basil Leaf-derived Callus During Proliferation. *International Food Research Journal*, 22 (2), 638.

- Wu, Q., Pagès, L., and Wu, J. (2016). Relationships Between Root Diameter, Root Length and Root Branching Along Lateral Roots in Adult, Field-grown Maize. *Annals of Botany*, 117 (3), 379-390.
- Wu, Y., Cui, S. W., Tang, J., and Gu, X. (2007). Optimization of Extraction Process of Crude Polysaccharides from Boat-fruited Sterculia Seeds by Response Surface Methodology. *Food Chemistry*, 105 (4), 1599-1605.
- Xie, J. H., Dong, C. J., Nie, S. P., Li, F., Wang, Z. J., Shen, M. Y., and Xie, M. Y. (2015). Extraction, Chemical Composition and Antioxidant Activity of Flavonoids from *Cyclocarya paliurus* (Batal.) Iljinskaja Leaves. *Food chemistry*, 186, 97-105.
- Xu, C., and Mou, B. (2018). Chitosan as Soil Amendment Affects Lettuce Growth, Photochemical Efficiency, and Gas Exchange. *HortTechnology*, 28 (4), 476-480.
- Xu, D., Li, H., Lin, L., Liao, M. A., Deng, Q., Wang, J., ... and Xia, H. (2020). Effects of Carboxymethyl Chitosan on the Growth and Nutrient Uptake in *Prunus davidiana* Seedlings. *Physiology and Molecular Biology of Plants*, 26 (4), 661-668.
- Yaldız, G., Çamlıca, M., Özen, F., and Eratalar, S. A. (2019). Effect of Poultry Manure on Yield and Nutrient Composition of Sweet Basil (*Ocimum basilicum* L.). *Communications in soil Science and Plant Analysis*, 50 (7), 838-852.
- Yang, J., Kloepper, J. W., and Ryu, C. M. (2009). Rhizosphere Bacteria Help Plants Tolerate Abiotic Stress. *Trends in Plant Science*, 14 (1), 1-4.
- Yermiyahu, U., Halpern, M., and Shtienberg, D. (2020). NH 4 Fertilization Increases Susceptibility of Sweet Basil (*Ocimum basilicum* L.) to Grey Mould (*Botrytis cinerea*) Due to Decrease in Ca Uptake. *Phytoparasitica*, 48 (5), 685-697.
- Yin, H., Fretté, X. C., Christensen, L. P., and Grevsen, K. (2012). Chitosan Oligosaccharides Promote the Content of Polyphenols in Greek Oregano (*Origanum vulgare* ssp. hirtum). *Journal of Agricultural and Food Chemistry*, 60 (1), 136-143.
- Yormaz, B., Kahraman, H., and Koksal, N. (2017). Evaluation of Bone Demineralization in Copd Phenotypes. ACTA Medica Mediterranea, 33 (4), 627-636.

- Yu, J., Ahmedna, M., and Goktepe, I. (2005). Effects of Processing Methods and Extraction Solvents on Concentration and Antioxidant Activity of Peanut Skin Phenolics. *Food Chemistry*, 90 (1-2), 199-206.
- Yu, J., Ahmedna, M., and Goktepe, I. (2005). Effects of Processing Methods and Extraction Solvents on Concentration and Antioxidant Activity of Peanut Skin Phenolics. *Food Chemistry*, 90 (1-2), 199-206.
- Yu, J., Wang, D., Geetha, N., Khawar, K. M., Jogaiah, S., and Mujtaba, M. (2021). Current Trends and Challenges in the Synthesis and Applications of Chitosan-based Nanocomposites for Plants: A Review. *Carbohydrate Polymers*, 261, 117904.
- Yu, M., Wang, B., Qi, Z., Xin, G., and Li, W. (2019). Response Surface Method Was Used to Optimize the Ultrasonic Assisted Extraction of Flavonoids from *Crinum asiaticum*. Saudi Journal of Biological Sciences, 26 (8), 2079-2084.
- Yu, M., Wang, B., Qi, Z., Xin, G., and Li, W. (2019). Response Surface Method Was Used to Optimize the Ultrasonic Assisted Extraction of Flavonoids from *Crinum asiaticum*. Saudi Journal of Biological Sciences, 26 (8), 2079-2084.
- Yuniastuti, E., Waliyyudin, Q., and Delfianti, M. N. I. (2021, January). Improvement Growth of Sapodilla (*Achras zapota* L.) by Chitosan. In *IOP Conference Series: Earth and Environmental Science*, Vol. 637, No. 1, p. 012095.
- Zayed, M. M., Elkafafi, S. H., Zedan, A. M., and Dawoud, S. F. (2017). Effect of Nano Chitosan on Growth, Physiological and Biochemical Parameters of *Phaseolus vulgaris* under Salt Stress. *Journal of Plant Production*, 8 (5), 577-585.
- Zhang, B., Hettiarachchy, N., Chen, P., Horax, R., Cornelious, B., and Zhu, D. (2006). Influence of the Application of Three Different Elicitors on Soybean Plants on the Concentrations of Several Isoflavones in Soybean Seeds. *Journal of Agricultural and Food Chemistry*, 54 (15), 5548-5554.
- Zhang, B., Yang, R., and Liu, C. (2008). Microwave-assisted Extraction of Chlorogenic Acid from Flower Buds of *Lonicera japonica* Thunb. *Separation and Purification Technology*, 62 (2), 480–483.
- Zhang, M., Hu, X., Zhu, M., Xu, M., and Wang, L. (2017). Transcription Factors NF-YA2 and NF-YA10 Regulate Leaf Growth via Auxin Signaling in Arabidopsis. *Scientific Reports*, 7 (1), 1-9.
- Zhang, Q. W., Lin, L. G., and Ye, W. C. (2018). Techniques for Extraction and Isolation of Natural Products: A Comprehensive Review. *Chinese Medicine*, 13 (1), 1-26.

- Zhang, Z. S., Li, D., Wang, L. J., Ozkan, N., Chen, X. D., Mao, Z. H., and Yang, H. Z. (2007). Optimization of Ethanol–water Extraction of Lignans from Flaxseed. Separation and Purification Technology, 57 (1), 17-24.
- Zhao, H., Cao, H. H., Pan, M. Z., Sun, Y. X., and Liu, T. X. (2017). The Role of Plant Growth Regulators in a Plant–aphid–parasitoid Tritrophic System. *Journal of Plant Growth Regulation*, 36 (4), 868-876.
- Zhao, J., Davis, L. C., and Verporte, R. (2005). Elicitor Signal Transduction Leading to Production of Plant Secondary Metabolites. *Biotechnology Advances*, 23, 283–333.
- Zhaohui, W., and Shengxiu, L. (2002). Effects of Water Deficit and Supplemental Irrigation at Different Growing Stage on Uptake and Distribution of Nitrogen, Phosphorus and Potassium in Winter Wheat. *Plant Nutrition and Fertitizer Science*, 8 (3), 265-270.
- Zhou, Y. G., Yang, Y. D., Qi, Y. G., Zhang, Z. M., Wang, X. J., and Hu, X. J. (2002). Effects of Chitosan on Some Physiological Activity in Germinating Seed of Peanut. *Journal of Peanut Science*, 31 (1), 22-25.
- Zong, H., Liu, S., Xing, R., Chen, X., and Li, P. (2017). Protective Effect of Chitosan on Photosynthesis and Antioxidative Defense System in Edible Rape (*Brassica rapa* L.) in the Presence of Cadmium. *Ecotoxicology and Environmental Safety*, 138, 271-278.
- Zuppini, A., Baldan, B., Millioni, R., Favaron, F., Navazio, L., and Mariani, P. (2003). Chitosan Induces Ca2+ mediated Programmed Cell Death in Soybean Cells. *New Phytologist, Phytologist*, 161, 557–568.