



UNIVERSITI PUTRA MALAYSIA

***REPRODUCTIVE BIOLOGY AND POPULATION DYNAMICS OF
Crassostrea (Magallana) saidii WONG & SIGWART, 2021 IN THE
ESTUARY OF SUNGAI MUAR, JOHOR, MALAYSIA***

ARUN CHANDRA BARMAN

FP 2022 35



REPRODUCTIVE BIOLOGY AND POPULATION DYNAMICS OF *Crassostrea (Magallana) saidii* WONG & SIGWART, 2021 IN THE ESTUARY OF SUNGAI MUAR, JOHOR, MALAYSIA

By

ARUN CHANDRA BARMAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

July 2022

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

To my parents, beloved wife (Doli Rani Kar) and lovely daughters (Ahenjita Barman Alo & Aratrika Barman Tory), who always kept praying for me day and night to achieve my goal

To my family members

and

To all my friends who supported me all these years



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

REPRODUCTIVE BIOLOGY AND POPULATION DYNAMICS OF *Crassostrea (Magallana) saidii* WONG & SIGWART, 2021 IN THE ESTUARY OF SUNGAI MUAR, JOHOR, MALAYSIA

By

ARUN CHANDRA BARMAN

July 2022

Chairman : Nur Leena Wong Wai Sin, PhD
Faculty : Agriculture

Crassostrea (Magallana) saidii Wong & Sigwart, 2021 was recently described as a new oyster species to science. It is an endemic species with its only known distribution limited to Muar estuary, Malaysia, and has been actively harvested from its natural habitat by the locals since at least 1858. This latest taxonomic development warrants a closer look into the reproductive biology and population structure of *C. (M.) saidii*, which was thought to be a morph-variant of other oyster species in the same vicinity but has a widespread distribution in the country. Additionally, concern arises as the population size is believed to reduce, implicating local fisheries. However, the objectives of this research were to provide information on the reproductive biology and population dynamics of *C. (M.) saidii* from the estuary of Sungai Muar, Johor, Malaysia. The first study was conducted to evaluate the sex ratio and reproductive phenology of *C. (M.) saidii* through gonad histology and biometry. Samples of 30 individual oysters were taken monthly from November 2018 to January 2020 from the sole population at Sungai Muar estuary, Johor, Malaysia. Histology revealed that among 450 oysters analyzed, male, female, hermaphrodite, and undifferentiated individuals were at 35.33%, 58%, 1.56%, and 5.11%, respectively. Overall male and female sex ratio was 1:1.64, with female numbers significantly higher. However, the sex ratio was different when the oysters were divided into groups according to shell height (SH) in intervals of 10 mm. The proportion of males decreased with increasing shell height, although the shell height of males and females greatly overlapped. The sex reversal pattern of the oyster might go from male to female and again female to male, with an indicator of rhythmical hermaphroditism. In the current study, statistically significant differences in the size of oogenic cells (mature > maturing > degenerating > immature > oogonia) and the spermatogenic cells (spermatogonia > spermatocyte > spermatid > spermatozoa) were observed. Gametogenesis occurred around the research period with remarkable time-based variations. The oyster spawned over the study period, with peaks in April (2019) and November (2018 and 2019). Condition index (CI) ranged from 70.03 (October 2019) to 31.90 (April 2019), exhibiting four peaks when most of the oysters were maturing

stage and subsequently decreases in CIs indicating spawning. Three distinct peaks in maturity indices (MIs) of the oyster were noted during March, August, and October 2019, when the majority of oysters were mature and ready for spawning. The second study was carried out to correlate environmental variables with the gonadal maturity of *C. (M.) saidii*. Thirty oysters were taken monthly from November 2018 to January 2020 from the Sungai Muar estuary. The findings showed that there was no significant correlation between water quality parameters like temperature, salinity, pH and total suspended solids, whereas a significant positive correlation was identified between upstream rainfall and spawning gonad ($r^2 = 0.534$, $p < 0.05$). A multiple regression between spawning individuals and environmental variables (temperature, pH, salinity, total suspended solids, and rainfall) indicated that rainfall ($\beta = 0.535$, $p < 0.05$) triggered the spawning of the oyster *C. (M.) saidii* in the estuary. The effects of salinity on the gonadal maturation of the oyster were observed in a controlled environment, conducted in the International Institute of Aquaculture and Aquatic Sciences (I-AQUAS), Universiti Putra Malaysia (UPM), between December 2020 and February 2021 with a duration of 60 days. The oysters were conditioned using three salinities of 10, 15, and 20 ppt. The study revealed that all the salinities were suitable for the maturation of the gonad of the oysters after 30 days and 60 days of the observation. The condition index was not affected by the salinity concentrations over the study period. The species' size-weight relationship and population structure were assessed in 2019 to determine its population dynamics. Results indicated that the gradient of the size-weight equation ($b = 2.5422$) was significantly different ($t = 38.93$, $p < 0.01$) from the isometric value (3), indicating negative allometric growth of the *C. (M.) saidii* in the estuary. The asymptotic shell height (SH_{∞}) and growth coefficient (K) were observed at 15.23 cm and 0.69 per year, respectively. The maximum observed height and predicted extreme height was 14.50 cm and 14.72 cm, respectively. The calculated growth performance index (ϕ') was 2.204. The annual total mortality (Z) using the length-converted catch curve was calculated at 3.02, annual fishing mortality (F) at 1.27, and annual natural mortality (M) at 1.75. The recruitment pattern of the oyster showed continuous recruitment, with two peaks over the study period. The exploitation level (E) of the oyster was computed at 0.42, while the allowable limit of exploitation (E_{max}) was 0.782 for the highest yield. The current exploitation level ($E = 0.42$) indicates that the oyster stock was exploited at almost the optimum ($E = 0.5$) from the estuary. The understanding of the reproductive biology and population dynamics could provide information for optimal and accurate management of the natural population of oyster *C. (M.) saidii* in its sole habitat. The identification of maturation and spawning season of this species will allow their conservation through aquaculture approach while reducing commercial pressures from the natural stock.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

BIOLOGI REPRODUKTIF DAN DINAMIK PENDUDUK *Crassostrea (Magallana) saidii* WONG & SIGWART, 2021 DI MUARA SUNGAI MUAR, JOHOR, MALAYSIA

Oleh

ARUN CHANDRA BARMAN

Julai 2022

Pengerusi : Nur Leena Wong Wai Sin, PhD
Fakulti : Pertanian

Crassostrea (Magallana) saidii Wong & Sigwart, 2021 baru-baru ini telah diklasifikasikan sebagai spesies tiram baharu. Ia adalah spesies endemik dengan taburan yang terhad di muara Muar, Malaysia, dan telah dituai secara aktif dari habitat semula jadi oleh penduduk tempatan sejak tahun 1858. Perkembangan taksonomi terkini dapat menentukan dengan lebih tepat berkenaan biologi pembiakan dan struktur populasi *C. (M.) saidii*, yang dianggap sebagai varian-morf bagi spesies tiram lain di kawasan yang sama tetapi mempunyai taburan yang meluas di negara ini. Selain itu, saiz populasi yang dipercayai berkurangan membabitkan perikanan tempatan menambahkan kebimbangan akan spesies ini. Walau bagaimanapun, objektif penyelidikan ini adalah untuk memberi maklumat tentang biologi pembiakan dan dinamik populasi *C. (M.) saidii* dari muara Sungai Muar, Johor, Malaysia. Kajian pertama dijalankan untuk menilai nisbah jantina dan fenologi pembiakan *C. (M.) saidii* melalui kaedah histologi gonad dan biometri. Sebanyak 30 sampel tiram telah diambil setiap bulan dari November 2018 hingga Januari 2020 daripada populasinya di muara Sungai Muar, Johor, Malaysia. Kajian histologi mendedahkan bahawa antara 450 tiram yang dianalisis, jantan, betina, hermafrodit, dan individu yang tidak dibezakan adalah masing-masing pada 35.33%, 58%, 1.56%, dan 5.11%. Nisbah jantina jantan dan betina keseluruhan ialah 1:1.64, dengan bilangan betina jauh lebih tinggi. Walau bagaimanapun, nisbah jantina adalah berbeza apabila tiram dibahagikan kepada kumpulan mengikut ketinggian cangkerang (*SH*) dalam selang 10 mm. Perkadaran jantan berkurangan dengan peningkatan ketinggian cangkerang, walaupun ketinggian cangkerang jantan dan betina banyak bertindih. Corak pembalikan jantina tiram mungkin berubah dari jantan ke betina, dan sekali lagi betina ke jantan, dengan penunjuk hermafroditisme berirama. Dalam kajian ini, perbezaan ketara secara statistik dalam saiz sel oogenik (matang > matang > merosot > tidak matang > oogonia) dan sel spermatogenik (spermatogonia > spermatosit > spermatid > spermatozoa) telah diperhatikan. Gametogenesis berlaku sekitar tempoh penyelidikan dengan variasi berasaskan masa. Tiram bertelur sepanjang tempoh kajian, dengan puncaknya pada April (2019) dan November (2018 dan 2019). Indeks keadaan (*CI*) berjulat dari 70.03 (Oktober

2019) hingga 31.90 (April 2019), mempamerkan empat kemuncak apabila kebanyakan tiram berada dalam peringkat matang dan seterusnya nilai *CI* menurun yang menunjukkan pemijahan. Tiga kemuncak yang berbeza dalam indeks kematangan (*MI*) tiram telah dicatatkan pada bulan Mac, Ogos dan Oktober 2019, apabila majoriti tiram telah matang dan bersedia untuk bertelur. Eksperimen kedua kemudiannya dijalankan untuk mengaitkan pembolehubah persekitaran dengan kematangan gonad *C. (M.) saidii*. Sebanyak 30 ekor tiram diambil setiap bulan dari November 2018 hingga Januari 2020 dari muara Sungai Muar. Dapatan kajian menunjukkan bahawa tidak terdapat perhubungan yang signifikan antara parameter kualiti air seperti suhu, kemasinan, pH dan jumlah pepejal terampai, manakala perhubungan positif yang signifikan telah dikenalpasti antara hujan dan pemijahan gonad ($r^2 = 0.534$, $p < 0.05$). Regresi berganda antara individu pemijahan dan pembolehubah persekitaran (suhu, pH, kemasinan, jumlah pepejal terampai, dan hujan) menunjukkan bahawa hujan ($\beta = 0.535$, $p < 0.05$) merangsang pemijahan tiram *C. (M.) saidii* dalam muara sungai. Kesan kemasinan ke atas pematangan gonad tiram diperhatikan dalam eksperimen terkawal, yang dijalankan di Institut Akuakultur dan Sains Akuatik Antarabangsa (I-AQUAS), Universiti Putra Malaysia (UPM), antara Disember 2020 dan Februari 2021 dalam tempoh 60 hari. Tiga tahap kemasinan 10, 15, dan 20 ppt telah ditetapkan untuk kajian ini. Kajian menunjukkan bahawa semua kemasinan adalah sesuai untuk pematangan gonad tiram selepas 30 hari dan 60 hari pemerhatian. Indeks keadaan tidak dipengaruhi oleh kepekatan kemasinan sepanjang tempoh kajian. Hubungan saiz-berat spesies dan struktur populasi telah dinilai pada 2019 untuk menentukan dinamik populasinya. Keputusan menunjukkan bahawa kecerunan persamaan saiz-berat ($b = 2.5422$) adalah berbeza secara ketara ($t = 38.93$, $p < 0.01$) daripada nilai isometrik (3), menunjukkan pertumbuhan alometri negatif bagi *C. (M.) saidii* dalam muara. Ketinggian cengkering asimptotik (SH_{∞}) dan pekali pertumbuhan (K) diperhatikan pada 15.23 cm dan 0.69 setahun. Ketinggian maksimum yang diperhatikan dan ketinggian paling tinggi yang diramalkan ialah 14.50 cm dan 14.72 cm. Indeks prestasi pertumbuhan yang dikira (ϕ) ialah 2.204. Jumlah kematian tahunan (Z) menggunakan keluk tangkapan tukar panjang dikira pada 3.02, kematian penangkapan ikan tahunan (F) pada 1.27, dan kematian semula jadi tahunan (M) pada 1.75. Corak pengambilan tiram menunjukkan pengambilan berterusan, dengan dua kemuncak sepanjang tempoh kajian. Tahap eksploitasi (E) tiram dikira pada 0.42, manakala had eksploitasi yang dibenarkan (E_{maks}) ialah 0.782 untuk hasil tertinggi. Tahap eksploitasi semasa ($E = 0.42$) menunjukkan bahawa stok tiram telah dieksploitasi pada kadar yang optimum ($E = 0.5$) dari muara. Pemahaman tentang biologi pembiakan dan dinamik populasi boleh membantu untuk menyediakan pengurusan yang optimum dan tepat bagi stok tersedia tiram *C. (M.) saidii* di habitat tunggalnya. Pengenalpastian musim kematangan dan pemijahan spesies ini akan membolehkan pemuliharaan tiram ini melalui pendekatan akuakultur sambil mengurangkan tekanan komersial daripada stok semula jadinya.

ACKNOWLEDGEMENTS

I would like to express my sincere gratitude to Dr. Nur Leena Wong Wai Sin, the Chairman of the Supervisory Committee, for her invaluable supervision, productive discussion, constructive criticisms, and continuous encouragement throughout the study. I am much grateful and indebted to Assoc. Professor Dr. Murni Marlina binti Abd. Karim and Dr. Mohd Zafri bin Hassan, members of the supervisory committee, for their constructive suggestions and guidance in all phases of this research project. I express my deep respect to all of the lecturers and professors in the Department of Aquaculture for their valuable suggestions and constructive comments.

I am grateful to the laboratory staff of the Department of Aquaculture, Faculty of Agriculture

I am indebted to my parents, wife, daughter, brothers, and sisters for their support, deep love, and tolerance during the whole period of my study.

Thankfully acknowledge the assistance of the National Agricultural Technology Program- Phase 2 project (NATP-2), Bangladesh Agricultural Research Council (BARC), Farmgate, Dhaka-1215, Bangladesh, for financial support and facilitating my study and Universiti Putra Malaysia for providing research facilities under the Aquaculture Department, Faculty of Agriculture.

I certify that a Thesis Examination Committee has met on 6 July 2022 to conduct the final examination of Arun Chandra Barman on his thesis entitled “Reproductive biology and population dynamics of *Crassostrea (Magallana) saidii* Wong & Sigwart, 2021 in the estuary of Sungai Muar, Johor, Malaysia” in accordance with the universities and university college act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15th March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Examination Committee were as follows:

Muta Harah Zakaria, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Aziz Arshad, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Annie Christianus, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

ZURIATI AHMAD ZUKARNAIN, PhD

Professor Ts. and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirements for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Nur Leena Wong Wai Sin, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Murni Marlina binti Abd. Karim, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Mohd Zafri bin Hassan, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 10 November 2022

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: Dr. Nur Leena Wong Wai Sin

Signature: _____
Name of Member
of Supervisory
Committee: Associate Professor
Dr. Murni Marlina binti Abd. Karim

Signature: _____
Name of Member
of Supervisory
Committee: Dr. Mohd Zafri bin Hassan

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	vii
LIST OF TABLES	xiii
LIST OF FIGURES	xv
LIST OF ABBREVIATIONS	xix
CHAPTER	
1 INTRODUCTION	1
1.1 Background of the study	1
1.2 Statement of problems	2
1.3 Significance of the study	2
1.4 Objectives	3
2 LITERATURE REVIEW	4
2.1 Taxonomy of <i>Crassostrea (Magallana) saidii</i>	4
2.2 Description of <i>Crassostrea (Magallana) saidii</i>	5
2.2.1 Morphological characteristics of <i>C. (M.) saidii</i>	5
2.2.2 Etymology of <i>C. (M.) saidii</i>	6
2.2.3 Ecological observations of <i>C. (M.) saidii</i>	6
2.2.4 Comparative remarks of <i>C. (M.) saidii</i> with co-occurring species	6
2.3 Habitat and distribution of oysters	6
2.4 Global oyster production	8
2.5 Potentiality of oysters in Malaysia	8
2.6 Biological aspects of oysters	9
2.6.1 Sex ratio of oysters	9
2.6.2 Length at first maturity	10
2.6.3 Fecundity and size of germ cell	10
2.6.4 Gonad development stages	11
2.6.5 Condition index (CI)	12
2.6.6 Maturity index (MI)	12
2.7 Water quality parameters	12
2.7.1 Temperature	13
2.7.2 Salinity	13
2.7.3 pH	14
2.7.4 Suspended organic matter	14
2.8 Rainfall influence on oyster maturity	14
2.9 Conditioning of oysters in the hatchery	15
2.10 Population dynamics	15
2.10.1 Length-weight relationship	15
2.10.2 Stock assessment	16

3	GENERAL METHODOLOGY	17
3.1	Sampling location	17
3.2	Samples collection	18
3.3	Oyster measurement and index	18
3.4	Collection and preservation of gonad sample	19
3.5	Histology	19
4	SEX RATIO AND GONADAL DEVELOPMENT OF <i>Crassostrea (Magallana) saidii</i> WONG AND SIGWART, 2021 THROUGH GONAD HISTOLOGY AND BIOMETRY FROM THE ESTUARY OF SUNGAI MUAR, JOHOR, MALAYSIA	20
4.1	Introduction	20
4.2	Materials and methods	21
4.2.1	Sex ratio estimation	21
4.2.2	Germinal cells identification and measurement	21
4.2.3	Identification of gonadal developmental phases	23
4.2.4	Condition index (CI)	25
4.2.5	Maturity index (MI)	25
4.2.6	Statistical analysis	26
4.3	Results	26
4.3.1	Biometry	26
4.3.2	Sex ratio	29
4.3.3	Size of germinal cells	32
4.3.4	Gonad maturation stages	34
4.3.5	Condition index (CI)	40
4.3.6	Maturity index (MI)	42
4.4	Discussion	43
4.5	Conclusion	45
5	CORRELATION OF ENVIRONMENTAL VARIABLES WITH THE GONADAL MATURITY OF <i>Crassostrea (Magallana) saidii</i> AT SUNGAI MUAR, JOHOR	47
5.1	Introduction	47
5.2	Materials and Methods	47
5.2.1	Water quality	47
5.2.2	Rainfall data	48
5.2.3	Statistical analysis	49
5.3	Results	50
5.3.1	<i>In-situ</i> water parameters	50
5.3.2	Rainfall	53
5.3.3	Correlation analysis outcome	55
5.3.4	Regression analysis outcome	58
5.4	Discussion	59
5.5	Conclusion	61
6	IMPACTS OF SALINITY ON GONAD MATURATION OF THE OYSTER <i>Crassostrea (Magallana) saidii</i> IN EX-SITU CONDITION	62
6.1	Introduction	62
6.2	Materials and methods	62
6.2.1	Experimental site and duration	62

6.2.2	Experimental design	63
6.2.3	Statistical analysis	64
6.3	Results	64
6.3.1	Biometry	64
6.3.2	Gonad maturation stages	66
6.3.3	Condition index (CI)	69
6.3.4	Maturity index (MI)	70
6.4	Discussion	71
6.5	Conclusion	73
7	SIZE-WEIGHT RELATIONSHIP AND POPULATION DYNAMICS OF THE OYSTER <i>Crassostrea (Magallana) saidii</i> COLLECTED FROM THE ESTUARY OF SUNGAI MUAR, JOHOR, MALAYSIA	74
7.1	Introduction	74
7.2	Materials and methods	74
7.2.1	Oyster measurement and index	74
7.2.2	Statistical analysis	75
7.3	Results	77
7.3.1	Size-weight relationship	77
7.3.2	Size-frequency distribution	77
7.3.3	Growth parameters	78
7.3.4	Mortality pattern, exploitation rate and exploitation size	81
7.3.5	Catch pattern (Virtual Population Analysis)	83
7.3.6	Recruitment pattern	83
7.3.7	Relative yield per recruitment and biomass per recruitment	84
7.4	Discussion	86
7.5	Conclusion	89
8	SUMMARY, CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE RESEARCH	90
8.1	Discussion and Conclusions	90
8.2	Recommendations for future research	92
	RERERENCES	93
	APPENDICES	113
	BIODATA OF STUDENT	117
	LIST OF PUBLICATIONS	118

LIST OF TABLES

Table		Page
4.1	Identification characteristics of female germinal cells (adapted and modified from Castilho-Westphal et al., (2015)	22
4.2	Identification characteristics (adopted and modified from Castilho-Westphal et al. (2015) of male germinal cells	22
4.3	Gonadal development stages of male and female oyster adopted and modified from Brousseau (1995)	24
4.4	Gonad maturation stages and their numerical Ranking Score used in the calculation of maturity index	26
4.5	Monthly samples (N = 30) analysis for <i>Crassostrea (M.) saidii</i> collected from the estuary of Sungai Muar, Johor	27
4.6	The biometric data according to the sex of the oyster <i>C. (M.) saidii</i> collected from the estuary of Sungai Muar, Johor, Malaysia, during the study period	30
4.7	Average (\pm S.E) and minimum and maximum Size of different oogenic cells of <i>C. (M.) saidii</i> collected from the estuary of Sungai, Muar, Johor	33
4.8	Average (\pm S.E), and minimum and maximum size in parentheses of spermatogenic germinal cells of <i>C. (M.) saidii</i> from the estuary of Sungai, Muar, Johor	34
4.9	Monthly gonadal maturation stages of the <i>C. (M.) saidii</i> collected from the estuary of Sungai Muar, Johor, Malaysia.	37
4.10	Correlation matrices (Spearman) among condition index (CI), maturity index (MI), and monthly gonad stages (%) of the <i>C. (M.) saidii</i> collected from the estuary of Sungai Muar, Johor over the 15-month study period	41
5.1	Correlation matrices (Spearman) among different water quality parameters and the rainfall over the 15-month study period in the estuary of Sungai Muar, Johor	56
5.2	Correlation between water quality parameters and gonad maturation stages in the estuary of Sungai Muar Johor	57
5.3	Correlation between gonad maturation stages of <i>C. (M.) saidii</i> and rainfall of different stations	58

5.4	Partial coefficients of multiple regression evaluating the relationship of the water quality parameters and rainfall of Kuala Pilah with the spawning gonad	59
6.1	The summary of the biometry measurement of the <i>C. (M.) saidii</i> used in the experiment	65
6.2	Pre-experiment gonadal maturation stages of the <i>C. (M.) saidii</i> collected from the estuary of Sungai Muar, Johor, Malaysia	66
6.3	The outcome of the Chi-Square Goodness of Fit Test between the number of spawning gonads of <i>C. (M.) saidii</i> recorded at pre-experiment and at different salinity concentrations after 30 days of observation	68
6.4	The outcome of the Chi-Square Goodness of Fit Test between the number of spawning gonads of <i>C. (M.) saidii</i> recorded at pre-experiment and at different salinity concentrations after 60 days of observation	68
6.5	The outcome of the Pearson's chi-square test among the gonadal stages of the <i>C. (M.) saidii</i> observed in <i>ex-situ</i> conditions during the study period	69
7.1	Population parameters of <i>C. (M.) saidii</i> in the estuary of Sungai Muar, Johor, Malaysia	86
7.2	Growth parameters of some <i>Crassostrea</i> species from different studies. Here L_{∞} = asymptotic length, $K \text{ yr}^{-1}$ = growth coefficient per year, and ϕ' = growth performance index	88

LIST OF FIGURES

Figure	Page
2.1	5
The internal and external appearance of <i>C. (M.) saidii</i> showing left valve (a, b); right valve (c, d); dorsal view showing flat hinge line (e); soft parts in live animal with right valve removed (f) (Sigwart et al., 2021)	
2.2	7
Distribution of some oyster species in Malaysia	
2.3	8
Global oyster production (in Million MT) by sector (2012–2018). Source: FAO (2020). Fishery and Aquaculture Commodities Statistics 2018/FAO Yearbook	
2.4	9
Quantity of export and import of oyster commodities during 2007-2018 (Department of Fisheries Malaysia, 2020)	
3.1	17
Map indicating the location of the study site at Sungai Muar estuary, Johor, Malaysia (Ellipse)	
3.2	18
Morphological measurements of the shell of the oyster. SH, shell height; SL, shell length and SW, shell width	
4.1	29
Photomicrograph depicts the cross section of different sexes of <i>C. (M.) saidii</i> . Here, A = male with spermatogenic cell (Sz), B = female with oogenic cell (Oo), C = undifferentiated gonad with storage connective tissue (SCT) but without sperm or egg, and D = hermaphrodite with both oogenetic (Oo) and spermatogenic (Sz) cells, Mag. 10x; Scale bar: 100 μ m, Staining: H&E	
4.2	31
The pie chart showing the percentage of different sexes of 450 <i>C. (M.) saidii</i> collected from the Sungai Muar estuary, Johor, observed through gonad histology	
4.3	32
Bar graph showing the percentage of the male and female subject to shell height of the <i>C. (M.) saidii</i> . At the top of the bar, the digit indicated the total number of oysters counted in the respective group	
4.4	33
Ovarian germ cells. A, different parts of oocyte (FW = follicle wall, Nu = nucleus, VtM = vitelline membrane, An = antrum); B, germ cells at different stages of development (Og = oogonia, IO = immature oocyte, OM = oocyte on maturation, M = mature oocyte and Do = degenerative oocyte). Mag. 40x, Scale bar: 30 μ m and 25 μ m for A and B, respectively, Staining: H&E	

4.5	Spermatogenic germ cells. A, spermatogonia (Sg), spermatid (St), B, spermatocyte (Sp) and spermatozoa (Sz). Mag. 40x, Scale bar: 100 μ m, Staining: H&E	34
4.6	Maturation stages of female <i>C. (M.) saidii</i> (A-E) and undifferentiated (F) oyster. (A) early development; (B) late development; (C) ripe; (D) spawning and (E) spent. Here, SCT = storage connective tissue; FL = follicle lumen; FW = follicle wall; Og = oogonia; IO = immature oocyte; FO = free oocyte inside lumen, OM = maturing oocyte; MO = mature oocyte; VC = void cells; RO = residual oocytes. Mag. 10x, Scale bar: 100 μ m, Staining: H&E	35
4.7	Maturation stages of male <i>C. (M.) saidii</i> (A-E) and hermaphrodite (F) oyster. (A) early development; (B) late development; (C) ripe; (D) spawning and (E) spent. Here, SPG = spermatogonia; GA = gonadal acinus; SCT = storage connective tissue; SPC = spermatocytes; SPZ = spermatozoa; VC = void cells; RS = residual spermatozoa; O = Oocyte; S = spermatozooids. Mag. 10x, Scale bar: 100 μ m, Staining: H&E	36
4.8	Temporal distribution of the gonadal maturation stages of the Muar Oyster <i>C. (M.) saidii</i> collected from the estuary Sungai Muar, Johor, between November 2018 and January 2020. Here A, described the overall gonad maturation stages, B, male gonad maturation stages, and C, female gonad maturation stages	39
4.9	Month-wise changes in condition index (Mean \pm S.E) of sampling <i>C. (M.) saidii</i> in the estuary Sungai Muar, Johor	40
4.10	Variations of condition index in relation to gonad stages of the <i>C. (M.) saidii</i> . Graphs represent mean size and interquartile range. Here, CI = condition index, ED = early development, LD = late development, Und = undifferentiated	41
4.11	Month-wise changes in maturity index of the <i>C. (M.) saidii</i> in the estuary Sungai Muar, Johor	42
5.1	Map depicted the three meteorological stations of Malaysia, including the sampling site. Here, KPS = Kuala Pilah station; MS = Melaka station; RA = Research area and BPS = Batu Pahat station	49
5.2	Monthly average temperature ($^{\circ}$ C) from November 2018 to January 2020 recorded from the sampling site during the study period	50
5.3	Monthly variations in salinity concentration (mean \pm S.E) during the study period in the estuary Sungai Muar, Johor	51
5.4	Monthly variation in pH (Mean \pm S.E) during the study period in the estuary of Sungai Muar Johor, Malaysia. The same letter above	52

bar indicates a statistically insignificant ($P > 0.05$) variation in monthly pH value using the Tukey HSD test

5.5	Monthly variation in TSS concentration (Mean \pm S.E) in the estuary of Sungai Muar, Johor during the study period. The same letter above the bar indicates statistically insignificant differences between the TSS level ($P > 0.05$), whereas different letters indicate significant differences ($p < 0.05$) using the Tukey HSD test	53
5.6	Daily rainfall data for November 2018 to January 2020 recorded at Kuala Pilah (A), Melaka (B) and Batu Pahat (C) meteorological station, Johor, obtained from the Malaysian Meteorological Department	54
5.7	Monthly rainfall data for November 2018 to January 2020 recorded at Kuala Pilah, Melaka and Batu Pahat meteorological station, Johor, obtained from the Malaysian Meteorological Department	55
6.1	Rearing techniques of <i>C. (M.) saidii</i> at different salinity concentrations in the experiment. Here, NN = nylon net that allows 50% light penetration in the rearing tank	63
6.2	Percentage of gonad stages of the <i>C. (M.) saidii</i> observed in different salinities after 30 days of the experimental period. Here, Und, undifferentiated; SP, spawning, LD, late development and ED, early development	66
6.3	Percentage of gonad stages of the <i>C. (M.) saidii</i> observed in different salinities after 60 days of the experimental period. Here, Und, undifferentiated; SP, spawning, LD, late development and ED, early development	67
6.4	Salinity-based changes in mean values of condition index of the oyster <i>C. (M.) saidii</i> during the experimental period. The same letter above the bars represents a statistically insignificant condition index in different salinity concentrations ($P > 0.05$)	70
6.5	Maturity index of the oyster <i>C. (M.) saidii</i> recorded in different salinity concentrations during the experiment	70
7.1	Size-weight relationship of the oyster <i>C. (M.) saidii</i> collected from the estuary of Sungai Muar, Johor, Malaysia	77
7.2	Month-wise height-frequency distribution of the oyster <i>C. (M.) saidii</i> collected from the estuary of Sungai Muar, Johor, Malaysia, between January 2019 to December 2019	78
7.3	The predicted maximum height for the <i>C. (M.) saidii</i> according to extreme value theory (Formacion et al., 1994). The predicted extreme height and the extreme height at 95% confidence were	79

	gained from the incision of the overall extreme height from the y and x, z lines, respectively	
7.4	The von Bertalanffy growth function fitted to size at age relationship for the <i>C. (M.) saidii</i> in the estuary of Sungai Muar, Johor	79
7.5	Restructured height-frequency distribution of <i>C. (M.) saidii</i> from the estuary of Sungai Muar Johor, Malaysia (L_{∞} = 15.23 cm and K = 0.69 per year). The white and black histograms were a negative and positive deviation from the weighted moving average of three length classes, representing pseudo-cohorts	80
7.6	K -scan routine for the best value of von Bertalanffy growth function (VBGF), asymptotic length (L_{∞}), and growth coefficients (K) of the <i>C. (M.) saidii</i> using ELEFAN-1	81
7.7	The size converted catch curve of the <i>C. (M.) saidii</i> . The least-square linear regression was used to determine the catch curve. The solids dots points were used in the estimation, while the open dots reflect the relative ages that were either not fully recruited or close to L_{∞}	82
7.8	Probability of capture of the oyster <i>C. (M.) saidii</i> in the estuary of Sungai Muar, Johor, Malaysia. The length classes indicated by cross-hatched bars were used to estimate the probability of capture of the oyster. In contrast, the open bars were the length classes that were not fully included in the probability calculation. The Red line indicates the trend of capture	82
7.9	Histograms depicting the mortality and catch pattern of the collected oyster <i>C. (M.) saidii</i>	83
7.10	Bimodal distribution of recruitment pattern of <i>C. (M.) saidii</i> from the estuary of Sungai Muar, Johor Malaysia. The backward projection of the restricted height-frequency data was used to obtain the recruitment pattern onto a one-year timescale	84
7.11	Beverton and Holt's Knife-edge selection to prepare a model on relative yield per recruitment (Y/R) and average biomass per recruitment (B/R) of the oyster <i>C. (M.) saidii</i> . The ascending curve indicated relative yield per recruitment, and the descending curve pointed to average biomass per recruitment. Estimating exploitation ratios (E) (on the x-axis) are useful for evaluating management options. Yellow dashed lines = E_{max} : exploitation rate which produces maximum yield, (Y/R) (g per year) and (B/R) (g). Green dashed lines $E_{0.1}$: exploitation rate at which the marginal increase in relative yield per-recruit is $1/10^{\text{th}}$ of its value at $E = 0$. Red dashed lines = $E_{0.5}$: the value of E below which the stock has been reduced to 50% of its unexploited biomass	85

LIST OF ABBREVIATIONS

CI	Condition index
cm	Centimeter
DoF	Department of Fisheries
E	Exploitation rate
ED	Early Development
ELEFAN	Electronic Length Frequency Analysis
F	Fishing mortality
FAO	Food and Agricultural Organization
FiSAT	FAO ICLARM Stock Assessment Tools
g	Gram
K	Growth coefficient of VBGF
Kg	Kilogram
Km	Kilometer
L	Liter
LD	Late Development
MI	Maturity index
mg/L	Milligram per liter
mm	Millimeter
NN	Nylon net
ppt	Parts per thousand
S.E	Standard error
Und	Undifferentiated
UPM	Universiti Putra Malaysia
μm	Micrometer

CHAPTER 1

INTRODUCTION

1.1 Background of the study

Oysters are ecologically keystone species in marine ecosystems by providing ecological services as filter feeders, clarifiers and reef-builders, and for the last reason, they are known as bioengineers (Smaal et al., 2019). They are listed as commercially important species within the invertebrates in many regions of the world (Piedra et al., 2014; Posada et al., 2014). However, in Malaysia, two oyster species are mainly used for commercial purposes, namely *Crassostrea iredalei* and *Crassostrea belcheri* (Suzana et al., 2011). Fishers in Malaysia usually collect spats from the wild to culture oysters on a commercial basis, but currently, the supply of seeds of the oyster has declined (Fakhrina et al., 2018) due to siltation, acidification, disease, overfishing, and lack of proper knowledge on management (Tan et al., 2014).

In Malaysia, marine science has received much attention in recent years, but marine molluscs studies are still overseen by many researchers (Wong & Arshad, 2011). However, some studies on the culture of oysters in Malaysia have been conducted, supported by the International Development and Research Canada (IDRC) (Tan et al., 2014). The larval development of local oyster species *C. belcheri* and *C. iredalei* larvae has been established in the country, considering substrates, storage temperature, and salinity (Devakie & Ali, 2002). The chemical interaction on larval settlement of the oysters has also been studied (Teh et al., 2012). Moreover, the relationship between environmental parameters on the development and survival of broodstock oysters was also documented (Hawkins et al., 1998; Izwandy, 2006). Genetic diversities in two cultured oyster species, *C. iredalei* and *C. belcheri* were assessed (Suzana et al., 2011). The use of genetics approaches of diploid and triploid (Rahim et al., 2008) and tetraploid induction in oysters, *C. iredalei* and *C. belcheri* also has been studied (Tan et al., 2017). Besides, gonad maturation, larval development, and settlement of the slipper cupped oyster *C. iredalei* was recorded at Sungai Terus, Batu Lintang, Kedah (Wahab, 2017). A systematics description of the Muar oyster *Crassostrea (Magallana) saidii* was reported in the estuary of Sungai Muar, Johor, Malaysia (Sigwart et al., 2021).

The oyster *C. (M.) saidii* has been marketed and consumed in Malaysia for over 160 years as an undescribed species. Recently, this oyster was named by Sigwart et al. (2021) according to a resident of Muar, Md Saidi Bin Mohamed, who has been actively promoting research and conservation for the sustainability of this species. The Muar oyster *C. (M.) saidii*, locally known as *Tiram Putih* (white oyster), co-occurring with the worldwide established species *C. belcheri* (*Tiram kapak*, Axe oyster). The oyster is distributed from the river mouth of Muar River, Johor, Malaysia to approximately 6 km upstream. The oyster *C. (M.) saidii* is morphologically and genetically distinct, limited to Sungai Muar, Malaysia, the only known population in the scientific record (Sigwart et al., 2021). Because of its creamy white meat and excellent quality, the species is

known as the white oyster among fisherfolk, and locals, and is more popular with oyster fans than the *C. belcheri* (Axe Oyster) species, which can also be found in the estuary of Muar (Sigwart et al., 2021). The oyster fishery in Muar estuary was first reported in 1858 by a British Captain who described Muar oyster as a delicacy reserved for the royals and high officials (Macpherson, 1858). It was then sold at the Singapore market, which is about 200 km away (Hanitsch, 1908), but later Muar oyster fishery was reported as unsustainable in 1929, probably due to the decline in the oyster population (Dover, 1929). Muar oysters have been farmed through bottom ranching by returning empty oyster shells to certain points of the Sungai Muar estuary, and it is still practiced to this day.

The oyster *C. (M.) saidii*, however, due to urbanization, is on under potential threat, and the production of this species is declining in the estuary (Sigwart et al., 2021). This declining led to the research on the *C. (M.) saidii*, especially in reproductive biology and population dynamics

1.2 Statement of problems

The oyster *Crassostrea (Magallana) saidii* is an important fishery in the estuary of Sungai Muar, Johor and the small population is threatened by urbanization, pollution, and habitat destruction (Sigwart et al., 2021). Moreover, in Malaysia, wild seeds are the mainly source for the farmers to culture this oyster. Again, there is no study conducted on the artificial conditioning of *C. (M.) saidii* which can facilitate its introduction in aquaculture. Although, it is important to know the reproductive biology and population structure to sustain its natural stock, there is no information on the biological characterization and *ex-situ* conditioning of the Muar Oyster *C. (M.) saidii*. Therefore, it is essential to find out the spawning environment, artificial conditioning, and population structure of the oyster in the respective area to facilitate its conservation and effective seed production for aquaculture. Furthermore, the population dynamics and stock assessment will provide the base to avoid overfishing and for the sustainable management of this species. Therefore, the current research will focus on the gonad development, correlation between environmental factors and gonadal maturation, impacts of salinity on the maturation of gonads in *ex-situ* conditions, and the population parameters of the *C. (M.) saidii* in the estuary of Sungai Muar, Johor, Malaysia.

1.3 Significance of the study

The information on gonad development could be useful for assessing the possible breeding behavior of *C. (M.) saidii*. Moreover, a thorough understanding of these reproductive cycles, condition index, and maturity index will allow farmers to determine the optimal moments for harvesting, intending to obtain oysters with a higher meat weight before spawning and seed collection. The knowledge of the relationship between the environmental variables and gonadal development are essential for the planning of breeding and seed collection for *C. (M.) saidii*. Besides, the information on the impacts of different salinities on gonad maturation in *ex-situ* conditions will provide information

in extending the seed production season of *C. (M.) saidii* for commercial purposes. The commercial importance of oysters in Malaysia is increasing gradually with the growing demand for oysters for human consumption. The research studies focusing on its population parameters will be of considerable necessity for future economic valorization and sustainable management of the species in Malaysian waters.

The present study investigated the biology and the population dynamics of the edible oyster *C. (M.) saidii* to define the biological characterization of the oyster population in Sungai Muar estuary. Data from the present study provide essential information for the sustainable management and proper utilization of the oyster *C. (M.) saidii* natural stocks, and possibly for the similar oyster stocks of the world.

1.4 Objectives

The objectives of the research were as follows:

1. To identify sex ratio and phenology of the oyster *C. (M.) saidii* through gonad histology and biometry
2. To determine the correlation between environmental variables and the gonadal maturity of the oyster *C. (M.) saidii* at Sungai Muar, Johor
3. To observe the impacts of salinity on the gonadal development of the oyster *C. (M.) saidii* in *ex-situ* condition.
4. To assess the length-weight relationship and population parameters of the oyster *C. (M.) saidii* at Sungai Muar.

REFERENCES

- Admodisastro, V. A., Ransangan, J., Ilias, N., & Tan, S. H. (2021). Oyster farming potential in Sabah, Malaysia. *International Journal of Aquatics Research*, 2(1), 17–22.
- Akélé, G. D., Agadjihouèdé, H., Mensah, G. A., & Lalèyè, P. A. (2015). Population dynamics of freshwater oyster *Etheria elliptica* (Bivalvia: Etheriidae) in the Pendjari River (Benin-Western Africa). *Knowledge and Management of Aquatic Ecosystems*, 416, 1–15. <https://doi.org/10.1051/kmae/2015002>
- Al-Barwani, S. M., Arshad, A., Amin, S. M. N., Japar, S. B., Siraj, S. S., & Yap, C. K. (2007). Population dynamics of the green mussel *Perna viridis* from the high spat-fall coastal water of Malacca, Peninsular Malaysia. *Fisheries Research*, 84(2), 147–152. <https://doi.org/10.1016/j.fishres.2006.10.021>
- Albuquerque, M. C. P., Ferreira, J. F., Salvador, G. C., & Turini, C. (2018). Influence of temperature and salinity on survival and growth of larvae of the pearl oyster *Pteria hirundo*. *Bulletin of the Fisheries Institute*, 38(3), 189–197.
- Alvarenga, L., & Nalesso, R. C. (2006). Preliminary assessment of the potential for mangrove oyster cultivation in Piraquê-açu river estuary (Aracruz, ES). *Brazilian Archives of Biology and Technology*, 49(1), 163–169. <https://doi.org/10.1590/S1516-89132006000100019>
- Amin, S. M. N., Zafar, M., & Halim, A. (2008). Age, growth, mortality and population structure of the oyster, *Crassostrea madrasensis*, in the Moheskhali Channel (southeastern coast of Bangladesh). *Journal of Applied Ichthyology*, 24(1), 18–25. <https://doi.org/10.1111/j.1439-0426.2007.01007.x>
- Andrews, J. D., Giese, A. C., & Pearse, J. S. (1979). Pelecypoda: ostreidae. *Reproduction of Marine Invertebrates*, 293–341.
- Angell, C. L. (1986). *The Biology and Culture of Tropical Oysters. ICLARM Studies and Reviews*. International Center for Living Aquatic Resources Management. Manila, Philippines.
- Antonio, Í., Sousa, A., Lenz, T., Funo, I., Lopes, R., & Figueiredo, M. (2021). Reproductive cycle of the mangrove oyster, *Crassostrea rhizophorae* (Bivalvia: Ostreidae) cultured in a macrotidal high-salinity zone on the Amazon mangrove coast of Brazil. *Acta Amazonica*, 51, 113–121.
- Bacon, P. R. (1970). Studies on the biology and cultivation of the mangrove oyster in Trinidad with notes on other shellfish resources. *Tropical Science*, 12(1), 265–278.

- Bagenda, D. K., Nishikawa, S., Kita, H., Kinai, Y., Terai, S., Kato, M., & Kasai, H. (2019). Impact of feeding on oyster depuration efficacy under conditions of high salinity and low temperature. *Aquaculture*, 500(July 2018), 135–140. <https://doi.org/10.1016/j.aquaculture.2018.10.009>
- Baghurst, B. C., & Mitchell, J. G. (2002). Sex-specific growth and condition of the Pacific oyster (*Crassostrea gigas* Thunberg). *Aquaculture Research*, 33(15), 1253–1263. <https://doi.org/10.1046/j.1365-2109.2002.00788.x>
- Barber, B. J., & Blake, N. J. (1991). Reproductive physiology. In S. E. Shumway (Editor), *Scallops; Biology, Ecology and Aquaculture*. Elsevier, Amsterdam, 41: 377-428.
- Barber, B. J., Ford, S. E., & Haskin, H. H. (1988). Effects of the parasite MSX (*Haplosporidium nelsoni*) on oyster (*Crassostrea virginica*) energy metabolism-II. Tissue biochemical composition. *Comparative Biochemistry and Physiology -- Part A: Physiology*, 91(3), 603–608. [https://doi.org/10.1016/0300-9629\(88\)90641-X](https://doi.org/10.1016/0300-9629(88)90641-X)
- Batista, F. (2007). *Assessment of the aquacultural potential of the Portuguese oyster Crassostrea angulata*. Instituto de Ciências Biomédicas Abel Salazar da Universidade do Porto. p. 302.
- Behmer, O. A., & Tolosa, E. M. C. de. (2003). Manual of techniques for normal and pathological histology. In *In Manual of techniques for normal and pathological histology* (p. 331).
- Bhromanonda, P. (1972). Biology and cultivation sataus of Coastal Aquaculture in Thailand. In: Thai fisheries in the future. Bangkok. *Thai Fisheries Gazette*, 31(3), 215–228.
- Boulais, M., Chenevert, K. J., Demey, A. T., Darrow, E. S., Robison, M. R., Roberts, J. P., & Volety, A. (2017). Oyster reproduction is compromised by acidification experienced seasonally in coastal regions. *Scientific Reports*, 7(1), 1–9. <https://doi.org/10.1038/s41598-017-13480-3>
- Breese, W. P., & Robinson, A. (1981). Razor clams, *Siliqua patula* (Dixon): gonadal development, induced spawning and larval rearing. *Aquaculture*, 22, 27–33.
- Brousseau, D. J. (1995). Gametogenesis and spawning in intertidal oysters (*Crassostrea virginica*) from Western long Island Sound. *Journal of Shellfish Research*, 14(2), 483–487.
- Buroker, N. E. (1983). Sexuality with respect to shell length and group size in the Japanese oyster *Crassostrea gigas*. *Malacologia*, 23(2), 271–279.
- Butler, P. A. (1949). Gametogenesis in the oyster under conditions of depressed salinity. *The Biological Bulletin*, 96(3), 263–269.

- Cadima, E. L. (2003). *Fish stock assessment manual* (Issue 393). Food & Agriculture Org.
- Carlander, K. (1977). *Handbook of freshwater fishery biology*. The Iowa State University press, Amesderdam, Netherland.
- Carrasco, M. A., Brito Manzano, N. P., Gómez Vázquez, A., & Cruz-Hernández, A. (2018). Effects of temperature and salinity on inducing spawning in the eastern oyster (*Crassostrea virginica*) under laboratory conditions. *Ecosistemas Y Recursos Agropecuarios*, 5(14), 239. <https://doi.org/10.19136/era.a5n14.1236>
- Casas, S. M., Lavaud, R., La Peyre, M. K., Comeau, L. A., Filgueira, R., & La Peyre, J. F. (2018). Quantifying salinity and season effects on eastern oyster clearance and oxygen consumption rates. *Marine Biology*, 165(5), 1–13.
- Castaños, C., Pascual, M., & Camacho, A. P. (2009). Reproductive biology of the nonnative oyster, *Crassostrea gigas* (thunberg, 1793), as a key factor for its successful spread along the rocky shores of Northern Patagonia, Argentina. *Journal of Shellfish Research*, 28(4), 837–847. <https://doi.org/10.2983/035.028.0413>
- Castilho-Westphal, G. G., Magnani, F. P., & Ostrensky, A. (2015). Gonad morphology and reproductive cycle of the mangrove oyster *Crassostrea brasiliiana* (Lamarck, 1819) in the baía de Guaratuba, Paraná, Brazil. *Acta Zoologica*, 96(1), 99–107. <https://doi.org/10.1111/azo.12055>
- Chávez-Villalba, J., Pommier, J., Andriamiseza, J., Pouvreau, S., Barret, J., Cochard, J. C., & Le Pennec, M. (2002). Broodstock conditioning of the oyster *Crassostrea gigas*: Origin and temperature effect. *Aquaculture*, 214(1–4), 115–130. [https://doi.org/10.1016/S0044-8486\(01\)00898-5](https://doi.org/10.1016/S0044-8486(01)00898-5)
- Chávez-Villalba, J., Villelas-Ávila, R., & Cáceres-Martínez, C. (2007). Reproduction, condition and mortality of the Pacific oyster *Crassostrea gigas* (Thunberg) in Sonora, México. *Aquaculture Research*, 38(3), 268–278.
- Chin, P. K., & Lim, A. L. (1975). Oyster culture development in Sabah (Malaysia). *Sabah Society Journal*, 6, 108–115.
- Choo, P. S. (1983). *Oyster culture. SAFIS Extension Manual No. 2* (pp. 1–16). Southeast Asian Fisheries Development Center, Bangkok.
- Christo, S. W., & Absher, T. M. (2006). Reproductive period of *Crassostrea rhizophorae* (Guilding, 1828) and *Crassostrea brasiliiana* (Lamarck, 1819)(Bivalvia: ostreidae) in Guaratuba bay, Paraná, Brazil. *Journal of Coastal Research*, 1215–1218.

- Chumkiew, S., Jaroensutasinee, M., Tina, F. W., Jaroensutasinee, K., & Koad, P. (2019). Physical factors affecting oyster diversity and distribution in Southern Thailand. *Journal of Environmental Biology*, *40*(1), 3–8.
- Chung, E.-Y., Seo, Y.-H., & Park, K. H. (1998). Sexual maturation, sex ratio and hermaphroditism of the Pacific oyster, *Crassostrea gigas*, on the west coast of Korea. *Fisheries and Aquatic Sciences*, *1*(1), 82–93.
- Cone, R. S. (1989). The need to reconsider the use of condition indices in fishery science. *Transactions of the American Fisheries Society*, *118*(5), 510–514.
- Costa, V. B. D., De Sousa, E. B., Pinheiro, S. C. C., Pereira, L. C. C., & da Costa, R. M. (2011). Effects of a high energy coastal environment on the structure and dynamics of phytoplankton communities (Brazilian Amazon littoral). *Journal of Coastal Research*, 354–358.
- Crosby, M. P., & Gale, D. L. (1990). A review and evaluation of bivalve condition index methodologies with a suggested standard method. *Journal of Shellfish Research*, *9*, 233–237.
- Dangwatanakul, R. (1992). *Reproductive biology of the large oyster Crassostrea belcheri in Thailand*. M S Thesis. Mahidul University, Thailand.
- Daria, E. B., Radonic, I., Arapov, J., Varezic, D., Zorica, B., Stagic'ic, N., Jozic, S., Peharda, M., Briski, E., Lin, Y., & S'egvic-Bubic, T. (2020). Reproductive cycle of the non-native Pacific oyster, *Crassostrea gigas*, in the Adriatic Sea. *Mediterranean Marine Science*, *21*(1), 146–156.
- Derbali, A., Kandeel, K. E., & Jarboui, O. (2020). Comparison of the dynamics between coastal and midshore populations of *Pinctada radiata* (Leach, 1814) (mollusca: Bivalvia) in the gulf of Gabes, Tunisia. *Turkish Journal of Fisheries and Aquatic Sciences*, *20*(4), 301–310. https://doi.org/10.4194/1303-2712-v20_4_06
- Devakie, M. N., & Ali, A. B. (2002). Effective use of plastic sheet as substrate in enhancing tropical oyster (*Crassostrea iredalei* Faustino) larvae settlement in the hatchery. *Aquaculture*, *212*(1–4), 277–287. [https://doi.org/10.1016/S0044-8486\(02\)00270-3](https://doi.org/10.1016/S0044-8486(02)00270-3)
- Devakie, M. N., Hall, R., & Angell, C. L. (1993). *Small-scale oyster culture on the West Coast of peninsular Malaysia*. BOBP/REP/63. GCP/RAS/118/MUL. BOBP, Madras, India.
- Dinamani, P. (1987). Gametogenic patterns in populations of Pacific oyster, *Crassostrea gigas*, in Northland, New Zealand. *Aquaculture*, *64*(1), 65–76.

- DoF. (2020). *Export and Import of Fishery Commodities by Country of Destination and Origin, 2007-2018*, Retrieved 10-28-2020 from <https://www.dof.gov.my/index.php/pages/view/3260>.
- Doinsing, J. W., & Ransangan, J. (2022). Population dynamics of the tropical oyster *Magallana bilineata* (Mollusca, Bivalvia, Ostreidae) in Mengkabong Bay, Tuaran, Malaysia. *Aquaculture and Fisheries*, June 2021, 1–11. <https://doi.org/10.1016/j.aaf.2022.04.005>
- Dore, I. (1991). *Shellfish: a guide to oysters, mussels, scallops, clams, and similar products for the commercial user*. Van Nostrand Reinhold.
- Doroudi, M. S., Southgate, P. C., & Mayer, R. J. (1999). The combined effects of temperature and salinity on embryos and larvae of the black-lip pearl oyster, *Pinctada margaritifera* (L.). *Aquaculture Research*, 30(4), 271–277.
- Dover, C. (1929). Oyster culture in Malaya. *Nature*, 124(3120), 264–265.
- Drummond, L., Mulcahy, M., & Culloty, S. (2006). The reproductive biology of the Manila clam, *Ruditapes philippinarum*, from the North-West of Ireland. *Aquaculture*, 254(1–4), 326–340.
- Dupuy, J. L., & Rivkin, S. (1972). The development of laboratory techniques for the production of cultch-free spat of the oyster, *Crassostrea virginica*. *Chesapeake Science*, 13(1), 45–52. <https://doi.org/10.2307/1350550>
- Dutertre, M., Beninger, P. G., Barillé, L., Papin, M., Rosa, P., Barillé, A. L., & Haure, J. (2009). Temperature and seston quantity and quality effects on field reproduction of farmed oysters, *Crassostrea gigas*, in Bourgneuf Bay, France. *Aquatic Living Resources*, 22(3), 319–329. <https://doi.org/10.1051/alr/2009042>
- Elamin, E. M., & Elamin, S. E. M. (2014). Biometric relationships of the mother of pearl oyster (*Pinctada margaritifera* var *erythraensis*). *International Journal of Science, Environment*, 3(3), 1193–1204.
- Enríquez-Díaz, M., Pouvreau, S., Chávez-Villalba, J., & Le Pennec, M. (2009). Gametogenesis, reproductive investment, and spawning behavior of the Pacific giant oyster *Crassostrea gigas*: Evidence of an environment-dependent strategy. *Aquaculture International*, 17(5), 491–506. <https://doi.org/10.1007/s10499-008-9219-1>
- Etchian, O. A., Pellerin, J., Audet, C., & Mathieu, M. (2004). Sexual maturation and related changes in aspartate transcarbamylase activity of gonad tissues in the soft shell clam (*Mya arenaria*). *Comparative Biochemistry and Physiology - B Biochemistry and Molecular Biology*, 139(2), 287–297. <https://doi.org/10.1016/j.cbpc.2004.08.006>

- Fabioux, C., Huvet, A., Le Souchu, P., Le Pennec, M., & Pouvreau, S. (2005). Temperature and photoperiod drive *Crassostrea gigas* reproductive internal clock. *Aquaculture*, 250(1–2), 458–470.
- Fakhrina, M. N., Christianus, A., & Ehteshamei, F. (2018). Production of tropical oyster seed in hatchery. *Journal of Survey in Fisheries Sciences*, 5(1), 7–19. <https://doi.org/10.18331/SFS2018.5.1.2>
- FAO. (2009). *FAO-Crassostrea gigas*. https://www.fao.org/fishery/FAOdocs/DOCUMENT/aquaculture/CulturedSpecies/file/en/en_pacific. accessed 2020-02-01.
- FAO. (2020). *The State of World Fisheries and Aquaculture 2020. Sustainability in action*. Rome. <https://doi.org/https://doi.org/10.4060/ca9229en>
- Ferreira, M. A. P., Paixão, L. F., Alcântara-Neto, C. P., Santos, S. S. D., & Rocha, R. M. (2006). Morphological and morphometric aspects of *Crassostrea rhizophorae* (Guilding, 1828) oocytes in three stages of the gonadal cycle. *International Journal of Morphology*, 24(3), 437–442. <https://doi.org/10.4067/S0717-95022006000400023>
- Formacion, S. P., Rongo, J. M., & Sambilay Jr, V. C. (1994). Extreme value theory applied to the statistical distribution of the largest lengths of fish. *UP Research Digest (Philippines)*.
- Fournier, M.-L. (1992). The reproductive biology of the tropical rocky oyster *Ostrea iridescens* (Bivalvia: Ostreidae) on the Pacific coast of Costa Rica. *Aquaculture*, 101(3–4), 371–378.
- Froese, R. (2004). Keep it simple: Three indicators to deal with overfishing. *Fish and Fisheries*, 5(1), 86–91. <https://doi.org/10.1111/j.1467-2979.2004.00144.x>
- Fujita, T. (1934). Note on the Japanese oyster larvae. *Fifth Pacific Science Congress*, 5, 4111–4117.
- Gallager, S. M., & Mann, R. (1986). Growth and survival of larvae of *Mercenaria mercenaria* (L.) and *Crassostrea virginica* (Gmelin) relative to broodstock conditioning and lipid content of eggs. *Aquaculture*, 56(2), 105–121. [https://doi.org/10.1016/0044-8486\(86\)90021-9](https://doi.org/10.1016/0044-8486(86)90021-9)
- Galtsoff, P. S. (1964). *The American Oyster Crassostrea virginica (Gmelin)*. (64th ed.). Fishery Bulletin, United States Government Printing Office.
- Galtsoff, P. S., Prytherch, H. F., & McMillin, H. C. (1930). *An experimental study in production and collection of seed oysters* (Issue 1088). US Government Printing Office.

- Galvao, M. S. N., Pereira, O. M., Machado, M. B., & Henrique, I. C. (2000). Aspectos reprodutivos da ostra *Crassostrea brasiliiana* de manguezais do estuário de Cananéia, SP (25°S; 48°W). *Boletim Do Instituto de Pesca, São Paulo*, 26(2), 147–162.
- Gaspar, M. B., Santos, M. N., & Vasconcelos, P. (2001). Weight–length relationships of 25 bivalve species (Mollusca: Bivalvia) from the Algarve coast (southern Portugal). *Journal of the Marine Biological Association of the United Kingdom*, 81(5), 805–807.
- Gauthier-Clerc, S., Pellerin, J., & Amiard, J. C. (2006). Estradiol-17 β and testosterone concentrations in male and female *Mya arenaria* (Mollusca bivalvia) during the reproductive cycle. *General and Comparative Endocrinology*, 145(2), 133–139. <https://doi.org/10.1016/j.ygcen.2005.08.004>
- Gayanilo, F. C., Sparre, P., & Pauly, D. (1996). *FAO-ICLARM stock assessment tools: user's manual*. Food and Agriculture Organization of the United Nations, Rome, Italy.
- George-Zamora, A., Sevilla-Hernández, M. L., & Aldana-Aranda, D. (2003). Ciclo gonádico del ostión americano *Crassostrea virginica* (Lamellibranchia: Ostreidae) en Mecoaacán, Tabasco, México. *Revista de Biología Tropical*, 51(4), 109–117.
- Gomes, C., Silva, F., Lopes, G., & Melo, C. (2014). The reproductive cycle of the oyster *Crassostrea gasar*. *Brazilian Journal of Biology*, 74(4), 967–976. <https://doi.org/10.1590/1519-6984.04912>
- Góngora-Gómez, A. M., Sepúlveda, C. H., Verdugo Escobar, H. A., Astorga Castro, O., Rodríguez-González, H., Domínguez-Orozco, A. L., Hernández-Sepúlveda, J. A., & García-Ulloa, M. (2020). Gonadal maturity of *Crassostrea corteziensis* cultivated in the Gulf of California. *Latin American Journal of Aquatic Research*, 48(3), 381–395. <https://doi.org/10.3856/vol48-issue3-fulltext-2422>
- Gosling, E. M. (2003). *Bivalve molluscs: biology, ecology, and culture*. Fishing News Books, Oxford. <http://library1.nida.ac.th/termpaper6/sd/2554/19755>.
- Grant, A., & Tyler, P. A. (1983). The analysis of data in studies of invertebrate reproduction. II. The analysis of oocyte size/frequency data, and comparison of different types of data. *International Journal of Invertebrate Reproduction*, 6(5–6), 271–283. <https://doi.org/10.1080/01651269.1983.10510053>
- Grave, C. (1912). *A Manual of Oyster Culture in Maryland*. Board Shellfish Community. Maryland, 4th Rep, pp 279-348.
- Guillou, E., Raymond, A., Krien, N., & Buschini, F. (2019). Oyster eaters: From consumer practices to the representation of risks. *Appetite*, 140(April), 105–113. <https://doi.org/10.1016/j.appet.2019.04.012>

- Gulland, J. A. (1965). Estimation of mortality rates. Annex to Arctic fisheries working group report ICES C.M./1965/D:3. (mimeo). In P. H. Cushing (Eds.), *Key papers on fish populations* (pp. 231-241). IRL Press, Oxford, UK.
- Gulland, J. A. (1971). *Fish Resources of the Ocean*. Fishing News Books, London. pp 255.
- Guo, X., Hedgecock, D., Hershberger, W. K., Cooper, K., & Allen, S. K. (1998). Genetic determinants of protandric sex in the Pacific oyster, *Crassostrea gigas* Thunberg. *Evolution*, 52(2), 394–402. <https://doi.org/10.1111/j.1558-5646.1998.tb01640.x>
- Hanitsch, R. (1908). *Guide to the Zoological Collections of the Raffles Museum*. Singapore Straits Times Press.
- Hassan, M. M., Qin, J. G., & Li, X. (2016). Spermatozeugmata structure and dissociation of the Australian flat oyster *Ostrea angasi*: Implications for reproductive strategy. *Tissue and Cell*, 48(3), 152–159. <https://doi.org/10.1016/j.tice.2016.03.002>
- Hassan, M. M., Qin, J. G., & Li, X. (2018). Gametogenesis, sex ratio and energy metabolism in *Ostrea angasi*: Implications for the reproductive strategy of spermcasting marine bivalves. *Journal of Molluscan Studies*, 84(1), 38–45. <https://doi.org/10.1093/mollus/eyx041>
- Hawkins, A. J. S., Smith, R. F. M., Tan, S. H., & Yasin, Z. B. (1998). Suspension-feeding behaviour in tropical bivalve molluscs: *Perna viridis*, *Crassostrea belcheri*, *Crassostrea iradelei*, *Saccostrea cucullata* and *Pinctada margarifera*. *Marine Ecology Progress Series*, 166, 173–185. <https://doi.org/10.3354/meps166173>
- Helm, M. M., & Bourne, N. (2004). *Hatchery culture of bivalves. FAO fisheries technical paper no. 471*. FAO Rome.
- Helm, M. M., Holland, D. L., & Stephenson, R. R. (1973). The effect of supplementary algal feeding of a hatchery breeding stock of *Ostrea edulis* L. on larval vigour. *Journal of the Marine Biological Association of the United Kingdom*, 53(3), 673–684.
- Hermawati, S., Sulistiono, & M., S. (2017). Distribution, Condition and Gonad Maturity of the Invasive Pacific Oysters (*Crassostrea gigas*, Thunberg 1793) in Cimanuk Delta, Indramayu, West Java, Indonesia. *Omni-Akuatika*, 13(2), 99–110.
- Hernández-Covarrubias, V., Luis, J., & Hugo, P. (2013). Inferencia multimodelo : cálculo de la talla media de madurez del ostión de roca *Striostrea prismatica* en Nayarit , México. *Ciencia Pesquera*, 22, 11–18.
- Hilborn, R., & Walters, C. J. (2013). *Quantitative fisheries stock assessment: choice, dynamics and uncertainty*. Springer Science & Business Media.

- His, E., Robert, R., & Dinet, A. (1989). Combined effects of temperature and salinity on fed and starved larvae of the mediterranean mussel *Mytilus galloprovincialis* and the Japanese oyster *Crassostrea gigas*. *Marine Biology*, 100(4), 455–463. <https://doi.org/10.1007/BF00394822>
- Hopkins, A. E. (1931). Factors influencing the spawning and Setting of oysters. *Bulletin of the Bureau of Fisheries*, 47, 55.
- Hornbach, D. J., & Childers, D. L. (1987). The effects of acidification on life-history traits of the freshwater clam *Musculium partumeium* (Say, 1822) (Bivalvia:Pisidiidae). *Canadian Journal of Zoology*, 65(1), 113–121. <https://doi.org/10.1139/z87-017>
- Hsiao, S. T., Chuang, S. C., Chen, K. S., Ho, P. H., Wu, C. L., & Chen, C. A. (2016). DNA barcoding reveals that the common cupped oyster in Taiwan is the Portuguese oyster *Crassostrea angulata* (Ostreoida; Ostreidae), not *C. gigas*. *Scientific Reports*, 6(March), 1–11. <https://doi.org/10.1038/srep34057>
- Hughes-Games, W. L. (1977). Growing the japanese oyster. *Elsevier Scientific Publishing Company*, 11, 217–229.
- Huo, Z., Wang, Z., Liang, J., Zhang, Y., Shen, J., Yao, T., Su, J., & Yu, R. (2014). Effects of salinity on embryonic development, survival, and growth of *Crassostrea hongkongensis*. *Journal of Ocean University of China*, 13(4), 666–670. <https://doi.org/10.1007/s11802-014-2206-4>
- Hutchinson, S., & Hawkins, L. E. (1992). Quantification of the physiological responses of the European flat Oyster *Ostrea edulis* L. to temperature and salinity. *Journal of Molluscan Studies*, 58(2), 215–226. <https://doi.org/10.1093/mollus/58.2.215>
- Izwandy, I. (2006). *The factors that influencing on environmental growth and survival of Crassostrea iredalei (Faustino) oyster commercial at the oyster farming area. case study: kg. Tenaga Nenas, Perak*. M S Thesis. Universiti Sains Malaysia.
- Joyce, A., & Vogeler, S. (2018). Molluscan bivalve settlement and metamorphosis: Neuroendocrine inducers and morphogenetic responses. *Aquaculture*, 487(January), 64–82. <https://doi.org/10.1016/j.aquaculture.2018.01.002>
- Jumatli, A., & Ismail, M. S. (2021). Promotion of Sustainable Aquaculture in Malaysia. *In Proceedings of the International Workshop on the Promotion of Sustainable Aquaculture, Aquatic Animal Health, and Resource Enhancement in Southeast Asia, December*, 31–40.
- Kandeel, K. E., Mohammed, S. Z., Mostafa, A. M., & Abd-Alla, M. E. (2013). Reproductive biology of the cockle *Cerastoderma glaucum* (Bivalvia: Cardiidae) from Lake Qarun, Egypt. *The Egyptian Journal of Aquatic Research*, 39(4), 249–260.

- Kennedy, A. V., & Battle, H. I. (1964). Cyclic changes in the gonad of the American oyster, *Crassostrea virginica* (Gmelin). *Canadian Journal of Zoology*, 42(2), 305–321.
- King, M. (2007). *Fisheries Biology, Assessment and Management, second ed.* Blackwell Scientific Publications, Oxford, UK.
- King, R. P. (1996). Length-weight relationships of Nigerian coastal water fishes. *Naga, The ICLARM Quarterly*, 19(4), 53–58 1996.
- Kuriakose, S., Mini, K. G., & Sathianandan, T. V. (2017). *Course Manual ICAR funded Summer School on Advanced Methods for Fish Stock Assessment and Fisheries Management.* CMFRI; Kochi. 367pp.
- Lam, K., & Morton, B. (2004). The oysters of Hong Kong (Bivalvia: Ostreidae and Gryphaeidae). *Raffles Bulletin of Zoology*, 52(1), 11–28.
- Lango-reynoso, F., Chávez-Villaba, J., & Le Pennec, M. (2006). Reproductive patterns of the pacific oyster *Crassostrea gigas* in France. *Invertebrate Reproduction and Development*, 49(1–2), 41–50. <https://doi.org/10.1080/07924259.2006.9652192>
- Lango-Reynoso, F., Chávez-Villalba, J., Cochard, J. C., & Le Pennec, M. (2000). Oocyte size, a means to evaluate the gametogenic development of the Pacific oyster, *Crassostrea gigas* (Thunberg). *Aquaculture*, 190(1–2), 183–199. [https://doi.org/10.1016/S0044-8486\(00\)00392-6](https://doi.org/10.1016/S0044-8486(00)00392-6)
- Lango-Reynoso, F., Devauchelle, N., Le Pennec, M., & Hatt, P.-J. (1999). Elements of reproductive strategy in oysters, *Crassostrea gigas*, from the “Rade de Brest”, France. *Invertebrate Reproduction & Development*, 36(1–3), 141–144.
- Lango Reynoso, F. (1999). *Détermination de la sexualité chez l’huître Crassostrea gigas (Thunberg, 1793).* Université de Bretagne Occidentale.
- Lannan, J. E. (1980). Broodstock management of *Crassostrea gigas*. IV. Inbreeding and larval survival. *Aquaculture*, 21(4), 353–356. [https://doi.org/10.1016/0044-8486\(80\)90070-8](https://doi.org/10.1016/0044-8486(80)90070-8)
- Lazar, N. (2015). *Training Course Curriculum on Fish Stock Assessment Methods Summary. The USAID Sustainable Fisheries Management Project (SFMP).* Narragansett, RI: Coastal Resources Center, Graduate School of Oceanography, University of Rhode Island.
- Lebata-Ramos, M. J. H. L., Dionela, C. S., Novilla, S. R. M., Sibonga, R. C., Solis, E. F. D., & Mediavilla, J. P. (2021). Growth and survival of oyster *Crassostrea iredalei* (Faustino, 1932): A comparison of wild and hatchery-bred spat in grow-out culture. *Aquaculture*, 534(October 2020), 736310. <https://doi.org/10.1016/j.aquaculture.2020.736310>

- Lee, J.-S., Lee, Y.-G., Kang, S.-W., Park, J.-S., Lee, D.-G., Jeon, M., & Ju, S.-M. (2010). Intersexuality of *Crassostrea gigas* and *Ruditapes philippinarum* in Southern coastal waters of Korea. *Environmental Health and Toxicology*, 25(4), 287–294.
- Lenz, T. D. M., & Boehs, G. (2011). Reproductive cycle of the mangrove oyster *Crassostrea rhizophorae* (Bivalvia: Ostreidae) in Camamu Bay, Bahia, Brasil. *Revista de Biología Tropical*, 59(1), 137–149.
- Li, A., Dai, H., Guo, X., Zhang, Z., Zhang, K., Wang, C., Wang, X., Wang, W., Chen, H., Li, X., Zheng, H., Li, L., & Zhang, G. (2021). Genome of the estuarine oyster provides insights into climate impact and adaptive plasticity. *Communications Biology*, 4(1), 1–12. <https://doi.org/10.1038/s42003-021-02823-6>
- Li, Q., Liu, W., Shirasu, K., Chen, W., & Jiang, S. (2006). Reproductive cycle and biochemical composition of the Zhe oyster *Crassostrea plicatula* Gmelin in an eastern coastal bay of China. *Aquaculture*, 261(2), 752–759. <https://doi.org/10.1016/j.aquaculture.2006.08.023>
- Lodeiros, C., Valentich-Scott, P., Chávez-Villalba, J., Mazón-Suástegui, J. M., & Grijalva-Chon, J. M. (2020). Tropical and Subtropical Ostreidae of the American Pacific: Taxonomy, Biology, Ecology, and Genetics. *Journal of Shellfish Research*, 39(2), 181–206. <https://doi.org/10.2983/035.039.0202>
- Loor, A., & Sonnenholzner, S. (2016). Reproductive cycle of the rock oyster, *Striostrea prismatica* (Gray, 1825) from two locations on the southern coast of Ecuador. *Aquaculture Research*, 47(5), 1432–1442. <https://doi.org/10.1111/are.12601>
- Lovatelli, A., Helm, M. M., & Bourne, N. (2004). Hatchery operation : broodstock conditioning , spawning and fertilization. In *Reading* (pp. 1–23).
- Ludi, P. A. (2011). Review: Spawning Induction in Bivalve. *Jurnal Penelitian Sains*, 14(D), 14207.
- Macpherson, R. (1858). Narrative of a trip to Dok in the Muar Territory by the Hon. Captain R. Macpherson, Madras Artillery, Resident Councillor at Malacca. *The Journal of the Indian Archipelago and Eastern Asia* (Logan J.R. Ed.), II, 295–300.
- Madrones-Ladja, J. A. (2002). Salinity effect on the embryonic development, larval growth and survival at metamorphosis of *Placuna placenta* Linnaeus (1758). *Aquaculture*, 214(1–4), 411–418.
- Mamat, N., Rasam, A. R. A., Adnan, N. A., & Abdullah, I. C. (2014). GIS-based multi-criteria decision making system for determining potential site of oyster aquaculture in Terengganu. *Proceedings - 2014 IEEE 10th International Colloquium on Signal Processing and Its Applications, CSPA 2014*, 71–76. <https://doi.org/10.1109/CSPA.2014.6805723>

- Mancera, E., & Mendo, J. (1996). Population dynamics of the oyster *Crassostrea rhizophorae* from the Cienaga Grande de Santa Marta, Colombia. *Fisheries Research*, 26(1–2), 139–148.
- Mann, R., Southworth, M., Carnegie, R. B., & Crockett, R. K. (2014). Temporal variation in fecundity and spawning in the eastern oyster, *Crassostrea virginica*, in the Piankatank River, Virginia. *Journal of Shellfish Research*, 33(1), 167–176. <https://doi.org/10.2983/035.033.0116>
- Marescaux, J., Falisse, E., Lorquet, J., Van Doninck, K., Beisel, J. N., & Descy, J. P. (2016). Assessing filtration rates of exotic bivalves: dependence on algae concentration and seasonal factors. *Hydrobiologia*, 777(1), 67–78. <https://doi.org/10.1007/s10750-016-2764-0>
- Mason, J. (1958). The breeding of the scallop, *Pecten maximus* (L.) in Manx waters. *Journal of the Marine Biological Association of the United Kingdom*, 37(3), 653–671.
- Mazón-Suástegui, J. M., Ruíz-García, M. C., Chávez-Villalba, J., Rodríguez-Jaramillo, C., & Saucedo, P. E. (2011). Analysis of growth and first reproduction of hatchery-reared juvenile Cortez oyster (*Crassostrea corteziensis*) in northwestern Mexico: proposal of a minimal fishing size. *Aquaculture Research*, 42(10), 1558–1568.
- McFarland, K., Donaghy, L., & Volety, A. K. (2013). Effect of acute salinity changes on hemolymph osmolality and clearance rate of the non-native mussel, *Perna viridis*, and the native oyster, *Crassostrea virginica*, in Southwest Florida. *Aquatic Invasions*, 8(3), 299–310. <https://doi.org/10.3391/ai.2013.8.3.06>
- Mohammed, S. Z., & Yassien, M. H. (2003). Population parameters of the pearl oyster *Pinctada radiata* (Leach) in Qatari waters, Arabian Gulf. *Doğa, Türk Zooloji Dergisi / Doğa, Turkish Journal of Zoology*, 27(4), 339–343.
- Mohd Saleh, M. ., Devakie, M. N., & Hadzley, H. (2010). Effect of Environmental Conditions on the Spat Recruitment Pattern of Green Mussel (*Perna viridis* L.) at Sebatu, Melaka, Peninsular Malaysia. *Malaysian Fisheries Journal*, 9(December), 71–85.
- Mohd Yatim, H. N. (1993). *A Guide to Oyster Culture in Malaysia*. Food and Agriculture organization of United Nation, Bay of Bengal Programme, Madras, India.
- Mok, T. K. (1973). Studies on spawning and setting of the oyster in relation to seasonal environmental changes in Deep Bay, Hong Kong. *Hong Kong Fish Buletin*, 3, 89–101.
- Morales-Alamo, R., & Mann, R. (1989). Anatomical features in histological sections of *Crassostrea virginica* (Gmelin, 1791) as an aid in measurements of gonad area for reproductive assessment. *Journal of Shellfish Research*, 8, 71–82.

- Moreau, J., & Cuende, F. X. (1991). On improving the resolution of the recruitment patterns of fishes. *Fishbyte*, 9(1), 45–46.
- Muar climate: *Climate-Data.org*. (n.d.). Retrieved February 3, 2022, from <https://en.climate-data.org/asia/malaysia/johor/muar-25944/>
- Muranaka, M. S., & Lannan, J. E. (1984). Broodstock management of *Crassostrea gigas*: environmental influences on broodstock conditioning. *Aquaculture*, 39(1–4), 217–228.
- Murua, H., & Saborido-Rey, F. (2003). Female reproductive strategies of marine fish species of the North Atlantic. *Journal of Northwest Atlantic Fisheries Science*, 33, 23–31.
- Nair, U. T., & Nair, B. N. (1986). Relation between weight and linear measurement of shell in *C. madrasensis* (Preston). *Fishery Technology*, 23(2), 120–124.
- Nascimento, I. A., & Lunetta, J. E. (1978). Ciclo sexual da ostra do mangue e sua importância para o cultivo. *Boletim Fisiologia Animal. São Paulo. Universidade de São Paulo*, 2, 63–98.
- Nascimento, I. A., & Pereira, S. A. (1980). Changes in the condition index for mangrove oysters (*Crassostrea rhizophorae*) from Todos os Santos Bay, Salvador, Brazil. *Aquaculture*, 20(1), 9–15.
- Nayar, K. N., & Rao, K. S. (1985). Molluscan fisheries of India. *Marine Fisheries Information Service, Technical and Extension Series*, 61, 1–7.
- Needler, A. B. (1932). *Sex reversal in Ostrea virginica* (Vol. 22, Issue 22, pp. 55–71). Duke University.
- Newman, S. J. (2002). Growth, age estimation of longevity and mortality in the mose (Indian ocean form), from continental western Australia. *Asian Fisheries Science*, 15(3), 283–294.
- Ng, F. O. (1979). Experimental culture of a flat oyster (*Ostrea folium* L.) in Malaysian waters. *Malaysian Agricultural Journal*.
- Ng, F. O. (1993). *Seed Production of the Oyster Crassostrea iredalei, in Malaysia*. Department of Fisheries, Ministry of Agriculture, Malaysia.
- Nguyen, C. V., An, N. T., & The, H. Van. (2014). *Biological Characteristics of the Oyster Crassostrea lugubris in Central Coastal Vietnam*. *World Aquaculture*, 52-54.

- Niogee, S. R., Tonni, K. F., Barman, A. C., Tanu, M. B., Sku, S., & Uddin, M. J. (2019). Ovarian cycle of freshwater Pearl Mussel, *Lamellidens Marginalis* (Lamarck, 1819) collected from a culture pond in Bangladesh. *Asian Fisheries Science*, 32(3), 117–123. <https://doi.org/10.33997/j.afs.2019.32.3.004>
- O'Connor, W. A., & Lawler, N. F. (2005). Emersion tolerance of pearl oyster, *Pinctada imbricate* Roding, spat and juveniles. *Asian Fisheries Science*, 18(3/4), 217.
- Octavina, C., Yulianda, F., & Krisanti, M. (2014). *Struktur komunitas tiram daging di perairan estuaria Kuala Gigieng , Kabupaten Aceh Besar , Provinsi Aceh* Population structure of oysters in estuary area of Kuala Gigieng , Aceh Besar District , Aceh Province. 3(2), 108–117.
- Oliveira, L. F. S., Ferreira, M. A. P., Juen, L., Nunes, Z. M. P., Pantoja, J. C. D., Paixão, L. F. da, Lima, M. de N. B. de, & Rocha, R. M. da. (2018). Influence of the proximity to the ocean and seasonality on the growth performance of farmed mangrove oysters (*Crassostrea gasar*) in tropical environments. *Aquaculture*, 495(June), 661–667. <https://doi.org/10.1016/j.aquaculture.2018.06.049>
- Osei, I. K., Yankson, K., Obodai, E. A., & Okyere, I. (2021). Implications of overlooked seasonal growth dynamics in tropical fisheries assessment: A test case of an oyster (*Crassostrea tulipa*) fishery in the Densu Delta, Ghana. *Fisheries Research*, 244, 106118. <https://doi.org/10.1016/J.FISHRES.2021.106118>
- Paixão, L., Ferreira, M. A., Nunes, Z., Fonseca-Sizo, F., & Rocha, R. (2013). Effects of salinity and rainfall on the reproductive biology of the mangrove oyster (*Crassostrea gasar*): Implications for the collection of broodstock oysters. *Aquaculture*, 380–383, 6–12. <https://doi.org/10.1016/j.aquaculture.2012.11.019>
- Pakhmode, P. K., Mohite, S. A., & Gurjar, U. R. (2020). Length-weight relationship of rock oyster, *Saccostrea cucullata* along Aare-Ware rocky shore of Ratnagiri, Maharashtra, India. *Journal of Experimental Zoology, India*, 23(1), 253–257.
- Pantoja, J. C. D., Oliveira, L. F. S., Ferreira, M. A. P., Silva, B. R. M., Nunes, Z. M. P., Mendes, Y. A., de Oliveira, R. S., & da Rocha, R. M. (2020). Salinity and rainfall as inducers of cell proliferation and apoptosis in mangrove oyster *Crassostrea gasar* spermatogenesis. *Regional Studies in Marine Science*, 39, 101411. <https://doi.org/10.1016/j.rsma.2020.101411>
- Park, J. J., Kim, H., Kang, S. W., An, C. M., Lee, S.-H., Gye, M. C., & Lee, J. S. (2012). Sex Ratio and Sex Reversal in Two-year-old Class of Oyster, *Crassostrea gigas* (Bivalvia: Ostreidae). *Development & Reproduction*, 16(4), 385–388. <https://doi.org/10.12717/dr.2012.16.4.385>
- Park, J. J., Kwang, & Choi, K.-S. (2004). Application of enzyme-linked immunosorbent assay for studying of reproduction in the Manila clam *Ruditapes philippinarum* (Mollusca: Bivalvia): I. Quantifying eggs. *Aquaculture*, 241(1–4), 667–687.

- Parker, L. M., O'Connor, W. A., Byrne, M., Dove, M., Coleman, R. A., Pörtner, H. O., Scanes, E., Virtue, P., Gibbs, M., & Ross, P. M. (2018). Ocean acidification but not warming alters sex determination in the Sydney rock oyster, *Saccostrea glomerata*. *Proceedings of the Royal Society B: Biological Sciences*, 285(1872). <https://doi.org/10.1098/rspb.2017.2869>
- Parsons, T. R., Maita, Y., & Lalli, C. M. (1984). *A manual of chemical and biological methods for seawater analysis*. Oxford: Pergamon Press.
- Paterson, K. J., Schreider, M. J., & Zimmerman, K. D. (2003). Anthropogenic effects on seston quality and quantity and the growth and survival of Sydney rock oyster (*Saccostrea glomerata*) in two estuaries in NSW, Australia. *Aquaculture*, 221(1–4), 407–426.
- Pauly, D. (1979). *Gill size and temperature as governing factors in fish growth: a generalization of von Bertalanffy's growth formula*. Berichte Institut Meereskunde, Kiel, Germany.
- Pauly, D. (1980). On the interrelationships between natural mortality, growth parameters, and mean environmental temperature in 175 fish stocks. *ICES Journal of Marine Science*, 39(2), 175–192. <https://doi.org/10.1093/icesjms/39.2.175>
- Pauly, D. (1983). *Some simple methods for the assessment of tropical fish stocks* (Issue 234). Food & Agriculture Org.
- Pauly, D. (1984). *Length-converted catch curves: a powerful tool for fisheries research in the tropics (III: conclusion)*.
- Pauly, D. (1987). A review of the ELEFAN system for analysis of length-frequency data in fish and aquatic invertebrates. *ICLARM Conference Proceedings*, 13(232), 7–34.
- Pauly, D., & Caddy, J. F. (1985). *A modification of Bhattacharya's method for the analysis of mixtures of normal distribution*. *FAO Fishery Circular*, 781.
- Pauly, D., & David, N. (1981). ELEFAN I, a BASIC program for the objective extraction of growth parameters from length-frequency data. *Meeresforschung*, 28(4), 205–211.
- Pauly, D., & Munro, J. L. (1984). Once more on the comparison of growth in fish and invertebrates. *Fishbyte*, 2(1), 1–21.
- Pauly, D., Soriano-Bartz, M., Jarre-Teichmann, A., & Moreau, J. (1992). A new model accounting for seasonal cessation of growth in fishes [Atlantic salmon; Norwegian pout]. *Australian Journal of Marine and Freshwater Research (Australia)*, 43(5), 1151–1161.

- Pereira, O. M., Henriques, M. B., & Fagundes, L. (1998). Viability of *Crassostrea gigas* oyster farming off the coast of the southeastern and southern regions of Brazil. *Informacoes Economicas- Government of the State of Sao Paulo Institute of Agricultural Economics*, 28, 7–24.
- Peters, R. (1983). *The ecological implications of body size*. Cambridge University Press, New York. doi:10.1017/CBO9780511608551.
- Piedra, A. R., Díaz, E., & Melo, J. M. (2014). *Invertebrados marinos de importancia comercial en la costa Pacífica de Costa Rica*. Posada, Juan M.
- Posada, J. M., Piedra, A., Ross, E., Díaz, J. M., Sánchez, G. N., Guerra, Z., & De Leon, M. (2014). *Invertebrados marinos de importancia comercial en la costa Pacífica de Panamá*. San Jose, Costa Rica: Fundación Mar Viva. 120pp.
- Pouladi, M., Paighambari, S. Y., Millar, R. B., & Babanezhad, M. (2020). Length-weight relationships and condition factor of five marine fish species from Bushehr Province, Persian Gulf, Iran. *Thalassas: An International Journal of Marine Sciences*, 36(2), 457–461.
- Pourmozaffar, S., Tamadoni Jahromi, S., Rameshi, H., Sadeghi, A., Bagheri, T., Behzadi, S., Gozari, M., Zahedi, M. R., & Abrari Lazarjani, S. (2020). The role of salinity in physiological responses of bivalves. *Reviews in Aquaculture*, 12(3), 1548–1566. <https://doi.org/10.1111/raq.12397>
- Pouvreau, S., Gangnery, A., Tiapari, J., Lagarde, F., Garnier, M., & Bodoy, A. (2000). Gametogenic cycle and reproductive effort of the tropical blacklip pearl oyster, *Pinctada margaritifera* (Bivalvia: Pteriidae), cultivated in Takapoto atoll (French Polynesia). *Aquatic Living Resources*, 13(1), 37–48.
- Qin, Y., Zhang, Y., Ma, H., Wu, X., Xiao, S., Li, J., Mo, R., & Yu, Z. (2018). Comparison of the biochemical composition and nutritional quality between diploid and triploid Hong Kong oysters, *Crassostrea hongkongensis*. *Frontiers in Physiology*, 9, 1674.
- Quayle, D. B. (1980). *Tropical oysters: culture and methods*. IDRC, Ottawa, Canada.
- Quinn, T. J., & Deriso, R. B. (1999). *Quantitative fish dynamics*. Oxford university Press, New York.
- Rahim, M. A., Taha, M. S. M., & Nair, D. M. (2008). Performance of mass produced triploid vs tropical oyster *Crassostrea iredalei* faustino. *Proceedings of the 5th National Fisheries Symposium 2008*.
- Ramos, C. de O., Ferreira, J. F., & de Melo, C. M. R. (2013). Maturação da ostra nativa *Crassostrea gasar* submetida a diferentes dietas em laboratório. *Boletim Do Instituto de Pesca*, 39(2), 107–120.

- Rao, K. V. (1951). Observations on the probable effects of salinity on the spawning, development and setting of the Indian backwater oyster, *Ostrea madrasensis* Preston. *Proceedings of the Indian Academy of Sciences - Section B*, 33(5), 231–256. <https://doi.org/10.1007/BF03050000>
- Rebelo, M. F., Amaral, M. C., & Pfeiffer, W. C. (2005). Oyster condition index in *Crassostrea rhizophorae* (Guilding 1828) from a heavy-metal polluted coastal lagoon. *Brazilian Journal of Biology = Revista Brasileira de Biologia*, 65(2), 345–351. <https://doi.org/10.1590/S1519-69842005000200019>
- Ricker, W. E. (1975). Computation and interpretation of biological statistics of fish populations. *Bulletin of the Fisheries Research Board of Canada*, 191, 1–382.
- Rodríguez-Jaramillo, C., Hurtado, M. A., Romero-Vivas, E., Ramírez, J. L., Manzano, M., & Palacios, E. (2008). Gonadal development and histochemistry of the tropical oyster, *Crassostrea corteziensis* (Hertlein, 1951) during an annual reproductive cycle. *Journal of Shellfish Research*, 27(5), 1129–1141. <https://doi.org/10.2983/0730-8000-27.5.1129>
- Rosell, N. C. (1991). The slipper-shaped oyster (*Crassostrea iredalei*) in the Philippines. *Eswarine and Marine Bivalve Mollusk Culture*, 307–313.
- Royer, J., Seguineau, C., Park, K. Il, Pouvreau, S., Choi, K. S., & Costil, K. (2008). Gametogenetic cycle and reproductive effort assessed by two methods in 3 age classes of Pacific oysters, *Crassostrea gigas*, reared in Normandy. *Aquaculture*, 277(3–4), 313–320. <https://doi.org/10.1016/j.aquaculture.2008.02.033>
- Scherrer, P. (1984). Biostatistique, Morin, Montreal, Paris. SPSS Inc., 1999. Systat version 9. SPSS. Inc., USA.
- Seed, R., & Suchanek, T. H. (1992). Population and community ecology of *Mytilus*. *The Mussel Mytilus: Ecology, Physiology, Genetics and Culture*, 25, 87–170.
- Šegvić-Bubić, T., Grubišić, L., Zrnčić, S., Jozić, S., Žužul, I., Talijančić, I., Oraić, D., Relić, M., & Katavić, I. (2016). Range expansion of the non-native oyster *Crassostrea gigas* in the adriatic sea. *Acta Adriatica*, 57(2), 321–330.
- Shaw, B. L., & Battle, H. I. (1957). The Gross and Microscopic Anatomy of the Digestive Tract of the Oyster *Crassostrea virginica*(Gmelin). *Canadian Journal of Zoology*, 35(3), 325–347. <https://doi.org/10.1139/z57-026>
- Siddique, M. A., Khatun, M. A., Rahman, M. M., Ahmed, G. U., Moniruzzaman, M., & Jasim Uddin, M. (2020). Annual gametogenic cycle of the freshwater pearl mussel, *Lamellidens marginalis* (Lamarck, 1819) collected from a perennial lentic habitat of Bangladesh. *Molluscan Research*, 40(1), 36–43.

- Siddiqui, G., & Ahmed, M. (2002). Histological basis of population structure of three species of *Crassostrea* from Pakistan (Northern Arabian Sea). *Pakistan Journal of Zoology*, 34(2), 129–137.
- Sigwart, J. D., Wong, N. L. W. S., & Esa, Y. (2021). Global controversy in oyster systematics and a newly described species from SE Asia (Bivalvia: Ostreidae: Crassostreinae). *Marine Biodiversity*, 51(5), 1–16. <https://doi.org/10.1007/s12526-021-01203-x>
- Singh, Y. T. (2019). Biometrics, condition index and meat yield of edible rock oyster, *Saccostrea cucullata* (Born, 1778). *Journal of the Marine Biological Association of the United Kingdom*, 99(2), 385–392. <https://doi.org/10.1017/S0025315418000309>
- Smaal, A. C., Ferreira, J. G., Grant, J., Petersen, J. K., & Strand, Ø. (2019). *Goods and services of marine bivalves*. Springer Nature, Cham, Switzerland.
- Steele, S., & Mulcahy, M. F. (1999). Gametogenesis of the oyster *Crassostrea gigas* in southern Ireland. *Journal of the Marine Biological Association of the United Kingdom*, 79(4), 673–686.
- Suedel, B. C., Clarke, J. U., Wilkens, J., Lutz, C. H., & Clarke, D. G. (2014). The Effects of a Simulated Suspended Sediment Plume on Eastern Oyster (*Crassostrea virginica*) Survival, Growth, and Condition. *Estuaries and Coasts*, 38(2), 578–589. <https://doi.org/10.1007/s12237-014-9835-0>
- Sunila, I. (1981). Reproduction of *Mytilus edulis* L. (Bivalvia) in a brackish water area, the Gulf of Finland. In *Annales Zoologici Fennici* (Vol. 18, pp. 121–128).
- Sussarellu, R., Fabioux, C., Le Moullac, G., Fleury, E., & Moraga, D. (2010). Transcriptomic response of the Pacific oyster *Crassostrea gigas* to hypoxia. *Marine Genomics*, 3(3–4), 133–143.
- Suzana, M., Lutfi, A. M., Hadi, A. A., Devakie, M. N., & Azizah, M. N. S. (2011). Genetic variation in Malaysian oysters: Taxonomic ambiguities and evidence of biological invasion. *Biological Invasions*, 13(8), 1893–1900. <https://doi.org/10.1007/s10530-011-0009-8>
- Tan, S.-H. A., Chang, G.-O., & Peng, P. K. Y. and T. C. (2014). Oyster culture in Malaysia: Opportunities and challenges. *Journal of Science and Technology in the Tropics*, 10(2), 99–108.
- Tan, S.-H. A., Teh, C. P., Chang, G. O., & Yasin, Z. (2017). Tetraploid induction in tropical oysters, *Crassostrea belcheri* (Sowerby) and *Crassostrea iredalei* (Faustino). *Aquaculture Research*, 48(4), 1406–1412. <https://doi.org/10.1111/are.12976>

- Taylor, J. J., Southgate, P. C., & Rose, R. A. (2004). Effects of salinity on growth and survival of silver-lip pearl oyster, *Pinctada maxima*, spat. *Journal of Shellfish Research*, 23(2), 375–377.
- Teh, C. P., Zulfigar, Y., & Tan, S. H. (2012). Epinephrine and l-DOPA promote larval settlement and metamorphosis of the tropical oyster, *Crassostrea iredalei* (Faustino, 1932): An oyster hatchery perspective. *Aquaculture*, 338–341, 260–263. <https://doi.org/10.1016/j.aquaculture.2012.01.014>
- Thomas, Y., Pouvreau, S., Alunno-Bruscia, M., Barillé, L., Gohin, F., Bryère, P., & Gernez, P. (2016). Global change and climate-driven invasion of the Pacific oyster (*Crassostrea gigas*) along European coasts: a bioenergetics modelling approach. *Journal of Biogeography*, 43(3), 568–579.
- Uddin, M. J., Yang, H. S., Park, J. J., Kang, C. K., Kang, H. S., & Choi, K. S. (2012). Annual reproductive cycle and reproductive efforts of the Manila clam *Ruditapes philippinarum* in Incheon Bay off the west coast of Korea using a histology-ELISA combined assay. *Aquaculture*, 364–365, 25–32. <https://doi.org/10.1016/j.aquaculture.2012.07.006>
- Vaschenko, M. A., Hsieh, H. L., & Radashevsky, V. I. (2013). Gonadal state of the oyster *Crassostrea angulata* cultivated in Taiwan. *Journal of Shellfish Research*, 32(2), 471–482. <https://doi.org/10.2983/035.032.0227>
- Venema, S. C., Christensen, J. M., & Pauly, D. (1988). Training in tropical fish stock assessment: a narrative of experience. *FAO Fisheries Technical Paper*, 389, 1–15.
- Vidya, R., Jenni, B., Aloyicious, P. S., Venkatesan, V., Sajikumar, K. K., Jestin Joy, K. M., Sheela, P. P., & Mohamed, K. S. (2020). *Oyster Farming Techniques* (pp. 79–92). Molluscan Fisheries Division, CMFRI, India.
- Wahab, N. I. B. A. (2017). *Gonad maturation, larval growth and settlement of the slipper cupped oyster Crassostrea iredalei Faustino, 1932 (Mollusca, peleypoda: ostreidei)*. PhD Thesis. Universiti Putra Malaysia.
- Wilderbuer, T. K., & Zhang, C. I. (1999). Evaluation of the population dynamics and yield characteristics of Alaska plaice, *Pleuronectes quadrituberculatus*, in the eastern Bering Sea. *Fisheries Research*, 41(2), 183–200.
- Wong, N. L. W. S. (2013). *Distribution, larval abundance, spawning and early development of saddle oyster Placuna ehippium (Philipsson, 1788) from merambong shoal*. PhD Thesis. Universiti Putra Malaysia.
- Wong, N. L. W. S., & Arshad, A. (2011). A brief review on marine shelled mollusca (Gastropoda and Bivalvia) record in Malaysia. *Journal of Fisheries and Aquatic Science*, 6(7), 669.

- Xu, F., Guo, X., Li, L., & Zhang, G. (2011). Effects of salinity on larvae of the oysters *Crassostrea ariakensis*, *C. sikamea* and the hybrid cross. *Marine Biology Research*, 7(8), 796–803. <https://doi.org/10.1080/17451000.2011.569555>
- Yasuoka, N., & Yusa, Y. (2016). Effects of size and gregariousness on individual sex in a natural population of the Pacific oyster *Crassostrea gigas*. *Journal of Molluscan Studies*, 82(July), 485–491. <https://doi.org/10.1093/mollus/eyw020>
- Yasuoka, N., & Yusa, Y. (2017). Direct evidence of bi-directional sex change in natural populations of the oysters *Saccostrea kegaki* and *S. Mordax*. *Plankton and Benthos Research*, 12(1), 78–81. <https://doi.org/10.3800/pbr.12.78>
- Yue, C., Li, Q., & Yu, H. (2018). Gonad Transcriptome Analysis of the Pacific Oyster *Crassostrea gigas* Identifies Potential Genes Regulating the Sex Determination and Differentiation Process. *Marine Biotechnology*, 20(2), 206–219. <https://doi.org/10.1007/s10126-018-9798-4>
- Zabel, R. W., Harvey, C. J., Katz, S. L., Good, T. P., & Levin, P. S. (2003). Ecologically sustainable yield: Marine conservation requires a new ecosystem-based concept for fisheries management that looks beyond sustainable yield for individual fish species. *American Scientist*, 91(2), 150–157.