



UNIVERSITI PUTRA MALAYSIA

***GROWTH AND MATURATION PROCESSES OF PURPLE PASSION
FRUIT (*Passiflora edulis Sims*) THROUGH PHYSICAL AND
METABOLOMICS APPROACHES***

SHAHIDAH BINTI MD. NOR

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By

SHAHIDAH BINTI MD. NOR

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

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The purple passion (*Passiflora edulis SIMS*) fruit has enormous potential to be cultivated on a mass scale in Malaysia since it gains high demand in the global market. This study is aimed to understand the growth and maturation process of passion fruit through physical and metabolomics approaches for producing high quality fruit for fresh fruit and varieties of industries. Growth and development of passion fruit were tracked starting from 7 days after anthesis (DAA) until the fruit detached from the plant. The first study was conducted to establish the fruit optimal harvesting stage. Logistic model showed all physical traits of passion fruit fitted well with single sigmoid curve with high regression coefficient (R^2) around 0.984-0.992. The anatomical study showed that passion fruit have three distinct growth phases; stage 1 (S1) (0-7 DAA), stage 2 (S2) (14-28 DAA) and stage 3 (S3) (35-63 DAA). Purple passion fruit obtained maturity at 49 DAA and ripened at 56 DAA. The S3 phase was investigated intensively in studies 2 and 3 because it is a vital period for maturation and ripening. In the second and third studies, metabolomics approaches have been implemented in studying the dispersion of metabolites during maturation and ripening process. In the study 2, juice and seed were subjected to ^1H NMR analysis as both compartments play significant role in fruit eating quality. Around 30 and 32 metabolites were found in juice and seed, respectively. Result showed the primary metabolites dominating both compartments consist of sugar (glucose, sucrose, fructose), organic acid (tartaric acid, citric acid, malic acid) and amino acid (lysine, threonine, methionine, leucine) while secondary metabolites present as minor compounds (chlorogenic acid, epicatechin and phenylacetic acid). Segregating the metabolites using principal component analysis (PCA) and partial least square discriminant analysis (PLS-DA) have outlined how the metabolites in both compartments changed dynamically throughout the maturation and ripening process. Variable importance in the projection (VIP) has sorted 13 and 18 metabolites as highly influential metabolites in juice and seed. Glycolysis, TCA cycle, shikimate pathway, ethylene and polyamine were detected as important biochemical

pathways that were responsible for the fruit's maturation and ripening. The third study analysed juice, seed and peel since all fruit compartments have equal benefits to be exploited as industrial products. Result showed that each fruit compartments have different secondary metabolites contents correlated with their antioxidant activity. During ripening, β -carotene and chlorophyll pigments degraded while anthocyanin accumulated, resulting peel in a deep purple. The peel was abundant with different types of phenolic acid (chlorogenic acid, caffeic acid and its derivatives) and flavonoid components (isorientin, rutinoid and vitexin-2"-O-rhamnoside) possessed the highest antioxidant activity determined by FRAP (732.91- 1089.61 $\mu\text{mol TE. } 100 \text{ g}^{-1} \text{ FW}$), ABTS (62.34-70.10 $\mu\text{mol TE. g}^{-1} \text{ FW}$) and DPPH (52.362- 67.66 $\mu\text{mol TE. g}^{-1} \text{ FW}$) as compared to seed and juice. All passion fruit compartments should be completely exploited for fresh consumption and product manufacturing. Hence, herein a comprehensive view of passion fruit growth and development may benefit Malaysia's agriculture and processing industries.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PROSES PERTUMBUHAN DAN KEMATANGAN BUAH MARKISA UNGU
(*Passiflora edulis Sims*) MELALUI PENDEKATAN FIZIKAL DAN
METABOLOMIK**

Oleh

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Buah markisa ungu (*Passiflora edulis SIMS*) mempunyai potensi besar untuk diusahakan secara besar-besaran di Malaysia memandangkan ia mendapat permintaan tinggi di pasaran global. Kajian ini bertujuan untuk memahami proses pertumbuhan dan kematangan buah markisa melalui pendekatan fizikal dan metabolomik untuk menghasilkan buah berkualiti tinggi bagi industri buah segar dan pelbagai. Pertumbuhan dan perkembangan buah markisa dijejaki bermula dari 7 hari selepas antesis (DAA) sehingga buah tertanggal daripada pokok. Kajian pertama telah dijalankan untuk menentukan peringkat penuaian buah yang optimum. Model logistik menunjukkan semua ciri fizikal buah markisa dipasang sesuai dengan keluk sigmoid tunggal dengan pekali regresi tinggi (R^2) sekitar 0.984-0.992. Kajian anatomi menunjukkan bahawa buah markisa mempunyai tiga fasa pertumbuhan yang berbeza; peringkat 1 (S1) (0-7 DAA), peringkat 2 (S2) (14-28 DAA) dan peringkat 3 (S3) (35-63 DAA). Buah markisa ungu memperoleh kematangan pada 49 DAA dan masak pada 56 DAA. Fasa S3 telah disiasat secara intesif dalam kajian 2 dan 3 kerana ia merupakan tempoh penting untuk proses matang dan masak. Dalam kajian kedua dan ketiga, kaedah metabolomik telah dilaksanakan bagi mengkaji penyebaran metabolit semasa proses pematangan dan masak. Dalam kajian 2, jus dan biji benih tertakluk kepada analisis ^1H NMR kerana kedua-dua bahagian memainkan peranan penting dalam penilaian kualiti buah. Sekitar 30 dan 32 metabolit dikenalpasti dalam jus dan biji, masing-masing. Keputusan menunjukkan metabolit primer yang mendominasi kedua-dua bahagian terdiri daripada gula (glukosa, sukrosa, fruktosa), asid organik (asid tartarik, asid sitrik, asid malik) dan asid amino (lisin, treonin, metionin, leucine) manakala metabolit sekunder hadir sebagai sebatian kecil. (asid klorogenik, epicatechin dan asid phenylacetik). Pengasingan metabolit menggunakan analisis komponen utama (PCA) dan analisis diskriminasi persegi terkecil separa (PLS-DA), telah mendapati metabolit dalam kedua-dua bahagian berubah secara dinamik sepanjang proses pematangan dan masak. Kepentingan berubah dalam unjuran (VIP) telah menyusun 13 dan 18 metabolit sebagai metabolit yang sangat berpengaruh dalam jus dan biji. Glikolisis, kitaran TCA, laluan shikimat, etilena dan poliamina telah dikesan sebagai jaluran biokimia penting yang bertanggungjawab untuk

kematangan dan masak buah. Kajian ketiga menganalisis jus, biji dan kulit kerana semua bahagian buah mempunyai manfaat yang sama untuk diproses sebagai produk industri. Keputusan menunjukkan setiap bahagian buah mempunyai kandungan metabolit sekunder yang berbeza yang berkaitan dengan aktiviti antioksidannya. Semasa masak, pigmen β -karotena dan klorofil terdegradasi manakala antosianin terkumpul, menghasilkan kulit berwarna ungu tua. Kulitnya kaya dengan pelbagai jenis asid fenolik (asid klorogenik, asid caffeic dan derivatifnya) dan komponen flavonoid (isorientin, rutinoid dan vitexin-2"-O-rhamnoside) mempunyai aktiviti antioksidan tertinggi yang ditentukan oleh FRAP (732.91- 1089.61 $\mu\text{mol TE. } 100 \text{ g}^{-1} \text{ FW}$), ABTS (62.34-70.10 $\mu\text{mol TE. g}^{-1} \text{ FW}$) dan DPPH (52.362- 67.66 $\mu\text{mol TE. g}^{-1} \text{ FW}$) berbanding dengan biji dan jus. Semua bahagian markisa harus dieksploitasi sepenuhnya untuk kegunaan segar dan pembuatan produk. Oleh itu, di sini pandangan menyeluruh tentang pertumbuhan dan pembangunan buah markisa yang boleh memberi manfaat kepada industri pertanian dan pemprosesan di Malaysia.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

$^1\text{H NMR}$	Proton nuclear magnetic resonance
1-MCP	1-methylcyclopropene
ABTS ^{•+}	2,2-azinobis (3-ethyl-benzothiazoline-6-sulfonic acid)
ACC	1-aminocyclopropane-1-carboxylic acid
ACO	ACC oxidase
ACS	ACC synthase
AdoMet	S-adenosyl-l-methionine
ANOVA	Analysis of variance
ANT	Anthocyanin
BC	β -Carotene
CA	Chlorogenic acid
CE	Cyanidin-3-glucoside equivalent
CLA	Chlorophyll A
CLB	Chlorophyll B
CME	Cellulose mixed ether-membrane
CRD	Completely randomised design
D ₂ O	Deuterium oxide
d ₄ -CD ₃ OD	Deuterated methanol
DAA	Days after anthesis
DAD	Diode array detector
DPPH	1,1-Diphenyl-2-picrylhydrazyl
DW	Dry weight
ESI	Electrospray ionisation interface

FeCl ₃ . 6H ₂ O	Ferric chloride hexahydrate
FRAP	Ferric reducing antioxidant power
FW	Fresh weight
GAE	Gallic acid equivalents
GC	Gas chromatography
HPLC	High performance liquid chromatography
HPO ₃	Metaphosphoric acid
KEGG	Kyoto encyclopaedia of genes and genome
KH ₂ PO ₄	Potassium dihydrogen phosphate
LSD	Least significant difference
M6PR	NADPH-mannose-6-phosphate reductase
MeOH	Methanol
MS	Mass spectrometry
MTA	5-Methylthioadenosine
MTBE	Methyl tert-butyl ether
Na ₂ CO ₃	Sodium carbonate
NaOH	Sodium hydroxide
NI	Neutral invertase
NPK	Nitrogen, phosphorous and potassium
PCA	Principal component analysis
PE	Pectinesterase
PG	Polygalacturonase
PL	Pectate lyase
PLS-DA	Partial least square discriminant analysis

ppm	Part per million
PRESAT	Pre-saturation
PRPP	Phosphoribosyl diphosphate
Q2	Cross-validate predictive ability
QE	Quercetin equivalents
R2	Total variance
RQ	Relative quantity
SAM	S-Adenosylmethionine
SPP	Sucrose-6-phosphate phosphatase
SPS	Sucrose-6-phosphate synthase
SSC	Soluble solids concentration
T6P	Trehalose-6-phosphatase
TA	Titrateable acidity
TCA cycle	Tricarboxylic acid cycle
TE	Trolox equivalent
TFC	Total flavonoid content
TPC	Total phenolic content
TPTZ	2,4,6-Tri-(2-pyridyl)-s-triozine
Trolox	6-Hydroxy2,5,7,8-tetramethyl-chroman-2-carboxylic acid
TSP	3-(Trimethylsilyl) propionic-2, 2, 3, 3-d ₄ acid sodium salt
USDA	United State Department of Agriculture
UV	Ultraviolet
V2R	Vitexin-2"-O-rhamnoside
VIP	Variable importance in the projection

vol

Volume

β-GAL

β-Galactosidase



CHAPTER 1

INTRODUCTION

1.1 Background of study

Fresh fruit is highly perishable, and the post-harvest loss occur naturally as soon after the fruit harvested from the parent plant. It is estimated about 18-28% of loss occur as soon after the harvesting stage (Md Nor et al. 2020). The loss continually develops during transportation, marketing, and trading levels, negatively impacts fruit farmers, consumer prices, nutritional quality produces, and income of the exporting countries (Md Nor et al. 2020). Due to this, Malaysia suffers around 17-35% of income loss, which mainly attributable to deteriorated fruit quality during local and global trading (Mahmud, 2017). The key strategy in producing high quality fruit is by determining the optimal harvesting. The Right harvesting stage is critical to make sure fruit achieves its optimal eating quality during harvesting and can maintain a longer post-harvest life (Mohammad and Ding, 2019; Tee et al. 2012).

Purple passion fruit (*Passiflora edulis* Sims.) is the focus of the current research. This fruit is classified as minor tropical fruit which grabbed increasing demand globally. The enticing flavour makes the fruit suitable for dessert or processed into a variety of confectionary products such as fresh juice, cordial, jams, and concentrates (Pham et al., 2019; Natnicha et al., 2019). In recent years, passion fruit has gained popularity since its applications have expanded beyond the fresh fruit and processed food industries. The peel and seed of the fruit are processed for nutraceutical, pharmaceutical, food ingredients, cosmetics, and skincare (Ramli et al., 2020; Oliveira et al., 2016; López-Vargas et al., 2013). The increasing demand for passion fruit in the world market should be viewed as an opportunity to strengthen the country's revenue. The passion fruit has immense potentials to be cultivated commercially in Malaysia since the fruit can be processed to multifunction products (The Insight Partners, 2020). In preparing Malaysia for passion fruit commercial cultivation, it is essential to produce high-quality fruit, whether fresh or processed.

Fruit maturity has profound effects on the final fruit quality. Fruit such as banana (Tee et al., 2012), mango (Syed et al., 2021), pomelo (Gupta et al., 2021) and pitaya (Magalhães et al., 2019) have dynamic physicochemical characteristics that affect determination of optimal harvesting stage. Non-optimal harvesting could compromise the fruit eating quality and post-harvest life (Ghosh et al., 2017). Commercial producer countries such as Colombia, Brazil, and India have conducted studies on the optimal harvesting stage for passion fruit. Nevertheless, most of the studies were only focus on physical and psychochemical aspects. None of the studies have ever integrated the concrete analysis from wide aspects, for example by anatomical perspective of passion fruit. Integrating anatomical research with physical and physicochemical features will allow for precise maturation and ripening stage determination, while also contribute to

fundamental knowledge in understanding the cellular process of growth and development.

Passion fruit has three main compartments: juice, seed and peel. In fresh fruit and food industries, the juice and seed are valuable compartments as they are graded by eating quality. Metabolomics is an efficient bioanalytical strategy that enables the visualisation of an organism's metabolites during a biological process by utilising comprehensive and high-throughput analysis (Lin et al., 2021). Metabolomics provides a holistic view of the physiological, structural, physical, and biochemical changes that occur in fruit during maturation and ripening. Primary metabolites such as sugars and organic acids are important determinants of fruit's eating quality (Tijero et al., 2021), whereas secondary metabolites such as phenolic and flavonoid compounds determining phytonutrients of a fruit (Del-Castillo-Alonso et al., 2021). Studying the fruit quality aspects analysing the juice and seed metabolome will understand the key metabolites that responsible for fruit maturation and ripening.

Numerous studies demonstrate that passion fruit have potential as useful source of natural antioxidants in neutralising or inhibiting free radicals activity that benefit to human health (He et al., 2020). Due to this, extensive research is being carried out to isolate and identify functional bioactive compounds that responsible for the antioxidant properties (Guimarães et al., 2020; Lin et al., 2021). All of the passion fruit compartment including peel may possess the antioxidant activity that can exploited beyond the fresh fruit and processing industries. Quantifying the primary and secondary metabolites will understand dynamic changes of antioxidant capacity during passion fruit growth and development. In addition, mechanism in pigment accumulation in passion fruit peel that responsible for colour remain unknown (Gioia et al., 2020). The study of the fruit metabolome and its correlation with antioxidant activity will help to understand the phenotypic development and functionality of each fruit compartment so that it can be fully exploited in industries.

1.2 Aims and objectives

The study aims to understand passion fruit's growth and maturation process through the physical and metabolomics approaches. Passion fruit is intended to be produced in optimal quality, with all fruit compartments being fully utilised in a variety of industries. The comprehensive study during growth and development will provide valuable knowledge in fruit's physiology as the fundamental knowledge in horticulture study.

The aims were achieved through the following set of objectives:

1. Characterise the growth and development and determine the optimal harvesting stage for passion fruit cultivated in Malaysia.
2. Identify and quantify metabolites that are responsible in fruit maturation and ripening.

3. Understand how targeting metabolites affect the fruit's phenotype and its relationship with antioxidant activity

1.3 Scope of thesis

This thesis is outlined in 6 chapters:

Chapter 1 provides an overview of the research background and aims of the study. The passion fruit quality, maturity of harvesting, and need for metabolomics study are briefly discussed.

Chapter 2 focuses on comprehensive review on literature related to research. The horticulture background about passion fruit, demand in global and fundamental knowledge in passion fruit development are highlighted. Study on metabolomics that discussing the differences between primary and secondary metabolites in plant metabolism are also incorporated.

Chapter 3 reports the optimal harvesting of passion fruit. The fruit's optimal harvesting stage was scrutinised from the data of physical, anatomical and physicochemical characteristics. Cellular study using scanning electron microscope was performed to distinguish the growth phases. Physically, fruit were rated for its firmness and colour, while physiology was rated according to respiration and ethylene production. The eating quality was rated according to biochemical analysis that commercially applied in post-harvest study. Fundamental knowledge on growth profile of passion fruit was established from this study, became the primary reference for the subsequent chapters.

Chapter 4 apply the nuclear magnetic resonance (NMR) spectroscopy and chemometric methods in achieving the metabolic fingerprint during passion fruit's maturation and ripening stage. Juice and seed were analysed as these compartments serve as major entities in determining the fruit eating quality. Through NMR analysis, majority of metabolites detected belong to primary metabolites while secondary metabolites detected as minor compound. The model of principle component analysis (PCA) and partial least square discriminant analysis (PLS-DA) were established to understand the segregation of metabolites in juice and seed that response toward different harvesting stage. The highly influential metabolites that responsible for maturation and ripening in juice and seed were identified using variance importance projection (VIP) and further quantified using relative quantification. These metabolites were denoted as key metabolites that influencing the maturation and ripening of passion fruit.

Chapter 5 provides extension study to chapter 4. Peel compartment was included besides the juice and seed. It because the peel can be further exploited as industrial products even it does not contribute to fruit eating quality. Calorimetric analysis was conducted to preliminary quantify the secondary metabolites content and asses the antioxidant activity. The specific secondary metabolites were identified and quantified using the high-performance liquid chromatography (HPLC). Relationship between

antioxidant content and its activity was established using Pearson's correlation. For seed and juice, a partial Least Square (PLS) regression analysis was performed to establish a correlation between antioxidant activity and the entire fruit metabolome, taking into account previous ^1H NMR data with secondary metabolites that were quantified by calorimetric and HPLC analysis.

Chapter 6 summarised the general conclusion obtain from the whole studies conducted in this thesis. Limitation of study and future perspective of result are embedded at the end of the conclusion.



REFERENCES

- Adaskaveg, J., Silva, C., Huang, P. and Blanco-Ulate, B. 2021. Single and double mutations in tomato ripening transcription factors have distinct effects on fruit development and quality traits. *Frontiers in Plant Science* 12: 647035.
- Aizat, W., Dias, D., Stangoulis, J.C., Able, J., Roessner, U. and Able, A. 2014. Metabolomics of capsicum ripening reveals modification of the ethylene related-pathway and carbon metabolism. *Postharvest Biology and Technology* 89: 19–31.
- Aizat, W.M., Able, J.A., Stangoulis, J.C. and Able, A. 2013. Characterisation of ethylene pathway components in non-climacteric capsicum. *BMC Plant Biology* 13(1): 1–14.
- Akamine, K. and Girolami, G. 1959. Pollination and fruit set in the yellow passion fruit. Hawaii Agricultural Experiment Station, University of Hawaii. 1959.
- Albrecht, G. and Mustrup, A. 2003. Localization of sucrose synthase in wheat roots: Increased in situ activity of sucrose synthase correlates with cell wall thickening by cellulose deposition under hypoxia. *Planta* 217(2): 252-260.
- Ali, M., Yusof, Y., Chin, N. and Ibrahim, M. 2016. Effect of different drying treatments on colour quality and ascorbic acid concentration of guava fruit. *International Food Research Journal* 23: 155–161.
- Al-Mekhlafi, N., Mediani, A., Ismail, N., Abas, F., Dymerski, T., Lubinska-Szczygeł, M., Vearasilp, S. and Gorinstein, S. 2021. Metabolomic and antioxidant properties of different varieties and origins of dragon fruit. *Microchemical Journal* 160: 105687.
- Alsmairat, N., Engelgau, P. and Beaudry, R. 2018. Changes in free amino acid content in the flesh and peel of ‘Cavendish’ banana fruit as related to branched-chain ester production, ripening, and senescence. *Journal of the American Society for Horticultural Science* 143(5): 370–380.
- Al-Taweel, A., Perveen, S., Ibrahim, S., Orfali, R., Aati, H., Nasser, E., Alghanem, B. and Shaibah, H. 2021. New flavane gallates from the aerial part of an African/Arabian medicinal plant *Plicosepalus curviflorus* by LC–MS and NMR based molecular characterization. *Journal of King Saud University-Science* 33: 101289.
- Altendorf, S. 2018. Minor tropical fruits. Retrieved 23 Jun 2021 from http://www.fao.org/fileadmin/templates/est/COMM_MARKETS_MONITORING/Tropical_Fruits/Documents/Minor_Tropical_Fruits_FoodOutlook_1_2018.pdf. <http://www.fao.org/3/CA0239EN/ca0239en.pdf>.

- Aluko, O., Li, C., Wang, Q. and Liu, H. 2021. Sucrose utilization for improved crop yields: A review article. *International Journal of Molecular Science* 22: 1-29.
- Angami, T., Kalita, H., Baruah, S. and Ronya, T. 2017. Dynamics of physico-biochemical changes in passion fruit varieties across maturity. *International Journal of Chemical Studies Chemical Studies* 5(6): 162–165.
- Aprèa, E., Charles, M., Endrizzi, I., Corollaro, M. and Betta, E. 2017. Sweet taste in apple: The role of sorbitol, individual sugars, organic acids and volatile compounds. *Scientific Report* 7: 44950.
- Araki, T., Mitano, M. and Eguchi, H. 2000. Dynamics of fruit growth and photoassimilate translocation in tomato plant (*Lycopersicon esculentum* Mill.) under controlled environment. *Acta Horticulturae* 534: 85-92.
- Argenta, L., Vidal, C., Brancher, T., Betinelli, K. and Nesi, C. 2020. Comparison of fruit maturation and quality of ‘Gala’ apple strains at harvest and after storage. *Revista Brasileira de Fruticultura* 43: e-285.
- Arias, F., Appelman, T., Echeverri, C., Arboleda, C. and Giraldo, L.F. 2020. Analysis of the world market of purple passion fruit. Retrieved 19 August 2021 from https://www.researchgate.net/publication/349849475_Analysis_of_the_world_market_of_purple_passion_fruit.
- Avalos-Soriano, A., De La Cruz-Cordero, R., Rosado, J. and Garcia-Gasca, T. 2016. 4-Hydroxyisoleucine from fenugreek (*Trigonella foenum-graecum*): Effects on insulin resistance associated with obesity. *Molecules* 21(11): 1–12.
- Awatif, S. and Alaeldin, A. 2013. Metabolic processes during seed germination. In *Advance in Seed Biology*. pp.137–144. United Kingdom: IntechOpen
- Awin, T., Mediani, A., Maulidiani, Leong, S., Muhd Faudzi, S., Shaari, K. and Abas, F. 2019. Phytochemical and bioactivity alterations of *Curcuma* species harvested at different growth stages by NMR-based metabolomics. *Journal of Food Composition and Analysis* 77: 66–76.
- Ayala-Silva, T., Schnell, R., Meerow, A., Winterstein, M., Cervantes, C. and Brown, S. 2005. Determination of colour and fruit traits of Halfsib families of mango (*Mangifera Indica* L.). *Proceedings of the Florida State Horticultural Society* 118: 253–257.
- Azizah, O. 2011. Fruit industry in Malaysia. In *Nutritious, Colourful, Yet Fragile Gifts of Nature*, ed. A.R. Shariah and M.F. Ahmad. pp. 17-20. Malaysia: Universiti Putra Malaysia Press.
- Bacher, A. and Eisenreich, W. 2010. Metabolic studies using the retrobiosynthesis concept—theory, technology, and examples. In *Comprehensive Natural Products II: Chemistry and Biology*. pp. 675–694. United Kingdom: Academic Press.

- Barbosa, L., Mira, S., Gonzalez-Benito, M., Souza, M., Meletti, M. and Perez-Garcia, F. 2013. Seed germination, desiccation tolerance and cryopreservation of *Passiflora* species. *Seed and Science Technology* 41: 89–97.
- Barbosa, S., de Araujo, F., Neto, A.F., de Freitas, S., de Souza Araújo, J., de Oliveira Vilar, S., Brito Araújo, A. and Lima, M. 2021. Phytochemical compounds and antioxidant activity of the pulp of two Brazilian passion fruit species: *Passiflora cincinnata* Mast. and *Passiflora edulis* Sims. *International Journal of Fruit Science* 21(1): 255-269.
- Baroja-Fernández, E., Muñoz, F., Montero, M., Etxeberria, E., Sesma, M., Ovecka, M., Bahaji, A., Ezquer, I., Li, J., Prat, S. and Pozueta-Romero, J. 2009. Enhancing sucrose synthase activity in transgenic potato (*Solanum tuberosum* L.) tubers results in increased levels of starch, ADP glucose and UDP glucose and total yield. *Plant and Cell Physiology* 50(9): 1651–1662.
- Barry, C. and Giovannoni, J. 2007. Ethylene and fruit ripening. *Journal of Plant Growth Regulation* 26(2): 143–159.
- Bartoshuk, L. and Klee, H. 2013. Better fruits and vegetables through sensory analysis. *Current Biology* 23(9): 374–378.
- Batista-Silva, W., Nascimento, V., Medeiros, D., Nunes-Nesi, A., Ribeiro, D., Zsögön, A. and Araújo, W. 2018. Modifications in organic acid profiles during fruit development and ripening: Correlation or causation? *Frontiers in Plant Science* 871: 1–20.
- Battino, M., Giampieri, F., Cianciosi, D., Ansary, J., Chen, X., Zhang, D., Gil, E. and Forbes-Hernández, T. 2021. The roles of strawberry and honey phytochemicals on human health: A possible clue on the molecular mechanisms involved in the prevention of oxidative stress and inflammation. *Phytomedicine* 86: 153170.
- Battistus, A.G., Fuchs, F., Felipe, R., Sousa, B. De, Matos, M. De, Alexandre, J., Dranski, L., Rampim, L., Bulegon, L.G., Guimarães, F., Moranza, T.M. and Müller, M.A. 2014. Physiological maturity of seeds and colorimetry of yellow passion fruit (*Passiflora edulis* f. *flavicarpa* Degener). *African Journal of Agriculture Research* 9(40): 3018–3024.
- Baud, S., Dubreucq, B., Miquel, M., Rochat, C. and Lepiniec, L. 2008. Storage reserve accumulation in *Arabidopsis*: Metabolic and developmental control of seed filling. In *The Arabidopsis Book*. pp. e0113. USA: The American Society of Plant Biologists.
- Beauvoit, B., Belouah, I., Bertin, N., Cakpo, C., Colombié, S., Dai, Z., Gautier, H., Génard, M., Moing, A., Roch, L., Vercambre, G. and Gibon, Y. 2018. Putting primary metabolism into perspective to obtain better fruits. *Annals of Botany* 122(1): 1–21.

- Becker, P., Sganzerla, M., Carolina, A., Teixeira, M. and Carlos, R. 2015. Carotenoids, tocopherols and ascorbic acid content in yellow passion fruit (*Passiflora edulis*) grown under different cultivation systems. *LWT-Food Science and Technology* 64: 259–263.
- Bento-Silva, A., Koistinen, V., Mena, P., Bronze, M., Hanhineva, K., Sahlstrøm, S., Kitrytė, V., Moco, S. and Aura, A. 2019. Factors affecting intake, metabolism and health benefits of phenolic acids: Do we understand individual variability? *European Journal of Nutrition* 59(4): 1275–1293.
- Bernacci, L.C., Soares-scott, M.D., Tadeu, N., Junqueira, V., Ribeiro, I., Silva, D.A., Maria, L. and Meletti, M. 2008. *Passiflora edulis* Sims: The correct taxonomic way to cite the yellow passion fruit (and of others colours). *Revista Brasileira de Fruticultura* 30(2): 566–576.
- Bestwick, C., Scobbie, L., Milne, L., Duncan, G., Cantlay, L. and Russell, W. 2020. Fruit-based beverages contain a wide range of phytochemicals and intervention targets should account for the individual compounds present and their availability. *Foods* 9(7): 891.
- Biale, J., Young, R. and Olmstead, A. 1954. Fruit respiration and ethylene production. *Plant Physiology* 29(2): 168–174.
- Bochkov, D., Sysolyatin, S.V., Kalashnikov, A. and Surmacheva, I. 2012. Shikimic acid: Review of its analytical, isolation, and purification techniques from plant and microbial sources. *Journal of Chemical Biology* 5(1): 5–17.
- Borbély, P., Bajkán, S., Poór, P. and Tari, I. 2019. Exogenous 1-aminocyclopropane-1-carboxylic acid controls photosynthetic activity, accumulation of reactive oxygen or nitrogen species and macroelement content in tomato in long-term experiments. *Journal of Plant Growth Regulation* 38(3): 1110–1126.
- Borsellino, V., Kaliji, S. and Schimmenti, E. 2020. COVID-19 drives consumer behaviour and agro-food markets towards healthier and more sustainable patterns. *Sustainability* 12: 1-26.
- Botoran, O., Ionete, R., Miricioiu, M., Costinel, D., Radu, G. and Popescu, R. 2019. Amino acid profile of fruits as potential fingerprints of varietal origin. *Molecules* 24(24): 1–11.
- Bower, J. and Cutting, J. 1988. Avocado fruit development and ripening physiology. In *Horticultural Reviews*, ed. J. Janick. pp. 229–271. Portland: Timber Press.
- Brandizzi, F. 2019. Divide, expand, differentiate- new insights on plant organ growth through cytokinin signaling. *Plant Journal* 97(5): 803–804.
- Brizzolara, S., Manganaris, G., Fotopoulos, V., Watkins, C. and Tonutti, P. 2020. Primary metabolism in fresh fruits during storage. *Frontiers in Plant Science* 11: 1–16.

- Bron, I.U. and Jacomino, A.P. 2006. Ripening and quality of “Golden” papaya fruit harvested at different maturity stages. *Brazilian Journal of Plant Physiology* 18(3): 389–396.
- Bürstenbinder, K., Rzewuski, G., Wirtz, M., Hell, R. and Sauter, M. 2007. The role of methionine recycling for ethylene synthesis in Arabidopsis. *Plant Journal* 49(2): 238–249.
- Camargo, L., José, A., Campos, D., Carlos, R., Ru, S. and Cisneros-Zevallos, L. 2017. Days after anthesis and postharvest behavior define maturity, harvesting time and nutraceutical content of camu–camu fruit. *Scientia Horticulturae* 224: 37–47.
- Cao, Q., Teng, J., Wei, B., Huang, L. and Xia, N. 2021. Phenolic compounds, bioactivity, and bioaccessibility of ethanol extracts from passion fruit peel based on simulated gastrointestinal digestion. *Food Chemistry* 356: 129682.
- Castillo, N., Melgarejo, L. and Blair, M. 2020. Seed structural variability and germination capacity in *Passiflora edulis* Sims f. *edulis*. *Frontiers in Plant Science* 11: 1–10.
- Che, G., Gu, R., Zhao, J., Liu, X., Song, X., Zi, H., Cheng, Z., Shen, J., Wang, Z., Liu, R., Yan, L., Weng, Y. and Zhang, X. 2020. Gene regulatory network of carpel number variation in cucumber. *The Company of Biologist* 147: 184788.
- Chen, D., Melton, L., Zujovic, Z. and Harris, P. 2019. Developmental changes in collenchyma cell-wall polysaccharides in celery (*Apium graveolens* L.) petioles. *BMC Plant Biology* 19(1): 1–19.
- Cheng, L., Zhou, R., Reidel, J., Sharkey, T. and Dandekar, A. 2005. Antisense inhibition of sorbitol synthesis leads to up-regulation of starch synthesis without altering CO₂ assimilation in apple leaves. *Planta* 220: 767–776.
- Choo, W. 2018. Fruit pigment changes during ripening. In *Encyclopedia of Food Chemistry*, ed. L. Melton, F. Shahidi, P. Varelis. pp. 117-123. United Kingdom: Academic Press.
- Codex Alimentarius, 2014. Standard for passion fruit codex stan 316-2014. Retrieved 23 Jun 2021 from http://www.fao.org/fao-who-codexalimentarius/sh-proxy/en/?lnk=1&url=https%253A%252F%252Fworkspace.fao.org%252Fsites%252Fcodex%252FStandards%252FCXS%2B316-2014%252FCXS_316e_2014.pdf.
- Coombe, B. 1976. The development of fleshy fruits. *Annual Review Plant Physiology* 27: 507–528.
- Cordenunsi-Lysenko, B., Nascimento, J.R., Castro-Alves, V., Purgatto, E., Fabi, J. and Peroni-Okyta, F.H. 2019. The starch is (not) just another brick in the wall: The

- primary metabolism of sugars during banana ripening. *Frontiers in Plant Science* 10: 391.
- Corrêa, R.C.G., Peralta, R.M., Haminiuk, C.W.I., Maciel, G.M., Bracht, A. and Ferreira, I.C.F.R. 2016. The past decade findings related with nutritional composition, bioactive molecules and biotechnological applications of *Passiflora* spp. (passion fruit). *Trends in Food Science and Technology* 58: 79-95.
- Corrêa, S., Pinheiro, A.C., Siqueira, H.E., Carvalho, E., Nunes, C.A. and Boas, E.D. 2014. Prediction of the sensory acceptance of fruits by physical and physical-chemical parameters using multivariate models. *LWT-Food Science and Technology* 59: 666–672.
- CoSeteng, M., McLellan, M. and Downing, D. 1989. Influence of titratable acidity and pH on intensity of sourness of citric, malic, tartaric, lactic and acetic acids solutions and on the overall acceptability of imitation apple juice. *Canadian Institute of Food Science and Technology Journal* 22(1): 46–51.
- Costa, G., Gazola, A., Madóglío, F., Silvana, M., Reginatto, F., Castellanos, L., Ramos, F., Duque, C., Schenkel, E., Costa, G., Gazola, A. and Madóglío, F. 2013. Vitexin derivatives as chemical markers in the differentiation of the closely related species *Passiflora alata* Curtis and *Passiflora quadrangularis* Linn. *Journal of Liquid Chromatography & Related Technologies* 36: 1697–1707.
- Crang, R., Lyons-Sobaski, L. and Wise, R. 2018. Parenchyma, Collenchyma, and Sclerenchyma. In *Plant Anatomy*. pp. 181–182. Switzerland: Springer.
- Cruz-Tejada, D., Acosta-Rojas, D. and Stevenson, P. 2018. Are seeds able to germinate before fruit colour ripening? Evidence from six Neotropical bird-dispersed plant species. *Ecosphere* 9(6): e02174.
- da Silva, P. and Savian, T. 2019. Chanter model: Nonlinear modelling of the fruit growth of cocoa. *Ciencia Rural* 49(11): 1–7.
- da Silva, S.R., Ferreira, T.H., Giunco, A.J., and Argandoña, E. 2021. Nutritional potential and effect of the solvent on the extraction of secondary metabolites from pulp and bark of baru (*Dipteryx alata*). *Journal of Food Measurement and Characterization* 15: 3453 - 3460.
- Dam, S., Laursen, B., Ørnfelt, J., Jochimsen, B., Stærfeldt, H., Friis, C., Nielsen, K., Goffard, N., Besenbacher, S., Krusell, L., Sato, S., Tabata, S., Thøgersen, I., Enghild, J. and Stougaard, J. 2009. The proteome of seed development in the model legume lotus japonicus. *Plant Physiology* 149(3): 1325–1340.
- Das, M.R., Hossain, T., Baset Mia, M.A., Ahmed, J.U., Sirajul Karim, A.J.M. and Hossain, M.M. 2013c. Blooming pattern of passion fruit flower (*Passiflora edulis* Sims) under diversified flashes. *American Journal of Agricultural and Biological Science* 8(3): 173–181.

- Das, M.R., Hossain, T., Mia, M.A.B., Ahmed, J.U. Kariman, A.J.M.S. and Hossain, M. 2013a. Fruit setting behaviour of passion fruit. *American Journal of Plant Sciences* 4(5): 1066–1073.
- Das, M.R., Mia, A.B., Hossain, T., Ahmed, J.U. and Haq, M.Z. 2013b. Floral attributes of passion fruits (*Passiflora edulis* Sims.) at varied flashes. *Pakistan Journal of Agricultural Sciences* 50(1): 11–16.
- Dasenaki, M.E and Thomaidis, N. 2019. Quality and authenticity control of fruit juices- A review. *Molecules* 24(6): 1014.
- David, A.B. 2014. Chapter 11 - Fruit growth, ripening and post-harvest physiology. *Plants in Action* 1st Edition. Retrieved at 24 Jun 2021 from <https://www.asps.org.au/wp-content/uploads/Chapter-11-Fruit-growth-ripening-and-post-harvest-physiology.pdf>.
- de Carvalho, K., Petkowicz, C.L., Nagashima, G., Filho, J.C., Viera, L.G., Pereira, L.F. and Domingues, D. 2014. Homeologous genes involved in mannitol synthesis reveal unequal contributions in response to abiotic stress in *Coffea arabica*. *Molecular Genetic Genomics* 289(5): 951-963.
- de Poel, B. and Stareten D.V.D, V. 2014. 1-aminocyclopropane-1-carboxylic acid (ACC) in plants: More than just the precursor of ethylene! *Frontiers in Plant Science* 5: 1–11.
- de Toledo, N., de Camargo, A., Ramos, P., Button, D., Granato, D. and Canniatti-Brazaca, S. 2018. Potentials and pitfalls on the use of passion fruit by-products in drinkable yogurt: Physicochemical, technological, microbiological, and sensory aspects. *Beverages* 4(3):47.
- Del-Castillo-Alonso, M., Monforte, L., Tomás-Las-Heras, R., Ranieri, A., Castagna, A., Martínez-Abaigar, J. and Núñez-Olivera, E. 2021. Secondary metabolites and related genes in *Vitis vinifera* L. cv. Tempranillo grapes as influenced by UV radiation and berry development. *Physiologia Plantarum* 2021: 1–16.
- Dhar, M., Sharma, R., Koul, A. and Kaul, S. 2014. Development of fruit colour in *Solanaceae*: A story of two biosynthetic pathways. *Briefing in functional genomics* 14(3): 199-212.
- Dick, E., Adopo, A., Camara, B. and Moudioh, E. 2009. Influence of maturity stage of mango at harvest on its ripening quality. *Fruits* 64(1): 13–18.
- Ding, P. and Mashah, N. 2016. Growth, maturation and ripening of underutilized *Carissa congesta* fruit. *Fruits* 71(3): 171–176.
- Ding, P. and Syazwani, S., 2016. Physicochemical quality, antioxidant compounds and activity of MD-2 pineapple fruit at five ripening stages. *International Food Research Journal* 23(2): 549–555.

- Ding, P., Ahmad, S., Razak, A., Saari N. and Mohamed, M.T. 2007. Plastid ultrastructure, chlorophyll contents, and colour expression during ripening of cavendish banana (*Musa acuminata* 'Williams') at 18 °C and 27 °C. *New Zealand Journal of Crop and Horticultural Science* 35(2): 201–210.
- Diop, B. and Benkeblia, N. 2014. Correlation between colour, firmness, dry matter, sugars and maturity in 'East Indian' mangos (*Mangifera indica*). *Acta Horticulturae* 1047: 121–126.
- Dörken, V., Nimsch, H. and Jagel, A. 2017. Morphology, anatomy and morphogenesis of seed cones of *Cupressus vietnamensis* (Cupressaceae) and the taxonomic and systematic implications. *Flora: Morphology, Distribution, Functional Ecology of Plants* 230: 47–56.
- dos Reis, L.C., Facco, E.M., Salvador, M., Flôres, S. and de Oliveira Rios, A. 2018. Antioxidant potential and physicochemical characterization of yellow, purple and orange passion fruit. *Journal of Food Science and Technology* 55(7): 2679–2691.
- Drewnowski, A. and Burton-freeman, B. 2020. A new category-specific nutrient rich food (NRF9f.3) score adds flavonoids to assess nutrient density of fruit. *Food Function* 11: 123–130.
- Drewnowski, A. and Gomez-Carneros, C. 2000. Bitter taste, phytonutrients and the consumer: A review. *American Journal of Clinical Nutrition* 72: 1424–1435.
- Enamorado, H.E., Finger, F.L., Barros, R.S. and Puschmann, R. 1995. Development and ripening of yellow passion fruit. *Journal of Horticultural Science* 70(4): 573–576.
- Erb, M. and Kliebenstein, D. 2020. Plant secondary metabolites as defenses, regulators, and primary metabolites: the blurred functional trichotomy. *Plant Physiology* 184(1): 39–52.
- Eriksson, L., Johansson, E., Kettaneh-Wold, N., Trygg, J., Wikström, C., Wold, S., 2006. Multivariate analysis. In *Multi- and megavariate data analysis: part II: advanced applications and method extensions*. pp. 24–56. Sweden: Umetric Academy.
- Etienne, A., Génard, M., Lobit, P., Mbéguié-A-Mbéguié, D. and Bugaud, C. 2013. What controls fleshy fruit acidity? A review of malate and citrate accumulation in fruit cells. *Journal of Experimental Botany* 64(6): 1451–1469.
- Everson, R., Cockburn, W. and Gibbs, M. 1967. Sucrose as a product of photosynthesis in isolated spinach chloroplasts. *Plant physiology* 42: 840–844.
- Fait, A., Angelovici, R., Less, H., Ohad, I., Urbanczyk-Wochniak, E., Fernie, A. and Galili, G. 2006. Arabidopsis seed development and germination is associated with temporally distinct metabolic switches. *Plant Physiology* 142(3): 839–854.

- Fait, A., Nesi, A., Angelovici, R., Lehmann, M., Pham, P., Song, L., Haslam, R., Napier, J., Galili, G. and Fernie, A. 2011. Targeted enhancement of glutamate-to- γ -aminobutyrate conversion in Arabidopsis seeds affects carbon-nitrogen balance and storage reserves in a development-dependent manner. *Plant Physiology* 157(3): 1026–1042.
- Falchi, R., Bonghi, C., Drincovich, M., Famiani, F., Lara, M., Walker, R. and Vizzotto, G. 2020. Sugar metabolism in stone fruit: Source-sink relationships and environmental and agronomical effects. *Frontiers in Plant Science* 11: 1–14.
- Famiani, F., Battistelli, A. and Moscatello, S. 2015. The organic acids that are accumulated in the flesh of fruits: Occurrence, metabolism and factors affecting their contents-A review. *Revista Chapingo Serie Horticultura* 21(2): 97–128.
- Fathizadeh, Z., Aboonajmi, M. and Hassan-Beygi, S. 2021. Non-destructive methods for determining the firmness of apple fruit flesh. *Information Processing in Agriculture. In press.*
- Feng, S., Wang, Y., Yang, S., Xu, Y. and Chen, X. 2010. Anthocyanin biosynthesis in pears is regulated by a *R2R3-MYB* transcription factor *PyMYB10*. *Planta* 232: 245–255.
- Fernandes, A., Matias, G., Sales, D., Henrique, P., Sousa, M., Maia, G. and Figueiredo, R.W. 2011. Chemical and physicochemical characteristics changes during passion fruit juice processing. *Ciência e Tecnologia de Alimentos* 31(3): 747–751.
- Fernandes, T., Pereira, A. and Muniz, J. 2017. Double sigmoidal models describing the growth of coffee berries. *Ciência Rural* 47(8): 1–7.
- Fischer, G., Melgarejo, L. and Cutler, J. 2018. Pre-harvest factors that influence the quality of passion fruit: A review. *Agronomia Colombiana* 36(3): 217–226.
- Fisher, G. and Miranda, D. 2021. Review on the ecophysiology of important Andean fruits: *Passiflora L.* *Revista Facultad Nacional de Agronomía Medellín* 74(2): 9471–9481.
- Fisher, G., Almanza-Merchán, P.J. and Ramírez, F. 2012. Source and sink relationship in fruit species: A review. *Revista Colombiana de Ciencias Hortícolas* 6(2): 238–253.
- Fitzgerald, M., Heinrich, M. and Booker, A. 2019. Medicinal plant analysis: A historical and regional discussion of emergent complex techniques. *Frontiers in Pharmacology* 10: 1480.
- Fleancu, M. 2007. Correlations among some physiological processes in apple fruit during growing and maturation processes. *International Journal of Agriculture & Biology* 9(4): 613–616.

- Florez, L., Vaillant, F., Hollander, H. and Ariza-Nieto, M. 2003. Passion fruit juice sacs: Biochemical characterization and enzymatic treatment. *Tropical Science* 43(1): 28–34.
- Forlani, S., Mizzotti, C. and Masiero, S. 2021. The NAC side of the fruit: Tuning of fruit development and maturation. *BMC Plant Biology* 21: 238.
- Franco, G., Valenzuela, J.R.C., Londoño, G.A.C., Rojano, B.A., Correa, A.M.P. and Arias, M.L. 2014. Physicochemical properties of gulupa fruits (*Passiflora edulis* Sims) during postharvest. *Revista Iberoamericana de Tecnología Postcosecha* 15(1): 15–30.
- Fraser, C. and Chapple, C. 2011. The phenylpropanoid pathway in *Arabidopsis*. In *The Arabidopsis Book*. pp. e0152. USA: The American Society of Plant Biologists.
- Fresh Plaza, 2021. Peru: Higher Brix levels key to passion fruit exports. Retrieved 25 August 2021 from <https://www.freshplaza.com/article/2193480/peru-higher-brix-levels-key-to-passion-fruit-exports/>.
- Fu, H., Qiao, Y., Wang, P., Mu, X., Zhang, J., Fu, B. and Du, J. 2021. Changes of bioactive components and antioxidant potential during fruit development of *Prunus humilis* Bunge. *Plos One* 16: 1–15.
- Génard, M., Lescourret, F., Gomez, L. and Habib, R. 2003. Changes in fruit sugar concentrations in response to assimilate supply, metabolism and dilution: A modelling approach applied to peach fruit (*Prunus persica*). *Tree Physiology* 23(6): 373–385.
- Gerhards, N., Neubauer, L., Tudzynski, P. and Li, S. 2014. Biosynthetic pathways of ergot alkaloids. *Toxins* 6(12): 3281–3295.
- Germano, T., Aguiar, R., Bastos, M.S., Moreira, R., Ayala-Zavala, J. and de Miranda, M.R. 2019. Postharvest biology and technology galactomannan-carnauba wax coating improves the antioxidant status and reduces chilling injury of ‘Paluma’ guava. *Postharvest Biology and Technology* 149: 9–17.
- Ghada, B., Pereira, E., Pinela, J., Prieto, M., Pereira, C., Calhelha, R., Stojkovic, D., Sokóvic, M., Zaghoudi, K., Barros, L. and Ferreira, I.C.F. 2020. Recovery of anthocyanins from passion fruit epicarp for food colorants: Extraction process optimization and evaluation of bioactive properties. *Molecules* 25(14): 3203.
- Ghosh, A., Dey, K., Bauri, F. and Dey, A. 2017. Physico-chemical changes during fruit growth and developmental stages in yellow type passion fruit (*Passiflora edulis* f. *Flavicarpa* Degener) accessions. *Journal of Applied and Natural Science* 9(4): 2026–2032.
- Giacomini, S.B., Lúcio, A.D., Santana, C.S., Olivoto, T., Diel, M.I and Ketzer K.D. 2019. Nonlinear growth models: An alternative to ANOVA in tomato trials evaluation. *European Journal of Agronomy* 104: 21–36.

- Giambanelli, E., Gómez-Caravaca, A., Ruiz-Torralba, A., Guerra-Hernández, E., Figueroa-Hurtado, J., García-Villanova, B. and Verardo, V. 2020. New advances in the determination of free and bound phenolic compounds of banana passion fruit pulp (*Passiflora tripartita*, var. Mollissima (Kunth) L.H. Bailey) and their in vitro antioxidant and hypoglycemic capacities. *Antioxidants* 9(7): 1–17.
- Gillaspy, G., Ben-David, H. and Gruissem, W. 1993. Fruits: A developmental perspective. *The Plant Cell* 5(10): 1439.
- Gioia, F., Tzortzakis, N., Roupael, Y., Kyriacou, M., Sampaio, S., Ferreira, I.C.F. and Petropoulos, S. 2020. Grown to be blue—antioxidant properties and health effects of coloured vegetables. Part II: Leafy, fruit, and other vegetables. *Antioxidants* 9(2): 1–42.
- Goldschmidt, E. and Huber, S. 1992. Regulation of photosynthesis by end-product accumulation in leaves of plants storing starch, sucrose, and hexose sugars. *Plant physiology* 99: 1443–1448.
- Gonçalves, M., Argenta, L. and Martin, M.S.D. 2017. Maturity and quality of apple fruit during the harvest period at apple industry. *Revista Brasileira De Fruticultura* 39(5): 1–10.
- Grafi, G., Florentin, A., Ransbotyn, V. and Morgenstern, Y. 2011. The stem cell state in plant development and in response to stress. *Frontiers in Plant Science* 2: 1–10.
- Graindorge, M., Giustini, C., Jacomin, A., Kraut, A., Curien, G. and Matringe, M. 2010. Identification of a plant gene encoding glutamate/aspartate-prephenate aminotransferase: The last homeless enzyme of aromatic amino acids biosynthesis. *FEBS Letters* 584(20): 4357–4360.
- Griffiths, C., Paul, M. and Foyer, C. 2016. Metabolite transport and associated sugar signalling systems underpinning source/sink interactions. *Biochimica et Biophysica Acta - Bioenergetics* 1857(10): 1715–1725.
- Guimarães, S., Lima, I. and Modolo, L. 2020. Phenolic content and antioxidant activity of parts of *Passiflora edulis* as a function of plant developmental stage. *Acta Botanica Brasilica* 34(1): 74–82.
- Guo, W., Rao, G. and Wen, X. 2021. Arabinogalactan in banana: Chemical characterization and pharmaceutical effects. *International Journal of Biological Macromolecules* 167: 1059–1065.
- Gupta, A., Medhi, M., Chakraborty, S., Yumnam, M. and Mishra, P. 2021. Development of rapid and non-destructive technique for the determination of maturity indices of pomelo fruit (*Citrus grandis*). *Journal of Food Measurement and Characterization* 15(2): 1463–1474.

- Hada, H., Hidema, J., Maekawa, M. and Kumagai, T. 2003. Higher amounts of anthocyanins and UV-absorbing compounds effectively lowered CPD photorepair in purple rice (*Oryza sativa* L.). *Plant, Cell and Environment* 26(10): 1691–1701.
- Hadacek, F., Bachmann, G., Engelmeier, D. and Chobot, V. 2010. Hormesis and a chemical raison d'être for secondary plant metabolites. *Dose-Response* 9: 79–116.
- Haminiuk, C.W., Maciel, G., Plata-Oviedo, M.S. and Peralta, R. 2012. Phenolic compounds in fruits - An overview. *International Journal of Food Science and Technology* 47(10): 2023–2044.
- Han, Y., Ban, Q., Hou, Y., Meng, K., Suo, J. and Rao, J. 2016. Isolation and characterization of two persimmon xyloglucan Endotransglycosylase/Hydrolase (XTH) genes that have divergent functions in cell wall modification and fruit postharvest softening. *Frontiers in Plant Science* 7: 1–12.
- Handa, A. and Mattoo, A. 2010. Differential and functional interactions emphasize the multiple roles of polyamines in plants. *Plant Physiology and Biochemistry* 48(7): 540–546.
- Harrison, E., Mcqueen-Mason, S. and Manning, K. 2001. Expression of six expansin genes in relation to extension activity in developing strawberry fruit. *Journal of Experimental Botany* 52: 1427–1446.
- Hartwig, P. 1994. Flavor Characterization of Selected Acidulants in Model and Food System. Master Thesis, Oregon State University.
- Hatfield, J. and Prueger, J. 2015. Temperature extremes: Effect on plant growth and development. *Weather and Climate Extremes* 10: 4–10.
- He, X., Luan, F., Yang, Y., Wang, Z., Zhao, Z., Fang, J. and Wang, M. 2020. *Passiflora edulis*: An insight into current researches on phytochemistry and pharmacology. *Frontier Pharmacology* 11:617.
- Hellal, K., Maulidiani, M., Ismail, I. and Tan, C. 2020. Antioxidant, α -Glucosidase, and nitric oxide inhibitory activities of six Algerian traditional medicinal plant extracts and $^1\text{H-NMR}$ -based metabolomics study of the active extract. *Molecules* 25(5):1247.
- Herremans, E., Verboven, P., Hertog, M.L.A.T., Cantre, D., van Dael, M., de Schryver, T., van Hoorebeke, L. and Nicolai, B. 2015. Spatial development of transport structures in apple (*Malus × domestica* Borkh.) fruit. *Frontiers in Plant Science* 6: 1–14.

- Herrera, J., Nixon, R., Marentes, M. and Díaz, M.P. 2019. Optimizing the extraction of anthocyanins from purple passion fruit peel using response surface methodology. *Journal of Food Measurement and Characterization* 14: 185-193.
- Hewitt, S. and Dhingra, A. 2020. Beyond ethylene: New insights regarding the role of alternative oxidase in the respiratory climacteric. *Frontiers in Plant Science* 11: 543958
- Hollender, C., Geretz, A., Slovin, J. and Liu, Z. 2012. Flower and early fruit development in a diploid strawberry. *Fragaria vesca*. *Planta* 235: 1123–1139.
- Hong, J., Cowan, A. and Lee, S. 2004. Glucose inhibits ACC Oxidase activity and ethylene biosynthesis in ripening tomato fruit glucose inhibits ACC oxidase activity and ethylene biosynthesis in ripening tomato fruit. *Plant Growth and Development* 43: 81–87.
- Honor, M., Belmonte-Ureña, L., Navarro-Velasco, A. and Camacho-Ferre, F. 2019. The production and quality of different varieties of papaya grown under greenhouse in short cycle in Continental Europe. *International Journal of Environmental Research and Public Health Article* (16): 1789.
- Hossain, A., Rana, M., Kimura, Y. and Roslan, H. 2014. Changes in biochemical characteristics and activities of ripening associated enzymes in mango fruit during the storage at different temperatures. *Hindawi Publishing Corporation* 2014: 1–11.
- Houben, M. and Van de Poel, B. 2019. 1-aminocyclopropane-1-carboxylic acid oxidase (ACO): The enzyme that makes the plant hormone ethylene. *Frontiers in Plant Science* 10: 695.
- Hu, Y., Zhang, Y., Yu, W., Hänninen, H., Song, L., Du, X., Zhang, R. and Wu, J. 2018. Novel insights into the influence of seed sarcotesta photosynthesis on accumulation of seed dry matter and oil content in *Torreya grandis* cv. “Merrillii.” *Frontiers in Plant Science* 8:2179.
- Huang, Y., Rasco, B. and Cavinato, A. 2009. Fruit Juices. In *Infrared Spectroscopy for Food Quality Analysis and Control*, ed. D.W. Sun. pp. 355–375. United Kingdom: Academic Press.
- Huber, D., Karakurt, Y. and Jeong, J. 2001. Pectin degradation in ripening and wounded fruits. *Revista Brasileira de Fisiologia Vegetal* 13(2): 224–241.
- Huberman, M., Zehavi, U., Stein, W., Etxeberria, E. and Goren, R. 2005. In vitro sugar uptake by grapefruit (*Citrus paradisi*) juice-sac cells. *Functional Plant Biology* 32(4): 357–366.
- Hussein, R. and El-Ansarry, A. 2013. Plants secondary metabolites: In *The Key Drivers of The Pharmacological Actions Of Medicinal Plants*. pp. 137-155. United Kingdom: IntechOpen.

- Hussin, M., Hamid, A., Abas, F., Ramli, N., Jaafar, A., Roowi, S., Majid, N. and Dek, M.D. 2019. NMR-based metabolomics profiling for radical scavenging and anti-aging properties of selected herbs. *Molecules* 24(17): 3208.
- Hwang, H., Kim, Y. and Shin, Y. 2020. Assessment of physicochemical quality, antioxidant content and activity, and inhibition of cholinesterase between unripe and ripe blueberry fruit. *Foods* 9: 690.
- Igamberdiev, A. and Eprintsev, A. 2016. Organic acids: The pools of fixed carbon involved in redox regulation and energy balance in higher plants. *Frontiers in Plant Science* 7: 1042.
- Ikegaya, A., Toyozumi, T., Ohba, S., Nakajima, T., Kawata, T., Ito, S. and Arai, E. 2019. Effects of distribution of sugars and organic acids on the taste of strawberries. *Food Science & Nutrition* 7(7): 1-8.
- Iloki, A.S., Lewis, L.L., Rivera-Castañeda, E., Gil-Salido, A., Acosta-Silva, A., Meza-Cueto, A. and Rubio-Pino, J. 2013. Effect of maturity and harvest season on antioxidant activity, phenolic compounds and ascorbic acid of *Morinda citrifolia* L. (noni) grown in Mexico (with track change). *African Journal of Biotechnology* 12(29): 4630–4639.
- Ingallina, C., Sobolev, A., Circi, S., Spano, M., Giusti, A. and Mannina, L. 2020. New hybrid tomato cultivars: An NMR-based chemical characterization. *Applied Sciences* 10(5):1887.
- Isah, T. 2019. Stress and defence responses in plant secondary metabolites production. *Biological Research* 52(1): 39.
- Jain, S., Purohit, S., Kumar, D. and Goud, V. 2021. Passion fruit seed extract as an antioxidant additive for biodiesel; shelf life and consumption kinetics. *Fuel* 289: 119906.
- Jan, N. and Kawabata, S. 2011. Relationship between fruit soluble solid content and the sucrose concentration of the phloem sap at different leaf to fruit ratios in tomato. *Journal of The Japanese Society for Horticultural Science* 80(3): 314–321.
- Jander, G. and Joshi, V. 2009. Aspartate-derived amino acid biosynthesis in *Arabidopsis thaliana*. In *The Arabidopsis Book*. pp. e0121. USA: The American Society of Plant Biologists.
- Jia, J., Zhang, F., Li, Z., Qin, X. and Zhang, L. 2015. Comparison of fruits of *Forsythia suspensa* at two different maturation stages by NMR-based metabolomics. *Molecules* 20(6): 10065–10081.
- John, W. 2012. Image Perception and Sensing. In *Multidimensional Signal, Image, and Video Processing and Coding (Second Edition)*, ed. J.W. Wood. pp. 193–221. United Kingdom: Academic Press.

- Joy, P.P. 2010. Passionfruit Production Technology (Adhoc). Kerala Agricultural University: India.
- Joymak, W., Ngamukote, S., Chantarasinlapin, P. and Adisakwattana, S. 2021. Unripe papaya by-product: From food wastes to functional ingredients in pancakes. *Foods* 10: 615.
- Juan-Cabot, A., Galmés, J. and Conesa, M. 2022. The tomato long shelf-life fruit phenotype: Knowledge, uncertainties and prospects. *Scientia Horticulturae* 291: 110578.
- Kadum, H., Hamid, A., Abas, F., Ramli, N., Mohammed, A., Muhialdin, B. and Jaafar, A. 2019. Bioactive compounds responsible for antioxidant activity of different varieties of date (*Phoenix dactylifera* L.) elucidated by ¹H- NMR based metabolomics. *International Journal of Food Properties* 22(1): 462–476.
- Kalayanamitra, K. and Sornsrivichai, J. 2005. Model for evaluation of maturity index of durian fruit (*Durio zibethinus* Murray 'Monthong'). *Acta Horticulturae* 682: 587–592.
- Karppinen, K., Zoratti, L., Nguyenquynh, N., Häggman, H. and Jaakola, L. 2016. On the developmental and environmental regulation of secondary metabolism in *Vaccinium* spp. Berries. *Frontiers in Plant Science* 7(5): 1–9.
- Ketsa, S., Wisutiamonkul, A., Palapol, Y. and Paull, R. 2020. The durian: botany, horticulture, and utilization. In *Horticultural Reviews*, ed. I. Warrington. pp. United States: John Wiley and Sons.
- Khefifi, H., Selmane, R., Mimoun, M., Tadeo, F., Morillon, R. and Luro, F. 2020. Abscission of orange fruit (*Citrus sinensis* (L.) Osb.) in the Mediterranean basin depends more on environmental conditions than on fruit ripeness. *Agronomy* 10(4): 591.
- Khoo, H., Azlan, A., Tang, S. and Lim, S. 2017. Anthocyanidins and anthocyanins: Coloured pigments as food, pharmaceutical ingredients, and the potential health benefits. *Food and Nutrition Research* 61(1). 1361779.
- Kim, H., Park, S., Hyun, S., Yang, S., Lee, J., Auh, J., Kim, J., Cho, S., Marriott, P. and Choi, H. 2011. Biochemical monitoring of black raspberry (*Rubus coreanus* Miquel) fruits according to maturation stage by ¹H NMR using multiple solvent systems. *Food Research International* 44(7): 1977–1987.
- Kiralj, R. and Ferreira, M.M. 2009. Basic validation procedures for regression models in QSAR and QSPR studies: Theory and application. *Journal of the Brazilian Chemical Society* 20(4): 770–787.
- Kishore, K., Pathak, K., Shukla, R. and Bharali, R. 2011. Effect of storage temperature on physico-chemical and sensory attributes of purple passion fruit (*Passiflora edulis* Sims). *Journal of Food Science and Technology* 48(4): 484–488.

- Kishore, K., Pathak, K., Yadav, D., Bujarbaruah, K., Bharali, R. Shukla, R. 2006. Passion Fruit. ICAR Research Complex for NEH region: Meghalaya.
- Klie, S., Osorio, S., Tohge, T., Drincovich, M., Fait, A., Giovannoni, J., Fernie, A. and Nikoloski, Z. 2014. Conserved changes in the dynamics of metabolic processes during fruit development and ripening across species. *Plant Physiology* 164(1): 55–68.
- Klopsch, R., Baldermann, S., Voss, A., Rohn, S., Schreiner, M. and Neugart, S. 2018. Bread enriched with legume microgreens and leaves - Ontogenetic and baking-driven changes in the profile of secondary plant metabolites. *Frontiers in Chemistry* 6(8): 1–19.
- Kok, S., Abdullah, M., Ee, G.W.W.C. and Namasivayam, P. 2015. A histological study of oil palm (*Elaeis guineensis*) endosperm during seed development. *Journal of Oil Palm Research* 27(2): 107–112.
- Kondo, T. and Higuchi, H. 2013. Acidity of passion fruit as affected by potassium fertilizer. *Acta Horticulturae* 984: 385–392.
- Kotani, A., Kusu, F., Takamura, K. and Hakamata, H. 2020. Review-A portable voltammetric sensor for determining titratable acidity in foods and beverages. *Journal of The Electrochemical Society* 167: 037517.
- Kour, R., Mandeep, S., Gill, P.P. and Jawandha, S. 2018. Ripening quality of Dusehri mango in relation to harvest time. *Journal of Food Science and Technology* 55(7): 2406–2411.
- Kozłowski, T. and Pallardy, S. 1997. Plant hormones and other endogenous growth regulators. In *Physiology of Woody Plants 3rd Edition*, ed. G.P. Stephen. pp. 367-377. United Kingdom: Academic Press.
- Kyriacou, M., Antoniou, C., Roupheal, Y., Graziani, G. and Kyratzis, A. 2021. Mapping the primary and secondary metabolomes of carob (*Ceratonia siliqua* L.) fruit and its postharvest antioxidant potential at critical stages of ripening. *Antioxidants* 10(1): 1-21.
- Lado, J., Gambetta, G. and Zacarias, L. 2018. Key determinants of citrus fruit quality: Metabolites and main changes during maturation. *Scientia Horticulturae* 233: 238–248.
- Lara-Espinoza, C., Carvajal-Millán, E., Balandrán-Quintana, R., López-Franco, Y. and Rascón-Chu, A. 2018. Pectin and pectin-based composite materials: Beyond food texture. *Molecules* 23(4): 942.
- Lee, S.K. and Kader, A.A., 2000. Preharvest and postharvest factors influencing vitamin content of horticultural crops. *Postharvest Biology and Technology* 20: 207–220.

- Leprince, O., Pellizzaro, A., Berriri, S. and Buitink, J. 2017. Late seed maturation: Drying without drying. *Journal of Experimental Botany* 68(4): 827–841.
- Leroux, O. 2012. Collenchyma: A versatile mechanical tissue with dynamic cell walls. *Annals of Botany* 110(6): 1083–1098.
- Li, J., Zhang, C., Liu, H., Liu, J. and Jiao, Z. 2020. Profiles of sugar and organic acid of fruit juices: A comparative study and implication for authentication. *Journal of Food Quality* 2020: 7236534.
- Li, M., Feng, F. and Cheng, L. 2012. Expression patterns of genes involved in sugar metabolism and accumulation during apple fruit development. *Plos One* 7(3): e33055.
- Li, M., Li, P., Ma, F., Dandekar, A. and Cheng, L. 2018a. Sugar metabolism and accumulation in the fruit of transgenic apple trees with decreased sorbitol synthesis. *Horticulture Research* 5(60): 1–11.
- Li, P., Wang, L., Liu, H. and Yuan, M. 2021. Impaired *SWEET* -mediated sugar transportation impacts starch metabolism in developing rice seeds. *The Crop Journal*. In press.
- Li, X., Zhu, X., Wang, H., Lin, X., Lin, H. and Chen, W. 2018b. Postharvest biology and technology postharvest application of wax controls pineapple fruit ripening and improves fruit quality. *Postharvest Biology and Technology* 136: 99–110.
- Li, Y., He, Q., Du, S., Guo, S., Geng, Z. and Deng, Z. 2018c. Study of methanol extracts from different parts of *Peganum harmala* L. Using ¹H-NMR plant metabolomics. *Journal of Analytical Methods in Chemistry* 2018: 1-9.
- Liang, J., Ren, Y., Wang, Y., Han, M., Yue, T., Wang, Z. and Gao, Z. 2021. Physicochemical, nutritional, and bioactive properties of pulp and peel from 15 kiwifruit cultivars. *Food Bioscience* 42: 101157.
- Liew, S.Q., Chin, N.L. and Yusof, Y.A. 2014. Extraction and characterization of pectin from passion fruit peels. *Agriculture and Agricultural Science Procedia* 2: 231–236.
- Lim, V., Gorji, S., Daygon, V. and Fitzgerald, M. 2020. Untargeted and targeted metabolomic profiling of Australian indigenous fruits. *Metabolites* 10(3): 114.
- Lima, R.E., Farias, L.F.D., Ferreira, J.F., Suarez, D. and Bezerra, M. 2020. Translocation of photoassimilates in melon vines and fruits under salinity using ¹³C isotope. *Scientia Horticulturae* 274(15): 109659.
- Lima-Neto, A.B., Marques, M.M., Mendes, F.N., Vieira, Í.G., Diniz, D. and Guedes, M.I. 2017. Antioxidant activity and physicochemical analysis of passion fruit (*Passiflora glandulosa* Cav.) pulp native to Cariri region. *Acta Scientiarum Biological Sciences* 39(4):417

- Lin, X., Gao, H., Ding, Z., Zhan, R., Zhou, Z. and Ming, J. 2021. Comparative metabolic profiling in pulp and peel of green and red pitayas (*Hylocereus polyrhizus* and *Hylocereus undatus*) reveals potential valorization in the pharmaceutical and food industries. *BioMed Research International* 2021: 6546170.
- Lina, M., Avraham, S. and Ron, P. 2014. Definition of minimum maturity indices for harvesting of early-season sweet pomegranate (*Punica granatum* L.) fruit. *The Journal of Horticultural Science and Biotechnology* 189: 17–22.
- Liu, M., Pirrello, J., Chervin, C., Roustan, J. and Bouzayen, M. 2015. Ethylene control of fruit ripening: Revisiting the complex network of transcriptional regulation. *Plant Physiology* 169(4): 2380–2390.
- Liu, Y., Wu, Y., Che, F., Zhang, Z. and Chen, B. 2019. Changes in “Ruaner” Pear (*Pyrus ussuriensis*) during freezing -thawing period. *Molecules* 24: 2611.
- López-Vargas, J., Fernández-López, J., Pérez-Álvarez, J. and Viuda-Martos, M. 2013. Chemical, physico-chemical, technological, antibacterial and antioxidant properties of dietary fiber powder obtained from yellow passion fruit (*Passiflora edulis* var. *flavicarpa*) co-products. *Food Research International* 51(2): 756–763.
- Lourith, N. and Kanlayavattanakul, M. 2013. Antioxidant activities and phenolics of *Passiflora edulis* seed recovered from juice production residue. *Journal of Oleo Science* 62(4): 235–240.
- Lytovchenko, A., Eickmeier, I., Pons, C., Osorio, S., Szecowka, M., Lehmborg, K., Arrivault, S., Tohge, T., Pineda, B., Anton, M., Hedtke, B., Lu, Y., Fisahn, J., Bock, R., Stütt, M., Grimm, B., Granell, A. and Fernie, A. 2011. Tomato fruit photosynthesis is seemingly unimportant in primary metabolism and ripening but plays a considerable role in seed development. *Plant Physiology* 157(4): 1650–1663.
- Ma, W., Xu, L., Gao, S., Lyu, X., Cao, X. and Yao, Y. 2021. Melatonin alters the secondary metabolite profile of grape berry skin by promoting VvMYB14-mediated ethylene biosynthesis. *Horticulture Research* 8: 43.
- Maduwanthi, S.D. and Marapana, R.A.U. 2019. Induced ripening agents and their effect on fruit quality of banana. *International Journal of Food Science* 2019: 1–8.
- Magalhães, D., da Silva, D., Ramos, J., Salles Pio, L., Pasqual, M., Vilas-Boas, E.V., Galvão, E. and de Melo, E. 2019. Changes in the physical and physico-chemical characteristics of red-pulp dragon fruit during its development. *Scientia Horticulturae* 253: 180–186.
- Magwaza, L. and Opara, U. 2015. Analytical methods for determination of sugars and sweetness of horticultural products-A review. *Scientia Horticulturae* 184: 179–192.

- Mahmud, T.M. 2017. Postharvest food losses and food waste, eds. A. Sahariah, M. Ahmad. In *Postharvest: An unsung solution for postharvest*. pp. 5-30. Serdang: UPM Publisher.
- Majumdar, R., Barchi, B., Turlapati, S., Gagne, M., Minocha, R., Long, S. and Minocha, S. 2016. Glutamate, ornithine, arginine, proline, and polyamine metabolic interactions: The pathway is regulated at the post-transcriptional level. *Frontiers in Plant Science* 7: 1–17.
- Makhai, M., Shakya, A. and Kale, R. 2021. Dietary phytochemicals: As a natural source of antioxidants. In *Antioxidants Benefits, Sources, Mechanisms of Action*, ed. V. Waisundara. pp. 1–25. United Kingdom: IntechOpen.
- Mamat, S., Azizan, K., Baharum, S., Noor, N. and Aizat, W. 2020. GC-MS and LC-MS analyses reveal the distribution of primary and secondary metabolites in mangosteen (*Garcinia mangostana* Linn.) fruit during ripening. *Scientia Horticulturae* 262: 109004.
- Manach, C., Williamson, G., Morand, C. and Scalbert, A. 2005. Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. *American Journal of Clinical Nutrition* 81: 230–242.
- Manjunatha, G., Gupta, K.J., Lokesh, V., Mur, L.A. and Neelwarne, B. 2012. Nitric oxide counters ethylene effects on ripening fruits. *Plant Signalling & Behaviour* 7(4): 476–483.
- Marchiosi, R., dos Santos, W., Constantin, R., de Lima, R., Soares, A., Finger-Teixeira, A., Mota, T., de Oliveira, D., Foletto-Felipe, M.D., Abrahão, J. and Ferrarese-Filho, O. 2020. Biosynthesis and metabolic actions of simple phenolic acids in plants. *Phytochemistry Reviews* 19(4): 865–906.
- María, L., Ferreyra, F., Sebastián, P. and Casati, P. 2012. Flavonoids: Biosynthesis, biological functions, and biotechnological applications. *Frontiers in Microbiology* 3(9): 1–15.
- Marini, Pengurus Jabatan Pertanian Melaka, Malaysia, pers. Comm. Jan 2018.
- Marowa, P., Ding, A. and Kong, Y. 2016. Expansins: Roles in plant growth and potential applications in crop improvement. *Plant Cell Reports* 35: 949–965.
- Marsh, K., Rossiter, K., Lau, K., Walker, S., Gunson, A. and Macrae, E. 2003. The use of fruit pulps to explore flavour in kiwifruit. *Acta Horticulturae* 610: 229–237.
- Martín, J., Navas, M., Jiménez-Moreno, A. and Asuero, A. 2013. Anthocyanin pigments: Importance, sample preparation and extraction. In *Phenolic Compounds Natural Sources, Importance and Applications metabolites*, ed. S.H. Marcos. pp. 137–144. United Kingdom: IntechOpen.

- Massot, C., Bancel, D., Lauri, F., Truffault, V., Baldet, P., Stevens, R. and Gautier, H. 2013. High temperature inhibits ascorbate recycling and light stimulation of the ascorbate pool in tomato despite increased expression of biosynthesis genes. *Plos One* 8(12): e84474.
- Matas, A., Agustí, J., Tadeo, F., Talón, M. and Rose, J.K. 2010. Tissue-specific transcriptome profiling of the citrus fruit epidermis and subepidermis using laser capture microdissection. *Journal of Experimental Botany* 61(12): 3321–3330.
- Matheyambath, A., Padmanabhan, P. and Paliyath, G. 2015. Citrus Fruits. In *Encyclopedia of Food and Health*, ed. B. Caballero, P. Finglas, and F. Toldra. pp. 136–140. United Kingdom: Academic Press.
- Mathieu, L., Joas, J. and Jahiel, M. 2009. How to predict the harvest date of tropical fruit: From simple methods to complex models. *Acta Horticulturae* 880: 175–182.
- Matsoukas, I., Jung, C., Albrechts, C., Jung, C. and Albrechts, C. 2014. Interplay between sugar and hormone signalling pathways modulate floral signal transduction. *Frontiers in Plant Science* 5: 1–12.
- Matusaka, Y. and Kawabata, J. 2010. Evaluation of antioxidant capacity of non-edible parts of some selected tropical fruits. *Food Science and Technology Research* 16(5): 467–472.
- Mazlan, O., Aizat, W., Baharum, S., Azizan, K. and Noor, N. 2018. Metabolomics analysis of developing *Garcinia mangostana* seed reveals modulated levels of sugars, organic acids and phenylpropanoid compounds. *Scientia Horticulturae* 233: 323–330.
- Mazur, E. and Friml, J., 2017. Vascular tissue development and regeneration in the model plant *Arabidopsis*. In: *Plant Engineering*, ed. S. Jurić. pp. 113–133. United Kingdom: IntechOpen.
- Md Nor, S. and Ding, P. 2020. Trends and advances in edible biopolymer coating for tropical fruit: A review. *Food Research International* 13: 109208.
- Mediani, A., Abas, F., Maulidiani, M., Khatib, A., Tan, C.P, Shaari, K. and Ismail, A.I. 2017. Characterization of metabolite profile in *Phyllanthus niruri* and correlation with bioactivity elucidated by nuclear magnetic resonance-based metabolomics. *Molecules* 22(6): 902.
- Medlicott, A. and Thompson, A. 1985. Analysis of sugars and organic acids in ripening mango fruits (*Mangifera indica* L. var Keitt) by high performance liquid chromatography. *Journal of the Science of Food and Agriculture* 36(7): 561–566.
- Meinhart, A., Damin, F., Caldeirão, L., de Jesus Filho, M., da Silva, L., da Silva Constant, L., Filho, J., Wagner, R. and Godoy, H. 2019. Chlorogenic and caffeic acids in 64 fruits consumed in Brazil. *Food Chemistry* 286(7): 51–63.

- Mekuria, W. 2018. The link between agricultural production and population dynamics in Ethiopia: A review. *Advances in Plants & Agriculture Research* 8(4): 336–346.
- Mellentín, J. and Crawford, K. 2008. Marketing healthy fruit. In *Improving the Health-Promoting Properties of Fruit and Vegetable Products*, ed. F.A, Tomás-Barberán and M.I., Gil. pp. 55-71. Sawston: Woodhead Publishing Limited.
- MetMalaysia, 2018. Annual Report 2018. Retrieved 25 August 2021 from <https://www.met.gov.my/content/pdf/penerbitan/laporantahunan/laporantahunan2018.pdf>.
- MetMalaysia, 2019. Annual Report 2019. Retrieved 25 August 2021 from <https://www.met.gov.my/content/pdf/penerbitan/laporantahunan/laporantahunan2019.pdf>.
- MetMalaysia, 2020. Annual Report 2021. Retrieved 25 August 2021 from <https://www.met.gov.my/content/pdf/penerbitan/laporantahunan/laporantahunan2021.pdf>.
- Michailides, T. and Manganaris, G. 2009. Harvesting and handling effects on postharvest decay. An international journal for reviews in postharvest biology and technology. *Stewart Postharvest Review* 2(3): 1–7.
- Michelle, G., Silva, C., Batista, W., Medeiros, D., Rodrigues, A., Helena, M., Cordeiro, M., Martins, N., Bortolini, D. and Polete, G. 2017. The chitosan affects severely the carbon metabolism in mango (*Mangifera indica* L. cv. Palmer) fruit during storage. *Revista Iberoamericana de Tecnología Postcosecha* 237: 372–378.
- Min, H., Mi, K., Ho, C., Hwa, B., Lee, J., Kong, G., Jin, S., Kim, S., Wook, S. and Chul, B. 2009. Mass spectrometry based metabolomic approaches in urinary biomarker study of women 's cancers. *Clinica Chimica Acta* 400: 63–69.
- Ministry Foreign Affair, 2021. The European market potential for exotic tropical fruit. Retrieved 25 August 2021 from <https://www.cbi.eu/market-information/fresh-fruit-vegetables/exotic-tropical-fruit/market-potential>.
- Miquel, G., Lina, T. and Rebecca, J. 2004. Plant respiration and elevated atmospheric CO₂ concentration: Cellular responses and global significance. *Annals of Botany* 94: 647–656.
- Mitalo, O., Tokiwa, S., Kondo, Y., Otsuki, T. and Galis, I. 2019. Low temperature storage stimulates fruit softening and sugar accumulation without ethylene and aroma volatile production in kiwifruit. *Frontiers in Plant Science* 10: 888.
- Mizrach, A. 2000. Determination of avocado and mango fruit properties by ultrasonic technique. *Ultrasonics* 38: 717–722.

- Mohammad, M. and Ding, P. 2019. Physico-textural and cellular structure changes of *Carissa congesta* fruit during growth and development. *Scientia Horticulturae* 246: 380–389.
- Moing, A., Renaud, C., Gaudillère, M., Raymond, P., Roudeillac, P. and Denoyes-Rothan, B. 2001. Biochemical changes during fruit development of four strawberry cultivars. *Journal of the American Society for Horticultural Science* 126(4): 394–440.
- Mokrzycki, W. and Tatol, M. 2012. Colour difference Delta E-A survey Colour. Olsztyn, Poland.
- Moneruzzaman, K.M., Hossain, A.B.M., Sani, W., Saifuddin, M. and Alenazi, M. 2009. Effect of harvesting and storage conditions on the post-harvest quality of tomato (*Lycopersicon esculentum* Mill) cv. Roma VF. *Australian Journal of Crop Science* 3(2): 113–121.
- Moreno, J., Tran, T., Augusto, L. and Lopez, B. 2021. Original article physicochemical and physiological changes during the ripening of Banana (*Musaceae*) fruit grown in Colombia. *International Journal of Food Science and Technology* 56: 1171–1183.
- Moya-León, M., Mattus-Araya, E. and Herrera, R. 2019. Molecular events occurring during softening of strawberry fruit. *Frontiers in Plant Science* 10: 615.
- Muñoz-Robredo, P., Robledo, P., Manríquez, D., Molina, R. and Defilippi, B. 2011. Characterization of sugars and organic acids in commercial varieties of table grapes. *Chilean Journal of Agricultural Research* 71(3): 452–458.
- Munteanu, I. and Apetrei, C. 2021. Analytical methods used in determining antioxidant activity: A Review. *International Journal of Molecular Science* 22: 3380.
- Muriel, Q., Angosto, T., Yuste-Lisbona, F., Blanchard-Gros, R., Bigot, S., Martinez, J. and Lutts, S. 2019. Tomato fruit development and metabolism. *Frontiers in Plant Science* 10: 1554.
- Muthuramalingam, P., Krishnan, S., Pandian, S., Mareeswaran, N., Aruni, W., Pandian, S. and Ramesh, M. 2018. Global analysis of threonine metabolism genes unravels key players in rice to improve the abiotic stress tolerance. *Scientific Reports* 8(1): 1–14.
- Nadjama, B., Abreu, C.B. De, Pinho, C.S., Junior, M.M.D.N., Silva, M.D., Espino, M., Silva, M.F. and Dias, F.D.S. 2022. Application of multivariate analysis to assess stress by Cd, Pb and Al in basil (*Ocimum basilicum* L.) using caffeic acid, rosmarinic acid, total phenolics, total flavonoids and total dry mass in response. *Food Chemistry* 367: 130682.

- Naosuke, N. and Coombe, B. 1990. Ultrastructural changes in the primordia of juice sacs of Satsuma Mandarin Fruits. *Journal of the Japanese Society for Horticultural Science* 59(1): 35–41.
- Natnicha, T., Kanyarat, L. and Kraiyot, S. 2019. Effect of passion fruit juice and pectin on characteristics of purple yard long bean jam. *IOP Conference Series: Materials Science and Engineering* 639(1): 4–10.
- Nedeva, D. and Nikolava, A. 1999. Fresh and dry weight changes and germination capacity of natural or premature desiccated developing wheat seeds. *Bulgarian Journal of Plant Physiology* 25(1–2): 3–15.
- Neugart, S., Krumbein, A. and Zrenner, R. 2016. Influence of light and temperature on gene expression leading to accumulation of specific flavonol glycosides and hydroxycinnamic acid derivatives in kale (*Brassica oleracea* var. *sabellica*). *Frontiers in Plant Science* 7: 1–16.
- Ng, J.K., Schröder, R., Sutherland, P., Hallett, I., Hall, M., Prakash, R., Smith, B., Melton, L. and Johnston, J. 2013. Cell wall structures leading to cultivar differences in softening rates develop early during apple (*Malus x domestica*) fruit growth. *BMC Plant Biology* 13: 183.
- Nouri, A., Heibati, F. and Heidarian, E. 2021. Gallic acid exerts anti-inflammatory, anti-oxidative stress, and nephroprotective effects against paraquat-induced renal injury in male rats. *Naunyn Schmiedebergs Arch Pharmacol*: 1–9.
- Nsubuga, U. 2021. Passion fruits, the money maker. Retrieved 23 Jun 2021 from <https://www.newvision.co.ug/news/1503865/passion-fruits-money-maker>.
- Oh, J., Yoon, D., Han, J., Choi, H. and Sung, G. 2018. ¹H NMR based metabolite profiling for optimizing the ethanol extraction of *Wolfiporia cocos*. *Saudi Journal of Biological Sciences* 25(6): 1128–1134.
- Oliveira, C., Gurak, P., Cladera-Olivera, F. and Marczak, L.D. 2016. Evaluation of physicochemical, technological and morphological characteristics of powdered yellow passion fruit peel. *International Food Research Journal* 23(4): 1653–1662.
- Opara, 2000. Fruit growth measurement and analysis. *Horticultural Reviews* 24:373-431.
- Ortiz, T.A. and Takahashi, L.S.A. 2015. Physical and chemical characteristics of pitaya fruits at physiological maturity. *Genetics and Molecular Research* 14(4): 14422–14439.
- Osorio, S., Scossa, F. and Fernie, A. 2013. Molecular regulation of fruit ripening. *Frontiers in Plant Science* 4: 198.

- Oyedele, O. and Gardner-lubbe, S. 2018. The PLS Biplot: Another graphical tool for multivariate data the PLS biplot. *International Science and Technology Journal of Namibia* 11: 15–27.
- Padmanabhan, P., Cheema, A. and Paliyath, G. 2015. Solanaceous fruits including tomato, eggplant, and peppers. In *Encyclopaedia of Food and Health* 1st edition, ed. B. Caballero, P. Finglas and F. Toldrá. pp. 291-331. United Kingdom: Academic Press.
- Paltrinieri, G. 2014. Handling of Fresh Fruits Vegetables and Root Crops. Food and Agriculture Organization of United Nations. Retrieved 22 September from <http://www.fao.org/3/a-au186e.pdf>.
- Paniagua, C., Pose, S., Morris, V., Kirby, A., Quesada, M. and Mercado, J. 2014. Fruit softening and pectin disassembly: An overview of nanostructural pectin modifications assessed by atomic force microscopy. *Annals of Botany* 114: 1375–1383.
- Parcheta, M. and Swisłocka, R. 2021. Recent developments in effective antioxidants: The structure and antioxidant properties. *Materials* 14: 1984.
- Pareek, S. 2015. Nutritional and biochemical composition of lychee (*Litchi chinensis* Sonn.) Cultivars. In *Nutritional Composition of Fruit Cultivars*. pp. 395-418. United Kingdom: Academic Press.
- Parthasarathy, A., Cross, P., Dobson, R.C., Adams, L., Savka, M. and Hudson, A. 2018. A three-ring circus: metabolism of the three proteogenic aromatic amino acids and their role in the health of plants and animals. *Frontiers in Molecular Biosciences* 5(4): 29.
- Patel, R.K., Singh, A., Prakash, J., Nath, A. and Deka, B.C. 2014. Physico-biochemical changes during fruit growth, development and maturity in passion fruit genotypes. *Indian Journal of Horticulture* 71(4): 486–493.
- Patrick, J., Botha, F. and Birch, R. 2013. Metabolic engineering of sugars and simple sugar derivatives in plants. *Plant Biotechnology Journal* 11(2): 142–156.
- Pattyn, J., Vaughan-Hirsch, J. and Van de Poel, B. 2020. The regulation of ethylene biosynthesis: A complex multilevel control circuitry. *New Phytologist* 229(2): 770–782.
- Paul, O., Julius, B.L., Joseph, B., Grace, K.R. and Collins, I.S. 2017. Non-destructive maturity assessment tools for commercially viable fruits and vegetables in Uganda. *African Journal of Plant Science* 11(6): 220–228.
- Paul, V. and Pandey, R. 2014. Role of internal atmosphere on fruit ripening and storability-A review. *Journal Food Science and Technology* 51(7): 1223–1250.

- Payasi, A. and Sanwal, G.G. 2010. Ripening of climacteric fruits and their control. *Journal of Food Biochemistry* 34(4): 679–710.
- Pereira, L.F., Galvao, R., Kobayashi, A., Cacao, S.M. and Veira, L.G. 2005. Ethylene production and ACC oxidase gene expression during fruit ripening of *Coffea arabica* L. Brazil. *Journal Plant Physiology* 17(3): 1–7.
- Pfeiffer, T., Summerfelt, S. and Watten, B. 2011. Comparative performance of CO₂ measuring methods: Marine aquaculture recirculation system application. *Aquacultural Engineering* 44(1): 1–9.
- Pham, D., Vu, N., Samhaber, W. and Nguyen, M. 2019. Physicochemical characteristics and aroma analysis of passion fruit juice and guava juice concentrated by a novel evaporation concept. *Chemical Engineering Transactions* 75: 43–48.
- Piasai, R., Chalmers, P., Piasai, O. and Khewkhom, N. 2021. Postharvest fungicide dips to control fruit rot of ‘Monthong’ durian (*Durio zibethinus*). *European Journal of Plant Pathology* 160: 325–336.
- Pilkington, S., Montefiori, M., Jameson, P. and Allan, A. 2012. The control of chlorophyll levels in maturing kiwifruit. *Planta* 236(5): 1615–1628.
- Pineapple Research Station, 2021. All about Passion Fruit & Guidelines for 134P Passion fruit Multilocation testing. Retrieved 15 September 2021 from <http://prsvkm.kau.in/announcement/guidelines-134p-passion-fruit-multilocation-testing>.
- Pinzón, I.M., Fisher, G. and Corredor, G. 2007. Determination of the maturity stages of purple passion fruit. *Agronomía Colombiana* 25(1): 83–95.
- Poel, B.V., Bulens, I., Oppermann, Y., Hertog, M.L.A.T., Bart, M., Sauter, M. and Geeraerd, A. 2013. S-adenosyl-L-methionine usage during climacteric ripening of tomato in relation to ethylene and polyamine biosynthesis and transmethylation capacity. *Physiologia Plantarum* 148: 176–188.
- Pongener, A., Sagar, V., Pal, R.K., Asrey, R., Sharma, R.R. and Singh, S.K. 2014. Physiological and quality changes during postharvest ripening of purple passion fruit (*Passiflora edulis* Sims). *Fruits* 69(1): 19–30.
- Pramai, P., Abdul Hamid, N., Mediani, A., Maulidiani, M., Abas, F. and Jiamyangyuen, S. 2018. Metabolite profiling, antioxidant, and α -Glucosidase inhibitory activities of germinated rice: nuclear-magnetic-resonance-based metabolomics study. *Journal of Food and Drug Analysis* 26(1): 47–57.
- Prasad, K., Jacob, S. and Siddiqui, M. 2018. Fruit maturity, harvesting, and quality standards. preharvest modulation of postharvest fruit and vegetable quality. In *Preharvest Modulation of Postharvest Fruit and Vegetable Quality*, ed. M. Siddiqui. pp. 41–69. United Kingdom: Academic Press.

- Prasertsri, P., Booranasuksakul, U., Naravoratham, K. and Trongtosak, P. 2019. Acute effects of passion fruit juice supplementation on cardiac autonomic function and blood glucose in healthy subjects. *Preventive Nutrition and Food Science* 24(3): 245–253.
- Radchanui, C. and Keawvongsri, P. 2017. Pattern and production of durian in Saikhao community, Kokpho District, Pattani Province Radchanui. *International Journal of Agricultural Technology* 13(6): 791–812.
- Radwanski, E. and Last, R. 1995. Tryptophan biosynthesis and metabolism: Biochemical and molecular genetics. *Plant Cell* 7(7): 921–934.
- Rafaela, Q.J.B. and Cazetta, J. 2016. Proline and trehalose in maize seeds germinating under low osmotic potentials. *Revista Brasileira de Engenharia Agrícola e Ambiental* 20(1): 22–28.
- Ramaiya, S., Bujang, J., Zakaria, M. and Saupi, N. 2019. Nutritional, mineral and organic acid composition of passion fruit (*Passiflora* species). *Food Research* 3(3): 231–240.
- Ramaiya, S.D., Bujang, J.S., Zakaria, M.H., King, W.S. and Shaffiq Sahrir, M.A. 2013. Sugars, ascorbic acid, total phenolic content and total antioxidant activity in passion fruit (*Passiflora*) cultivars. *Journal of the Science of Food and Agriculture* 93(5): 1198–1205.
- Ramli, A.N., Manap, N.W., Bhuyar, P. and Azelee, N.I. 2020. Passion fruit (*Passiflora edulis*) peel powder extract and its application towards antibacterial and antioxidant activity on the preserved meat products. *SN Applied Sciences* 2(10): 1–11.
- Ramos, M. and de Toda, F. 2020. Variability in the potential effects of climate change on phenology and on grape composition of Tempranillo in three zones of the Rioja Doca (Spain). *European Journal of Agronomy* 115: 126014.
- Rathor, P., Borza, T., Liu, Y., Qin, Y., Stone, S., Zhang, J., Hui, J.P., Berrue, F., Groisillier, A., Tonon, T., Yurgel, S., Potin, P. and Prithiviraj, B. 2020. Low mannitol concentrations in *Arabidopsis thaliana* expressing *Ectocarpus* genes improve salt tolerance. *Plants* 9(11): 1–18.
- Raza, A., Razzaq, A., Mehmood, S., Zou, X., Zhang, X., Lv, Y. and Xu, J. 2019. Impact of climate change on crops adaptation and strategies to tackle its outcome: A review. *Plants* 8(2): 34.
- Rebaya, A., Belghith, S., Baghdikian, B., Leddet, V., Mabrouki, F., Olivier, E., Cherif, J. and Ayadi, M. 2015. Total phenolic, total flavonoid, tannin content, and antioxidant capacity of *Halimium halimifolium* (Cistaceae). *Journal of Applied Pharmaceutical Science* 5(1): 052–057.

- Reis-Giada, M.D.L. 2013. Food phenolic compounds: Main classes, sources, and their antioxidant power. In *Oxidative stress and chronic degenerative diseases—A role for antioxidants*, ed. J.A.M. Gonzales. pp. 87–112, United Kingdom: IntechOpen.
- Renaudin, J., Deluche, C., Cheniclet, C., Chevalier, C. and Frangne, N. 2017. Cell layer-specific patterns of cell division and cell expansion during fruit set and fruit growth in tomato pericarp. *Journal of Experimental Botany* 68(7): 1613–1623.
- Reyes, F., DeJong, T., Franceschi, P., Tagliavini, M. and Gianelle, D. 2016. Maximum growth potential and periods of resource limitation in apple tree. *Frontiers in Plant Science* 7:233.
- Rienth, M., Torregrosa, L., Sarah, G., Ardisson, M., Brillouet, J. and Romieu, C. 2016. Temperature desynchronizes sugar and organic acid metabolism in ripening grapevine fruits and remodels their transcriptome. *BMC Plant Biology* 16(1): 1–23.
- Rocky, T and Goutam, M. 2017. Production preference and importance of passion fruit (*Passiflora Edulis*): A Review. *Journal of Agricultural Engineering and Food Technology* 4(1): 27-30.
- Rodeo, A., Castro, A. and Esguerra, E. 2018. Postharvest handling of dragon fruit (*Hylocereus* spp.) in the Philippines. Dragon Fruit Regional Network Initiation Workshop: London.
- Rodrigues, C., Carvalho, V., Mapeli, A. and Oliveira, A.B. 2021. Anatomical characterization of *Passiflora cincinnata* Mast. fruit subjected to refrigeration Carolina. *Revista Brasileira de Fruticultura* 43: e-698.
- Rodriguez, C., Bustamante, C.A., Budde, C., Müller, G., Drincovich, M. and Lara, M. 2019. Peach fruit development: A comparative proteomic study between endocarp and mesocarp at very early stages underpins the main differential biochemical processes between these tissues. *Frontiers in Plant Science* 10: 715.
- Rokaya, P., Baral, D., Gautam, D., Shrestha, A. and Paudyal, K. 2016. Effect of altitude and maturity stages on quality attributes of mandarin (*Citrus reticulata* Blanco). *American Journal of Plant Sciences* 7(6): 958–966.
- Romes, N., Wahab, R. and Hamid, M. 2021. The role of bioactive phytoconstituents-loaded nanoemulsions for skin improvement: A review. *Biotechnology & Biotechnological Equipment* 35(1): 712–730.
- Rozhan, 2019. Trends in production, trade and consumption of tropical fruit in Malaysia. Retrieved 25 August 2021 from <https://ap.fftc.org.tw/article/1381>.

- Rubashiny, V. and Haron, N. 2015. Macromorphological and micromorphological studies of four selected *Passiflora* species in Peninsular Malaysia. *Pakistan Journal of Botany* 47(2): 485–492.
- Sadka, A., Shlizerman, L., Kamara, I. and Blumwald, E. 2019. Primary metabolism in citrus fruit as affected by its unique structure. *Frontiers in Plant Science* 10: 1167.
- Salgó, A., Gergely, S. and Juhász, R. 2005. Characterizing the maturation and germination processes in wheat by NIR methods. Proceedings from the 12th International ICC Cereal and Bread Congress. Harrogate, United Kingdom.
- Sampaio, B., Edrada-Ebel, R. and Da Costa, F. 2016. Effect of the environment on the secondary metabolic profile of *Tithonia diversifolia*: A model for environmental metabolomics of plants. *Scientific Reports* 6: 1–11.
- Sara, D., Pott, D., Osorio, S. and Vallarino, J. 2020. Sugar signalling during fruit ripening. *Frontiers in Plant Science* 11: 564917.
- Schmidt, S., Zietz, M., Schreiner, M., Rohn, S., Kroh, L. and Krumbein, A. 2010. Identification of complex, naturally occurring flavonoid glycosides in kale (*Brassica oleracea* var. *sabellica*) by high-performance liquid chromatography diode-array detection/electrospray ionization multi-stage mass spectrometry. *Rapid Communications in Mass Spectrometry* 24(14): 2009–2022.
- Schotsmans, W. C. and Fischer, G. 2011. Passion fruit (*Passiflora edulis* Sims). In *Postharvest Biology and Technology of Tropical and Subtropical Fruits*, ed. E.M. Yahia. pp. 125-142. United Kingdom: Woodhead Publishing
- Sepúlveda, P., Costa, G.M., Aragón, D.M., Ramos, F. and Castellanos, L. 2018. Analysis of vitexin in aqueous extracts and commercial products of Andean *Passiflora* species by UHPLC-DAD. *Journal of Applied Pharmaceutical Science* 8(9): 081–086.
- Shahbani, N., Ramaiya, S. and Saupi, N. 2020. Reproductive biology and fruit setting of *Passiflora quadrangularis* L. (Giant Granadilla) in East Malaysia. *Pertanika Agricultural Science* 43(4): 637–652.
- Shahidi, F. and Zhong, Y. 2015. Measurement of antioxidant activity. *Journal of Functional Foods* 18: 757–781.
- Shameer, S., Vallarino, J., Fernie, A., Ratcliffe, R. and Sweetlove, L. 2020. Flux balance analysis of metabolism during growth by osmotic cell expansion and its application to tomato fruits. *Plant Journal* 103(1): 68–82.
- Shiomi, S., Wamocho, L.S. and Agong, S.G. 1996. Ripening characteristics of purple passion fruit on and off the vine. *Postharvest Biology and Technology* 7(1–2): 161–170.

- Shiu, J., Slaughter, D., Boyden, L. and Barrett, D. 2016. Correlation of descriptive analysis and instrumental puncture testing of watermelon cultivars. *Journal of Food Science* 81(6): S1506–S1514.
- Silva, A.C. and Jorge, N. 2017. Data report bioactive compounds of oils extracted from fruits seeds obtained from agroindustrial waste. *European Journal of Lipid Science and Technology* 19(4): 1–5.
- Silva, W., Silva, G.M., Santana, D., Salvador, A.R., Medeiros, D., Belghith, I., da Silva, N., Cordeiro, M.H. and Misobutsi, G. 2018. Chitosan delays ripening and ROS production in guava (*Psidium guajava* L.) fruit. *Food Chemistry* 242: 232–238.
- Simpson, M. 2010. Diversity and classification of flowering plants: Eudicots. In *Plant Systematics*, ed. M. G. Simpson. pp. 275–448. United Kingdom: Academic Press.
- Singh, V., Chahal, T., Grewal, S. and Gill, P. 2021. Effect of fruit development stages on antioxidant properties and bioactive compounds in peel, pulp and juice of grapefruit varieties. *Journal of Food Measurement and Characterization* 15(3): 2531–2539.
- Singh, V., Weksler, A. and Friedman, H. 2017. Different preclimacteric events in apple cultivars with modified ripening physiology. *Frontiers in Plant Science* 8(9): 1–14.
- Smith, H., Mcausland, L. and Murchie, E. 2017. Don't ignore the green light: Exploring diverse roles in plant processes. *Journal of Experimental Botany* 68(9): 2099–2110.
- Sobolev, A., Brosio, E., Gianferri, R. and Segre, A. 2005. Metabolic profile of lettuce leaves by high-field NMR spectra. *Magnetic Resonance in Chemistry* 43: 625–638.
- Soh, O. 2018. Durian dash: How investors are eyeing the world's most controversial fruit. Retrieved 23 Jun 2021 from <https://www.straitstimes.com/business/durian-dash-how-investors-are-eyeing-the-worlds-most-controversial-fruit>.
- Solovchenko, A. and Schmitz-Eiberger, M. 2003. Significance of skin flavonoids for UV-B-protection in apple fruits. *Journal of Experimental Botany* 54(389): 1977–1984.
- Song, F., Gan, R., Zhang, Y., Xiao, Q., Kuang, L. and Li, H. 2010. Total phenolic contents and antioxidant capacities of selected chinese medicinal plants. *International Journal of Molecular Sciences* 11(6): 2362–2372.
- Srivastava, L. 2002. Seed development and maturation. In *Plant growth and development. Hormones and the environment*, ed. J. Roberts. pp. 431–446. United Kingdom: Academic Press.

- Stein, O. and Granot, D. 2019. An overview of sucrose synthases in plants. *Frontiers in Plant Science* 10: 1–14.
- Strelin, M. and Aizen, M. 2018. The interplay between ovule number, pollination and resources as determinants of seed set in a modular plant. *PeerJ* 6: e5384.
- Suleria, H.A.R., Barrow, C.J. and Dunshea, F.R. 2020. Screening and characterization of phenolic compounds and their antioxidant capacity in different fruit peels. *Foods* 9: 1206.
- Sun, Y., Qian, M., Wu, R., Niu, Q., Teng, Y. and Zhang, D. 2014. Postharvest biology and technology postharvest pigmentation in red Chinese sand pears (*Pyrus pyrifolia* Nakai) in response to optimum light and temperature. *Postharvest Biology and Technology* 91: 64–71.
- Syed, S.A., Zeb, A., Qureshi, W., Malik, A., Tiwana, M., Walsh, K., Amin, M., Alasmery, W. and Alanazi, E. 2021. Mango maturity classification instead of maturity index estimation: A new approach towards handheld NIR spectroscopy. *Infrared Physics and Technology* 115: 103639.
- Taghvaei, M. and Jafari, S. 2015. Application and stability of natural antioxidants in edible oils in order to substitute synthetic additives. *Journal of Food Science and Technology* 52(3): 1272–1282.
- Takac, A., Popovic, V., Glogovac, S., Dokic, V. and Kovac, D. 2015. Effects of fruit maturity stages and seed extraction time on the seed quality of eggplant (*Solanum melongena* L.). *Ratarstvo I Povrtarstvo* 52(1): 7–13.
- Takehiko, S., Ryohei, N., Vladimir, S., Avi, S. and Eduardo, B. 2014. Vacuolar citrate / H⁺ symporter of citrus juice cells. *Planta* 224: 472–480.
- Takshak, S. and Agrawal, S. 2014. Secondary metabolites and phenylpropanoid pathway enzymes as influenced under supplemental ultraviolet-B radiation in *Withania somnifera* Dunal, an indigenous medicinal plant. *Journal of Photochemistry and Photobiology B: Biology* 140: 332–343.
- Tambellini, N., Zaremborg, V., Turner, R. and Weljie, A. 2013. Evaluation of extraction protocols for simultaneous polar and non-polar yeast metabolite analysis using multivariate projection methods. *Metabolites* 3(3): 592–605.
- Tang, N., Deng, W., Hu, N., Chen, N. and Li, Z. 2016. Postharvest biology and technology metabolite and transcriptomic analysis reveals metabolic and regulatory features associated with Powell orange pulp deterioration during room temperature and cold storage. *Postharvest Biology and Technology* 112: 75–86.
- Tao, X., Wu, Q., Li, J., Cai, L., Mao, L., Luo, Z. and Li, L. 2021. Postharvest biology and technology exogenous methyl jasmonate regulates sucrose metabolism in

- tomato during postharvest ripening. *Postharvest Biology and Technology* 181: 111639.
- Tee, Y., Ding, P. and Nor Aini, A. 2012. Determination of optimum harvest maturity and physico-chemical quality of Rastali banana (*Musa AAB Rastali*) during fruit ripening. *Journal of the Science of Food and Agriculture* 92(1): 171–176.
- Tee, Y., Ding, P. and Nor Ani, A. 2011. Physical and cellular structure changes of Rastali banana (*Musa AAB*) during growth and development. *Scientia Horticulturae* 129: 382–389.
- The Insight Partner, 2020. The European market potential for exotic tropical passion fruit concentrate market forecast to 2027-COVID-19 impact and global analysis by source (Organic and conventional) and end use (Infant food, beverages, bakery and snacks, ice cream and yogurt, and others) fruit. Retrieved 25 August 2021 from <https://www.theinsightpartners.com/reports/passion-fruit-concentrate-market>.
- Thomas, B. 2016. Sources and sinks. In *Encyclopedia of Applied Plant Sciences*, ed. B. Thomas, B.G., Murray and D.J. Murphy. pp. 724-734. United Kingdom: Academic Press.
- Tijero, V., Girardi, F. and Botton, A. 2021. Fruit development and primary metabolism in apple. *Agronomy* 11: 1160.
- Tohge, T., Alseekh, S. and Fernie, A.R. 2014. On the regulation and function of secondary metabolism during fruit development and ripening. *Journal of Experimental Botany* 65(16): 4599–4611.
- Torres, G.A.M. 2018. Seed dormancy and germination of two cultivated species of *Passifloraceae*. *Boletín Científico Centro De Museos Museo De Historia Natural* 22(1): 15–27.
- Tran, X., Parks, S., Roach, P., Golding, J. and Nguyen, M. 2016. Effects of maturity on physicochemical properties of Gac fruit (*Momordica cochinchinensis* Spreng.). *Food Science and Nutrition* 4: 305–314.
- Tsegay, Z. 2020. Total titratable acidity and organic acids of wines produced from cactus pear (*Opuntia-ficus-Indica*) fruit and *Lantana camara* (*L. camara*) fruit blended fermentation process employed response surface optimization. *Food Science & Nutrition* 8: 4449–4462.
- Twelfth Malaysian Plan. 2022. Retrieved 2 February 2022 from <https://rmke12.epu.gov.my/bm>.
- Uda, M.N., Gopinath, S.C., Hashim, U., Hakimi, A., Anuar, A., Bakar, M.A. and Parmin, N. 2020. Harumanis mango: Perspectives in disease management and advancement using interdigitated electrodes (IDE) nano- biosensor. *IOP Conf. Series: Materials Science and Engineering* 864: 012180.

- Ukalska, J. and Jastrzębowski, S. 2019. Sigmoid growth curves, a new approach to study the dynamics of the epicotyl emergence of oak. *Folia Forestalia Polonica* 61(1): 30–41.
- Umer, M., Safdar, L., Gebremeskel, H., Zhao, S., Yuan, P., Zhu, H., Kaseb, M., Anees, M., Lu, X., He, N., Gong, C. and Liu, W. 2020. Identification of key gene networks controlling organic acid and sugar metabolism during watermelon fruit development by integrating metabolic phenotypes and gene expression profiles. *Horticulture Research* 7: 193.
- Usenik, V., Stampar, F. and Kastelec, D. 2013. Phytochemicals in fruits of two *Prunus domestica* L. plum cultivars during ripening. *Journal of the Science of Food and Agriculture* 93(3): 681–692.
- Utsunomiya, N. 1992. Effect of temperature on shoot growth, flowering and fruit growth of purple passionfruit (*Passiflora edulis* Sims var. *edulis*). *Scientia Horticulturae* 52: 63–68.
- Vaillant, F., Jeanton, E., Dornier, M., O'Brien, G., Reynes, M. and Decloux, M. 2001. Concentration of passion fruit juice on an industrial pilot scale using osmotic evaporation. *Journal of Food Engineering* 47: 195–202.
- Valenzuela, C., Régulo, J., Londoño, C., Antonio, G., Franco, G., Régulo, J., Valenzuela, C., Antonio, G., Londoño, C., Rojano, B., María, A., Correa, P. and Arias, M. 2014. Physicochemical properties of gulupa fruits (*Passiflora edulis* Sims) during pre and postharvest. *Revista Iberoamericana de Tecnología Postcosecha* 15(1): 15–30.
- Venkatachalam, K. 2016. Postharvest physiology and handling of longkong fruit: A review. *Fruits* 71(5): 289–298.
- Vidović, M., Morina, F., Milić, S., Zechmann, B., Albert, A., Winkler, J. and Veljović, J. 2015. Ultraviolet-B component of sunlight stimulates photosynthesis and flavonoid accumulation in variegated *Plectranthus coleoides* leaves depending on background light. *Plant, Cell and Environment* 38(5): 968–979.
- Villa-Ruano, N., Velásquez-Valle, R., Zepeda-Vallejo, L., Pérez-Hernández, N., Velázquez-Ponce, M., Arcos-Adame, V. and Becerra-Martínez, E. 2018. ¹H NMR-based metabolomic profiling for identification of metabolites in *Capsicum annuum* cv. *mirasol* infected by beet mild curly top virus (BMCTV). *Food Research International* 106: 870–877.
- Wang, Y., Ji, S., Dai, H., Kong, X., Hao, J., Wang, S. and Zhou, X. 2019. Changes in membrane lipid metabolism accompany pitting in blueberry during refrigeration and subsequent storage at room temperature. *Frontiers in Plant Science* 10:829.
- Wijeratnam, S. 2015. Passion Fruit. In *Encyclopedia of Food and Health* (Edition.1), ed. B. Caballero, P. Finglas, and F. Toldrá. pp. 23-234. United Kingdom: Academic Press.

- Willson, M.F. and Whelan, C.J. 1990. The evolution of fruit colour in fleshy-fruited plants. *American Naturalist* 136(6): 790–809.
- Wojdyło, A. and Oszmiański, J. 2020. Antioxidant activity modulated by polyphenol contents in apple and leaves during fruit development and ripening. *Antioxidants* 9: 567.
- Wu, J., Xu, Z., Zhang, Y., Chai, L., Yi, H. and Deng, X. 2014. An integrative analysis of the transcriptome and proteome of the pulp of a spontaneous late-ripening sweet orange mutant and its wild type improves our understanding of fruit ripening in citrus. *Journal of Experimental Botany* 65(6): 1651–1671.
- Wu, X. and Prior, R. 2005. Systematic Identification and characterization of anthocyanins by HPLC-ESI-MS/MS in common foods in the United States: Fruits and Berries. *Journal Agricultural Food Chemistry* 53: 2589–2599.
- Wu, Y., Xu, L., Liu, X., Hasan, K.M., Li, H., Zhou, S., Zhang, Q. and Zhou, Y. 2021. Effect of thermosonication treatment on blueberry juice quality: Total phenolics, flavonoids, anthocyanin, and antioxidant activity. *LWT-Food Science and Technology* 150: 112021.
- Wyatt, L., Strickler, S., Mueller, L. and Mazourek, M. 2016. Comparative analysis of *Cucurbita pepo* metabolism throughout fruit development in acorn squash and oilseed pumpkin. *Horticulture Research* 3: 1–12.
- Xie, X., Chen, C. and Fu, X. 2020. Study on the bioaccessibility of phenolic compounds and bioactivities of passion fruit juices from different regions in vitro digestion. *Journal of Food Processing and Preservation* 45: e15056.
- Xu, X., Chen, Q. and Cai, Y. 2020. Physiological effects of artificial light environment on rice in ecological landscape. *IOP Conf. Series: Earth and Environmental Science* 598: 1–7.
- Yaday, R., Goyal, R. and Dhankar, S. 2014. Post-harvest technology of horticultural crops. Doi: 10.13140/RG.2.2.28507.98089.
- Yang, Z., Zou, X., Li, Z., Huang, X., Zhai, X., Zhang, W., Shi, J. and Tahir, H. 2019. Improved postharvest quality of cold stored blueberry by edible coating based on composite gum arabic/roselle extract. *Food and Bioprocess Technology* 12(9): 1537–1547.
- Yasuko, K., Miyawaki, K., Noji, S. and Tanakashi, A. 2013. Phototropin 2 is involved in blue light-induced anthocyanin accumulation in *Fragaria x ananassa* fruits. *Journal of Plant Research* 126: 847–857.
- Yepes, A., Ochoa-Bautista, D., Murillo-Arango, W., Quintero-Saumeth, J., Bravo, K. and Osorio, E. 2021. Purple passion fruit seeds (*Passiflora edulis* f. *edulis* Sims) as

- a promising source of skin anti-aging agents: Enzymatic, antioxidant and multi-level computational studies. *Arabian Journal of Chemistry* 14: 102905.
- Yoo, H., Widhalm, J., Qian, Y., Maeda, H., Cooper, B., Jannasch, A., Gonda, I., Lewinsohn, E., Rhodes, D. and Dudareva, N. 2013. An alternative pathway contributes to phenylalanine biosynthesis in plants via a cytosolic tyrosine: phenylpyruvate aminotransferase. *Nature Communications* 4:1–11.
- Youn, J., Kim, Y., Na, H., Jung, H., Song, C., Kang, S. and Kim, J. 2019. Antioxidant activity and contents of leaf extracts obtained from *Dendropanax morbifera* LEV are dependent on the collecting season and extraction conditions. *Food Science and Biotechnology* 28(1): 201–207.
- Yu, S., Lo, S. and Ho, T.H. 2015. Source-sink communication: Regulated by hormone, nutrient and stress. *Trends in Plant Science* 20(12): 844–857.
- Yu, W., Peng, F., Wang, W., Liang, J., Xiao, Y. and Yuan, X. 2021. *SnRK1* phosphorylation of *SDH* positively regulates sorbitol metabolism and promotes sugar accumulation in peach fruit. *Tree Physiology* 41(6): 1077–1086.
- Zainal, B., Ding, P., Ismail, I. and Saari, N. 2019b. ¹H NMR metabolomics profiling unveils the compositional changes of hydro-cooled rockmelon (*Cucumis melo* L. *reticulatus* cv glamour) during storage related to in vitro antioxidant activity. *Scientia Horticulturae* 246: 618–633.
- Zainal, B., Ding, P., Safinar, I. and Saari, N. 2019a. Postharvest biology and technology physico-chemical and microstructural characteristics during postharvest storage of hydrocooled rockmelon (*Cucumis melo* L. *reticulatus* cv. Glamour). *Postharvest Biology and Technology* 152: 89–99.
- Zanetti, L.H., Murakami, A., Diaz-Vargas, M., Guerra, A.F., Garcia, A.F., Ospina-Rojas, I., do Nascimento, G. R., dos Santos, T., Pintro, P. and Matumoto, P. 2017. By-product of passion fruit seed (*Passiflora edulis*) in the diet of broilers. *Canadian Journal of Animal Science* 98(1): 109–118.
- Zeng, A., Yan, J., Song, L., Gao, B., Y, Z., Li, J, Liu, H., Hou, X. and Li, Y. 2015. Induction and development of microspore-derived embryos in broccoli 3 white-headed cabbage hybrids microspore culture. *Euphytica*: 261–272.
- Zhang, X., Wei, X., Ali, M., Rizwan, H., Li, B., Li, H., Jia, K., Yang, X., Ma, S., Li, S. and Chen, F. 2021. Changes in the content of organic acids and expression analysis of citric acid accumulation-related genes during fruit development of yellow (*Passiflora edulis* f. *flavicarpa*) and Purple (*Passiflora edulis* f. *edulis*) Passion Fruits. *International Journal of Molecular Sciences* 22: 5765.
- Zhang, Y., Li, P. and Cheng, L. 2010. Developmental changes of carbohydrates, organic acids, amino acids, and phenolic compounds in ‘Honeycrisp’ apple flesh. *Food Chemistry* 123(4): 1013–1018.

- Zhao, J., Li, H., Xi, W., An, W., Niu, L., Cao, Y., Wang, H., Wang, Y. and Yin, Y. 2015. Changes in sugars and organic acids in wolfberry (*Lycium barbarum* L.) fruit during development and maturation. *Food Chemistry* 173: 718–724.
- Zhao, J., Li, Y., Ding, L., Yan, S., Liu, M., Jiang, L., Zhao, W., Wang, Q., Yan, L., Liu, R. and Zhang, X. 2016. Phloem transcriptome signatures underpin the physiological differentiation of the pedicel, stalk and fruit of cucumber (*Cucumis sativus* L.). *Plant and Cell Physiology* 57(1): 19–34.
- Zhao, M. and Li, J. 2020. Molecular events involved in fruitlet abscission in Litchi. *Plants* 9(2): 1–11.
- Zhou, X., Liu, L., Li, Y., Li, K., Liu, X., Zhou, J., Yang, C., Liu, X., Fang, C. and Luo, J. 2020. Integrative metabolomic and transcriptomic analyses reveal metabolic changes and its molecular basis in rice mutants of the strigolactone pathway. *Metabolites* 10(11): 425.
- Zoecklein, B.W., Tech, V., Fugelsang, K.C. and Gump, B.H. 2010. Practical methods of measuring grape quality. In *Managing Wine Quality*, ed. G.R. Andrew. pp. 107–133. United Kingdom: Woodhead Publishing Publisher Limited.
- Zolkeflee, N.K., Ismail, N., Maulidiani, M., Abdul Hamid, N., Ramli, N., Azlan, A. & Abas, F. 2020. Metabolite variations and antioxidant activity of *Muntingia calabura* leaves in response to different drying methods and ethanol ratios elucidated by NMR-based metabolomics. *Phytochemical Analysis* 32(1): 69-83.
- Zucolotto, S., Fagundes, C., Reginatto, H., Ramos, F., Castellanos, L., Duque, C. and Schenkel, P. 2012. Analysis of C-glycosyl flavonoids from South American *Passiflora* species by HPLC-DAD and HPLC-MS. *Phytochemical Analysis* 23: 232–239.