



UNIVERSITI PUTRA MALAYSIA

EVALUATION OF WOUND HEALING PROPERTIES OF *Clinicanthus nutans* (Burm.F.) Lindau (BELALAI GAJAH) AND ITS RELATED BIOCHEMICAL ACTIVITIES ON MALE SPRAGUE-DAWLEY RATS.

SATHIAVANI A/P RAMASAMY

FS 2022 15



EVALUATION OF WOUND HEALING PROPERTIES OF *Clinicanthus nutans* (Burm.F.) Lindau (*BELALAI GAJAH*) AND ITS RELATED BIOCHEMICAL ACTIVITIES ON MALE SPRAGUE-DAWLEY RATS.

By

SATHIAVANI A/P RAMASAMY

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the degree of Master of Science

May 2021

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

EVALUATION OF WOUND HEALING PROPERTIES OF *Clinacanthus nutans* (Burm.F.) Lindau (BELALAI GAJAH) AND ITS RELATED BIOCHEMICAL ACTIVITIES ON MALE SPRAGUE-DAWLEY RATS.

By

SATHIAVANI A/P RAMASAMY

May 2021

Chair: Meenakshii Nallappan, PhD
Faculty: Science

Clinacanthus nutans Lindau (*C. nutans*), also known as Sabah Snake Grass, of the Acanthaceae family has been widely used in traditional medicine and scientific investigations of this plant have shown multitude therapeutic benefits ranging from anticancer, antioxidant, antidiabetic, antimicrobial, immunomodulatory, anti-inflammatory and analgesic activities. *C. nutans* has also shown promising properties in preliminary wound healing rat models by the integrity of epidermal tissue at the lesion sites. The objective of this study is to determine the most potent solvent extracts of *C. nutans* to develop a topical herbal cream that will be efficacious on the wound lesions in Sprague Dawley rat models along the lines of biochemical and histological assessments. The plant leaves were subjected to hot solvent extraction and crude extracts were derived from ethanol, methanol, chloroform and water. The extracts were subjected to both phytochemicals assessments which comprised of phenols, flavonoids and tannins and antioxidant activity assessments by (2,2-diphenyl-1-picrylhydrazyl) DPPH and (Azino-bis-3-ethylbenzthiazoline-6-sulphonic acid) ABTS assays prior to conducting the *in vivo* excision wound experiment in rat models. Methanol extract demonstrated the highest phenolic, flavonoid and tannin content of (12.96 ± 1.91 mg GAE/gm), (10.06 ± 1.68 mg QUE/gm) and (7.15 ± 0.64 mg TAE/gm). It was found that, methanol extract could scavenge DPPH with the maximum scavenging activity of $56.37 \pm 0.70\%$ and with an IC₅₀ of 222.69 ± 0.29 μ g/ml and for ABTS, aqueous extract showed highest scavenging activity of $57.50 \pm 0.52\%$ with an IC₅₀ of 149.45 ± 0.24 μ g/ml respectively. The rats were arranged into 6 groups, each group consist of 6 rats except the negative control group which consist of 3 rats. Group 1 (control), group 2 (standard / Dettol antiseptic cream), group 3 was aqueous extract of *C. nutans* with dose 1 (12.5 mg/ml of aqueous extract of *C. nutans*), dose 2 (25.0 mg/ml of aqueous extract of *C. nutans*) and dose 3 (50.0 mg/ml of aqueous extract of *C.*

nutans). Group 4 was methanolic extract of *C. nutans* with dose 1 (12.5 mg/ml methanolic extract of *C. nutans*), dose 2 (25.0 mg/ml methanolic extract of *C. nutans*) and dose 3 (50.0 mg/ml methanolic extract of *C. nutans*) group 5 (cream base), group 6 (herbal cream). The most effective dosages were 12.5 mg/ml of methanol extract and 50.0 mg/ml of aqueous extract, combined and produced a topical herbal cream which was tested *in vivo* using rat models again. Subsequently, the mechanism of action of the *C. nutans* topical herbal cream was assessed using biochemical parameters like hydroxyproline, collagen and hexosamine and histopathological changes. Topical herbal cream by incorporating methanol extract 12.5 mg/ml and aqueous extract 50.0 mg/ml showed a significant wound repair and skin regeneration about (99.5 ± 0.22%) wound contraction at Day 15 compared to control groups. Biochemical parameters like hydroxyproline (64.11 ± 0.01 mg/g), hexosamine (31.37 ± 1.79 mg/g) and collagen (478.24 ± 1.12 mg/g) turnover were found to be significantly increased ($p < 0.05$) in test topical herbal cream treated animals as compared to controls. All statistical analysis was performed using SPSS statistical version 20.0 software package and *P*-value was set as < 0.05 for all analysis. Histopathological findings revealed that the cream confirmed that the epidermis layers with complete re-epithelialization and dermis show dense bundles of collagen fibers. In conclusion, all these investigations indicate significant wound healing potential of *C. nutans* topical herbal cream in Sprague Dawley excision wound models.

Keywords: *C. nutans*; wound healing; rat; biochemical changes; antioxidant; excision wound model;

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

**PENILAIAN AKTIVITI PENYEMBUHAN LUKA *Clinacanthus nutans*
(Burm.F.) Lindau (BELALAI GAJAH) DAN AKTIVITI BIOKIMIA YANG
BERKAITAN PADA TIKUS SPRAGUE-DAWLEY LELAKI.**

Oleh

SATHIAVANI A/P RAMASAMY

Mei 2021

Pengerusi: Meenakshii Nallappan, PhD
Fakulti: Sains

Clinacanthus nutans Lindau (*C. nutans*), yang juga dikenali sebagai belalai gajah, dari keluarga Acanthaceae yang banyak digunakan dalam perubatan tradisional dan kajian saintifik tumbuhan ini telah menunjukkan banyak faedah terapeutik yang terdiri daripada antikanser, antioksidan, antidiabetik, antimikrobial, imunomodulator dan analgesik. *C. nutans* juga menunjukkan ciri-ciri yang menjanjikan dalam model tikus penyembuhan luka oleh integriti tisu epidermis pada luka. Objektif kajian ini adalah untuk menentukan ekstrak pelarut yang paling efektif dari *C. nutans* untuk merumuskan salap herba yang akan berefektif pada luka pada model tikus Sprague Dawley dengan mekanisme biokimia. Daun tumbuhan tertakluk kepada pengekstrakan pelarut panas dan ekstrak mentah diperolehi daripada etanol, metanol, kloroform dan air. Ekstrak tersebut tertakluk kepada penilaian fitokimia iaitu fenol, flavonoid dan tanin dan penilaian aktiviti antioksidan oleh (2,2-diphenyl-1-picrylhydrazyl) DPPH dan (Asid Azino-bis-3-ethylbenzothiazoline-6-sulfonat) sebelum melakukan eksperimen luka *in vivo* pada tikus. Ekstrak metanol menunjukkan kandungan fenolik, flavonoid dan tanin (12.96 ± 1.91 mg GAE/gm), (10.06 ± 1.68 mg QUE/gm) dan (7.15 ± 0.64 mg TAE/gm). Kajian ini menunjukkan bahawa ekstrak metanol dengan DPPH menunjukkan aktiviti maksimum $56.37 \pm 0.70\%$ dan dengan IC50 222.69 ± 0.29 $\mu\text{g} / \text{ml}$ dan ABTS, ekstrak air menunjukkan aktiviti penimbunan tertinggi $57.50 \pm 0.52\%$ dengan IC50 149.45 ± 0.24 $\mu\text{g} / \text{ml}$. Dos yang paling berkesan adalah 12.5 mg/kg ekstrak metanol dan 50.0 mg/kg ekstrak air, digabungkan dan menghasilkan salap herba yang diuji di *in vivo* menggunakan model tikus sekali lagi. Seterusnya, mekanisme tindakan salap herba *C. nutans* dinilai dengan menggunakan parameter biokimia seperti hidroksiprolin, kolagen dan heksosamine dan perubahan histopatologi. Perumusan herba dengan menggabungkan ekstrak metanol 12.5 mg/ml dan ekstrak air 50.0 mg/ml menunjukkan pembedahan luka yang signifikan dan penyembuhan kulit ($99.5 \pm 0.22\%$) pengecutan luka pada 15 hari berbanding kontrol. Parameter biokimia

seperti hidrokisprolin (64.11 ± 0.01 mg/g), heksosamine (31.37 ± 1.79 mg/g) dan kolagen (478.24 ± 1.12 mg/g) didapati meningkat dengan ketara ($p < 0.05$) berbanding dengan kawalan. Semua analisis statistik dilaksanakan menggunakan pakej perisian versi statistik SPSS 20.0 dan nilai P ditetapkan sebagai < 0.05 untuk semua analisis. Kesimpulan, kajian ini mendapati menunjukkan potensi penyembuhan luka dari salap herba *C. nutans* dalam model luka Sprague Dawley.

Kata kunci: *C.nutans*; penyembuhan luka; tikus; perubahan boikimia; antioksidan; model luka eksisi;



ACKNOWLEDGEMENTS

I praise to the almighty GOD for giving me the strength and patience throughout my research work and showers of blessings to complete this research successfully. I have taken actions in this project. Nevertheless, it would not have been feasible without the kind of support and aid of many persons.

I would like to express my genuine thanks to all of them. I am greatly in debt to my supervisor Dr. Meenakshii Nallappan for her guidance and continual supervision, advise, encouragement as well as for providing relevant information regarding the project and also for her support in successfully completing the study. Her dynamisms vision, sincerity and motivation have deeply inspired me. It was a privilege and honor to work and study under her guidance. Furthermore, thank you to my co-supervisors, Dr. Siti Farah Tohid and Prof. Shamarina bt Shohaimi who taught me the methodology to carry out the research works.

I am extremely grateful and would like to convey my special appreciation and thanks towards my parents and siblings who became the heartbeat of this study and for their love, prayers, caring, understanding, support, motivation and sacrifices throughout this study and educating me for my future. I would like to extend my sincere gratitude to my cousins as well who gave continuous encouragement and support.

Also, I express my thanks to my good friends and laboratory mates in developing the study and people who have voluntarily helped me out with their capabilities and also those who helped directly and indirectly to complete this research.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Meenakshii a/p Nallappan, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Siti Farah binti Md Tohid, PhD

Senior Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Shamarina binti Shohaimi, PhD

Associate Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 8 September 2022

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research and the writing of this thesis were done under our supervision;
- supervisory responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2015-2016) are adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: Dr. Meenakshii A/P Nallappan

Signature: _____
Name of Member of
Supervisory
Committee: Dr. Siti Farah Binti Md Tohid

Signature: _____
Name of Member of
Supervisory
Committee: Prof. Madya Dr. Shamarina Bt
Shohaimi

TABLE OF CONTENTS

		Page
ABSTRACT		i
ABSTRAK		iii
ACKNOWLEDGEMENTS		v
APPROVAL		vi
DECLARATION		viii
LIST OF TABLES		xiii
LIST OF FIGURES		xiv
LIST OF ABBREVIATIONS		xv
CHAPTER		
1	INTRODUCTION	1
	1.1 Backgrounds	1
	1.2 Problem statement	2
	1.3 Hypothesis	3
	1.4 Objectives	3
	1.4.1 General objective	3
	1.4.2 Specific objective	3
2	LITERATURE REVIEW	4
	2.1 <i>Clinacanthus nutans</i> (Burm. f.) Lindau	4
	2.1.1 Vernacular names	4
	2.1.2 Taxonomy of <i>C. nutans</i>	4
	2.1.3 Geographical distribution	4
	2.1.4 Botany of <i>C. nutans</i>	4
	2.2 Traditional medicinal uses of <i>C. nutans</i>	5
	2.3 Pharmacological activities	6
	2.3.1 Antioxidant activity	6
	2.3.2 Anti-inflammatory activity and immune-modulatory effects	7
	2.3.3 Cytotoxic activity	8
	2.3.4 Antitumorigenic activity	8
	2.3.5 Antimicrobial activity	8
	2.3.6 Antiviral activity	9
	2.3.7 Antinociceptive	10
	2.3.8 Antidiabetic effects	10
	2.3.9 Toxicity	11
	2.4 The Skin	12
	2.4.1 Wound healing process	12
	2.5 1,1-diphenyl-2-picrylhydrazyl radical (DPPH) and 3.1 2,2'-Azino-bis (3-Ethylbenzothiazoline-6-Sulfonic Acid) (ABTS) Assays	14
	2.6 Traditional medicine and Modern medicine	14
	2.6.1 Potential of traditional medicine in wound healing	15

3	MATERIALS AND METHODS / METHODOLOGY	17
3.1	Methods	17
3.1.1	Animals	17
3.1.2	Collection of authentications of plant materials	17
3.1.3	<i>C.nutans</i> extraction	17
3.2	Determination of total flavonoids, phenols and tannins	18
3.2.1	Total phenolic content	18
3.2.2	Total flavonoid content	18
3.2.3	Total tannin content	18
3.3	Determination of antioxidant properties	19
3.3.1	DPPH assay	19
3.3.2	ABTS assay	19
3.4	Excision wound model	19
3.4.1	Preparation of herbal ointment	21
3.4.2	Quality control parameters of formulation	22
3.4.3	Biochemical Parameters	23
3.4.3.1	Estimation of Hydroxyproline	23
3.4.3.2	Estimation of Collagen	23
3.4.3.3	Estimation of Hexosamine	23
3.4.4	Histopathology	24
3.5	Statistical Analysis	24
4	RESULTS AND DISCUSSION	25
4.1	Hot extraction of <i>C.nutans</i> leaves in various solvents via Soxhlet apparatus	25
4.2	Quantitative phytochemical analysis of <i>C.nutans</i>	26
4.2.1	Total phenolic content of <i>C.nutans</i>	26
4.2.2	Total flavonoid content of <i>C.nutans</i>	27
4.2.3	Total tannin content of <i>C.nutans</i>	29
4.3	Antioxidant properties of <i>C.nutans</i>	32
4.3.1	DPPH Free Radical Scavenging Activity	32
4.3.2	ABTS Radical Scavenging Activity	33
4.4	Rat excision wound model	35
4.4.1	Efficacy testing on <i>C.nutans</i> extracts	35
4.4.2	Herbal formulation <i>in vivo</i> rat wound healing models	42
4.5	Quality control formulation	43
4.6	Biochemistry of wound healing	43
4.6.1	Hydroxyproline	43
4.6.2	Collagen	44
4.6.3	Hexosamine	44
4.7	Histopathological observation	45

5	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	48
5.1	Conclusions	48
5.2	Future recommendation	49
	REFERENCES	50
	APPENDICES	61
	BIODATA OF STUDENT	74



LIST OF TABLES

Table		Page
3.1	Treatment group of excision wound model	20
4.1	Extraction yield of <i>C.nutans</i> with respect to different solvents	25
4..2	Phytochemical analysis of <i>C. nutans</i> at 1.5mg/ml and 2.5 mg/ml (n=3)	31
4.3	Antioxidant activity of <i>C. nutans</i> solvent extracts	34
4.4	Wound contraction of efficacy testing of <i>C.nutans</i> methanol and water extracts	40
4.5	Percentage wound contraction between control groups and herbal formulation group	43
4.6	Biochemical parameters between control group and herbal formulation group	45

LIST OF FIGURES

Figure		Page
2.1	<i>Clinacanthus nutans</i> (Burm.F.) Lindau leaves	5
2.2	Skin structure	12
2.3	Stages of wound healing	13
3.1	Counting squares on grid tracing	21
3.2	A 100g of <i>C. nutans</i> ointment	22
4.1	Control and positive groups on excision wound model	36
4.2	Efficacy testing of <i>C. nutans</i> methanol extract	37
4.3	Efficacy testing of <i>C. nutans</i> water extract.	38
4.4	Wound contraction between control groups and herbal formulation group	42
4.5	(A) Hematoxylin and eosin (H&E) and (B) Masson trichrome (x40) of negative control group tissue	46
4.6	(A) Hematoxylin and eosin (H&E) and (B) Masson trichrome (x40) of positive control group tissue	46
4.7	(A) Hematoxylin and eosin (H&E) and (B) Masson trichrome (x40) of herbal formulation group tissue	47

LIST OF ABBREVIATIONS

ABTS	2,2'-azino-bis-3-ethylbenzothiazoline-6-sulphonic acid
AlCl ₃	Aluminum chloride
ANOVA	Analysis of Variance
DNA	Deoxyribonucleic acid
DMSO	Dimethyl sulfoxide
DPPH	1,1- diphenyl-2- picrylhydrazyl
ECM	Extracellular matrix
GAE	Gallic acid equivalent
HGF	Human gingival fibroblast
IC ₅₀	Inhibitory concentration at 50%
MPO	Myeloperoxidase
NaNO ₃	Sodium nitrate
NaOH	Sodium hydroxide
PBMC	Peripheral blood mononuclear cells
QE	Quercetin equivalent
ROS	Reactive oxygen species
SD	Standard deviation
Sdn Bhd	Sendrian Berhad
SPSS	Statistical Package of Social Sciences
TAE	Tannic acid equivalent
VZV	Varicella-zoster
WHO	World Health Organization
<	Less than
%	Percentage

&	And
±	Plus and/or minus
µg/ml	Microgram per millilitre
µl	Microliter
g	Gram
mg	Milligram
GAE/gm	Milligram gallic acid equivalent per gram
QE/gm	Milligram quercetin equivalent per gram
TAE/gm	Milligram tannic acid equivalent per gram
mg/ml	Milligram per milliliter
mg/kg	Milligram per kilogram
ml	milliliter
mM	Millimolar
nm	nanometer
°C	Degree Celsius
®	Registered trademark

CHAPTER 1

INTRODUCTION

1.1 Background

Clinacanthus nutans (Burm. f) Lindau, otherwise called Sabah Snake Grass, from the family of Acanthaceae is a notable therapeutic plant in Thai folklore medicinal practices (Yong et al., 2013). It is known that the fresh leaves of *C. nutans* can be taken as unrefined vegetable or mixed in with other fresh squeezes, for instance, crushed apple and sugarcane press, or green tea, and filled in as restoring drinks. The dried leaves soak up high temperature water and prepared as tea concoctions (Shim et al., 2013). *C. nutans* is medicinal plant traditionally used in treating skin abnormalities like skin rashes, snakebite and burns (Aslam et al., 2015). Additionally, this plant has been utilized traditionally to treat malignancy in Malaysia. Nevertheless, it lacks verifiable studies backing the statements (Roosita et al., 2008; Shim et al., 2013).

Scientific studies on *C. nutans* have its potential along the lines of chemotherapeutics and chemoprevention, antidiabetic, antimicrobial, immunomodulatory, wound healing, analgesics and most as of late for the treatment and anticipation of malignant growth infections (P'ng et al., 2012); (Mustapa et al., 2015). Scientific studies evinced *C. nutans* to have an extensive antimicrobial and mitigating action towards genital herpes and varicella-zoster contamination (VZV) bruises analyzed in immunocompromised patients and effectively retards of unfavorable reactions due to insect bites (Sakdarat et al., 2009). *C. nutans* has likewise been for the treatment of hepatitis, skin-rashes and dermatitis (Mustapa et al., 2015).

Observation of all illnesses is the primary distinction between contemporary medicine and ancient medicine. Modern medicine heals illnesses with ease, yet it does not address the underlying causes. It also has the negative effect of causing the body of the living person to develop additional illnesses. Diseases may take a long time to heal with traditional medicine, but the core cause of the illness is destroyed (Fauzi et al., 2013). Thus, evidence-based medicine may be adopted by the whole medical sector, allowing for the reduction of adverse effects and the maximization of benefits from scientific study for patient care (More, 2016).

1.2 Problem Statement

Bioactive chemicals found in medicinal plants have been utilized to maintain human health, including prevention, diagnosis, and improvement or treatment of a wide range of human physical diseases for thousands of years (Zainol et al., 2019). Moderately, about eighty percent of resident's dependent on Asia, Africa and the Middle East upon customary prescriptions. There are 350000 higher plant species in nature, and around 80000 contain therapeutic qualities, according to recent reports. Bioactive chemicals and biological activities of just 5000 of the plants have been thoroughly investigated (Dar et al., 2017; Yuan et al., 2016). Globally, medical plants-derived medicines are popular since they are less expensive and have fewer negative effects (Marušek et al., 2010). Complex and lengthy treatments are causing an increased burden on healthcare expenses (Nuutila et al., 2014). *C. nutans* has been generally utilized in Asia for quite a long time for its restorative profile (Fong, 2015). Besides that, *C. nutans* improves the physiological appearance of wound. It will be cost effective since the number of healing days are lesser compared to modern methods (Ramasamy, 2016). Preliminary studies indicate detailed research should be done to show the properties of wound healing with *C. nutans* solvent extracts (Ramasamy, 2016).

Various customary restorative utilizes for *C. nutans* have appeared, yet clinical and scientific information bolster just a portion of these discoveries. Basically, the leaves of *C. nutans* exclusively utilized in customary prescriptions (Zulkipli et al., 2017). The use of this medicinal plant to treat Herpes virus contaminations is upheld by the aftereffects of various scientific studies and clinical preliminaries (Lipipun et al., 2011, Kunsorn et al., 2013). According to Yong et al. (2013), *C. nutans* extricates are cancer prevention agents with antiproliferative efficacies on a range of human malignant growth cell lines explicitly HepG2, IMR32, NCL-H23, SNU-1, Hela, LS-174T, K562, Raji, and IMR32. It was uncovered that the chloroform concentrate of this plant leaves have the majority basic constituents that equipped for rummaging free radicals and hinders the development of refined malignant growth cells. Notable inhibition on the generation of superoxide anion and elastase discharge by initiated neutrophils by the 80% ethanol concentrate of aerial part of *C. nutans*. The restraint was delivered by 10 mg/mL ethanolic concentrate of *C. nutans* at 68.33% (Tu et al., 2014). Arullappan et al. (2014), demonstrated that the fraction 7 with most noteworthy enemy of microbial action, with minimum inhibitory concentration (MIC) of 1.39mg/ml against the *Bacillus cereus*, *Escherichia coli*, *Salmonella enterica Typhium* and *Candida albicans*.

Anti-inflammatory activity revealed with significantly reduced oedema after 3 hours in Carrageenan-induced oedema by extract, in a portion subordinate way (Wanikiat et al., 2008). Preliminary *in vivo* screening of *C. nutans* aqueous extracts have shown significant wound healing in mice models (Ramasamy, 2016). Since radical scavenging activity is also an indicator for wound healing

activity potencies, the investigator seeks to further study the efficacies of *C. nutans* solvent concentrates for wound recuperating properties

1.3 Hypothesis

It is hypothesized that the herbal ointment formulation of *C. nutans* to possess phytochemicals, wound healing properties and as well as exhibiting antioxidants properties.

1.4 Objective

1.4.1 General objective

To investigate the efficacy of *C. nutans* extracts and formulated herbal ointment on wound healing employing rat excision wound model and the underlying biochemical changes.

1.4.2 Specific Objective

- 1) To determine total flavonoids (TF), total phenolic compounds (TPC) and tannins (TTC) and antioxidant properties using DPPH and ABTS assays of *C. nutans* extracts.
- 2) To determine the efficacy of the *C. nutans* extracts on wound healing in excision wound of Sprague-Dawley rats
- 3) To evaluate the biochemical parameters with regards to hydroxyproline, collagen and hexosamine content on excision wounds in Sprague-Dawley rats between control groups using antiseptic Dettol cream and treatment group employing *C. nutans* herbal formulation.
- 4) To compare and characterize the histopathological changes of skin restoration between herbal ointment formulation of *C. nutans* and control groups in wound of Sprague-Dawley rats.

REFERENCES

- Abdul Rahim, M. H., Zakaria, Z. A., Sani, M. H., Omar, M. H., Yakob, Y., Cheema, M. S., Kadir, A. A. (2016). Methanolic Extract of *Clinacanthus nutans* Exerts Antinociceptive Activity via the Opioid/Nitric Oxide-Mediated, but cGMP-Independent, Pathways. *Evidence-Based Complementary and Alternative Medicine*, 2016, 1-11. doi:10.1155/2016/1494981
- Agyare, C., Boakye, Y. D., Bekoe, E. O., Hensel, A., Dapaah, S. O., & Appiah, T. (2016). Review: African medicinal plants with wound healing properties. *Journal of Ethnopharmacology*, 177, 85-100. doi:10.1016/j.jep.2015.11.008
- Ajuru, M. G., Williams, L. F., & Ajuru, G. (2017). Qualitative and quantitative phytochemical screening of some plants used in ethnomedicine in the Niger Delta region of Nigeria. *Journal of Food and Nutrition Sciences*, 5(5), 198-205.
- Akiyama, H., Fujii, K., Yamasaki, O., Oono, T., & Iwatsuki, K. (2001). Antibacterial action of several tannins against *Staphylococcus aureus*. *Journal of antimicrobial chemotherapy*, 48(4), 487-491.
- Alam, A., Ferdosh, S., Ghafoor, K., Hakim, A., Juraimi, A. S., Khatib, A., & Sarker, Z. I. (2016). *Clinacanthus nutans*: A review of the medicinal uses, pharmacology and phytochemistry. *Asian Pacific Journal of Tropical Medicine*, 9(4), 402-409. doi: 10.1016/j.apjtm.2016.03.011
- Arullappan, S., Rajamanickam, P., Thevar, N., & Kodimani, C. (2014). In Vitro Screening of Cytotoxic, Antimicrobial and Antioxidant Activities of *Clinacanthus nutans* (Acanthaceae) leaf extracts. *Tropical Journal of Pharmaceutical Research*, 13(9), 1455. doi:10.4314/tjpr.v13i9.11
- Aslam, M. S., Ahmad, M. S., Mamat, A. S., Ahmad, M. Z., & Salam, F. (2016). Antioxidant and wound healing activity of polyherbal fractions of *Clinacanthus nutans* and *Elephantopus scaber*. *Evidence-based Complementary and Alternative Medicine*, 2016.
- Aslam, M. S., Ahmad, M. S., & MAMAT, A. S. (2015). A review on phytochemical constituents and pharmacological activities of *Clinacanthus nutans*. *world*, 2, 4.
- Altromin International. (2019, October 31). Retrieved August 3, 2021, from altromin.com website: https://altromin.com/products/standarddiets/rats/1310#produkt_1314
- Azlim Almey, A. A., Ahmed Jalal Khan, C., Syed Zahir, I., Mustapha Suleiman, K., Aisyah, M. R., & Kamarul Rahim, K. (2010). Total phenolic content and primary antioxidant activity of methanolic and ethanolic extracts of aromatic plants' leaves. *International Food Research Journal*, 17(4).

- Barek, M. L., Hasmadi, M., Zaleha, A. Z., & Fadzelly, A. M. (2015). Effect of different drying methods on phytochemicals and antioxidant properties of unfermented and fermented teas from Sabah Snake Grass (*Clinacanthus nutans* Lind.) leaves. *International Food Research Journal*, 22(2), 661.
- Bowden, L. G., Byrne, H. M., Maini, P. K., & Moulton, D. E. (2016). A morphoelastic model for dermal wound closure. *Biomechanics and modeling in mechanobiology*, 15(3), 663-681.
- Chandran, P. K., & Kuttan, R. (2008). Effect of *Calendula officinalis* flower extract on acute phase proteins, antioxidant defense mechanism and granuloma formation during thermal burns. *Journal of clinical biochemistry and nutrition*, 43(2), 58-64.
- Charuwichitratana, S., Wongrattanapasson, N., Timpatanapong, P., & Bunjob, M. (1996). Herpes zoster: treatment with *Clinacanthus nutans* cream. *International journal of dermatology*, 35(9), 665-666.
- Siew, C. K., & Ch'ng, Y. Y. (2018). Effects of Solvent System, Drying and Storage on the Total Phenolic Content and Antioxidant Activities of *Clinacanthus nutans* Lindau (Sabah Snake Grass). *Pertanika Journal of Tropical Agricultural Science*, 41(4)
- Cheeptham, N., & Towers, G. H. N. (2002). Light-mediated activities of some Thai medicinal plant teas. *Fitoterapia*, 73(7-8), 651-662.
- Chelyn, J. L., Omar, M. H., Mohd Yousof, N. S., Ranggasamy, R., Wasiman, M. I., & Zakiah, I. (2014). Analysis of Flavone C-Glycoside in the Leaves of *Clinacanthus nutans* (Burm. f.) Lindau by HPTLC and HPLC- UV/DAD. *The Scientific World Journal*, 1-6.
- Chomnawang, M. T., Surassmo, S., Wongsariya, K., & Bunyapraphatsara, N. (2009). Antibacterial activity of Thai medicinal plants against methicillin-resistant *Staphylococcus aureus*. *Fitoterapia*, 80(2), 102-104.
- Christodouleas, D., Fotakis, C., Nikokavoura, A., Papadopoulos, K., & Calokerinos, A. (2014). Modified DPPH and ABTS Assays to Assess the Antioxidant Profile of Untreated Oils. *Food Analytical Methods*, 8(5), 1294-1302. doi: 10.1007/s12161-014-0005-6
- Coger, V., Million, N., Rehbock, C., Sures, B., Nachev, M., Barcikowski, S., ... & Vogt, P. M. (2019). Tissue concentrations of zinc, iron, copper, and magnesium during the phases of full thickness wound healing in a rodent model. *Biological trace element research*, 191(1), 167-176.
- Dar, R. A., Shahnawaz, M., & Qazi, P. H. (2017). General overview of medicinal plants: A review. *The Journal of Phytopharmacology*, 6(6), 349-351.
- Datta, H., Mitra, S., & Patwardhan, B. (2011). Wound Healing Activity of Topical Application Forms Based on Ayurveda. *Evidence-Based Complementary And Alternative Medicine*, 2011, 1-10. doi: 10.1093/ecam/nep015

- Devasagayam, T. P. A., Tilak, J. C., Bloor, K. K., Sane, K. S., Ghaskadbi, S. S., & Lele, R. D. (2004). Free radicals and antioxidants in human health: current status and future prospects. *Japi*, 52(794804), 4.
- Dorai, A. A. (2012). Wound care with traditional, complementary and alternative medicine. *Indian Journal of plastic surgery*, 45(02), 418-424.
- Dorsett-Martin, W. A. (2004). Rat models of skin wound healing: a review. *Wound Repair and Regeneration*, 12(6), 591-599.
- Dunn, G., Koebel, C., & Schreiber, R. (2006). Interferons, immunity and cancer immunoediting. *Nature Reviews Immunology*, 6:836-848.
- Dwivedi, D., Dwivedi, M., Malviya, S., & Singh, V. (2017). Evaluation of wound healing, anti-microbial and antioxidant potential of *Pongamia pinnata* in wistar rats. *Journal Of Traditional And Complementary Medicine*, 7(1), 79-85. doi: 10.1016/j.jtcme.2015.12.002
- Ene, I. (2020, January 20). What Happens During the Stages of Wound Healing? *MBBCH Health Encyclopedia*. <https://mbbch.com/health/stages-of-wound-healing/>
- Farsi, E., Esmaili, K., Shafaei, A., Khaniabadi, P. M., Hindi, B. A., Ahamed, M. B., Majid, A. S. (2016). Mutagenicity and preclinical safety assessment of the aqueous extract of *Clinacanthus nutans* leaves. *Drug and Chemical Toxicology*, 39(4), 461-473. doi:10.3109/01480545.2016.1157810
- Fauzi, F. M., Koutsoukas, A., Lowe, R., Joshi, K., Fan, T. P., Glen, R. C., & Bender, A. (2013). Linking Ayurveda and Western medicine by integrative analysis. *Journal of Ayurveda and integrative medicine*, 4(2), 117.
- Fong, S. Y. (2015). Genetic, phytochemical and bioactivity studies of *Clinacanthus nutans* (Burm. f.) Lindau (Acanthaceae). (PhD thesis, RMIT University). Retrieved from <https://researchbank.rmit.edu.au/eserv/rmit:161489/Fong.pdf>
- Formagio, A., Volobuff, C., Santiago, M., Cardoso, C., Vieira, M., & Pereira, Z. (2014). Evaluation of Antioxidant Activity, Total Flavonoids, Tannins and Phenolic Compounds in *Psychotria* Leaf Extracts. *Antioxidants*, 3:745-757.
- Geethalakshmi, R., Sakravarthi, C., Kritika, T., Arul Kirubakaran, M., & Sarada, D. V. L. (2013). Evaluation of antioxidant and wound healing potentials of *Sphaeranthus amaranthoides* Burm. f. *BioMed research international*, 2013.
- Hajiaghaalipour, F., Kanthimathi, M. S., Abdulla, M. A., & Sanusi, J. (2013). The effect of *Camellia sinensis* on wound healing potential in an animal model. *Evidence-Based Complementary and Alternative Medicine*, 2013.

- Healthcare. (2018, March 28). Modern Medicine vs. Traditional Medicine (A Breakdown). Retrieved from Automated Future website: <https://theautomatedfuture.com/modern-medicine-vs-traditional-medicine-a-breakdown/>
- Huang, D., Guo, W., Gao, J., Chen, J., & Olatunji, J. (2015). Clinacanthus nutans (Burm. f.) Lindau Ethanol Extract Inhibits Hepatoma in Mice through Upregulation of the Immune Response. *Molecules*, 20(9), 17405-17428. doi:10.3390/molecules200917405
- Huang, D., Ou, B., & Prior, R. L. (2005). The chemistry behind antioxidant capacity assays. *Journal of agricultural and food chemistry*, 53(6), 1841-1856.
- Iqbal, Z., Kamran, Z., Sultan, J. I., Ali, A., Ahmad, S., Shahzad, M. I., & Sohail, M. U. (2015). Replacement effect of vitamin E with grape polyphenols on antioxidant status, immune, and organs histopathological responses in broilers from 1-to 35-d age. *Journal of Applied Poultry Research*, 24(2), 127-134.
- James, W. D. 1., Berger, T. G., Elston, D. M., & Odom, R. B. (2006). *Andrews' diseases of the skin: Clinical dermatology* (10th ed.). Philadelphia: Saunders Elsevier.
- Jayvasu, C., Dechatiwongse, T., & Balachandra, K. (2013). The virucidal activity of clinacanthus nutans lindau extracts against herpes simplex virus type-2: an in vitro. *Bull Dep Med Sc*, 34(4), 153-8.
- Kanitakis, J. (2002). Anatomy, histology and immunohistochemistry of normal human skin. *European journal of dermatology*, 12(4), 390-401.
- Khoo, L. W., Mediani, A., Zolkeflee, N. K. Z., Leong, S. W., Ismail, I. S., Khatib, A., ... & Abas, F. (2015). Phytochemical diversity of Clinacanthus nutans extracts and their bioactivity correlations elucidated by NMR based metabolomics. *Phytochemistry Letters*, 14, 123-133.
- Kong, H. S., Musa, K. H., Kasim, Z. M., & Sani, N. A. (2019). Qualitative and Quantitative Phytochemical Analysis and Antioxidant Properties of Leaves and Stems of Clinacanthus nutans (Burm. f.) Lindau from Two Herbal Farms of Negeri Sembilan, Malaysia. *ASM Science Journal*, 12.
- Krishnaiah, D., Sarbatly, R., & Nithyanandam, R. (2011). A review of the antioxidant potential of medicinal plant species. *Food and Bioproducts Processing*, 89(3), 217-233.
- Kumari, M., & Jain, S. (2012). Tannins: An antinutrient with positive effect to manage diabetes. *Research Journal of Recent Sciences* ISSN, 2277, 2502.

- Kunsorn, P., Ruangrunsi, N., Lipipun, V., Khanboon, A., & Rungsihirunrat, K. (2013). The identities and anti-herpes simplex virus activity of *Clinacanthus nutans* and *Clinacanthus siamensis*. *Asian Pacific journal of tropical biomedicine*, 3(4), 284-290.
- Lau, K. W., Lee, S. K., & Chin, J. H. (2014). Effect of the methanol leaves extract of *Clinacanthus nutans* on the activity of acetylcholinesterase in male mice. *Journal of Acute Disease*, 3(1), 22-25.
- Lee, K., Lee, B., Lee, M. H., Kim, B., Chinannai, K. S., Ham, I., & Choi, H. Y. (2015). Effect of *Ampelopsis Radix* on wound healing in scalded rats. *BMC complementary and alternative medicine*, 15(1), 1-9.
- Lindsay, D. G., & Astley, S. B. (2002). European research on the functional effects of dietary antioxidants. *EUROFEDA. Molecular Aspects of Medicine*, 1(23), 1-38..
- Lipipun, V., Kurokawa, M., Rutt, S., Taweechoitipatr, P., Pramyothin, P., Hattori, M., & Shiraki, K. (2003). Efficacy of Thai medicinal plant extracts against herpes simplex virus type 1 infection in vitro and in vivo. *Antiviral Research*, 60:175-180.
- Lipipun, V., Sasivimolphan, P., Yoshida, Y., Daikoku, T., Sritularak, B., Ritthidej, G., ... & Shiraki, K. (2011). Topical cream-based oxyresveratrol in the treatment of cutaneous HSV-1 infection in mice. *Antiviral research*, 91(2), 154-160.
- Lobo, R., Sodde, V., Dashora, N., Gupta, N., & Prabhu, K. (2011). Quantification of flavonoid and phenol content from *Macrosolen parasiticus* (L.) Danser. *J. Nat. Prod. Plant Resour*, 11, 96-99.
- Lutz Slomianka. (2021). Blue Histology - Integumentary System. Retrieved August 2, 2021, from Free.fr website: <http://lecannabiculteur.free.fr/SITES/UNIV%20W.AUSTRALIA/mb140/Cor ePages/Integumentary/Integum.htm>
- Magalhães, L. M., Segundo, M. A., Reis, S., & Lima, J. L. (2008). Methodological aspects about in vitro evaluation of antioxidant properties. *Analytica chimica acta*, 613(1), 1-19.
- Majumder, P., & M Paridhavi, P. (2019). A Novel Poly-herbal Formulation Hastens Diabetic Wound Healing with Potent Antioxidant Potential: A Comprehensive Pharmacological Investigation. *Pharmacognosy Journal*, 11(2), 324-331. doi: 10.5530/pj.2019.11.48
- Mat Ali, R., Abu Samah, Z., Mustapha, N., & Hussein, N. (2010). ASEAN Herbal and Medicinal Plants. (Internet). Available from: <http://www.asean.org/uploads/archive/publications/aseanherbal2010.p df>

- Mishra, K., Ojha, H., & Chaudhury, N. K. (2012). Estimation of antiradical properties of antioxidants using DPPH assay: A critical review and results. *Food chemistry*, 130(4), 1036-1043.
- More, B. (2016). Overview of medicine—Its importance and impact. *DJ Int J Med Res*, 1, 1-8.
- Moron, M. S., Depierre, J. W., & Mannervik, B. (1979). Levels of glutathione, glutathione reductase and glutathione S-transferase activities in rat lung and liver. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 582(1), 67-78.
- Muhamad, S. H. A., On, S., Sanusi, S. N., Hashim, A. A., & Zai, M. A. (2019, November). Antioxidant activity of Camphor leaves extract based on variation solvent. In *Journal of Physics: Conference Series* (Vol. 1349, No. 1, p. 012102). IOP Publishing.
- Murthy, S., Gautam, M. K., Goel, S., Purohit, V., Sharma, H., & Goel, R. K. (2013). Evaluation of in vivo wound healing activity of *Bacopa monniera* on different wound model in rats. *BioMed Research International*, 2013.
- Murugesu, S., Ibrahim, Z., Ahmed, Q. U., Uzir, B. F., Yusoff, N. I. N., Perumal, V., ... & Khatib, A. (2019). Identification of α -glucosidase inhibitors from *Clinacanthus nutans* leaf extract using liquid chromatography-mass spectrometry-based metabolomics and protein-ligand interaction with molecular docking. *Journal of pharmaceutical analysis*, 9(2), 91-99.
- Mustapa, A. N., Martin, Á, Mato, R. B., & Cocero, M. J. (2015). Extraction of phytochemicals from the medicinal plant *Clinacanthus nutans* Lindau by microwave-assisted extraction and supercritical carbon dioxide extraction. *Industrial Crops and Products*, 74, 83-94. doi:10.1016/j.indcrop.2015.04.035
- Nagar, H., Srivastava, A., Srivastava, R., Kurmi, M., Chandel, H., & Ranawat, M. (2016). Pharmacological Investigation of the Wound Healing Activity of *Cestrum nocturnum*(L.) Ointment in Wistar Albino Rats. *Journal of Pharmaceutics*, 2016, 1-8. doi: 10.1155/2016/9249040
- Nayak, B. S., Anderson, M., & Pereira, L. P. (2007). Evaluation of wound-healing potential of *Catharanthus roseus* leaf extract in rats. *Fitoterapia*, 78(7-8), 540-544.
- Nazarni, R., Purnama, D., Umar, S., & Eni, H. (2016). The effect of fermentation on total phenolic, flavonoid and tannin content and its relation to antibacterial activity in jaruk tigarun (*Crataeva nurvala*, Buch HAM). *International Food Research Journal*, 23(1): 309-315.
- Neuman, R. E., & Logan, M. A. (1950). The determination of collagen and elastin in tissues. *Journal of Biological Chemistry*, 186(2), 549-556.

- Ninan, N., Thomas, S., & Grohens, Y. (2015). Wound healing in urology. *Advanced drug delivery reviews*, 82, 93-105.
- Nichols, E. L. I. Z. A. B. E. T. H. (2015). Wound assessment part 1: how to measure a wound. *Wound Essentials*, 10(2), 51-5.
- Nithya, V., Brinda, P., & Anand, K. (2011). Wound Healing Activity Of *Leonotis Nepetaefolia* R.Br., In Wistar Albino Rats. *Asian Journal of Pharmaceutical and Clinical Research*, 23-26.
- Norshazila, S., Syed Zahir, I., Mustapha Suleiman, K., Aisyah, M. R., & Kamarul Rahim, K. (2010). Antioxidant levels and activities of selected seeds of malaysian tropical fruits. *Malaysian Journal of Nutrition*, 16(1).
- Nurliyana, R. D., Syed Zahir, I., Mustapha Suleiman, K., Aisyah, M. R., & Kamarul Rahim, K. (2010). Antioxidant study of pulps and peels of dragon fruits: a comparative study. *International Food research journal*, 17(2).
- Nuutila, K., Katayama, S., Vuola, J., & Kankuri, E. (2014). Human Wound-Healing Research: Issues and Perspectives for Studies Using Wide- Scale Analytic Platforms. *Advances in Wound Care*, 3(3), 264-271. doi:10.1089/wound.2013.0502
- Oates, J. (2020, June 30). Benefits of traditional medicine in the modern world. Retrieved August 2, 2021, from WaysTo.digital website: <https://waysto.digital/benefits-of-traditional-medicine-in-the-modern-world/>
- Ostermeyer, K. (2018, July 3). Pros & Cons of Alternative Medicine, Modern Medicine, & Traditional Medicine. Retrieved August 2, 2021, from Elite Learning website: <https://www.elitecme.com/resource-center/nursing/pros-cons-of-alternative-medicine-modern-medicine-traditional-medicine>
- Ordonez, A., Gomez, J., Vattuone, M., & Lsla, M. (2006). Antioxidant activities of *Sechium edule* (Jacq.) Swartz extracts. *Food Chemistry*, 97(3), 452–458. doi: 10.1016/j.foodchem.2005.05.024
- Ozgok Kangal, M. K., & Regan, J.-P. (2021). Wound Healing. Retrieved August 2, 2021, from PubMed website: <https://pubmed.ncbi.nlm.nih.gov/30571027/>
- Pannangpetch, P., Laupattarakasem, P., Kukongviriyapan, V., Kukongviriyapan, U., Kongyingoes, B., & Aromdee, C. (2007). Antioxidant activity and protective effect against oxidative hemolysis of *Clinacanthus nutans* (Burm.f) Lindau. *Songklanakarin J. Sci Technol.*, 1-9.
- Petkova, N , Ognyanov, M , Todorova, M , Denev, P. (2015) Ultrasound-assisted extraction and characterisation of inulin-type fructan from roots of *elecampane* (*Inula helenium* L.). *Acta Scientifica Naturalis*, 1, 225– 235.

- Peng, T. W., Han, C. J., & Akowuah, G. A. (2015). Effect of methanol extract of *Clinacanthus nutans* on serum biochemical parameters in rats. *Journal of Applied Pharmacy*, 6, 77-86.
- P'ng, X. W., Akowuah, G. A., & Chin, J. H. (2013). Evaluation of the sub-acute oral toxic effect of methanol extract of *Clinacanthus nutans* leaves in rats. *Journal of Acute Disease*, 2(1), 29-32. doi:10.1016/s2221- 6189(13)60090-6
- Priya, K. S., Babu, M., & Wells, A. (2004). 136 *Celosia argentea* Linn. Leaf Extract Improves Wound Healing in Rat Burn Wound Model. *Wound Repair and Regeneration*, 12(2), A35-A35.
- Qader, S. W., Abdulla, M. A., Chua, L. S., Najim, N., Zain, M., & Hamdan, S. (2011). Antioxidant, total phenolic content and cytotoxicity evaluation of selected Malaysian plants. *Molecules*, 16(4), 3433-3443.
- Ramasamy, S. (2016). Evaluation of *Clinacanthus nutans* (burm. F.) Lindau dried extract on wound healing activity in mice (Unpublished master's thesis). Management and Science University.
- Rane, M. M., & Mengi, S. A. (2003). Comparative effect of oral administration and topical application of alcoholic extract of *Terminalia arjuna* bark on incision and excision wounds in rats. *Fitoterapia*, 74(6), 553-558. doi: 10.1016/s0367-326x(03)00118-7
- Rashid, A., Qureshi, M., Raza, S., William, J., & Arshad, M. (2010). Quantitative determination of antioxidant potential of *artemisia persica*. *Ars Docendi Publishing House*, 23-30.
- Rathee, P., Chaudhary, H., Rathee, S., Rathee, D., Kumar, V., & Kohli, K. (2009). Mechanism of Action of Flavonoids as Anti-inflammatory Agents: A Review. *Inflammation & Allergy - Drug Targets*, 8(3), 229- 235. doi:10.2174/187152809788681029
- Roeslan MO, Na Ayudhya TD, Koontongkaew S. (2012) Characteristics of *Clinacanthus nutans* extraction from Thailand and Indonesia (Preliminary study). *Sci-Health* 002
- Rohn,S.(2018, July 1). How does the difference happen between ABTS and DPPH radical scavenging activity? Retrieved from https://www.researchgate.net/post/How_does_the_difference_happen_between_ABTS_and_DPPH_radical_scavenging_activity
- Roosita, K., Kusharto, C. M., Sekiyama, M., Fachrurozi, Y., & Ohtsuka, R. (2008). Medicinal plants used by the villagers of a Sundanese community in West Java, Indonesia. *Journal of ethnopharmacology*, 115(1), 72-81.
- Sakdarat, S., Shuyprom, A., Pientong, C., Ekalaksananan, T., & Thongchai, S. (2009). Bioactive constituents from the leaves of *Clinacanthus nutans* Lindau. *Bioorganic & medicinal chemistry*, 17(5), 1857-1860.

- Sangkitporn, S. (1993). Treatment of recurrent genital herpes simplex virus infections with *Clinacanthus nutans* extract. *Bull Dept Med Serv*, 18, 226-231.
- Sarega, N., Imam, M. U., Ooi, D.-J., Chan, K. W., Esa, N. M., Zawawi, N., & Ismail, M. (2016). Phenolic Rich Extract from *Clinacanthus nutans* Attenuates Hyperlipidemia-Associated Oxidative Stress in Rats. *Oxidative Medicine and Cellular Longevity*, 2016, 1–16. doi: 10.1155/2016/4137908
- Sekar, M., & Rashid, N. A. (2016). Formulation, evaluation and antibacterial properties of herbal ointment containing methanolic extract of *Clinacanthus nutans* leaves. *International Journal of Pharmaceutical and Clinical Research*, 8(8), 1170-1174.
- Shim, S. Y., Aziana, I., & Khoo, B. Y. (2013). Perspective and insight on *Clinacanthus nutans* Lindau in traditional medicine. *International Journal of Integrative Biology*, 14(1), 7-9.
- Sittiso, S., Ekalaksananan, T., Pientong, C., Sakdarat, S., Charoensri, N., & Kongyingoes, B. (2010). Effects of compounds from *Clinacanthus Nutans* on Dengue Virus Type 2 Infection. *Srinagarind Med J*, (25)1, 272- 275.
- Sreeramulu, D., & Raghunath, M. (2010). Antioxidant activity and phenolic content of roots, tubers and vegetables commonly consumed in India. *Food Research International*, 43(4), 1017-1020.
- Sriwanthana B, Chavalittumrong P, Chompuk L. (1996) Effect of *Clinacanthus nutans* on Human cell-mediated immune response in vitro. *Thai J Pharm Sci* 20(4), 261-267.
- Sullivan, T. (2011, August 16). Modern Medicine vs. Alternative Medicine: Different Levels of Evidence. Retrieved from Policy & Medicine website: <https://www.policymed.com/2011/08/modern-medicine-vs-alternative-medicine-different-levels-of-evidence.html>
- Talekar, Y. P., Apte, K. G., Paygude, S. V., Tondare, P. R., & Parab, P. B. (2017). Studies on wound healing potential of polyherbal formulation using in vitro and in vivo assays. *Journal of Ayurveda and Integrative Medicine*, 8(2), 73-81. doi:10.1016/j.jaim.2016.11.007
- Teixeira, B., Marques, A., Ramos, C., Neng, N. R., Nogueira, J. M., Saraiva, J. A., & Nunes, M. L. (2013). Chemical composition and antibacterial and antioxidant properties of commercial essential oils. *Industrial Crops and Products*, 43, 587-595.
- Thakur, R., Jain, N., Pathak, R., & Sandhu, S. (2011). Practices in Wound Healing Studies of Plants. *Evidence-Based Complementary And Alternative Medicine*, 2011, 1-17. doi: 10.1155/2011/438056

- Thongchai S, Ekalaksananan T, Pientong C, Aromdee C, Seubsasana S, Sukpol C & Kongyingyoes B. (2008). Anti-herpes simplex virus Type 1 activity of crude ethyl acetate extract derived from leaves of *Clinacanthus nutans* Lindau. *J Sci Technol* (27)4, 318-326.
- Thongrakard V, Tencomnao T. (2010). Modulatory effects of Thai medicinal plant extract on proinflammatory cytokines-induced apoptosis in human keratinocyte HaCaT cells. *African J Biotech* 93(1), 4999-5003.
- Timpawat, S., & Vajrabhaya, L. (1994). Clinical evaluation of *Clinacanthus nutans* Lindau in orabase in the treatment of recurrent aphthous stomatitis. *Mahidol Dental Journal*, 14(1), 10-16
- Tropicos. (2021). Tropicos. <https://www.tropicos.org/name/50058310> [Accessed 12 Aug. 2014]
- Tu, S. F., Liu, R. H., Cheng, Y. B., Hsu, Y. M., Du, Y. C., El-Shazly, M., & Chang, F. R. (2014). Chemical constituents and bioactivities of *Clinacanthus nutans* aerial parts. *Molecules*, 19(12), 20382-20390.
- Wanikiat, P., Panthong, A., Sujayanon, P., Yoosook, C., Rossi, A. G., & Reutrakul, V. (2008). The anti-inflammatory effects and the inhibition of neutrophil responsiveness by *Barleria lupulina* and *Clinacanthus nutans* extracts. *Journal of Ethnopharmacology*, 116(2), 234-244. doi:10.1016/j.jep.2007.11.035
- World Health Organization: WHO. (2019, November 25). Traditional, Complementary and Integrative Medicine. Who.int; World Health Organization: WHO. https://www.who.int/health-topics/traditional-complementary-and-integrative-medicine#tab=tab_1
- Witschi, H. P. (1986). Enhanced tumour development by butylated hydroxytoluene (BHT) in the liver, lung and gastro-intestinal tract. *Food and Chemical Toxicology* 24(10), 1127-1130
- Wong F, Yong A, Ting EP, Khoo S, Ong H, Chai T. (2014) Antioxidant, metal chelating, anti-glucosidase activities and phytochemical analysis of selected tropical medicinal plants. *Iranian J Pharm Res* 13(4), 1409- 1415.
- Yadav, K. S., Yadav, N. P., Rawat, B., Rai, V. K., Shanker, K., & Venkateswara Rao, C. (2014). An assessment of wound healing potential of *Argyrea speciosa* leaves. *The Scientific World Journal*, 2014.
- Yang, H. S., Peng, T. W., Madhavan, P., Shukkoor, M. A., & Akowuah, G. A. (2013). Phytochemical analysis and antibacterial activity of methanolic extract of *Clinacanthus nutans* leaf. *Int J Drug Dev Res*, 5, 349-355.
- Yahaya, R., Dash, G. K., Abdullah, M. S., & Mathews, A. (2015). *Clinacanthus nutans* (burm. F.) Lindau: An useful medicinal plant of south-east Asia. *International Journal of Pharmacognosy and Phytochemical Research*, 7(6), 1244-1250.

- Yong, Y. K., Tan, J. J., Teh, S. S., Mah, S. H., Ee, G. C., Chiong, H. S., & Ahmad, Z. (2013). Clinacanthus nutans Extracts Are Antioxidant with Antiproliferative Effect on Cultured Human Cancer Cell Lines. Evidence-Based Complementary and Alternative Medicine, 2013, 1-8. doi:10.1155/2013/462751
- Shiuan, J. M. P. Y., Wang, H. L. J., Lin, C. C., & Liang, J. Y. (2012). Effects of Clinacanthus nutans (Burm. f) Lindau leaf extracts on protection of plasmid DNA from riboflavin photoreaction. MC-Transaction on Biotechnology, 4(1), e5.
- Yousef, H., Alhadj, M., & Sharma, S. (2017). Anatomy, skin (integument), epidermis.
- Zainol, H., & Mansor, H. (2019). A Review of Therapeutic Potentials of Clinacanthus nutans as Source for Alternative Medicines. Sains Malaysiana, 48(12), 2683–2691. <https://doi.org/10.17576/jsm-2019-4812-09>
- Zakaria, Y., Yee, L. W., & Nik Hassan, N. (2017). Anti-Cancer Effects of Clinacanthus nutans. Journal of Biomedical and Clinical Sciences (JBACS), 2 (1), 11-19.
- Zhang, H., Chen, J., & Cen, Y. (2018). Burn wound healing potential of a polysaccharide from Sanguisorba officinalis L. in mice. International journal of biological macromolecules, 112, 862-867.
- Zulkipli, I. N., David, S. R., Rajabalaya, R., & Idris, A. (2015). Medicinal Plants: A Potential Source of Compounds for Targeting Cell Division. Drug Target Insights, 9. doi:10.4137/dti.s24946
- Zulkipli, I. N., Rajabalaya, R., Idris, A., Sulaiman, N. A., & David, S. R. (2017). Clinacanthus nutans: a review on ethnomedicinal uses, chemical constituents and pharmacological properties. Pharmaceutical Biology, 55(1), 1093-1113. doi:10.1080/13880209.2017.128874