

UNIVERSITI PUTRA MALAYSIA

PHYTOCHEMICAL SURVEY AND IN VITRO ANTIBACTERIAL ACTIVITY OF ETHNOMEDICINAL PLANTS TO TREAT GASTROINTESTINAL AILMENTS IN YOBE STATE, NIGERIA

ABDALLAH MUHAMMAD SALIHU

FS 2022 33



PHYTOCHEMICAL SURVEY AND *IN VITRO* ANTIBACTERIAL ACTIVITY OF ETHNOMEDICINAL PLANTS TO TREAT GASTROINTESTINAL AILMENTS IN YOBE STATE, NIGERIA



ABDALLAH MUHAMMAD SALIHU

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

July 2021

COPYRIGHT

All material contained inside the thesis, including without limitation text, logos, icons, photographs, and all other artwork, are copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

I dedicated this work to my late father (Salihu Abdallah Gamoji) and my lovely mother (Hajiya Maryam Abubakar Bello).



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in Fulfilment of the requirement for the degree of Doctor of Philosophy

PHYTOCHEMICAL SURVEY AND *IN VITRO* ANTIBACTERIAL ACTIVITY OF ETHNOMEDICINAL PLANTS TO TREAT GASTROINTESTINAL AILMENTS IN YOBE STATE, NIGERIA

By

ABDALLAH MUHAMMAD SALIHU

July 2021

Chairperson : Rusea Go, PhD Faculty : Science

Many plants in Yobe State, Nigeria have potentialities in curing many diseases. Rural and urban people made use of medicinal plants as their curative measures with the fact that, conventional drugs are quite expensive as well as contribute to the antimicrobial drug resistance. Abundant chemical constituents played vital roles which enabled the fight against any disease and ailments. This study aim to: Assess the most commonly used plant extracts in curing gastrointestinal ailments across Yobe State, Nigeria; evaluate their antibacterial efficacy against some enteric isolates using disk and well diffusion; determine phenolic and flavonoid contents of these plants extracts (Vachelia nilotica pods extract, Sclerocarya birrea stem (bark) extract, Guiera senegalensis leaves extract and Leptadenia hastata leaves extract) and lastly, determine the bioassay guided fractionating substances of the selected plant stem (bark), and detect compounds present in Sclerocarya birrea (A.Rich.) Hochst stem (bark) fraction capable of actions against the isolate using LC-MS. Thus, a semi structured questionnaires was used to collect initial information on the plants. Their antibacterial efficacy were tested using isolates; Escherichia coli, Salmonella typhi, Bacillus cereus and Staphylococcus aureus, where inhibition zones were measured. Folin-Ciocalteu's reagent procedure was adopted for the phenolic content, absorbance recorded at 760 nm that of flavonoid measured at 510 nm spectrophotometrically. The data was subjected to Chi-square (x^2) comparisons using SPSS version 22 and Graph pad prism version 8. The surveyed plants, were mostly Fabaceae. It has also recorded that 41-50 years were many into practice with their Pvalue 0.13. The majority of the respondents, were illiterates (*P-value* 0.06). However, ailments were ranged 0.69 - 0.75 informant consensus factors. Moreover, plant species with 0.34, 0.27 values were the highest Relative frequency citations. The (Methanolic and ethanolic) stem (bark) extracts reveal sound inhibition zones; S. typhi (16.3 mm); E. coli (15 mm); B. cereus (17 mm), on crude S. birrea stem (bark) where, 50mg/ml was the MIC without MBC. S. typhi (18 mm); E. coli (15 mm) were inhibited with V. nilotica (L.) Willd. ex Delile pods ethanolic extract, and exerted its MICs on (12.5 and 25 mg/ml & 50mg/ml).Nevertheless, organisms were resistant to aqueous extracts using Ciprofloxacin (19.7 mm - 33 mm) as the positive control, where by, no inhibition zones procured from Leptadenia hastata (Pers.) Decne leaves extracts against the isolates. Extracts were significantly different according to Tukey at $p \ge 0.05$. Nevertheless, Gallic acid increased tremendously in V. *nilotica* pods extracts with a curve ($R^2 = 0.9958$). While a large Catechin increase noticed in S. birrea stem (bark) extracts and followed by V. nilotica pods extracts with a curve ($R^2 = 0.9993$), all were significantly different at P-value < 0.0001 across the extracts which turned to have low contents especially L. hastata leaves extracts as compared. Their respective correlations were clearly denote phenolic contents with a curve ($R^2 = 0.5025$) and flavonoid contents ($R^2 = 0.7089$). Subsequently, 10mg/ml of the S. birrea stem (bark) ethyl acetate fraction extract was the MIC and no MBC on the fraction extract, therefore, inhibited the growth of the S. typhi. Statistically showed that, the isolate was susceptible to the positive control (Ciprofloxacin 30.33±0.0) as the highest inhibition zone followed by the ethyl acetate fraction extract at 10 mg/ml (9.7 \pm 0.0) and resistant as well at P< 0.0001 Tukey. Subsequently, LC-HRMS results of the most active fraction identified 16 compounds with various structures include: 1,8-Diazabicyclo [5.4.0]undec-7ene;Epigallocatechingallate;2,Amino1,3,4octadecanetriol;Gentisic acid; Vidarabine as well as DL-Isoleucine among others. Lastly, reasonable amount of chemical compounds determined the actions of individual plants, notably towards development of many valuable pharmaceutical products. S. birrea stem (bark) extract was found to be very useful in Yobe State, which is in accordance with the present study by having very active compounds for the efficacy including Vidarabine as the novel one.

Keywords: gastrointestinal, Informant consensus factor, medicinal plants, Bioassay guided fractionation, solvent partitioning, *Salmonella typhi*, *Sclerocarya birrea*, LC-MS

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

TINJAUAN FITOKIMIA DAN AKTIVITI ANTIBAKTERIA IN *VITRO* TUMBUHAN ETHNOMEDICINAL UNTUK MERAWAT PENYAKIT GASTROINTESTINAL DI NEGERI YOBE, Nigeria

Oleh

ABDALLAH MUHAMMAD SALIHU

Julai 2021

Pengerusi : Rusea Go, PhD Fakulti : Sains

Banyak tanaman di Negeri Yobe, Nigeria berpotensi menyembuhkan banyak penyakit. Penduduk desa dan bandar menggunakan tanaman ubat sebagai langkah penyembuhan mereka dengan fakta bahawa, ubat konvensional cukup mahal dan juga menyumbang kepada ketahanan terhadap ubat antimikrob. Kandungan bahan kimia yang banyak memainkan peranan penting yang membolehkan memerangi penyakit dan penyakit. Tujuan kajian ini adalah untuk: menilai ekstrak tumbuhan yang paling biasa digunakan dalam menyembuhkan penyakit gastrointestinal penyakit di seluruh Negeri Yobe, Nigeria; menilai keberkesanan antibakteria mereka terhadap beberapa isolat enterik menggunakan cakera dan penyebaran dengan baik; tentukan kandungan fenolik dan flavonoid ekstrak tumbuhan ini (ekstrak polong Vachelia nilotica, ekstrak batang Sclerocarya birrea (kulit kayu), ekstrak daun Guiera senegalensis dan ekstrak daun Leptadenia hastata) dan terakhir, tentukan bahan pecahan berpandu bioassay tumbuhan terpilih batang (kulit kayu),dan menentukan kandungan fenolik dan flavonoid ekstrak tumbuhan ini dan terakhir; menentukan bahan pecahan berpandu bioassay dari batang tanaman terpilih (kulit), dan mengesan sebatian yang terdapat dalam Sclerocarya birrea (A.Rich.) Batang Hochst (kulit) pecahan yang mampu bertindak terhadap isolat menggunakan LC MS. Oleh itu, soal selidik separa berstruktur digunakan untuk mengumpulkan maklumat awal mengenai tanaman. Keberkesanan antibakteria mereka diuji menggunakan isolat; Escherichia coli, Salmonella typhi, Bacillus cereus dan Staphylococcus aureus, di mana zon penghambatan diukur. Prosedur reagen Folin Ciocalteu diadopsi untuk kandungan fenolik, serapan direkodkan pada 760nm dari flavonoid yang diukur pada 510 nm secara spektrofotometrik. Data tersebut dilakukan perbandingan Chi square (x2) menggunakan SPSS versi 22 dan grafik prisma versi 8. Tumbuhan yang disurvei, kebanyakannya adalah Fabaceae. Ia juga mencatatkan bahawa 41-50 tahun banyak berlatih dengan nilai P mereka 0,13. Majoriti responden, buta huruf (nilai P- 0.06). Walau bagaimanapun, penyakit berjumlah 0.69 - 0.75 faktor permuafakatan informan. Lebih-lebih lagi, spesies tumbuhan dengan nilai 0.34, 0.27 adalah petikan frekuensi Relatif tertinggi. Ekstrak batang (Metanol dan etanol) kulit kayu menunjukkan zon penghambatan bunyi; S. typhi (16.3 mm); E. coli (15mm); B. cereus

(17mm), pada batang S. birrea kasar (kulit kayu) di mana, 50mg / ml adalah MIC tanpa MBC. S. typhi (18 mm); E. coli (15 mm) dihambat dengan ekstrak etanolik pod nilotica V., dan menggunakan MIC pada (25 mg / ml & 50 mg / ml).Walau bagaimanapun, organisma tahan kepada ekstrak akueus. Siprofloksasin (19.7 mm-33 mm) sebagai kawalan positif, di mana, tiada zon perencatan mendapat dari Leptadenia hastata (Pers.) Decne, ekstrak daun terhadap penyisihan. Ekstrak nyata sekali berbeza menurut Tuki pada $p \ge 0.05$. Meskipun asid galik sungguh bertambah dalam lenggai V. nilotica ekstrak dengan satu lengkungan ($R^2 = 0.9958$). Manakala Catechin besar bertambah dalam ekstrak batang (kulit) S. birrea dan diikuti oleh lenggai V. nilotica ekstrak dengan satu lengkungan ($\mathbb{R}^2 = 0.9993$), semua nyata sekali berbeza pada Nilai P < 0.0001 merentasi ekstrak yang bertukar untuk mempunyai isian rendah khususnya ekstrak daun L. hastata seperti yang jika dibandingkan.menghubung kait mereka dengan jelas menunjukkan kandungan fenolik dengan satu lengkungan ($R^2 = 0.5025$) dan kandungan flavonoid (R = 0.7089). Berikutnya, 10 mg / ml etil asetat batang (kulit) S. birrea pecahan ekstrak ialah MICs dan tiada MBCs oleh itu, menghalang pertumbuhan S. typhi. Secara statistik menunjukkan bahawa, mengasingkan rentan kepada kawalan positif (Ciprofloxacin 30.33±0.0) apabila zon perencatan tertinggi diikuti oleh pecahan ekstrak etil asetat pada 10 mg / ml (9.7 ± 0.0) dan tahan juga pada P< 0.0001 Tuki. Kemudiannya, LC-HRMS menyebabkan pecahan yang paling aktif mengenal pasti 16 sebatian dengan pelbagai Diazabicyclo struktur merangkumi: 1.8 [5.4.0] undec 7 ene; Epigallocatechingallate;2Amino1,3,4octadecanetriol;Asid Gentisic; Vidarabine dan juga DL Isoleucine antara lain. Akhir sekali, sebilangan besar sebatian kimia menentukan tindakan setiap tumbuhan, terutamanya ke arah pengembangan banyak produk farmaseutikal yang berharga. Ekstrak batang S. birrea (kulit kayu) didapati sangat berguna di Negeri Yobe, yang sesuai dengan kajian ini dengan mempunyai sebatian yang sangat aktif untuk keberkesanannya termasuk Vidarabine sebagai yang baru.

Kata Kunci: gastrousus, faktor konsensus Informant, tumbuhan ubatan, Bioassay berpandukan pemeringkatan, pemetakan pelarut, *Salmonella typhi*, *Sclerocarya birrea*, LC-MS

ACKNOWLEDGEMENTS

All praises and salutations are surrendered to Almighty Allah (S.W.T) for given me this opportunity to run this program successfully. My sincere appreciations to Prof. Rusea Go, Chairperson of this work, for her tireless patience, guidance, couching towards the completion of this work always there for me day and night. I also thank Associate Prof. Dr. Muskhazli Mustafa for his support and guidance and also Dr. Meenakshii A/P Nallappan for her supports in carrying out this work, all of them did well towards the completion of this study. My appreciations also to the entire Department of Biology, University Putra Malaysia academic and non-academic staff for their kind help and supports in one way or the other, and to my colleagues in seminar classes for their inputs especially Musa Ibrahim. I also appreciate the efforts of Dr. Norsha from the Institute of Bioscience, University Putra Malaysia. Moreover, I also appreciate the efforts of my institution, Yobe State University, Damaturu, Nigeria for their support through the Tertiary Education Trust Fund, in taking care of my tuition fees and up keeps throughout my study. My sincere appreciations also to Prof. Yakubu Mukhtar, University of Maiduguri and Associate Prof. Ali Usman Manzo, Yobe State University, Damaturu for their guidance and advises.

Furthermore, I duly appreciate the assistance given to me by the traditional medicine healers across Yobe State, my research field team; Yahaya Jibrin Damo, Muhammad Inuwa, Bashir Ararrabi, Sadisu Suleiman Dan jaji maji, Baba Zallah and Muhammad Ganjala throughout the survey of this study. I do acknowledge the efforts of my beloved mother, Hajiya Maryam Abubakar Bello, my beloved wife, Nafisa Abubakar, my kids; Salihu Muhammad Salihu, Maryam Muhammad Salihu and Aisha Muhammad Salihu, my sister Asma'u Salihu Abdallah for their unequalled perseverance, support and encouragement throughout the study.

Lastly, I also appreciate the rest of my family members, neighbours and friends for their prayers and encouragement.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Rusea Go, PhD

Professor Faculty of Science Universiti Putra Malaysia (Chairperson)

Muskhazli bin Mustafa, PhD Associate Professor Faculty of Sciences

Universiti Putra Malaysia (Member)

Meenakshii A/P Nallappan, PhD

Senior Lecturer Faculty of Sciences Universiti Putra Malaysia (Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 09 December 2021

Declaration by graduate student

I hereby confirm that:

- This thesis is my original work.
- Quotations, illustrations, and citations have been duly referenced.
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions.
- Intellectual property from the thesis and copyright of thesis are fully owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012.
- Written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:
Name and Matric No:	

Declaration by Members of Supervisory Committee

This is to confirm that:

- The research conducted and the writing of this thesis was under our supervision.
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of		
Chairman of		
Supervisory		
Committee:	Professor Dr. Rusea Go	
Signature:		
Name of		
Member of		
Supervisory		
Committee:	Associate Professor Dr. Muskhazli Mustafa	
Signature:		
Name of		
Member of		
Supervisory		
Committee:	Dr. Meenakshii A/P Nallappan	

TABLE OF CONTENTS

			Page	
ABG	траст		i	
	TRACI		iii	
ACI	V			
APF	ROVAL		vi	
DEC	LARATIO	N	viii	
LIS	Γ OF TABI	LES	xiii	
LIS	r of figu	RES	xiv	
LIS	Г OF APPE	INDICES	xvi	
LIS	FOF ABB	REVIATIONS	xvii	
CH	PTER			
1	INTROD	UCTION	1	
	1.1 Ba	ckground of the study	1	
	1.2 Pr	oblem statements	1	
	1.3 Re	esearch Justification	2	
	1.4 Re	esearch objectives	3	
•				
2	LITERA	TURE REVIEW	4	
	2.1 CC	onsumption of plants as medicines	5	
	2.2 Pla	ant therapeutic effects on organisms	/	
	2.3 DI	versity of gastrointestinal species on different plants	7	
	24 Ge	moral overview on modicinal plants action	13	
	2.4 00	1.1 Ethnobotanical study	15	
	2.5 M	ostly consumed medicinal plants in the study area	16	
	2.5 101	5.1 Vachellia nilotica overview	16	
	2.	5.2 An overview on <i>Sclerocarva birrea</i> (Marula)	19	
	2.4	5.3 An overview on Guiera senegalensis	26	
	2.4	5.4 Laptadenia hastata overview	29	
			_>	
3	ETHNO	BOTANICAL KNOWLEDGE OF THE MOST		
	COMMO	ONLY USED PLANTS IN THE MANAGEMENT OF		
	GASTRO	DINTESTINAL AILMENTS IN YOBE STATE,		
	NIGERL	A	32	
	3.1 Int	roduction	32	
	3.2 M	aterials and methods	34	
	3.2	2.1 Study Area:	34	
	3.2	2.2 Data collection	34	
	3.2	2.3 Collection of plant Materials and identification:	35	
	3.2	2.4 Data organization:	35	
	3.2	2.5 Fidelity level:	36	
	3.2	2.6 Informant consensus factor:	36	
	3.2	2./ Relative frequency of citations:	36	
	3.3 Re	esuits and discussion	31	

	3.3.1 Medicinal plant species collected from the survey area	
	with their usage on different gastrointestinal ailments	37
	3.3.2 Socio-demographic details of the respondents in Yobe	
	State Nigeria	40
	3.3.3 Fidelity level values of most commonly used modicinal	40
	5.5.5 Fidenty level values of most commonly used medicinal	
	plants as remedy to gastrointestinal aliments in Yobe	
	State	42
	3.3.4 Informant consensus factor values of medicinal plants	
	used as remedy for gastrointestinal ailments in Yobe	
	State Nigeria	44
	3.3.5 Relative frequency citations values of the surveyed	
	medicinal plants in Yobe State, Nigeria	45
3.4	4 Conclusion	47
4 NG	OTEWORTHY THREATENED PLANT SPECIES IN THE	
	HEI DECION NICEDIA	18
	Introduction	48
4.1		48
4.2	2 Loss of Biodiversity	49
	4.2.1 Plants species utilization patterns	50
	4.2.2 Other threatened species	51
4.3	3 NEEM TREE	51
	4.3.1 Neems Characteristics	52
	4.3.2 Neem as part of the West African trees	52
	4.3.3 Uses of Neem tree in the Sahel	53
	4.3.4 Timber	53
	435 Fuel	53
	A 3.6 Medicinal values of Neem	53
	4.3.7 Near needed a toucher law of protection	53
4	4.5.7 Neem needed a toughet law of protection	54
4.4		54
	4.4.1 Collection of plant Materials and identification:	54
	4.4.2 Data organization:	55
	4.4.3 Relative frequency of citations:	55
4.5	5 Results and discussion	55
4	4.5.1 Medicinal plant species collected from the survey area	
	with their medicinal values	55
	4.5.2 Relative frequency of citations	57
4.6	5 Conclusion	57
5 AF	NTIBACTERIAL EFFICACY OF SOME LISEFUL	
C M	FDICINAL PLANTS AGAINST SOME ENTERIC ISOLATES	
	SINC DISC AND WELL DIFFUSION	60
5 1	Introduction	60
5.1	Materials and methods	60
5.2		62
5.:	3 Collection of plant Materials and identification:	62
	5.3.1 Preparation of Extracts:	62
5.4	4 Cell cultures	63
	5.4.1 Efficacy of the extracts against the test isolates	63
	5.4.2 Sensitivity disc preparation	63
	5.4.3 Preparation of concentrate for sensitivity test	63
5.5	5 Antimicrobial Assay	64
	5.5.1 Disc diffusion method	64

xi

	5.5.2	Well diffusion method	64
	5.5.3	(MICs)	64
	5.5.4	Determination of Minimum Bactericidal Concentrations (MBCs)	65
5.6	Statisti	cal analysis	65
5.7	Results		65
5.8	Discuss	sion	66
5.9	Conclu	sion	78
6 DET USE FRA BIRI ITS	FUL MI CTIONA REA (A. EFFICA	ATION OF PHENOLICS, FLAVONOIDS OF SOME EDICINAL PLANTS AND BIOASSAY GUIDED ATING SUBSTANCES FROM SCLEROCARYA RICH) HOCHST STEM (BARK) EXTRACT, AND CY AGAINST SALMONELLA TYPHI	79
6.1	Materia	cuoli	79 80
0.2	6 2 1	Collection of Plant Materials and Their Identifications	80
	6.2.1	Sample Propagation:	80
	623	Total Phenolic Contents	80
	62.3	Total Elavonoid Contents	81
	625	Preparation of the Ethanol Extract and the Eractions	81
	626	Solvent Partitioning	81
	627	Testing Isolate Using the fraction extracts	82
	628	I C-MS Procedure	82
	629	Statistical Analyses	84
63	Results	Statistical 7 maryses	84
0.5	6.3.1	Quantification of Phenolic Contents of Sclerocarya birrea, Vachellia nilotica, Guiera senegalensis, and Leptadenia lanceolata extracts	84
	6.3.2	Quantification of Flavonoid Contents of Sclerocarya birrea, Vachellia nilotica, Guiera senegalensis, and	01
	6.3.3	Leptadenia lanceolata Extracts Bioassay-guided fractionating substances of <i>Sclerocarya birrea</i> (A. Rich) Hochst. stem (bark)	85
	6.3.4	extract against <i>Salmonella typhi</i> LC-MS analysis of ethyl acetate fraction of <i>S. birrea</i> stem (bark) extract identified by MZCloud and ChamSpider	86
64	Discuss	zion	91
6.5	Conclu	sions	95
		N AND DECOMMENDATIONS	04
	Conclu	of and RECOMMENDATIONS	90
(C) 7.1 7.2	Recom	mendations	96 97
REFEREN	CES		98
APPENDI	CES		113
BIODATA	OF STU	DENT	136
LIST OF P	UBLICA	TIONS	137

LIST OF TABLES

Table		Page
3.1	Medicinal plant species collected from the survey area with their usage on different gastrointestinal ailments	38
3.2	Socio-demographic details of the respondents in Yobe State, Nigeria	40
3.3	Fidelity level values of most commonly used medicinal plants as remedy to gastrointestinal ailments in Yobe State.	43
3.4	Informant consensus factor values of medicinal plants used as remedy for gastrointestinal ailments in Yobe State Nigeria	45
3.5	Relative frequency citations values of the surveyed medicinal plants in Yobe State, Nigeria	46
4.1	Medicinal plant species collected from the survey area with their medicinal values	56
4.2	Relative frequency citations values of the surveyed medicinal plants in	57
5.1	Antibacterial activity of plant extracts against Salmonella typhi	68
5.2	Antibacterial activity of plant extracts against <i>E. coli</i>	69
5.3	Antibacterial activity of plant extracts against Staphylococcus aureus	70
5.4	Antibacterial activity of plant extracts against B.cereus	71
5.5	Minimum inhibitory concentrations and minimum bactericidal concentrations of the various plants extracts against <i>S. typhi</i> (Concentration mg/ml)	72
5.6	Antibacterial activity of plant extracts against Salmonella typhi	74
5.7	Antibacterial activity of plant extracts against E. coli	75
5.8	Antibacterial activity of plant extracts against <i>Staphylococcus aureus</i>	76
5.9	Antibacterial activity of plant extracts against B. cereus	77

LIST OF FIGURES

	Figure		Page
	2.1	Distribution cases in the Local Government Areas similar to the spread of the Cholera Epidemic in Western Kenya	11
	2.2	N, N-Dimethyl aniline chemical structure	23
	2.3	Vidarabine chemical structure	24
	2.4	Isoleucine chemical structure	24
	2.5	DL-Norleucine chemical structure	24
	2.6	Gallic acid chemical structure	25
	3.1	Map of Yobe State., where by Damaturu represent senatorial district A, Potiskum represent senatorial district B and Nguru represent senatorial district C.	35
	3.2	(a.) chart compared the distribution between male and female respondents; (b.) explained the age distribution of the respondent; (c.) explained the literacy levels of the respondents; (d). It shows how the practice was carried out among the respondents. (e.) Indicates how formal training was undergone.	42
	3.3	Informant consensus factor values of medicinal plants used as remedy for gastrointestinal ailments in Yobe State Nigeria	45
	3.4	Relative frequency citations values of the surveyed medicinal plants in Yobe State, Nigeria	47
	4.1	Map of Yobe State, Nigeria (Sahel region)	49
	4.2	Threatened useful plants from the Sahel region in Nigeria. A. <i>Sclerocarya birrea</i> , B. <i>Acacia seyal</i> , C. <i>Azadirachta indica</i> , D. <i>Acacia nilotica</i> , E. <i>Balanites aegyptiaca</i> , F. <i>Detarium microcarpum</i> , and G. <i>Tamarindus indica</i>	59
	4.3	Major caused of depleting plant resources in the Sahel region. A. Demand for timber, B. Bush fire	59
	6.1	Fraction preparation flow chart	82

Graphical presentations of various plants' total phenolic contents. The following are the nomenclature of the used plants, $SME = Sclerocarya \ birrea$ methanolic extract; SEE = $Sclerocarya \ birrea$ ethanolic extract; $SCE = Sclerocarya \ birrea$ chloroform extract; $SAE = Sclerocarya \ birrea$ water extract; $VME = Vachellia \ nilotica$ methanolic extract; VEE = Vachellianilotica ethanolic extract; $VCE = Vachellia \ nilotica$ chloroform extract; $VAE = Vachellia \ nilotica$ water extract; GME = Gueirasenegalensis methanolic extract; $GEE = Gueira \ senegalensis$ ethanolic extract; $GCE = Gueira \ senegalensis$ chloroform extract; $GAE = Gueira \ senegalensis$ water extract; LME = $Leptadenia \ lanceolata$ methanolic extract; $LEE = Leptadenia \ lanceolata$ chloroform extract and $LAE = Leptadenia \ lanceolata$ water extract

- Graphical presentations of various plants' total flavonoid contents. The following are the nomenclature of the used plants, $SME = Sclerocarya \ birrea$ methanolic extract; $SEE = Sclerocarya \ birrea$ ethanolic extract; $SCE = Sclerocarya \ birrea$ chloroform extract; $SAE = Sclerocarya \ birrea$ water extract; $VME = Vachellia \ nilotica$ methanolic extract; VEE = Vachellianilotica ethanolic extract; $VCE = Vachellia \ nilotica$ chloroform extract; $VAE = Vachellia \ nilotica$ water extract; GME = Gueirasenegalensis methanolic extract; $GEE = Gueira \ senegalensis$ ethanolic extract; $GCE = Gueira \ senegalensis$ chloroform extract; $GAE = Gueira \ senegalensis$ water extract; LME = $Leptadenia \ lanceolata \ methanolic extract; \ LCE = Leptadenia \ lanceolata \ chloroform extract \ and \ LAE = Leptadenia \ lanceolata \ water$ extract.
- 6.4

6.5

Inhibition zones of different *Sclerocarya birrea* stem (bark) fraction extracts against *Salmonella typhi*

Identified compounds from *Sclerocarya birrea* ethyl acetate fraction extracts using LC MS/LC-HRMS.

XV

90

86

85

84

6.3

ŀ

LIST OF APPENDICES

Appendix		Page
A 1	Questionnaire for the ethnobotanical survey	113
A 2	Traditionsal healers' across the world	115
A 3	Traditional healers across Yobe State	115



 (\mathbf{G})

LIST OF ABBREVIATIONS

AGE	acute gastroenteritis
BCAAs	branched-chain amino acids
СсрА	Responds to a preferred carbon source
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acids
ET OH	Ethanolic
ETEC	Enterotoxigenic Escherichia coli
FL	Fidelity level
FtsZ	Filamenting temperature-sensitive mutant Z
GIT	Gastrointestinal
HUS	haemolytic uremic syndrome
HPLC	High performance liquid chromatography
ICF	Informant consensus factor
LB Agar	Luria Broth Agar
LC-MS	Liquid chromatography mass spectrometry
MBC	Minimum bactericidal concentrations
MHA	Mueller Hinton agar
MG	Milligram
MIC	Minimum inhibitory concentrations
MLSB	Macrolides, lincosamide and streptogramin B antibiotics
ML	Milliliter
MRSA	Methicillin resistant Staphylococcus aureus
MSA32	Muhammad Salihu Abdallah voucher number 32
NANTMP	National Association of Nigerian Traditional Medicine Practitioners

NC	Negative control
NUC	number of used citations
NS	number of used species for each citation
PC	Positive control
PCR	Polymerase chain reaction
PUFAs	Polyunsaturated fatty acids
PWD	Post weaning diarrhoea
QE	Quercetin equivalent
R&D	Research and Development
RFC	Relative frequency of citation
RNA	Ribonucleic acids
RpiR	responds to pentose phosphate pathway intermediates
SCFAs	Short-chain fatty acids
SF	Specific frequency for a specific ailment.
STEC	Shiga toxin-producing Escherichia coli
Stx	Shiga toxins
TF	Total number of citations of that very species
TFC	Total flavonoids contents
TPC	Total phenolic contents
UV	Ultraviolet light
μL	Microliter
μg	Microgram
WHO	World Health Organization

xviii

CHAPTER 1

INTRODUCTION

1.1 Background of the study

Rich biodiversity that Nigeria has and the number of plants used in traditional medicine of certain ailments were still not documented, such as on antimalarial plants covered in some part of Africa, as such traditional knowledge also need to be documented by conserving reports of valuable resources contained in the plants for the medicinal purposes (Tefera & Kim, 2019). Majority of the traditional herbal medications used in Africa are provided by practitioners who live within the populations. They trusted the system over time and are often willing to assist the patients with their knowledge and skills. Most of these herbal medicines are procured in their crude forms although some pharmaceutical pre-packaged forms also exist. Interestingly, unaffected interest in various traditional practices now exists among practitioners of traditional, indigenous or alternative systems (Aliyu et al., 2015). It has been proved that a certain reasonable number has been documented to be the rate on how indigenous traditional medicines were made from plants' natural products. Despite the fact that some of the traditional knowledge were not used in a larger society.

Many important compounds found in plants were identified through phytochemical analysis so as to know the suitable one for a proper medication. Plants species played important roles and due to that, they need to be conserved for the betterment of populace to properly attack the problems, serious destruction may lead to extinction of the plants species. Thus, many plants were tested on different health problems and worked wonderfully due to having sound natural products (Kumari et al., 2017).

1.2 Problem statements

Many plants in Nigeria have potentialities in curing many diseases. Moreover, many researches have been conducted scientific studies. With the advent of many orthodox, it has been difficult to utilize them due to the cost implication in curing different ailments. Plants became useful being the reservoir of many ingredients to alter the actions of microbial pathogens, which is being earlier stated by Madara et al. (2018). Based on the many researches surfaced in the world of traditional medicine, African traditional systems of medication have come up with a model so as to cure many diseases in a cheaper rate and total submission from the populace (Mahomoodally, 2013). The ethno medicinal plants survey has been revitalized in many aspect of screening the compounds for an alternative medication (Ashidi et al., 2010).

However, some of the Nigerian medicinal plants are helpful to different organs of the body in curing so many disorders and infections that do affect them. Generally, they do possessed ; fats, proteins, vitamins, carbohydrates and other bioactive compounds,

among them include: (*Aspilia africana, Bryophyllum pinnatum, Garcinia kola, Spondias mombim* and *Uvaria chamea*) they have been used also in curing tumor viruses as highlighted by (Okwu, 2007). It has been proved that ortho, cyanins, lignans, flavonoids and lignins were the phytoconstituents that made up of the phenolic groups which contributed towards; anticlotting, anti-inflammatory, immune enhancers as well as antioxidants, which were active as disease preventives as well as protect plants and humans damage (Okwu, 2007). The *G. kola* showed successful inhibitory activities against microbes, especially bacteria (*Staphylococcus aureus* and *Pseudomonas aeruginosa*) and fungi (*Candida albican*), which was in similar with some antibiotics (Okwu, 2003) and such in conformity with that of (Akoachere *et al.* 2002).

Many other plant species showed activities on both gram positive and other gram negative bacteria, such as; *E. coli, S. typhi, S. aureus, Klebsiella* spp, *Shigella flexneri, B. subtilis* etc., due to the presence of phenolic compounds which were used widely for the purpose of curative and prevention of many ailments and disorders (Ofokansi *et al.*, 2005). Both primary and secondary metabolites with disparity in their action on microbes and other disorders since in the olden days (Kumari et al., 2017). During AIDS pathogenesis and diarrheic diseases, gastrointestinal tract plays a vital role in the worsening the system, where such an issue mostly happened in a rural areas as well as some part of cities in African region (Cimerman et al., 1999). More so, more than a billion people worldwide in under developing countries are affected with parasitic infections specifically with *Ascaris lumbricoides* which triggered not only adult but to even children by causing malnutrition as pinpointed by (Ijaz, 2013).

The gastrointestinal tract infections in form of gastroenteritis, enteritis or enterocolitis which most common intestinal parasites such as *Giardia lamblia (Giardia duodenalis, Giardia intestinalis)*, *Cryptosporidium hominis* or *Cryptosporidium parvum* as well as *Entemoeba histolytic*. Infections caused by the parasitic organisms manifest with an abdominal pain, diarrhoea, dysentery, watery diarrhoea, dysentery or bloody stool may be caused by the parasites as well (Garcia et al., 2018). It has been known through ancient knowledge that, *Moringa oleifera* cures so many diseases and disorders due to the fact that, possessed so many natural products that do away with the abnormalities, such as anaemia, asthma, rheumatism, diarrhoea, among others as pinpointed by (Gupta et al., 2018). It has been compounded that, herbal formulations in traditional medicine were active due to the presence of bioactive compounds (Mustapa et al., 2018).

1.3 Research Justification

Medicinal plants are regarded as valuable and most useful natural resources used for the invention of new novel drugs. Many compounds such as; saponins, terpenoids, flavonoids, coumarins etc., were better known as attribute to the efficacies of the used medicinal plants in treating many ailments caused by microbes. Infectious diseases caused by bacterial pathogens have caused many deaths toll across the globe which was a major health-threat, approximately millions were killed. Some Gram- negative bacteria (*Escherichia coli* and *Klebsiella* spp.) were recorded to be responsible, as such, many metabolites inhibited their growths (Hajrah et al., 2018).

Nevertheless, much screened compounds from the plants extracts have shown anticancer activities for more than forty years, whereby, African plants leads some part of other continents like; China and India. Nigerian government has recently set aside US\$1billion for the development of drugs from the medicinal plants, this came up due to the fact that, some part of the country like south-western part, used plants for their medications by Ashidi et al. (2010). Report has shown that, many plants species were destroyed in search of medicine as well as for the fire wood. There will be a time the valuable medicinal plants will be disappeared if care is not taking, with such actions on the plants, so many diseases may lead to death of a larger number of both local and urban populace (Salisu et al., 2015).

Polyherbal formulations of medicinal plants is in line with synergistic effect of phytochemicals, where did a tremendous effort in curing many ailments, Mustapa (et al., 2018). Generally people in Nigeria and specifically Yobe State, made use of medicinal plants as their curative measures as well as raw materials for their day to day activities as shown in appendix A2, more importantly, combat various microbial and other disorders for their well-being (Elekwa et al., 2017). Moreover, some plant species have shown a larger portion of percentages for having the anti-gastrointestinal characteristics. The most common one include; Parasitosis, diarrhoea, constipation, colic, stomach ulcer, vomiting and lack of appetite (Ouachinou et al., 2019). It has been assessed in India and other regions that plants species revealed a myriad number of benefits.

1.4 Research objectives

This study aims to:

- 1. To assess the most commonly used plant extracts in curing gastrointestinal ailments across Yobe State, Nigeria.
- 2. To evaluate their antibacterial efficacy against some enteric isolates using disk and well diffusion.
- 3. To determine the phenolic and flavonoid contents of the four selected plants extracts (*V. nilotica* pods extract, *S. birrea* stem (bark) extract, *G. senegalensis* leaves extract and *L. hastata* leaves extract).
- 4. To determine the bioassay-guided fractionating substances of the selected plant stem (bark), and detect compounds present in *Sclerocarya birrea* (A. Rich.) Hochst stem (bark) fraction capable of actions against the isolate using LC-MS.

REFERENCES

- Abdallah, M. S., Mustafa, M., Nallappan, M. & Go, R., (2019). Review on Some Plants' Therapeutic Effects against Gastrointestinal Microbes. *Annual Review And Research In Biology*; 31(6), 1–8.
- Abadallah, M. S., Imam, I. U., & Ali, M. (2019). Identification and Evaluation of Antibiotic Sensitivity Pattern of Bacteria Associated with Gastro-Intestinal Disorder in Damaturu Yobe State, Nigeria. *Journal Allergy Research*, 1(2), 19-24.
- Abdallah, M. S. (2018). Comparative Study of Antibacterial and Phytochemical Screening of Ethanolic Extracts of *Citrus aurentifolia* and *Psidium guajava* on Some Clinical Isolates (*Pseudomonas aeruginosa* and *Escherichia coli*) of Patients Attending General Hospital Damagum, Yobe, *East African Scholars Journal of Medical Sciences*, 4421(3), 2617-7188.
- Abdu Zakari, Dimas Kubmarawa, Said Jibril, Sani Aliyu, and Shuaibu (2020). *In vitro* Antifungal, Antibacterial and Cytotoxicity Studies of Solvents Extracts of Some Selected Medicinal Plants Collected in Girei Adamawa State –Nigeria. *Official Journal of Department of Applied Chemistry*, *3*, 2705-2362.
- Abdullah, M. S., Nas, F. S., & Ali, M. (2019). Antibacterial Activity of Psidium guajava Leaf and Stem Bark Extracts against Clinical Isolates of Staphylococcus aureus and Salmonella typhi, International Journal of Research in Pharmacy and Biosciences, 6(5) 2394-5893, 11-17.
- AD, Olasupo, Aborisade, A. B., & Olagoke, O. V. (2018). Phytochemical analysis and antibacterial activities of spinach leaf. *American Journal Phytomedic Clinical Therapy*, 2, 1-4.
- ADOUM, O. A. (2016). Screening of Medicinal Plants Native To Kano and Jigawa States of Northern Nigeria, Using Artemia Cysts (Brine Shrimp Test). American Journal of Pharmacological Sciences, 4(1), 7–10.
- Al-Bishri, W., & Nabil Danial, E. (2013). Comparative study on the antioxidant, antimicrobial activities and total phenolic content of selected seeds from Saudi Arabia. *Journal of Food, Agriculture and Environment*, 11(2), 202–207.
- Aliyu, M., Yaro, A. H., Chedi, B. A. Z., & Salisu, A. I. (2015). Median lethal dose (LD50) evaluation of some polyherbal formulations marketed in Northern Nigeria. *International Journal of Herbs and Pharmacological Research*, 4(1), 18-23.
- Amerikova, M., El-tibi, I. P., Maslarska, V., & Tachkov, K. (2019). Antimicrobial activity, mechanism of action, and methods for stabilisation of defensins as new therapeutic agents. *Biotechnology & Biotechnological Equipment*, 33(1), 671–682.

- Amézquita-López, B. A., Soto-Beltrán, M., Lee, B. G., Yambao, J. C., & Quiñones, B. (2018). Isolation, genotyping and antimicrobial resistance of Shiga toxinproducing *Escherichia coli*. Journal of Microbiology, Immunology and Infection, 51(4), 425–434.
- Anka, Z. M., Singh, V., Gimba, S. N., & Singh, G. (2020). Antitoxic, Antifungal and Phytochemical Analysis of Medicinal Compounds of *Guiera senegalensis* Leaves, *Journal of Drug Delivery & Therapeutics*, 10(2), 148–152.10(2).
- Arthur, T. D., Ajami, N. J., Stewart, C. J., Lähdesmäki, H., Huttenhower, C., Krischer, J. P., Vehik, K. (2018). The human gut microbiome in early-onset type 1 diabetes from the TEDDY study. *Nature*, 562(7728), 589–594.
- Aryal, S., Baniya, M. K., Danekhu, K., Kunwar, P., Gurung, R., & Koirala, N. (2019). Total phenolic content, flavonoid content and antioxidant potential of wild vegetables from Western Nepal. *Plants*, 8(4), 96.
- Ashidi, J. S., Houghton, P. J., Hylands, P. J., & Efferth, T. (2010). Ethnobotanical survey and cytotoxicity testing of plants of South-western Nigeria used to treat cancer, with isolation of cytotoxic constituents from *Cajanus cajan* Millsp. leaves *Journal* of *Ethnopharmacology*, *128*(2), 501–512.
- Aslam, H., Green, J., Jacka, F. N., Collier, F., Berk, M., Pasco, J., & Dawson, S. L. (2020). Fermented foods, the gut and mental health: a mechanistic overview with implications for depression and anxiety. *Nutritional neuroscience*, 23(9), 659-671.
- Ave, N. (2020). Evaluation of antibacterial activity of medicinal plant extracts against clinical isolates of pathogens from children with acute gastroenteritis at katutura state hospital windhoek, namibia. *African, Journal Complement Alternative Medicine*, *16* (2), 15-23.
- Baldé, A. M., Traoré, M. S., Baldé, M. A., Barry, M. S., Diallo, A., Camara, M., & Oularé, K. (2016). Ethnomedical and ethnobotanical investigations on the response capacities of Guinean traditional health practioners in the management of outbreaks of infectious diseases: The case of the Ebola virus epidemic. *Journal of ethnopharmacology*, 182, 137-149.
- Bbosa, G. S., Kyegombe, D. B., Ogwal-Okeng, J., Bukenya-Ziraba, R., Odyek, O., & Waako, P. (2007). Antibacterial activity of *Mangifera indica* (L.). *African Journal* of Ecology, 45, 13-16.
- Bhat, K., Sankeshwari, R. M., Ankola, A. V., & Hullatti, K. (2018). Soxhlet versus cold maceration: Which method gives better antimicrobial activity to licorice extract against *Streptococcus mutans*. *Journal of the Scientific Society*, *45*(2), 67.
- Bi, G., Landry, G., Brice, K. A., Kevin, C., Marc, L., Akhanovna, M. J., & Yves-alain,
 B. (2018). Bio-guided anti-cariogenic and phytochemical valorization of *Guiera* senegalensis and Pseudocedrela kotschyi stem extracts, 12(28), 500–507.

- Binns, J., Parsons, S., & McIntyre, G. J. (2016). Accurate hydrogen parameters for the amino acid 1-leucine. Acta Crystallographica Section B: Structural Science, Crystal Engineering and Materials, 72(6), 885-892.
- Biswas, B., Rogers, K., McLaughlin, F., Daniels, D., & Yadav, A. (2013). Antimicrobial activities of leaf extracts of guava (*Psidium guajava* L.) on two gram-negative and gram-positive bacteria. *International journal of microbiology*,2013, 1-7.
- Bottone, E. J. (2010). *Bacillus cereus*, a volatile human pathogen. *Clinical microbiology reviews*, 23(2), 382-398.
- Bridson, D. and L. Forman (1999). *The Herbarium Handbook*. Royal Botanic Gardens, Kew. 3rd Edn (Edi.), pp: 4.
- Brunton, N. P., Russo, D., Kenny, O., Smyth, T. J., Milella, L., Hossain, M. B., & Diop, M. S., (2013). Profiling of phytochemicals in tissues from *Sclerocarya birrea* by HPLC-MS and their link with antioxidant activity. *International Scholarly Research Notices*, 2013,1-11.
- Cádiz-Gurrea, M. D. L. L., Lozano-Sánchez, J., Fernández-Ochoa, Á., & Segura-Carretero, A. (2019). Enhancing the Yield of Bioactive Compounds from *Sclerocarya birrea* Bark by Green Extraction Approaches. *Molecules*, 24(5), 966.
- Chakraborty, S. P., Mahapatra, S. K., & Roy, S. (2011). Biochemical characters and antibiotic susceptibility of *Staphylococcus aureus* isolates, *1*(3), 212–216.
- Cheikhyoussef, A., Shapi, M., Matengu, K., & Ashekele, H. M. (2011). Ethnobotanical study of indigenous knowledge on medicinal plant use by traditional healers in Oshikoto region ,Namibia. *Journal of Ethnobiology and Ethnomedicine*, 7(1),10.
- Chinsamy, M., Finnie, J. F., & Van Staden, J. (2014). Anti-inflammatory, antioxidant, anti-cholinesterase activity and mutagenicity of South African medicinal orchids. *South African Journal of Botany*, *91*, 88–98.
- Choudhury, S., Sharan, L., & Sinha, M. P. (2012). Phytochemical and Antimicrobial Screening of *Psidium Guajava* L. Leaf Extracts against Clinically Important Gastrointestinal Pathogens. *Journal of Natural Product Plant Resource*, 2(4), 524– 529.
- Irvine, F. R. (1952). Supplementary and emergency food plants of West Africa. *Economic Botany*, 6(1), 23-40.
- Chan, K.W., Iqbal, S., Khong, N.M.H., Ooi, D-J. & Ismail, M. 2014. Antioxidant activity of phenolicssaponins rich fraction prepared from defatted kenaf seed meal. *LWT-Food, Science & Technology*, 56, 181-186.
- Cimerman, S., Cimerman, B., & Lewi, D. S. (1999). Enteric parasites and AIDS. São Paulo Medical Journal = Revista Paulista de Medicina, 117(6), 266–273.

- Clavel, T., Gomes-Neto, J. C., Lagkouvardos, I., & Ramer-Tait, A. E. (2017). Deciphering interactions between the gut microbiota and the immune system via microbial cultivation and minimal microbiomes. *Immunological Reviews*, 279(1), 8–22.
- Cunha, L. C. S., Morais, S. A. L. De, Aquino, F. J. T. De, Chang, R., Oliveira, A. De, Martins, M. M., Evandro, A. (2017). Bioassay-guided fractionation and antimicrobial and cytotoxic activities of *Cassia bakeriana* extracts. *Revista Brasileira de Farmacognosia*, 27(1), 91–98.
- Dash, P. K., Sahoo, S., & Bal, S. (2008). Ethnobotanical Studies on Orchids of Niyamgiri Hill Ranges, Orissa, India. *Ethnobotanical Leaflets*, 12, 70–78.
- De Rauw, K., Thiry, D., Caljon, B., Saulmont, M., Mainil, J., & Piérard, D. (2019). Characteristics of Shiga toxin producing- and enteropathogenic *Escherichia coli* of the emerging serotype O80:H2 isolated from humans and diarrhoeic calves in Belgium. *Clinical Microbiology and Infection*, 25(1).
- Drioiche, A., Benhlima, N., Kharchouf, S., El-makhoukhi, F., Mehanned, S., Aaziz, H., Sciences, B. (2020). Research team on the chemistry of bioactive molecules and environment Moulay Ismaïl University, *Faculty of Science*, *16*, 1–14.
- Dubois, A. (1995). Spiral bacteria in the human stomach: The gastric helicobacters. *Emerging Infectious Diseases*, 1(3), 79–85.
- Dwyer, D. J., Belenky, P. A., Yang, J. H., MacDonald, I. C., Martell, J. D., Takahashi, N., ... & Collins, J. J. (2014). Antibiotics induce redox-related physiological alterations as part of their lethality. *Proceedings of the National Academy of Sciences*, 111(20), E2100-E2109.
- Elekwa, I., Ugbogu, A., Okereke, S., & Okezie, E. (2017). A Review of Selected Medicinal Plants with Potential Health Benefits in South-Eastern. *International Journal of Pharmaceutical and Chemical Sciences*, 6(4), 162–171.
- Ene, A. C., & Atawodi, S. E. (2012). Ethnomedicinal survey of plants used by the Kanuris of North-eastern Nigeria. *Indian Journal of Traditional Knowledge*, 11(4), 640–645.
- Esimone, C. O., Iroha, Ibezim, E. C., Okeh, C. O., & Okpana, E. M. (2006). *In vitro* evaluation of the interaction between tea extracts and penicillin G against staphylococcus aureus. *African Journal of Biotechnology*, *5*(11), 1082–1086.
- Ezuruike, U. F., & Prieto, J. M. (2014). The use of plants in the traditional management of diabetes in Nigeria: Pharmacological and toxicological considerations. *Journal of Ethnopharmacology*, 155(2), 857–924.
- Faruque, M. O., Uddin, S. B., Barlow, J. W., Hu, S., Dong, S., Cai, Q., & Hu, X. (2018). Quantitative ethnobotany of medicinal plants used by indigenous communities in the Bandarban District of Bangladesh. *Frontiers in pharmacology*, 9, 40.

- Fatiregun, A., Olowookere, S., Isere, E., & Ayede, A. (2017). Epidemiology of an outbreak of cholera in a south-west state of Nigeria. Southern African Journal of Epidemiology and Infectiossn, 27(4), 201–204.
- Fletcher, S., Sibbritt, D., Stark, D., Harkness, J., Rawlinson, W., Andresen, D., Ellis, J. (2015). Descriptive epidemiology of infectious gastrointestinal illnesses in Sydney, Australia, 2007-2010. Western Pacific Surveillance and Response Journal : WPSAR, 6(4), 7–16.
- Fratamico, P. M., Kim, M., Brinkac, L., Radune, D., Feng, P. C. H., Losada, L., Sanka, R. (2015). Whole genome sequencing of diverse Shiga toxin-producing and nonproducing *Escherichia coli* strains reveals a variety of virulence and novel antibiotic resistance plasmids. *Plasmid*, 83, 8–11.
- GARBA, A. (1997). Useful plants in the Chad region of North-East Nigeria. L'homme et le milieu végétal dans le bassin du lac Tchad. Paris, Orstom, 113-121.
- Garcia, L. S., Arrowood, M., Kokoskin, E., Paltridge, G. P., Pillai, D. R., Procop, G. W., Visvesvara, G. (2018). Laboratory diagnosis of parasites from the gastrointestinal tract. *Clinical Microbiology Reviews*, 31(1), 1–81.
- Gmaraldeen, S. M., Magzoub, A. A., Badri, A. M., & Garbi, M. I. (2016). Antibacterial activity of Acacia nilotica fruits extract against pathogenic bacteria, *International Journal of Applied Research*, 2(6): 103-106.
- Gouwakinnou, G. N., Lykke, A. M., Assogbadjo, A. E., & Sinsin, B. (2011). Local knowledge, pattern and diversity of use of *Sclerocarya birrea*. Journal of *Ethnobiology and Ethnomedicine*,7(18), 1–9.

Grinberg, A., & Data, R. U. S. A. (2013). (12) United States Patent, 27(12), 67-82.

- Gupta, S., Jain, R., Kachhwaha, S., & Kothari, S. L. (2018). Nutritional and medicinal applications of *Moringa oleifera* Lam.Review of current status and future possibilities. *Journal of Herbal Medicine*, 11, 1–11.
- Gurama, H. M., Maude, F. M., Jibrin, M. U., Oluwatovi, O. S., Sani, A. A., Inuwa, M. A., & Chikere, U. P. (2020). Phytochemical Analysis, Cytotoxcity and Antifungal Activities of *Guiera Senegalensis* Leaves Extract Review. *Chemical & Pharmaceutical Research*, 2(1), 1-4.
- Hannan, A., Asghar, S., Naeem, T., Ikram Ullah, M., Ahmed, I., Aneela, S., & Hussain, S. (2013). Antibacterial effect of mango (*Mangifera indica* Linn.) leaf extract against antibiotic sensitive and multi-drug resistant Salmonella typhi. Pakistan Journal of Pharmaceutical Sciences, 26(4), 715–719.
- Hassan, N., Wang, D., Zhiwei, Z., Nisar, M., & Zhu, Y. (2017). Determination and analysis of informant consensus factor of medicinal plant species used as remedy in Northern Pakistan. *Journal of Biodiversity and Environmental Sciences*, 11(2), 117-133.

- Ho, S.K.; Tan, C.P.; Thoo, Y.Y.; Abas, F.; Ho, C.W.(2014). Ultrasound-Assisted Extraction of Antioxidants in Misai Kucing (Orthosiphon stamineus). Molecules, 19, 12640-12659.
- Hua, X., Yang, Q., Zhang, W., Dong, Z., Yu, S., Schwarz, S., & Liu, S. (2018). Antibacterial activity and mechanism of action of aspidinol against multi-drugresistant methicillin-resistant *Staphylococcus aureus*. *Frontiers in pharmacology*, 9, 619.
- Huang, L. Z., Huang, B. K., Ye, Q., & Qin, L. P. (2011). Bioactivity-guided fractionation for anti-fatigue property of *Acanthopanax senticosus*. *Journal of ethnopharmacology*, 133(1), 213-219.
- Idrees, S., Qureshi, R., Bibi, Y., Ishfaq, A., Khalid, N., Iftikhar, A., Ahmad, N. (2016). Ethnobotanical and biological activities of *Leptadenia pyrotechnica* (forssk.) decne.: A review. *African Journal of Traditional, Complementary and Alternative Medicines*, 13(4), 88–96.
- Idris, A., Ali, M., & Yahaya, A. (2017). Antibacterial Activity and Phytochemical Screening of *Carica papaya* on some Enteric Bacterial Isolates of Public Health Importance. *Greener Journal of Biological Sciences*, 7(1), 001–007.
- Ijaz, M. K. (2013). Natural contamination of human hands with enteric parasites in Indian Subcontinent. World Journal of Clinical Infectious Diseases, 3(2), 13.
- Isa, M. A. (2016). Phytochemical And Antibacterial Activity of Leave Extracts of Guiera Senegalensis Lam on Selected Species of gram positive and gram Negative Bacteria, International Journal of Environment, 2(1), 262-268.
- Jaafar, H. Z. E., Karimi, E., Ibrahim, M. H., & Ghasemzadeh, A. (2013). Phytochemical screening and antioxidant activity assessment of the leaf stem and root of (*Labisia* paucifolia). Australian Journal of Crop Science, 7(2), 276–280.
- Jackson, M., & Gradmann, C. (2018). Medical Bacteriology. The Routledge History of Disease, (September), 378–401.
- Jamil, M., Haq, I., Mirza, B., & Qayyum, M. (2012). Isolation of antibacterial compounds from *Quercus dilatata* L. through bioassay guided fractionation, *Annual Clinical Microbiology*, 1(11), 1-11.
- Jiyil, M. K., Mafuyai, C. E., Shago, M. I., & Inuwa, H. M. (2019). Antimicrobial Activity of Methanolic, Aqueous and Partially Purified Protein of Young and Matured Leaves of *Guiera senegalensis (Moshi Medicine)*, 21(4), 1–11.
- Joffré, E., von Mentzer, A., Svennerholm, A. M., & Sjöling, Å. (2016). Identification of new heat-stable (STa) enterotoxin allele variants produced by human enterotoxigenic *Escherichia coli* (ETEC). *International Journal of Medical Microbiology*, 306 (7), 586–594.

- Johari, M. A., & Khong, H. Y. (2019). Total phenolic content and antioxidant and antibacterial activities of *Pereskia bleo. Advances in Pharmacological Sciences*, 2019, 1-4.
- Justine, V. T., Mustafa, M., & Go, R. (2018). Effect of Antimicrobial Activities on the Various Solvents Extracts of Leaves of *Scurrula Ferruginea* (Jack) Danser (Loranthaceae). *Tropical Agricultural Science*, 41(2), 677–686.
- Kaiser, J. C., & Heinrichs, D. E. (2018). Branching out: alterations in bacterial physiology and virulence due to branched-chain amino acid deprivation. *MBio*, 9(5), 01188-18.
- Kahkeshani N, Farzaei F, Fotouhi M, Alavi SSH, Bahramsoltani R, Naseri R, Momtaz S, Abbasabadi Z, Rahimi R, Farzaei MH, Bishayee A.(2019). Pharmacological effects of gallic acid in health and diseases: A mechanistic review. *Iran Journal of Basic Medical Science*, 22, 225-237.
- Kankara, S. S., Ibrahim, M. H., Mustafa, M., & Go, R. (2015). Ethnobotanical survey of medicinal plants used for traditional maternal healthcare in Katsina state, Nigeria. *South African Journal of Botany*, 97, 165–175.
- Kantati, Y. T., Kodjo, K. M., Dogbeavou, K. S., Vaudry, D., Leprince, J., & Gbeassor, M. (2016). Ethnopharmacological survey of plant species used in folk medicine against central nervous system disorders in Togo. *Journal of Ethnopharmacology*, 181, 214–220.
- Karim, M. T., Khanum, H., & Musa, S. (2018). Occurrence of enteric parasites and their risk factors among the female inhabitants of lower socioeconomic groups in Dhaka city. Asian Journal of Medical and Biological Research, 4(4), 343–350.
- Karimi, E., Jaafar, H. Z. E., & Ahmad, S. (2011). Phytochemical analysis and antimicrobial activities of methanolic extracts of leaf, stem and root from different varieties of *Labisa pumila* benth. *Molecules*, 16(6), 4438–4450.
- Khameneh, B., Iranshahy, M., Soheili, V., & Bazzaz, B. S. F. (2019). Review on plant antimicrobials: A mechanistic viewpoint. Antimicrobial Resistance & Infection Control, 8(1), 1-28.
- Kwaga, B. T. Martha, E.M., Ali, A. and Khobe, D. (2017). Evaluation Of The Growth Performance of lands snail *Achatina Achatina* Fed With Different Plant Materials In Sangere, Girei Local Government Of Adamawa State, Nigeria. *Journal of American Science*, 13(8):93-98.
- Koné, W. M., & Atindehou, K. K. (2008). Ethnobotanical inventory of medicinal plants used in traditional veterinary medicine in Northern Côte d'Ivoire (West Africa). South African Journal of Botany, 74 (1), 76-84.
- Kumari, P., Kumari, C., & Singh, P. S. (2017). Phytochemical Screening of Selected Medicinal Plants for Secondary Metabolites. *International. Journal of Life Science Scientific Research*, 3(4), 1151–1157.

- Lee, Y. S., Lee, D. Y., Kim, Y. B., Lee, S. W., Cha, S. W., Park, H. W., & Han, S. H. (2015). The mechanism underlying the antibacterial activity of shikonin against methicillin-resistant *Staphylococcus aureus*. *Evidence-based complementary and alternative medicine*.
- Mohammed, S. Y. (2013). Quantitative phytochemical and elemental analysis of *Guiera* senegalensis leaf extract. Journal of Pharmacognosy and Phytotherapy, 5(12), 204-207.
- Louis, H., Linus, M. N., Israt, A., Innocent, J., Amos, P. I., & Magu, T. O. (2018). Antimicrobial Activity of Stem, Leave and Root Plant Extract of Sclerocarya birrea and Sterculia setigera against Some Selected Microorganisms, World Scientific News, 92 (2), 309-326.
- Lulekal, E., Asfaw, Z., Kelbessa, E., & Van Damme, P. (2013). Ethnomedicinal study of plants used for human ailments in Ankober District, North Shewa Zone, Amhara Region, Ethiopia. *Journal of Ethnobiology and Ethnomedicine*, 9 (1), 63.
- M. Moyo, J.F. Finnie, J. Van Staden (2011). Antimicrobial and cyclooxygenase enzyme inhibitory activities of *Sclerocarya birrea* and *Harpephyllum caffrum* (Anacardiaceae) plant extracts. *South African Journal of Botany*, 77, 592–597.
- Madara, A. A., Abah, R. O., & Elkanah, O. S. (1857). *The revelator. Fudma Journal of Sciences*, 2, 2616-1370.
- Mahomoodally, M. F. (2013). Traditional medicines in Africa: an appraisal of ten potent African medicinal plants. *Evidence-Based Complementary and Alternative Medicine*.
- Mahdi, H. J., Khan, N. A., Mahmud, R., Asmawi, M. Z., Vikneswaran, A., & Murugaiyah, L. (2017). LC/MS, GC/MS screening and in vivo anti-inflammatory activity of Malaysian Moringa oleifera Lam leaf extracts and fractions against carrageenan-induced paw oedema in rats. *Journal of Innovative Pharmarcy and Biological Science*, 4, 48-54.
- Malgwi, S. A., Zango, M. K., Mbaya, A. W., Dennis, G., Kyari, F., Sanda, K. A., Bwala, A. D. (2019). Anti-trypanosomal activity of crude root extract of *Leptadenia hastata* (Pers) decne in Wistar rats infected with Trypanosoma brucei brucei and associated hematological changes. *Journal of Advanced Veterinary and Animal Research*, 6 (2), 241–246.
- Malviya, N., & Malviya, S. (2017). Bioassay guided fractionation-an emerging technique influence the isolation, identification and characterization of lead phytomolecules. *Hospital. Pharmacy*, *2* (5).
- Manzo, L. M., Moussa, I., & Ikhiri, K. (2017a). Antibacterial Evaluation of Ethnomedicinal Plants Used Against Diarrhea in Niger, Western Africa, *International Journal of Enteric Pathogens*, 5 (2), 54-58.

- Manzo, L. M., Moussa, I., & Ikhiri, K. (2017b). Phytochemical screening of selected medicinal plants used against diarrhea in Niger, West Africa. *International Journal* of Herbal Medicine; 5 (4), 32-385.
- Maria Kura 'nska, Milena Leszczy 'nska, El 'zbieta Malewska, Aleksander Prociak and Joanna Ryszkowska (2019). Implementation of Circular Economy Principles in the Synthesis of Polyurethane Foams. *Polymers*, *12*, 2068.
- Mariod, A. A., & Abdelwahab, S. I. (2010). *Sclerocarya birrea*: Biochemical Composition, Nutritional and Medicinal Uses A Review. *Natural Products:* Research Reviews, *1*, 28:375-388.
- Maroyi, A. (2013). Local knowledge and use of Marula (*Sclerocarya birrea*). (A. Rich Hochst).*Indian Journal of Traditional Knowledge*, *12* (3), 398-403.
- Mc, K., Jacquees, V., & Gerhard, P. (2017). Discrimination of the Geographical Origin of *Sclerocarya birrea* by 1 HNMR and LC-MS Based Metabolite Profiling Metabolomics : Open Access, 7 (2), 1–7.
- Milia, E., Usai, M., Szotáková, B., Elstnerová, M., Králová, V., Guy, D., Bortone, A. (2020). The Pharmaceutical Ability of *Pistacia lentiscus* L. Leaves Essential Oil Against *Periodontal* Bacteria and *Candida* spp and Its Anti-Inflammatory Potential, 1–16.
- Mlambo, V., Dlamini, B. J., Nkambule, M. T., Mhazo, N., & Sikosana, J. L. N. (2011). Nutritional evaluation of marula (*Sclerocarya birrea*) seed cake as a protein supplement for goats fed grass hay. *Tropical Agriculture*, 88(1), 35–43.
- Mohamed, S. T., Mamdou, A. B., Aïssata, C., Paul, C., Louis, M., & Luc, P. (2015). This item is the archived peer-reviewed author-version of: The malaria co-infection challenge: an investigation into the antimicrobial activity of selected Guinean medicinal plants. *Journal of ethnopharmacology*, 174, 576–581.
- Mohammed Jajere, S., Rabana Lawal, J., Mohammed Bello, A., Wakil, Y., Aliyu Turaki, U., & Waziri, I. (2016). Risk factors associated with the occurrence of gastrointestinal helminths among indigenous donkeys (Equus asinus) in Northeastern Nigeria. *Scientifica*, 2016,(1-7).
- Mohammed, S., Naziru, A., Mohammed, K., Sa'idu, H., Muntari, M., Zhigila, D. A., & Isa, S. (2016). Evaluation of bacteriostatic effect of methanolic extract of guiera senegalensis on some clinical bacteria. *Journal of Advanced Research in Materials Science*, 18(1), 10-17.
- Mohammed, S., Danjuma, M. N., & Abdulkarim, I. A. (2015). Ethnobotanical survey of medicinal plants in metropolitan Kano, Nigeria. *International journal of public health research*, 3(6), 345-351.
- Monajemzadeh, M., Abbasi, A., Tanzifi, P., Taba Taba Vakili, S., Irani, H., & Kashi, L. (2014). The Relation between *Helicobacter pylori* Infection and Acute Bacterial Diarrhea in Children . *International Journal of Pediatrics*,1–5.

- Moyo, M., Finnie, J. F., & Staden, J. Van. (2011). Antimicrobial and cyclooxygenase enzyme inhibitory activities of *Sclerocarya birrea* and *Harpephyllum caffrum* (Anacardiaceae) plant extracts. *South African Journal of Botany*, 77(3), 592–597.
- Munazir, M., Qureshi, R., Arshad, M. and Gulfraz, M. (2012). Antibacterial activity of root and fruit extracts of *Leptadenia pyrotechnica* (Asclepiadaceae) from *Pakistan*. *Pakistan Journal Botany*, 44, 1209-1213.
- Mushore, J., & Matuvhunye, M. (2013). Antibacterial properties of *Mangifera indica* on *Staphylococcus aureus*. *African Journal of Clinical and Experimental Microbiology*, *14* (2).
- Mustafa, N. K., Tanira, M. O. M., Dar, F. K., & Nsanze, H. (1999). Antimicrobial activity of *Acacia nilotica* subspp. *nilotica* fruit extracts. *Pharmacy and Pharmacology Communications*, 5 (9), 583–586.
- Mustapa, A. N., Martin, Á., Mato, R. B., Cocero, M. J., Mamun, M., Rashid, O., Manuhara, Y. S. W. (2018). Traditional West African pharmacopeia, plants and derived compounds for cancer therapy. *Journal of Ethnopharmacology*, 3 (1), 1– 5.
- Nakamoto, N., Sasaki, N., Aoki, R., Miyamoto, K., Suda, W., Teratani, T., Kanai, T. (2019). Gut pathobionts underlie intestinal barrier dysfunction and liver T helper 17 cell immune response in primary sclerosing cholangitis. *Nature Microbiology*, *4* (3), 492–503.
- Nciki, S., Vuuren, S., van Eyk, A., & de Wet, H. (2016). Plants used to treat skin diseases in northern Maputaland, South Africa: antimicrobial activity and *in vitro* permeability studies. *Pharmaceutical Biology*, *54* (11), 2420-2436.
- Njume C., Afolayan A. J., Samie A. and Ndip R. N.(2011). *In-vitro* anti-*Helicobacter* pylori activity of acetone, ethanol and methanol extracts of the stem bark of Combretum molle (Combretaceae). *Journal of Medicinal Plants Research*, 5 (14), pp. 3210-3216.
- Nn, A. (2015). A Review on the Extraction Methods Use in Medicinal Plants, Principle, Strength and Limitation. *Medicinal & Aromatic Plants*, 04 (03), 3–8.
- Nneka, N. I., Anthony, A. A., Kwaliafon, S. M., Charles, O. E., & Kennedy, F. C. (2016). Antimicrobial activity of *Psidium guajava* Linn. stem extracts against methicillinresistant *Staphylococcus aureus*. *African Journal of Biotechnology*, *11* (89), 15556–15559.
- Ojewole, J. A., Mawoza, T., Chiwororo, W. D., & Owira, P. M. (2010). Sclerocarya birrea (A. Rich) Hochst.['Marula'](Anacardiaceae): a review of its phytochemistry, pharmacology and toxicology and its ethnomedicinal uses. Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives, 24 (5), 633-639.

- Okhale, S. E., Ugbabe, G. E., Bamidele, O., Ajoku, G. A., & Egharevba, H. O. (2014). Phytochemical and antimicrobial studies on extractives of *Calyptrochilum emarginatum* (SW) Schltr (Orchidaceae) growing in Nigeria. Journal of Medicinal Plants Research, 8(4), 223-228.
- Okon, K. (2014). Enteropathogens Associated with Childhood Diarrheal Cases seen at a Tertiary Hospital in Nguru, Yobe State of Nigeria. *British Microbiology Research Journal*, 4(5), 583–591.
- Okwu, D. E., & Josiah, C. (2007). Nigerian medicinal plants I. Med Aromat Plant Sci Biotechnol, 1(1), 90-6. Stefanović, O. D. (2018). Synergistic activity of antibiotics and bioactive plant extracts: A study against Gram-positive and Gram-negative bacteria. Bacterial Pathogenesis and Antibacterial Control, 23, 23-48.
- Olgica D. Stefanović (2018), Synergistic Activity of Antibiotics and Bioactive Plant Extracts: A Study Against Gram-Positive and Gram- Negative Bacteria, *Bacterial Pathogenesis and Antibacterial Control*, 23, 23-48.
- Ouachinou, J. A., Dassou, G. H., Idohou, R., Adomou, A. C., & Yédomonhan, H. (2019). National inventory and usage of plant-based medicine to treat gastrointestinal disorders with cattle in Benin (West Africa). *South African Journal of Botany*, 122, 432-446.
- Panezai, M., Taj, M. K., Nawaz, I., Taj, I., Panezai, M., Panezai, N., Muhammade, G. (2018). Research article isolation and identification of *salmonella paratyphi* from enteric fever patients at different hospitals of quetta city. *Pakistan Journal of Biological Sciences*, 21 (9), 469–474.
- Parvez, M. K., Al-dosari, M. S., Arbab, A. H., Al-rehaily, A. J., & Abdelwahid, M. A. S. (2020). Bioassay-guided isolation of anti-hepatitis B virus flavonoid myricetin-3- O -rhamnoside along with quercetin from *Guiera senegalensis* leaves. *Saudi Pharmaceutical Journal*, 28 (5), 550–559.
- Paudel, M. R., Rajbanshi, N., Sah, A. K., Acharya, S., & Pant, B. (2018). Antibacterial activity of selected Dendrobium species against clinically isolated multiple drug resistant bacteria. *African Journal of Microbiology Research*, 12 (18), 426-432.
- Peng, M., & Biswas, D. (2017). Short chain and polyunsaturated fatty acids in host gut health and foodborne bacterial pathogen inhibition. *Critical Reviews in Food Science and Nutrition*, 57 (18), 3987–4002.
- Pirker, H., Haselmair, R., Kuhn, E., Schunko, C., & Vogl, C. R. (2012). Transformation of traditional knowledge of medicinal plants: the case of Tyroleans (Austria) who migrated to Australia, Brazil and Peru. *Journal of Ethnobiology and Ethnomedicine*, 8 (1), 1-26.
- Poblocka-Olech, L., Inkielewicz-Stepniak, I., & Krauze-Baranowska, M. (2019). Antiinflammatory and antioxidative effects of the buds from different species of Populus in human gingival fibroblast cells: Role of bioflavanones. *Phytomedicine*, 56, 1–9.

- Pouya, S. R. H., Moloudizargari, M., Asghari, M. H., Babaei, E., Ghobadi, E., & Mehdikhani, F. (2017). The healing effects of herbal preparations from *Sambucus ebulus* and *Urtica dioica* in full-thickness wound models. *Asian Pacific Journal of Tropical Biomedicine*, 7 (5), 421–427.
- Rahimi, E., Abdos, F., Momtaz, H., Torki Baghbadorani, Z., & Jalali, M. (2013). Bacillus cereus in infant foods: prevalence study and distribution of enterotoxigenic virulence factors in Isfahan Province, Iran. *The Scientific World Journal*, 2013,1-5.
- Regina, T., Lopes, M., Oliveira, F. R. De, Filocreão, F., Andrade, M. A. De, Monteiro, M. C.,Gonc, B. (2015). Antimicrobial bioassay-guided fractionation of a methanol extract of Eupatorium triplinerve Antimicrobial bioassay-guided fractionation of a methanol extract of *Eupatorium triplinerve*, 0209. *Pharmaceutical Biology*, 53 (6), 897-903.
- Roncarati, D., Pinatel, E., Fiore, E., Peano, C., Loibman, S., & Scarlato, V. (2019). *Helicobacter pylori* stress-response:Definition of the HrcA regulon. *Microorganisms*, 7 (10), 436.
- Ryan, U., Paparini, A., & Oskam, C. (2017). New Technologies for Detection of Enteric Parasites. *Trends in Parasitology*, *33* (7), 532–546.
- Sadiq, M. B., Tarning, J., Cho, T. Z. A., & Anal, A. K. (2017). Antibacterial activities and possible modes of action of *Acacia nilotica* (L.) Del. Against multidrugresistant *Escherichia coli* and *Salmonella* spp. *Molecules*, 22 (1).
- Sagar, S., Kaur, M., & Minneman, K. P. (2010). Antiviral lead compounds from marine sponges. *Marine drugs*, 8 (10), 2619-2638.
- Sani, H. Danbatta and Aliyu, B.(2011). In-Vitro Antibacterial Activity of Anogeissus Leiocarpus Dc (Stem Bark) Extracts Against Escherichia coli and Staphylococcus aureus Bayero Journal of Pure and Applied Sciences, 4 (2),56–59.
- Sarkar, R., Chaudhary, S. K., Sharma, A., Yadav, K. K., Nema, N. K., Sekhoacha, M., & Sen, T. (2014). Anti-biofilm activity of Marula–a study with the standardized bark extract. *Journal of Ethnopharmacology*, 154 (1), 170-175.
- Sawadogo, W. R., Schumacher, M., Teiten, M. H., Dicato, M., & Diederich, M. (2012). Traditional West African pharmacopeia, plants and derived compounds for cancer therapy. *Biochemical pharmacology*, 84(10), 1225-1240.
- Sibanda, T., & Okoh, A. I. (2007). The challenges of overcoming antibiotic resistance: Plant extracts as potential sources of antimicrobial and resistance modifying agents. *African Journal of Biotechnology*, 6 (25).
- Shoko, T., Maharaj, V. J., Naidoo, D., Tselanyane, M., Nthambeleni, R., Khorombi, E., & Apostolides, Z. (2018). Anti-aging potential of extracts from *Sclerocarya birrea* (A. Rich.) Hochst and its chemical profiling by UPLC-Q-TOF-MS. *BMC complementary and alternative medicine*, 18 (1), 1-14.

- Singh, J. (2015). Maceration, Percolation and Infusion Techniques of Extraction of Medicinal and Aromatic Plants (MAPs), 1–21. Ugwah, M. O., Etuk, E. U., Bello, S. O., Aliero, A. A., & Chinenye, J. (2013). Comparative studies of antiulcerogenic activities of three Nigerian medicinal plants: A preliminary evaluation. *Journal of Medicinal Plants Research*, 7 (9), 490–495.
- Skarpeli-Liati, M., Pati, S. G., Bolotin, J., Eustis, S. N., & Hofstetter, T. B. (2012). Carbon, hydrogen, and nitrogen isotope fractionation associated with oxidative transformation of substituted aromatic N-alkyl amines. *Environmental science & technology*, 46 (13), 7189-7198.
- Somboro, A. A., Patel, K., Diallo, D., Sidibe, L., Chalchat, J. C., Figueredo, G., & Chalard, P. (2011). An ethnobotanical and phytochemical study of the African medicinal plant Guiera senegalensis JF Gmel. *Journal of Medicinal Plants Research*, 5 (9), 1639-1651.
- Soni, N., & Singh, V. K. (2019). Traditional, nutraceutical and pharmacological approaches of Tamarindus indica (Imli). *European Journal of Biological Research*, 9 (3), 141-154.
- Suliman, M. B., Satti, N. M. E., Suliman, M. B., & Jagdouj, A. S. (2020). Identification of phenolic compounds of Adansonia digitata different parts via LC-MS / QTOF and their antioxidant activity, International Journal of Botany Studies, 5, (4), 108-111.
- Sunday, Ene-OjoAtawodi, Gboyega, Suleiman, Onaolapo, Brown, A. N. E. (2010). Comparative in vitro antioxidant potential of different parts of *Ipomoea asarifolia*, *Roemer & Schultes, Guiera senegalensis, JF Gmel and Anisopus mannii* NE Brown. *Brazilian Journal of Pharmaceutical Sciences*, 46 (2), 245-250.
- Tambunan, A. P., Bahtiar, A., Tjandrawinata, R. R., Tambunan, A. P., & Bahtiar, A. (2017). Influence of Extraction Parameters on the Yield, Phytochemical, TLC-Densitometric Quantification of Quercetin, and LC-MS Profile, and how to Standardize Different Batches for Long Term from Ageratum conyoides L. Leaves, 9 (6), 767–774.
- Tanih, N. F., & Ndip, R. N. (2012). Evaluation of the acetone and aqueous extracts of mature stem bark of *Sclerocarya birrea* for antioxidant and antimicrobial properties. *Evidence-Based Complementary and Alternative Medicine*, 2012, 1-7.
- Te, P. Jallali, I., Smaoui, A., Mérillon, J. M., Abdelly, C., & Ksouri, R (2020). Bioguided fractionation and characterization of powerful antioxidant compounds from the *halophyte Inula crithmoïdes*, *Arabian Journal of Chemistry*, 13(1), 2680-2688.
- Tefera, B. N., & Kim, Y. D. (2019). Ethnobotanical study of medicinal plants used as antimalarial and repellent by Sidama people of Hawassa Zuria district, Southern Ethiopia. J Complement Med Res, 10(1), 13-26.

- Teh, S. S., Ee, G. C. L., Mah, S. H., Yong, Y. K., Lim, Y. M., Rahmani, M., & Ahmad, Z. (2013). In vitro cytotoxic, antioxidant, and antimicrobial activities of Mesua beccariana (Baill.) Kosterm., Mesua ferrea Linn., and Mesua congestiflora extracts. BioMed research international, 517072, 9.
- Teke, G. N., Kuiate, J., Kueté, V., Teponno, R. B., Tapondjou, L. A., & Tane, P. (2011). Bio-guided isolation of potential antimicrobial and antioxidant agents from the stem bark of *Trilepisium madagascariense*. South African Journal of Botany, 77(2), 319–327.
- Tumpa, S. I., Hossain, I., & Ishika, T. (2015). Papaya and Mangifera indica against some gram positive and gram negative bacteria. Journal of Pharmacognosy and Phytochemistry, 3(6), 125–129.
- Ugwah, M. O., Etuk, E. U., Bello, S. O., Aliero, A. A., & Chinenye, J. (2013). Comparative studies of anti-ulcerogenic activities of three Nigerian medicinal plants: A preliminary evaluation. *Journal of Medicinal Plants Research*, 7(9), 490– 495.
- Umair, M., Altaf, M., & Abbasi, A. M. (2017). An ethnobotanical survey of indigenous medicinal plants in Hafizabad district, Punjab-Pakistan. *PLoS ONE*, *12*(6), 1–22.
- Valgas, C., Souza, S. M. D., Smânia, E. F., & Smânia Jr, A. (2007). Screening methods to determine antibacterial activity of natural products. *Brazilian journal of microbiology*, 38, 369-380.
- Vitetta, L., Briskey, D., Hayes, E., Shing, C., & Peake, J. (2012). A review of the pharmacobiotic regulation of gastrointestinal inflammation by probiotics, commensal bacteria and prebiotics. *Inflammopharmacology*, 20(5), 251-266.
- Van Vuuren, S. F. (2008). Antimicrobial activity of South African medicinal plants. *Journal of ethnopharmacology*, *119*(3), 462-472.
- Walker, M. Y., Pratap, S., Southerland, J. H., Farmer-Dixon, C. M., Lakshmyya, K., & Gangula, P. R. (2018). Role of oral and gut microbiome in nitric oxide-mediated colon motility. *Nitric Oxide*, 73, 81-88.
- Weshaya, H. M., & Sir, R. T. Al. (2017). Antimicrobial Activity of Acacia nilotica (Qaradh) Extract on Bacterial Species causing Wound Infections, African Journal of Medical Sciences, 2(5), 1–6.
- Wolfe, M. S. (1992). Giardiasis. Clinical microbiology reviews, 5(1), 93-100.
- Wong, B. Y., Tan, C. P., & Ho, C. W. (2013). Effect of solid-to-solvent ratio on phenolic content and antioxidant capacities of "Dukung Anak" (*Phyllanthus niruri*). *International Food Research Journal*, 20(1), 325–330.
- Yang, J. H., Bening, S. C., & Collins, J. J. (2017). ScienceDirect Antibiotic efficacy context matters. *Current Opinion in Microbiology*, 39, 73–80.

- Zhang Z, Guo Y, Liu Y, Chen M, Zhang D, Tian C, Liu M, Jiang G (2020). The Antibacterial Activity and Mechanism of Action of Luteolin Against *Trueperella pyogenes*. *Infection and Drug Resistance*, *13*, 1697–1711.
- Zeratsky, A. F. K. (2009). Nutrition and healthy eating. *Mayo Clinic. com. Mayo Clinic*, 27.
- Zhou, H., Gai, C., Ye, G., An, J., Liu, K., Xu, L., & Cao, H. (2019). Aeromonas *hydrophila*, an Emerging Causative Agent of Freshwater-farmed whiteleg shrimp *Litopenaeus vannamei*. *MDPI Microorganisms*, 7(10), 450.

