



UNIVERSITI PUTRA MALAYSIA

***DOCETAXEL-LOADED MAGNETIC NANOSTRUCTURED LIPID
CARRIER FUNCTIONALIZED WITH FISH OIL-COATED IRON OXIDE
NANOPARTICLES INTENDED FOR LUNG CANCER TREATMENT***

AUNI HAMIMI BINTI IDRIS

FS 2022 28



**DOCETAXEL-LOADED MAGNETIC NANOSTRUCTURED LIPID CARRIER
FUNCTIONALIZED WITH FISH OIL-COATED IRON OXIDE
NANOPARTICLES INTENDED FOR LUNG CANCER TREATMENT**

By

AUNI HAMIMI BINTI IDRIS

**Thesis submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of
Philosophy**

October 2021

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**DOCETAXEL-LOADED MAGNETIC NANOSTRUCTURED LIPID CARRIER
FUNCTIONALIZED WITH FISH OIL-COATED IRON OXIDE
NANOPARTICLES INTENDED FOR LUNG CANCER TREATMENT**

By

AUNI HAMIMI IDRIS

October 2021

Chair :Mohd Basyaruddin bin Abdul Rahman, PhD
Faculty :Science

Lung cancer is currently the most prevalent cause of cancer mortality due to late diagnosis and lack of curative therapies. Docetaxel (Dtx) is clinically proven to be effective, but poor aqueous solubility and non-selective cytotoxicity limit its therapeutic efficacy. Increasing the bioavailability of Dtx while potentially monitoring the therapeutic response via Magnetic Resonance Imaging is an appropriate strategy for effective drug delivery. In this work, a nanostructured lipid carrier (NLC) loaded with iron oxide nanoparticles (IONP) and Dtx (Dtx-MNLC) was developed as a potential theranostic agent for lung cancer treatment. The IONP was synthesised from thermal decomposition of iron oxyhydroxide (Fe(O)OH) and functionalised with Menhaden fish oil (MFO). Its physicochemical properties, cytotoxicity, and potential as contrast agents were then evaluated. The NLC was optimised using Response Surface Methodology. The amount of IONP and Dtx loaded into the Dtx-MNLC was quantified using Inductively Coupled Plasma Optical Emission Spectroscopy and high-performance liquid chromatography. Dtx-MNLC was then subjected to assessment of physicochemical characteristics, in vitro drug release, and cytotoxicity. IONP having 10 nm size was synthesised at 60 minutes aging time and 400 rpm stirring rate. The MFO-coated IONP (MFO-IONP) showed excellent aqueous dispersibility and good negative contrast with transverse relaxation rate of $9.85 \text{ mM}^{-1}\text{s}^{-1}$. MFO-IONP exhibited dose-dependent toxicity with higher toxicity on human lung carcinoma cells ($\text{IC}_{50} = 41 \text{ }\mu\text{g/mL}$) than human lung fibroblast cells ($\text{IC}_{50} = 494 \text{ }\mu\text{g/mL}$) within 72 hours exposure. The RSM model suggested the NLC formulated with 6% w/w lipid (MCT/ Precirol ATO 5) and 7.7% w/w emulsifier (TPGS/ Lipoid S75), with 20 minutes stirring time and 400 rpm stirring rate to achieve 187 nm particle size. Dtx loading percentage was determined at 3.98% w/w, and 0.36 mg/mL MFO-IONP was loaded into the Dtx-MNLC. The formulation showed a biphasic drug release in a simulated cancer cell environment, where 40% of Dtx was released for the first 6 hours, and 80% cumulative release was achieved after 48 hours. Dtx-MNLC exhibited higher

cytotoxicity to A549 cells than MRC5 in a dose-dependent manner. Furthermore, the toxicity of Dtx-MNLC to MRC5 was lower compared to the commercial formulation. In conclusion, Dtx-MNLC shows the efficacy to inhibit lung cancer cells growth, yet reduced toxicity on healthy lung cells and potentially capable as a theranostic agent for lung cancer treatment.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**MAGNETIK LIPID BERSTRUKTUR NANO MENGANDUNGI DOCETAXEL
DAN FERUM OKSIDA BERSALUT MINYAK IKAN SEBAGAI AGEN
RAWATAN KANSER PARU-PARU**

Oleh

AUNI HAMIMI IDRIS

Oktober 2021

Pengerusi : Mohd Basyaruddin bin Abdul Rahman, PhD
Fakulti : Sains

Kanser paru-paru menjadi punca kematian yang tertinggi di antara jenis kanser yang lain disebabkan kelewatan diagnosis dan kekurangan terapi penyembuhan yang berkesan. Docetaxel (Dtx) ialah antikanser yang berkesan secara klinikal, tetapi sifat tidak larut air dan kesitotoksikan yang tidak selektif telah menghadkan keberkesannya. Satu strategi yang efektif ialah dengan meningkatkan bioketersediaan Dtx dan memantau kesan terapi terhadap tumor melalui pengimejan resonans magnetik. Dalam kajian ini, Dtx dan magnetik ferum oksida nanopartikel (IONP) dimuatkan dalam lipid berstruktur nano (NLC) dan dijadikan Dtx-MNLC sebagai agen teranostik yang berpotensi merawat kanser paru-paru. IONP disintesis melalui penguraian terma ferum oksihidroksida ($\text{Fe}(\text{O})\text{OH}$) dan permukaannya difungsikan dengan minyak ikan Menhaden (MFO-IONP). Sifat fizikokimia, kesitotoksikan dan ciri agen kontras kemudiannya dinilai. Formulasi NLC dioptimakan melalui Metodologi Respons Permukaan manakala jumlah kandungan IONP dan Dtx ditentukan menggunakan plasma induksi optik pelepasan spektrometri dan kromatografi cecair prestasi tinggi. Pencirian fizikokimia, kadar pelepasan dos dan kesitotoksikan kemudian dilakukan. IONP bersaiz 10 nm disintesis dalam masa 60 minit dan kadar pengadukan 400 rpm. MFO-IONP dapat diserakkan dalam air dan menunjukkan kontras negatif dan kadar relaksasi melintang sebanyak $9.85 \text{ mM}^{-1}\text{s}^{-1}$. Kesitotoksikan MFO-IONP bergantung kepada dos, dan ketoksikannya lebih tinggi kepada sel karsinoma paru-paru manusia ($\text{IC}_{50} = 41 \mu\text{g}/\text{mL}$) berbanding sel fibroblas paru-paru ($\text{IC}_{50} = 494 \mu\text{g}/\text{mL}$) dalam tempoh 72 jam. Berdasarkan model RSM, formulasi NLC dihasilkan secara optima dengan 6% lipid (MCT/Precirol ATO 5) dan 7.7% pengemulsi (TPGS/Lipoid S75) dengan 20 minit masa pengadukan dan 400 rpm kadar pengadukan untuk mendapatkan saiz nanopartikel sebesar 187 nm. Sebanyak 3.98% b/b Dtx dan 0.36 mg/mL MFO-IONP berjaya dimuatkan dalam Dtx-MNLC. Formulasi ini melepaskan dos secara dwifasa dalam persekitaran sel kanser simulasi, iaitu 40% Dtx dilepaskan pada 6 jam pertama dan 80% pelepasan

kumulatif sepanjang 48 jam. Bergantung kepada dos, Dtx-MNLC adalah lebih toksik kepada A549 berbanding MRC5. Dtx-MNLC juga kurang toksik kepada MRC5 berbanding formulasi komersial. Kesimpulannya, Dtx-MNLC berkesan untuk merencatkan pertumbuhan sel kanser tetapi kurang toksik kepada sel tubuh yang sihat dan berpotensi sebagai agen teranostik untuk rawatan kanser paru-paru.



ACKNOWLEDGEMENTS

With the name of Allah, the Most Compassionate and the Most Merciful. All praise and gratitude to Allah, who gave me the opportunity, strength, health, and passion for completing this study.

I would like to express my gratitude to my main supervisor, Prof. Dr. Mohd Basyaruddin Abdul Rahman, my co-supervisors Associate Prof. Dr Che Azurahaman Che Abdullah and Prof. Dr Nor Azah Yusof for their excellent supervision. Thank you for your encouragement and patience in helping me along this journey. Without their guidance and counsel, this work would be impossible to complete. My deepest appreciation goes to Prof Basya for his support and constructive advice, and Dr Che Azurahaman for her insightful feedback, emotional encouragement and sisterly concern. I am also grateful for the help and expertise of Dr. Nur Khatijah Mohd Zin during the toxicity studies at UPM-MAKNA lab. Thank you for your support and assistance.

My appreciation is extended to all my lab mates in Lab 105, 401 and 236 (Faculty of Science, UPM) for their assistance and encouragement. Your friendship and accompaniment have made this journey colourful and memorable. Not to forget, many thanks to all Integrated Chemical BioPhysics Research committee, friends, and staff members of the Faculty of Science, Universiti Putra Malaysia, although not individually acknowledged here for their direct or indirect effort and contribution to this study. Some special thanks go to my friends Fazriyana and Akmarina for lending their helping hands and being a listening ear throughout this journey. Thank you for reminding me to take breaks and have fun when I've been stressed out.

I gratefully acknowledge the financial support from Universiti Malaysia Pahang and Ministry of Higher Education Malaysia under SLAB scheme during my PhD candidature. I would also like to thank Malaysian Institute for Innovative Nanotechnology (NanoMITE) for funding this research under LRGS research grant (RU029-2014;5526306). Thank you for the trust and opportunity granted.

Last but not least, to my parents Idris bin Ahmad and Maznon binti Ishak, my parents-in-law Abdul Razak bin Daud and Hamidah binti Rahman, my brothers and sisters and my sisters-in-law, thank you for all your prayers, supports and motivations all these years. I am indebted to my dearest husband Amir bin Abdul Razak, for the love and constant support, and for giving me the lift I needed when I feel down. Finally, to my lovely kids, Raihan, Zahra, Zayd and Ammar, who blessed me with a life full of joy, and taught me to move forward no matter what comes.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Mohd Basyaruddin bin Abdul Rahman, PhD

Professor, ChM.
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Nor Azah binti Yusof, PhD

Professor, ChM.
Faculty of Science
Universiti Putra Malaysia
(Member)

Che Azurahaman binti Che Abdullah, PhD

Associate Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 19 May 2022

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Auni Hamimi Binti Idris

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research and the writing of this thesis were done under our supervision;
- supervisory responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2015-2016) are adhered to.

Signature: _____

Name of Chairman
of Supervisory

Committee: Prof. ChM. Dr. Mohd Basyaruddin bin Abdul Rahman

Signature: _____

Name of Member of
Supervisory

Committee: Prof. ChM. Dr. Nor Azah binti Yusof

Signature: _____

Name of Member of
Supervisory

Committee: Assoc. Prof. Dr. Che Azurahaman binti Che Abdullah

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xv
LIST OF ABBREVIATIONS	xix
CHAPTER	
1 INTRODUCTION	1
1.1 Research Background	1
1.2 Research Objectives	2
1.3 Scope of the Study	3
1.4 Limitation of the Study	3
2 LITERATURE REVIEW	4
2.1 Lung cancer: epidemiology, classification and biology	4
2.1.1 Current state of diagnosis and treatment challenges of lung cancer	6
2.1.2 Moving forward: application of nanotechnology in diagnosis and treatment of lung cancer	8
2.2 Nanostructured lipid carriers (NLC) in drug delivery system	10
2.2.1 General ingredients in NLC formulation	11
2.2.2 Preparation and physicochemical characterisation of nanostructured lipid carrier	13
2.2.3 Challenges and issues in the preparation of NLC	14
2.3 Iron oxide-based theranostic formulation	15
2.3.1 Synthesis of iron oxide nanoparticles	16
2.3.2 Surface functionalization of IONP for biomedical purposes	17
2.3.3 IONP in Magnetic Resonance Imaging (MRI)	19
3 MATERIALS AND METHODOLOGY	23
3.1 Materials	23
3.2 Modified synthesis of iron oxide nanoparticles	24
3.3 Substitution of oleic acid coating on IONP with Menhaden fish oil (MFO) via ligand exchange	25
3.4 Physicochemical characterization of IONP	26

3.4.1	Morphology analysis	26
3.4.2	Studies on magnetic properties of IONP	27
3.4.3	Structural analysis and phase identification	27
3.4.4	Identification of functional groups on IONP surface	28
3.4.5	Thermogravimetric (TGA) analysis	28
3.4.6	Magnetic Resonance (MR) imaging and relaxometry	28
3.5	Screening of excipients for development of docetaxel-loaded magnetic nanostructured lipid carrier (Dtx-MNLC)	30
3.5.1	Screening of oil	30
3.5.2	Screening of solid lipid	30
3.5.3	Physical compatibility of solid lipid and oil	31
3.5.4	Ratio of solid lipid and oil	31
3.5.5	Screening of emulsifiers	31
3.5.6	Screening of organic solvent	32
3.6	Formulation optimization of Dtx-MNLC using Central Composite Design	33
3.6.1	Design of Experiments	33
3.6.2	Fitting and evaluation of the model	34
3.6.3	Model validation	35
3.7	Optimisation of IONP and drug loading	35
3.8	Selection of purification methods	37
3.9	Preparation of docetaxel-loaded magnetic nanostructured lipid carrier	37
3.10	Physicochemical characterizations of Dtx-MNLC	38
3.10.1	HPLC method for quantification of Dtx	38
3.10.2	Particle size distribution and zeta potential analysis	39
3.10.3	Differential Scanning Calorimetry (DSC) analysis	39
3.10.4	Morphology analysis	39
3.10.5	Crystallinity study of MNLC	39
3.10.6	Fourier Transform Infrared (FTIR) Spectroscopy	40
3.10.7	Magnetic properties of Dtx-MNLC	40
3.11	In vitro drug release studies	40
3.12	In vitro safety and efficacy studies of IONP and Dtx-MNLC	43
3.12.1	Cell propagation method	43
3.12.2	Cell seeding	44
3.12.3	Treatment of cells	46
3.12.4	XTT assay	47
3.12.5	Statistical analysis	47
3.13	Selection of cryoprotectant concentration for freeze-drying	48

3.14	Stability assessment of Dtx-MNLC	48
4	RESULTS AND DISCUSSION	49
4.1	Synthesis optimisation of oleic acid-coated iron oxide nanoparticles	49
4.1.1	Effect of aging time	49
4.1.2	Effect of stirring rate	53
4.2	Substitution of oleic acid ligand to fish oil	57
4.2.1	Surface modification of OA-IONP to MFO-IONP	57
4.2.2	Colloidal stability of MFO-IONP in aqueous buffer	62
4.2.3	Thermogravimetric analysis of coating on IONP surface	64
4.2.4	Magnetic Resonance Imaging of MFO-IONP	67
4.2.5	In vitro cytotoxicity of MFO-IONP	70
4.3	Formulation of magnetic nanostructured lipid carrier loaded with docetaxel	73
4.3.1	Screening of lipid components	73
4.3.2	Compatibility of solid lipid with oil	76
4.3.3	Screening of emulsifier composition	79
4.3.4	Composition of organic solvent	81
4.4	Optimisation of NLC using Central Composite Design (CCD)	83
4.4.1	Model fitting	83
4.4.2	Interactions of independent variables on particle size and polydispersity index (PDI)	83
4.4.3	Optimisation of nanostructured lipid carrier formulation	88
4.5	Optimisation of IONP and drug loading	88
4.6	Selection of purification method	89
4.7	Physicochemical characterisation of Dtx-MNLC	91
4.8	In vitro drug release profiles	98
4.9	In vitro cytotoxicity of Dtx-MNLC	100
4.10	Selection of cryoprotectant concentration for lyophilisation	104
4.11	Storage stability studies of Dtx-MNLC	105
5	CONCLUSIONS AND RECOMMENDATION FOR FUTURE WORKS	109
5.1	Conclusions	109
5.2	Recommendations for future research	110
	REFERENCES	112
	APPENDICES	139
	BIODATA OF STUDENT	144
	LIST OF PUBLICATIONS	145

LIST OF TABLES

Table		Page
2.1	Classification of drugs, mechanism of actions and side effects associated with chemotherapeutics	7
2.2	Lipids commonly used in the preparation of NLC	12
3.1	List of chemicals	23
3.2	Experiment parameters for synthesis of IONP	25
3.3	Ratio of IONP to MFO in ligand exchange	26
3.4	Parameters for MR-imaging and relaxometry	28
3.5	Parameters for T2- weighted images	29
3.6	Independent variables and their corresponding levels in CCD model	33
3.7	Central Composite Design experimental array	34
3.8	Composition of cell culture media	44
4.1	Particle diameter, particle dispersity and saturation magnetisation of OA-IONP synthesised at four different aging time.	52
4.2	Particle diameter, particle dispersity and saturation magnetisation of OA-IONP synthesised at four different stirring rates.	54
4.3	Band assignments for FTIR spectra of oleic acid and OA-IONP	59
4.4	Band assignments of MFO and MFO-IONP	62
4.5	Dispersibility and pH of MFO-IONP in PBS.	63
4.6	Dispersibility of OA-IONP and MFO15-IONP in different simulated biological media.	64
4.7	Summary of thermal analysis of OA-IONP	66
4.8	Thermal analysis of MFO-IONP after ligand exchange	67

4.9	IC50 values of MFO-IONP on MRC5 and A549 at 24H, 48H and 72H	72
4.10	Onset temperature, melting temperature and heat of fusion for binary lipid mix with various ratios determined using DSC.	79
4.11	Regression coefficient of the reduced model of nanostructured lipid carrier without Dtx and IONP	85
4.12	Analysis of variance of the reduced quadratic model on particle size and Pdl of nanostructured lipid carrier	85
4.13	Actual and predicted response values for optimal condition of nanostructured lipid carrier	88
4.14	Concentration of IONP, percentage of entrapment efficiency, drug loading, particle size and Pdl of MNLC 1 and MNLC 2.	89
4.15	Particle size and Pdl of the Dtx-MNLC before purification and after purification with three different methods gel filtration, centrifugation and centrifugation/ dialysis.	90
4.16	Onset melting temperature and heat of fusion of Precirol ATO 5, binary mixture of Precirol/MCT (60:40 w/w), ternary mixture of Precirol/MCT/Dtx and Dtx-MNLC analysed using DSC.	96
4.17	Model fitting for release kinetics of Dtx-MNLC	100
4.18	IC50 value of MRC5 and A549 cell lines after 24 H and 48 H exposure to CFDtx or Dtx-MNLC	103
4.19	Particle size, and Pdl of the Dtx-MNLC after addition of sucrose as cryoprotectant according to the percentage added into the formulation.	105
4.20	Peak temperature and heat of fusion of freshly prepared Dtx-MNLC and after 6 months storage at 27 °C (Dtx-MNLC RT) or at a cooler temperature of 2 8 °C (Dtx-MNLC FR) analysed using DSC.	107

LIST OF FIGURES

Figure		Page
2.1	Global cancer mortality for both sexes combined at all ages according to cancer types in 2020.	4
2.2	Illustration of lung cancer staging according to tumour size, tumour localisation and metastatic degree of the tumour cells	5
2.3	Illustration of normal vasculature of healthy tissues compared with disorganized vasculature in the tumour tissue.	9
2.4	Crystal structure of magnetite (left) and maghemite (right).	16
2.5	Most abundant fatty acid in Menhaden fish oil.	18
3.1	Synthesis procedure of OA-IONP.	24
3.2	Two-step ligand exchange procedure.	26
3.3	Schematic of Dtx-MNLC preparation.	37
3.4	Schematic of drug dissolution setup.	41
3.5	Chemical structure of tetrazolium salt (left) and orange formazan dye (right)	43
3.6	Cell counting using hemocytometer	45
4.1	HRTEM images of OA-IONP samples with four different aging time at 50KX magnification.	50
4.2	Field-dependent hysteresis loops (M–H) measured at room temperature for OA IONP obtained with different aging time.	51
4.3	X-ray diffraction pattern of OA-IONP obtained with four different aging time.	52
4.4	HRTEM images of IONP samples synthesised with different stirring rates at 50KX magnification.	54
4.5	Field-dependent hysteresis loops (M–H) at room temperature for OA IONP obtained with different stirring rates.	55

4.6	X-ray diffraction pattern of IONP obtained with different stirring rates.	56
4.7	Chemical structure of oleic acid	57
4.8	FTIR spectra of oleic acid and oleic acid-coated IONP	58
4.9	Schematic of chelating bidentate coordination mode between Fe atom and carboxylate ion.	58
4.10	FTIR spectra of OA-IONP before ligand exchange (black line) and IONP with temporary NOBF ₄ ligand after the first step of ligand exchange (red line).	60
4.11	FTIR spectra comparison of MFO (bottom) and MFO-IONP after the ligand exchange (top) with four different weight ratio of MFO to IONP.	60
4.12	Dispersibility of 1 mg/mL MFO- in PBS (pH 7.4). The MFO-IONP was sonicated for 10 minutes in a bath sonicator to disperse.	63
4.13	TGA of OA-IONP showing mass loss vs temperature.	65
4.14	TGA curve of MFO-IONP showing weight loss vs temperature.	66
4.15	T ₂ -weighted images of MFO-IONP with various concentrations measured using a 3 T MRI scanner	68
4.16	HRTEM image of MFO15-IONP at 50KX magnification showing particle aggregation after ligand exchange	69
4.17	Relaxation rate of MFO-IONP as a function of concentration in mg/mL to determine relaxivity, r ₂ , of MFO15-IONP	70
4.18	Percentage of cell viability of MRC5 and A549 analysed by XTT assay after treatments with MFO-IONP at seven different concentrations.	71
4.19	Amount of Dtx partitioned in five different solid lipids (Precirol ATO 5, Compritol 888 ATO, myristic acid, palmitic acid and stearic acid).	74
4.20	Solubility of Dtx in MCT, Pkoe, olive oil, safflower oil and soybean oil determined using HPLC-UV after dilution with methanol.	75

4.21	Microscopic images of Compritol 888 ATO/ MCT mixture (left) and Precirol ATO 5/ MCT (right) observed at 4KX magnification.	77
4.22	Effect of increasing percentage of Precirol ATO 5 in the binary mixture (Precirol ATO 5 and MCT) on its melting behaviour.	78
4.23	Particle size and Pdl of nanoparticles produced with Lipoid S75/ Poloxamer 188 and Lipoid S75/ TPGS.	81
4.24	Particle size and Pdl of lipid nanoparticles with increasing ratio of ethanol to acetone (v/v).	82
4.25	Three-dimensional response surface plot of process parameters on particle size of nanostructured lipid carrier.	86
4.26	Three-dimensional response surface plot of process parameters on polydispersity index (Pdl) of nanostructured lipid carrier.	87
4.27	HRTEM image of Dtx-MNLC at 50KX magnification.	91
4.28	X-ray diffraction pattern of Precirol ATO5, binary mixture Precirol/MCT, Dtx and Dtx-MNLC.	92
4.29	ATR-FTIR spectra of Dtx and Dtx-MNLC.	94
4.30	DSC thermogram of Precirol ATO5, binary mixture of Precirol/MCT, ternary mixture of Precirol/MCT/Dtx and final formulation of Dtx-MNLC.	95
4.31	Hysteresis curve of MFO-IONP and Dtx-MNLC.	97
4.32	Proposed illustration of Dtx-MNLC structure.	98
4.33	Drug release profiles of CFDtx and Dtx-MNLC using dialysis membrane in SLF at two different pH.	99
4.34	Percentage of cell viability of human lung fibroblast cell lines (MRC5) and human lung carcinoma cell lines (A549) analysed by XTT assay after treatments with commercialised formulation of Dtx or MNLC formulation at seven different concentrations of Dtx.	101
4.35	Percentage of cell viability of human lung fibroblast cell lines (MRC5) and human lung carcinoma cell lines (A549) analysed by XTT assay after treatments with	

	commercialised formulation of Dtx or Dtx-MNLC formulation at seven different concentrations of Dtx.	102
4.36	Schematic illustration of improved therapeutic efficacy by inhibiting P-glycoprotein (P-gp) pump to prevent drug efflux.	104
4.37	DSC thermogram for Dtx-MNLC at day 0 and after 6 months storage at either room temperature (27 °C) or in the refrigerator (2 8 °C).	106
4.38	PXRD diffractogram of Dtx, sucrose and Dtx-MNLC at day 0 and 6 months of storage at either room temperature (27 °C) or in the refrigerator (2 8 °C).	108

LIST OF ABBREVIATIONS

A549	Human lung carcinoma cell
ADC	Adenocarcinoma
ALK	Anaplastic lymphoma kinase
AMF	Alternating magnetic field
ANOVA	Analysis of Variance
ATCC	American Type Culture Collection
ATR-FTIR	Attenuated total reflection - fourier transform infrared spectroscopy
CCD	Central Composite Design
CCM	Cell Culture Media
CT	Computed Tomography
DDS	Drug Delivery System
DL	Drug loading
DLS	Dynamic Light Scattering
DMF	Dimethylformamide
DMSO	Dimethyl sulfoxide
DSC	Differential scanning calorimetry
Dtx	Docetaxel
EE	Entrapment efficiency
EGFR	Epidermal Growth Factor Receptor
EPR	Enhanced permeability and retention
FDA	Food and Drug Administration
GRAS	Generally Recognized as Safe

HLB	Hydrophile-lipophile balance
HPLC	High Performance Liquid Chromatography
HRTEM	High resolution transmission electron microscopy
ICP-OES	Inductively Coupled Plasma Optical Emission Spectroscopy
IONP	Iron oxide nanoparticles
ISO	International Organization of Standardization
LBF	Lipid-based formulation
LCC	Large cell carcinoma
LP	Lipoid S75
MAR	Motion averaging regime
MCT	Medium chain triglyceride
MFO	Menhaden fish oil
MFO-IONP	Menhaden fish oil-coated iron oxide nanoparticle
MNLC	Magnetic nanostructured lipid carrier
MPS	Mononuclear phagocytic system
MRC5	Human lung fibroblast cell
MRI	Magnetic resonance imaging
MWCO	Molecular weight cut-off
NLC	Nanostructured lipid carrier
NSCLC	Non-small cell lung cancer
OA	Oleic acid
OA-IONP	Oleic acid-coated iron oxide nanoparticle
PBS	Phosphate buffer saline
PdI	Polydispersity index

PET	Positron emission tomography
PMS	phenazine methosulfate
PS	Particle size
PTFE	Polytetrafluoroethylene
PXRD	Powder X-ray diffraction
R2	Transverse relaxation rate
RES	Reticuloendothelial system
RF	Radiofrequency
RPMI	Roswell Park Memorial Institute
RSM	Response Surface Methodology
SBRT	Stereotactic body radiotherapy
SCC	Squamous cell carcinoma
SCLC	Small cell lung cancer
SDR	Static dephasing regime
SLF	Simulated lung fluid
SLN	Solid lipid nanoparticle
SMR	Slow motion regime
T1	Longitudinal relaxation time
T2	Transverse relaxation time
TFA	Trifluoroacetic acid
TGA	Thermogravimetric analysis
TME	Tumour microenvironment
TNM	Tumour-Node-Metastasis
TPGS	D- α -tocopheryl polyethylene glycol succinate

VSM

Vibrating sample magnetometer

WHO

World Health Organization



© COPYRIGHT UPM

CHAPTER 1

INTRODUCTION

1.1 Research Background

Until 2020, global lung cancer prevalence was estimated to increase to 2.2 million cases compared to 2.1 million cases in 2018 (Bray et al., 2018). The exact cause of lung cancer is unknown, but high-risk lifestyle such as cigarette smoking has been identified as a major risk factor. Symptoms of lung cancer are similar to other respiratory and lung diseases, which complicates a correct diagnosis; thus, it is often confirmed when the patients have reached stage III and IV. According to Sung et al. (2021), it was estimated that most lung cancer patients die within one year of diagnosis, and only 18% survived within five years due to poor prognosis and less effective treatment.

Standard therapy for lung cancer patients includes surgery, chemotherapy, and radiotherapy. Surgery and radiotherapy are often combined with chemotherapy to avoid recurrent tumour progression for more effective treatment. Anticancer agents such as doxorubicin, cisplatin, and docetaxel (Dtx) have been shown to inhibit solid tumor progression in the lungs, breasts, and prostates, among others (Montero et al., 2005). Nevertheless, clinical applications of these drugs are limited due to low aqueous solubility, serious side effects, and non-specific distribution of the body. For example, the commercial formulation of Dtx (Taxotere®) contains a high amount of surfactant (Tween 80) to increase its aqueous solubility. Although this formulation is proven effective in cancer therapy, it has caused dose-limiting toxicity, and hypersensitivity reactions in patients, as both cancer cells and healthy cells are exposed to the toxicity effects, leading to severe adverse effects such as neurotoxicity and neutropenia (Ho & Mackey, 2014).

The development of nanotechnology in pharmaceutical sciences has opened up endless possibilities for improving cancer treatment. Various colloidal drug delivery systems (DDS) such as micelles, polymeric nanoparticles, dendrimer, and solid lipid nanoparticles (SLN) have shown promising results to entrap hydrophobic and hydrophilic chemotherapeutics for improved biodistribution and better therapeutic response. Among these nanoparticulate delivery systems, lipid-based formulations designed as respirable nanocarriers, such as liposomes (Lin et al., 2017), nanoemulsions (Asmawi et al., 2019), solid lipids nanoparticles (SLN) (Bakhtiary et al., 2017), and nanostructured lipid carrier (NLC) (Ong et al., 2020) were reported to have good lung tolerability and low toxicity due to the biocompatible lipid used in the formulations.

Nanoparticles formulated from solid lipid, e.g., SLN, present high stability in vivo and often demonstrate controlled release kinetics (Duan et al., 2020). However, premature drug leakage from their solid lipid during storage and low drug loading appear to limit their potential as DDS. In 2002, Müller et al. started incorporating liquid lipid in the solid lipid to form NLC, which resulted in higher loading capacity and better stability than SLN. The types of lipid and the ratio of solid and liquid lipid selected to produce NLC were observed to significantly impact their polymorphism, possible existence of supercooled melts, and presence of other colloidal species. Therefore, careful selection of formulation excipients and investigations on their physicochemical properties are required to develop a suitable formulation designed as lung cancer therapy.

Current chemotherapy effectiveness can only be determined upon treatment completion by physical examinations, X-ray/ Computed Tomography (CT) scan, or blood tests. The treatment protocol would be changed later if inadequate chemotherapy response was measured, which consequently could cause the tumour to progress further before an effective treatment regime can be prescribed. Theranostic in oncology offers unique opportunities to provide diagnostic imaging and therapeutic molecules in a single platform. Functionalisation of nanocarriers with magnetic nanoparticles such as iron oxide nanoparticles (IONP) will enable visualisation of tumours and metastases in various organs such as the liver, spleen, and lymph nodes using Magnetic Resonance Imaging (MRI) (Dadfar et al., 2019). A theranostic approach to continuously monitor the treatment response is beneficial for measuring chemotherapy effectiveness.

1.2 Research Objectives

This work describes the development of magnetic nanostructured lipid carrier (MNLC) loaded with Dtx for potential theranostic application in lung cancer treatment. This study is designed to achieve the following objectives:

- I. To optimise the synthesis protocol of oleic acid-coated iron oxide nanoparticles (OA-IONP) using thermal decomposition method based on their measured size and magnetic properties
- II. To determine the physicochemical properties of Menhaden fish oil-coated IONP (MFO-IONP) after ligand exchange
- III. To assess the suitability of MFO-IONP for Magnetic Resonance Imaging (MRI) using agarose phantom
- IV. To develop the formulation of MNLC loaded with MFO-IONP and Dtx with an understanding of its physicochemical properties, release kinetics of drugs and short-term storage stability of formulated MNLC

- V. To study the in vitro cytotoxicity of MFO-IONP and MNLC on non-small cell lung carcinoma (A549) and lung fibroblast cells (MRC5) using colorimetric assay

1.3 Scope of the Study

IONP were used as magnetic nanoparticles in this work. The IONP was chemically synthesised using thermal decomposition of iron oxyhydroxide in the presence of oleic acid as the capping agent and 1-octadecene. The oleic acid coating was then substituted with Menhaden fish oil to improve the biocompatibility of the IONP. The IONP was further embedded in lipid vesicles composed of Precirol ATO 5 as solid lipid, medium chain triglyceride (MCT) as oil, Vitamin E TPGS (TPGS) and soy lecithin (Lipoid S75) as the emulsifier. Dtx, an antineoplastic agent, was used as active pharmaceutical ingredient in this formulation and was solubilised in the lipid component to form MNLC.

Various characterisation techniques were used to investigate the physicochemical properties of the IONP and MNLC. Particle size and dispersity were determined using High Resolution Transmission Electron Microscope (HRTEM) and dynamic light scattering (DLS). Chemical bonding was analysed by Attenuated Reflection-Fourier Transform Infrared spectroscopy (ATR-FTIR). Crystallinity and solid state of the nanoparticles were studied using powder X-ray diffraction (PXRD), thermogravimetric analysis (TGA) and differential scanning calorimetry (DSC). Magnetic properties of IONP and MNLC were characterised by Vibrating Sample Magnetometry (VSM). Concentration of Dtx in the formulation was determined using high performance liquid chromatography equipped with UV detector (HPLC-UV).

In vitro tests were carried out to study the drug release profile of MNLC in simulated lung fluid (SLF) at pH 7.4 and in pH 6.0 to simulate the cancerous microenvironment. Cytotoxicity tests of IONP and MNLC were performed on the normal human fibroblast lung cell (MRC-5) and adenocarcinomic human alveolar basal epithelial cells (A549) using colorimetric cell viability assay.

1.4 Limitation of the Study

This study is only limited to the development, physicochemical analysis, in vitro drug release and in vitro cytotoxicity of IONP and MNLC. The assessment of their aerodynamic properties for inhalational delivery route, in vivo toxicity and MRI-monitored magnetic targeting performance are beyond the scope of this work.

REFERENCES

- Abakumov, M. A., Semkina, A. S., Skorikov, A. S., Vishnevskiy, D. A., Ivanova, A. v., Mironova, E., Davydova, G. A., Majouga, A. G., & Chekhonin, V. P. (2018). Toxicity of iron oxide nanoparticles: Size and coating effects. *Journal of Biochemical and Molecular Toxicology*, 32(12), e22225. <https://doi.org/10.1002/jbt.22225>
- Abdel-Mottaleb, M. M. A., & Lamprecht, A. (2011). Standardized in vitro drug release test for colloidal drug carriers using modified USP dissolution apparatus I. *Drug Development and Industrial Pharmacy*, 37(2), 178–184. <https://doi.org/10.3109/03639045.2010.502534>
- Abdel-Razeq, H. N., Mansour, R. A., Ammar, K. S., Abdel-Razeq, R. H., Zureigat, H. Y., Yousef, L. M., & Shahin, O. A. (2020). Amenorrhea, fertility preservation, and counseling among young women treated with anthracyclines and taxanes for early-stage breast cancer, a retrospective study. *Medicine*, 99(11), e19566. <https://doi.org/10.1097/MD.00000000000019566>
- Alcantara, K. P., Zulfakar, M. H., & Castillo, A. L. (2019). Development, characterization and pharmacokinetics of mupirocin-loaded nanostructured lipid carriers (NLCs) for intravascular administration. *International Journal of Pharmaceutics*, 571, 118705. <https://doi.org/10.1016/j.ijpharm.2019.118705>
- Alipour, A., Soran-Erdem, Z., Utkur, M., Sharma, V. K., Algin, O., Saritas, E. U., & Demir, H. V. (2018). A new class of cubic SPIONs as a dual-mode T1 and T2 contrast agent for MRI. *Magnetic Resonance Imaging*, 49, 16–24. <https://doi.org/10.1016/j.mri.2017.09.013>
- Almalik, A., Alradwan, I., Kalam, M. A., & Alshamsan, A. (2017). Effect of cryoprotection on particle size stability and preservation of chitosan nanoparticles with and without hyaluronate or alginate coating. *Saudi Pharmaceutical Journal*, 25(6), 861–867. <https://doi.org/10.1016/j.jsps.2016.12.008>
- Almousallam, M., Moia, C., & Zhu, H. (2015). Development of nanostructured lipid carrier for dacarbazine delivery. *International Nano Letters*, 5(4), 241–248. <https://doi.org/10.1007/s40089-015-0161-8>
- Alskär, L. C., Porter, C. J. H., & Bergström, C. A. S. (2016). Tools for Early Prediction of Drug Loading in Lipid-Based Formulations. *Molecular Pharmaceutics*, 13(1), 251–261. <https://doi.org/10.1021/acs.molpharmaceut.5b00704>
- Amin, M. L. (2013). P-glycoprotein Inhibition for Optimal Drug Delivery. *Drug Target Insights*, 7, 27–34. <https://doi.org/10.4137/DTI.S12519>

- Arora, M. (2013). Cell Culture Media: A Review. *Materials and Methods*, 3. <https://doi.org/10.13070/mm.en.3.175>
- Asif, S., Kaur, G., Sharma, S., & Awasthi, V. (2020). Oleic acid magnetic iron oxide nanoparticles improve iron uptake by the modification of NADH-HCF (III) oxidoreductase without affecting cellular viability. *Gene Reports*, 21, 100837. <https://doi.org/10.1016/j.genrep.2020.100837>
- Asmawi, A. A. (2020). *Development and Characterization of Docetaxel and Curcumin Loaded Aerosolized Nanoemulsion For Pulmonary Cancer*.
- Asmawi, A. A., Salim, N., Ngan, C. L., Ahmad, H., Abdulmalek, E., Masarudin, M. J., & Abdul Rahman, M. B. (2019). Excipient selection and aerodynamic characterization of nebulized lipid-based nanoemulsion loaded with docetaxel for lung cancer treatment. *Drug Delivery and Translational Research*, 9(2), 543–554. <https://doi.org/10.1007/s13346-018-0526-4>
- Attia, M. F., Anton, N., Wallyn, J., Omran, Z., & Vandamme, T. F. (2019). An overview of active and passive targeting strategies to improve the nanocarriers efficiency to tumour sites. *Journal of Pharmacy and Pharmacology*, 71(8), 1185–1198. <https://doi.org/https://doi.org/10.1111/jphp.13098>
- Aw, D. C.-W., Tan, E. H., Chin, T. M., Lim, H. L., Lee, H. Y., & Soo, R. A. (2018). Management of epidermal growth factor receptor tyrosine kinase inhibitor-related cutaneous and gastrointestinal toxicities. *Asia-Pacific Journal of Clinical Oncology*, 14(1), 23–31. <https://doi.org/10.1111/ajco.12687>
- Babes, L., Denizot, B., Tanguy, G., Le Jeune, J. J., & Jallet, P. (1999). Synthesis of Iron Oxide Nanoparticles Used as MRI Contrast Agents: A Parametric Study. *Journal of Colloid and Interface Science*, 212(2), 474–482. <https://doi.org/10.1006/jcis.1998.6053>
- Bae, K. H., Lee, J. Y., Lee, S. H., Park, T. G., & Nam, Y. S. (2013). Optically Traceable Solid Lipid Nanoparticles Loaded with siRNA and Paclitaxel for Synergistic Chemotherapy with In situ Imaging. *Advanced Healthcare Materials*, 2(4), 576–584. <https://doi.org/10.1002/adhm.201200338>
- Bailly, C. (2019). Irinotecan: 25 years of cancer treatment. *Pharmacological Research*, 148, 104398. <https://doi.org/10.1016/j.phrs.2019.104398>
- Bakhtiary, Z., Barar, J., Aghanejad, A., Saei, A. A., Nemati, E., Ezzati Nazhad Dolatabadi, J., & Omidi, Y. (2017). Microparticles containing erlotinib-loaded solid lipid nanoparticles for treatment of non-small cell lung cancer. *Drug Development and Industrial Pharmacy*, 43(8), 1244–1253. <https://doi.org/10.1080/03639045.2017.1310223>
- Balavandy, S. K., Shameli, K., Biak, D. R. B. A., & Abidin, Z. Z. (2014). Stirring time effect of silver nanoparticles prepared in glutathione mediated by

green method. *Chemistry Central Journal*, 8(1), 11.
<https://doi.org/10.1186/1752-153X-8-11>

- Ball, R., Bajaj, P., & Whitehead, K. (2016). Achieving long-term stability of lipid nanoparticles: examining the effect of pH, temperature, and lyophilization. *International Journal of Nanomedicine*, Volume 12, 305–315.
<https://doi.org/10.2147/IJN.S123062>
- Banerjee, P., Geng, T., Mahanty, A., Li, T., Zong, L., & Wang, B. (2019). Integrating the drug, disulfiram into the vitamin E-TPGS-modified PEGylated nanostructured lipid carriers to synergize its repurposing for anti-cancer therapy of solid tumors. *International Journal of Pharmaceutics*, 557, 374–389. <https://doi.org/10.1016/j.ijpharm.2018.12.051>
- Barenholz, Y. (Chezy). (2012). Doxil® — The first FDA-approved nano-drug: Lessons learned. *Journal of Controlled Release*, 160(2), 117–134.
<https://doi.org/https://doi.org/10.1016/j.jconrel.2012.03.020>
- Bashir, M. R., Bhatti, L., Marin, D., & Nelson, R. C. (2015). Emerging applications for ferumoxytol as a contrast agent in MRI. *Journal of Magnetic Resonance Imaging: JMRI*, 41(4), 884–898. <https://doi.org/10.1002/jmri.24691>
- Beck, W., & Suenkel, K. (1988). Metal complexes of weakly coordinating anions. Precursors of strong cationic organometallic Lewis acids. *Chemical Reviews*, 88(7), 1405–1421. <https://doi.org/10.1021/cr00089a017>
- Becker, K., Salar-Behzadi, S., & Zimmer, A. (2015). Solvent-Free Melting Techniques for the Preparation of Lipid-Based Solid Oral Formulations. *Pharmaceutical Research*, 32(5), 1519–1545.
<https://doi.org/10.1007/s11095-015-1661-y>
- Belaïd, S., Stanicki, D., vander Elst, L., Muller, R. N., & Laurent, S. (2018). Influence of experimental parameters on iron oxide nanoparticle properties synthesized by thermal decomposition: size and nuclear magnetic resonance studies. *Nanotechnology*, 29(16), 165603.
<https://doi.org/10.1088/1361-6528/aaae59>
- Bestebreurtje, P., Roeleveld, N., Knibbe, C. A. J., van Sorge, A. A., Plötz, F. B., & de Wildt, S. N. (2020). Development and Stability Study of an Omeprazole Suppository for Infants. *European Journal of Drug Metabolism and Pharmacokinetics*, 45(5), 627–633. <https://doi.org/10.1007/s13318-020-00629-1>
- Bodratti, A., & Alexandridis, P. (2018). Formulation of Poloxamers for Drug Delivery. *Journal of Functional Biomaterials*, 9(1), 11.
<https://doi.org/10.3390/jfb9010011>
- Borges, G. S. M., Silva, J. de O., Fernandes, R. S., de Souza, Â. M., Cassali, G. D., Yoshida, M. I., Leite, E. A., de Barros, A. L. B., & Ferreira, L. A. M. (2019). Sclareol is a potent enhancer of doxorubicin: Evaluation of the free

- combination and co-loaded nanostructured lipid carriers against breast cancer. *Life Sciences*, 232, 116678. <https://doi.org/10.1016/j.lfs.2019.116678>
- Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R. L., Torre, L. A., & Jemal, A. (2018). Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: A Cancer Journal for Clinicians*, 68(6), 394–424. <https://doi.org/10.3322/caac.21492>
- Brian C. Smith. (2018). *The C=O Bond, Part VI: Esters and the Rule of Three*. Spectroscopy. <https://www.spectroscopyonline.com/view/co-bond-part-vi-esters-and-rule-three>
- Bruschi, M. L. B. T.-S. to M. the D. R. from P. S. (Ed.). (2015). Mathematical models of drug release. In *Strategies to Modify the Drug Release from Pharmaceutical Systems* (pp. 63–86). Woodhead Publishing. <https://doi.org/https://doi.org/10.1016/B978-0-08-100092-2.00005-9>
- Bulte, J. W. M. (2019). Superparamagnetic iron oxides as MPI tracers: A primer and review of early applications. *Advanced Drug Delivery Reviews*, 138, 293–301. <https://doi.org/10.1016/j.addr.2018.12.007>
- Cano, M., Sbagoud, K., Allard, E., & Larpent, C. (2012). Magnetic separation of fatty acids with iron oxide nanoparticles and application to extractive deacidification of vegetable oils. *Green Chemistry*, 14(6), 1786–1795. <https://doi.org/10.1039/c2gc35270b>
- Cao, X., Ganti, A. K., Stinchcombe, T., Wong, M. L., Ho, J. C., Shen, C., Liu, Y., Crawford, J., Pang, H., & Wang, X. (2020). Predicting risk of chemotherapy-induced severe neutropenia: A pooled analysis in individual patients data with advanced lung cancer. *Lung Cancer*, 141, 14–20. <https://doi.org/10.1016/j.lungcan.2020.01.004>
- Chalasanani, R., & Vasudevan, S. (2011). Form, content, and magnetism in iron oxide nanocrystals. *Journal of Physical Chemistry C*, 115(37), 18088–18093. <https://doi.org/10.1021/jp204697f>
- Charron, D. M., Chen, J., & Zheng, G. (2015). Theranostic Lipid Nanoparticles for Cancer Medicine. In C. A. Mirkin, T. J. Meade, S. H. Petrosko, & A. H. Stegh (Eds.), *Nanotechnology-Based Precision Tools for the Detection and Treatment of Cancer* (pp. 103–127). Springer International Publishing. https://doi.org/10.1007/978-3-319-16555-4_5
- Coates, J. (2006). Interpretation of Infrared Spectra, A Practical Approach. In *Encyclopedia of Analytical Chemistry*. John Wiley & Sons, Ltd. <https://doi.org/10.1002/9780470027318.a5606>
- Correia, A., Costa, C. P., Silva, V., Silva, R., Lobo, J. M. S., & Silva, A. C. (2020). Pessaries containing nanostructured lipid carriers (NLC) for prolonged vaginal delivery of progesterone. *European Journal of Pharmaceutical*

- Costo, R., Heinke, D., Grüttner, C., Westphal, F., Morales, M. P., Veintemillas-Verdaguer, S., & Gehrke, N. (2019). Improving the reliability of the iron concentration quantification for iron oxide nanoparticle suspensions: a two-institutions study. *Analytical and Bioanalytical Chemistry*, 411(9), 1895–1903. <https://doi.org/10.1007/s00216-018-1463-2>
- Cryer, A. M., & Thorley, A. J. (2019). Nanotechnology in the diagnosis and treatment of lung cancer. *Pharmacology & Therapeutics*, 198, 189–205. <https://doi.org/10.1016/j.pharmthera.2019.02.010>
- Czajkowska-Kośnik, A., Szymańska, E., Czarnomysy, R., Jacyna, J., Markuszewski, M., Basa, A., & Winnicka, K. (2021). Nanostructured Lipid Carriers Engineered as Topical Delivery of Etodolac: Optimization and Cytotoxicity Studies. *Materials*, 14(3), 596. <https://doi.org/10.3390/ma14030596>
- Dadfar, S. M., Camozzi, D., Darguzyte, M., Roemhild, K., Varvarà, P., Metselaar, J., Banala, S., Straub, M., Güvener, N., Engelmann, U., Slabu, I., Buhl, M., van Leusen, J., Kögerler, P., Hermanns-Sachweh, B., Schulz, V., Kiessling, F., & Lammers, T. (2020). Size-isolation of superparamagnetic iron oxide nanoparticles improves MRI, MPI and hyperthermia performance. *Journal of Nanobiotechnology*, 18(1), 22. <https://doi.org/10.1186/s12951-020-0580-1>
- Dadfar, S. M., Roemhild, K., Drude, N. I., von Stillfried, S., Knüchel, R., Kiessling, F., & Lammers, T. (2019). Iron oxide nanoparticles: Diagnostic, therapeutic and theranostic applications. *Advanced Drug Delivery Reviews*, 138, 302–325. <https://doi.org/10.1016/j.addr.2019.01.005>
- Danaei, M., Dehghankhold, M., Ataei, S., Hasanzadeh Davarani, F., Javanmard, R., Dokhani, A., Khorasani, S., & Mozafari, M. (2018). Impact of Particle Size and Polydispersity Index on the Clinical Applications of Lipidic Nanocarrier Systems. *Pharmaceutics*, 10(2), 57. <https://doi.org/10.3390/pharmaceutics10020057>
- Das, S., Diyali, S., Vinothini, G., Perumalsamy, B., Balakrishnan, G., Ramasamy, T., Dharumadurai, D., & Biswas, B. (2020). Synthesis, morphological analysis, antibacterial activity of iron oxide nanoparticles and the cytotoxic effect on lung cancer cell line. *Heliyon*, 6(9), e04953. <https://doi.org/https://doi.org/10.1016/j.heliyon.2020.e04953>
- Date, P. V., Samad, A., & Devarajan, P. V. (2010). Freeze Thaw: A Simple Approach for Prediction of Optimal Cryoprotectant for Freeze Drying. *AAPS PharmSciTech*, 11(1), 304–313. <https://doi.org/10.1208/s12249-010-9382-3>

- de Haan, H. W. (2011). Mechanisms of proton spin dephasing in a system of magnetic particles. *Magnetic Resonance in Medicine*, 66(6), 1748–1758. <https://doi.org/10.1002/mrm.22966>
- de Jonge, M. J. A., Slingerland, M., Loos, W. J., Wiemer, E. A. C., Burger, H., Mathijssen, R. H. J., Kroep, J. R., den Hollander, M. A. G., van der Biessen, D., Lam, M.-H., Verweij, J., & Gelderblom, H. (2010). Early cessation of the clinical development of LiPlaCis, a liposomal cisplatin formulation. *European Journal of Cancer*, 46(16), 3016–3021. <https://doi.org/https://doi.org/10.1016/j.ejca.2010.07.015>
- de Oliveira, I. F., Barbosa, E. J., Peters, M. C. C., Henostroza, M. A. B., Yukuyama, M. N., dos Santos Neto, E., Löbenberg, R., & Bou-Chacra, N. (2020). Cutting-edge advances in therapy for the posterior segment of the eye: Solid lipid nanoparticles and nanostructured lipid carriers. *International Journal of Pharmaceutics*, 589, 119831. <https://doi.org/https://doi.org/10.1016/j.ijpharm.2020.119831>
- D'Eliseo, D., & Velotti, F. (2016). Omega-3 Fatty Acids and Cancer Cell Cytotoxicity: Implications for Multi-Targeted Cancer Therapy. *Journal of Clinical Medicine*, 5(2), 15. <https://doi.org/10.3390/jcm5020015>
- Demirer, G. S., Okur, A. C., & Kizilel, S. (2015). Synthesis and design of biologically inspired biocompatible iron oxide nanoparticles for biomedical applications. *J. Mater. Chem. B*, 3(40), 7831–7849. <https://doi.org/10.1039/C5TB00931F>
- Deshpande, A., Mohamed, M., Daftardar, S. B., Patel, M., Boddu, S. H. S., & Nesamony, J. (2017). Chapter 12 - Solid Lipid Nanoparticles in Drug Delivery: Opportunities and Challenges. In A. K. Mitra, K. Cholkar, & A. B. T.-E. N. for D. Mandal Drug Delivery and Medical Devices (Eds.), *Micro and Nano Technologies* (pp. 291–330). Elsevier. <https://doi.org/https://doi.org/10.1016/B978-0-323-42978-8.00012-7>
- Dewhirst, M. W., & Secomb, T. W. (2017). Transport of drugs from blood vessels to tumour tissue. *Nature Reviews Cancer*, 17(12), 738–750. <https://doi.org/10.1038/nrc.2017.93>
- Di Marco, M., Sadun, C., Port, M., Guilbert, I., Couvreur, P., & Dubernet, C. (2007). Physicochemical characterization of ultrasmall superparamagnetic iron oxide particles (USPIO) for biomedical application as MRI contrast agents. *International Journal of Nanomedicine*, 2(4), 609–622.
- do Prado, A. H., Araújo, V. H. S., Eloy, J. O., Fonseca-Santos, B., Pereira-da-Silva, M. A., Peccinini, R. G., & Chorilli, M. (2020). Synthesis and Characterization of Nanostructured Lipid Nanocarriers for Enhanced Sun Protection Factor of Octyl p-methoxycinnamate. *AAPS PharmSciTech*, 21(4), 125. <https://doi.org/10.1208/s12249-019-1547-0>

- Dolatabadi, S., Karimi, M., Nasirizadeh, S., Hatamipour, M., Golmohammadzadeh, S., & Jaafari, M. R. (2021). Preparation, characterization and in vivo pharmacokinetic evaluation of curcuminoids-loaded solid lipid nanoparticles (SLNs) and nanostructured lipid carriers (NLCs). *Journal of Drug Delivery Science and Technology*, 62, 102352. <https://doi.org/10.1016/j.jddst.2021.102352>
- Dong, A., Ye, X., Chen, J., Kang, Y., Gordon, T., Kikkawa, J. M., & Murray, C. B. (2011). A generalized ligand-exchange strategy enabling sequential surface functionalization of colloidal nanocrystals. *Journal of the American Chemical Society*, 133(4), 998–1006. <https://doi.org/10.1021/jal08948z>
- Duan, Y., Dhar, A., Patel, C., Khimani, M., Neogi, S., Sharma, P., Siva Kumar, N., & Vekariya, R. L. (2020). A brief review on solid lipid nanoparticles: part and parcel of contemporary drug delivery systems. *RSC Advances*, 10(45), 26777–26791. <https://doi.org/10.1039/D0RA03491F>
- Dulińska-Litewka, J., Łazarczyk, A., Hałubiec, P., Szafranski, O., Karnas, K., & Karewicz, A. (2019). Superparamagnetic Iron Oxide Nanoparticles—Current and Prospective Medical Applications. *Materials*, 12(4), 617. <https://doi.org/10.3390/ma12040617>
- Duong, V.-A., Nguyen, T.-T.-L., & Maeng, H.-J. (2020). Preparation of Solid Lipid Nanoparticles and Nanostructured Lipid Carriers for Drug Delivery and the Effects of Preparation Parameters of Solvent Injection Method. *Molecules*, 25(20), 4781. <https://doi.org/10.3390/molecules25204781>
- Dwivedi, S., Siddiqui, M. A., Farshori, N. N., Ahamed, M., Musarrat, J., & Al-Khedhairi, A. A. (2014). Synthesis, characterization and toxicological evaluation of iron oxide nanoparticles in human lung alveolar epithelial cells. *Colloids and Surfaces B: Biointerfaces*, 122, 209–215. <https://doi.org/https://doi.org/10.1016/j.colsurfb.2014.06.064>
- Edmondson, R., Broglie, J. J., Adcock, A. F., & Yang, L. (2014). Three-dimensional cell culture systems and their applications in drug discovery and cell-based biosensors. *Assay and Drug Development Technologies*, 12(4), 207–218. <https://doi.org/10.1089/adt.2014.573>
- Eid, R. K., Ashour, D. S., Essa, E. A., el Maghraby, G. M., & Arafa, M. F. (2020). Chitosan coated nanostructured lipid carriers for enhanced in vivo efficacy of albendazole against *Trichinella spiralis*. *Carbohydrate Polymers*, 232, 115826. <https://doi.org/10.1016/j.carbpol.2019.115826>
- Emami, J., Mohiti, H., Hamishehkar, H., & Varshosaz, J. (2015). Formulation and optimization of solid lipid nanoparticle formulation for pulmonary delivery of budesonide using Taguchi and Box-Behnken design. *Research in Pharmaceutical Sciences*, 10(1), 17–33.
- Evrard, B., Amighi, K., Beten, D., Delattre, L., & Moës, A. J. (1999). Influence of Melting and Rheological Properties of Fatty Binders on the Melt

Granulation Process in a High-Shear Mixer. *Drug Development and Industrial Pharmacy*, 25(11), 1177–1184. <https://doi.org/10.1081/DDC-100102285>

Fang, G., Tang, B., Chao, Y., Zhang, Y., Xu, H., & Tang, X. (2015). Improved oral bioavailability of docetaxel by nanostructured lipid carriers: in vitro characteristics, in vivo evaluation and intestinal transport studies. *RSC Advances*, 5(117), 96437–96447. <https://doi.org/10.1039/c5ra14588k>

Feng, Q., Liu, Y., Huang, J., Chen, K., Huang, J., & Xiao, K. (2018). Uptake, distribution, clearance, and toxicity of iron oxide nanoparticles with different sizes and coatings. *Scientific Reports*, 8(1), 1–13. <https://doi.org/10.1038/s41598-018-19628-z>

Ferreira, M., Chaves, L. L., Lima, S. A. C., & Reis, S. (2015). Optimization of nanostructured lipid carriers loaded with methotrexate: A tool for inflammatory and cancer therapy. *International Journal of Pharmaceutics*, 492(1–2), 65–72. <https://doi.org/10.1016/j.ijpharm.2015.07.013>

Fessi, H. C., Devissaguet, J.-P., & F. Puisieux, C. T. (1992). *Process for the preparation of dispersible colloidal systems of a substance in the form of nanoparticles* (Patent No. 5118528). US Patent.

Frampton, J. E. (2020). Liposomal Irinotecan: A Review in Metastatic Pancreatic Adenocarcinoma. *Drugs*, 80(10), 1007–1018. <https://doi.org/10.1007/s40265-020-01336-6>

Fundamentals of Magnetic Resonance Imaging. (2008). In *Breast MRI: Fundamentals and Technical Aspects* (pp. 1–17). Springer New York. https://doi.org/10.1007/978-0-387-73507-8_1

Gala, U. H., Miller, D. A., & Williams, R. O. (2020). Harnessing the therapeutic potential of anticancer drugs through amorphous solid dispersions. *Biochimica et Biophysica Acta (BBA) - Reviews on Cancer*, 1873(1), 188319. <https://doi.org/https://doi.org/10.1016/j.bbcan.2019.188319>

García-Fernández, C., Fornaguera, C., & Borrós, S. (2020). Nanomedicine in Non-Small Cell Lung Cancer: From Conventional Treatments to Immunotherapy. *Cancers*, 12(6), 1609. <https://doi.org/10.3390/cancers12061609>

Gattefossé. (2020). *Labrafac™ lipophile WL 1349*. Product Information. <https://www.gattefosse.com/pharmaceuticals-products/labrafac-lipophile-wl-1349>

Gazdar, A. F., Bunn, P. A., & Minna, J. D. (2017). Small-cell lung cancer: what we know, what we need to know and the path forward. *Nature Reviews Cancer*, 17(12), 725–737. <https://doi.org/10.1038/nrc.2017.87>

- Gordillo-Galeano, A., & Mora-Huertas, C. E. (2018). Solid lipid nanoparticles and nanostructured lipid carriers: A review emphasizing on particle structure and drug release. *European Journal of Pharmaceutics and Biopharmaceutics*, 133, 285–308. <https://doi.org/10.1016/j.ejpb.2018.10.017>
- Grover, V. P. B., Tognarelli, J. M., Crossey, M. M. E., Cox, I. J., Taylor-Robinson, S. D., & McPhail, M. J. W. (2015). Magnetic Resonance Imaging: Principles and Techniques: Lessons for Clinicians. *Journal of Clinical and Experimental Hepatology*, 5(3), 246–255. <https://doi.org/10.1016/j.jceh.2015.08.001>
- Gu, L., Zhong, D., Yu, T., Tang, P., Meng, F., & Qin, Q. (2019). Retrospective study of the efficacy and toxicity of lobaplatin-etoposide chemotherapy in small cell lung cancer. *Thoracic Cancer*, 10(2), 226–233. <https://doi.org/10.1111/1759-7714.12936>
- Guadagnini, R., Moreau, K., Hussain, S., Marano, F., & Boland, S. (2015). Toxicity evaluation of engineered nanoparticles for medical applications using pulmonary epithelial cells. *Nanotoxicology*, 9(sup1), 25–32. <https://doi.org/10.3109/17435390.2013.855830>
- Guo, Y., Luo, J., Tan, S., Otieno, B. O., & Zhang, Z. (2013). The applications of Vitamin e TPGS in drug delivery. *European Journal of Pharmaceutical Sciences*, 49(2), 175–186. <https://doi.org/10.1016/j.ejps.2013.02.006>
- Hajesmaeelzadeh, F., Shanehsazzadeh, S., Grüttner, C., Daha, F. J., & Oghabian, M. A. (2016). Effect of coating thickness of iron oxide nanoparticles on their relaxivity in the MRI. *Iranian Journal of Basic Medical Sciences*, 19(2), 166–171.
- Hamdani, J., Moës, A. J., & Amighi, K. (2003). Physical and thermal characterisation of Precirol® and Compritol® as lipophilic glycerides used for the preparation of controlled-release matrix pellets. *International Journal of Pharmaceutics*, 260(1), 47–57. [https://doi.org/10.1016/S0378-5173\(03\)00229-1](https://doi.org/10.1016/S0378-5173(03)00229-1)
- Han, F., Li, S., Yin, R., Liu, H., & Xu, L. (2008). Effect of surfactants on the formation and characterization of a new type of colloidal drug delivery system: Nanostructured lipid carriers. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 315(1–3), 210–216. <https://doi.org/10.1016/j.colsurfa.2007.08.005>
- Han, L., Zhan, H., Sun, X., Zhang, Z.-R., & Deng, L. (2019). A Density-Changing Centrifugation Method for Efficient Separation of Free Drugs from Drug-Loaded Particulate Delivery Systems. *The AAPS Journal*, 21(3), 33. <https://doi.org/10.1208/s12248-019-0306-1>
- Hassan, S. A., Palaskas, N., Kim, P., Iliescu, C., Lopez-Mattei, J., Mouhayar, E., Mougdil, R., Thompson, K., Banchs, J., & Yusuf, S. W. (2018).

Chemotherapeutic Agents and the Risk of Ischemia and Arterial Thrombosis. *Current Atherosclerosis Reports*, 20(2), 10. <https://doi.org/10.1007/s11883-018-0702-5>

Helgason, T., Awad, T. S., Kristbergsson, K., McClements, D. J., & Weiss, J. (2009). Effect of surfactant surface coverage on formation of solid lipid nanoparticles (SLN). *Journal of Colloid and Interface Science*, 334(1), 75–81. <https://doi.org/https://doi.org/10.1016/j.jcis.2009.03.012>

Heydenreich, A. V., Westmeier, R., Pedersen, N., Poulsen, H. S., & Kristensen, H. G. (2003). Preparation and purification of cationic solid lipid nanospheres—effects on particle size, physical stability and cell toxicity. *International Journal of Pharmaceutics*, 254(1), 83–87. [https://doi.org/10.1016/S0378-5173\(02\)00688-9](https://doi.org/10.1016/S0378-5173(02)00688-9)

Hientz, K., Mohr, A., Bhakta-Guha, D., & Efferth, T. (2017). The role of p53 in cancer drug resistance and targeted chemotherapy. *Oncotarget*, 8(5), 8921–8946. <https://doi.org/10.18632/oncotarget.13475>

Ho, M. Y., & Mackey, J. R. (2014). Presentation and management of docetaxel-related adverse effects in patients with breast cancer. *Cancer Management and Research*, 6, 253–259. <https://doi.org/10.2147/CMAR.S40601>

Holder, C. F., & Schaak, R. E. (2019). Tutorial on Powder X-ray Diffraction for Characterizing Nanoscale Materials. *ACS Nano*, 13(7), 7359–7365. <https://doi.org/10.1021/acsnano.9b05157>

Hufschmid, R., Arami, H., Ferguson, R. M., Gonzales, M., Teeman, E., Brush, L. N., Browning, N. D., & Krishnan, K. M. (2015). Synthesis of phase-pure and monodisperse iron oxide nanoparticles by thermal decomposition. *Nanoscale*, 7(25), 11142–11154. <https://doi.org/10.1039/c5nr01651g>

Hussain, S. (2016). Nanomedicine for Treatment of Lung Cancer. In *Lung Cancer and Personalized Medicine: Novel Therapies and Clinical Management* (Vol. 890, pp. 137–147). https://doi.org/10.1007/978-3-319-24932-2_8

Huynh, L., Grant, J., Leroux, J.-C., Delmas, P., & Allen, C. (2008). Predicting the Solubility of the Anti-Cancer Agent Docetaxel in Small Molecule Excipients using Computational Methods. *Pharmaceutical Research*, 25(1), 147–157. <https://doi.org/10.1007/s11095-007-9412-3>

Hyeon, T., Lee, S. S., Park, J., Chung, Y., & Na, H. Bin. (2001). Synthesis of Highly Crystalline and Monodisperse Maghemite Nanocrystallites without a Size-Selection Process. *Journal of the American Chemical Society*, 123(51), 12798–12801. <https://doi.org/10.1021/ja016812s>

Ibarra-Sánchez, J. J., Fuentes-Ramírez, R., Roca, A. G., del Puerto Morales, M., & Cabrera-Lara, L. I. (2013). Key Parameters for Scaling up the Synthesis of Magnetite Nanoparticles in Organic Media: Stirring Rate and Growth

Kinetic. *Industrial & Engineering Chemistry Research*, 52(50), 17841–17847. <https://doi.org/10.1021/ie403250p>

International Organization for Standardization [ISO]. (2009). *Biological evaluation of medical devices — Part 5: Tests for in vitro cytotoxicity (ISO 10993-5:2009)*.

Islan, G. A., Gonçalves, L. M. D., Marto, J., Duarte, A., Alvarez, V. A., Castro, G. R., & Almeida, A. J. (2021). Effect of α -tocopherol on the physicochemical, antioxidant and antibacterial properties of levofloxacin loaded hybrid lipid nanocarriers. *New Journal of Chemistry*, 45(2), 1029–1042. <https://doi.org/10.1039/d0nj03781h>

Jaiswal, P., Gidwani, B., & Vyas, A. (2016). Nanostructured lipid carriers and their current application in targeted drug delivery. *Artificial Cells, Nanomedicine, and Biotechnology*, 44(1), 27–40. <https://doi.org/10.3109/21691401.2014.909822>

Jayasankar, K., Pandey, A., Mishra, B. K., & Das, S. (2016). Evaluation of microstructural parameters of nanocrystalline Y2O3 by X-ray diffraction peak broadening analysis. *Materials Chemistry and Physics*, 171, 195–200. <https://doi.org/10.1016/j.matchemphys.2016.01.005>

Jensen, C., & Teng, Y. (2020). Is It Time to Start Transitioning From 2D to 3D Cell Culture? *Frontiers in Molecular Biosciences*, 7, 33. <https://doi.org/10.3389/fmolb.2020.00033>

Jeon, M., Halbert, M. V., Stephen, Z. R., & Zhang, M. (2020). Iron Oxide Nanoparticles as T 1 Contrast Agents for Magnetic Resonance Imaging: Fundamentals, Challenges, Applications, and Prospectives. *Advanced Materials*, 1906539. <https://doi.org/10.1002/adma.201906539>

Jhaveri, K. D., Wanchoo, R., Sakhiya, V., Ross, D. W., & Fishbane, S. (2017). Adverse Renal Effects of Novel Molecular Oncologic Targeted Therapies: A Narrative Review. *Kidney International Reports*, 2(1), 108–123. <https://doi.org/10.1016/j.ekir.2016.09.055>

Jiang, K., Zhang, L., & Bao, G. (2021). Magnetic iron oxide nanoparticles for biomedical applications. *Current Opinion in Biomedical Engineering*, 20, 100330. <https://doi.org/10.1016/j.cobme.2021.100330>

Jindal, A. B., & Devarajan, P. V. (2015). Asymmetric lipid–polymer particles (LIPOMER) by modified nanoprecipitation: role of non-solvent composition. *International Journal of Pharmaceutics*, 489(1), 246–251. <https://doi.org/https://doi.org/10.1016/j.ijpharm.2015.04.073>

Joos, A., Löwa, N., Wiekhorst, F., Gleich, B., & Haase, A. (2017). Size-dependent MR relaxivities of magnetic nanoparticles. *Journal of Magnetism and Magnetic Materials*, 427, 122–126. <https://doi.org/10.1016/j.jmmm.2016.11.021>

- Joshi, B. L., Zielbauer, B. I., & Vilgis, T. A. (2020). Comparative Study on Mixing Behavior of Binary Mixtures of Cocoa Butter/Tristearin (CB/TS) and Cocoa Butter/Coconut Oil (CB/CO). *Foods*, 9(3), 327. <https://doi.org/10.3390/foods9030327>
- Joshi, M. D., & Müller, R. H. (2009). Lipid nanoparticles for parenteral delivery of actives. *European Journal of Pharmaceutics and Biopharmaceutics*, 71(2), 161–172. <https://doi.org/https://doi.org/10.1016/j.ejpb.2008.09.003>
- Jović Orsini, N., Babić-Stojić, B., Spasojević, V., Calatayud, M. P., Cvjetičanin, N., & Goya, G. F. (2018). Magnetic and power absorption measurements on iron oxide nanoparticles synthesized by thermal decomposition of Fe(acac)₃. *Journal of Magnetism and Magnetic Materials*, 449, 286–296. <https://doi.org/10.1016/j.jmmm.2017.10.053>
- Kan, C. S., & Chan, K. M. J. (2016). A Review of Lung Cancer Research in Malaysia. *The Medical Journal of Malaysia*, 71(Suppl 1), 70–78.
- Kan, P., Chen, Z.-B., Lee, C.-J., & Chu, I.-M. (1999). Development of nonionic surfactant/phospholipid o/w emulsion as a paclitaxel delivery system. *Journal of Controlled Release*, 58(3), 271–278. [https://doi.org/10.1016/S0168-3659\(98\)00164-3](https://doi.org/10.1016/S0168-3659(98)00164-3)
- Karn-orachai, K., Smith, S. M., Saesoo, S., Treethong, A., Puttipipatkachorn, S., Pratontep, S., & Ruktanonchai, U. R. (2016). Surfactant effect on the physicochemical characteristics of γ -oryanol-containing solid lipid nanoparticles. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 488, 118–128. <https://doi.org/https://doi.org/10.1016/j.colsurfa.2015.10.011>
- Kasongo, K. W., Müller, R. H., & Walker, R. B. (2012). The use of hot and cold high pressure homogenization to enhance the loading capacity and encapsulation efficiency of nanostructured lipid carriers for the hydrophilic antiretroviral drug, didanosine for potential administration to paediatric patients. *Pharmaceutical Development and Technology*, 17(3), 353–362. <https://doi.org/10.3109/10837450.2010.542163>
- Kaur, G., & Mehta, S. K. (2017). Developments of Polysorbate (Tween) based microemulsions: Preclinical drug delivery, toxicity and antimicrobial applications. *International Journal of Pharmaceutics*, 529(1), 134–160. <https://doi.org/https://doi.org/10.1016/j.ijpharm.2017.06.059>
- Kelidari, H. R., Saeedi, M., Hajheydari, Z., Akbari, J., Morteza-Semnani, K., Akhtari, J., Valizadeh, H., Asare-Addo, K., & Nokhodchi, A. (2016). Spironolactone loaded nanostructured lipid carrier gel for effective treatment of mild and moderate acne vulgaris: A randomized, double-blind, prospective trial. *Colloids and Surfaces B: Biointerfaces*, 146, 47–53. <https://doi.org/https://doi.org/10.1016/j.colsurfb.2016.05.042>

- Keng, P. S., Basri, M., Zakaria, M. R. S., Rahman, M. B. A., Ariff, A. B., Rahman, R. N. Z. A., & Salleh, A. B. (2009). Newly synthesized palm esters for cosmetics industry. *Industrial Crops and Products*, 29(1), 37–44. <https://doi.org/10.1016/j.indcrop.2008.04.002>
- Khan, A., Mudassir, J., Akhtar, S., Murugaiyah, V., & Darwis, Y. (2019). Freeze-Dried Lopinavir-Loaded Nanostructured Lipid Carriers for Enhanced Cellular Uptake and Bioavailability: Statistical Optimization, in Vitro and in Vivo Evaluations. *Pharmaceutics*, 11(2), 97. <https://doi.org/10.3390/pharmaceutics11020097>
- Khan, M. I., Mohammad, A., Patil, G., Naqvi, S. A. H., Chauhan, L. K. S., & Ahmad, I. (2012). Induction of ROS, mitochondrial damage and autophagy in lung epithelial cancer cells by iron oxide nanoparticles. *Biomaterials*, 33(5), 1477–1488. <https://doi.org/https://doi.org/10.1016/j.biomaterials.2011.10.080>
- Khosa, A., Reddi, S., & Saha, R. N. (2018). Nanostructured lipid carriers for site-specific drug delivery. *Biomedicine & Pharmacotherapy*, 103, 598–613. <https://doi.org/10.1016/j.biopha.2018.04.055>
- Kim, C. H., Kang, T. H., Kim, B. D., Lee, T. H., Yoon, H. Y., Goo, Y. T., Choi, Y. S., Kang, M. J., & Choi, Y. W. (2020). Enhanced docetaxel delivery using sterically stabilized RIPL peptide-conjugated nanostructured lipid carriers: In vitro and in vivo antitumor efficacy against SKOV3 ovarian cancer cells. *International Journal of Pharmaceutics*, 583, 119393. <https://doi.org/10.1016/j.ijpharm.2020.119393>
- Kim, E. S. (2016). Chemotherapy Resistance in Lung Cancer. In A. Ahmad & S. Gadgeel (Eds.), *Lung Cancer and Personalized Medicine. Advances in Experimental Medicine and Biology*, vol. 893 (pp. 189–209). Springer, Cham. https://doi.org/10.1007/978-3-319-24223-1_10
- Kobayashi, H., Watanabe, R., & Choyke, P. L. (2014). Improving Conventional Enhanced Permeability and Retention (EPR) Effects; What Is the Appropriate Target? *Theranostics*, 4(1), 81–89. <https://doi.org/10.7150/thno.7193>
- Köhler, T., Feoktystov, A., Petravic, O., Kentzinger, E., Bhatnagar-Schöffmann, T., Feygenson, M., Nandakumaran, N., Landers, J., Wende, H., Cervellino, A., Rücker, U., Kovács, A., Dunin-Borkowski, R. E., & Brückel, T. (2021). Mechanism of magnetization reduction in iron oxide nanoparticles. *Nanoscale*, 13(14), 6965–6976. <https://doi.org/10.1039/D0NR08615K>
- Komaiko, J. S., & McClements, D. J. (2016). Formation of Food-Grade Nanoemulsions Using Low-Energy Preparation Methods: A Review of Available Methods. *Comprehensive Reviews in Food Science and Food Safety*, 15(2), 331–352. <https://doi.org/10.1111/1541-4337.12189>

- Korpany, K. V., Majewski, D. D., Chiu, C. T., Cross, S. N., & Blum, A. S. (2017). Iron Oxide Surface Chemistry: Effect of Chemical Structure on Binding in Benzoic Acid and Catechol Derivatives. *Langmuir*, 33(12), 3000–3013. <https://doi.org/10.1021/acs.langmuir.6b03491>
- Kovacevic, A., Savic, S., Vuleta, G., Müller, R. H., & Keck, C. M. (2011). Polyhydroxy surfactants for the formulation of lipid nanoparticles (SLN and NLC): Effects on size, physical stability and particle matrix structure. *International Journal of Pharmaceutics*, 406(1–2), 163–172. <https://doi.org/10.1016/j.ijpharm.2010.12.036>
- Kucheryavy, P., He, J., John, V. T., Maharjan, P., Spinu, L., Goloverda, G. Z., & Kolesnichenko, V. L. (2013). Superparamagnetic Iron Oxide Nanoparticles with Variable Size and an Iron Oxidation State as Prospective Imaging Agents. *Langmuir*, 29(2), 710–716. <https://doi.org/10.1021/la3037007>
- Kumar, S., Umar, M., Saifi, A., Kumar, S., Augustine, S., Srivastava, S., & Malhotra, B. D. (2019). Electrochemical paper based cancer biosensor using iron oxide nanoparticles decorated PEDOT:PSS. *Analytica Chimica Acta*, 1056, 135–145. <https://doi.org/10.1016/j.aca.2018.12.053>
- Kupetz, E., & Bunjes, H. (2014). Lipid nanoparticles: Drug localization is substance-specific and achievable load depends on the size and physical state of the particles. *Journal of Controlled Release*, 189, 54–64. <https://doi.org/https://doi.org/10.1016/j.jconrel.2014.06.007>
- Kwon, S. G., & Hyeon, T. (2008). Colloidal Chemical Synthesis and Formation Kinetics of Uniformly Sized Nanocrystals of Metals, Oxides, and Chalcogenides. *Accounts of Chemical Research*, 41(12), 1696–1709. <https://doi.org/10.1021/ar8000537>
- LaConte, L. E. W., Nitin, N., Zurkiya, O., Caruntu, D., O'Connor, C. J., Hu, X., & Bao, G. (2007). Coating thickness of magnetic iron oxide nanoparticles affects R2 relaxivity. *Journal of Magnetic Resonance Imaging*, 26(6), 1634–1641. <https://doi.org/10.1002/jmri.21194>
- Lai, C. W., Low, F. W., Tai, M. F., & Abdul Hamid, S. B. (2018). Iron oxide nanoparticles decorated oleic acid for high colloidal stability. *Advances in Polymer Technology*, 37(6), 1712–1721. <https://doi.org/10.1002/adv.21829>
- Lassenberger, A., Grünewald, T. A., van Oostrum, P. D. J., Rennhofer, H., Amenitsch, H., Zirbs, R., Lichtenegger, H. C., & Reimhult, E. (2017). Monodisperse Iron Oxide Nanoparticles by Thermal Decomposition: Elucidating Particle Formation by Second-Resolved in Situ Small-Angle X-ray Scattering. *Chemistry of Materials*, 29(10), 4511–4522. <https://doi.org/10.1021/acs.chemmater.7b01207>
- Lassoued, A., Dkhil, B., Gadri, A., & Ammar, S. (2017). Control of the shape and size of iron oxide (α -Fe₂O₃) nanoparticles synthesized through the

chemical precipitation method. *Results in Physics*, 7, 3007–3015. <https://doi.org/https://doi.org/10.1016/j.rinp.2017.07.066>

- Lee, M. K., Kim, M. Y., Kim, S., & Lee, J. (2009). Cryoprotectants for freeze drying of drug nano-suspensions: Effect of freezing rate. *Journal of Pharmaceutical Sciences*, 98(12), 4808–4817. <https://doi.org/10.1002/jps.21786>
- Lee, M. S., Dees, E. C., & Wang, A. Z. (2017). Nanoparticle-Delivered Chemotherapy: Old Drugs in New Packages. *Oncology (Williston Park, N.Y.)*, 31(3), 198–208.
- Li, J., Wang, X., Zhang, T., Wang, C., Huang, Z., Luo, X., & Deng, Y. (2015). A review on phospholipids and their main applications in drug delivery systems. *Asian Journal of Pharmaceutical Sciences*, 10(2), 81–98. <https://doi.org/10.1016/j.ajps.2014.09.004>
- Lin, C., Wong, B. C. K., Chen, H., Bian, Z., Zhang, G., Zhang, X., Kashif Riaz, M., Tyagi, D., Lin, G., Zhang, Y., Wang, J., Lu, A., & Yang, Z. (2017). Pulmonary delivery of triptolide-loaded liposomes decorated with anti-carbonic anhydrase IX antibody for lung cancer therapy. *Scientific Reports*, 7(1), 1097. <https://doi.org/10.1038/s41598-017-00957-4>
- Lu, C., Ji, J., Zhu, X., Tang, P., Zhang, Q., Zhang, N., Wang, Z., Wang, X.-J., Chen, W., Hu, J., Du, Y.-Z., & Yu, R.-S. (2017). T2-Weighted Magnetic Resonance Imaging of Hepatic Tumor Guided by SPIO-Loaded Nanostructured Lipid Carriers and Ferritin Reporter Genes. *ACS Applied Materials & Interfaces*, 9(41), 35548–35561. <https://doi.org/10.1021/acsami.7b09879>
- Lu, Y., Yue, Z., Xie, J., Wang, W., Zhu, H., Zhang, E., & Cao, Z. (2018). Micelles with ultralow critical micelle concentration as carriers for drug delivery. *Nature Biomedical Engineering*, 2(5), 318–325. <https://doi.org/10.1038/s41551-018-0234-x>
- Lv, X., Zhang, L., Xing, F., & Lin, H. (2016). Controlled synthesis of monodispersed mesoporous silica nanoparticles: Particle size tuning and formation mechanism investigation. *Microporous and Mesoporous Materials*, 225, 238–244. <https://doi.org/10.1016/j.micromeso.2015.12.024>
- Ma, D., Shi, M., Li, X., Zhang, J., Fan, Y., Sun, K., Jiang, T., Peng, C., & Shi, X. (2020). Redox-Sensitive Clustered Ultrasmall Iron Oxide Nanoparticles for Switchable T₂/T₁-Weighted Magnetic Resonance Imaging Applications. *Bioconjugate Chemistry*, 31(2), 352–359. <https://doi.org/10.1021/acs.bioconjchem.9b00659>
- Mahmoudi, M., Laurent, S., Shokrgozar, M. A., & Hosseinkhani, M. (2011). Toxicity Evaluations of Superparamagnetic Iron Oxide Nanoparticles: Cell “Vision” versus Physicochemical Properties of Nanoparticles. *ACS Nano*, 5(9), 7263–7276. <https://doi.org/10.1021/nn2021088>

- Majumder, J., & Minko, T. (2021). Multifunctional and stimuli-responsive nanocarriers for targeted therapeutic delivery. *Expert Opinion on Drug Delivery*, 18(2), 205–227. <https://doi.org/10.1080/17425247.2021.1828339>
- Malvindi, M. A., De Matteis, V., Galeone, A., Brunetti, V., Anyfantis, G. C., Athanassiou, A., Cingolani, R., & Pompa, P. P. (2014). Toxicity Assessment of Silica Coated Iron Oxide Nanoparticles and Biocompatibility Improvement by Surface Engineering. *PLoS ONE*, 9(1), e85835. <https://doi.org/10.1371/journal.pone.0085835>
- Mangal, S., Gao, W., Li, T., & Zhou, Q. (Tony). (2017). Pulmonary delivery of nanoparticle chemotherapy for the treatment of lung cancers: challenges and opportunities. *Acta Pharmacologica Sinica*, 38(6), 782–797. <https://doi.org/10.1038/aps.2017.34>
- Manohar, S., & Leung, N. (2018). Cisplatin nephrotoxicity: a review of the literature. *Journal of Nephrology*, 31(1), 15–25. <https://doi.org/10.1007/s40620-017-0392-z>
- Marashdeh, M. W., Ababneh, B., Lemine, O. M., Alsadig, A., Omri, K., El Mir, L., Sulieman, A., & Mattar, E. (2019). The significant effect of size and concentrations of iron oxide nanoparticles on magnetic resonance imaging contrast enhancement. *Results in Physics*, 15, 102651. <https://doi.org/10.1016/j.rinp.2019.102651>
- Matarazzo, A. P., Elisei, L. M. S., Carvalho, F. C., Bonfílio, R., Ruela, A. L. M., Galdino, G., & Pereira, G. R. (2021). Mucoadhesive nanostructured lipid carriers as a cannabidiol nasal delivery system for the treatment of neuropathic pain. *European Journal of Pharmaceutical Sciences*, 159. <https://doi.org/10.1016/j.ejps.2020.105698>
- Matsumoto, Y., & Jasanoff, A. (2008). T2 relaxation induced by clusters of superparamagnetic nanoparticles: Monte Carlo simulations. *Magnetic Resonance Imaging*, 26(7), 994–998. <https://doi.org/10.1016/j.mri.2008.01.039>
- Mbeh, D. A., Mireles, L. K., Stanicki, D., Tabet, L., Maghni, K., Laurent, S., Sacher, E., & Yahia, L. (2015). Human Alveolar Epithelial Cell Responses to Core–Shell Superparamagnetic Iron Oxide Nanoparticles (SPIONs). *Langmuir*, 31(13), 3829–3839. <https://doi.org/10.1021/la5040646>
- Mehnert, W., & Mäder, K. (2012). Solid lipid nanoparticles. *Advanced Drug Delivery Reviews*, 64, 83–101. <https://doi.org/10.1016/j.addr.2012.09.021>
- Mi, P. (2020). Stimuli-responsive nanocarriers for drug delivery, tumor imaging, therapy and theranostics. *Theranostics*, 10(10), 4557–4588. <https://doi.org/10.7150/thno.38069>
- Mikuła-Pietrasik, J., Witucka, A., Pakuła, M., Uruski, P., Begier-Krasińska, B., Niklas, A., Tykarski, A., & Książek, K. (2019). Comprehensive review on

how platinum- and taxane-based chemotherapy of ovarian cancer affects biology of normal cells. *Cellular and Molecular Life Sciences*, 76(4), 681–697. <https://doi.org/10.1007/s00018-018-2954-1>

- Mohammadi, H., Nekobahr, E., Akhtari, J., Saeedi, M., Akbari, J., & Fathi, F. (2021). Synthesis and characterization of magnetite nanoparticles by coprecipitation method coated with biocompatible compounds and evaluation of in-vitro cytotoxicity. *Toxicology Reports*, 8, 331–336. <https://doi.org/10.1016/j.toxrep.2021.01.012>
- Montero, A., Fossella, F., Hortobagyi, G., & Valero, V. (2005). Docetaxel for treatment of solid tumours: a systematic review of clinical data. *The Lancet Oncology*, 6(4), 229–239. [https://doi.org/10.1016/S1470-2045\(05\)70094-2](https://doi.org/10.1016/S1470-2045(05)70094-2)
- Mosafer, J., Abnous, K., Tafaghodi, M., Jafarzadeh, H., & Ramezani, M. (2017). Preparation and characterization of uniform-sized PLGA nanospheres encapsulated with oleic acid-coated magnetic-Fe₃O₄ nanoparticles for simultaneous diagnostic and therapeutic applications. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 514, 146–154. <https://doi.org/10.1016/j.colsurfa.2016.11.056>
- Mrad, K., Schoenstein, F., Nong, H. T. T., Anagnostopoulou, E., Viola, A., Mouton, L., Mercone, S., Ricolleau, C., Jouini, N., Abderraba, M., Lacroix, L.-M., Viau, G., & Piquemal, J.-Y. (2017). Control of the crystal habit and magnetic properties of Co nanoparticles through the stirring rate. *CrystEngComm*, 19(25), 3476–3484. <https://doi.org/10.1039/C7CE00714K>
- Mulens-Arias, V., Rojas, J. M., Sanz-Ortega, L., Portilla, Y., Pérez-Yagüe, S., & Barber, D. F. (2019). Polyethylenimine-coated superparamagnetic iron oxide nanoparticles impair in vitro and in vivo angiogenesis. *Nanomedicine: Nanotechnology, Biology and Medicine*, 21, 102063. <https://doi.org/10.1016/j.nano.2019.102063>
- Müller, R. H., Radtke, M., & Wissing, S. A. (2002a). Nanostructured lipid matrices for improved microencapsulation of drugs. *International Journal of Pharmaceutics*, 242(1–2), 121–128. [https://doi.org/10.1016/S0378-5173\(02\)00180-1](https://doi.org/10.1016/S0378-5173(02)00180-1)
- Müller, R. H., Radtke, M., & Wissing, S. A. (2002b). Solid lipid nanoparticles (SLN) and nanostructured lipid carriers (NLC) in cosmetic and dermatological preparations. *Advanced Drug Delivery Reviews*, 54 Suppl 1, S131-55.
- Müllertz, A., Ogonna, A., Ren, S., & Rades, T. (2010). New perspectives on lipid and surfactant based drug delivery systems for oral delivery of poorly soluble drugs. *Journal of Pharmacy and Pharmacology*, 62(11), 1622–1636. <https://doi.org/10.1111/j.2042-7158.2010.01107.x>

- Musika, J., & Chudapongse, N. (2020). Development of Lipid-Based Nanocarriers for Increasing Gastrointestinal Absorption of Lupinifolin. *Planta Medica*, 86(5), 364–372. <https://doi.org/10.1055/a-1095-1129>
- Nafee, N., Makled, S., & Boraie, N. (2018). Nanostructured lipid carriers versus solid lipid nanoparticles for the potential treatment of pulmonary hypertension via nebulization. *European Journal of Pharmaceutical Sciences*, 125, 151–162. <https://doi.org/https://doi.org/10.1016/j.ejps.2018.10.003>
- Naguib, Y. W., Rodriguez, B. L., Li, X., Hursting, S. D., Williams, R. O., & Cui, Z. (2014). Solid lipid nanoparticle formulations of docetaxel prepared with high melting point triglycerides: In vitro and in vivo evaluation. *Molecular Pharmaceutics*, 11(4), 1239–1249. <https://doi.org/10.1021/mp4006968>
- Nakamoto, K. (2008a). *Infrared and Raman Spectra of Inorganic and Coordination Compounds: Part A: Theory and Applications in Inorganic Chemistry* (Sixth). John Wiley & Sons, Inc. <https://doi.org/10.1002/9780470405840>
- Nakamoto, K. (2008b). *Infrared and Raman Spectra of Inorganic and Coordination Compounds: Part B: Applications in Coordination, Organometallic, and Bioinorganic Chemistry* (Sixth). John Wiley & Sons, Inc. <https://doi.org/10.1002/9780470405888>
- Naqvi, S., Samim, M., Abdin, M., Ahmad, F. J., Maitra, A., Dinda, K., & Prashant, C. (2010). Concentration-dependent toxicity of iron oxide nanoparticles mediated by increased oxidative stress. *International Journal of Nanomedicine*, 5, 983–989. <https://doi.org/10.2147/IJN.S13244>
- Negi, L. M., Jaggi, M., & Talegaonkar, S. (2014). Development of protocol for screening the formulation components and the assessment of common quality problems of nano-structured lipid carriers. *International Journal of Pharmaceutics*, 461(1–2), 403–410. <https://doi.org/10.1016/j.ijpharm.2013.12.006>
- Neophytou, C. M., & Constantinou, A. I. (2015). Drug Delivery Innovations for Enhancing the Anticancer Potential of Vitamin E Isoforms and Their Derivatives. *BioMed Research International*, 2015(APRIL), 1–16. <https://doi.org/10.1155/2015/584862>
- Nguyen, T.-D. (2013). From formation mechanisms to synthetic methods toward shape-controlled oxide nanoparticles. *Nanoscale*, 5(20), 9455. <https://doi.org/10.1039/c3nr01810e>
- Niepel, M., Hafner, M., Mills, C. E., Subramanian, K., Williams, E. H., Chung, M., Gaudio, B., Barrette, A. M., Stern, A. D., Hu, B., Korkola, J. E., Gray, J. W., Birtwistle, M. R., Heiser, L. M., Sorger, P. K., Shamu, C. E., Jayaraman, G., Azeloglu, E. U., Iyengar, R., ... Devlin, K. (2019). A Multi-center Study

on the Reproducibility of Drug-Response Assays in Mammalian Cell Lines. *Cell Systems*, 9(1), 35-48.e5. <https://doi.org/10.1016/j.cels.2019.06.005>

- Obeidat, W. M., Schwabe, K., Müller, R. H., & Keck, C. M. (2010). Preservation of nanostructured lipid carriers (NLC). *European Journal of Pharmaceutics and Biopharmaceutics*, 76(1), 56–67. <https://doi.org/https://doi.org/10.1016/j.ejpb.2010.05.001>
- Odularu, A. T. (2018). Metal Nanoparticles: Thermal Decomposition, Biomedical Applications to Cancer Treatment, and Future Perspectives. *Bioinorganic Chemistry and Applications*, 2018, 1–6. <https://doi.org/10.1155/2018/9354708>
- Oehlert, G. W. (2010). *A First Course in Design and Analysis of Experiments*. Retrieved from the University of Minnesota Digital Conservancy.
- Olerile, L. D., Liu, Y., Zhang, B., Wang, T., Mu, S., Zhang, J., Selotlegeng, L., & Zhang, N. (2017). Near-infrared mediated quantum dots and paclitaxel co-loaded nanostructured lipid carriers for cancer theragnostic. *Colloids and Surfaces B: Biointerfaces*, 150, 121–130. <https://doi.org/https://doi.org/10.1016/j.colsurfb.2016.11.032>
- Oles, V. (1992). Shear-induced aggregation and breakup of polystyrene latex particles. *Journal of Colloid And Interface Science*, 154(2), 351–358. [https://doi.org/10.1016/0021-9797\(92\)90149-G](https://doi.org/10.1016/0021-9797(92)90149-G)
- Oliveira, R. R., Carrião, M. S., Pacheco, M. T., Branquinho, L. C., de Souza, A. L. R., Bakuzis, A. F., & Lima, E. M. (2018). Triggered release of paclitaxel from magnetic solid lipid nanoparticles by magnetic hyperthermia. *Materials Science and Engineering C*, 92, 547–553. <https://doi.org/10.1016/j.msec.2018.07.011>
- Ong, Y. S., Bañobre-López, M., Costa Lima, S. A., & Reis, S. (2020). A multifunctional nanomedicine platform for co-delivery of methotrexate and mild hyperthermia towards breast cancer therapy. *Materials Science and Engineering: C*, 116, 111255. <https://doi.org/https://doi.org/10.1016/j.msec.2020.111255>
- Orsavova, J., Misurcova, L., Ambrozova, J. V., Vicha, R., & Mlcek, J. (2015). Fatty Acids Composition of Vegetable Oils and Its Contribution to Dietary Energy Intake and Dependence of Cardiovascular Mortality on Dietary Intake of Fatty Acids. *International Journal of Molecular Sciences*, 16(6), 12871–12890. <https://doi.org/10.3390/ijms160612871>
- Oumzil, K., Ramin, M. A., Lorenzato, C., Hémadou, A., Laroche, J., Jacobin-Valat, M. J., Mornet, S., Roy, C.-E., Kauss, T., Gaudin, K., Clofent-Sanchez, G., & Barthélémy, P. (2016). Solid Lipid Nanoparticles for Image-Guided Therapy of Atherosclerosis. *Bioconjugate Chemistry*, 27(3), 569–575. <https://doi.org/10.1021/acs.bioconjchem.5b00590>

- Ozawa, Y., Koda, K., Akahori, D., Matsui, T., Hasegawa, H., Kakutani, T., Amano, T., Tanahashi, M., Niwa, H., Kunimoto, Y., Yamada, K., Yokomura, K., & Suda, T. (2018). Preexisting Interstitial Lung Disease and Lung Injury Associated with Irinotecan in Patients with Neoplasms. *Anticancer Research*, 38(10), 5937–5941. <https://doi.org/10.21873/anticancerres.12939>
- Pardeike, J., Weber, S., Haber, T., Wagner, J., Zarfl, H. P., Plank, H., & Zimmer, A. (2011). Development of an Itraconazole-loaded nanostructured lipid carrier (NLC) formulation for pulmonary application. *International Journal of Pharmaceutics*, 419(1–2), 329–338. <https://doi.org/10.1016/j.ijpharm.2011.07.040>
- Paunovic, J., Vučević, D., Radosavljević, T., Mandić-Rajčević, S., & Pantić, I. (2020). Iron-based nanoparticles and their potential toxicity: Focus on oxidative stress and apoptosis. *Chemico-Biological Interactions*, 316, 108935. <https://doi.org/https://doi.org/10.1016/j.cbi.2019.108935>
- Petrov, P. A., Ali, A., & Potter, D. K. (2017). Diamagnetic Behavior in Nanoparticle Hematite? *Journal of Modern Physics*, 08(07), 1013–1019. <https://doi.org/10.4236/jmp.2017.87063>
- Pinheiro, M., Ribeiro, R., Vieira, A., Andrade, F., & Reis, S. (2016). Design of a nanostructured lipid carrier intended to improve the treatment of tuberculosis. *Drug Design, Development and Therapy*, 10, 2467–2475. <https://doi.org/10.2147/DDDT.S104395>
- Pinto, F., de Barros, D. P. C., Reis, C., & Fonseca, L. P. (2019). Optimization of nanostructured lipid carriers loaded with retinoids by central composite design. *Journal of Molecular Liquids*, 293, 111468. <https://doi.org/https://doi.org/10.1016/j.molliq.2019.111468>
- Pooley, R. A. (2005). Fundamental Physics of MR Imaging. *RadioGraphics*, 25(4), 1087–1099. <https://doi.org/10.1148/rg.254055027>
- Prat, D., Hayler, J., & Wells, A. (2014). A survey of solvent selection guides. *Green Chem.*, 16(10), 4546–4551. <https://doi.org/10.1039/C4GC01149J>
- Qiao, H., Chen, L., Rui, T., Wang, J., Chen, T., Fu, T., Li, J., & Di, L. (2017). Fabrication and in vitro/in vivo evaluation of amorphous andrographolide nanosuspensions stabilized by D- α -tocopheryl polyethylene glycol 1000 succinate/sodium lauryl sulfate. *International Journal of Nanomedicine*, Volume 12, 1033–1046. <https://doi.org/10.2147/IJN.S120887>
- Rahmawati, R., Permana, M. G., Harison, B., Nugraha, Yulianto, B., Suyatman, & Kurniadi, D. (2017). Optimization of Frequency and Stirring Rate for Synthesis of Magnetite (Fe₃O₄) Nanoparticles by Using Coprecipitation-Ultrasonic Irradiation Methods. *Procedia Engineering*, 170, 55–59. <https://doi.org/10.1016/j.proeng.2017.03.010>

- Rajadurai, P., How, S. H., Liam, C. K., Sachithanandan, A., Soon, S. Y., & Tho, L. M. (2020). Lung Cancer in Malaysia. *Journal of Thoracic Oncology*, 15(3), 317–323. <https://doi.org/10.1016/j.jtho.2019.10.021>
- Rampino, A., Borgogna, M., Blasi, P., Bellich, B., & Cesàro, A. (2013). Chitosan nanoparticles: Preparation, size evolution and stability. *International Journal of Pharmaceutics*, 455(1–2), 219–228. <https://doi.org/10.1016/j.ijpharm.2013.07.034>
- Rathod, S., Bahadur, P., & Tiwari, S. (2021). Nanocarriers based on vitamin E-TPGS: Design principle and molecular insights into improving the efficacy of anticancer drugs. *International Journal of Pharmaceutics*, 592, 120045. <https://doi.org/https://doi.org/10.1016/j.ijpharm.2020.120045>
- Ren, X., Chen, H., Yang, V., & Sun, D. (2014). Iron oxide nanoparticle-based theranostics for cancer imaging and therapy. *Frontiers of Chemical Science and Engineering*, 8(3), 253–264. <https://doi.org/10.1007/s11705-014-1425-y>
- Rodenak-Kladniew, B., Islan, G. A., de Bravo, M. G., Durán, N., & Castro, G. R. (2017). Design, characterization and in vitro evaluation of linalool-loaded solid lipid nanoparticles as potent tool in cancer therapy. *Colloids and Surfaces B: Biointerfaces*, 154, 123–132. <https://doi.org/https://doi.org/10.1016/j.colsurfb.2017.03.021>
- Rodrigues da Silva, G. H., Geronimo, G., García-López, J. P., Ribeiro, L. N. M., de Moura, L. D., Breikreitz, M. C., Feijóo, C. G., & de Paula, E. (2020). Articaine in functional NLC show improved anesthesia and anti-inflammatory activity in zebrafish. *Scientific Reports*, 10(1), 19733. <https://doi.org/10.1038/s41598-020-76751-6>
- Roonasi, P., & Holmgren, A. (2009). A Fourier transform infrared (FTIR) and thermogravimetric analysis (TGA) study of oleate adsorbed on magnetite nano-particle surface. *Applied Surface Science*, 255(11), 5891–5895. <https://doi.org/10.1016/j.apsusc.2009.01.031>
- Rouco, H., Diaz-Rodriguez, P., Gaspar, D. P., Gonçalves, L. M. D., Cuerva, M., Remuñán-López, C., Almeida, A. J., & Landin, M. (2020). Rifabutin-Loaded Nanostructured Lipid Carriers as a Tool in Oral Anti-Mycobacterial Treatment of Crohn's Disease. *Nanomaterials*, 10(11), 2138. <https://doi.org/10.3390/nano10112138>
- Sah, S. K., Karn, A., Shah, A., Paudel, B. D., Adhikari, K., Acharya, B., & Chapagain, S. (2019). Incidence and attributes of chemotherapy induced myelotoxicity, anemia and neutropenia in adults with cancer in Nepal: A cross-sectional observational study. *Journal of Oncology Pharmacy Practice*, 25(8), 1823–1830. <https://doi.org/10.1177/1078155218817815>
- Salminen, H., Helgason, T., Aulbach, S., Kristinsson, B., Kristbergsson, K., & Weiss, J. (2014). Influence of co-surfactants on crystallization and stability

of solid lipid nanoparticles. *Journal of Colloid and Interface Science*, 426, 256–263. <https://doi.org/https://doi.org/10.1016/j.jcis.2014.04.009>

Salunkhe, A., Khot, V., Patil, S. I., Tofail, S. A. M., Bauer, J., & Thorat, N. D. (2020). MRI Guided Magneto-chemotherapy with High-Magnetic-Moment Iron Oxide Nanoparticles for Cancer Theranostics. *ACS Applied Bio Materials*, 3(4), 2305–2313. <https://doi.org/10.1021/acsabm.0c00077>

Sánchez-Cabezas, S., Montes-Robles, R., Gallo, J., Sancenón, F., & Martínez-Máñez, R. (2019). Combining magnetic hyperthermia and dual T₁/T₂ MR imaging using highly versatile iron oxide nanoparticles. *Dalton Transactions*, 48(12), 3883–3892. <https://doi.org/10.1039/C8DT04685A>

Schneider, C. A.; Rasband, W. S. & Eliceiri, K. W. (2012). NIH Image to ImageJ: 25 years of image analysis. *Nature Methods*, 9(7), 671–675.

Schubert, M. A., & Müller-Goymann, C. C. (2003). Solvent injection as a new approach for manufacturing lipid nanoparticles – evaluation of the method and process parameters. *European Journal of Pharmaceutics and Biopharmaceutics*, 55(1), 125–131. [https://doi.org/10.1016/S0939-6411\(02\)00130-3](https://doi.org/10.1016/S0939-6411(02)00130-3)

Schwartzberg, L. S., & Navari, R. M. (2018). Safety of Polysorbate 80 in the Oncology Setting. *Advances in Therapy*, 35(6), 754–767. <https://doi.org/10.1007/s12325-018-0707-z>

Schwertmann, U., & Cornell, R. M. (2000). Iron Oxides in the Laboratory. In U. Schwertmann & R. M. Cornell (Eds.), *Wiley-VCH Verlag GmbH* (2nd ed.). Wiley-VCH Verlag GmbH. <https://doi.org/10.1002/9783527613229>

Severino, P., Pinho, S. C., Souto, E. B., & Santana, M. H. A. (2011). Polymorphism, crystallinity and hydrophilic–lipophilic balance of stearic acid and stearic acid–capric/caprylic triglyceride matrices for production of stable nanoparticles. *Colloids and Surfaces B: Biointerfaces*, 86(1), 125–130. <https://doi.org/10.1016/j.colsurfb.2011.03.029>

Sezer, N., Ari, İ., Biçer, Y., & Koç, M. (2021). Superparamagnetic nanoarchitectures: Multimodal functionalities and applications. *Journal of Magnetism and Magnetic Materials*, 538, 168300. <https://doi.org/10.1016/j.jmmm.2021.168300>

Sigma-Aldrich. (2020). *Product specification: Fish oil from Menhaden*. Product Specification. https://api.sigmaaldrich.com/deepweb/assets/sigmaaldrich/quality/spec/204/656/F8020-BULK____SIGMA____.pdf

Smolensky, E. D., Park, H.-Y. E., Zhou, Y., Rolla, G. A., Marjańska, M., Botta, M., & Pierre, V. C. (2013). Scaling laws at the nanosize: the effect of particle size and shape on the magnetism and relaxivity of iron oxide nanoparticle

- contrast agents. *Journal of Materials Chemistry B*, 1(22), 2818. <https://doi.org/10.1039/c3tb00369h>
- Soares, P. I. P., Laia, C. A. T., Carvalho, A., Pereira, L. C. J., Coutinho, J. T., Ferreira, I. M. M., Novo, C. M. M., & Borges, J. P. (2016). Iron oxide nanoparticles stabilized with a bilayer of oleic acid for magnetic hyperthermia and MRI applications. *Applied Surface Science*, 383, 240–247. <https://doi.org/10.1016/j.apsusc.2016.04.181>
- Sosa Iglesias, V., Giuranno, L., Dubois, L. J., Theys, J., & Vooijs, M. (2018). Drug Resistance in Non-Small Cell Lung Cancer: A Potential for NOTCH Targeting? *Frontiers in Oncology*, 8. <https://doi.org/10.3389/fonc.2018.00267>
- Sou, T., & Bergström, C. A. S. (2021). Contemporary Formulation Development for Inhaled Pharmaceuticals. *Journal of Pharmaceutical Sciences*, 110(1), 66–86. <https://doi.org/10.1016/j.xphs.2020.09.006>
- Suciati, T., Rachmawati, P., Soraya, E., Mahardhika, A. B., Satrialdi, Hartarti, R., & Anggadiredja, K. (2018). A novel acemannan-chitosan modified lipid nanoparticles as intracellular delivery vehicles of antibiotic. *Journal of Applied Pharmaceutical Science*, 8(12), 1–11. <https://doi.org/10.7324/JAPS.2018.81201>
- Sun, M., Nie, S., Pan, X., Zhang, R., Fan, Z., & Wang, S. (2014). Quercetin-nanostructured lipid carriers: Characteristics and anti-breast cancer activities in vitro. *Colloids and Surfaces B: Biointerfaces*, 113, 15–24. <https://doi.org/10.1016/j.colsurfb.2013.08.032>
- Sun, S., & Zeng, H. (2002). Size-Controlled Synthesis of Magnetite Nanoparticles. *Journal of the American Chemical Society*, 124(28), 8204–8205. <https://doi.org/10.1021/ja026501x>
- Sung, H., Ferlay, J., Siegel, R. L., Laversanne, M., Soerjomataram, I., Jemal, A., & Bray, F. (2021). Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: A Cancer Journal for Clinicians*. <https://doi.org/10.3322/caac.21660>
- Svedberg, A., Björn, N., Sigurgeirsson, B., Pradhananga, S., Brandén, E., Koyi, H., Lewensohn, R., De Petris, L., Apellániz-Ruiz, M., Rodríguez-Antona, C., Lundeberg, J., & Gréen, H. (2020). Genetic association of gemcitabine/carboplatin-induced leukopenia and neutropenia in non-small cell lung cancer patients using whole-exome sequencing. *Lung Cancer*, 147, 106–114. <https://doi.org/10.1016/j.lungcan.2020.07.005>
- Ta, H. T., Li, Z., Wu, Y., Cowin, G., Zhang, S., Yago, A., Whittaker, A. K., & Xu, Z. P. (2017). Effects of magnetic field strength and particle aggregation on relaxivity of ultra-small dual contrast iron oxide nanoparticles. *Materials Research Express*, 4(11), 116105. <https://doi.org/10.1088/2053-1591/aa96e3>

- Tapeinos, C., Marino, A., Battaglini, M., Migliorin, S., Brescia, R., Scarpellini, A., de Julián Fernández, C., Prato, M., Drago, F., & Ciofani, G. (2019). Stimuli-responsive lipid-based magnetic nanovectors increase apoptosis in glioblastoma cells through synergic intracellular hyperthermia and chemotherapy. *Nanoscale*, *11*(1), 72–88. <https://doi.org/10.1039/c8nr05520c>
- Teeranachaideekul, V., Boonme, P., Souto, E. B., Müller, R. H., & Junyaprasert, V. B. (2008). Influence of oil content on physicochemical properties and skin distribution of Nile red-loaded NLC. *Journal of Controlled Release*, *128*(2), 134–141. <https://doi.org/10.1016/j.jconrel.2008.02.011>
- Tetyczka, C., Hodzic, A., Kriechbaum, M., Juraić, K., Spirk, C., Hartl, S., Pritz, E., Leitinger, G., & Roblegg, E. (2019). Comprehensive characterization of nanostructured lipid carriers using laboratory and synchrotron X-ray scattering and diffraction. *European Journal of Pharmaceutics and Biopharmaceutics*, *139*, 153–160. <https://doi.org/10.1016/j.ejpb.2019.03.017>
- Thermo Fisher Scientific. (2018). *Docetaxel, 98%, ACROS Organics™*. Safety Data Sheet. <https://www.fishersci.com/shop/products/docetaxel-98-acros-organics-2/AC456262500>
- Thermo Fisher Scientific. (2020). *Counting Cells in a Hemocytometer*. Cell Culture Protocols. <https://www.thermofisher.com/my/en/home/references/gibco-cell-culture-basics/cell-culture-protocols/counting-cells-in-a-hemocytometer.html>
- Thorat, N. D., Lemine, O. M., Bohara, R. A., Omri, K., El Mir, L., & Tofail, S. A. M. (2016). Superparamagnetic iron oxide nanocargoes for combined cancer thermotherapy and MRI applications. *Physical Chemistry Chemical Physics*, *18*(31), 21331–21339. <https://doi.org/10.1039/C6CP03430F>
- Tyn, M. T., & Calus, W. F. (1975). Temperature and concentration dependence of mutual diffusion coefficients of some binary liquid systems. *Journal of Chemical & Engineering Data*, *20*(3), 310–316. <https://doi.org/10.1021/je60066a009>
- Umeyor, C. E., Okoye, I., Uronnachi, E., Okeke, T., Kenechukwu, F., & Attama, A. (2020). Repositioning miconazole nitrate for malaria: Formulation of sustained release nanostructured lipid carriers, structure characterization and in vivo antimalarial evaluation. *Journal of Drug Delivery Science and Technology*, 102125. <https://doi.org/10.1016/j.jddst.2020.102125>
- Unni, M., Uhl, A. M., Savliwala, S., Savitzky, B. H., Dhavalikar, R., Garraud, N., Arnold, D. P., Kourkoutis, L. F., Andrew, J. S., & Rinaldi, C. (2017). Thermal Decomposition Synthesis of Iron Oxide Nanoparticles with Diminished Magnetic Dead Layer by Controlled Addition of Oxygen. *ACS Nano*, *11*(2), 2284–2303. <https://doi.org/10.1021/acsnano.7b00609>

- van der Zanden, S. Y., Qiao, X., & Neefjes, J. (2020). New insights into the activities and toxicities of the old anticancer drug doxorubicin. *The FEBS Journal*, febs.15583. <https://doi.org/10.1111/febs.15583>
- Varshosaz, J., Eskandari, S., & Tabbakhian, M. (2012). Freeze-drying of nanostructure lipid carriers by different carbohydrate polymers used as cryoprotectants. *Carbohydrate Polymers*, 88(4), 1157–1163. <https://doi.org/10.1016/j.carbpol.2012.01.051>
- Velmurugan, R., & Selvamuthukumar, S. (2016). Development and optimization of ifosfamide nanostructured lipid carriers for oral delivery using response surface methodology. *Applied Nanoscience*, 6(2), 159–173. <https://doi.org/10.1007/s13204-015-0434-6>
- Vuong, Q. L., Berret, J.-F., Fresnais, J., Gossuin, Y., & Sandre, O. (2012). A Universal Scaling Law to Predict the Efficiency of Magnetic Nanoparticles as MRI T2-Contrast Agents. *Advanced Healthcare Materials*, 1(4), 502–512. <https://doi.org/10.1002/adhm.201200078>
- Wang, H., Jordan, V. C., Ramsay, I. A., Sojoodi, M., Fuchs, B. C., Tanabe, K. K., Caravan, P., & Gale, E. M. (2019). Molecular Magnetic Resonance Imaging Using a Redox-Active Iron Complex. *Journal of the American Chemical Society*, 141(14), 5916–5925. <https://doi.org/10.1021/jacs.9b00603>
- Wang, T., Liu, K., Shi, X., Ye, L., Gu, W., & Yan, C. (2017). Tuning of synthesis conditions by thermal decomposition towards gadolinium-doped manganese carbonate nanoparticles with uniform size and high relaxivity. *New Journal of Chemistry*, 41(1), 225–230. <https://doi.org/10.1039/C6NJ02739C>
- Warashina, S., Nakamura, T., Sato, Y., Fujiwara, Y., Hyodo, M., Hatakeyama, H., & Harashima, H. (2016). A lipid nanoparticle for the efficient delivery of siRNA to dendritic cells. *Journal of Controlled Release*, 225, 183–191. <https://doi.org/10.1016/j.jconrel.2016.01.042>
- Wauthoz, N., Rosière, R., & Amighi, K. (2020). Inhaled cytotoxic chemotherapy: clinical challenges, recent developments, and future prospects. *Expert Opinion on Drug Delivery*, 1–22. <https://doi.org/10.1080/17425247.2021.1829590>
- Weber, S., Zimmer, A., & Pardeike, J. (2014). Solid Lipid Nanoparticles (SLN) and Nanostructured Lipid Carriers (NLC) for pulmonary application: A review of the state of the art. *European Journal of Pharmaceutics and Biopharmaceutics*, 86(1), 7–22. <https://doi.org/10.1016/j.ejpb.2013.08.013>
- Wei, H., Bruns, O. T., Kaul, M. G., Hansen, E. C., Barch, M., Wiśniowska, A., Chen, O., Chen, Y., Li, N., Okada, S., Cordero, J. M., Heine, M., Farrar, C. T., Montana, D. M., Adam, G., Ittrich, H., Jasanoff, A., Nielsen, P., & Bawendi, M. G. (2017). Exceedingly small iron oxide nanoparticles as

- positive MRI contrast agents. *Proceedings of the National Academy of Sciences*, 114(9), 2325–2330. <https://doi.org/10.1073/pnas.1620145114>
- Wei, R., Zhou, T., Sun, C., Lin, H., Yang, L., Ren, B. W., Chen, Z., & Gao, J. (2018). Iron-oxide-based twin nanoplates with strong T₂ relaxation shortening for contrast-enhanced magnetic resonance imaging. *Nanoscale*, 10(38), 18398–18406. <https://doi.org/10.1039/C8NR04995E>
- Wei, W., Zhaohui, W., Taekyung, Y., Changzhong, J., & Woo-Sik, K. (2015). Recent progress on magnetic iron oxide nanoparticles: synthesis, surface functional strategies and biomedical applications. *Science and Technology of Advanced Materials*, 16(2), 23501. <https://doi.org/10.1088/1468-6996/16/2/023501>
- Wetterskog, E., Tai, C.-W., Grins, J., Bergström, L., & Salazar-Alvarez, G. (2013). Anomalous Magnetic Properties of Nanoparticles Arising from Defect Structures: Topotaxial Oxidation of Fe_{1-x}O/Fe_{3-δ}O₄ Core/Shell Nanocubes to Single-Phase Particles. *ACS Nano*, 7(8), 7132–7144. <https://doi.org/10.1021/nn402487q>
- Wood, M. H., Casford, M. T., Steitz, R., Zorbakhsh, A., Welbourn, R. J. L., & Clarke, S. M. (2016). Comparative Adsorption of Saturated and Unsaturated Fatty Acids at the Iron Oxide/Oil Interface. *Langmuir*, 32(2), 534–540. <https://doi.org/10.1021/acs.langmuir.5b04435>
- Xia, Q., Hao, X., Lu, Y., Xu, W., Wei, H., Ma, Q., & Gu, N. (2008). Production of drug-loaded lipid nanoparticles based on phase behaviors of special hot microemulsions. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 313–314, 27–30. <https://doi.org/10.1016/j.colsurfa.2007.04.067>
- Xie, W., Guo, Z., Gao, F., Gao, Q., Wang, D., Liaw, B., Cai, Q., Sun, X., Wang, X., & Zhao, L. (2018). Shape-, size- and structure-controlled synthesis and biocompatibility of iron oxide nanoparticles for magnetic theranostics. *Theranostics*, 8(12), 3284–3307. <https://doi.org/10.7150/thno.25220>
- Yang, C., Wu, T., Qi, Y., & Zhang, Z. (2018). Recent Advances in the Application of Vitamin E TPGS for Drug Delivery. *Theranostics*, 8(2), 464–485. <https://doi.org/10.7150/thno.22711>
- Yang, K., Peng, H., Wen, Y., & Li, N. (2010). Re-examination of characteristic FTIR spectrum of secondary layer in bilayer oleic acid-coated Fe₃O₄ nanoparticles. *Applied Surface Science*, 256(10), 3093–3097. <https://doi.org/10.1016/j.apsusc.2009.11.079>
- Yu, M., Huang, S., Yu, K. J., & Clyne, A. M. (2012). Dextran and Polymer Polyethylene Glycol (PEG) Coating Reduce Both 5 and 30 nm Iron Oxide Nanoparticle Cytotoxicity in 2D and 3D Cell Culture. In *International Journal of Molecular Sciences* (Vol. 13, Issue 5). <https://doi.org/10.3390/ijms13055554>

- Yu, W. W., Falkner, J. C., Yavuz, C. T., & Colvin, V. L. (2004). Synthesis of monodisperse iron oxide nanocrystals by thermal decomposition of iron carboxylate salts. *Chemical Communications*, 20, 2306. <https://doi.org/10.1039/b409601k>
- Zhang, L., He, R., & Gu, H. C. (2006). Oleic acid coating on the monodisperse magnetite nanoparticles. *Applied Surface Science*, 253(5), 2611–2617. <https://doi.org/10.1016/j.apsusc.2006.05.023>
- Zhang, P., Zhang, Y., Ding, X., Shen, W., Li, M., Wagner, E., Xiao, C., & Chen, X. (2020). A Multistage Cooperative Nanoplatform Enables Intracellular Co-Delivery of Proteins and Chemotherapeutics for Cancer Therapy. *Advanced Materials*, 32(46), 2000013. <https://doi.org/10.1002/adma.202000013>
- Zhao, Z., Zhou, Z., Bao, J., Wang, Z., Hu, J., Chi, X., Ni, K., Wang, R., Chen, X., Chen, Z., & Gao, J. (2013). Octapod iron oxide nanoparticles as high-performance T2 contrast agents for magnetic resonance imaging. *Nature Communications*, 4(1), 2266. <https://doi.org/10.1038/ncomms3266>
- Zhou, Z., Ding, Y., Zu, X., & Deng, Y. (2011). ZnO spheres and nanorods formation: their dependence on agitation in solution synthesis. *Journal of Nanoparticle Research*, 13(4), 1689–1696. <https://doi.org/10.1007/s11051-010-9922-5>
- Zhou, Z., Zhu, X., Wu, D., Chen, Q., Huang, D., Sun, C., Xin, J., Ni, K., & Gao, J. (2015). Anisotropic Shaped Iron Oxide Nanostructures: Controlled Synthesis and Proton Relaxation Shortening Effects. *Chemistry of Materials*, 27(9), 3505–3515. <https://doi.org/10.1021/acs.chemmater.5b00944>
- Zhu, X., Deng, X., Lu, C., Chen, Y., Jie, L., Zhang, Q., Li, W., Wang, Z., Du, Y., & Yu, R. (2018). SPIO-loaded nanostructured lipid carriers as liver-targeted molecular T2-weighted MRI contrast agent. *Quantitative Imaging in Medicine and Surgery*, 8(8), 770–780. <https://doi.org/10.21037/qims.2018.09.03>