



UNIVERSITI PUTRA MALAYSIA

***CELLULAR AND MOLECULAR CHARACTERIZATION OF
ATHEROTHROMBOSIS-RELATED MARKERS IN SPRAGUE DAWLEY
RATS FED WITH HIGH CHOLESTEROL DIET SUPPLEMENTED WITH
HERBAL EXTRACTS***

IBRAHIM KALLE KWAIFA

FPSK(p) 2022 30



**CELLULAR AND MOLECULAR CHARACTERIZATION OF
ATHEROTHROMBOSIS-RELATED MARKERS IN SPRAGUE DAWLEY
RATS FED WITH HIGH CHOLESTEROL DIET SUPPLEMENTED WITH
HERBAL EXTRACTS**

By

IBRAHIM KALLE KWAIFA

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

September 2021

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

This thesis is dedicated to Almighty Allah (SWT), who saved me, gave me health, strength and wisdom throughout the study period



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

**CELLULAR AND MOLECULAR CHARACTERIZATION OF
ATHEROTHROMBOSIS-RELATED MARKERS IN SPRAGUE DAWLEY
RATS FED WITH HIGH CHOLESTEROL DIET SUPPLEMENTED WITH
HERBAL EXTRACTS**

By

IBRAHIM KALLE KWAIFA

September 2021

Chairman : Associate Professor Sabariah Md Noor, MD, MPath, AM
Faculty : Medicine and Health Sciences

Atherothrombotic cardiovascular diseases are the most common cause of death worldwide. Although antiplatelet, anticoagulant, simvastatin and other related drugs globally prescribed for these disorders have been available, some are commonly associated with several adverse effects. Intervention with natural-based preparation has been recognized as an alternative. This study utilised a combination of herbal extracts for the treatment of atherothrombosis in Sprague Dawley (SD) rats fed with a high cholesterol diet (HCD). The study investigated the therapeutic potentials of herbal extracts; a combination of *Zingiber officinale* (ginger), *Allium sativum* (garlic), *Citrus Limon* (lemon), *Malus Domestica* (Apple)/ apple cider vinegar and Honey (ZACAH) on cellular and molecular atherothrombosis-related markers in SD rats fed with HCD. Identification of the related phytochemicals present in ZACAH extracts was performed by ultra-high performance liquid chromatography-mass spectrophotometry (UHPLC-MS) profiling at both negative and positive models. Thirty-six male SD rats were randomly divided into six groups. The normal diet (ND), HCD, treatment with simvastatin (TRTSM) at 10mg/kg of body weight (BW), treatment with ZACAH extracts (TRT1) at 1mg, 3mg (TRT2), and 5mg (TRT3) per kilogram of BW. At the end of the 12 weeks of experiments, the rats were sacrificed, and the collected blood was subjected to serum lipid profile, protein expressions, and platelet ultrastructural analysis by scanning electron microscope (SEM). The atherogenic index (AI) and % protection were calculated based on the results obtained from the lipid profile. SEM was also used to examine the aorta endothelial ultrastructural changes. The aorta and liver tissues were utilised for histopathological examinations by haematoxylin & eosin (H & E) and Masson's trichrome (MT) stains. The tissues were also processed for the mRNA expression profiling of lipid metabolism atherothrombosis-related genes, endothelial prothrombotic and coagulation related genes, and the genes of the fibrinolytic system. Ten related phytochemical compounds present in ZACAH extracts were identified, including gingerol (retention time (RT); 18.62, compounds %; 5.6), hesperidin (11.19,

12.1), naringin (10.07, 13.1), sulindac (13.96, 9.7), scoparone (15.86, 8.0), hesperetin (11.18, 12.1), rutin (9.90, 13.2), limonin (17.87, 6.2), carboxylic acid (3.30, 18.9) and citric acid (2.18, 20). Increased BW and food intake were recorded in all the experimental groups, while a significant difference was observed in the HCD group compared to ND, TRTSM and ZACAH extracts treatment groups ($p<0.05$). The total cholesterol (TC), triglycerides (TG) and low-density lipoprotein (LDL) levels were significantly increased, and a decrease in high-density lipoprotein (HDL) was observed in the HCD group compared to ND, TRTSM and ZACAH extracts treatment groups ($p<0.05$). Significant differences were observed in the AI of HCD compared to ND and ZACAH extracts treatment groups ($p<0.05$). The % protection was improved at TRTSM (18%), TRT1 (55.6%), TRT2 (59.7%), and TRT3 (72.8%) in dose-dependent manner. The SEM analysis revealed organized atherothrombosis, smooth muscle cells proliferation in the intima, and significantly activated platelets in the HCD group. Histopathological examinations revealed a significant increase in the intima-media ratio, marked increase in collagen fibres and intense hepatic steatosis in the HCD group compared to ND, TRTSM and ZACAH extracts treatment groups ($p<0.05$). The mRNA expression of lipid metabolism genes, including 3-hydroxy-3-methyl-glutaryl-coenzyme A reductase (HMG-CoA-R), proprotein convertase subtilizing Kexin-9 (PCSK-9) and sterol regulatory element-binding protein-2 (SREBP-2) were upregulated while the expression of low-density lipoprotein-receptor (LDL-R) gene was downregulated in the HCD compared to ND, TRTSM and ZACAH extracts treatment groups ($p<0.05$). The proteins levels of nitric oxide (NO) and tissue-type plasminogen activator (t-PA) were significantly downregulated while the tissue factor (TF) and plasminogen activator inhibitor-1 (PAI-1) protein concentrations were upregulated in the HCD group compared to ND, TRTSM and ZACAH extracts treatment groups ($p<0.05$). The mRNA expression of endothelial prothrombotic, coagulation related genes and the genes of the fibrinolytic system, including TF, PAI-1 and thrombin-activatable fibrinolysis inhibitor (TAFI) were upregulated, whereas the expression of endothelial nitric oxide synthase (eNOS), tissue factor pathway inhibitor (TFPI) and t-PA genes were downregulated in the HCD group compared to ND, TRTSM and ZACAH extracts treatment groups ($P<0.05$). No significant difference was recorded between ZACAH extracts treatment groups, ND and TRTSM treatment groups at $p<0.05$. The extent of the relationship between related proteins and genes expression profile determined by bivariate Pearson's product-moment correlation coefficient (r) was positively correlated across the groups. The findings suggested that ZACAH extracts supplementation could ameliorate the dysfunctional endothelial, activated platelet, hepatic steatosis and molecular atherothrombotic changes in SD rats fed with HCD, and could be considered a potential candidate for the treatment of hyperlipidaemia-related atherothrombotic cardiovascular diseases.

Keywords: High cholesterol diet, atherothrombotic proteins and genes, ZACAH extracts,

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENCIRIAN SELULAR DAN MOLEKULAR TERHADAP
PENANDA BERKAIT ATEROTROMBOSIS KE ATAS TIKUS SPRAGUE
DAWLEY YANG DIBERI MAKAN DIET BERKOLESTEROL TINGGI
DISUPLEMEN DENGAN EKSTRAK HERBA**

Oleh

IBRAHIM KALLE KWAIFA

September 2021

Pengerusi : Profesor Madya Sabariah Md Noor, MD, MPath, AM
Fakulti : Perubatan dan Sains Kesihatan

Penyakit kardiovaskular aterotrombotik adalah punca kematian yang paling lazim diseluruh dunia. Walaupun antiplatelet, antikoagulan, simvastatin dan lain-lain kumpulan ubat-ubatan berkaitan yang dipreskrip secara global untuk gangguan ini telah tersedia, sesetengahnya biasanya dikaitkan dengan beberapa kesan buruk. Intervensi berasaskan sumber semula jadi telah diperakui sebagai alternatif. Kajian ini menggunakan gabungan ekstrak herba untuk rawatan aterotrombosis dalam tikus Sprague Dawley (SD) yang diberi makan diet berkolesterol tinggi (HCD). Kajian itu menyiasat potensi terapeutik ekstrak herba; gabungan daripada; *Zingiber officinale* (halia), *Allium sativum* (bawang putih), *Citrus Limon* (lemon), *Malus Domestica* (epal) / cuka sari epal dan Madu (ZACAH) pada penanda berkaitan aterotrombosis secara selular dan molekular ke atas tikus SD yang diberi makan HCD. Pengenalpastian sebatian fitokimia utama yang terdapat dalam ekstrak ZACAH telah dilakukan dengan pemprofilan kromatografi cecair berprestasi tinggi-spektrofotometri jisim (UHPLC-MS) pada kedua-dua model negatif dan positif. Tiga puluh enam tikus SD jantan dibahagikan secara rawak kepada enam kumpulan mengikut jenis diet yang diberikan; diet biasa (ND), HCD, rawatan dengan simvastatin (TRTSM) pada kadar 10mg/kg mengikut berat badan (BW), rawatan dengan ekstrak ZACAH (TRT1) pada 1mg, 3mg (TRT2), dan 5mg (TRT3) sekilogram mengikut berat badan. Pada akhir minggu ke 12 eksperimen, tikus telah dikorbankan dan darah terkumpul telah dilakukan ujian profil lipid, ekspresi protein dan analisis ultrastruktur platelet dengan scanning electron microscope (SEM). Indeks aterogenik (AI) dan peratus perlindungan dikira berdasarkan keputusan yang diperolehi daripada profil lipid. SEM juga digunakan untuk memeriksa perubahan ultrastruktur endothelial aorta. Aorta dan tisu hati juga telah digunakan untuk pemeriksaan histopatologi menggunakan pewarnaan haematoxylin & eosin (H & E) dan trichrome masson (MT) nada. Tisu-tisu ini juga di proses untuk meneliti ekspresi mRNA bagi gen berkaitan aterotrombosis metabolisme lipid, protrombotik dari endothelial dan gen berkaitan pembekuan, dan gen sistem fibrinolitik. Sepuluh sebatian fitokimia

berkaitan yang terdapat dalam ekstrak ZACAH telah dikenal pasti, termasuk gingerol (masa pengekal (RT); 18.62, sebatian%; 5.6), hesperidin (11.19, 12.1), naringin (10.07, 13.1), sulindac (13.96, 9.7), scoparone (15.86, 8.0), hesperetin (11.18, 12.1), rutin (9.90, 13.2), limonin (17.87, 6.2), asid karboksilik (3.30, 18.9) dan asid sitrik (2.18, 20). Peningkatan BW dan pengambilan makanan telah turut direkodkan. Perbezaan yang ketara diperhatikan dalam kumpulan HCD berbanding kumpulan rawatan ekstrak ND, TRTSM dan ZACAH ($p < 0.05$). Jumlah kolesterol (TC), trigliserida (TG) dan lipoprotein berketumpatan (LDL) rendah telah meningkat dengan ketara dan penurunan lipoprotein berketumpatan tinggi (HDL) telah diperhatikan dalam kumpulan HCD berbanding kumpulan rawatan ekstrak ND, TRTSM dan ZACAH ($p < 0.05$). Perbezaan ketara diperhatikan dalam AI HCD berbanding kumpulan rawatan ekstrak ND dan ZACAH ($p < 0.05$). Peratus perlindungan telah dipertingkatkan pada TRTSM (18%), TRT1 (55.6%), TRT2 (59.7%) dan TRT3 (72.8%) dalam cara yang bergantung kepada dos. Analisa SEM menunjukkan aterotrombosis yang teratur, penambahan sel otot licin dalam intima, dan platelet yang teraktif dengan ketara dalam kumpulan HCD. Pemeriksaan histopatologi menunjukkan peningkatan ketara dalam nisbah intima-media, peningkatan gentian kolagen dan steatosis hepatic ketara dalam kumpulan HCD berbanding kumpulan rawatan ekstrak ND, TRTSM dan ZACAH ($p < 0.05$). Ekspresi mRNA metabolisme lipid gen, termasuk *3-hydroxy-3-methyl-glutaryl-coenzyme A reductase (HMG-CoA-R)*, *proprotein convertase subtilizing Kexin-9 (PCSK-9)* and *sterol regulatory element-binding protein-2 (SREBP-2)* dikawal selia manakala ekspresi gen reseptor lipoprotein berketumpatan rendah (LDL-R) berkurangan dalam HCD berbanding kumpulan rawatan ekstrak ND, TRTSM dan ZACAH ($p < 0.05$). Tahap protein oksida nitrat (NO) dan pengaktif plasminogen jenis tisu (t-PA) telah di bawah kawal selia dengan signifikan, manakala factor tisu (TF) dan kepekatan protein penghambat pengaktif plasminogen-1 (PAI-1) telah di atas kawal selia dalam kumpulan HCD berbanding kepada kumpulan rawatan ekstrak ND, TRTSM dan ZACAH ($p < 0.05$). Ekspresi mRNA protrombotik endothelial, gen berkaitan pembekuan dan gen sistem fibrinolitik, termasuk TF, PAI-1 dan *thrombin-activatable fibrinolysis inhibitor (TAFI)* telah di dikawal selia, manakala ekspresi gen mRNA *endothelial nitric oxide synthase (eNOS)*, perencat laluan faktor tisu (TFPI) dan t-PA berkurangan dalam kumpulan HCD berbanding kumpulan rawatan ekstrak ND, TRTSM dan ZACAH ($P < 0.05$). Tiada perbezaan yang ketara direkodkan antara kumpulan rawatan ekstrak ZACAH, kumpulan rawatan ND dan TRTSM pada $p < 0.05$. Tahap hubungan antara profil ekspresi protein dan gen yang berkaitan yang ditentukan oleh pekali korelasi momen-produk bivariat Pearson (r) telah dikaitkan secara positif merentas kumpulan. Penemuan ini mencadangkan bahawa suplemen ekstrak ZACAH berupaya memperbaiki endothelial disfungsi, hiperaktif platelet, steatosis hepatic dan perubahan aterotrombotik molekul dalam tikus SD yang diberi HCD dan mungkin dianggap sebagai pilihan yang berpotensi untuk rawatan penyakit kardiovaskular aterotrombotik yang berkaitan dengan hiperlipidaemia.

Kata kunci: Diet tinggi kolesterol, protein dan gen aterotrombotik, ekstrak ZACAH

ACKNOWLEDGEMENTS

All praises are to Almighty Allah (SWT), the lord of the world, the Most Gracious and Most Merciful, who saves my life and bestows on my strength, wisdom and of all blessing, for protecting, sustaining and sparing my life to conduct this research to completion, despite several challenges. May He bless the acquired certificate, make it beneficial to humanity and pray through His infinite mercy for continuous blessing till the end of my life. Ameen thumma Ameen.

My profound gratitude goes to Assoc. Prof. Dr Sabariah Md Noor. The head of my supervisory committee, my adviser and mentor, for her unrelenting guidance, close supervision, simplicity, accessibility, enthusiasm, suggestions, comments and corrections, despite her tight clinical schedules, making it possible for this research to reach its zenith. May Allah, in his infinity mercy, reward you and grant you all the best your heart desires for remodelling me to appreciate scientific research, particularly in the field of atherothrombosis.

My appreciation also goes to my co-supervisors, Assoc. Prof. Dr Hasnah Bahari and Assoc. Prof. Dr Yoke Keong Yong for their giant contributions to the design, implementation and reporting of this study. Indeed, their teamwork and mutual understanding made this thesis a possible reality. I also highly appreciate Miss Kokila Vani Perumal of Fakulti Perubatan Dan Sains Kesihatan (FPSK), UPM and Dr Santhra Segaran Balan of Management and Science University (MSU), Malaysia, for their wonderful contributions. To Dr Zolkapli Eshak and Noorhayati, Faculty of Pharmacy, University Institute Technology, Mara (UiTM), Malaysia, for their wonderful contributions during my SEM histopathological studies.

I would also like to appreciate the Dean and Deputy Dean, Faculty of Medicine and Health Sciences, the Dean and Deputy Dean, School of Graduate Studies, UPM for the training environment, a series of Putra Sarjana seminars and other learning opportunities given to me during my study period. I also thank the management of Usmanu Danfodiyo University Sokoto (UDUS) and TETFUND for the opportunity and supports given to me throughout my study period, despite the limited manpower in my department.

I wholeheartedly acknowledged the immeasurable support given by our haematology laboratory (UPM) staff, particularly, Amrina, Saidatu and Ahmad. Also, to the staff of the Anatomy Department, especially Miss Sharmala, and Histopathology lab, more appreciably, Mrs Juita and Mrs Noor for their tireless supports. My special thanks to all my colleagues here in UPM, including Dr Ismaila Gombe, Dr Hassan Yahaya, Dr Raji, Dr Abubakar Dankumo Gombe, Dr Mallam Buhari, Dr Abbas Kazaure, Dr Ibrahim Sani Kankara, Dr Auwalu Gana, Dr Auwalu Muhammad (Nursing), Dr Ardo, Dr Baba Ali Alanguburo, Dr Mahmud Katsina, Dr Abdullahi Katsina and Dr Sharif, it has been a pleasure to be part of you all and making my staying in UPM and Malaysia a happy one. To all my friends and colleagues back home in Nigeria, including Alh. Yusuf Kanji,

Mallam Garba Abdullahi, too many to mention, that I cherish all of you for many years of friendship, for the wonderful moments we had together.

I will not forget the striving efforts of my beloved late father, Alh. Musa Kalle, I owe you special prayer and respect for guiding me to reach this level in my life. To my dearest and beloved late mother, Halimatu Musa Kalle, I have no enough words in this world to describe how important you were to me or to thank you for your understanding, caring, and tolerance which has nurtured me from the cradle with caring hands and gentle song for soothing a child to make me what I am today. Inna, the person I like most in my life, you taught me, held me, stood by my side at any given condition, help me on the right path, encouraged me to face the challenges of this world and most importantly loved me unconditionally. I owe you a debt that I can't ever pay back except with prayers to Almighty Allah (SWT) to reward you abundantly, forgive you and grant you a house in Jannatul-Firdausi, Ameen thumma Ameen. Your commitments shall never be in vain.

I am also very grateful to appreciate the efforts of my children and the entire family, particularly Muhammad Fodiyo Ibrahim, who greatly functions like a father in the house. The daily prayers from my children, Mansur Ibrahim Kwaifa, Muhammad Fodiyo Ibrahim, Muqaffah Ibrahim, Muhaisin Ibrahim, Nana Aishatu Ibrahim, Maryam Ibrahim Kalle and the last but not the least Khadijatulkubura Ibrahim Kalle, including Hajaratu Ibrahim Mailalle, are highly appreciated. May Allah rewards you all abundantly. I love you all, May Allah bless you, Ameen thumma Ameen.

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Sabariah Md Noor, MD, MPath, AM

Associate Professor (Medical)
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Hasnah binti Bahari, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Yoke Keong Yong, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 10 February 2022

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____ Date: _____

Name and Matric No: Ibrahim Kalle Kwaifa

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee:

Associate Professor Dr. Sabariah Md Noor

Signature: _____

Name of Member
of Supervisory
Committee:

Associate Professor Dr. Hasnah Bahari

Signature: _____

Name of Member
of Supervisory
Committee:

Associate Professor Dr. Yoke Keong Yong

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vii
DECLARATION	ix
LIST OF TABLES	xv
LIST OF FIGURES	xvi
LIST OF APPENDICES	xxi
LIST OF ABBREVIATIONS	xxiii
CHAPTER	
1 INTRODUCTION	1
1.1 Atherothrombosis	1
1.2 Problem Statement	2
1.3 Study Justification	3
1.4 The objective of the Study	4
1.4.1 General Objective	4
1.4.2 Specific Objectives	4
1.5 Research Hypothesis	4
2 LITERATURE REVIEW	6
2.1 Atherosclerosis and Atherothrombosis: An Overview	6
2.2 Prevalence of Atherothrombosis	6
2.3 Risk Factors for Atherothrombosis	8
2.3.1 The Modifiable Risk Factors	8
2.3.2 Non-Modifiable Risk Factors	9
2.3.3 Other Risk Factors of Atherothrombosis:	9
2.4 Clinical Manifestations of Atherothrombosis	11
2.5 Pathogenesis of Atherothrombosis	12
2.5.1 Contribution of Total Cholesterol to the Progress of Atherothrombosis	13
2.5.2 Endothelial Involvement	20
2.5.3 Platelet Involvement	27
2.5.4 Haemostasis	28
2.5.5 Thrombosis	31
2.6 The Molecular Biomarker of Atherothrombosis	33
2.7 Treatment of Atherothrombosis	34
2.8 The Contributions of Natural Products in the Discovery of New Medicine	35
2.8.1 Medicinal Plants Used for the Treatment of Atherothrombosis	36
2.9 Rat Model for Hyperlipidaemia-Related Atherothrombosis	37
2.10 Summary and Conclusion of the Literature Review	38

3	IDENTIFICATION OF THE MAJOR PHYTOCHEMICALS IN ZACAH EXTRACTS BY ULTRA-HIGH PERFORMANCE LIQUID CHROMATOGRAPHY-MASS SPECTROPHOTOMETRY (UHPLC-MS) AND ANIMAL STUDY	40
3.1	Introduction	40
3.2	Material and Methods	41
3.2.1	Ethical Consideration	41
3.2.2	Reagents and consumables	41
3.2.3	Equipment	42
3.2.4	Composition of ZACAH Extracts	42
3.2.5	Preparations of High Cholesterol Diet, ZACAH Extract and Simvastatin (Standard Drug).	42
3.2.6	Animal	43
3.2.7	Experimental Design	43
3.2.8	Feeding	43
3.2.9	Sample Collection and preparations	44
3.2.10	Calculation of Atherogenic Index and % Protection	46
3.2.11	Total RNA Extraction	46
3.2.12	RNA Concentration, Purity and Integrity	46
3.2.13	Complementary DNA (cDNA) Synthesis	47
3.2.14	Primer Design and Synthesis for qPCR	47
3.2.15	Endogenous Control for Relative Quantification (reference genes)	47
3.2.16	Primers Optimization for qPCR	48
3.2.17	Validation of Efficiency of Primers for qPCR	48
3.2.18	Quantitative RT-qPCR	48
3.2.19	Assay for Reactive Oxygen Species	49
3.2.20	Identification of the Phytochemicals Present in ZACAH Extracts	49
3.3	Statistical Analysis	51
3.4	Results	51
3.4.1	ORAC Values, Elastase Inhibitory Activity, Lipoygenase Inhibitory Assay, 2,2-diphenyl-1-picrylhydrazyl (DPPH) Free Radical Scavenging Activity and Total Phenolic Content (TPC) of ZACAH Extracts	51
3.4.2	Identified Phytochemicals Present in ZACAH Extracts (Methanol)	51
3.4.3	UHPLC Method Validation	58
3.4.4	Linearity	60
3.4.5	Accuracy	60
3.5	Discussion	62
3.6	Conclusion	69

4	EFFECTS OF ZACAH EXTRACTS ON BODY AND LIVER WEIGHTS, FOOD INTAKE, LIPID PROFILE AND HEPATIC HISTOPATHOLOGICAL CHANGES IN SD RATS FED WITH HCD	70
4.1	Introduction	70
4.2	Material and Methods	71
4.2.1	Determination of Body and Liver Weights (g), and Food Intake (g).	71
4.2.2	Biochemical Studies	71
4.2.3	Haematoxylin and Eosin (H & E) Staining of the Liver	71
4.2.4	Histopathological Scoring of the Liver	71
4.3	Statistical Analysis	72
4.4	Results	72
4.4.1	Effect of ZACAH Extracts on Body Weight and Food Intake	72
4.4.2	Effects of ZACAH Extracts on Liver Weight and Liver Ratio	73
4.4.3	Effects of ZACAH Extracts on Lipid Profile	73
4.4.4	The Mean Hepatic Score of the Liver	75
4.5	Discussion	79
4.6	Conclusion	82
5	ZACAH EXTRACTS ATTENUATED PLATELET AND ENDOTHELIAL HISTOPATHOLOGICAL CHANGES IN SD RATS FED WITH HCD	83
5.1	Introduction	83
5.2	Materials and Methods	84
5.2.1	Scanning Electron Microscopy	84
5.2.2	Histopathological Analysis	85
5.2.3	Scoring of the Activated Platelet	85
5.2.4	Histopathological Scoring of the Aorta	86
5.2.5	Statistical Analysis	87
5.3	Results	87
5.3.1	SEM Examination	87
5.3.2	Histopathological Changes	95
5.4	Discussion	103
5.5	Conclusion	106
6	EFFECTS OF ZACAH EXTRACTS ON MRNA EXPRESSION PROFILING OF THE LIPID METABOLISM AND HAEMOSTASIS-RELATED GENES IN SD RATS FED WITH HCD	107
6.1	Introduction	107
6.2	Material and Methods	111
6.2.1	Precautionary Measures	111
6.2.2	Statistical Analysis	111
6.3	Results	111
6.3.1	RT-qPCR Optimization	111

6.3.2	ZACAH Extracts Attenuated Cholesterol Biosynthesis by Suppressing the mRNA Expression of HMG-CoA-R, SREBP-2 and PCSK-9 Genes.	112
6.3.3	To analyse the Levels of Atherothrombosis-Related Proteins in SD Rats Fed with HCD Supplemented with ZACAH Extracts.	114
6.3.4	Evaluation of the mRNA Expression Profiling of Endothelial Prothrombotic and Coagulation-Related Genes in SD Rats Fed with HCD Supplemented with ZACAH Extracts	118
6.3.5	Determination of the mRNA Expression Profiling of Fibrinolytic Genes in SD Rats Fed with HCD Supplemented with ZACAH Extracts.	120
6.3.6	To Correlate the Activities of Atherothrombosis-Related Proteins with the Expression Profile of Atherothrombosis-related Genes, and the Genes Expression Patterns in the SD Rats Fed with HCD Supplemented with ZACAH Extracts.	122
6.4	Discussion	132
6.5	Conclusion	139
7	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	
7.1	Research Summary	140
7.2	General Conclusion	142
7.3	Recommendations for Future Research	144
	REFERENCES	145
	APPENDICES	176
	BIODATA OF STUDENT	219
	LIST OF PUBLICATIONS	220

LIST OF TABLES

Table		Page
2.1	Classification of Lipoproteins	15
3.1	Accurate Mass Measurement of the Phytochemical Compounds Observed in the ESI-MS and ESI-MS/MS spectra with RT	53
3.2	UHPLC Method Selectivity Validation	60
3.3	% Recovery for the Phytochemicals Present in ZACAH Extracts	61
4.1	The Hepatic Steatosis Scoring of the Liver	72
4.2	Effects of ZACAH Extracts on Body Weight and Food Intake	73
4.3	Liver Weight and Liver Ratio of the Experimental SD Rats	73
4.4	Effects of ZACAH Extracts on Lipid Profile of the Experimental SD Rats	74
4.5	Effects of ZACA Extracts and Simvastatin on Atherogenic Index (AI) and % Protection	74
5.1	Scoring of Activated Platelet	86
5.2	Histopathological Scoring of Aorta	86

LIST OF FIGURES

Figure		Page
1.1	Research Concept Map	5
2.1	World Distribution of Ischemic Heart Disease	7
2.2	Major Risk Factors for Atherothrombosis	11
2.3	Clinical Presentation of Atherothrombosis	12
2.4	Mechanisms of Atherothrombotic Cardiovascular Diseases	13
2.5	Spectrum of lipoprotein particles	14
2.6	The Three basic Pathways Involved in Lipid Metabolism	17
2.7	Regulation and Synthesis of Cholesterol	18
2.8	Main Functions of the Endothelial Cells	21
2.9	Some Anti-atherothrombotic Potential of the Normal Vascular Endothelium	22
2.10	The Key Mechanisms Associated with the Development of Atherosclerosis	25
2.11	Metabolisms of Collagen and Elastin in the Plaque's Fibrous Cap	26
2.12	Basic Mechanisms of Platelet Activation, Adhesion and Aggregation. PAF	28
2.13	Coagulation Cascade and its Interrelationship with the Fibrinolytic System	30
2.14	Mechanisms of Virchow Triad in the Pathophysiology of Thrombus Formation	32
2.15	The Main Differences between Arterial and Venous Thrombosis	33
2.16	The Percentage of Natural Products Values by the Years	35
2.17	The Processes Involved in the Development of Atherothrombotic CVDs	39
3.1	Combination of ZACAH Extracts	50

3.2	UHPLC Chromatogram	59
4.1 A	The H & E Photomicrographs of the Liver of the Normal Diet (ND) Group	75
4.1 B	The H & E Photomicrographs of the Liver of High Cholesterol Diet (HCD) Group	76
4.1 C	The H & E Photomicrographs of the Liver of Simvastatin Treatment at 10mg/kg of Body Weight (TRTSM)	76
4.1 D	The H & E Photomicrographs of the Liver of ZACAH Extracts Treatment at 1mg/kg of body weight (TRT1)	77
4.1 E	The H & E Photomicrographs of the Liver of ZACAH Extracts Treatment at 3mg/kg of body weight (TRT2)	77
4.1 F	H & E Photomicrographs of the Liver of ZACAH Extracts Treatment at 5mg/kg of body weight (TRT3) Group	78
4.2	The Mean Hepatic Steatosis Scoring of the Liver	78
5.1	Mechanism of Plaque Rupture and the Formation of Atherothrombosis	84
5.2A	Photomicrograph of Platelet Ultrastructure of the Normal Diet (ND) Group	88
5.2B	Photomicrograph of Platelet Ultrastructure of the High Cholesterol Diet (HCD) Group	88
5.2C	Photomicrograph of Platelet Ultrastructure of the Simvastatin Treatment at 10mg/kg of Body Weight (TRTSM)	89
5.2D	Photomicrograph of Platelet Ultrastructure of the ZACAH Extracts Treatment at 1mg/kg of Body Weight (TRT1)	89
5.2E	Photomicrograph of Platelet Ultrastructure of the ZACAH Extracts Treatment at 3mg/kg of Body Weight (TRT2)	90
5.2F	Photomicrograph of Platelet Ultrastructure of the ZACAH Extracts Treatment at 5mg/kg of Body Weight (TRT3)	90
5.3	The SEM Mean Value Scored of Activated Platelet	91
5.4A	Photomicrograph of the Aorta of the Normal Diet (ND) Group	92
5.4B	Photomicrograph of Aorta of the High Cholesterol Diet (HCD) Group	92

5.4C	Photomicrograph of Aorta of the Simvastatin Treatment at 10mg/kg of Body Weight (TRTSM)	93
5.4D	Photomicrograph of Aorta of the ZACAH Extracts Treatment at 1mg/kg of Body Weight (TRT1)	93
5.4E	Photomicrograph of Aorta of the ZACAH Extracts Treatment at 3mg/kg of Body Weight (TRT2)	94
5.4F	Photomicrograph of Aorta of the ZACAH Extracts Treatment at 5mg/kg of Body Weight (TRT3)	94
5.5	The SEM Intima-Media Ratio of the Aorta	95
5.6A	The H & E Photomicrograph of Aorta of the Normal Diet (ND) Group	96
5.6B	The H & E Photomicrograph of Aorta of the High Cholesterol Diet (HCD) Group	96
5.6C	The H & E Photomicrograph of Aorta of the Simvastatin Treatment at 10mg/kg of Body Weight (TRTSM)	97
5.6D	The H & E Photomicrograph of Aorta of the ZACAH Extracts Treatment at 1mg/kg of Body Weight (TRT1)	97
5.6E	The H & E Photomicrograph of Aorta of the ZACAH Extracts Treatment at 3mg/kg of Body Weight (TRT2)	98
5.6F	The H & E Photomicrograph of Aorta of the ZACAH Extracts Treatment at 5mg/kg of Body Weight (TRT3) Group	98
5.7	The H & E Intima-Media Ratio of the Aorta	99
5.8A	The MT Stain Photomicrograph of Aorta of the Normal Diet (ND) Group	100
5.8B	The MT Stain Photomicrograph of Aorta of the High Cholesterol Diet (HCD) Group	100
5.8C	The MT Stain Photomicrograph of Aorta of the Simvastatin Treatment at 10mg/kg of Body Weight (TRTSM)	101
5.8D	The MT Stain Photomicrograph of Aorta of the ZACAH extracts Treatment at 1mg/kg of body weight (TRT1)	101
5.8E	The MT Stain Photomicrograph of Aorta of the ZACAH extracts Treatment at 3mg/kg of Body Weight (TRT2)	102

5.8F	The MT Stain Photomicrograph of Aorta of the ZACAH extracts Treatment at 5mg/kg of Body Weight (TRT3)	102
5.9	The Masson's Trichrome Mean Value of the Collagen Score of the Aorta in Area (%)	103
6.1	Proposed Mechanisms of Action of ZACAH Extracts on Lipid Metabolism Atherothrombosis-Related Genes	109
6.2	Possible Mechanisms of Action of ZACAH Extracts	110
6.3	Agarose Gel Electrophoresis Results of the Lipid Metabolism Atherothrombosis-related and reference Genes	112
6.4	Effects of ZACAH Extracts on Lipid Metabolism Atherothrombosis-Related Genes.	113
6.5	The Standard Curve of each Atherothrombosis-Related Protein and Coefficient of Determination (R^2)	115
6.6	Expression of Atherothrombosis-Related Proteins	117
6.7	Agarose Gel Electrophoresis Results of the Endothelial Prothrombotic and Coagulation-Related Gene	118
6.8	Effects of ZACAH Extracts on Endothelial Prothrombotic and Coagulation-Related Genes	119
6.9	Agarose Gel Electrophoresis Results of Fibrinolytic Genes.	120
6.10	Effects of ZACAH Extracts on Fibrinolytic Genes	121
6.11	Correlation between the Levels of NO Protein and eNOS Gene	123
6.12	Correlation between the Level of TF Protein and TF Gene	124
6.13	Correlation between the t-PA Protein and t-PA Gene	125
6.14	Correlation between the PAI-1 Protein and PAI-1 Gene	126
6.15	Gel Expression Patterns of the Related Genes	127
6.16	Correlation between the Levels of NO Protein and eNOS Gene Expression Patterns	128
6.17	Correlation between the t-PA Protein and t-PA Gene Expression Patterns	129

6.18	Correlation between the PAI-1 Protein and PAI-1 Gene Expression Patterns	130
6.19	Correlation between the Level of TF Protein and TF Gene Expression Patterns. Results are expressed as mean \pm SEM (n = 6)	131
7.1	Findings of the entire project	143



LIST OF APPENDICES

Appendix	Page	
A1	MeSTIC Certificate	176
A2	Agi-Food & Veterinary Authority Singapore	177
A3	Certificate of Authentication	178
A4	Certificate of Analysis	179
A5	Protocol for the Identification of Phytochemical Compounds Present in ZACAH Extracts	180
A6	Nutritional Composition of ZACAH Extracts	181
A7	Chromatogram of the Phytochemical Compounds Present in ZACAH Extracts at Positive Mode	182
A8	Chromatogram of the Phytochemical Compounds Present in ZACAH Extracts at Negative Mode	183
A9	Linear Regression Coefficient Curves for the Ten Identified Phytochemicals	194
B1	Animal Ethical Approval	195
B2	Supporting Grant (GP-IPS/2020/9686500)	196
B3	Dosage Calculations	199
B4	Study Design	201
B5	Hyperlipidaemia Male Sprague Dawley Rat Model	202
C1	Procedure for Scanning Electron Microscope (SEM)	203
C2	Procedure for Haematoxylin and Eosin (H & E)	204
C3	Procedure for Masson's Trichrome (MT) Staining	206
D1	Table 6.1: Specific Primer Sets for the Lipid Metabolism and Haemostasis Atherothrombosis-related and Reference Genes	207
D2	Protocol for RNA Extraction from Tissues (Aorta and Liver)	208
D3	Procedure for NanoDrop:	210

D4	Quantification Results of the Extracted RNA by NanoDrop Spectrophotometry	211
D5	Procedure for cDNA Synthesis	212
D6	Procedure for Conventional PCR and Gel Electrophoresis	213
D7	Banding of Atherothrombosis and Reference Genes	214
D8	Procedure for qPCR	215
D9	Standard Curve	216
D10	Amplification Plot	216
D11	Melting Curve	216
E1	Procedure for Enzyme-linked Immunosorbent Assay (ELISA)	217
E2	Procedure for MyAssays	218

LIST OF ABBREVIATIONS

8-OHdG	8-Hydroxy-2-Deoxyguanosine
AA	Arachidonic Acid
AB	Antibody
ABCA1	ATP-Binding Cassette Transporter A1
ABCG1	ATP-Binding Cassette Transporter G1
ABI	Ankle-Brachial Index
ABPI	Ankle-Brachial Blood Pressure Index
Abs	Absorbance
ACAS	Acetyl CoA Synthase
ACS	Acute Coronary Syndrome
ACV	Apple Cider Vinegar
AD	Adventitia
ADAMA	Asymmetric Dimethyl-L-Arginine
ADP	Adenosine Diphosphate
AGES	Advanced Glycation End-Product
AHA	American Heart Association
ALP	Alkaline Phosphatase
ALT	Alanine Transferase
AMI	Acute Myocardial Infarction
AMPK	Activated-Mitogen Protein Kinase
Ang II	Angiotensin-II
ANOVA	Analysis of Variance
AP-1	Activated Protein-1

APC	Activated Protein C
Apo AI	Apolipoprotein AI
Apo B48	Apolipoprotein B [(%)]
Apo B100	Apolipoprotein B [100 (%)]
Apo C	Apolipoprotein C
Apo E	Apolipoprotein E
ARAS	Atomic Resonance Absorption Spectrometry
AST	Aspartate aminotransferase
AT	Arterial Thrombosis
ATIII	Antithrombin III
ATE	Arterial Thromboembolism
ATP	Adenosine Triphosphate
BD	Bile Ducts
BFGF	Basic Fibroblast Growth Factor
BH ₄	Tetrahydrobiopterin
BM	Bone Marrow
BMI	Body Mass Index
BP	Blood Pressure
Ca ²⁺	Calcium Ion
CAC	Coronary Artery Calcium
CAD	Coronary Arterial Disease
CaMKK-2	Ca ²⁺ + Calmodulin Dependent Protein Kinase Kinase-2
CAMs	Cells Adhesion Molecules
CARM-1	Coactivator-Associated Methyltransferase-1

CCL	Chemokine (C-C Motif) Ligand
CCL ₄	Carbon Tetrachloride
CD	Cluster of Differentiation
CDCP	Centre for Disease Control and Prevention
cDNA	Complementary Deoxyribonucleic Acid
CD40L	CD40 Ligand
C/EBPs	CCAAT/Enhancer-Binding Proteins
CE	Cholesterol Ester
CETP	Cholesterol Ester Transferring Protein
cGMP	Cyclic Guanidine Monophosphate
CHDs	Coronary Heart Diseases
CM	Chylomicron
CO ₂	Carbon Dioxide
COX	Cyclooxygenase
Cr	Creatinine
CRP	C-Reactive Protein
CS	Cigarette Smoking
CV	Coefficient of Variation
CVDs	Cardiovascular Diseases
DEPC-treated water	Diethylpyrocarbonate-treated water
DIC	Disseminated Intravascular Coagulopathy
DM	Diabetes Mellitus
DMIQE	Digital Minimum Information for Publication of Quantitative RT-PCR Experiments
DNL	<i>De Novo</i> Lipogenesis

DPX	Distyrene Plasticizer and Xylene
DVT	Deep Vein Thrombosis
DVTE	Deep Vein Thromboembolism
DW	Distilled Water
ECs	Endothelial Cells
ECM	Extracellular Matrix
EDHF	Endothelium-Derived Hyperpolarizing Factor
EDTA	Ethylenediaminetetraacetic Acid
EGF	Epidermal Growth Factor
ELISA	Enzyme-Linked Immunosorbent Assay
EMCV	Encephalomyocarditis
EMCs	Electron Microscopes
eNOS	Endothelial Nitric Oxide Synthase
EPCR	Endothelial Protein C Receptor
EPI	Extrinsic Pathway Inhibitor
ESI	Electrospray Ionization
ET1	Endothelin-1
FCHL	Familial Combined Hyperlipidaemia
FDA	Food And Drugs Administration
FDPs	Fibrin Degradation Products
FFAs	Free Fatty Acids
FGF	Fibroblast Growth Factor
FH	Family History
FHC	Familial Hypercholesterolemia

FHL	Familial Hyperlipidaemia
FMD	Flow Mediated Dilation
FN	Fibrinogen
FPA	Fibrinopeptide A
FPB	Fibrinopeptide B
FPSK	Fakulti Perubatan Dan Sains Kesihatan
FI	Fibrinogen
FII	Prothrombin
FIII	Thromboplastin (Tissue Factor)
FIV	Calcium Ion
FV	Labile Factor/ Proaccelerin
FV Leiden	Factor V Leiden
FVI	Accelerin
FVII	Stable Factor/ Proconvertin
FVIII	Anti-Haemophilia A Factor
FIX	Christmas Factor/ Anti-Haemophilia B
FX	Stuart Prower Factor
FXI	Anti-Haemophilia C Factor/ Plasma Thromboplastin Antecedent
FXII	Hageman Factor
FXIII	Fibrin Stabilizing Factor
GIT	Gastrointestinal Track
GOT	Gene of Interest
GP	Glycoprotein
GPIb	Glycoprotein Ib

GPx	Glutathione Peroxidase
GPVI	Glycoprotein VI
GSH	Glutathione
GWAS	Genome-Wide Association Study
HA	Hepatic Artery
Hb	Haemoglobin
HBP	High Blood Pressure
HBV	Hepatitis B Virus
HCD	High Cholesterol Diet
HCV	Hepatitis C Virus
HDL	High Density Lipoprotein
HDL-C	High Density Lipoprotein-Cholesterol
H & E	Haematoxylin and Eosin
HESI	Heat Electrospray Ionisation
HF	History of The Family
HFD	High Fat Diet
HGF	Hepatocyte Growth Factor
HIV	Human Immune Virus
HL	Hepatic Lipase
HMDS	Hexamethyldisilane
HMG-CoA-R	3-Hydroxy-3-Methylglutaryl Coenzyme A-Reductase
HMWK	High Molecular Weight Kininogen
HOCl	Hypochlorous Acid
HPLC	High Performance Liquid Chromatography

HPS	Hormone Sensitive Lipase
HRP	Horse Reddish Peroxidase
HSPG	Heparan Sulphate Proteoglycans
H ₂ O ₂	Hydrogen Peroxide
H ₂ S	Hydrogen Sulphide
HT	Hormone Therapy
5-HT	5-hydroxytryptamine
I	Intima
ICAM-1	Intercellular Adhesion Molecule-1
IDL	Intermediate Lipoprotein
IGF-1	Insulin-Like Growth Factor-1
IHD	Ischaemic Heart Disease
IKK	Inhibitor of Kappa B Kinase
IL	Interleukin
IL-1 β	Interleukin-1 β
IL-6	Interleukin-6
IL-10	Interleukin-10
IMT	Intimal Media Thickness
INF- γ	Interferon-Gamma
iNOS	Inducible Nitric Oxide Synthase
INR	International Normalised Ratio
IS	Ischemic Stroke
JAM	Junctional Adhesion Molecule
JNK – c-Jun	C-Jun N-Terminal Kinase

KLF	Kruppel-Like Factor
L	Lumen
LC	Liquid Chromatography
LCAT	Lectin-Cholesterol-Acyltransferase
LC-MS	Liquid Chromatography- Mass Spectrophotometry
LD	Lipid Droplet
LDL	Low Density Lipoprotein
LDL-C	Low Density Lipoprotein-Cholesterol
LDL-R	Low Density Lipoprotein-Receptor
LOX-1	Lectin-Like Oxidized LDL Receptor-1
Lp(a)	Lipoprotein(a)
LPL	Lipoprotein Lipase
LRP-1	LDL-Receptor Related Protein-1
M	Media
MAPK	Mitogen-Activated Protein Kinase
MAPKP	Mitogen-Activated Protein Kinase Pathway
MCP-1	Monocyte Chemoattractant Protein-1
M-CSF	Monocyte Colony Stimulating Factor
MDA	Malondialdehyde
MI	Myocardial Infarction
MIP-1 α	Macrophages Inflammatory Protein-1 α
MiRNAs	Micro-RNAs
MMP-1	Matrix Metalloproteinase-1
MMP-2	Matrix Metalloproteinase-2

MMP-9	Matrix Metalloproteinase-9
MPV	Mean Platelet Volume
MS	Mass Spectrophotometry
MT	Masson's Trichrome
NAFLD	Non-Alcoholic Fatty Liver Disease
NASH	Non-Alcoholic Steatohepatitis
NCBI	National Centre for Biotechnology Information
ND	Normal Diet
NF-KB	Nuclear Factor Kappa-Light-Chain Enhancer of Activated B-Cells B
NHMS	National Health and Morbidity Survey
NIH	National Institutes of Health
NLM	National Library of Medicine
NO	Nitric Oxide
NS	Normal Saline
NSAID	Nonsteroidal Anti-inflammatory Drugs
OH	Hydroxyl
ONOO-	Peroxyneitrite
Ox-LDL	Oxide-Low Density Lipoprotein
Ox-LDL-Th	Oxide-Low Density Lipoprotein-T-Helper Lymphocyte
PACE-4	Proprotein Convertase Cleaving Enzyme-4
PADs	Peripheral Arterial Diseases
PAF	Platelet-Activating Factor
PAI-1	Plasminogen Activator Inhibitor-1
PAP	Physiologic Anticoagulation Pathway

PAPase	Phosphatidate Phosphohydrolase
PAR	Protease-Activated Receptor-1
PAR-1	Protease Activator Receptor-2
PBS	Phosphate Buffer Saline
PC	Protein C
PC-1	Proprotein Convertase-1
PCL	Prostacyclin
PCSK-9	Proprotein Convertase Subtilisin/Kexin Type-9
PDI	Protein Disulphide Isomerases
PDGF	Platelet-Derived Growth Factor
PE	Pulmonary Embolism
PECAM-1	Platelet Endothelia Cell Adhesion Molecule-1
PF4	Platelet Factor-4
PGH ₂	Prostaglandin H ₂
PGL	Prostaglandin
PGs	Proteoglycan
PKA	Protein Kinase A
PKR	Protein Kinase-R
PPARs	Peroxisome Proliferator-Activated Receptors
PS	Protein S
PUFAs	Polyunsaturated Fatty Acid
PVDs	Peripheral Vascular Diseases
QTOF	Quadrupole Time of Flight
R ²	Coefficient of Determination

r	Correlation Coefficient
RANTES	Regulated on Activating, Normal T-Cell Expressed and Secreted
RAS	Rat Sarcoma
RBCs	Red Blood Cells
Rho	Ras Homologous
RNA	Ribonucleic Acid
RNS	Reactive Nitrogen Species
ROS	Reactive Oxygen Species
rpm	Revolution Per Minute
RT-qPCR	Real-Time Quantitative Polymerase Chain Reaction
SKI-1	Subtilisin Kexin Isoenzyme-1
SLE	Systemic Lupus Erythematosus
SMCs	Smooth Muscle Cells
SNPs	Single Nucleotide Polymorphisms
SO ₂	Sulphur Dioxide
SPSS	Statistical Package for Social Sciences
SRs	Scavenger Receptors
SR-B1	Scavenger Receptor B Type 1
SREBP-2	Sterol Regulatory Element Binding Protein-2
Svcam-1	Soluble Vascular Cell Adhesion Molecule-1
TAE	Tris-Acetate-EDTA
TAFI	Thrombin-Activatable Fibrinolytic Inhibitor
TBE	Tris Borate-EDTA
TC	Total Cholesterol

TCFA	Thin-Cap Fibroatheroma
T2DM	Type-II Diabetes Mellitus
TEM	Transmission Electron Microscope
TF	Tissue Factor
TF-MPs	Tissue Factor-Bearing Microparticles
TFPI	Tissue Factor Pathway Inhibitor
TG	Triglycerides
TGF-B R-II	Transforming Growth Factor-B Receptor-II
TGRP	Triglyceride-Rich Lipoprotein Particles
Th	T-Helper Cells
TIA	Transient Ischaemic Attack
TLRs	Toll-Like Receptors
TM	Thrombomodulin,
TNF- α	Tumour Necrosis Factor-Alpha
TNF-1 β	Tumour Necrosis Factor-1 β
t-PA	Tissue-Type Plasminogen Activator
TPC	Total Phenolic Content
TRs	Thromboxane Receptors
TRT1	Treatment at 1mg/Kg Of Body Weight
TRT2	Treatment at 3mg/Kg Of Body Weight
TRT3	Treatment at 5mg/Kg Of Body Weight
TRTSM	Treatment with Simvastatin at 10mg/Kg Of Body Weight
T-TM	Thrombin-Thrombomodulin
TXA ₂	Thromboxane A ₂

TXB2	Thromboxane B2
UHPLC	Ultra-High Pressure Liquid Chromatography
ulvWF	Ultra-Large von Willebrand Factor
uPA	Urokinase Plasminogen Activator
uPA-R	Urokinase Plasminogen Activator-Receptor
VCAM-1	Vascular Cell Adhesion Molecule-1
VEGF	Vascular Endothelial Growth Factor
VKA	Vitamin K-Antagonists
VKAs	Vitamin K-Dependent Anticoagulants
VLDL	Very Low-Density Lipoprotein
VLDL-C	Very Low-Density Lipoprotein Cholesterol
VSMC	Vascular Smooth Muscle Cells
VT	Venous Thrombosis
VTE	Venous Thromboembolism
vWF	von Willebrand Factor
WHO	World Health Organization
WPB	Weibel-Palade Bodies
α_2 -AP	α_2 -Antiplasmin
ZACAH	Zingiber Officinale (Ginger), Allium Sativum (Garlic), Citrus Limon (Lemon), Malus Domestica (Apple)/ Apple Cider Vinegar and Honey.

CHAPTER 1

INTRODUCTION

1.1 Atherothrombosis

Atherothrombosis is an atherosclerotic lesion with thrombus formation and has been recognised as the major cause of ischemic heart disease and other cardiovascular diseases (CVDs), associated with global deaths (Grover & Mackman, 2020; Roth et al., 2020). The initial pathophysiology that gives rise to IHD is atherosclerosis, an inflammatory condition of the arterial vessels associated with the accumulation of lipids and metabolic modifications caused by several risk factors. Atherosclerosis is the underlying cause of CVDs characterized by plaque formation in the endothelial lumen. Atherosclerotic plaque is composed of accumulated lipids, cellular elements, including vascular smooth muscle cells (VSMCs), endothelial cells (ECs), immune mobilizing cells, such as monocyte, macrophage, T-cell and B-cell from blood circulation, calcium ions, and other formed blood elements (Shapiro & Fazio, 2017). Under normal physiology, cells are protected from the intracellular deposition of lipids by constantly regulated synthesis, through the influx and efflux of cholesterol. Lipids are organic compounds, insoluble in water, necessarily needed for normal metabolic processes. They are the basic components of the cell membrane, functioning as an energy reserve, participating in the absorption of fat-soluble vitamins, forming an essential part of various hormones, contributing effectively as co-factors, intracellular messengers, and maintaining the integrity of the cell membrane (Ahmed et al., 2021; Verma, 2016). Accumulation of lipids in the bloodstream might be attenuated by the endothelium, which participates significantly in the regulation of blood flow through its anti-inflammatory, anticoagulant and profibrinolytic activities. Compromising these mechanisms could lead to excessive deposition of cholesterol, leading to organs toxicity, associated with apoptosis and enhanced necrosis. Excessive accumulation of lipids in the bloodstream could result in several metabolic conditions, including hyperlipidaemia and obesity (Verma, 2016).

Existing report has demonstrated that hyperlipidaemia is associated with the increased serum concentrations of total cholesterol (TC), triglycerides (TG), low-density lipoprotein (LDL), and a corresponding decrease in the serum level of high-density lipoprotein (HDL), in high cholesterol-induced SD rats (Fattepur et al., 2018). Hyperlipidaemia has been recognised as the most common risk factor to atherothrombotic CVDs, such as coronary heart diseases (CHDs), cerebral stroke, myocardial infarction (MI), and renal dysfunction (Karam et al., 2018; Badimon et al., 2012). Several factors, including vascular endothelial lumen, thrombogenicity, local haemorrhology, systemic thrombogenicity, and fibrinolytic activities are known to mediate atherothrombotic propagation and formation (Asada, 2020). Arterial thrombi have been identified to contain aggregated platelets due to high blood velocity. However, established atherothrombosis from disrupted plaques may contain not only aggregated platelets but also a significant amount of fibrin. Plaque disruption facilitates both the activations of platelet and coagulation cascade at the injured vascular surface, leading to

a pronounced atherothrombotic process (Asada, 2020). Previous study has demonstrated high blood cholesterol and LDL levels after feeding SD rats with 4% cholesterol for four weeks (Balkanci et al., 2012). Therefore, a high cholesterol diet (HCD) could be utilized to develop high cholesterol SD rats model, targeting hyperlipidaemia (Rosenbaum & Chaudhuri, 2014). Several natural-based products have been used to treat various metabolic conditions, including CVDs (Kirichenko et al., 2020). Herbal extracts as natural sources contained *Zingiber officinale* (ginger), *Allium sativum* (garlic), *Citrus limon* (lemon), *Malus domestica* (Apple)/ apple cider vinegar and honey (ZACAH extracts), formulated to treat high blood pressure (HBP) and reduce cholesterol levels. This study utilises the above-mentioned mechanisms to investigate the therapeutic potentials of ZACAH extracts on atherothrombosis-related markers in SD rats fed with a high cholesterol diet (HCD). At present, there is a lack of information on the therapeutic potentials of ZACAH extracts on hyperlipidaemia-related atherothrombotic CVDs. Therefore, this present study investigated the mechanisms of action of ZACAH extracts on cellular and molecular atherothrombosis-related markers in SD rats fed with HCD. The efficacy of the phytochemicals present in ZACAH extracts would be compared with simvastatin, to provide experimental evidence for utilizing ZACAH extracts, to treat hyperlipidaemia-related atherothrombotic complications.

1.2 Problem Statement

The incidence and severity of IHD and other atherothrombosis-related CVDs are grossly increasing worldwide (Karam et al., 2018). Pharmacological agents, including statin, antiplatelet and anticoagulant, are the gold standard drugs prescribed to treat hyperlipidaemia-related atherothrombotic CVDs. The hallmark of these drugs is to inhibit the excess accumulation of lipids, activation of platelets, endothelial cells (ECs), and coagulation cascade while promoting profibrinolytic activities by facilitating the clearance of TG-rich lipoprotein and enhancing endothelial nitric oxide bioavailability (Karam et al., 2017). Although the present anti-atherothrombotic drugs available are safe and effective, the morbidity and mortality caused by atherothrombosis are still unacceptably high (Dias et al., 2018). Many of these drugs are mostly associated with several side effects (Khan et al., 2018). Statin, such as simvastatin, a lipid-lowering agent, also known as 3-hydroxy-3-methyl-glutaryl coenzyme A reductase inhibitor, is associated with several side effects, including fever, headache, gastric irritation, myositis, hyperuricaemia, rhabdomyolysis, myalgia, renal and hepatic dysfunctions (Hussain et al., 2019; Fattepur et al., 2018; Khan et al., 2018). Acetylsalicylic acid, including aspirin, are antiplatelet synthetic drugs widely prescribed to treat inflammation, headache, fever and atherothrombosis-related CVDs (Heart et al., 2019). Aspirin, in particular, has been reported to inhibit cyclooxygenase (COX), a potent enzyme that catalyses prostaglandin formation by blocking the synthesis of thromboxane A₂ (TXA₂), an essential mediator of blood clotting (Karam et al., 2019a). However, aspirin and other related antiplatelet drugs were reported to give recurrent thromboembolic vascular events (aspirin intolerance), including dizziness, nausea, abdominal pain or patients may suffer from increased risk of bleeding (Heart et al., 2019; Karam et al., 2019b). Coumarin and its derivatives are class of vitamin K-dependent anticoagulants (VKAs). Warfarin is the most common anticoagulant agent currently in use. Coumarin and its derivatives are class of vitamin K-dependent anticoagulants (VKAs). Warfarin is the most common anticoagulant agent currently in use. It functions

to inhibit vitamin K epoxide reductase (VKOR), which is necessarily needed for the gamma-carboxylation of vitamin K-dependent factor, including factors II, VII, IX, X and protein C and S. Inhibition of vitamin K carboxylation triggers the decreased hepatic synthesis activity of clotting factors, leading to an anticoagulated state. Bleeding is the most common complication associated with warfarin therapy and is related to exponentially higher international normalised ratio (INR) values. The goal of the management is to reduce the INR back to a therapeutic safe level (Heart et al., 2019).

Existing reports have shown that plant extracts have analgesic, antioxidant, antibacterial, antihyperlipidemic, anti-inflammatory, and antitumor properties, while the documented evidence concerning the anti-atherothrombotic potentials of many plant extracts are still lacking (Kirichenko et al., 2020; Khan et al., 2018; Fattepur et al., 2018). Adequate clinical investigations and scientific validations are still lacking for the authentication and recommendation of the plant extracts, to be used in the treatment of hyperlipidaemia-related atherothrombotic CVDs (Balaji et al., 2016).

1.3 Study Justification

Treatment failure associated with synthetic-based therapies posed a challenge to the researchers to look for alternative medications. In this regard, the development of natural-based products to augment conventional synthetic drugs is essential. They are more effective in reducing the serum levels of TC, TG and LDL while elevating HDL serum levels and enhancing nitric oxide (NO) bioavailability (Newman & Cragg, 2016).

1.4 The objective of the Study

1.4.1 General Objective

To investigate the mechanisms of action of ZACAH extracts on cellular and molecular atherothrombosis-related markers in SD rats fed with a high cholesterol diet (HCD).

1.4.2 Specific Objectives

1. To identify the major phytochemicals present in ZACAH extracts by UHPLC-MS profiling and animal study
2. To determine the effects of ZACAH extracts on body and liver weights, food intake, lipid profile and hepatic histopathological changes in SD rats fed with HCD.
3. To examine the endothelial and platelet ultrastructural and histopathological changes in SD rats fed with HCD supplemented with ZACAH extracts.
4. To determine the mRNA expression profiling of lipid metabolism atherothrombosis-related genes in SD rats fed with HCD supplemented with ZACAH extracts.
5. To evaluate the levels of atherothrombosis-related proteins, the mRNA expression profiling of the endothelial prothrombotic, coagulation and fibrinolytic genes in SD rats fed with HCD supplemented with ZACAH extracts.
6. To correlate the activities of related proteins with the expression profile of atherothrombosis-related genes, and the genes expression pattern in SD rats fed with HCD supplemented with ZACAH extracts.

1.5 Research Hypothesis

ZACAH extracts have inhibitory effects on excess cholesterol, activated platelets and endothelial cells, abnormal coagulation and fibrinolytic system through modulation of the proteins and genes involved in lipid metabolism, endothelial prothrombotic, coagulation and fibrinolytic system.

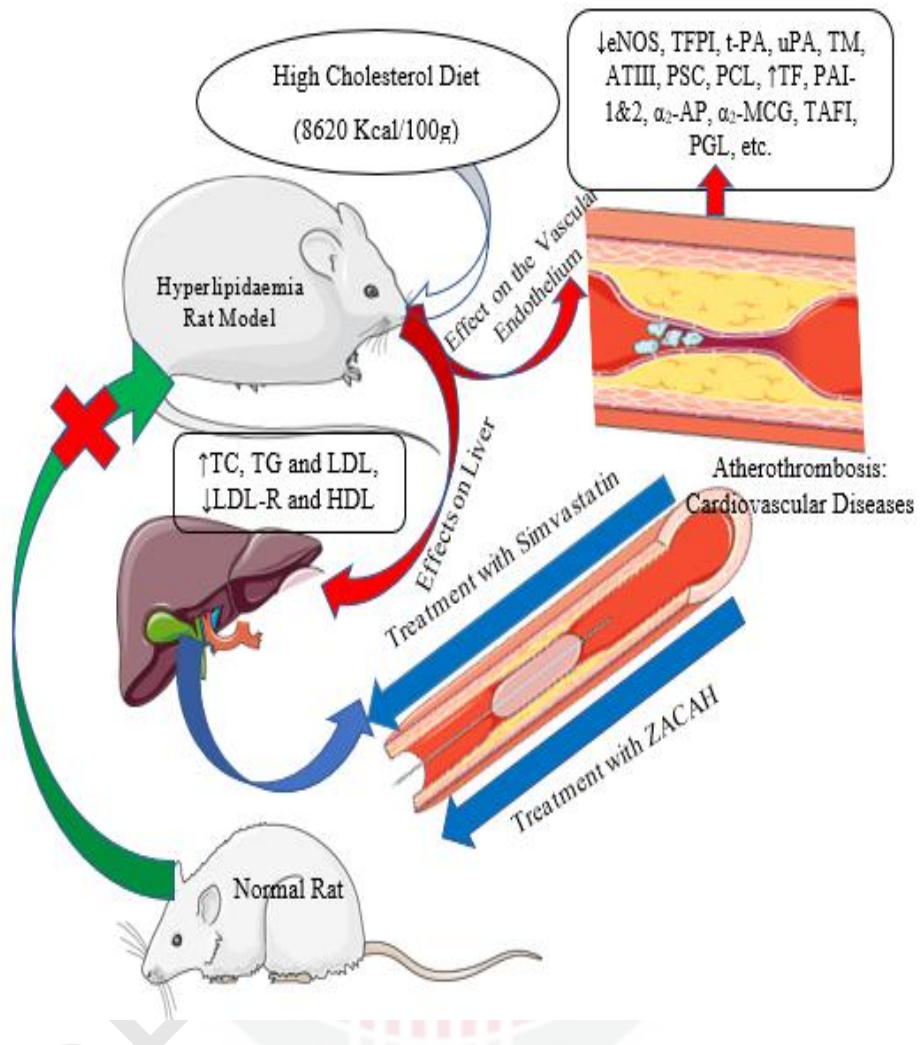


Figure 1.1 : Research Concept Map: Therapeutic potentials of ZACAH on atherothrombosis-related markers in SD rats fed with HCD

REFERENCES

- Adab, Z., Eghtesadi, S., Vafa, M. R., Heydari, I., Shojaii, A., Haqqani, H., ... Eghtesadi, M. (2019). Effect of turmeric on glycemic status, lipid profile, hs-CRP, and total antioxidant capacity in hyperlipidemic type 2 diabetes mellitus patients. *Phytother. Res.*, 33(4), 1173–1181. doi: 10.1002/ptr.6312.
- Abualhasan, M. N., Al- Masri, M. Y., Manasara, R., Yadak, L., & Abu-Hasan, N. S. (2020). Anti-Inflammatory and Anticoagulant Activities of Synthesized NSAID Prodrug Esters. *Scientifica*, 2020(Figure 3). <https://doi.org/10.1155/2020/9817502>.
- Afshin, A., Forouzanfar, M. H., Reitsma, M. B., Sur, P., Estep, K., Lee, A., Marczak, L., Mokdad, A. H., Moradi-Lakeh, M., Naghavi, M., Salama, J. S., Vos, T., Abate, K. H., Abbafati, C., Ahmed, M. B., Al-Aly, Z., Alkerwi, A., Al-Raddadi, R., Amare, A. T., ... Murray, C. J. L. (2017). Health effects of overweight and obesity in 195 countries over 25 years. *New England Journal of Medicine*, 377(1), 13–27. <https://doi.org/10.1056/NEJMoa1614362>.
- Ahmed, S., Shah, P., & Ahmed, O. (2021). Biochemistry, Lipids. [Updated 2021 May 9]. In: Stat Pearls [Internet]. Treasure Island (FL): Stat Pearls Publishing; 2021
- Alam, P., Alam, A., Anwer, M. K., & Alqasoumi, S. I. (2014). Quantitative estimation of hesperidin by HPTLC in different varieties of citrus peels. *Asian Pacific Journal of Tropical Biomedicine*, 4(4), 262–266. <https://doi.org/10.12980/APJTB.4.2014C1007>.
- Alhawiti, N. M. (2018). Antiplatelets and profibrinolytic activity of Citrullus colocynthis in control and high-fat diet-induced obese rats: mechanisms of action. *Arch Physiol Biochem.*, 124(2), 156-166. doi: 10.1080/13813455.2017.1369999. Epub.
- Al-Mashhadi, R. H., Sorensen, C. B., Kragh, P. M., Christoffersen, C., Mortensen, M. B., Tolbod, L. P., ... Bentzon, J. F. (2013). Familial hypercholesterolemia and atherosclerosis in cloned minipigs created by DNA transposition of a human PCSK9 gain-of-function mutant. *Sci Transl Med*, 5, 166ra1. doi:10.1126/scitranslmed.3004853.
- Alsharari, S. D., Al-rejaie, S. S., Abuhashish, H. M., Ahmed, M. M., & Hafez, M. M. (2016). *Rutin Attenuates Hepatotoxicity in High-Cholesterol-Diet-Fed Rats. 2016*.
- Anastasia, P. N., Eugene, A. K., Maria, Z., Sergey, S., Vladimir, V. S., Natalia, V. I., ... Anton, Y. (2020). *Diseases of the circulatory system in Disease Pathways*.
- Anna, N. B., & Avia, R.-D. (2015). Hypercholesterolemia Effect on Potassium Channels. *We Are IntechOpen, the World's Leading Publisher of Open Access Books Built by Scientists, for Scientists.*, <http://dx>.

- Argyaki, A., Markvart, M., Stavnsbjerg, C., Kragh, K. N., Ou, Y., Bjørndal, L., Bjarnsholt, T., Petersen, P. M., Muramoto, Y., Kimura, M., Nouda, S., Solution, I. N., Takayoshi, T., Takuya, M., Jun, S., Norimichi, N., Kenji, T., Hideki, H., Germicidal, U., ... Kang, D. (2018). (生物用) HHS Public Access. 2015 *IEEE Summer Topicals Meeting Series, SUM 2015*, 10(1), 1–13. <https://doi.org/10.1038/s41598-019-39414-9>.
- Arunkumar, E., Bhuvanewari, S., & Anuradha, C. V. (2012). An intervention study in obese mice with astaxanthin, a marine carotenoid-effects on insulin signaling and pro-inflammatory cytokines. *Food Funct.*, 3, 120–126.
- Asada, Y. (2020). *Pathophysiology of atherothrombosis: Mechanisms of thrombus formation on disrupted atherosclerotic plaques*. February, 309–322. <https://doi.org/10.1111/pin.12921>.
- Asdaq, S. M. B. (2015). Antioxidant and hypolipidemic potential of aged garlic extract and its constituent, s-allyl cysteine, in rats. *Evid Based Complement Alternat Med*, 2015, 328545–328552.
- Atashak, S., Peeri, M., Azarbayjani, M. A., & Stannard, S. R. (2014). Effects of ginger (*Zingiber officinale* Roscoe) supplementation and resistance training on some blood oxidative stress markers in obese men. *Journal of Exercise Science and Fitness*, 12(1), 26–30. <https://doi.org/10.1016/j.jesf.2014.01.002>.
- Athanasίου, L. S., Michalis, L. K., & Charac-, A. P. (2017). *Atherogenesis Learn more about Atherogenesis Introduction*.
- Ayman M. M., Hernández Bautista, R. J., Sandhu, M. A., & Hussein, O. E. (2019). Beneficial effects of citrus flavonoids on cardiovascular and metabolic health. *Oxidative Medicine and Cellular Longevity*, 2019. <https://doi.org/10.1155/2019/5484138>.
- Bäck, M. (2017). Omega-3 fatty acids in atherosclerosis and coronary artery disease. *Future Science OA*, 3(4). <https://doi.org/10.4155/fsoa-2017-0067>.
- Badimon, L., Padró, T., & Vilahur, G. (2012). Atherosclerosis, Platelets and Thrombosis in Acute Ischaemic Heart Disease. *Eur. Heart J. Acute Cardiovasc. Care*, 1, 60–74.
- Badimon, L. (2012). *European Heart Journal: Acute Cardiovascular*. May 2014. <https://doi.org/10.1177/2048872612441582>.
- Bagoly, Z., Koncz, Z., Hársfalvi, J., & Muszbek., L. (2012). Factor XIII, clot structure, thrombosis. *Thromb Res*, 129(3), 382–7.
- Bakogiannis, C., Sachse, M., Stamatelopoulos, K., & Stellos, K. (2019). Platelet-derived chemokines in inflammation and atherosclerosis. *Cytokine*, 122(154157).

- Balaji, M., Ganjaji, M. S., Kumar, G. E. N., Parim, B. N., Mopuri, R., & Dasari, S. (2016). A review on possible therapeutic targets to contain obesity: The role of phytochemicals. *Obesity Research and Clinical Practice*, 10(4), 363–380. <https://doi.org/10.1016/j.orcp.2015.12.004>.
- Baliga, M. S., Haniadka, R., Pereira, M. M., Thilakhand, K. R., Rao, S., & Arora, A. (2012). Radioprotective effects of *Zingiber officinale* Roscoe (ginger): past, present and future. *Food Funct*, 3, 714–723.
- Balkanci, Z. D., Pehlivanoglu, B., Bayrak, S., Karabulut, I., & Karaismailoglu, S. E. A. (2012). The effect of hypercholesterolemia on carbachol-induced contractions of the detrusor smooth muscle in rats: increased role of L-type Ca²⁺ channels. *Naunyn-Schmiedeberg Arch. Pharmacol.*, 385(11), 1141–8.
- Banu, K. S., & Cathrine, L. (2015). *General Techniques Involved in Phytochemical Analysis*. 2(4), 25–32.
- Barale, C., & Russo, I. (2020). *Influence of Cardiometabolic Risk Factors on Platelet Function*. CV, 1–27.
- Barale, C., Frascaroli, C., Senkeev, R., Cavalot, F., & Russo, I. (2018). Simvastatin Effects on Inflammation and Platelet Activation Markers in Hypercholesterolemia. *Biomed. Res. Int.*, 6508709.
- Barrachina, M. N., Sueiro, A. M., Izquierdo, I., Hermida-Nogueira, L., Guitián, E., Casanueva, F. F., ... Garcia, A. (2019). GPVI surface expression and signalling pathway activation are increased in platelets from obese patients: Elucidating potential anti-atherothrombotic targets in obesity. *Atherosclerosis* 2019, 281, 62–70., 281, 62–70.
- Bart, H. J., & Pilz, S. (2011). Industrial Scale Natural Products Extraction. *Industrial Natural Products Extraction*, 1–26.
- Benes, L. B., Bassi, N. S., & Davidson, M. H. (2016). Omega-3 carboxylic acids monotherapy and combination with statins in the management of dyslipidemia. *Vascular Health and Risk Management*, 12, 481–490. <https://doi.org/10.2147/VHRM.S58149>.
- Bentzon, J. F., Otsuka, F., Virmani, R., & Falk, E. (2014). *Mechanisms of Plaque Formation and Rupture*. 1852–1866. <https://doi.org/10.1161/CIRCRESAHA.114.302721>
- Berillis, P. (2021). *The Role of Collagen in the Aorta 's Structure The Role of Collagen in the Aorta's Structure*. May 2014. <https://doi.org/10.2174/1877382601306010001>.
- Bin-Jumah, M. N. (2018). Monolluma quadrangula protects against oxidative stress and modulates ldl receptor and fatty acid synthase gene expression in hypercholesterolemic rats. *Oxidative Medicine and Cellular Longevity*, 2018. <https://doi.org/10.1155/2018/3914384>.

- Bjorklund, M. M., Hollensen, A. K., Hagensen, M. K., Dagnaes-Hansen, F., Christoffersen, C., Mikkelsen, J. G., & Bentzon, J. F. (2014). Induction of atherosclerosis in mice and hamsters without germline genetic engineering. *Circ Res*, *114*(11), 1684–1689. doi:10.1161/circresaha.114.302937.
- Blokhin, I. O., Lentz, S. R., & City, I. (2015). *Mechanisms of thrombosis in obesity*. *20*(5), 437–444. <https://doi.org/10.1097/MOH.0b013e3283634443.Mechanisms>.
- Borissoff, J. I., Joosen, I. A., Versteyleen, M. O., Brill, A., Fuchs, T. A., Savchenko, A. S., ... Kietseleer, B. L. J. H. (2013). Elevated levels of circulating DNA and chromatin are independently associated with severe coronary atherosclerosis and a prothrombotic state. *Arterioscler Thromb Vasc Biol*, *33*, 2032–2040.
- Brahmanaidu, P. (2017). Reversal of endothelial dysfunction in aorta of streptozotocinnicotinamide-induced type-2 diabetic rats by SALLYcysteine. *Molecular and Cellular Biochemistry*, *432*(1–2), 25–32.
- Budak, N. H., Kumbul, D. D., Savas, C. M., Seydim, A. C., Kok, T. T., Ciris, M. I., & Guzel-Seydim, Z. B. (2011). Effects of apple cider vinegars produced with different techniques on blood lipids in high-cholesterol-fed rats. *J Agric Food Chem.*, *59*(12), 6638–6644.
- Calixto, J. B. (2019). The role of natural products in modern drug discovery. *Anais Da Academia Brasileira de Ciencias*, *91*, 1–7. <https://doi.org/10.1590/0001-3765201920190105>.
- Camera, M., Rossetti, L., Barbieri, S. S., Zanotti, I., Canciani, B., Trabattoni, D., ... Ferri, N. (2018). PCSK9 as a Positive Modulator of Platelet Activation. *J. Am. Coll. Cardiol.*, *71*, 952–854.
- Campbell, L. A., & Rosenfeld, M. E. (2015). Infection and atherosclerosis development. *Arch Med Res*, *46*, 339–350.
- Carlos, J.-S. (2015). Thrombolysis in Pulmonary Embolism. *Library of Congress Control Number: 2015943833. Springer Cham Heidelberg New York Dordrecht London. Springer International Publishing Switzerland. ISBN 978-3-319-19706-7 ISBN 978-3-319-19707-4 (EBook). DOI 10.1007/978-3-319-19707-4*.
- Carreiro, A. L., & Buhman, K. K. (2019). Absorption of Dietary Fat and Its Metabolism in Enterocytes. In *The Molecular Nutrition of Fats* (Vol. 36). Elsevier Inc. <https://doi.org/10.1016/B978-0-12-811297-7.00003-2>.
- Catapano, A. L., Graham, I. G., De Backer, G., Wiklund, O., Chapman, M. J., Drexel, H., ... Cooney, M.-T. (2016). “2016 ESC/EAS guidelines for the management of dyslipidaemias: the task force for the management of dyslipidaemias of the European Society of Cardiology (ESC) and European Atherosclerosis Society (EAS) developed with the special contribution of the Europe. *Atherosclerosis*, *253*, 291–344.

- Charo, I. F., & Taub, R. (2011). Anti-inflammatory therapeutics for the treatment of atherosclerosis. *Nature Reviews Drug Discovery*, *10*(5), 365–375.
- Chaudhary, H. R., & Brocks, D. R. (2013). The single dose poloxamer 407 model of hyperlipidemia; systemic effects on lipids assessed using pharmacokinetic methods, and its effects on adipokines. *Journal of Pharmacy and Pharmaceutical Sciences*, *16*(1), 65–73.
- Chen, T., Huang, J. B., Dai, J., Zhou, Q. R. J., & Zhou, G. (2018). PAI-1 is a novel component of the miR-17~92 signaling that regulates pulmonary artery smooth muscle cell phenotypes. *Am. J. Physiol. Lung Cell Mol. Physiol*, *315*(2), L149–L161.
- Chen, R., Qi, Q. L., Wang, M. T., & Li, Q. Y. (2016). Therapeutic potential of naringin: an overview. *Pharmaceutical Biology*, *54*(12), 3203–3210. <https://doi.org/10.1080/13880209.2016.1216131>.
- Chen, B., Lu, Y., Chen, Y., & Cheng, J. (2015). The role of Nrf2 in oxidative stress-induced endothelial injuries. *J Endocrinol Invest*, *225*(R83-99).
- Chernysh, I. N., Nagaswami, C., Kosolapova, S., Peshkova, A. D., Cuker, A., Cines, D. B., Cambor, C. L., Litvinov, R. I., & Weisel, J. W. (2020). The distinctive structure and composition of arterial and venous thrombi and pulmonary emboli. *Scientific Reports*, *10*(1), 1–12. <https://doi.org/10.1038/s41598-020-59526-x>.
- Chistiakov, D. A., Orekhov, A. N., & Bobryshev, Y. V. (2015). Endothelial barrier and its abnormalities in cardiovascular disease. *Frontiers in Physiology*, *6*(DEC), 1–11. <https://doi.org/10.3389/fphys.2015.00365>.
- Cianciosi, D., Forbes-Hernández, T. Y., Afrin, S., Gasparrini, M., Reboredo-Rodríguez, P., Manna, P. P., ... Battino, M. (2018). Phenolic compounds in honey and their associated health benefits: a review. *Molecules*, *23*, pii:E2322. doi: 10.3390/molecules 23092322.
- Clay, F. S., Goldberg, A. C., & Goldberg, I. J. (2016). Disorders of Lipid Metabolism 2016. *Textbook of Endocrinology (Thirteenth Edition)*.
- Crea, F., & Libby, P. (2017). Acute coronary syndromes: the way forward from mechanisms to precision treatment. *Circulation*, *136*, 1155–1166. doi: 10.1161/CIRCULATIONAHA.117.029870.
- Cristina, M., Sena, C. M., Fernanda, C., & Raquel, M. S. (2018). Endothelial Dysfunction in Type 2 Diabetes: Targeting Inflammation. *We Are IntechOpen , the World ' s Leading Publisher of Open Access Books Built by Scientists , for Scientists TOP 1 % 2018*. <Http://Dx.Doi.Org/10.5772/Intechopen.76994>.
- Dalibalta, S., Majdalawieh, A. F., & Manjikian, H. (2020). Health benefits of sesamin on cardiovascular disease and its associated risk factors. *Saudi Pharmaceutical Journal*, *28*(10), 1276–1289. <https://doi.org/10.1016/j.jsps.2020.08.018>.

- Danielli, M., Marrone, J., Capiglioni, A. M., & Marinelli, R. A. (2019). Mitochondrial aquaporin-8 is involved in SREBP-controlled hepatocyte cholesterol biosynthesis. *Free Radic. Biol. Med.*, *131*, 370–375.
- Danish, I., Salman, K., Mohd, S. K., Saheem, A., Sarfaraj, H. M., & Mohd, A. (2015). Bioactivity guided fractionation and hypolipidemic property of a novel HMG-CoA reductase inhibitor from *Ficus virens* Ait. *Lipids Health Dis*, 14–15.
- Deng, Z., Shan, W., Wang, S., Hu, M., & Chen, Y. (2017). Effects of astaxanthin on blood coagulation, fibrinolysis and platelet aggregation in hyperlipidemic rats. *Pharmaceutical Biology*, *0(0)*, 000. <https://doi.org/10.1080/13880209.2016.1261905>.
- Denise, R. F. (2017). Lippincott Illustrated Review: Biochemistry. 7th Edition. Wolters Kluwer., 1014963444, 173–219.
- Dettlaff-Pokora, A., Sucaszys-Szulc, E., & Sledzinski, T. (2019). Up-regulation of PCSK9 gene expression and diminished level of LDL-receptor in rat liver as a potential cause of post-lipectomy hypercholesterolemia. *Mol. Cell. Biochem.*, *455(1–2)*, 207–217.
- Dhananjayan, R., Koundinya, K. S. S., Malati, T., & Kutala, V. K. (2016). Endothelial Dysfunction in Type 2 Diabetes Mellitus. *Indian Journal of Clinical Biochemistry*, *31(4)*, 372–379. <https://doi.org/10.1007/s12291-015-0516-y>.
- Dias, S., Paredes, S., & Ribeiro, L. (2018). Review Article Drugs Involved in Dyslipidemia and Obesity Treatment : Focus on Adipose Tissue. 2018.
- Digennaro, F., & Dominici, F. P. (2012). Nebivolol : impact on cardiac and endothelial function and clinical utility. *March*. <https://doi.org/10.2147/VHRM.S20669>.
- Doddapattar, P., Dhanesha, N., Chorawala, M. R., Tinsman, C., Jain, M., Nayak, M. K., ... Chauchan, A. K. (2018). Endothelial cell-derived von Willebrand factor, but not platelet-derived, promotes atherosclerosis in apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol.*, *38*, 520–528. doi: 10.1161/ATVBAHA.117.309918.
- Driest, K. D., Sturm, M. S., O'Brien, S. H., Spencer, C. H., Stanek, J. R., & Ardion, S. P. (2016). Factors associated with thrombosis in pediatric patients with systemic lupus erythematosus. *Lupus*, *25*, 749–753.
- Earnest, O. E., & Daniel, L. A. (2014). Guidelines on dosage calculation and stock solution preparation in experimental animals' studies. *Journal of Natural Sciences Research Wwww*, *4(18)*, 2225–2921.
- Ebert, J., Wilgenbus, P., Teiber, J. F., Jurk, K., Schwierczek, K., Dohrmann, M., ... Horke, S. (2017). Paraoxonase-2 regulates coagulation activation through endothelial tissue factor. *Blood.*, doi: 10.1182/blood-2017-09-807040. <http://www.blood.org>.

- Ebrahimi, F., Torbati, M., Mahmoudi, J., & Valizadeh, H. (2020). Medicinal plants as potential hemostatic agents. *Journal of Pharmacy and Pharmaceutical Sciences*, 23(1), 10–23. <https://doi.org/10.18433/jpps30446>.
- El-Zayat, M. M., Eraqi, M. M., Alrefai, H., El-Khateeb, A. Y., Ibrahim, M. A., Aljohani, H. M., ... Elshaer, M. M. (2021). The Antimicrobial, Antioxidant, and Anticancer Activity of Greenly Synthesized Selenium and Zinc Composite Nanoparticles Using Ephedra aphylla Extract. *Biomolecules*, 11(3), 470. <https://doi.org/10.3390/biom11030470>.
- Engelmann, B., & Massberg, S. (2013). Thrombosis as an intravascular effector of innate immunity. *Nat Rev Immunol*, 13, 34–45.
- Espley, R. V., Butts, C. A., Laing, W. A., Martell, S., Smith, H., McGhie, T. K., ... Hellens, R. P. (2014). Dietary flavonoids from modified apple reduce inflammation markers and modulate gut microbiota in mice. *J Nutr*, 144(2), 146–154.
- Estruel-Amades, S., Massot-Cladera, M., Perez-Cano, F. J., Franch, A., Castell, M., & Camps-Bossacoma, M. (2019). Hesperidin effects on gut microbiota and gut-associated lymphoid tissue in healthy rats. *Nutrients*, 11(E324).
- Ewelina, D., Michal, M., Margorzata, O., & Marek, J. D. (2018). Atherothrombosis as a Leading Cause of Acute Coronary Syndromes and Stroke: The Main Killers in Developed Countries. In *Intech*. <https://www.intechopen.com/books/advanced-biometric-technologies/liveness-detection-in-biometrics>.
- Fan, S., Zhang, C., Luo, T., Wang, J., Tang, Y., Chen, Z., & Yu, L. (2019). *Limonin: A Review of Its Pharmacology, Toxicity, and Pharmacokinetics*. Table 1, 1–22.
- Fan, J. G. (2013). Epidemiology of alcoholic and nonalcoholic fatty liver disease in China. *J Gastroenterol Hepatol*, 28(1), 11–17.
- Fang, P., Zhang, D., Cheng, Z., Yan, C., Jiang, X., Kruger, W. D., ... Wang, H. (2014). Hyperhomocysteinaemia potentiates hyperglycaemia-induced inflammatory monocyte differentiation and atherosclerosis. *Diabetes*, 14(8), 10–15.
- Fattepur, S., Nilugal, K. C., Rajendran, R., Asmani, F., & Yusuf, E. (2018). Anti-Hyperlipidemic Activity of Methanolic Extract of Boesenbergia Pandurata (Finger Root) in Experimental Induced Hypercholesterolemic Sprague Dawley Rats. *Asian Journal of Pharmaceutical and Clinical Research*, 11(15), 8. <https://doi.org/10.22159/ajpcr.2018.v11s3.29962>.
- FBMR. (2017). Foundation for Biomedical Research. [[Www.Fbresearch.Org](http://www.Fbresearch.Org)].

- Ference, B. A., Ginsberg, H. N., Graham, I., Ray, K. K., Packard, C. J., Bruckert, E., Hegele, R. A., Krauss, R. M., Raal, F. J., Schunkert, H., Watt, G. F., Borén, J., Fazio, S., Horton, J. D., Masana, L., Nicholls, S. J., Nordestgaard, B. G., Van De Sluis, B., Taskinen, M. R., ... Catapano, A. L. (2017). "Low-density lipoproteins cause atherosclerotic cardiovascular disease. 1. Evidence from genetic, epidemiologic, and clinical studies. A consensus statement from the European Atherosclerosis Society Consensus Panel." *European Heart Journal*, 38(32), 2459–2472.
- Fischer, A. H., Jacobson, K. A., Rose, J., & Zeller, R. (2008). Hematoxylin and eosin staining of tissue and cell sections. *CSH Protoc.*, *pdb.prot49*.
- Frezza, C., Venditti, A., De Vita, D., Sciubba, F., Tomai, P., Franceschin, M., ... Bianco, A. (2021). Phytochemical analysis and biological activities of the ethanolic extract of daphne sericea vahl flowering aerial parts collected in central italy. *Biomolecules*, 11(3), 1–17. <https://doi.org/10.3390/biom11030379>.
- Gan, L. (2015). *Molecular mechanisms of fatty liver in obesity*. 9(3), 275–287. <https://doi.org/10.1007/s11684-015-0410-2>
- Ganeshpurkar, A., & Saluja, A. K. (2017). The Pharmacological Potential of Rutin. *Saudi Pharmaceutical Journal*, 25(2), 149–164. <https://doi.org/10.1016/j.jsps.2016.04.025>.
- Ganguly, P., & Alam, S. F. (2015). Role of homocysteine in the development of cardiovascular disease. *Nutrition Journal*, 14(1), 1–10. <https://doi.org/10.1186/1475-2891-14-6>.
- Gholipour, S., Sewell, R. D. E., Lorigooini, Z., & Rafieian-Kopaei, M. (2018). Medicinal Plants and Atherosclerosis: A Review on Molecular Aspects. *Current Pharmaceutical Design*, 24(26), 3123–3131. <https://doi.org/10.2174/1381612824666180911121525>.
- Gilani, A. H., Mandukhail, S. U., Iqbal, J., Yasinzai, M., Aziz, N. K. A., & Ehman, N. (2010). Antispasmodic and vasodilator activities of Morinda citrifolia root extract are mediated through blockade of voltage dependent calcium channels. *BMC Complement Altern Med*, 10, 2.
- Goliasch, G., Dabas, P., Wang, V., & Fayad, Z. A. (2014). *Brain Imaging Changes Associated With Risk Factors for Cardiovascular and Cerebrovascular Disease in Asymptomatic Patients*. 7(10). <https://doi.org/10.1016/j.jcmg.2014.06.014>.
- Gremmel, T., Frelinger, A. L., & Michelson, A. D. (2016). Platelet physiology. *Seminars in Thrombosis and Hemostasis*, 42(3), 191–204. <https://doi.org/10.1055/s-0035-1564835>.
- Grover, S. P., & Mackman, N. (2020). Tissue factor in atherosclerosis and atherothrombosis. *Atherosclerosis*, 307(July), 80–86. <https://doi.org/10.1016/j.atherosclerosis.2020.06.003>.

- Haidan, Y., Qianqian, M., Li, Y., & Gguangchun, P. (2016). *The Traditional Medicine and Modern Medicine from Natural Products*. <https://doi.org/10.3390/molecules21050559>.
- Halim, S. S. A., Ahmad, Z., Shamsudin, L., Abdullah, M. N. H., & Rashid, S. A. (2018). An alternative organic diet in preventing hyperlipidemia in cholesterolfed Sprague-Dawley rats by *Arthrospira* (*Spirulina*) *platensis*. *Songklanakarin Journal of Science and Technology*, *40*(6), 1259–1264. <https://doi.org/10.14456/sjst-psu.2018.154>.
- Han, Q., Yeung, S. C., Ip, M. S. M., & Mak, J. C. W. (2018). Dysregulation of cardiac lipid parameters in high-fat high-cholesterol diet-induced rat model. *Lipids in Health and Disease*, *17*(1), 1–10. <https://doi.org/10.1186/s12944-018-0905-3>.
- Hansel, B., Roussel, R., Elbez, Y., Marre, M., Krempf, M., Ikeda, Y., ... Steg, P. G. (2015). Cardiovascular risk in relation to body mass index and use of evidence-based preventive medications in patients with or at risk of atherothrombosis. *European Heart Journal*, *36*(40), 2716–2728. <https://doi.org/10.1093/eurheartj/ehv347>.
- Hansson, G. K., Libby, P., & Tabas, I. (2015). Inflammation and Plaque Vulnerability. *J. Int. Med.*, *278*(483–493).
- Harron, D. W. G. (2013). Technical Requirements for Registration of Pharmaceuticals for Human Use: The ICH Process. *The Textbook of Pharmaceutical Medicine*, *1994*(November), 447–460. <https://doi.org/10.1002/9781118532331.ch23>.
- Hashimoto, H., Usui, G., Tsugeno, Y., Sugita, K., Amori, G., Morikawa, T., & Inamura, K. (2019). Cerebral thromboembolism after lobectomy for lung cancer: Pathological diagnosis and mechanism of thrombus formation. *Cancers*, *11*(4), 1–17. <https://doi.org/10.3390/cancers11040488>.
- He, Y., Li, Z., Wang, W., Sooranna, S. R., Shi, Y., Chen, Y., ... Xie, H. (2018). Chemical profiles and simultaneous quantification of *aurantii fructus* by use of hplc-q-tof-ms combined with gc-ms and hplc methods. *Molecules*, *23*(9), 1–18. <https://doi.org/10.3390/molecules23092189>.
- Heart, T., Benziger, C. P., Moran, A. E., & Roth, G. A. (2020). *AN OVERVIEW OF GLOBAL HEALTH IN 2013*. 386(C), 1990–2013.
- Heart, T., Falk, E., & Fuster, V. (2019). *Chapter 32: atherothrombosis: disease burden, activity, and vulnerability*. 1–19.
- Heijnen, H., & van der Sluijs, P. (2015). Platelet secretory behaviour: As diverse as the granules or not? *J. Thromb. Haemost.*, *13*, 2141–2151.
- Hernández-brenes, C., Rodríguez-sánchez, D. G., Pacheco, A., Villarreal-, R., Ramos-gonzález, M. R., Ramos-parra, P. A., ... García-, G. (2019). *Accepted Payment Methods 1. June*, 99920.

- Hinojosa-Azaola, A., Romero-Diaz, J., Vargas-Ruiz, A. G., Nuñez-Alvarez, C.A., Cicero-Casarrubias, A., Ocampo-Torres, M. C., & Sanchez-Guerrero, J. (2016). Venous and arterial thrombotic events in systemic Lupus Erythematosus. *J Rheumatol*, *43*, 576–586.
- Holy, E. W., Stämpfli, S. F., Akhmedov, A., Holm, N., Camici, G. G., Lüscher, T. F., & Tanner, C. F. (2010). Laminin receptor activation inhibits endothelial tissue factor expression. *J Mol Cell Cardiol.*, *48*, 1138–1145.
- Homayouni, F., Haidari, F., Hedayati, M., Zakerkish, M., & Ahmadi, K. (2018). Blood pressure lowering and anti-inflammatory effects of hesperidin in type 2 diabetes: a randomized double-blind controlled clinical trial. *Phytother Res*, *32*, 1073–1079.
- Huda, N., Nor, M., Othman, F., Rahayu, E., Tohit, M., & Noor, S. (2016). *Medicinal Herbs with Antiplatelet Properties Benefit in Coronary Atherothrombotic Diseases. 2016.*
- Huo, J., Ma, R., Chai, X., Liang, H.-J., Jiang, P., Zhu, X.-L., ... Su, B. (2019). Inhibiting a spinal cord signaling pathway protects against ischemia injury in rats. *J. Thorac. Cardiovasc. Surg.*, *157*(2), 494–503.
- Hurt-camejo, E. (2018). *ApoB-100 Lipoprotein Complex Formation with Intima Proteoglycans as a Cause of Atherosclerosis and Its Possible Ex Vivo Evaluation as a Disease Biomarker. Ldl.* <https://doi.org/10.3390/jcdd5030036>.
- Hussain, M. S., Jahan, N., Rashid, M. M., Hossain, M. S., Chen, U., & Rahman, N. (2019). Antihyperlipidemic screening and plasma uric acid reducing potential of Momordica charantia seeds on Swiss albino mice model. *Heliyon*, *5*(5), e01739. <https://doi.org/10.1016/j.heliyon.2019.e01739>.
- Hutter, R., Valdiviezo, C., Sauter, b. v., Savontaus, M., Chereshev, I., Carrick, F. E., Bauriedel, G., Luderitz, B., Fallon, J. T. & Fuster, V. (2004). Caspase-3 and tissue factor expression in lipid-rich plague macrophages evidence for apoptosis as link between inflammation and atherothrombosis. *Circulation*, *109*(16), 2001–2008.
- Huxley, R. R., Barzi, F., Woo, J., Giles, G., Lam, T. H., Rahimi, K., ... Woodward, M. (2014). A comparison of risk factors for mortality from heart failure in asian and non-asian populations: An overview of individual participant data from 32 prospective cohorts from the asi-pacific region. *BMC Cardiovascular Disorders*, *14*, 61-71. Doi: 10.1186/1471-2261-14-61.
- Igor, A., & Sobenin, I. V. (2015). Anti-atherosclerotic effects of garlic preparation in freeze injury model of atherosclerosis in cholesterol-fed rabbits Andrianova, K. Yu. Lakunin, Alexander Orekhov. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*, *23*(11), DOI: 10.1016/j.phymed.

- Institute for Public Health. National Health and Morbidity Survey 2015 (NHMS 2015). Vol. II: (2015). Non-Communicable Diseases, Risk Factors & Other Health Problems. Malaysia: Ministry of Health, Malaysia; 2015. *Institute for Public Health, 11*.
- Jia, Y. J., Liu, J., Guo, Y. L., Xu, R. X., Sun, J., & Li, J. J. (2013). Dyslipidemia in rat fed with high-fat diet is not associated with PCSK9-LDL-receptor pathway but ageing. *Journal of Geriatric Cardiology*, *10*(4), 361–368. <https://doi.org/10.3969/j.issn.1671-5411.2013.04.007>.
- Jin, D., Yi, X., Mei, X., Meng, Q., Gao, Y., Li, B., & Tu, Y. (2013). Anti-obesity and lipid lowering effects of theaflavins on high fat diet induced obese rats. *J Functional Foods*, *5*(3), 1142–1150.
- Kajal, A., Kishore, L., Kaur, N., Gollen, R., & Singh, R. (2016). Therapeutic agents for the management of atherosclerosis from herbal sources. *Beni-Suef University Journal of Basic and Applied Sciences*, *5*(2), 156–169. <https://doi.org/10.1016/j.bjbas.2016.02.004>.
- Kalaivani, A., Sathibabu, U. V. V., Brahmanaidu, P., Saravanan, G., Nivedha, P. R., Sushma, C. K., ... Vadivukkarasi, S. (2018). enzymes and inflammatory markers expressions in rats Reversal of high fat diet-induced obesity through modulating lipid metabolic enzymes and. *Archives of Physiology and Biochemistry*, *0*(0), 1–7. <https://doi.org/10.1080/13813455.2018.1452036>.
- Kamel, E. M., Mahmoud, A. M., Ahmed, S. A., & Lamsabhi, A. M. (2016). “A phytochemical and computational study on flavonoids isolated from *Trifolium resupinatum* L. and their novel hepatoprotective activity.” *Food & Function*, *7*(4), 2094–2106.
- Kang, Y. E., Kim, J. M., Joung, K. H., Lee, J. H., You, B. R., Choi, M. J., ... Kim, H. J. (2016). The roles of adipokines, proinflammatory cytokines, and adipose tissue macrophages in obesity-associated insulin resistance in modest obesity and early metabolic dysfunction. *PLoS ONE*, *11*(4), 1–14. <https://doi.org/10.1371/journal.pone.0154003>.
- Karagodin, V. P., Sobenin, I. A., & Orekhov, A. N. (2016). Antiatherosclerotic and Cardioprotective Effects of Time-Released Garlic Powder Pills. *Curr. Pharm., Des.* *22*, 196–213. doi: 10.2174/1381612822666151112153351.
- Karam, I., Ma, N., Liu, X. W., Li, J.-Y., & Yang, Y.-J. (2019a). Effect of Aspirin on Hyperlipidemia in Rats. *JSM Pharmacol Clin Toxicol*, *1*, 6.
- Karam, I., Ma, N., Liu, X., Li, J., & Yang, Y. (2019b). *Short Review on Hyperlipidemia Journal of Blood Transfusions and Short Review on Hyperlipidemia. August*.
- Karam, I., Ma, N., Yang, Y.-J., & Li, J.-Y. (2018). Induce Hyperlipidemia in Rats Using High Fat Diet Investigating Blood Lipid and Histopathology. *Journal of Hematology and Blood Disorders*, *4*(1), 1–5. <https://doi.org/10.15744/2455-7641.4.104>.

- Karam, I., Yang, Y.-J., & Li, J.-Y. (2017). *SM Gr up SM Atherosclerosis Hyperlipidemia Background and Progress*. September.
- Karimi, G., Sabran, M. R., Jamaluddin, R., Parvaneh, K., Mohtarrudin, N., Ahmad, Z., ... Khodavandi, A. (2015). The anti-obesity effects of *Lactobacillus casei* strain Shirota versus Orlistat on high fat diet-induced obese rats. *Food and Nutrition Research*, 59(1), 1–9. <https://doi.org/10.3402/fnr.v59.29273>.
- Katakami, N. (2018). Mechanism of Development of Atherosclerosis and Cardiovascular Disease in Diabetes Mellitus. *J. Atheroscler Thromb.*, 25(doi: 10.5551/jat.RV17014.), 27–39.
- Kenneth, R., & Feingold, M. D. (2021). *Introduction to Lipids and Lipoproteins STRUCTURE OF LIPOPROTEINS (2). 1*, 1–16.
- Khamis, A. A., Salama, A. F., Kenawy, M. E., & Mohamed, T. M. (2017). Regulation of hepatic hydroxy methyl glutarate - CoA reductase for controlling hypercholesterolemia in rats. *Biomed. Pharmacother.*, 95, 1242–1250.
- Khan, M. A., Hashim, M. J., Mustafa, H., Baniyas, M. Y., Al Suwaidi, S. K. B. M., AlKatheeri, R., ... Lootah, S. N. A. H. (2020). Global Epidemiology of Ischemic Heart Disease: Results from the Global Burden of Disease Study. *Cureus*, 12(7). <https://doi.org/10.7759/cureus.9349>.
- Khan, S. J., Afroz, S., & Khan, R. A. (2018). Antihyperlipidemic and anti-hyperglycemic effects of *Cymbopogon jwarancusa* in high-fat high-sugar Diet model. *Pakistan Journal of Pharmaceutical Sciences*, 31(4), 1341–1345.
- Khari, N., Aisha, A. F. A., & Ismail, Z. (2014). Reverse phase high performance liquid chromatography for the quantification of eurycomanone in *Eurycoma longifolia* jack (*Simaroubaceae*) extracts and their commercial products. *Tropical Journal of Pharmaceutical Research*, 13(5), 801–807. <https://doi.org/10.4314/tjpr.v13i5.22>.
- Khosravani, M., Azarbayjani, M. A., Abolmaesoomi, M., Yusof, A., Abidin, N. Z., Rahimi E., & Dehghan, F. (2016). Ginger extract and aerobic training reduces lipid profile in high-fat fed diet rats. *Eur. Rev. Med. Pharmaco.*, 20, 1617–1622.
- Kim, J., Wie, M. B., Ahn, M., Tanaka, A., Matsuda, H., & Shin, T. (2019a). Benefits of hesperidin in central nervous system disorders: A review. *Anatomy and Cell Biology*, 52(4), 369–377. <https://doi.org/10.5115/acb.19.119>.
- Kim, J. Y., & Shim, S. H. (2019b). Medicinal herbs effective against atherosclerosis: Classification according to mechanism of action. *Biomolecules and Therapeutics*, 27(3), 254–264. <https://doi.org/10.4062/biomolther.2018.231>.
- King, R. J., & Ajjan, R. A. (2017). *Vascular risk in obesity : Facts , misconceptions and the unknown*. 2–13. <https://doi.org/10.1177/1479164116675488>.

- Kirichenko, T. V., Sukhorukov, V. N., Markin, A. M., Nikiforov, N. G., Liu, P. Y., Sobenin, I. A., ... Aliev, G. (2020). Medicinal Plants as a Potential and Successful Treatment Option in the Context of Atherosclerosis. *Frontiers in Pharmacology*, 11(April), 1–15. <https://doi.org/10.3389/fphar.2020.00403>.
- Knaapen, M., Kootte, R. S., Zoetendal, E. G., Vos, W. M., De Levi, M., Stroes, E. S., & Nieuwdorp, M. (2013). Obesity , non-alcoholic fatty liver disease , and atherothrombosis : a role for the intestinal microbiota? *European Society of Clinical Infectious Diseases*, 19(4), 331–337. <https://doi.org/10.1111/1469-0691.12170>.
- Koniari, I., Mavrilas, D., Apostolakis, E., Papadimitriou, E., Papadaki, H., Papalois, A., ... Alexopoulos, D. (2016). Inhibition of atherosclerosis progression, intimal hyperplasia, and oxidative stress by simvastatin and ivabradine may reduce thoracic aorta's stiffness in hypercholesterolemic rabbits. *Journal of Cardiovascular Pharmacology and Therapeutics*, 21(4), 412–422. <https://doi.org/10.1177/1074248415617289>.
- Korou, L., Pergialiotis, V., Misiakos, E. P., & Rizos, I. (2016). *Pre-treatment with simvastatin prevents the induction of diet-induced atherosclerosis in a rabbit model. October*. <https://doi.org/10.3892/br.2016.780>.
- Koupenova, M., Kehrel, B. E., Corkrey, H. A., & Freedman, J. E. (2017). Thrombosis and platelets: An update. *European Heart Journal*, 38(11), 785–791. <https://doi.org/10.1093/eurheartj/ehw550>
- Koupenova, M., Vitseva, O., MacKay, C. R., Beaulieu, L. M., Benjamin, E. J., Mick, E., ... Freedman, J. E. (2014). Platelet-TLR7 mediates host survival and platelet count during viral infection in the absence of platelet-dependent thrombosis. *Blood*, 124, 791–802.
- Kozera, B., & Repacz, M. (2013). (2013). Reference genes in real-time PCR. *Journal of Applied Genetics*, 54(4), 391–406.
- Kumar, R., Akhtar, F., & Rizvi, S. I. (2020). Hesperidin attenuates altered redox homeostasis in an experimental hyperlipidaemic model of rat. *Clinical and Experimental Pharmacology and Physiology*, 47(4), 571–582. <https://doi.org/10.1111/1440-1681.13221>.
- Lee, H., Woo, M., Kim, M., Noh, J. S., & Song, Y. O. (2018). *Antioxidative and Cholesterol-Lowering Effects of Lemon Essential Oil in Hypercholesterolemia-Induced Rabbits*. 23(August 2017), 8–14.
- Lehmann, M., Schoeman, R. M., Krohl, P. J., Wallbank, A. M., Samaniuk, J. R., Jandrot-Perrus, M., & Neeves, K. B. (2018). Platelets drive thrombus propagation in a hematocrit and glycoprotein VI-dependent manner in an in vitro venous thrombosis model. *Arterioscler Thromb Vasc Biol.*, 38, 1052–1062. doi: 10.1161/ATVBAHA.118.310731.

- Lentz, S. R. (2019). *Review Article Thrombosis in the setting of obesity or inflammatory bowel disease*. *128*(20), 180–187. <https://doi.org/10.1182/blood-2016-05-716720>.
- Leong, X. F., Ng, C. Y., & Jaarin, K. (2015). Animal Models in Cardiovascular Research: Hypertension and Atherosclerosis. *BioMed Research International*, *2015*(ii). <https://doi.org/10.1155/2015/528757>.
- Levi, M., & van der Poll, T. (2017). Coagulation and sepsis. *Thrombosis Research*, *149*, 38–44. <https://doi.org/10.1016/j.thromres.2016.11.007>.
- Li, C., & Schluesener, H. (2017). Health-promoting effects of the citrus flavanone hesperidin. *Critical Reviews in Food Science and Nutrition*, *57*(3), 613–631, DOI: 10.1080/10408398.2014.906382.
- Li, W., Wu, N., Shu, W., Jia, D., & Jia, P. (2015). Pharmacological preconditioning and postconditioning with nicorandil attenuates ischemia/reperfusion-induced myocardial necrosis and apoptosis in hypercholesterolemic rats. *Exp Ther Med*, *10*, 2197–205.
- Liang, N., Sang, Y., Liu, W., Yu, W., & Wang, X. (2018). *Anti-Inflammatory Effects of Gingerol on Lipopolysaccharide-Stimulated RAW 264. 7 Cells by Inhibiting NF- κ B Signaling Pathway*. *41*(3). <https://doi.org/10.1007/s10753-018-0737-3>.
- Libby, P., & Hansson, G. K. (2015). Inflammation and immunity in diseases of the arterial tree: players and layers. *Circ Res*, *116*, 307–311.
- Lim, J. U., Lee, J. H., Kim, J. S., Hwang, Y. Il., Kim, T. H., Lim, S. Y., ... Rhee, C. K. (2017). Comparison of World Health Organization and Asia-Pacific body mass index classifications in COPD patients. *International Journal of COPD*, *12*, 2465–2475. <https://doi.org/10.2147/COPD.S141295>.
- Lim, S. S., VoS, T., Flaxman, A. D., Danaei, G., Shibuya, K., Adair-Rohani, H., AlMazroa, M. A., Amann, M., Anderson, R., Andrews, K. G., Aryee, M., Atkinson, C., Bacchus, L. J., Bahalim, A. N., Balakrishnan, K., Barker-Collo, S., Baxter, A., Bell, M. L., Blore, J., ... Ezzati, M. (2012). A comparative risk assessment of burden of disease and injury attributable to 67 risk factors and risk factor clusters in 21 regions, 1990-2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet*, *380*, 2224–2260.
- Ling, Y., Shi, Z., Yang, X., Cai, Z., Wang, L., Wu, X., ... Jiang, J. (2020). Hypolipidemic effect of pure total flavonoids from peel of Citrus (PTFC) on hamsters of hyperlipidemia and its potential mechanism. *Experimental Gerontology*, *130*, 110786. <https://doi.org/10.1016/j.exger.2019.110786>.
- Liu, B., Wu, Y., Wang, Y., Cheng, Y., Yao, L., Liu, Y., ... Shen, F. (2018). NF-kappaB p65 Knock-down inhibits TF, PAI-1 and promotes activated protein C production in lipopolysaccharide-stimulated alveolar epithelial cells type II. *Exp. Lung Res*, *44*(4–5), 241–251.

- Liu, T., Zhang, L., Joo, D., & Sun, S. C. (2017). NF- κ B signaling in inflammation. *Signal Transduction and Targeted Therapy*, 2(April). <https://doi.org/10.1038/sigtrans.2017.23>.
- Liu, Y., Wu, J., Shi, Q., Guo, H., Ying, H., & Xu, N. (2014). Primary genetic investigation of a hyperlipidemia model: Molecular characteristics and variants of the apolipoprotein e gene in Mongolian gerbil. *BioMed Research International*, 2014. <https://doi.org/10.1155/2014/410480>.
- Longstaff, C., & Kolev, K. (2015). Basic mechanisms and regulation of fibrinolysis. *J. Thromb. Haemost*, 13(1), S98–S105.
- Lunagariya, N. A., Patel, N. K., Jagtap, S. C., & Bhutani, K. (2014). Inhibitors of pancreatic lipase: state of the art and clinical perspectives. *EXCLI J*, 13, 897–921.
- Mackman, N. (2012). New insights into the mechanisms of venous thrombosis. *J Clin Invest*, 122, 2331–2336.
- Mahmoodi, B. K., Veeger, N. J., Middeldorp, S., Lijfering, W. M., Brouwer, J. L., Berg, J. T., ... Meijer, K. (2016). Interaction of hereditary thrombophilia and traditional cardiovascular risk factors on the risk of arterial thromboembolism: pooled analysis of four family cohort studies. *Circ Cardiovasc Genet*, 9, 79–85.
- Mahmoud, A. M., Hernández, B. R. J., Sandhu, M. A., & Hussein, O. E. (2019). Beneficial effects of citrus flavonoids on cardiovascular and metabolic health. *Oxidative Medicine and Cellular Longevity*, 2019(March). <https://doi.org/10.1155/2019/5484138>.
- Mahmoud, A. M., Mohammed, H. M., Khadrawy, S. M., & Galaly, S. R. (2017). “Hesperidin protects against chemically induced hepatocarcinogenesis via modulation of Nrf2/ARE/HO-1, PPAR γ and TGF- β 1/Smad3 signaling, and amelioration of oxidative stress and inflammation.” *Chemico-Biological Interactions*, 277, 146–158.
- Mahmoud, A. M. (2014). “Hesperidin protects against cyclophosphamide-induced hepatotoxicity by upregulation of PPAR γ and abrogation of oxidative stress and inflammation.” *Canadian Journal of Physiology and Pharmacology*, 92(9), 717–724.
- Maino, A., Siegerink, B., Algra, A., Martinelli, I., Peyvandi, F., & Rosendaal, F. (2016). Pregnancy loss and risk of ischaemic stroke and myocardial infarction. *Br J Haematol*, 174, 302–309.
- Malakul, W., Pengnet, S., Kumchoom, C. C., & Tunsophon, S. S. (2018). “Naringin ameliorates endothelial dysfunction in fructose-fed rats. *Experimental and Therapeutic Medicine*, 15(3), 3140–3146.

- Mamatha, A. (2011). Quantitative HPLC analysis of andrographolide in *Andrographis paniculata* obtained from different geographical sources (India). *International Journal of Pharmacy and Pharmaceutical Sciences*, 3(2), 42–44.
- Mancini, G. B. J., Baker, S., Bergeron, J., Fitchett, D., Frohlich, J., Genest, J., ... Yashakkor, A. Y. (2016). “Diagnosis, prevention, and management of statin adverse effects and intolerance: canadian consensus working group update.” *Canadian Journal of Cardiology*, 32(7), 535–565.
- Mangold, A., Alias, S., Scherz, T., Hofbauer, T., Jakowitsch, J., Panzenbock, A., ... Lang, I. M. (2015). Coronary neutrophil extracellular trap burden and deoxyribonuclease activity in ST-elevation acute coronary syndrome are predictors of ST-segment resolution and infarct size. *Circ Res*, 116, 1182–1192.
- Manoj, W., & Jolanta, U. W. (2018). *Lipid Disorders in Obesity in Practical Guide to Obesity Medicine*.
- Mao, Q. Q., Xu, X. Y., Cao, S. Y., Gan, R. Y., Corke, H., Beta, T., & Li, H. B. (2019). Bioactive compounds and bioactivities of ginger (*zingiber officinale roscoe*). *Foods*, 8(6), 1–21. <https://doi.org/10.3390/foods8060185>.
- Margetic, S. (2012). Review Inflammation and haemostasis. *Inflammation*, 22(1), 49–62.
- Marques, C., Meireles, M., Norberto, S., Leite, J., Freitas, J., Pestana, D., ... Calhau, C. (2016). High-fat diet-induced obesity Rat model : a comparison between Wistar and Sprague-Dawley Rat. *Adipocyte*, 5(1), 11–21. <https://doi.org/10.1080/21623945.2015.1061723>.
- Martinod, K., & Wagner, D. (2014). Thrombosis: tangled up in NETs. *Blood*, 123, 2768–2776.
- Masarone, M., Federico, A., Abenavoli, L., Loguercio, C., & Persico, M. (2014). Non alcoholic fatty liver: epidemiology and natural history. *Rev Recent Clin Trials*, 9(3), 126–133.
- Mashhadi, N. S., Ghiasvand, R., Askari, G., Hariri, M., Darvishi, L., & Mofid, M. R. (2013). Anti-oxidative and anti-inflammatory effects of ginger in health and physical activity: review of current evidence. *Int J Prev Med*, 4(1), S36- S42.
- McFadyen, J. D., Schaff, M., & Peter, K. (2018). Current and future antiplatelet therapies: emphasis on preserving haemostasis. *Nat Rev Cardiol*, 15, 181–191.
- Meadows, T. A., & Bhatt, D. L. (2007). Clinical aspects of platelet inhibitors and thrombus formation. *Circulation Research*, 100(9), 1261–1275. <https://doi.org/10.1161/01.RES.0000264509.36234.51>.

- Mechelinck, M., Kupp, C., Krüger, J. C., Habigt, M. A., Helmedag, M. J., Tolba, R. H., Hein, M. (2019). Oxygen inhalation improves postoperative survival in ketamine-xylazine anaesthetised rats: An observational study. *PLoS ONE*, *14*(12), 1–14. <https://doi.org/10.1371/journal.pone.0226430>.
- Meng, Q., Wang, W., Yu, X., Li, W., Kong, L., Qian, A., ... Li, X. (2015). Upregulation of microRNA-126 contributes to endothelial progenitor cell function in deep vein thrombosis via its target PIK3R2. *J Cell Biochem.*, *116*, 1613– 1623. doi: 10.1002/jcb.25115.
- Michael, S., Kiran, V., Melissa, C., Robert, R., Xuming, D., George, A. S., & Michael, Y. (2017). Ratio of systolic blood pressure to left ventricular end- diastolic pressure at the time of primary percutaneous coronary intervention predicts in-hospital mortality in patients with ST- elevation myocardial infarction. *Catheterization and Cardiovascular Interventions.*, *90*(3), 389–395.
- Ministry of Health Malaysia. (2017). CPG 5th Edition: Management of Dyslipidaemia 2017. *MOH Malaysia, 5th Editio*(July), 1–107. <https://doi.org/10.1136/hrt.2003.021287>.
- Mittendorfer, B., Yoshino, M., Patterson, B. W., & Klein, S. (2016). VLDL triglyceride kinetics in lean, overweight, and obese men and women. *J. Clin. Endocrinol. Metab.*, *101*, 4151–4160.
- Mopuri, R., Ganjaji, M., Banavathy, K. S., Parim, B. N., & Meriga, B. (2015). *Evaluation of anti-obesity activities of ethanolic extract of Terminalia paniculata bark on high fat diet-induced obese rats.* 1–11. <https://doi.org/10.1186/s12906-015-0598-3>.
- Moran, A. E., Forouzanfar, M. H., Roth, G. A., Mensah, G. A., Ezzati, M., Murray, C. J. L., & Naghavi, M. (2014). Temporal trends in ischemic heart disease mortality in 21 world regions, 1980 to 2010: the Global Burden of Disease 2010 study. *Circulation.*, *129*, 1483–1492. [10.1161/CIRCULATIONAHA.113.004042](https://doi.org/10.1161/CIRCULATIONAHA.113.004042).
- Mozaffarian, D., Benjamin, E. J., Go, S. A., Arnett, D. K., Blaha, M. J., Cushman, M., Das, S. R., de Ferranti, S., Després, J., Fullerton, H. J., Howard, V. J., Huffman, M. D., Isasi, C. R., Jiménez, M. C., Judd, S. E., Kissela, B. M., Lichtman, J. H., Lisabeth, L. D., Liu, S., ... Turner, M. B. (2016). Heart disease and stroke statistics—2016 update: a report from the American Heart Association. *Circulation*, *133*(4), e36–360.
- Muhammad, S. A., & Abubakar, S. M. (2016). *Qualitative and Quantitative Determination of Phytochemicals In Aqueous Extract of Chrysophyllum albidum Seed Kernel.* *13*(June), 1201–1206.
- Munshi, R. P., Joshi, S. G., & Rane, B. N. (2019). *Development of an experimental diet model in rats to study hyperlipidemia and insulin resistance , markers for coronary heart disease.* *46*(3), 270–276. <https://doi.org/10.4103/0253-7613.132156>.

- Mushtaq, R., Mushtaq, R., & Khan, T. Z. (2011). Effects of natural honey on lipid profile and body weight in normal weight and obese adults: a randomized clinical trial. *Pak J Zool*, 43, 161–9.
- Mussbacher, M., Salzmann, M., Brostjan, C., Hoesel, B., Schoergenhofer, C., Datler, H., ... Schmid, J. A. (2019). Cell Type-Specific Roles of NF-κB Linking Inflammation and Thrombosis. *Front. Immunol.*, 10.
- myAssay. (2012). “Four Parameter Logistic Curve” online data analysis tool, MyAssays Ltd., 25th October. <http://www.myassays.com/four-parameter-logistic-cu.>
- Nagasawa, S-Y., Okamura, T., Iso, H., Tamakoshi, A., Yamada, M., Watanabe, M., ... Ueshima, H. (2012). “Relation between serum total cholesterol level and cardiovascular disease stratified by sex and age group: a pooled analysis of 65 594 individuals from 10 cohort studies in Japan.” *Journal of the American Heart Association*, 1(5), e001974.
- Navarese, E. P., Kolodziejczak, M., Winter, M.-P., Alimohammadi, A., Lang, I. M., Buffon, A., ... Siller-Matula, J. M. (2017). Association of PCSK9 with platelet reactivity in patients with acute coronary syndrome treated with prasugrel or ticagrelor: The PCSK9-REACT study. *Int. J. Cardiol.*, 227, 644–649.
- Nelson, R. H. (2013). Hyperlipidaemia as a risk factor for cardiovascular disease. *Primary Care*, 40(1), 195-211. doi:10.1016/j.pop.2012.11.003.
- Neumüller, J., Meisslitzer-Ruppitsch, C., Ellinger, A., Pavelka, M., Jungbauer, C., Renz, R., ... Wagner, T. (2013). Monitoring of platelet activation in platelet concentrates using transmission electron microscopy. *Transfusion Medicine and Hemotherapy*, 40(2), 101–107. <https://doi.org/10.1159/000350034>.
- Newman, D. J., & Cragg, G. M. (2016). Natural Products as Sources of New Drugs from 1981 to 2014. *Journal of Natural Products*, 79(3), 629–661. <https://doi.org/10.1021/acs.jnatprod.5b01055>.
- Ogiwara, K., Nogami, K., Matsumoto, T., & Shima, M. (2014). Tissue factor pathway inhibitor in activated prothrombin complex concentrates (aPCC) moderates the effectiveness of therapy in some severe hemophilia A patients with inhibitor. *Int. J. Hematol*, 99(5), 577–587.
- Olie, R. H., van der Meijden, P. E. J., & Ten Cate, H. (2018). The coagulation system in atherothrombosis: Implications for new therapeutic strategies. *Research and Practice in Thrombosis and Haemostasis*, 2(2), 188–198. <https://doi.org/10.1002/rth2.12080>.
- Olivieri, O., Martinelli, N., Baroni, M., Branchini, A., Girelli, D., Friso, S., ... Bernard, F. (2013). Factor II activity is similarly increased in patients with elevated apolipoprotein CIII and in carriers of the factor II 20210A allele. *J Am Heart Assoc.*, 2:e000440. doi: 10.1161/JAHA.113.000440.

- Omar, S. H. (2017). *Garlic and Cardiovascular Diseases* (Issue December). <https://doi.org/10.1007/978-3-642-22144-6>.
- Oppi, S., Lüscher, T. F., & Stein, S. (2019). Mouse Models for Atherosclerosis Research—Which Is My Line? *Frontiers in Cardiovascular Medicine*, 6(April), 1–8. <https://doi.org/10.3389/fcvm.2019.00046>.
- Orekhov, A. N., & Ivanova, E. A. (2016). Cellular models of atherosclerosis and their implication for testing natural substances with anti-atherosclerotic potential. *Phytomedicine*, 23, 1190–1197.
- Othman, Z. A., Wan, G. W. S., Noordin, L., Omar, N., Mohd, Y. N. A., & Mohamed, M. (2019). Protective effects of orlistat on lipid profile, cardiac oxidative stress biomarkers and histology in high-fat diet-induced obese rats. *International Medical Journal Malaysia*, 18(2), 23–28.
- Ouchi, N., Parker, J. L., Lugus, J. J., & Walsh, K. (2011). Adipokines in inflammation and metabolic disease. *Nat Rev Immunol.*, 11, 85–97.
- Padala, S., & Thompson, P. D. (2013). “Statins as a possible cause of inflammatory and necrotizing myopathies.” *Atherosclerosis*, 222(1), 15–21.
- Papapanagiotou, A., Siasos, G., Kassi, E., Gargalionis, A. N., & Papavassiliou, A. G. (2015). Novel inflammatory markers in hyperlipidaemia. *Clin. Implic. Curr. Med. Chem*, 22, 2727–2743.
- Pareja, J., & Restrepo, J. (2016). Métodos diagnósticos en hipertensión portal. *Rev Col Gastroenterol.*, 31, 135–145.
- Pari, L., Karthikeyan, A., & Karthika, P. (2015). *Protective effects of hesperidin on oxidative stress , dyslipidaemia and histological changes in iron-induced hepatic and renal toxicity in rats.* 2, 46–55.
- Park, S., Kim, J. K., Oh, C. J., Choi, S. H., Jeon, J. H., & Lee, I. K. (2015). Scoparone interferes with STAT3-induced proliferation of vascular smooth muscle cells. *Experimental and Molecular Medicine*, 47(November 2014). <https://doi.org/10.1038/emm.2014.113>.
- Park, T., & Kim, Y. (2011). Phytochemicals as potential agents for prevention and treatment of obesity and metabolic diseases. In: *Anti-obesity drug discovery and development. Dubai: Bentham*, 150–185.
- Pastori, D., Nocella, C., Farcomeni, A., Bartimoccia, S., Santulli, M., Vasaturo, F., ... Pignatelli, P. (2017). Relationship of PCSK9 and Urinary Thromboxane Excretion to Cardiovascular Events in Patients with Atrial Fibrillation. *J. Am. Coll. Cardiol.*, 70, 1455–1462.

- Patocka, J., Bhardwaj, K., Klimova, B., Nepovimova, E., Wu, Q., Landi, M., ... Wu, W. (2020). *Malus domestica*: A review on nutritional features, chemical composition, traditional and medicinal value. *Plants*, 9(11), 1–19. <https://doi.org/10.3390/plants9111408>.
- Patterson, K. A., Zhang, X., Wroblewski, S. K., Hawley, A. E., Lawrence, D. A., Wakefield, T. W., ... Diaz, J. A. (2013). Rosuvastatin reduced deep vein thrombosis in ApoE gene deleted mice with hyperlipidaemia through non-lipid lowering effects. *Thromb Res.*, 131(3), 268–276.
- Pengnet, S., Prommaouan, S., Sumarithum, P., & Malakul, W. (2019). *Naringin Reverses High-Cholesterol Diet-Induced Vascular Dysfunction and Oxidative Stress in Rats via Regulating LOX-1 and NADPH Oxidase Subunit Expression*. 2019.
- Perumal, K. V., Ja'afar, N. L., Balan, S. S., Zainal, A. A., Arapoc, D. J., Shafie, N. H., & Bahari, H. (2019). Preventive effect of *Elateriospermum tapos* seed extract against obese Sprague Dawley rats. *Oriental Pharmacy and Experimental Medicine*, 1–7. <https://doi.org/10.1007/s13596-019-00394-w>.
- Petta, S., Gastaldelli, A., Rebelos, E., Bugianesi, E., Messa, P., Miele, L., ... Bonino, F. (2016). Pathophysiology of non alcoholic fatty liver disease. *International Journal of Molecular Sciences*, 17(12). <https://doi.org/10.3390/ijms17122082>.
- Prabhakaran, D., Jeemon, P., Sharma, M., Roth, T. A., Johnson, C., Harikrishnan, S., Gupta, R., Pandian, J., Naik, N., Roy, A., Dhaliwal, R., Xavier, D., Kumar, R., Tandon, N., Mathur, P., Shukla, D., Mehrotra, R., Venugopal, K., Kumar, A. G., ... Dandona, L. L. (2018). The changing patterns of cardiovascular diseases and their risk factors in the states of India: the Global Burden of Disease Study 1990-2016. *Lancet Glob Health*, 6, 1339-1351. [10.1016/S2214-109X\(18\)30407-8](https://doi.org/10.1016/S2214-109X(18)30407-8).
- Prasetyo, R. H., & Safitri, E. (2016). Effect of honey to mobilize endogenous stem cells in efforts intestinal and ovarian tissue regeneration in rats with proteinenergy malnutrition. *Asian Pac J Reprod*, 5, 198-203. <https://doi.org/10.1016/j.apjr.2016.04.0>.
- Procknow, J. D., Staiculescu, M. C., Walji, T. A., & Craft, C. S. (2017). *accumulation in Ldlr - / - mice*. 22–29. <https://doi.org/10.1016/j.atherosclerosis.2016.03.022>.Hypertension.
- Psarros, C., Economou, E. K., & Koutsilieris, M. (2015). *Statins as Pleiotropic Modifiers of Vascular Oxidative Stress and Inflammation*. 1(2), 43–54. <https://doi.org/10.1515/jccm-2015-0007>.
- Qadir, M. I., Manzoor, A., & Akash, M. S. H. (2018). Potential role of medicinal plants for anti-atherosclerosis activity. *Bangladesh Journal of Pharmacology*, 13(1), 59–66. <https://doi.org/10.3329/bjp.v13i1.33478>.

- Raal, F. J., Giugliano, R. P., Sabatine, M. S., Koren, M. J., Blom, D., Seidah, N. G., ... Stein, E. A. (2016). PCSK9 inhibition-mediated reduction in Lp(a) with evolocumab: an analysis of 10 clinical trials and the LDL receptor's role. *J Lipid Res*, *57*, 1086–1096. doi:10.1194/jlr.p065334.
- Rahman, M., Gan, S. H., & Khalil, M. (2014). Neurological effects of honey: current and future prospects. *Evid Based Complement Altern Med*, *2014*, 958721. doi: 10.1155/2014/958721.
- Raluca, E. H., Roxana, II., Doina, B., & Veronica, M. (2013). Flaxseed prevents leukocyte and platelet adhesion to endothelial cells in experimental atherosclerosis by reducing sVCAM-1 and vWF. *ScientificWorldJournal*, *2013*(Article ID 303950), 6.
- Rameshreddy, P., Uddandrao, V. V. S., Brahmanaidu, P., Vadivukkarasi, S., Ravindarnaik., R., Suresh, P., ... Saravanan, G. (2017). Obesity-alleviating potential of asiatic acid and its effects on ACC1, UCP2, and CPT1 mRNA expression in high fat diet-induced obese Sprague–Dawley rats. *Molecular and Cellular Biochemistry*., doi: 10.1007/s11010-017-3199-2.
- Ramli, N. Z., Chin, K. Y., Zarkasi, K. A., & Ahmad, F. (2018). A review on the protective effects of honey against metabolic syndrome. *Nutrients*, *10*(1009), doi: 10.3390/nu10081009.
- Ramos, C., Adrián Santoyo, H. C. S., Martínez, C., Olarte, I., & Adolfo Martínez. (2017). Insuficiencia hepática crónica y hemostasia Chronic Liver Failure and Hemostasis. *Rev Colomb Gastroenterol* /, *32*(4), 349–358.
- Reyes, G. M. (2019). Overview of the Coagulation System. In *Transfusion Medicine and Hemostasis* (Third Edit). Elsevier Inc. <https://doi.org/10.1016/b978-0-12-813726-0.00091-x>.
- Riccioni, G., Speranza, L., Pesce, M., Cusenza, S., D'Orazio, N., & Glade, M. (2012). Novel phytonutrient contributors to antioxidant protection against cardiovascular disease. *Nutrition*., *28*, 605–610.
- Robier, C. (2020). Platelet morphology. *Journal of Laboratory Medicine*, *44*(5), 231–239. <https://doi.org/10.1515/labmed-2020-0007>.
- Robinson, J. G., & Stone, N. J. (2013). “The 2013 ACC/AHA guideline on the treatment of blood cholesterol to reduce atherosclerotic cardiovascular disease risk: a new paradigm supported by more evidence,.” *EuropeanHeart Journal*, *36*(31), 2110–2118.
- Rocha, D. M., Caldas, A. P., Oliveira, L. L., Bressan, J., & Hermsdorff, H. H. (2016). Saturated fatty acids trigger TLR4-mediated inflammatory response. *Atherosclerosis*, *244*, 211–215.

- Rodriguez-Leyva, D., Malik, A., & Tappia, P. S. (2011). Gender-related gene expression in response to dietary fatty acids and predisposition to atherosclerosis and cardiovascular disease. *Clinical Lipidology*, 6(6), 653–664. <https://doi.org/10.2217/clp.11.62>.
- Rondina, M. T., Tatsumi, K., Bastarache, J. A., & Mackman, N. (2016). Microvesicle tissue factor activity and interleukin-8 levels are associated with mortality in patients with influenza A/H1N1 infection. *Crit Care Med*, 44, e574–e578.
- Roohbakhsh, A., Parhiz, H., Soltani, F., Rezaee, R., & Iranshahi, M. (2015a). Antioxidant and anti inflammatory properties of the citrus flavonoids hesperidin and hesperetin: an updated review of their molecular mechanisms and experimental models. *Phytother Res*, 29, 323–331.
- Roohbakhsh, A., Parhiz, H., Soltani, F., Rezaee, R., & Iranshihi, M. (2015b). Molecular mechanisms behind the biological effects of hesperidin and hesperetin for the prevention of cancer and cardiovascular diseases. *Life Sci*, 124(64–74).
- Roohbakhsh, A., Parhiz, H., Soltani, F., Rezaee, R., & Iranshahi, M. (2014). Neuropharmacological properties and pharmacokinetics of the citrus flavonoids hesperidin and hesperetin: a mini-review. *Life Sci*, 113, 1–6.
- Rosenbaum, M. A., Chaudhuri, P., & Graham, L. M. (2014). Hypercholesterolemia inhibits re-endothelialization of arterial injuries by TRPC channel activation. *J Vasc Surg*, pii S0741-.
- Rosenson, R. S., & Grundy, S. M. (2019). *Chapter 29: hyperlipidemia*. 1–55.
- Rosenson, R. S., Davidson, M. H., Hirsh, B. J., Kathiresan, S., & Gaudet, D. (2014). Genetics and causality of triglyceride-rich lipoproteins in atherosclerotic cardiovascular disease. *J* 2014;64(23):2525–2540. *Am Coll Cardiol.*, 64(23), 2525–2540.
- Roth, G. A., Mensah, G. A., Johnson, C. O., Addolorato, G., Ammirati, E., Baddour, L. M., Barengo, N. C., Beaton, A. Z., Benjamin, E. J., Benziger, C. P., Bonny, A., Brauer, M., Brodmann, M., Cahill, T. J., Carapetis, J., Catapano, A. L., Chugh, S. S., Cooper, L. T., Coresh, J., ... Fuster, V. (2020). Global Burden of Cardiovascular Diseases and Risk Factors, 1990–2019. *Journal of the American College of Cardiology*, 76(25), 2982–3021. <https://doi.org/10.1016/j.jacc.2020.11.010>.
- Ryu, S. K., King, T. J., Fujioka, K., Pattison, J., Pashkow, F. J., & Tsimikas, S. (2012). Effect of an oral astaxanthin prodrug (CDX-085) on lipoprotein levels and progression of atherosclerosis in LDLR(+/-) and ApoE(+/-) mice. *Atherosclerosis*, 222, 99–105.
- Saes, J. L., Schols, S. E. M., van Heerde, W. L., & Nijziel, M. R. (2018). Hemorrhagic disorders of fibrinolysis: a clinical review. *Journal of Thrombosis and Haemostasis*, 16(8), 1498–1509. <https://doi.org/10.1111/jth.14160>.

- Saito, J., Yokoyama, U., Nicho, N., Zheng, Y.-W., Ichikawa, Y., Ito, S., ... Ishikawa, Y. (2018). Tissue-type plasminogen activator contributes to remodeling of the rat ductus arteriosus. *PLoS ONE*, *13*(1), e0190871.
- Samad, F., & Ruf, W. (2017). *Review Article Inflammation, obesity, and thrombosis*. *122*(20), 3415–3423. <https://doi.org/10.1182/blood-2013-05-427708.coagulation>.
- Sampasa-Kanyinga, H., & Lewis, R. (2015). Frequent use of social networking sites is associated with poor psychological functioning among children and adolescents. *Cyberpsychology Behav Soc Netw.*, *18*, 380-385. [10.1089/cyber.2015.0055](https://doi.org/10.1089/cyber.2015.0055).
- Sampathkumar, M. T., Kasetti, R. B., Nabi, S. A., Sudarshan, P. R., Swapna, S., & Apparao, C. (2011). Antihyperlipidemic and antiatherogenic activities of *Terminalia pallida* Linn. fruits in high fat diet-induced hyperlipidemic rats. *Journal of Pharmacy and Bioallied Sciences*, *3*(3), 449–452. <https://doi.org/10.4103/0975-7406.84464>.
- Samsudin, S. (2016). The Prevalence of Non-Communicable Diseases among older age group in Malaysia and Its Effects on Health Care Demand. *International Journal of Public Health Research.*, *6*, 741–749.
- Sanguesa, G., Shaligram, S., Akther, F., Roglans, N., Laguna, J. C., Rahimian, R., & Algret, M. (2017). Type of supplemented simple sugar, not merely calorie intake, determines adverse effects on metabolism and aortic function in female rats. *Am. J. Physiol. Heart Circ. Physiol.*, *312*(2), H289–H304.
- Saravanakumar, M., Raja, B., Manivannan, J., Silambarasan, T., Prahalathan, P., Kumar, S., & Mishra, S. K. (2015). *Oral administration of veratric acid, a constituent of vegetables and fruits, prevents cardiovascular remodelling in hypertensive rats: a functional evaluation*. 1385–1394. <https://doi.org/10.1017/S0007114515003086>.
- Saravanan, G., Ponnuragan, P., Deepa, A., & Senthilkumar, B. (2014). *Anti-obesity action of gingerol: effect on lipid profile, insulin, leptin, amylase and lipase in male obese rats induced by a high-fat diet*. January. <https://doi.org/10.1002/jsfa.6642>.
- Sarkady, F., Szegedi, I., Fekete, I., Fekete, K., Csiba, L., & Bagoly, Z. (2020). *Low α 2-plasmin Inhibitor Antigen Levels on Admission Predict Unfavorable Outcomes in Acute Ischemic Stroke Patients Treated with Intravenous Thrombolysis*. 3–4.
- Sathibabu, U. V. V. (2017). Beneficial role of some natural products to attenuate the diabetic cardiomyopathy through Nrf2 pathway in cell culture and animal models. *Cardiovascular Toxicology.*, *doi: 10.10*.

- Satoh, T., Satoh, K., Yaoita, N., Kikuchi, N., Omura, J., Kurosawa, R. N., ... Shimokawa, H. (2017). Activated TAFI Promotes the Development of Chronic Thromboembolic Pulmonary Hypertension: A Possible Novel Therapeutic Target. *Circ. Res.*, *120*(8), 1246–1262.
- Sazlina, S. G., Sooryanarayana, R., Ho, B. K., Azahadi, O. M., Krishnapillai, A. D., Tohit, N. M., ... Ahmad, N. A. (2020). Cardiovascular disease risk factors among older people: Data from the National Health and Morbidity Survey 2015. *PLoS ONE*, *15*(10 October), 1–11. <https://doi.org/10.1371/journal.pone.0240826>.
- Schipke, J., Brandenberger, C., Rajces, A., Manninger, M., Alogna, A., Post, H., & Mühlfeld, C. (2017). Assessment of cardiac fibrosis: A morphometric method comparison for collagen quantification. *Journal of Applied Physiology*, *122*(4), 1019–1030. <https://doi.org/10.1152/jappphysiol.00987.2016>.
- Schuetze, S., Manig, A., Ribes, S., & Nau, R. (2019). Aged mice show an increased mortality after anesthesia with a standard dose of ketamine/xylazine. *Laboratory Animal Research*, *35*(1), 1–7. <https://doi.org/10.1186/s42826-019-0008-y>.
- Schulz, R., & Schlüter, K. D. (2017). PCSK9 targets important for lipid metabolism. *Clinical Research in Cardiology Supplements*, *12*(February), 2–11. <https://doi.org/10.1007/s11789-017-0085-0>.
- Scott, C. H., Hassan, M., & Yaish, M. D. (2019). Pediatric Thromboembolism eMedicine. <https://Emedicine.Medscape.Com/Article/959501-Print>.
- Sedaghat, A., Shahbazian, H., Rezazadeh, A., Haidari, F., Jahanshahi, A., Latifi, S. M., & Shirbeigi, E. (2019). The effect of soy nut on serum total antioxidant, endothelial function and cardiovascular risk factors in patients with type 2 diabetes. *Diabetes Metab. Syndr.*, *13*(2), 1387–1391. doi: 10.1016/j.dsx.2019.01.057.
- Segovia, S. A., Vickers, M. H., & Reynolds, C. M. (2019). The impact of maternal obesity on inflammatory processes and consequences for later offspring health outcomes. *J. Dev. Orig. Health Dis*, *8*(doi : 10.1017/S2040174417000204.), 529–540.
- Segovia, S. A., Vickers, M. H., Gray, C., & Reynolds, C. M. (2014). Maternal Obesity, Inflammation, and Developmental Programming. *Biomed. Res. Int.*, *Biomed. 418975*. doi : 10.1155/2014/418975.
- Seidah, N. G., Awan, Z., Chretien, M., & Mbikay, M. (2014). PCSK9: a key modulator of cardiovascular health. 114.301621. *Circ Res*, *114*, 1022–1036. doi:10.1161/circresaha.

- Semmler, L., Weberruß, H., Baumgartner, L., Pirzer, R., & Oberhoffer-Fritz, R. (2020). Vascular diameter and intima-media thickness to diameter ratio values of the carotid artery in 642 healthy children. *European Journal of Pediatrics*. <https://doi.org/10.1007/s00431-020-03785-3>.
- Sena, M., & Sena, C. M. (2018). *We are IntechOpen , the world ' s leading publisher of Open Access books Built by scientists , for scientists TOP 1 %*.
- Seo, E., Seo, K. W., Gil, J., Ha, Y., Yeom, E., Lee, S., & Lee, S. J. (2014). *Biophysicochemical properties of endothelial cells cultured on bio-inspired collagen films*. 1–16.
- Shao, Y., Yu, Y., Li, C., Yu, J., Zong, R., & Pie, C. (2016). Synergistic effect of quercetin and 6- gingerol treatment in streptozotocin induced type 2 diabetic rats and poloxamer P- 407 induced hyperlipidemia. *Rsc Adv*, 6, 12235–12242.
- Shapiro, M. D., & Fazio, S. (2017). Apolipoprotein B-containing lipoproteins and atherosclerotic cardiovascular disease. *F1000Res*, 6, 134.
- Shen, L., Sun, Z., Nie, P., Yuan, R., Cai, Z., Wu, C., ... He, B. (2019). Sulindac-derived retinoid X receptor- α modulator attenuates atherosclerotic plaque progression and destabilization in ApoE $^{-/-}$ mice. *British Journal of Pharmacology*, 176(14), 2559–2572. <https://doi.org/10.1111/bph.14682>.
- Shi, J., Li, R., Liu, Y., Lu, H., Yu, L., & Zhang, F. (2019). Shuangyu tiaozhi granule attenuates hypercholesterolemia through the reduction of cholesterol synthesis in rat fed a high cholesterol diet. *BioMed Research International*, 2019. <https://doi.org/10.1155/2019/4805926>.
- Shlomo, M., Ronald, K., Clifford, R., Richard, A., & Allison, G. (2019). Williams Textbook of Endocrinology,. *14th Edition. Elsevier., ISBN: 9780*.
- Smith, D. D., Tan, X., Raveendran, V. V., Tawfik, O., Stechschulte, D. J., & Dileepan, K. N. (2012). Mast cell deficiency attenuates progression of atherosclerosis and hepatic steatosis in apolipoprotein E-null mice. *American Journal of Physiology - Heart and Circulatory Physiology*, 302(12), 2612–2621. <https://doi.org/10.1152/ajpheart.00879.2011>.
- Soneye, M. A., Adekanmi, A. J., Obajimi, M. O., & Aje, A. (2019). Intima-media thickness of femoral arteries and carotids among an adult hypertensive Nigerian population: A case-control study to assess their use as surrogate markers of atherosclerosis. *Annals of African Medicine*, 158–166., 18(3), 158–166, <https://doi.org/10.4103/aam.aam5718>.
- Song, C., Burgess, S., Eicher, J. D., O'Donnell, C. J., Johnson, A. D., Huang, J., Sabater-Lleal, M., Asselbergs, F. W., Tregouet, D., Shin, S. Y., Ding, J., Baumert, J., Oudot-Mellakh, T., Folkersen, L., Smith, N. L., Williams, S. M., Ikram, M. A., Kleber, M. E., Becker, D. M., ... Cheng, Y. C. (2017). Causal effect of plasminogen activator inhibitor type 1 on coronary heart disease. *Journal of the American Heart Association*, 6(6). <https://doi.org/10.1161/JAHA.116.004918>.

- Sonneveld, M. A., de Maat, M. P., Portegies, M. L., Kavousi, M., Hofman, A., Turecek, P. L., ... Leebeek, F. W. G. (2015). Low ADAMTS13 activity is associated with an increased risk of ischemic stroke. *Blood*, *126*, 2739–2746.
- Sprague, Dawley. (2017). "Charles River". [10] ([Http://Www.Criver.Com/Products-Services/Basic-Research/Find-a-Model/Sprague-Dawley-Rat](http://www.Criver.Com/Products-Services/Basic-Research/Find-a-Model/Sprague-Dawley-Rat)).
- Stanisic, D., Liu, L. H. B., Dos Santos, R. V., Costa, A. F., Durán, N., & Tasic, L. (2020). New sustainable process for hesperidin isolation and anti-ageing effects of hesperidin nanocrystals. *Molecules*, *25*(19), 1–18. <https://doi.org/10.3390/molecules25194534>.
- Statistics on Causes of Death, Malaysia, 2017. (2017).
- Sun, Y. Z., Chen, J. F., Shen, L. M., Zhou, J., & Wang, C. F. (2017). Anti-atherosclerotic effect of hesperidin in LDLr^{-/-} mice and its possible mechanism. *European Journal of Pharmacology*, *815*(June), 109–117. <https://doi.org/10.1016/j.ejphar.2017.09.010>.
- Sun, G. Z., Li, Z., Guo, L., Zhou, Y., Yang, H. M., & Sun, Y. X. (2014). High prevalence of dyslipidemia and associated risk factors among rural Chinese adults. *Lipids in Health and Disease*, *13*(1). <https://doi.org/10.1186/1476-511X-13-189>.
- Sunil, C., Ignacimuthu, S., & Kumarappan, C. (2012). Hypolipidemic activity of *Symplocos cochinchinensis* S. Moore leaves in hyperlipidemic rats. *Journal of Natural Medicines*, *66*(1), 32–38. <https://doi.org/10.1007/s11418-011-0548-4>.
- Suvik, A. (2012). The use of modified Masson's trichrome staining in collagen evaluation in wound healing study. *Malaysian Journal of Veterinary Research (Malaysia)*, *January 2012*, 39–47.
- Swieringa, F., Baaten, C. C., Verdoold, R., Mastenbroek, T. G., Rijnveld, N., van der Laan, K. O., ... van der Meijden, P. E. J. (2016). Platelet control of fibrin distribution and microelasticity in thrombus formation under flow. *Arterioscler Thromb Vasc Biol.*, *36*, 699. doi: 10.1161/ATVBAHA.115.306537.
- Tang, X., Liu, J., Dong, W., Li, P., Li, L., Lin, C., ... Li, D. (2013). The cardioprotective effects of citric acid and L-malic acid on myocardial ischemia/reperfusion injury. *Evidence-Based Complementary and Alternative Medicine*, *2013*. <https://doi.org/10.1155/2013/820695>.
- Tarkin, J. M., Joshi, F. R., & Ruud, J. H. F. (2014). PET imaging of inflammation in atherosclerosis. *Nat Rev Cardiol*, *11*, 443–457.
- Tasneem, Z., Naqvi, M. D., & S. Lee, M.-S. (2014). *Carotid Intima-Media Thickness and Plaque in Cardiovascular Risk Assessment*. *7*(10). <https://doi.org/10.1016/j.jcimg.2013.11.014>.

- Tatsumi, K., & Mackman, N. (2015). Tissue factor and atherothrombosis. *Journal of Atherosclerosis and Thrombosis*, 22(6), 543–549. <https://doi.org/10.5551/jat.30940>.
- Testai, L., & Calderone, V. (2017). Nutraceutical value of citrus flavanones and their implications in cardiovascular disease. *Nutrients*, 9(5), 1–13. <https://doi.org/10.3390/nu9050502>.
- Tomaiuolo, M., Litvinov, R. I., Weisel, J. W., & Stalker, T. J. (2020). Use of electron microscopy to study platelets and thrombi. *Platelets*, 31(5), 580–588. <https://doi.org/10.1080/09537104.2020.1763939>.
- Torisu, T., Torisu, K., Lee, I. H., Liu, J., Malide, D., Combs, C. A., ... Finkel, T. (2013). Autophagy regulates endothelial cell processing, maturation and secretion of von Willebrand factor. *Nat Med*, 19, 1281–1287.
- Tousoulis, D., Kampoli, A.-M., Tentolouris, C., Papageorgiou, N., & Stefanadis, C. (2011). The Role of Nitric Oxide on Endothelial Function. *Current Vascular Pharmacology*, 10(1), 4–18. <https://doi.org/10.2174/157016112798829760>.
- Tripathi, S., & Mazumder, P. M. (2020). Apple cider vinegar (ACV) and their pharmacological approach towards alzheimer's disease (AD): A review. *Indian Journal of Pharmaceutical Education and Research*, 54(2), S67–S74. <https://doi.org/10.5530/ijper.54.2s.62>.
- Umeno, A., Horie, M., Murotomi, K., Nakajima, Y., & Yoshida, Y. (2016). Antioxidative and antidiabetic effects of natural polyphenols and isoflavones. *Molecules*, 21(E708).
- Van Der Stoep, M., Korporaal, S. J. A., & Van Eck, M. (2014). High-density lipoprotein as a modulator of platelet and coagulation responses. *Cardiovascular Research*, 103(3), 362–371. <https://doi.org/10.1093/cvr/cvu137>.
- Vedder, V. L. (2020). *Dare to Compare . Development of Atherosclerotic Lesions in Human , Mouse , and Zebrafish*. 7(June). <https://doi.org/10.3389/fcvm.2020.00109>.
- Verma, N. (2016). Introduction To Hyperlipidemia and Its Treatment: a Review. *International Journal of Current Pharmaceutical Research*, 9(1), 6. <https://doi.org/10.22159/ijcpr.2017v9i1.16616>.
- Vieira, A. J., Pereira, F., Souza, M. C., Totti, B. M., & Rozza, A. L. (2018). Limonene: Aroma of innovation in health and disease. *Chemico-Biological Interactions* 283, DOI: 10.1016/j.cbi.2018.02.007.
- Vilahur, G., Ben-aicha, S., & Badimon, L. (2017). New insights into the role of adipose tissue in thrombosis. *Thrombosis and Haemostasis*. 1046–1054. <https://doi.org/10.1093/cvr/cvx086>.

- Virani, S. S., Alonso, A., Benjamin, E. J., Bittencourt, M. S., Callaway, C. W., Carson, A. P., Chamberlain, A. M., Chang, A. R., Cheng, S., Delling, F. N., Djousse, L., Elkind, M. S. V., Ferguson, J. F., Fornage, M., Khan, S. S., Kissela, B. M., Knutson, K. L., Kwan, T. W., Lackland, D. T., ... Heard, D. G. (2020). Heart disease and stroke statistics—2020 update: A report from the American Heart Association. *Global Burden of Disease High Blood Cholesterol and other Lipids*. *Cvd*, 9-11. In *Circulation*. <https://doi.org/10.1161/CIR.0000000000000757>.
- Vorkapic, E., Dugic, E., Vikingsson, S., Roy, J., Mäyränpää, M. I., Eriksson, P., & Wågsäter, D. (2016). Imatinib treatment attenuates growth and inflammation of angiotensin II induced abdominal aortic aneurysm. *Atherosclerosis*, 249, 101–109. <https://doi.org/10.1016/j.atherosclerosis.2016.04.006>.
- Wang, S., Ye, Q., Zeng, X., & Qiao, S. (2019). Functions of macrophages in the maintenance of intestinal homeostasis. *Journal of Immunology Research*, 2019. <https://doi.org/10.1155/2019/1512969>.
- Wang, M., Hao, H., Leeper, N. J., & Zhu, L. (2018a). Thrombotic Regulation From the Endothelial Cell Perspectives. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 38(6), e90–e95. <https://doi.org/10.1161/ATVBAHA.118.310367>.
- Wang, W., Zhao, L., Huang, H., Yao, J., Zhou, L., Wang, D., & Qiu, X. (2018b). Development of an Ultra-High Performance Liquid Chromatography Method for Simultaneous Determination of Six Active Compounds in Fructus aurantii and Rat Plasma and Its Application to a Comparative Pharmacokinetic Study in Rats Administered with Different . *Journal of Analytical Methods in Chemistry*, 2018. <https://doi.org/10.1155/2018/7579136>.
- Wang, Z. T., Wang, Z., & Hu, Y. W. (2016). Possible roles of platelet-derived microparticles in atherosclerosis. *Atherosclerosis*, 248, 10–16. <https://doi.org/10.1016/j.atherosclerosis.2016.03.004>.
- Wang, Y., Huang, Y., Hobbs, H. H., & Cohen, J. C. (2012). Molecular characterization of proprotein convertase subtilisin/kexin type 9-mediated degradation of the LDLR. *J Lipid Res*, 53, 1932–1943. doi:10.1194/ jlr.m028563.
- Wang, X., Hasegawa, J., Kitamura, Y., Wang, Z., Matsuda, A., Shinoda, W., ... Kimura, K. (2011). Effects of hesperidin on the progression of hypercholesterolemia and fatty liver induced by high-cholesterol diet in rats. *Journal of Pharmacological Sciences*, 117(3), 129–138. <https://doi.org/10.1254/jphs.11097FP>.
- Weibel, G. L., Drazul-Schrader, D., Shivers, D. K., Wade, A. N., Rothblat, G. H., Reilly, M. P., & de la Llera-Moya, M. (2014). Importance of evaluating cell cholesterol influx with efflux in determining the impact of human serum on cholesterol metabolism and atherosclerosis. *Arterioscler Thromb Vasc Biol.*, 34, 17–25.
- Weibel, E. R., & Palade, G. E. (1964). New cytoplasmic components in arterial endothelia. *J. Cell Biol.*, 23, 101–112. doi:10.1083/jcb. 23.1.101.

- Wentzel, J. J., Chatzizisis, Y. S., Gijzen, F. J., Giannoglou, G. D., Feldman, C. L., & Stone, P. H. (2012). Endothelial shear stress in the evolution of coronary atherosclerotic plaque and vascular remodelling: current understanding and remaining questions. *Cardiovasc Res.*, *96*(234–243).
- Whale, A. S., De Spiegelaere, W., Trypsteen, W., Nour, A. A., Bae, Y.-K., Benes, V., Burke, D., Cleveland, M., Corbisier, P., Devonshire, A. S., Dong, L., Drandi, D., Foy, C. A., Garson, J. A., He, H.-J., Hellemans, J., Kubista, M., Lievens, A., Makrigiorgos, M. G., ... Huggett, J. F. (2020). The Digital MIQE Guidelines Update: Minimum Information for Publication of Quantitative Digital PCR Experiments for 2020. *Clinical Chemistry*, *66*(8), 1012–1029. <https://doi.org/10.1093/clinchem/hvaa125>.
- Williams, M. S., Cushman, M., & Ouyang, P. (2016). *Association of Serum Sex Hormones with Hemostatic Factors in Women On and Off Hormone Therapy: The Multiethnic Study of Atherosclerosis*. *25*(2), 166–172. <https://doi.org/10.1089/jwh.2015.5465>.
- Wiśniewska, A., Olszanecki, R., Totoń-Żurańska, J., Kuś, K., Stachowicz, A., Suski, M., ... Korbuc, R. (2017). Anti-Atherosclerotic Action of Agmatine in ApoE-Knockout Mice. *Int. J. Mol. Sci.*, *18*(1706).
- Wong, N. D. (2012). Is diabetes really a coronary heart disease risk equivalent? *Cardiovascular Endocrinology*, *1*(14), 65–67.
- World Health Organization. (2020). WHO reveals leading causes of death and disability worldwide: 2000-2019. *WHO Reveals Leading Causes of Death and Disability Worldwide: 2000-2019*.
- World Health Organisation. (2017). World Health Organization. Cardiovascular diseases (CVDs) [Internet]. *World Health Organization*, Geneva; 2017. http://www.who.int/cardiovascular_.
- World Health Organization. (2016). Obesity and overweight. *Fact Sheet, Available Online at: Http://Www.Who.Int/Mediacentre/Factsheets/Fs311/En/*,.
- Wu, H. C., Horng, C. T., Tsai, S. C., Lee, Y. L., Hsu, S. C., Tsai, Y. J., ... Yang, J. S. (2018a). Relaxant and vasoprotective effects of ginger extracts on porcine coronary arteries. *International Journal of Molecular Medicine*, *41*(4), 2420–2428. <https://doi.org/10.3892/ijmm.2018.3380>.
- Wu, Z., Xia, Z., Wu, B., Wei, F., Lv, X., Xie, Y., ... Huang, F. (2018b). Lipid profiling in serum from apolipoprotein E-knock out mice fed with different diets and its application to the study of the regulatory effect on lipid metabolism. *Food Funct.*, *9*(10), 5103–5114.
- Wu, M. Y., Li, C. J., Hou, M. F., & Chu, P. Y. (2017). New insights into the role of inflammation in the pathogenesis of atherosclerosis. *International Journal of Molecular Sciences*, *18*(10). <https://doi.org/10.3390/ijms18102034>.

- Xiong, H., Wang, J., Ran, Q., Lou, G., Peng, C., Gan, Q., ... Huang, Q. (2019). Hesperidin: A therapeutic agent for obesity. In *Drug Design, Development and Therapy* (Vol. 13, pp. 3855–3866). <https://doi.org/10.2147/DDDT.S227499>.
- Xu, C., Mathews, A. E., Rodrigues, C., Eudy, B. J., Rowe, C. A., O'Donoghue, A., & Percival, S. S. (2018). Aged garlic extract supplementation modifies inflammation and immunity of adults with obesity: A randomized, double-blind, placebocontrolled clinical trial. *Clin. Nutr. ESPEN*, *24*, 148–155. doi: 10.1016/j.clnesp.2017.11.01.
- Xu, X. R., Carrim, N., Neves, M. A. D., McKeown, T., Stratton, T. W., Coelho, R. M. P., ... Ni, H. (2016). Platelets and platelet adhesion molecules: Novel mechanisms of thrombosis and anti-thrombotic therapies. *Thrombosis Journal*, *14*(Suppl 1). <https://doi.org/10.1186/s12959-016-0100-6>.
- Xu, D., Wang, Z., Zhang, Y., Jiang, W., Pan, Y., Song, B.-L., & Chen, Y. (2015). “PAQR3 modulates cholesterol homeostasis by anchoring Scap/SREBP complex to the Golgi apparatus.” *Nature Communications*, *6*, 8100.
- Xu, Q.-Y., Liu, Y.-H., Zhang, Q., Ma, B., Yang, Z.-D., Liu, L., ... Wu, Z.-M. (2014). Metabolomic analysis of simvastatin and fenofibrate intervention in high-lipid diet-induced hyperlipidemia rats. *Acta Pharmacol Sin*, *35*, 1265–1273.
- Yahaya, R., Dash, G. K., Abdullah, M. S., & Mathews, A. (2015). *Clinacanthus nutans* (burm. F.) Lindau : An Useful Medicinal Plant of. *7*(6), 1244–1250.
- Yang, C., Li, L., Yang, L., Lu, H., Wang, S., & Sun, G. (2018). Anti-obesity and Hypolipidemic effects of garlic oil and onion oil in rats fed a high-fat diet. *Nutrition and Metabolism*, *15*(1), 4–11. <https://doi.org/10.1186/s12986-018-0275-x>.
- Yau, J. W., Teoh, H., & Verma, S. (2015). Endothelial cell control of thrombosis. *BMC Cardiovascular Disorders*, *15*(1), 1–11. <https://doi.org/10.1186/s12872-015-0124-z>
- Yida, Z., Imam, M. U., Ismail, M., Ismail, N., & Ideris, A. (2015). High fat diet-induced inflammation and oxidative stress are attenuated by N-acetylneuraminic acid in rats. *Journal of Biomedical Science*, 1–10. <https://doi.org/10.1186/s12929-015-0211-6>.
- Yin, T., Wang, G., Ye, T., & Wang, Y. (2016). Sulindac, a non-steroidal anti-inflammatory drug, mediates breast cancer inhibition as an immune modulator. *Scientific Reports*, *6*(October 2015), 1–8. <https://doi.org/10.1038/srep19534>.
- Zaragoza, C., Gomez-Guerrero, C., Martin-Ventura, J. L., Blanco-Colio, L., Lavin, B., Mallavia, B., ... Egido, J. (2011). Animal models of cardiovascular diseases. *Journal of Biomedicine and Biotechnology*, *2011*. <https://doi.org/10.1155/2011/497841>.

- Zeng, Z., Cao, B., Guo, X., Li, W., Li, S., Chen, J., ... Wei, Y. (2018). Apolipoprotein B-100 peptide 210 antibody inhibits atherosclerosis by regulation of macrophages that phagocytize oxidized lipid. *American Journal of Translational Research*, 10(6), 1817–1828.
- Zhang, Y. X., Wang, Z. X., Zhao, J. S., & Chu, Z. H. (2017). The current prevalence and regional disparities in general and central obesity among children and adolescents in Shandong, China. *International Journal of Cardiology*, 227, 89–93. <https://doi.org/10.1016/j.ijcard.2016.11.135>.
- Zhang, A., Qiu, S., Sun, H., Zhang, T., Guan, Y., Han, Y., ... Wang, X. (2016). Scoparone affects lipid metabolism in primary hepatocytes using lipidomics. *Scientific Reports*, 6(May), 1–8. <https://doi.org/10.1038/srep28031>.
- Zhao, L., Liu, B., & Li, C. (2015). *Progress in research into the genes associated with venous thromboembolism*. 6(2), 100–104. <https://doi.org/10.5847/wjem.j.1920>.
- Zhu, C., Dong, Y., Liu, H., Ren, H., & Cui, Z. (2017). Hesperetin protects against H₂O₂-triggered oxidative damage via upregulation of the Keap1-Nrf2/HO-1 signal pathway in ARPE-19 cells. *Biomed Pharmacother*, 88(124–33).
- Zhu, J., Wang, C. G., & Xu, Y.-G. (2011). Lycopene attenuates endothelial dysfunction in streptozotocin-induced diabetic rats by reducing oxidative stress. *Pharm Biol*, 49, 1144–1149.
- Zuurbier, S. M., Arnold, M., Middeldorp, S., Broeg-Morvaj, A., Silvis, S. M., Heldner, M. R., ... Coutinho, J. M. (2016). Risk of cerebral venous thrombosis in obese women. *JAMA Neurol*, 73, 579–584.