



UNIVERSITI PUTRA MALAYSIA

***HOMING PEPTIDES (RGD/iRGD) FUNCTIONALIZED ZEOLITIC
IMIDAZOLATE FRAMEWORK-8 FOR TARGETED DELIVERY OF
GEMCITABINE TO LUNG CANCER CELL***

NURUL AKMARINA BINTI MOHD ABDUL KAMAL

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By

NURUL AKMARINA BINTI MOHD ABDUL KAMAL

**Thesis Submitted to School of Graduate Studies, Universiti Putra
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Philosophy**

March 2022

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

HOMING PEPTIDES (RGD/iRGD) FUNCTIONALIZED ZEOLITIC IMIDAZOLATE FRAMEWORK-8 FOR TARGETED DELIVERY OF GEMCITABINE TO LUNG CANCER CELL

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NURUL AKMARINA BINTI MOHD ABDUL KAMAL

March 2022

Chair: Mohd Basyaruddin Abdul Rahman, PhD
Faculty: Science

Lung cancer is a serious threat to human health, with metastasis being the top cause of cancer-related mortality. Gemcitabine (GEM) used in treating lung cancer operates in a non-selective manner tending to accumulate in normal tissue when cancer patients face a long duration of treatment. To mitigate the non-selective action of the GEM on the healthy tissues, there is a vital necessity to develop targeted nano delivery systems capable of regulating optimum doses selectively to cancer cells and minimizing untoward toxicity to normal tissues. Herein, a reticular nanoparticle zeolitic imidazolate framework-8 (nZIF-8) encapsulating GEM was surface-functionalized with selective homing systems (RGD; sequence Arg-Gly-Asp and iRGD; sequence Cys-Arg-Gly-Asp-Lys-Gly-Pro-Asp-Cys, respectively) through a straightforward, one-pot solvothermal reaction. Successful surface functionalization of nZIF-8 encapsulated GEM (GEM_cnZIF-8) with RGD and iRGD, respectively, were characterized and systematically interpreted as a function of newly-formed functional groups, particle size, surface structure, surface topography, surface area, and surface charge. These functionalized GEM_cRGD@nZIF-8 and GEM_ciRGD@nZIF-8 not only responsive to an acidic environment but also controlled the GEM dissolution released rate (57.6 and 56.2%, respectively) after 48 h compared to non-functionalized GEM_cnZIF-8 (76.0%). Both functionalized nanoparticles successfully increased the cellular uptake within cancerous human adenocarcinoma alveolar epithelial cells (A549), compared with non-functionalized nanoparticles. The highest uptake was shown by iRGD functionalized nZIF-8. The GEM_cRGD@nZIF-8 and GEM_ciRGD@nZIF-8 experienced not only efficient uptake within A549, but also induced obvious cytotoxicity (75 and 73%, respectively at 10 $\mu\text{g mL}^{-1}$) and apoptosis (62 and 74.9%) post 48 h treatment when compared to the GEM_cnZIF-8. The apoptosis study verified the ability of functionalized GEM_ciRGD@nZIF-8 to intensify the

apoptotic population in A549 while minimizing cell death (11%) in normal human lung fibroblast cells (MRC-5). Both functionalized nanoparticles reflect a potential outstanding treatment efficacy when GEM@RGD@nZIF-8 and GEM@iRGD@nZIF-8 demonstrated selective cytotoxicity (selective index, SI > 2) towards A549 than MRC-5. The follow-up study using healthy zebrafish embryos demonstrated enhanced permeation by both functionalized nanoparticles. At the lethal endpoint (96 h), all nanoparticles at all concentrations (7.81-250 $\mu\text{g mL}^{-1}$) did not elicit toxicity during embryonic and larvae stages. With the absence of nZIF-8, pristine GEM at a concentration of 250 $\mu\text{g mL}^{-1}$ exhibited 33% of pericardially and yolk sac edema indicating adverse side effects of the chemotherapeutic agent alone towards healthy tissue. Taken together, it is believed that surface functionalization of zeolitic imidazolate framework-8 with RGD/iRGD holds great selective targeting and enhances the optimum chemotherapeutic dosage to human lung cancer with less unwarranted toxicity to normal human lung fibroblast.

Abstrak tesis yang dikemukakan kepada Senat of Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Doktor Falsafah

**KERANGKA ZEOLITIK IMIDAZOLAT-8 BERFUNGSIKAN PEPTIDA PANDU
TUJU (RGD/iRGD) UNTUK HANTARAN GEMCITABINE SECARA
BERSASAR KEPADA SEL BARAH PARU-PARU**

Oleh

NURUL AKMARINA BINTI MOHD ABDUL KAMAL

Mac 2022

Pengerusi: Mohd Basyaruddin Abdul Rahman, PhD
Fakulti: Sains

Barah paru-paru merupakan ancaman serius bagi kesihatan manusia, dengan metastasis menjadi punca utama kematian. Gemcitabine (GEM) yang digunakan untuk merawat barah paru-paru berfungsi secara tidak-berpilih lebih cenderung berkumpul dalam tisu normal apabila pesakit barah menjalani rawatan dalam tempoh yang panjang. Bagi mengurangkan tindakan tidak-berpilih oleh GEM ke atas tisu normal, adalah menjadi keperluan penting untuk membangunkan sistem hantaran nano bersasar yang boleh mengawal selia dos optimum secara berpilih kepada sel barah dan meminimumkan ketoksikan yang tidak diingini pada tisu normal. Di sini, satu kerangka retikular nanopartikel zeolitik imadazolot-8 (nZIF-8) mengandungi GEM telah difungsikan permukaannya dengan sistem panduan yang berpilih (RGD; urutan Arg-Gly-Asp and iRGD; urutan Cys-Arg-Gly-Asp-Lys-Gly-Pro-Asp-Cys, masing-masing) melalui tindak balas mudah solvoterma secara pukal. Kejayaan memungsikan permukaan nZIF-8 yang mengandungi GEM (GEM@nZIF-8) dengan RGD dan iRGD, masing-masing, telah dicirikan dan dianalisis secara sistematik dalam bentuk kumpulan berfungsi baru yang terhasil, saiz partikel, struktur permukaan, topografi permukaan, luas permukaan, dan cas permukaan. GEM@RGD@nZIF-8 and GEM@iRGD@nZIF-8 yang telah difungsikan bukan sahaja bertindak balas terhadap persekitaran berasid tetapi juga mengawal kadar pelepasan pelarutan GEM (57.6 dan 56.2%, masing-masing) selepas 48 jam berbanding dengan GEM@nZIF-8 (76.0%) yang tidak berfungsi. Kedua-dua nanopartikel yang difungsikan berjaya meningkatkan pengambilan selular oleh sel barah manusia epitelial alveolar adenokarsinoma (A549), berbanding dengan nanopartikel yang tidak difungsikan. Pengambilan tertinggi ditunjukkan oleh iRGD berfungsi nZIF-8. GEM@RGD@nZIF-8 and GEM@iRGD@nZIF-8 tidak hanya menunjukkan kecekapan pengambilan oleh A549, tetapi juga mengaruh sitotoksik (75 dan 73%, masing-masing pada 10 $\mu\text{g mL}^{-1}$) dan apoptosis (62 and 74.9%) selepas rawatan 48 jam berbanding

GEM@nZIF-8. Kajian apoptosis mengesahkan kemampuan GEM@iRGD@nZIF-8 yang difungsikan untuk meningkatkan populasi apoptosis pada A549 sementara meminimumkan juga kematian (11%) pada sel normal fibroblas paru-paru manusia (MRC-5). Kedua-dua nanopartikel yang difungsikan menunjukkan potensi keberkesanan rawatan yang luar biasa apabila GEM@RGD@nZIF-8 dan GEM@iRGD@nZIF-8 menunjukkan kepilihan sitotoksik (indeks kepilihan, SI>2) terhadap A549 lebih dari MRC-5. Kajian susulan menggunakan embrio ikanzebra yang sihat menunjukkan peningkatan resapan oleh kedua-dua nanopartikel yang difungsikan. Pada titik akhir maut (96 jam), semua nanopartikel pada semua kepekatan (7.81-250 $\mu\text{g mL}^{-1}$) tidak menimbulkan ketoksikan semasa peringkat embrio dan larva. Dengan ketiadaan nZIF-8, GEM tulen pada kepekatan 250 $\mu\text{g mL}^{-1}$ menunjukkan 33% edema pada kantung perikardi dan kuning telur menandakan kesan sampingan buruk dari agen kemoterapi tulen terhadap tisu yang sihat. Secara menyeluruh, adalah diyakini bahawa permukaan berfungsi RGD/iRGD pada kerangka imidazolat zeolitik-8 menunjukkan penyasaran berpilih yang sangat baik dan meningkatkan dos kemoterapeutik secara optimum kepada barah paru-paru manusia dengan mengurangkan ketoksikan yang tidak diinginkan pada normal fibroblas paru-paru manusia.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Mohd Basyaruddin bin Abdul Rahman, PhD

Professor, ChM.
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Emilia binti Abd Malek, PhD

Associate Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

Sharida binti Fakurazi, PhD

Professor Datin
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 21 July 2022

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Signature: _____

Name of Chairman of
Supervisory
Committee:

Prof. ChM Dr. Mohd Basyaruddin bin Abdul
Rahman

Signature: _____

Name of Member of
Supervisory
Committee:

Assoc. Prof. Dr. Emilia binti Abd Malek

Signature: _____

Name of Member of
Supervisory
Committee:

Prof. Datin Dr. Sharida binti Fakurazi

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LIST OF ABBREVIATIONS

AFM	Atomic force microscope
A459	Human adenocarcinoma alveolar epithelial cells
BET	Brunauer-Emmett-Teller
CendR	C-end Rule
DLS	Dynamic light scattering
ECM	Extracellular matrix
EPR	Enhanced permeability and retention
GEM	Gemcitabine
f_2	Similarity factor
FDA	Food and Drug Administration
FTIR-ATR	Fourier transform infrared-attenuated total reflection
FlcnZIF-8	Fluorescein encapsulated nZIF-8
FlcRGD@nZIF-8	Fluorescein encapsulated RGD functionalized nZIF-8
FlciRGD@nZIF-8	Fluorescein encapsulated iRGD functionalized nZIF-8
GEMcnZIF	Gemcitabine encapsulated nZIF-8
GEMcRGD@nZIF-8	Fluorescein encapsulated RGD functionalized nZIF-8
GEMciRGD@nZIF-8	Fluorescein encapsulated iRGD functionalized nZIF-8
HPLC	High-performance liquid chromatography
HRTEM	High-resolution transmission electron microscope
iRGD	Cyclic RGD
MOFs	MOF Metal-Organic Frameworks
MRC-5	normal human lung fibroblast cells
NRP-1	Neuropilin-1

NSCLC	Non-small cell lung cancer
nZIF-8	nanoparticle zeolitic imidazolate framework-8
PDT	Photodynamic therapy
PBS	Phosphate buffer saline
PXRD	Powder X-ray diffraction analysis
RGD	Arg-Gly-Asp
SEC	Size exclusion-chromatography
SI	Selective index
SLF	Simulated lung fluid
TGA	Thermal gravimetric analysis
WHO	World Health Organization
ZIFs	Zeolitic imidazolate frameworks

CHAPTER 1

INTRODUCTION

1.1 Research background

Lung cancer has one of the highest prevalence rates with the poorest prognosis amongst cancer types (Globocan, 2020). Over 50% of those diagnosed with lung cancer die within their first year of diagnosis, beyond which time, less than 18% will survive after the fifth year (Cruz *et al.*, 2011). Nevertheless, lung cancer survival rates are also influenced by the stage of cancer. For cases diagnosed when the cancer is still localized, the five-year survival rate for lung cancer is 56%. Unfortunately, only 16% of lung cancer are diagnosed at an early stage and when the tumor metastasizes, only 5% will survive after the fifth year (NIH 2018). In Malaysia, lung cancer accounting approximately 10% of all malignancies, with only 11% of the five-year relative survival rate. Nearly 90% of lung cancer patients in Malaysia are diagnosed with stage III or IV disease. At this stage, patients are commonly treated with various chemotherapeutic agents available (MySCan, 2018).

Traditionally, the primary approach for treating late-stage lung cancer remains chemotherapy, in which gemcitabine (GEM) is employed alone or in combination with other platinum-based chemotherapeutic agents such as cisplatin and carboplatin (Cavalcante & Monteiro, 2014; Ciccolini *et al.*, 2016; Manegold, 2004; Zappa & Mousa, 2016). Gemcitabine is a nucleoside analog that mediates its cytotoxic effects by inhibiting DNA synthesis. However, the non-selective action towards cancer cells and tends to accumulate with high doses leads to severe toxicity in normal cells (Golombek *et al.*, 2018; Hryciuk *et al.*, 2018). As such, there exists an urgent need to rectify this situation by developing targeted chemotherapeutic delivery systems that are capable of regulating optimal doses specifically to cancer cells without harming adjacent and ever-present normal, healthy cells (Cheok, 2012; Harrington & Smith, 2008; Wahgiman *et al.*, 2019).

To realize such systems, research attention has been placed on the creation and use of nanocarriers (Barenholz, 2012). Different types of nanocarriers such as liposomes, chitosan, iron oxide, and albumin have been formulated with varying degrees of success (Singh *et al.*, 2019). Indeed, several classes have exhibited excellent performance in clinical phase trials (Grodzinski *et al.*, 2019; He *et al.*, 2019). As a representative example, polyethylene glycol (PEG) have been clinically approved as a result of their ability to improve the low solubilities of hydrophobic chemotherapeutic agents (Gothwal *et al.*, 2016). Others based on lipids have also successfully improved the loading capacity of both hydrophobic and hydrophilic chemotherapeutic agents (Asmawi *et al.*, 2018; Chen *et al.*, 2020; Naderinezhad *et al.*, 2017). The general mechanism by which untargeted

nanocarriers operate is most often one of a passive-targeting strategy (R. J. Browning *et al.*, 2017), whereby the nanocarriers take advantage of solid tumor conditions such as inadequate lymphatic drainage. This leads to nanocarrier accumulation in the cancer cells for a longer period (Yu, B., and Tai, H.C, 2010). However, this mechanism is inefficient due to high interstitial fluid pressure that restricts the nanocarriers accumulation and is removed outward. Therefore, the use of ligand can promote the nanocarriers binding by attaching it to the cell membrane and preventing the nanocarriers from being removed from the tumor cells (Izci *et al.*, 2021).

To advance the effectiveness of chemotherapeutic nanocarriers, it is essential to focus on developing such systems based on active-targeting mechanisms (Kan *et al.*, 2011; H. Li *et al.*, 2018; Tamam *et al.*, 2019). The most straightforward method for achieving this is to functionalize the surface of the nanocarrier with active ligands that specifically target certain cancer cells; examples of which include antibodies and peptides (Egeblad *et al.*, 2010). In general, peptides are deemed increasingly promising as certain sequences (i.e. Arg-Gly-Asp; RGD and CRGDKGPDC; iRGD) are capable of selectively interacting with specific integrin receptors ($\alpha v \beta 3$) overexpress on the tumor endothelial (Pridgen *et al.*, 2013). Furthermore, instead of integrins receptors interaction, iRGD peptide can also bind to neuropilin-1 (NRP-1) receptors via the CendR fragment (Sugahara *et al.*, 2010). The general mechanism of such an interaction is akin to a 'homing device', whereby the functionalized nanocarrier navigates to a cancer cell, docks as a result of selective interactions with the specific integrin and neuropilin-1 (NRP-1) receptors, and then releases the chemotherapeutic agent (Attia *et al.*, 2019; Freund *et al.*, 2018; Kang *et al.*, 2020; Muhamad *et al.*, 2018)

In the quest to realize a nanoparticle for the active and autonomous targeting of cancer cells for delivery of a chemotherapeutic payload, two requisite structural characteristics were considered: (i) high porosity, which affords the ability to load and regulate the chemotherapeutic agents; and (ii) surface customizability to ensure the active-targeting of cancer cells by the nanoparticles. Reticular materials that are governed by reticular chemistry meet both structural important requirements. Zeolitic imidazolate frameworks (ZIFs), are a subclass of reticular materials, known for their high porosity, well-defined structures due to their crystallinity, and structural diversity (Furukawa *et al.*, 2013). ZIFs have been demonstrated capable of loading optimal quantities of chemotherapeutic agents within their pores, their surfaces have proven modifiable by a wide range of moieties, and their biocompatibility has been established (Maleki *et al.*, 2020). Accordingly, ZIF-8, an archetypal reticular material adopting a sodalite structure, can be surface functionalized and can be efficiently loaded within and released from its pores (Dong *et al.*, 2019a; Lin *et al.*, 2020; Zheng *et al.*, 2017). The exclusive properties of ZIF-8 and good targeting manner owned by homing targeting peptides are the revolutionary ideas to solve the non-selective and accumulation issue of GEM.

1.2 Justification and hypothesis of the study

The reasons for pursuing reticular nanoparticles zeolitic imidazolate framework-8 (nZIF-8) as GEM nanocarrier is the fact that they are well-known to embody high porosity, large and modifiable surface areas, tailorable particle size, pH-responsive, and often enjoy inherent biocompatibility, which can improve the non-selective manner of GEM. The active modified nanoparticles with RGD or iRGD are able to direct to the cancer cells selectively, ligand-receptors bind, then offload the chemotherapeutic agent, GEM. Previous studies have shown the advantages of nZIF-8 after encapsulation and functionalization reactions in treating cancer cells, including breast and cervical cancer cells (Dong *et al.*, 2019b; Hao *et al.*, 2021; Lin *et al.*, 2020). Therefore, this study anticipated that these active nanoparticles would improve therapeutic effectiveness by selectively regulating the chemotherapeutic agent to lung cancer cells while causing less harm to healthy cells than those currently available.

1.3 Objectives

The aim of this research is to investigate the possibility of enhancing the non-selective activity of chemotherapeutic, gemcitabine (GEM) by encapsulating it into zeolitic imidazolate framework-8 nanoparticles (nZIF-8) that functionalized by linear (RGD) or cyclic homing peptide (iRGD). Therefore, this research focuses on the following objectives:

1. To encapsulate the GEM and surface functionalize the nZIF-8 using RGD or iRGD homing peptides.
2. To characterize the physicochemical properties of functionalized nanoparticles.
3. To evaluate the *in-vitro* GEM dissolution profile of functionalized nanoparticles.
4. To evaluate the *in-vitro* cellular uptake, cytotoxicity level, selectivity ability, and apoptotic induction of functionalized nanoparticles on lung cancer and normal cells lines.
5. To assess the permeability and potential toxicity developed by functionalized nanoparticles on healthy zebrafish embryos.

REFERENCES

- Abadeer, N. S., & Murphy, C. J. (2016). Recent Progress in Cancer Thermal Therapy Using Gold Nanoparticles. *Journal of Physical Chemistry C*, 120(9), 4691–4716.
- Ablooglu, A. J., Kang, J., Handin, R. I., Traver, D., & Shattil, S. J. (2007). The zebrafish vitronectin receptor: Characterization of integrin αV and $\beta 3$ expression patterns in early vertebrate development. *Developmental Dynamics*, 236(8), 2268–2276.
- Abramenko, N., Deyko, G., Abkhalimov, E., Isaeva, V., Pelgunova, L., Krysanov, E., & Kustov, L. (2021). Acute toxicity of cu-mof nanoparticles (Nanohkust-1) towards embryos and adult zebrafish. *International Journal of Molecular Sciences*, 22(11).
- Ahmad, S., Raemy, D. O., Loader, J. E., Kailey, J. M., Neeves, K. B., White, C. W., Ahmad, A., Gehr, P., & Rothen-Rutishauser, B. M. (2012). Interaction and localization of synthetic nanoparticles in healthy and cystic fibrosis airway epithelial cells: Effect of ozone exposure. *Journal of Aerosol Medicine and Pulmonary Drug Delivery*, 25(1), 7–15.
- Ahn, H. K., Jung, M., Sym, S. J., Shin, D. B., Kang, S. M., Kyung, S. Y., Park, J. W., Jeong, S. H., & Cho, E. K. (2014). A phase II trial of Cremorphor EL-free paclitaxel (Genexol-PM) and gemcitabine in patients with advanced non-small cell lung cancer. *Cancer Chemotherapy and Pharmacology*, 74(2), 277–282.
- Akbari, M., Ghasemzadeh, M. A., & Fadaeian, M. (2020). Synthesis and Application of ZIF-8 MOF Incorporated in a TiO₂@Chitosan Nanocomposite as a Strong Nanocarrier for the Drug Delivery of Acyclovir. *ChemistrySelect*, 5(46), 14564–14571.
- Aksorn, N., & Chanvorachote, P. (2019). Integrin as a molecular target for anti-cancer approaches in lung cancer. *Anticancer Research*, 39(2), 541–548.
- Ali, M. K., Saber, S. P., Taite, D. R., Emadi, S., & Irving, R. (2017). The Protective Layer of Zebrafish Embryo Changes Continuously with Advancing Age of Embryo Development (AGED). *Journal of Toxicology and Pharmacology*, 1(2), 009.
- Ali, S., van Mil, H. G. J., & Richardson, M. K. (2011). Large-Scale assessment of the zebrafish embryo as a possible predictive model in toxicity testing. *PLoS ONE*, 6(6).
- Allen, D. D., Caviedes, R., Cárdenas, A. M., Shimahara, T., Segura-Aguilar, J., & Caviedes, P. A. (2005). Cell lines as in vitro models for drug screening and toxicity studies. *Drug Development and Industrial Pharmacy*, 31(8), 757–768.

- Ananikov, V. P., Eremin, D. B., Yakukhnov, S. A., Dilman, A. D., Levin, V. V., Egorov, M. P., Karlov, S. S., Kustov, L. M., Tarasov, A. L., Greish, A. A., Shesterkina, A. A., Sakharov, A. M., Nysenko, Z. N., Sheremetev, A. B., Stakheev, A. Y., Mashkovsky, I. S., Sukhorukov, A. Y., Ioffe, S. L., Terent'ev, A. O., ... Nifantiev, N. E. (2017). Organic and hybrid systems: from science to practice. *Mendeleev Communications*, 27(5), 425–438.
- Asmawi, A. A., Salim, N., Ngan, C. L., Ahmad, H., Abdulmalek, E., Masarudin, M. J., & Abdul Rahman, M. B. (2018). Excipient selection and aerodynamic characterization of nebulized lipid-based nanoemulsion loaded with docetaxel for lung cancer treatment. *Drug Delivery and Translational Research*.
- Attia, M. F., Anton, N., Wallyn, J., Omran, Z., & Vandamme, T. F. (2019). An overview of active and passive targeting strategies to improve the nanocarriers efficiency to tumour sites. *Journal of Pharmacy and Pharmacology*, 71(8), 1185–1198.
- Babu, A., Amreddy, N., Muralidharan, R., Pathuri, G., Gali, H., Chen, A., Zhao, Y. D., Munshi, A., & Ramesh, R. (2017). Chemodrug delivery using integrin-targeted PLGA-Chitosan nanoparticle for lung cancer therapy. *Scientific Reports*, 7(1), 1–17.
- Bai, C., & Tang, M. (2020). Toxicological study of metal and metal oxide nanoparticles in zebrafish. *Journal of Applied Toxicology*, 40(1), 37–63.
- Barenholz, Y. (2012). Doxil® - The first FDA-approved nano-drug: Lessons learned. *Journal of Controlled Release*, 160(2), 117–134.
- Barosova, H., Meldrum, K., Karakocak, B. B., Balog, S., Doak, S. H., Petri-Fink, A., Clift, M. J. D., & Rothen-Rutishauser, B. (2021). Inter-laboratory variability of A549 epithelial cells grown under submerged and air-liquid interface conditions. *Toxicology in Vitro*, 75, 105178.
- Baskar, R., Lee, K. A., Yeo, R., & Yeoh, K. W. (2012). Cancer and radiation therapy: Current advances and future directions. *International Journal of Medical Sciences*, 9(3), 193–199.
- Baumann, A. E., Burns, D. A., Liu, B., & Thoi, V. S. (2019). Metal-organic framework functionalization and design strategies for advanced electrochemical energy storage devices. *Communications Chemistry*, 2(1), 1–14.
- Beg, S., Rahman, M., Jain, A., Saini, S., Midoux, P., Pichon, C., Ahmad, F. J., & Akhter, S. (2017). Nanoporous metal organic frameworks as hybrid polymer–metal composites for drug delivery and biomedical applications. *Drug Discovery Today*, 22(4), 625–637.

- Bender, E. (2014). The Dominant Malignancy. *Nature*, 513, S2–S3.
- Benyumov, A., Gurchich, V. J., Lis, L. G., Williams, B. W., & Kirstein, M. N. (2011). Combinatorial Pharmacologic Effects of Gemcitabine and its Metabolite dFdU. *ChemMedChem*, 6(3), 457–464.
- Berghmans, S., Butler, P., Goldsmith, P., Waldron, G., Gardner, I., Golder, Z., Richards, F. M., Kimber, G., Roach, A., Alderton, W., & Fleming, A. (2008). Zebrafish based assays for the assessment of cardiac, visual and gut function - potential safety screens for early drug discovery. *Journal of Pharmacological and Toxicological Methods*, 58(1), 59–68.
- Bernard F. Hoskins and Richard Robson. (1996). Infinite Polymeric Frameworks Consisting of Three Dimensionally Linked Rod-like Segment. *Acta Paediatrica, International Journal of Paediatrics*, 85(10), 1228–1231.
- Bertrand, N., Wu, J., Xu, X., Kamaly, N., & Farokhzad, O. C. (2014). The impact of passive and active targeting in the era of modern cancer biology. *Cancer Nanotechnology*, 6(2), 2–25.
- Bi, J., Lu, Y., Dong, Y., & Gao, P. (2018). Synthesis of Folic Acid-Modified DOX @ ZIF-8 Nanoparticles for Targeted Therapy of Liver Cancer. *Journal of Nanomaterials*, 2018, 2–7.
- Bianconi, D., Unseld, M., & Prager, G. W. (2016). Integrins in the spotlight of cancer. *International Journal of Molecular Sciences*, 17(12).
- Blanco, E., Shen, H., & Ferrari, M. (2015). Principles of nanoparticle design for overcoming biological barriers to drug delivery. *Nature Biotechnology*, 33(9), 941–951.
- Bo Yu, Heng Chiat Tai, W. X. (2010). Receptor-targeted nanocarriers for therapeutic delivery to cancer. *International Journal of Nanomedicine*, 27(7), 286–298.
- Braga, C. B., Kido, L. A., Lima, E. N., Lamas, C. A., Cagnon, V. H. A., Ornelas, C., & Pilli, R. A. (2020). Enhancing the Anticancer Activity and Selectivity of Goniotalamin Using pH-Sensitive Acetalated Dextran (Ac-Dex) Nanoparticles: A Promising Platform for Delivery of Natural Compounds. *ACS Biomaterials Science & Engineering*, 6(5), 2929–2942.
- Braunbeck, T., Böttcher, M., Hollert, H., Kosmehl, T., Lammer, E., Leist, E., Rudolf, M., & Seitz, N. (2005). Towards an alternative for the acute fish LC50 test in chemical assessment: The fish embryo toxicity test goes multi-species - An update. *Altex*, 22(2), 87–102.
- Browning, L. M., Lee, K. J., Huang, T., Nallathamby, P. D., Lowman, J. E., & Nancy Xu, X. H. (2009). Random walk of single gold nanoparticles in zebrafish embryos leading to stochastic toxic effects on embryonic developments. *Nanoscale*, 1(1), 138–152.

- Browning, R. J., Reardon, P. J. T., Parhizkar, M., Pedley, R. B., Edirisinghe, M., Knowles, J. C., & Stride, E. (2017). Drug Delivery Strategies for Platinum-Based Chemotherapy. *ACS Nano*, *11*(9), 8560–8578.
- Burdett, S., Burdett, S., Stephens, R., Stewart, L., Tierney, J., Auperin, A., Le Chevalier, T., Le Pechoux, C., Pignon, J. P., Arriagada, R., Higgins, J., Johnson, D., Van Meerbeeck, J., Parmar, M., Souhami, R., Bell, D., Cartei, G., Cormier, Y., Cullen, M., ... Williamson, I. (2008). Chemotherapy in addition to supportive care improves survival in advanced non-small-cell lung cancer: A systematic review and meta-analysis of individual patient data from 16 randomized controlled trials. *Journal of Clinical Oncology*, *26*(28), 4617–4625.
- Cai, W., Chu, C. C., Liu, G., & Wang, Y. X. J. (2015). Metal-Organic Framework-Based Nanomedicine Platforms for Drug Delivery and Molecular Imaging. *Small*, *11*(37), 4806–4822.
- Calderwood, D. A. (2004). Integrin activation. *Journal of Cell Science*, *117*(5), 657–666.
- Canossa, S., & Wuttke, S. (2020). Functionalization Chemistry of Porous Materials. *Advanced Functional Materials*, *30*(41).
- Cariati, E., Pizzotti, M., Roberto, D., Tessore, F., & Ugo, R. (2006). Coordination and organometallic compounds and inorganic-organic hybrid crystalline materials for second-order non-linear optics. *Coordination Chemistry Reviews*, *250*(11–12), 1210–1233.
- Carvalho, P. M., Felício, M. R., Santos, N. C., Gonçalves, S., & Domingues, M. M. (2018). Application of light scattering techniques to nanoparticle characterization and development. *Frontiers in Chemistry*, *6*(June), 1–17.
- Caswell, H., & Zarulli, V. (2018). Matrix methods in health demography: A new approach to the stochastic analysis of healthy longevity and DALYs. *Population Health Metrics*, *16*(1).
- Chan, L. L. Y., Rice, W. L., & Qiu, J. (2020). Observation and quantification of the morphological effect of trypan blue rupturing dead or dying cells. *PLoS ONE*, *15*(1), 1–17.
- Chen, H., Zhang, W., Zhu, G., Xie, J., & Chen, X. (2017). Rethinking cancer nanotheranostics. *Nature Reviews Materials*, *2*(May).
- Chen, T., Song, X., Gong, T., Fu, Y., Yang, L., Zhang, Z., & Gong, T. (2017). nRGD modified lycobetaine and octreotide combination delivery system to overcome multiple barriers and enhance anti-glioma efficacy. *Colloids and Surfaces B: Biointerfaces*, *156*, 330–339.
- Chen, W. H., Yu, X., Ceconello, A., Sohn, Y. S., Nechushtai, R., & Willner, I. (2017). Stimuli-responsive nucleic acid-functionalized metal-organic

- framework nanoparticles using pH- and metal-ion-dependent DNAzymes as locks. *Chemical Science*, 8(8), 5769–5780.
- Chen, Z. J., Yang, S. C., Liu, X. L., Gao, Y., Dong, X., Lai, X., Zhu, M. H., Feng, H. Y., Zhu, X. Di, Lu, Q., Zhao, M., Chen, H. Z., Lovell, J. F., & Fang, C. (2020). Nanobowl-Supported Liposomes Improve Drug Loading and Delivery. *Nano Letters*, 20(6), 4177–4187.
- Cheng, J., Flahaut, E., & Shuk, H. C. (2007). Effect of carbon nanotubes on developing zebrafish (*Danio rerio*) embryos. *Environmental Toxicology and Chemistry*, 26(4), 708–716.
- Cheok, C. F. (2012). Protecting normal cells from the cytotoxicity of chemotherapy. *Cell Cycle*, 11(12), 2227.
- Choi, J. Y., Huang, R., Uribe-romo, F. J., Chae, H. K., Park, K. S., Ni, Z., Co, A. P., Keeffe, M. O., & Yaghi, O. M. (2006). Exceptional chemical and thermal stability of zeolitic imidazolate frameworks. *103(27)*, 8–13.
- Chowdhuri, A. R., Laha, D., Pal, S., Karmakar, P., & Sahu, S. K. (2016). One-pot synthesis of folic acid encapsulated upconversion nanoscale metal organic frameworks for targeting, imaging and pH responsive drug release. *Dalton Transactions*, 45(45), 18120–18132.
- Chowdhury, M. A. (2017). Metal-organic-frameworks for biomedical applications in drug delivery, and as MRI contrast agents. *Journal of Biomedical Materials Research - Part A*, 105(4), 1184–1194.
- Ciccolini, J., Serdjebi, C., Peters, G. J., & Giovannetti, E. (2016). Pharmacokinetics and pharmacogenetics of Gemcitabine as a mainstay in adult and pediatric oncology: an EORTC-PAMM perspective. *Cancer Chemotherapy and Pharmacology*, 78(1), 1–12.
- Coudert, F. X. (2017). Molecular Mechanism of Swing Effect in Zeolitic Imidazolate Framework ZIF-8: Continuous Deformation upon Adsorption. *ChemPhysChem*, 18(19), 2732–2738.
- Cruz, C. S. Dela, Tanoue, L. T., & Matthay, R. A. (2011). Lung Cancer: Epidemiology, Etiology, and Prevention. *Clinics in Chest Medicine*, 32(4), 11.
- Cui, Y., Yue, Y., Qian, G., & Chen, B. (2012). Luminescent functional metal-organic frameworks. *Chemical Reviews*, 112(2), 1126–1162.
- Curran, W. J., Paulus, R., Langer, C. J., Komaki, R., Lee, J. S., Hauser, S., Movsas, B., Wasserman, T., Rosenthal, S. A., Gore, E., MacHtay, M., Sause, W., & Cox, J. D. (2011). Sequential vs concurrent chemoradiation for stage iii non-small cell lung cancer: Randomized phase III trial RTOG 9410. *Journal of the National Cancer Institute*, 103(19), 1452–1460.

- D'Souza, S. E., Ginsberg, M. H., & Plow, E. F. (1991). Arginyl-glycyl-aspartic acid (RGD): a cell adhesion motif. *Trends in Biochemical Sciences*, *16*(C), 246–250.
- Damjanovich, L., Albelda, S. M., Mette, S. A., & Buck, C. A. (1992). Distribution of integrin cell adhesion receptors in normal and malignant lung tissue. *American Journal of Respiratory Cell and Molecular Biology*, *6*(2), 197–206.
- Danhier, F., Breton, A. Le, & Pr eat, V. (2012). RGD-based strategies to target alpha(v) beta(3) integrin in cancer therapy and diagnosis. *Molecular Pharmaceutics*, *9*(11), 2961–2973.
- Danhier, F., Feron, O., & Pr eat, V. (2010). To exploit the tumor microenvironment: Passive and active tumor targeting of nanocarriers for anti-cancer drug delivery. *Journal of Controlled Release*, *148*(2), 135–146.
- de Moura Ferraz, L. R., Tabosa, A.  . G. A., da Silva Nascimento, D. D. S., Ferreira, A. S., de Albuquerque Wanderley Sales, V., Silva, J. Y. R., J nior, S. A., Rolim, L. A., de Souza Pereira, J. J., & Rolim-Neto, P. J. (2020). ZIF-8 as a promising drug delivery system for benzimidazole: development, characterization, in vitro dialysis release and cytotoxicity. *Scientific Reports*, *10*(1), 1–14.
- Deffieux, A., & Schappacher, M. (2012). Synthesis and Properties of Macrocyclic Polymers. In *Polymer Science: A Comprehensive Reference*, 10 Volume Set (Vol. 6). Elsevier B.V.
- Desai, N. (2012). Challenges in development of nanoparticle-based therapeutics. *AAPS Journal*, *14*(2), 282–295.
- Dhas, N. L., Ige, P. P., & Kudarha, R. R. (2015). Design, optimization and in-vitro study of folic acid conjugated-chitosan functionalized PLGA nanoparticle for delivery of bicalutamide in prostate cancer. *Powder Technology*, *283*, 234–245.
- Ding, S., Chen, G., Zhang, W., Xing, C., Xu, X., Xie, H., Lu, A., Chen, K., Guo, H., Ren, Z., Zheng, S., & Zhou, L. (2015). MRC-5 fibroblast-conditioned medium influences multiple pathways regulating invasion, migration, proliferation, and apoptosis in hepatocellular carcinoma. *Journal of Translational Medicine*, *13*(1), 1–13.
- Dings, R. P. M., Miller, M. C., Griffin, R. J., & Mayo, K. H. (2018). Galectins as molecular targets for therapeutic intervention. *International Journal of Molecular Sciences*, *19*(3), 1–22.
- Dong, K., Wang, Z., Zhang, Y., Ren, J., & Qu, X. (2018). Metal-Organic Framework-Based Nanoplatfor for Intracellular Environment-Responsive Endo/Lysosomal Escape and Enhanced Cancer Therapy. *ACS Applied Materials and Interfaces*, *10*(38), 31998–32005.

- Dong, K., Zhang, Y., Zhang, L., Wang, Z., Ren, J., & Qu, X. (2019a). Facile preparation of metal-organic frameworks-based hydrophobic anticancer drug delivery nanoplatform for targeted and enhanced cancer treatment. *Talanta*, *194*, 703–708.
- Dong, K., Zhang, Y., Zhang, L., Wang, Z., Ren, J., & Qu, X. (2019b). Facile preparation of metal-organic frameworks-based hydrophobic anticancer drug delivery nanoplatform for targeted and enhanced cancer treatment. *Talanta*, *194*, 703–708.
- Du, X. J., Wang, J. L., Iqbal, S., Li, H. J., Cao, Z. T., Wang, Y. C., Du, J. Z., & Wang, J. (2018). The effect of surface charge on oral absorption of polymeric nanoparticles. *Biomaterials Science*, *6*(3), 642–650.
- Eaton, P., Quaresma, P., Soares, C., Neves, C., de Almeida, M. P., Pereira, E., & West, P. (2017). A direct comparison of experimental methods to measure dimensions of synthetic nanoparticles. *Ultramicroscopy*, *182*, 179–190.
- Egeblad, M., Nakasone, E. S., & Werb, Z. (2010). Tumors as organs: Complex tissues that interface with the entire organism. *Developmental Cell*, *18*(6), 884–901. <https://doi.org/10.1016/j.devcel.2010.05.012>
- Ellis, L. M. (2006). The role of neuropilins in cancer. *Molecular Cancer Therapeutics*, *5*(5), 1099–1107.
- Esfahanian, M., Ghasemzadeh, M. A., & Razavian, S. M. H. (2019). Synthesis, identification and application of the novel metal-organic framework Fe₃O₄@PAA@ZIF-8 for the drug delivery of ciprofloxacin and investigation of antibacterial activity. *Artificial Cells, Nanomedicine and Biotechnology*, *47*(1), 2024–2030.
- Fan, G., Dundas, C. M., Zhang, C., Lynd, N. A., & Keitz, B. K. (2018). Sequence-Dependent Peptide Surface Functionalization of Metal-Organic Frameworks [Research-article]. *ACS Applied Materials and Interfaces*, *10*(22), 18601–18609.
- Fang, J., Nakamura, H., & Maeda, H. (2011). The EPR effect: Unique features of tumor blood vessels for drug delivery, factors involved, and limitations and augmentation of the effect. *Advanced Drug Delivery Reviews*, *63*(3), 136–151.
- Fantin, A., Vieira, J. M., Plein, A., Denti, L., Fruttiger, M., Pollard, J. W., & Ruhrberg, C. (2013). NRP1 acts cell autonomously in endothelium to promote tip cell function during sprouting angiogenesis. *Blood*, *121*(12), 2352–2362.
- Fenaroli, F., Westmoreland, D., Benjaminsen, J., Kolstad, T., Skjeldal, F. M., Meijer, A. H., Van Der Vaart, M., Ulanova, L., Roos, N., Nyström, B., Hildahl, J., & Griffiths, G. (2014). Nanoparticles as drug delivery system against tuberculosis in zebrafish embryos: Direct visualization and treatment. *ACS Nano*, *8*(7), 7014–7026.

- Feng, S. S., & Chien, S. (2003). Chemotherapeutic engineering: Application and further development of chemical engineering principles for chemotherapy of cancer and other diseases. *Chemical Engineering Science*, 58(18), 4087–4114.
- Férey, G., & Serre, C. (2009). Large breathing effects in three-dimensional porous hybrid matter: Facts, analyses, rules and consequences. *Chemical Society Reviews*, 38(5), 1380–1399.
- Forbes, B., Richer, N. H., & Buttini, F. (2015). Dissolution: A Critical Performance Characteristic of Inhaled Products? In *Pulmonary Drug Delivery* (Issue May 2015, pp. 223–240).
- Foroozandeh, P., & Aziz, A. A. (2018). Insight into Cellular Uptake and Intracellular Trafficking of Nanoparticles. *Nanoscale Research Letters*, 6, 1–12.
- Freund, R., Lächelt, U., Gruber, T., Rühle, B., & Wuttke, S. (2018). Multifunctional Efficiency: Extending the Concept of Atom Economy to Functional Nanomaterials. *ACS Nano*, 12(3), 2094–2105.
- Furukawa, H., Cordova, K. E., O’Keeffe, M., & Yaghi, O. M. (2013). The chemistry and applications of metal-organic frameworks. *Science*, 341(6149).
- Gaspar, M. M., Radomska, A., Gobbo, O. L., Bakowsky, U., Radomski, M. W., & Ehrhardt, C. (2012). Targeted delivery of transferrin-conjugated liposomes to an orthotopic model of lung cancer in nude rats. *Journal of Aerosol Medicine and Pulmonary Drug Delivery*, 25(6), 310–318.
- Gazdar, A. F., Girard, L., Lockwood, W. W., Lam, W. L., & Minna, J. D. (2010). Lung cancer cell lines as tools for biomedical discovery and research. *Journal of the National Cancer Institute*, 102(17), 1310–1321.
- Gibson, J. (2014). *A functional characterisation of the dimerisation motif in fibronectin, in vivo and in vitro.*
- Globocan. (2020). The Global Cancer Observatory - All cancers. *International Agent for Research on Cancer - WHO*, 419, 199–200.
- GLOBOCAN. (2020). *Globocan*. 418, 1–2. <https://gco.iarc.fr/today/data/factsheets/populations/458-malaysia-factsheets.pdf>
- Gohel, M. C., Sarvaiya, K. G., Shah, A. R., & Brahmabhatt, B. K. (2009). Mathematical approach for the assessment of similarity factor using a new scheme for calculating weight. *Indian Journal of Pharmaceutical Sciences*, 71(2), 142–144.

- Gothwal, A., Khan, I., & Gupta, U. (2016). Polymeric Micelles: Recent Advancements in the Delivery of Anticancer Drugs. *Pharmaceutical Research*, 33(1), 18–39.
- Grodzinski, P., Kircher, M., Goldberg, M., & Gabizon, A. (2019). Integrating Nanotechnology into Cancer Care. *ACS Nano*, 13(7), 7370–7376.
- Gustafson, H. H., Holt-Casper, D., Grainger, D. W., & Ghandehari, H. (2015). Nanoparticle uptake: The phagocyte problem. *Nano Today*, 10(4), 487–510.
- Gutiérrez-Lovera, C., Vázquez-Ríos, A. J., Guerra-Varela, J., Sánchez, L., & de la Fuente, M. (2017). The potential of zebrafish as a model organism for improving the translation of genetic anticancer nanomedicines. *Genes*, 8(12), 1–20.
- Han, H., Li, S., Zhong, Y., Huang, Y., Wang, K., Jin, Q., Ji, J., & Yao, K. (2021). Emerging pro-drug and nano-drug strategies for gemcitabine-based cancer therapy. *Asian Journal of Pharmaceutical Sciences*.
- Hao, J., Eken, Z. B., Mravak-stipetic, M., & Paveli, K. (2021). Effects of Zeolite as a Drug Delivery System on Cancer Therapy : A Systematic Review. *Molecules*.
- Haque, S., Pouton, C. W., McIntosh, M. P., Ascher, D. B., Keizer, D. W., Whittaker, M. R., & Kaminskis, L. M. (2020). The impact of size and charge on the pulmonary pharmacokinetics and immunological response of the lungs to PLGA nanoparticles after intratracheal administration to rats. *Nanomedicine: Nanotechnology, Biology, and Medicine*, 30(xxxx), 102291.
- Harrington, S. E., & Smith, T. J. (2008). The Role of Chemotherapy at the End of Life: “When is Enough, Enough?” *The Journal of the American Medical Association*, 299(22), 2667–2678.
- Haute, D. Van, & Berlin, J. M. (2017). Challenges in realizing selectivity for nanoparticle biodistribution and clearance: Lessons from gold nanoparticles. *Therapeutic Delivery*, 8(9), 763–774.
- Hayat, M. A. (2007). *Methods of Cancer Diagnosis , Therapy* , (Vol. 8).
- He, H., Liu, L., Morin, E. E., Liu, M., & Schwendeman, A. (2019). Survey of Clinical Translation of Cancer Nanomedicines - Lessons Learned from Successes and Failures. *Accounts of Chemical Research*, 52(9), 2673–2683.
- He, Y., Xiong, T., He, S., Sun, H., Huang, C., Ren, X., Wu, L., Patterson, L. H., & Zhang, J. (2021). Pulmonary Targeting Crosslinked Cyclodextrin Metal–Organic Frameworks for Lung Cancer Therapy. *Advanced Functional Materials*, 31(3), 1–13.
- Hill, A. J., Teraoka, H., Heideman, W., & Peterson, R. E. (2005). Zebrafish as a model vertebrate for investigating chemical toxicity. *Toxicological Sciences*,

86(1), 6–19.

- Hirschle, P., Preiß, T., Auras, F., Pick, A., Völkner, J., Valdepérez, D., Witte, G., Parak, W. J., Rädler, J. O., & Wuttke, S. (2016). Exploration of MOF nanoparticle sizes using various physical characterization methods – is what you measure what you get? *CrystEngComm*, 18(23), 4359–4368.
- Hong, M. (2007). Inorganic-organic hybrid coordination polymers: A new frontier for materials research. *Crystal Growth and Design*, 7(1), 10–14.
- How, S. H., Ng, T. H., Kuan, Y. C., Jamalludin, A. R., & Fauzi, A. R. (2015). Survival of lung cancer patients in a resource-limited country. *Asia-Pacific Journal of Clinical Oncology*, 11(3), 221–227.
- Howe, K., Clark, M. D., Torroja, C. F., Torrance, J., Berthelot, C., Muffato, M., Collins, J. E., Humphray, S., McLaren, K., Matthews, L., McLaren, S., Sealy, I., Caccamo, M., Churcher, C., Scott, C., Barrett, J. C., Koch, R., Rauch, G. J., White, S., ... Stemple, D. L. (2013). The zebrafish reference genome sequence and its relationship to the human genome. *Nature*, 496(7446), 498–503.
- Hu, C., Huang, Y., & Chen, Y. (2019). Targeted Modification of the Cationic Anticancer Peptide HPRP-A1 with iRGD to Improve Specificity, Penetration, and Tumor-Tissue Accumulation [Research-article]. *Molecular Pharmaceutics*, 16(2), 561–572.
- Hu, C. M. J., Zhang, L., Aryal, S., Cheung, C., Fang, R. H., & Zhang, L. (2011). Erythrocyte membrane-camouflaged polymeric nanoparticles as a biomimetic delivery platform. *Proceedings of the National Academy of Sciences of the United States of America*, 108(27), 10980–10985.
- Hu, Y. L., Qi, W., Han, F., Shao, J. Z., & Gao, J. Q. (2011). Toxicity evaluation of biodegradable chitosan nanoparticles using a zebrafish embryo model. *International Journal of Nanomedicine*, 6, 3351–3359.
- Huang, M. S., Liu, J. Y., Xia, X. B., Liu, Y. Z., Li, X., Yin, J. Y., Peng, J. B., Wu, L., Zhang, W., Zhou, H. H., & Liu, Z. Q. (2019). Hsa_circ_0001946 Inhibits Lung Cancer Progression and Mediates Cisplatin Sensitivity in Non-small Cell Lung Cancer via the Nucleotide Excision Repair Signaling Pathway. *Frontiers in Oncology*, 9(JUN), 1–12.
- Huxford, R. C., Della Rocca, J., & Lin, W. (2010). Metal-organic frameworks as potential drug carriers. *Current Opinion in Chemical Biology*, 14(2), 262–268.
- Huynh, E. (2015). Cancer nanomedicine: addressing the dark side of the enhanced permeability and retention effect. *Nanomedicine (Lond.)*, 10(13), 1993–1995.
- Hyun, S. M., Lee, J. H., Jung, G. Y., Kim, Y. K., Kim, T. K., Jeoung, S., Kwak, S. K., Moon, D., & Moon, H. R. (2016). Exploration of Gate-Opening and

- Breathing Phenomena in a Tailored Flexible Metal-Organic Framework. *Inorganic Chemistry*, 55(4), 1920–1925.
- ICH Q1A. (2003). Stability Testing of New Drug Substances and Products. *Handbook of Pharmaceutical Manufacturing Formulations, August 2003*, 31–40.
- ICH Q1A (R2). (2003). Guidance for Industry of New Drug Substances and Products Guidance for Industry Q1A (R2) Stability Testing of New Drug Substances and Products. *Food and Drug Administration, November*.
- ICH5(R3). (2020). International Council for Harmonisation of Technical Requirements for Pharmaceuticals for Human Use - DETECTION OF REPRODUCTIVE AND DEVELOPMENTAL TOXICITY FOR HUMAN PHARMACEUTICALS. *ICH Safety Guideline*, 41(February), 1–120.
- Iinuma, H., Maruyama, K., Okinaga, K., Sasaki, K., Sekine, T., Ishida, O., Ogiwara, N., Johkura, K., & Yonemura, Y. (2002). Intracellular targeting therapy of cisplatin-encapsulated transferrin-polyethylene glycol liposome on peritoneal dissemination of gastric cancer. *International Journal of Cancer*, 99(1), 130–137.
- Innes, E., Yiu, H. H. P., McLean, P., Brown, W., & Boyles, M. (2021). Simulated biological fluids—a systematic review of their biological relevance and use in relation to inhalation toxicology of particles and fibres. *Critical Reviews in Toxicology*, 51(3), 217–248.
- Izci, M., Maksoudian, C., Manshian, B. B., & Soenen, S. J. (2021). The Use of Alternative Strategies for Enhanced Nanoparticle Delivery to Solid Tumors. *Chemical Reviews*, 121(3), 1746–1803.
- Jakoby, J., Beuschlein, F., Mentz, S., Hantel, C., & Süß, R. (2015). Liposomal doxorubicin for active targeting: Surface modification of the nanocarrier evaluated in vitro and in vivo - challenges and prospects. *Oncotarget*, 6(41), 43698–43711.
- Jo, D. H., Kim, J. H., Lee, T. G., & Kim, J. H. (2015). Size, surface charge, and shape determine therapeutic effects of nanoparticles on brain and retinal diseases. *Nanomedicine: Nanotechnology, Biology, and Medicine*, 11(7), 1603–1611.
- Jurewicz, A., Ilyas, S., Uppal, J. K., Ivandic, I., Korsching, S., & Mathur, S. (2020). Evaluation of Magnetite Nanoparticle-Based Toxicity on Embryo-Larvae Stages of Zebrafish (*Danio rerio*). *ACS Applied Nano Materials*, 3(2), 1621–1629.
- Kan, P., Tsao, C.-W., Wang, A.-J., Su, W.-C., & Liang, H.-F. (2011). A Liposomal Formulation Able to Incorporate a High Content of Paclitaxel and Exert Promising Anticancer Effect. *Journal of Drug Delivery*, 2011, 1–9.

- Kang, S., Lee, S., & Park, S. (2020). iRGD peptide as a tumor-penetrating enhancer for tumor-targeted drug delivery. *Polymers*, 12(9), 1–27.
- Kapara, A., Brunton, V., Graham, D., & Faulds, K. (2020). Investigation of cellular uptake mechanism of functionalised gold nanoparticles into breast cancer using SERS †. *Chemical Science*, 11, 5819–5829.
- Kapp, T. G., Rechenmacher, F., Neubauer, S., Maltsev, O. V., Cavalcanti-Adam, E. A., Zarka, R., Reuning, U., Notni, J., Wester, H. J., Mas-Moruno, C., Spatz, J., Geiger, B., & Kessler, H. (2017). A comprehensive evaluation of the activity and selectivity profile of ligands for RGD-binding integrins. *Scientific Reports*, 7(August 2016), 1–13.
- Karimia, M., Bahramia, S., Ravaric, S. B., Zangabadd, P. S., Mirshekarie, H., Bozorgomidf, M., Shahrezag, S., Soria, M., & Hamblin, M. R. (2017). Albumin nanostructures as advanced drug delivery systems. *Physiology & Behavior*, 176(3), 139–148.
- Khatamian, M., Divband, B., & Farahmand-Zahed, F. (2016). Synthesis and characterization of Zinc (II)-loaded Zeolite/Graphene oxide nanocomposite as a new drug carrier. *Materials Science and Engineering C*, 66, 251–258.
- Kim, D. W., Kim, S. Y., Kim, H. K., Kim, S. W., Shin, S. W., Kim, J. S., Park, K., Lee, M. Y., & Heo, D. S. (2007). Multicenter phase II trial of Genexol-PM, a novel Cremophor-free, polymeric micelle formulation of paclitaxel, with cisplatin in patients with advanced non-small-cell lung cancer. *Annals of Oncology*, 18(12), 2009–2014.
- Kinoshita, Y., Matsubara, I., & Saito, Y. (1959). The Crystal of Bis (succinonitrilo) copper (I) Nitrate. *Bulletin of the Chemical Society of Japan*, 32(7), 741–747.
- Knights, O. B., & McLaughlan, J. R. (2018). Gold nanorods for light-based lung cancer theranostics. *International Journal of Molecular Sciences*, 19(11).
- Kobayashi, H., Watanabe, R., & Choyke, P. L. (2014). Improving conventional enhanced permeability and retention (EPR) effects; What is the appropriate target? *Theranostics*, 4(1), 81–89.
- Kush, P., Bajaj, T., Kaur, M., Madan, J., Jain, U. K., Kumar, P., Deep, A., & Kim, K. H. (2020). Biodistribution and Pharmacokinetic Study of Gemcitabine Hydrochloride Loaded Biocompatible Iron-Based Metal Organic Framework. *Journal of Inorganic and Organometallic Polymers and Materials*, 30(8), 2827–2841.
- Lackey, A., & Donington, J. (2013). Surgical management of lung cancer. *Seminars in Interventional Radiology*, 30(2), 133–140.
- Läecheit, U., Röder, R., Preiß, T., Hirschle, P., Steinborn, B., Zimpel, A., Höhn, M., Rädler, J. O., Bein, T., Wagner, E., & Wuttke, S. (2017). Multifunctional Nanoparticles by Coordinative Self-Assembly of His-Tagged Units with

Metal-Organic Frameworks. *Journal of the American Chemical Society*, 139(6), 2359–2368.

- Lammer, E., Carr, G. J., Wendler, K., Rawlings, J. M., Belanger, S. E., & Braunbeck, T. (2009). Is the fish embryo toxicity test (FET) with the zebrafish (*Danio rerio*) a potential alternative for the fish acute toxicity test? *Comparative Biochemistry and Physiology - C Toxicology and Pharmacology*, 149(2), 196–209.
- Langheinrich, U. (2003). Zebrafish: A new model on the pharmaceutical catwalk. *BioEssays*, 25(9), 904–912.
- Lee, X. J., Lim, H. N., Gowthaman, N. S. K., Rahman, M. B. A., Che Abdullah, C. A., & Muthoosamy, K. (2020). In-situ surface functionalization of superparamagnetic reduced graphene oxide – Fe₃O₄ nanocomposite via *Ganoderma lucidum* extract for targeted cancer therapy application. *Applied Surface Science*, 512(October 2019), 145738.
- Lewis, J. M., Truong, T. N., & Schwartz, M. A. (2002). *Integrins regulate the apoptotic response to DNA damage through modulation of p53*. 99(6).
- Li, H., Jin, H., Wan, W., Wu, C., & Wei, L. (2018). Cancer nanomedicine: Mechanisms, obstacles and strategies. *Nanomedicine*, 13(13), 1639–1656. <https://doi.org/10.2217/nnm-2018-0007>
- Li, J. R., Kuppler, R. J., & Zhou, H. C. (2009). Selective gas adsorption and separation in metal-organic frameworks. *Chemical Society Reviews*, 38(5), 1477–1504.
- Li, N., Qiu, S., Fang, Y., Wu, J., & Li, Q. (2021). Comparison of Linear vs. Cyclic RGD Pentapeptide Interactions with Integrin $\alpha\beta3$ by Molecular Dynamics Simulations. *Biology*, 10(7), 688.
- Li, S., Wang, K., Shi, Y., Cui, Y., Chen, B., He, B., Dai, W., Zhang, H., Wang, X., Zhong, C., Wu, H., Yang, Q., & Zhang, Q. (2016). Novel Biological Functions of ZIF-NP as a Delivery Vehicle: High Pulmonary Accumulation, Favorable Biocompatibility, and Improved Therapeutic Outcome. *Advanced Functional Materials*, 26(16), 2715–2727.
- Li, X., Salzano, G., Qiu, J., Menard, M., Berg, K., Theodossiou, T., Ladavière, C., & Gref, R. (2020). Drug-Loaded Lipid-Coated Hybrid Organic-Inorganic “Stealth” Nanoparticles for Cancer Therapy. *Frontiers in Bioengineering and Biotechnology*, 8(September), 1–12.
- Li, Y., Zheng, Y., Lai, X., Chu, Y., & Chen, Y. (2018). Biocompatible surface modification of nano-scale zeolitic imidazolate frameworks for enhanced drug delivery. *RSC Advances*, 8(42), 23623–23628.
- Lin, Y., Zhong, Y., Chen, Y., Li, L., Chen, G., Zhang, J., Li, P., Zhou, C., Sun, Y., Ma, Y., Xie, Z., & Liao, Q. (2020). Ligand-modified erythrocyte membrane-cloaked metal-organic framework nanoparticles for targeted antitumor therapy. *Molecular Pharmaceutics*, 17(9), 3328–3341.

- Liu, B. R., Chan, M.-H., Chen, H.-H., Lo, S.-Y., Huang, Y.-W., & Lee, H.-J. (2013). Effects of Surface Charge and Particle Size of Cell-Penetrating Peptide / Nanoparticle Complexes on Cellular Internalization. *Cell Membrane*, 44.
- Liu, J., Yu, M., Zhou, C., & Zheng, J. (2013). Renal clearable inorganic nanoparticles: A new frontier of bionanotechnology. *Materials Today*, 16(12), 477–486.
- Liu, S. (2009). Radiolabeled Cyclic RGD Peptides as Integrin $\alpha v\beta 3$ -Targeted.pdf. *Bioconjugate Chemistry*, 20(12), 2199–2213.
- Liu, Shouchun, Thomas, S. M., Woodside, D. G., Rose, D. M., Kiosses, W. B., Pfaff, M., & Ginsberg, M. H. (1999). *Vacuolar protein sorting Binding of paxillin to $\alpha 4$ integrins modifies integrin-dependent biological responses.* 402(December).
- Liu, X., Zhu, X., Qi, X., Meng, X., & Xu, K. (2021). Co-Administration of iRGD with Sorafenib-Loaded Iron-Based Metal-Organic Framework as a Targeted Ferroptosis Agent for Liver Cancer Therapy. *International Journal of Nanomedicine*, Volume 16, 1037–1050.
- Luo, B. H., Carman, C. V., & Springer, T. A. (2007). Structural basis of integrin regulation and signaling. *Annual Review of Immunology*, 25, 619–647.
- Lutz, S. T., Jones, J., & Chow, E. (2014). Role of radiation therapy in palliative care of the patient with cancer. *Journal of Clinical Oncology*, 32(26), 2913–2919.
- Luzuriaga, M. A., Benjamin, C. E., Gaertner, M. W., Lee, H., Herbert, F. C., Mallick, S., & Gassensmith, J. J. (2019). ZIF-8 degrades in cell media , serum , and some — but not all — common laboratory bu ff ers. *Supramolecular Chemistry*, 00(00), 1–6.
- Lyu, F., Zhang, Y., Zare, R. N., Ge, J., & Liu, Z. (2014). *One-Pot Synthesis of Protein-Embedded Metal – Organic Frameworks with Enhanced Biological Activities.*
- M. Christopher, A. M. L. S. (2016). Principles of nanoparticle design for overcoming biological. *Physiology & Behavior*, 176(1), 100–106.
- Malatesta, M. (2016). Transmission electron microscopy for nanomedicine: Novel applications for long-established techniques. *European Journal of Histochemistry*, 60(4), 8–12.
- Maleki, A., Shahbazi, M. A., Alinezhad, V., & Santos, H. A. (2020). The Progress and Prospect of Zeolitic Imidazolate Frameworks in Cancer Therapy, Antibacterial Activity, and Biomineralization. *Advanced Healthcare Materials*, 9(12), 1–42.

- Manegold, C. (2004). Gemcitabine (Gemzar®) in non-small cell lung cancer. *Expert Review of Anticancer Therapy*, 4(3), 345–360.
- Masood, F. (2016). Polymeric nanoparticles for targeted drug delivery system for cancer therapy. *Materials Science and Engineering C*, 60, 569–578.
- Mendoza, A., Izadifar, M., & Chen, X. (2017). Evaluation of PBS Treatment and PEI Coating Effects on Surface Morphology and Cellular Response of 3D-Printed Alginate Scaffolds. *Journal of Functional Biomaterials*, 8(48), 1–13.
- Meng, H., Leong, W., Leong, K. W., Chen, C., & Zhao, Y. (2018). Walking the line: The fate of nanomaterials at biological barriers. *Biomaterials*, 174, 41–53.
- Mezu-Ndubuisi, O. J., & Maheshwari, A. (2020). The role of integrins in inflammation and angiogenesis. *Pediatric Research*, June 2020.
- Mi, H.-Y., Xin, J., Thomsom, J. A., & Turng, L.-S. (2018). Promoting endothelial cell affinity and antithrombogenicity of polytetrafluoroethylene (PTFE) by mussel-inspired modification and RGD/heparin grafting. *Journal of Materials Chemistry B*, 6(21), 3485–3485.
- Molina, J. R., Yang, P., Cassivi, S. D., Schild, S. E., & Adjei, A. A. (2008). Non-small cell lung cancer: Epidemiology, risk factors, treatment, and survivorship. *Mayo Clinic Proceedings*, 83(5), 584–594.
- Mphuthi, L. E., Erasmus, E., & Langner, E. H. G. (2021). Metal Exchange of ZIF-8 and ZIF-67 Nanoparticles with Fe(II) for Enhanced Photocatalytic Performance. *ACS Omega*, 6(47), 31632–31645.
- Muhamad, N., Plengsuriyakarn, T., & Na-Bangchang, K. (2018). Application of active targeting nanoparticle delivery system for chemotherapeutic drugs and traditional/herbal medicines in cancer therapy: A systematic review. *International Journal of Nanomedicine*, 13, 3921–3935.
- Muthukumar, T., Prabhavathi, S., Chamundeeswari, M., & Sastry, T. P. (2014). Bio-modified carbon nanoparticles loaded with methotrexate Possible carrier for anticancer drug delivery. *Materials Science and Engineering C*, 36(1), 14–19.
- MySCan. (2018). MALAYSIAN STUDY ON CANCER SURVIVAL (MySCan). In *National Cancer Institute, Ministry of Health Malaysia* (Vol. 4).
- Naderinezhad, S., Amoabediny, G., & Haghirsadat, F. (2017). Co-delivery of hydrophilic and hydrophobic anticancer drugs using biocompatible pH-sensitive lipid-based nano-carriers for multidrug-resistant cancers. *RSC Advances*, 7(48), 30008–30019.
- Nasir, I., Lundqvist, M., & Cabaleiro-Lago, C. (2015). Size and surface chemistry of nanoparticles lead to a variant behavior in the unfolding dynamics of human carbonic anhydrase. *Nanoscale*, 7(41), 17504–17515.

- Ni, K., Lan, G., Veroneau, S. S., Duan, X., Song, Y., & Lin, W. (2018). Nanoscale metal-organic frameworks for mitochondria-targeted radiotherapy-radiodynamic therapy. *Nature Communications*, 9(1).
- Nieberler, M., Reuning, U., Reichart, F., Notni, J., Wester, H. J., Schwaiger, M., Weinmüller, M., Räder, A., Steiger, K., & Kessler, H. (2017). Exploring the role of RGD-recognizing integrins in cancer. *Cancers*, 9(9), 1–33.
- Niu, Y., Yu, M., Meka, A., Liu, Y., Zhang, J., Yang, Y., & Yu, C. (2015). Understanding the contribution of surface roughness and hydrophobic modification of silica nanoparticles to enhanced therapeutic protein delivery. *Journal of Materials Chemistry B*, 4(2), 212–219.
- Oberdörster, G., Oberdörster, E., & Oberdörster, J. (2005). Nanotoxicology: An emerging discipline evolving from studies of ultrafine particles. *Environmental Health Perspectives*, 113(7), 823–839.
- OECD. (2013). *Guidelines for the testing chemicals, Section 2: Effects on biotic systems, Test No. 236: Fish, Embryo Acute Toxicity (FET) Test*. July, 1–22.
- Outtandy, P., Russell, C., Kleta, R., & Bockenbauer, D. (2019). Zebrafish as a model for kidney function and disease. *Pediatric Nephrology*, 34(5), 751–762.
- Overchuk, M., & Zheng, G. (2018). Overcoming obstacles in the tumor microenvironment: Recent advancements in nanoparticle delivery for cancer theranostics. *Biomaterials*, 156, 217–237.
- Paarakh, M. P., Jose, P. A. N. I., Setty, C. M., & Peter, G. V. (2019). Release Kinetics – Concepts and Applications. *International Journal of Pharmacy Research & Technology*, 8(1), 12–20.
- Pang, H. B., Braun, G. B., Friman, T., Aza-Blanc, P., Ruidiaz, M. E., Sugahara, K. N., Teesalu, T., & Ruoslahti, E. (2014). An endocytosis pathway initiated through neuropilin-1 and regulated by nutrient availability. *Nature Communications*, 5.
- Pang, J., Xing, H., Sun, Y., Feng, S., & Wang, S. (2020). Non-small cell lung cancer combination therapy: Hyaluronic acid modified, epidermal growth factor receptor targeted, pH sensitive lipid-polymer hybrid nanoparticles for the delivery of erlotinib plus bevacizumab. *Biomedicine and Pharmacotherapy*, 125(December 2019), 109861.
- Park, K. S., Ni, Z., Côté, A. P., Choi, J. Y., Huang, R., Uribe-Romo, F. J., Chae, H. K., O’Keeffe, M., & Yaghi, O. M. (2006). Exceptional chemical and thermal stability of zeolitic imidazolate frameworks. *Proceedings of the National Academy of Sciences of the United States of America*, 103(27), 10186–10191.

- Parodi, A., Quattrocchi, N., Van De Ven, A. L., Chiappini, C., Evangelopoulos, M., Martinez, J. O., Brown, B. S., Khaled, S. Z., Yazdi, I. K., Enzo, M. V., Isenhardt, L., Ferrari, M., & Tasciotti, E. (2013). Synthetic nanoparticles functionalized with biomimetic leukocyte membranes possess cell-like functions. *Nature Nanotechnology*, 8(1), 61–68.
- Patton, E. E., Zon, L. I., & Langenau, D. M. (2021). Zebrafish disease models in drug discovery: from preclinical modelling to clinical trials. *Nature Reviews Drug Discovery*, 20(8), 611–628.
- Peller, M., Böll, K., Zimpel, A., & Wuttke, S. (2018). Metal-organic framework nanoparticles for magnetic resonance imaging. *Inorganic Chemistry Frontiers*, 5(8), 1760–1779.
- Pensado-López, A., Fernández-Rey, J., Reimunde, P., Crecente-Campo, J., Sánchez, L., & Torres Andón, F. (2021). Zebrafish models for the safety and therapeutic testing of nanoparticles with a focus on macrophages. *Nanomaterials*, 11(7), 1–34.
- Pfister, D. G., Johnson, D. H., Azzoli, C. G., Sause, W., Smith, T. J., Baker, S., Olak, J., Stover, D., Strawn, J. R., Turrisi, A. T., & Somerfield, M. R. (2004). American Society of Clinical Oncology treatment of unresectable non-small-cell lung cancer guideline: Update 2003. *Journal of Clinical Oncology*, 22(2), 330–353.
- Pietrojusti, A., Campagnolo, L., & Fadeel, B. (2013). Interactions of engineered nanoparticles with organs protected by internal biological barriers. *Small*, 9(9–10), 1557–1572.
- Pisano, C., Cecere, S. C., Di Napoli, M., Cavaliere, C., Tambaro, R., Facchini, G., Scaffa, C., Losito, S., Pizzolorusso, A., & Pignata, S. (2013). Clinical Trials with Pegylated Liposomal Doxorubicin in the Treatment of Ovarian Cancer. *Journal of Drug Delivery*, 2013, 1–12.
- Prendiville, J., & O'Brien, M. (1997). Gemcitabine. *British Journal of Hospital Medicine*, 57(8), 405–409. <https://doi.org/10.2165/00128415-201214300-00064>
- Pridgen, E. M., Alexis, F., Kuo, T. T., Levy-Nissenbaum, E., Karnik, R., Blumberg, R. S., Langer, R., & Farokhzad, O. C. (2013). Transepithelial transport of Fc-targeted nanoparticles by the neonatal Fc receptor for oral delivery. *Science Translational Medicine*, 5(213).
- Proenza, Y. G., & Longo, R. L. (2020). Simulation of the Adsorption and Release of Large Drugs by ZIF-8. *Journal of Chemical Information and Modeling*, 60(2), 644–652.
- Psarra, E., König, U., Müller, M., Bittrich, E., Eichhorn, K. J., Welzel, P. B., Stamm, M., & Uhlmann, P. (2017). In Situ Monitoring of Linear RGD-Peptide Bioconjugation with Nanoscale Polymer Brushes. *ACS Omega*, 2(3), 946–958.

- Pyati, U. J., Look, A. T., & Hammerschmidt, M. (2007). Zebrafish as a powerful vertebrate model system for in vivo studies of cell death. *Seminars in Cancer Biology*, 17(2), 154–165.
- Ramos, A. P., Cruz, M. A. E., Tovani, C. B., & Ciancaglini, P. (2017). Biomedical applications of nanotechnology. *Biophysical Reviews*, 9(2), 79–89.
- Rasooly, R. S., Henken, D., Freeman, N., Tompkins, L., Badman, D., Briggs, J., & Hewitt, A. T. (2003). Genetic and Genomic Tools for Zebrafish Research: The NIH Zebrafish Initiative. *Developmental Dynamics*, 228(3), 490–496.
- Rawson, D. M., Zhang, T., Kalicharan, D., & Jongebloed, W. L. (2000). Field emission scanning electron microscopy and transmission electron microscopy studies of the chorion, plasma membrane and syncytial layers of the gastrula-stage embryo of the zebrafish *Brachydanio rerio*: A consideration of the structural and functional. *Aquaculture Research*, 31(3), 325–336.
- Rennekamp, A. J., & Peterson, R. T. (2015). 15 Years of Zebrafish Chemical Screening. *Current Opinion in Chemical Biology*, 24, 58–70.
- Riss, T. L., Moravec, R. A., Niles, A. L., Duellman, S., Benink, H. A., Worzella, T. J., & Minor, L. (2004). Cell Viability Assays. *Assay Guidance Manual*, Md, 1–36.
- Rizvi, S. A. A., & Saleh, A. M. (2018). Applications of nanoparticle systems in drug delivery technology. *Saudi Pharmaceutical Journal*, 26(1), 64–70.
- Rocca, J. Della, & Lin, W. (2010). Nanoscale metal-organic frameworks: Magnetic resonance imaging contrast agents and beyond. *European Journal of Inorganic Chemistry*, 24, 3725–3734.
- Röder, R., Preiß, T., Hirschle, P., Steinborn, B., Zimpel, A., Höhn, M., Rädler, J. O., Bein, T., Wagner, E., Wuttke, S., & Lächelt, U. (2017). Multifunctional Nanoparticles by Coordinative Self-Assembly of His-Tagged Units with Metal-Organic Frameworks. *Journal of the American Chemical Society*, 139(6), 2359–2368.
- Rodriguez, P. L., Harada, T., Christian, D. A., Pantano, D. A., Tsai, R. K., & Discher, D. E. (2013). Minimal “self” peptides that inhibit phagocytic clearance and enhance delivery of nanoparticles. *Science*, 339(6122), 971–975.
- Roger W. Russell. (1991). Essential roles for animal models in understanding human toxicities. *Neuroscience & Biobehavioral Reviews*, 7–11.
- Rogge, S. M. J., Bavykina, A., Hajek, J., Garcia, H., Olivos-Suarez, A. I., Sepúlveda-Escribano, A., Vimont, A., Clet, G., Bazin, P., Kapteijn, F., Daturi, M., Ramos-Fernandez, E. V., Llabrés Xamena, F. X. I., Van Speybroeck, V., & Gascon, J. (2017). Metal-organic and covalent organic frameworks as single-site catalysts. *Chemical Society Reviews*, 46(11),

- Ruoslahti, E. (2012). Peptides as targeting elements and tissue penetration devices for nanoparticles. *Advanced Materials*, 24(28), 3747–3756.
- Sarin, H., Kanevsky, A. S., Wu, H., Sousa, A. A., Wilson, C. M., Aronova, M. A., Griffiths, G. L., Leapman, R. D., & Vo, H. Q. (2009). Physiologic upper limit of pore size in the blood-tumor barrier of malignant solid tumors. *Journal of Translational Medicine*, 7, 1–13.
- Senapati, S., Mahanta, A. K., Kumar, S., & Maiti, P. (2018). Controlled drug delivery vehicles for cancer treatment and their performance. *Signal Transduction and Targeted Therapy*, 3(1), 1–19.
- Shaikh, A., Kohale, K., Ibrahim, M., & Khan, M. (2019). Teratogenic effects of aqueous extract of *Ficus glomerata* leaf during embryonic development in zebrafish (*Danio rerio*). *Journal of Applied Pharmaceutical Science*, 9(5), 107–111.
- Shen, J., Meng, Q., Sui, H., Yin, Q., Zhang, Z., Yu, H., & Li, Y. (2013). iRGD Conjugated TPGS Mediates Codelivery of Paclitaxel and Survivin shRNA for the Reversal of Lung Cancer Resistance. *Molecular Pharma*.
- Sieber, S., Grossen, P., Bussmann, J., Campbell, F., Kros, A., Witzigmann, D., & Huwyler, J. (2019). Zebrafish as a preclinical in vivo screening model for nanomedicines. *Advanced Drug Delivery Reviews*, 151–152, 152–168.
- Singh, A. P., Biswas, A., Shukla, A., & Maiti, P. (2019). Targeted therapy in chronic diseases using nanomaterial-based drug delivery vehicles. *Signal Transduction and Targeted Therapy*, 4(1), 1–21.
- Soo Choi, H., Liu, W., Misra, P., Tanaka, E., Zimmer, J. P., Iyiti Ipe, B., Bawendi, M. G., & Frangioni, J. V. (2007). Renal clearance of quantum dots. *Nature Biotechnology*, 25(10), 1165–1170.
- Steichen, S. D., Caldorera-Moore, M., & Peppas, N. A. (2013). A review of current nanoparticle and targeting moieties for the delivery of cancer therapeutics. *European Journal of Pharmaceutical Sciences*, 48(3), 416–427.
- Strebhardt, K., & Ullrich, A. (2008). Paul Ehrlich's magic bullet concept: 100 Years of progress. *Nature Reviews Cancer*, 8(6), 473–480.
- Su, C., Li, J., Zhang, L., Wang, H., & Wang, F. (2020). *The Biological Functions and Clinical Applications of Integrins in Cancers*. 11(September), 1–14.
- Sugahara, K. N., Teesalu, T., Karmali, P. P., Kotamraju, V. R., Agemy, L., Girard, O. M., Hanahan, D., Mattrey, R. F., & Ruoslahti, E. (2009). Tissue-Penetrating Delivery of Compounds and Nanoparticles into Tumors. *Cancer Cell*, 16(6), 510–520.

- Sugahara, K. N., Teesalu, T., Prakash Karmali, P., Ramana Kotamraju, V., Agemy, L., Greenwald, D. R., & Ruoslahti, E. (2010). Coadministration of a tumor-penetrating peptide enhances the efficacy of cancer drugs. *Science*, *328*(5981), 1031–1035.
- Sun, C. Y., Qin, C., Wang, X. L., Yang, G. S., Shao, K. Z., Lan, Y. Q., Su, Z. M., Huang, P., Wang, C. G., & Wang, E. B. (2012). Zeolitic imidazolate framework-8 as efficient pH-sensitive drug delivery vehicle. *Dalton Transactions*, *41*(23), 6906–6909.
- Sun, T., Zhang, Y. S., Pang, B., Hyun, D. C., Yang, M., & Xia, Y. (2014). Engineered nanoparticles for drug delivery in cancer therapy. *Angewandte Chemie - International Edition*, *53*(46), 12320–12364.
- Takada, Y., Ye, X., & Simon, S. (2007). The integrins. *Genome Biology*, *8*(5). <https://doi.org/10.1186/gb-2007-8-5-215>
- Tamam, H., Park, J., Gadalla, H. H., Masters, A. R., Abdel-Aleem, J. A., Abdelrahman, S. I., Abdelrahman, A. A., Lyle, L. T., & Yeo, Y. (2019). Development of Liposomal Gemcitabine with High Drug Loading Capacity. *Molecular Pharmaceutics*, *16*, 2858–2871.
- Teesalu, T., Sugahara, K. N., & Ruoslahti, E. (2013). Tumor-penetrating peptides. *Frontiers in Oncology*, *3* AUG(August), 1–8.
- Temming, K., Schiffelers, R. M., Molema, G., & Kok, R. J. (2005). RGD-based strategies for selective delivery of therapeutics and imaging agents to the tumour vasculature. *Drug Resistance Updates*, *8*(6), 381–402.
- Thanh, N. T. K., Maclean, N., & Mahiddine, S. (2014). Mechanisms of nucleation and growth of nanoparticles in solution. *Chemical Reviews*, *114*(15), 7610–7630.
- Tibbetts, I., & Kostakis, G. E. (2020). Recent bio-advances in metal-organic frameworks. *Molecules*, *25*(6), 1–32.
- Tiwari, A., Singh, A., Garg, N., & Randhawa, J. K. (2017a). Curcumin encapsulated zeolitic imidazolate frameworks as stimuli responsive drug delivery system and their interaction with biomimetic environment. *Scientific Reports*, *7*(1), 1–12.
- Tiwari, A., Singh, A., Garg, N., & Randhawa, J. K. (2017b). Curcumin encapsulated zeolitic imidazolate frameworks as stimuli responsive drug delivery system and their interaction with biomimetic environment. *Scientific Reports*, *7*(1), 1–12.
- Tobin, D. M., May, R. C., & Wheeler, R. T. (2012). Zebrafish: A see-through host and a fluorescent toolbox to probe host-pathogen interaction. *PLoS Pathogens*, *8*(1), 8–11.

- Torchilin, V. (2011). Tumor delivery of macromolecular drugs based on the EPR effect. *Advanced Drug Delivery Reviews*, 63(3), 131–135.
- USP 35. (2012). (1090) Assessment of drug performance - bioavailability, bioequivalence, and dissolution. *United State Pharmacopeia (USP 35)*, 669–675.
- Van Agthoven, J. F., Xiong, J. P., Alonso, J. L., Rui, X., Adair, B. D., Goodman, S. L., & Arnaout, M. A. (2014). Structural basis for pure antagonism of integrin $\alpha V\beta 3$ by a high-affinity form of fibronectin. *Nature Structural and Molecular Biology*, 21(4), 383–388.
- Vasconcelos, I. B., Da Silva, T. G., Militão, G. C. G., Soares, T. A., Rodrigues, N. M., Rodrigues, M. O., Da Costa, N. B., Freire, R. O., & Junior, S. A. (2012). Cytotoxicity and slow release of the anti-cancer drug doxorubicin from ZIF-8. *RSC Advances*, 2(25), 9437–9442.
- Velásquez-Hernández, M. D. J., Ricco, R., Carraro, F., Limpoco, F. T., Linares-Moreau, M., Leitner, E., Wiltche, H., Rattenberger, J., Schröttner, H., Frühwirt, P., Stadler, E. M., Gescheidt, G., Amenitsch, H., Doonan, C. J., & Falcaro, P. (2019). Degradation of ZIF-8 in phosphate buffered saline media. *CrystrEngComm*, 21(31), 4538–4544.
- Veldman, M. B., & Lin, S. (2008). Zebrafish as a Developmental Model Organism for Pediatric Research. *Pediatric Research*, 64(5), 470–476.
- Velozo-Sá, V. S., Pereira, L. R., Lima, A. P., Mello-Andrade, F., Rezende, M. R. M., Goveia, R. M., Pires, W. C., Silva, M. M., Oliveira, K. M., Ferreira, A. G., Ellena, J., Deflon, V. M., Grisolia, C. K., Batista, A. A., & Silveira-Lacerda, E. P. (2019). In vitro cytotoxicity and in vivo zebrafish toxicity evaluation of Ru(II)/2-mercaptopyrimidine complexes. *Dalton Transactions*, 48(18), 6026–6039.
- Vokes, E. E., & Bitran, J. D. (1994). Non-small-cell lung cancer: Toward the next plateau. *Chest*, 106(3), 659–661.
- von Roemeling, C., Jiang, W., Chan, C. K., Weissman, I. L., & Kim, B. Y. S. (2017). Breaking Down the Barriers to Precision Cancer Nanomedicine. *Trends in Biotechnology*, 35(2), 159–171.
- Wahgiman, N. A., Salim, N., Rahman, M. B. A., & Ashari, S. E. (2019). Optimization of nanoemulsion containing gemcitabine and evaluation of its cytotoxicity towards human fetal lung fibroblast (MRC5) and human lung carcinoma (A549) cells. *International Journal of Nanomedicine*, 14, 7323–7338.
- Wang, J., Li, W., Lu, Z., Zhang, L., Hu, Y., Li, Q., Du, W., Feng, X., Jia, H., & Liu, B. F. (2017). The use of RGD-engineered exosomes for enhanced targeting ability and synergistic therapy toward angiogenesis. *Nanoscale*, 9(40), 15598–15605.

- Wang, Q., Sun, Y., Li, S., Zhang, P., & Yao, Q. (2020). Synthesis and modification of ZIF-8 and its application in drug delivery and tumor therapy. *RSC Advances*, *10*(62), 37600–37620.
- Wang, R., Liu, J., Liu, Y., Zhong, R., Yu, X., Liu, Q., Zhang, L., Lv, C., Mao, K., & Tang, P. (2020). The cell uptake properties and hyperthermia performance of Zn_{0.5}Fe_{2.5}O₄/SiO₂ nanoparticles as magnetic hyperthermia agents. *Royal Society Open Science*, *7*(1).
- Wang, S., Chen, Y., Wang, S., Li, P., Mirkin, C. A., & Farha, O. K. (2019). DNA-Functionalized Metal-Organic Framework Nanoparticles for Intracellular Delivery of Proteins. *Journal of the American Chemical Society*, *141*(6), 2215–2219.
- Wang, S., McGuirk, C. M., d'Aquino, A., Mason, J. A., & Mirkin, C. A. (2018). Metal-Organic Framework Nanoparticles. *Advanced Materials*, *30*(37), 1–14.
- Wang, X., Chen, X. Z., Alcântara, C. C. J., Sevim, S., Hoop, M., Terzopoulou, A., de Marco, C., Hu, C., de Mello, A. J., Falcaro, P., Furukawa, S., Nelson, B. J., Puigmartí-Luis, J., & Pané, S. (2019). MOFBOTS: Metal-Organic-Framework-Based Biomedical Microrobots. *Advanced Materials*, *1901592*, 2–8.
- Wang, X. G., Zhang, X. Z., Dong, Z. Y., Cheng, H., Wan, S. S., Chen, W. H., Zou, M. Z., Huo, J. W., & Deng, H. X. (2015). A multifunctional metal-organic framework based tumor targeting drug delivery system for cancer therapy. *Nanoscale*, *7*(38), 16061–16070.
- WHO. (2015). *April*.
- Wu, C., You, J., & Wang, X. (2018). Thermal decomposition mechanism and kinetics of gemcitabine. *Journal of Analytical and Applied Pyrolysis*, *130*(January), 249–255.
- Wu, M., Guo, H., Liu, L., Liu, Y., & Xie, L. (2019). Size-dependent cellular uptake and localization profiles of silver nanoparticles. *International Journal of Nanomedicine*, *14*, 4247–4259.
- Wu, W., Luo, L., Wang, Y., Wu, Q., Dai, H. Bin, Li, J. S., Durkan, C., Wang, N., & Wang, G. X. (2018). Endogenous pH-responsive nanoparticles with programmable size changes for targeted tumor therapy and imaging applications. *Theranostics*, *8*(11), 3038–3058.
- Xia, Y., Hong, Y., Geng, R., Li, X., Qu, A., Zhou, Z., & Zhang, Z. (2020). Amine-Functionalized ZIF-8 as a Fluorescent Probe for Breath Volatile Organic Compound Biomarker Detection of Lung Cancer Patients. *ACS Omega*, *5*(7), 3478–3486.
- Xie, R., Yang, P., Peng, S., Cao, Y., Yao, X., Guo, S., & Yang, W. (2020). A phosphorylcholine-based zwitterionic copolymer coated ZIF-8 nanodrug

with a long circulation time and charged conversion for enhanced chemotherapy. *Journal of Materials Chemistry B*, 8(28), 6128–6138.

- Xing, Q., Pan, Y., Hu, Y., & Wang, L. (2020). Review of the Biomolecular Modification of the Metal-Organ-Framework. *Frontiers in Chemistry*, 8(July), 1–10.
- Yaghi, O. M., Eddaoudi, M., & M. O., & Li, H. (1999). Design and synthesis of an exceptionally stable and highly porous metal-organic framework. *Nature*, 402, 276–279.
- Yaghi, Omar M. (2016). Reticular Chemistry - Construction, Properties, and Precision Reactions of Frameworks. *Journal of the American Chemical Society*, 138(48), 15507–15509.
- Yameen, B., Choi, W. I., Vilos, C., Swami, A., Shi, J., & Farokhzah, O. C. (2008). Insight into nanoparticle cellular uptake and intracellular targeting. *Journal of Controlled Release*, 23(1), 1–7.
- Yan, H., You, Y., Li, X., Liu, L., Guo, F., Zhang, Q., Liu, D., Tong, Y., Ding*, S., & Wang, and J. (2020). Preparation of RGD Peptide / Folate Acid Double-Targeted Mesoporous Silica Nanoparticles and Its Application in Human Breast Cancer. *Frontiers in Pharmacology*, 11(June), 1–10.
- Yan, J., Zhang, H., Cheng, F., He, Y., Su, T., Zhang, X., Zhang, M., Zhu, Y., Li, C., Cao, J., & He, B. (2018). Highly stable RGD/disulfide bridge-bearing starshaped biodegradable nanocarriers for enhancing drug-loading efficiency, rapid cellular uptake, and on-demand cargo release. *International Journal of Nanomedicine*, 13, 8247–8268.
- Yan, L., Chen, X., Wang, Z., Zhang, X., Zhu, X., Zhou, M., Chen, W., Huang, L., Roy, V. A. L., Yu, P. K. N., Zhu, G., & Zhang, W. (2017). Size Controllable and Surface Tunable Zeolitic Imidazolate Framework-8-Poly(acrylic acid sodium salt) Nanocomposites for pH Responsive Drug Release and Enhanced in Vivo Cancer Treatment. *ACS Applied Materials and Interfaces*, 9(38), 32990–33000.
- Yang, K., Yang, K., Chao, S., Wen, J., Pei, Y., & Pei, Z. (2018). A supramolecular hybrid material constructed from pillar[6]arene-based host-guest complexation and ZIF-8 for targeted drug delivery. *Chemical Communications*, 54(70), 9817–9820.
- Yang, L., Ho, N. Y., Alshut, R., Legradi, J., Weiss, C., Reischl, M., Mikut, R., Liebel, U., Müller, F., & Strähle, U. (2009). Zebrafish embryos as models for embryotoxic and teratological effects of chemicals. *Reproductive Toxicology*, 28(2), 245–253.
- Yun, Y. H., Lee, B. K., Park, K., & Lafayette, W. (2016). Controlled Drug Delivery: Historical perspective for the next generation. *Journal of Controlled*

- Zappa, C., & Mousa, S. A. (2016). Non-small cell lung cancer: Current treatment and future advances. *Translational Lung Cancer Research*, 5(3), 288–300.
- Zeng, R., Chen, Y., Zhao, S., & Cui, G. H. (2012). Autophagy counteracts apoptosis in human multiple myeloma cells exposed to oridonin in vitro via regulating intracellular ROS and SIRT1. *Acta Pharmacologica Sinica*, 33(1), 91–100.
- Zhang, K., Na, T., Wang, L., Gao, Q., Yin, W., Wang, J., & Yuan, B. Z. (2014). Human diploid MRC-5 cells exhibit several critical properties of human umbilical cord-derived mesenchymal stem cells. *Vaccine*, 32(50), 6820–6827.
- Zhang, Q., Zhang, Y., Li, K., Wang, H., Li, H., & Zheng, J. (2015). A novel strategy to improve the therapeutic efficacy of Gemcitabine for non-small cell lung cancer by the tumor-penetrating peptide iRGD. *PLoS ONE*, 10(6), 1–14.
- Zhang, R., Qin, X., Kong, F., Chen, P., & Pan, G. (2019). Improving cellular uptake of therapeutic entities through interaction with components of cell membrane. *Drug Delivery*, 26(1), 328–342.
- Zhang, Y, Jia, Y., & Hou, L. (2018). Synthesis of zeolitic imidazolate framework-8 on polyester fiber for PM 2.5 removal. *RSC Advances*, 8(55), 31471–31477.
- Zhang, Yongyong, Jia, Y., Li, M., & Hou, L. (2018). Influence of the 2-methylimidazole/zinc nitrate hexahydrate molar ratio on the synthesis of zeolitic imidazolate framework-8 crystals at room temperature. *Scientific Reports*, 8(1), 1–7.
- Zhang, Yujing, & Chang, C. H. (2020). Metal-organic framework thin films: Fabrication, modification, and patterning. *Processes*, 8(3).
- Zhao, F., Zhao, Y., Liu, Y., Chang, X., Chen, C., & Zhao, Y. (2011). Cellular uptake, intracellular trafficking, and cytotoxicity of nanomaterials. *Small*, 7(10), 1322–1337.
- Zhen, Z., Tang, W., Chen, H., Lin, X., Todd, T., Wang, G., Cowger, T., Chen, X., & Xie, J. (2013). RGD-modified apoferritin nanoparticles for efficient drug delivery to tumors. *ACS Nano*, 7(6), 4830–4837.
- Zheng, C., Wang, Y., Phua, S. Z. F., Lim, W. Q., & Zhao, Y. (2017). ZnO-DOX@ZIF-8 Core-Shell Nanoparticles for pH-Responsive Drug Delivery. *ACS Biomaterials Science and Engineering*, 3(10), 2223–2229.
- Zheng, S., Wang, X., Weng, Y. H., Jin, X., Ji, J. L., Guo, L., Hu, B., Liu, N., Cheng, Q., Zhang, J., Bai, H., Yang, T., Xia, X. H., Zhang, H. Y., Gao, S., & Huang, Y. (2018). siRNA Knockdown of RRM2 Effectively Suppressed Pancreatic

Tumor Growth Alone or Synergistically with Doxorubicin. *Molecular Therapy - Nucleic Acids*, 12(5), 805–816.

Zhuang, J., Kuo, C. H., Chou, L. Y., Liu, D. Y., Weerapana, E., & Tsung, C. K. (2014). Optimized metal-organic-framework nanospheres for drug delivery: Evaluation of small-molecule encapsulation. *ACS Nano*, 8(3), 2812–2819.

Zuo, H. (2019). iRGD: A Promising Peptide for Cancer Imaging. *Journal of Oncology*, 2019.

