



UNIVERSITI PUTRA MALAYSIA

***SUPPRESSION OF LIGNOHEMICELLULOSIC DEGRADATION CAUSED
BY *Ganoderma boninense* IN OIL PALM DURING BASAL STEM ROT
ESTABLISHMENT USING BENZOIC ACID***

ROZI FERNANDA

IKP 2022 2



**SUPPRESSION OF LIGNOHEMICELLULOSIC DEGRADATION CAUSED BY
Ganoderma boninense IN OIL PALM DURING BASAL STEM ROT
ESTABLISHMENT USING BENZOIC ACID**

By

ROZI FERNANDA

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

April 2022

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

“Special dedication to my late father; Nazaruddin bin Mansyur, mother; Netti Hernawati, brother and sisters; Rahmat Hidayat, Lidya Widya, Nurul Nadia, Jerli Yanita for their endless love, prayers, motivation, continuous support, and encouragement throughout my study”



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Master of Science

**SUPPRESSION OF LIGNOHEMICELLULOSIC DEGRADATION CAUSED BY
Ganoderma boninense IN OIL PALM DURING BASAL STEM ROT
ESTABLISHMENT USING BENZOIC ACID**

By

ROZI FERNANDA

April 2022

Chair : Assoc. Prof. Khairulmazmi Ahmad, PhD
Institute : Plantation Studies

The palm oil industry is one of the most successful industries in the world in terms of agricultural crops towards economic growth and development in its producing countries. Indonesia and Malaysia have become the main producers of palm oil dominated approximately 85% of the total world production of palm oil. However, a decline in the production of this oil has been observed that occurred due to the presence of white-rot fungus *Ganoderma boninense* the causal pathogen of basal stem rot (BSR) disease causing the degradation in the lower part of the trunk. *G. boninense* degrades the major components of wood in the lower trunk makes a loss of its support to the palm, which eventually caused the palm to collapse. Inefficacy in controlling this disease could be the lack of understanding of how the pathogen establishes itself in the host and degrade the structural components of oil palm trees. One of the potential compounds that can be used to suppress the growth of *G. boninense* is benzoic acid (BA). BA belong to aromatic carboxylic acid group that has antifungal effect against many pathogenic fungi. Therefore, this study aimed to understand a step-by-step mechanism of structural degradation of oil palm by *G. boninense* and the effect of BA on suppression of *G. boninense* growth and enzymatic activities while maintaining the structural components of oil palm wood (OPW). The antifungal activity of BA at different concentrations against the growth and enzymatic activities of *G. boninense* were tested using OPW as the substrate. Further, the use of BA to strengthen the lignin structure in oil palm seedlings was evaluated. BA at a concentration above 5 mM successfully inhibited the growth of *G. boninense* in *in-vitro* study. The mechanism of pathogen suppression by BA was related to the disruption and depolarization of fungal cell membrane lead to the loss in membrane permeability and integrity and finally caused the alteration in morphological, anatomical, and ultracellular structure of fungal mycelia. The study on wood degradation by *G. boninense* in response to BA treatment

showed that *G. boninense* simultaneously degrade all the components of OPW. A significant ($P < 0.05$) reduction in wood weight loss and wood chemical component was observed in the BA-treated woodblock at 5mM and above. BA significantly reduced the degradation of OPW depending on its concentration. Microscopic observation of wood inoculated with *G. boninense* showed that the severe damage in wood structure of untreated wood block in compare with treated wood block showing least damage in the wood structure. This was due to the suppression of the ligno-hemicellulolytic activity of *G. boninense* by BA. The observation at the nursery level showed that during the disease establishment, *G. boninense* colonized the plant's root tissue 10 to 14 days after inoculation, while BA-treated seedlings showed the slow disease progression that colonized the root tissues in 7 to 8 weeks after inoculation. BA affect the morphological, anatomical, and cellular structure of *G. boninense* unable them to produce lignohemicellulolytic enzymes for degradation of wood lignohemicellulose. In addition, BA induced the activity of defense-related enzymes that increase lignin biosynthesis in the oil palm seedlings. These findings could be useful in understanding the mechanism of *G. boninense* infection in OPW and contribute to the development of sustainable control of BSR disease.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

**PENINDASAN DEGRADASI LIGNOHEMISELULOSA DISEBABKAN OLEH
Ganoderma boninense PADA KELAPA SAWIT SEMASA FASA
PENUBUHAN PENYAKIT REPUT PANGKAL BATANG MENGGUNAKAN
ASID BENZOIK**

Oleh

ROZI FERNANDA

April 2022

Pengerusi : Prof. Madya. Khairulmazmi Ahmad, PhD
Institut : Kajian Perladangan

Industri minyak sawit adalah salah satu industri yang paling berjaya di dunia dari segi tanaman pertanian ke arah pertumbuhan ekonomi dan pembangunan di negara pengeluar. Indonesia dan Malaysia adalah pengeluar terbesar minyak sawit yang mendominasi kira-kira 85% daripada jumlah pengeluaran minyak sawit dunia. Walau bagaimanapun, terdapat penurunan dalam pengeluaran minyak berlaku disebabkan oleh kulat *Ganoderma boninense*, patogen penyakit reput pangkal batang (BSR) yang menyebabkan degradasi di bahagian bawah batang sawit. *G. boninense* mendegradasi komponen utama kayu yang membuat kehilangan sokongan kepada pokok sawit yang akhirnya menyebabkan pokok sawit tumbang. Ketidakterkesanan dalam mengawal penyakit ini disebabkan oleh kekurangan pemahaman mekanisme tentang bagaimana patogen ini menjangkiti pokok sawit dan penguraian komponen binaan struktur pokok kelapa sawit. Asid benzoik dapat digunakan sebagai salah satu antikulat untuk menindas pertumbuhan *G. boninense*. Asid benzoik tergolong dalam kumpulan asid karboksilik yang mempunyai kesan antikulat terhadap banyak patogen. Oleh itu, kajian ini bertujuan untuk memahami mekanisme penguraian struktur lignohemiselulosa kelapa sawit oleh *G. boninense* dan kesan asid benzoik pada kepekatan optimum untuk menindas aktiviti enzim dan pertumbuhan patogen ini sambil mengekalkan komponen binaan struktur batang kelapa sawit. Aktiviti antikulat dari asid benzoik pada kepekatan yang berbeza terhadap pertumbuhan dan aktiviti enzimasi *G. boninense* telah diuji dengan menggunakan kayu kelapa sawit sebagai substrat. Keberkesanan asid benzoik dalam mengukuhkan struktur lignin di dalam anak benih kelapa sawit juga dinilai. Asid benzoik pada kepekatan di atas 5 mM berjaya menghalang pertumbuhan *G. boninense* dalam kajian in-vitro. Mekanisma penindasan pertumbuhan patogen oleh asid benzoik adalah

berkaitan dengan gangguan dan depolarisasi membran sel kulit yang mengakibatkan kehilangan kebolehtelapan dan integriti membran yang akhirnya menyebabkan perubahan dalam struktur morfologi, anatomi, dan ultraselular miselia kulit. Kajian mengenai degradasi kayu sebagai tindak balas kepada rawatan asid benzoik menunjukkan bahawa *G. boninense* memusnahkan semua komponen kayu kelapa sawit secara serentak. Pengurangan yang ketara ($P \leq 0.05$) dalam penurunan berat kayu dan komponen kimia kayu diperhatikan pada kayu yang dirawat oleh asid benzoik pada 5 mM dan ke atas. Asid benzoik dengan ketara berupaya mengurangkan degradasi kayu kelapa sawit bergantung kepada kepekatannya. Pemerhatian mikroskopik kayu setelah degradasi *G. boninense* menunjukkan bahawa kerosakan teruk dalam struktur kayu yang tidak dirawat di dalam perbandingan dengan kayu yang dirawat menunjukkan kerosakan paling rendah terhadap struktur kayu. Ini disebabkan oleh penindasan aktiviti lignohemiselolitik *G. boninense* oleh asid benzoik. Pemerhatian di peringkat nurseri menunjukkan bahawa semasa perkembangan penyakit, *G. boninense* menakluki tisu akar tumbuhan pada 10 hingga 14 hari selepas inokulasi, sementara anak benih yang dirawat dengan asid benzoik menunjukkan perkembangan penyakit yang perlahan dalam menjangkiti tisu akar dalam tempoh 7 hingga 8 minggu selepas inokulasi. Asid benzoik menyebabkan kerosakan struktur morfologi, anatomi, dan selular *G. boninense* sehingga tidak berupaya menghasilkan enzim lignohemiselolitik untuk degradasi komponen kayu sawit. Tambahan, asid benzoik mendorong aktiviti enzim perumah berkaitan sistem pertahanan dengan meningkatkan aktiviti biosintesis lignin pada anak pokok kelapa sawit. Penemuan ini bermanfaat dalam memahami mekanisme jangkitan *G. boninense* dan pertahanan pada kayu kelapa sawit seterusnya menyumbang kepada pembangunan kawalan mampan penyakit reput pangkal batang sawit.

ACKNOWLEDGEMENTS

In the name of ALLAH, the Most Beneficent, Most Gracious, Most Merciful.

Alhamdulillah, all praises and thanks only for Allah, the one who by His blessing and favor the perfected works are accomplished. First, I would like to express my sincere gratitude to my supervisor Assoc. Prof. Dr. Khairulmazmi Ahmad and Dr. Yasmeeen Siddique for their continuous guidance, advice, motivation, encouragement, critics, and patience towards me. I would like to appreciate their valuable guidance from writing up the proposal till the final thesis submission. I would like to extend my sincere gratitude to my co-supervisor Dr. Kong Lih Ling and Dr. Yuvarani Naidu for their help, guidance, knowledge, and advice throughout my study.

I would like to thank my labmate, Daarshini Ganapathy who help me throughout my study especially in completing the first objective of my study. Also Mrs. Manori Kuruppu for her support, motivation, and guidance during my work.

My utmost appreciation to all the Laboratory staff members Institute of Agriculture and Food Security for their help during my study. I also would like to thank Mr. Rosmidi and Dr. Yuvarani Naidu from Plant Pathology and Biosecurity Unit, Malaysia Palm Oil Board (MPOB), Bangi, Selangor, who guided me at the beginning of Nursery study and their help in obtaining the samples for my study.

Special acknowledgment is given to the Ministry of Higher Education of Malaysia for funding this research project and giving me sponsorship through Special Graduate Research Assistantship (SGRA).

Finally, my deepest gratitude goes to all concerned persons who helped, assisting as well as supported me along this journey.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Khairulmazmi bin Ahmad, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Kong Lih Ling, PhD

Research Officer
Institute of Plantation Studies
Universiti Putra Malaysia
(Member)

Yuvarani Naidu Raju Naidu, PhD

Research Officer
Biology and Sustainability Research Division
Malaysian Palm Oil Board
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 11 August 2022

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman of
Supervisory
Committee: Assoc. Prof. Dr. Khairulmazmi Ahmad

Signature: _____
Name of Member of
Supervisory
Committee: Dr. Kong Lih Ling

Signature: _____
Name of Member of
Supervisory
Committee: Dr. Yuvarani Naidu Raju Naidu

TABLE OF CONTENTS

	Page
ABSTRACT	I
ABSTRAK	iii
ACKNOWLEDGEMENTS	V
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xiv
LIST OF ABBREVIATIONS	xvi
CHAPTER	
1 INTRODUCTION	
1.1. Background	1
1.2. Problemstatement	2
1.3. Hypothesis	2
1.4. Research objectives	3
2 LITERATURE REVIEW	
2.1 Oil palm and its economic importance	4
2.2 Basal stem rot disease of Oil Palm	5
2.2.1 Mode of infection of BSR	6
2.2.1 Management strategies of BSR	6
2.3 General composition of wood	8
2.3.1 Cellulose and hemicellulose	9
2.3.2 Lignin	9
2.3.3 Starch	10
2.4 Degradation of wood components	10
2.4.1 Enzymatic degradation of wood	11
2.4.2 Ligno-hemicellulolytic enzymes	12
2.4.3 Role of ligno-hemicellulolytic enzymes	13
2.4.3.1 Pathogenesis	13
2.4.3.2 Detoxification	14
2.5 Lignification	14
2.6 Benzoic acid	15
3 MECHANISM OF PATHOGEN SUPPRESSION AND PATTERN OF WOOD DEGRADATION BY <i>Ganoderma boninense</i> IN RESPONSE TO BENZOIC ACID	
3.1 Introduction	17
3.2 Materials and Methods	18
3.2.1 Chemicals	18
3.2.2 Fungal strain and culture conditions	18

3.2.3	Screening of benzoic acid against the growth of <i>Ganoderma boninense</i> using “food poison” technique	18
3.2.4	Morphological and ultrastructural alterations of <i>G. boninense</i> mycelia	18
3.2.4.1	Scanning Electron Microscopy (SEM)	19
3.2.4.2	High-Resolution Transmission Electron Microscopy (HR-TEM)	19
3.2.5	Flow Cytometry	19
3.2.5.1	Membrane depolarization assay	19
3.2.5.2	Propidium iodide influx assay	20
3.2.6	Biodegradation of oil palm wood blocks	20
3.2.7	Scanning electron microscopy analysis	20
3.2.8	Statistical analysis	21
3.3	Results	21
3.3.1	Growth of <i>G. boninense</i> on the media incorporated with benzoic acid	21
3.3.2	Alteration in the ultrastructure of <i>G. boninense</i>	22
3.3.3	Cell Membrane integrity and depolarization	24
3.3.4	Biodegradation of oil palm wood blocks	25
3.3.5	Alteration in oil palm wood structure by <i>G. boninense</i>	27
3.4	Discussion	29
3.5	Conclusion	32

4 ALTERATION OF LIGNOHEMICELLULOSIC COMPONENTS OF *G. boninense* INFECTED OIL PALM WOOD AND ITS RESPONSE TO BENZOIC ACID TREATMENT

4.1	Introduction	33
4.2	Materials and Methods	34
4.2.1	Chemicals	34
4.2.2	Culture conditions and treatments	34
4.2.3	Sample preparation	34
4.2.4	Determination of oil palm wood chemical composition degraded by <i>G. boninense</i>	34
4.2.5	Spectroscopic characterization of oil palm wood	35
4.2.6	Production and extraction of ligninolytic and hemicellulolytic enzymes	35
4.2.7	Bradford assay	36
4.2.8	Determination of lignohemicellulolytic enzymes activity	36
4.2.8.1	Laccase enzyme assay	36
4.2.8.2	Lignin peroxidase enzyme assay	36
4.2.8.3	Manganese peroxidase enzyme assay	36
4.2.8.4	Cellulase, xylanase, and amylase enzymes assay	37
4.2.7	Statistical analysis	37
4.3	Results	37

4.3.1	Weight loss due to degradation of lignohemicellulose components by <i>G. boninense</i>	37
4.3.2	Chemical composition of oil palm wood	38
4.3.3	Spectroscopic analysis	40
4.3.4	Ligninolytic enzymes activity of <i>G. boninense</i>	43
4.3.5	Hemicellulolytic enzymes activity of <i>G. boninense</i>	43
4.4	Discussion	46
4.5	Conclusion	49
5	ROLE OF BENZOIC ACID ON STRENGTHENING ROOTS LIGNIN STRUCTURE AS A DEFENSE AGAINST <i>Ganoderma boninense</i> ESTABLISHMENT AND BASAL STEM ROT DISEASE DEVELOPMENT IN OIL PALM	
5.1	Introduction	50
5.2	Materials and Methods	51
5.2.1	Chemicals	51
5.2.2	Treatment selection and culture condition	51
5.2.3	Preparation of inoculum	51
5.2.4	Pre-treatment and artificial inoculation of oil palm seedlings	51
5.2.5	Oil palm growth and disease assessment	52
5.2.6	Determination of defense related enzymes activity	53
5.2.7	Anatomical changes of roots lignin structure in response to <i>G. boninense</i> and benzoic acid treatment during disease progression	54
5.2.8	Statistical analysis	54
5.3	Results	54
5.3.1	Pathogenicity assessment	54
5.3.2	Physiological growth of oil palm seedlings	58
5.3.3	The activity of defense related enzymes by the host during BSR establishment	62
5.3.4	Effect of fungal growth progression to the lignin distribution in the roots system	65
5.4	Discussion	66
5.5	Conclusion	69
6	SUMMARY, CONCLUSION, AND RECOMMENDATION FOR FUTURE RESEARCH	
6.1	Summary and Conclusion	70
6.2	Recommendation for future research	71
	REFERENCES	72
	APPENDICES	88
	BIODATA OF STUDENT	95
	LIST OF PUBLICATIONS	96

LIST OF TABLES

Table		Page
3.1	Effect of different concentrations of benzoic acid on Percentage Inhibition of Diameter Growth (PIDG) of <i>G. boninense</i>	22
4.1	Chemical composition of Healthy oil palm wood	37
4.2	FT-IR analysis of OPW at different stages of degradation	41
5.1	List of treatments for nursery evaluation	49
5.2	Disease severity scale and descriptions of internal and external symptoms of BSR	50
5.3	Area under disease progress curve (AUDPC) of treated oil palm seedlings in response to <i>Ganoderma</i> infection eight months after inoculation	55
5.4	Disease reduction (DR) of treated oil palm seedlings in response to <i>Ganoderma</i> infection eight months after inoculation	55

LIST OF FIGURES

Figure		Page
3.1	Scanning electron micrograph of <i>G. boninense</i> mycelia grown on PDA media.	23
3.2	Ultra-structure observation through HR-Transmission electron micrograph of <i>G. boninense</i> mycelia.	23
3.3	The percentage of membrane stained with Dibac4(3) for detection of membrane depolarization of <i>G. boninense</i> cell.	25
3.4	<i>G. boninense</i> inoculated wood block after 120 days of inoculation.	26
3.5	Percentage of weight loss in untreated and BA-treated OPW block degraded by <i>G. boninense</i> at different incubation times.	27
3.6	Scanning Electron Micrographs of oil palm wood blocks inoculated with <i>G. boninense</i> .	28
4.1	Percentage weight loss of OPW degraded by <i>G. boninense</i> in response to benzoic acid treatment.	37
4.2	The percentage loss of lignohemicellulosic materials of OPW block degraded by <i>G. boninense</i> at different stages of degradation.	39
4.3	FT-IR spectrum of lignohemicellulosic materials of OPW degraded by <i>G. boninense</i> at different stages of degradation	40
4.4	Enzymes production during the degradation of lignohemicellulose components of OPW block at different stages of decay.	44
5.1	The external symptoms of BSR disease on oil palm seedlings observed eight MAI.	53
5.2	The internal symptoms of BSR disease in oil palm seedlings observed eight MAI.	54
5.3	Measurement of physiological parameters of treated and untreated seedlings in response to <i>G. boninense</i> infection.	57
5.4	Measurement of fresh and dry root weight of treated and untreated seedlings in response to <i>G. boninense</i>	59

infection.

- 5.5 The activity of defense-related enzymes; Phenylalanine ammonia-lyase, Peroxidase, and Polyphenol oxidase, in response to *G. boninense* infection and benzoic acid treatments. 61
- 5.6 Alteration in the anatomical structure of oil palm roots infected by *G. boninense* and its response to BA treatment. 63



LIST OF ABBREVIATIONS

Ø	Diameter
%	Percentage
ANOVA	Analysis of variance
AUDPC	Area under disease progress curve
BA	Benzoic acid
BSR	Basal stem rot
BSA	Basal serum albumin
cm	Centimeter
CMC	Carboxymethyl cellulose
DAI	Day after inoculation
DNS	Dinitrosalicylic acid
DMP	2,6-dimethoxyphenol
DS	Disease severity
DR	Disease reduction
FT-IR	Fourier transform infrared
g	Gram
h	Hours
HR-TEM	High-resolution transmission electron microscope
L	Liter
LiP	Lignin peroxidase
MAI	Month after inoculation
MEA	Malt extract agar
M	Molar
mM	Millimolar

mm	Millimeter
mg	Milligram
Min	Minute
MnP	Manganese peroxidase
MPOB	Malaysian Palm Oil Board
OD	Optical density
OP	Oil palm
OPT	Oil Palm trunk
OPW	Oil palm wood
PAL	Phenyl ammonia-lyase
PDA	Potato dextrose agar
PIDG	Percentage inhibition of diameter growth
POD	Peroxidase
PPO	Polyphenol oxidase
RCBD	Randomized Complete Block Design
rpm	Rotations per minute
SA	Salicylic acid
SEM	Scanning electron microscope
WAI	Week after inoculation
WRF	White rot fungi

CHAPTER 1

INTRODUCTION

1.1 Background

Oil palm (*Elaeis guineensis*) is the most productive vegetable oil-producing crop in the world. It is one of the important contributors to economic development of Malaysia and Indonesia as the major producers of palm oil. Indonesia palm oil responsible for 60% of global production followed by Malaysia at 25% (USDA, 2019). Production of oil palm (OP) increases in Indonesia by 65.9% annually (Purnomo et al., 2020). In 2018, OP has significantly contributed to the Gross Domestic Product (GDP) of Malaysia agricultural sector at 37.9% (Uzir, 2019). To fulfil global needs of this oil, as the main producer, Indonesia expand the land for OP cultivation every year (Purnomo et al., 2020).

Having an significant role in economic growth and development and rapidly increasing global demand for palm oil, the palm oil industry faced some constraints (Susila and Supriono, 2001). The major constraint in OP production is the presence of diseases in many oil palm cultivation (Basiron and Weng, 2004). Having disease and unproductive plants can reduce the yield and productivity of OP that lead to significant economic loss to the palm oil industry (Assis et al., 2016). There are many diseases caused by different types of organism such as insect pests, bacteria, and fungi have been identified to cause the disease in OP plantations. Basal stem rot (BSR) disease caused by basidiomycetes fungus, *Ganoderma boninense* is considered as the most serious disease (Wong et al., 2012) that reduces the production of palm oil (Flood et al., 2010 ; Rakib et al., 2014).

The BSR is reported to cause an economic loss of up to USD 365 million per annum in Malaysia (Seman, 2018). It has been estimated the BSR disease caused a yield loss of 68.73% of obtainable yield per year from all the infected palm (Kamu et al., 2021). The infection rate of BSR has been increased from 1.5% in 1995 to 7.4% in 2017 (Ildris et al., 2019). In 2020, the total oil palm planted area infected by *G. boninense* estimated around 443,430 ha in Malaysia (Bharudin et al., 2022). In one of the plantation in North Sumatera BSR devastated thousands of hectares of the palm tree caused 31 to 67% loss of tree per hectare in 25 years oil palm plantation (Riyanto et al., 2020). Estimated the loss at USD 256 million for every 1% incidence in Indonesia (Darmono, 2011). These yield losses were estimated not only from the dead tree but also from the disease infected tree (Kannan et al., 2017). Fruit bunch yield is continuously reduced until the fungus kills the tree (Olaniyi & Szulczyk, 2020).

1.2 Problem statement

The BSR disease caused by *G. boninense* is the most destructive disease of OP industry in Southeast Asia (Idris 1999; Susanto et al., 2005; Wong et al., 2012). BSR has already resulted in a significant loss of income in Malaysian and Indonesian OP industry. To control the *G. boninense* in the plantation, clean clearing, chemical, and biological control methods, and improving disease resistance have been practiced. However, none of those was promising. Some of the chemicals that are applied by trunk injection require technical skills and intensive equipment for their use in the plantation (Lam and Chiu, 1993; Kalidas, 2012).

G. boninense causes degradation in oil palm wood (OPW) by their enzymatic activities leads to a reduction in plant yield and productivity. The degradation of lignohemicellulose components by enzymatic activity of *G. boninense* are associated with BSR infection of OP yet this aspect has been overlooked. The lignin component is destroyed by the fungus, allowing it to attack the cellulose and cause the OP to collapse because it can no longer support the weight of the oil palm. Inefficacy in controlling this devastating disease might be due to the lack of understanding on pathogen establishment in the host and its role in wood component degradation. A recent study reported that phenolic compounds has successfully suppressed the lignin-degrading enzyme produced by the pathogen (Surendran et al., 2018b). Amongst the tested phenolic compounds, benzoic acid (BA) was the most potent inhibitor (Surendran et al., 2017). Plant produce BA naturally in response to environmental stresses (Williams et al., 2003). BA has an antifungal activity towards many pathogens. The mechanism of pathogen suppression by BA is related with the disruption in fungal cellular structure that damaged the organelles and affect the membrane function such as cell septation during cell division (Ganapathy et al., 2021; Nahela et al., 2021). Knowing its ability as an inhibitor in fungal growth, the use of BA could be one of the potential methods for controlling pathogenic fungi. Therefore, this study was designed to understand the fundamental role of *G. boninense* in wood degradation causing BSR and patterns of wood degradation in BSR epidemiology. Additionally, the emphasis is given on the use of BA as prevention of lignohemicellulose degradation caused by *G. boninense* as a potential compound for BSR suppression.

1.3 Hypothesis

BA could be used as an antifungal compound to control BSR disease by understanding the mechanism of lignohemicellulose degradation due to *G. boninense* establishment.

1.4 Research objectives

The general objective is to understand the mechanism of lignohemicellulose degradation due to *G. boninense* establishment and evaluate the use of BA for BSR suppression. The specific objectives of the study are as follow:

1. To determine the role of BA in altering the morphology and cellular structure of *G. boninense* and to evaluate the patterns of wood degradation by *G. boninense* and alterations due to BA treatment in OPW blocks.
2. To determine the changes in wood chemical components and enzymatic activities of *G. boninense* during lignohemicellulosic degradation of OPW blocks in response to BA treatment.
3. To determine the potential use of BA in strengthening the OPW lignin structure and suppression of *G. boninense* in oil palm seedlings during BSR establishment.

REFERENCES

- Agrios, G. N. (2005). *Plant Pathology 5th ed*. Elsevier-Academic Press.
- Alcazar-Fuoli, L., and Mellado, E. (2012). Ergosterol biosynthesis in *Aspergillus fumigatus*: Its relevance as an antifungal target and role in antifungal drug resistance. *Frontiers in Microbiology*. <https://doi.org/10.3389/fmicb.2012.00439>
- Alexander, A., Dayou, J., Abdullah, S., and Chong, K. P. (2017a). The changes of oil palm roots cell wall lipids during pathogenesis of *Ganoderma boninense*. In *IOP Conference Series: Earth and Environmental Science*. <https://doi.org/10.1088/1755-1315/77/1/012014>
- Alexander, A., Sipaut, C. S., Dayou, J., and Chong, K. P. (2017b). Oil palm roots colonisation by *Ganoderma boninense*: An insight study using scanning electron microscopy. *Journal of Oil Palm Research*, 29(2), 262–266. <https://doi.org/10.21894/jopr.2017.2902.10>
- Al-Obaidi, J., Mohd-Yusuf, Y., Razali, N., and Jayapalan, J. (2014). Identification of proteins of altered abundance in oil palm infected with *Ganoderma boninense*. *International Journal of Molecular Science*.
- Amborabé, B.-E., Fleurat-Lessard, P., Chollet, J.-F., and Roblin, G. (2002). Antifungal effects of salicylic acid and other benzoic acid derivatives towards *Eutypa lata*: structure–activity relationship. *Plant Physiology and Biochemistry*, 40(12), 1051–1060.
- Andlar, M., Rezić, T., Mardetko, N., Kracher, D., Ludwig, R., and Šante, K.B. (2018). Lignocellulose degradation: an overview of fungi and fungal enzymes involved in lignocelluloses degradation. *Eng Life Sci* 18: 768–77.
- Aoun, M. (2017). Host defense mechanisms during fungal pathogenesis and how these are overcome in susceptible plants: A review. *International Journal of Botany*. <https://doi.org/10.3923/ijb.2017.82.102>
- ASEAN Secretariat. (2003). *Implementation of the ASEAN Policy on Zero Burning Guidelines for the Implementation of the ASEAN Policy on Zero Burning*. [https://doi.org/ISBN 43\\$3\\$4134](https://doi.org/ISBN 43$3$4134)
- Assis, K., Chong, K. P., Idris, A. S., and Ho, C. M. (2016). Economic loss due to *Ganoderma* disease in oil palm. *International Journal of Social, Behavioral, Educational, Economic, Business and Industrial Engineering*, 10(2), 604–608. Retrieved from <http://waset.org/publications/10003869>
- Baird, R., and Carling, D. (1998). Survival of parasitic and saprophytic fungi on intact senescent cotton roots. *Journal of Cotton Science*.
- Bari, E., Nazarnezhad, N., Kazemi, S. M., Tajick Ghanbary, M. A., Mohebby, B., Schmidt, O., and Clausen, C. A. (2015). Comparison between degradation capabilities of the white rot fungi *Pleurotus ostreatus* and

- Trametes versicolor in beech wood. *International Biodeterioration and Biodegradation*. <https://doi.org/10.1016/j.ibiod.2015.03.033>
- Bari, E., Mohebby, B., Naji, R. N., Oladi, R., Yilgor, N., Nazarnezhad, N., Ohno, K. M., and Nicholas, D. D. (2018). Monitoring the cell wall characteristics of degraded Beech wood by white-rot fungi: anatomical, chemical, and photochemical study. *Madeias. ciencia y tecnologia* 20(1): 35-56.
- Basiron, Y., and Weng, C. K. (2004). The Oil Palm and its Sustainability. *Journal of Oil Palm Research*, 16(1): 1–10.
- Bharudin, I., Ab Wahab, A.F.F., Abd Samad, M.A., Xin Yie, N., Zairun, M.A., Abu Bakar, F.D., and Abdul Murad, A.M. (2022). Review Update on the Life Cycle, Plant–Microbe Interaction, Genomics, Detection and Control Strategies of the Oil Palm Pathogen *Ganoderma boninense*. *Biology*, 11, 251. <https://doi.org/10.3390/biology11020251>
- Bucher, V. V. C., Hyde, K. D., Pointing, S. B., and Reddy, C. A. (2004). Production of wood decay enzymes, mass loss and lignin solubilization in wood by marine ascomycetes and their anamorphs. *Fungal Diversity*, 15, 1–14.
- Buchanan, P. K. (2001). Aphyllophorales in Australasia. *Australian Systematic botany*, 14(3): 417-437.
- Bhuiyan, N. H., Selvaraj, G., Wei, Y., and King, John. (2009). Role of lignification in plant defense. *Plant Signaling and Behavior* 4:2, 158-159.
- Brigham, C. (2018). “Biopolymers: biodegradable alternatives to traditional plastics,” in *Green Chemistry: an Inclusive Approach*, eds B. Török and T. Dransfield (Amsterdam: Elsevier Inc.), 753–770.
- Campbell, C. L., and Madden, L. V. (1990). Introduction to plant disease epidemiology. *New York: Wiley-Interscience*
- Castoldi, R., Bracht, A., de Moraes, G. R., Baesso, M. L., Correa, R. C. G., Peralta, R. A., ... Peralta, R. M. (2014). Biological pretreatment of Eucalyptus grandis sawdust with white-rot fungi: Study of degradation patterns and saccharification kinetics. *Chemical Engineering Journal*. <https://doi.org/10.1016/j.cej.2014.07.090>
- Chong, K. P., Dayou, J., and Alexander, A. (2017). Detection and Control of *Ganoderma boninense* in Oil Palm Crop. *Springer International Publishing*
- Chu, J., Li, W. F., Cheng, W., Lu, M., Zhou, K. H., Zhu, H. Q., ... Zhou, C. Z. (2015). Comparative analyses of secreted proteins from the phytopathogenic fungus *Verticillium dahliae* in response to nitrogen starvation. *Biochimica et Biophysica Acta - Proteins and Proteomics*. <https://doi.org/10.1016/j.bbapap.2015.02.004>
- Chung, G. F. (2011). Management of *Ganoderma* diseases in oil palm plantations. *The planter*, 87(1022): 325-339.

- Coelho-Moreira, J. d. S. , Maciel, G. M. , Rafael Castoldi, R., Mariano, S. d. S. , Inácio, F. D. , Adelar Bracht, A., and Peralta, R. M. (2013). Involvement of Lignin-Modifying Enzymes in the Degradation of Herbicides. *IntechOpen*. <https://doi.org/10.5772/55848>
- Dambolena, J. S., López, A. G., Meriles, J. M., Rubinstein, H. R., and Zygadlo, J. A. (2012). Inhibitory effect of 10 natural phenolic compounds on *Fusarium verticillioides*. A structure-property-activity relationship study. *Food Control*. <https://doi.org/10.1016/j.foodcont.2012.05.008>
- Darwish, S.S., El Hadidi, N., and Mansour, M. (2013). The effect of fungal decay on *Ficus sycomoris* wood. *International Journal Conservation Science* 4:271-282
- Darmono, T. (2011). Strategi berperang melawan *Ganoderma* pada perkebunan kelapa sawit Proc. Symp. Nasional & Lokakarya *Ganoderma* "Sebagai Patogen Penyakit Tanaman & Bahan Baku Obat Tradisional" (Bogor: IPB Press)
- Dashtban, M., Schraft, H., Syed, T. A., and Qin, W. (2010). Fungal biodegradation and enzymatic modification of lignin. *International Journal of Biochemistry and Molecular Biology*, 1(1), 36–50.
- Dawood, M. G., Sadak, M. S., Bakry, B. A., and El Karamany, M. F. (2019). Comparative studies on the role of benzoic, t-cinnamic, and salicylic acids on growth, some biochemical aspects, and yield of three flax cultivars grown under sandy soil conditions. *Bulletin of the National Research Centre*, 43(1). <https://doi.org/10.1186/s42269-019-0152-4>
- Del Río, J. C., Gutiérrez, A., and Martínez, Á. T. (2004). Identifying acetylated lignin units in non-wood fibers using pyrolysis-gas chromatography/mass spectrometry. *Rapid Communications in Mass Spectrometry*, 18(11): 1181–1185. <https://doi.org/10.1002/rcm.1457>
- Ding, P., and Ding, Y. (2020). Stories of Salicylic Acid: A Plant Defense Hormone. *Trends in Plant Science*. <https://doi.org/10.1016/j.tplants.2020.01.004>
- Donaldson, L. A. (2001). Lignification and lignin topochemistry - An ultrastructural view. *Phytochemistry*, 57(6): 859–873.
- Durmaz, S., Özgenç, Ö., Boyacı, I. H., Yildiz, Ü. C., and Erişir, E. (2016). Examination of the chemical changes in spruce wood degraded by brown-rot fungi using FT-IR and FT-Raman spectroscopy. *Vibrational Spectroscopy*. <https://doi.org/10.1016/j.vibspec.2016.04.020>
- Durner, J., Shah, J., and Klessig, D. F. (1997). Salicylic acid and disease resistance in plants. *Trends in Plant Science*. [https://doi.org/10.1016/S1360-1385\(97\)86349-2](https://doi.org/10.1016/S1360-1385(97)86349-2)
- El-Fawy M. M., and Abo-Elyousr, K. A. M. (2016). Efficacy of certain chemical compounds on common bean rust disease. *Archives of Phytopathology and Plant Protection* 49:19-20.

- Erwin, Takemoto, S., Hwang, W. J., Takeuchi, M., Itoh, T., and Imamura, Y. (2008). Anatomical characterization of decayed wood in standing light red meranti and identification of the fungi isolated from the decayed area. *Journal of Wood Science*, 54(3): 233–241.
- Fackler, K., and Schwanninger, M. (2012). How spectroscopy and microspectroscopy of degraded wood contribute to understand fungal wood decay. *Applied Microbiology and Biotechnology*, 96(3), 587–599. <https://doi.org/10.1007/s00253-012-4369-5>
- Fernandes, L., Loguercio-Leite, C., Esposito, E., and Reis, M. M. (2005). In vitro wood decay of *Eucalyptus grandis* by the basidiomycete fungus *Phellinus flavomarginatus*. *International Biodeterioration and Biodegradation*, 55(3): 187–193.
- Flood, J., Hasan, Y., and Foster, H. (2002). *Ganoderma* diseases of oil palm— an interpretation from Bah Lias Research Station. *Planter*, 78(921): 689–710.
- Flood, J., Cooper, R., Rees, R., Potter, U., and Hasan, Y. (2010). Some Latest R & D on *Ganoderma* Diseases in Oil Palm. *IOPRI/MPOB Seminar: Advances in the Controlling of Devastating Disease of Oil Palm (Ganoderma) in South East Asia*, (October 2014), 1–21.
- Freeman. (2008). An Overview of Plant Defenses against Pathogens and Herbivores. *The Plant Health Instructor*. <https://doi.org/10.1094/phi-i-2008-0226-01>
- Ganapathy, D., Siddiqui, Y., Ahmad, K., Adzmi, F., and Ling, K.L. (2021). Alterations in Mycelial Morphology and Flow Cytometry Assessment of Membrane Integrity of *Ganoderma boninense* Stressed by Phenolic Compounds. *Biology*, 10, 930. <https://doi.org/10.3390/biology10090930>
- Garcia-Rubio, R., de Oliveira, H. C., Rivera, J., and Trevijano-Contador, N. 2020. The fungal cell wall: *Candida*, *Cryptococcus*, and *Aspergillus* species. *Front. Microbiol*, 10, 3389. <https://doi.org/10.3389/fmicb.2019.02993>
- Gawade, D.B., Perane, R.R., Surywanshi, A.P., and Deokar, C.D. (2017). Extracellular enzymes activity determining the virulence of *Rhizoctonia bataticola*, causing root rot in soybean. *Physiol. Mol. Plant Pathol*, 100: 49–56.
- Govender, N., and Wong, M. Y. (2017). Detection of oil palm root penetration by agrobacterium-mediated transformed *ganoderma boninense*, expressing green fluorescent protein. *Phytopathology*, 107(4): 483–490. <https://doi.org/10.1094/PHYTO-02-16-0062-R>
- H'ng, P. S., Wong, L. J., Chin, K. L., Tor, E. S., Tan, S. E., Tey, B. T., and Mammski, M. (2011). Oil palm (*Elaeis guineensis*) trunk as a resource of starch and other sugars. *Journal of Applied Sciences*. <https://doi.org/10.3923/jas.2011.3053.3057>
- Hamid, N. H., Sulaiman, O., Mohammad, A., and Ahmad Ludin, N. (2012). The

Decay Resistance and Hyphae Penetration of Bamboo *Gigantochloa scortechinii* Decayed by White and Brown Rot Fungi . *International Journal of Forestry Research*. <https://doi.org/10.1155/2012/572903>

- Hayat, Q., Hayat, S., Alyemeni, M. N., and Ahmad, A. (2012). Salicylic acid mediated changes in growth, photosynthesis, nitrogen metabolism and antioxidant defense system in *Cicer arietinum* L. *Plant Soil Environ*, 58 (9): 417-423
- Hou, R., Shi, J., Ma, X., Wei, H., Hu, J., Tsang, Y. F., and Gao, M. T. (2020). Effect of Phenolic Acids Derived from Rice Straw on *Botrytis cinerea* and Infection on Tomato. *Waste and Biomass Valorization*. <https://doi.org/10.1007/s12649-020-00938-1>
- Ho, C. L., and Tan, Y. C. (2015). Molecular defense response of oil palm to *Ganoderma* infection. *Phytochemistry*, 114: 168-177.
- Husaini, A., Fisol, F. A., Yun, L. C., Hasnain, M., Muid, S., and Roslan, H. A. (2011). Lignocellulolytic enzymes produced by tropical white rot fungi during biopulping of *Acacia mangium* wood chips. *Journal of Biochemical Technology*, 3(2): 245–250.
- Hushiarian, R., Yusof, N. A., and Dutse, S. W. (2013). Detection and control of *Ganoderma boninense*: Strategies and perspectives. *SpringerPlus*, 2(1): 1–12. <https://doi.org/10.1186/2193-1801-2-555>
- Idris, A. S. (1999). *Basal Stem Rot (BSR) of oil palm (Elaeis guineensis Jacq.) In Malaysia: Factors associated with variation in Disease Severity*. PhD Thesis. University of London, UK.
- Idris, A. S. (2009). Basal stem rot in Malaysia - Biology, epidemiology, economic importance, detection and control.
- Idris, A., Kushairi, D., Ariffin, D., and Basri, M. (2006). Technique for inoculation of oil palm germinated seeds with *Ganoderma*. *MPOB inf Ser* 314:1-4
- Idris, A.S., Mohd Shukri, I., Izzuddin, M.A., Norman, K., Khairuman, H., Ramle, M., Iptizam, N., and Dayang, N.S. *Survey on Status of Ganoderma Disease of Oil Palm Estates and Smallholders in Malaysia*. In Proceedings of the MPOB International Palm Oil Congress and Exhibition (PIPOC) 2019, Kuala Lumpur, Malaysia, 19–21 November 2019; Volume 2, pp. 548–557.
- Industri, P., Sawit, M., and Lestari, P. E. (2017). The role of Malaysian palm oil industry in the Malaysian sustainable economic development. *International Journal of the Malay World and Civilisation (Jurnal Antarabangsa Alam Dan Tamadun Melayu)*, 5(1): 11–18.
- Jordá, T., and Puig, S. (2020). Regulation of ergosterol biosynthesis in *saccharomyces cerevisiae*. *Genes*. <https://doi.org/10.3390/genes11070795>
- Kadir, G., Parveez, A., Lip, K., Charles Hill, T., Arn, T. Y., and Kushairi, A. (2019). *Journal of Oil Palm, Environment & Health* An official publication

of the Malaysian Palm Oil Council (MPOC) Sustainable Oil Palm Cultivation In Malaysia-Are Peatlands A Suitable Choice?. *Environment & Health*, 10(May), 13–18. <https://doi.org/10.5366/jope.2019.03>

- Kalidas, P. (2012). Pest problems of oil palm and management strategies for sustainability. *Agrotechnol* S11:001. <https://doi.org/10.4172/2168-9881.S11-001>
- Kamu, A. (2020). Estimating The Yield Loss Of Oil Palm Due To Ganoderma Basal Stem Rot Disease By Using Bayesian Model Averaging. *Journal of Oil Palm Research*, (November). <https://doi.org/10.21894/jopr.2020.0061>
- Kamu, A., Khim Phin, C., Abu Seman, I., Gabda, D., and Chong Mun, H. (2021). ESTIMATING The YIELD LOSS Of OIL PALM DUE To Ganoderma BASAL STEM ROT DISEASE By USING BAYESIAN MODEL AVERAGING. *Journal of Oil Palm Research*, 33(1): 46–55. <https://doi.org/10.21894/jopr.2020.0061>
- Kamu, A., Mun, H. C., Phin, C. K., and Seman, I. A. (2018). Identifying the early visible symptoms of the ganoderma-infected oil palms: A case study on the infected palms which collapsed within twelve months after disease census. *ASM Science Journal*, 11(Special Issue 2), 156–163.
- Kang, Z., and Buchenauer, H. (2000). Ultrastructure and cytochemical studies on the infection of wheat spikes by *Fusarium culmorum* as well as on degradation of cell wall components and localization of mycotoxins in the host tissue. *Mycotoxin Research*, 16 Suppl (S1): 1-5.
- Kannan, P., Peng, T. S., Ahmad, S. M., Seman, I. A., Rahman, A. K. b, Hashim, K., ... Omar, W. (2017). Knowledge Assessment of Basal Stem Rot Disease of Oil Palm and its Control Practices among Recipients of Replanting Assistance Scheme in Malaysia. *International Journal of Agricultural Research*, 12(2): 73–81.
- Karim, N. F. A., Mohd, M., Nor, N. M. I. M., and Zakaria, L. (2016). Saprophytic and potentially pathogenic fusarium species from peat soil in perak and pahang. *Tropical Life Sciences Research*.
- Kaur, H., Salh, P. K., and Singh, B. (2017). Role of defense enzymes and phenolics in resistance of wheat crop (*Triticum aestivum* L.) towards aphid complex. *Journal of Plant Interactions*, 12(1): 304–311. <https://doi.org/10.1080/17429145.2017.1353653>
- Kazan, K., and Gardiner, D. M. (2017). Targeting pathogen sterols: Defence and counter defence?. *PLoS Pathogens*. <https://doi.org/10.1371/journal.ppat.1006297>
- Khalil, H. P. S. A., Alwani, M. S., Ridzuan, R., Kamarudin, H., and Khairul, A. (2008). Chemical composition, morphological characteristics, and cell wall structure of Malaysian oil palm fibers. *Polymer - Plastics Technology and Engineering*, 47(3): 273–280.
- King, B. C., Waxman, K. D., Nenni, N. V., Walker, L. P., Bergstrom, G. C., and Gibson, D. M. (2011). Arsenal of plant cell wall degrading enzymes

- reflects host preference among plant pathogenic fungi. *Biotechnology for Biofuels*, 4: 1–14. <https://doi.org/10.1186/1754-6834-4-4>
- Koyani, R. D., Bhatt, I. M., Patel, H. R., Vasava, A. M., and Rajput, K. S. (2016). Evaluation of *Schizophyllum commune* Fr. potential for biodegradation of lignin: A light microscopic analysis. *Wood Material Science & Engineering*, 11(1): 46–56.
- Kubicek, C. P., Starr, T. L., and Glass, N. L. (2014). Plant Cell Wall–Degrading Enzymes and Their Secretion in Plant-Pathogenic Fungi. *Annual Review of Phytopathology*, 52(1): 427–451.
- Kumar, S., Abedin, M. M., Singh, A. K., and Das, S. (2020). Role of phenolic compounds in plant-defense mechanism. In R, Lone et al. (Eds). *Plant Phenolic in Sustainable Agriculture* (pp. 517-532). Singapore: Springer Nature.
- Kushairi, A., Loh, S. K., Azman, I., Hishamuddin, E., Ong-Abdullah, M., Izuddin, Z. B. M. N., ... Parveez, G. K. A. (2018). Oil palm economic performance in Malaysia and r&d progress in 2017. *Journal of Oil Palm Research*, 30(2): 163–195. <https://doi.org/10.21894/jopr.2018.0030>
- Kushairi, A., Ong-Abdullah, M., Nambiappan, B., Hishamuddin, E., Bidin, M. N. I. Z., Ghazali, R., ... Parveez, G. K. A. (2019). Oil palm economic performance in Malaysia and r&d progress in 2018. *Journal of Oil Palm Research*, 31(2): 165–194. <https://doi.org/10.21894/jopr.2019.0026>
- Lacan, I. (2018). Wood Decay Fungi Found in Failed Wood. *Landscape Disease Symposium*, 12–13.
- Lam, C., and Chiu, S. (1993). hexaconazole (anvil5SC), a cost-effective fungicide for controlling white rot disease in immature rubber. *Planter* 69, 465-474.
- Lamaming, J., Hashim, R., Leh, C. P., Sulaiman, O., Sugimoto, T., and Nasir, M. (2015). Isolation and characterization of cellulose nanocrystals from parenchyma and vascular bundle of oil palm trunk (*Elaeis guineensis*). *Carbohydrate Polymers*. <https://doi.org/10.1016/j.carbpol.2015.08.017>
- Latifah, J., Nurrul-Atika, M., Sharmiza, A., and Rushdan, I. (2020). Extraction of nanofibrillated cellulose from Kelempayan (*Neolamarckia cadamba*) and its use as strength additive in papermaking. *Journal of Tropical Forest Science* 32(2):170-178.
- Lattanzio, V., Veronica M. T. Lattanzio, and A. C. (2015). *Role of phenolics in the resistance mechanisms of plants against fungal pathogens and insects*. *Phytochemistry* (Vol. 661).
- Lee, J. S. H., Ghazoul, J., Obidzinski, K., and Koh, L. P. (2014). Oil palm smallholder yields and incomes constrained by harvesting practices and type of smallholder management in Indonesia. *Agronomy for Sustainable Development*, 34(2): 501–513. <https://doi.org/10.1007/s13593-013-0159-4>
- Lee, W., and Lee, D. G. (2014). An antifungal mechanism of curcumin lies in

- membrane-targeted action within *Candida albicans*. *IUBMB Life*. <https://doi.org/10.1002/iub.1326>
- Li, S. M., Hua, G. G., Liu, H. X., and Guo, J. H. (2008). Analysis of defence enzymes induced by antagonistic bacterium *Bacillus subtilis* strain AR12 towards *Ralstonia solanacearum* in tomato. *Annals of Microbiology*, 58(4): 573–578. <https://doi.org/10.1007/BF03175560>
- Liu, J., Zong, Y., Qin, G., Li, B., and Tian, S. (2010). Plasma membrane damage contributes to antifungal activity of silicon against *penicillium digitatum*. *Current Microbiology*. <https://doi.org/10.1007/s00284-010-9607-4>
- Liu, Q., Luo, L., and Zheng, L. (2018). Lignins: Biosynthesis and biological functions in plants. *International Journal of Molecular Sciences*. <https://doi.org/10.3390/ijms19020335>
- Loyd, A. L., Linder, E. R., Anger, N. A., Richter, B. S., Blanchette, R. A., and Smith, J. A. (2018). Pathogenicity of ganoderma species on landscape trees in the Southeastern United States. *Plant Disease*, 102(10): 1944–1949.
- Lundell, T. K., Mäkelä, M. R., de Vries, R. P., and Hildén, K. S. (2014). Genomics, lifestyles and future prospects of wood-decay and litter-decomposing basidiomycota. In *Advances in Botanical Research*. <https://doi.org/10.1016/B978-0-12-397940-7.00011-2>
- Lyu, X., Shen, C., Fu, Y., Xie, J., Jiang, D., Li, G., and Cheng, J. (2015). Comparative genomic and transcriptional analyses of the carbohydrate-active enzymes and secretomes of phytopathogenic fungi reveal their significant roles during infection and development. *Sci Rep*, 5: 15565.
- Manavalan, T., Manavalan, A., Thangavelu, K. P., and Heese, K. (2012). Secretome analysis of *Ganoderma lucidum* cultivated in sugarcane bagasse. *Journal of Proteomics*, 77: 298–309.
- Martínez, Á. T., Ruiz-Dueñas, F. J., Martínez, M. J., del Río, J. C., and Gutiérrez, A. (2009). Enzymatic delignification of plant cell wall: from nature to mill. *Current Opinion in Biotechnology*. <https://doi.org/10.1016/j.copbio.2009.05.002>
- Martínez, Á. T., Speranza, M., Ruiz-Dueñas, F. J., Ferreira, P., Camarero, S., Guillén, F., ... Del Río, J. C. (2005). Biodegradation of lignocellulosics: Microbial, chemical, and enzymatic aspects of the fungal attack of lignin. *International Microbiology*, 8(3): 195–204.
- Mandels, M., and Reese, E. T. (1965). Inhibition of cellulases. *Annual review of Phytopathology*, 3(1): 85-102.
- Midot, F., Lau, S. Y. L., Wong, W. C., Tung, H. J., Yap, M. L., Lo, M. L., ... Melling, L. (2019). Genetic diversity and demographic history of *Ganoderma boninense* in oil palm plantations of Sarawak, Malaysia inferred from ITS regions. *Microorganisms*, 7(10): 1–17.

- Mikulic-Petkovsek, M., Schmitzer, V., Stampar, F., Veberic, R., and Koron, D. (2014). Changes in phenolic content induced by infection with *Didymella applanata* and *Leptosphaeria coniothyrium*, the causal agents of raspberry spur and cane blight. *Plant Pathology*. <https://doi.org/10.1111/ppa.12081>
- Murphy, D. J., Goggin, K., and Paterson, R. R. M. (2021). Oil palm in the 2020s and beyond: challenges and solutions. *CABI Agriculture and Bioscience* 2, 39.
- Mohammed, C. L., Rimbawanto, A., and Page, D. E. (2014). Management of basidiomycete root- and stem-rot diseases in oil palm, rubber and tropical hardwood plantation crops. *Forest Pathology*. <https://doi.org/10.1111/efp.12140>
- Moore, C. W., Mckov, J., Del Valle, R., Armstrong, D., Bernard, E. M., Katz, N., and Gordon, R. E. (2003). Fungal cell wall septation and cytokinesis are inhibited by bleomycins. *Antimicrob Agents Chemother*, 47, 3281-3289
- Naher, L., Yusuf, U. K., Ismail, A., Tan, S. G., and Mondal, M. M. A. (2013). Ecological status of *Ganoderma* and basal stem rot disease of oil palms (*Elaeis guineensis* Jacq.). *Australian Journal of Crop Science*.
- Naidu, Y., Idris, A. S., Nusaibah, S. A., Norman, K., and Siddiqui, Y. (2015). In vitro screening of biocontrol and biodegradation potential of selected hymenomycetes against *Ganoderma boninense* and infected oil palm waste. *Forest Pathology*, 45(6): 474–483.
- Naidu, Yuvarani, Siddiqui, Y., Rafii, M. Y., Saud, H. M., and Idris, A. S. (2017). Investigating the effect of white-rot hymenomycetes biodegradation on basal stem rot infected oil palm wood blocks: Biochemical and anatomical characterization. *Industrial Crops and Products*, 108(September): 872–882.
- Naidu, Yuvarani, Siddiqui, Y., Rafii, M. Y., Saud, H. M., and Idris, A. S. (2018). Inoculation of oil palm seedlings in Malaysia with white-rot hymenomycetes: Assessment of pathogenicity and vegetative growth. *Crop Protection*. <https://doi.org/10.1016/j.cropro.2018.02.018>
- Najmie, M., Khalid, K., Sidek, A., and Jusoh, M. (2011). Density and ultrasonic characterization of oil palm trunk infected by *Ganoderma boninense* disease. *Measurement Science Review*. <https://doi.org/10.2478/v10048-011-0026-x>
- Nambiappan, B., Ismail, A., Hashim, N., Ismail, N., Shahari, D. N., Idris, N. A. N., ... Kushairi, A. (2018). Malaysia: 100 years of resilient palm oil economic performance. *Journal of Oil Palm Research*, 30(1): 13–25. <https://doi.org/10.21894/jopr.2018.0014>
- Nehela, Y., Taha, N. A., Elzaawely, A. A., Xuan, T. D., Amin, M. A., Ahmed, M. E., and El-Nagar, A. (2021). Benzoic acid and its hydroxylated derivatives suppress early blight of tomato (*Alternaria solani*) via the induction of salicylic acid biosynthesis and enzymatic and nonenzymatic antioxidant defense machinery. *Journal of Fungi*, 7(8): 1–26.

- Nirwan, B., Choudhary, S., Sharma, K., and Singh, S. (2016). In vitro studies on management of root rot disease caused by *Ganoderma lucidum* in *Prosopis cineraria*. *Current Life Sciences*, 2(4): 118–126. <https://doi.org/10.5281/zenodo.166773>
- Noor, M. M. (2003). Zero Burning Techniques in Oil Palm Cultivation: an Economic Perspective. *Oil Palm Industry Economic Journal*, 3(1): 16–24.
- O'Connell, R., Thon, M., Hacquard, S., Amyotte, S. G., Kleemann, J., ... Vailancourt, L. J. (2012). Lifestyle transitions in plant pathogenic *Colletotrichum* fungi deciphered by genome and transcriptome analyses. *Nat Genet*, 44: 1060–1065.
- Olaniyi, O. N., and Szulczyk, K. R. (2020). Estimating the economic damage and treatment cost of basal stem rot striking the Malaysian oil palms. *Forest Policy and Economics*. <https://doi.org/10.1016/j.forpol.2020.102163>
- Ommelna, B. G., Jennifer, A. N., and Chong, K. P. (2012). The potential of chitosan in suppressing *Ganoderma boninense* infection in oil-palm seedlings. *Journal of Sustainability Science and Management*.
- Padró, M. D. A., Caboni, E., Morin, K. A. S., Mercado, M. A. M., and Olalde-Portugal, V. (2021). Effect of *Bacillus subtilis* on antioxidant enzyme activities in tomato grafting. *PeerJ*, 9: 1–28.
- Parihar, S. P., Om Prakash, ., and Punetha, H. (2012). Investigation on defensive enzymes activity of *Brassica juncea* genotypes during pathogenesis of *Alternaria* blight. *Nature and Science*, 10(2), 63–68.
- Paterson, R., Meon, S., Abidin, M. A., and Lima, N. (2008). Prospects for Inhibition of Lignin Degrading Enzymes to Control *Ganoderma* White Rot of Oil Palm. *Current Enzyme Inhibition*, 4(4): 172–179.
- Paterson, R. R. M. (2007). *Ganoderma* disease of oil palm-A white rot perspective necessary for integrated control. *Crop Protection*, 26(9): 1369–1376. <https://doi.org/10.1016/j.cropro.2006.11.009>
- Pattathil, S., Hahn, M. G., Dale, B. E., and Chundawat, S. P. S. (2015). Insights into plant cell wall structure, architecture, and integrity using glycome profiling of native and AFEXTM-pretreated biomass. *Journal of Experimental Botany*. <https://doi.org/10.1093/jxb/erv107>
- Pratyusha, S. (2022). Phenolic compounds in the plant development and defense: An overview. In Hasanuzzaman, M., and Nahar, K. *Plant stress Physiology-Perspective in Agriculture*. IntechOpen. <https://doi.org/10.5772/intechopen.102873>
- Peralta, R. M., da Silva, B. P., Gomes Côrrea, R. C., Kato, C. G., Vicente Seixas, F. A., and Bracht, A. (2017). Enzymes from Basidiomycetes-Peculiar and Efficient Tools for Biotechnology. *Biotechnology of Microbial Enzymes: Production, Biocatalysis and Industrial Applications*. <https://doi.org/10.1016/B978-0-12-803725-6.00005-4>

- Pizzolitto, R. P., Barberis, C. L., Dambolena, J. S., Herrera, J. M., Zunino, M. P., Magnoli, C. E., ... Dalcerro, A. M. (2015). Inhibitory effect of natural phenolic compounds on *Aspergillus parasiticus* growth. *Journal of Chemistry*. <https://doi.org/10.1155/2015/547925>
- Pozdnyakova, N. N. (2012). Involvement of the Ligninolytic System of White-Rot and Litter-Decomposing Fungi in the Degradation of Polycyclic Aromatic Hydrocarbons. *Biotechnology Research International*, 1–20. <https://doi.org/10.1155/2012/243217>
- Podobnik, B., Stojan, J., Lah, L., and Krasevec, N. (2008). CYP53A15 of *Cochliobolus lunatus*, a Target for natural antifungal compounds. *Journal of Medicinal Chemistry*.
- Purnomo, H., Okarda, B., Dermawan, A., Ilham, Q. P., Pacheco, P., Nurfatriani, F., and Suhendang, E. (2020). Reconciling oil palm economic development and environmental conservation in Indonesia: A value chain dynamic approach. *Forest Policy and Economics*, 111(January), 102089. <https://doi.org/10.1016/j.forpol.2020.102089>
- Qualley, A. V., Widhalm, J. R., Adebesein, F., Kish, C. M., and Dudareva, N. (2012). Completion of the core Beta-oxidative pathway of benzoic acid biosynthesis in plants. *Proc. Natl. Acad. Sci. USA*. 109(40): 16383-16388.
- Rahmati, Y., Nourmohammadi, K., Naghdi, R., and Kartoolinejad, D. (2019). Effect of fungal degradation on physicochemical properties of exploited stumps of oriental beech over a 25-year felling period and the obtained Kraft pulp properties. *Journal of Forest Science*. <https://doi.org/10.17221/93/2018-JFS>
- Rakib, M. R. M., Bong, C. F. J., Khairulmazmi, A., and Idris, A. S. (2014). Genetic and morphological diversity of *Ganoderma* species isolated from infected oil palms (*Elaeis guineensis*). *International Journal of Agriculture and Biology*, 16(4): 691–699.
- Ramzi, A. B., Me, M. L. C., Ruslan, U. S., Baharum, S. N., and Muhammad, N. A. N. (2019). Insight into plant cell wall degradation and pathogenesis of *Ganoderma boninense* via comparative genome analysis. *PeerJ*, 12. <https://doi.org/10.7717/peerj.8065>
- Rees, R. W., Flood, J., Hasan, Y., and Cooper, R. M. (2007). Effects of inoculum potential, shading and soil temperature on root infection of oil palm seedlings by the basal stem rot pathogen *Ganoderma boninense*. *Plant Pathology*, 56(5): 862-870.
- Rees, R. W., Flood, J., Hasan, Y., Potter, U., and Cooper, R. M. (2009). Basal stem rot of oil palm (*Elaeis guineensis*); Mode of root infection and lower stem invasion by *Ganoderma boninense*. *Plant Pathology*, 58(5): 982–989. <https://doi.org/10.1111/j.1365-3059.2009.02100.x>
- Riyanto., Sartini., and Nasution, J. (2020). Oil palm in related to plant density and *Ganoderma boninense* infection in Simalangun and Asahan Plantations, North Sumatera, Indonesia. *European Journal of Biology and Medical Science Research*, Vol.8, No. 5, pp. 1-7

- Roslan, A., and Idris, A. S. (2012). Economic Impact of Ganoderma Incidence on Malaysian Oil Palm Plantation – A Case Study in Johor. *Oil Palm Industry Economic Journal*, 12(1): 24–30.
- Sahebi, M., Hanafi, M. M., Wong, M. Y., Idris, A. S., Azizi, P., Jahromi, M. F., ... Mohidin, H. (2015). Towards immunity of oil palm against Ganoderma fungus infection. *Acta Physiologiae Plantarum*, 37(10). <https://doi.org/10.1007/s11738-015-1939-z>
- Sanghvi, G. V., Koyani, R. D., and Rajput, K. S. (2013). Anatomical characterisation of teak (*Tectona grandis*) wood decayed by fungus *Chrysosporium asperatum*. *Journal of Tropical Forest Science*.
- Sanderson, F. R. (2005). An insight into spore dispersion of *Ganoderma boninense* on oil palm. *Mycopathologia*, 159(1): 139-141.
- Seman, I.B. R&D on biology, detection and management of *Ganoderma* disease in oil palm. In Workshop on Basal Stem Rot (BSR) of Oil Palm, Universiti Putra Malaysia, Selangor, Malaysia, 2018; Volume 12.
- Scheller, H. V., Ulvskov, P. (2010). Hemicellulose_composition. *Annu. Rev. Plant Biol.* .
- Schirp, A., and Wolcott, M. P. (2005). Influence of fungal decay and moisture absorption on mechanical properties of extruded wood-plastic composites. *Wood and Fiber Science*.
- Schneider, W. D. H., Fontana, R. C., Baudel, H. M., de Siqueira, F. G., Rencoret, J., Gutiérrez, A., ... Camassola, M. (2020). Lignin degradation and detoxification of eucalyptus wastes by on-site manufacturing fungal enzymes to enhance second-generation ethanol yield. *Applied Energy*. <https://doi.org/10.1016/j.apenergy.2020.114493>
- Schoenherr, S., Ebrahimi, M., and Czermak, P. (2018). Lignin Degradation Processes and the Purification of Valuable Products. *Lignin - Trends and Applications*. <https://doi.org/10.5772/intechopen.71210>
- Schwartz, M., Perrot, T., Aubert, E., Dumarçay, S., Favier, F., Gérardin, P., ... Gelhaye, E. (2018). Molecular recognition of wood polyphenols by phase II detoxification enzymes of the white rot *Trametes versicolor*. *Scientific Reports*. <https://doi.org/10.1038/s41598-018-26601-3>
- Schwarze, F. W. M. R. (2007). Wood decay under the microscope. *Fungal Biology Reviews*, 21(4): 133–170.
- Sharma, Y. K., León, J., Raskin, I., and Davis, K. R. (1996). Ozone-induced responses in *Arabidopsis thaliana*: The role of salicylic acid in the accumulation of defense-related transcripts and induced resistance. *Proceedings of the National Academy of Sciences of the United States of America*. <https://doi.org/10.1073/pnas.93.10.5099>
- Shibata, M., Varman, M., Tono, Y., Miyafuji, H., and Saka, S. (2008). Characterization in chemical composition of the oil palm (*Elaeis guineensis*). *Nihon Enerugi Gakkaishi/Journal of the Japan Institute of*

Energy, 87(5): 383–388. <https://doi.org/10.3775/jie.87.383>

- Siddiqui, Y., Surendran, A., Masya, E., Agricultural, F., and Sdn, S. (2019). Inhibition of Lignin Degrading Enzymes of Ganoderma spp.: An Alternative Control of Basal Stem Rot Disease of Oil Palm gth Article Inhibition of Lignin Degrading Enzymes of Ganoderma spp.: An Alternative Control of Basal Stem Rot Disease of Oil Palm, (July), 523–530. <https://doi.org/10.17957/IJAB/15.1095>
- Siddiqui, Y., Surendran, A., Paterson, R. R. M., Ali, A., and Ahmad, K. (2021). Current strategies and perspectives in detection and control of basal stem rot of oil palm. *Saudi J. Biol. Scie* 28: 2840-2849.
- Sigoillot, J. C., Berrin, J. G., Bey, M., Lesage-Meessen, L., Levasseur, A., Lomascolo, A., ... Uzan-Boukhris, E. (2012). *Fungal Strategies for Lignin Degradation. Advances in Botanical Research* (1st ed., Vol. 61). Elsevier Ltd. <https://doi.org/10.1016/B978-0-12-416023-1.00008-2>
- Singh Arora, D., and Kumar Sharma, R. (2010). Ligninolytic fungal laccases and their biotechnological applications. *Applied Biochemistry and Biotechnology*, 160(6): 1760–1788. <https://doi.org/10.1007/s12010-009-8676-y>
- Somai-Jemmali, L., Siah, A., Harbaoui, K., Fergaoui, S., Randoux, B., Magnin-Robert, M., ... Hamada, W. (2017). Correlation of fungal penetration, CWDE activities and defense-related genes with resistance of durum wheat cultivars to *Zymoseptoria tritici*. *Physiological and Molecular Plant Pathology*. <https://doi.org/10.1016/j.pmpp.2017.08.003>
- Song, W., Han, X., Qian, Y., Liu, G., Yao, G., Zhong, Y., and Qu, Y. (2016). Proteomic analysis of the biomass hydrolytic potentials of *Penicillium oxalicum* lignocellulolytic enzyme system. *Biotechnology for Biofuels*, 9(1): 1–15. <https://doi.org/10.1186/s13068-016-0477-2>
- Song, Z., Vail, A., Sadowsky, M. J., and Schilling, J. S. (2012). Competition between two wood-degrading fungi with distinct influences on residues. *FEMS Microbiology Ecology*, 79(1): 109–117.
- Srivastava, Seweta; Kumar, R. (2014). Wood Decaying Fungi. *Lambert Academic Publishing*, 1–4.
- Suga, H., and Hyakumachi, M. (2004). Genomics of phytopathogenic fusarium. *Applied Mycology and Biotechnology* (Vol. 4). Elsevier Masson SAS. [https://doi.org/10.1016/S1874-5334\(04\)80009-1](https://doi.org/10.1016/S1874-5334(04)80009-1)
- Sumathi, S., Chai, S., and Mohamed, A. (2008). Utilization of oil palm as a source of renewable energy in Malaysia. *Renew. Sustain. Energy Rev*, 12: 2404-2421.
- Surendran, A., Siddiqui, Y., Ali, N. S., and Manickam, S. (2018a). Inhibition and kinetic studies of cellulose- and hemicellulose-degrading enzymes of *Ganoderma boninense* by naturally occurring phenolic compounds. *Journal of Applied Microbiology*, 124(6): 1544–1555.

- Surendran, A., Siddiqui, Y., Saud, H. M., Ali, N. S., and Manickam, S. (2018b). Inhibition and kinetic studies of lignin degrading enzymes of *Ganoderma boninense* by naturally occurring phenolic compounds. *Journal of Applied Microbiology*, 125(3): 876–887.
- Surendran, Arthy, Siddiqui, Y., Ahmad, K., and Fernanda, R. (2021). Deciphering the physicochemical and microscopical changes in *Ganoderma boninense*-infected oil palm woodblocks under the influence of phenolic compounds. *Plants*, 10(9): 0–15.
- Surendran, Arthy, Siddiqui, Y., Manickam, S., and Ali, A. (2018). Role of benzoic and salicylic acids in the immunization of oil palm seedlings-challenged by *Ganoderma boninense*. *Industrial Crops and Products*, 122(March), 358–365. <https://doi.org/10.1016/j.indcrop.2018.06.005>
- Surendran, Arthy, Siddiqui, Y., Saud, H. M., Ali, N. S., and Manickam, S. (2017). The antagonistic effect of phenolic compounds on ligninolytic and cellulolytic enzymes of *Ganoderma boninense*, causing basal stem rot in oil palm. *International Journal of Agriculture and Biology*, 19(6): 1437–1446.
- Surendran, A. (2018). *Effect of naturally occurring phenolic compound on cell wall degrading enzymes and suppression of Ganoderma boninense infection in oil palm seedlings*. Doctoral thesis. University of Putra Malaysia, Malaysia.
- Suryantini, R., and Wulandari, R. S. (2018). Diversity of *Ganoderma* pathogen in Pontianak, West Kalimantan: Characteristics, virulence and ability to infect *Acacia mangium* seedlings. *Biodiversitas*, 19(2): 465–471. <https://doi.org/10.13057/biodiv/d190213>
- Susanto, A., Sudharto, P. S., and Purba, R. Y. (2005). Enhancing biological control of basal stem rot disease (*Ganoderma boninense*) in oil palm plantations. In *Mycopathologia*. <https://doi.org/10.1007/s11046-004-4438-0>
- Susila, W. R., and Supriono, A. (2001). Industri CPO: peluang terbuka, hambatan masih menghadang. *Kompas*, 2 Oktober 2001.
- Tappi. (2011). Lignin in Wood and Pulp. T222 Om-02, 1–7.
- ten Have, A., Tenberge, K. B., Benen, J. A. E., Tudzynski, P., Visser, J., and van Kan, J. A. L. (2002). The Contribution of Cell Wall Degrading Enzymes to Pathogenesis of Fungal Plant Pathogens. *Agricultural Applications*, 341–358. https://doi.org/10.1007/978-3-662-03059-2_17
- Tian, C., Beeson, W. T., Iavarone, A. T., Sun, J., Marletta, M. A., Cate, J. H. D., and Glass, N. L. (2009). Systems analysis of plant cell wall degradation by the model filamentous fungus *Neurospora crassa*. *Proceedings of the National Academy of Sciences of the United States of America*, 106(52): 22157–22162.
- Tomimura, Y. (1992). Chemical Characteristics and Utilization of Oil Palm Trunk. Jarq, (1992). Chemical Characteristics and Utilization of Oil Palm Trunk. Jarq,

- 25: 283–288. Retrieved from <https://www.jircas.affrc.go.jp/english/publication/jarq/25-4/25-4-283-288.pdf>.
- Tripathi, A., Upadhyay, R. C., and Singh, S. (2012). Extracellular Lignolytic Enzymes in *Bjerkandera adusta* and *Lentinus squarrosulus*. *Indian Journal of Microbiology*, 52(3): 381–387.
- Tzin, V., and Galili. G. (2010). The Biosynthetic Pathways for Shikimate and Aromatic Amino Acids in *Arabidopsis thaliana*. *The arabidopsis book* vol. 8 doi:10.1199/tab.0132
- USDA. (2019). Indonesia Oilseeds and Products Annual 2019. *Global Agricultural Information Network*, 20.
- USDAFAS. (2022). Palm oil explorer: World Palm Oil Production of 2021. <https://ipad.fas.usda.gov/cropexplorer/cropview/commodityView.aspx?cropid=4243000>. (accessed March 5, 2022).
- Uzir, M. (2019). Department of statistics Malaysia: Press release Malaysia economic performance fourth quarter 2018. *Department of Statistics Malaysia*, (February), 1–5.
- van Wyk, J. P. (2001). Biotechnology and the utilization of biowaste as a resource for bioproduct development. *Trends in biotechnology*, 19(5): 172-177.
- Villavicencio, E. V., Mali, T., Mattila, H. K., and Lundell, T. (2020). Enzyme activity profiles produced on wood and straw by four fungi of different decay strategies. *Microorganisms*, 8(1): 4–9.
- Wang, X., Zhou, X., Cai, Z., Guo, L., Chen, X., Chen, X., ... Wang, A. (2021). A biocontrol strain of *Pseudomonas aeruginosa* CQ-40 promote growth and control *Botrytis cinerea* in tomato. *Pathogens*, 10(1): 1–17. <https://doi.org/10.3390/pathogens10010022>
- Widhalm, J. R., and Dudareva, N. (2015). A familiar ring to it: Biosynthesis of plant benzoic acids. *Molecular Plant*, 8(1): 83–97. <https://doi.org/10.1016/j.molp.2014.12.001>
- Wildermuth, M. C. (2006). Variations on a theme: synthesis and modification of plant benzoic acids. *Current Opinion in Plant Biology*, 9(3): 288–296. <https://doi.org/10.1016/j.pbi.2006.03.006>
- Williams, M., Senaratna, T., Dixon, K., and Sivasithamparan, K. (2003). Benzoic acid induces tolerance to biotic stress caused by *Phytophthora cinnamomi* in *Banksia attenuata*. *Plant Growth Regulation*. <https://doi.org/10.1023/A:1027355604096>
- Winandy, J.E., and Rowell, R.M. (1984). The chemistry of wood strength in The chemistry of solid wood Chapter 5. American Chemical Society, 5; 211-255
- Wise, L. E., Murphy, M., and D'Addieco, A. A. (1946). Chlorite holocellulose:

Its fractionation and bearing on summative wood analysis and on studies on the hemicelluloses. *Paper Trade Journal*, 122(2): 35-43.

- Wong, L. C., Bong, C. F. J., and Idris, A. S. (2012). *Ganoderma* species associated with basal stem rot disease of oil palm. *American Journal of Applied Sciences*, 9(6): 879–885.
- Yang, C., Hamel, C., Vujanovic, V., and Gan, Y. (2011). Fungicide: Modes of Action and Possible Impact on Nontarget Microorganisms. *ISRN Ecology*. <https://doi.org/10.5402/2011/130289>
- Yoon, M.-Y., Seo, K.-H., Lee, S.-H., Choi, G.-J., Jang, K.-S., Choi, Y.-H., ... Kim, J.-C. (2012). Antifungal Activity of Benzoic Acid from *Bacillus subtilis* GDYA-1 against Fungal Phytopathogens. *Research in Plant Disease*, 18(2): 109–116. <https://doi.org/10.5423/rpd.2012.18.2.109>
- You, Z., Zhang, C., and Ran, Y. (2020). The effects of clioquinol in morphogenesis, cell membrane and ion homeostasis in *Candida albicans*. *BMC Microbiology*. <https://doi.org/10.1186/s12866-020-01850-3>
- Zaynab, M., Fatima, M., Abbas, S., Sharif, Y., Umair, M., Zafar, M. H., and Bahadar, K. (2018). Role of secondary metabolites in plant defense against pathogens. *Microbial Pathogenesis*, 124: 198-202
- Zhang, L., and Van Kan, J. A. L. (2013). Pectin as a barrier and nutrient source for fungal plant pathogens. In *Agricultural Applications, 2nd Edition*. https://doi.org/10.1007/978-3-642-36821-9_14
- Zhensheng, K., and Buchenauer, H. (2002). Studies on the infection process of *Fusarium culmorum* in wheat spikes: Degradation of host cell wall components and localization of Trichothecene toxins in infected tissue. *European Journal of Plant Pathology*, 108(7): 653-660.