

# **UNIVERSITI PUTRA MALAYSIA**

# DRUG REPURPOSING OF CLINICALLY-APPROVED DRUGS TO TARGET EPITHELIAL-MESENCHYMAL TRANSITION USING IN SILICO ANALYSIS

**ONG CHUN HAO** 

FPSK(m) 2022 17



# DRUG REPURPOSING OF CLINICALLY-APPROVED DRUGS TO TARGET EPITHELIAL-MESENCHYMAL TRANSITION USING *IN SILICO* ANALYSIS



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

December 2021

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

#### DRUG REPURPOSING OF CLINICALLY-APPROVED DRUGS TO TARGET EPITHELIAL-MESENCHYMAL TRANSITION USING *IN SILICO* ANALYSIS

By

### **ONG CHUN HAO**

December 2021

# Chair: Professor Daud Ahmad Israf Ali, PhDFaculty: Medicine and Health Science

Epithelial-mesenchymal transition (EMT) is a process where epithelial cells transform into mesenchymal cells type like fibroblasts and myofibroblasts. In the process, the epithelial cells lose their epithelial phenotype with reduced apicalbasal polarity, while acquiring new mesenchymal traits with increased invasiveness. Accumulation of the mesenchymal cells also leads to the deposition of collagen in the extracellular matrix (ECM). It is well established that EMT contributes to the progression of fibrosis and cancer diseases. Therefore, a therapeutic method that inhibits the EMT process would be required. Transforming growth factor-beta (TGF- $\beta$ ) is a potent inducer of the EMT process. Previous studies had demonstrated that inhibition of the TGF-ß receptor type 1 (also named ALK5) could inhibit EMT. However, current progress on the clinical development of novel ALK5 inhibitor has not been encouraging, often due to safety concerns of the novel drug leads. Since traditional de novo drug discovery comes with high risks, pharmaceutical companies have begun to use drug repurposing strategy for drug development. Drug repurposing or repositioning is a strategy of finding new therapeutic purposes for current existing drugs in the clinical market. Due to the fact that these drugs had been established to be safe for use, it would reduce the concerns of safety risks in human. In this study, drug repurposing approach was used to identify clinically approved drugs that can inhibit the EMT process via targeting TGF- $\beta$  activity. Initial computational screening of clinically approved drugs via molecular docking had revealed several drugs with strong binding affinity (-10.8 to -9.6 kcal/mol) to ALK5 based on the reference range of known ALK5 inhibitors ranging from -11.2 to -9.5 kcal/mol. The shortlisted drug candidates include ergotamine, telmisartan, saquinavir, indinavir, nelfinavir and celecoxib. Subsequently, these drugs were tested experimentally in normal human bronchial epithelial cell line, BEAS-2B induced by TGF- $\beta$ 1. In the experiments, the

morphology changes from cobblestone shape of epithelial cells towards elongated shape of mesenchymal cells were not prevented by the drug treatments. In addition, the drugs did not exhibit inhibitory effects on the downregulation of epithelial proteins (E-cadherin) and upregulation of mesenchymal proteins (vimentin and  $\alpha$ -smooth muscle actin). Based on these observations, it is postulated that the results from molecular docking were false positives. It is recommended that future studies involving molecular docking method would require better optimization and improvement by performing cross-docking validation prior to screening and including the negative controls during screening. The tested drugs in this study could serve as negative controls in future screening against ALK5 protein.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

#### REPOSISI DADAH KLINIKAL UNTUK MENSASARKAN PERALIHAN EPITELIUM KEPADA MESENKIMA DENGAN ANALISIS *IN SILIKO*

Oleh

#### **ONG CHUN HAO**

Disember 2021

#### Pengerusi : Professor Daud Ahmad Israf Ali, PhD Fakulti : Perubatan dan Sains Kesihatan

Peralihan epitelium-mesenkima (EMT) merupakan satu proses dimana sel-sel epitelium berubah menjadi sel-sel mesenkima seperti fibroblas dan myofibroblas. Di dalam proses tersebut, sel-sel epitelium kehilangan fenotip epitelium mereka disebabkan oleh pengurangan polariti apikal-basal, sementara memperolehi ciri mesenkima yang baru dengan peningkatan invasif. Pengumpulan sel-sel mesenkima turut membawa kepada pemendapan kolagen di dalam matriks ekstrasel (ECM). Seperti yang diketahui umum, EMT merupakan penyumbang kepada perkembangan penyakit fibrosis dan kanser. Oleh itu, satu kaedah terapeutik yang boleh merencat proses EMT amat diperlukan. Transformasi faktor pertumbuhan-beta (TGF-β) merupakan pendorong kuat dalam proses EMT. Kajian terdahulu telah menunjukkan bahawa perencatan reseptor TGF- β jenis 1 (juga dipanggil ALK5) boleh merencat proses EMT. Namun begitu, kemajuan semasa perkembangan klinikal perencat baharu ALK5 tidak memberangsangkan, selalunya berpunca daripada masalah keselamatan mengenai dadah yang baharu. Memandangkan penemuan ubat secara tradisional de novo mempunyai risiko tinggi, syarikat farmaseutikal telah mula menggunakan strategi penggunaan semula ubat bagi pembuatan ubat. Penggunaan semula atau reposisi ubat merupakan strategi untuk mencari penggunaan terapeutik baru bagi ubat yang sedia ada di pasaran klinikal. Oleh kerana ubat-ubatan tersebut telah terbukti selamat untuk digunakan, ini akan mengurangkan kebimbangan terhadap risiko keselamatan pada manusia. Dalam kajian ini, penggunaan semula ubat telah digunakan untuk mengenal pasti ubat-ubat yang telah diluluskan secara klinikal yang dapat merencatkan proses EMT dengan mensasarkan aktiviti TGF-β. Saringan awal komputasi ubat yang telah diluluskan secara klinikal melalui pendokkan molekular mendedahkan beberapa ubat-ubatan mempunyai ikatan afiniti yang kuat dengan ALK5, termasuk ergotamine, telmisartan, saguinavir, indinavir, nelfinavir dan celecoxib. Seterusnya, ubat-ubatan tersebut diuji secara eksperimen pada sel bronkus manusia normal, BEAS-2B yang dirangsang oleh TGF- $\beta$ 1. Di dalam eksperimen ini, rawatan ubat tidak dapat menghalang perubahan morfologi daripada epitilum sel berbentuk batu buntar ke bentuk sel mesenkima yang memanjang. Tambahan lagi, ubat-ubatan tersebut tidak menunjukkan kesan perencatan terhadap penurunan regulasi protein epitelium (E-cadherin) dan peningkatan regulasi protein mesenkima (vimentin dan otot lembut α-aktin). Berdasarkan pemerhatian ini, disimpulkan bahawa keputusan daripada pendokkan molekular merupakan positif palsu. Adalah disarankan agar kajian akan datang melibatkan pendokkan molekular memerlukan pengoptimuman dan peningkatan yang lebih baik. Ubat-ubatan yang diuji di dalam kajian ini boleh berfungsi sebagai kontrol negatif di dalam saringan terhadap protein ALK5 pada masa akan datang.



#### ACKNOWLEDGEMENTS

I would like to express my gratitude to my supervisor, Prof. Daud Ahmad Israf Ali for his advice and guidance throughout the course of my degree. His experiences had helped me to overcome many hardships in the project. His encouragement had also kept me motivated during the difficult time in my study.

I would also like to thank my co-supervisor, Dr. Tham Chau Ling and Dr. Hanis Hazeera Harith for their valuable inputs and suggestions in my research project. Their constructive feedbacks had led to many improvements to the project over the course of my study. A special thanks to Dr. Siti Farah Binti Md Tohid for her lesson on molecular docking.

Next, I would like to express my appreciation to Mr. Zulkhairi Zainol for his technical insistence during the usage of any equipment in the Cell Signaling Laboratory. Many thanks to my seniors, Nazmi and Amy for their guidance and advices in cell culture experiments. I also want to give thanks to the other lab mates, Aida, Audrey, Kong Yen, Yee Han, Vivi and Fatiah for their support.

Lastly, I would like to express my deepest thanks to my family who have given their continuous love and support for me. Without them, I will not be able to overcome many challenges that I faced throughout the study. This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

#### Daud Ahmad bin Israf Ali, PhD

Professor Faculty of Medicine and Health Science Universiti Putra Malaysia (Chairman)

## Tham Chau Ling, PhD

Associate Professor Faculty of Medicine and Health Science Universiti Putra Malaysia (Member)

#### Hanis Hazeera binti Harith, PhD

Senior Lecturer Faculty of Medicine and Health Science Universiti Putra Malaysia (Member)

### ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 19 May 2022

# Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman	
Committee:	Prof. Dr. Daud Ahmad Israf Ali
Signature: Name of Member of	
Supervisory Committee:	Assoc. Prof. Dr. Tham Chau Ling
Signature: Name of Member of	
Supervisory Committee:	Dr. Hanis Hazeera Harith

# TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	vi
APPROVAL	vii
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xv

# CHAPTER

1	INTRODUCTION			1
	1.1	General o	biective	3
	1.2	Specific o	bjective	3
2	LITE		REVIEW	4
	2.1	TGF-β	physiological function and	4
	2.2	TGF-β:	from activation to signalling	5
		2.2.1	Synthesis of TGF-β	5
		2.2.2	Activation of latent TGF-β	6
		2.2.3	TGF-β receptor activation	6
		2.2.4	TGF-β intracellular signalling	7
	2.3	Epitheli	al-mesenchymal transition	8
	2.4	Preclini	cal and clinical studies on ALK5	9
		inhibito	rs	
		2.4.1	Preclinical studies on ALK5 inhibitors	9
		2.4.2	Clinical studies on ALK5	12
			inhibitors	
	2.5	Drug re	purposing and molecular docking	12
3	MET	<b>THODOLO</b>	GY	15
	3.1	Materia	lls	15
	3.2	Method	ls	15
		3.2.1	Protein structure retrieval	15
		3.2.2	Ligand structure retrieval	15
		3.2.3	Preparation of proteins and	16
			ligands	
		3.2.4	Molecular docking	16
		3.2.5	Cell culture	18
		3.2.6	Cell proliferation, cell viability	18
			and cytotoxicity (MTS assay)	
		3.2.7	Cell treatment	18
		3.2.8	Western blot	19

		3.2.9	Statistical analysis	20
4	RES	SULTS AN	D DISCUSSION	21 21
	4.1	Molecular	docking	21
		4.2.1	Selection of 3TZM	21
		4.2.2	Validation of docking protocol	21
		4.2.3	Protein-ligand interaction of known ALK5 inhibitor	23
		4.2.4	Protein-ligand interaction of clinically approved drugs	28
	4.3	Cytotoxici	ty of drug candidates	32
	4.4	Determini	ng optimal induction time	34
	4.5	Effects of	drug candidates on cell	35
	4.6	morpholog Effects of markers	gy drug candidates on EMT	40
5	SUN REC RES	MMARY, C COMMEND SEARCH	ONCLUSION AND ATIONS FOR FUTURE	48
REFEREI APPEND BIODATA LIST OF	NCES ICES A OF S PUBLIC	TUDENT CATIONS		49 61 62 63

 $\bigcirc$ 

# LIST OF TABLES

Table		Page
2.1	Summary of ALK5 inhibitors	11
2.2	Summary of computational methods used in drug repurposing	14
4.1	Binding affinity and data of interactions resulting from the molecular docking of known ALK5 inhibitors into TGF-β receptor type 1 protein	24
4.2	Binding affinity and data of interactions resulting from the molecular docking of clinically approved drugs into TGF-β receptor type 1 protein	28

# LIST OF FIGURES

Figure	9	Page
2.1	Schematic representation of the TGF- $\beta$ activation pathway	8
3.1	Interface of Drug ReposER	16
3.2	Flowchart of in silico methodology	17
4.1	Poseview 2D interaction diagram of (A) Co- crystallised SB431542 and (B) Redocked SB431542	22
4.2	Magnified ribbon representation of superimposed co-crystal structure of ALK5-SB431542 complex and the redocked ligand	23
4.3	Poseview 2D interaction diagram of known ALK5 inhibitors	27
4.4	Poseview 2D interaction diagram of top six drugs	30
4.5	Cell viability assay of BEAS-2B cells after treatment of (A) Ergotamine (B) Telmisartan (C) Saquinavir (D) Indinavir (E) Nelfinavir and (F) Celecoxib	33
4.6	Effect of different TGF-β1 induction time on the EMT markers in BEAS-2B cells	35
4.7	Effect of drugs on the cell morphology in TGF- $\beta$ 1-induced BEAS-2B cells	39
4.8	Effect of Ergotamine on EMT markers in TGF- $\beta$ 1-induced BEAS-2B cells	40
4.9	Effect of Telmisartan on EMT markers in TGF- $\beta$ 1-induced BEAS-2B cells	41
4.10	Effect of Saquinavir on EMT markers in TGF- $\beta$ 1-induced BEAS-2B cells	42
4.11	Effect of Indinavir on EMT markers in TGF- $\beta$ 1-induced BEAS-2B cells	43
4.12	Effect of Nelfinavir on EMT markers in TGF-β1- induced BEAS-2B cells	44

4.13	Effect of Celecoxib on EMT markers in TGF-β1- induced BEAS-2B cells	45
6.1	Cell seeding optimization for MTS assay	61
6.2	Standard curve for BCA assay	61



 $\bigcirc$ 

# LIST OF ABBREVIATIONS

ADT	Autodock Tools
AIDS	Acquired immunodeficiency syndrome
ALK	Activin receptor-like kinase
ASO	Antisense oligonucleotide
BCA	Bicinchoninic acid assay
BDL	Bile duct ligation
BEGM	Bronchial Epithelial Growth Medium
CCL4	Carbon tetrachloride
Co-Smad	Common partner Smad
COX-2	Cyclooxygenase 2
СТВР	C-terminal binding protein
DMSO	Dimethylsulfoxide
DNA	Deoxyribonucleic acid
ECM	Extracellular matrix
EDTA	Ethylenediaminetetraacetic acid
EGF	Epidermal growth factor
ELISA	Enzyme-linked immunosorbent assay
ЕМТ	Epithelial-mesenchymal transition
FHD	First-in human dose
GSI	Glomerular Sclerosis Index
IL	Interleukin
I-Smad	Inhibitory Smad
LAP	Latency-associated peptide
LTBP	Latent TGF-β binding protein

5

xv

MMP	Matrix metalloproteinase
mRNA	Messenger ribonucleic acid
MTS	3-(4,5-Dimethylthiazol-2-yl)-5-(3- carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H- tetrazolium
PBS	Phosphate buffer saline
PDB	Protein Data Bank
PVDF	Polyvinylidene fluoride
RIPA	Radioimmunoprecipitation assay
RMSD	Root mean square division
R-Smad	Receptor-regulated Smad
SDS-PAGE	Sodium dodecyl Sulfate-Polyacrylamide Gel Electrophoresis
shRNA	Short hairpin ribonucleic acid
TF	Transcription factor
TGF-β	Transforming growth factor-beta
Treg	Regulatory T cell
TSP	Thrombospondin
ΤβR	TGF-β receptor
uPA	Urokinase-type plasminogen activator
υυο	Unilateral urethral obstruction
ZO	Zona occludens
α-sma	Alpha-smooth muscle actin

 $\mathbf{C}$ 

#### CHAPTER 1

#### INTRODUCTION

Epithelial-mesenchymal transition (EMT) is a process where epithelial cells acquire mesenchymal traits while losing their epithelial phenotype. This process was first recognised in embryonic development, where the pluripotent epiblast forms the mesoderm and endoderm via EMT (Hay, 1995; Kim et al., 2018). The EMT process also plays a major role in wound healing. During the reepithelialization phase of wound healing, the EMT process allows the epithelial cells to acquire the invasive phenotype of mesenchymal cells, so that the cells can migrate and close the wounds (Stone et al., 2016). The EMT process can be involved in pathological conditions such as fibrosis and cancer metastasis when it is not adequately regulated (Barriere et al., 2015). When EMT is adequately regulated, the epithelial cells transform into mesenchymal cell-types such as fibroblasts and myofibroblasts for the production and deposition of extracellular matrix (ECM) components that would help in the wound healing process. However, pathologically prolonged induction of EMT would result in persistent myofibroblast activation and excessive deposition of ECM components, which eventually lead to fibrosis (Stone et al., 2016). In cancer, the epithelial cells transform to a mesenchymal cell type with the ability of invasion and migration through ECM, which allows them to metastasise via the blood and lymphatics (Ramos et al., 2017).

Transforming growth factor-beta (TGF- $\beta$ ) is a multifunctional cytokine that is known to be a potent inducer of the EMT process. TGF- $\beta$  has many important physiological roles including regulation of inflammation, proliferation, differentiation, and apoptosis (Prud'homme, 2007). However, it can become problematic if the level of TGF- $\beta$  goes unchecked. Chronic inflammation and elevated levels of TGF- $\beta$  can lead to excessive induction of the EMT process. As a result, the mesenchymal cells and ECM components will replace the normal parenchymal tissue. While TGF- $\beta$  can suppress the tumour growth in the early phase of neoplasia, excessive TGF- $\beta$  induction of EMT can promote cancer metastasis in the later phase of malignant tumour development (Prud'homme, 2007). All these problems have highlighted the importance to develop a new therapy that can inhibit the EMT process by targeting TGF- $\beta$  activity.

To date, various research projects have tried to develop novel inhibitors of TGF- $\beta$  for the treatment of fibrosis and cancer, but none so far have reached the market. This highlights the difficulty of developing a new drug, which is costly, time-consuming and has a low success rate (Xu et al., 2015). In order to overcome this issue, the drug discovery fraternity have turned to another strategy, known as drug repurposing. Drug repurposing or repositioning is a process of identifying new therapeutic uses for existing clinically-approved drugs (GNS et al., 2019). Since an existing drug has already been tested in humans, it is less likely to fail in clinical trials due to toxicity issues. Moreover, numerous preclinical and clinical data would be available for an existing drug. As a result,

the repurposing process will be relatively less time-consuming and also less costly as compared to the traditional *de novo* drug discovery process. Aside from the benefits of saving valuable resources in drug development, drug repurposing can help to deepen the understanding of the mechanism of action for old drugs and sometimes also lead to discovery of novel therapeutic targets of the diseases. One notable example of drug repurposing is sildenafil, a phosphodiesterase type 5A inhibitor that was originally developed for angina pectoris. Its initial clinical trial revealed little efficacy on the cardiovascular system, and a side effect of penile erection was reported at high doses. Eventually, the investigators repurposed sildenafil for the treatment of erectile dysfunction, and later successfully marketed it as Viagra (Kass et al., 2007).

Taking into consideration that the success rate from development to approval for novel drugs (11%) is lower than repurposed drugs (30%) globally (Fetro and Scherman, 2020), increased efforts have been focused on the repurposing of clinically approved drugs in the recent years. There are computational and experimental approaches that could be used in order to identify the drug candidate for a new indication of interest. Recently, Ab Ghani et al. (2019) have designed a web server known as Drug Repositioning Exploration Resource (Drug ReposER) that can identify potential alternative targets of known drugs by comparison of the three dimensional amino acid arrangement of known drug binding site from the Protein Data Bank (PDB) repository with the query protein. The concept is that when the binding site of two different proteins share similarity in amino acid arrangement, a drug that is known to bind to the first protein would likely bind to the other protein as well. Molecular docking is another computational tool that can predict how two molecules (for example a ligand and a receptor) can form stable binding. This method utilise docking algorithms to predict the binding affinity of a ligand to the binding site of the protein target. Therefore, a large number of drugs can be screened against a protein of interest that is involved in a disease by using the molecular docking tool. This would allow the identification of drugs with the best affinity (in comparison to positive controls) towards the protein of interest based on the result of molecular docking prediction. Subsequently, experiments can be performed to verify the results of molecular docking. This can be carried out with cell-based assays to demonstrate whether the selected drugs can affect the cell phenotype relevant to the disease model. In this study, computational methods (in silico) were used to screen clinically approved drugs for potential inhibition of the TGF-β type I receptor. The identified drugs were further examined in subsequent cell-based assays that employed a TGF-\u00df1-induced model of the bronchial epithelial cell line, BEAS-2B. In past studies, molecular docking and in vitro screening of new compounds were used as the first steps in the development of TGF-β inhibitors but many hit compounds eventually failed at the clinical stage. In current study, a new drug repurposing approach by combining the bioinformatics-based method (Drug ReposER) and molecular docking was used to screen for potential TGF- $\beta$  inhibitors among the clinically approved drugs. This approach would expedite the development of new TGF-ß inhibitors after successful identification of potential drug candidate.

# 1.1 General objective

To identify existing drugs that inhibit TGF- $\beta$  activity upon the bronchial epithelial cell line, BEAS-2B

### 1.2 Specific objectives

- i. To identify potential drug candidates with high binding affinity to TGF- $\beta$  type 1 receptor by using molecular docking
- ii. To determine the effects of selected drug candidates on the cell morphology of TGF-β1-induced BEAS-2B cells
- iii. To determine the effect of selected drug candidates on the EMT markers of TGF-β1-induced BEAS-2B cells

#### REFERENCES

- Ab Ghani, N.S., Ramlan, E.I. and Firdaus-Raih, M. (2019). Drug ReposER: a web server for predicting similar amino acid arrangements to known drug binding interfaces for potential drug repositioning. *Nucleic Acids Research* 47: 350-356.
- Annes, J.P., Munger, J.S. and Rifkin, D.B. (2003). Making sense of latent TGFβ activation. *Journal of Cell Science* 116: 217–224.
- Araujo, S.C., Maltarollo, V.G., Almeida, M.O., Ferreira L.L., Andricopulo, A.D. and Honorio, K.M. (2020). Structure-based virtual screening, molecular dynamics and binding free energy calculations of hit candidates as ALK5 inhibitors. *Molecules* 25: 264.
- Barriere, G., Fici, P., Gallerani, G., Fabbri, F., and Rigaud, M. (2015). Epithelial Mesenchymal Transition: a double-edged sword. *Clinical and Translational Medicine* 4: 1-6.
- Battle, E. and Massague, J. (2019). Transforming Growth Factor-b Signaling in Immunity and Cancer. *Immunity* 50.
- Bhojwani, H.R. and Joshi, U.J. (2019). Selecting Protein Structure/S for Docking-Based Virtual Screening: a Case Study on Type II Inhibitors of Vegfr-2 Kinase. International Journal of Pharmaceutical Sciences and Research 10: 2998–3011.
- Bissantz, C., Folkers, G. and Rognan, D. (2000). Protein-based virtual screening of chemical databases. 1. Evaluation of different docking/scoring combinations. *Journal of Medicinal Chemistry* 43: 4759–4767.
- Bocca, C., Bozzo, F., Cannito, S., Parola, M. and Miglietta, A. (2012). Celecoxib inactivates epithelial-mesenchymal transition stimulated by hypoxia and/or epidermal growth factor in colon cancer cells. *Molecular Carcinogenesis* 51: 783–795.
- Brabletz, T., Pfeuffer, I., Schorr, E., Siebelt, F., Wirth, T. and Serfling, E. (1993). Transforming growth factor beta and cyclosporin A inhibit the inducible activity of the interleukin-2 gene in T cells through a noncanonical octamerbinding site. *Molecular and Cellular Biology* 13: 1155-1162.
- Bueno, L., de Alwis, D.P., Pitou, C., Yingling, J., Lahn, M., Glatt, S. and Trocóniz, I. F. (2008). Semi-mechanistic modelling of the tumour growth inhibitory effects of LY2157299, a new type I receptor TGF-β kinase antagonist, in mice. *European Journal of Cancer* 44: 142–150.
- Bugge, T.H., Flick, M.J., Daugherty, C.C. and Degen, J.L. (1995). Plasminogen deficiency causes severe thrombosis but is compatible with development and reproduction. *Genes and Development* 9: 794–807.

- Byfield, S.D., Major, C., Laping, N.J. and Roberts, A.B. (2004). SB-505124 Is a Selective Inhibitor of Transforming Growth Factor-β Type I Receptors ALK4, ALK5, and ALK7. *Molecular Pharmacology* 65: 744–752.
- Callahan, J.F., Burgess, J.L., Fornwald, J.A., Gaster, L.M., Harling, J.D., Harrington, F.P., Heer, J., Kwon, C., Lehr, R., Mathur, A., Olson, B.A., Weinstock, J. and Laping, N.J. (2002). Identification of novel inhibitors of the transforming growth factor β1 (TGF-β1) type 1 receptor (ALK5). *Journal of Medicinal Chemistry* 45: 999–1001.
- Cano, A., Pérez-Moreno, M.A., Rodrigo, I., Locascio, A., Blanco, M.J., Del Barrio, M.G., Portillo, F. and Nieto, M.A. (2000). The transcription factor Snail controls epithelial-mesenchymal transitions by repressing E-cadherin expression. *Nature Cell Biology* 2: 76–83.
- Chang, Y.C., Tsai, C.H., Lai, Y.L., Yu, C.C., Chi, W.Y., Li, J.J. and Chang, W.W. (2014). Arecoline-induced myofibroblast transdifferentiation from human buccal mucosal fibroblasts is mediated by ZEB1. *Journal of Cellular and Molecular Medicine* 18: 698–708.
- Chen, W.J., Jin, W., Hardegen, N., Lei, K.J., Li, L., Marinos, N., McGrady, G. and Wahl, S.M. (2003). Conversion of Peripheral CD4+CD25- Naive T Cells to CD4+CD25+ Regulatory T Cells by TGF-β Induction of Transcription Factor Foxp3. *Journal of Experimental Medicine* 198: 1875–1886.
- Connor, T.B., Roberts, A.B., Sporn, M.B., Danielpour, D., Dart, L.L., Michels, R.G., Bustros, S., Enger, C., Kato, H. and Lansing, M. (1989). Correlation of fibrosis and transforming growth factor-β type 2 levels in the eye. *Journal of Clinical Investigation* 83: 1661–1666.
- Corbet, C., Bastien, E., Santiago de Jesus, J.P., Dierge, E., Martherus, R., Vander Linden, C., Doix, B., Degavre, C., Guilbaud, C., Petit, L., Michiels, C., Dessy, C., Larondelle, Y. and Feron, O. (2020). TGFβ2-induced formation of lipid droplets supports acidosis-driven EMT and the metastatic spreading of cancer cells. *Nature Communications* 11: 1-15.
- Coutts, A., Chen, G., Stephens, N., Hirst, S., Douglas, D., Eichholtz, T. and Khalil, N. (2001). Release of biologically active TGF-β from airway smooth muscle cells induces autocrine synthesis of collagen. *American Journal of Physiology - Lung Cellular and Molecular Physiology* 280: 999-1008.
- Das, S., Becker, B.N., Hoffmann, F.M. and Mertz, J.E. (2009). Complete reversal of epithelial to mesenchymal transition requires inhibition of both ZEB expression and the Rho pathway. *BMC Cell Biology* 10: 1-18.
- Dituri, F., Mazzocca, A., Peidrò, F.J., Papappicco, P., Fabregat, I., De Santis, F., Paradiso, A., Sabba, C. and Giannelli, G. (2013). Differential Inhibition of the TGF-β Signaling Pathway in HCC Cells Using the Small Molecule Inhibitor LY2157299 and the D10 Monoclonal Antibody against TGF-β Receptor Type II. *PLoS ONE* 8: e67109.

- Dotolo, S., Cervellera, C., Russo, M., Russo, G.L. and Facchiano, A. (2021). Virtual screening of natural compounds as potential pi3k-Akt1 signaling pathway inhibitors and experimental validation. *Molecules* 46: 492.
- Dudley, J. T., Sirota, M., Shenoy, M., Pai, R. K., Roedder, S., Chiang, A. P., Morgan, A.A., Sarwal, M.M., Pasricha, P.J. and Butte, A.J. (2011). Computational repositioning of the anticonvulsant topiramate for inflammatory bowel disease. *Science Translational Medicine* 3: 96ra76.
- Erickson, J.A., Jalaie, M., Robertson, D.H., Lewis, R.A. and Vieth, M. (2004). Lessons in Molecular Recognition: The Effects of Ligand and Protein Flexibility on Molecular Docking Accuracy. *Journal of Medicinal Chemistry* 47: 45–55.
- Felber, R. and Bencheqroun, H. (2014). Ergotamine-Induced Tricuspid Valvulopathy Presenting as Recurrent Pleural Effusions. *Chest* 145: 63A.
- Fetro, C. and Scherman, D. (2020). Drug repurposing in rare diseases: Myths and reality. *Therapie* 75: 157–160.
- Flanders, K.C. and Wakefield, L. M. (2009). Transforming growth Factor-βs and mammary gland involution; functional roles and implications for cancer progression. *Journal of Mammary Gland Biology and Neoplasia* 14: 131–144.
- Gilbert, R.W.D., Vickaryous, M.K. and Viloria-Petit, A.M. (2016). Signalling by transforming growth factor beta isoforms in wound healing and tissue regeneration. *Journal of Developmental Biology* 4: 21.
- GNS, H.S., Saraswathy, G.R., Murahari, M. and Krishnamurthy, M. (2019). An update on Drug Repurposing: Re-written saga of the drug's fate. *Biomedicine and Pharmacotherapy* 110: 700–716.
- Gosse, P. (2006). A review of telmisartan in the treatment of hypertension: Blood pressure control in the early morning hours. *Vascular Health and Risk Management* 2: 195–201.
- Grainger, D.J., Mosedale, D.E. and Metcalfe, J. C. (2000). TGF-β in blood: A complex problem. *Cytokine and Growth Factor Reviews* 11: 133–145.
- Gregory, P.A., Bert, A.G., Paterson, E.L., Barry, S.C., Tsykin, A., Farshid, G., Vadas, M.A., Khew-Goodfall, Y. and Goodall, G.J. (2008). The miR-200 family and miR-205 regulate epithelial to mesenchymal transition by targeting ZEB1 and SIP1. *Nature Cell Biology* 10: 593–601.
- Grooteclaes, M.L. and Frisch, S.M. (2000). Evidence for a function of CtBP in epithelial gene regulation and anoikis. *Oncogene* 19: 3823–3828.
- Guan, L., Rui, W., Bai, R., Zhang W., Zhang, F. and Ding, W. (2016). Effects of Size-Fractionated Particulate Matter on Cellular Oxidant Radical Generation in Human Bronchial Epithelial BEAS-2B Cells. *International Journal of Environmental Research and Public Health* 13(5): 483.

- Hata, A. and Chen, Y.G. (2016). TGF-β signaling from receptors to smads. *Cold Spring Harbor Perspectives in Biology* 8: a022061.
- Hay, E.D. (1995). An overview of epithelio-mesenchymal transformation. *Acta Anatomica* 154: 8–20.
- Heldin, C.H. and Moustakas, A. (2016). Signaling receptors for TGF-β family members. *Cold Spring Harbor Perspectives in Biology*, 8: a022053.
- Horan, G.S., Wood, S., Ona, V., Dan, J.L., Lukashev, M.E., Weinreb, P.H., Simon, K.J., Hahm, K., Allaire, N.E., Rinaldi, N.J. and Violette, S.M. (2008).
  Partial inhibition of integrin αvβ6 prevents pulmonary fibrosis without exacerbating inflammation. *American Journal of Respiratory and Critical Care Medicine* 177: 56–65.
- Huang, Y. xin, Zhao, J., Song, Q. hang, Zheng, L. hua, Fan, C., Liu, T., Bao, Y., Sun, L., Zhang, L. and Li, Y. (2016). Virtual screening and experimental validation of novel histone deacetylase inhibitors. *BMC Pharmacology and Toxicology* 17: 1–14.
- Ikenouchi, J., Matsuda, M., Furuse, M. and Tsukita, S. (2003). Regulation of tight junctions during the epithelium-mesenchyme transition: Direct repression of the gene expression of claudins/occludin by Snail. *Journal of Cell Science* 116: 1959–1967.
- Inman, G.J., Nicolás, F.J., Callahan, J.F., Harling, J.D., Gaster, L.M., Reith, A.D., Laping, N.J. and Hill, C.S. (2002). SB-431542 is a potent and specific inhibitor of transforming growth factor-β superfamily type I activin receptorlike kinase (ALK) receptors ALK4, ALK5, and ALK7. *Molecular Pharmacology* 62: 65–74.
- Jin, C.H., Krishnaiah, M., Sreenu, D., Subrahmanyam, V.B., Rao, K.S., Lee, H. J., Park, S.J., Park, H.J., Lee, K., Sheen, Y.Y. and Kim, D.K. (2014). Discovery of N-((4-([1,2,4]triazolo[1,5-a]pyridin-6-yl)-5-(6-methylpyridin-2-yl)-1H-imidazol-2-yl)methyl)-2-fluoroaniline (EW-7197): A highly potent, selective, and orally bioavailable inhibitor of TGF-β type I receptor kinase as cancer immunotherapeutic. *Journal of Medicinal Chemistry* 57: 4213–4238.
- Jung, M., Lee, C.K., Kim, H.S., Ock, C.Y., Bum, B.J., Lee, J., Kang, D.W., Hwang, S., Kim, S.J., Chung, H.C. and Rha, S.Y. (2020). Safety and preliminary antitumor activity of the transforming growth factor beta (TGFβ) receptor I kinase inhibitor, vactosertib, in combination with paclitaxel in patients with metastatic gastric adenocarcinoma. *Journal of Clinical Oncology* 38: e16505.
- Kalluri, R. and Weinberg, R.A. (2009). Review series The basics of epithelialmesenchymal transition. *The Journal of Clinical Investigation*, 119: 1420-1428.

- Kandagalla S., Sharath, B.S., Bharath B.R., Hani, U. and Manjunatha H. (2017). Molecular docking analysis of curcumin analogues against kinase domain of ALK5. *In silico Pharmacology* 5(1): 15.
- Kass, D.A., Champion, H.C. and Beavo, J.A. (2007). Phosphodiesterase type 5: Expanding roles in cardiovascular regulation. *Circulation Research* 101: 1084-1095.
- Keedy, V.L., Bauer, T.M., Clarke, J.M., Hurwitz, H., Baek, I., Ha, I., Ock, C.Y., Nam, S.Y., Kim, M., Park, N., Kim, J.Y. and Kim, S.J. (2018). Association of TGF-β responsive signature with anti-tumor effect of vactosertib, a potent, oral TGF-β receptor type I (TGFBRI) inhibitor in patients with advanced solid tumors. *Journal of Clinical Oncology* 36: 3031.
- Kelley, R.K., Gane, E., Assenat, E., Siebler, J., Galle, P.R., Merle, P., Hourmand, I.O., Cleverly, A., Zhao, Y., Gueorguieva, I., Lahn, M., Faivre, S., Benhadji, K.A. and Giannelli, G. (2019). A phase 2 study of galunisertib (TGF-b1 Receptor Type i Inhibitor) and sorafenib in patients with advanced hepatocellular carcinoma. *Clinical and Translational Gastroenterology* 10: e00056.
- Khalil, N. (1999). TGF-β: From latent to active. *Microbes and Infection* 1: 1255– 1263.
- Khorramdelazad, H., Hassanshahi, G., Ahmadabadi, B. and Arababadi, K.M. (2012). High serum levels of TGF-β in Iranians with chronic HBV infection. *Hepatitis Monthly* 12: 7581.
- Kim, D.H., Xing, T., Yang, Z., Dudek, R., Lu, Q. and Chen Y.H. (2018). Epithelial Mesenchymal Transition in Embryonic Development, Tissue Repair and Cancer: A Comprehensive Overview. *Journal of Clinical Medicine* 7(1): 1.
- Kim, H.S., Ahn, J.H., Kim, J.E., Hong, J.Y., Lee, J., Kim, S.H., Lee, J., Hwang, S., Jeon, M.K. and Kim, S.J. (2020). A phase I study of TGF-β inhibitor, vactosertib in combination with imatinib in patients with advanced desmoid tumor (aggressive fibromatosis). *Journal of Clinical Oncology* 38: 11557.
- Knust, E. (2002). Regulation of epithelial cell shape and polarity by cell-cell adhesion (Review). *Molecular Membrane Biology* 19: 113–120.
- Ko, T.C., Yu, W., Sakai, T., Sheng, H., Shao, J., Beauchamp, R.D. and Thompson, E.A. (1998). TGF-β1 effects on proliferation of rat intestinal epithelial cells are due to inhibition of cyclin D1 expression. *Oncogene* 16: 3445-3454.
- Kobayashi, T., Kim, H.J., Liu, X., Sugiura, H., Kohyama, T., Fang, Q., Wen, F.Q., Abe, S., Wang, X., Atkinson, J.J., Shipley, J.M., Senior, R.M. and Rennard, S.I. (2014). Matrix metalloproteinase-9 activates TGF-β and stimulates fibroblast contraction of collagen gels. *American Journal of Physiology -Lung Cellular and Molecular Physiology* 306: 1006-1015.

- Koh, R.Y., Lim, C.L., Uhal, B.D., Abdullah, M., Vidyadaran, S., Ho, C.C. and Seow, H.F. (2015). Inhibition of transforming growth factor-β via the activin receptor-like kinase-5 inhibitor attenuates pulmonary fibrosis. *Molecular Medicine Reports* 11: 3808–3813.
- Kulkarni, A.B. and Karlsson, S. (1993). Transforming growth factor-β1 knockout mice: A mutation in one cytokine gene causes a dramatic inflammatory disease. *American Journal of Pathology* 143: 3–9.
- Kuntz, I.D., Blaney, J.M., Oatley, S.J., Langridge, R. and Ferrin, T.E. (1982). A geometric approach to macromolecule-ligand interactions. *Journal of Molecular Biology* 161: 269–288.
- Lee, Y.Z., Yap, H.M., Shaari, K., Tham, C.L., Sulaiman, M.R. and Israf, D.A. (2017). Blockade of Eosinophil-Induced Bronchial Epithelial-Mesenchymal Transition with a Geranyl Acetophenone in a Coculture Model. *Frontiers in Pharmacology* 8: 837.
- Liao, Y., Zhang, M. and Lonnerdal B. (2012). Growth factor TGF-β induces intestinal epithelial cell (IEC-6) differentiation: miR-146b as a regulatory component in the negative feedback loop. *Genes & Nutrition* 8: 69-78.
- Li, H., Wang, H., Wang, F., Gu, Q. and Xu, X. (2011). Snail involves in the transforming growth factor β1-mediated epithelial-mesenchymal transition of retinal pigment epithelial cells. *PLoS ONE* 6: e23322.
- Li, L., Li, H., Zhang, Z., Zheng, J., Shi, Y., Liu, J., Cao, Y., Yuan, X. and Chu, Y. (2018). Recombinant truncated TGF-β receptor II attenuates carbon tetrachloride-induced epithelial-mesenchymal transition and liver fibrosis in rats. *Molecular Medicine Reports* 17: 315–321.
- Liu, L.J., Leung, K.H., Chan, D.S.H., Wang, Y.T., Ma, D.L. and Leung, C.H. (2014). Identification of a natural product-like STAT3 dimerization inhibitor by structure-based virtual screening. *Cell Death and Disease* 5: e1293.
- Liu, X., Wu, Y., Zhou, Z., Huang, M., Deng, W., Wang, Y., Zhou, X., Chen, L., Li, Y., Zeng, T., Wang, G. and Fu, B. (2019). Celecoxib inhibits the epithelialto-mesenchymal transition in bladder cancer via the miRNA-145/TGFBR2/Smad3 axis. *International Journal of Molecular Medicine* 44: 683–693.
- Liu, Z., Zhao, J., Li, W., Shen, L., Huang, S., Tang, J., Duan, J., Fang, F., Huang, Y., Chang, H., Chen, Z. and Zhang, R. (2016). Computational screen and experimental validation of anti-influenza effects of quercetin and chlorogenic acid from traditional Chinese medicine. *Scientific Reports* 6: 1– 9.
- Lu, Z. (2008, July). Second generation HIV protease inhibitors against resistant virus. *Expert Opinion on Drug Discovery* 3: 775–786.

- Luangmonkong, T., Suriguga, S., Bigaeva, E., Boersema, M., Oosterhuis, D., de Jong, K.P., Schuppan, D., Mutsaers, H.A.M. and Olinga, P. (2017). Evaluating the antifibrotic potency of galunisertib in a human ex vivo model of liver fibrosis. *British Journal of Pharmacology* 174: 3107–3117.
- Massagué, J. (2012). TGFβ signalling in context. *Nature Reviews Molecular Cell Biology* 13: 616–630.
- McCabe, R.P., Secrist, H., Botney, M., Egan, M. and Peters, M.G. (1993). Cytokine mRNA expression in intestine from normal and inflammatory bowel disease patients. *Clinical Immunology and Immunopathology* 66: 52–58.
- Meng, X.Y., Zhang, H.X., Mezei, M. and Cui, M. (2011). Molecular Docking: A powerful approach for structure-based drug discovery. *Current Computer-Aided Drug Design* 7: 146-157.
- Minshall, E.M., Leung, D.Y.M., Martin, R.J., Song, Y.L., Cameron, L., Ernst, P. and Hamid, Q. (1997). Eosinophil-associated TGF-β1 mRNA Expression and Airways Fibrosis in Bronchial Asthma. *American Journal of Respiratory Cell and Molecular Biology* 17: 326–333.
- Miyoshi, S., Kudo, M., Shitara, K., Yamauchi, M., Doi, T. and Matsumura, Y. (2016). TGF- $\beta$  inhibitor LY2157299 (galunisertib) in combination with standard chemotherapy and inhibition of signaling to pSmad and EMT and suppression of tumor growth in gastric cancer. *Journal of Clinical Oncology* 34: 50.
- Molina-Molina, M., Lario, S., Luburich, P., Ramírez, J., Carrión, M.T. and Xaubet, A. (2006). Quantifying Plasma Levels of Transforming Growth Factor β1 in Idiopathic Pulmonary Fibrosis. *Archivos de Bronconeumología ((English Edition))* 42: 380–383.
- Moreno-Bueno, G., Cubillo, E., Sarrió, D., Peinado, H., Rodríguez-Pinilla, S.M., Villa, S., Bolós, V., Jorda, M., Fabra, A., Portillo, F., Palacios, J. and Cano, A. (2006). Genetic profiling of epithelial cells expressing E-cadherin repressors reveals a distinct role for snail, Slug, and E47 Factors in epithelial- mesenchymal transition. *Cancer Research* 66: 9543–9556.
- Mori, Y., Ishida, W., Bhattacharyya, S., Li, Y., Platanias, L.C. and Varga, J. (2004). Selective inhibition of activin receptor-like kinase 5 signaling blocks profibrotic transforming growth factor  $\beta$  responses in skin fibroblasts. *Arthritis and Rheumatism* 50: 4008–4021.
- Morris, G.M., Ruth, H., Lindstrom, W., Sanner, M.F., Belew, R.K., Goodsell, D.S. and Olson, A.J. (2009). Software news and updates AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. *Journal of Computational Chemistry* 30: 2785–2791.

- Munger, J.S., Huang, X., Kawakatsu, H., Griffiths, M.J.D., Dalton, S.L., Wu, J., Pittet, J.F., Kaminski, N., Garat, C., Matthay, M.A., Rifkin, D.B. and Sheppard, D. (1999). The integrin  $\alpha\nu\beta6$  binds and activates latent TGF $\beta1$ : A mechanism for regulating pulmonary inflammation and fibrosis. *Cell* 96: 319–328.
- Munger, J.S. and Sheppard, D. (2011). Cross talk among TGF-β signaling pathways, integrins, and the extracellular matrix. *Cold Spring Harbor Perspectives in Biology* 3: a0050107.
- Murakami, K., Takemura, T., Hino, S. and Yoshioka, K. (1997). Urinary transforming growth factor-β in patients with glomerular diseases. *Pediatric Nephrology* 11: 334–336.
- Murphy-Ullrich, J.E. and Suto, M.J. (2018). Thrombospondin-1 regulation of latent TGF-β activation: A therapeutic target for fibrotic disease. *Matrix Biology* 68: 28–43.
- Mysinger, M.M., Carchia, M., Irwin, J.J. and Shoichet, B.K. (2012). Directory of useful decoys, enhanced (DUD-E): Better ligands and decoys for better benchmarking. *Journal of Medicinal Chemistry* 55: 6582–6594.
- Ogunjimi, A.A., Zeqiraj, E., Ceccarelli, D.F., Sicheri, F., Wrana, J.L. and David, L. (2012). Structural Basis for Specificity of TGF beta Family Receptor Small Molecule Inhibitors. *Cell Signal* 24: 476–483.
- Olsson, P.O., Gustafsson, R., Salnikov, A.V., Gothe, M., Zeller, K.S., Friman, T., Baldetorp, B., Koopman, L.A., Weinreb, P.H., Violette S.M., Kalamajski, S., Heldin, N.E. and Rubin, K. (2018). Inhibition of integrin αVβ6 changes fibril thickness of stromal collagen in experimental carcinomas. *Cell Communication and Signaling* 16:36.
- Pakyari, M., Farrokhi, A., Maharlooei, M.K. and Ghahary, A. (2013). Critical Role of Transforming Growth Factor Beta in Different Phases of Wound Healing. *Advances in Wound Care* 2: 215–224.
- Park, S.A., Kim, M.J., Park, S.Y., Kim, J.S., Lee, S.J., Woo, H.A., Kim, D.K., Nam, J.S. and Sheen, Y.Y. (2015). EW-7197 inhibits hepatic, renal, and pulmonary fibrosis by blocking TGF-β/Smad and ROS signaling. *Cellular and Molecular Life Sciences* 72: 2023–2039.
- Petersen, M., Thorikay, M., Deckers, M., Van Dinther, M., Grygielko, E.T., Gellibert, F., Gouville, A.C., Huet, S., Dijke, P. and Laping, N.J. (2008). Oral administration of GW788388, an inhibitor of TGF-β type I and II receptor kinases, decreases renal fibrosis. *Kidney International* 73: 705–715.
- Postigo, A.A., Depp, J.L., Taylor, J.J. and Kroll, K.L. (2003). Regulation of Smad signaling through a differential recruitment of coactivators and corepressors by ZEB proteins. *EMBO Journal* 22: 2453–2462.

- Prud'homme, G.J. (2007). Pathobiology of transforming growth factor  $\beta$  in cancer, fibrosis and immunologic disease, and therapeutic considerations. *Laboratory Investigation* 87: 1077-1091.
- Pushpakom, S., Iorio, F., Eyers, P. A., Escott, K. J., Hopper, S., Wells, A., Doig, A., Guiliams, T., Latimer, J., McNamee, C., Norris, A., Sanseau, P., Cavalla, D. And Pirmohamed, M. (2018). Drug repurposing: Progress, challenges and recommendations. *Nature Reviews Drug Discovery* 18: 41– 58.
- Ramírez, D. and Caballero, J. (2018). Is It Reliable to Take the Molecular Docking Top Scoring Position as the Best Solution without Considering Available Structural Data? *Molecules* 23: 1038.
- Ramos, F.S., Wons, L., Cavalli, I.J., and Ribeiro, E.M.S.F. (2017). Epithelialmesenchymal transition in cancer: An overview. *Integrative Cancer Science and Therapeutics* 4: 1-5.
- Réau, M., Langenfeld, F., Zagury, J.F., Lagarde, N. and Montes, M. (2018). Decoys selection in benchmarking datasets: Overview and perspectives. *Frontiers in Pharmacology* 9: 11.
- Roberts, A.B., Sporn, M.B., Assoian, R.K., Smith, J.M., Roche, N.S., Wakefield, L.M., Heine, U.I., Liotta, L.A., Falanga, V. and Kehrl, J.H. (1986). Transforming growth factor type β: Rapid induction of fibrosis and angiogenesis in vivo and stimulation of collagen formation in vitro. *Proceedings of the National Academy of Sciences of the United States of America* 83: 4167–4171.
- Robertson, I.B., Horiguchi, M., Zilberberg, L., Dabovic, B., Hadjiolova, K. and Rifkin, D.B. (2015). Latent TGF-β-binding proteins. *Matrix Biology* 47: 44–53.
- Rodon, J., Carducci, M.A., Sepulveda-Sánchez, J.M., Azaro, A., Calvo, E., Seoane, J., Brana, I., Sicart, E., Gueorguieva, I., Cleverly, A.L., Pillay, N.S., Desaiah, D., Estrem, S.T., Paz-Ares, L., Holdoff, M., Blakeley, J., Lahn, M.M. and Baselga, J. (2015). First-in-human dose study of the novel transforming growth factor-β receptor I kinase inhibitor LY2157299 monohydrate in patients with advanced cancer and glioma. *Clinical Cancer Research* 21: 553–560.
- Sanchez, C.G., Molinski, S.V., Gongora, R., Sosulski, M., Fuselier, T., MacKinnon, S.S., Mondal, D. and Lasky, J.A. (2018). The Antiretroviral Agent Nelfinavir Mesylate: A Potential Therapy for Systemic Sclerosis. *Arthritis and Rheumatology* 70: 115–126.
- Santibanez, J.F., Obradović, H., Kukolj, T. and Krstić, J. (2018). Transforming growth factor-β, matrix metalloproteinases, and urokinase-type plasminogen activator interaction in the cancer epithelial to mesenchymal transition. *Developmental Dynamics* 247: 382–395.

- Sawyer, J.S., Beight, D.W., Britt, K.S., Anderson, B.D., Campbell, R.M., Goodson, T., Herron, D.K., Li, H.Y., McMillen, W.T., Mort, N., Parsons, S., Smith, E.C.R., Wagner, J.R., Yan, L., Zhang, F. and Yingling, J.M. (2004). Synthesis and activity of new aryl- and heteroaryl-substituted 5,6-dihydro-4H-pyrrolo[1,2-b]pyrazole inhibitors of the transforming growth factor-β type I receptor kinase domain. *Bioorganic and Medicinal Chemistry Letters* 14: 3581–3584.
- Shi, M., Zhu, J., Wang, R., Chen, X., Mi, L., Walz, T. and Springer, T.A. (2011). Latent TGF-β structure and activation. *Nature* 474: 343-351.
- Shull, M.M., Ormsby, I., Kier, A.B., Pawlowski, S., Diebold, R.J., Yin, M., Allen, Y., Sidman, C., Proetzel, G., Calvin, D., Annunziata, N. and Doetschman, T. (1992). Targeted disruption of the mouse transforming growth factor-β1 gene results in multifocal inflammatory disease. *Nature* 359: 693–699.
- Si, D., Moritz, S.A., Pfab, J., Hou, J., Cao, R., Wang, L., Wu, T., and Cheng, J. (2020). Deep Learning to Predict Protein Backbone Structure from High-Resolution Cryo-EM Density Maps. *Scientific Reports* 10: 4282.
- Smith, B.N., Burton, L.J., Henderson, V., Randle, D.D., Morton, D.J., Smith, B.A., Taliaferro-Smith, L., Nagappan, P., Yates, C., Zayzafoon, M., Chung, L.W.K. and Odero-Marah, V. A. (2014). Snail Promotes Epithelial Mesenchymal Transition in Breast Cancer Cells in Part via Activation of Nuclear ERK2. *PLoS ONE* 9: e104987.
- Son, J.Y., Park, S.Y., Kim, S.J., Lee, S.J., Park, S.A., Kim, M.J., Kim, S.W., Kim, D.K., Nam, J.S. and Sheen, Y.Y. (2014). EW-7197, a novel ALK-5 kinase inhibitor, potently inhibits breast to lung metastasis. *Molecular Cancer Therapeutics* 13: 1704–1716.
- Song, K.M., Chung, D.Y., Choi, M.J., Ghatak, K., Nguyen, N.M., Limanjaya, A., Kwon, M.H., Ock, J., Yin, G.N., Kim, D.K., Ryu, J.K. and Suh, J.K. (2019). Vactosertib, a Novel, Orally Bioavailable Activin Receptor-Like Kinase 5 Inhibitor, Promotes Regression of Fibrotic Plaques in a Rat Model of Peyronie's Disease. *The World Journal of Men's Health* 38: 552-563.
- Stierand, K. and Rarey, M. (2010). PoseView molecular interaction patterns at a glance. *Journal of Cheminformatics* 2: 50.
- Stone, R.C., Pastar, I., Ojeh, N., Chen, V., Liu, S., Garzon, K.I., and Tomic-Canic, M. (2016). Epithelial-mesenchymal transition in tissue repair and fibrosis. *Cell and Tissue Research* 365: 495–506.
- Sutherland, J.J., Nandigam, R.K., Erickson, J.A. and Vieth, M. (2007). Lessons in molecular recognition. 2. Assessing and improving cross-docking accuracy. *Journal of Chemical Information and Modeling* 47: 2293–2302.
- Suzuki, D., Pinto, F. and Senoo, M. (2017). Inhibition of TGF-β signaling supports high proliferative potential of diverse p63+ mouse epithelial progenitor cells in vitro. *Scientific Reports* 7: 6089.

- Terashima, H., Aonuma, M., Tsuchida, H., Sugimoto, K., Yokoyama, M. and Kato, M. (2019). Attenuation of pulmonary fibrosis in type I collagentargeted reporter mice with ALK-5 inhibitors. *Pulmonary Pharmacology and Therapeutics* 54: 31–38.
- Terashima, H., Kato, M., Ebisawa, M., Kobayashi, H., Suzuki, K., Nezu, Y. and Sada, T. (2014). R-268712, an orally active transforming growth factor-β type i receptor inhibitor, prevents glomerular sclerosis in a Thy1 nephritis model. *European Journal of Pharmacology* 734: 60–66.
- Tfelt-Hansen, P. (2000). Ergotamine in the acute treatment of migraine: A review and European consensus. *Brain* 123: 9–18.
- Trott, O. and Olson, A.J. (2009). AutoDock Vina: Improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. *Journal of Computational Chemistry* 31: 455-461.
- Verkhivker, G.M., Bouzida, D., Gehlhaar, D.K., Rejto, P.A., Freer, S.T. and Rose, P. W. (2002). Complexity and simplicity of ligand-macromolecule interactions: The energy landscape perspective. *Current Opinion in Structural Biology* 12: 197–203.
- Wang, Y.L., Zhao, X.M., Shuai, Z.F., Li, C.Y., Bai, Q.Y., Yu, X.W. and Wen, Q.T. (2015). Snail promotes epithelial-mesenchymal transition and invasiveness in human ovarian cancer cells. *International Journal of Clinical and Experimental Medicine* 8: 7388–7393.
- Wang, X.J., Liefer, K.M., Tsai, S., O'Malley, B.W. and Roop, D.R. (1999). Development of gene-switch transgenic mice that inducibly express transforming growth factor β1 in the epidermis. *Proceedings of the National Academy of Sciences of the United States of America* 96: 8483–8488.
- Xu, H., Aldrich, M.C., Chen, Q., Liu, H., Peterson, N.B., Dai, Q., Levy, M., Shah, A., Han, X., Ruan, X., Jiang, M., Li, Y., Julien, J., Warner, J., Friedman, C., Roden, D.M. and Denny, J.C. (2015). Validating drug repurposing signals using electronic health records: A case study of metformin associated with reduced cancer mortality. *Journal of the American Medical Informatics Association* 22: 179–191.
- Xu, H., Qu, J., Wang, J., Han, K., Li, Q., Bi., W. and Liu, R. (2020). Discovery of pulmonary fibrosis inhibitor targeting TGF-β RI in Polygonum cuspidatum by high resolution mass spectrometry with in silico strategy. *Journal of Pharmaceutical Analysis*.
- Yingling, J. (2005). Targeting the TGF-ß RI kinase with LY2157299: A PK/PDdriven drug discovery and clinical development program. *Cancer Research* 46: 1463.

- Yumin, C., Qiong, L., Zibo, X., Wei, L., Li, C. and Zuying, X. (2012). Telmisartan counteracts TGF-β1 induced epithelial-to-mesenchymal transition via PPAR-γ in human proximal tubule epithelial cells. *International Journal of Clinical and Experimental Pathology* 5: 522–529.
- Zeisberg, M. and Neilson, E. G. (2009). Biomarkers for epithelial-mesenchymal transitions. *Journal of Clinical Investigation* 119: 1429–1437.
- Zha, D., Wu, S., Gao, P. and Wu, X. (2019). Telmisartan Attenuates Uric Acid-Induced Epithelial-Mesenchymal Transition in Renal Tubular Cells. *BioMed Research International* 2019: 3851718.
- Zhao, B.M. and Hoffmann, F.M. (2006). Inhibition of transforming growth factorβ1-induced signaling and epithelial-to-mesenchymal transition by the Smad-binding peptide aptamer Trx-SARA. *Molecular Biology of the Cell* 17: 3819–3831.
- Zulfiqar, Z., Shah, F.A., Shafique, S., Alattar, A., Ali, T., Alvi, A.M., Rashid, S. and Li, S. (2020). Repurposing fda approved drugs as jnk3 inhibitor for prevention of neuroinflammation induced by mcao in rats. *Journal of Inflammation Research* 13: 1185–1205.