



UNIVERSITI PUTRA MALAYSIA

***Saccharum officinarum L. BY-PRODUCTS (PITH AND RIND) AS
POTENTIAL SOURCE OF PREBIOTIC IN VITRO***

DINA ABDUL GHANI ZIDAN

FPSK(m) 2021 42



***Saccharum officinarum* L. BY-PRODUCTS (PITH AND RIND) AS POTENTIAL
SOURCE OF PREBIOTIC *IN VITRO***

By

DINA ABDUL GHANI ZIDAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in
Fulfilment of the Requirements for the Degree of Master of Science**

November 2019

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of
the requirement for the degree of Master of Science

***Saccharum officinarum L. BY-PRODUCTS (PITH AND RIND) AS POTENTIAL
SOURCE OF PREBIOTIC IN VITRO***

By

DINA ABDUL GHANI ZIDAN

November 2019

Chairman : Mohd Redzwan bin Sabran, PhD
Faculty : Medicine and Health Sciences

Agricultural residues are normally present in massive amounts, and their costs of disposal are very high. Notwithstanding their rich nutritional values, the wastes remain under-utilized commercially due to inadequate study. Sugarcane (*Saccharum officinarum L.*) by-products; rind (SR) and pith (SP) promote well-being, probably as natural ingredients for functional and medical food industry. The aims of the research were to evaluate the health benefits and functional properties of Sugarcane by-products (SB) through the estimation of their chemical compositions, antioxidant capacities (DPPH and FRAP assays) and phytochemical compounds (phenolic and flavonoids) as well as to evaluate the prebiotic properties of enzymatically produced xylooligosaccharide (XOS) including; non-digestibility experiment using α -amylase and artificial gastric acid juice and selective fermentation using pure cultures namely: *Lactobacillus casei* Shirota (LcS), *Bifidobacterium animals* subsp. *Lactis* and *Escherichia coli*. Both SR and SP were found to have high total dietary fiber contents (58.91% to 69.10% DW, respectively), with high proportion of insoluble dietary fiber. The available carbohydrate content (AC) in SR ($31.72\% \pm 0.47$ DW) was significantly ($p<0.05$) higher compared to SP ($23.07\% \pm 0.45$ DW). The high content of dietary fiber and available carbohydrate are characteristics that made SB as fermentable potential food. The result of antioxidant activities revealed that SR had significantly higher radical scavenging activity ($62\% \pm 1.06$) than SP ($51.32\% \pm 1.18$) using the DPPH assay and higher reducing power activity (6.98 ± 0.30 mg FeSO₄/100g DW), than SP (2.77 ± 0.32 mg FeSO₄/100g DW) using the FRAP assay. In addition, SB showed high total phenolic contents (SR= 469 ± 3.25 ; SP= 273 ± 4.27 mg GA/100g), and total flavonoids contents (SR= 269 ± 16.80 ; SP= 169 ± 4.36 mg QE /100g), being significantly higher in SR ($p<0.05$). This study showed that SB contained XOS known as prebiotic agent. In the

present study, XOS mixture was successfully produced from SR and SP (XOS-SR and XOS-SP) respectively. XOS-SR was composed of 1.78 ± 1.01 and 1.62 ± 0.02 mg/mL xylobiose and xylotriose respectively, while XOS-SP was composed of 1.75 ± 0.05 , 1.04 ± 0.06 and 0.19 ± 0.02 mg/mL xylobiose, xylotetrose and xylotriose, respectively. Based on the non-digestibility test, around 84 to 88.24% of XOS consumed would reach the colon and then be utilized by the probiotic as small amounts of the saccharides were hydrolyzed by α -amylase (8.29 to 12.87%) and by gastric juice (3.47 to 4.07%). The fermentation test demonstrated that XOS derived from SB is a potential prebiotic ingredient due to the evidence that probiotic bacteria (LcS and *Bifidobacterium*) increased significantly ($p < 0.05$), while *E. coli* growth was suppressed *in vitro* condition. In addition, the probiotic bacteria supplemented with XOS produced high amount of SCFAs, known as fermentation end products during the fermentation, indicating the ability of tested bacteria to utilize XOS-SR and XOS-SP, where acetic acid was the predominant end product, followed by propionic acid and lactic acid. In conclusion, sugarcane by-products are potential ingredients for development of functional foods.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

Saccharum officinarum L. (EMPULUR DAN KULIT) SEBAGAI SUMBER BERPOTENSI PREBIOTIK IN VITRO

Oleh

DINA ABDUL GHANI ZIDAN

September 2019

Pengerusi : Mohd Redzwan bin Sabran, PhD
Fakulti : Perubatan dan Sains Kesihatan

Residu buah-buahan dan produk akhir mereka biasanya boleh diperoleh dalam kuantiti yang banyak, dan kos pelupusan mereka adalah sangat tinggi. Walaubagaimana pun nilai pemakanan mereka kaya, sisa mereka tidak dieksplotasi secara komersial sepenuhnya disebabkan kekurangan penyelidikan. Produk akhir tebu (*Saccharum officinarum L.*) iaitu, kulit (SR) dan empulur (SP) memberikan ciri penggalakan kesihatan, berpotensi sebagai bahan semula jadi baharu bagi industri industri makanan fungsian dan kesihatan. Objektif kajian ini adalah untuk menilai manfaat kesihatan dan ciri fungsian produk akhir tebu melalui penganggaran komposisi kimia, kapasiti antioksidan (ujian DPPH dan FRAP) dan sebatian fitokimia mereka (fenolik dan flavonoid) di samping untuk menilai ciri prebiotik hasil enzim *Xylooligosaccharide* (XOS) termasuk; eksperimen ketidakcerneaan menggunakan α -amylase dan jus gastrik tiruan dan penapaian selektif menggunakan kultur tulen iaitu, *Lactobacillus casei* Shirota (LcS), *Bifidobacterium animals* subsp. *Lactis* dan *Escherichia coli*. Kedua-dua SR dan SP didapati mempunyai kandungan jumlah serat makanan yang tinggi (masing-masing 58.91% hingga 69.10% DW), dengan perkadaruan serat tidak larut yang tinggi. Kandungan karbohidrat tersedia (AC) di dalam SR ($31.72\% \pm 0.47$ DW) adalah tinggi secara signifikan ($p < 0.05$) daripada SP ($23.07\% \pm 0.45$ DW). Kandungan serat makanan yang tinggi dan karbohidrat tersedia merupakan ciri yang menjadikan produk akhir tebu sebagai makanan berpotensi untuk penapaian. Dapatkan aktiviti antioksidan menunjukkan bahawa SR mempunyai aktiviti radikal hapus sisa yang secara signifikan lebih tinggi ($62\% \pm 1.06$) daripada SP ($51.32\% \pm 1.18$) menggunakan ujian DPPH. Demikian juga, SR menunjukkan aktiviti pengurangan kuasa yang secara signifikan lebih tinggi (6.98 ± 0.30 mg FeSO₄/100g DW), daripada SP (2.77 ± 0.32 FeSO₄/100g DW) menggunakan ujian FRAP. Tambahan pula, SB menunjukkan kandungan fenolik total yang tinggi (SR= 469 ± 3.25 ; SP= $273 \pm$

4.27 GA mg/100g), dan kandungan flavonoid total (SR= 269 ± 16.80 ; SP= 169 ± 4.36 QE mg/100g), adalah secara signifikan lebih tinggi dalam SR ($p<0.05$). Kajian ini menunjukkan bahawa Produk akhir tebu mengandungi XOS, dikenali sebagai agen prebiotik. Untuk kajian ini, campuran XOS telah berjaya dihasilkan daripada SR dan SP (XOS-SR dan XOS-SP). XOS-SR terdiri daripada masing-masing, 1.78 ± 1.01 dan 1.62 ± 0.02 mg/mL xylobiosa dan xylotriosa, manakala XOS-SP terdiri daripada, masing-masing, 1.75 ± 0.05 , 1.04 ± 0.06 dan 0.19 ± 0.02 mg/mL xylobiosa, xylotetrosa dan xylotriosa. Berdasarkan eksperimen ketidakceraan, sekitar 84 hingga 88.24% XOS yang dimakan akan sampai ke kolon dan kemudian digunakan oleh populasi probiotik disebabkan jumlah kecil sakrida telah dicerna oleh α -amylase (8.29 hingga 12.87%) dan jus gastrik (3.47 hingga 4.07%). Ujian fermentasi memperlihatkan bahawa XOS yang diperoleh daripada SB merupakan bahan berpotensi prebiotik disebabkan bukti yang menyatakan bahawa bakteria probiotik (LcS dan *Bifidobacterium*) meningkat secara signifikan ($p<0.05$), manakala pertumbuhan *E. coli* telah disekat *in vitro*. Tambahan pula, bakteria probiotik yang digabung dengan XOS menghasilkan jumlah SCFA yang tinggi ketika fermentasi, memperlihatkan kebolehan bakteria yang diuji untuk menggunakan XOS-SR dan XOS-SP, di mana asid asetik merupakan produk akhir yang dominan dihasilkan, diikuti oleh asid propionik dan asid laktik. Kesimpulannya, produk akhir tebu merupakan bahan yang berpotensi untuk pembangunan makanan fungsian.

ACKNOWLEDGEMENTS

First of all, I would like to express my sincere and heartfelt gratitude to the chairman of my supervisory committee Dr. Mohd Redzwan bin Sabran for his invaluable guidance and ideal leadership in the course of this research. Without his help, this work would not have been possible. I would also appreciate the guidance and supervision of Dr. Nurul Shazini Ramli.

I acknowledge the support of UPM that funded this project under the Putra Graduate Initiative (IPS) grant (Project No: GP-IPS/2018/9615800).

I am deeply thankful to others: Nutrition and Dietetics Laboratory staffs, En. Syed Hasbullah Syed, En. Shahidan bin Sulaiman, En. Eddy Ghadaffie Jamiauddin, Mrs. Che Maznah Ahmad for their enthusiastic assistance and support in using the various laboratory and equipment. I acknowledge En. Mohd Sawal Taib and Mrs. Fatimah Mokhtar for their help during my work in Food Engineering Laboratory; Mrs. Noridah Md and Mrs. Siti Nur Aishah Ramlee for their assistance during my work in Anatomy & Histology Research Laboratory.

I would like to thank Winnie Liew and Nurul Adilah Zainuddin as researchers under the supervision of Dr. Mohd Redzwan for their friendship and help over the years. My gratitude also goes to my post graduate friends who I met during my years as a student in Malaysia who offered warm friendship, encouragement, wisdom and help particularly Mr. Ali Abdulqader Mahdi, Mr. AbdulRahman Muthanna, Mrs. Lee Siew Siew and Choo Sulin.

I want to seize this opportunity to immensely thank my beloved parents and other family members for their kind support, encouragement, understanding and patience in this journey.

Finally, I would love to extend my warm regards to my pretty country, Palestine Enthusiastic, Revolutionary Youth and Soul of Martyrs.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mohd Redzwan bin Sabran, PhD

Senior Lecturer

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Chairman)

Nurul Shazini Ramli, PhD

Senior Lecturer

Faculty of Food Science and Technology

Universiti Putra Malaysia

(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 14 January 2021

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of
Chairman of
Supervisory
Committee: _____
Dr. Mohd Redzwan bin Sabran

Signature: _____

Name of Member
of Supervisory
Committee: _____
Dr. Nurul Shazini Ramli

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iv
ACKNOWLEDGEMENTS	vii
APPROVAL	x
DECLARATION	xii
LIST OF TABLES	xv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xvii
CHAPTER	
1 INTRODUCTION	1
1.1 Background	1
1.2 Problem Statement	2
1.3 Significance of Study	2
1.4 Objectives	3
1.4.1 General Objective	3
1.4.2 Specific Objectives	3
2 LITERATURE REVIEW	4
2.1 Saccharum spp.	4
2.1.1 General Information	4
2.1.2 Sugarcane Varieties	6
2.1.3 Processing of Sugarcane and its Products	7
2.1.4 Nutritional Composition and Bioactive Compounds of Sugarcane	7
2.2 Antioxidant activity	8
2.3 Prebiotic	9
2.3.1 Definition and Criteria of Prebiotic	9
2.3.2 Oligosaccharides	10
2.3.3 Type and Sources of Prebiotic	11
2.3.4 Digestibility of prebiotic	12
2.3.5 Fermentability and Selectivity of Prebiotic	13
2.3.6 Prebiotic and gut system	14
2.3.7 Short chain fatty acids (SCFAs)	15
2.4 Agriculture by-Products as Prebiotic Source	16
2.4.1 Xylooligosaccharide as prebiotic	17
2.4.2 Lignocellulose biomass for xylan fractionation	18
2.4.3 Production of Xylooligosaccharide	19
2.5 Probiotic	22
2.5.1 Definition and Criteria of Probiotic	22
2.5.2 <i>Lactobacillus</i> and <i>Bifidobacterium</i>	24
3 MATERIALS AND METHODS	26
3.1 Materials and Chemicals	26

3.1.1	Chemical and Reagents	26
3.1.2	Sampling of <i>S. officinarum</i> L.	26
3.2	Methods	28
3.2.1	Proximate Composition of <i>Saccarum officinarum</i> L. by-Products	28
3.2.2	Total phenolic content, total flavonoids content and antioxidant capacities of <i>S. officinarum</i> L. by-products	33
3.2.4	Prebiotic Properties of Xylooligosaccharide Derived from <i>S. officinarum</i> L. by-Products	37
3.3	Statistical and Analysis	42
4	RESULTS AND DISCUSSION	43
4.1	Proximate Composition	43
4.2	Bioactive Compounds and Antioxidant Capacities	44
4.2.1	Diphenyl-l-picrylhydrazyl (DPPH) Radical Scavenging Activity	44
4.2.2	Ferric Ion Reducing Antioxidant Power (FRAP)	45
4.2.3	Total Phenolic (TPC) and Flavonoids Content (TFC)	47
4.3	Enzymatic Production of xylooligosaccharide Derived from <i>S. officinarum</i> L. by-Products	49
4.3.1	Extraction of Xylan	49
4.3.2	Enzymatic Production of Xylooligosaccharide	50
4.4	Prebiotic Properties of Xylooligosaccharide Derived from <i>S. officinarum</i> L. Rind (XOS-SR) and Pith (XOS-SP)	52
4.4.1	Effect of α -Amylase Hydrolysis	52
4.4.2	Effect of Human Gastric Acid Juice Hydrolysis	54
4.4.3	Selectivity of Fermentation of Xylooligosaccharide	56
4.4.4	Acidifying Activity and Production of Short Chain Fatty Acids (SCFAs)	60
5	CONCLUSION, LIMITATIONS AND RECOMMENDATIONS	64
5.1	Conclusion	64
5.2	Limitations	65
5.3	Recommendation	65
REFERENCES		66
APPENDICES		84
BIODATA OF STUDENT		94

LIST OF TABLES

Table		Page
2.1	Characteristics of Six Species of <i>Saccharum</i>	6
2.2	Oligosaccharides with Bifidogenic Action	11
2.3	Natural Prebiotic Compounds	12
2.4	Potential Probiotic Strains	25
3.1	Components of Different Media Broth	39
3.2	Sample Used As Substrates in Fermentation	40
4.1	Proximate Composition of <i>Saccharum Officinarum L.</i> By-Products (G Per100g)	43
4.2	TPC, TFC, FRAP, DPPH of <i>S. officinarum L.</i> by-products.	46
4.3	Concentration of Carbohydrate During Enzymatic Production of Xylooligosaccharide from <i>S. officinarum L.</i> by-Products	49
4.4	The Growth of Selected Probiotic and <i>E. coli</i> in XOS, Inulin and Glucose Over Fermentation Time (log CFU/mL)	57
4.5	The pH Level and Organic Acid Concentration (mM) After 48h of Fermentation Selected Probiotic and <i>E. coli</i> with Different Carbon Source	61

LIST OF FIGURES

Figure		Page
2.1	Production Share of Sugarcane by Region	4
2.2	Portion of Stem Showing Details of Node and Internode	5
2.3	Mechanism of Action Prebiotic	15
2.4	Production of Xylooligosaccharide from Agriculture Waste	20
3.1	Processing of <i>S. officinarum</i> L. by-Products	27
3.2	Analytical Scheme for Soluble and Insoluble Dietary Fiber Determination Procedures	32
3.3	Research Flow Chart	36
3.4	Diagram of Fermentation of Selected Strains in Media, Inulin, XOS-SR, XOS-SP	41
4.1	Scavenging Activity of <i>S. officinarum</i> L by-Products at Different Concentrations (0.2, 0.4, 0.6, 0.8, 1 mg/mL)	45
4.2	Reducing Power Capacities of <i>S. officinarum</i> L. by-Products	47
4.3	HPLC Chromatograms of XOS Produced from <i>S. officinarum</i> L. by-Products	51
4.4	Degree of Hydrolysis of XOS and Inulin by α -Amylase	53
4.5	Degree of hydrolysis of XOS and inulin by Artificial Human Gastric Juice	55

LIST OF ABBREVIATIONS

%	Percentage
μ L	Microliter
AC	Available carbohydrate
ANOVA	Analysis of Variance
AOAC	Association of Official Analytical Chemists
ATCC	American Type Culture Collection
<i>B.</i>	<i>Bifidobacterium</i>
BHT	Butylated Hydroxytoluene
CFU	Colony Forming Unit
DP	Degree of Polymerization
DPPH	2,2- Diphenyl-1-Picrylhydrazyl
DW	Dry weight
<i>E.</i>	<i>Escherichia</i>
FAO	Food and Agriculture Organization
FOS	Fructooligosaccharide
FRAP	Ferric Reducing Antioxidant Power
g	Gram
h	Hour
IDF	Insoluble Dietary Fiber
kg	Kilogram
L	Liter
<i>L.</i>	<i>Lactobacillus</i>
LAB	Lactic Acid Bacteria
LcS	<i>Lactobacillus casei</i> Shirota
Log	Logarithm
M	Molar
mg	milligram
min	Minute
mL	Milliliter
mM	Millimolar
MRS	De Man Rogosa Sharpe

MW	Molecular Weight
nd	Not detected
RCM	Reinforced Clostridial Medium
rpm	Route Per Minute
SB	<i>S. officinarum</i> L. By-products
SCFAs	Short Chain Fatty Acids
SD	Standard Deviation
SDF	Soluble Dietary Fiber
SP	<i>S. officinarum</i> L. Pith
SR	<i>S. officinarum</i> L. Rind
ssp.	Sub-species
TDF	Total Dietary Fiber
TFC	Total Flavonoids Content
TPC	Total Phenolic Content
v/v	volume/ volume
w/v	weight/ volume
WHO	World Health Organization
XOS	Xylooligosaccharide
XOS-SP	Xylooligosaccharide from <i>S. officinarum</i> L. Pith
XOS-SR	Xylooligosaccharide from <i>S. officinarum</i> L. Rind
α -	Alpha-
β -	Beta-

CHAPTER 1

INTRODUCTION

1.1 Background

Sugarcane (*Saccharum* spp.) is one of the most crucial crops in tropical and subtropical areas with immense industrial applications especially in sugar and alcohol industries (Henry & Kole, 2010). Over 1000 million tons of sugarcane are harvested annually and the level of its production excelled among the major grain food crops, i.e. wheat, rice and maize, which have annual production of around 600 million tons (Cordeiro *et al.*, 2007). The harvesting and processing of sugarcane to its different products produces massive amount of bagasse residues including rind (outer layer) and pith (inner layer) of plant stalk (Lee & Mariatti, 2008). The bagasse residues are rich in polysaccharides (Jayapal *et al.*, 2013; Leang & Saw, 2011; Wang *et al.*, 2019c) and oligosaccharides (Reddy & Krishnan, 2016; Bian *et al.*, 2013) which are known as prebiotic ingredients. Besides, other beneficial nutrients such as phytochemical compounds (Zhao *et al.*, 2015; Feng *et al.*, 2015) which may be involved in scavenging of free radicals are also present. Therefore, they are ideal functional food ingredients that improve human health.

Over the last two decades, consumers adopted food products that are well known to promote wellness and health (Cencic & Chingwaru, 2010). As recorded by Institute of Medicine's Food and Nutrition Board, "functional foods" are food products that can enhance health beyond its basic nutritional requirements at the same time alleviate risks toward chronic disease (Al-Sheraji *et al.*, 2013). It is commonly known that prebiotic is one among the most commonly type of functional foods (Al-Sheraji *et al.*, 2013). Dietary prebiotic has been identified as "a selectively fermented ingredient which cause particular alterations in the constitution and/or activity of the gut microbiota, therefore according benefit(s) upon host well-being" (Gibson *et al.*, 2017). Thus, the prebiotic compounds should not undergo hydrolysis nor absorption within the upper section of the gut; and reach the colon to particularly stimulate the development of colonic microbiota (Rastall & Gibson, 2015). The use of prebiotics is not just limited to the regulation of gastrointestinal tract, but also could be used as therapeutic agents against various infections such as reducing the risk of colon carcinogenesis (Laparra & Sanz, 2010), and obesity (Cani *et al.*, 2009), enhancing mineral absorption (Whisner *et al.*, 2013), improving lipid metabolism and glucose homeostasis (Beserra *et al.*, 2015) and modification of inflammatory conditions like Irritable Bowel Syndrome (IBS) (Silk *et al.*, 2009).

Nevertheless, large scale study on prebiotics has promoted the desire to explore other possibility and inexpensive origins of prebiotics production in order to meet the market and consumers' requirements (Singdevsachan *et al.*, 2016). Xylooligosaccharide (XOS) is sugar polymers with a great prebiotic potential produced from the hydrolysis of xylan and consist of xylose units linked by β -(1,4) bonds, with degree of polymerization (DP) counting from 2 to 10. They are termed according to their number of xylose in their

structure such as xylobiose, xylotriose, xylotetraose and so on (Aachary & Prapulla, 2011). XOS as prebiotic has comparative advantages over fructooligosaccharide (FOS), inulin and galactooligosaccharide (GOS) with respect to their high resistance against heat and acidity. In fact, the recommended daily dose of XOS is only 0.7g, compared to FOS of 3g indicating that XOS can exert prebiotic effect at a lower dosage than FOS (Singh *et al.*, 2015). Agricultural wastes are among the most widely used sources for XOS production due to their high content of plant cell wall xylan as their utilization can relieve the environmental damage associated with the disposal of waste. Recently, different agricultural materials have been investigated for their XOS content as well as their prebiotic functions such as corn cobs (Samanta *et al.*, 2012b), wheat bran (Immerzeel *et al.*, 2014) and corn stover (Buruiana *et al.*, 2017). Nevertheless, there is insufficient information on prebiotic function of *S. officinarum* L. by-products (rind and pith).

1.2 Problem Statement

While most of the recent studies are focusing mainly on biological activity of fruit or their products, a handful of research investigated residues and by-products of fruit or food processing. Agricultural waste obtained from the food industry is plentiful in Malaysia, while its utilization is still limited. A great amount of untreated wastes can contaminate water, air and land by volatilization, leaching and dusting. On the other hand, various studies demonstrated that these wastes are rich in insoluble and soluble dietary fiber, polyphenol compound, oligosaccharides and others components which can be exploited for human consumption. (Leang & Saw, 2011; Kong *et al.*, 2015).

In year 2017, the global generation of sugarcane was calculated to be about 1.9 billion, that was produced on nearly 27.2 million hectares (FAOSTAT, 2017). The production of sugarcane in Malaysia was estimated at 28038 and 29990 tons in 2016 and 2017, respectively. Typically, over 90 countries produce sugarcane and of 100 tons of sugarcane processing in a factory, 30–34 tons of bagasse are obtained (Yadav & Solomon, 2006). Sugarcane remnants are burnt in sugar mills for energy production or stockpiled on site posing environmental issues without recognizing its potential beneficial for a variety of purposes, including for human dietary purpose (Attard *et al.*, 2015; Mandelli *et al.*, 2014). In Malaysia, there is lack of scientific information of sugarcane by-products in relation to their functional and health properties, and largely uncollected, which require more investigation to find other alternative uses in commercial food formulations. Therefore, to put an end to wasting resources, harnessing these residues for modern means of functional compounds for food application might be a commendable alternative for the health and economic sector.

1.3 Significance of Study

Several organizations (Food and Drug Administration and European Food Safety Authority) have recognized the proven health benefits of agricultural waste obtained from food industry. Therefore, the alternative uses of *S. officinarum* L. wastes in the formulation of different food products would be vital, both from the view of commercial

application and food science investigation. Due to unavailability of studies or investigation about the functional and health properties of the *S. officinarum* L. wastes, the present study will offer the methodological information to researchers and provide information regarding the ideal and effective use for each material based on its properties.

From the view of industrial application, the food factories would gain from the conversion of wastes into commercial products either as raw material for intermediate foods ingredients, or as ingredients of new products. Besides, using such inexpensive material as functional ingredients in food products can facilitate the product's availability in affordable and cheaper price to large number of customers. Based on that, *S. officinarum* L. wastes are ideal source of a variety of phytochemical compounds, and indigestible poly- or oligosaccharides which have exhibited different health benefits (Takara *et al.*, 2007; Kadam *et al.*, 2008; Reddy & Krishnan, 2016). Therefore, this alternative use of *S. officinarum* L. waste could not only reduce the amount of agricultural waste contaminating the environment, but also provide a variety of benefits for the human health and food industry.

1.4 Objectives

1.4.1 General Objective

To determine the ability of Xylooligosaccharide derived from *Saccharum officinarum* L. by-products (pith and rind) as a potential prebiotic to promote the growth of selected probiotics.

1.4.2 Specific Objectives

- a. To estimate the chemical composition, antioxidant activity, total phenolic, and total flavonoids content of *Saccharum officinarum* L. by-products.
- b. To determine the resistance of xylooligosaccharide derived from *Saccharum officinarum* L. by-products against human hydrolysis using α - amylase and artificial human gastric juice *in vitro*.
- c. To determine the effect of xylooligosaccharide derived from *Saccharum officinarum* L. by-products on two probiotic strains namely, *Lactobacillus casei* Shirota and *Bifidobacterium animals* subsp. *Lactis*.
- d. To quantify the primary end products of fermentation of xylooligosaccharide derived from *Saccharum officinarum* L. by-products particularly lactic, acetic and propionic acids.

REFERENCES

- Aachary, A. A., & Prapulla, S. G. (2011). Xylooligosaccharides (XOS) as an emerging prebiotic: microbial synthesis, utilization, structural characterization, bioactive properties, and applications. *Comprehensive Reviews in Food Science and Food Safety*, 10, 2-16.
- Abbas, S. R., Sabir, S. M., Ahmad, S. D., Boligon, A. A., & Athayde, M. L. (2014). Phenolic profile, antioxidant potential and DNA damage protecting activity of sugarcane (*Saccharum officinarum*). *Food Chemistry*, 147, 10-16.
- Abbasiliasi, S., Marikkar, M. N., Ariff, A., Amid, M., Lamasudin, D. U., Manap, M. Y. A., & Mustafa, S. (2017). Defatted coconut residue crude polysaccharides as potential prebiotics: study of their effects on proliferation and acidifying activity of probiotics in vitro. *Journal of Food Science and Technology*, 54, 164-173.
- Abdulqader, A., Ali, F., Ismail, A., & Esa, N. M. (2019). Antioxidant compounds and capacities of Gac (*Momordica cochinchinensis* Spreng) fruits. *Asian Pacific Journal of Tropical Biomedicine*, 9, 158.
- Ahmed, D., Khan, M., & Saeed, R. (2015). Comparative analysis of phenolics, flavonoids, and antioxidant and antibacterial potential of methanolic, hexanic and aqueous extracts from *Adiantum caudatum* leaves. *Antioxidants*, 4, 394-409.
- Akbari-Alavijeh, S., Soleimanian-Zad, S., Sheikh-Zeinoddin, M., & Hashmi, S. (2018). Pistachio hull water-soluble polysaccharides as a novel prebiotic agent. *International Journal of Biological Macromolecules*, 107, 808-816.
- Akpınar, O., Erdogan, K., & Bostancı, S. (2009). Enzymatic production of xylooligosaccharide from selected agricultural wastes. *Food and Bioproducts Processing*, 87, 145-151.
- Akpınar, O., Gunay, K., Yilmaz, Y., Levent, O., & Bostancı, S. (2010). Enzymatic processing and antioxidant activity of agricultural waste autohydrolysis liquors. *BioResources*, 5, 699-711.
- Alam, M. N., Bristi, N. J., & Rafiquzzaman, M. (2013). Review on in vivo and in vitro methods evaluation of antioxidant activity. *Saudi Pharmaceutical Journal*, 21, 143-152.
- Albishi, T., John, J. A., Al-Khalifa, A. S., & Shahidi, F. (2013). Antioxidant, anti-inflammatory and DNA scission inhibitory activities of phenolic compounds in selected onion and potato varieties. *Journal of Functional Foods*, 5, 930-939.
- Albuquerque, M. A. C. D., Levit, R., Beres, C., Bedani, R., de Moreno de LeBlanc, A., Saad, S. M. I., & LeBlanc, J. G. (2019). Tropical fruit by-products water extracts as sources of soluble fibres and phenolic compounds with potential antioxidant, anti-inflammatory, and functional properties. *Journal of Functional Foods*, 52, 724-733.

- Al-Sheraji, S. H., Ismail, A., Manap, M. Y., Mustafa, S., Yusof, R. M., & Hassan, F. A. (2013). Prebiotics as functional foods: A review. *Journal of Functional Foods*, 5, 1542-1553.
- Al-sheraji, S. H., Ismail, A., Manap, M. Y., Mustafa, S., Yusof, R. M., & Hassan, F. A. (2012). Fermentation and non-digestibility of Mangifera pajang fibrous pulp and its polysaccharides. *Journal of Functional Foods*, 4, 933-940.
- Álvarez, C., González, A., Negro, M. J., Ballesteros, I., Oliva, J. M., & Sáez, F. (2017). Optimized use of hemicellulose within a biorefinery for processing high value-added xylooligosaccharides. *Industrial Crops and Products*, 99, 41-48.
- AOAC. (1999). *Official methods of analysis of AOAC international* (16th ed). Gaithersburg: AOAC international.
- Attard, T. M., McElroy, C. R., Rezende, C. A., Polikarpov, I., Clark, J. H., & Hunt, A. J. (2015). Sugarcane waste as a valuable source of lipophilic molecules. *Industrial Crops and Products*, 76, 95-103.
- Azmi, A. F. M. N., Mustafa, S., Hashim, D. M., & Manap, Y. A. (2012). Prebiotic activity of polysaccharides extracted from Gigantochloa levis (Buluh beting) shoots. *Molecules*, 17, 1635-1651.
- Bach Knudsen, K. E. (2015). Microbial degradation of whole-grain complex carbohydrates and impact on short-chain fatty acids and health. *Advances in Nutrition*, 6, 206-213.
- Bakar, M. F. A., Mohamed, M., Rahmat, A., & Fry, J. (2009). Phytochemicals and antioxidant activity of different parts of bambangan (Mangifera pajang) and tarap (Artocarpus odoratissimus). *Food Chemistry*, 113, 479-483.
- Belorkar, S. A., & Gupta, A. K. (2016). Oligosaccharides: a boon from nature's desk. *Amb Express*, 6, 82.
- Beserra, B. T., Fernandes, R., do Rosario, V. A., Mocellin, M. C., Kuntz, M. G., & Trindade, E. B. (2015). A systematic review and meta-analysis of the prebiotics and synbiotics effects on glycaemia, insulin concentrations and lipid parameters in adult patients with overweight or obesity. *Clinical Nutrition*, 34, 845-858.
- Bhardwaj, N., Kumar, B., Agarwal, K., Chaturvedi, V., & Verma, P. (2019). Purification and characterization of a thermo-acid/alkali stable xylanases from Aspergillus oryzae LC1 and its application in Xylo-oligosaccharides production from lignocellulosic agricultural wastes. *International Journal of Biological Macromolecules*, 122, 1191-1202.
- Bian, J., Peng, F., Peng, X. P., Peng, P., Xu, F., & Sun, R. C. (2013). Structural features and antioxidant activity of xylooligosaccharides enzymatically produced from sugarcane bagasse. *Bioresource Technology*, 127, 236-241.
- Bian, J., Peng, P., Peng, F., Xiao, X., Xu, F., & Sun, R. C. (2014). Microwave-assisted

acid hydrolysis to produce xylooligosaccharides from sugarcane bagasse hemicelluloses. *Food Chemistry*, 156, 7-13.

Buruiana, C. T., Gómez, B., Vizireanu, C., & Garrote, G. (2017). Manufacture and evaluation of xylooligosaccharides from corn stover as emerging prebiotic candidates for human health. *LWT-Food Science and Technology*, 77, 449-459.

Can-cauich, C. A., Sauri-Duch, E., Betancur-Ancona, D., Chel-Guerrero, L., González-Aguilar, G. A., Cuevas-Glory, L. F., & Moo-Huchin, V. M. (2017). Tropical fruit peel powders as functional ingredients: Evaluation of their bioactive compounds and antioxidant activity. *Journal of Functional Foods*, 37, 501-506.

Cani, P. D., Lecourt, E., Dewulf, E. M., Sohet, F. M., Pachikian, B. D., Naslain, D., & Delzenne, N. M. (2009). Gut microbiota fermentation of prebiotics increases satietogenic and incretin gut peptide production with consequences for appetite sensation and glucose response after a meal. *The American Journal of Clinical Nutrition*, 90, 1236-1243.

Cencic, A., & Chingwaru, W. (2010). The role of functional foods, nutraceuticals, and food supplements in intestinal health. *Nutrients*, 2, 611-625.

Chapla, D., Pandit, P., & Shah, A. (2012). Production of xylooligosaccharides from corncob xylan by fungal xylanase and their utilization by probiotics. *Bioresource Technology*, 115, 215-221.

Charalampopoulos, D., & Rastall, R. A. (2012). Prebiotics in foods. *Current Opinion in Biotechnology*, 23, 187-191.

Chau, C. F., & Huang, Y. L. (2004). Characterization of passion fruit seed fibres—a potential fibre source. *Food Chemistry*, 85, 189-194.

Chau, C. F., Huang, Y. L., & Lin, C. Y. (2004). Investigation of the cholesterol-lowering action of insoluble fibre derived from the peel of Citrus sinensis L. cv. Liucheng. *Food Chemistry*, 87, 361-366.

Cheavegatti-Gianotto, A., de Abreu, H. M. C., Arruda, P., Bespalhok Filho, J. C., Burnquist, W. L., Creste, S., & de Fátima Grossi-de-Sá, M. (2011). Sugarcane (*Saccharum X officinarum*): a reference study for the regulation of genetically modified cultivars in Brazil. *Tropical Plant Biology*, 4, 62-89.

Chimtong, S., Saenphoom, P., Karageat, N., & Somtua, S. (2016). Oligosaccharide Production from Agricultural Residues by Non-Starch Polysaccharide Degrading Enzymes and Their Prebiotic Properties. *Agriculture and Agricultural Science Procedia*, 11, 131-136.

Chong, R. W. W., Ball, M., McRae, C., & Packer, N. H. (2019). Comparing the chemical composition of dietary fibres prepared from sugarcane, psyllium husk and wheat dextrin. *Food Chemistry*, 298, 125032.

Chung, Y. C., Hsu, C. K., Ko, C. Y., & Chan, Y. C. (2007). Dietary intake of

- xylooligosaccharides improves the intestinal microbiota, fecal moisture, and pH value in the elderly. *Nutrition Research*, 27, 756-761.
- Colombo, R., Lanças, F. M., & Yariwake, J. H. (2006). Determination of flavonoids in cultivated sugarcane leaves, bagasse, juice and in transgenic sugarcane by liquid chromatography-UV detection. *Journal of Chromatography A*, 1103, 118-124.
- Cordeiro, G., Amouyal, O., Elliott, F., & Henry, R. (2007). Sugarcane. In: C. Kole (Ed.), *Pulses, sugar and tuber crops* (pp 175–203). Heidelberg: Springer.
- Cruz-Guerrero, A., Hernández-Sánchez, H., Rodríguez-Serrano, G., Gómez-Ruiz, L., García-Garibay, M., & Figueroa-González, I. (2014). Commercial probiotic bacteria and prebiotic carbohydrates: a fundamental study on prebiotics uptake, antimicrobials production and inhibition of pathogens. *Journal of the Science of Food and Agriculture*, 94, 2246-2252.
- Cuello-Garcia, C. A., Brożek, J. L., Fiocchi, A., Pawankar, R., Yepes-Nuñez, J. J., Terracciano, L., & Schünemann, H. J. (2015). Probiotics for the prevention of allergy: a systematic review and meta-analysis of randomized controlled trials. *Journal of Allergy and Clinical Immunology*, 136, 952-961.
- Dasari, S., Kathera, C., Janardhan, A., Kumar, A. P., & Viswanath, B. (2017). Surfacing role of probiotics in cancer prophylaxis and therapy: A systematic review. *Clinical Nutrition*, 36, 1465-1472.
- De Melo Pereira, G. V., de Oliveira Coelho, B., Júnior, A. I. M., Thomaz-Soccol, V., & Soccol, C. R. (2018). How to select a probiotic? A review and update of methods and criteria. *Biotechnology Advances*, 36, 2060-2076.
- Delgado, G. T. C., Tamashiro, W. M. D. S. C., Junior, M. R. M., Moreno, Y. M. F., & Pastore, G. M. (2011). The putative effects of prebiotics as immunomodulatory agents. *Food Research International*, 44, 3167-3173.
- Delzenne, N. M., Neyrinck, A. M., Bäckhed, F., & Cani, P. D. (2011). Targeting gut microbiota in obesity: effects of prebiotics and probiotics. *Nature Reviews Endocrinology*, 7, 639.
- Derakhshan, Z., Ferrante, M., Tadi, M., Ansari, F., Heydari, A., Hosseini, M. S., & Sadrabad, E. K. (2018). Antioxidant activity and total phenolic content of ethanolic extract of pomegranate peels, juice and seeds. *Food and Chemical Toxicology*, 114, 108-111.
- Dhingra, D., Michael, M., Rajput, H., & Patil, R. T. (2012). Dietary fibre in foods: a review. *Journal of Food Science and Technology*, 49, 255-266.
- Dias, M. I., Sousa, M. J., Alves, R. C., & Ferreira, I. C. (2016). Exploring plant tissue culture to improve the production of phenolic compounds: A review. *Industrial Crops and Products*, 82, 9-22.
- Díaz-Vela, J., Totosaus, A., Cruz-Guerrero, A. E., & de Lourdes Pérez-Chabela, M.

- (2013). In vitro evaluation of the fermentation of added-value agroindustrial by-products: cactus pear (*O. puncticarpa* L.) peel and pineapple (*A. comosus*) peel as functional ingredients. *International Journal of Food Science & Technology*, 48, 1460-1467.
- Diether, N. E., & Willing, B. P. (2019). Microbial fermentation of dietary protein: An important factor in diet-microbe-host interaction. *Microorganisms*, 7, 19.
- Ding, C. H., Jiang, Z. Q., Li, X. T., Li, L. T., & Kusakabe, I. (2004). High activity xylanase production by *Streptomyces olivaceoviridis* E-86. *World Journal of Microbiology and Biotechnology*, 20, 7-10.
- Ding, W., Shi, C., Chen, M., Zhou, J., Long, R., & Guo, X. (2017). Screening for lactic acid bacteria in traditional fermented Tibetan yak milk and evaluating their probiotic and cholesterol-lowering potentials in rats fed a high-cholesterol diet. *Journal of Functional Foods*, 32, 324-332.
- Dixit, Y., Wagle, A., & Vakil, B. (2016). Patents in the field of probiotics, prebiotics, synbiotics: a review. *Journal of Food: Microbiology Safety & Hygiene*, 1, 1-2.
- Doron, S., & Snydman, D. R. (2015). Risk and safety of probiotics. *Clinical Infectious Diseases*, 60, S129-S134.
- Drakoularakou, A., Tzortzis, G., Rastall, R. A., & Gibson, G. R. (2010). A double-blind, placebo-controlled, randomized human study assessing the capacity of a novel galacto-oligosaccharide mixture in reducing travellers' diarrhoea. *European Journal of Clinical Nutrition*, 64, 146.
- Duary, R. K., Rajput, Y. S., Batish, V. K., & Grover, S. (2011). Assessing the adhesion of putative indigenous probiotic lactobacilli to human colonic epithelial cells. *The Indian Journal of Medical Research*, 134, 664.
- Elleuch, M., Besbes, S., Roiseux, O., Blecker, C., & Attia, H. (2007). Quality characteristics of sesame seeds and by-products. *Food Chemistry*, 103, 641-650.
- Esgalhado, M., Kemp, J. A., Damasceno, N. R., Fouque, D., & Mafra, D. (2017). Short-chain fatty acids: A link between prebiotics and microbiota in chronic kidney disease. *Future Microbiology*, 12, 1413-1425.
- FAOSTAT (2017). FAO (Food and Agriculture Organization of the United Nations). FAOSTAT Statistical Database, Rome, Italy. <http://www.fao.org/faostat/en>
- Feng, S., Luo, Z., Tao, B., & Chen, C. (2015). Ultrasonic-assisted extraction and purification of phenolic compounds from sugarcane (*Saccharum officinarum* L.) rinds. *LWT-Food Science and Technology*, 60, 970-976.
- Feng, S., Luo, Z., Zhang, Y., Zhong, Z., & Lu, B. (2014). Phytochemical contents and antioxidant capacities of different parts of two sugarcane (*Saccharum officinarum* L.) cultivars. *Food Chemistry*, 151, 452-458.

- Fernandes, R., do Rosario, V. A., Mocellin, M. C., Kuntz, M. G., & Trindade, E. B. (2017). Effects of inulin-type fructans, galacto-oligosaccharides and related synbiotics on inflammatory markers in adult patients with overweight or obesity: A systematic review. *Clinical Nutrition*, 36, 1197-1206.
- Fernández, J., Redondo-Blanco, S., Gutierrez-del-Rio, I., Miguélez, E. M., Villar, C. J., & Lombo, F. (2016). Colon microbiota fermentation of dietary prebiotics towards short-chain fatty acids and their roles as anti-inflammatory and antitumour agents: A review. *Journal of Functional Foods*, 25, 511-522.
- Figuerola, F., Hurtado, M. L., Estévez, A. M., Chiffelle, I., & Asenjo, F. (2005). Fibre concentrates from apple pomace and citrus peel as potential fibre sources for food enrichment. *Food Chemistry*, 91, 395-401.
- Finegold, S. M., Li, Z., Summanen, P. H., Downes, J., Thames, G., Corbett, K., & Heber, D. (2014). Xylooligosaccharide increases bifidobacteria but not lactobacilli in human gut microbiota. *Food & Function*, 5, 436-445.
- Fontana, L., Bermudez-Brito, M., Plaza-Diaz, J., Munoz-Quezada, S., & Gil, A. (2013). Sources, isolation, characterisation and evaluation of probiotics. *British Journal of Nutrition*, 109, S35-S50.
- Frost, G., Sleeth, M. L., Sahuri-Arisoylu, M., Lizarbe, B., Cerdan, S., Brody, L., & Carling, D. (2014). The short-chain fatty acid acetate reduces appetite via a central homeostatic mechanism. *Nature Communications*, 5, 3611.
- Fung, K. Y., Cosgrove, L., Lockett, T., Head, R., & Topping, D. L. (2012). A review of the potential mechanisms for the lowering of colorectal oncogenesis by butyrate. *British Journal of Nutrition*, 108, 820-831.
- Galisteo, M., Duarte, J., & Zarzuelo, A. (2008). Effects of dietary fibers on disturbances clustered in the metabolic syndrome. *The Journal of Nutritional Biochemistry*, 19, 71-84.
- Gibson, G. R. (2004). From probiotics to prebiotics and a healthy digestive system. *Journal of Food Science*, 69, 141-143.
- Gibson, G. R., & Roberfroid, M. B. (1995). Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. *The Journal of Nutrition*, 125, 1401-1412.
- Gibson, G. R., Hutkins, R., Sanders, M. E., Prescott, S. L., Reimer, R. A., Salminen, S. J., & Verbeke, K. (2017). The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nature Reviews Gastroenterology & Hepatology*, 14, 491-502.
- Gibson, G. R., Probert, H. M., Van Loo, J., Rastall, R. A., & Roberfroid, M. B. (2004). Dietary modulation of the human colonic microbiota: updating the concept of prebiotics. *Nutrition Research Reviews*, 17, 259-275.

- Gibson, G. R., Scott, K. P., Rastall, R. A., Tuohy, K. M., Hotchkiss, A., Dubert-Ferrandon, A., & Macfarlane, S. (2010). Dietary prebiotics: current status and new definition. *Food Science and Technology Bulletin: Functional Foods*, 7, 1-19.
- Gilad, O., Jacobsen, S., Stuer-Lauridsen, B., Pedersen, M. B., Garrigues, C., & Svensson, B. (2010). Combined transcriptome and proteome analysis of *Bifidobacterium animalis* subsp. *lactis* BB-12 grown on xylo-oligosaccharides and a model of their utilization. *Applied and Environmental Microbiology*, 76, 7285-7291.
- Gobinath, D., Madhu, A. N., Prashant, G., Srinivasan, K., & Prapulla, S. G. (2010). Beneficial effect of xylo-oligosaccharides and fructo-oligosaccharides in streptozotocin-induced diabetic rats. *British Journal of Nutrition*, 104, 40-47.
- Goldenberg, J. Z., Lytvyn, L., Steurich, J., Parkin, P., Mahant, S., & Johnston, B. C. (2015). Probiotics for the prevention of pediatric antibiotic-associated diarrhea. *Cochrane Database of Systematic Reviews*, 463-466.
- Gómez, B., Gullón, B., Yáñez, R., Schols, H., & Alonso, J. L. (2016). Prebiotic potential of pectins and pectic oligosaccharides derived from lemon peel wastes and sugar beet pulp: A comparative evaluation. *Journal of Functional Foods*, 20, 108-121.
- Gómez, B., Míguez, B., Yáñez, R., & Alonso, J. L. (2017). Extraction of Oligosaccharides With Prebiotic Properties From Agro-Industrial Wastes. In H. D. González & M. J. G. (Eds.), *water Extraction of Bioactive Compounds* (pp. 131-161). Elsevier.
- Gourbeyre, P., Denery, S., & Bodinier, M. (2011). Probiotics, prebiotics, and synbiotics: impact on the gut immune system and allergic reactions. *Journal of Leukocyte Biology*, 89, 685-695.
- Guarner, F., & Malagelada, J. R. (2003). Gut flora in health and disease. *The Lancet*, 361, 512-519.
- Gullón, B., Gullón, P., Tavaría, F., Pintado, M., Gomes, A. M., Alonso, J. L., & Parajó, J. C. (2014). Structural features and assessment of prebiotic activity of refined arabinoxyloligosaccharides from wheat bran. *Journal of Functional Foods*, 6, 438-449.
- Gullon, B., Pintado, M. E., Fernández-López, J., Pérez-Álvarez, J. A., & Viuda-Martos, M. (2015). In vitro gastrointestinal digestion of pomegranate peel (*Punica granatum*) flour obtained from co-products: Changes in the antioxidant potential and bioactive compounds stability. *Journal of Functional Foods*, 19, 617-628.
- Gullón, P., González-Muñoz, M. J., & Parajó, J. C. (2011). Manufacture and prebiotic potential of oligosaccharides derived from industrial solid wastes. *Bioresource Technology*, 102, 6112-6119.
- Gullón, P., Moura, P., Esteves, M. P., Girio, F. M., Domínguez, H., & Parajó, J. C. (2008). Assessment on the fermentability of xylooligosaccharides from rice husks

- by probiotic bacteria. *Journal of Agricultural and Food Chemistry*, 56, 7482-7487.
- Han, K. H., Kobayashi, Y., Nakamura, Y., Shimada, K. I., Aritsuka, T., Ohba, K., & Fukushima, M. (2014). Comparison of the effects of longer chain inulins with different degrees of polymerization on colonic fermentation in a mixed culture of swine fecal bacteria. *Journal of Nutritional Science and Vitaminology*, 60, 206-212.
- Han, S., Gao, J., Zhou, Q., Liu, S., Wen, C., & Yang, X. (2018). Role of intestinal flora in colorectal cancer from the metabolite perspective: a systematic review. *Cancer Management and Research*, 10, 199-206.
- Healey, G., Murphy, R., Butts, C., Brough, L., Rosendale, D., Blatchford, P., & Coad, J. (2017). Variability in gut microbiota response to an inulin-type fructan prebiotic within an in vitro three-stage continuous colonic model system. *Bioactive Carbohydrates and Dietary Fibre*, 11, 26-37.
- Henry, R. J., & Kole, C. (2010). *Genetics, genomics and breeding of sugarcane*. United States of America: CRC Press.
- Hernández-Hernández, O., Muthaiyan, A., Moreno, F. J., Montilla, A., Sanz, M. L., & Ricke, S. C. (2012). Effect of prebiotic carbohydrates on the growth and tolerance of Lactobacillus. *Food Microbiology*, 30, 355-361.
- Hill, C., Guarner, F., Reid, G., Gibson, G. R., Merenstein, D. J., Pot, B., & Calder, P. C. (2014). The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nature Reviews Gastroenterology & Hepatology*, 11, 506-514.
- Holscher, H. D. (2017). Dietary fiber and prebiotics and the gastrointestinal microbiota. *Gut Microbes*, 8, 172-184.
- Hosseini, E., Grootaert, C., Verstraete, W., & Van de Wiele, T. (2011). Propionate as a health-promoting microbial metabolite in the human gut. *Nutrition Reviews*, 69, 245-258.
- Hromádková, Z., Paulsen, B. S., Polovka, M., Košťálová, Z., & Ebringerová, A. (2013). Structural features of two heteroxylan polysaccharide fractions from wheat bran with anti-complementary and antioxidant activities. *Carbohydrate Polymers*, 93, 22-30.
- Hu, J. L., Nie, S. P., Li, C., Wang, S., & Xie, M. Y. (2018). Ultrasonic irradiation induces degradation and improves prebiotic properties of polysaccharide from seeds of Plantago asiatica L. during in vitro fermentation by human fecal microbiota. *Food Hydrocolloids*, 76, 60-66.
- Huebner, J., Wehling, R. L., & Hutkins, R. W. (2007). Functional activity of commercial prebiotics. *International Dairy Journal*, 17, 770-775.
- Ibrahim, A. H. (2009). *Physico-chemical and Health-promoting Properties of Dietary*

Fibre Powder from Pink Guava By-products (Unpublished doctoral dissertation). Universiti Putra, Malaysia.

- Immerzeel, P., Falck, P., Galbe, M., Adlercreutz, P., Karlsson, E. N., & Stålbrand, H. (2014). Extraction of water-soluble xylan from wheat bran and utilization of enzymatically produced xylooligosaccharides by *Lactobacillus*, *Bifidobacterium* and *Weissella* spp. *LWT-Food Science and Technology*, 56, 321-327.
- Jagtap, S., Deshmukh, R. A., Menon, S., & Das, S. (2017). Xylooligosaccharides production by crude microbial enzymes from agricultural waste without prior treatment and their potential application as nutraceuticals. *Bioresource Technology*, 245, 283-288.
- Jahurul, M. H. A., Zaidul, I. S. M., Beh, L., Sharifudin, M. S., Siddiquee, S., Hasmadi, M., & Jinap, S. (2019). Valuable components of bambangan fruit (*Mangifera pajang*) and its co-products: A review. *Food Research International*, 115, 105-115.
- James, G. (2004). *Sugarcane*. United Kingdom: Blackwell's
- Jayapal, N., Samanta, A. K., Kolte, A. P., Senani, S., Sridhar, M., Suresh, K. P., & Sampath, K. T. (2013). Value addition to sugarcane bagasse: xylan extraction and its process optimization for xylooligosaccharides production. *Industrial Crops and Products*, 42, 14-24.
- Jeddou, K. B., Bouaziz, F., Helbert, C. B., Nouri-Ellouz, O., Maktouf, S., Ellouz-Chaabouni, S., & Ellouz-Ghorbel, R. (2018). Structural, functional, and biological properties of potato peel oligosaccharides. *International Journal of Biological Macromolecules*, 112, 1146-1155.
- Jenkins, G. H. (2013). *Introduction to cane sugar technology*. Elsevier.
- Kadam, U. S., Ghosh, S. B., De, S., Suprasanna, P., Devasagayam, T. P. A., & Bapat, V. A. (2008). Antioxidant activity in sugarcane juice and its protective role against radiation induced DNA damage. *Food Chemistry*, 106, 1154-1160.
- Kim, H. J., & White, P. J. (2009). In vitro fermentation of oat flours from typical and high β -glucan oat lines. *Journal of Agricultural and Food Chemistry*, 57, 7529-7536.
- Klinchongkon, K., Khuwijitjaru, P., Wiboonsirikul, J., & Adachi, S. (2017). Extraction of oligosaccharides from passion fruit peel by subcritical water treatment. *Journal of Food Process Engineering*, 40, e12269.
- Kolida, S., Meyer, D., & Gibson, G. R. (2007). A double-blind placebo-controlled study to establish the bifidogenic dose of inulin in healthy humans. *European Journal of Clinical Nutrition*, 61, 1189.
- Kong, F., Yu, S., Zeng, F., & Wu, X. (2015). Preparation of Antioxidant and Evaluation of the Antioxidant Activities of Antioxidants Extracted From Sugarcane

- Products. *Journal of Food and Nutrition Research*, 3, 458-463.
- Kumar, C. A. S., Varadharajan, R., Muthumani, P., Meera, R., Devi, P., & Kameswari, B. (2010). Psychopharmacological studies on the stem of Saccharum spontaneum. *International Journal of PharmTech Research*, 2, 319-321.
- Kumar, V., & Satyanarayana, T. (2015). Generation of xylooligosaccharides from microwave irradiated agroresidues using recombinant thermo-alkali-stable endoxylanase of the polyextremophilic bacterium *Bacillus halodurans* expressed in *Pichia pastoris*. *Bioresource Technology*, 179, 382-389.
- Kurdi, P., & Hansawadi, C. (2015). Assessment of the prebiotic potential of oligosaccharide mixtures from rice bran and cassava pulp. *LWT-Food Science and Technology*, 63, 1288-1293.
- Laparra, J. M., & Sanz, Y. (2010). Interactions of gut microbiota with functional food components and nutraceuticals. *Pharmacological research*, 61, 219-225.
- Layden, B. T., Yalamanchi, S. K., Wolever, T. M., Dunaif, A., & Lowe Jr, W. L. (2012). Negative association of acetate with visceral adipose tissue and insulin levels. *Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy*, 5, 49.
- Leang, Y. H., & Saw, H. Y. (2011). Proximate and functional properties of sugarcane bagasse. *Agro Food Industry Hi Tech*, 22, 5.
- Lebeer, S., Vanderleyden, J., & De Keersmaecker, S. C. (2008). Genes and molecules of lactobacilli supporting probiotic action. *Microbiology and Molecular Biology Reviews*, 72, 728-764.
- Lee, C. P., Chen, Z. T., Yu, P. Y., Wang, Y. C., & Duh, P. D. (2012). Identification of bioactive compounds and comparison of apoptosis induction of three varieties of sugarcane leaves. *Journal of Functional Foods*, 4, 391-397.
- Lee, S. C., & Mariatti, M. (2008). The effect of bagasse fibers obtained (from rind and pith component) on the properties of unsaturated polyester composites. *Materials Letters*, 62, 2253-2256.
- Leroy, F., & De Vuyst, L. (2004). Lactic acid bacteria as functional starter cultures for the food fermentation industry. *Trends in Food Science & Technology*, 15, 67-78.
- Lim, H. J., Kim, S. Y., & Lee, W. K. (2004). Isolation of cholesterol-lowering lactic acid bacteria from human intestine for probiotic use. *Journal Veterinary Science*, 5, 391-395.
- Liu, Z., Lin, X., Huang, G., Zhang, W., Rao, P., & Ni, L. (2014). Prebiotic effects of almonds and almond skins on intestinal microbiota in healthy adult humans. *Anaerobe*, 26, 1-6.
- Ma, J., Zhang, J., Li, Q., Shi, Z., Wu, H., Zhang, H., & Sun, X. (2019). Oral administration of a mixture of probiotics protects against food allergy via induction

- of CD103+ dendritic cells and modulates the intestinal microbiota. *Journal of Functional Foods*, 55, 65-75.
- Maathuis, A., Hoffman, A., Evans, A., Sanders, L., & Venema, K. (2009). The effect of the undigested fraction of maize products on the activity and composition of the microbiota determined in a dynamic in vitro model of the human proximal large intestine. *Journal of the American College of Nutrition*, 28, 657-666.
- Macfarlane, S., & Macfarlane, G. T. (2003). Regulation of short-chain fatty acid production. *Proceedings of the Nutrition Society*, 62, 67-72.
- Mandelli, F., Brenelli, L. B., Almeida, R. F., Goldbeck, R., Wolf, L. D., Hoffmann, Z. B., & Squina, F. M. (2014). Simultaneous production of xylooligosaccharides and antioxidant compounds from sugarcane bagasse via enzymatic hydrolysis. *Industrial Crops and Products*, 52, 770-775.
- Manisseri, C., & Gudipati, M. (2012). Prebiotic activity of purified xylobiose obtained from Ragi (Eleusine coracana, Indaf-15) Bran. *Indian Journal of Microbiology*, 52, 251-257.
- Matsuo, Y., Miura, L. A., Araki, T., & Yoshie-Stark, Y. (2019). Proximate composition and profiles of free amino acids, fatty acids, minerals and aroma compounds in Citrus natsudaidai peel. *Food Chemistry*, 279, 356-363.
- McFarland, L. V., & Goh, S. (2019). Are probiotics and prebiotics effective in the prevention of travellers' diarrhea: A systematic review and meta-analysis. *Travel Medicine and Infectious Disease*, 27, 11-19.
- Meier, R. F. (2009). Basics in clinical nutrition: Fibre and short chain fatty acids. *e-SPEN, the European e-Journal of Clinical Nutrition and Metabolism*, 2, e69-e71.
- Míguez, B., Gómez, B., Gullón, P., Gullón, B., & Alonso, J.L. (2016). Pectic oligosaccharides and other emerging prebiotics. In V. Rao (Ed.), *Prebiotics and Probiotics in Human Nutrition and Health* (pp. 301). United Kingdom: IntechOpen's Academic.
- Moo-Huchin, V. M., Moo-Huchin, M. I., Estrada-León, R. J., Cuevas-Glory, L., Estrada-Mota, I. A., Ortiz-Vázquez, E., & Sauri-Duch, E. (2015). Antioxidant compounds, antioxidant activity and phenolic content in peel from three tropical fruits from Yucatan, Mexico. *Food Chemistry*, 166, 17-22.
- Moore, P. H., & Botha, F. C. (2013). *Sugarcane: physiology, biochemistry and functional biology*. United states: John Wiley & Sons.
- Moreno, F. J., Corzo, N., Montilla, A., Villamiel, M., & Olano, A. (2017). Current state and latest advances in the concept, production and functionality of prebiotic oligosaccharides. *Current Opinion in Food Science*, 13, 50-55.
- Moure, A., Gullón, P., Domínguez, H., & Parajó, J. C. (2006). Advances in the manufacture, purification and applications of xylo-oligosaccharides as food

- additives and nutraceuticals. *Process Biochemistry*, 41, 1913-1923.
- Mugahed, S. H. A. S. (2012). *Characterization of Mangifera Pajang Kort, Fruit Fibres and Polysaccharides and Their Roles in Survival and Hypocholesterolemic Effects of Bifidobacteria*. (Unpublished doctoral dissertation). Universiti Putra, Malaysia.
- Nicolucci, A. C., Hume, M. P., Martínez, I., Mayengbam, S., Walter, J., & Reimer, R. A. (2017). Prebiotics reduce body fat and alter intestinal microbiota in children who are overweight or with obesity. *Gastroenterology*, 153, 711-722.
- Nieto-Domínguez, M., de Eugenio, L. I., York-Durán, M. J., Rodríguez-Colinas, B., Plou, F. J., Chenoll, E., & Martínez, M. J. (2017). Prebiotic effect of xylooligosaccharides produced from birchwood xylan by a novel fungal GH11 xylanase. *Food Chemistry*, 232, 105-113.
- OECD, (2016) "Sugarcane (Saccharum spp.)", in *Safety Assessment of Transgenic Organisms in the Environment*, Volume 6: OECD Consensus Documents. Paris: OECD Publishing. <https://doi.org/10.1787/9789264253421-5-en>.
- Otieno, D. O., & Ahring, B. K. (2012). A thermochemical pretreatment process to produce xylooligosaccharides (XOS), arabinooligosaccharides (AOS) and mannooligosaccharides (MOS) from lignocellulosic biomasses. *Bioresource Technology*, 112, 285-292.
- Pallavi, R., Elakkiya, S., Tennety, S. S. R., & Devi, P. S. (2012). Anthocyanin analysis and its anticancer property from sugarcane (*Saccharum officinarum* L) peel. *International Journal of Research in Pharmacy and Chemistry*, 2, 338-45.
- Pandey, A., Soccol, C. R., Nigam, P., & Soccol, V. T. (2000). Biotechnological potential of agro-industrial residues. I: sugarcane bagasse. *Bioresource Technology*, 74, 69-80.
- Patel, S., & Goyal, A. (2011). Functional oligosaccharides: production, properties and applications. *World Journal of Microbiology and Biotechnology*, 27, 1119-1128.
- Pereira, G. A., Arruda, H. S., Molina, G., & Pastore, G. M. (2018). Extraction optimization and profile analysis of oligosaccharides in banana pulp and peel. *Journal of Food Processing and Preservation*, 42, e13408.
- Plongbunjong, V., Grajist, P., Knudsen, K. E. B., & Wichienchot, S. (2017). Starch-based carbohydrates display the bifidogenic and butyrogenic properties in pH-controlled faecal fermentation. *International Journal of Food Science & Technology*, 52, 2647-2653.
- Quigley, E. M. (2012). Prebiotics and probiotics: their role in the management of gastrointestinal disorders in adults. *Nutrition in Clinical Practice*, 27, 195-200.
- Raghavendra, S. N., Swamy, S. R., Rastogi, N. K., Raghavarao, K. S. M. S., Kumar, S., & Tharanathan, R. N. (2006). Grinding characteristics and hydration properties of coconut residue: A source of dietary fiber. *Journal of Food Engineering*, 72, 281-

- Rahman, N. F. A., Shamsudin, R., Ismail, A., Shah, N. N. A. K., & Varith, J. (2018). Effects of drying methods on total phenolic contents and antioxidant capacity of the pomelo (*Citrus grandis* (L.) Osbeck) peels. *Innovative Food Science & Emerging Technologies*, 50, 217-225.
- Rastall, R. A., & Gibson, G. R. (2015). Recent developments in prebiotics to selectively impact beneficial microbes and promote intestinal health. *Current Opinion in Biotechnology*, 32, 42-46.
- Ratnadewi, A. A. I., Santoso, A. B., Sulistyaningsih, E., & Handayani, W. (2016). Application of cassava peel and waste as raw materials for xylooligosaccharide production using endoxylanase from *Bacillus subtilis* of soil termite abdomen. *Procedia Chemistry*, 18, 31-38.
- Raudone, L., Raudonis, R., Liaudanskas, M., Janulis, V., & Viskelis, P. (2017). Phenolic antioxidant profiles in the whole fruit, flesh and peel of apple cultivars grown in Lithuania. *Scientia horticulturae*, 216, 186-192.
- Reddy, S. S., & Krishnan, C. (2016). Production of high-pure xylooligosaccharides from sugarcane bagasse using crude β -xylosidase-free xylanase of *Bacillus subtilis* KCX006 and their bifidogenic function. *LWT-Food Science and Technology*, 65, 237-245.
- Resende, L. M., Franca, A. S., & Oliveira, L. S. (2019). Buriti (*Mauritia flexuosa* L. f.) fruit by-products flours: Evaluation as source of dietary fibers and natural antioxidants. *Food Chemistry*, 270, 53-60.
- Riciputi, Y., Diaz-de-Cerio, E., Akyol, H., Capanoglu, E., Cerretani, L., Caboni, M. F., & Verardo, V. (2018). Establishment of ultrasound-assisted extraction of phenolic compounds from industrial potato by-products using response surface methodology. *Food Chemistry*, 269, 258-263.
- Rohin, M. A. K., Bakar, A., Abdullah, C., & Ali, A. M. (2014). Isolation and characterization of oligosaccharides composition in organically grown red pitaya, white pitaya and papaya. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6, 131-136.
- Romagnolo, D. F., & Selmin, O. I. (2012). Flavonoids and cancer prevention: a review of the evidence. *Journal of Nutrition in Gerontology and Geriatrics*, 31, 206-238.
- Russell, D. A., Ross, R. P., Fitzgerald, G. F., & Stanton, C. (2011). Metabolic activities and probiotic potential of bifidobacteria. *International Journal of Food Microbiology*, 149, 88-105.
- Saad, N., Delattre, C., Urdaci, M., Schmitter, J. M., & Bressollier, P. (2013). An overview of the last advances in probiotic and prebiotic field. *LWT-Food Science and Technology*, 50, 1-16.

- Sajib, M., Falck, P., Sardari, R. R., Mathew, S., Grey, C., Karlsson, E. N., & Adlercreutz, P. (2018). Valorization of Brewer's spent grain to prebiotic oligosaccharide: production, xylanase catalyzed hydrolysis, in-vitro evaluation with probiotic strains and in a batch human fecal fermentation model. *Journal of Biotechnology*, 268, 61-70.
- Samanta, A. K., Jayapal, N., Jayaram, C., Roy, S., Kolte, A. P., Senani, S., & Sridhar, M. (2015). Xylooligosaccharides as prebiotics from agricultural by-products: production and applications. *Bioactive Carbohydrates and Dietary Fibre*, 5, 62-71.
- Samanta, A. K., Jayapal, N., Kolte, A. P., Senani, S., Sridhar, M., Suresh, K. P., & Sampath, K. T. (2012a). Enzymatic production of xylooligosaccharides from alkali solubilized xylan of natural grass (*Sehima nervosum*). *Bioresource Technology*, 112, 199-205.
- Samanta, A. K., Senani, S., Kolte, A. P., Sridhar, M., Sampath, K. T., Jayapal, N., & Devi, A. (2012b). Production and in vitro evaluation of xylooligosaccharides generated from corn cobs. *Food and Bioproducts Processing*, 90, 466-474. 1
- Sangeetha, A. V., Mahadevamma, S., Begum, K., & Sudha, M. L. (2011). Influence of processed sugarcane bagasse on the microbial, nutritional, rheological and quality characteristics of biscuits. *International Journal of Food Sciences and Nutrition*, 62, 457-464..
- Sangnark, A., & Noomhorm, A. (2003). Effect of particle sizes on functional properties of dietary fibre prepared from sugarcane bagasse. *Food Chemistry*, 80, 221-229.
- Sarker, S., Seraj, S., Sattar, M. M., Haq, W. M., Chowdhury, M. H., Ahmad, I., & Rahmatullah, M. (2011). Medicinal plants used by folk medicinal practitioners of six villages in Thakurgaon district, Bangladesh. *American-Eurasian Journal of Sustainable Agriculture*, 11, 1-22.
- Sáyago-Ayerdi, S. G., Zamora-Gasga, V. M., & Venema, K. (2017). Prebiotic effect of predigested mango peel on gut microbiota assessed in a dynamic in vitro model of the human colon (TIM-2). *Food Research International*, 118, 89-95.
- Serban, D. E. (2014). Gastrointestinal cancers: influence of gut microbiota, probiotics and prebiotics. *Cancer letters*, 345, 258-270.
- Shahidi, F. (2015). *Handbook of antioxidants for food preservation*. Woodhead Publishing.
- Shakirin, F. H., Prasad, K. N., Ismail, A., Yuon, L. C., & Azlan, A. (2010). Antioxidant capacity of underutilized Malaysian Canarium odontophyllum (dabai) Miq. fruit. *Journal of Food Composition and Analysis*, 23, 777-781.
- Shao, Y., Gao, Z., Marks, P. A., & Jiang, X. (2004). Apoptotic and autophagic cell death induced by histone deacetylase inhibitors. *Proceedings of the National Academy of Sciences*, 101, 18030-18035.

- Shirazi, O. U., Khattak, M. A. K., Shukri, N. A. M., & Nasyriq, M. N. (2014). Determination of total phenolic, flavonoid content and free radical scavenging activities of common herbs and spices. *Journal of Pharmacognosy and Phytochemistry*, 3, 104-108.
- Shukla, G., Verma, A., Singh, J., & Yadav, H. (2014). Prebiotic inulin alters the colonic mass, pH, microflora and short chain fatty acids in 1, 2-dimethylhydrazine dihydrochloride induced early colon carcinogenesis in male Laca mice. *J. Probiotics Health*, 2, 1-6.
- Silk, D. B. A., Davis, A., Vulevic, J., Tzortzis, G., & Gibson, G. R. (2009). Clinical trial: the effects of a trans-galactooligosaccharide prebiotic on faecal microbiota and symptoms in irritable bowel syndrome. *Alimentary Pharmacology & Therapeutics*, 29, 508-518.
- Singdevsachan, S. K., Auroshree, P., Mishra, J., Baliarsingh, B., Tayung, K., & Thatoi, H. (2016). Mushroom polysaccharides as potential prebiotics with their antitumor and immunomodulating properties: A review. *Bioactive Carbohydrates and Dietary Fiber*, 7, 1-14.
- Singh, H., & Gallier, S. (2014). Processing of food structures in the gastrointestinal tract and physiological responses. In M. Boland, M. Golding & H. Singh (Eds), *Food Structures, Digestion and Health* (pp. 51-81). United States: Academic Press.
- Singh, R. D., Banerjee, J., & Arora, A. (2015). Prebiotic potential of oligosaccharides: A focus on xylan derived oligosaccharides. *Bioactive Carbohydrates and Dietary Fibre*, 5, 19-30.
- Singh, R. D., Banerjee, J., Sasmal, S., Muir, J., & Arora, A. (2018). High xylan recovery using two stage alkali pre-treatment process from high lignin biomass and its valorisation to xylooligosaccharides of low degree of polymerisation. *Bioresource Technology*, 256, 110-117.
- Singh, S. P., Jadaun, J. S., Narnoliya, L. K., & Pandey, A. (2017). Prebiotic oligosaccharides: Special focus on fructooligosaccharides, its biosynthesis and bioactivity. *Applied Biochemistry and Biotechnology*, 183, 613-635.
- Singh, S., & Singh, R. P. (2008). In vitro methods of assay of antioxidants: an overview. *Food Reviews International*, 24, 392-415.
- Slover, C. M., & Danziger, L. (2008). Lactobacillus: a review. *Clinical Microbiology Newsletter*, 30, 23-27.
- Sophonputtanaphoca, S., Pridam, C., Chinnak, J., Nathong, M., & Juntipwong, P. (2018). Production of non-digestible oligosaccharides as value-added by-products from rice straw. *Agriculture and Natural Resources*, 52, 169-175.
- Surek, E., & Buyukkileci, A. O. (2017). Production of xylooligosaccharides by autohydrolysis of hazelnut (*Corylus avellana* L.) shell. *Carbohydrate Polymers*, 174, 565-571.

- Suttirak, W., & Manurakchinakorn, S. (2014). In vitro antioxidant properties of mangosteen peel extract. *Journal of Food Science and Technology*, 51, 3546-3558.
- Takara, K., Ushijima, K., Wada, K., Iwasaki, H., & Yamashita, M. (2007). Phenolic compounds from sugarcane molasses possessing antibacterial activity against cariogenic bacteria. *Journal of Oleo Science*, 56, 611-614.
- Teng, C., Yan, Q., Jiang, Z., Fan, G., & Shi, B. (2010). Production of xylooligosaccharides from the steam explosion liquor of corncobs coupled with enzymatic hydrolysis using a thermostable xylanase. *Bioresource Technology*, 101, 7679-7682.
- Thammarutwasik, P., Hongpattarakere, T., Chantachum, S., Kijoongrojana, K., Itharat, A., Reanmongkol, W., & Ooraikul, B. (2009). Prebiotics-A Review. *Songklanakarin Journal of Science & Technology*, 31, 401-408.
- Trollope, K. M., van Wyk, N., Kotjomela, M. A., & Volschenk, H. (2015). Sequence and structure-based prediction of fructosyltransferase activity for functional subclassification of fungal GH32 enzymes. *The FEBS journal*, 282, 4782-4796.
- Tuo, Y., Yu, H., Ai, L., Wu, Z., Guo, B., & Chen, W. (2013). Aggregation and adhesion properties of 22 Lactobacillus strains. *Journal of Dairy Science*, 96, 4252-4257.
- Van Laere, K. M., Hartemink, R., Bosveld, M., Schols, H. A., & Voragen, A. G. (2000). Fermentation of plant cell wall derived polysaccharides and their corresponding oligosaccharides by intestinal bacteria. *Journal of Agricultural and Food Chemistry*, 48, 1644-1652.
- Vallejos, M. E., Felissia, F. E., Kruyeniski, J., & Area, M. C. (2015). Kinetic study of the extraction of hemicellulosic carbohydrates from sugarcane bagasse by hot water treatment. *Industrial Crops and Products*, 67, 1-6.
- Verbeke, K. A., Boobis, A. R., Chiodini, A., Edwards, C. A., Franck, A., Kleerebezem, M., & Tuohy, K. M. (2015). Towards microbial fermentation metabolites as markers for health benefits of prebiotics. *Nutrition Research Reviews*, 28, 42-66.
- Verheyen, W. (ED.). (2010). *Soils, plant growth and crop production Volume 2*. United Kingdom: EOLSS.
- Wang, J., Sun, B., Cao, Y., & Wang, C. (2010). In vitro fermentation of xylooligosaccharides from wheat bran insoluble dietary fiber by Bifidobacteria. *Carbohydrate Polymers*, 82, 419-423.
- Wang, M., Wichienchot, S., He, X., Fu, X., Huang, Q., & Zhang, B. (2019a). In vitro colonic fermentation of dietary fibers: Fermentation rate, short-chain fatty acid production and changes in microbiota. *Trends in Food Science & Technology*, 88, 1-9.
- Wang, X., Huang, M., Yang, F., Sun, H., Zhou, X., Guo, Y., & Zhang, M. (2015). Rapeseed polysaccharides as prebiotics on growth and acidifying activity of

- probiotics in vitro. *Carbohydrate Polymers*, 125, 232-240.
- Wang, Y., Guo, Q., Goff, H. D., & LaPointe, G. (2019b). Oligosaccharides: Structure, Function and Application. *Encyclopedia of Food Chemistry*, 202-207.
- Wang, Y., Hou, F., Xu, H., Li, J., Miao, C., Lu, S., & Sun, C. (2019c). Integrated methane production improvement from sugarcane rind by microwave coupled calcium hydroxide pretreatment. *Fuel*, 246, 402-407.
- Whelan, K. (2011). Probiotics and prebiotics in the management of irritable bowel syndrome: a review of recent clinical trials and systematic reviews. *Current Opinion in Clinical Nutrition & Metabolic Care*, 14, 581-587.
- Whisner, C. M., Martin, B. R., Schoterman, M. H., Nakatsu, C. H., McCabe, L. D., McCabe, G. P., & Weaver, C. M. (2013). Galacto-oligosaccharides increase calcium absorption and gut bifidobacteria in young girls: a double-blind cross-over trial. *British Journal of Nutrition*, 110, 1292-1303.
- Wichienchot, S., Jatupornpipat, M., & Rastall, R. A. (2010). Oligosaccharides of pitaya (dragon fruit) flesh and their prebiotic properties. *Food Chemistry*, 120, 850-857.
- Williams, I. O., Onyenweaku, E. O., & Atangwho, I. J. (2016). Nutritional and antimicrobial evaluation of *Saccharum officinarum* consumed in Calabar, Nigeria. *African Journal of Biotechnology*, 15, 1789-1795.
- Xu, Y., Zheng, Z., Xu, Q., Yong, Q., & Ouyang, J. (2016). Efficient conversion of inulin to inulooligosaccharides through endoinulinase from *Aspergillus niger*. *Journal of Agricultural and Food Chemistry*, 64, 2612-2618.
- Yadav, R. L., & Solomon, S. (2006). Potential of developing sugarcane by-product based industries in India. *Sugar Tech*, 8, 104-111.
- Yahfoufi, N., Mallet, J. F., Graham, E., & Matar, C. (2018). Role of probiotics and prebiotics in immunomodulation. *Current Opinion in Food Science*, 20, 82-91.
- Yasmin, A., Butt, M. S., Afzaal, M., van Baak, M., Nadeem, M. T., & Shahid, M. Z. (2015). Prebiotics, gut microbiota and metabolic risks: Unveiling the relationship. *Journal of Functional Foods*, 17, 189-201.
- Yu, T., Zheng, Y. P., Tan, J. C., Xiong, W. J., Wang, Y., & Lin, L. (2017). Effects of prebiotics and synbiotics on functional constipation. *The American Journal of the Medical Sciences*, 353, 282-292.
- Yue, P. P., Hu, Y. J., Fu, G. Q., Sun, C. X., Li, M. F., Peng, F., & Sun, R. C. (2017). Structural differences between the lignin-carbohydrate complexes (LCCs) from 2- and 24-month-old bamboo (*Neosinocalamus affinis*). *International Journal of Molecular Sciences*, 19, 1.
- Zeng, H., Xue, Y., Peng, T., & Shao, W. (2007). Properties of xylanolytic enzyme system in bifidobacteria and their effects on the utilization of xylooligosaccharides. *Food*

Chemistry, 101, 1172-1177.

- Zhang, H., Yang, Y. F., & Zhou, Z. Q. (2018a). Phenolic and flavonoid contents of mandarin (*Citrus reticulata Blanco*) fruit tissues and their antioxidant capacity as evaluated by DPPH and ABTS methods. *Journal of Integrative Agriculture*, 17, 256-263.
- Zhang, W., You, Y., Lei, F., Li, P., & Jiang, J. (2018b). Acetyl-assisted autohydrolysis of sugarcane bagasse for the production of xylo-oligosaccharides without additional chemicals. *Bioresource Technology*, 265, 387-393.
- Zhang, Y. J., Li, S., Gan, R. Y., Zhou, T., Xu, D. P., & Li, H. B. (2015). Impacts of gut bacteria on human health and diseases. *International Journal of Molecular Sciences*, 16, 7493-7519.
- Zhao, Y., Chen, M., Zhao, Z., & Yu, S. (2015). The antibiotic activity and mechanisms of sugarcane (*Saccharum officinarum L.*) bagasse extract against food-borne pathogens. *Food Chemistry*, 185, 112-118.
- Zhao, Z., Yan, H., Zheng, R., Khan, M. S., Fu, X., Tao, Z., & Zhang, Z. (2018). Anthocyanins characterization and antioxidant activities of sugarcane (*Saccharum officinarum L.*) rind extracts. *Industrial Crops and Products*, 113, 38-45.
- Zheng, R., Su, S., Li, J., Zhao, Z., Wei, J., Fu, X., & Liu, R. H. (2017). Recovery of phenolics from the ethanolic extract of sugarcane (*Saccharum officinarum L.*) bagasse and evaluation of the antioxidant and antiproliferative activities. *Industrial Crops and Products*, 107, 360-369.