



**UNIVERSITI PUTRA MALAYSIA**

***METABOLIC SIGNATURE OF PROTEIN ENERGY WASTING AMONG  
MALAYSIAN HEMODIALYSIS PATIENTS***

**FATIN ATHIRAH BINTI PAUZI**

**FPSK(m) 2020 32**



**METABOLIC SIGNATURE OF PROTEIN ENERGY WASTING AMONG  
MALAYSIAN HEMODIALYSIS PATIENTS**

By

**FATIN ATHIRAH BINTI PAUZI**

**Thesis Submitted to the School of Graduate Studies, Universiti  
Putra Malaysia, in Fulfilment of the Requirements for the Degree of  
Master of Science**

**January 2020**

## COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

## **METABOLIC SIGNATURE OF PROTEIN ENERGY WASTING AMONG MALAYSIAN HEMODIALYSIS PATIENTS**

By

**FATIN ATHIRAH BT PAUZI**

**January 2020**

**Chair : Zulfetri 'Azuan bin Mat Daud, PhD**  
**Faculty : Medicine and Health Sciences**

End-stage renal disease (ESRD) patients undergoing maintenance hemodialysis (HD) often experience a state of metabolic and nutritional derangements that is highly associated with morbidity and mortality due to the protein energy wasting (PEW) disorder. Identification of PEW was recommended by International Society of Renal Nutrition and Metabolism (ISRNM) with a reported prevalence of 18-75%. However, direct quantification of muscle wasting remains unsolved for this population besides there are no consistent guidelines and standardized scoring tools in a diverse patient-wide population that making it difficult to diagnose PEW. Thus, it is prudent to identify biomarkers signature of PEW using proton nuclear magnetic resonance ( $^1\text{H-NMR}$ ) based metabolomics approach which will assist in diagnosis of PEW. In this cross-sectional study, a total of 106 subjects of ESRD patients undergoing chronic HD were stratified into PEW ( $n=53$ ) and Non-PEW (NPEW) ( $n=53$ ) based on the established ISRNM criteria. Fasting predialysis plasma samples with filtration were analyzed by  $^1\text{H-NMR}$  and the data acquired through by the combination of presaturation (PRESAT) and the Carr–Purcell–Meiboom–Gill (CPMG) pulse sequence were further processed using Chenomx. Multivariate data analysis and the statistical analysis were performed using SIMCA-P and the SPSS softwares to discriminate the metabolomics profiles between the two groups.

Orthogonal partial least square discriminant analysis (OPLS-DA) demonstrated the difference in metabolomics profiles between PEW and NPEW patients by the principal component 1 and indicates that PEW syndrome induced specific metabolic pattern alteration. Combination of loading plots and variable importance in projection (VIP) plots of OPLS-DA model provides the important variable that contributed to the group separation. A total of 32 metabolites were identified and quantified using Chenomx, with 18 metabolites belong to PEW

groups and 14 metabolites represented NPEW groups. Among all the metabolites analyzed using Mann-Whitney U Test, 3-hydroxybutyrate ( $p < 0.001$ ), acetate ( $p = 0.027$ ), arabinose ( $p = 0.029$ ), maltose ( $p = 0.021$ ), ribose ( $p = 0.041$ ), sucrose ( $p = 0.008$ ), and tartrate ( $p = 0.018$ ) were significantly increased ( $p < 0.05$ ) in PEW subjects whilst creatinine ( $p < 0.001$ ) was significantly decreased.

However, analysis of covariance (ANCOVA) revealed that the contribution of arabinose, maltose, ribose, sucrose and tartrate on the group discrimination might be affected by age and dialysis vintage. Besides, creatinine was also showed to be significantly correlated ( $p < 0.01$ ) with the anthropometric mid-arm muscle area (MAMA) ( $r_s = -0.426$ ), lean tissue mass (LTM) ( $r_s = 0.362$ ) and albumin ( $r_s = -0.315$ ). Our results signified that patients who experienced PEW had different metabolic profiles as opposed to NPEW patients. Pathway analysis indicated that PEW-related metabolites triggered perturbation in fatty acid mechanism and induced gluconeogenesis as well as glyoxylate and dicarboxylate metabolism. These results provide primary data in understanding metabolic perturbation of PEW and corresponding abnormal metabolites that potentially serve as biomarker of PEW.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Master Sains

## **METABOLIK KHUSUS BAGI PENYUSUTAN PROTIN DAN TENAGA DALAM KALANGAN PESAKIT HEMODIALISIS DI MALAYSIA**

Oleh

**FATIN ATHIRAH BT PAUZI**

Januari 2020

**Pengerusi : Zulfetri 'Azuan Mat Daud, PhD**  
**Fakulti : Perubatan dan Sains Kesihatan**

Pesakit kegagalan ginjal peringkat akhir (ESRD) yang menjalani rawatan hemodialisis (HD) sering mengalami satu keadaan di mana berlaku gangguan metabolik dan nutrisi yang berkait dengan morbiditi dan mortaliti yang disebabkan oleh sindrom penyusutan protin dan tenaga (PEW). Identifikasi PEW telah dicadangkan oleh Persatuan Nutrisi dan Metabolisme Renal Antarabangsa (ISRNM) dengan prevalens yang dilaporkan sebanyak 18-75%. Walau bagaimanapun, kuantifikasi secara langsung bagi penyusutan otot masih tidak dapat diatasi bagi populasi ini selain ketiadaan garis panduan yang konsisten dan alat penskoran yang seragam bagi populasi pesakit yang berbeza-beza yang mengakibatkan kesukaran untuk mendiagnosis PEW. Oleh itu, adalah bernas untuk mengenalpasti biomarker khusus bagi PEW dengan menggunakan pendekatan metabolomik berasaskan proton resonans magnet nukleus ( $^1\text{H-NMR}$ ) yang akan membantu dalam diagnosis PEW. Di dalam kajian keratan-rentas ini, seramai 106 subjek pesakit ESRD yang menjalani HD kronik dibahagikan kepada PEW ( $n=53$ ) dan bukan PEW (NPEW) ( $n=53$ ) berdasarkan kriteria ISRNM yang telah ditetapkan. Sampel plasma pradialisis dari pesakit yang berpuasa yang telah ditapis, dianalisis menggunakan  $^1\text{H-NMR}$  dan data yang diperolehi melalui kombinasi presertasi (PRESAT) dan *Carr-Purcell-Meiboom-Gill* (CPMG) rangkaian pulsa seterusnya diproses menggunakan Chenomx. Analisis data multivariat dan analisis statistik melalui perisian SIMCA-P dan SPSS digunakan untuk mendiskriminasi profil metabolomik di antara dua kumpulan.

Analisis *orthogonal partial least square discriminant* (OPLS-DA) menunjukkan perbezaan pada profil metabolomik di antara pesakit PEW dan NPEW pada prinsipal komponen 1 dan menunjukkan bahawa sindrom PEW menyebabkan perubahan khusus pada pola metabolit. Gabungan plot pemuatan dan plot *variable importance in projection* (VIP) bagi OPLS-DA menunjukkan variabel

penting yang menyumbang kepada pemisahan kumpulan. Sebanyak 32 metabolit dikenalpasti dan dikira kuantiti menggunakan Chenomx, di mana 18 metabolit dimiliki oleh kumpulan PEW dan 14 metabolit mewakili kumpulan NPEW. Di antara kesemua metabolit yang dianalisis menggunakan *Mann-Whitney U Test*, 3- hidroksibutirat ( $p < 0.001$ ), asetat ( $p = 0.027$ ), arabinosa ( $p = 0.029$ ), maltosa ( $p = 0.021$ ), ribosa ( $p = 0.041$ ), sukrosa ( $p = 0.008$ ), dan tartrat ( $p = 0.018$ ) menunjukkan peningkatan signifikan ( $p < 0.05$ ) pada subjek PEW manakala kreatinin ( $p = 0.000$ ) menunjukkan pengurangan signifikan.

Walaubagaimanapun, analisis kovarians (ANCOVA) menunjukkan bahawa pengaruh arabinosa, maltosa, ribosa, sukrosa dan tartrat pada pemisahan kumpulan mungkin dipengaruhi oleh umur and panjang masa dialisis. Selain itu, kreatinin juga menunjukkan signifikan korelasi ( $p < 0.01$ ) dengan kawasan otot pertengahan lengan antropometrik (MAMA) ( $r_s = 0.426$ ), jisim tisu tanpa lemak (LTM) ( $r_s = 0.362$ ) dan albumin ( $r_s = -0.315$ ). Hasil kajian ini menandakan bahawa pesakit yang mengalami PEW mempunyai perubahan metabolik yang berbeza berbanding pesakit NPEW. Analisis laluan menunjukkan bahawa metabolit yang berhubung kait dengan PEW mencetuskan perturbasi dalam mekanisme asid lemak serta mendorong glukoneogenesis dan metabolisme glioksilat dan dikarboksilat. Dapatan ini menyediakan data primer dalam memahami perubahan metabolik PEW dan metabolit abnormal sepadan yang mempunyai potensi untuk berfungsi sebagai biopenanda PEW.

## ACKNOWLEDGEMENT

First and foremost, I am deeply grateful to Allah SWT for all His blessings and as a true source of strength for me throughout this research journey.

I would like to express my greatest appreciation and sincere gratitude to my supervisor, Assoc. Prof. Zulfitri 'Azuan Mat Daud for his generous guidance, continuous encouragement, patience and motivation throughout my research study. I am very grateful especially for the opportunity that he gave me that opened the door for metabolomics analysis. Besides, I would like to thank my co-supervisors, Assoc. Prof. Dr. Faridah Abas and Dr. Nor Fadhlina Zakaria for their technical advices, valuable suggestion and support during my study. Their critical appraisal had widened my research from various perspectives.

The special gratitude should be given to Mr. Salehuddin who is in-charged in handling Nuclear Magnetic Resonance (NMR) spectrometer at Institute of Bioscience (IBS) for his training and technical assistant on the metabolomics work. Not to forget, Ministry of Higher Education for the research grant and scholarship that support me financially during my research period. To all of my postgraduate friends – thank you for the valuable encouragement and assistance throughout my study years. Without their help and support, I would not have found pleasure in this journey as much as I do today.

Last but not least, a very special appreciation to my dear husband, thank you for always encourage me to never give up and lend me emotional support during ups and downs of this research journey. To my beloved parents - thank you for the continuous prayers and moral support throughout my study. To my siblings – thank you for the prayers and love. This accomplishment would not have been possible without all of them. My regards and blessing also goes to all of those who supported me in any aspect until the completion of this study.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master Science. The members of the Supervisory Committee were as follows:

**Zulfitri 'Azuan bin Mat Daud, PhD**

Associate Professor  
Faculty Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Faridah binti Abas, PhD**

Professor  
Faculty of Food Science and Technology  
Universiti Putra Malaysia  
(Member)

**Nor Fadhlina Zakaria, MMed**

Senior Medical Lecturer  
Faculty Medicine and Health Sciences  
Universiti Putra Malaysia  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 12 November 2020

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_  
Name of Chairman  
of Supervisory  
Committee: Assoc. Prof. Zulfitri 'Azuan bin Mat Daud

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory  
Committee: Prof. Dr Faridah binti Abas

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory  
Committee: Dr. Nor Fadhlina binti Zakaria

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiii
<b>LIST OF FIGURES</b>	xiv
<b>LIST OF ABBREVIATIONS</b>	xv
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
1.1 Research background	1
1.2 Problem statement	3
1.3 Significance of the study	4
1.4 Objectives	4
1.4.1 General objective	4
1.4.2 Specific objectives	4
<b>2 LITERATURE REVIEW</b>	<b>5</b>
2.1 End-stage renal disease	5
2.1.1 Hemodialysis	7
2.1.2 Hemodialysis procedure	7
2.1.3 Complications of hemodialysis	8
2.2 Protein Energy Wasting	9
2.2.1 Etiology of PEW	9
2.2.2 Effects of PEW	11
2.2.3 Diagnosis of PEW using ISRN parameter	12
2.3 Metabolomics	14
2.3.1 Metabolomics technologies	15
2.3.2 Analytical technique of NMR	15
2.3.3 Metabolomics in CKD researches	16
2.3.4 Application of metabolomics as a new approach for diagnosis of PEW	18
<b>3 METHODOLOGY</b>	<b>19</b>
3.1 Subject selection and study design	19
3.2 Sample size determination	20
3.3 Data collection	20

3.4	Blood samples collection and measurements	27
3.5	Chemicals and reagents	27
3.6	Equipment and instrument	27
3.7	Metabolomics analyses	28
3.7.1	Buffer preparation	28
3.7.2	Sample preparation	28
3.7.3	NMR analysis	28
3.7.4	Multivariate data analysis	29
3.7.5	Metabolites identification and quantification	30
3.8	Statistical analyses	32
<b>4</b>	<b>RESULTS</b>	<b>33</b>
4.1	Demographic, anthropometric, biochemical, clinical and nutritional profiles	33
4.2	Nuclear Magnetic Resonance (NMR) spectral analysis	36
4.2.1	Metabolomics profiles separation between the groups using multivariate data analysis	38
4.2.2	Partial Least Squares Discriminant Analysis	40
4.2.3	Orthogonal Partial Least Square Discriminant Analysis	43
4.3	Metabolites identification and quantification	47
4.3.1	Identification and quantification of metabolites using Chenomx	47
4.3.2	Identification of PEW biomarkers signature	48
4.4	Metabolic pathway analysis	49
4.5	Analysis of covariance	51
4.6	Correlation of PEW potential biomarker with ISRNM PEW criteria	52
<b>5</b>	<b>DISCUSSION</b>	<b>55</b>
5.1	Postulated mechanism based on altered metabolites in PEW	56
5.1.1	Metabolites involve in gluconeogenesis	56
5.1.2	Metabolites involve in ketogenesis	57
5.1.3	Metabolites involve in carbohydrate metabolism	59
5.2	Overall postulation	60

<b>6</b>	<b>CONCLUSION, LIMITATIONS AND RECOMMENDATIONS</b>	<b>62</b>
6.1	Conclusion	62
6.2	Limitations and recommendations	62
	<b>REFERENCES</b>	<b>64</b>
	<b>BIODATA OF STUDENT</b>	<b>81</b>



## LIST OF TABLES

Table		Page
2.1	CKD stages based on level of GFR	5
2.2	Contributing factors of developing PEW in CKD patients	11
2.3	ISRNM recommended criteria for diagnosis of PEW	12
2.4	Comparison of the strengths and limitations of NMR and MS technique in metabolomics	16
3.1	Inclusion and exclusion criteria of the study	19
3.2	Characteristics used to identify PEW based on ISRNM criteria	25
4.1	Characteristics of the study population	34
4.2	CV ANOVA of PLS-DA model discriminating PEW and NPEW groups	41
4.3	Comparison on the mean concentration of metabolites identified from plasma samples of PEW and NPEW HD patients	47
4.4	Comparison on the relative concentration of discriminant metabolites between PEW and NPEW identified by <sup>1</sup> H-NMR	49
4.5	Analysis of covariance (ANCOVA) between selected factors on significant metabolites with controlled covariates	53
4.6	Spearman's rank correlation coefficient between potential biomarkers and selected parameters in ISRNM diagnostic criteria	54

## LIST OF FIGURES

Figure		Page
3.1	Study design	21
3.2	Graphical conceptual framework	26
3.3	Steps in metabolomics analysis	31
4.1	Representative of <sup>1</sup> H-NMR Carr-Purcell-Meiboom-Gill (CPMG) spectra of plasma samples obtained from a) PEW subjects and b) NPEW subjects	37
4.2	PCA-X score plot for NMR spectra acquired from plasma samples of PEW and NPEW HD patients	38
4.3	PCA-X score plot with 9 outliers removed for NMR spectra acquired from plasma samples of PEW and NPEW HD patients	39
4.4	PLS-DA score plot for NMR spectra acquired from plasma samples of PEW and NPEW HD patients	40
4.5	Permutation test for validation of PLS-DA model for a) PEW groups and b) NPEW groups	42
4.6	OPLS-DA score plot for NMR spectra acquired from plasma samples of PEW and NPEW HD patients	44
4.7	OPLS-DA loading plot with identified metabolites for NMR spectra acquired from plasma samples of PEW and NPEW HD patients	45
4.8	Variable importance in the projection (VIP) plot	46
4.9	Corresponding loading plot of OPLS-DA with a confluence of VIP results	46
4.10	Proposed metabolic pathway of altered metabolites in PEW metabolic profile	50

## LIST OF ABBREVIATIONS

<sup>1</sup> H-NMR	Proton Nuclear Magnetic Resonance Spectroscopy
ANOVA	Analysis of variance
ANCOVA	Analysis of covariance
BMI	Body mass index
B/P	Blood pressure
CKD	Chronic Kidney Disease
CPMG	Carr-Purcell-Meiboom-Gill
CVD	Cardiovascular disease
DEI	Dietary energy intake
DPI	Dietary protein intake
ESRD	End Stage Renal Disease
FTM	Fat tissue mass
GC-MS	Gas Chromatography-Mass Spectrometry
GNRI	Geriatric Nutritional Risk Index
HD	Haemodialysis
HDLC	High density lipoprotein cholesterol
HGS	Handgrip strength
Hgb	Hemoglobin
hsCRP	high-sensitivity C-reactive protein
Hz	Hertz
ISRNM	International Society of Renal Nutrition and Metabolism
KDIGO	Kidney Disease: Improving Global Outcomes
Kt/V	Index of dialysis adequacy
LC-MS	Liquid Chromatography-Mass Spectrometry
LDLC	Low density lipoprotein cholesterol
LTM	Lean tissue mass
MAMA	Mid-arm muscle area
MAMC	Mid-arm muscle circumference
MHz	Megahertz
MIS	Malnutrition Inflammation Score
MVDA	Multivariate data analysis
NKF	National Kidney Foundation
NMR	Nuclear Magnetic Resonance
NOESY	Nuclear Overhauser Effect Spectroscopy
OPLS-DA	Orthogonal Partial Least Squares-Discriminant Analysis
PATCH	Palm Tocotrienols in Chronic Hemodialysis Patients
PCA	Partial Component Analysis
PEW	Protein Energy Wasting
PLS	Partial Least Square
PLS-DA	Partial Least Square-Discriminant Analysis
ppm	parts per million
QOL	Quality of Life
SD	Standard deviation
SE	Standard error
SGA	Subjective Global Assessment
SIMCA	Soft Independent Modelling of Class Analogy
SPSS	Statistical Package for the Social Sciences



TAG	Triacylglyceride
TC	Total cholesterol
TSF	Triceps skinfold
TSP	Trimethylsilylpropanoic acid
WC	Waist circumferences
$\delta$	Chemical shift in ppm



# CHAPTER 1

## INTRODUCTION

### 1.1 Research background

End-stage renal disease (ESRD) is an advanced stage of chronic kidney disease (CKD) that has been acknowledged as a public health problem worldwide (Yokote & Yokoo, 2017). Malaysia is reported as one of the country with ESRD prevalence per million populations that has steadily increased by 163.7% within 14 years (US Renal Data System, 2018). Hemodialysis (HD) continues to be the most common mode of therapy worldwide including Malaysia (Wong & Goh, 2018). While the dialysis therapy can assist in improving serum creatinine, albumin and prealbumin, normalises the protein catabolic rate (nPCR) as well as increases the dietary intake of patients (Mehrotra et al., 2002; Pupim et al., 2002), it is also highly associated with malnutrition and lower the quality of life (QOL) of the patients (Liu et al., 2006; Rutledge & McMahon, 2000). In fact, high incidence of malnutrition is a major contributor to morbidity and mortality in advanced CKD patients undergoing HD.

The Malaysian Dialysis and Transplant Registry rely on two nutrition-related parameters, namely BMI and serum albumin to screen HD patients for malnutrition. However, these nutritional parameters are inadequate to identify an alternate view of malnutrition known as protein energy malnutrition, or recently known as protein energy wasting (PEW) syndrome. PEW has a wide prevalent in dialysis population with 18-75% of CKD patients undergoing maintenance dialysis therapy exhibit wasting according to International Society of Renal Nutrition and Metabolism (ISRNM) proposed criteria and this has been associated with increased risk of mortality and adverse clinical outcomes in this population (Leinig et al., 2011; Kalantar-Zadeh, Ikizler, Block, Avram, & Kopple, 2003; Mehrotra & Kopple, 2001; Kopple, 1997). PEW is referred as multiple nutrition and catabolic alterations that caused a chronic decreased of protein stores and energy fuels in the body, exhibited by low body protein and fat masses (Carrero et al., 2013). Causes of PEW in HD patients are multifactorial including abnormalities in energy and protein metabolism due to uremic toxicity, metabolic acidosis, chronic inflammation, co-morbid illnesses, multiple endocrine disorder, dialysis related hypermetabolism as well as nutrient losses into dialysate that eventually leads to excess catabolism of muscle and fat (Carrero et al., 2013; Fouque et al., 2008; Gracia-Iguacel et al., 2014; Ikizler et al., 1994). Besides, ageing, poor physical activity, frailty, gender differences and patients' socioeconomic status can also contributed to the presence of PEW in dialysis patients (Carrero et al., 2013; Ikizler et al., 2013). Consequently, PEW patients usually experienced nutritional and metabolic derangement that include persistent inflammation, metabolic acidosis, gradual

weight loss and a state of hypercatabolic leading to break down of muscle and fat tissues as well as reduction of serum proteins (Carrero et al., 2013; Avesani et al., 2006). According to Carrero et al., (2013) even though dialysis reverses uremia, residual metabolic derangements, inflammation, comorbid conditions and the dialysis procedure itself may allow PEW to develop or worsen.

PEW is common in patients with CKD and is associated with adverse clinical outcomes, particularly in individuals with CKD stage G3b, G4 and G5 (eGFR<45 ml/min/1.73 m<sup>2</sup> of the body surface area) (Obi, Qader, Kovesdy, & Kalantar-Zadeh, 2015). This syndrome amongst HD and peritoneal dialysis (PD) patients has been a worldwide incidence since the 1990s (Kopple, 1997; Cianciaruso et al., 1995) and is correlated with the increment of mortality and morbidity cases (Sung Hee Chung, Lindholm, & Lee, 2003; Marcen, Teruel, De La Cal, & Gamez, 1997). Therefore, in 2008, the diagnosis criteria of PEW were presented by the ISRNM. According to ISRNM, PEW is diagnosed if three out of four characteristics are present; (i) low levels of body serum (albumin, transthyretin, or cholesterol); (ii) reduced body mass (low or reduced body or fat mass); (iii) reduced muscle mass (muscle wasting or sarcopenia, reduced mid-arm muscle circumference) and (iv) unintentional low energy and protein intake (Fouque et al., 2008). However, emerging studies are challenging the application of these parameters (Gracia-Iguacel et al., 2014; Drechsler et al., 2011; Leinig et al., 2011; Vegine, Fernandes, Torres, Silva, & Avesani, 2011) as this diagnostic criteria lacks biological validation and more intensive research to identify reliable PEW biomarkers are required.

Although no study on identifying PEW using the metabolomics approach has been carried out to date, current literature across multiple metabolomics studies have been able to identify several potential biomarkers linked to progression of CKD (Jia et al., 2008; Mutsaers et al., 2013; Qi et al., 2012; Rhee et al., 2010; Schefold et al., 2009) enabling CKD detection and prediction at much earlier stage than conventional test. This method offers the potential for a holistic approach to clinical medicine, as well as improvements in disease diagnoses and the understanding of pathological mechanisms. Therefore, we propose to use the cutting edge proton nuclear magnetic resonance (<sup>1</sup>H-NMR)-based metabolomics approach to facilitate identification of plasma metabolites associated with the catabolism of muscle and fat tissues related to the facets of PEW development in Malaysian HD patients. The principal of the metabolomics techniques is to identify global small-molecule metabolite profiles from complex samples like biofluids and tissue using a non-targeted and unbiased approach (Zhao, 2013). Metabolomics measures metabolites within a biological system at a given time, and profiling of these metabolites can provide detailed information on how the disease affects the metabolites physiologically or pathologically and represents a useful tool for biomarker detection (Friedrich, 2012; Zhang et al., 2014). In view of this, metabolomics approach will serve as the primary data for a more in depth knowledge on PEW specifically related to its mechanistic explanation.

## 1.2 Problem statement

Traditional nutritional status measurements indicate that prevalence of PEW can be as high as 75% in dialysis population (Leinig et al., 2011). In fact, prevalence of PEW has been reported to occur in 59% of HD population in two major hospitals in Malaysia (Harvinder et al., 2016). PEW is not only interrelated with infection, CVD, frailty, and depression, but these complications may also increase the extent of PEW itself in a vicious cycle. Even worse, this syndrome of detrimental changes in nutrition and body composition lead to heightened morbidity and mortality particularly in dialysis population (Carrero et al., 2013). Due to the fact that PEW is caused by multifactorial conditions, this signify the need for a global diagnostic approach that consider a panel of biomarkers as opposed to a single biomarker in a traditional system.

The ISRNM provided guidelines in identifying PEW. However, a few studies reported that they identified less malnourished patients using diagnostic criteria proposed by ISRNM compared to the other tools (Gracia-Iguacel et al., 2014; Drechsler et al., 2011; Leinig et al., 2011; Vegine et al., 2011). According to Obi et al. (2015), studies that applied serial assessment of nutritional status using scoring tools to identified PEW patients, including the subjective global assessment (SGA), the malnutrition inflammation score (MIS) and geriatric nutritional risk index (GNRI) might have better association with hospitalization and mortality as well as nutritional and inflammatory parameters in the final stage CKD patients. These emerging studies are challenging the application of the ISRNM diagnostic criteria which have not been validated against biomarkers of metabolic changes that are underlying the catabolism of muscle and fat tissue associated with PEW. In fact, extrapolation of ISRNM diagnostic criteria to different HD population has also been questioned (Gracia-Iguacel et al., 2014).

On that account, the use of  $^1\text{H-NMR}$  based metabolomics approach is postulated to facilitate identification of plasma metabolites changes associated with PEW and explain the PEW mechanism in HD population. Because metabolomics in renal practice is relatively new, literature linking metabolic parameters assessed through metabolomics platform is scarce. To the best of our knowledge, no study on identifying PEW using the metabolomics approach has been carried out to date. Therefore, it is of interest to find out the prevalence of PEW in Malaysian HD patients using the diagnostic criteria proposed by the ISRNM that works simultaneously with the metabolomics techniques. This research is also attempts to investigate the relationships of the plasma metabolic profile with conventional parameters with regards on identification of PEW.

### **1.3 Significance of the study**

As there is no study has reported on metabolomics approach to identify PEW in HD patients, this finding will serve as the pilot and primary data for a more in depth knowledge on PEW specifically related to its mechanistic explanation and the differences in plasma metabolites between PEW and Non-PEW HD patients. The baselines data of plasma metabolic profiles of PEW patients can be established together with a discovery of new biomarkers that eventually could be used for the identification of therapeutic targets and selection of the most suitable treatment that are critical to the health status of HD patients. Likewise, new PEW metabolic diagnostic criteria will provide the basis for future work to increase awareness, identify research needs, and screen for malnutrition in patients with CKD. This is in line with Ministry of Health's National Strategic Plan for Non-Communicable Disease that emphasized on prevention, early detection and diagnosis as a cost-effective approach. This innovation will impact the society, economy and nation in the positive perspectives. It is important to search actively for diagnostic and therapeutic approach of PEW since early diagnosis and treatment can improve the prognosis for CKD patients and reduce the monetary costs connected with treatment.

### **1.4 Objectives**

#### **1.4.1 General objective**

To identify biomarkers signature of hemodialysis patients experiencing protein energy wasting using the <sup>1</sup>H-NMR based metabolomics approach.

#### **1.4.2 Specific objective**

1. To determine demographic, anthropometric, biochemical, clinical and nutritional profiles of PEW and Non-PEW (NPEW) patients.
2. To characterize and discriminate metabolic profiles of PEW and NPEW HD patients using non-targeted metabolomics approach.
3. To identify potential biomarker of PEW and its association with ISRNM diagnostic criteria.

## REFERENCES

- An, L., Shi, Q., & Feng, F. (2015). Metabolomics approach to identify therapeutically potential biomarkers of the Zhi-Zi-Da-Huang decoction effect on the hepatoprotective mechanism. *RSC Advances*, 5(102), 84048–84055.
- Andrews, R., Greenhaff, P., Curtis, S., Perry, A., & Cowley, A. J. (1998). The effect of dietary creatine supplementation on skeletal muscle metabolism in congestive heart failure. *European Heart Journal*, 19(4), 617–622.
- Aoki, J., Taira, A., Takanezawa, Y., Kishi, Y., Hama, K., Kishimoto, T., ... Arai, H. (2002). Serum Lysophosphatidic Acid Is Produced through Diverse Phospholipase Pathways. *Journal of Biological Chemistry*, 277(50), 48737–48744.
- Arnold, M. H. M. (1975). *Acidulants for foods and beverages*. London: Food Trade Press.
- Avesani, C. M., Carrero, J. J., Axelsson, J., Qureshi, A. R., Lindholm, B., & Stenvinkel, P. (2006). Inflammation and wasting in chronic kidney disease: partners in crime. *Kidney International*, 70, S8–S13.
- Axelsson, J., Qureshi, A. R., Divino-Filho, J. C., Bárány, P., Heimbürger, O., Lindholm, B., & Stenvinkel, P. (2006). Are insulin-like growth factor and its binding proteins 1 and 3 clinically useful as markers of malnutrition, sarcopenia and inflammation in end-stage renal disease? *European Journal of Clinical Nutrition*, 60(6), 718-726.
- Bailey, J. L., Wang, X., England, B. K., Price, S. R., Ding, X., & Mitch, W. E. (1996). The acidosis of chronic renal failure activates muscle proteolysis in rats by augmenting transcription of genes encoding proteins of the ATP-dependent ubiquitin-proteasome pathway. *The Journal of Clinical Investigation*, 97(6), 1447–1453.
- Beckonert, O. P., Keun, H. C., Ebbels, T. M. D., Bundy, J. G., Holmes, E., Lindon, J. C., & Nicholson, J. K. (2007). Metabolic profiling, metabolomic and metabonomic procedures for NMR spectroscopy of urine, plasma, serum and tissue extracts. *Nature Protocols*, 2(11), 2692–2703.
- Berg, J. M., Tymoczko, J. L., & Stryer, L. (2002). Food intake and starvation induce metabolic changes. In *Biochemistry* (5th ed.). New York: W H Freeman.
- Bergstrom, J., & Lindholm, B. (1998). Malnutrition, cardiac disease, and mortality: an integrated point of view. *American Journal of Kidney Diseases*, 32(5), 834–841.
- Bhagavan, N. V. (2002). CHAPTER 22 - Metabolic Homeostasis. In N.V.

- Vhagavan (Ed.) *Medical Biochemistry*, (pp. 485–519). San Diego: Academic Press.
- Bharadwaj, S., Ginoya, S., Tandon, P., Gohel, T. D., Guirguis, J., Vallabh, H., ... Hanouneh, I. (2016). Malnutrition: Laboratory markers vs nutritional assessment. *Gastroenterology Report*, 4(4), 272–280.
- Bilbrey, G. L. (1989). Identification and treatment of protein calorie malnutrition in chronic hemodialysis patients. *Dialysis and Transplantation*, 18, 669–678.
- Brereton, R. G., & Lloyd, G. R. (2014). Partial least squares discriminant analysis: Taking the magic away. *Journal of Chemometrics*, 28(4), 213–225.
- Burgess, K., Rankin, N., & Weidt, S. (2014). Chapter 10 - Metabolomics. In S. Padmanabhan (Ed.), *Handbook of Pharmacogenomics and Stratified Medicine* (pp. 181–205). San Diego: Academic Press.
- Butch, C., Cope, E. D., Pollet, P., Gelbaum, L., Krishnamurthy, R., & Liotta, C. L. (2013). Production of tartrates by cyanide-mediated dimerization of glyoxylate: A potential abiotic pathway to the citric acid cycle. *Journal of the American Chemical Society*, 135(36), 13440–13445.
- Bylesjö, M., Rantalainen, M., Cloarec, O., Nicholson, J. K., Holmes, E., & Trygg, J. (2006). OPLS discriminant analysis: combining the strengths of PLS-DA and SIMCA classification. *Journal of Chemometrics: A Journal of the Chemometrics Society*, 20(8-10), 341–351.
- Cameron, N. (2012). The measurement of human growth. In N. Cameron & B. Bogin (Eds.), *Human growth and development* (pp. 487–513). San Diego: Academic Press.
- Carrero, J. J., Stenvinkel, P., Cuppari, L., Ikizler, T. A., Kalantar-Zadeh, K., Kaysen, G., ... Franch, H. A. (2013). Etiology of the Protein-Energy Wasting Syndrome in Chronic Kidney Disease: A Consensus Statement From the International Society of Renal Nutrition and Metabolism (ISRNM). *Journal of Renal Nutrition*, 23(2), 77–90.
- Casadei, K., & Kiel, J. (2019). Anthropometric Measurement. In *StatPearls*. Treasure Island (FL): StatPearls Publishing.
- Chak, C. M., Lacruz, M. E., Adam, J., Brandmaier, S., Covic, M., Huang, J., ... Wang-Sattler, R. (2019). Ageing investigation using two-time-point metabolomics data from KORA and CARLA studies. *Metabolites*, 9(3), 44.
- Chattaway, F. D., & Ray, F. E. (1921). II.—The decomposition of tartaric acid by heat. *Journal of the Chemical Society, Transactions*, 119, 34–37.
- Chazot, C., Shahmir, E., Matias, B., Laidlaw, S., & Kopple, J. D. (1997). Dialytic nutrition: provision of amino acids in dialysate during hemodialysis.

*Kidney International*, 52(6), 1663–1670.

Chertow, G. M., Johansen, K. L., Lew, N., Lazarus, J. M., & Lowrie, E. G. (2000). Vintage, nutritional status, and survival in hemodialysis patients. *Kidney International*, 57(3), 1176–1181.

Chmielewski, M., Carrero, J. J., Qureshi, A. R., Axelsson, J., Heimbürger, O., Berglund, L., ... Stenvinkel, P. (2009). Temporal discrepancies in the association between the apoB/apoA-I ratio and mortality in incident dialysis patients. *Journal of Internal Medicine*, 265(6), 708–716.

Chung, S. H., Lindholm, B., & Lee, H. B. (2003). Is malnutrition an independent predictor of mortality in peritoneal dialysis patients? *Nephrology Dialysis Transplantation*, 18(10), 2134–2140.

Chung, S., Koh, E. S., Shin, S. J., & Park, C. W. (2012). Malnutrition in patients with chronic kidney disease. *Open Journal of Internal Medicine*, 02(02), 89–99.

Cianciaruso, B., Brunori, G., Kopple, J. D., Traverso, G., Panarello, G., Enia, G., ... Viglino, G. (1995). Cross-sectional comparison of malnutrition in continuous ambulatory peritoneal dialysis and hemodialysis patients. *American Journal of Kidney Diseases*, 26(3), 475–486.

Dagley, S., & Trudgill, P. W. (1963). The metabolism of tartaric acid by a *Pseudomonas*. A new pathway. *Biochemical Journal*, 89(1), 22-31.

Davis, E. J. (1968). On the nature of malonate-insensitive oxidation of pyruvate and glutamate by heart sarcosomes. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, 162(1), 1–10.

Dong, J., Li, Y., Xu, Y., & Xu, R. (2011). Daily protein intake and survival in patients on peritoneal dialysis. *Nephrology Dialysis Transplantation*, 26(11), 3715–3721.

Down, W. H., Sacharin, R. M., Chasseaud, L. F., Kirkpatrick, D., & Franklin, E. R. (1977). Renal and bone uptake of tartaric acid in rats: comparison of L (+) and DL-forms. *Toxicology*, 8(3), 333–346.

Drechsler, C., Grootendorst, D. C., Pilz, S., Tomaschitz, A., Krane, V., Dekker, F., ... Wanner, C. (2011). Wasting and sudden cardiac death in hemodialysis patients: a post hoc analysis of 4D (Die Deutsche Diabetes Dialyse Studie). *American Journal of Kidney Diseases*, 58(4), 599–607.

Eknoyan, G., Lameire, N., Barsoum, R., Eckardt, K.-U., Levin, A., Levin, N., & Locatelli, F. (2004). The burden of kidney disease: Improving global outcomes. *Kidney International*, 66, 1310–1314.

Emery, P. W. (2005). Metabolic changes in malnutrition. *Eye*, 19(10), 1029-1034.



- Eriksson, L., Antti, H., Gottfries, J., Holmes, E., Johansson, E., Lindgren, F., ... Wold, S. (2004). Using chemometrics for navigating in the large data sets of genomics, proteomics, and metabonomics (gpm). *Analytical and Bioanalytical Chemistry*, 380(3), 419–429.
- Eriksson, L., Johansson, E., Kettaneh-Wold, N., Trygg, J., Wikstrom, C., & Wold, S. (2006). *Multi- and Megavariate Data Analysis. Part I Basic Principles and Applications* (Second rev). Umea Sweden: MKS Umetrics AB.
- Eriksson, Lennart, Trygg, J., & Wold, S. (2008). CV-ANOVA for significance testing of PLS and OPLS® models. *Journal of Chemometrics*, 22(11–12), 594–600.
- Fiehn, O. (2002). Metabolomics—the link between genotypes and phenotypes. *Plant Molecular Biology*, 48(1-2), 155–171.
- Finkle, P. (1933). The fate of tartaric acid in the human body. *Journal of Biological Chemistry*, 100, 349–355.
- Fituri, N., Allawi, N., Bentley, M., & Costello, J. (1985). The effect of a high intake of tartaric acid on urinary and plasma oxalate. In P.O Schwille, L.H. Smith, W.G. Robertson & W. Vahlensieck (Eds), *Urolithiasis and Related Clinical Research* (pp. 379–382). Boston: Springer.
- Floegel, A., Stefan, N., Yu, Z., Mühlenbruch, K., Drogan, D., Joost, H.-G., ... Pischon, T. (2013). Identification of Serum Metabolites Associated With Risk of Type 2 Diabetes Using a Targeted Metabolomic Approach. *Diabetes*, 62(2), 639–648.
- Fonville, J. M., Maher, A. D., Coen, M., Holmes, E., Lindon, J. C., & Nicholson, J. K. (2010). Evaluation of Full-Resolution J-Resolved <sup>1</sup>H NMR Projections of Biofluids for Metabonomics Information Retrieval and Biomarker Identification. *Analytical Chemistry*, 82(5), 1811–1821.
- Fouque, D., Kalantar-Zadeh, K., Kopple, J., Cano, N., Chauveau, P., Cuppari, L., ... Wanner, C. (2008). A proposed nomenclature and diagnostic criteria for protein–energy wasting in acute and chronic kidney disease. *Kidney International*, 73(4), 391–398.
- Fouque, D., Peng, S., Shamir, E., & Kopple, J. (2000). Recombinant human insulin-like growth factor-1 induces an anabolic response in malnourished CAPD patients. *Kidney International*, 57(2), 646–654.
- Friedrich, N. (2012). Metabolomics in diabetes research. *Journal of Endocrinology*, 215(1), 29–42.
- Gama-Axelsson, T., Heimbürger, O., Stenvinkel, P., Bárány, P., Lindholm, B., & Qureshi, A. R. (2012). Serum albumin as predictor of nutritional status in patients with ESRD. *Clinical Journal of the American Society of Nephrology*, 7(9), 1446–1453.

- Geamanu, A., Gupta, S. V., Bauerfeld, C., & Samavati, L. (2016). Metabolomics connects aberrant bioenergetic, transmethylation, and gut microbiota in sarcoidosis. *Metabolomics*, 12(2), 35.
- Gordon, A. (2015). Biochemistry of Hypoglycin and Toxic Hypoglycemic Syndrome. In A. Gordon (Ed.), *Food Safety and Quality Systems in Developing Countries* (pp. 47–61). San Diego: Academic Press.
- Gotch, F. A., & Sargent, J. A. (1985). A Mechanistic analysis of the National Cooperative Study (NCDS). *Kidney International*, 28, 526–534.
- Gracia-Iguacel, C., González-Parra, E., Barril-Cuadrado, G., Sánchez, R., Egido, J., Ortiz-Arduán, A., & Carrero, J. J. (2014). Defining protein-energy wasting syndrome in chronic kidney disease: prevalence and clinical implications. *Nefrología*, 34(4), 507–519.
- Gracia-Iguacel, C., González-Parra, E., Pérez-Gómez, M. V., Mahillo, I., Egido, J., Ortiz, A., & Carrero, J. J. (2013). Prevalence of protein-energy wasting syndrome and its association with mortality in haemodialysis patients in a centre in Spain. *Nefrología (English Edition)*, 33(4), 495–505.
- Gregory, J. F., Park, Y., Lamers, Y., Bandyopadhyay, N., Chi, Y. Y., Lee, K., ... Jones, D. P. (2013). Metabolomic Analysis Reveals Extended Metabolic Consequences of Marginal Vitamin B-6 Deficiency in Healthy Human Subjects. *PLoS ONE*, 8(6), e0063544
- Guarnieri, G., Toigo, G., Situlin, R., Faccini, L., Coli, U., Landini, S., ... Campanacci, L. (1983). Muscle biopsy studies in chronically uremic patients: evidence for malnutrition. *Kidney International. Supplement*, 16, S187-S193.
- Hakim, R. M. (2014). Hemodialysis. In S. Gilbert & D. Weiner (Eds.), *National Kidney Foundation Primer on Kidney Diseases* (pp. 508–519). Philadelphia: Saunders.
- Harvinder, G. S., Swee, W. C. S., Karupaiah, T., Sahathevan, S., Chinna, K., Ahmad, G., ... Goh, B. L. (2016). Dialysis malnutrition and malnutrition inflammation scores: Screening tools for prediction of dialysis - related protein-energy wasting in Malaysia. *Asia Pacific Journal of Clinical Nutrition*, 25(1), 26–33.
- Heimbürger, O. (2010). Peritoneal Physiology. In J. Himmelfarb & M.H. Sayegh (Eds.), *Chronic Kidney Disease, Dialysis, and Transplantation* (pp. 387–404). Philadelphia: Saunders.
- Himmelfarb, J., & Ikizler, T. A. (2010). Hemodialysis. *New England Journal of Medicine*, 363(19), 1833–1845.
- Hoher, B., & Adamski, J. (2017). Metabolomics for clinical use and research in chronic kidney disease. *Nature Reviews Nephrology*, 13(5), 269–284.

- Horgan, R. P., & Kenny, L. C. (2011). Omic technologies: proteomics and metabolomics. *The Obstetrician & Gynaecologist*, 13, 189–195.
- Hurlbert, R. E., & Jakoby, W. B. (1965). Tartaric acid metabolism I. Subunits of L (+)-tartaric acid dehydrase. *Journal of Biological Chemistry*, 240(7), 2772–2777.
- Ikizler, T. A., Flakoll, P. J., Parker, R. A., & Hakim, R. M. (1994). Amino acid and albumin losses during hemodialysis. *Kidney International*, 46(3), 830–837.
- Ikizler, T. A., Greene, J. H., Wingard, R. L., Parker, R. A., & Hakim, R. M. (1995). Spontaneous dietary protein intake during progression of chronic renal failure. *Journal of the American Society of Nephrology*, 6(5), 1386 – 1391.
- Ikizler, T., Cano, N., Franch, H., Fouque, D., Himmelfarb, J., Kalantar-Zadeh, K., ... Teta, D. (2013). Prevention and treatment of protein energy wasting in chronic kidney disease patients: a consensus statement by the International Society of Renal Nutrition and Metabolism. *Kidney International*, 84(6), 1096–1107.
- Inker, L. A., Schmid, C. H., Tighiouart, H., Eckfeldt, J. H., Feldman, H. I., Greene, T., ... Levey, A. S. (2012). Estimating Glomerular Filtration Rate from Serum Creatinine and Cystatin C. *New England Journal of Medicine*, 367(1), 20–29.
- Ishikawa, M., Maekawa, K., Saito, K., Senoo, Y., Urata, M., Murayama, M., ... Saito, Y. (2014). Plasma and Serum Lipidomics of Healthy White Adults Shows Characteristic Profiles by Subjects' Gender and Age. *PLOS ONE*, 9(3), e91806.
- Jacob, V., Le Carpentier, J. E., Salzano, S., Naylor, V., Wild, G., Brown, C. B., & El Nahas, A. M. (1990). IGF-I, a marker of undernutrition in hemodialysis patients. *The American Journal of Clinical Nutrition*, 52(1), 39–44.
- Jacobsen, N. E. (2007). *NMR spectroscopy explained: simplified theory, applications and examples for organic chemistry and structural biology*. Hoboken: John Wiley & Sons Inc.
- Jarrett, I. G., Filsell, O. H., & Ballard, F. J. (1974). Metabolic and endocrine interrelationships in normal and diabetic sheep. *Hormone and Metabolic Research, Suppl 4*, 111–116.
- Ji, Y., Hebring, S., Zhu, H., Jenkins, G. D., Biernacka, J., Snyder, K., ... Weinshilboum, R. M. (2011). Glycine and a glycine dehydrogenase (GLDC) SNP as citalopram/escitalopram response biomarkers in depression: pharmacometabolomics-informed pharmacogenomics. *Clinical Pharmacology & Therapeutics*, 89(1), 97–104.

- Jia, L., Chen, J., Yin, P., Lu, X., & Xu, G. (2008). Serum metabolomics study of chronic renal failure by ultra performance liquid chromatography coupled with Q-TOF mass spectrometry. *Metabolomics*, 4(2), 183–189.
- Johnson, C. H., Ivanisevic, J., & Siuzdak, G. (2016). Metabolomics: Beyond biomarkers and towards mechanisms. *Nature Reviews Molecular Cell Biology*, 17(7), 451–459.
- Jones, C. H., Wolfenden, R. C., & Wells, L. M. (2004). Is subjective global assessment a reliable measure of nutritional status in hemodialysis? *Journal of Renal Nutrition*, 14(1), 26–30.
- Kalantar-Zadeh, K., Ikizler, T. A., Block, G., Avram, M. M., & Kopple, J. D. (2003). Malnutrition-inflammation complex syndrome in dialysis patients: causes and consequences. *American Journal of Kidney Diseases*, 42(5), 864–881.
- Kalantar-Zadeh, K., & Kopple, J. D. (2001). Relative contributions of nutrition and inflammation to clinical outcome in dialysis patients. *American Journal of Kidney Diseases*, 38(6), 1343–1350.
- Kalantar-Zadeh, K., Kopple, J. D., Block, G., & Humphreys, M. H. (2001). A malnutrition-inflammation score is correlated with morbidity and mortality in maintenance hemodialysis patients. *American Journal of Kidney Diseases*, 38(6), 1251–1263.
- Kalantar-Zadeh, K., Kopple, J. D., Humphreys, M. H., & Block, G. (2004). Comparing outcome predictability of markers of malnutrition–inflammation complex syndrome in haemodialysis patients. *Nephrology Dialysis Transplantation*, 19(6), 1507–1519.
- Kamimura, M. A., Draibe, S. A., Dalboni, M. A., Cendoroglo, M., Avesani, C. M., Manfredi, S. R., ... Cuppari, L. (2007). Serum and cellular interleukin-6 in haemodialysis patients: relationship with energy expenditure. *Nephrology Dialysis Transplantation*, 22(3), 839–844.
- Kaysen, G. A. (2005). Diabetes, a cause of progressive sarcopenia in dialysis patients? *Kidney International*, 68(5), 2396–2397.
- Kipp, R., & Kellerman, P. S. (2009). Chronic Kidney Disease. In A.V. Moorthy (Ed.), *Pathophysiology of Kidney Disease and Hypertension* (pp. 145–157). Philadelphia: Saunders.
- Kirwan, G. M., Johansson, E., Kleemann, R., Verheij, E. R., Wheelock, Å. M., Goto, S., ... Wheelock, C. E. (2012). Building multivariate systems biology models. *Analytical Chemistry*, 84(16), 7064–7071.
- Knight, J., Assimos, D. G., Easter, L., & Holmes, R. P. (2010). Metabolism of fructose to oxalate and glycolate. *Hormone and Metabolic Research*, 42(12), 868–873.
- Knowles, S., Jarrett, I., Filsell, O., & Ballard, F. (1974). Production and

- utilization of acetate in mammals. *Biochemical Journal*, 142(2), 401–411.
- Kohn, L. D. (1968). Tartaric acid metabolism VIII. Crystalline tartronic semialdehyde reductase. *Journal of Biological Chemistry*, 243(17), 4426–4433.
- Kopple, J. D. (1994). Effect of nutrition on morbidity and mortality in maintenance dialysis patients. *American Journal of Kidney Diseases*, 24(6), 1002–1009.
- Kopple, J. D. (1997). McCollum Award Lecture, 1996: protein-energy malnutrition in maintenance dialysis patients. *The American Journal of Clinical Nutrition*, 65(5), 1544–1557.
- Kopple, J. D., Cianciaruso, B., & Massry, S. G. (1980). Does parathyroid hormone cause protein wasting? In G.M. Berlyne & M.M. Avram (Eds.), *Parathyroid Hormone in Kidney Failure* (Vol. 20, pp. 138–148). Basel: Karger Publishers.
- Kovesdy, C. P., George, S. M., Anderson, J. E., & Kalantar-zadeh, K. (2009). *Outcome predictability of biomarkers of protein-energy wasting and inflammation in moderate and advanced chronic kidney disease 1 – 4*. 407–414.
- Krebs, H. A. (1942). The effect of inorganic salts on the ketone decomposition of oxaloacetic acid. *The Biochemical Journal*, 36(3–4), 303–305.
- Krumsiek, J., Mittelstrass, K., Do, K. T., Stücker, F., Ried, J., Adamski, J., ... Kastenmüller, G. (2015). Gender-specific pathway differences in the human serum metabolome. *Metabolomics*, 11(6), 1815–1833.
- Lajis, N., Maulidiani, M., Abas, F., & Ismail, I. S. (2017). Metabolomics Approach in Pharmacognosy. In S.Badal & R. Delgoda (Eds.), *Pharmacognosy* (pp. 597-616). Cambridge: Academic Press.
- Lecker, S. H., Jagoe, R. T., Gilbert, A., Gomes, M., Baracos, V., Bailey, J., ... Goldberg, A. L. (2004). Multiple types of skeletal muscle atrophy involve a common program of changes in gene expression. *The FASEB Journal*, 18(1), 39–51.
- Lee, K. M., Jeon, J. Y., Lee, B. J., Lee, H., & Choi, H. K. (2017). Application of metabolomics to quality control of natural product derived medicines. *Biomolecules and Therapeutics*, 25(6), 559–568.
- Leinig, C. E., Moraes, T., Ribeiro, S., Riella, M. C., Olandoski, M., Martins, C., & Pecoits-Filho, R. (2011). Predictive Value of Malnutrition Markers for Mortality in Peritoneal Dialysis Patients. *Journal of Renal Nutrition*, 21(2), 176–183.
- Litbarg, N. O. (2018). Chronic Kidney Disease. In D. Rakel (Ed.), *Integrative Medicine*, (pp. 411–421). Philadelphia: Elsevier.

- Liu, W. J., Chew, T. F., Chiu, A. S., & Zaki, M. (2006). Quality of life of dialysis patients in Malaysia. *The Medical Journal of Malaysia*, 61(5), 540–546.
- Liu, X., Hoene, M., Wang, X., Yin, P., Häring, H., & Lehmann, R. (2018). Serum or plasma, what is the difference? Investigations to facilitate the sample material selection decision making process for metabolomics studies and beyond. *Analytica Chimica Acta*, 1037, 292-300.
- Locatelli, F., Manzoni, C., & Di Filippo, S. (2002). The importance of convective transport. *Kidney International*, 61, S115–S120.
- Lopes, A. A., Bragg-Gresham, J. L., Elder, S. J., Ginsberg, N., Goodkin, D. A., Pifer, T., ... Akizawa, T. (2010). Independent and joint associations of nutritional status indicators with mortality risk among chronic hemodialysis patients in the Dialysis Outcomes and Practice Patterns Study (DOPPS). *Journal of Renal Nutrition*, 20(4), 224–234.
- Mak, R. H. (1996). Insulin resistance but IGF-I sensitivity in chronic renal failure. *American Journal of Physiology-Renal Physiology*, 271(1), F114–F119.
- Mamoun, A.-H., Anderstam, B., Södersten, P., Lindholm, B., & Bergström, J. (1996). Influence of peritoneal dialysis solutions with glucose and amino acids on ingestive behavior in rats. *Kidney International*, 49(5), 1276–1282.
- Marcen, R., Teruel, J. L., De La Cal, M. A., & Gamez, C. (1997). The impact of malnutrition in morbidity and mortality in stable haemodialysis patients. Spanish Cooperative Study of Nutrition in Hemodialysis. *Nephrology, Dialysis, Transplantation: Official Publication of the European Dialysis and Transplant Association-European Renal Association*, 12(11), 2324–2331.
- Maulidiani, Abas, F., Khatib, A., Perumal, V., Suppaiah, V., Ismail, A., ... Lajis, N. H. (2016). Metabolic alteration in obese diabetes rats upon treatment with *Centella asiatica* extract. *Journal of Ethnopharmacology*, 180, 60–69.
- Mehrotra, R., Berman, N., Alistwani, A., & Kopple, J. D. (2002). Improvement of nutritional status after initiation of maintenance hemodialysis. *American Journal of Kidney Diseases*, 40(1), 133–142.
- Mehrotra, R., & Kopple, J. D. (2001). Nutritional management of maintenance dialysis patients: why aren't we doing better? *Annual Review of Nutrition*, 21(1), 343–379.
- Moodalbil, D. G., & Hooper, S. R. (2017). Neurocognitive Functioning in Children Undergoing Dialysis. In A.R. Nissenson & R.N. Fine (Eds.), *Handbook of Dialysis Therapy* (955-964). Philadelphia: Elsevier.
- Morais, A. A. C., Silva, M. A. T., Faintuch, J., Vidigal, E. J., Costa, R. A., Lyrio, D. C., ... Pitanga, K. K. (2005). Correlation of nutritional status and food

intake in hemodialysis patients. *Clinics*, 60(3), 185–192.

- Moshage, H. J., Janssen, J. A., Franssen, J. H., Hafkenscheid, J. C., & Yap, S. H. (1987). Study of the molecular mechanism of decreased liver synthesis of albumin in inflammation. *The Journal of Clinical Investigation*, 79(6), 1635–1641.
- Moyle, G. J., Daar, E. S., Gertner, J. M., Kotler, D. P., Melchior, J.-C., O'Brien, F., ... Team, S. 9037 S. (2004). Growth hormone improves lean body mass, physical performance, and quality of life in subjects with HIV-associated weight loss or wasting on highly active antiretroviral therapy. *JAIDS Journal of Acquired Immune Deficiency Syndromes*, 35(4), 367–375.
- Mutsaers, H. A. M., Engelke, U. F. H., Wilmer, M. J. G., Wetzels, J. F. M., Wevers, R. A., van den Heuvel, L. P., ... Masereeuw, R. (2013). Optimized metabolomic approach to identify uremic solutes in plasma of stage 3–4 chronic kidney disease patients. *PLoS One*, 8(8), e71199.
- Nagana Gowda, G., & Raftery, D. (2013). Biomarker Discovery and Translation in Metabolomics. *Current Metabolomics*, 1(3), 227–240.
- Neyra, R., Chen, K. Y., Sun, M., Shyr, Y., Hakim, R. M., & Ikizler, T. A. (2003). Increased resting energy expenditure in patients with end-stage renal disease. *Journal of Parenteral and Enteral Nutrition*, 27(1), 36–42.
- Ni, Y., Su, M., Lin, J., Wang, X., Qiu, Y., Zhao, A., ... Jia, W. (2008). Metabolic profiling reveals disorder of amino acid metabolism in four brain regions from a rat model of chronic unpredictable mild stress. *FEBS Letters*, 582(17), 2627–2636.
- Nicholson, J. K., Connelly, J., Lindon, J. C., & Holmes, E. (2002). Metabonomics: a platform for studying drug toxicity and gene function. *Nature Reviews Drug Discovery*, 1(2), 153–161.
- Nicholson, J. K., Lindon, J. C., & Holmes, E. (1999). "Metabonomics": understanding the metabolic responses of living systems to pathophysiological stimuli via multivariate statistical analysis of biological NMR spectroscopic data. *Xenobiotica*, 29(11), 1181–1189.
- Nikiforova, V. J., Giesbertz, P., Wiemer, J., Bethan, B., Looser, R., Liebenberg, V., Ruiz Noppinger, P., Daniel, H., & Rein, D. (2014). Glyoxylate, a new marker metabolite of type 2 diabetes. *Journal of diabetes research*, 2014, 1–9.
- Obi, Y., Qader, H., Kovesdy, C. P., & Kalantar-Zadeh, K. (2015). Latest Consensus and Update on Protein Energy-Wasting in Chronic Kidney Disease. *Current Opinion in Clinical Nutrition and Metabolic Care*, 18(3), 254–262.
- Park, J., Mehrotra, R., Rhee, C. M., Molnar, M. Z., Lukowsky, L. R., Patel, S. S., ... Kalantar-Zadeh, K. (2013). Serum creatinine level, a surrogate of

muscle mass, predicts mortality in peritoneal dialysis patients. *Nephrology Dialysis Transplantation*, 28(8), 2146–2155.

Pearson, D. J., & Tubbs, P. K. (1967). Carnitine and derivatives in rat tissues. *Biochemical Journal*, 105(3), 953–963.

Pinto, R. C., Trygg, J., & Gottfries, J. (2012). Advantages of orthogonal inspection in chemometrics. *Journal of Chemometrics*, 26(6), 231–235.

Pupim, L. B., Cuppari, L., & Ikizler, T. A. (2006). Nutrition and Metabolism in Kidney Disease. *Seminars in Nephrology*, 26(2), 134–157.

Pupim, L. B., Kent, P., Caglar, K., Shyr, Y., Hakim, R. M., & Ikizler, T. A. (2002). Improvement in nutritional parameters after initiation of chronic hemodialysis. *American Journal of Kidney Diseases*, 40(1), 143–151.

Qi, S., Ouyang, X., Wang, L., Peng, W., Wen, J., & Dai, Y. (2012). A Pilot Metabolic Profiling Study in Serum of Patients with Chronic Kidney Disease Based on <sup>1</sup>H-NMR-Spectroscopy. *Clinical and Translational Science*, 5(5), 379–385.

Qureshi, A. R., Alvestrand, A., Divino-Filho, J. C., Gutierrez, A., Heimbürger, O., Lindholm, B., & Bergström, J. (2002). Inflammation, malnutrition, and cardiac disease as predictors of mortality in hemodialysis patients. *Journal of the American Society of Nephrology*, 13(suppl 1), S28–S36.

Reisinger, N., & Berkoben, M. (2019). CHAPTER 71 - Volume disorders and assessment. In E.V. Lerma, M.A. Sparks & J.M. Topf (Eds.), *Nephrology Secrets* (pp. 487–500). Philadelphia: Elsevier.

Rhee, E. P., Souza, A., Farrell, L., Pollak, M. R., Lewis, G. D., Steele, D. J. R., ... Gerszten, R. E. (2010). Metabolite Profiling Identifies Markers of Uremia. *Journal of the American Society of Nephrology*, 21(6), 1041–2051.

Rhee, Eugene P, Souza, A., Farrell, L., Pollak, M. R., Lewis, G. D., Steele, D. J. R., ... Gerszten, R. E. (2010). Metabolite profiling identifies markers of uremia. *Journal of the American Society of Nephrology*, 21(6), 1041–2051.

Rist, M. J., Roth, A., Frommherz, L., Weinert, C. H., Krüger, R., Merz, B., ... Watzl, B. (2017). Metabolite patterns predicting sex and age in participants of the Karlsruhe Metabolomics and Nutrition (KarMeN) study. *PLoS ONE*, 12(8), 1–21.

Rocco, M. V. (2012). Hemodialysis. In E.V. Lerma, M.A. Sparks & J.M. Topf (Eds.) *Nephrology Secrets* (pp. 353–362). Philadelphia: Elsevier

Ruiz, S., Pergola, P. E., Zager, R. A., & Vaziri, N. D. (2013). Targeting the transcription factor Nrf2 to ameliorate oxidative stress and inflammation in chronic kidney disease. *Kidney International*, 83(6), 1029–1041.



- Rutledge, C., & McMahon, L. P. (2000). Relationship between dialysis and nutritional adequacy in haemodialysis patients. *Nephrology*, 5(1–2), 27–32.
- Sadeghi-Bazargani, H., Bangdiwala, S., Mohammad, K., Maghsoudi, H., & Mohammadi, R. (2010). Compared application of the new OPLS-DA statistical model versus partial least squares regression to manage large numbers of variables in an injury case-control study. In *Scientific Research and Essays* 6(20), 4369-4377.
- Särnblad, S., Magnuson, A., Ekelund, U., & Åman, J. (2016). Body fat measurement in adolescent girls with type 1 diabetes: a comparison of skinfold equations against dual-energy X-ray absorptiometry. *Acta Paediatrica*, 105(10), 1211–1215.
- Schefold, J. C., Zeden, J.-P., Fotopoulou, C., von Haehling, S., Pschowski, R., Hasper, D., ... Reinke, P. (2009). Increased indoleamine 2, 3-dioxygenase (IDO) activity and elevated serum levels of tryptophan catabolites in patients with chronic kidney disease: a possible link between chronic inflammation and uraemic symptoms. *Nephrology Dialysis Transplantation*, 24(6), 1901–1908.
- Schutte, J. E., Longhurst, J. C., Gaffney, F. A., Bastian, B. C., & Blomqvist, C. G. (1981). Total plasma creatinine: an accurate measure of total striated muscle mass. *Journal of Applied Physiology*, 51(3), 762–766.
- Sherwin, R. S., Bastl, C., Finkelstein, F. O., Fisher, M., Black, H., Hendler, R., & Felig, P. (1976). Influence of uremia and hemodialysis on the turnover and metabolic effects of glucagon. *The Journal of Clinical Investigation*, 57(3), 722–731.
- Shetty, P. S. (1999). Adaptation to low energy intakes: the responses and limits to low intakes in infants, children and adults. *European Journal of Clinical Nutrition*, 53(s1), s14–s33.
- Shi, L., Campbell, G., Jones, W. D., Campagne, F., Wen, Z., Walker, S. J., ... Puzstai, L. (2010). The MicroArray Quality Control (MAQC)-II study of common practices for the development and validation of microarray-based predictive models. *Nature Biotechnology*, 28(8), 827-838.
- Shommu, N. S., Jenne, C. N., Blackwood, J., Joffe, A. R., Martin, D. A., Thompson, G. C., & Vogel, H. J. (2018). Metabolomic and inflammatory mediator based biomarker profiling as a potential novel method to aid pediatric appendicitis identification. *PLoS ONE*, 13(3), 1–13.
- Singh, A. K. (2010). Chronic Kidney Disease. In S.B. Mushlin & H.L. Greene (Eds.), *Decision Making in Medicine* (pp. 352–353). Philadelphia: Mosby Elsevier.
- Sloan, G., Ali, A., & Webster, J. (2017). A rare cause of metabolic acidosis:

ketoacidosis in a non-diabetic lactating woman. *Endocrinology, Diabetes & Metabolism Case Reports*, 2017(1), 17-0073.

Snoswell, A. M., & Koundakjian, P. P. (1972). Relationships between carnitine and coenzyme A esters in tissues of normal and alloxan-diabetic sheep. *Biochemical Journal*, 127(1), 133–141.

Song, S. (2000). Can the glyoxylate pathway contribute to fat-induced hepatic insulin resistance? *Medical Hypotheses*, 54(5), 739–747.

Song, X., Wang, J., Wang, P., Tian, N., Yang, M., & Kong, L. (2013). 1H NMR-based metabolomics approach to evaluate the effect of Xue-Fu-Zhu-Yu decoction on hyperlipidemia rats induced by high-fat diet. *Journal of Pharmaceutical and Biomedical Analysis*, 78, 202–210.

Song, Y.-H., Li, Y., Du, J., Mitch, W. E., Rosenthal, N., & Delafontaine, P. (2005). Muscle-specific expression of IGF-1 blocks angiotensin II-induced skeletal muscle wasting. *The Journal of Clinical Investigation*, 115(2), 451–458.

Srivastava, N., Kumar, A., Singh, S., Mishra, C. P., Mishra, R., & Singh, R. G. (2012). Protein energy wasting in chronic kidney disease patients: a hospital based study. 43(4), 389–395.

Stegmayr, B. (2017). Dialysis Procedures Alter Metabolic Conditions. *Nutrients*, 9(6), 548.

Stenvinkel, P., Barany, P., Chung, S. H., Lindholm, B., & Heimbürger, O. (2002). A comparative analysis of nutritional parameters as predictors of outcome in male and female ESRD patients. *Nephrology Dialysis Transplantation*, 17(7), 1266–1274.

Stevens, J. (2003). Body mass index and mortality in Asian populations: implications for obesity cut-points. *Nutrition Reviews*, 61(3), 104-107.

Stevens, L. A., Schmid, C. H., Greene, T., Zhang, Y. (Lucy), Beck, G. J., Froissart, M., ... Levey, A. S. (2010). Comparative Performance of the CKD Epidemiology Collaboration (CKD-EPI) and the Modification of Diet in Renal Disease (MDRD) Study Equations for Estimating GFR Levels Above 60 mL/min/1.73 m<sup>2</sup>. *American Journal of Kidney Diseases*, 56(3), 486–495.

Struijk, D. G., Krediet, R. T., Koomen, G. C., Boeschoten, E. W., & Arisz, L. (1994). The effect of serum albumin at the start of continuous ambulatory peritoneal dialysis treatment on patient survival. *Peritoneal Dialysis International*, 14(2), 121–126.

Sung, C.-C., Hsu, Y.-C., Chen, C.-C., Lin, Y.-F., & Wu, C.-C. (2013). Oxidative stress and nucleic acid oxidation in patients with chronic kidney disease. *Oxidative Medicine and Cellular Longevity*, 2013, 1-15.

- Suri, R. S., & Klinger, A. S. (2010). Frequent Hemodialysis: Physiological, Epidemiological, and Practical Aspects. In J. Himmelfarb & M.H. Sayegh (Eds), *Chronic Kidney Disease, Dialysis, and Transplantation* (pp. 370–384). Philadelphia: Saunders.
- Tam, Z. Y., Ng, S. P., Tan, L. Q., Lin, C.-H., Rothenbacher, D., Klenk, J., ... Group, A. S. (2017). Metabolite profiling in identifying metabolic biomarkers in older people with late-onset type 2 diabetes mellitus. *Scientific Reports*, 7(1), 4392.
- Teerlink, T., Nijveldt, R. J., de Jong, S., & van Leeuwen, P. A. M. (2002). Determination of Arginine, Asymmetric Dimethylarginine, and Symmetric Dimethylarginine in Human Plasma and Other Biological Samples by High-Performance Liquid Chromatography. *Analytical Biochemistry*, 303(2), 131–137.
- Thongprayoon, C., Cheungpasitporn, W., & Kashani, K. (2016). Serum creatinine level, a surrogate of muscle mass, predicts mortality in critically ill patients. *Journal of Thoracic Disease*, 8(5), E305–E311.
- Triba, M. N., Le Moyec, L., Amathieu, R., Goossens, C., Bouchemal, N., Nahon, P., ... Savarin, P. (2015). PLS/OPLS models in metabolomics: The impact of permutation of dataset rows on the K-fold cross-validation quality parameters. *Molecular BioSystems*, 11(1), 13–19.
- Trygg, J., & Wold, S. (2002). Orthogonal projections to latent structures (O-PLS). *Journal of Chemometrics: A Journal of the Chemometrics Society*, 16(3), 119–128.
- Tulchinsky, T. H., & Varavikova, E. A. (2014). Non-Communicable Diseases and Conditions. In T.H. Tulchinsky & E.A. Varavikova (Eds.), *The New Public Health* (pp. 237-309). San Diego: Academic Press.
- United States Renal Data System. (2018). *2018 USRDS annual data report: Epidemiology of kidney disease in the United States* (Vol. 2). Bethesda, MD.
- Unruh, M. L., & Sanders, M. H. (2011). Sleep in Chronic Kidney Disease. *Principles and Practice of Sleep Medicine*, 1462–1472.
- Urbanczyk-Wochniak, E., Luedemann, A., Kopka, J., Selbig, J., Roessner-Tunali, U., Willmitzer, L., & Fernie, A. R. (2003). Parallel analysis of transcript and metabolic profiles: a new approach in systems biology. *EMBO Reports*, 4(10), 989–993.
- Utaka, S., Avesani, C. M., Draibe, S. A., Kamimura, M. A., Andreoni, S., & Cuppari, L. (2005). Inflammation is associated with increased energy expenditure in patients with chronic kidney disease. *The American Journal of Clinical Nutrition*, 82(4), 801–805.
- Van Tonder, E., Mace, L., Steenkamp, L., Tydeman-Edwards, R., Gerber, K., &

- Friskin, D. (2018). Mid-upper arm circumference (MUAC) as a feasible tool in detecting adult malnutrition. *South African Journal of Clinical Nutrition*, 1–6.
- van Valkengoed, I. G. M., Argmann, C., Ghauharali-van der Vlugt, K., Aerts, J. M. F. G., Brewster, L. M., Peters, R. J. G., ... Houtkooper, R. H. (2017). Ethnic differences in metabolite signatures and type 2 diabetes: a nested case-control analysis among people of South Asian, African and European origin. *Nutrition & Diabetes*, 7(12), 300.
- Vanholder, R., Van Laecke, S., & Glorieux, G. (2008). What is new in uremic toxicity? *Pediatric nephrology*, 23(8), 1211-1221.
- Vegine, P. M., Fernandes, A. C. P., Torres, M. R. S. G., Silva, M. I. B., & Avesani, C. M. (2011). Assessment of methods to identify protein-energy wasting in patients on hemodialysis. *Brazilian Journal of Nephrology*, 33(1), 55–61.
- Von Korff, R. W. (1967). Substrate Transformations dependent on Respiratory States of Mitochondria: Changes in Metabolic Control Sites of Rabbit Heart Mitochondria. *Nature*, 214, 20–23.
- Wang, A. Y.-M., Sea, M. M.-M., Tang, N., Sanderson, J. E., Lui, S.-F., Li, P. K.-T., & Woo, J. (2004). Resting energy expenditure and subsequent mortality risk in peritoneal dialysis patients. *Journal of the American Society of Nephrology*, 15(12), 3134–3143.
- Wang, H., Liu, A., Zhao, W., Zhao, H., Gong, L., Chen, E., ... Jiang, H. (2018). Metabolomics Research Reveals the Mechanism of Action of Astragalus Polysaccharide in Rats with Digestive System Disorders. *Molecules*, 23(12), 3333.
- Wang, X., Sun, H., & Zhang, A. (2015). Metabolic Profiling and Biomarkers Analysis of XinQiXu Syndrome. In X.Wang, A. Zhang & H. Sun (Eds.), *Chinmedomics: The Integration of Serum Pharmacochimistry and Metabolomics to Elucidate the Scientific Value of Traditional Chinese Medicine*. (pp. 233-242). San Diego, CA: Academic Press.
- Wehrens, R., & Engelke, U. (2013). Chemometric Methods in Nuclear Magnetic Resonance–Based Body Fluid Analysis. In N. Lutz, J. Sweedler, & R. Wevers (Eds.), *Methodologies for Metabolomics: Experimental Strategies and Techniques* (pp. 244-256). Cambridge: Cambridge University Press.
- Westerhuis, J. A., Hoefsloot, H. C. J., Smit, S., Vis, D. J., Smilde, A. K., van Velzen, E. J. J., ... van Dorsten, F. A. (2008). Assessment of PLSDA cross validation. *Metabolomics*, 4(1), 81–89.
- Westerhuis, J. A., van Velzen, E. J. J., Hoefsloot, H. C. J., & Smilde, A. K. (2010). Multivariate paired data analysis: Multilevel PLSDA versus OPLSDA. *Metabolomics*, 6(1), 119–128.

- Wettersten, H. I., Ganti, S., & Weiss, R. H. (2014). Metabolomic Profiling of Tumor-Bearing Mice. *Methods in Enzymology*, 543, 275–296.
- Wheelock, Å. M., & Wheelock, C. E. (2013). Trials and tribulations of 'omics data analysis: Assessing quality of SIMCA-based multivariate models using examples from pulmonary medicine. *Molecular BioSystems*, 9(11), 2589–2596.
- Wiklund, S. (2008). *Multivariate data analysis for Omics*. Umea: Umetrics.
- Wolfson, M., Jones, M. R., & Kopple, J. D. (1982). Amino acid losses during hemodialysis with infusion of amino acids and glucose. *Kidney International*, 21(3), 500–506.
- Wong, H. S., & Goh, B. L. (2018). *Twenty Forth Report of the Malaysian Dialysis and Transplant 2016*. Kuala Lumpur: The Natinal Renal Registry.
- Wong, J. S., Port, F. K., Hulbert-Shearon, T. E., Carroll, C. E., Wolfe, R. A., Agodoa, L. Y. C., & Daugirdas, J. T. (1999). Survival advantage in Asian American end-stage renal disease patients. *Kidney International*, 55(6), 2515–2523.
- Wright, M., Woodrow, G., O'Brien, S., King, N., Dye, L., Blundell, J., ... Turney, J. (2003). Disturbed appetite patterns and nutrient intake in peritoneal dialysis patients. *Peritoneal Dialysis International*, 23(6), 550–556.
- Yeun, J. Y., & Depner, T. A. (2010). Principles of Hemodialysis. In J. Himmelfarb & M.H. Sayegh (Eds.), *Chronic Kidney Disease, Dialysis, and Transplantation* (pp. 277–302). Philadelphia: Saunders.
- Yokote, S., & Yokoo, T. (2017). Embryonic Organoid Transplantation. In G. Orlando, G. Remuzzi, D.F. Williams (Eds.), *Kidney Transplantation, Bioengineering, and Regeneration: Kidney Transplantation in the Regenerative Medicine Era* (pp. 1163–1166). Cambridge: Academic Press.
- Yu, Z., Kastenmüller, G., He, Y., Belcredi, P., Möller, G., Prehn, C., ... Wang-Sattler, R. (2011). Differences between Human Plasma and Serum Metabolite Profiles. *PLOS ONE*, 6(7), e21230.
- Yu, Z., Zhai, G., Singmann, P., He, Y., Xu, T., Prehn, C., ... Wang-Sattler, R. (2012). Human serum metabolic profiles are age dependent. *Aging Cell*, 11(6), 960–967.
- Zhang, A., Yan, G., Han, Y., & Wang, X. (2014). Metabolomics approaches and applications in prostate cancer research. *Applied Biochemistry and Biotechnology*, 174(1), 6–12.
- Zhang, L., Du, J., Hu, Z., Han, G., Delafontaine, P., Garcia, G., & Mitch, W. E. (2009). IL-6 and serum amyloid A synergy mediates angiotensin II-induced muscle wasting. *Journal of the American Society of Nephrology*,

20(3), 604–612.

Zhao, Y. (2013). Metabolomics in chronic kidney disease. *Clinica Chimica Acta*, 422, 59–69.

Zimmermann, J., Herrlinger, S., Pruy, A., Metzger, T., & Wanner, C. (1999). Inflammation enhances cardiovascular risk and mortality in hemodialysis patients. *Kidney International*, 55(2), 648–658.



© COPYRIGHT UPM