

UNIVERSITI PUTRA MALAYSIA

EXPRESSION OF SURVIVAL AND APOPTOTIC MOLECULES IN RESPONSE TO EARLY STAGE CHEMOTHERAPY IN ACUTE MYELOID LEUKAEMIA

STEPHNIE YIAU KANG XIAN

FPSK(m) 2014 58



EXPRESSION OF SURVIVAL AND APOPTOTIC MOLECULES IN RESPONSE TO EARLY STAGE CHEMOTHERAPY IN ACUTE MYELOID LEUKAEMIA



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

December 2013

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

To my beloved family and cherished friends, Your support and encouragements were the pillars of my strength to finish this journey



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

EXPRESSION OF SURVIVAL AND APOPTOTIC MOLECULES IN RESPONSE TO EARLY STAGE CHEMOTHERAPY IN ACUTE MYELOID LEUKAEMIA

By

STEPHNIE YIAU KANG XIAN

December 2013

Chairman: Maha Abdullah, PhDFaculty: Medicine and Health Sciences

Acute myeloid leukaemia (AML) is a haematological malignancy characterised by a predominance of myeloid precursor cells that leads to death if not treated. A major factor in the failure of AML chemotherapy is due to the acquisition of multidrug resistance (MDR) by the malignant myeloblast. Factors such as cytokines, activation of signalling pathway mediators and Bcl-2 family proteins contributes to cell survival, thus leading to MDR. Bone marrow aspirate was collected a month after induction therapy to determine complete remission (CR). It is postulated that the treatment outcome could be determined during early induction therapy through the expression pattern of cellular molecules in peripheral blood. The expression patterns of stem cell marker, CD34, c-Kit receptor (CD117), cytokines and their receptors, signalling mediators of the PI3K/Akt and MAP kinase pathway, and Bcl-2 family proteins were observed in the peripheral blood of AML patients collected before and/or during Day 3 of induction therapy with anthracycline and cytarabine arabinoside (Ara-C). Expression were measured using flow cytometry for the percentage of cells expressing and geometric mean fluorescent intensity (MFI), as well asRT-PCR. Results showed that the percentage of cells expressing IL-1 β (p=0.028), the MFI of IL-18R α (p=0.01), MFI (p=0.007) and mRNA levels (p=0.038) of TNF- α , MFI of DR5 (p=0.02) and MFI of pAkt-T308 (p=0.038) was found to be significantly higher in samples of sensitive patients prior to induction therapy. Untreated resistant samples were found to have significantly higher MFI for pFKHR (p=0.05). During induction therapy, we found that in sensitive samples, IL-18Rα was found to be significantly higher in the percentage of expressing cells (p=0.014) and the MFI (p=0.02)was higher as well. These treated sensitive samples were also found to have significantly higher pp38 MFI (p=0.039). Treated resistant samples were found to have significantly higher percentage of CD34 (p=0.028) and pBAD (p=0.014) expressing cells. Spearman Rank-order correlation analysis showed that there is no correlation between the MFI and the mRNA transcript of cytokine and its receptors. Similar correlations analysis

were also observed between cytokines and receptors with signalling mediators and apoptotic molecules, pBAD and Bim. Some of these correlations corresponded to the cytokines role in apoptosis and survival but there are some contradictions. Molecules such as DR5, CD34, pFKHR and pBAD are potential prognostic markers of treatment outcome but some might not be suitable markers of treatment outcome at the early stages of induction therapy.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

EXPRESI MOLEKUL KEMANDIRAN DAN APOPTOTIK SEBAGAI TINDAK BALAS TERHADAP KEMOTERAPI PERINGKAT AWAL LEUKEMIA MIELOID AKUT

Oleh

STEPHNIE YIAU KANG XIAN

Disember 2013

Pengerusi : Maha Abdullah, PhD Fakulti : Perubatan dan Sains Kesihatan

Leukemia mieloid akut (AML) adalah kanser hematologi yang dicirikan oleh penguasaan sel-sel pelopor mieloid yang membawa maut jika tidak dirawat. Sebab utama dalam kegagalan kemoterapi AML adalah kekebalan sel myeloblast malignan terhadap beraneka jenis ubat, suatu fenomena yang dikenal sebagai MDR. Faktorfaktor seperti sitokina, pengaktifan pengantara laluan transduksi isyarat dan kumpulan protein Bcl-2 menyumbang kepada kemandirian sel dan seterusnya MDR. Sampel kandungan sumsum tulang diambil sebulan selepas terapi induksi untuk menentukan status sembuh sempurna (CR). Diandaikan bahawa keputusan akhir hasil daripada rawatan dapat ditentukan semasa terapi induksi awal melalui tindak balas biologi antara molekul-molekul sel dalam darah. Corak ekspresi penanda sel induk, CD34, reseptor c-Kit (CD117), sitokina dan reseptor-reseptornya, pengantara isyarat laluan PI3K/Akt dan MAP kinase, dan kumpulan protein Bcl-2 dikaji daripada darah pesakit AML yang dikumpul sebelum dan/atau semasa hari ketiga terapi induksi dijalankan dengan antrasiklin dan sitosina arabinosida (Ara-C) dengan menggunakan kaedah sitometri aliran dan RT-PCR. Keputusan mendedahkan bahawa sebelum terapi induksi, peratusan sel yang menunjukkan IL-1β, MFI IL-18Rα, MFI dan mRNA TNFa, DR5 dan MFI pAkt-T308 adalah lebih tinggi dalam sampel sensitif. Didapati sampel rintang sebelum rawatan menunjukkan MFI pFKHR yang jauh lebih tinggi. Semasa terapi induksi, sampel sensitif yang mempunyai IL-18Ra memaparkan peratusan bilangan sel dan MFI yang lebih tinggi. Sampel sensitif yang dirawati ini juga menunjukkan pp38 MFI yang lebih tinggi. Sampel rintang yang dirawati didapati mempunyai jauh lebih tinggi peratusan sel-sel yang expresi CD34 dan pBAD. Penelitian menunjukkan bahawa hubungan antara sitokina dan reseptor tidak berkaitan dengan transkrip mRNA sitokina. Hubungan antara sitokina dan reseptor dengan pengantara isyarat dan molekul apoptotik, pBAD dan Bim, juga ditinjau. Sesetengah analisis hubungan tersebut sepadan dengan peranan sitokina dalam apoptosis dan kemandirian tetapi terdapat beberapa keputusan yang bercanggahan. Didapati bahawa



tahap rendah DR5 dan tahap tinggi pFKHR sebelum terapi induksi berpotensi menunjukkan rintangan-kimo manakala tahap tinggi CD34 dan pBAD semasa induksi menunjukkan rintangan-kimo. Oleh itu, molekul seperti DR5, CD34, pFKHR dan pBAD adalah potensi ramalan penanda untuk hasil rawatan manakala sesetengah mungkin tidak sesuai penanda hasil rawatan pada awal terapi induksi.



ACKNOWLEDGEMENTS

First and foremost, I would like to express my deepest gratitude to my supervisor, Prof Dr. Maha Abdullah, Dato' Dr. Chang Kian Meng and Dr. Eusni Rahayu Mohd. Tohit for their unfailing support, guidance and patience throughout the whole project. I would like to especially thank Dr. Maha for her encouragements, tolerance and guidance for the past 4 years.

I would like to express my appreciation and love towards my family. They have unwaveringly and unquestioningly supported my pursuit for knowledge in this field.

I am extremely grateful to all the nurses in Hospital Ampang, especially Sr. Normah, Sr, Rodiyah, Sr. Masitoh, Sr. Fazilah and Sr. Halimah for their support and help in the sample collections from both Haematological Wards. I would also like to express my appreciation to the staff of Immunology Lab, Marsitah, Kak Aishah, Kak Ezura and Uncle Anthony for their hardwork in keeping the lab in tip top working condition which aids the efficiency of the work.

To my collegues, Fara, Zeha, James, LJ, Ram and Fatemeh for their support and help throughout the project, be it in support, advice or technical help, they were there and ready to extend their hands.

Finally, to Shins, Zul, Shi Wei, Tong, Catherine and Cindee, thanks very much for all the loving patience and support you all have extended to me. All of you have looked out for me and stood by me regardless of the situation. Your presence in my life have made my Masters degree a memory that will be cherished eternally.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Maha Abdullah, PhD

Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Chairman)

Eusni Rahayu Mohd Tohit, MBBS (Malaya), MPath (UKM, Mal)

Senior Medical Lecturer Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

Dato' Dr. Chang Kian Meng, MBBS (Malaya), MRCP (UK), FRCP (London), FRCPA (Haem)

Consultant Hematologist and Head of Department Hospital Ampang (Member)

BUJANG BIN KIM HUAT, PhD Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 03 April 2014

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: Name of Chairman of Supervisory Committee:	Professor Dr. Maha Abdullah
Signature: Name of Member of Supervisory Committee:	Dr. Eusni Rahayu Mohd Tohit
Signature: Name of Member of Supervisory Committee:	Dato' Dr. Chang Kian Meng,

TABLE OF CONTENTS

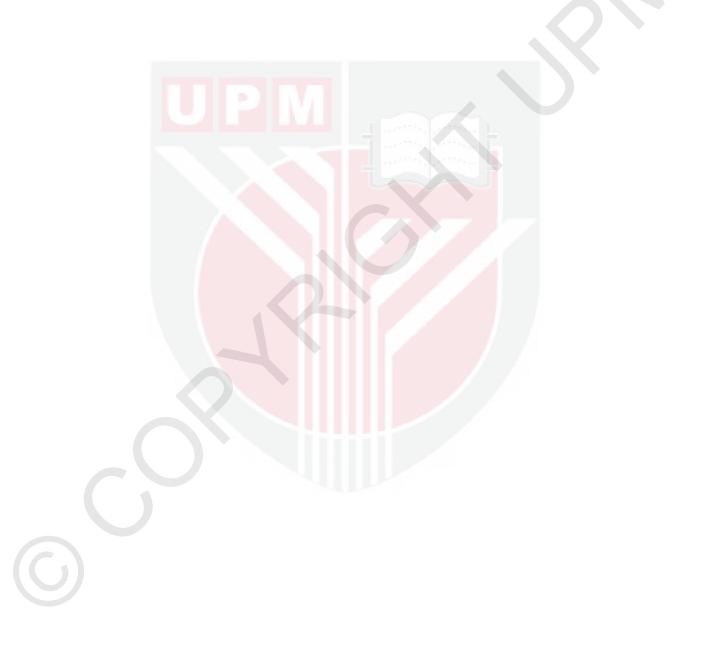
			1	Page
AB AC AP DE LIS LIS	PROVA CLARA ST OF 1 ST OF F	LED L ATIO ABL TGUI	ES	i iii v vi viii xiii xviii xvii xix
СН	IAPTER	2		
	1 I	NTR	ODUCTION	1
			RATURE REVIEW	4
	2	.1	Acute Myeloid Leukaemia (AML)	4
			2.1.1 Diagnosis	4
			2.1.2 Classifications	5 5
			2.1.2.1 FAB Classification	5
			2.1.2.2 WHO Classification	6
			2.1.3 Treatment and Treatment Outcome	7
	2	.2	Chemotherapeutic Drugs	8
			2.2.1 Daunorubicin and Idarubicin	8
			2.2.2 Cytarabine Arabinoside (Ara-C)	9
			2.2.3 Mitoxantrone	9
		.3	Drug Resistance	10
	2	.4	Signalling Pathways	10
			2.4.1 PI3K/Akt Pathways	11
			2.4.1.1 Akt	12
			2.4.1.2 Forkhead Transcript Factors	12
			2.4.2 MAP Kinase Pathways	13
			2.4.2.1 MAPK/ERK	13
			2.4.2.2 p38 MAPK	14
			2.4.2.3 c-Jun N-terminal Kinase (JNK)	15
		_	2.4.2.4 Cross-talk between JNK and p38 Pathways	
	2	.5	Cytokines	16
			2.5.1 Role of Cytokine in Cytokine Regulation	17
			2.5.2 Role of Cytokines In Regulating Signalling Pathways	17
		-	2.5.3 Dual Roles of Cytokines –IL-6 and TNF- α	18
	2	.6	Cell Death Related Proteins	19
			2.6.1 Bcl-2 Family Proteins	19
			2.6.1.1 BAD	19
			2.6.1.2 Bim	20
			2.6.2 TRAIL and TRAIL Receptors	21
	2	.7	Stem Cell Markers	22

Stem Cell Markers 2.7

	3	МАТ	ERIALS AND METHODS	24
	3	3.1		24 24
		5.1	Sample Collection	24 24
			3.1.1 Study Population3.1.2 Experimental Design	24 24
		3.2		24 25
		3.3	Isolation of Peripheral Blood Mononuclear Cells	23 26
		5.5	Flow Cytometry	
			3.3.1 Antibodies and Reagents	26 30
			3.3.2 Protein Transport Inhibition	30 30
			3.3.3 Surface Staining3.3.4 Intracellular Staining	30 30
			3.3.4.1 Intracellular Cytokines Staining	30 30
				30
				31
		3.4	3.3.5 Data Acquisition and Analysis RNA Isolation	31
		3.5	DNase Treatment	31
		3.6	Quantitation and Quality Assessment of RNA	32
		3.7	Reverse Transcription	32
		3.8	PCR	33
		5.0	3.8.1 Primer Pairs	33
			3.8.2 Multiplex PCR	33
		3.9	Gel Electrophoresis	33 34
		3.10	Data Analysis	34
		5.10		51
	4	RESU	JLTS	35
		4.1	Clinical Samples	35
		4.2	Treatment Outcome	38
		4.3	Immunophenotyping	38
		4.4	Cytokines and Receptors	41
			4.4.1 Interleukin1 β (IL-1 β)	42
			4.4.2 Interleukin-18 Receptor-α (IL-18Rα) and Interleukin-	
			18 (IL-18)	43
			4.4.3 Interleukin-6 (IL-6)	43
			4.4.4 Tumour Necrosis Factor α (TNF- α)	43
			4.4.5 TRAIL-R2 (DR5)	44
		4.5	Signal Transduction Pathways	44
			4.5.1 PI3K/Akt Pathway	45
			4.5.2 MAP Kinase Pathway	46
		4.6	Phosphorylated BAD (pBAD) and Bim	46
		4.7	Correlations Analysis	48
	5	DISC	USSION	54
	5	5.1	Clinical Samples	54 54
		5.2	Immunophenotyping	55
		5.3	Correlation Analyses	60
		5.4	Limitations and Improvements	65
		5.1		05
	6	CON	CLUSION	66

xi

REFERENCES	68
APPENDICES	85
BIODATA OF STUDENT	140
PUBLICATION	141



LIST OF TABLES

Table		Page
2.1	FAB classifications of AML and its association with various cytogenetics abnormalities, with modification from	6
2.2	WHO classification of AML with description	7
3.1	List of surface and intracellular antigens studied	27
3.2	Four-colour to six-colour antibody combination within tube for flow cytometry analysis	28
3.3	Alternative antibody combination: Four to six-colour antibody combination within tube for flow cytometry analysis	29
3.4	Primer sequences and expected amplicon size of cDNA	33
3.5	Thermocycling conditions for multiplex PCR	34
4.1	Clinical characteristics of AML patients in Hospital Ampang (N=16)	37
4.2	Significant correlations between the percentage of expressing cells and MFI of CD117 and CD34 and signalling mediators, cytokines and Bcl-2 family proteins	48
4.3	Significant correlations between the percentage of expressing cells of cytokines and cytokine receptors and CD34, CD117, signalling mediators and Bcl-2 family proteins	49
4.4	Significant correlations between MFI of cytokines and cytokine receptors and CD34, CD117, signalling mediators and Bcl-2 family proteins	49
4.5	Significant correlations between cytokines and cytokine receptors mRNA expression and CD34, CD117, signalling mediators and Bcl-2 family proteins	51
4.6	Significant correlations between the percentages of expressing cells and mean fluorescence intensity of MAP Kinase signalling mediators and CD34, CD117, cytokines and Bcl-2 family proteins	51
4.7	Significant correlations between the percentages of expressing cells and MFI of PI3K/Akt signalling mediators and CD34, CD117, cytokines and Bcl-2 family proteins	52
4.8	Significant correlations between the percentages of expressing cells of Bcl-2 family proteins and CD34, CD117, signalling mediators and cytokines	53

A 1	Clinical data, subclassification and treatment outcome of AML patients from Hospital Ampang	86
A 2	Immunophenotyping of treated and untreated AML samples	87
A 3	Protein analysis of cytokines and cytokine receptors percentage of cells and MFI) in untreated AML samples	88
A 4	Protein analysis of cytokines and cytokine receptors (percentage of cells and MFI) in treated AML samples	89
A 5	mRNA expression of housekeeping gene, B2M and cytokines and receptors of treated and untreated AML samples	90
A 6	Protein analysis survival pathway signalling mediators (percentage of cells and MFI) in untreated AML samples	91
A 7	Protein analysis survival pathway signalling mediators and Bcl-2 proteins (percentage of cells and MFI) in treated AML samples	92
A 8	Untreated Sample: Mann-Whitney T-test of comparing mean of cytokines and receptors (percentage of cells and MFI) of sensitive and resistant samples	93
A 9	Treated Sample: Mann-Whitney T-test of comparing mean of cytokines and receptors (percentage of cells and MFI) of sensitive and resistant samples	93
A 10	Mann-Whitney T-test of comparing mean of cytokines and receptors mRNA expression of sensitive and resistant samples (Untreated and treated)	94
A 11	Untreated Sample: Mann-Whitney T-test of comparing mean of signalling mediators (percentage of cells and MFI) of sensitive and resistant samples	94
A 12	Treated Sample: Mann-Whitney T-test of comparing mean of signalling mediators (percentage of cells and MFI) of sensitive and resistant samples	95
A 13	Untreated Sample: Mann-Whitney T-test of comparing mean of CD34, CD117 and Bcl-2 family proteins (percentage of cells and MFI) of sensitive and resistant samples	95
A 14	Treated Sample: Mann-Whitney T-test of comparing mean of CD34, CD117 and Bcl-2 family proteins (percentage of cells and MFI) of sensitive and resistant samples	96

	A 15	Untreated samples: Correlation of percentage of cells expressing CD117 and CD34 with signalling mediators, cytokines and Bcl-2 family proteins	96
	A 16	Treated samples: Correlation of percentage of cells expressing CD117 and CD34 with signalling mediators, cytokines and Bcl-2 family proteins	97
	A 17	Untreated samples: Correlation of MFI of CD117 and CD34 with signalling mediators, cytokines and Bcl-2 family proteins	97
	A 18	Treated samples: Correlation of MFI of CD117 and CD34 with signalling mediators, cytokines and Bcl-2 family proteins	98
	A 19	Untreated and Treated samples: Correlation of MFI of CD117 and CD34 with cytokine mRNA	98
	A 20	Untreated samples: Correlation of percentage of cells expressing cytokines and cytokine receptors with CD34, CD117, signalling mediators and Bcl-2 family proteins	99
A 22	A 21	Treated samples: Correlation of percentage of cells expressing cytokines and cytokine receptors with CD34, CD117, signalling mediators and Bcl-2 family proteins	100
	A 22	Untreated samples: Correlation of MFI of cytokines and cytokine receptors with CD34, CD117, signalling mediators and Bcl-2 family proteins	101
	A 23	Treated samples: Correlation of MFI of cytokines and cytokine receptors with CD34, CD117, signalling mediators and Bcl-2 family proteins	102
	A 24	Untreated and treated samples: Correlation of MFI of cytokines and cytokine receptors with cytokine mRNA	103
	A 25	Untreated and Treated samples: Correlation of cytokine mRNA	104
	A 26	Untreated samples: Correlation of percentage of cells expressing mediators of MAP Kinase and PI3K/Akt signalling mediators with CD34, CD117, cytokines and Bcl-2 family proteins	105
	A 27	Treated samples: Correlation of percentage of cells expressing mediators of MAP Kinase and PI3K/Akt signalling mediators with CD34, CD117, cytokines and Bcl-2 family proteins	106
	A 28	Untreated samples: Correlation MFI of mediators of MAP Kinase and PI3K/Akt signalling mediators with CD34, CD117, cytokines and Bcl-2 family proteins	107

A 29	Treated samples: Correlation MFI of mediators of MAP Kinase and PI3K/Akt signalling mediators with CD34, CD117, cytokines and Bcl-2 family proteins	108
A 30	Untreated and Treated samples: Correlation MFI of mediators of MAP Kinase and PI3K/Akt signalling mediators with cytokine mRNA	109
A 31	Untreated samples: Correlation of percentage of cells expressing Bcl- 2 family proteins with CD34, CD117, signalling mediators and cytokines	110
A 32	Treated samples: Correlation of percentage of cells expressing Bcl-2 family proteins with CD34, CD117, signalling mediators and cytokines	110
A 33	Untreated samples: Correlation of MFI of Bcl-2 family proteins with CD34, CD117, signalling mediators and cytokines	111
A 34	Treated samples: Correlation of MFI of Bcl-2 family proteins with CD34, CD117, signalling mediators and cytokines	111
A 35	Untreated and Treated samples: Correlation MFI of mediators of Bcl- 2 family proteins with cytokine mRNA	112

C

LIST OF FIGURES

	Figur	Figure			
	2.1	Summary of nucleus and mitochondria targeted signalling of JNK	15		
	3.1	Blood separation layers in peripheral blood mononuclear cells extraction before and after Ficoll-paque PLUS centrifugation method	25		
	4.1	Distribution of patient samples collected from Hospital Ampang, Selangor (N=60)	36		
4.2		Flow cytometry staining of AML samples upon diagnosis (left) and during Day 3 of induction therapy (right)	39		
	4.3	Percentage of cell expressing c-Kit receptor, CD117 and myeloid haemapoietic stem cell marker, CD34 expression in AML samples	40		
	4.4	Protein expression of cytokines/cytokine receptor and DR5	41		
	4.5	Agarose gel images of multiplex mRNA results in AML samples	42		
4.6 4.7 4.8 5.1		The relative mRNA expression of cytokines and DR5	42		
		Protein phosphorylation of signalling mediators of the PI3K/Akt pathway and MAP kinase	45		
		Phosphorylation of pBAD and the expression of whole Bim in AML samples	47		
		The relationship between molecules and cytokines before chemotherapy in both sensitive and resistant AML samples	63		
5.2	5.2	The relationship between signalling molecules and cytokines on Day 3 of chemotherapy in sensitive AML samples	64		
	5.3	The relationship between signalling molecules and cytokines on Day 3 of chemotherapy in resistant AML samples	65		
	A 1	Untreated Resistant (Samples No. 32): Flow cytometry analysis plots	113		
	A 2	Treated Resistant (Samples No. 32): Flow cytometry analysis plots	114		
	A 3	Untreated Sensitive (Samples No. 33): Flow cytometry analysis plots	115		
	A 4	Treated sensitive (Samples No. 33): Flow cytometry analysis plots	116		
	A 5	Untreated resistant (Samples No. 35): Flow cytometry analysis plots	117		

A	6	Treated resistant (Samples No. 35): Flow cytometry analysis plots	118
A	7	Untreated resistant (Samples No. 51): Flow cytometry analysis plots	119
А	8	Treated resistant (Samples No. 51): Flow cytometry analysis plots	120
A	9	Untreated resistant (Samples No. 54): Flow cytometry analysis plots	121
А	10	Treated resistant (Samples No. 54): Flow cytometry analysis plots	122
А	11	Untreated resistant (Samples No. 56): Flow cytometry analysis plots	123
А	12	Treated resistant (Samples No. 56): Flow cytometry analysis plots	124
А	13	Untreated resistant (Samples No. 60): Flow cytometry analysis plots	125
А	14	Treated resistant (Samples No. 60): Flow cytometry analysis plots	126
А	15	Untreated resistant (Samples No. 61): Flow cytometry analysis plots	127
А	16	Treated resistant (Samples No. 61): Flow cytometry analysis plots	128
А	17	Untreated sensitive (Samples No. 63): Flow cytometry analysis plots	129
А	18	Untreated sensitive (Samples No. 64): Flow cytometry analysis plots	130
А	19	Untreated sensitive (Samples No. 65): Flow cytometry analysis plots	131
A	20	Treated sensitive (Samples No. 65): Flow cytometry analysis plots	132
A	21	Untreated sensitive (Samples No. 70): Flow cytometry analysis plots	133
A	22	Treated sensitive (Samples No. 70): Flow cytometry analysis plots	134
A	23	Untreated sensitive (Samples No. 71): Plots of flow cytometry analysis	135
A	24	Treated sensitive (Samples No. 71): Plots of flow cytometry analysis	136
A	25	Treated sensitive (Samples No. 73): Plots of flow cytometry analysis	137
	26	Treated Resistant (Samples No.74: Plots of flow cytometry analysis	138
A	27	Agarose gel images of multiplex MRNA results in AML samples	139

LIST OF ABBREVIATIONS

	AML	acute myeloid leukaemia
	Ara-C	cytarabine arabinoside
	Ara-CTP	cytarabine arabinoside triphosphate
	Ara-CMP	cytarabine arabinoside monophosphate
	APML	acute promyeloid leukaemia
	B2M	beta-2-microglobulin
	BAD	Bcl-2-associated death promoter
	BCP	2-Bromo-Phenol-Chloroform
	BD	Becton and Dickinson
	BMA	bone marrow aspirate
	CALGB	Cancer and Leukaemia Group B
	CO ₂	carbon dioxide
	CR	complete remission
	dCyc	deoxycytidine kinase
	DFS	disease-free survival
	DNA	deoxyribonucleic acid
	DNase	deoxyribonuclease
	dNTP	deoxyribonucleotide triphosphates
	DNR	Daunorubicin
	EDTA	Ethylenediaminetetraacetic acid
	FAB	French-American-British
	FBS	foetal bovine serum
	FITC	Fluorescein isothiocyanate
	FKHR	forkhead in rhabdomyosarcoma
	HIDAC	high dose intermittent Ara-C

IDA	Idarubicin
IL	interleukin
JNK	c-Jun N-terminal kinase
MIDAC	Mitoxontrone and arabinoside cytarabine
MAP	mitogen activated kinase
MFI	mean fluorescence intensity
min(s)	minute(s)
МКК	MAP kinase kinase
МККК	MAP kinase kinase
MMLV	murine leukaemia virus
PBS	Phosphate Buffered Saline
PI3K	Phosphoinositide 3-kinase
PE	Phycoerythrin
PerCP	Peridinin Chlorophyll
RNA	ribonucleic acid
RT	room temperature
s	serine
Т	threonine
TAE	tris-acetate EDTA
TNF	tumour necrosis factor
WHO	World Health Organization

CHAPTER 1

INTRODUCTION

Acute myeloid leukaemia (AML) is an infrequent but extremely malignant neoplasm which is responsible for a large number of cancer-related deaths. AML is a heterogeneous group of malignant haemapoietic precursor of the myeloid lineage. AML can be characterised by the predominance of immature cells as well as the loss of regular haemapoietic functions. If the disease is left untreated, the patient progresses towards death within weeks to months of clinical presentation (Johnstone et al., 2002).

In the United States of America, the age-adjusted incident of AML was reported to be 3.4 case/100,000 persons (Kanno et al., 2004b).According to the 2007 National Cancer Registry 3.1 percent of new cancer cases were reported to be myeloid leukaemia (Ariffin and Saleha, 2011). Childhood AML is rare (Karin et al., 1997) but the incidence increases at adulthood with a higher risk of occurance among the elderly (Johansson and Harrison, 2009).

Diagnosing AML successfully has always been a challenge for pathologists as the treatment hinges on the diagnosis. Various classifications for AML has been set, from the traditional FAB classification that depends on morphological and immunophenotypic evaluations, to the more recent WHO classification that utilises morphology, immunophenotyping and genetic abnormalities (Bennett et al., 1976; Vardiman et al., 2002).

Chemotherapy remains the standard therapeutic approach to treat the malignancy but little of the treatment regimen has changed over the past three decades ((Kimby et al., 2001). The chemotherapeutic drugs exert its cytotoxic effects on the cells by damaging DNA, lipid membrane and cellular bodies (Keesler et al., 1998). The targeted myeloblast is then killed by inducing apoptosis through a cascade of biochemical events (Kim et al., 2010).

Around 80% of patients who underwent the standard induction regimen achieved complete remission (CR) and is adequate for the younger patients (Stone et al., 2004). However, it is observed that elderly patients (>60 years) are highly resistant to the chemotherapeutic treatment and have poor overall outcomes (Leith et al., 1997b). However, the overall survival of AML from 2003 to 2009 was reported to be 22 to 25.7% with men having lower overall survival percentage (Howlander, 2012).

The current method of detecting drug resistance in AML is through the painful and invasive procedure of obtaining the patient's bone marrow aspiration. It is usually done approximately 3-4 weeks from the first day of induction therapy to observe the presence of malignant myeloblast in the marrow. Clinicians also tries to predict drug

resistance through several other prognostic markers such as age, white blood count and cytogenetic markers before chemotherapy treatment.

Multidrug resistance (MDR) remains a major hurdle to overcome in achieving CR in AML patients especially among the elderly patients. This is because the myeloblast are able to escape the drug induced apoptosis through overexpression of anti-apoptotic molecules. Previous studies has implicated immature phenotypic markers (Kim et al., 2006), a consecutive activation and up-regulation of mediator of survival pathway e.g. PI3K/Akt(Tazzari et al., 2008) and MAP kinase (Abedini et al., 2009) as well as increased expression of anti-apoptotic Bcl-2 family (Cory et al., 2003)protein as factors of drug resistance.

Previous study has shown that chemotherapy induces a change in the proteomic expression profile due to the induction of molecules and mediators of survival and apoptotic pathways (Devemy et al., 2001; Maha et al., 2009). Maha (2009) has observed that induction therapy is capable of inducing changes in expression of key signalling molecules of PI3K/Akt and MAP kinase pathway. Similarly, it was also observed that the cytokine expression changes when the patient undergoes induction therapy (Devemy et al., 2001; Maha et al., 2009). These changes in intracellular molecule expression are discovered to have an association with the patient's treatment outcome.

It is believed that the changes of these intracellular molecules caused by induction therapy are detectable in AML cells found in the peripheral blood and respond early in treatment. Therefore, they have the potential of being markers to monitor tumour cell response and predicting treatment outcome at the early stages of induction therapy.

This research attempts to study the peripheral blood of expression of surface phenotypic markers, various signalling mediators from survival pathways, cytokines and receptors; and Bcl-2 family protein in association with drug resistance at early induction of AML patients.

The objectives of this research are as follows:

- To determine the expression levels of surface markers, cytokines, signalling mediators and apoptotic related molecules in the peripheral blood of acute leukaemia samples before and during chemotherapy using flow cytometry;
- To determine the level of cytokine mRNA expression and correlate with cytokine protein expression in leukaemia samples before and during chemotherapy; and
- To correlate the expressionlevels of these molecules with treatment outcome.

In this study, it is hypothesised that:

- 1. Cytokines, IL-1β, IL-18, and IL-18 receptor are significantly increased in resistant AML samples during chemotherapy.
- 2. Cytokines, IL-6 and TNF-αare significantly decreased in resistant AML samples during chemotherapy.
- 3. Pro-survival mediators of signalling pathway including Akt, p38 and JNK are significantly increased in resistant AML samples during chemotherapy...
- 4. Pro-apoptotic mediators of signalling pathways including BAD, Bim, and FKHR are significantly decreased in resistant AML samples during chemotherapy.
- 5. Surface markers including CD34 and CD117 are significantly increased in resistant AML samples during chemotherapy.



REFERENCES

- Abedini M, Muller E, Bergeron R, Gray D, Tsang B. 2009. Akt promotes chemoresistance in human ovarian cancer cells by modulating cisplatin-induced, p53-dependent ubiquitination of FLICE-like inhibitory protein. Oncogene 29:11-25.
- Accili D, Arden KC. 2004. FoxOs at the Crossroads of Cellular Metabolism, Differentiation, and Transformation. Cell 117:421-426.
- Aho TLT, Sandholm J, Peltola KJ, Mankonen HP, Lilly M, Koskinen PJ. 2004. Pim-1 kinase promotes inactivation of the pro-apoptotic Bad protein by phosphorylating it on the Ser112 gatekeeper site. FEBS Letters 571:43-49.
- Aichberger KJ, Mayerhofer M, Krauth M-T, Vales A, Kondo R, Derdak S, Pickl WF, Selzer E, Deininger M, Druker BJ, Sillaber C, Esterbauer H, Valent P. 2005. Low-Level Expression of Proapoptotic Bcl-2–Interacting Mediator in Leukemic Cells in Patients with Chronic Myeloid Leukemia: Role of BCR/ABL, Characterization of Underlying Signaling Pathways, and Reexpression by Novel Pharmacologic Compounds. Cancer research 65:9436-9444.
- Alberts B, Bray D, Hopkins K, Johnson A. 1998. Essential Cell Biology: with CD: An Introduction to the Molecular Biology of the Cell: Garland Pub.
- Appelbaum FR, Gundacker H, Head DR, Slovak ML, Willman CL, Godwin JE, Anderson JE, Petersdorf SH. 2006. Age and acute myeloid leukemia. Blood 107:3481-3485.
- Apte RN, Dotan S, Elkabets M, White MR, Reich E, Carmi Y, Song X, Dvozkin T, Krelin Y, Voronov E. 2006. The involvement of IL-1 in tumorigenesis, tumor invasiveness, metastasis and tumor-host interactions. Cancer and Metastasis Reviews 25:387-408.
- Arber DA, Stein AS, Carter NH, Ikle D, Forman SJ, Slovak ML. 2003. Prognostic Impact of Acute Myeloid Leukemia Classification Importance of Detection of Recurring Cytogenetic Abnormalities and Multilineage Dysplasia on Survival. American journal of clinical pathology 119:672-680.
- Arrifin OZ, Saleha ITN. 2011. National Cancer Registry Report 2007. In: Ministry of Health, Malaysia.
- Bachur NR, Yu F, Johnson R, Hickey R, Wu Y, Malkas L. 1992. Helicase inhibition by anthracycline anticancer agents. Molecular pharmacology 41:993-998.
- Bai D, Ueno L, Vogt PK. 2009a. Akt-mediated regulation of NFκB and the essentialness of NFκB for the oncogenicity of PI3K and Akt. International Journal of Cancer 125:2863-2870.

- Bai D, Ueno L, Vogt PK. 2009b. Akt- mediated regulation of NFκB and the essentialness of NFκB for the oncogenicity of PI3K and Akt. International Journal of Cancer 125:2863-2870.
- Balkwill F. 2006. TNF-α in promotion and progression of cancer. Cancer and Metastasis Reviews 25:409-416.
- Barksby H, Lea S, Preshaw P, Taylor J. 2007. The expanding family of interleukin-1 cytokines and their role in destructive inflammatory disorders. Clinical & Experimental Immunology 149:217-225.
- Bast RC, Kufe DW, Pollock RE, Weichselbaum RR, Holland JF, Frei E, Andreeff M, Goodrich DW, Pardee AB. 2000. Cell Proliferation, Differentiation, and Apoptosis.
- Benderra Z, Faussat AM, Sayada L, Perrot J-Y, Tang R, Chaoui D, Morjani H, Marzac C, Marie J-P, Legrand O. 2005. MRP3, BCRP, and P-glycoprotein activities are prognostic factors in adult acute myeloid leukemia. Clinical cancer research 11:7764-7772.
- Bennett JM, Catovsky D, Daniel MT, Flandrin G, Galton DA, Gralnick HR, Sultan C. 1976. Proposals for the classification of the acute leukaemias. French-American-British (FAB) co-operative group. Br J Haematol 33:451-458.
- Bennett JM, Catovsky D, Daniel MT, Flandrin G, Galton DAG, Gralnick HR, Sultan C. 1985. Proposed Revised Criteria for the Classification of Acute Myeloid Leukemia A Report of the French-American-British Cooperative Group. Annals of Internal Medicine 103:620-625.
- Boissel N, Leroy H, Brethon B, Philippe N, De Botton S, Auvrignon A, Raffoux E, Leblanc T, Thomas X, Hermine O. 2006. Incidence and prognostic impact of c-Kit, FLT3, and Ras gene mutations in core binding factor acute myeloid leukemia (CBF-AML). Leukemia 20:965-970.
- Bower JE, Ganz PA, Aziz N, Olmstead R, Irwin MR, Cole SW. 2007. Inflammatory responses to psychological stress in fatigued breast cancer survivors: relationship to glucocorticoids. Brain, behavior, and immunity 21:251-258.
- Breuleux M, Klopfenstein M, Stephan C, Doughty CA, Barys L, Maira S-M, Kwiatkowski D, Lane HA. 2009. Increased AKT S473 phosphorylation after mTORC1 inhibition is rictor dependent and does not predict tumor cell response to PI3K/mTOR inhibition. Molecular Cancer Therapeutics 8:742-753.
- Brown P, McIntyre E, Rau R, Meshinchi S, Lacayo N, Dahl G, Alonzo TA, Chang M, Arceci RJ, Small D. 2007. The incidence and clinical significance of nucleophosmin mutations in childhood AML. Blood 110:979-985.
- Bruserud O, Tjonnfjord G, Gjertsen BT, Foss B, Ernst P. 2000. New strategies in the treatment of acute myelogenous leukemia: mobilization and transplantation of autologous peripheral blood stem cells in adult patients. Stem Cells 18:343-351.

- Cai B, Chang SH, Becker EB, Bonni A, Xia Z. 2006. p38 MAP kinase mediates apoptosis through phosphorylation of BimEL at Ser-65. Journal of Biological Chemistry 281:25215-25222.
- Cai J, Damaraju VL, Groulx N, Mowles D, Peng Y, Robins MJ, Cass CE, Gros P. 2008. Two distinct molecular mechanisms underlying cytarabine resistance in human leukemic cells. Cancer research 68:2349-2357.
- Cairoli R, Beghini A, Grillo G, Nadali G, Elice F, Ripamonti CB, Colapietro P, Nichelatti M, Pezzetti L, Lunghi M. 2006. Prognostic impact of c-KIT mutations in core binding factor leukemias: an Italian retrospective study. Blood 107:3463-3468.
- Campo E, Swerdlow SH, Harris NL, Pileri S, Stein H, Jaffe ES. 2011. The 2008 WHO classification of lymphoid neoplasms and beyond: evolving concepts and practical applications. Blood 117:5019-5032.
- Cappellini A, Mantovani I, Tazzari PL, Grafone T, Martinelli G, Cocco L, Martelli AM. 2005. Application of flow cytometry to molecular medicine: detection of tumor necrosis factor-related apoptosis-inducing ligand receptors in acute myeloid leukaemia blasts. International journal of molecular medicine 16:1041.
- Carter BZ, Mak DH, Schober WD, Dietrich MF, Pinilla C, Vassilev LT, Reed JC, Andreeff M. 2008. Triptolide sensitizes AML cells to TRAIL-induced apoptosis via decrease of XIAP and p53-mediated increase of DR5. Blood 111:3742-3750.
- Carvalho F, Coelho A, Domingues C, Carvalho J, Ribeiro A, Costa S, Goncalves A, Alves V, Dourado M, Sarmento-Ribeiro A. 2010. Evaluation of the therapeutic potential of recombinant TRAIL in leukemia- a study in cell lines in culture In: Haematologica: Ferrata Storti Foundattion via Guisepee Belli 4m 27100 Pavia, Italy. p 691-691.
- Casas SI, Ollila J, Aventín A, Vihinen M, Sierra J, Knuutila S. 2003. Changes in apoptosis-related pathways in acute myelocytic leukemia. Cancer genetics and cytogenetics 146:89-101.
- Castaigne S, Chevret S, Archimbaud E, Fenaux P, Bordessoule D, Tilly H, de Revel T, Simon M, Dupriez B, Renoux M. 2004. Randomized comparison of double induction and timed-sequential induction to a "3+ 7" induction in adults with AML: long-term analysis of the Acute Leukemia French Association (ALFA) 9000 study. Blood 104:2467-2474.
- Chen C-C, Yang C-F, Yang M-H, Lee K-D, Kwang W-K, You J-Y, Yu Y-B, Ho C-H, Tzeng C-H, Chau W-K. 2005a. Pretreatment prognostic factors and treatment outcome in elderly patients with de novo acute myeloid leukemia. Annals of oncology 16:1366-1373.
- Chen CC, Yang CF, Yang MH, Lee KD, Kwang WK, You JY, Yu YB, Ho CH, Tzeng CH, Chau WK, Hsu HC, Gau JP. 2005b. Pretreatment prognostic factors and

treatment outcome in elderly patients with de novo acute myeloid leukemia. Ann Oncol 16:1366-1373.

- Chen K-H, Weng M-S, Lin J-K. 2007. Tangeretin suppresses IL-1β-induced cyclooxygenase (COX)-2 expression through inhibition of p38 MAPK, JNK, and AKT activation in human lung carcinoma cells. Biochemical pharmacology 73:215-227.
- Cheong J-W, Eom JI, Maeng H-Y, Lee ST, Hahn JS, Ko YW, Min YH. 2003. Constitutive phosphorylation of FKHR transcription factor as a prognostic variable in acute myeloid leukemia. Leukemia research 27:1159-1162.
- Chiang C-W, Kanies C, Kim KW, Fang WB, Parkhurst C, Xie M, Henry T, Yang E. 2003. Protein phosphatase 2A dephosphorylation of phosphoserine 112 plays the gatekeeper role for BAD-mediated apoptosis. Molecular and cellular biology 23:6350-6362.
- Chittenden T, Harrington EA, O'Connor R, Remington C, Lutz RJ, Evan GI, Guild BC. 1995. Induction of apoptosis by the Bcl-2 homologue Bak.
- Chiu CP, Lee F. 1989. IL-6 is a differentiation factor for M1 and WEHI-3B myeloid leukemic cells. The Journal of Immunology 142:1909-1915.
- Ciechomska I, Pyrzynska B, Kazmierczak P, Kaminska B. 2003. Inhibition of Akt kinase signalling and activation of Forkhead are indispensable for upregulation of FasL expression in apoptosis of glioma cells. Oncogene 22:7617-7627.
- Cochet O, Teillaud J-L, Sautès C. 1998. Immunological techniques made easy: John Wiley & Sons.
- Cory S, Huang DC, Adams JM. 2003. The Bcl-2 family: roles in cell survival and oncogenesis. Oncogene 22:8590-8607.
- Costello RT, Mallet F, Gaugler B, Sainty D, Arnoulet C, Gastaut J-A, Olive D. 2000. Human acute myeloid leukemia CD34+/CD38- progenitor cells have decreased sensitivity to chemotherapy and Fas-induced apoptosis, reduced immunogenicity, and impaired dendritic cell transformation capacities. Cancer research 60:4403-4411.
- Craig FE, Foon KA. 2008. Flow cytometric immunophenotyping for hematologic neoplasms. Blood 111:3941-3967.
- Creutzig U, Ritter J, Zimmermann M, Hermann J, Gadner H, Sawatzki DB, Niemeyer C, Schwabe D, Selle B, Boos J. 2001. Idarubicin improves blast cell clearance during induction therapy in children with AML: results of study AML-BFM 93. AML-BFM Study Group. Leukemia 15:348-354.
- Datta SR, Katsov A, Hu L, Petros A, Fesik SW, Yaffe MB, Greenberg ME. 2000. 14-3-3 proteins and survival kinases cooperate to inactivate BAD by BH3 domain phosphorylation. Molecular cell 6:41-51.

- Deng H, Zhang J, Yoon T, Song D, Li D, Lin A. 2011. Phosphorylation of Bclassociated death protein (Bad) by erythropoietin-activated c-Jun N-terminal protein kinase 1 contributes to survival of erythropoietin-dependent cells. The International Journal of Biochemistry & Cell Biology 43:409-415.
- Devemy E, Li B, Tao M, Horvath E, Chopra H, Fisher L, Nayini J, Creech S, Venugopal P, Yang J. 2001. Poor prognosis acute myelogenous leukemia: 3 biological and molecular biological changes during remission induction therapy. Leukemia research 25:783-791.
- Dhanasekaran DN, Reddy EP. 2008. JNK signaling in apoptosis. Oncogene 27:6245-6251.
- Dijkers PF, Lammers J-WJ, Koenderman L, Coffer PJ. 2000. Expression of the proapoptotic Bcl-2 family member Bim is regulated by the forkhead transcription factor FKHR-L1. Current Biology 10:1201-1204.
- Dinarello C. 2002. The IL-1 family and inflammatory diseases. Clinical and experimental rheumatology 20:S1-S13.
- Dinarello CA. 1996. Biologic basis for interleukin-1 in disease. Blood 87:2095-2147.
- Dohner H, Estey EH, Amadori S, Appelbaum FR, Buchner T, Burnett AK, Dombret H, Fenaux P, Grimwade D, Larson RA, Lo-Coco F, Naoe T, Niederwieser D, Ossenkoppele GJ, Sanz MA, Sierra J, Tallman MS, Lowenberg B, Bloomfield CD. 2010. Diagnosis and management of acute myeloid leukemia in adults: recommendations from an international expert panel, on behalf of the European LeukemiaNet. Blood 115:453-474.
- Dolado I, Nebreda AR. 2008. Regulation of tumorigenesis by p38α MAP kinase. In: Stress-Activated Protein Kinases: Springer. p 99-128.
- Dolcet X, Llobet D, Pallares J, Matias-Guiu X. 2005. NF-kB in development and progression of human cancer. Virchows Archiv 446:475-482.
- Dorr R, Karanes C, Spier C, Grogan T, Greer J, Moore J, Weinberger B, Schiller G, Pearce T, Litchman M. 2001. Phase I/II study of the P-glycoprotein modulator PSC 833 in patients with acute myeloid leukemia. Journal of clinical oncology 19:1589-1599.
- Doyle LA, Ross DD. 2003. Multidrug resistance mediated by the breast cancer resistance protein BCRP (ABCG2). Oncogene 22:7340-7358.
- Dumka D, Puri P, Carayol N, Lumby C, Balachandran H, Schuster K, Verma AK, Terada LS, Platanias LC, Parmar S. 2009. Activation of the p38 Map kinase pathway is essential for the antileukemic effects of dasatinib. Leukemia & lymphoma 50:2017-2029.
- Edling CE, Hallberg B. 2007. c-Kit—a hematopoietic cell essential receptor tyrosine kinase. The International Journal of Biochemistry & Cell Biology 39:1995-1998.

- Elmore S. 2007. Apoptosis: a review of programmed cell death. Toxicologic pathology 35:495-516.
- Fang X, Yu S, Eder A, Mao M, Bast Jr RC, Boyd D, Mills GB. 1999. Regulation of BAD phosphorylation at serine 112 by the Ras-mitogen-activated protein kinase pathway. Oncogene 18:6635-6640.
- Fleming Y, Armstrong C, MORRICE N, PATERSON A, GOEDERT M, COHEN P. 2000. Synergistic activation of stress-activated protein kinase 1/c-Jun Nterminal kinase (SAPK1/JNK) isoforms by mitogen-activated protein kinase kinase 4 (MKK4) and MKK7. Biochem J 352:145-154.
- Fortune JM, Osheroff N. 2000. Topoisomerase II as a target for anticancer drugs: when enzymes stop being nice. Progress in nucleic acid research and molecular biology 64:221-253.
- Fu Z, Tindall D. 2008. FOXOs, cancer and regulation of apoptosis. Oncogene 27:2312-2319.
- Galmarini CM, Thomas X, Calvo F, Rousselot P, Jafaari AE, Cros E, Dumontet C. 2002. Potential mechanisms of resistance to cytarabine in AML patients. Leukemia research 26:621-629.
- Gewirtz D. 1999. A critical evaluation of the mechanisms of action proposed for the antitumor effects of the anthracycline antibiotics adriamycin and daunorubicin. Biochemical pharmacology 57:727-741.
- Grandage VL, Gale RE, Linch DC, Khwaja A. 2005. PI3-kinase/Akt is constitutively active in primary acute myeloid leukaemia cells and regulates survival and chemoresistance via NF-kappaB, Mapkinase and p53 pathways. Leukemia 19:586-594.
- Grethe S, Coltella N, Di Renzo MF, Pörn-Ares MI. 2006. p38 MAPK downregulates phosphorylation of Bad in doxorubicin-induced endothelial apoptosis. Biochemical and biophysical research communications 347:781-790.
- Grimwade D. 2001. The clinical significance of cytogenetic abnormalities in acute myeloid leukaemia. Best Pract Res Clin Haematol 14:497-529.
- Grivennikov S, Karin E, Terzic J, Mucida D, Yu G-Y, Vallabhapurapu S, Scheller J, Rose-John S, Cheroutre H, Eckmann L. 2009. IL-6 and Stat3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. Cancer cell 15:103-113.
- Guo S, Rena G, Cichy S, He X, Cohen P, Unterman T. 1999. Phosphorylation of serine 256 by protein kinase B disrupts transactivation by FKHR and mediates effects of insulin on insulin-like growth factor-binding protein-1 promoter activity through a conserved insulin response sequence. Journal of Biological Chemistry 274:17184-17192.

- Hande KR. 2008. Topoisomerase II inhibitors. Update on Cancer Therapeutics 3:13-26.
- Harada H, Quearry B, Ruiz-Vela A, Korsmeyer SJ. 2004. Survival factor-induced extracellular signal-regulated kinase phosphorylates BIM, inhibiting its association with BAX and proapoptotic activity. Proceedings of the National Academy of Sciences of the United States of America 101:15313-15317.
- Harris AL, Hochhauser D. 1992. Mechanisms of multidrug resistance in cancer treatment. Acta Oncologica 31:205-213.
- Hayat M, Hurteloup P, Parmentier C, Carde P, Pico JO, Schlumberger M, Chahine G, Kamioner D. 1984. Phase I trial of Idarubicin (4-demethoxydaunorubicin) in adult acute leukemia. Invest New Drugs 2:375-379.
- Hazlehurst LA, Argilagos RF, Dalton WS. 2007. β1 integrin mediated adhesion increases Bim protein degradation and contributes to drug resistance in leukaemia cells. British Journal of Haematology 136:269-275.
- Hazzalin CA, Cano E, Cuenda A, Barratt MJ, Cohen P, Mahadevan LC. 1996. p38/RK is essential for stress-induced nuclear responses: JNK/SAPKs and c-Jun/ATF-2 phosphorylation are insufficient. Current Biology 6:1028-1031.
- Heinrich P, Behrmann I, Haan S, Hermanns H, Muller-Newen G, Schaper F. 2003. Principles of interleukin (IL)-6-type cytokine signalling and its regulation. Biochem J 374:1-20.
- Henderson B, Dougherty W, James V, Tilley L, Noble J. 1982. Safety assessment of a new anticancer compound, mitoxantrone, in beagle dogs: comparison with doxorubicin. I. Clinical observations. Cancer treatment reports 66:1139-1143.
- Hideshima T, Nakamura N, Chauhan D, Anderson KC. 2001. Biologic sequelae of interleukin-6 induced PI3-K/Akt signaling in multiple myeloma. Oncogene 20:5991-6000.
- Hui L, Bakiri L, Mairhorfer A, Schweifer N, Haslinger C, Kenner L, Komnenovic V,
 Scheuch H, Beug H, Wagner EF. 2007. p38α suppresses normal and cancer cell proliferation by antagonizing the JNK–c-Jun pathway. Nature genetics 39:741-749.
- Hui RC-Y, Gomes AR, Constantinidou D, Costa JR, Karadedou CT, de Mattos SF, Wymann MP, Brosens JJ, Schulze A, Lam EW-F. 2008. The forkhead transcription factor FOXO3a increases phosphoinositide-3 kinase/Akt activity in drug-resistant leukemic cells through induction of PIK3CA expression. Molecular and cellular biology 28:5886-5898.
- Jaffrézou J-P, Levade T, Bettaieb A, Andrieu N, Bezombes C, Maestre N, Vermeersch S, Rousse A, Laurent G. 1996. Daunorubicin-induced apoptosis: triggering of ceramide generation through sphingomyelin hydrolysis. The EMBO journal 15:2417.

- Johansson B, Harrison CJ. 2009. Acute Myeloid Leukemia. In: Heim S, Mitelman F, editors. Cancer Cytogenetics, 3 eds ed. New Jersey: John Wiley&Sons Inc.
- Johnstone RW, Ruefli AA, Lowe SW. 2002. Apoptosis: a link between cancer genetics and chemotherapy. Cell 108:153-164.
- Kanno S-i, Higurashi A, Watanabe Y, Shouji A, Asou K, Ishikawa M. 2004a. Susceptibility to cytosine arabinoside (Ara-C)-induced cytotoxicity in human leukemia cell lines. Toxicology Letters 152:149-158.
- Kanno S-i, Shouji A, Hirata R, Asou K, Ishikawa M. 2004b. Effects of naringin on cytosine arabinoside (Ara-C)-induced cytotoxicity and apoptosis in P388 cells. Life sciences 75:353-365.
- Karin M, Liu Z-g, Zandi E. 1997. AP-1 function and regulation. Current opinion in cell biology 9:240-246.
- Keesler GA, Bray J, Hunt J, Johnson DA, Gleason T, Yao Z, Wang S-W, Parker C, Yamane H, Cole C. 1998. Purification and activation of recombinant p38 isoforms α , β , γ , and δ . Protein expression and purification 14:221-228.
- Kelekar A, Chang BS, Harlan JE, Fesik SW, Thompson CB. 1997. Bad is a BH3 domain-containing protein that forms an inactivating dimer with Bcl-XL. Molecular and cellular biology 17:7040-7046.
- Kim J-H, Lee SC, Ro J, Kang HS, Kim HS, Yoon S. 2010. Jnk signaling pathwaymediated regulation of Stat3 activation is linked to the development of doxorubicin resistance in cancer cell lines. Biochemical pharmacology 79:373-380.
- Kim J, Kim C, Kim TS, Bang SI, Yang Y, Park H, Cho D. 2006. IL-18 enhances thrombospondin-1 production in human gastric cancer via JNK pathway. Biochemical and biophysical research communications 344:1284-1289.
- Kim K, Fisher MJ, Xu SQ, el-Deiry WS. 2000. Molecular determinants of response to TRAIL in killing of normal and cancer cells. Clin Cancer Res 6:335-346.
- Kimby E, Nygren P, Glimelius B. 2001. A systematic overview of chemotherapy effects in acute myeloid leukaemia. Acta Oncol 40:231-252.
- Kishimoto T. 2010. IL-6: from its discovery to clinical applications. International immunology 22:347-352.
- Kolch W. 2000. Meaningful relationships: the regulation of the Ras/Raf/MEK/ERK pathway by protein interactions. Biochem J 351:289-305.
- Kornblau SM, Singh N, Qiu Y, Chen W, Zhang N, Coombes KR. 2010. Highly Phosphorylated FOXO3A Is an Adverse Prognostic Factor in Acute Myeloid Leukemia. Clinical Cancer Research 16:1865-1874.

- Krutzik PO, Trejo A, Schulz KR, Nolan GP. 2011. Phospho flow cytometry methods for the analysis of kinase signaling in cell lines and primary human samples. In: Hawley TS, Hawley RG, editors. Flow Cytometry Protocols, 3rd Editon ed. Totowa, NJ: Humana Press. p 179-202.
- L. Y. 2005. Disfunction og the apoptotic pathway in cancer cells. In: Application of apoptosis to cancer treatment: Springer.
- Laurent G, Jaffrézou J-P. 2001. Signaling pathways activated by daunorubicin. Blood 98:913-924.
- Le PT, Lazorick S, Whichard LP, Yang YC, Clark SC, Haynes BF, Singer KH. 1990. Human thymic epithelial cells produce IL-6, granulocyte-monocyte-CSF, and leukemia inhibitory factor. The Journal of Immunology 145:3310-3315.
- Lee M-W, Park SC, Yang YG, Yim SO, Chae HS, Bach J-H, Lee HJ, Kim KY, Lee WB, Kim SS. 2002. The involvement of reactive oxygen species (ROS) and p38 mitogen-activated protein (MAP) kinase in TRAIL/Apo2L-induced apoptosis. FEBS Letters 512:313-318.
- Legrand-Poels S, Schoonbroodt S, Piette J. 2000. Regulation of interleukin-6 gene expression by pro-inflammatory cytokines in a colon cancer cell line. Biochem J 349:765-773.
- Legrand O, Simonin G, Beauchamp-Nicoud A, Zittoun R, Marie J-P. 1999. Simultaneous activity of MRP1 and Pgp is correlated with in vitro resistance to daunorubicin and with in vivo resistance in adult acute myeloid leukemia. Blood 94:1046-1056.
- Lei K, Davis RJ. 2003. JNK phosphorylation of Bim-related members of the Bcl2 family induces Bax-dependent apoptosis. Proceedings of the National Academy of Sciences 100:2432-2437.
- Leith CP, Kopecky KJ, Godwin J, McConnell T, Slovak ML, Chen I-M, Head DR, Appelbaum FR, Willman CL. 1997a. Acute myeloid leukemia in the elderly: assessment of multidrug resistance (MDR1) and cytogenetics distinguishes biologic subgroups with remarkably distinct responses to standard chemotherapy. A Southwest Oncology Group study. Blood 89:3323-3329.
- Leith CP, Kopecky KJ, Godwin J, McConnell T, Slovak ML, Chen IM, Head DR, Appelbaum FR, Willman CL. 1997b. Acute myeloid leukemia in the elderly: assessment of multidrug resistance (MDR1) and cytogenetics distinguishes biologic subgroups with remarkably distinct responses to standard chemotherapy. A Southwest Oncology Group study. Blood 89:3323-3329.
- Ley R, Ewings K, Hadfield K, Cook S. 2005. Regulatory phosphorylation of Bim: sorting out the ERK from the JNK. Cell Death & Differentiation 12:1008-1014.

- Lizcano J, MORRICE N, COHEN P. 2000. Regulation of BAD by cAMP-dependent protein kinase is mediated via phosphorylation of a novel site, Ser155. Biochem J 349:547-557.
- Lotem J, Sachs L. 2002. Cytokine control of developmental programs in normal hematopoiesis and leukemia. Oncogene 21:3284-3294.
- Lowenberg B, Downing JR, Burnett A. 1999. Acute myeloid leukemia. N Engl J Med 341:1051-1062.
- Lu J, Quearry B, Harada H. 2006. p38-MAP kinase activation followed by BIM induction is essential for glucocorticoid-induced apoptosis in lymphoblastic leukemia cells. FEBS Letters 580:3539-3544.
- Luciano F, Jacquel A, Colosetti P, Herrant M, Cagnol S, Pages G, Auberger P. 2003. Phosphorylation of Bim-EL by Erk1/2 on serine 69 promotes its degradation via the proteasome pathway and regulates its proapoptotic function. Oncogene 22:6785-6793.
- Łukaszewicz M, Mroczko B, Szmitkowski M. 2007. Clinical significance of interleukin-6 (IL-6) as a prognostic factor of cancer disease. Pol Arch Med Wewn 117:247-251.
- Macdonald A, Campbell DG, Toth R, McLauchlan H, Hastie CJ, Arthur JSC. 2006. Pim kinases phosphorylate multiple sites on Bad and promote 14-3-3 binding and dissociation from Bcl-XL. BMC cell biology 7:1.
- MacEwan DJ. 2002. TNF receptor subtype signalling: differences and cellular consequences. Cellular signalling 14:477-492.
- Maekawa T, Metcalf D, Gearing DP. 1990a. Enhanced suppression of human myeloid leukemic cell lines by combinations of IL-6, LIF, GM-CSF and G-CSF. International Journal of Cancer 45:353-358.
- Maekawa T, Metcalf D, Gearing DP. 1990b. Enhanced suppression of human myeloid leukemic cell lines by combinations of IL- 6, LIF, GM- CSF and G- CSF. International Journal of Cancer 45:353-358.
- Maha A. 2003. Prognostic marker of resistance and relapse in acute leukemia. In: Faculty of Medicine and Health Science. Kuala Lumpur: University Putra Malaysia.
- Maha A, Cheong S-K, Leong C-F, Seow H-F. 2009. Molecular responses during chemotherapy in acute myeloid leukemias in predicting poor-response to standard chemotherapy. Malaysian J Pathol 31:81-91.
- Manzo F, Nebbioso A, Miceli M, Conte M, De Bellis F, Carafa V, Franci G, Tambaro FP, Altucci L. 2009. TNF-related apoptosis-inducing ligand: Signalling of a '< i> smart</i>'molecule. The International Journal of Biochemistry & Cell Biology 41:460-466.

- Marchion DC, Cottrill HM, Xiong Y, Chen N, Bicaku E, Fulp WJ, Bansal N, Chon HS, Stickles XB, Kamath SG. 2011. BAD phosphorylation determines ovarian cancer chemosensitivity and patient survival. Clinical cancer research 17:6356-6366.
- Mason KD, Juneja SK, Szer J. 2006. The immunophenotype of acute myeloid leukemia: is there a relationship with prognosis? Blood reviews 20:71-82.
- Masters SC, Yang H, Datta SR, Greenberg ME, Fu H. 2001. 14-3-3 inhibits Badinduced cell death through interaction with serine-136. Molecular pharmacology 60:1325-1331.
- McCubrey JA, Steelman LS, Chappell WH, Abrams SL, Wong EW, Chang F, Lehmann B, Terrian DM, Milella M, Tafuri A. 2007a. Roles of the Raf/MEK/ERK pathway in cell growth, malignant transformation and drug resistance. Biochimica et Biophysica Acta (BBA)-Molecular Cell Research 1773:1263-1284.
- McCubrey JA, Steelman LS, Chappell WH, Abrams SL, Wong EW, Chang F, Lehmann B, Terrian DM, Milella M, Tafuri A, Stivala F, Libra M, Basecke J, Evangelisti C, Martelli AM, Franklin RA. 2007b. Roles of the Raf/MEK/ERK pathway in cell growth, malignant transformation and drug resistance. Biochim Biophys Acta 1773:1263-1284.
- McIlwain C, Townsend D, Tew K. 2006. Glutathione S-transferase polymorphisms: cancer incidence and therapy. Oncogene 25:1639-1648.
- Mendelsohn J, Howley PM, Israel MA, Liotta LA. 2001. The molecular basis of cancer: Saunders.
- Milella M, Kornblau SM, Estrov Z, Carter BZ, Lapillonne H, Harris D, Konopleva M, Zhao S, Estey E, Andreeff M. 2001. Therapeutic targeting of the MEK/MAPK signal transduction module in acute myeloid leukemia. Journal of Clinical Investigation 108:851-859.
- Min Y, Eom J, Cheong J, Maeng H, Kim J, Jeung H, Lee S, Lee M, Hahn J, Ko Y.
 2003. Constitutive phosphorylation of Akt/PKB protein in acute myeloid leukemia: its significance as a prognostic variable. Leukemia 17:995-997.
- Moore JO, George SL, Dodge RK, Amrein PC, Powell BL, Kolitz JE, Baer MR, Davey FR, Bloomfield CD, Larson RA, Schiffer CA. 2005. Sequential multiagent chemotherapy is not superior to high-dose cytarabine alone as postremission intensification therapy for acute myeloid leukemia in adults under 60 years of age: Cancer and Leukemia Group B Study 9222. Blood 105:3420-3427.
- Neame PB, Soamboonsrup P, Browman GP, Meyer RM, Benger A, Wilson WE, Walker IR, Saeed N, McBride JA. 1986. Classifying acute leukemia by immunophenotyping: a combined FAB-immunologic classification of AML. Blood 68:1355-1362.

- O'Connor L, Strasser A, O'Reilly LA, Hausmann G, Adams JM, Cory S, Huang DC. 1998. Bim: a novel member of the Bcl-2 family that promotes apoptosis. The EMBO journal 17:384-395.
- Olson JM, Hallahan AR. 2004. p38 MAP kinase: a convergence point in cancer therapy. Trends in molecular medicine 10:125-129.
- Ottilie S, Diaz J-L, Horne W, Chang J, Wang Y, Wilson G, Chang S, Weeks S, Fritz LC, Oltersdorf T. 1997. Dimerization properties of human BAD identification of a BH-3 domain and analysis of its binding to mutant bcl-2 and bcl-xl proteins. Journal of Biological Chemistry 272:30866-30872.
- Padron E, Fernandez H. 2012. Anthracycline dose intensification in young adults with acute myeloid leukemia. Therapeutic advances in hematology 3:17-27.
- Park S, Cheon S, Cho D. 2007. The dual effects of interleukin-18 in tumor progression. Cell Mol Immunol 4:329-335.
- Pawson T, Scott JD. 1997. Signaling through scaffold, anchoring, and adaptor proteins. Science 278:2075-2080.
- Pelengaris S, Khan M. 2013. The Molecular Biology of Cancer: A Bridge from Bench to Bedside: Wiley. com.
- Peyssonnaux C, Eychène A. 2001. The Raf/MEK/ERK pathway: new concepts of activation. Biology of the Cell 93:53-62.
- Plo I, Bettaïeb A, Payrastre B, Mansat-De Mas V, Bordier C, Rousse A, Kowalski-Chauvel A, Laurent G, Lautier D. 1999. The phosphoinositide 3-kinase/Akt pathway is activated by daunorubicin in human acute myeloid leukemia cell lines. FEBS Letters 452:150-154.
- Porter GW, Khuri FR, Fu H. 2006. Dynamic 14-3-3/client protein interactions integrate survival and apoptotic pathways. Seminars in Cancer Biology 16:193-202.
- Putcha GV, Le S, Frank S, Besirli CG, Clark K, Chu B, Alix S, Youle RJ, LaMarche A, Maroney AC, Johnson Jr EM. 2003. JNK-Mediated BIM Phosphorylation Potentiates BAX-Dependent Apoptosis. Neuron 38:899-914.
- Rastogi RC. 2003. Cell Biology, 3rd ed. New Delhi: New Age International Publishers.
- Rena G, Guo S, Cichy SC, Unterman TG, Cohen P. 1999. Phosphorylation of the transcription factor forkhead family member FKHR by protein kinase B. Journal of Biological Chemistry 274:17179-17183.
- Riccioni R, Senese M, Diverio D, Riti V, Mariani G, Boe A, LoCoco F, Foà R, Peschle C, Sporn M. 2008. Resistance of acute myeloid leukemic cells to the triterpenoid

CDDO-Imidazolide is associated with low caspase-8 and FADD levels. Leukemia research 32:1244-1258.

- Roberts KG, Smith AM, McDougall F, Carpenter H, Horan M, Neviani P, Powell JA, Thomas D, Guthridge MA, Perrotti D. 2010a. Essential requirement for PP2A inhibition by the oncogenic receptor c-KIT suggests PP2A reactivation as a strategy to treat c-KIT+ cancers. Cancer research 70:5438-5447.
- Roberts KG, Smith AM, McDougall F, Carpenter H, Horan M, Neviani P, Powell JA, Thomas D, Guthridge MA, Perrotti D, Sim ATR, Ashman LK, Verrills NM. 2010b. Essential Requirement for PP2A Inhibition by the Oncogenic Receptor c-KIT Suggests PP2A Reactivation as a Strategy to Treat c-KIT+ Cancers. Cancer research 70:5438-5447.
- Rock KL, Lai J-J, Kono H. 2011a. Innate and adaptive immune responses to cell death. Immunological Reviews 243:191-205.
- Rock KL, Lai JJ, Kono H. 2011b. Innate and adaptive immune responses to cell death. Immunological reviews 243:191-205.
- Roy SK, Srivastava RK, Shankar S. 2010. Inhibition of PI3K/AKT and MAPK/ERK pathways causes activation of FOXO transcription factor, leading to cell cycle arrest and apoptosis in pancreatic cancer. Journal of molecular signaling 5:10.
- Rubnitz JE, Gibson B, Smith FO. 2010. Acute myeloid leukemia. Hematol Oncol Clin North Am 24:35-63.
- Samuel L, Cummings J, Shaw P. 1998. Daunorubicin cardiotoxicity in childhood cancer. The Lancet 352:1150.
- Saxena R, Anand H. 2008. Flow cytometry in acute leukemia. Indian Journal of Hematology and Blood Transfusion 24:146-150.
- Schindler R, Mancilla J, Endres S, Ghorbani R, Clark S, Dinarello CA. 1990.
 Correlations and interactions in the production of interleukin-6 (IL-6), IL-1, and tumor necrosis factor (TNF) in human blood mononuclear cells: IL-6 suppresses IL-1 and TNF. Blood 75:40-47.
- Schoch C, Schnittger S, Klaus M, Kern W, Hiddemann W, Haferlach T. 2003. AML with 11q23/MLL abnormalities as defined by the WHO classification: incidence, partner chromosomes, FAB subtype, age distribution, and prognostic impact in an unselected series of 1897 cytogenetically analyzed AML cases. Blood 102:2395-2402.
- Secchiero P, Gonelli A, Mirandola P, Melloni E, Zamai L, Celeghini C, Milani D, Zauli G. 2002. Tumor necrosis factor–related apoptosis-inducing ligand induces monocytic maturation of leukemic and normal myeloid precursors through a caspase-dependent pathway. Blood 100:2421-2429.

- Secchiero P, Zauli G. 2008. Tumor necrosis factor-related apoptosis-inducing ligand and the regulation of hematopoiesis. Current opinion in hematology 15:42-48.
- Shipley JL, Butera JN. 2009. Acute myelogenous leukemia. Experimental hematology 37:649-658.
- Sims JE. 2002. IL-1 and IL-18 receptors, and their extended family. Current opinion in immunology 14:117-122.
- Smeets M, Raymakers R, Vierwinden G, Pennings A, Boezeman J, Minderman H, De Witte T. 1999. Idarubicin DNA intercalation is reduced by MRP1 and not Pgp. Leukemia 13:1390-1398.
- Steelman L, Pohnert S, Shelton J, Franklin R, Bertrand F, McCubrey J. 2004. JAK/STAT, Raf/MEK/ERK, PI3K/Akt and BCR-ABL in cell cycle progression and leukemogenesis. Leukemia 18:189-218.
- Steelman LS, Blalock WL, Wang X-Y, Moye PW, Lee JT, Shelton JG, Navolanic PM, Davis JM, Knapp SL, Franklin RA. 2003. Fibroblastic, hematopoietic, and hormone responsive epithelial cell lines and culture conditions for elucidation of signal transduction and drug resistance pathways by gene transfer. In: Cancer Cell Signaling: Springer. p 185-201.
- Steptoe A, Hamer M, Chida Y. 2007. The effects of acute psychological stress on circulating inflammatory factors in humans: a review and meta-analysis. Brain, behavior, and immunity 21:901-912.
- Stone RM, O'Donnell MR, Sekeres MA. 2004. Acute myeloid leukemia. Hematology Am Soc Hematol Educ Program:98-117.
- Sujobert P, Bardet V, Cornillet-Lefebvre P, Hayflick JS, Prie N, Verdier F, Vanhaesebroeck B, Muller O, Pesce F, Ifrah N. 2005. Essential role for the p110δ isoform in phosphoinositide 3-kinase activation and cell proliferation in acute myeloid leukemia. Blood 106:1063-1066.
- Sunayama J, Tsuruta F, Masuyama N, Gotoh Y. 2005. JNK antagonizes Akt-mediated survival signals by phosphorylating 14-3-3. The Journal of cell biology 170:295-304.
- Sunters A, de Mattos SF, Stahl M, Brosens JJ, Zoumpoulidou G, Saunders CA, Coffer PJ, Medema RH, Coombes RC, Lam EW-F. 2003. FoxO3a transcriptional regulation of Bim controls apoptosis in paclitaxel-treated breast cancer cell lines. Journal of Biological Chemistry 278:49795-49805.
- Suvannasankha A, Minderman H, O'Loughlin KL, Nakanishi T, Ford LA, Greco WR, Wetzler M, Ross DD, Baer MR. 2004. Breast cancer resistance protein (BCRP/MXR/ABCG2) in adult acute lymphoblastic leukaemia: frequent expression and possible correlation with shorter disease- free survival. British Journal of Haematology 127:392-398.

- Tazzari PL, Tabellini G, Ricci F, Papa V, Bortul R, Chiarini F, Evangelisti C, Martinelli G, Bontadini A, Cocco L. 2008. Synergistic proapoptotic activity of recombinant TRAIL plus the Akt inhibitor Perifosine in acute myelogenous leukemia cells. Cancer research 68:9394-9403.
- Teiten MH, Eifes S, Reuter S, Duvoix A, Dicato M, Diederich M. 2009. Gene Expression Profiling Related to Anti- inflammatory Properties of Curcumin in K562 Leukemia Cells. Annals of the New York Academy of Sciences 1171:391-398.
- Thomas X, Archimbaud E. 1997. Mitoxantrone in the treatment of acute myelogenous leukemia: a review. Hematol Cell Ther 39:63-74.
- Tournier C, Dong C, Turner TK, Jones SN, Flavell RA, Davis RJ. 2001. MKK7 is an essential component of the JNK signal transduction pathway activated by proinflammatory cytokines. Genes & development 15:1419-1426.
- Van Dongen J, Macintyre E, Gabert J, Delabesse E, Rossi V, Saglio G, Gottardi E, Rambaldi A, Dotti G, Griesinger F. 1999. Standardized RT-PCR analysis of fusion gene transcripts from chromosome aberrations in acute leukemia for detection of minimal residual disease Report of the BIOMED-I Concerted Action: Investigation of minimal residual disease in acute leukemia. Leukemia 13:1901-1928.
- Van Snick J. 1990. Interleukin-6: an overview. Annual review of immunology 8:253-278.
- Vardiman JW, Harris NL, Brunning RD. 2002. The World Health Organization (WHO) classification of the myeloid neoplasms. Blood 100:2292-2302.
- Vardiman JW, Thiele J, Arber DA, Brunning RD, Borowitz MJ, Porwit A, Harris NL, Le Beau MM, Hellstrom-Lindberg E, Tefferi A, Bloomfield CD. 2009. The 2008 revision of the World Health Organization (WHO) classification of myeloid neoplasms and acute leukemia: rationale and important changes. Blood 114:937-951.
- Ventura JJ, Tenbaum S, Perdiguero E, Huth M, Guerra C, Barbacid M, Pasparakis M, Nebreda AR. 2007. p38α MAP kinase is essential in lung stem and progenitor cell proliferation and differentiation. Nature genetics 39:750-758.
- Vivanco I, Sawyers CL. 2002. The phosphatidylinositol 3-kinase–AKT pathway in human cancer. Nature Reviews Cancer 2:489-501.
- Voronov E, Shouval DS, Krelin Y, Cagnano E, Benharroch D, Iwakura Y, Dinarello CA, Apte RN. 2003. IL-1 is required for tumor invasiveness and angiogenesis. Proceedings of the National Academy of Sciences 100:2645-2650.
- Wang J, Zhou J-Y, Wu GS. 2011. Bim protein degradation contributes to cisplatin resistance. Journal of Biological Chemistry 286:22384-22392.

- Wang S, El-Deiry WS. 2003. TRAIL and apoptosis induction by TNF-family death receptors. Oncogene 22:8628-8633.
- Wen J, Ramadevi N, Nguyen D, Perkins C, Worthington E, Bhalla K. 2000. Antileukemic drugs increase death receptor 5 levels and enhance Apo-2L– induced apoptosis of human acute leukemia cells. Blood 96:3900-3906.
- Wong GW, Manaf A, Seow H-F. 1999. Cloning, expression of human interleukin-18 expression and the effects of chemical stimuli. Asia Pac J Biotechnol Mol Biol 7:1-11.
- Xu Q, Simpson S-E, Scialla TJ, Bagg A, Carroll M. 2003. Survival of acute myeloid leukemia cells requires PI3 kinase activation. Blood 102:972-980.
- Yamaguchi H, Wang H-G. 2002. Bcl-XL protects BimEL-induced Bax conformational change and cytochrome C release independent of interacting with Bax or BimEL. Journal of Biological Chemistry 277:41604-41612.
- Yang L. 2005. Disfunction of the Apoptotic Pathway in Cancer Cells. In: Application of Apoptosis to Cancer Treatment: Springer Netherlands. p 1-28.
- Yang X, Fraser M, Abedini M, Bai T, Tsang B. 2008. Regulation of apoptosisinducing factor-mediated, cisplatin-induced apoptosis by Akt. British journal of cancer 98:803-808.
- Yang X, Fraser M, Moll UM, Basak A, Tsang BK. 2006. Akt-mediated cisplatin resistance in ovarian cancer: modulation of p53 action on caspase-dependent mitochondrial death pathway. Cancer research 66:3126-3136.
- Yen M-L, Tsai H-F, Wu Y-Y, Hwa H-L, Lee B-H, Hsu P-N. 2008. TNF-related apoptosis-inducing ligand (TRAIL) induces osteoclast differentiation from monocyte/macrophage lineage precursor cells. Molecular immunology 45:2205-2213.
- Yoo JK, Kwon H, Khil L-Y, Zhang L, Jun H-S, Yoon J-W. 2005. IL-18 induces monocyte chemotactic protein-1 production in macrophages through the phosphatidylinositol 3-kinase/Akt and MEK/ERK1/2 pathways. The Journal of Immunology 175:8280-8286.
- Yoshimura A. 2006. Signal transduction of inflammatory cytokines and tumor development. Cancer science 97:439-447.
- Yu C, Minemoto Y, Zhang J, Liu J, Tang F, Bui TN, Xiang J, Lin A. 2004. JNK suppresses apoptosis via phosphorylation of the proapoptotic Bcl-2 family protein BAD. Molecular cell 13:329-340.
- Zamarron BF, Chen W. 2011. Dual roles of immune cells and their factors in cancer development and progression. International journal of biological sciences 7:651.

- Zebisch A, Czernilofsky AP, Keri G, Smigelskaite J, Sill H, Troppmair J. 2007. Signaling through RAS-RAF-MEK-ERK: from basics to bedside. Current medicinal chemistry 14:601-623.
- Zha J, Harada H, Yang E, Jockel J, Korsmeyer SJ. 1996. Serine Phosphorylation of Death Agonist BAD in Response to Survival Factor Results in Binding to 14-3-3 Not BCL-XL. Cell 87:619-628.
- Zhang B, Ma X-T, Zheng G-G, Li G, Rao Q, Wu K-F. 2003. Expression of IL-18 and its receptor in human leukemia cells. Leukemia research 27:813-822.
- Zhang B, Wang Y, Zheng G-G, Ma X-T, Li G, Zhang F-K, Wu K-F. 2002. Clinical significance of IL-18 gene over-expression in AML. Leukemia research 26:887-892.
- Zhang X, Tang N, Hadden TJ, Rishi AK. 2011. Akt, FoxO and regulation of apoptosis. Biochimica et Biophysica Acta (BBA) - Molecular Cell Research 1813:1978-1986.
- Zhao S, Konopleva M, Cabreira-Hansen M, Xie Z, Hu W, Milella M, Estrov Z, Mills G, Andreeff M. 2003. Inhibition of phosphatidylinositol 3-kinase dephosphorylates BAD and promotes apoptosis in myeloid leukemias. Leukemia 18:267-275.
- Zhou LJ, Tedder TF. 1995. A distinct pattern of cytokine gene expression by human CD83+ blood dendritic cells. Blood 86:3295-3301.
- Zhu Y, Swanson BJ, Wang M, Hildeman DA, Schaefer BC, Liu X, Suzuki H, Mihara K, Kappler J, Marrack P. 2004. Constitutive association of the proapoptotic protein Bim with Bcl-2-related proteins on mitochondria in T cells. Proceedings of the National Academy of Sciences of the United States of America 101:7681-7686.