

UNIVERSITI PUTRA MALAYSIA

DEVELOPMENT OF IN VITRO PROPAGATION TECHNIQUES FOR ENDOSPERMUM MALACCENSE M.A AND SHOREA PARVIFOLIA DYER

AZIAH MOHD YUSOFF

FH 2003 13

DEVELOPMENT OF *IN VITRO* PROPAGATION TECHNIQUES FOR ENDOSPERMUM MALACCENSE M.A AND SHOREA PARVIFOLIA DYER

By

AZIAH MOHD YUSOFF

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

April 2003



Abstract of thesis submitted to the Senate of University Putra Malaysia in fulfilment of the requirements for the degree of Doctor of Philosophy

DEVELOPMENT OF IN VITRO PROPAGATION TECHNIQUES FOR ENDOSPERMUM MALACCENSE M.A AND SHOREA PARVIFOLIA DYER

by

AZIAH BINTI MOHD YUSOFF

April 2003

Chairman: Associate Prof. Dr. Nor Aini Ab. Shukor

Forestry

Faculty:

Endospermum malaccense M.A and Shorea parvifolia Dyer are two commercially important timber species identified as potential plantation species. The procurement and storage of their seeds are difficult and is a major hindrance to plantation establishment. Development of micropropagation techniques is being pursued to provide an alternative in planting stock production.

Micropropagation of *E. malaccense* was achieved through *in vitro* production of plants through axillary shoots development from nodal segment explants of an elite tree. The explants were initially washed in 10 changes of sterile distilled water, followed by 5 changes of 0.05% (v/v) Tween 20 solution for 10 minutes each. This was then followed by rinsing in 10 changes of sterile distilled water and subsequently sterilised in a solution comprising 60 % (v/v) Clorox and 0.05% v/v Tween 20 for 10 minutes. After



which they were rinsed 10 times in 300ml sterile distilled water and finally immersed in 70% (v/v) ethanol for 1 minute. Shoots were induced on the nodal segment explants in MS basal salts supplemented with 22.2 x 10^{-6} M or 44.4 x 10^{-6} M. For shoot multiplication, the best medium is MS supplemented with 44.4 x 10^{-6} M BAP and solidified with a mixture of 1.7 g Gelrite and 4g Bacto agar per liter. *In vivo* rooting with Seradix 3 was more suitable for *in vitro* produced shoots of *E. malaccense*, compared with an *in vitro* rooting technique.

For *S. parvifolia*, high contamination was observed in all explant types. Multiple shoots were induced on nodal segments culture in WPM solid medium supplemented with 10⁻⁵ M BAP or 10⁻⁶ TDZ but were non-amenable to further subculture. Callus developed from immature seeds with gelatinous endosperm termed as embryonic masses in WPM supplemented with 10⁻⁴M CPA induced callus formation. Globular shaped callus developed upon subculture. Histological examination of the globular shaped callus showed no evidence of somatic embryos formation. The globular structure was similar to the development of nodules.

The contaminants found on the immature seeds included a fungus, *Collectotrycum* spp. and a range of bacteria which are as follows: *-Kleibsella planticola*, *Enterobacter* agglomerans, Erwina spp. (E. uredora or E. herbicola), Serratia odorifera, Serratia marcesens, Serratia proteomaculans, Morganella moragnii, Kluyera ascorbata. A fern, Asplenium nidus was found contaminating the nodal segment cultures.

Abstrak tesis yang dikemukakan kepada Senat Universityi Putra Malaysia sebagai memenuhi keperluan ijazah Doktor Falsafah

PEMBENTUKAN TEKNIK MIKRO PERAMBATAN UNTUK ENDOSPERMUM MALACCENSE M.A DAN SHOREA PARVIFOLIA DYER

Oleh

AZIAH BINTI MOHD YUSOFF

April 2003

Pengerusi: Prof. Madya Dr. Nor Aini Ab. Shukor

Fakulti:

Perhutanan

Endospermum malaccense M.A dan Shorea parvifolia Dyer, merupakan dua sepsis balak yang berpotensi ditanam secara ladang. Masalah pengutipan dan penyimpanan bijibenih menghindar kemajuan peladangan hutan secara komersial. Pembentukan teknik mikroperambatan dapat mengatasi masalah pembekalan bahan tanaman.

Mikroperambatan *E. Malaccense* tercapai apabila pembentukan pucuk aksil terbentuk ke atas eksplan bahagian nod pokok elit. Eksplan dibasmikuman dengan mencuci 10 kali dengan air suling steril, diikuti dengan 5 pencucian dalam larutan 0.05% (isipadu/isipadu) Tween 20 selama 10 minit, diikuti pula dengan 10 pertukaran air suling steril. Kemudian dicuci pula dalam larutan 60% (isipadu/isipadu) Clorox dicampur dengan 0.05% (isipadu/isipadu) Tween 20 selama 10 minit. Kemudian dicuci dengan 10 kali dengan 300ml air suling steril dan akhir sekali direndam didalam 70% (isipadu/isipadu) etanol. Media MS dicampur dengan 22.2 x 10⁻⁶ M or 44.4 x 10⁻⁶ M sesuai dalam pengaruhan pembentukan pucuk. Gandaan pucuk tercapai dalam campuran medium MS dan 44.4 x 10⁻⁶ M BAP yang dipepejalkan dengan jel campuran 1.7 g Gelrite and 4g Bacto agar se liter. Teknik pengakaran *in vivo* dengan Seradix 3 lebih sesuai dibandingkan dengan teknik *in vitro*.

Untuk *S. parvifolia*, kadar kontaminasi tinggi dihadapi dalam kultur semua jenis eksplan. Pembentukan pucuk berbilang tercapai apabila bahagian nod di kulturkan didalam medium WPM yang ditambah dengan 10⁻⁵ M BAP atau 10⁻⁶ M TDZ. Pertambahan pucuk tidak tercapai dalam semua jenis medium. Pembentukan kalus terjadi ke atas eksplan endoperma cair di dalam medium WPM ditambah dengan 10⁻⁴ M CPA. Pengsubkulturan menghasilkan kalus berbentul globul. Kajian histology menunjukkan bahawa struktur ini adalah nodul.

Bijibenih muda dikontaminasikan oleh sejenis kulat, Collectotrycum spp. Dan pelbagai jenis bacteria seperti berikut:--Kleibsella planticola, Enterobacter agglomerans, Erwina spp. (E. uredora or E. herbicola), Serratia odorifera, Serratia marcesens, Serratia proteomaculans, Morganella moragnii, Kluyera ascorbata. Sejenis paku penumpang Asplenium nidus juga didapati menkontaminasikan kultur.



ACKNOWLEDGEMENTS

In the name of Allah S.W.T. The Most Gracious. The Most Merciful.

First and foremost I would like to convey my appreciation to my present employer the Golden Hope Research Sdn. Bhd. (GHRSB), for allowing me to continue my studies till completion. My appreciation goes to the Executive Director of Golden Hope Research Sdn. Bhd., Dr. Mohd Hashim Tajudin, for his support and to Dr. Mohd Nor Ghani, the Research Controller of the BioScience Division for his continuous encouragement. I am indebted to the management of the Forest Research Institute of Malaysia (FRIM), my former employer, for the permission to pursue my studies during my tenure in FRIM. Special thanks goes to the Director General Dato' Abd. Razak bin Mohd Ali, who has been very supportive of my pursuit to complete my studies.

I deeply appreciate the support of my supervisor Assoc. Prof. Dr. Nor Aini Shukor for her patience, encouragement and guidance. My heartfelt thanks go to the members supervisory committee, Assoc. Prof. Dr. Kamis Awang and Assoc. Prof. Dr. Midhzar Abdul Kadir for their suggestions and comments leading to the completion of this thesis.

To my former Research Assistant in FRIM, Pn. Sharifah Maulana, I thank her for assisting me in my experiments as well as her patience and fruitful suggestions. To my



present assistant at GHRSB, Pn. Ruziah Ahmad, I thank her for her assistance in the final layout and final touches of my thesis and to Asmida of FRIM, I am grateful for her assistance in the preparation of the colour prints for this thesis.

A million thanks goes to the members of the tissue culture laboratory at FRIM, Pn Fadhilah, Pn. Haliza, Pn. NorAsmah, Pn Halilah, Pn Normah, Pn Rozidah, Cik Sabariah, Pn Naemah, Pn Rukiah and Nizam for their cooperation.

To my former colleagues at FRIM, Dr. Ab. Rasip, Mohd Nor, David McKellar, Norhara, Dr. Ilham and numerous others, I thank you all for your support and suggestions.

To my beloved family, Mansor, my husband and children, Murni, Maizura, Marlina and Muhd. Isa, and last but not least to my mother, Cik An, my deepest appreciation, for their continuous encouragement, understanding and most of all, for their sacrifices in bearing all the shortcomings during these trying period, in order for me to complete this thesis. Thank you.



TABLE OF CONTENTS

Page

	S2 •
ABSTRACT	
ABSTRAK	iv
ACKNOWLEDGEMENTS	vi
APPROVAL SHEETS	viii
DECLARATION FORM	х
TABLE OF CONTENTS	xi
LISTS OF TABLES	xviii
LISTS OF FIGURES	xxiii
LISTS OF ABBREVIATIONS	xxix
TERMINOLOGY	xxxi

CHAPTER I

Π

INTRODUCTION

1 ·

LITERATURE REVIEW	7
Shorea parvifolia Dyer	7
Botany and Taxonomy	7
Distribution and Ecology	7
Uses	7
Propagation of S. parvifolia	8
Potential as Plantation Species and the Improvement	8
Activity	8
Endospermum malaccense M.A	9
Botany and Taxonomy	9
Distribution and Ecology	9
Uses	10
Propagation of <i>E. malaccense</i>	10
Status of Global Forest Plantation	10
Plantation Forestry in Malaysia	12
Plantation Establishment of Indigenous Species	14
Planting Stock Production of Indigenous Species	16
Vegetative Propagation	19
In Vitro Propagation of Tropical Forest Species	21
Explant Selection and Sterilisation Procedures	24
Microbial Contaminants of Plant Tissue Culture	30
Plant Production Through The Multiplication of Axillary	
Shoots	32
Factors Affecting Plant Production Through the Multiplication	
of Axillary Shoots	33
Genotype of Explants	33
Media Composition	34
Plant Growth Regulators	38
Sugars as sources of carbohydrates	42
Plant Production Through Somatic Embryogenesis	42
Factors Affecting the Production and Development of	
Embryogenic Callus.	44



Factors Associated with Explants	45
Effects of Media Composition	48
Effects of Plant Growth Regulators and Plant Growth	
Factors	50
Effects of Carbohydrates Source	53
Effects of Light	54
Maturation and Germination of Somatic Embryos	55
Effects of Abscisic Acid	56
Effects of Carbohydrates	57
Dessication	57
Rooting and Establishment of Plantlets	58
Media	59
Sugars	59
Plants Growth Regulators	60
Light and Temperature	62
Explant Support System	64
Factors Associated with Explants	65
Problems Faced in In Vitro Propagation of Plants	67
Browning of Cultures	67
Lag Time of Response from Explants of Mature	
Donors	68
Vitrification of shoots	68
MATERIALS AND METHODS	70
MATERIALS AND METHODS Introduction	70 70
Introduction Preparation of Media	70
Introduction Preparation of Media Murashige and Skoog Media, MS (1962)	70 70
Introduction Preparation of Media	70 70
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962) Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium.	70 70 70
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962) Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown,	70 70 70
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962) Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980).	70 70 70 70 70 71
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962) Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980). Preparation of Stock Solution of Growth Substances	70 70 70 70 70 71 71
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962) Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980). Preparation of Stock Solution of Growth Substances. Description of Explants Used.	70 70 70 70 71 71 71 72
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962) Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980). Preparation of Stock Solution of Growth Substances Description of Explants Used. Identification of Contaminants.	70 70 70 70 71 71 71 72 73
 Introduction. Preparation of Media Murashige and Skoog Media, MS (1962). Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980). Preparation of Stock Solution of Growth Substances. Description of Explants Used. Identification of Contaminants. Histological Techniques. 	70 70 70 70 71 71 71 72 73 75
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962) Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980). Preparation of Stock Solution of Growth Substances. Description of Explants Used. Identification of Contaminants. Histological Techniques. Resin Embedding.	70 70 70 71 71 71 72 73 75 75
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962) Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980). Preparation of Stock Solution of Growth Substances. Description of Explants Used. Identification of Contaminants. Histological Techniques. Resin Embedding. Fixation.	70 70 70 71 71 71 72 73 75 75
 Introduction. Preparation of Media Murashige and Skoog Media, MS (1962). Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980). Preparation of Stock Solution of Growth Substances. Description of Explants Used. Identification of Contaminants. Histological Techniques. Resin Embedding. Fixation. Dehydration. 	70 70 70 71 71 71 72 73 75 75 75 75
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962) Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980). Preparation of Stock Solution of Growth Substances. Description of Explants Used. Identification of Contaminants. Histological Techniques. Resin Embedding. Fixation. Dehydration. Infiltration.	70 70 70 71 71 71 72 73 75 75 75 75 75 77
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962). Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980). Preparation of Stock Solution of Growth Substances. Description of Explants Used. Identification of Contaminants. Histological Techniques. Resin Embedding. Fixation. Dehydration. Infiltration. Embedding.	70 70 70 70 71 71 71 72 73 75 75 75 75 77 77
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962). Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980). Preparation of Stock Solution of Growth Substances. Description of Explants Used. Identification of Contaminants. Histological Techniques. Resin Embedding. Fixation. Dehydration. Infiltration. Embedding. Cutting.	70 70 70 71 71 71 72 73 75 75 75 75 75 77 77 77 77
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962) Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980) Preparation of Stock Solution of Growth Substances. Description of Explants Used. Identification of Contaminants. Histological Techniques. Resin Embedding. Fixation. Dehydration. Infiltration. Embedding. Cutting. Staining.	70 70 70 71 71 71 72 73 75 75 75 75 75 77 77 77 77 77
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962) Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980). Preparation of Stock Solution of Growth Substances. Description of Explants Used. Identification of Contaminants. Histological Techniques. Resin Embedding. Fixation. Dehydration. Infiltration. Embedding. Cutting. Staining. Mounting.	70 70 70 70 71 71 71 72 73 75 75 75 75 75 75 77 77 77 77 78 79 80
Introduction. Preparation of Media Murashige and Skoog Media, MS (1962) Gamborg B5 medium and Schenk and Hilderbrandt (SH) medium. Woody Plant Medium, WPM (Lloyd and Mc Cown, 1980) Preparation of Stock Solution of Growth Substances. Description of Explants Used. Identification of Contaminants. Histological Techniques. Resin Embedding. Fixation. Dehydration. Infiltration. Embedding. Cutting. Staining.	70 70 70 71 71 71 72 73 75 75 75 75 75 77 77 77 77 77

III



MICROPROPAGATION OF ENDOSPERMUM	
MALACCENSE	
Introduction	
Establishment of Sterile Explants and Determination	
of Suitable Media Compositions	
Sterilisation of Explants	
Determination of suitable media for Establishment of	
Cultures	
Optimisation of Media for the Multiplication of	
Shoots	
Overcoming Vitrification	
Rooting Experiments and Acclimatisation of Shoots	
Results and Discussions	
Establishment of Sterile Explants and Determination	
of Suitable Media Compositions	
Sterilisation of Explants	
Determination of suitable media	
Multiplication of Shoots	
Overcoming Vitrification Problems	
Rooting And Acclimatisation Of Shoots	
Summary of Results	
SHOREA PARVIFOLIA Introduction	
Explant Sources	
In Vitro Germinated Seedlings	
Immature Seeds.	
Explants from Seedlings Germinated in Non-Sterile	
Conditions	
Explants from Nursery Raised Seedlings	
Nodal segments and Shoot – Tips from Mature	
Nodal segments and Shoot – Tips from Mature Trees	
Nodal segments and Shoot –Tips from Mature Trees Establishment of Sterile Culture and The Induction of	
Nodal segments and Shoot –Tips from Mature Trees Establishment of Sterile Culture and The Induction of Propagules On Various Explants of <i>S. parvifolia</i>	
Nodal segments and Shoot –Tips from Mature Trees Establishment of Sterile Culture and The Induction of Propagules On Various Explants of <i>S. parvifolia</i> Development of Sterilisation Protocol for Mature	
Nodal segments and Shoot –Tips from Mature Trees Establishment of Sterile Culture and The Induction of Propagules On Various Explants of <i>S. parvifolia</i> Development of Sterilisation Protocol for Mature Seeds.	
Nodal segments and Shoot –Tips from Mature Trees Establishment of Sterile Culture and The Induction of Propagules On Various Explants of <i>S. parvifolia</i> Development of Sterilisation Protocol for Mature	
Nodal segments and Shoot –Tips from Mature Trees Establishment of Sterile Culture and The Induction of Propagules On Various Explants of <i>S. parvifolia</i> Development of Sterilisation Protocol for Mature Seeds.	
Nodal segments and Shoot –Tips from Mature Trees Establishment of Sterile Culture and The Induction of Propagules On Various Explants of <i>S. parvifolia</i> Development of Sterilisation Protocol for Mature Seeds <i>In Vitro</i> Germination of Mature <i>S. parvifolia</i>	
Nodal segments and Shoot –Tips from Mature Trees Establishment of Sterile Culture and The Induction of Propagules On Various Explants of <i>S. parvifolia</i> Development of Sterilisation Protocol for Mature Seeds <i>In Vitro</i> Germination of Mature <i>S. parvifolia</i> Seeds	
Nodal segments and Shoot –Tips from Mature Trees. Establishment of Sterile Culture and The Induction of Propagules On Various Explants of S. parvifolia. Development of Sterilisation Protocol for Mature Seeds. In Vitro Germination of Mature S. parvifolia Seeds. Induction of Propagules on Explants Excised from	
Nodal segments and Shoot –Tips from Mature Trees. Establishment of Sterile Culture and The Induction of Propagules On Various Explants of S. parvifolia. Development of Sterilisation Protocol for Mature Seeds. In Vitro Germination of Mature S. parvifolia Seeds. Induction of Propagules on Explants Excised from In Vitro Germinated Seedlings. Effects of MS and WPM Media in Combination	
Nodal segments and Shoot –Tips from Mature Trees Establishment of Sterile Culture and The Induction of Propagules On Various Explants of S. parvifolia Development of Sterilisation Protocol for Mature Seeds	

V

IV

Effects of MS, WPM, SH and B5 Media in Combination with TDZ on Explants Excised from *in vitro* Germinated Seedlings.....

Establishment of Sterile Culture for Explants from	
Seedlings Germinated in Non-Sterile Conditions	125
Sterilisation Protocols and Media Used in	
Establishment of Sterile Culture for Explants	
from Seedlings in Non-sterile Conditions	125
Response of explants Excised from In Vivo	
Germinated Seedlings to Sterilisation Protocol	
and Media Used in the Induction of	
Propagagules	126
Establishment of Sterile Cultures for Shoot Tips and	
Nodal Segment Explants from Nursery Raised	
Seedlings	137
Development of Sterilisation Protocol for Shoot	157
Tips and Nodal Segment Explants from Nursery	
Raised Seedlings	137
Effects of Different Sterilisation Protocols on	157
Nodal Segments and Shoot Tip Explants from	140
Nursery Raised Seedlings	140
Experiments with Fungicides and Antibiotic Pre-	140
Treated Nursery Raised Seedlings	142
Pretreatment Protocols and Media Used for	
Explants from Pre-Treated Nursery Raised	1.40
Seedlings	142
Response of Explants Excised from Nursery	
Raised Seedlings to Pre-Treatment	144
Response of Nodal Segments, Shoot Tip	
Explants Excised from Nursery-Raised	
Seedlings Pre-Treated with Fungicide and	
Antibiotic to Different Plant Growth	
Regulators	146
The Effect of Different Medium Formulations on	
Growth Responses of Nodal Segments and Shoot	
Tip Explants of S. parvifolia Cultured in	
vitro	149
The Effects of Supplementing BAP in WPM	
Medium on Nodal Segment and Leaf Explants	
Excised from Nursery Raised Seedlings	155
The Effects of Supplementing Different	
Combinations of BAP with Kinetin in WPM	
Medium on Nodal Segments and Leaf Explants	
Excised from Nursery Raised Seedlings	156
The Effects of Supplementing WPM Medium	
with TDZ on Shoot Tip and Nodal Segment	
Explants from Nursery Raised Seedlings	157
The Effects of Supplementing WPM Medium	
with NAA or 2,4-D on Explants Excised from	
Shoots of Nursery Raised Seedlings	159
Establishment of Sterile Cultures for Root Tip Explants	
from Nursery-Raised Seedlings	171
	1/1



Sterilisation of Root Tip Explants from Nursery	
Raised Seedlings	171
Induction of Propagules in Root Tip Explants	
from Nursery Raised Seedlings	172
Establishment of Sterile Culture for Explants from	
Mature Trees	175
Sterilisation of Explants from Mature Trees	176
Induction o Propagules on Nodal Segments and	
Shoot Tips from Mature Trees	178
Establishment of Sterile Culture of Immature Seeds	
Sterilisation of Immature Seeds	180
Induction of Propagules on Explants Excised	
from Immature Seeds	184
Multiplication of Propagules	191
Multiplication of Propagules on Nodal Segment and	
Shoot Tip Explants Excised from in vitro and in vivo	
Germinated Seedlings	191
Multiplication of Shoots	191
Multiplication of Callus	197
Multiplication of Propagules on Nodal Segment and	
Shoot Tip Explant Excised from Nursery Raised	
Seedlings	200
Multiplication of Shoots	200
Multiplication of Callus	201
Callus Multiplication On Root Tip Explants	208
Effects of Several Different Types of PGRs on	
Callus Originating from Root Tips Explants	208
Effects of WPM Medium Supplemented with	
BAP and Kinetin on Callus Originating from	
Root Tips Explants	209
Effects of Sucrose, Abscisic Acid and Proline on	
Callus Originating from Root Tips Explants	216
Multiplication Of Callus On Immature Seeds Explants	217
Contaminants Found in <i>S. parvifolia</i> Cultures	222
Fungal Contamination.	222
Bacterial Contaminants	222
Contamination with Epiphytes	223
Histological Examination of Embryogenic Callus from	
Immature Seeds	227
Summary of Results	231

VI	DISCUSSIONS	237
----	-------------	-----



VII	CONCLUSIONS AND RECOMMENDATIONS	245
	REFERENCES	247
	APPENDICES	272

VITA/BIODATA OF THE AUTHOR	304



LISTS OF TABLES

Table		Page
2.1	Annual plantation rates and plantation areas by region and species group	12
3.1	Tabulation of the plant growth regulators used in the experiments, their molecular weights, the desired quantity to prepare 50ml. Of 10 ⁻³ M stock solutions, their initial solvents and distilled water requirements	72
3.2	Glutaraldehyde-Paraformaldehyde-Caffeine fixative	75
3.3	Components of Phosphate buffer pH 7.2, 0.2 M	76
4.1	Effects of using four different sterilization protocol on nodal explants of <i>Endospermum maccalense</i> . Scoring was conducted as follows: $+++=>61\%$, $++=31-60\%$, $+=$ 1-30\%, $-=0\%$. Data were recorded 4 weeks after culture	90
4.2	Effects of the four different media formulations on development of nodal explants of <i>Endospermum</i> maccalense. Scoring was conducted as follows: $+++ = >61\%, ++=31-60\%, +=1-30\%, -+0\%$. Data were recorded 4 weeks after sub-culture	90
4.3	Summary of the t-Test on data from shoot multiplication of <i>Endospermum maccalense</i> in MS medium supplemented with either 2.22 or 4.44 x 10 ⁻⁶ M BAP	91
4.4	The effects of using different gelling agents on rates of vitrification of Endospermum maccalense shoots arising from nodal explants	93
4.5	The Effects of Commercial Rooting Powder Seradix 2 and Seradix 3 on in vitro Rooting and the Effects of the Solid MS Medium Supplemented with IBA and 10-6M and 10-5 M on the <i>in vitro</i> Rooting of <i>E.malaccense</i> Shoots	93
5.1	Effects of different sterilization protocols on percentage browning, percentage germination, and percentage contamination of <i>Shorea parvifolia</i> seeds. Data were recorded after four weeks in culture. Values followed by the same letter within a column are not significantly different (P<0.05; Duncan's Multiple Range Test)	102
5.1	browning, percentage germination, and percentage contamination of <i>Shorea parvifolia</i> seeds. Data were recorded after four weeks in culture. Values followed by the same letter within a column are not significantly	1



5.2	Effects of different media on percentage browning, percentage germination, and percentage contamination of <i>Shorea parvifolia</i> seeds. Observations were made after four week in culture. Values followed by the same letter within a column are not significantly different (P<0.05; Duncan's Multiple Range Test)	108
5.3	Number of hypocotyls, leaf, nodal segment, root and shoot explants cultured in either MS or WPM basal medium, regardless of the combination of BAP and 2,4-D used	110
5.4	Responses of explants from in vitro seedlings of Shorea parvifolia cultured on Murashige and Skoog medium containing benzylamino purine (at 10^{-4} M, 10^{-5} M, 10^{-6} M, 10^{-7} M) alone, or in combination with 2,4-dichlorophenoxyacetic acid (at 10^{-4} M) after 4 weeks in culture	117
5.5	Responses of explants from <i>in vitro</i> germinated seedlings of <i>Shorea parvifolia</i> cultured on Woody Plant Medium (1980) containing benzylamino purine (at 10^{-4} M, 10^{-5} M, 10^{-6} M, 10^{-7} M) alone, or in combination with 2,4-dichlorophenoxyacetic acid (at 10^{-4} M) after 4 weeks in culture	119
5.6	Responses of explants from <i>in vitro</i> germinated seedlings of <i>Shorea parvifolia</i> cultured on Murashige and Skoog MS (1962). Schenk and Hiderbrandt, SH (1972), Gamborg et al, B5, (1968) and Woody Plant Medium (WPM), (Llyod and Mc Cown, 1980) in combination with 10-8 M to 10-6 M thidiazuron (TDZ) after 4 weeks in culture	122
5.7	Responses of explants from <i>in vivo</i> germinated seedlings of <i>Shorea parvifolia</i> cultured on Murashige and Skoog medium containing benzylamino purine (at 10^{-4} M, 10^{-5} M, 10^{-6} M, 10^{-7} M) alone, or in combination with 2,4- dichlorophenoxyacetic acid (at 10^{-4} M) after 4 weeks in culture	129
		-



5.8	Responses of explants from <i>in vivo</i> germinated seedlings of <i>Shorea parvifolia</i> cultured on Woody Plant Medium containing benzylamino purine (at 10^4 M, 10^{-5} M, 10^{-6} M, 10^{-7} M) alone, or in combination with 2,4-dichlorophenoxyacetic acid (at 10^4 M) after 4 weeks in culture	133
5.9	Effects of different sterilization protocols on nodal and shoot tip explants excised from nursery raised seedlings of <i>Shorea parvifolia</i> . Percentage contamination was observed after 4 weeks in WPM medium	141
5.10	Effects of pre-treatment of <i>Shorea parvifolia</i> nursery raised seedlings by root drenching and/or spraying with antibiotics (streptomycin and tetracycline) alone or in combination with the funicide Benlate. Seedlings were treated once a week for 1, 2, 3, or 4 weeks, before excision of nodal segments, leaves and shoot tips. Observations were made after 4 weeks in culture on WPM medium supplemented with TDZ, BAP, kinetin, NAA or 2,4-d alone	146
5.11	Effects of different concentrations of 2,4-D, NAA, BAP, Kinetin and thidiazuron (TDZ) on callus formation, shoot elongation and browning of leaf, nodal segments and shoot tip explants excised from nursery raised seedlings of <i>Shorea parvifolia</i>	149
5.12	Response of explants of <i>Shorea parvifolia</i> explants from nursery raised seedlings to different basal medium compositions. Data were recorded after 4 weeks in culture. (There was no significance difference at α =0.05 for all the means tested)	151
5.13	Response of explants of Shorea parvifolia explants from nursery raised seedlings to WPM medium supplemented with 10-4 M, 10-5 M, 10-6 M and 10-7 M BAP. Data were recorded after 4 weeks in culture. (There was no significance difference at α =0.05 for all the means	157
	tested)	156



5.14	Responses of explants of <i>Shorea parvifolia</i> explants from nursery raised seedlings to WPM medium supplemented with BAP(10^{-7} M to 10^{-4} M) in combination with kinetin (10^{-5} M to 10^{-7} M). Data were recorded after 4 weeks in culture. (There was no significance difference at α =0.05 for all the means tested)	157
5.15	tested) Response of <i>Shorea parvifolia</i> nodal segments and shoot tip explants excised from nursery raised seedlings to WPM medium supplemented with TDZ (10-9M to 10^{-5} M). Data were recorded after 4 weeks in culture. (There was no significance difference at α =0.05 for all the means tested)	157
5.16	Response of all explant types (mid-rib, nodal segment, leaf, petiole and shoot tips) from nursery raised seedlings to WPM medium supplemented with 10^{-7} M to 10^{-4} M 2,4-D. (There was no significance at α =0.05 for all the means tested)	160
5.17	Response of leaf, mid-rib, nodal segment, petiole and shoot tip explants initially cultured in WPM media supplemented with 2,4-D at concertrations (10-7 M to 10-4M) to WPM medium supplemented with 10-7M to 10-8 M 2,4-D	164
5.18	Response of all explant types (mid-rib), nodal segment, leaf, petiole and shoot tips) from nursery raised seedlings to WPM supplemented with 10^{-7} M to 10^{-4} M NAA	168
5.19	Response of leaf, mid-rib, nodal segment, petiole and shoot tip explants initially cultured in WPM media supplemented with NAA at concentrations (10-7 M to 10-4M) to WPM medium supplemented with 10-7M to 10-8 M NAA	170
5.20	Results of the effect different concerntration of Mercuric chloride solution on root explants in terms of non- contamination and response. The different concentration of mercuric chloride used were $S1=0.1\%$ (w/v), $S2=0.2\%$ (w/v), $S3=0.3\%$ (w/v). (There was no significance difference at $\alpha=0.05$ for all the means	
	tested)	172



Results of the effect of supplementing 2,4-D or NAA to Woody Plant Medium on root explants excised from nursery grown seedling. (There was no significnce difference at α =0.05 for all the means tested)	175
Results of different treatments used during collection of <i>Shorea parvifolia</i> explants from a 40-year old tree within FRIM campus. Shoot tips and nodal segment were cultured in half strength MS, half-strength WPM, WPM (NH ₄ NO ₃), SH and B5 media. Explants were sterilized according to method 1 (page 173)	178
Responses of nodal segments and shoot tip explants of <i>Shorea parvifolia</i> to three sterilization protocols when cultured on half-strength Murashige and Skoog medium (MS), half-strength Woody Plant Medium (WPM), WPM lacking ammonium nitrate, Shenck and Hildebrandt Medium (SH) or Gamborg's B5 medium (B5) after 4 weeks in culture	180
Response of immature <i>S. parvifolia</i> seeds to four different sterilization treatments and cultured in media listed on page 72, four weeks after culture. (There was no significance difference at α =0.05 for all the means tested)	183
Types of Plant Growth Regulators Used and Their Abbreviations	184
Response of cotyledon, embryonic mass and whole seeds* of <i>S. parvifolia</i> to WPM medium supplemented with 10-7 to 10-4 to several different types of Plant Growth Regulators (PGR) added alone after 4 weeks in culture	187
Results on multiplication of shoots induced on nodal segments and shoot tip explants excised from <i>in vitro</i> and <i>in vivo</i> germinated seedlings of <i>S. parvifolia</i> cultured in WPM medium supplemented with BAP (10 ⁻⁵ M -10^{-7} M) or TDZ (10 ⁻⁹ M-10 ⁻⁵ M). Observation was carried out four weeks after culture. For each treatment six explants were used	192
	 Woody Plant Medium on root explants excised from nursery grown seedling. (There was no significance difference at α=0.05 for all the means tested) Results of different treatments used during collection of <i>Shorea parvifolia</i> explants from a 40-year old tree within FRIM campus. Shoot tips and nodal segment were cultured in half strength MS, half-strength WPM, WPM (NH₄NO₃), SH and B5 media. Explants were sterilized according to method 1 (page 173) Responses of nodal segments and shoot tip explants of <i>Shorea parvifolia</i> to three sterilization protocols when cultured on half-strength Murashige and Skoog medium (MS), half-strength Woody Plant Medium (WPM), WPM lacking ammonium nitrate, Shenck and Hildebrandt Medium (SH) or Gamborg's B5 medium (B5) after 4 weeks in culture Response of immature <i>S. parvifolia</i> seeds to four different sterilization treatments and cultured in media listed on page 72, four weeks after culture. (There was no significance difference at α=0.05 for all the means tested) Types of Plant Growth Regulators Used and Their Abbreviations Response of cotyledon, embryonic mass and whole seeds* of <i>S. parvifolia</i> to WPM medium supplemented with 10-7 to 10-4 to several different types of Plant Growth Regulators (PGR) added alone after 4 weeks in culture Results on multiplication of shoots induced on nodal segments and shoot tip explants excised from <i>in vitro</i> and <i>in vivo</i> germinated seedlings of <i>S. parvifolia</i> cultured in WPM medium supplemented with BAP (10⁻⁵ M - 10⁻⁷M) or TDZ (10⁻⁹M-10⁻⁵ M). Observation was



5.28	Results on multiplication of callus induced on leaf, cotyledon, hypocotyly, nodal segment and shoot tip explants excised from <i>in vitro</i> and <i>in vivo</i> germinated seedlings of <i>S. parvifolia</i> cultured in WPM medium supplemented with 2,4-D $(10^{4}M - 10^{-7}M)$ or TDZ $(10^{8}M-10^{-4}M)$. Observation was carried out six weeks	
	after culture	198
5.29	The number of tubes used per treatment in callus multiplication experiment. The callus were classified according to the explant origin of callus	201
5.30	Effects of solid WPM medium supplemented with 2,4-D, NAA, BAP, kinetin, NOA, 4-CPA, MCPA, Dicamba and Picloran at levels $(10^{-6} \text{ M to } 10^{-4} \text{ M})$ after six weeks in culture	210
5.31	Effects of solid WPM medium supplemented with a combination of BAP and Kinetin on callus originating from roots of <i>S. parvifolia</i> six weeks after culture. Both PGRs were used at 10 ⁻⁷ M, 10 ⁻⁶ M and 10 ⁻⁵ M	213
5.32	Effects of different levels of Abscisic acid (ABA), sucrose and proline on callus originating from roots	217
5.33	Effects of 2,4-D, NAA, Dicamba, MCPA, NOA, 4-CPA and Picloram on callus originating from embryos	219



LIST OF FIGURES

Figure		Page
2.1	Different phases in somatic embryogenesis (Bornman, 1993)	45
4.1	In vitro rooting of Endospermum malaccense by dipping shoots bases in either Seradix 2 or Seradix 3 and placing shoots in seed trays containing sand, with 10 shoots per tray	88
4.2	In vitro Rooting of Endospermum malaccense in half- strength MS medium solidified with mixture of Bacto- agar and Gelrite and supplemented with either 10 ⁻⁵ M IBA or 10 ⁻⁶ M IBA	88
5.1	Mature seeds of Shorea parvifolia	96
5.2	In vitro germination of <i>S. parvifolia</i> seeds in liquid medium with filter paper support which was prepared by folding two pieces 15 cm disc of Whatman No.1 filter paper folded length-wise. Each fold was 1 cm wide and wavelike fan folds was created and the seeds were placed in between the folds	104
5.3	Response of Hypocotyl, Leaf, Nodal segments and Root Explants Origninating from <i>in vitro</i> germinated seedlings after 4 weeks in WPM and MS media	112
5.4	Response of Hypocotyl, Leaf, Nodal segments Explants Originating from <i>in vivo</i> germinated seedlings after 4 weeks in MS and WPM media	128
5.5	Formation of callus on leaf explants excised from S. <i>parvifolia</i> nursery raised seedlings four weeks after culture in 10^{-7} M TDZ	
		148



5.6	Response of shoot tip explants excised from nursery raised seedlings of S. parvifolia to different media basal used after 4 weeks in culture. (B5 = Gamborg Medium (1968), MS = Murashige & Skoog (1962), SH = Schenk Hilderbrandt (1972), WPM = Woody Plant Medium (Lloyd & Mc Cown, 1980), wpm- = WPM = Woody Plant Medium (Lloyd & Mc Cown, 1980) without NH_4NO_3	
		153
5.7	Response of nodal segment explants excised from nursery raised seedlings of <i>S. parvifolia</i> to different basal media used after 4 weeks in culture. (B5 = Gamborg Medium (1968), MS = Murashige & Skoog (1962), SH = Schenk Hilderbrandt (1972), WPM = Woody Plant Medium (Lloyd & Mc Cown, 1980), wpm- = WPM = Woody Plant Medium (Lloyd & Mc Cown, 1980) without NH ₄ NO ₃	
		154
5.8	Response of leaf, petiole, nodal segment (ns), shoot-tip (st) and mid-rib explants to WPN medium supplemented with 10^{-7} M to 10^{-4} M 2, 4-D after 4 weeks in culture. (D4 = 10^{-4} M 2, 4-D, D5 = 10^{-5} M 2, 4-D, D^ = 10^{-6} M 2, 4-D and D7 = 10^{-7} M 2, 4-D)	163
5.9	Roots developed on callus formed on nodal segment explants. Callus was induced on nodal segment explants in media supplemented 10^{-4} M 2, 4-D after six weeks in culture. Development of roots were observed after transfer to media supplemented with 10^{-8} M 2, 4-D	165
5.10	Development of roots on nodal segment explants excised from nursery raised germinated seedlings of <i>S. parvifolia</i> , when the explant were transferred WPM medium supplemented with 10^{-4} M 2, 4-D to WPM medium containing 10^{-7} M 2, 4-D	166
5.11	Development of roots on nodal segment explants excised from nursery raised germinated seedlings of <i>S. parvifolia</i> , when the explant were transferred WPM medium supplemented with 10^{-4} M 2, 4-D to WPM medium	
	containing 10 ⁻⁷ M 2, 4-D	166

