



UNIVERSITI PUTRA MALAYSIA

**EFFECT OF ENDOSULFAN AND CARBOFURAN ON ENZYME
ACTIVITIES IN CLARIAS GARIEPINUS (AFRICAN CATFISH)**

DARYANI

FSAS 2003 11



**EFFECT OF ENDOSULFAN AND CARBOFURAN ON ENZYME
ACTIVITIES IN *CLARIAS GARIEPINUS* (AFRICAN CATFISH)**

By

DARYANI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Partial Fulfillment of the Requirements for the Degree of Master of Science**

March 2003



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirements for the degree of Master of Science

EFFECT OF ENDOSULFAN AND CARBOFURAN ON ENZYME ACTIVITIES IN *CLARIAS GARIEPINUS* (AFRICAN CATFISH)

BY

DARYANI

March 2003

Chairperson : Assoc. Prof. Dr. Mohd Arif Syed

Faculty : Science and Environmental Studies

Determination of lethal concentration (LC₅₀) of carbofuran and the effect of a sub lethal concentration of endosulfan and carbofuran on glutathione S-transferase (GST), glutathione peroxidase (GPx) and ethoxyresorufin O-deethylase (EROD) activities were investigated in the liver and intestine (proximal, medial and distal) of *Clarias gariepinus* (African catfish).

The 96-h LC₅₀ for carbofuran in present study was 1.63 mg/l. Endosulfan and carbofuran treatment resulted in a significant increase ($p < 0.05$) of EROD, GST and GPx in liver and intestine (proximal, medial and distal) at the sub-lethal concentrations of 10.8 µg/l and 0.82 mg/l, respectively. Endosulfan enhanced GST, GPx and EROD enzymes activity in the liver and intestine (proximal, medial and distal) of treated fish. The fish were examined every four days for twenty days. The results showed that all the enzyme activities were maximum after day four exposure and decreased until the termination of the experiment. The activity of GST in all of the organs of treated fish exposed to carbofuran decreased linearly after 2 hours



exposure until day 20. However GPx activity in all of the organs of treated fish exposed to carbofuran decreased after day 4 followed by an increase up to day 16 and decreased further until the termination of the experiment. The EROD activity in all of the organs exposed to carbofuran has a similar trend with that when exposed to endosulfan. Both activities, exposed to carbofuran and endosulfan increased until day 4 followed by a gradual decrease until day 20. In general, GST and GPx activity in all parts of the intestine of *Clarias gariepinus* (African catfish) is significantly higher than those in the liver. Compared to GST and GPx, EROD activity in the liver showed significantly higher activity than that found in the intestinal organs.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk mendapatkan ijazah Master Sains

**KESAN ENDOSULFAN DAN CARBOFURAN TERHADAP AKTIVITI
ENZYME *CLARIAS GARIEPINUS* (IKAN KELI AFRIKA)**

Oleh

DARYANI

Mac 2003

Pengerusi : Prof. Madya Dr. Mohd. Arif Syed

Fakulti : Sains dan Pengajian Alam Sekitar

Pengenalpastian ketoksikan oral akut (LC₅₀) carbofuran dan kesan kepekatan endosulfan dan carbofuran terhadap aktiviti enzim glutathione S-transferase (GST), glutathione peroxidase (GPx) dan ethoxyresorufin O-deethylase (EROD) telah dikaji didalam organ hati dan bahagian usus (proximal, medial dan distal) *Clarias gariepinus* (ikan keli Afrika).

Kesan pendedahan selepas 96 jam oleh carbofuran ialah 1.63 mg/l. Keputusan bagi rawatan endosulfan dan carbofuran jelas menunjukkan peningkatan aktiviti ($p < 0.05$) bagi GST, GPx dan EROD dalam hati dan bahagian usus (proximal, medial dan distal) pada kepekatan sublethal 10.8 µg/l and 0.82 mg/l secara berturutan. Endosulfan telah merangsangkan peningkatan aktiviti enzim GST, GPx dan EROD di dalam hati dan bahagian usus (proximal, medial dan distal) pada ikan yang dirawat. Aktiviti enzim pada ikan diuji setiap 4 hari selama 20 hari. Keputusan menunjukkan aktiviti enzim mencapai tahap maksimum selepas 4 hari pendedahan

dan semakin menurun sehingga tamat tempoh pendedahan. Aktiviti GST di dalam kesemua organ pada ikan yang dirawat dengan carbofuran menunjukkan penurunan garis lurus selepas pendedahan 2 jam sehingga hari ke-20. Aktiviti GPx di dalam semua organ pada ikan yang telah dirawat dengan carbofuran menunjukkan penurunan selepas 4 hari diikuti peningkatan sehingga hari ke-16 dan penurunan semula sehingga tamat tempoh pendedahan. Aktiviti EROD di dalam organ yang telah dirawat dengan carbofuran mempunyai keluk yang sama dengan endosulfan. Kedua-dua aktiviti terhadap rawatan dengan carbofuran dan endosulfan meningkat sehingga hari ke-4 dan diikuti penurunan secara perlahan sehingga hari ke-20. Secara umumnya aktiviti GST dan GPx di dalam kesemua bahagian usus ikan *Clarias gariepinus* (keli Africa), adalah lebih tinggi dari hati. Manakala aktiviti EROD dalam hati menunjukkan lebih tinggi berbanding di dalam bahagian organ usus.

ACKNOWLEDGEMENTS

I would like to express my great appreciation and sincere gratitude to Associate Professor Dr. Mohd. Arif Syed, for their sponsoring and supervision, exactly their help made my dream of pursuing higher education come true. I am indebted to my co-supervisor Associate Professor Dr. Nor Aripin Shamaan, Associate Professor Dr. Juzu Hayati Arshad and Associate Professor Dr. Johari Ramli, for his patience and valuable guidance and advise throughout this study.

I would like to thank to Miss Suryani, Miss Sukiran, Miss Fazilah, Mrs. Neni, Mrs. Paricher and all my lab-mates for their tremendous assistance and encouragement during the whole study period.

My appreciation goes to my closest friend Gamma and Ninit for the amity, patience and encouragement and to all my country-mates Harini Boekhari, Lailan Saufina, Mayastri, Penni Ismiati, Rini, Faiq and Radniyah Zakaria for the kindly cares, helps and their religious guidance and to all my college-mates Ernawati, Neila, Mirna, Sifa, Tesa, Ayu and Maesaroh who always be beside me whenever I needed them.

I also to acknowledge Bimo, Eni, Hena, Emi, Arif Sugandi, Arief Fadhilah, Hari Priyadi, Ediyatno, Pak Taufik, Pak Bambang, Dewi yanti, Melissa, Dian Permata, Meuthia naim, Irene Pardede and Eva Zulfiah, for their friendship, enthusiastic support and companionship.



I thank the technical staff of the Department of Biochemistry and Microbiology, UPM; Mohd Zamros, Ibrahim, Khalid, Onn and Husain and technical staff of the Aquatic Resources Laboratory of Institute of Bioscience, UPM; Zasni, Zaidi, Mohammad and Krisnan.

Finally, but not least, I most greatly appreciate my late mother, my father my brothers and sister, my aunt, my uncle and my cousins; and my fiance for their support and encouragement.



I certify that an Examination Committee met on 5th March 2003 to conduct the final examination of Daryani on her Master of Science thesis entitled “ Effect of Endosulfan and Carbofuran on Enzyme Activities in *Clarias gariepinus* (African Catfish)” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulation 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination committee are as follows:

Radzali Muse. Ph.D.

Associate Professor,
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Chairman)

Mohd. Arif Syed. Ph.D.

Associate Professor,
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Member)

Nor Aripin Shamaan. Ph.D.

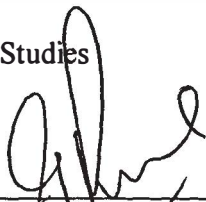
Associate Professor,
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Member)

Juzu Hayati Arshad. Ph.D.

Associate Professor,
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Member)

Johari Ramli. Ph.D.

Associate Professor,
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Member)



GULAM RUSUL RAHMAT ALI, Ph.D.

Professor/Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 29 MAR 2003



This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee are as follows:

Mohd. Arif Syed. Ph.D.

Associate Professor,
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Chairman)

Nor Aripin Shamaan. Ph.D.

Associate Professor,
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Member)

Juzu Hayati Arshad. Ph.D.

Associate Professor,
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Member)

Johari Ramli. Ph.D.

Associate Professor,
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Member)



AINI IDERIS, Ph.D.
Professor/Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 8 MAY 2003



DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations, which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.



DARYANI

Date: 17 Mac 2003

TABLE OF CONTENTS

	Page
ABSTRACT	ii
ABSTRAK	iv
ACKNOWLEDGEMENT	vi
APPROVAL	viii
DECLARATION	x
LIST OF TABLES	xiv
LIST OF FIGURES	xv
LIST OF ABBREVIATIONS	xvii
CHAPTER	
I INTRODUCTION	1
II LITERATURE REVIEW	5
Pesticides contamination in the environment in Malaysia	6
Endosulfan	7
Carbofuran	11
Molecular and Biochemical Indicators	13
Cytochrome P450 (CYP450)	17
Cytochrome P450 in fish	18
Glutathione S-Transferase (GST)	20
Glutathione Peroxidase (GPx)	23
Target Organ Toxicity	26
Liver	27
Intestine	30
III METHODOLOGY	35
Chemicals	35
Acclimatisation and Treatment of the Test Organism	35
Pesticides Administration	37
Determination of LC ₅₀ of Carbofuran	37
Preparation of Cytosolic and Microsomes	38
The Glutathione S-Transferase (GST) Assay	40
The Glutathione Peroxidase (GPx) Assay	41
The 7-Ethoxyresorufin O-Deethylase (EROD) Assay	42
Protein Determination	43



	Statistical Analysis	43
IV	RESULTS	
	Determination of the LC ₅₀ value of Carbofuran	45
	Effect of Endosulfan	48
	GST, GPx and EROD Activity in Fish Exposed to Endosulfan	47
	GST Activity Levels in Fish Exposed to Endosulfan	47
	GPx Activity in Fish Exposed to Endosulfan	53
	EROD Activity in Fish Exposed to Endosulfan	56
	Effect of Carbofuran	62
	GST, GPx and EROD Activity in Fish Exposed to Carbofuran	62
	GST Activity in Fish Exposed to Carbofuran	62
	GPx Activity in Fish Exposed to Carbofuran	67
	EROD Activity in Fish Exposed to Carbofuran	71
	Comparison of Enzyme Activity in Fish Exposed to Endosulfan versus Carbofuran	75
	Comparison of GST Activity in Fish Exposed to Endosulfan versus Carbofuran	75
	Comparison of GPx Activity in Fish Exposed to Endosulfan versus Carbofuran	81
	Comparison of EROD Activity in Fish Exposed to Endosulfan versus Carbofuran	87
V	DISCUSSION	93
	Determination of the LC ₅₀ value of Carbofuran	93
	Effect of Endosulfan on GST, GPx and EROD activities	96
	Effect of Endosulfan on GST Activity	98
	Effect of Endosulfan on GPx Activity	102
	Effect of Endosulfan on EROD activity	103
	Effect of Carbofuran on GST, GPx and EROD Activities	104
	Effect of Carbofuran on GST Activity	104
	Effect of Carbofuran on GPx Activity	105
	Effect of Carbofuran on EROD Activity	106
	Comparison of Enzyme Activity in Fish Exposed to Endosulfan versus Carbofuran	108
	Comparison of GST Activity in Fish Exposed to Endosulfan versus Carbofuran	108
	Comparison of GPx Activity in Fish Exposed to Endosulfan versus Carbofuran	109



	Comparison of EROD Activity in Fish Exposed to Endosulfan versus Carbofuran	111
VI	CONCLUSION	112
	BIBLIOGRAPHY	115
	APPENDICES	128
	VITA	133



LIST OF TABLE

Table		Page
1	Properties of xenobiotic-metabolizing enzymes in the uninduced catfish.	28
2	Cells of control teleost liver and associated markers.	31
3	Major gastrointestinal functions.	32
4	Percentage of dead fish, empirical probit and approximately expected probits for estimation of LC ₅₀ of carbofuran for <i>C.gariepinus</i> .	45
5	GST activities in liver and intestine (proximal,medial and distal) of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan.	50
6	GPx activities in liver and intestine (proximal, medial and distal) of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan.	54
7	EROD activities in liver and intestine (proximal, medial and distal) of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan.	60
8	GST activities in liver and intestine (proximal, medial and distal) of <i>Clarias gariepinus</i> (African catfish) exposed to carbofuran.	64
9	GPx activities in liver and intestine (proximal, medial and distal) of <i>Clarias gariepinus</i> (African catfish) exposed to carbofuran.	69
10	EROD activities in liver and intestine (proximal, medial and distal) of <i>Clarias gariepinus</i> (African catfish) exposed to carbofuran.	73



LIST OF FIGURE

Figure		Page
1	A chemical formula of endosulfan.	8
2	A chemical formula of carbofuran.	11
3	Integration of histopathologic biomarkers with physiologic and biochemical approaches.	16
4	Hypothesized induction mechanism of CYP1A, commonly referred to as the Ah-receptor theory.	20
5	Thioether formation with substrate CDNB	22
6	peroxidative destruction of a polyunsaturated lipid initiated by a free radical attack such as by the hydroxyl radical (OH^\bullet).	25
7	Lateral view of the internal anatomy of the channel catfish	33
8	Summary diagram of the methods used to prepare samples for measuring the specific activities of marker enzymes.	39
9	Determination of the LC_{50} value of carbofuran for <i>C. gariepinus</i> .	46
10	GST activities in liver (A) and intestine (proximal (B), medial (C) and distal (D)) of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan.	49
11	GPx activities in liver (A) and intestine (proximal (B), medial (C) and distal (D)) of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan.	54
12	EROD activities in liver (A) and intestine (proximal (B), medial (C) and distal (D)) of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan.	59
13	GST activities in liver (A) and intestine (proximal (B), medial (C) and distal (D)) of <i>Clarias gariepinus</i> (African catfish) exposed to carbofuran.	63
14	GPx activities in liver (A) and intestine (proximal (B), medial (C) and distal (D)) of <i>Clarias gariepinus</i> (African catfish) exposed to carbofuran.	68

15	EROD activities in liver (A) and intestine (proximal (B), medial (C) and distal (D)) of <i>Clarias gariepinus</i> (African catfish) exposed to carbofuran.	72
16	GST activity in liver of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan and carbofuran.	77
17	GST activity in proximal intestine of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan and carbofuran.	78
18	GST activity in medial intestine of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan and carbofuran.	79
19	GST activity in distal intestine of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan and carbofuran.	80
20	GPx activity in liver <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan and carbofuran.	81
21	GPx activity in proximal intestine of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan and carbofuran.	84
22	GPx activity of in medial intestine of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan and carbofuran.	85
23	GPx activity in distal intestine of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan and carbofuran.	86
24	EROD activity in liver of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan and carbofuran.	89
25	EROD activity in proximal intestine of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan and carbofuran.	90
26	EROD activity in medial intestine of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan and carbofuran.	91
27	EROD activity in distal intestine of <i>Clarias gariepinus</i> (African catfish) exposed to endosulfan and carbofuran.	92
28	Model of reaction mechanism of GST exposure to endosulfan.	101



LIST OF ABBREVIATIONS

AChE	Acetylcholinesterase
AHH	Aryl hydrocarbon hydroxylase
AhR	Aryl hydrocarbon receptor
ARNT	Aryl hydrocarbon nuclear transferase
BSA	Bovine Serum Albumin
BaP	Benzo(a)pyrene
BPB	Bromophenol blue
CDNB	1-chloro-2,4-dinitrobenzene
CO	Carbon monoxide
CYP	Cytochrome P450
DCNB	1,2-Dichloro-4-nitrobenzene
DEN	Diethylnitrosamine
DNA	Deoxyribonucleic acid
e.u.	Enzyme per unit
EA	Ethacrynic acid
EDTA	Ethylenediaminetetra acetic acid
ER	Endoplasmic reticulum
EROD	7-ethoxyresorufin O-deethylase
g	gram
GSH	Glutathione (reduced form)
GSSG	Glutathione disulphate (oxidised)
GPx	Glutathione peroxidase
GST	Glutathione S-transferase
HCl	Hydrochloric acid
HSP90	Heat-shock protein 90
H ₃ PO ₄	Orthophosphoric acid
H ₂ O ₂	Hydrogen peroxide
K ₂ HPO ₄	dipotassium hydrogen phosphate
KH ₂ PO ₄	Potassium dihydrogen phosphate
KCl	Potassium chloride
L	liter
MACA	Malaysian Agriculture Chemical Association
MARDI	Malaysian Agricultural Research and Development Institute
mg	Milligram
mL	Millilitre
mM	Millimolar
MO	Monooxygenase
mRNA	messenger ribonucleic acid



Mw	Molecular weight
M	Molar
NADH	Reduced nicotinamide adenine dinucleotide
NaN ₃	Sodium azide
NaOH	Sodium hydroxide
PAH	Poly aromatic hydrocarbon
PB	Phenobarbital
PCbs	Polychlorinated biphenyls
ppb	Part per billion
ppm	part per million
SDS	Sodium dodecyl sulphate
TCDD	2,3,7,8-tetra chlorodibenzo-p-dioxin
Tris	Tris (hydroxymethyl) aminomethane
U	Unit
UV	Ultra violet
µg	Microgram
µl	Microliter
v/v	Volume per volume
w/v	Weight per volume
w/w	Weight per weight
XRE	Xenobiotic regulatory element



CHAPTER I

INTRODUCTION

Agricultural production of both arable crops and livestock is an activity of primary importance to developing countries. The growing world demand for food due to the increased population had led to greatly increased crop and animal production in these countries. It has led farmers to use sophisticated agricultural technology in which pesticides play a critical role. Residual levels of pesticides can constitute, in some cases, an important source of pollution, mainly if they have been used for a long time. Their mobility through air and water, their accumulation and transformation in the environment and finally, their biomagnification, constitute a real risk to human health, wildlife and the environment (Ruzafa et al., 2000). Pesticides can enter the aquatic environment by a number of routes including spillages, inappropriate disposal of diluted pesticides and runoff into drains and rivers. In the aquatic ecosystem, there is a continuous interchange of pesticides between land, sediment, sediment-water interface, interstitial waters and air-water interstitial and aquatic organisms. Data from various countries, through directed research on water-quality monitoring systems and accident reports, indicate that incidents of pollution in the aquatic environment by pesticides are frequent (OECD, 1986).

Utilization of biochemical factors to evaluate responses to pollutants, especially to carcinogenic compounds, such as PAHs and PCBs, has increased



considerably over the past 15 years. Alteration in biochemical systems are often more sensitive indicators than those at higher levels of biological organization. Depending upon the function of the systems affected and the nature of the responses, biochemical perturbations can indicate whether additional effects (at the organ or individual level) are likely to occur (Stegeman, et al., 1992). Relationships between exposure to chemical contamination and alterations in several biochemical processes in fish may allow the use of the latter as biomarkers of exposure and early responses to chemical contaminants (Stien et al., 1998).

The fish can provide excellent models to study the *in vivo* and *in vitro* effects of mutagens and carcinogens in the natural environment. It has been reported that feral population of fishes which exhibit extraordinary high prevalence of hepatic tumours, preneoplastic lesions and nonneoplastic hepatic lesions, have been identified at highly urban and industrialized sites in North America (Harshbarger and Clark, 1990). Freshwater catfish from industrialized tributaries of the Great Lakes exhibited hepatic tumours frequencies that ranged from 28-44% and frequent occurrences of epidermal papillomas were reported in the catfish as well as white suckers *Catostomus commersoni* from same geographic area (Wirgin and Waldman, 1998). A marine flatfish species at highly contaminated sites in Puget Sound, WA, showed hepatic tumour levels that ranged from 15 to 30% (Malins et al, 1988).

A number of such biomarkers was used in the present study. Cytochrome P450-dependent mono-oxygenase enzymes (CYPIA), conveniently measured as 7-ethoxyresorufin O-deethylase (EROD) and glutathione S-transferase (GST), are two

biomarkers that have been used extensively as they are induced by poly aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs) and certain dioxins (Stegemen, 1981; Stegemen and Lech, 1991; Stegemen et al., 1992). Enhanced levels of antioxidant biomarkers, e.g. glutathione peroxidase (GPx) and the associated reduced glutathione (GSH), function to remove oxyradicals derived from exposure to organic xenobiotics, including aromatic diols and quinines, nitroaromatics, bipyridyl herbicides and metal (Stegemen et al., 1992). Exposure to PAHs, PCBs, chlorinated hydrocarbons and metals have been shown to correlate with the increased activity of antioxidants in fish (Livingstone, 2001).

Cytochrome-P450 dependent monooxygenases (MO) system participates in the phase I detoxification of exogenous compounds; 7-ethoxyresorufin is a model substrate for the cytochrome-P450IA dependent MO action, called 7-ethoxyresorufin O-deethylase (EROD), which can be specifically induced by compounds with relatively flat molecular structures, such as planar polychlorinated biphenyls (PCBs) and polycyclic aromatic hydrocarbons (PAHs). EROD activity in several fish species has been shown to be a good indicator of exposure to organic micro pollutants, because of the strong correlation observed with doses of toxicants both in laboratory and field studies (Eggens et al., 1995). CYP1A1 expression is transcriptionally regulated in fish and has been quantified in biomarker. Studies at the mRNA, CYP1A protein and associated catalytic activities of EROD and AHH levels. The mechanisms of CYP1A transcription in fish are probably very similar to those in mammals (Wirgin and Waldman, 1998).



This study is undertaken to study the toxicity of endosulfan and carbofuran on *Clarias gariepinus* and the effect of sub lethal concentrations (LC₅₀) of the carbofuran and endosulfan on the activities of glutathione S-transferase (GST), glutathione peroxidase (GPx) and 7-ethoxyresorufin O-deethylase (EROD). In this study, *Clarias gariepinus* (African catfish) a tropical freshwater fish was chosen. The choice of the fish in this study was based on its wide distribution in tropical and sub-tropical areas and availability of the cultures species which is never being exposed to any pesticide contamination. This study was expected to determine aquatic environment pollution level of pesticides using suitable biomarker. It was also expected that the pollution level could be reduced and the human health risk exposed by this pollution could also be mitigated.

CHAPTER II

LITERATURE REVIEW

Growing demand for food as a result of increased human population has led to a substantial increase in the production and use of agro-chemicals, such as pesticides and fertilizers. The increased use of various types of pesticides has led to concerns regarding the potential for contamination of environmental media (i.e., water, sediment, and biota) and the associated effects on wildlife and human health.

Approximately 1000 pesticide formulations are used throughout the world today for virtually all forms of agricultural commodity. The annual worldwide agricultural use of pesticides has been estimated to be in the order of 5 million tons, of which about 70 per cent is used for agriculture and the remainder by public health agencies and government agencies for vector control and for domestic purposes (Shaw and Chadwick, 1998). Despite the use of pesticides, about 35 per cent of the crops are lost. Nearly 50 per cent of food in the world may be lost annually despite all pest control procedures. It has been estimated that 0.1 per cent of the pesticides applied to crops reach the target pests; thus, more than 99 per cent of the applied pesticides have the potential to affect non- target organisms and to become widely dispersed in the environment. Although not yet well documented, pesticide-induced chronic toxicity is emerging as a public health concern including cancer, reproductive impairment and irreversible neurotoxicity (Shaw and Chadwick, 1998).



Pesticides Contamination in the Environment in Malaysia

In Malaysia, the usage of pesticides is extensive and the number of pesticides which are available on the market is large (Balasubramaniam and Yunus, 1974). Currently there are over 300 retail outlets involved in the manufacture, formulation and packaging of pesticides (MACA, 1989). In 1980, sale of pesticides amounted to RM 160 million. This increased to RM 237 million in 1984 and to nearly RM 300 million in 1988 (MACA, 1989). In Malaysia, pesticides are predominantly used in the plantation industries (rubber and oil palm), vegetable growing and public health control and in rice cultivation. The major insecticides used are γ -BHC, endosulfan, chlorpyrifos, carbofuran and carbaryl in rice field (Ooi, 1974), while malathion, resmethrin and abate are used for public health control (Yap, 1974).

Some herbicides have been shown to be toxic to paddy field fish in Malaysia (Ooi and Lo, 1990). Paddy field fish forms an important protein source for the rural people in west Malaysia (Gill, 1980). The estimated per capita consumption of fish in Malaysia was at 30 kg per annum in 1986. This consumption rate is higher than in 1974 where the value was at 26 kg per annum. Given this estimate, fish production has to increase substantially in order to meet the national requirement (Omar et al., 1990). Widespread use of pesticides in paddy fields has resulted in the decline of paddy field fish production. Paddy field fish production in Krian, Perak has been reported to decline over the past few years because of a pesticide (Ooi and Lo, 1990). It was also reported that fish killed in the Muda river was caused by the contamination of the river water with pesticide residues from effluents of pesticides