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ENZYMATIC HYDROLYSIS OF SAGO STARCH FOR THE PRODUCTION OF MALTODEXTRINS

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ENZYMATIC HYDROLYSIS OF SAGO STARCH FOR THE PRODUCTION OF MALTODEXTRINS

By CH'NG YAN ENG

Thesis Submitted in Fulfilment of the Requirements for the Degree of Master of Science in the Faculty of Food Science and Biotechnology,
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Specially for.....

My respected parents,

Ong,

sisters Daisy, Judy and Janet

Prof. Mahyuddin and Dr. Nasir



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TABLE OF CONTENTS

]	Page
ACI	KNOWLEDGEMENTS			iii
LIST	T OF TABLES			ix
LIST	T OF FIGURES			Х
LIS	T OF ABBREVIATIONS			xiii
ABS	STRACT			xiv
ABS	STRAK			xvii
СНА	APTER			
1	INTRODUCTION	٠	. ,	1
2	LITERATURE REVIEW			4
	Maltodextrins			4
	Manufacturing Process			5
	Effect of Processing Parameters			14
	Type of Starches			24
	Type of Maltodextrins			28
	Characteristics and Properties			29
	Sago starch			37
	Production and Processing			38
	Quality of Sago Starch			39
	Chemistry			42
	Utilisation of Sago Starch			42



3	TO LOW DE SYRUPS	44
	Materials and methods	46
	Quality of Different Bags of Sago Starch	46
	Measurement of Starch Damage	48
	Enzymatic Susceptibility of Sago Starch	50
	Measurement of Reducing Sugars	51
	Measurement of Total Sugars	52
	Scanning Electron Microscopy (SEM)	52
	Effect of Enzymes on Brabender Viscosity Profile of Sago Starch	53
	Effect of Sago Quality on Hydrolysis to Low Dextrose Equivalent (DE) Syrup	53
	Presence of Alpha-amylase Inhibitors	54
	Results and Discussion	55
4	PRODUCTION OF MALTODEXTRINS	89
	Materials and Methods	91
	Quality of Sago Starch	91
	Enzymatic Hydrolysis of Sago Starch to Maltodextrins	91
	Effect of Starch Concentration	92
	Effect of pH on Hydrolysis	92
	Effect of Alpha-amylases and Their Dosage on Hydrolysis	92
	Refining and Clarification	93
	Quality of Maltodextrins Produced	93
	Results and Discussion	94



5	CHARACTERISTICS OF MALTODEXTRINS FROM SAGO STARCH	112
	Materials and Methods	115
	Analysis of Oligosaccharide Profile by HPLC	115
	Solubility of Maltodextrins	117
	Viscosity of Maltodextrin Solutions	117
	Results and Discussion	118
6	SUMMARY AND CONCLUSIONS	130
	Summary	130
	Conclusions	132
BIBLIOGRAPHY		
BIOC	GRAPHICAL SKETCH	144



LIST OF TABLES

Table		Page
1	Enzyme Products for Starch Hydrolysis: Alpha-amylase Products for Starch Liquefaction	. 17
2	Impurity Levels in Native Granular Starches	. 27
3	Typical Oligosaccharide Profile of Several Commercial Maltodextrins	. 33
4	Viscosity of Maltodextrin Solutions	. 36
5	Solubility of Maltodextrins	. 36
6	Quality of Sago Starch	. 56
7	Flow Time of 200 ml Sago Starch Hydrolysate	. 85
8	Comparison of Oligosaccharide Profiles of Maltodextrins	124



LIST OF FIGURES

Figure	Pa	ge
1	A Schematic Diagram of Stages of Maltodextrin Production	6
2	A Single Stage Acid or Enzyme Hydrolysis of Starch to Maltodextrins	9
3	A Dual Stage Acid-Enzyme or Enzyme-Enzyme Hydrolysis of Starch to Maltodextrins	12
4	Functional Properties of Maltodextrins in Relation to its DE Vaule	30
5	Modern Sago Starch Extraction Process	10
6	Traditional Sago Starch Extraction Process	10
7	Peak Gelatinisation Viscosity of Sago Starch from Different Bags	50
8	Degree of Starch Damage in Sago Starch from Different Bags	52
9	Effect of Degree of Starch Damage on Peak Gelatinisation Viscosity of Sago Starch	54
10	Susceptibility of Sago Starch from Different Bags to Alpha-amylase Termamyl 120L	56
11	Susceptibility of Sago Starch to Alpha-amylase Termamyl 120L	57
12	Susceptibility of Sago Starch from Different Bags to Alpha-amylase BAN 240L	5 9
13	Susceptibility of Sago Starch to	70



14	Effect of Viscozyme 120L on Hydrolysis of Sago Starch by Alpha-amylase Termamyl 120L
15	Effect of Viscozyme 120L on Hydrolysis of Sago Starch by Alpha-amylase BAN 240L
16	Susceptibility of Gelatinised Sago Starch to Termamyl 120L, BAN 240L and Viscozyme 120L
17	SEM of Sago Starch
18	Effect of Viscozyme 120L and/or Termamyl 120L on Peak Gelatinisation Viscosity of Sago Starch
19	Effect of Viscozyme 120L and/or BAN 240L on Peak Gelatinisation Viscosity of Sago Starch
20	Effect of Sago Starch of Different Quality on DE Values of Hydrolysates During Hydrolysis with Termamyl 120L 81
21	Effect of Starch Damage on DE Values of Sago Starch Hydrolysate 83
22	Effect of Viscozyme 120L and/or Termamyl 120L on DE Values of Sago Starch Hydrolysate
23	Effect of Sago Starch Concentration on DE Values of Sago Starch Hydrolysate (produced using 0.05% w/w dsb of Termamyl 120L)
24	Effect of Sago Starch Concentration on DE Values of Sago Starch Hydrolysate (produced using 0.10% w/w dsb of Termamyl 120L)
25	Effect of Processing pH on DE Values of Sago Starch Hydrolysate
26	Effect of Different Dosages of Termamyl 120L on DE Values of Sago Starch Hydrolysate



27	Effect of Different Dosages of BAN 240L on DE Values of Sago Starch Hydrolysate	106
28	Low Molecular Weight (DP 1-7) Saccharides Profile of DE 10.0 and DE 19.4 Maltodextrins (produced using Termamyl 120L)	119
29	Low Molecular Weight (DP 1-7) Saccharides Profile of DE 10.2 and DE 20.3 Maltodextrins (produced using BAN 240L)	120
30	Viscosity of Maltodextrin Solutions at Various Solids Concentrations	126
31	Effect of Temperature on Viscosity of Maltodextrin Solutions (40 % solids)	128



LIST OF ABBREVIATIONS

BU Brabender unit

°C degree Centigrade

DE Dextrose Equivalent

DP Degree of Polymerisation

dsb dry starch basis

gm gram

hr hour

M Molar

min minute(s)

mg milligram

ml millimeter

nm nanometer

ppm parts per million

u micron

ul microliter

um micrometer

w/v weight per volume

w/w weight per weight

< less than

% percent



Abstract of the Thesis Presented to the Senate of Universiti Pertanian Malaysia in Fulfilment of the Requirements for the Degree of Master of Science.

ENZYMATIC HYDROLYSIS OF SAGO STARCH FOR THE PRODUCTION OF MALTODEXTRINS

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Maltodextrin is a partial starch hydrolysis product used widely in food. It has been produced from corn, tapioca and potato starches. This project was designed to study the production of maltodextrin from refined sago starch and to characterise the maltodextrins produced. The refined sago starch used was obtained from different sago factories and labelled as Bag A, B, C, D, E, F, G, H and I. Studies carried out showed that there were variations in the quality of the starch of different bags. The moisture content of the starch varied from 10.05 to 15.40% (w/w) while the ash and crude fibre contents were 0.110 to 0.930 % (w/w) and 0.040 to 0.560% (w/w) respectively. The sago starch showed greyish colour with Hunter Lab L values of 87.70 to 90.17 and pH values of 4.77 to 7.58. Particle size analysis showed that more than 99.3% (w/w) of the starch passed through the 125 um pore size sieve. The peak gelatinisation viscosity of the starch varied between 458 to 680 BU. The sago starch showed a common pasting temperature of 74°C and an



extremely low level of protein of less than 0.010% w/w. The measurement of the degree of starch damage showed that sago starch of Bags D, E, F and G were undamaged, while starch of the remaining bags had damage of up to 8% w/w. The undamaged sago starch was of two types: the high, and the low viscosity starch with peak gelatinisation viscosities of about 680 BU and 485 BU respectively. Enzymatic susceptibility studies showed that undamaged sago starch was relatively unsusceptible to alpha-amylases Termamyl 120L and BAN 240L. The degrees of hydrolysis achieved after prolonged incubation for 12 hours at 60°C were 8.6 to 12.1% w/w, and 1.8 to 2.3% w/w respectively. The raw damaged and the gelatinised undamaged sago starch had increased susceptibilities to these alpha-amylases. The degrees of hydrolysis were 17.5 to 22.1% w/w, and about 3.0% w/w respectively, for damaged starch, and 88.3% and 76.5% (w/w) respectively, for the gelatinised starch. Viscozyme 120L when used alone (at a dosage of 0.05% w/w) or in combination with either Termamyl 120L or BAN 240L (at a dosage of 0.10% w/w) had the ability of reducing the viscosity of gelatinised sago starch. It was unable to hydrolyse either the raw or gelatinised starch. Damaged sago starch when used for enzymatic hydrolysis (by Termamyl 120L) produced unstable low DE syrups which retrograded on cooling. Sago starch with high crude fibre content produced low DE syrups that were difficult to filter. Both Termamyl 120L and **BAN** 240L found to be suitable for the production of maltodextrin from sago starch. A starch concentration of 20% (w/v) was suitable for hydrolysis at pH 5.5 to pH 6.5. Optimum Termamyl 120L dosages were 0.08 to 0.10% (w/w) for the production of DE 6 to 20 maltodextrins. A dosage of 0.04% (w/w) of BAN 240L was suitable for



producing DE 8 to 11 maltodextrins while 0.06% (w/w) and 0.08% (w/w) of the amylase were suitable for producing DE 12 to 15, and DE 16 to 20 maltodextrins respectively. Maltodextrins with similar DE value derived from Termamyl 120L and BAN 240L had different oligosaccharide profiles. The DE 10.0 maltodextrin (from Termamyl 120L hydrolysis) had predominant amount of maltoheptose, maltopentose and maltotriose with very low level of maltohexose. The DE 10.2 maltodextrin (from BAN 240L hydrolysis) had prominent amount of maltopentose and maltoheptose and a higher level of maltohexose than the aforementioned DE 10.0 maltodextrin. Maltodextrins produced from sago starch by using Termamyl 120L had much superior solution stability and solubility characteristics than those produced by using BAN 240L. The viscosity of solutions of maltodextrins produced were comparable to that reported for commercial Maltrin maltodextrins with similar DE value.



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HYDROLYSIS ENZIMATIK KANJI SAGU UNTUK PENGHASILAN MALTODEKSTRIN

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Maltodekstrin adalah produk hasilan hidrolisis separa kanji yang digunakan secara meluas dalam makanan. Ia telah dihasilkan daripada kanji jangung, ubi kayu dan ubi kentang. Projek ini dijalankan untuk mengkaji penghasilan dan pencirian maltodekstrin daripada kanji sagu. Kanji sagu yang digunakan didapati daripada kilang yang berlainan dan dilabelkan sebagai Beg A, B, C, D, E, F, G, H dan I. Kajian yang dijalankan menunjukkan bahawa terdapat perbezaan pada kualiti kanji sagu tersebut. Kandungan airnya adalah di antara 10.05 hingga 15.40% (w/w). Kandungan abu dan fiber kasar dalam kanji sagu ialah masing-masingnya sebanyak 0.110 hingga 0.930% (w/w) dan 0.040 hingga 0.560% (w/w). Kanji sagu didapati berwarna kelabu dan bernilai L Hunter Lab sebanyak 87.70 hingga 90.17 L sementara nilai pH kanji berada di antara 4.77 hingga 7.58. Kajian saiz partikel menunjukkan bahawa lebih daripada 99.3% (w/w) kanji tersebut boleh melepasi penyaring bersaiz lubang sebesar 125 um. Kanji sagu didapati mempunyai kelikatan pengelatinan yang



berbeza-beza di antara 458 hingga 680 BU. Kandungan protinnya adalah sangat rendah sekitar < 0.010% w/w. Kanji sagu ini menunjukkan suhu 'pasting' serupa sebanyak 74°C. Kajian darjah kerosakkan kanji menunjukkan bahawa kanji sagu pada Beg D, E, F and G adalah sempurna tanpa kerosakkan granul, sementara kanji beg-beg lain mempunyai kerosakan setinggi 8% (w/w). Terdapat dua jenis kanji sagu tanpa kerosakkan berdasarkan nilai kelikatan pengelatinan tertingginya: iaitu kanji berkelikatan tinggi sebanyak 680 BU, dan kanji berkelikatan rendah sekadar 485 BU. Kajian enzimatik menunjukkan bahawa kanji sagu tanpa kerosakkan mempunyai kerintangan terhadap hidrolisis oleh alfa-amilase komersial Termamyl 120L dan BAN 240L. Pengeraman selama 12 jam pada 60°C menghasilkan darjah penghadaman sebanyak 8.6 hingga 12.1% (w/w), dan 1.8 hingga 2.3% (w/w). Kanji sagu dengan kerosakkan menunjukkan peningkatan darjah penghadaman sekitar 17.5 hingga 22.1% (w/w), dan 3.0% (w/w) masing-masingnya. Kanji sagu sempurna tergelatin mempunyai darjah penghadaman oleh kedua-dua amilase tersebut sebanyak 88.3% (w/w) dan 76.5% (w/w) masing-masingnya. Viscozyme 120L didapati berupaya menurunkan kelikatan pengelatinan tertinggi kanji sagu bila digunakan secara bersendirian (pada dos 0.05% w/w) atau bersama salah satu amilase tersebut (pada dos 0.10% w/w). Ia tidak berupaya menghidrolisiskan kanji sagu baik mentah mahupun yang tergelatin. Sirup bernilai DE rendah yang dihasilkan daripada kanji sagu yang rosak adalah tidak stabil dan mudah mengalami retrogradasi. Kanji sagu berkandungan fiber kasar yang tinggi menghasilkan sirup bernilai DE rendah yang sukar ditapis. Termamyl 120L dan BAN 240L didapati sesuai untuk penghasilan maltodekstrin dari kanji sagu. Kepekatan sluri kanji sebanyak 20% w/v and pH bernilai 5.5 hingga 6.5 adalah



CHAPTER 1

INTRODUCTION

Maltodextrin is a partial starch hydrolysis product widely used as bulking, bodying and thickening agents, spray drying and encapsulating aids, and fat replacers in many food systems and other applications (Meyer and Everett, 1972; Lloyd and Nelson, 1984; Blanchard, 1992a; Fullbrook, 1984; Alexander, 1992). Many types of maltodextrins have been produced from different starches with varying degree of hydrolysis measured in terms of dextrose equivalent value (DE) and through different manufacturing processes (Alexander, 1992; Katz, 1986; Lloyd and Nelson, 1984; Slott and Madsen, 1975, Coker and Venkatasubramaniam, 1984; Morehouse and Sander, 1987). All these offer a multitude of useful common functional properties. However, due to manufacturing variations, these maltodextrins have unique characteristic differences particularly in their oligosaccharide profiles (Alexander, 1992; Katz, 1986).

Maltodextrins have been produced mainly in America, Mexico, Europe and Australia from starches that represent cheap carbohydrate sources of these regions. These products are therefore mostly derived from corn, potato and tapioca starch (Alexander, 1992; Katz, 1986). Consequently, technical information regarding the manufacturing process and the various processing parameters are based on hydrolysis of these starches (Slott and Madsen, 1975; Coker and Venkatasubramaniam, 1984; Morehouse and Sander, 1987; Morehouse et al., 1972; Armbruster, 1981).



In Malaysia, maltodextrins are being imported for utilisation in the local food industries. Although there are no figures in the literature regarding the amount and type of maltodextrins being imported and consumed, the amount is assumed to be large as can be seen from the quantity of local products produced that contain maltodextrins as an ingredient. To date, there is no factory producing commercial maltodextrins for local utilisation or for export. Therefore, if maltodextrins can be manufactured using one of the local indigenous starches, this will not only save revenue but also diversify the utilisation of such starch and related industries apart from the possibility of exporting the maltodextrins produced. For these reasons, one such possible indigenous starch is sago starch of which Malaysia is the largest producer and exporter (Azudin and Lim, 1991).

The sago starch industry has improved tremendously with the production of better quality sago starch for utilisation in the food and biotechnological industries (Azudin and Lim, 1991; Zulpilip et al., 1991). Various starch hydrolysis products typically produced from corn starch have also been produced from sago starch (Azudin and Lim, 1991; Azudin and Ali, 1991; Zulpilip et al., 1991; Solichien, 1994; Pranamuda et al., 1994; Haska, 1994; Ghani et al., 1994). However, although the starch quality has improved greatly, the main problem of quality variation still persists (Azudin and Lim, 1991). This significantly affects its utilisation in a bioconversion process, like for example the production of maltodextrins where a rather constant starch quality is required (Katz, 1986).



At present there is no report on the suitability of sago starch of various quality for hydrolysis or bioconversion process. More importantly, there is no information available on the manufacture of maltodextrins from sago starch. Therefore, the objectives of this study are:

- 1. To characterise the quality of refined sago starch and to determine its suitability for enzymatic hydrolysis
- 2. To determine the optimum conditions for enzymatic hydrolysis of sago starch to maltodextrins
- 3. To characterise the maltodextrins (DE 10 and DE 19) produced



CHAPTER 2

LITERATURE REVIEW

Maltodextrins

Maltodextrin is defined as non-sweet, nutritive saccharide polymers that consist of D-glucose units linked primarily by alpha-1,4 glucosidic bonds having a dextrose equivalent (DE) value of less than 20. It is prepared as a white powder or concentrated solution by partial hydrolysis of starch with safe and suitable acids and/or enzymes. Maltodextrin is generally recognised as safe as a direct human food ingredient (Blanchard, 1992b; Long, 1985; Fullbrook, 1984; Lloyd and Nelson, 1984; Alexander, 1992; Katz, 1986; FDA CRF 184.1444). Typically, maltodextrins are used as bulking, bodying and thickening agents, spray drying and encapsulating aids, and fat replacers in many food systems and other applications (Meyer and Everett, 1972; Lloyd and Nelson, 1984; Blanchard, 1992b; Fullbrook, 1984; Alexander, 1992).

Although a wide variety of commercial maltodextrins made from corn, waxy corn, tapioca and potato starch is available, the information on acidic and/or enzymatic processes manufacturing them are rather limited (Alexander, 1992; Katz, 1986). Moreover, most of the limited information available regarding the production of these maltodextrins, particularly technical information, is derived from pilot-scale patented processes (Slott and Madsen, 1975; Morehouse and Sander, 1987, Coker and Venkatasubramaniam, 1984). This chapter presents and discusses, in general, various aspects of



maltodextrin, and in particular its enzymatic production, quality and characterisation. It also presents currently available information on sago starch.

Manufacturing Process

The industrial process of making maltodextrin (Figure 1) can basically be divided into five main stages which comprise of, starch gelatinisation, liquefaction, partial dextrinisation and stabilisation, refining and clarification, and finally spray drying (Alexander, 1992; Fullbrook, 1984). Native, ungelatinised starch is insoluble and relatively unsusceptible to hydrolysis (Swinkels, 1985). Gelatinisation, therefore, yields solubilised starch containing amylose and amylopectin molecules. This is thinned and hydrolysed during liquefaction, and dextrinisation stages to yield a stabilised low DE hydrolysate. It is then purified by refining to produce a concentrated maltodextrin solution or spraydried to give a white, dry maltodextrin powder (Alexander, 1992; Lloyd and Nelson, 1984).

Two main hydrolysis modes are utilised for the liquefaction, and dextrinisation stages (Lloyd and Nelson, 1984; Fullbrook, 1984; Blanchard, 1992b). Generally, total acidic hydrolysis previously used yields maltodextrins of poorer quality that haze easily (Lloyd and Nelson, 1984; Fullbrook, 1984; Schenck and Hebeda, 1992). Therefore, enzymatic hydrolysis with or without an initial acid hydrolysis stage is now more commonly employed (Armbruster and Harjes, 1971; Morehouse et al., 1972; Armbruster and Kooi, 1974; Alexander, 1992; Blanchard, 1992b). This mode which uses a thermal stable bacterial endo alpha-amylase has several advantages (Reilly, 1985; Teague



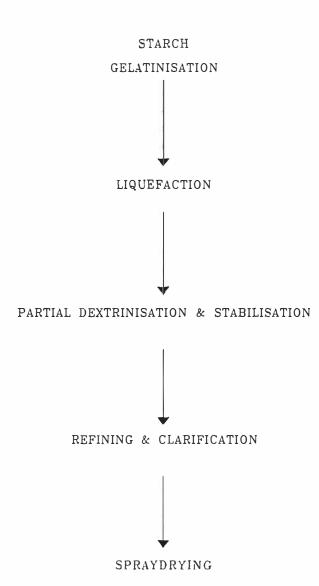


Figure 1. A Schematic Diagram of Stages of Maltodextrin Production

