



UNIVERSITI PUTRA MALAYSIA

**EXTRACTION, CHARACTERIZATION AND STORAGE STABILITY
OF OILS FROM SELECTED PLANT SEEDS**

NYAM KAR LIN

FSTM 2009 25



**EXTRACTION, CHARACTERIZATION AND STORAGE STABILITY
OF OILS FROM SELECTED PLANT SEEDS**

By

NYAM KAR LIN

Thesis Submitted in Fulfilment of the Requirements for the Degree of Doctor of
Philosophy in the Faculty of Food Science and Technology Universiti Putra
Malaysia

November 2009



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Doctor of Philosophy

**EXTRACTION, CHARACTERIZATION AND STORAGE STABILITY
OF OILS FROM SELECTED PLANT SEEDS**

By

NYAM KAR LIN

November 2009

Chairman: Assoc. Prof. Dr. Tan Chin Ping, PhD

Faculty: Food Science and Technology

There is a great demand for renewable sources of raw materials that have nutritional and industrial potential. To meet the increasing demand for vegetable oils, improvements are being made with conventional crops as well as with selected plant species that have the ability to produce unique, desirable fats and oils.

The physicochemical properties and chemical composition of oil extracted from five varieties of plant seeds (bitter melon, Kalahari-melon, kenaf, pumpkin and roselle) were examined by established methods. Most of the quality indices and fatty acid compositions showed significant ($P < 0.05$) variations among the extracted oils. The oils were rich in tocopherols, with γ -tocopherol as the major component in all oil samples. Among the phytosterols, β -sitosterol was the major phytosterol extracted from the five plant-seed oils.



Enzymatic extraction of oil from Kalahari-melon seeds was investigated and evaluated by response surface methodology. Two commercial protease enzyme products were separately used: Neutrase® 0.8 L and Flavourzyme® 1000 L from Novozymes (Bagsvaerd, Denmark). Response surface methodology (RSM) was used to model and optimize the reaction conditions, namely concentration of enzyme (2-5 g/100 g of seed mass), initial pH of mixture (pH 5-9), incubation temperature (40-60 °C), and incubation times (12-36 h). The optimal conditions for Neutrase 0.8 L were enzyme concentration of 2.5 g/100 g, initial pH of 7, temperature at 58°C and incubation time of 31 h, yielding an oil recovery of $68.58 \pm 3.39\%$. The optimal conditions for Flavourzyme 1000 L were: enzyme concentration of 2.1 g/100 g, initial pH of 6, temperature at 50 °C and incubation time of 36 h, yielding a $71.55 \pm 1.28\%$ oil recovery.

The physicochemical properties of oil from Kalahari-melon seed were determined following extraction with petroleum ether and aqueous-enzymatic methods. The free fatty acid, peroxide, iodine and saponification values of the oils extracted using these two methods were found to be significantly ($P < 0.05$) different. No significant ($P > 0.05$) difference was observed between the melting points of the oils obtained from solvent and aqueous-enzymatic extractions. Enzyme-extracted oil tended to be light-colored and more yellow in color, compared with solvent-extracted oil. Fatty acids and phenolic acids in enzyme-extracted oils were comparable to the solvent-extracted oil. The oils extracted with these two methods differed in the composition of their phytosterol and tocopherol contents, but no significant ($P > 0.05$) difference between the two enzyme-extracted oils was observed.

Supercritical carbon dioxide extraction of oil from Kalahari-melon and roselle-seeds were investigated in this study. Response surface methodology (RSM) was used to model and optimize the extraction conditions, namely pressure (200-400 bar), temperature (40-80 °C) and supercritical fluid flow rate (10-20 mL/min). The optimal processing conditions for Kalahari-melon-seed oil recovery and phytosterol concentration were pressure of 300 bar, temperature of 40 °C and supercritical fluid flow rate of 12 mL/min. These optimal conditions yielded a 76.3% oil recovery and 836.5 mg/100 g of phytosterol concentration. The results indicate that the roselle-seed oil recovery was optimal, with a recovery of 102.61% and a phytosterol composition of 727 mg/100 g at the relatively low temperature of 40 °C, a high pressure of 400 bar and at a high supercritical fluid flow rate of 20 mL/min.

Tocopherol-enriched oil from Kalahari-melon and roselle-seeds was extracted by supercritical fluid extraction with carbon dioxide (SFE-CO₂). The optimal SFE-CO₂ conditions for the extraction of tocopherol-enriched oil from Kalahari-melon seeds were extraction pressure of 290 bar, extraction temperature of 58 °C and flow rate of carbon dioxide of 20 mL/min. The optimum conditions for roselle-seeds were extraction pressure of 200 bar, extracting temperature of 80 °C and flow rate of carbon dioxide of 20 mL/min. These optimum conditions yielded a tocopherol concentration of 274.74 and 89.75 mg/100 g oil from Kalahari-seed and roselle-seed, respectively.

During 6 months of storage of Kalahari-melon-seed and roselle-seed oils at both 4 °C and room temperature in the darkness, changes occurred in the

content of fatty acids, phytosterols and tocopherols, and in the presence of primary and secondary oxidative products. These seed oils were obtained from the seeds of Kalahari melon (*Citrullus lanatus*) and roselle (*Hibiscus sabdariffa* Linn.) by supercritical carbon dioxide (SC-CO₂). As expected, statistically significant differences were observed in the content of fatty acids, phytosterols and tocopherols, and in the presence of primary and secondary oxidative products in Kalahari-melon-seed and roselle-seed oils throughout the storage. The quality indices peroxide and anisidine values increased during the 6 months storage time. After storage, degradation parameters may change because of lipid oxidation.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**PENGESTRAKAN, PENCIRIAN DAN KESTABILAN
PENYIMPANAN BAGI MINYAK DARIPADA BIJI BENIH TUMBUHAN
TERTENTU**

Oleh

NYAM KAR LIN

November 2009

Pengerusi: Prof. Madya Dr. Tan Chin Ping, PhD

Fakulti: Sains dan Teknologi Makanan

Sumber bahan mentah boleh dibaharui yang mempunyai potensi pemakanan dan perindustrian amat diperlukan. Untuk memenuhi permintaan yang semakin meningkat terhadap minyak-minyak sayuran, perbaikan telah dilakukan untuk tanaman lazim, begitu juga dengan spesies tumbuhan terpilih yang mempunyai kemampuan untuk menghasilkan lemak serta minyak yang unik dan diingini.

Sifat fiziko-kimia dan komposisi kimia bagi minyak yang diekstrak daripada lima jenis biji benih tumbuhan (peria, tembikai Kalahari, kenaf, labu dan roselle) dikaji dengan menggunakan kaedah yang telah ditetapkan. Kebanyakan indeks kualiti dan komposisi asid lemak menunjukkan variasi yang nyata ($P < 0.05$) antara minyak-minyak yang diekstrakkan. Minyak-minyak yang diekstrak kaya dengan tocoferol di mana γ -tocoferol merupakan komposisi yang

utama dalam minyak-minyak tersebut. β -sitosterol merupakan fitosterol yang utama dalam kelima-lima minyak biji tumbuhan.

Pengekstrakan minyak biji tembikai Kalahari dengan enzim dikaji dan dinilai dengan metodologi tindakbalas permukaan. Dua produk komersial enzim protease telah digunakan secara berasingan iaitu Neutrased[®] 0.8 L and Flavourzyme[®] 1000 L dari Novozymes (Bagsvaerd, Denmark). Metodologi tindakbalas permukaan telah digunakan untuk model dan keadaan reaksi bernama kepekatan enzim (2-5 g/100 g daripada berat biji), pH campuran awal (pH 5-9), suhu penderaman (40-60 °C) dan tempoh penderaman (12-36 h). Keadaan optimum bagi Neutrased[®] 0.8 L ialah kepekatan enzim 2.5 g/100 g , campuran awal pH 7, suhu penderaman 58 °C dan tempoh penderaman 31 jam dengan perolehan minyak sebanyak $68.58 \pm 3.39\%$. Keadaan optimum bagi Flavourzyme[®] 1000 L ialah kepekatan enzim 2.1 g/100 g , campuran awal pH 6, suhu penderaman 50 °C dan tempoh penderaman 36 jam dengan perolehan minyak sebanyak $71.55 \pm 1.28\%$.

Sifat fiziko-kimia minyak biji tembikai Kalahari yang diekstrak dengan kaedah petroleum eter dan enzim berair telah dikaji. Asid lemak bebas, nilai peroksida, iodin dan saponifikasi dalam minyak yang diekstrak dengan menggunakan kaedah-kaedah tersebut didapati berbeza dengan nyata ($P < 0.05$). Takat lebur minyak yang diekstrak dengan kaedah-kaedah tersebut didapati tiada perbezaan yang nyata ($P < 0.05$). Minyak yang diekstrak dengan enzim adalah lebih cerah dan warnanya lebih kuning daripada minyak yang diekstrak dengan pelarut. Asid lemak dan asid fenolik dalam minyak yang diekstrak dengan enzim

adalah setanding dengan minyak yang diekstrak dengan pelarut. Minyak-minyak yang diekstrak dengan dua kaedah ini adalah berbeza dalam kandungan fitosterol dan tocoferol dari segi komposisi, tetapi tiada perbezaan yang nyata dalam kedua-dua minyak yang diekstrak dengan enzim.

Minyak-minyak biji tembikai Kalahari dan roselle yang diekstrak dengan supergending karbon dioksida telah dikaji. Metodologi tindakbalas permukaan telah digunakan dalam model dan keadaan pengekstrakan dioptimumkan bernama tekanan (200-400 bar), suhu 40, 60 dan 80 °C dan aliran cecair supergending 10-20 mL/min. Keadaan proses yang optimum bagi perolehan minyak biji tembikai Kalahari dan kepekatan fitosterol ialah tekanan 300 bar, suhu operasi 40 °C dan aliran cecair supergending 12 mL/min. Keadaan optimum ini dapat memperoleh 76.3% minyak biji tembikai Kalahari dan kepekatan fitosterol 836.5 mg/100 g. Keputusan menunjukkan bahawa perolehan minyak biji roselle adalah optimum dengan 102.61% dengan kehadiran komposisi fitosterol 727 mg/100 g dalam keadaan suhu yang rendah 40 °C, tekanan yang tinggi 400 bar dan aliran cecair supergending yang tinggi 20 mL/min.

Minyak yang kaya dengan tocoferol telah diekstrak dengan pengekstrakkan cecair supergending oleh karbon dioksida dari biji-biji tembikai Kalahari dan roselle. Keadaan optimum bagi pengekstrakkan minyak yang kaya dengan tocoferol dari biji tembikai Kalahari ialah tekanan pengekstrakkan 290 bar, suhu pengekstrakkan 58 °C dan pengaliran karbon dioksida 20 mL/min. Keadaan optimum bagi pengekstrakkan minyak biji roselle adalah tekanan pengekstrakkan 200 bar, suhu pengekstrakkan 80 °C dan pengaliran karbon dioksida 20 mL/min. Keadaan optimum ini memperoleh kepekatan tocoferol

274.74 dan 89.75 mg/100 g minyak daripada biji-biji tembikai Kalahari dan roselle masing-masing.

Semasa penyimpanan minyak-minyak biji tembikai Kalahari dan roselle selama 6 bulan pada suhu 4 °C dan suhu bilik dalam kegelapan, perubahan berlaku dalam kandungan asid lemak, fitosterol, tocoferol, kehadiran produk pengoksidaan pertama dan kedua. Minyak-minyak tersebut adalah diperolehi daripada biji-biji tembikai Kalahari dan roselle dengan pengekstrakkan supergenting karbon dioksida. Seperti yang dijangkakan, perbezaan yang nyata dalam kandungan asid lemak, fitosterol, tocoferol, kehadiran produk pengoksidaan pertama dan kedua dalam minyak-minyak biji tembikai Kalahari dan roselle telah diperhatikan sepanjang penyimpanan. Kualiti indeks nilai peroksida dan anisidin telah meningkat semasa penyimpanan 6 bulan. Parameter degradasi mungkin berubah akibat pengoksidaan minyak selepas penyimpanan.

ACKNOWLEDGEMENTS

First of all, I would like to express my deepest gratitude and respect to my kind supervisor, Associate Professor Dr. Tan Chin Ping for his understanding, guidance, encouragement, and support throughout my study. I would also like to extend my appreciation and gratitude to the members of the advisory committee Professor Dr. Yaakob Bin Che Man, Associate Professor Dr. Lai Oi Ming and Dr. Kamariah Long for their invaluable contributions and support.

My sincere gratitude is also extended to the financial support provided by the Science Fund for this research, which was awarded to Associate Professor Dr. Tan Chin Ping. I am also indebted to all the staff of the Faculty of Food Science and Technology for their generous cooperation. Acknowledgement is also due to all my colleagues and laboratory assistants who had given me the moral encouragement and support to complete my graduate study.

Last but not the least, I also wish to express my deepest appreciation to my beloved parents, elder brother and younger brothers who have given me encouragement and support in one way or another during the many years of my seemingly never ending pursue for knowledge.



I certify that an Examination Committee has met on 16 November 2009 to conduct the final examination of Nyam Kar Lin on her Doctor of Philosophy thesis entitled “Extraction, Characterization and Storage Stability of Oils from Selected Plant Seeds” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the student be awarded the degree of Doctor of Philosophy.

Members of the Examination Committee were as follows:

Mohd Yazid Manap, Ph.D.,
Professor,
Faculty of Food Science and Technology,
Universiti Putra Malaysia.
(Chairman)

Md Zaidul Islam Sarker, Ph.D.,
Associate Professor,
Faculty of Food Science and Technology,
Universiti Putra Malaysia.
(Internal Examiner)

Lasekan Olusegun, Ph.D.,
Associate Professor,
Faculty of Food Science and Technology,
Universiti Putra Malaysia.
(Internal Examiner)

David B Min, Ph.D.,
Professor,
Department of Food Science and Technology,
The Ohio State University,
Columbus.
(External Examiner)

BUJANG KIM HUAT, PH.D.,
Professor and Deputy Dean
School of Graduate Studies,
Universiti Putra Malaysia

Date:



The thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy; the members of the Supervisory Committee were as follows:

Tan Chin Ping, PhD
Associate Professor,
Faculty of Food Science and Technology,
Universiti Putra Malaysia.
(Chairman)

Yaakob Bin Che Man, PhD
Professor,
Faculty of Food Science and Technology,
Universiti Putra Malaysia.
(Member)

Lai Oi Ming, PhD
Associate Professor,
Faculty of Biotechnology and Biomolecular Sciences,
Universiti Putra Malaysia.
(Member)

Kamariah Long, PhD
Doctor,
Malaysian Agricultural Research & Development Institute (MARDI)
(Member)

HASANAH MOHD GHAZALI, PHD
Professor and Dean
School of Graduate Studies,
Universiti Putra Malaysia

Date: 14 January 2010



Declaration Form

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

NYAM KAR LIN

Date:

TABLE OF CONTENTS

	Page
ABSTRACT	ii
ABSTRAK	v
ACKNOWLEDGEMENTS	viii
APPROVAL	ix
DECLARATION	xi
LIST OF TABLES	xii
LIST OF FIGURES	xviii
CHAPTER	
I. GENERAL INTRODUCTION	1
II. LITERATURE REVIEW	8
Oilseeds	8
Bitter melon	8
Kalahari-melon	9
Kenaf	10
Pumpkin	11
Roselle	13
Bioactive Compounds	15
Phenolic compounds	15
Phytosterols	20
Tocopherols and tocotrienols	24
Lipid Oxidation	27
Free radicals	28
Antioxidants	30
Important antioxidants present in food	31
Antioxidants allowed in food	33
Health Benefits of Antioxidants	35
Extraction of Oil	37
Solvent extraction	37
Aqueous enzymatic extraction	38
Supercritical fluid extraction	39
Oxidative Stability	46
Methods for the determination of lipid oxidation	47
III. PHYSICOCHEMICAL PROPERTIES AND BIOACTIVE COMPOUNDS OF SELECTED SEED OILS	50
Introduction	50
Materials and Methods	52
Proximate analysis of plant seeds	53
Oil extraction	53
Physical analysis of crude oil	54



	Thermal behavior	54
	CIE L*a*b*coordinates	54
	Chemical analysis	55
	Fatty acid composition	55
	Phenolic acids	56
	Gas chromatography analysis of sterols and squalene	58
	Chromatographic analysis of α -, β -, γ -, δ -Tocopherols	59
	Statistical analysis	59
	Results and Discussion	60
	Conclusion	75
IV.	ENZYME-ASSISTED AQUEOUS EXTRACTION OF KALAHARI-MELON-SEED OIL: OPTIMIZATION USING RESPONSE SURFACE METHODOLOGY	78
	Introduction	78
	Materials and Methods	79
	Experimental designs	81
	Aqueous enzymatic oil extraction from Kalahari-melon-seed	82
	Results and Discussion	83
	Conclusion	92
V.	PHYSICOCHEMICAL PROPERTIES OF KALAHARI-MELON-SEED OIL FOLLOWING EXTRACTIONS USING SOLVENT AND AQUEOUS ENZYMATIC METHOD	93
	Introduction	93
	Materials and Methods	94
	Oil extraction	95
	Physical analysis of crude oil	96
	Thermal behavior	96
	CIE L*a*b*coordinates	96
	Chemical analysis	97
	Fatty acid composition	97
	Phenolic acids	98
	Gas chromatography analysis of sterols	98
	Chromatographic analysis of α -, β -, γ -, δ -tocopherols	100
	Statistical analysis	100
	Results and Discussion	100
	Conclusion	115
VI.	OPTIMIZATION OF SUPERCRITICAL CO₂ EXTRACTION OF PHYTOSTEROL-ENRICHED OIL FROM KALAHARI-MELON-SEEDS	116
	Introduction	116
	Materials and Methods	118
	Experimental designs	119

	Extraction procedures	120
	Soxhlet extraction	120
	Gas chromatography analysis of sterols	121
	Results and Discussion	123
	Conclusion	134
VII.	OPTIMIZATION OF SUPERCRITICAL FLUID EXTRACTION OF PHYTOSTEROL FROM ROSELLE-SEEDS WITH A CENTRAL COMPOSITE DESIGN MODEL	136
	Introduction	136
	Materials and Methods	138
	Supercritical fluid extraction	139
	Soxhlet extraction	140
	Gas chromatography analysis of sterols	141
	Results and Discussion	142
	Conclusion	154
VIII.	EXTRACTION OF TOCOPHEROL-ENRICHED OILS FROM KALAHARI-MELON AND ROSELLE-SEEDS BY SUPERCRITICAL FLUID EXTRACTION (SFE-CO₂)	155
	Introduction	155
	Materials and Methods	156
	Experimental designs	157
	Extraction procedures	158
	Determination of tocopherol concentration of the extract	159
	Results and Discussion	159
	Conclusion	175
IX.	CHANGES OCCURRING IN FATTY ACIDS, PHYTOSTEROLS, TOCOPHEROLS COMPOSITION AND OXIDATIVE STABILITY OF <i>CITRULLUS LANATUS</i> AND <i>HIBISCUS SABDARIFFA</i> LINN. SEED OILS DURING STORAGE	177
	Introduction	177
	Materials and Methods	179
	Extraction procedures	180
	Storage experiments	181
	Chemical analysis	181
	Fatty acid composition	182
	Gas chromatography analysis of sterols	183
	Determination of tocopherol concentration of the extract	184
	Statistical analysis	184
	Results and Discussion	185
	Conclusion	197
X.	SUMMARY, CONCLUSION AND RECOMMENDATIONS	199
	Summary	199

Conclusion and Recommendations	202
REFERENCES	205
BIODATA OF STUDENT	229
LIST OF PUBLICATION	230



LIST OF TABLES

Table		Page
1	Phenolic classes in plants	18
2	Some reported sterol concentrations in selected vegetable oils (mg/ 100 g) (USDA, 1999)	23
3	Approximate content of tocopherol and tocotrienol found in vegetable oils (Schuler, 1990)	26
4	Antioxidants permitted in foods	34
5	Range values of several physicochemical properties of gases, liquids and supercritical fluids	41
6	Proximate analysis (g/100 g) of bitter melon, Kalahari-melon, kenaf, pumpkin and roselle-seeds ^A	61
7	Chemical properties of bitter melon, Kalahari-melon, kenaf, pumpkin and roselle-seed oils ^A	63
8	Relative percent composition of fatty acid in bitter melon, Kalahari-melon, kenaf, pumpkin and roselle-seed oils	64
9	Crystallization and melting behaviour of bitter melon, Kalahari-melon, kenaf, pumpkin and roselle-seed oils	69
10	Phenolic acids of oilseeds (mg/100 g, mean \pm SD) ^A	72
11	Sterols and squalene of oilseeds (mg/100 g, mean \pm SD) ^A	73
12	Tocopherols of oilseeds (mg/100 g, mean \pm SD) ^A	75
13	Experimental data and the observed response values with different combinations of enzyme concentration (g/100 g) (X_1), initial pH of mixture (X_2), incubation temperature ($^{\circ}$ C) (X_3) and incubation times (h) (X_4) for aqueous enzymatic oil extraction by Neutrase 0.8 L and Flavourzyme 1000 L	84
14	Analysis of variance for response surface quadratic model for aqueous enzymatic oil extraction by Neutrase 0.8 L	86
15	Analysis of variance for response surface quadratic model for aqueous enzymatic oil extraction by Flavourzyme 1000 L	86

16	Regression coefficients and P-values for aqueous enzymatic oil extraction by Neutrase 0.8 L and Flavourzyme 1000 L after backward elimination	88
17	Chemical properties of Kalahari-melon-seed oils extracted using various methods ^A	103
18	Relative percent composition of fatty acid in Kalahari-melon-seed oils extracted using various methods ^A	105
19	Crystallization and melting behaviour of Kalahari-melon-seed oils	107
20	Phenolic acids of Kalahari-melon-seed oils extracted using various methods (mg/100 g, mean \pm SD) ^A	113
21	Sterols of Kalahari-melon-seed oil extracted using various methods (mg/100 g, mean \pm SD) ^A	113
22	Tocopherol of Kalahari-melon-seed oil extracted using various methods (mg/100 g, mean \pm SD) ^A	115
23	Experimental and the observed response values with different combinations of pressure (X_1), temperature (X_2) and flow rate (X_3) for Kalahari-melon-seed oil extraction by SFE	124
24	Analysis of variance for response surface quadratic model for oil recovery by supercritical fluid extraction	126
25	Analysis of variance for response surface quadratic model for phytosterol concentration in Kalahari-melon-seed oil by using supercritical fluid extraction	126
26	The differences between the observed and predicted values for oil recovery and phytosterol concentration in extracted oil	132
27	Experimental and the observed response values with different combinations of pressure (X_1), temperature (X_2) and flow rates (X_3) for roselle-seed oil extraction by SFE	143
28	Analysis of variance for response surface quadratic model for oil recovery by supercritical fluid extraction	145
29	Analysis of variance for response surface quadratic model for phytosterol concentration in roselle-seed oil by using supercritical fluid extraction	145
30	Regression coefficients and P-values for oil recovery by supercritical fluid extraction after backward elimination	146

31	Regression coefficients and P-values for phytosterol concentration in roselle-seed oil by supercritical fluid extraction after backward elimination	150
32	The differences between the observed and predicted values for oil recovery and phytosterol concentration in extracted oil	153
33	Experimental and the observed response values with different combinations of pressure (X_1), temperature (X_2) and flow rate (X_3) for extraction of tocopherol from Kalahari-melon-seed oil by SFE	161
34	Experimental and the observed response values with different combinations of pressure (X_1), temperature (X_2) and flow rate (X_3) for extraction of tocopherol from roselle-seed oil by SFE	162
35	Analysis of variance for response surface quadratic model for tocopherol concentration from Kalahari-melon seed by supercritical fluid extraction	163
36	Analysis of variance for response surface quadratic model for tocopherol concentration from roselle-seed by supercritical fluid extraction	164
37	Regression coefficients and P-values for tocopherol concentration from Kalahari-melon-seed by supercritical fluid extraction after backward elimination	165
38	Regression coefficients and P-values for tocopherol concentration from roselle-seed by supercritical fluid extraction after backward elimination	165
39	The differences between the observed and predicted values for tocopherol concentration in Kalahari-melon-seed oil	174
40	The differences between the observed and predicted values for tocopherol concentration in roselle-seed oil	175
41	Changes in fatty acid content (%) of Kalahari-melon-seed oil during 6 months storage period at 4 °C and room temperature	189
42	Changes in fatty acid content (%) of roselle-seed oil during 6 months storage period at 4°C and room temperature	190

LIST OF FIGURES

Figure	Page
1 Chemical Structures of Phenolic Acids Analysed in This Study. (A) Derivatives of Benzoic Acid; (B) Derivatives of Cinnamic Acid	17
2 Chemical Structures of Cholesterol and the Studied Phytosterols	22
3 DSC Cooling Curves for (A) Bitter melon, (B) Kalahari-Melon, (C) Kenaf, (D) Pumpkin and (E) Roselle-Seed Oils	67
4 DSC Heating Curves for (A) Bitter melon, (B) Kalahari-Melon, (C) Kenaf, (D) Pumpkin and (E) Roselle-Seed Oils	68
5 The CIE L*a*b*coordinates for Bitter melon (1), Kalahari-Melon (2), Kenaf (3), Pumpkin (4) and Roselle (5) Seed Oils, Respectively	71
6 DSC Cooling Curves for Kalahari-Melon-Seed Oils Extracted Using (a) Solvent (b) Flavourzyme 1000 L (c) Neutrased 0.8 L	108
7 DSC Heating Curves for Kalahari-Melon-Seed Oils Extracted Using (a) Solvent (b) Flavourzyme 1000 L (c) Neutrased 0.8 L	109
8 The CIE L*a*b*Coordinates for Kalahari-Melon-Seed Oils Extracted Using (a) Solvent (b) Flavourzyme 1000 L (c) Neutrased 0.8 L	111
9 Response Surface Plot of Interaction Between Pressure (X_1) and Supercritical Fluid Flow Rate (X_3) at Low Level of Temperature (X_2) on Oil Recovery	128
10 Response Surface Plot of Interaction Between Temperature (X_2) and Supercritical Fluid Flow Rate (X_3) at Central Level of Pressure (X_1) on Phytosterol Concentration of Kalahari-Melon-Seed Oil	130
11 Response Surface Plot of Interaction Between Pressure (X_1) and Supercritical Fluid Flow Rates (X_3) at Low Level of Temperature (X_2) on Oil Recovery	147
12 Response Surface Plot of Interaction Between Pressure (X_1) and Supercritical Fluid Flow Rates (X_4) at Low Level of Temperature (X_2) on Phytosterol Concentration of Roselle-Seed Oil	151
13(a) Response Surface Plot of Interaction between Pressure (X_1) and Temperature (X_2) at High Level of Flow Rate of Carbon Dioxide (X_3) on Tocopherol Concentration of Kalahari-Melon-Seed Oil	168



13(b)	Response Surface Plot of Interaction between Temperature (X_2) and Supercritical Fluid Flow Rate (X_3) at Central Level of Pressure (X_1) on Tocopherol Concentration of Kalahari-Melon-Seed Oil	169
14(a)	Response Surface Plot of Interaction between Pressure (X_1) and Supercritical Fluid Flow Rate (X_3) at High Level of Temperature (X_2) on Tocopherol Concentration of Roselle-Seed Oil	170
14(b)	Response Surface Plot of Interaction between Temperature (X_2) and Supercritical Fluid Flow Rate (X_3) at Low Level of Pressure (X_1) on Tocopherol Concentration of Roselle-Seed Oil	171
15(a)	Changes in Phytosterol Concentration (mg/100 g) of Kalahari-Melon-Seed Oil During 6 Months Storage Period at 4°C and Room Temperature	186
16(a)	Changes in Tocopherol Concentration (mg/100 g) of Kalahari-Melon-Seed Oil During 6 Months Storage Period at 4°C and Room Temperature	187
15(b)	Changes in Phytosterol Concentration (mg/100 g) of Roselle-Seed Oil During 6 Months Storage Period at 4°C and Room Temperature	191
16(b)	Changes in Tocopherol Concentration (mg/100 g) of Roselle-Seed Oil During 6 Months Storage Period at 4°C and Room Temperature	192
17(a)	Changes in Peroxide Value (meq/kg) of Roselle-Seed Oil During 6 Months Storage Period at 4°C and Room Temperature	193
17(b)	Changes in Anisidine Value of Roselle-Seed Oil During 6 Months Storage Period at 4°C and Room Temperature	194
17(C)	Changes in TOTOX Value of Roselle-Seed Oil During 6 Months Storage Period at 4°C and Room Temperature	195

CHAPTER 1

GENERAL INTRODUCTION

Recently, more attention has been focused on the utilization of food processing by-products and wastes, as well as under-utilized agricultural products. Obviously, such utilization would contribute to maximizing available resources and result in the production of various new foods. Simultaneously, waste disposal problems could be minimized.

The problems of industrial waste are becoming harder to solve, and much effort will be needed to develop the nutritional and industrial potential of by-products, waste and under-utilized agricultural products. Only a small portion of plant material is utilized directly for human consumption (El-Adawy *et al.*, 1999). A portion of the remaining material may be converted into nutrients for either food or feed, or into fertilizer, making possible an important contribution to food resources or industrial products (El-Adawy *et al.*, 1999; Kamel *et al.*, 1982). For example, the seeds of the bitter melon, Kalahari-melon, kenaf, pumpkin and roselle could be used; these seeds are present in large quantities as waste products after the removal of the pulp, peel and flesh of these plants.

Bitter melon (*Momordica charantia* L.), also known as bitter gourd, is a monoecious climbing vine. It is a tropical crop, grown throughout Asia for food and



medicinals (Chakravarty, 1990). The seeds contain oil in which the major fatty acid is eleostearic acid (ESA), which is a major component of oil from tung nuts and is the constituent responsible for the “drying” characteristic of tung oil. The latter is used extensively in paints, coatings and inks.

Kalahari-melon (*Citrullus lanatus*) is the most important source of water in the Kalahari during dry months of the year when no surface water is available. The fruit is cut open at the one end and the first piece of flesh is eaten. The remaining contents are pounded with a stick, and are then eaten and drunk. Seeds are roasted and ground into meal—a nutritious food with a pleasant, nutty taste. The leaves and young fruit are utilized as green vegetables (Van Wyk and Gericke, 2000). The peels of the fruit are traditionally used for making jam. The cultivated watermelon is a popular summer fruit in all parts of the world.

Kenaf (*Hibiscus cannabinus* L.) is a warm season annual belonging to the Malvaceae family, which also includes cotton (*Gossypium* spp.) and okra (*Abelmoschus esculentus* L. Moench). It has been used for thousands of years in Africa and parts of Asia as a source of fiber for making clothes, rugs, ropes and other product. The commercial uses of kenaf continues to diversify from its historical role as cordage to its various new applications, including paper products, building materials, absorbents and livestock feeds (Webber and Bledsoe, 1993; Sullivan, 2003). Seeds from kenaf fruit may provide an excellent oil resource. The oil has chemical

