

UNIVERSITI PUTRA MALAYSIA

TOXICOLOGICAL AND IMMUNOLOGICAL EFFECTS OF DIAZINON ON THE GREAT STURGEON (*HUSO HUSO*) OF NORTHERN IRAN

HOSSEIN ALI KHOSHBAVAR ROSTAM FPV 2008 4



TOXICOLOGICAL AND IMMUNOLOGICAL EFFECTS OF DIAZINON ON THE GREAT STURGEON (*HUSO HUSO*) OF NORTHERN IRAN

HOSSEIN ALI KHOSHBAVAR ROSTAMI

DOCTOR OF PHILOSOPHY UNIVERSITI PUTRA MALAYSIA

2008



TOXICOLOGICAL AND IMMUNOLOGICAL EFFECTS OF DIAZINON ON THE GREAT STURGEON (*HUSO HUSO*) OF NORTHERN IRAN

BY

HOSSEIN ALI KHOSHBAVAR ROSTAMI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in the Fulfilment of Requirements for the Degree of Doctor of Philosophy

March 2008



DEDICATION

WITH LOVE AND APPRECIATION TO:

My parents: Mohamad Ali and Soghra Javan My wife: Ommolbanin Kardar My sons: Mohamad Ali and Mohsen My daughter: Farzaneh



Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

TOXICOLOGICAL AND IMMUNOLOGICAL EFFECTS OF DIAZINON ON THE GREAT STURGEON (*HUSO HUSO*) OF NORTHERN IRAN

By

HOSSEIN ALI KHOSHBAVAR ROSTAMI

March 2008

Chairman: Associate Professor Hassan Hj. Mohd. Daud, PhD

Faculty: Veterinary Medicine

Great sturgeon (*Huso huso*) is one of the highly valuable commercial fish species and the major economic resources of Northern Iran. Currently limited data are available on the immunophysiological responses of this valuable species. Unfortunately, within the last decade the average annual sturgeon harvest including great sturgeon from the South Caspian Sea has drastically reduced due to increase pollution problem. Diazinon is one of the most important organophosphorus pesticide groups commonly used in Iranian agriculture, including northern Iran in which both are the natural habitat and aquaculture sites of great sturgeon. In addition also, previous studies had shown that motile *Aeromonas* septicemia disease is one of the main factors in high mortality outbreaks in the sturgeon farming, particularly whenever the fish immune system is suppressed by some toxicants. Thus, the main objectives of this study were: (i) purification and characterization of great sturgeon immunoglobulin (IgM); (ii) determination of 96-hour LC₅₀ diazinon in great



sturgeon; (iii) assessment of some immunophysiological variables of fish following exposure to diazinon; and (iv) assessment of some immunoresponses of fish following treatment with some immunostimulators i.e. glucan and Aeromonas hydrophila antigen. In the immune response assessment, the IgM purified by affinity chromatography under non-reduced condition was found to be 870 kDa, as estimated by SDS-PAGE, while the MWt of the heavy and light chains under reduced condition were estimated at 77-84 and 28-30 kDa, respectively. In the diazinon toxicity study, the LC_{50} value at 96-hour in fish weighing ca. 14 g under static water quality condition at 22°C was calculated as at 5.63 mg/L. Examination of haematological indices and tissue lysozyme of kidney, liver and spleen showed that exposure to diazinon at sublethal concentration of 1.5 mg/mL as long-term bath caused an effect similar to anaemia. Also, there were significant and insignificant changes in some blood parameters including immunocompetent cell populations, AST, ALP, ALT and LDH enzymes as well as chemiluminescence response of leucocytes at different days post-exposure to diazinon. An almost similar finding was observed when the glucan-injected fish (0.3 mg/kg bwt.) and fish immunized intraperitoneally with a single dose of formalin-killed A.hydrophila (5×10^7 cells/fish) were exposed to long-term, sublethal concentration of diazinon. However, some of these immunophysiological responses including respiratory burst, immunocompetent cell counts and lysozyme content were enhanced in both glucan-injected fish and immunized fish without diazinon bath. Also, the microagglutination antibody titer in immunized fish was higher than unimmunized fish.

In light microscope examinations there were congestion, haemorrhages, focal and generalized necrosis, cellular infiltration, hyperplasia and lamellae fusion as major



histopathological changes in the tissues of liver, kidney, spleen and gills of the fish exposed to sublethal dose of diazinon. Also, there were an increase in droplet materials blocking of club cell surfaces in nostrils and barbels tissues; a reduction in excretion of amorphous proteinaceous materials, increase in vesicle numbers and blockage of nostrils epithelial cell surfaces in diazinon exposed fish under scanning electron microscope examination.

In conclusion, short and long-term exposure of great sturgeon to diazinon at acute and chronic concentrations changes the basic blood cells constituents causing leucopenia, lymphopenia, neutrophila, erythropenia, hyperglycemia, hypoproteinemia and decreased in both specific antibody production and leucocytes respiratory burst. Thus, avoiding the exposure of endangered juvenile fish to this pollutant is highly recommended.

Key words: great sturgeon, *Huso huso*, diazinon, glucan, *A. hydrophila*, lysozyme, chemiluminescence response, antibody, immunocompent cells, immunoglobulin, LC₅₀ 96-hour.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

KESAN DIAZINON TERHADAP TOKSIKOLOGIKAL DAN IMUNOFISIOLOGIKAL DALAM STURGEON GERGASI, *HUSO HUSO* (BRANDT, 1869)

Oleh

HOSSEIN ALI KHOSHBAVAR ROSTAMI

Mac 2008

Pengerusi: Profesor Madya Hassan Hj. Mohd. Daud, PhD

Fakulti: Perubatan Veterinar

Sturgeon gergasi (*Huso huso*) adalah salah satu spesis ikan komersial bernilai tinggi dan menjadi sumber utama ekonomi di utara Iran. Pada masa ini, data mengenai tindakbalas imunofisiologi spesis yang bernilai ini amat terbatas. Walaubagaimana pun, dalam dekad kebelakangan ini, purata hasil tangkapan tahunan ikan sturgeon, termasuk sturgeon gergasi dari Laut Caspian Selatan telah menurun secara drastik kerana peningkatan masalah pencemaran. Diazinon merupakan salahsatu dari racun serangga penting kumpulan organofosfat yang digunakan secara meluas dalam pertanian Iran, termasuk di utara Iran yang mana merupakan habitat semulajadi dan tempat akuakultur sturgeon gergasi. Tambahan juga, kajian terdahulu telah menunjukkan bahawa penyakit septisemia *Aeromonas* motil adalah faktor utama dalam kejadian wabak kematian tinggi dalam penternakan sturgeon, terutamanya



apabila sistem imun ikan tertekan oleh setengah bahan toksik. Olehitu objektif utama kajian ini adalah: (i) menyuling dan menciri imunoglobulin (IgM) sturgeon gergasi; (ii) menentu LC₅₀ pada 96 jam diazinon dalam sturgeon gergasi; (iii) menilai beberapa pembolehubah imunofisiologi ikan selepas pendedahan terhadap diazinon; dan (iv) menilai beberapa tindakbalas imun ikan selepas rawatan dengan peransang imun iaitu glukan dan antigen Aeromonas hydrophila. Dalam penilaian tindakbalas imun, IgM yang ditulinkan dengan kromatografi keafinan dibawah keadaan tak berkurangan didapati seberat 870 kDa, sebagaimana dianggarkan dengan SDS-PAGE. Manakala berat molekul rantai berat dan rantai ringan adalah dianggarkan masing-masing pada 77-84 dan 28-30 kDa. Dalam ujian ketoksikan diazinon pada ikan seberat 14 g, nilai LC₅₀ pada 96 jam, dalam keadaan air statik pada suhu 22°C adalah 5.63 mg/L. Pemeriksaan indek hematologikal dan lisozim tisu ginjal, hepar dan limfa menunjukkan bahawa pendedahan terhadap diazinon pada kepekatan subletal 1.5 mg/mL sebagai mandian jangka panjang menyebabkan kesan sama seperti anemia. Juga terdapat perubahan bererti dan tidak bererti dalam beberapa parameter darah termasuk populasi sel imunomampu, AST, ALP, ALT dan enzim LDH, dan juga tindakbalas kimipendarcahaya leukosit pada hari berbeza pasca pendedahan pada diazinon. Keputusan yang hampir sama dapat dilihat dalam ikan yang disuntik glukan (0.3 mg/kg berat badan) dan ikan yang diimun dengan satu dos A. hydrophila yang dimati dengan formalin secara intraperitoneum (5×10^7) sel/ikan) apabila didedahkan secara jangka masa panjang pada kepekatan subletal diazinon. Walaubagaimana pun, beberapa tindakbalas imunofisiologikal termasuk letusan penafasan leukosit, bilangan sel imunomampu dan kandungan lisozim dipertingkatkan dalam ikan yang disuntik dengan glukan dan yang diimun, tanpa



mandian diazinon. Didapati juga, mikroagglutinasi titer antibodi dalam ikan diimun adalah lebih tinggi dari ikan tidak diimun.

Dalam pemeriksaan mikroskop cahaya, perubahan histopatologi yang utama adalah kongesi, hemoraj, nekrosis fokal dan umum, infiltrasi sel dalam tisu hepar, ginjal, limfa dan hiperplasia dan penyantuman lamella dalam insang ikan yang terdedah pada dos subletal diazinon. Didapati juga, pemerhatian mikroskop elektron imbasan menunjukkan terdapat penambahan bahan titisan dan penutupan permukaan sel belantan dalam tisu nostril dan barbel, pengurangan dalam perkumuhan bahan protein amorfus, peningkatan jumlah vesikel dan penutupan permukaan sel epitelial nostril dalam ikan yang didedahkan pada diazinon.

Sebagai rumusan, pendedahan samaada secara jangka pendek atau panjang sturgeon gergasi pada diazinon pada kepekatan akut dan kronik mengubah konstitusi asas sel darah menyebabkan leukopenia, limfopenia, neutrofila, eritropenia, hiperglisemia, hipoproteinemia dan pengurangan dalam pengeluaran antibodi spesifik dan letusan penafasan leukosit. Olehitu, mengelakkan pendedahan ikan juvenil yang terancam pada bahan pencemaran ini adalah amat disyorkan.

Perkataan kunci: sturgeon gergasi, *Huso huso*, diazinon, glukan, *A. hydrophila*, lisozim, tindakbalas kemopendafluor, antibodi, sel imunomampu, imunoglobulin, LC₅₀ pada 96 jam.



ACKNOWLEDGEMENTS

Praise is to the Almighty ALLAH. Lord of all creations, for his heavenly, luxurious blessings over me throughout my life and the period of this study. I would like to express my heartfelt gratitude and appreciation to my main Supervisor, Associate Professor Dr. Hassan Hj. Mohd Daud, for his valuable guidance and constructive suggestions throughout the research program. I sincerely appreciate the innumerable hours he spent reading the draft and the suggestions made to improve the thesis. I wish to express my deepest thanks to my co-supervisors: Professor Dr. Mehdi Soltani, Associate Professor Dr. Abdul Rahman Omar, for their valuable suggestions and kind assistance throughout this study.

A very special acknowledgement is given to Dr. Sohrab Rezvani and Dr. Abas Motallebi Head of Iranian Fisheries Research Organization (IFRO), for their cooperation during the process of conducting the study. I would also like to thank Dr. Ahmadian, Dr. Bahram Kazemi and Dr. Iesa Sharifpour for their assistance with electron microscopy, purification and characterization of beluga IgM and histopathological slides reading.

I am also grateful to the staff of Mazandaran Fisheries Research Center, Faculty of Veterinary Medicine, University of Tehran, and Research Deputy of University of Tehran for their co-operation. I would like to express my thanks to all my friends whom I obtained their assistance during this study especially Mr Ali Mokarmi for his invaluble help and assistance. I wish to express my deepest thanks to my wife, my sons and my daughter for their supports and patience during my study.Lastly, I



would like to record my gratitude to many others whose name do not appear here, who have helped me during my study period.



I certify that an Examination Committee met on 14th March 2008 to conduct the final examination of HOSSEIN ALI KHOSHBAVAR ROSTAMI on his Doctor of Philosophy thesis entitled "Toxicological and immunophysiological effects of diazinon on the great sturgeon, *Huso huso* (Brandt)" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The committee recommends that the candidate be awarded the relevant degree. Members of Examination Committee are as follows:

Mohd. Hair Bejo, Ph.D.

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

Mohamed Ali Rajion, Ph.D.

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Internal Examiner)

Rasedee Abdullah, Ph.D. Professor

Faculty of Veterinary Medicine Universiti Putra Malaysia (Internal Examiner)

Benjamin, R. Mikryakov, Ph.D.

Professor Institute for Biology of Inland Waters Russian Academy of Sciences (External Examiner)

HASANAH MOHD GAZALI, PhD

Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia



This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfilment of the requirements for the degree of Doctor of Philosophy. The members of the Supervisory Committee are as follows:

Hassan Hj Mohd Daud, Ph.D.

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

Mehdi Soltani, Ph.D.

Professor Faculty of Veterinary Medicine University of Tehran (Member)

Abdul Rahman Omar, Ph.D.

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

AINI IDERIS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia



DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

HOSSEIN ALI KHOSHBAVAR ROSTAMI

Date :



TABLE OF CONTENTS

DEDICATION	ii
ABSTRACT	Error! Bookmark not defined.
ABSTRAK	Error! Bookmark not defined.
ACKNOWLEDGEMENTS	ix
APPROVAL	xi
DECLARATION	xiii
LIST OF TABLES	xviii
LIST OF FIGURES	xxi
LIST OF ABBREVIATIONS	xxvi
CHAPTER 1	1
INTRODUCTION	1
CHAPTER 2	7
LITERATURE REVIEW	7
2.1. Sturgeon immunity	7
2.1.1. Non-specific immunity	7
2.1.2. Specific infinumity 2.2. Effect of toxic chemicals on fish immunophysi	ological responses 11
2.2.1. Effect of diazinon on fish immunophysiolo	ogical responses 14
2.3. Motile Aeromonas septiceamia in sturgeons	19
2.3.1. The Disease (in Fish)	20
2.3.2. Therapy	22
2.5.5. The Disease (in numans) 2.4. Glucans as fish immunostimulator	22
2.5. Conclusion	25

CHAPTER 3	27
PURIFICATION AND PARTIAL CHARACTERIZATION OF SERUM	
IMMUNOGLOBULINS FROM CASPIAN SEA STURGEONS	27
3.1. Introduction	27
3.2. Materials and Methods	28
3.2.1. Immunization of fish	28



3.2.2. Affinity chromatography	28
3.2.3. SDS-PAGE	29
3.3. Results	30
3.4. Discussion	32

CHAPTER 4

33

ACUTE TOXICITY AND SOME HEAMATOLOGICAL AND BIOCH	EMICAL
CHANGES IN GREAT STURGEON (HUSO HUSO) EXPOSED TO	
DIAZINON	33
4.1. Introduction	33
4.2. Materials and Methods	34
4.2.1. Fish	34
4.2.2. Toxicant	34
4.2.3. Acute toxicity	34
4.2.3.1. Determination of survival rate:	34
4.2.3.2. Determination of lethal concentration of diazinon	35
4.2.3.3. Determination of LC_{50} at 96 hours	35

4.2.4. Haematological and biochemical studies	36
4.2.5. Data analysis	37
4.3. Results	37
4.3.1. Acute toxicity test	37
4.3.1.1. Survival rate	37
4.3.1.2. Determination of the lethal concentration of diazinon	37
4.3.1.3. Determination of LC ₅₀ of diazinon	38

4.3.1.3. Determination of LC_{50} of diazinon	38
4.3.1.4. Behavioral signs	39
4.3.2. Haematological profile	39
4.3.3. Plasma biochemistry	41
4.4. Discussion	42

CHAPTER 5

CHAPTER 5	45
SUBLETHAL TOXICITY OF DIAZINON ON SOME IMMUNOPHYSIOLOGICAL VARIABLES OF GREAT STURGEON	45
5.1. Introduction	45
5.2. Materials and methods	47
5.2.1. Fish	47
5.2.2. Application of diazinon	47
5.2.3. Sample collection and assays	48
5.2.4. Haematology and biochemistry	48
5.2.5. Lysozyme assay	48
5.2.6. Chemiluminescent (CL) assay	49
5.2.7. Light microscopy study	50
5.2.8. Electron microscopy	50
5.2.9. Statistical analysis	52
5.3. Results	52
5.3.1. Haematology	52



5.3.2. Biochemistry	54
5.3.3. Differential count	56
5.3.4. Lysozyme	58
5.3.5. Chemiluminescence (CL) response	60
5.3.6. Light microscopy examination	61
5.3.7. Electron microscope examination	77
5.3.7.1. Scanning electron microscopic	77
5.3.7.2. Transmission electron microscopic	86
5.4 Discussion	88

CHAPTER 6

94

ASSESSMENT OF SOME IMMUNOPHYSIOLOGICAL VARIABLES OF GLUCAN INJECTED GREAT STURGEON AFTER LONG-TERM	
EXPOSURE TO SUBLETHAL CONCENTRATION OF DIAZINON	94
6.1. Introduction	94
6.2. Materials and methods	97
6.2.1. Fish	97
6.2.2. Application of glucan and diazinon	97
6.2.3. Sample collection and assays	98
6.2.3.1. Haematology and biochemistry	98
6.2.3.2. Lysozyme assay	99
6.2.3.3. Chemiluminescent (CL) assay	99
6.3. Results	99
6.3.1. Haematology	99
6.3.2. Biochemistry	102
6.3.3. Differential count	105
6.3.4. Lysozyme	107
6.3.5. Chemiluminescence (CL) response	109
6.4. Discussion	110

CHAPTER 7

115

IMMUNE RESPONSE OF GREAT STURGEON TO AEROMONAS HYDROPHILA BACTERIN AND IN THE PRESCNCE OF LONG-TERM EXPOSURE TO SUBLETHAL CONCENTRATION OF DIAZINON

XPOSURE TO SUBLETHAL CONCENTRATION OF DIAZINON	115
7.1. Introduction	115
7.2. Materials and methods	117
7.2.1. Fish	117
7.2.2. Antigen preparation and immunization	118
7.2.3. Application of diazinon	118
7.2.4. Sample collection and assays	118
7.2.4.1. Haematology and biochemistry	119
7.2.4.2. Lysozyme assay	119
7.2.4.3. Chemiluminescent (CL) assay	120
7.2.4.4. Antibody titration	120
7.2.5. Statistical analysis	120
7.3. Results	120



731 Haematology	120
7.3.2. Biochemistry	123
7.3.3. Differential count	125
7.3.4. Lysozyme	128
7.3.5. Agglutination titration	130
7.3.6.Chemiluminescence (CL) response	131
7.4. Discussion	132

CHAPTER 8	138
GENERAL DISCUSSION AND CONCLUSION	138
8.1. General discussion	138
8.2. Summary and conclusion	150
REFERENCES	153
APPENDICES	169
BIODATA OF THE AUTHOR	172
LIST OF PUBLICATIONS	174



LIST OF TABLES

Table 4.1	The lethal concentration of diazinon for great sturgeon	Page 38
4.2	Cumulative mortality of great sturgeon at 24, 48, 72 and 96 hrs post- exposure to various concentrations of diazinon.	38
4.3	Determination of diazinon lethal concentration (mg/L) for great sturgeon)	39
4.4	Erythrocyte profile of great sturgeon following exposure to diazinon (5.63 mg/L, 96h) at $21\pm2^{\circ}C$	40
4.5	Leukocyte profile of great sturgeon following exposure to diazinon (5.63 mg/L, 96h) at $21\pm2^{\circ}$ C.	41
4.6	The effect of diazinon following exposure (5.63 mg/L, 96h) on enzyme activities, and total protein of blood plasma of great sturgeon at $21\pm2^{\circ}$ C.	41
5.1	Erythroyte profile of <i>Huso huso</i> following exposure to a constant sublethal concentration of diazinon at 1.5mg/L at 22°C. (Mean \pm SD). A= Fish without diazinon bath, D= Fish with diazinon bath. *Indicating significant difference at P<0.05).	53
5.2	The effect of constant sublethal concentration of diazinon at 1.5mg/mg/L on enzyme activities, glucose level and total protein concentrations of blood plasma of Huso huso at 22°C. (Mean \pm SD) A= Fish without diazinon bath, , D= Fish with diazinon bath *Indicating significant difference at P<0.05).	55
5.3	Leucocyte profile of <i>Huso huso</i> following continuous exposure to sublethal concentration of diazinon at 1.5 mg L-1 at 22°C. (Mean \pm St. Dev., n=5). A=Fish without diazinon exposure, D= Fish with diazinon exposure, *Indicating values are significantly different at P<0.05).	57
5.4	Lysozyme contents in tissues of liver, kidney, spleen and serum of Huso huso following continuous exposure to sublethal concentration of diazinon at 1.5 mg L-1 at 22°C. (Mean±St. Dev., n=5) A=Fish without diazinon bath, B= Fish with diazinon exposur. *Indicating significant difference at (P<0.05).	59

- 5.5 Histopathological scores of beluga's liver exposed to continuous 62 exposure of diazinon at 20-22°C.
- 5.6 Histopathological scores of beluga's kidney exposed to continuous 66 exposure of diazinon at 20-22°C.
- 5.7 Histopathological scores of beluga's gills exposed to continuous 70



exposure of diazinon at 20-22°C.

- 5.8 Histopathological scores of beluga's spleen exposed to continuous 74 exposure of diazinon at 20-22°C.
- 6.1 Erythrocyte profile of great sturgeon following exposure to a constant 102 sublethal concentration of diazinon at 1.5mg/L at 22°C. (Mean ±SD).A=Fish without diazinon bath or glucan (normal control), B=Glucan injected fish without diazinon bath, C= Glucan injected fish with diazinon bath.
- 6.2 The effect of constant sublethal concentration of diazinon at 105 1.5mg/mg/L on enzyme activities, glucose level and total protein concentrations of blood plasma of great sturgeon at 22°C. (Mean± SD), A= normal control, B=Glucan injected fish without diazinon bath, C= Glucan injected fish with diazinon bath.
- 6.3 Leucocyte profile of great sturgeon following continous exposure to 107 sublethal concentration of diazinon at 1.5 mg L-1 at 22°C. (Mean±St. Dev., n=5), A= normal control, B=Glucan injected fish without diazinon exposure, C= Glucan injected fish with diazinon exposure.
- 6.4 Lysozyme contents in tissues of liver, kidney, spleen and serum of 109 great sturgeon following continous exposure to sublethal concentration of diazinon at 1.5 mg /L at 22°C. (Mean±St. Dev., n=5), A= normal control, B=Glucan injected fish without diazinon exposure, C= Glucan injected fish with diazinon exposure.
- 7.1 Erythrocyte profile of *Huso huso* following exposing to a constant 123 sublethal concentration of diazinon at 1.5mg/L at 22°C. (Mean ±St.Dev.), A=Normal control, E=Immunized fish without diazinon bath, F=Immunized fish with diazinon bath.

The effect of constant sublethal concentration of diazinon at 125 1.5mg/mg/L on enzyme activities, glucose level and total protein
7.2 concentrations of blood plasma of *Huso huso* at 22°C. (Mean± St. Dev.), A=Normal control, E=Immunized fish without diazinon bath, F=Immunized fish with diazinon bath..

- 7.3 The effect of constant sublethal concentration of diazinon at 1.5mg/L 128 on differential count of blood of Huso huso at 22°C. (Mean± St. Dev.), A=Normal control, E=Immunized fish without diazinon bath, F=Immunized fish with diazinon bath.
- 7.4 Lysozyme contents in tissues of liver, kidney, spleen and serum of 130 Huso huso following continous exposure to sublethal concentration of diazinon at 1.5 mg /L at 22°C. (Mean±St. Dev., n=5), A=Normal



control, E=Immunized fish without diazinon bath, F=Immunized fish with diazinon bath.



LIST OF FIGURES

Figure

- 3.1 SDS-PAGE with 8% polyacrylamide gel stained with Coomassie Blue 31 under non-reduceding conditions for estimation of molecular weight of the Caspian Sea sturgeon species IgM. Lanes 1 through 5: affinity-purified IgMs from Huso huso, Acipenser guldenstedti, Acipenser nudiventris, Acipenser stellatus and Acipenser persicus, respectively, M: marker
- 3.2 SDS-PAGE under reduceding conditions for estimation of molecular 32 weight of IgMs of the Caspian Sea sturgeons. Lanes 1 through 5: affinity-purified Igs from Huso huso, Acipenser guldenstedti, Acipenser nudiventris, Acipenser stellatus and Acipenser persicus, respectively, M: marker
- 5.1 .Mean spontaneous chemiluminescence response of great sturgeon 60 blood leucocyte following a continues exposure to sublethal concentration of diazinon at 1.5 mg L-1 at 22°C. (Mean ±St. Dev., n=5) A= Negative control (fish without diazinon bath), B=Fish with diazinon bath. *Indicating values are significantly different at (P<0.05).
- 5.2 Peak of spontaneous chemiluminescence response of great sturgeon 61 blood leucocyte following a continues exposure to sublethal concentration of diazinon at 1.5 mg L-1 at 22°C. (Mean ±St. Dev., n=5) A= Negative control (fish without diazinon bath), B= Fish with diazinon bath.
- 5.3 Generalized liver degeneration as indicated by pyknotic nuclei of 63 hepatocytes (P), vacuolation of cytoplasm (V) and melanomacrophage center (MMC) in fish exposed to 1.5 mg/L diazinon at 1 day post-exposure (HandE, 1075).
- 5.4 15 days post exposing to 1.5 mg/L diazinon in liver of beluga as 63 indicated by pyknotic nuclei of hepatocytes (P), congestion of blood vessel (CV) and melanomacrophage center (MMC), (HandE, 860).
- 5.5 28 days post exposing to 1.5 mg/L diazinon in liver of great sturgeon 64 as indicated by pyknotic nuclei of hepatocytes (P), necrosis of liver tissue (N) and melanomacrophage center (MMC) (HandE,860).
- 5.6 42 days post exposing to 1.5 mg/L diazinon in liver of great sturgeon 64 as indicated by expansion of sinusoid (E), congestion of vessle (CV), degeneration of hepatocytes (DH) and melanomacrophage center (MMC) (HandE, 344).
- 5.7 63 days post exposing to 1.5 mg/L diazinon in liver of great sturgeon 65 indicated by melanomacrophage center, degeneration of hepatocytes, pyknotic and necrosis of hepatocytes and rupture of liver tissue (HandE, 860).



- 5.8 Congestion of glomerulus (CG) and dilation of bowmen capsule(DB) 67 in tissue of kidney at 1 day post-exposure to 1.5 mg/L diazinon (HandE, 860).
- 5.9 congestion of glomerulus (CG), dilation of bowmen capsule(DB), 67 necrosis of glomerulus (NG) and necrosis of tubular (NT) in tissue of kidney at 15 day post-exposure to 1.5 mg/L diazinon (HandE, 860).
- 5.10 thicken of basal membrane of bowmen capsule (TB), dilation of 68 bowmen capsule(DB), necrosis of glomerulus (NG) and protein in tubule (P) in tissue of kidney at 28 day post-exposure to 1.5 mg/L diazinon (HandE, 752).
- 5.11 thicken of basal membrane of bowmen capsule (TB), necrosis of 68 glomerulus (NG) and congestion of glomerulus (CG) in tissue of kidney at 42 day post-exposure to 1.5 mg/L diazinon (HandE, 860).
- 5.12 thicken of basal membrane of bowmen capsule (TB), dilation of 69 bowmen capsule (DB), degeneration of tubules (DT), hemorrhage of kidney (H) and congestion of glomerulus (CG) in tissue of kidney at 63 day post-exposure to 1.5 mg/L diazinon (HandE, 860).
- 5.13 Gills lamellae of beluga at 1 day post-exposure to 1.5 mg/L diazinon 71 showing congestion of secondary lamellae (SC). Congestion of primary lamellae (PC) and existing of melanin in blood vessel (M) (HandE, 1075).
- 5.14 Gills lamellae of beluga at 14 day post-exposure to 1.5 mg/L diazinon 71 showing congestion of secondary lamellae (SC).congestion of primary lamellae (PC) and proliferation of mucus cells in secondary lamellae (PM) (HandE, 860).
- 5.15 Gills lamellae of beluga at 28 day post-exposure to 1.5 mg/L diazinon 72 showing clubbing of secondary lamellae (CL), Hyperplasia (H)and proliferation of mucus cells in secondary lamellae (PM) (picture A) and existing of melanin in blood vessel (M), congestion of secondary lamellae (SC) (picture B) (HandE, A 537 and B 430).
- 5.16 Gills lamellae of great sturgeon at 42 day post-exposure to 1.5 mg/L 72 diazinon showing Separation of epiderm from the base of secondary lamellae (arrow head). (HandE, 677).
- 5.17 Gills lamellae of great sturgeon at 42 day post-exposure to 1.5 mg/L 73 diazinon showing congestion of secondary lamellae (SC), Congestion of primary lamellae (PC), dilation of secondary lamellae(D), proliferation of mucus cells in secondary lamellae (PM), hyperplasia (H) and Separation of epiderm from the base of primary lamellae (SP). (HandE, 677).



- 5.18 spleen of great sturgeon at 1 day post-exposure to 1.5 mg/L diazinon 75 showing Congestion of red pulp (arrow head) and existing of eosinophil in white pulp (arrow) (HandE, 860).
- 5.19 spleen of great sturgeon at 14 day post-exposure to 1.5 mg/L diazinon 75 showing Congestion of spleen(arrow head) and necrosis in tissue of spleen(arrow) (HandE, 433).
- 5.20 spleen of great sturgeon at 28 day post-exposure to 1.5 mg/L diazinon 76 showing cogestion (arrow head) and necrosis in tissue of spleen (arrow) (HandE, 433).
- 5.21 spleen of great sturgeon at 42 day post-exposure to 1.5 mg/L diazinon 76 showing hypercellularity and necrosis of spleen tissue (HandE, 433).
- 5.22 spleen of great sturgeon at 63 day post-exposure to 1.5 mg/L diazinon 77 showing hypercellularity, disconfiguration and necrosis of spleen tissue (HandE, 433).
- 5.23 SEM micrographs of nostril epithelial cells of normal beluga showing 78 amorphous proteinaceous materials, vesicles and cell surface canals, Mag: A = x500, B = x1000, C = x2500, D = x5000.
- 5.24 SEM micrograph of nostril epithelial cells of great sturgeon 1 day post-79 exposing to 1.5 ppm diazinon showing an increase in droblet on the cell surface. Mag: A= x2500, B= x5000, C= x500, D= x1000.
- 5.25 SEM micrograph of nostril epithelial cells of great sturgeon 28 day 80 post- exposing to 1.5 ppm diazinon showing a reduction in excretion of amorphous proteinaceous materials, vesicles numbers and blockage of cell surface canals. Mag: A= x2500, B= x5000, C= x500, D= x1000.
- 5.26 SEM micrograph of nostril epithelial cells of great sturgeon 63 day 81 post- exposing to 1.5 ppm diazinon showing a severe reduction in excretion of amorphous proteinaceous materials and vesicles. The cell surface canals were blocked. Mag: A= x2000, B= x7000, C= x2000, D= x2500.
- 5.27 SEM micrographs of barbell epithelial cells of normal great sturgeon 82 showing amorphous proteinaceous materials, vesicles and cell surface canals, Mag: A = x200, B = x100, C = x1000, D = x1000.
- 5.28 SEM micrograph of barbell epithelial cells of great sturgeon 1 day postexposing to 1.5 ppm diazinon showing an increase in droblet on the cell surface. Mag: A= x500, B= x1000, C= x1000, D= x2000.
- 5.29 SEM micrograph of barbell epithelial cells of great sturgeon 28 day 84 post- exposing to 1.5 ppm diazinon showing a reduction in excretion of amorphous proteinaceous materials, vesicles numbers and blockage of cell surface canals. Mag: A= x2500, B= x5000, C= x500, D= x1000.

