



UNIVERSITI PUTRA MALAYSIA

**MOLECULAR CHARACTERIZATION OF HUMAN GROUP A
ROTAVIRUS ISOLATES FROM MALAYSIA AND DEVELOPMENT OF A
COLORIMETRIC PCR-BASED TEST FOR THE
DETECTION OF P[8] GENOTYPE**

ZURIDAH HASSAN

FPV 2004 11



**MOLECULAR CHARACTERIZATION OF HUMAN GROUP A
ROTAVIRUS ISOLATES FROM MALAYSIA AND DEVELOPMENT OF A
COLORIMETRIC PCR-BASED TEST FOR THE
DETECTION OF P[8] GENOTYPE**

By

ZURIDAH HASSAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia
for the Degree of Doctor of Philosophy**

September 2004



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment
of the requirement for the degree of Doctor of Philosophy

**MOLECULAR CHARACTERIZATION OF HUMAN GROUP A
ROTAVIRUS ISOLATES FROM MALAYSIA AND DEVELOPMENT OF A
COLORIMETRIC PCR-BASED TEST FOR THE DETECTION OF P[8]
GENOTYPE**

By

ZURIDAH HASSAN

September 2004

Chairman: Professor Abdul Rani Bahaman, Ph.D.

Faculty : Veterinary Medicine

Rotavirus has been recognized as a leading cause of the diarrhoeal illness in children under 5 years of age in the developing world. Latex agglutination test was used to detect group A rotavirus from 157 in-patients from different hospitals in Malaysia during 2000 to 2001. Diarrhoea was detected in 31 (19.7%) children and majority were under two years of age.

When viewed under electron microscope by negative staining, rotavirus was seen as both double-shelled and single-shelled particles. Thirty one rotavirus antigen



positive samples with typical group A electropherotype were further characterized into their G or P types by polymerase chain reaction (PCR) assay. The two common electropherotypes were IIC (51.6%) and IIG (35.5%). The most prevalent VP4 genotype was 25 (80.6%) P[8] and 1 (3.2 %) P[6]. Genotype P[4] and P[9] were not isolated and 5 (16.1 %) were P untypable (P^{UT}). Regarding the VP7 genotype, G4 was the most prevalent (64.5 %), followed by G1 (6.45%), G2 (6.45%) and G3 (3.2 %). Neither G8 nor G9 was found and 6 (19.4 %) were G untypable (G^{UT}). Studies in many countries found that G1P[8], G4P[8], G2P[4] and G3P[8] are the group A rotavirus strains more commonly seen in children. However from this present study, the common strains in Malaysia were G4P[8], G1P[8] and G3P[8]. One G^{UT}P[6] strain (designated as 7W) was identified for the first time in Kuala Lumpur. Restriction endonuclease *Hae*III and *Sau*96I were also used to characterize the VP7 gene of the local 7W strain. However a restriction profile could not be assigned. The P[8] and P[6] local strains (represented by 67F and 7W, respectively) were also characterized by nucleotide sequence analysis. Phylogenetic analysis revealed that the VP4 genes of the 67F and 7W formed a distinct lineage.

The P[8] and P[6] are encoded by distinct VP4 gene alleles. The main diagnostic problem is the genetic diversity of these alleles among different rotavirus



strains. To overcome this problem, a method that employs non-radioactive dot hybridization was successfully developed for P[8] and P[6]. VP4 cDNA rotavirus-specific probes were prepared and labelled with digoxigenin (DIG). Anti-DIG-alkaline phosphatase and the substrate NBT/BCIP were used to detect the binding of the probe to target sequence.

A simple, practical, sensitive and specific assay based on polymerase chain reaction (PCR) and a colorimetric detection method (ELISA) for the typing of rotavirus in infected faeces has been developed successfully. A set of oligonucleotides was employed for a single-tube reverse-transcription nested PCR (RT-nPCR). Upon synthesis of the first strand cDNA, a first stage of 10 cycles of PCR amplification was run to generate an 876-bp dsDNA from the 5' terminal third of gene 4. The process was completed in the same tube by performing another 35 cycles of second stage amplification incorporating a biotinylated and digoxigenin 5'-end labelled primers. The RT-nPCR produced a 180-bp amplicon representing the VP4 P[8] type. The sensitivity of the RT-nPCR method was compared to non-nested PCR method and nested PCR was found to be 100 times more sensitive. To further increase the sensitivity, the enzyme-linked immunoassay (ELISA) was incorporated into the system. Streptavidin-coated microtitre plate was used to capture the biotinylated PCR-amplified products. This RT-nPCR ELISA was able



to detect RNA as low as 4 pg nucleic acid. It was designed to type the single most epidemiologically important human rotavirus VP4 P[8] type which is often associated with rotavirus G1, G3 and G4 types. Monoclonal antibodies (Mabs) were used for G serotyping, but no Mabs were available for P serotyping. Therefore, the RT-nPCR ELISA method is a very useful technique to detect rotavirus.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**PENCIRIAN MOLEKULAR ROTAVIRUS KUMPULAN A DARI MANUSIA
DI MALAYSIA DAN PEMBENTUKAN UJIAN KOLORIMETRIK
BERASASKAN PCR UNTUK MENGESAN P[8] GENOTIP**

Oleh

ZURIDAH HASSAN

September 2004

Pengerusi : Profesor Abdul Rani Bahaman, Ph.D.

Fakulti : Perubatan Veterinar

Rotavirus telah dikenalpasti sebagai penyebab utama penyakit cirit-birit di kalangan kanak-kanak berumur kurang dari 5 tahun di negara membangun. Agglutinasia latex telah digunakan untuk mengenalpasti rotavirus kumpulan A dari 157 pesakit dari berlainan hospital di Malaysia sepanjang 2000 ke 2001. Cirit-birit telah dipencil daripada 31 (19.7%) kanak-kanak dan kebanyakannya adalah berumur kurang dari dua tahun.

Apabila dilihat menggunakan mikroskop elektron kaedah pencelupan negatif, rotavirus yang mengandungi dua selaput dan satu selaput dapat dilihat. Tiga puluh satu sampel rotavirus antigen positif yang menunjukkan elektroferotaip



khusus untuk group A telah dicirikan kepada jenis G atau P melalui reaksi rantaian polimerasi (PCR). Dua elektroferotaip yang sering ditemui adalah IIC (51.6%) dan IIG (35.5%). VP4 genotip yang terbanyak adalah 25 P[8] (80.6%) dan 1 (3.2%) P[6]. Genotip P[4] dan P[9] tidak ditemui dan 5 (16.1%) adalah P yang tidak boleh digenotip (P^{UT}). Berkaitan VP7 genotip, G4 adalah terbanyak (64.5%), diikuti oleh G1 (6.45%), G2 (6.45%) dan G3 (3.2%). G8 dan G9 tidak ditemui dan 6 (19.4%) adalah G yang tidak boleh digenotip (G^{UT}). Kajian di beberapa negara menunjukkan bahawa G1P[8], G4P[8], G2P[4] dan G3P[8] rotavirus kumpulan A sering ditemui dikalangan kanak-kanak. Walau bagaimanapun, dari kajian ini jenis yang selalu ditemui di Malaysia adalah G4P[8], G1P[8] dan G3P[8]. Satu strain $G^{UT}P[6]$ (dikenalpasti sebagai 7W) dari kajian ini telah dilaporkan buat pertama kalinya dari Kuala Lumpur. Enzim pembatas *HaeIII* dan *Sau96I* juga telah digunakan untuk mengkaji gen VP7 strain tempatan 7W tetapi profil pembatas tidak dapat ditentukan. Strain P[8] dan P[6] tempatan (diwakili sebagai 67F dan 7W) juga dikaji menggunakan jujukan nukleotid. Analisis filigenesis menunjukkan gen VP4 67F dan 7W membentuk kumpulan berlainan.

P[8] dan P[6] dienkod oleh gen VP4 allele yang berlainan. Masalah utama untuk mengenalpasti ialah kepelbagaian allele genetic di dalam strain. Untuk mengatasinya, dot hybridization tanpa-radioaktif telah dicipta untuk P[8] dan P[6]. Prob spesifik VP4 cDNA telah disediakan dan dilabelkan dengan digoxigenin (DIG).



Anti-DIG-alkaline phosphatase dan substrat NBT/BCIP telah digunakan untuk mengesan prob yang terlekat pada jujukan yang disasarkan.

Satu lagi kaedah yang menggunakan teknik reaksi rantaian polimerasi (PCR) dan pengesanan 'colorimetrik' (ELISA) yang mudah, praktikal, sensitif dan spesifik untuk mengesan rotavirus telah berjaya dibentuk. Satu set oligonukleotida telah diguna dalam kaedah 'single-tube reverse-transcription nested polymerase chain reaction' (RT-nPCR). Di dalam pembentukan stran pertama cDNA di dalam tindakbalas RT, amplifikasi PCR pertama sebanyak 10 pusingan menghasilkan 876 bp dsDNA dari pangkal 5' gen 4. Proses ini disempurnakan dengan 35 pusingan amplifikasi kedua di mana primers yang dilabel dengan biotin dan DIG pada pangkal 5' digunakan. RT-nPCR menghasilkan amplicon 180 bp mewakili VP4 P[8]. Sensitiviti kaedah RT-nPCR didapati 100 kali lebih sensitif berbanding dengan kaedah tanpa-nPCR. Untuk menambahkan sensitiviti, kaedah ELISA telah dimasukkan. Plat mikrotiter yang disalut dengan streptavidin telah digunakan untuk memerangkap produk PCR yang berbiotin. RT-nPCR ELISA boleh mengesan RNA sehingga 4 pg. Ia telah dicipta untuk mengesan VP4 gen P[8] yang mempunyai kepentingan epidemiologi dan lazimnya dikaitkan dengan rotavirus G1, G3 dan G4. Sebelum ini antibodi monoklonal (Mabs) digunakan untuk G serotyping, tetapi Mabs untuk P serotyping belum diwujudkan lagi. Maka, dengan itu, RT-nPCR ELISA adalah satu kaedah penting untuk mengesan rotavirus.



ACKNOWLEDGEMENTS

I would like to express my appreciation to Prof. Dr. Abdul Rani Bahaman, Prof. Dr. Mohd Azmi Lila and Assoc. Prof. Dr. Abdul Rahim Mutalib for their invaluable advice. Their untiring assistance, support, constructive comments, understanding and encouragement motivate me to complete this study.

Special thanks to Norumon Sumalee who always share the tasks together, to Lai KY and Zeenathul Nazariah Allaudin for extending their vast molecular biology knowledge and frequent lift to KTM Serdang and to other laboratory mates, Cheng, Tam, Sandy, Do Yew, Dr Khairani, Liza, Kamarudin, Zainudin and staff of Institute of Bioscience, UPM for various assistance.

I would also like to thank various individuals and institutions who have helped me during my study:

the pathologists and microbiologists from various hospitals in Malaysia for providing the test samples;

Dr Jon R Gentsch, Centre of Disease Control, Atlanta, Georgia, for providing the reference rotavirus strains ;

En Fauzi for the photography work, Ms Azilah and Mr Ho for EM work;

Director of Health (Tan Sri Dr Mohd Taha), Deputy Director of Health, Dato Dr Ahmad Tajuddin, Jabatan Perkhidmatan Awam Malaysia and Dr Jamil



Dolkadir, Pathologist, Hospital Umum Sarawak, for granting the study leave;

I dedicate this work to my husband, Syed Abdul Razak Syed Aziz, my daughter (Syarifah Hafsa) and my son (Syed Munawir) and my parents (Hj Hassan Saman and Hjh Ainiyah Saad) for their love, support and understanding.

This study was sponsored by Jabatan Perkhidmatan Awam, Malaysia and by IRPA Grant No. 54001.



I certify that an Examination Committee met on 22nd September 2004 to conduct the final examination of Zuridah Hassan on her Doctor of Philosophy thesis entitled “Molecular Characterization of Human Group A Rotavirus Isolates From Malaysia and Development of a Colorimetric PCR-based Test for the Detection of P[8] Genotype” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

Aini Ideris, Ph.D.

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Datin Khatijah Mohd Yusoff, Ph.D.

Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Abdul Rahman Omar, Ph.D.

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Yap Kok Leong, Ph.D.

Professor
Department of Biomedical Science
Faculty of Allied Health Sciences
Universiti Kebangsaan Malaysia Kuala Lumpur
(Independent Examiner)

.....
GULAM RUSUL RAHMAT ALI, Ph.D.

Professor/Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:



This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirements for the degree of Doctor of Philosophy. The members of the Supervisory Committee are as follows:

Abdul Rani Bahaman, Ph.D.
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Mohd Azmi Mohd Lila, Ph.D.
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Abdul Rahim Mutalib, Ph.D.
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

AINI IDERIS, Ph.D.
Professor/ Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:



DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

ZURIDAH HASSAN

Date: 2004



TABLE OF CONTENTS

	Page
ABSTRACT	ii
ABSTRAK	vi
ACKNOWLEDGEMENTS	ix
APPROVAL	xi
DECLARATION	xiii
LIST OF TABLES	xviii
LIST OF FIGURES	xix
LIST OF ABBREVIATIONS	xxiii
CHAPTER	
I INTRODUCTION	1
II LITERATURE REVIEW	6
Rotavirus Infection in Humans	6
Characteristics of Rotavirus	7
Rotavirus Protein Structure and Function	10
Rotavirus VP4 and VP7 serotypes	12
Rotavirus Serotypes/Genotypes and Epidemiology Worldwide	15
Epidemiology of Rotavirus Infection in Malaysia	18
Immunity to Rotavirus Infection	20
Clinical Manifestation of Rotavirus Infection	21
Rotavirus Vaccines	22
Laboratory Testing for Rotaviruses	24
Electron Microscopy	25
Detection of Rotavirus Antigen	24
Electropherotyping	27
Hybridization Assay	29
Reverse-transcriptase Nested Polymerase Chain Reaction	30
PCR ELISA Detection Method	34
Restriction Endonuclease Analysis of Rotaviruses	35
Sequence Analysis of the Gene Coding for VP4 Protein	36



III	DETECTION, IDENTIFICATION AND GENOTYPING OF ROTAVIRUS IN MALAYSIA	39
	Introduction	39
	Materials and Methods	42
	Patients and Stool Specimens	42
	Latex Agglutination to Detect Rotavirus Group A Antigen	44
	Electron Microscopy	44
	Extraction and Detection of Rotavirus RNA by Polyacrylamide Gel Electrophoresis	45
	RNA Extraction	45
	RNA Polyacrylamide Gel Electrophoresis (RNA-PAGE)	45
	Reference Rotavirus Strains for RNA-PAGE	46
	Rotavirus G and P Genotyping by RT-PCR	47
	Test Samples	47
	RNA Extraction, Purification and Removal of Inhibitors	49
	G-typing Method for Gene 9	51
	Primers	51
	RT-PCR From dsRNA	52
	First PCR Reaction	53
	Second PCR Reaction	53
	P-Typing for Gene 4	54
	Primers	54
	RT-PCR From dsRNA	55
	First and Second PCR Reaction	56
	Detection of PCR Product	57
	Results	58
	Occurrence of Rotavirus in Different Hospitals in Malaysia As Detected by Latex Agglutination Test	58
	Incidence of Rotavirus Infection in Different Age Groups	59
	Detection of Rotavirus by Electron Microscope	61
	RNA Polyacrylamide Gel Electrophoresis (RNA-PAGE)	62
	Rotavirus G and P Typing Results	64
	Distribution of VP7 and VP4 Genotypes	67
	Distribution of G and P Types and their association with RNA-PAGE patterns	69
	Distribution of Rotavirus Antigen, Electropherotypes, VP7 and VP4 Types In Different Locations	71
	Discussion	72



IV	MOLECULAR CHARACTERIZATION OF THE LOCAL ROTAVIRUS ISOLATES USING RESTRICTION ENZYME ANALYSIS, DOT HYBRIDISATION AND NUCLEOTIDE SEQUENCE ANALYSIS	82
	Introduction	82
	Materials and Methods	86
	Restriction Endonuclease Analysis of the VP7 Genes of Human Rotaviruses	86
	Viruses for Restriction Endonuclease Analysis	86
	Nucleotide sequence analysis	86
	Dot Blot Hybridisation	87
	Preparation of Probes by Random primed DNA Labeling with digoxigenin-dUTP	87
	Fixation of samples	88
	Hybridization	88
	Stringency Washes	89
	Immunological detection	89
	Results	91
	Restriction profiles of VP7 amplicons	91
	Dot hybridization	93
	Nucleotide Sequence Analysis	96
	Discussion	100
V	DEVELOPMENT OF RT-NESTED PCR ELISA METHOD FOR THE DETECTION OF ROTAVIRUS INFECTION	106
	Introduction	106
	Materials and Methods	110
	Primers	110
	Sample Processing and dsRNA extraction	111
	cDNA synthesis and First Amplification Reaction of One-tubed Nested PCR	111
	Second amplification step	113
	Optimisation of PCR ELISA	113
	Analysis of PCR Products	116
	Agarose Gel Electrophoresis	116
	Colorimetric detection (ELISA) of PCR Product	117



Sensitivity and Specificity of RT-nested PCR	117
Sensitivity of RT-nested PCR-ELISA	118
Reproducibility of RT-PCR-ELISA	118
Efficacy of the developed system on clinical samples	118
Results	119
RT-nested PCR Optimization	119
Optimisation and Standardization of RT-nested PCR ELISA	120
Sensitivity and specificity of RT-nested PCR (RT-nPCR)	125
Sensitivity of RT-nested PCR ELISA	127
Reproducibility of RT-nested PCR ELISA	131
Efficacy of the Developed Method to detect Rotavirus	131
Discussion	133
VI GENERAL DISCUSSION AND CONCLUSION	139
REFERENCES	150
APPENDIX A	179
BIODATA OF THE AUTHOR	180
PUBLICATIONS	181



LIST OF TABLES

Table		Page
1.1	Rotavirus SA11 genes coding assignments	11
3.1	History of rotavirus isolates	48
3.2.	Oligonucleotides primers for PCR amplification of rotavirus gene 9	52
3.3.	Oligonucleotides primers for PCR amplification of rotavirus gene 4	55
3.4.	Combinations of rotavirus G and P types in Malaysia in the 31 RNA-PAGE positive rotavirus isolates	68
3.5.	Relationship between genotypes and RNA electropherotypes in Malaysia	70
4.1	Comparison of the sequence of the VP4 gene local strains 67F and 7W and the corresponding reference strains ...	97



LISTS OF FIGURES

Figure	Page
2.1. Schematic diagram showing rotavirus gene products. Left, RNA segments and gene coding assignments. Right, the viral structural proteins on different shells of the virus particles (from Kapikian and Chanock, 1990)	8
2.2 Schematic diagram of electrophoretic migration of RNA segments 10 and 11, which are characteristic of human rotavirus long (II) pattern (reference strain SA11) and short (I) pattern (reference strain DS1). The positions of RNA segments 1 through 11 are indicated on the left. The classification type (see text) of the major electropherotype is shown at the top of the figure....	28
3.1 Map of Malaysia showing the locations of the hospitals included in the study. The numbers indicate total samples and percentage positive; and the type of electropherotypes in square bracket	43
3.2 Incidence of rotavirus infection according to age group and corresponding electropherotypes	60
3.3 Electron micrograph of rotavirus (60,000 x magnification). The double-shelled particle (dsp) and single-shelled particle (ssp) are shown (negative staining)	61
3.4 Comparative genome profile of Malaysian group A human rotavirus isolates and reference strain WI61. The position of RNA segments 1 through 11 are indicated on the left and the type of electropherotypes is shown at the bottom of the gel. The WI61 reference strain is used to represent rotavirus with ‘long’ or II electropherotype	63



- 3.5. A: Amplification of VP7 and VP4 gene of human rotavirus from clinical samples (Table 3.1). Lanes 2-3, product of VP7 gene (1,062-bp) (primers Beg 9/End 9); lanes 6 to 9 , product of VP4 gene (876-bp) (primers con 3/con 2); M, 100-bp DNA ladder (New England, Biolabs). B: PCR G typing (second PCR amplification) of VP7 gene. Lanes 1 and 4, G1 (primers RVG 9/aBT1); lane 2, G4 (primers RVG 9/aDT4); lane 3, G2 (primers RVG 9/aCT2); lane 5, G3 (primers RVG 9/aET3). C: PCR P typing (second PCR amplification) and confirmation of P types. Lanes 1 and 4, rotavirus dsRNA was reversed transcribed, amplified and confirmed as P[8] (primers 1C-1/1C-2); lane 2, second PCR amplification of P[8] (primers con 3/1T-1); lane 3, second PCR amplification of P[6] (primers con 3/3T-1); lane 5, confirmation of P[6] (primers 3C-1/3C-2) 65
- 4.1. Restriction profiles of VP7 gene performed in 2.5 % NuSieve-1% SeaKem agarose gel electrophoresis. In gel A, G4 type reference strain ST3 were digested with *Hae*III (lane 2). Lanes 3 and 4 were loaded with 67F and 7W, respectively. Lane 6 was loaded with undigested reference strain ST3. In gel B, ST3 reference strain digested with *Sau*96I was loaded into lane 1. Lanes 2 and 3 were loaded with 7W and 67F, respectively. Lane M was loaded with bacteriophage ΦX174 DNA marker (Boehringer Mannheim, Germany) 92
- 4.2. Dot blot hybridization of rotavirus dsRNA extracted from stool samples with VP4 P[8] and VP4 P[6] probes. (A) The VP4 P[8] probe was hybridized with Wa(P[8] reference strain), and P[8] samples obtained from clinical samples (5KB, CFB and 1K – Table 3.1). Other control included nucleic acid extracted from a rotavirus-antigen negative (Rota –ve) sample. (B) The VP4 P[6] probe was hybridized with P[6] reference strain, ST3; P[6] sample obtained from patient 7W and clinical samples 5KB, CFB and 1K 95



4.3.	CLUSTAL W analysis of local strains 7W with P[8] type and 67F with P[6] type. Nucleotide sequence of the VP4 gene were compared by the CLUSTAL W program and are presented as a phylogram. The local strains were compared with the reference P[8] type (strains 1076, ST3, RV3 and M37) and with the reference P[6] type (strains KU and Wa)	99
5.1	Human rotavirus gene 4 showing the positions and directions of amplification relative to those of the plus (mRNA) sense genomic strand for the outer primers con3 and con2 and for the gene inner (nested) primers 1C-1 and 1C-2	110
5.2	Schematic diagram showing the nested PCR-ELISA. B, biotin; D, digoxigenin; V, streptavidin; E, horse-radish peroxidase conjugated to anti-digoxigenin antibody; ABTS, 2,2,'-azinodi-ethyl-benzothiazolinesulphonate	114
5.3	Various concentrations of streptavidin titrated against various concentration of conjugate. Absorbance values of streptavidin gradually increased with increased conjugate concentration. The 10 ug/ml and 5 ug/ml did not differ markedly but the 1 ug/ml gave a lower absorbance values	121
5.4	Determination of the optimum conjugate concentrations for ELISA PCR. Various dilutions of PCR products were titrated against different conjugate concentration. Absorbance was recorded up to 1:32,000 PCR dilutions for both the 1:5,000 and 1:10,000 conjugate concentrations. At this same PCR dilutions, the 1:15,000 and 1:30,000 conjugate dilutions failed to record a positive signal	123



- 5.5 Sensitivity of RT-nested PCR (product 180 bp), (B) Sensitivity of non-nested RT-PCR using the outer primers (con3/con2, product 876 bp), and (C) Sensitivity of non-nested RT-PCR using the inner primers (1C-1/1C-2, product 180 bp). The PCR products (10 ul) were loaded in 2 % agarose gel, run at 80 V for 1.2 h, and photographed under UV light. Different band intensities were produced. The amplification was up to 10^{-6} dilution in the RT-nested PCR but only up to 10^{-4} in the non-nested PCR. Lane M, 100 bp DNA ladder, lanes 1-7 represented a serial 10-fold dilution of virus after clarification of stool and extraction of RNA 126
- 5.6 Specificity of the RT-nested PCR assay in differentiating rotavirus from other virus samples and human cell line in ethidium bromide-stained 2 % agarose gel. Lane M, 100 bp DNA ladder; lanes 1,5-9, 11-14, rotavirus samples; lane 2, NDV; lane 3, IBV; lane 4, negative control; and lane 10, RNA from human cell line ...127
- 5.7 Detection of amplified rotavirus RNA by the ELISA method. Samples (lanes) 1 to 7 correspond to 10-fold serial dilutions of the RNA virus (10^{-2} to 10^{-8}), starting at 40 ng, 20 ng, 4 ng, 0.4 ng, 0.04 ng, 0.004 ng (4 pg) and < 4 pg of RNA as template in the PCR. The cutoff was defined as the mean of the negative control plus three times the standard deviations 128
- 5.8 (A). Detection of 10-fold amplified RNA by PCR from rotavirus by the ELISA method with a positive signal up to 1:1000 dilutions. (B) Agarose gel electrophoresis, with very faint positive bands in the 1:100 dilutions (lane 3). Lane M, 100 bp DNA ladder, lanes 1 to 5 correspond to the dilution of PCR product in A 130
- 5.9 Detection of rotavirus PCR product by RT-nested PCR from those samples that were previously screened by LA and RNA-PAGE (using ethidium bromide-stained 2% agarose gel. Lane M, 100-bp DNA ladder; lanes 1 to 14, stool samples, clarified and amplified by RT-nested PCR. B. The same clinical in A were examined by ELISA. The absorbance values (OD_{405}) and the interpretation of the tests were tabulated below (the numbers corresponded to those in panel A, cutoff = 0.113 132



LISTS OF ABBREVIATIONS

aa	-	amino acid
ABTS	-	2,2'-Azido-di(3-ethyl)benzthiazoline sulphonic acid
Arg	-	arginine
bp	-	base pairs
CaCl ₂	-	calcium chloride
cDNA	-	complementary deoxyribonucleic acid
CF11	-	cellulose fiber
COOH	-	carboxy end
CsCl ₂	-	caesium chloride
DNA	-	deoxyribonucleic acid
dNTP	-	deoxyribonucleic acid
ds	-	double stranded
dsp	-	double shelled particle
DEPC	-	diethyl pyrocarbonate
DIG	-	digoxigenin
EDTA	-	ethylene diamine tetra acetic acid
ELISA	-	enzyme-linked immunoassay
EM	-	electron microscope
e-type	-	electropherotype
<i>HaeIII</i>	-	<i>Haemophilus influenzae</i> III
HCl	-	hydrochloric acid
H ₂ O ₂	-	hydrogen peroxide
IBDV	-	Infectious Bursal Disease Virus
KCl	-	potassium chloride
KDa	-	kilodalton
Mg	-	magnesium
MgCl ₂	-	magnesium chloride
MTP	-	microtiter plate
NaCl	-	sodium chloride
Na ₂ CO ₃	-	sodium carbonate
NaHCO ₃	-	sodium bicarbonate
NaOH	-	sodium hydroxide
NBT-BCIP	-	nitroblue tetrazolium 5-bromo-4-chloro-3-indolyl-phosphate
NH ₂ terminal	-	amino terminal
NDV	-	Newcastle disease virus
OD	-	optical density
ORF	-	open reading frame
PBS	-	phosphate buffered saline



PCR	-	polymerase chain reaction
PD	-	primer dimer
pH	-	negative logarithm of hydrogen ion
P ³²	-	phosphorous ³²
rpm	-	rotation per minute
RNA	-	ribonucleic acid
RNA-PAGE	-	RNA polyacrylamide gel electrophoresis
RT-nPCR	-	Reverse-transcription nested Polymerase Chain Reaction
RT-PCR	-	reverse transcriptase polymerase chain reaction
<i>Sau961</i>	-	<i>Staphylococcus aureus</i> 961
SD	-	standard deviation
SDS-PAGE	-	sodium dodecyl sulphate polyacrylamide gel electrophoresis
ssp	-	single shelled particle
STE	-	saline tris EDTA
TAE	-	Tris acetic acid
TBE	-	Tris borate EDTA
TBS	-	tris buffered saline
TE	-	Tris EDTA
TEMED	-	tetramethylethylenediamine
<i>UT</i>	-	untypable
UPM	-	Universiti Putra Malaysia
VP	-	viral protein

