



UNIVERSITI PUTRA MALAYSIA

***OCCURRENCE OF CAMPYLOBACTER SPP. AND THEIR ANTIBIOTIC
RESISTANCE PROFILES IN CATTLE AND FARM ENVIRONMENT***

WINT WINT AUNG

FPV 2014 26



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By

WINT WINT AUNG

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Master of
Veterinary Science**

June 2014

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DEDICATION

This thesis is especially dedicated to:

My beloved parents,

**U AUNG MYINT
and
DAW THAN AYE**

My beloved husband and daughter,

**DR. SWE MYINT OO
KAY ZIN LEI**

Who always supported and encourage me to do the best

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Veterinary Science

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WINT WINT AUNG

June 2014

Chairman: Prof. Saleha Abdul Aziz, PhD

Faculty : Veterinary Medicine

Campylobacter, principally *C. jejuni* and *C. coli*, have been recognized as one of the important causal agents of gastrointestinal infections in humans all over the world. The major source of human infection is raw or undercooked poultry meat but beef, pork, raw milk and water have also been associated with the infection. Most of the studies in Malaysia were on poultry and poultry products. The work on occurrence of *Campylobacter* in cattle, beef and milk is very scarce. Thus, the objectives of this study were to determine the occurrence of *Campylobacter* in cattle, farm environment, milk and meat, to identify the *Campylobacter* isolates by phenotypic method and multiplex PCR assay and to study the antibiotic resistance patterns of the isolates. One hundred and eighty (180) rectal swab samples from cattle, 68 samples from cattle farm environments, 36 raw milk samples from six farms and 30 beef samples from four markets were collected. All samples were cultured on selective media and isolated *Campylobacter* species were confirmed and identified using multiplex PCR. The overall prevalence of *Campylobacter* in dairy and beef cattle was 47 (26.1%) out of 180 samples. Eleven cattle were colonized by two *Campylobacter* species. The prevalence was higher in beef cattle 18 out of 57 samples (31.6%) compared to dairy cattle 29 out of 123 samples (23.6%) but the difference was not significant ($p=0.256$). The prevalence was significantly higher in calves 16 out of 40 samples (40%) than adult cattle 31 out of 140 samples (22.1%) ($p=0.023$). The isolation of *Campylobacter* from cattle was more at incubation temperature of 42°C (25.0%) compared to at 37°C (21.1%), however the difference was not significant ($p=0.381$) and kappa test statistic showed almost perfect agreement between the two different temperatures ($\text{kappa}>0.8$). Six *Campylobacter* species were identified at both temperatures; the most frequent isolated species was *C. jejuni* 23 (39.6%) and followed by *C. fetus* 13 (22.4%), *C. upsaliensis* 8 (13.8%), *C. coli* 5 (8.6%), *C. hyointestinalis* subsp. *hyointestinalis* 4 (6.9%) and the least prevalent species was *C. lari* 3 (5.2%). However, two isolates were unidentified *Campylobacter* species. From a total of 68 environmental samples, 19 (27.9%) *Campylobacter* isolates were isolated, namely from 10 out of 27 water samples (37.0%), four out of 16 flies samples (25.0%), one out of seven feed samples (14.3%), three out of nine

floors of the cattle houses samples (33.3%) and one out of nine water trough samples (11.1%) which are considered as the risk factors for *Campylobacter* in cattle. Flies could be an essential vector for transmission of *Campylobacter* from contaminated environment to cattle in the farms or from infected animals to the environment. The occurrence of *Campylobacter* in feed, floor, drinking water and water trough could be contaminated via flies and animal faeces. Ten (10) isolates (27.8%) of the 36 raw milk samples were *Campylobacter* positive, however none of the 30 retail beef samples were positive. The occurrence of *Campylobacter* in milk could have resulted from contamination during milking. The absence of *Campylobacter* in retail beef probably suggests they were not contaminated at processing and poor resistance of *Campylobacter* to atmospheric oxygen and other environmental pressures during storage, transportation and retailing may cause *Campylobacter* to convert to viable but non culturable (VBNC) form. The overall isolation rate of *Campylobacter* from cattle, environment samples, beef and milk when incubated under two different temperatures was higher at 42°C (22.6%) when compared to 37°C (18.5%); however, the difference was not significant ($p=0.199$) and kappa test statistic showed good agreement between the two different incubation temperatures ($0.6 \leq k < 0.8$) and six *Campylobacter* species were isolated at both temperatures.

The *Campylobacter* isolates were tested for antibiotic resistance using standard disc diffusion method and Minimum Inhibitory Concentration (M.I.C) method. The *Campylobacter* isolates were tested against 12 antibiotics and showed resistance to clindamycin and nalidixic acid (50.9%) each, cefotaxime (49.1%), sulfamethoxazole-trimethoprim (40%), ampicillin (38.2%), ciprofloxacin (23.6%), enrofloxacin and streptomycin (21.8%), tetracycline (20%), erythromycin (18.2%), chloramphenicol (16.4%) and gentamicin (12.7%) by disc diffusion method. For M.I.C method using M.I.C.Evaluator strips, the isolates were tested against four antibiotics. The isolates were found resistant to ampicillin and tetracycline (26.3%), ciprofloxacin (21%) and erythromycin (15.8%). All the isolated *Campylobacter* spp. in this study were resistant to five antibiotics namely ampicillin, clindamycin, nalidixic acid, streptomycin and cefotaxime. The resistance rates between the two methods for four antibiotics were found comparable. There is almost perfect agreement of kappa test statistic for ampicillin, erythromycin and ciprofloxacin ($kappa > 0.8$) and also good agreement for tetracycline ($0.6 \leq k < 0.8$) between both methods. Multidrug resistance, that is resistant to three or more antibiotic classes, was high, at 52.7%. Multidrug resistant *Campylobacter* isolates poses a significant risk if they are resistant to the drugs of choice and alternative drugs for treatment.

It can be concluded from this study that *Campylobacter* species are quite prevalent at 26.1% in cattle in the farms. The presence of *Campylobacter* in cattle and milk could be a potential source of human infections and environmental contamination. Hence, it is recommended that good animal husbandry practices (GAHP) and good milking procedures must be practiced at the farms and good manufacturing procedures (GMP) at abattoirs where it may reduce the risk to humans through meat, milk and environment. The use of antibiotics in animals should also be controlled and monitored to reduce antibiotic resistance.

Abstrak tesis dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan bagi Ijazah Sarjana Sains Veterinar

KEHADIRAN *CAMPYLOBACTER* SPP. DAN PROFIL KERINTANGAN ANTIBIOTIK DI DALAM LEMBU DAN PERSEKITARAN LADANG

Oleh

WINT WINT AUNG

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Campylobacter, terutama *C. jejuni* dan *C. coli* telah dikenali sebagai salah satu agen penyebab jangkitan gastrousus pada manusia di seluruh dunia. Sumber utama jangkitan pada manusia adalah daging ayam mentah atau kurang dimasak, tetapi daging lembu, daging babi, susu segar dan air telah juga dikaitkan dengan jangkitan tersebut. Kebanyakan kajian yang telah dilakukan di Malaysia adalah pada ayam dan produk ayam. Kajian kehadiran *Campylobacter* pada lembu, daging lembu dan susu adalah kurang dan amat sukar diperolehi. Objektif kajian ini adalah untuk menentukan kehadiran *Campylobacter* pada lembu, persekitaran ladang, susu dan daging, mengenalpasti isolat *Campylobacter* mengguna kaedah fenotipik dan asei m-PCR dan juga mengkaji corak kerintangan antibiotik isolat. Satu ratus lapan puluh (180) sampel calitan rektal lembu, 68 sampel persekitaran ladang lembu, 36 sampel susu segar daripada enam buah ladang dan 30 sampel daging lembu di empat pasar basah telah diambil. Kesemua sampel telah dikultur pada media selektif dan spesis *Campylobacter* yang diasingkan telah dikenalpasti dan dispesis menggunakan PCR multipleks. Prevalenkeseluruhan spesis *Campylobacter* pada lembu tenusu dan lembu pedaging adalah 26.1%. Prevalen adalah lebih tinggi pada lembu pedaging (31.6%) berbanding lembu tenusu (23.6%) tetapi perbezaannya adalah tidak signifikan ($p=0.256$). Prevalen adalah secara signifikan lebih tinggi pada anak lembu (40%) daripada lembu dewasa (22.1%) ($p=0.023$). *Campylobacter* lebih banyak diasingkan pada suhu 42°C (25.0%) berbanding pada 37°C (21.1%), walau bagaimanapun perbezaannya adalah tidak signifikan ($p=0.381$) dan ujian statistik kappa menunjukkan persetujuan hampir sempurna di antara dua suhu tersebut ($\text{kappa}>0.8$). Enam spesis *Campylobacter* telah dikenalpasti pada kedua-dua suhu; spesis yang paling kerap diasingkan adalah *C. jejuni* (39.6%) dan diikuti oleh *C. fetus* (22.4%), *C. upsaliensis* (13.8%), *C. coli* (8.6%), *C. hyointestinalis* subsp. *hyointestinalis* (6.9%) dan paling kurang adalah *C. lari* (5.2%). Walau bagaimanapun, dua isolat spesis *Campylobacter* tidak dapat dikenalpasti. Daripada sampel persekitaran, sejumlah 27.9% spesis *Campylobacter* telah diasingkan, iaitu daripada air (37.0%), lalat (25.0%), makanan ternakan (14.3%), lantai kandang (33.3%) dan bekas minuman (11.1%) yang telah dianggap sebagai faktor-faktor berisiko bagi

Campylobacter pada lembu. Lalat boleh menjadi vektor penting bagi pemindahan *Campylobacter* dari persekitaran tercemar kepada tenakan lembu di ladang atau dari haiwan terjangkau kepada persekitaran. Kehadiran *Campylobacter* pada makanan haiwan, lantai, air minuman dan bekas minuman boleh melalui pencemaran lalat dan tinja haiwan. Dua puluh tujuh persepuluh lapan (27.8%) sampel susu segar didapati positif *Campylobacter*, walau bagaimanapun tiada satu pun daripada 30 sampel daging lembu yang positif. Kehadiran *Campylobacter* pada susu boleh terhasil daripada pencemaran semasa pemerahan susu. Ketiadaan *Campylobacter* pada daging lembu mungkin ianya tidak dicemari semasa pemprosesan, dan juga kerintangan lemah *Campylobacter* terhadap atmosfera oksigen dan lain-lain tekanan persekitaran semasa penyimpanan, pengangkutan dan penjualan boleh menyebabkan *Campylobacter* bertukar kepada bentuk berdaya hidup tetapi tidak boleh dikultur (VBNC). Kadar keseluruhan pengasingan *Campylobacter* pada lembu, sampel persekitaran, daging lembu dan susu apabila diinkubasi di bawah dua suhu berbeza adalah lebih tinggi pada 42°C (22.6%) berbanding 37°C (18.5%); walaubagaimanapun, tiada perbezaan signifikan ($p=0.199$) dan ujian statistik kappa menunjukkan persetujuan baik di antara dua suhu inkubasi yang berbeza ($0.6 \leq k < 0.8$). Enam spesies *Campylobacter* telah diasingkan pada kedua-dua suhu.

Isolat *Campylobacter* telah diuji kerintangan antibiotik dengan menggunakan kaedah disc diffusion dan kaedah *Minimum Inhibitory Concentration (M.I.C)*. Isolat *Campylobacter* telah diuji terhadap 12 antibiotik dan menunjukkan kerintangan terhadap setiap satu clindamycin dan nalidixic acid (50.9%), cefotaxime (49.1%), sulfamethoxazole-trimethoprim (40%), ampicillin (38.2%), ciprofloxacin (23.6%), enrofloxacin dan streptomycin (21.8%), tetracycline (20%), erythromycin (18.2%), chloramphenicol (16.4%) dan gentamicin (12.7%) melalui kaedah disc diffusion. Bagi kaedah *M.I.C* menggunakan strip *M.I.C. Evaluator*, isolat telah diuji terhadap empat antibiotik. Isolat telah didapati rintang terhadap ampicillin dan tetracycline (26.3%), ciprofloxacin (21%) dan erythromycin (15.8%). Kesemua isolat *Campylobacter* spp. di dalam kajian ini adalah rintang terhadap lima antibiotik iaitu ampicillin, clindamycin, nalidixic acid, streptomycin dan cefotaxime. Kadar kerintangan di antara dua kaedah bagi empat antibiotik didapati setanding. Terdapat persetujuan hampir sempurna bagi ujian statistik kappa bagi ampicillin, erythromycin dan ciprofloxacin ($kappa > 0.8$) dan juga persetujuan baik bagi tetracycline ($0.6 \leq k < 0.8$) di antara kedua-dua kaedah. Kerintangan multidrug, iaitu rintang kepada tiga atau lebih kelas antibiotik, adalah tinggi, 52.7%. Isolat multidrug rintang *Campylobacter* boleh menyebabkan risiko signifikan sekiranya ia rintang kepada drug pilihan dan drug alternatif untuk rawatan.

Daripada kajian ini, dapat disimpulkan bahawa spesies *Campylobacter* adalah agak prevalen 26.1% pada ternakan lembu di ladang. Kehadiran *Campylobacter* pada lembu dan susu boleh menjadi sumber yang berpotensi menyebabkan jangkitan pada manusia dan pencemaran persekitaran. Oleh yang demikian, Amalan Penternakan Haiwan Baik (GAHP) dan prosedur pemerahan susu yang baik perlu dilaksanakan di ladang serta Prosedur Pengeluaran Baik (GMP) di rumah sembelih yang dapat mengurangkan risiko kepada manusia melalui daging, susu dan persekitaran. Penggunaan antibiotik pada haiwan perlu dikawal dan dipantau bagi mengurangkan kerintangan antibiotik.

ACKNOWLEDGEMENTS

My warmest appreciation goes to my supervisor Prof. Dr. Saleha Abdul Aziz, the chairman of Supervisory Committee for her continuous encouragement, care and excellent scientific guidance during the course of this study. I deeply appreciate her patience, understanding and invaluable advice. I would like to express my special thanks to my co-supervisors, Assoc. Prof. Dr. Zunita Zakaria and Dr. Murugaiyah Marimuthu for their compassion, supervision, encouragement, valuable comments and suggestions.

I would like to take this opportunity to acknowledge my appreciation to SEARCA (South East Asian Regional Centre for Graduate Study and Research in Agriculture), for the financial assistance.

I would like to express my appreciation and special gratitude to Brigadier General U Ohn Myint (Minister, Ministry of Livestock Fisheries and Rural Development), U Khin Mg Aye, Dr. Aung Myat Oo, U Tin Ngwe (Deputy Ministers, Ministry of Livestock Fisheries and Rural Development), Colonel Dr. Myint Than (Director General, Livestock Breeding and Veterinary Department, Ministry of Livestock Fisheries and Rural Development), Prof. Dr. Myint Thein (formerly Director General, LBVD), Dr. Khin Zaw (Deputy Director General, LBVD), Dr. Win Myint (Director, Animal Health and Development Section), Dr. Khin Mg Aye (Deputy Director, Animal Health and Development Section), Dr. Khin Mg Oo and Dr. Yin Yin San (Assistant Directors, Artificial Insemination and Research and Development Section) for allowing me to pursue this postgraduate programme. My thanks to all my colleagues in department for their kind taking over my duties during my postgraduate study.

Grateful acknowledgement and sincere appreciation are extended to Puan Fauziah Nordin, staff of Veterinary Public Health Laboratory and all of my lab mates, Rasheed, Emelia, Teguh, Dauda Goni, Abdelrahman, Yousif, and Jalo for sharing their knowledge and experience with me, for their generous help and kindness which enabled me to finish my project smoothly. In addition, I am indeed thankful to Krish and all the staff of Veterinary Bacteriology laboratory for their kindness, guidance, teaching and generous helps to finish the project of my research. My sincere thanks to all colleagues, staff of the faculty who contributed one way or another towards completion of my study.

Last but not least, I express the most gratitude to my beloved parents, my sisters, my brother and my parent-in-laws for their love, understanding, encouragement, support and affection. Special thanks to my friend Khin Thida Khaing who give me endless patience and care during the time stay together in Malaysia and also my sister-in-laws Lei Lei Swe and Toe Toe Lwin who take care of my daughter when I was studying. My deepest gratitude goes to my dearest and nearest: my husband for his endless love and encouragement, and also to my daughter for being understanding when Mum was working and not around as much as you were previously used to. Without their support, surely I would not be able to give attention on my studies.

I certify that a Thesis Examination Committee has met on 13th June 2014 to conduct the final examination of WINT WINT AUNG on her thesis titled “Occurrence of *Campylobacter* in Cattle and Its Farm Environment and their Antibiotic Resistance Profiles” in accordance with Universities and University College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The committee recommends that the student be awarded the Master of Veterinary Science. Members of the Examination Committee are as follows:

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LIST OF ABBREVIATIONS

ATCC	American Type Culture Collection
bp	Base pairs
CBA	Columbia Blood Agar
CCUG	Culture Collection of the University of Goteborg
ceuE	Siderophore enterochelin
CLSI	Clinical Laboratory Standard Institute
cstA	Cystatin-A
°C	Degree Celcius
DNA	Deoxyribonucleic acid
EDTA	Ethylenediaminetetraacetic acid
flaA	Flagellin A gene
glyA	Serine hydroxyl methyl transferase gene
g	Gram (s)
h	Hour (s)
hip	hippuricase gene
lpxA	UDP-N-acetyl glucosamine acetyltransferase
ml	Milliliter
mg	Milligram (s)
min	Minute (s)
mm	Millimeter
MDR	Multidrug resistance
mCCDA	Modified Charcoal Cefoperazone Deoxycholate Agar
MIC	Minimum inhibitory concentration
MICE	Minimum Inhibitory Concentration Evaluator
mPCR	Multiplex Polymerase Chain Reaction
PFGE	Pulsed Field Gel Electrophoresis
RNA	Ribonucleic acid
rRNA	Ribosomal RNA
s	Second (s)
Spp.	Species
TBE	Tris-borate EDTA
UV	Ultraviolet

V	Volt
WHO	World Health Organization
μL	Micro Liter
μg	Micro Gram
μM	Micro Molar



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CHAPTER 1

INTRODUCTION

Campylobacter species are important in veterinary and public health due to their zoonotic nature, colonizing a large variety of reservoir hosts and being environmental persistence (Hannon et al., 2009). In humans, these *Campylobacter* species are well-known causes of food-borne gastroenteritis (Allos, 2001), thus of major public health importance worldwide particularly in industrialized countries (Adhikari et al., 2004). In many European countries, the prevalence of campylobacteriosis continues to increase and today it exceeds the number of salmonellosis cases (Silva et al., 2011). Most of the human foodborne diseases are caused by *Campylobacter jejuni* and *Campylobacter coli* (Uaboi-Egbenni et al., 2012). *Campylobacter* species can be found in the reproductive organs, gastrointestinal tracts, and oral cavities of animals and humans (Dadi & Asrat, 2008). *Campylobacter* species colonize various species of wild and farm animals, principally poultry and birds, as part of their gut microbiota (Neimann et al., 2003; Van Vliet & Ketley, 2001) without causing infection. *Campylobacter* infection is also one of the causes of reproductive disorders in cattle such as poor calving in southern Africa (Schmidt et al., 2010).

In human campylobacteriosis, poultry meat has long been regarded as the major source and cattle may also play an important reservoir host species (Stanley & Jones, 2003). Contamination of human food can arise at any step from the slaughter house, to processing plant to the consumer (Neimann et al., 2003). Detection of *C. jejuni* and *C. coli* on the carcasses is mainly due to contamination from the gastrointestinal contents of slaughtered healthy animals (Nonga et al., 2010). Besides thermophilic *Campylobacter* spp. which included *C. jejuni*, *C. coli*, *C. lari*, *C. hyointestinalis* and *C. lanienae* in cattle may have implication on public health (Sanad et al., 2011; Humphrey et al., 2007; Acik & Cetinkaya, 2005; Logan et al., 2000). Many studies have observed identical strain types between *Campylobacter* species isolated from cattle faeces or from contaminated bovine origin food products and those from infected human (Hakkinen et al., 2009; Gilpin et al., 2008b).

Apart from beef, the existence of foodborne pathogens in milk is also a potential hazard to public health, principally among milk manufacturers, farm workers and their families and those keen on consuming unpasteurized milk (Ryser, 1998). Besides chicken meat, cattle and beef have been implicated in human campylobacteriosis outbreaks and sporadic cases, were generally associated with drinking of unpasteurized milk and consumption of beef (Nielsen, 2002). The contact with cattle faeces via environmental contamination is also regarded as a threat to humans (Garrett et al., 2007). Furthermore, cattle have been involved in the environmental transmission of *Campylobacter* to water (Clark et al., 2003). *Campylobacter* from the faeces of warm blooded animals, birds and infected humans can get into the water and food (Scotter et al., 1993) and that water is not only common as vehicle of *Campylobacter* spread to humans but also to cattle (Besser et al., 2005).

There is increasing scientific confirmation, especially in developed countries concerning the widespread antibiotic usage in food animals that leads to the development of resistant pathogenic microorganisms that can get to humans through the food chain (Marshall & Levy, 2011; Philips et al., 2004). Treatment with antibiotics for uncomplicated *Campylobacter* infection is not common. On the other hand, *Campylobacter* have been increasingly reported to be resistant to antibiotics used for treatment (principally macrolides and fluoroquinolones) (Aarestrup & Engberg, 2001). Antibiotic therapy is mostly considered in severe cases. The frequency of resistance to macrolides among *Campylobacter* spp. is considerable since the 1990s, and it has since been identified as an emerging public health problem (Engberg et al., 2001). Numerous studies have revealed that human diseases with fluoroquinolone-resistant (FQR) *Campylobacter* have increased worldwide, corresponding with the use of fluoroquinolones in animal agriculture (Serichantalergs et al., 2007; Gupta et al., 2004; Engberg et al., 2001).

The occurrence of *Campylobacter* species in cattle has been studied in countries such as United States, Turkey, New Zealand, Nigeria, Southern Chile, Canada, UK, Tanzania, USA (Sanad et al., 2011; Grove-White et al., 2010; Nonga et al., 2010; Salihu et al., 2009; Fernández & Hirschfeld, 2009; Hannon et al., 2009; Gilpin et al., 2008b; Bae et al., 2005; Açık & Cetinkaya, 2005) but there is very few information on the occurrence of *Campylobacter* in cattle in Malaysia. There is a need to know the extent of *Campylobacter* infection in cattle and the presence of *Campylobacter* in farm environment, milk and meat.

Thus, the objectives of this study were:

1. to determine the occurrence of *Campylobacter* in dairy and beef cattle, their farm environment, milk and meat.
2. to identify the *Campylobacter* isolates by phenotypic method and multiplex PCR assay.
3. to determine the antibiotic resistance patterns among *Campylobacter* isolates.

REFERENCES

- Aarestrup, F. M., & Engberg, J. (2001). Antimicrobial resistance of thermophilic *Campylobacter*. *Veterinary Research*, 32(3-4), 311–321.
- Abe, T., Haga, S., Yokoyama, K., & Watanabe, N. (2008). An outbreak of *Campylobacter jejuni* subsp. *jejuni* infection via tap water. *Japanese Journal of Infectious Diseases*, 61(4), 327.
- Abu-Halaweh, M., Bates, J., & Patel, B. K. (2005). Rapid detection and differentiation of pathogenic *Campylobacter jejuni* and *Campylobacter coli* by real-time PCR. *Research in microbiology*, 156(1), 107-114.
- Abulreesh, H. H., Paget, T.A., & Goulder, R. (2005). Recovery of thermophilic *Campylobacter* from pond water and sediment and the problem of interference by background bacteria in enrichment culture. *Water Research*, 39(13), 2877-82.
- Açik, M. N., & Cetinkaya, B. (2005). The heterogeneity of *Campylobacter jejuni* and *Campylobacter coli* strains isolated from healthy cattle. *Letters in Applied Microbiology*, 41(5), 397–403.
- ACMSF, (2004). Second Report on *Campylobacter*. London: Advisory Committee on the Microbiological Safety of Food.
- Adak, G. K., Long, S. M., & O'Brien, S. J. (2002). Trends in indigenous foodborne disease and deaths, England and Wales: 1992 to 2000. *Gut*, 51(6), 832–41.
- Adhikari, B., Madie, P., Connolly, J., Davies, P., Layland, M., & Rogers, L. (2002). Wild Birds, Flies, and Rodents as Reservoirs of *Campylobacter* spp on Dairy Farm. *MAF, Technical paper, Ministry of Agriculture and Forestry, New Zealand*, (November).
- Adhikari, B., Connolly, J., Madie, P., & Davies, P. (2004). Prevalence and clonal diversity of *Campylobacter jejuni* from dairy farms and urban sources. *New Zealand Veterinary Journal*, 52(6), 378–383.
- Alfredson, D. A., & Korolik, V. (2007). Antibiotic resistance and resistance mechanisms in *Campylobacter jejuni* and *Campylobacter coli*. *FEMS Microbiology Letters*, 277(2), 123–32.
- Ali, A. M., Qureshi, A. H., Rafi, S., Roshan, E., Khan, I., Malik, A. M., & Shahid, S. A. (2003). Frequency of *Campylobacter jejuni* in diarrhoea/dysentery in children in Rawalpindi and Islamabad. *JPMA. The Journal of the Pakistan Medical Association*, 53(11), 517–20.
- Allos, B. M. (1997). Association between *Campylobacter* Infection and Guillain-Barre. *The Journal of Infectious Diseases*, 176(Suppl 2), 125–128.

- Allos, B. M. (2001). *Campylobacter jejuni* Infections: update on emerging issues and trends. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America*, 32(8), 1201–6.
- Alm, R. A, Guerry, P., & Trust, T. J. (1993). The *Campylobacter* sigma 54 flaB flagellin promoter is subject to environmental regulation. *Journal of Bacteriology*, 175(14), 4448–55.
- Altekruse, S. F., Swerdlow, D. L., & Stern, N. J. (1998). Microbial food borne pathogens. *Campylobacter jejuni*. *The Veterinary clinics of North America. Food Animal Practice*, 14(1), 31.
- Altekruse, S. F., & Tollefson, L. K. (2003). Human campylobacteriosis: a challenge for the veterinary profession. *Journal of the American Veterinary Medical Association*, 223(4), 445-452.
- Anonymous,(2007). Notifiable and other diseases in New Zealand Annual Report 2006. Institute of Environmental Science and Research Limited, Kenepuru, New Zealand. Available at: <http://www.surv.esr.cri.nz>.
- Arsenault, J., Letellier, A., Quessy, S., Normand, V., & Boulianne, M. (2007). Prevalence and risk factors for *Salmonella* spp. and *Campylobacter* spp. caecal colonization in broiler chicken and turkey flocks slaughtered in Quebec, Canada. *Preventive Veterinary Medicine*, 81(4), 250–64.
- Atabay, H. I., & Corry, J. E. (1998). The isolation and prevalence of *Campylobacter* from dairy cattle using a variety of methods. *Journal of Applied Microbiology*, 84(5), 733–40.
- Bae, W., Kaya, K. N., Hancock, D. D., Call, D. R., Park, Y. H., & Besser, T. E. (2005). Prevalence and Antimicrobial Resistance of Thermophilic *Campylobacter* spp . from Cattle Farms in Washington State. *Applied and Environmental Microbiology*, 71(1), 169–174.
- Bates, A. C., Hiatt, K. L., & Stern, N. J. (2004). Relationship of *Campylobacter* Isolated from Poultry and from Darkling Beetles in New Zealand. *Avian Diseases*, 48(1), 138–147.
- Belanger, A. E., & Shryock, T. R. (2007). Macrolide-resistant *Campylobacter*: the meat of the matter. *The Journal of Antimicrobial Chemotherapy*, 60(4), 715–23.
- Berry, E. D., Wells, J. E., Archibeque, S. L., Ferrell, C. L., Freetly, H. C., & Miller, D. N. (2006). Influence of genotype and diet on steer performance, manure odor, and carriage of pathogenic and other fecal bacteria. II. Pathogenic and other fecal bacteria. *Journal of animal science*, 84(9), 2523–32.
- Besser, T. E., Lejeune, J. T., Rice, D. H., Berg, J., Stilborn, R. P., Kaya, K., & Hancock, D. D. (2005). Increasing Prevalence of *Campylobacter jejuni* in Feedlot Cattle through the Feeding Period. *Applied and Environmental Microbiology*, 71(10), 5752–5758.

- Blaser, M. J., Hardesty, H. L., Powers, B., & Wang, W. L. (1980). Survival of *Campylobacter fetus* subsp. *jejuni* in biological milieus. *Journal of Clinical Microbiology*, 11(4), 309–313.
- Bolton, F. J., Hutchinson, D. N., & Coates, D. (1984). Blood-free selective medium for isolation of *Campylobacter jejuni* from feces. *Journal of Clinical Microbiology*, 19(2), 169–71.
- Bolton, F. J., Hutchinson, D. N., & Parker, G. (1988). Reassessment of selective agars and filtration techniques for isolation of *Campylobacter* species from faeces. *European Journal of Clinical Microbiology and Infectious Diseases*, 7(2), 155-160.
- Boonmar, S., Yingsakmongkon, S., Songserm, T., Hanhaboon, P., & Passadurak, W. (2007). Detection of *Campylobacter* in duck using standard culture method and multiplex polymerase chain reaction. *The Southeast Asian Journal of Tropical Medicine and Public Health*, 38(4), 728–731.
- Bull, S. A., Allen, V. M., Domingue, G., Jørgensen, F., Ure, R., Whyte, R., & Frost, J. A. (2006). Sources of *Campylobacter* spp . Colonizing Housed Broiler Flocks during Rearing. *Applied and Environmental Microbiology*, 72(1), 645–652.
- Butzler, J.P. (2004). *Campylobacter*, from obscurity to celebrity. *Clinical Microbiology and Infection: the official publication of the European Society of Clinical Microbiology and Infectious Diseases*, 10(10), 868–76.
- Bywater, R., Deluyker, H., Deroover, E., De Jong, A., Marion, H., McConville, M., & Walters, J. (2004). A European survey of antimicrobial susceptibility among zoonotic and commensal bacteria isolated from food-producing animals. *The Journal of Antimicrobial Chemotherapy*, 54(4), 744–54.
- Cabrita, J., Rodrigues, J., Braganca, F., Morgado, C., Pires, I., & Gonçalves, A. P.(1992). Prevalence, biotypes, plasmid profile and antimicrobial resistance of *Campylobacter* isolated from wild and domestic animals from northeast Portugal. *Journal of Applied Microbiology*, 73(4), 279-285.
- Calciati, E., Lafuente, S., De Simó, M., Balfagon, P., Bartolomé, R., & Caylà, J. (2012). A *Campylobacter* outbreak in a Barcelona school. *Enfermedades Infecciosas y Microbiología Clínica*, 30(5), 243–5.
- Campana, R., Patrone, V., Federici, S., Fulvi, S., & Baffone, W. (2010). Antibiotic Resistance of *Campylobacter* Spp Isolated From Chickens and Humans in Central Italy. *Journal of Food Safety*, 30(4), 924–940.
- Carrique-Mas, J., Andersson, Y., Hjertqvist, M., Svensson, A., Torner, A., & Giesecke, J. (2005). Risk factors for domestic sporadic campylobacteriosis among young children in Sweden. *Scandinavian Journal of Infectious Diseases*, 37(2), 101–10.

- Casadémont, I., Chevrier, D., & Guesdon, J. L. (1998). Cloning of a sapB homologue (sapB2) encoding a putative 112-kDa *Campylobacter fetus* S-layer protein and its use for identification and molecular genotyping. *FEMS Immunology & Medical Microbiology*, 21(4), 269-281.
- Chai, L. C., Robin, T., Ragavan, U. M., Gunsalam, J. W., Bakar, F. A., Ghazali, F. M., & Kumar, M. P. (2007). Thermophilic *Campylobacter* spp. in salad vegetables in Malaysia. *International Journal of Food Microbiology*, 117(1), 106–11.
- Chai, L. C., Ghazali, F. M., Bakar, F. A., Lee, H. Y., Suhaimi, L. R., Talib, S. A., & Radu, S. (2009). Occurrence of thermophilic *Campylobacter* spp. contamination on vegetable farms in Malaysia. *Journal of Microbiology and Biotechnology*, 19(11), 1415-1420.
- Chatre, P., Haenni, M., Meunier, D., Botrel, M. A., Calavas, D., & Madec, J. Y. (2010). Prevalence and antimicrobial resistance of *Campylobacter jejuni* and *Campylobacter coli* isolated from cattle between 2002 and 2006 in France. *Journal of Food Protection*®, 73(5), 825-831.
- Choo, L. C., Saleha, A. A., Wai, S. S., & Fauziah, N. (2011). Isolation of *Campylobacter* and *Salmonella* from houseflies (*Musca domestica*) in a university campus and a poultry farm in Selangor, Malaysia. *Tropical Biomedicine*, 28(1), 16-20.
- Chu, Y., Chu, M., Luey, K., Ngan, Y., Tsang, K., & Kam, K. (2004). Genetic Relatedness and Quinolone Resistance of *Campylobacter jejuni* Strains Isolated in 2002 in Hong Kong. *Journal of Clinical Microbiology*, 42(7), 3321–3323.
- Clark, C. G., Price, L., Ahmed, R., Woodward, D. L., Melito, P. L., Rodgers, F. G., & Ellis, A. (2003). Characterization of Waterborne Outbreak – associated Walkerton, Ontario. *Emerging Infectious Diseases*, 9(10), 1232–1241.
- CLSI, (2012). Performance standards for antimicrobial susceptibility testing; Twenty-second informational supplement. M100-S22, vol 32, No.3.
- CLSI, (2013). Performance standards for antimicrobial disk and dilution susceptibility tests for bacteria isolated from animals; Second Informational Supplement. VET01-S2, vol.33, No.8.
- Coker, A. O., Isokpehi, R. D., Thomas, B. N., Amisu, K. O., & Obi, C. L. (2002). Human campylobacteriosis in developing countries. *Emerging Infectious Diseases*, 8(3), 237–44.
- Cools, I., D'Haese, E., Uyttendaele, M., Storms, E., Nelis, H. J., & Debevere, J. (2005). Solid phase cytometry as a tool to detect viable but non-culturable cells of *Campylobacter jejuni*. *Journal of Microbiological Methods*, 63, 107–114.

- Corcoran, D., Quinn, T., Cotter, L., Whyte, P., & Fanning, S. (2006). Antimicrobial resistance profiling and fla-typing of Irish thermophilic *Campylobacter* spp. of human and poultry origin. *Letters in Applied Microbiology*, 43(5), 560–5.
- Cornelius, A. J., Nicol, C., & Hudson, J. A. (2005). *Campylobacter* spp. in New Zealand raw sheep liver and human campylobacteriosis cases. *International Journal of Food Microbiology*, 99(1), 99–105.
- Corry, J.E.L., Post, D.E., Colin, P., & Laisney, M. J. (1995). Culture media for isolation of *Campylobacter*. *International Journal of Food Microbiology*, 26, 43–76.
- Dadi, L., & Asrat, D. (2008). Prevalence and antimicrobial susceptibility profiles of thermotolerant *Campylobacter* strains in retail raw meat products in Ethiopia. *Ethiopian Journal of Health Development*, 22(2), 195–200.
- Debruyne, L., Samyn, E., De Brandt, E., Vandenberg, O., Heyndrickx, M., & Vandamme, P. (2008a). Comparative performance of different PCR assays for the identification of *Campylobacter jejuni* and *Campylobacter coli*. *Research in Microbiology*, 159(2), 88–93.
- Debruyne, L., Gevers, D., & Vandamme, P. (2008b). Taxonomy of the family *Campylobacteraceae*. In eds Nachamkin, I., Szymanski, C. M., & Blaser, M. J. *Campylobacter*, (Ed. 3), Washington, DC: ASM Press, 3-25.
- Debruyne, L., On, S. L. W., De Brandt, E., & Vandamme, P. (2009). Novel *Campylobacter lari*-like bacteria from humans and molluscs: description of *Campylobacter peloridis* sp. nov., *Campylobacter lari* subsp. *concheus* subsp. nov. and *Campylobacter lari* subsp. *lari* subsp. nov. *International Journal of Systematic and Evolutionary Microbiology*, 59(Pt 5), 1126–32.
- Debruyne, L., Broman, T., Bergström, S., Olsen, B., On, S. L. W., & Vandamme, P. (2010a). *Campylobacter subantarcticus* sp. nov., isolated from birds in the sub-Antarctic region. *International Journal of Systematic and Evolutionary Microbiology*, 60(Pt 4), 815–9.
- Debruyne, L., Broman, T., Bergström, S., Olsen, B., On, S. L. W., & Vandamme, P. (2010b). *Campylobacter volucris* sp. nov., isolated from black-headed gulls (*Larus ridibundus*). *International Journal of Systematic and Evolutionary Microbiology*, 60(Pt 8), 1870–5.
- Denis, M., Soumet, C., Rivoal, K., Ermel, G., Blivet, D., Salvat, G., & Colin, P. (1999). Development of a m-PCR assay for simultaneous identification of *Campylobacter jejuni* and *C. coli*. *Letters in Applied Microbiology*, 29(6), 406–10.
- Denis, M., Tanguy, M., Chidaine, B., Laisney, M.-J., Mégraud, F., & Fravallo, P. (2011). Description and sources of contamination by *Campylobacter* spp. of river water destined for human consumption in Brittany, France. *Pathologie-Biologie*, 59(5), 256–63.

- Devlin, H. R., & McIntyre, L. (1983). *Campylobacter fetus* subsp. *fetus* in homosexual males. *Journal of Clinical Microbiology*, 18(4), 999-1000.
- Diker, K. S., Diker, S., & Özlem, M. B. (1990). Bovine diarrhea associated with *Campylobacter hyointestinalis*. *Journal of Veterinary Medicine, Series B*, 37(1-10), 158-162.
- El-Sharoud, W. M. (2009). Prevalence and survival of *Campylobacter* in Egyptian dairy products. *Food Research International*, 42(5-6), 622–626.
- Ellis-Iversen, J., Cook, A. J. C., Smith, R. P., Pritchard, G. C., & Nielsen, M. (2009a). Temporal patterns and risk factors for *Escherichia coli* O157 and *Campylobacter* spp, in young cattle. *Journal of Food Protection*, 72(3), 490–6.
- Ellis-Iversen, J., Pritchard, G. C., Wooldridge, M., & Nielsen, M. (2009b). Risk factors for *Campylobacter jejuni* and *Campylobacter coli* in young cattle on English and Welsh farms. *Preventive Veterinary Medicine*, 88(1), 42–8.
- Engberg, J., On, S. L., Harrington, C. S., & Gerner-Smidt, P. (2000a). Prevalence of *Campylobacter*, *Arcobacter*, *Helicobacter* and *Sutterella* spp. in Human Fecal Samples as Estimated by a Reevaluation of Isolation Methods for *Campylobacter*. *Journal of Clinical Microbiology*, 38(1), 286-291.
- Engberg J, Hansen J. L., & Gerner-Smidt, P. (2000b). Examination of optimal culture temperatures for thermophilic *Campylobacter* species. (Danish). *Nytm Mikrobiologi*, 52.
- Engberg, J, Aarestrup, F. M., Taylor, D. E., Gerner-Smidt, P., & Nachamkin, I. (2001). Quinolone and macrolide resistance in *Campylobacter jejuni* and *C. coli*: resistance mechanisms and trends in human isolates. *Emerging Infectious Diseases*, 7(1), 24–34.
- Engberg, J. (2006). Contributions to the epidemiology of *Campylobacter* infections. A review of clinical and microbiological studies. *Danish Medical Bulletin*, 53(4), 361–89.
- Englen, M. D., Hill, A. E., Dargatz, D. A, Ladely, S. R., & Fedorka-Cray, P. J. (2007). Prevalence and antimicrobial resistance of *Campylobacter* in US dairy cattle. *Journal of Applied Microbiology*, 102(6), 1570–7.
- Engvall, A. (2001). May organically farmed animals pose a risk for *Campylobacter* infections in humans? *Acta Veterinaria Scandinavica Supplementum*, 95, 85–7.
- Eyers, M., Chapelle, S., Van Camp, G., Goossens, H., & De Wachter, R. (1993). Discrimination among thermophilic *Campylobacter* species by polymerase chain reaction amplification of 23S rRNA gene fragments. *Journal of Clinical Microbiology*, 32(6), 1623.

- Feierl, G., Berghold, C., Furab, T., & Marth, E. (1999). Further increase in ciprofloxacin-resistant *Campylobacter jejuni/coli* in Styria, Austria. *Clinical Microbiology and Infection*, 5, 59–60.
- Fernandez, H., Salazar, R., & Landskron, E. (1993). Occurrence of thermotolerant species of *Campylobacter* in three groups of hens maintained under different environmental conditions. *Revista de Microbiologia*, 24(4), 265-8.
- Fernández, H., & Hitschfeld, M. (2009). Occurrence of *Campylobacter jejuni* and *Campylobacter coli* and their biotypes in beef and dairy cattle from the South of Chile. *Brazilian Journal of Microbiology*, 40(3), 450–454.
- Fitzgerald, C., Whichard, J., & Nachamkin, I. (2008). Diagnosis and antimicrobial susceptibility of *Campylobacter* species. In eds Nachamkin, I., Szymanski, C. M., & Blaser, M. J. *Campylobacter*, (Ed. 3), Washington, DC: ASM Press 227-243.
- French, N. P., Midwinter, A., Holland, B., Collins-Emerson, J., Pattison, R., Colles, F., & Carter, P. (2009). Molecular epidemiology of *Campylobacter jejuni* isolates from wild-bird fecal material in children's playgrounds. *Applied and Environmental Microbiology*, 75(3), 779–83.
- Friedman, C. R., Hoekstra, R. M., Samuel, M., Marcus, R., Bender, J., Shiferaw, B., & Tauxe, R. V. (2004). Risk factors for sporadic *Campylobacter* infection in the United States: A case-control study in FoodNet sites. *Clinical Infectious Disease*, 38(Suppl 3), S285–96.
- Gallay, A., De Valk, H., Cournot, M., Ladeuil, B., Hemery, C., Castor, C., & Desenclos, J. C. (2006). A large multi-pathogen waterborne community outbreak linked to faecal contamination of a groundwater system, France, 2000. *Clinical Microbiology and Infection*, 12(6), 561–70.
- Gallay, A., Bousquet, V., Siret, V., Prouzet-Mauléon, V., De Valk, H., Vaillant, V., & Desenclos, J. C. (2008). Risk factors for acquiring sporadic *Campylobacter* infection in France: results from a national case-control study. *The Journal of Infectious Diseases*, 197(10), 1477–84.
- Garcia, M. M., Ruckerbauer, G. M., Eaglesome, M. D., & Boisclair, W. E. (1983). Detection of *Campylobacter fetus* in artificial insemination bulls with a transport enrichment medium. *Canadian Journal of Comparative Medicine*, 47(3), 336.
- Garcia, M. M., Lior, H., Stewart, R. B., Ruckerbauer, G. M., Trudel, J. R., & Skljarevski, A. (1985). Isolation, characterization, and serotyping of *Campylobacter jejuni* and *Campylobacter coli* from slaughter cattle. *Applied and Environmental Microbiology*, 49(3), 667–72.
- Garénaux, A., Jugiau, F., Rama, F., De Jonge, R., Denis, M., Federighi, M., & Ritz, M. (2008). Survival of *Campylobacter jejuni* strains from different origins

- under oxidative stress conditions: effect of temperature. *Current Microbiology*, 56(4), 293–7.
- Garrett, N., Devane, M. L., Hudson, J. A., Nicol, C., Ball, A., Klena, J. D., & Savill, M. G. (2007). Statistical comparison of *Campylobacter jejuni* subtypes from human cases and environmental sources. *Journal of Applied Microbiology*, 103(6), 2113–21.
- Gaudreau, C., & Gilbert, H. (1998). Antimicrobial Resistance of Clinical Strains of *Campylobacter jejuni* subsp. *jejuni* Isolated from 1985 to 1997 in Quebec, Canada. *Antimicrobial Agents and Chemotherapy*, 42(8), 2106–2108.
- Gaudreau, C., & Gilbert, H. (2003). Antimicrobial Resistance of *Campylobacter jejuni* subsp. *jejuni* Strains Isolated from Humans in 1998 to 2001 in Montréal, Canada. *Antimicrobial Agents and Chemotherapy*, 47(6), 2027–2029.
- Gblossi Bernadette, G., Eric Essoh, A., Elise Solange, K.N., Natalie, G., Souleymane, B., Lamine Sébastien, N., & Mireille, D. (2012). Prevalence and Antimicrobial Resistance of Thermophilic *Campylobacter* Isolated from Chicken in Côte d'Ivoire. *International Journal of Microbiology*, 2012, 150612.
- Ge, B., Bodeis, S., Walker, R.D., White, D. G., Zhao, S., McDermott, P. F., & Meng, J. (2002). Comparison of the Etest and agar dilution for in vitro antimicrobial susceptibility testing of *Campylobacter*. *Journal of Antimicrobial Chemotherapy*, 50(4), 487–494.
- Ge, B., White, D. G., McDermott, P. F., Girard, W., Zhao, S., Hubert, S., & Meng, J. (2003). Antimicrobial-Resistant *Campylobacter* Species from Retail Raw Meats. *Applied and Environmental Microbiology*, 69(5), 3005–3007.
- Gebhart, C. J., Edmonds, P., Ward, G. E., Kurtz, H. J., & Brenner, D. O. N. J. (1985). "*Campylobacter hyointestinalis*" sp. nov.: a new species of *Campylobacter* found in the intestines of pigs and other animals. *Journal of Clinical Microbiology*, 21(5), 715–720.
- Gee, B., Nye, K. J., Fallon, D., Messer, S., Howe, S., Warren, R. E., & Andrews, N. (2002). Effect of incubation temperature on the isolation of thermophilic species of *Campylobacter* from faeces. *Communicable Disease and Public Health/PHLS*, 5(4), 282.
- Gill, C. O., & Harris, L. M. (1982). Contamination of red-meat carcasses by *Campylobacter fetus* subsp. *jejuni*. *Applied and Environmental Microbiology*, 43(5), 977–80.
- Gilpin, B. J., Scholes, P., Robson, B., & Savill, M. G. (2008a). The transmission of thermotolerant *Campylobacter* spp. to people living or working on dairy farms in New Zealand. *Zoonoses and Public Health*, 55(7), 352–60.
- Gilpin, B. J., Thorrold, B., Scholes, P., Longhurst, R. D., Devane, M., Nicol, C., & Savill, M. (2008 b). Comparison of *Campylobacter jejuni* genotypes from dairy

cattle and human sources from the Matamata-Piako District of New Zealand. *Journal of Applied Microbiology*, 105(5), 1354–60.

- Gonzalez, I., Grant, K. A., Richardson, P. T., Park, S. F., & Collins, M. D. (1997). Specific identification of the enteropathogens *Campylobacter jejuni* and *Campylobacter coli* by using a PCR test based on the *ceuE* gene encoding a putative virulence determinant. *Journal of Clinical Microbiology*, 35(3), 759–763.
- Gorkiewicz, G., Feierl, G., Zechner, R., & Zechner, E. L. (2002). Transmission of *Campylobacter hyointestinalis* from a Pig to a Human. *Journal of Clinical Microbiology*, 40(7), 2601–2605.
- Gorkiewicz, G., Feierl, G., Schober, C., Dieber, F., Ko, J., Zechner, R., & Zechner, E. L. (2003). Species-Specific Identification of *Campylobacter* by Partial 16S rRNA Gene Sequencing. *Journal of Clinical Microbiology*, 41(6), 2537–2546.
- Griffiths, P. L., & Park, R. W. (1990). *Campylobacter* associated with human diarrhoeal disease. *The Journal of Applied Bacteriology*, 69(3), 281–301.
- Grove-White, D. H., Leatherbarrow, A. J. H., Cripps, P. J., Diggle, P. J., & French, N. P. (2010). Temporal and farm-management-associated variation in the faecal-pat prevalence of *Campylobacter jejuni* in ruminants. *Epidemiology and Infection*, 138(4), 549–58.
- Gupta, P. S., Nair, G. B., Mondal, S., Gupta, D. N., Sen, D., Sikdar, S. N., & Pal, S. C. (1991). Epidemiology of campylobacteriosis in a cohort of rural population near Calcutta. *Epidemiology and Infection. London, New York NY*, 106(3), 507–512.
- Gupta, A., Nelson, J. M., Barrett, T. J., Tauxe, R. V, Rossiter, S. P., Friedman, C. R., & Working, N. (2004). Antimicrobial resistance among *Campylobacter* strains, United States, 1997–2001. *Emerging Infectious Diseases*, 10(6), 1102–1109.
- Hakkinen, M., Heiska, H., & Hänninen, M.L. (2007). Prevalence of *Campylobacter* spp. in cattle in Finland and antimicrobial susceptibilities of bovine *Campylobacter jejuni* strains. *Applied and Environmental Microbiology*, 73(10), 3232–8.
- Hakkinen, M., Nakari, U. M., & Siitonen, A. (2009). Chickens and cattle as sources of sporadic domestically acquired *Campylobacter jejuni* infections in Finland. *Applied and Environmental Microbiology*, 75(16), 5244–5249.
- Hald, B., Bang, D. D., Pedersen, K., Dybdahl, J., Madsen, M., & Study, T. (2004). Flies and *Campylobacter* Broiler Flocks. *Emerging Infectious Diseases*, 10(8), 1490–1492.
- Hanninen, M., Niskanen, M., & Korhonen, L. (1998). Water as a Reservoir for *Campylobacter jejuni* Infection in Cows Studied by Serotyping and Pulsed-field Gel Electrophoresis. *Journal of Veterinary Medicine*, 45(2), 37–42.

- Hannis, J. C., Manalili, S. M., Hall, T. A., Ranken, R., White, N., Sampath, R., & Hofstadler, S. A. (2008). High-resolution genotyping of *Campylobacter* species by use of PCR and high-throughput mass spectrometry. *Journal of Clinical Microbiology*, 46(4), 1220–5.
- Hannon, S. J., Allan, B., Waldner, C., Russell, M. L., Potter, A., Babiuk, L. A., & Townsend, H. G. G. (2009). Prevalence and risk factor investigation of *Campylobacter* species in beef cattle feces from seven large commercial feedlots in Alberta, Canada. *Canadian Journal of Veterinary Research*, 4(403), 275–282.
- Hannu, T., Mattila, L., Rautelin, H., Pelkonen, P., Lahdenne, P., Siitonen, A., & Leirisalo-Repo, M. (2002). *Campylobacter*-triggered reactive arthritis: a population-based study. *Rheumatology (Oxford, England)*, 41(3), 312–8.
- Hassanain, N. A. (2011). Antimicrobial Resistant *Campylobacter jejuni* Isolated from Humans and Animals in Egypt. *Global Veterinaria*, 6(2), 195–200.
- He, Y., Yao, X., Gunther, N. W., Xie, Y., Tu, S.I., & Shi, X. (2010). Simultaneous Detection and Differentiation of *Campylobacter jejuni*, *C. coli*, and *C. lari* in Chickens Using a Multiplex Real-Time PCR Assay. *Food Analytical Methods*, 3(4), 321–329.
- Heuvelink, A. E., Valkenburgh, S. M., Tilburg, J. J. H. C., Van Heerwaarden, C., Zwartkruis-Nahuis, J. T. M., & De Boer, E. (2007). Public farms: hygiene and zoonotic agents. *Epidemiology and Infection*, 135(7), 1174–83.
- Heuvelink, A. E., Van Heerwaarden, C., Zwartkruis-Nahuis, A., Tilburg, J. J., Bos, M. H., Heilmann, F. G., & De Boer, E. (2009). Two outbreaks of campylobacteriosis associated with the consumption of raw cows' milk. *International Journal of Food Microbiology*, 134(1), 70-74.
- Hindiyeh, M., Jense, S., Hohmann, S., Benett, H., Edwards, C., Aldeen, W., & Carroll, K. C. (2000). Rapid Detection of *Campylobacter jejuni* in Stool Specimens by an Enzyme Immunoassay and Surveillance for *Campylobacter upsaliensis* in the Greater Salt Lake City Area. *Journal of Clinical Microbiology*, 38(8), 3076-3079.
- Hong, J., Kim, J. M., Jung, W. K., Kim, S. H., Bae, W., Koo, H. C., & Park, Y. H. (2007). Prevalence and antibiotic resistance of *Campylobacter* spp. isolated from chicken meat, pork, and beef in Korea, from 2001 to 2006. *Journal of Food Protection*, 70(4), 860–6.
- Hrudey, S. E., & Hrudey, E. J. (2005). *Safe drinking water: Lessons from recent outbreaks in affluent nations*, P.486. IWA Publishing, London.
- Huat, J. T. Y., Aziz, S. A., Abu, J., Ghazali, F. M., Zainazor, T., & Radu, S. (2010). Thermophilic *Campylobacter* spp. Occurrence on Chickens at Farm, Slaughter House and Retail. *International Journal of Poultry Science*, 9(2), 134–138.

- Hudson, J. A., Nicol, C., Wright, J., Whyte, R., & Hasell, S. K. (1999). Seasonal variation of *Campylobacter* types from human cases, veterinary cases, raw chicken, milk and water. *Journal of Applied Microbiology*, 87(1), 115-124.
- Hum, S., Quinn, K., & Brunner, J. (1997). Evaluation of a PCR assay for identification and differentiation of *Campylobacter fetus* subspecies. *Australian Veterinary Journal*, 75(11), 827-831.
- Humphrey, B. Y. T. J., & Beckett, P. (1987). *Campylobacter jejuni* in dairy cows and raw milk. *Epidemiology and Infection*, 98(3), 263-269.
- Humphrey, T., O'Brien, S., & Madsen, M. (2007). *Campylobacter* as zoonotic pathogens: a food production perspective. *International Journal of Food Microbiology*, 117(3), 237-57.
- Hussain, I., Shahid Mahmood, M., Akhtar, M., & Khan, A. (2007). Prevalence of *Campylobacter* species in meat, milk and other food commodities in Pakistan. *Food Microbiology*, 24(3), 219-22.
- Inglis, G. D., Kalischuk, L. D., & Busz, H. W. (2003a). A survey of *Campylobacter* species shed in faeces of beef cattle using polymerase chain reaction. *Canadian Journal of Microbiology*, 49(11), 655-661.
- Inglis, G. D., & Kalischuk, L. D. (2003b). Use of PCR for Direct Detection of *Campylobacter* Species in Bovine Feces†. *Applied and Environmental Microbiology*, 69(6), 3435-3447.
- Inglis, G. D., Kalischuk, L. D., & Busz, H. W. (2004). Chronic shedding of *Campylobacter* species in beef cattle. *Journal of Applied Microbiology*, 97(2), 410-20.
- Inglis, G. D., McAllister, T. A., Busz, H. W., Yanke, L. J., Morck, D. W., Olson, M. E., & Read, R. R. (2005). Effects of Subtherapeutic Administration of Antimicrobial Agents to Beef Cattle on the Prevalence of Antimicrobial Resistance in *Campylobacter jejuni* and *Campylobacter hyointestinalis*†. *Applied and Environmental Microbiology*, 71(7), 3872-3881.
- Inglis, G. D., Morck, D. W., McAllister, T. A., Entz, T., Olson, M. E., Yanke, L. J., & Read, R. R. (2006). Temporal prevalence of antimicrobial resistance in *Campylobacter* spp. from beef cattle in Alberta feedlots. *Applied and Environmental Microbiology*, 72(6), 4088-95.
- Inglis, G. D., Hoar, B. M., Whiteside, D. P., & Morck, D. W. (2007). *Campylobacter canadensis* sp. nov., from captive whooping cranes in Canada. *International Journal of Systematic and Evolutionary Microbiology*, 57(Pt 11), 2636-44.
- Isenbarger, D. W., Hoge, C. W., Srijan, A., Pitarangsi, C., Bodhidatta, L., Hickey, K. W., & Cam, P. D. (2002). Comparative antibiotic resistance of diarrheal pathogens from Vietnam and Thailand, 1996-1999. *Emerging Infectious Diseases*, 8(2), 175-180.

- Jakopanec, I., Borgen, K., Vold, L., Lund, H., Forseth, T., Hannula, R., & Nygård, K. (2008). A large waterborne outbreak of campylobacteriosis in Norway: the need to focus on distribution system safety. *BMC Infectious Diseases*, 128(8).
- Jayarao, B. M., Donaldson, S. C., Straley, B. A., Sawant, A. A., Hegde, N. V., & Brown, J. L. (2006). A survey of foodborne pathogens in bulk tank milk and raw milk consumption among farm families in pennsylvania. *Journal of Dairy Science*, 89(7), 2451–8.
- Johnsen, G., Zimmerman, K., Lindstedt, B.A., Vardund, T., Herikstad, H., & Kapperud, G. (2006). Intestinal carriage of *Campylobacter jejuni* and *Campylobacter coli* among cattle from South-western Norway and comparative genotyping of bovine and human isolates by amplified-fragment length polymorphism. *Acta Veterinaria Scandinavica*, 48(1), 4.
- Jørgensen, F., Bailey, R., Williams, S., Henderson, P., Wareing, D. R. A., Bolton, F. J., & Humphrey, T. J. (2002). Prevalence and numbers of *Salmonella* and *Campylobacter* spp. on raw, whole chickens in relation to sampling methods. *International Journal of Food Microbiology*, 76(1-2), 151–64.
- Kassa, T., Gebre-selassie, S., & Asrat, D. (2005). The prevalence of thermotolerant *Campylobacter* species in food animals in Jimma Zone , southwest Ethiopia. *Ethiopian Journal of Health Development*, 19(3), 225–229.
- Kassa, T., Gebre-Selassie, S., & Asrat, D. (2007). Antimicrobial susceptibility patterns of thermotolerant *Campylobacter* strains isolated from food animals in Ethiopia. *Veterinary Microbiology*, 119(1), 82–7.
- Kaur, T., Singh, J., Huffman, M. A., Petrzelková, K. J., Taylor, N. S., Xu, S., & Fox, J. G. (2011). *Campylobacter troglodytis* sp. nov., isolated from feces of human-habituated wild chimpanzees (*Pan troglodytes schweinfurthii*) in Tanzania. *Applied and Environmental Microbiology*, 77(7), 2366–73.
- Kazemeini, H., & Valizade, Y. (2011). Prevalence of *Campylobacter* Species in Raw Bovine Milk in Isfahan , Iran. *Middle-East Journal of Scientific Research*, 10(5), 664–666.
- Kemp, R., Leatherbarrow, A. J. H., Williams, N. J., Hart, C. A., Clough, H. E., Turner, J., & French, N. P. (2005). Prevalence and Genetic Diversity of *Campylobacter* spp . in Environmental Water Samples from a 100-Square-Kilometer Predominantly Dairy Farming Area. *Applied and Environmental Microbiology*, 71(4), 1876–1882.
- Ketley, J. M. (1997). Pathogenesis of enteric infection by *Campylobacter*. *Microbiology*, 143, 5–21.
- Klena, J. D., Parker, C. T., Knibb, K., Ibbitt, J. C., Devane, P. M. L., Horn, S. T., & Konkel, M. E. (2004). Differentiation of *Campylobacter coli*, *Campylobacter jejuni*, *Campylobacter lari*, and *Campylobacter upsaliensis* by a multiplex PCR

developed from the nucleotide sequence of the lipid A gene *lpxA*. *Journal of Clinical Microbiology*, 42(12), 5549–5557.

- Krueger, N. A., Anderson, R. C., Krueger, W. K., Horne, W. J., Wesley, I. V., Callaway, T. R., & Nisbet, D. J. (2008). Prevalence and concentration of *Campylobacter* in rumen contents and feces in pasture and feedlot-fed cattle. *Foodborne Pathogens and Disease*, 5(5), 571–577.
- Larson, D.J., Wesley, I. V., & Hoffman, L. J.(1992). Use of Oligodeoxynucleotide Probes to Verify *Campylobacter jejuni* as a Cause of Bovine Abortion. *Journal of Veterinary Diagnostic Investigation*, 4(3), pp.348–351.
- Lastovica, A. J., & Allos, B. M. (2008). Clinical significance of *Campylobacter* and related species other than *Campylobacteri jejuni* and *Campylobacter coli*. In eds Nachamkin, I., Szymanski, C. M., & Blaser, M. J. *Campylobacter*, (Ed. 3), Washington, DC: ASM Press, 123–149.
- Lee, A., O'Rourke, J. L., Barrington, P. J., & Trust, T. J. (1986). Mucus colonization as a determinant of pathogenicity in intestinal infection by *Campylobacter jejuni*: a mouse cecal model. *Infection and Immunity*, 51(2), 536–46.
- Leedom, J. M. (2006). Milk of nonhuman origin and infectious diseases in humans. *Clinical Infectious Diseases*, 43(5), 610–5.
- Lim, V. K. E. (2007). Infectious Diarrhoea. *Medical Journal of Malaysia*, 62(3).
- Linton, D., Owen, R. J., & Stanley, J. (1996). Rapid identification by PCR of the genus *Campylobacter* and of five *Campylobacter* species enteropathogenic for man and animals. *Research in Microbiology*, 147(9), 707–18.
- Linton, D., Lawson, A. J., & Owen, R. J. (1997). PCR Detection , Identification to Species Level , and Fingerprinting of *Campylobacter jejuni* and *Campylobacter coli* Direct from Diarrheic Samples. *Journal of Clinical Microbiology*, 35(10), 2568–2572.
- Logan, J. M., Burnens, A., Linton, D., Lawson, A. J., & Stanley, J. (2000). *Campylobacter lanienae* sp. nov., a new species isolated from workers in an abattoir. *International Journal of Systematic and Evolutionary Microbiology*, 50, 865–72.
- Luangtongkum, T., Morishita, T. Y., El-Tayeb, A. B., Ison, A. J., & Zhang, Q. (2007). Comparison of antimicrobial susceptibility testing of *Campylobacter* spp. by the agar dilution and the agar disk diffusion methods. *Journal of Clinical Microbiology*, 45(2), 590–4.
- Luangtongkum, T., Jeon, B., Han, J., Plummer, P., Logue, C. M., & Zhang, Q. (2009). Antibiotic resistance in *Campylobacter*: emergence, transmission and persistence. *Future Microbiology*, 4(2), 189–200.

- Lucey, B., Cryan, B., O'Halloran, F., Wall, P. G., Buckley, T., & Fanning, S. (2002). Trends in antimicrobial susceptibility among isolates of *Campylobacter* species in Ireland and the emergence of resistance to ciprofloxacin. *Veterinary Record*, *151*(11), 317-320.
- Lund, M., Wedderkopp, A., Wainø, M., Nordentoft, S., Bang, D. D., Pedersen, K., & Madsen, M. (2003). Evaluation of PCR for detection of *Campylobacter* in a national broiler surveillance programme in Denmark. *Journal of Applied Microbiology*, *94*(5), 929–35.
- Lynch, O. A., Cagney, C., McDowell, D. A., & Duffy, G. (2011). Occurrence of fastidious *Campylobacter* spp. in fresh meat and poultry using an adapted cultural protocol. *International Journal of Food Microbiology*, *150*(2-3), 171–7.
- Mabote, K. I., Mbewe, M., & Ateba, C. N. (2011). Prevalence of *Campylobacter* Contamination in Fresh Chicken Meat and Milk Obtained from Markets in the North-West Province , South Africa. *Journal of Human Ecology*, *36*(1), 23–28.
- Madden, R. H., Murray, K. A., & Gilmour, A. (2007). Carriage of four bacterial pathogens by beef cattle in Northern Ireland at time of slaughter. *Letters in Applied Microbiology*, *44*(2), 115–119.
- Mansouri-najand, L., Saleha, A. A., & Wai, S. S. (2012). Prevalence of multidrug resistance *Campylobacter jejuni* and *Campylobacter coli* in chickens slaughtered in selected markets, Malaysia. *Tropical Biomedicine*, *29*(2), 231–8.
- Marshall, B. M., & Levy, S. B. (2011). Food animals and antimicrobials: impacts on human health. *Clinical Microbiology Reviews*, *24*(4), 718-733.
- Mazick, A., Ethelberg, S., Møller, N. E., Mølbak, K., & Lisby, M. (2006). An outbreak of *Campylobacter jejuni* associated with consumption of chicken, Copenhagen, 2005. *Euro Surveillace*, *11*(5), 137–139.
- McGill, K., Kelly, L., Madden, R. H., Moran, L., Carroll, C., O'Leary, A., & Whyte, P. (2009). Comparison of disc diffusion and epsilometer (E-test) testing techniques to determine antimicrobial susceptibility of *Campylobacter* isolates of food and human clinical origin. *Journal of Microbiological Methods*, *79*(2), 238-241.
- Michaud, S., Ménard, S., & Arbeit, R. D. (2004). Campylobacteriosis, Eastern Townships, Que ´bec. *Emerging Infectious Diseases*, *10*(10), 1844–1847.
- Miller, W. G., On, S. L., Wang, G., Fontanoz, S., Lastovica, A. J., & Mandrell, R. E. (2005). Extended multilocus sequence typing system for *Campylobacter coli*, *C. lari*, *C. upsaliensis*, and *C. helveticus*. *Journal of Clinical Microbiology*, *43*(5), 2315-2329.
- Milnes, A. S., Stewart, I., Clifton-Hadley, F.A, Davies, R. H., Newell, D. G., Sayers, A. R., & Paiba, G. A. (2008). Intestinal carriage of verocytotoxigenic *Escherichia coli* O157, *Salmonella*, thermophilic *Campylobacter* and *Yersinia*

enterocolitica, in cattle, sheep and pigs at slaughter in Great Britain during 2003. *Epidemiology and Infection*, 136(6), 739–51.

- Minihan, D., Whyte, P., O'Mahony, M., Fanning, S., McGill, K., & Collins, J. D. (2004). *Campylobacter* spp. in Irish feedlot cattle: a longitudinal study involving pre-harvest and harvest phases of the food chain. *Journal of Veterinary Medicine. B, Infectious Diseases and Veterinary Public Health*, 51(1), 28–33.
- Moore, J. E., Caldwell, P. S., Millar, B. C., & Murphy, P. G. (2001a). Occurrence of *Campylobacter* spp. in water in Northern Ireland: implications for public health. *The Ulster Medical Journal*, 70(2), 102–7.
- Moore, J. E., Crowe, M., & Heaney, N. (2001b). Antibiotic resistance in *Campylobacter* spp. isolated from human faeces (1980–2000) and foods (1997–2000) in Northern Ireland: an update. *Journal of Antimicrobial Chemotherapy*, 48(3), 455–457.
- Moore, J. E., Orcorانب, D. C., Ooleyc, J. S. G. D., Anningd, S. F., Uceye, B. L., Atsudaf, M. M., & Hyted, P. W. (2005). *Campylobacter*. *Veterinary Research*, 36, 351–382.
- Müller, W., Hotzel, H., & Schulze, F. (2003). Identification and differentiation of *Campylobacter fetus* subspecies by PCR. *Deutsche Tierärztliche Wochenschrift*, 110(2), 55.
- Nachamkin, I., Ung, H., & Li, M. (2002). Increasing fluoroquinolone resistance in *Campylobacter jejuni*, Pennsylvania, USA, 1982– 2001. *Emerging Infectious Diseases*, 8(12), 1501– 1503.
- Nannapaneni, R., Story, R., Wiggins, K. C., & Johnson, M. G. (2005). Concurrent Quantitation of Total *Campylobacter* and Total Ciprofloxacin-Resistant *Campylobacter* Loads in Rinses from Retail Raw Chicken Carcasses from 2001 to 2003 by Direct Plating at 42°C. *Applied and Environmental Microbiology*, 71(8), 4510–4515.
- Nayak, R., Stewart, T. M., & Nawaz, M. S. (2005). PCR identification of *Campylobacter coli* and *Campylobacter jejuni* by partial sequencing of virulence genes. *Molecular and Cellular Probes*, 19(3), 187–93.
- Neimann, J., Engberg, J., Mølbak, K., & Wegener, H. C. (2003). A case-control study of risk factors for sporadic *Campylobacter* infections in Denmark. *Epidemiology and Infection*, 130(3), 353–66.
- Nelson, J. M., Smith, K. E., Vugia, D. J., Rabatsky-Ehr, T., Segler, S. D., Kassenborg, H. D., & Angulo, F. J. (2004). Prolonged diarrhea due to ciprofloxacin-resistant *Campylobacter* infection. *The Journal of Infectious Diseases*, 190(6), 1150–1157.

- Nesbakken, T., Eckner, K., & Røtterud, O.-J. (2008). The effect of blast chilling on occurrence of human pathogenic *Yersinia enterocolitica* compared to *Campylobacter* spp. and numbers of hygienic indicators on pig carcasses. *International Journal of Food Microbiology*, 123(1-2), 130–3.
- Nichols, G. L. (2005). Fly transmission of *Campylobacter*. *Emerging Infectious Diseases*, 11(3), 361–4.
- Nielsen, E. M., Engberg, J., Fussing, V., Petersen, L., Brogren, C. H., & On, S. L. (2000). Evaluation of phenotypic and genotypic methods for subtyping *Campylobacter jejuni* isolates from humans, poultry, and cattle. *Journal of Clinical Microbiology*, 38(10), 3800–10.
- Nielsen, E. M. (2002). Occurrence and strain diversity of thermophilic *Campylobacter* in cattle of different age groups in dairy herds. *Letters in Applied Microbiology*, 35(1), 85–9.
- Nonga, H. E., Sells, P., & Karimuribo, E. D. (2010). Occurrences of thermophilic *Campylobacter* in cattle slaughtered at Morogoro municipal abattoir, Tanzania. *Tropical Animal Health and Production*, 42(1), 73–8.
- Noormohamed, A., & Fakhr, M. K. (2013). A higher prevalence rate of *Campylobacter* in retail beef livers compared to other beef and pork meat cuts. *International Journal of Environmental Research and Public Health*, 10(5), 2058–68.
- Ogden, I. D., MacRae, M., Johnston, M., Strachan, N. J. C., Cody, A. J., Dingle, K. E., & Newell, D. G. (2007). Use of multilocus sequence typing to investigate the association between the presence of *Campylobacter* spp. in broiler drinking water and *Campylobacter* colonization in broilers. *Applied and Environmental Microbiology*, 73(16), 5125–9.
- OIE, (2008). World organisation for animal health manual of diagnostic tests and vaccines (mammals , birds and bees) Sixth Edition. OIE *Terrestrial Manual*, Volume 2.
- OIE, (2014). List of Antimicrobial Agents of Veterinary Importance. http://www.oie.int/fileadmin/Home/eng/Our_scientific_expertise/docs/pdf/OIE_list_antimicrobials.pdf
- Olah, P., Sherwood, J., Elijah, L., Dockter, M., Doetkott, C., Miller, Z., & Logue, C. (2004). Comparison of antimicrobial resistance in *Salmonella* and *Campylobacter* isolated from turkeys in the Midwest USA. *Food Microbiology*, 21(6), 779–789.
- On, S. L., & Holmes, B. (1991). Effect of inoculum size on the phenotypic characterization of *Campylobacter* species. *Journal of Clinical Microbiology*, 29(5), 923–6.

- On, S. L. (2001). Taxonomy of *Campylobacter*, *Arcobacter*, *Helicobacter* and related bacteria: current status, future prospects and immediate concerns. *Symposium series (Society for Applied Microbiology)*, 90(S6), 1S–15S.
- On, S. L. W., & Jordan, P. J. (2003). Evaluation of 11 PCR Assays for Species-Level Identification of *Campylobacter jejuni* and *C. coli*. *Journal of Clinical Microbiology*, 41(1), 330–336.
- Oporto, B., Esteban, J. I., Aduriz, G., Juste, R. A., & Hurtado, A. (2007). Prevalence and strain diversity of thermophilic *Campylobacter* in cattle, sheep and swine farms. *Journal of Applied Microbiology*, 103(4), 977–84.
- Oporto, B., Juste, R. A., & Hurtado, A. (2009). Phenotypic and Genotypic Antimicrobial Resistance Profiles of *Campylobacter jejuni* Isolated from Cattle, Sheep, and Free-Range Poultry Faeces. *International Journal of Microbiology*, Article ID456573.
- Oyarzabal, O. A., Backert, S., Nagaraj, M., Miller, R. S., Hussain, S. K., & Oyarzabal, E. A. (2007). Efficacy of supplemented buffered peptone water for the isolation of *Campylobacter jejuni* and *C. coli* from broiler retail products. *Journal of Microbiological Methods*, 69(1), 129–36.
- Padungton, P., & Kaneene, J. B. (2003). *Campylobacter* spp in human, chickens, pigs and their antimicrobial resistance. *The Journal of Veterinary Medical Science / the Japanese Society of Veterinary Science*, 65(2), 161–70.
- Parisi, A., Lanzilotta, S. G., Addante, N., Normanno, G., Di Modugno, G., Dambrosio, A., & Montagna, C. O. (2007). Prevalence, molecular characterization and antimicrobial resistance of thermophilic *Campylobacter* isolates from cattle, hens, broilers and broiler meat in south-eastern Italy. *Veterinary Research Communications*, 31(1), 113–23.
- Paulsen, P., Kanzler, P., Hilbert, F., Mayrhofer, S., Baumgartner, S., & Smulders, F. J. M. (2005). Comparison of three methods for detecting *Campylobacter* spp. in chilled or frozen meat. *International Journal of Food Microbiology*, 103(2), 229-33.
- Pellegrin, A. O., Lage, A.P., Serens, J.R.B., Ravaglia, E., Costa, M. S., & Leite, R. (2002). Bovine Genital Campylobacteriosis in Pantanal , State of Mato Grosso do Sul, Brazil. *Revue d'Évage et de Médecine Vétérinaire des pays Tropicaux*, 55(3), 169–173.
- Pena, L. A., & Fishbein, M. C. (2007). Fatal myocarditis related to *Campylobacter jejuni* infection: a case report. *Cardiovascular Pathology*, 16(2), 119-121.
- Phillips, I., Casewell, M., Cox, T., De Groot, B., Friis, C., Jones, R., & Waddell, J. (2004). Does the use of antibiotics in food animals pose a risk to human health? A critical review of published data. *The Journal of Antimicrobial Chemotherapy*, 53(1), 28–52.

- Praakle-Amin, K., Roasto, M., Korkeala, H., & Hänninen, M.-L. (2007). PFGE genotyping and antimicrobial susceptibility of *Campylobacter* in retail poultry meat in Estonia. *International Journal of Food Microbiology*, 114(1), 105–12.
- Puthucheary, S. D., Parasakthi, N., Liew, S. T., & Chee, Y. W. (1994). *Campylobacter* enteritis in children: clinical and laboratory findings in 137 cases. *Singapore Medical Journal*, 35(5), 453.
- Quiñones, B., Parker, C. T., Janda, J. M., Miller, W. G., & Mandrell, R. E. (2007). Detection and genotyping of *Arcobacter* and *Campylobacter* isolates from retail chicken samples by use of DNA oligonucleotide arrays. *Applied and Environmental Microbiology*, 73(11), 3645–55.
- Ragimbeau, C., Salvat, G., Colin, P., & Ermel, G. (1998). Development of a multiplex PCR gene fingerprinting method using *gyrA* and *pflA* polymorphisms to identify genotypic relatedness within *Campylobacter jejuni* species. *Journal of Applied Microbiology*, 85(5), 829–38.
- Rahimi, E., Ameri, M., & Kazemeini, H. R. (2010). Prevalence and antimicrobial resistance of *Campylobacter* species isolated from raw camel, beef, lamb, and goat meat in Iran. *Foodborne Pathogens and Disease*, 7(4), 443-447.
- Rejab, S. B. M., Zessin, K.-H., Fries, R., & Patchanee, P. (2012). *Campylobacter* in chicken carcasses and slaughterhouses in Malaysia. *The Southeast Asian Journal of Tropical Medicine and Public Health*, 43(1), 96–104.
- Riddle, M. S., Sanders, J. W., Putnam, S. D., & Tribble, D. R. (2006). Incidence, etiology, and impact of diarrhea among long-term travelers (US military and similar populations): a systematic review. *The American Journal of Tropical Medicine and Hygiene*, 74(5), 891–900.
- Rollins, D. M., & Colwell, R. R. (1986). Viable but nonculturable stage of *Campylobacter jejuni* and its role in survival in the natural aquatic environment. *Applied and Environmental Microbiology*, 52(3), 531–8.
- Rönner, A.C., Engvall, E. O., Andersson, L., & Kaijser, B. (2004). Species identification by genotyping and determination of antibiotic resistance in *Campylobacter jejuni* and *Campylobacter coli* from humans and chickens in Sweden. *International Journal of Food Microbiology*, 96(2), 173–9.
- Rossi, M., Debruyne, L., Zanoni, R. G., Manfreda, G., Revez, J., & Vandamme, P. (2009). *Campylobacter avium* sp. nov., a hippurate-positive species isolated from poultry. *International Journal of Systematic and Evolutionary Microbiology*, 59(Pt 9), 2364–9.
- Ryser, E. T. (1998) Public health concerns. In: Marth EH, Steele JL (eds) Applied Dairy Microbiology. Marcel Dekker Inc., New York, pp 263-404.
- Sáenz, Y., Zarazaga, M., Lantero, M., Gastanares, M. J., Baquero, F., & Torres, C. (2000). Antibiotic resistance in *Campylobacter* strains isolated from animals,

- foods, and humans in Spain in 1997-1998. *Antimicrobial Agents and Chemotherapy*, 44(2), 267–71.
- Said, B., Wright, F., Nichols, G. L., Reacher, M., & Rutter, M. (2003). Outbreaks of infectious disease associated with private drinking water supplies in England and Wales 1970-2000. *Epidemiology and Infection*, 130(3), 469–79.
- Saito, S., Yatsuyanagi, J., Harata, S., Ito, Y., Shinagawa, K., Suzuki, N., & Enomoto, K. (2005). *Campylobacter jejuni* isolated from retail poultry meat, bovine feces and bile, and human diarrheal samples in Japan: comparison of serotypes and genotypes. *FEMS Immunology and Medical Microbiology*, 45(2), 311–9.
- Salama, S. M., Tabor, H., Richter, M., & Taylor, D. E. (1992). Pulsed-field gel electrophoresis for epidemiologic studies of *Campylobacter hyointestinalis* isolates. *Journal of Clinical Microbiology*, 30(8), 1982–4.
- Saleha, A.A. (2002). Isolation and characterization of *Campylobacter jejuni* from broiler chickens in Malaysia. *Poultry Science*, 1(4), 94–7.
- Saleha, A.A. (2003). Overview of *Campylobacter* in poultry, other animal species and in meat in reference to Malaysia. *Jurnal Veterinar Malaysia*, 15(1-2), 1-6.
- Salihu, M. D., Abdulkadir, J. U., Oboegbulem, S. I., Egwu, G. O., Magaji, A. A., Lawal, M., & Hassan, Y. (2009). Isolation and prevalence of *Campylobacter* species in cattle from Sokoto state, Nigeria. *Veterinaria Italiana*, 45(4), 501–5.
- Salihu, M. D., Junaidu, A. U., Magaji, A. A., & Rabi, Z. M. (2010). Study of *Campylobacter* in Raw Cow Milk in Sokoto State , Nigeria. *British Journal of Dairy Sciences*, 1(1), 1–5.
- Salihu, M. D, Junaidu, A. U, Magaji, A. A, Faleke, O. O, Abubakar, M. B, Tambuwal, F. M, Ahmad, I., Ahmed, A., & Yakubu, Y. (2011). Prevalence and antibiotic resistance of thermophilic *Campylobacter* spp. isolates from raw beef, mutton and camel meat in Sokoto, Nigeria. *Research Opinions in Animal & Veterinary Sciences*, 1(7), 401-405.
- Samie, A., Ramalivhana, J., Igumbor, E. O., & Obi, C. L. (2007). Prevalence, haemolytic and haemagglutination activities and antibiotic susceptibility profiles of *Campylobacter* spp. isolated from human diarrhoeal stools in Vhembe District, South Africa. *Journal of Health, Population, and Nutrition*, 25(4), 406–13.
- Sammarco, M. L., Ripabelli, G., Fanelli, I., Grasso, G. M., & Tamburro, M. (2010). Prevalence and biomolecular characterization of *Campylobacter* spp. isolated from retail meat. *Journal of Food Protection*, 73(4), 720-728.
- Sanad, Y. M., Kassem, I. I., Abley, M., Gebreyes, W., LeJeune, J. T., & Rajashekara, G. (2011). Genotypic and phenotypic properties of cattle-associated *Campylobacter* and their implications to public health in the USA. *PloS One*, 6(10), e25778.

- Sato, K., Bartlett, P. C., Kaneene, J. B., & Downes, F. P. (2004). Comparison of Prevalence and Antimicrobial Susceptibilities of *Campylobacter* spp. Isolates from Organic and Conventional Dairy Herds in Wisconsin. *Applied and Environmental Microbiology*, 70(3), 1442–1447.
- Scates, P., Moran, L., & Madden, R. H. (2003). Effect of Incubation Temperature on Isolation of *Campylobacter jejuni* Genotypes from Foodstuffs Enriched in Preston Broth. *Applied and Environmental Microbiology*, 69(8), 4658–4661.
- Schildt, M., Savolainen, S., & Hänninen, M.L. (2006). Long-lasting *Campylobacter jejuni* contamination of milk associated with gastrointestinal illness in a farming family. *Epidemiology and Infection*, 134(2), 401–5.
- Schmidt, T., Venter, E. H., & Picard, J. A. (2010). Evaluation of PCR assays for the detection of *Campylobacter fetus* in bovine preputial scrapings and the identification of subspecies in South African field isolates. *Journal of the South African Veterinary Association*, 81(2), 87–92.
- Scotter, S. L., Humphrey, T. J., & Henley, A. (1993). Methods for the detection of thermotolerant *Campylobacter* in foods: results of an inter-laboratory study. *Journal of Applied Bacteriology*, 74(2), 155–163.
- Senok, A., Yousif, A., Mazi, W., Sharaf, E., Bindayna, K., Elnima, E.A., & Botta, G. (2007). Pattern of antibiotic susceptibility in *Campylobacter jejuni* isolates of human and poultry origin. *Japanese Journal of Infectious Diseases*, 60(1), 1-4.
- Serichantalergs, O., Jensen, L. B., Pitarangsi, C., Mason C. J., & Dalsgaard, A. (2007). A possible mechanism of macrolide resistance among multiple resistant *Campylobacter jejuni* and *Campylobacter coli* isolated from Thai children during 1991–2000. *Southeast Asian Journal of Tropical Medicine and Public Health*, 38(3), 501–506.
- Shane, S. M. (1992). The significance of *Campylobacter jejuni* infection in poultry: A review. *Avian Pathology*, 21(2), 189–213.
- Shin, E., & Lee, Y. (2007). Antimicrobial resistance of 114 porcine isolates of *Campylobacter coli*. *International Journal of Food Microbiology*, 118(2), 223–7.
- Silva, J., Leite, D., Fernandes, M., Mena, C., Gibbs, P. A., & Teixeira, P. (2011). *Campylobacter* spp. as a Foodborne Pathogen: A Review. *Frontiers in Microbiology*, 2(September), 200.
- Skirrow, M. B. (1991). Epidemiology of *Campylobacter* enteritis. *International Journal of Food Microbiology*, 12(1), 9-16.
- Skirrow, M. B. (1994). Diseases due to *Campylobacter*, *Helicobacter* and related bacteria. *Journal of Comparative Pathology*, 111(2), 113-149.

- Skirrow, M. B. (2006). John McFadyean and the centenary of the first isolation of *Campylobacter* species. *Clinical Infectious Diseases*, 43(9), 1213–7.
- Skov, M. N., Spencer, A. G., Hald, B., Petersen, L., Nauerby, B., Carstensen, B., & Madsen, M. (2004). The role of litter beetles as potential reservoir for *Salmonella enterica* and thermophilic *Campylobacter* spp. between broiler flocks. *Avian Diseases*, 48(1), 9–18.
- Smith, K. E., Besser, J. M., Hedberg, C. W., Leano, F. T., Bender, J. B., Wicklund, J. H., & Osterholm, M. T. (1999). Quinolone-resistant *Campylobacter jejuni* infections in Minnesota, 1992–1998. *New England Journal of Medicine*, 340(20), 1525–1532.
- Snelling, W. J., Matsuda, M., Moore, J. E., & Dooley, J. S. G. (2005). *Campylobacter jejuni*. *Letters in Applied Microbiology*, 41(4), 297–302.
- Solomon, E. B., & Hoover, D. G. (1999). *Campylobacter jejuni*: a bacterial paradox. *Journal of Food Safety*, 19(2), 121–136.
- Stanley, K. N., Wallace, J. S., Currie, J. E., Diggle, P. J., & Jones, K. (1998a). The seasonal variation of thermophilic *Campylobacter* in beef cattle, dairy cattle and calves. *Journal of Applied Microbiology*, 85(3), 472–80.
- Stanley, K., Cunningham, R., & Jones, K. (1998b). Isolation of *Campylobacter jejuni* from groundwater. *Journal of Applied Microbiology*, 85(1), 187–91.
- Stanley, K., & Jones, K. (2003). Cattle and sheep farms as reservoirs of *Campylobacter*. *Journal of Applied Microbiology*, 94 Suppl(Skirrow 1977), 104S–113S.
- Steele, M. L., McNab, W. B., Poppe, C., Griffiths, M. W., Chen, S., Degrandis, S. A., & Odumeru, J. A. (1997). Survey of Ontario bulk tank raw milk for food-borne pathogens. *Journal of Food Protection*, 60(11), 1341–1346.
- Steinbrueckner, B., Haerter, G., Pelz, K., & Kist, M. (1999). Routine identification of *Campylobacter jejuni* and *Campylobacter coli* from human stool samples. *FEMS Microbiology Letters*, 179, 227–232.
- Stucki, U. R. S., Frey, J., Nicolet, J., & Burnens, A. P. (1995). Identification of *Campylobacter jejuni* on the basis of a species-specific gene that encodes a membrane protein. *Journal of Clinical Microbiology*, 33(4), 855–859.
- Studahl, A., & Andersson, Y. (2000). Risk factors for indigenous *Campylobacter* infection: a Swedish case-control study. *Epidemiology and Infection*, 125(2), 269–75.
- Tambur, Z., Stojanov, I., Konstantinovic, S., Jovanovic, D., Cenic-Milosevic, D., & Opacic, D. (2010). Multi drug resistance of *Campylobacter jejuni* and *Campylobacter coli* to tested antibiotics in strains originating from humans, poultry and swine. *Zbornik Matice srpske za prirodne nauke*, (118), 27–35.

- Tan, Y.F, Haresh, K.K., Chai, L.C., Ghazali, F.M., & Son, R. (2008). Prevalence of *Campylobacter* spp . in retailed ready-to-eat sushi. *International Food Research Journal*, 15(3), 1–6.
- Taremi, M., Mehdi Soltan Dallal, M., Gachkar, L., MoezArdalan, S., Zolfagharian, K., & Reza Zali, M. (2006). Prevalence and antimicrobial resistance of *Campylobacter* isolated from retail raw chicken and beef meat, Tehran, Iran. *International Journal of Food Microbiology*, 108(3), 401–3.
- Teuber, M. (2001). Veterinary use and antibiotic resistance. *Current Opinion in Microbiology*, 4(5), 493–9.
- Uaboi-Egbenni, P. O., Bessong, P. O., Samie, A., & Obi, C. L. (2012). Potentially pathogenic *Campylobacter* species among farm animals in rural areas of Limpopo province, South Africa: A case study of chickens and cattles. *African Journal of Microbiology Research*, 6(12), 2835–2843.
- Vandamme, P. (2000). "Taxonomy of the family *Campylobacteraceae*," in *Campylobacter*, I. Namchamkin & M.J. Blaser (Washington, DC: ASM), 3-27.
- Vandamme, P., Debruyne, L., De Brandt, E., & Falsen, E. (2010). Reclassification of *Bacteroides ureolyticus* as *Campylobacter ureolyticus* comb. nov., and emended description of the genus *Campylobacter*. *International Journal of Systematic and Evolutionary Microbiology*, 60(Pt 9), 2016–22.
- Van de Giessen, A. W., Bloemberg, B. P., Ritmeester, W. S., & Tilburg, J. J. (1996). Epidemiological study on risk factors and risk reducing measures for *Campylobacter* infections in Dutch broiler flocks. *Epidemiology and Infection*, 117(2), 245–50.
- Van den Bogaard, A. E., & Stobberingh, E. E. (2000). Epidemiology of resistance to antibiotics. Links between animals and humans. *International Journal of Antimicrobial Agents*, 14(4), 327–35.
- Van Vliet, A. H., & Ketley, J. M. (2001). Pathogenesis of enteric *Campylobacter* infection. *Symposium Series (Society for Applied Microbiology)*, 90, 45S–56S.
- Velayudhan, J., & Kelly, D. J. (2002). Analysis of gluconeogenic and anaplerotic enzymes in *Campylobacter jejuni*: an essential role for phosphoenolpyruvate carboxykinase. *Microbiology (Reading, England)*, 148(Pt 3), 685–94.
- Vereen, E., Lowrance, R. R., Cole, D. J., & Lipp, E. K. (2007). Distribution and ecology of *Campylobacter* in coastal plain streams (Georgia, United States of America). *Applied and Environmental Microbiology*, 73(5), 1395–403.
- Vilardo, M. D. C. B., Thomé, J. D. D. S., Esteves, W. T. C., Filgueiras, A. L. L., & De Oliveira, S. S. (2006). Application of biochemical and polymerase chain reaction assays for identification of *Campylobacter* isolates from non-human primates. *Memórias do Instituto Oswaldo Cruz*, 101(5), 499–501.

- Wainø, M., Bang, D. D., Lund, M., Nordentoft, S., Andersen, J. S., Pedersen, K., & Madsen, M. (2003). Identification of campylobacteria isolated from Danish broilers by phenotypic tests and species-specific PCR assays. *Journal of Applied Microbiology*, 95(4), 649-655.
- Wang, G., Clark, C. G., Taylor, T. M., Pucknell, C., Barton, C., Price, L., & Rodgers, F. G. (2002). Colony multiplex PCR assay for identification and differentiation of *Campylobacter jejuni*, *C. coli*, *C. lari*, *C. upsaliensis*, and *C. fetus* subsp. *fetus*. *Journal of Clinical Microbiology*, 40(12), 4744-4747.
- Wassenaar, T. M. (2000). MINIREVIEW Genotyping of *Campylobacter* spp. *Applied and Environmental Microbiology*, 66(1), 1-9.
- Wegmüller, B., Lüthy, J., & Candrian, U. (1993). Direct polymerase chain reaction detection of *Campylobacter jejuni* and *Campylobacter coli* in raw milk and dairy products. *Applied and Environmental Microbiology*, 59(7), 2161-2165.
- Werno, A. M., Klena, J. D., Shaw, G. M., & Murdoch, D. R. (2002). Fatal Case of *Campylobacter lari* Prosthetic Joint Infection and Bacteremia in an Immunocompetent Patient. *Journal of Clinical Microbiology*, 40(3), 1053-1055.
- Wesley, I. V, Wells, S. J., Harmon, K. M., Green, a, Schroeder-Tucker, L., Glover, M., & Siddique, I. (2000). Fecal shedding of *Campylobacter* and *Arcobacter* spp. in dairy cattle. *Applied and Environmental Microbiology*, 66(5), 1994-2000.
- Whyte, P., McGill, K., Cowley, D., Madden, R. H., Moran, L., Scates, P., & Cormican, M. (2004). Occurrence of *Campylobacter* in retail foods in Ireland. *International Journal of Food Microbiology*, 95(2), 111-8.
- Wieczorek, K., Szewczyk, R., & Osek, J. (2012). Prevalence , antimicrobial resistance , and molecular characterization of *Campylobacter jejuni* and *C. coli* isolated from retail raw meat in Poland. *Veterinarni Medicina*, 57(6), 293-299.
- Williams, L. K., Jørgensen, F., Grogono-Thomas, R., & Humphrey, T. J. (2009). Enrichment culture for the isolation of *Campylobacter* spp: Effects of incubation conditions and the inclusion of blood in selective broths. *International Journal of Food Microbiology*, 130(2), 131-4.
- Willis, W. L., & Murray, C. (1997). *Campylobacter jejuni* seasonal recovery observations of retail market broilers. *Poultry Science*, 76(2), 314-7.
- Wilson, D. J., Gabriel, E., Leatherbarrow, A. J., Cheesbrough, J., Gee, S., Bolton, E., & Diggle, P. J. (2008). Tracing the source of campylobacteriosis. *PLoS Genetics*, 4(9), e1000203.
- Wong, T. L., Hollis, L., Cornelius, A., Nicol, C., Cook, R., & Hudson, J. A. (2007). Prevalence, numbers, and subtypes of *Campylobacter jejuni* and *C. coli* in uncooked retail meat samples. *Journal of Food Protection*, 70(3), 566-573.

- Wysok, B., Wiszniewska-Łaszczych, A., Uradziński, J., & Sztejn, J. (2011). Prevalence and antimicrobial resistance of *Campylobacter* in raw milk in the selected areas of Poland. *Polish Journal of Veterinary Sciences*, 14(3), 473-477.
- Yamazaki-Matsune, W., Taguchi, M., Seto, K., Kawahara, R., Kawatsu, K., Kumeda, Y., & Tsukamoto, T. (2007). Development of a multiplex PCR assay for identification of *Campylobacter coli*, *C. fetus*, *C. hyointestinalis* subsp. *hyointestinalis*, *C. jejuni*, *C. lari* and *C. upsaliensis*. *Journal of Medical Microbiology*, 56(Pt 11), 1467-73.
- Yang, C., Jiang, Y., Huang, K., Zhu, C., & Yin, Y. (2003). Application of real-time PCR for quantitative detection of *Campylobacter jejuni* in poultry, milk and environmental water. *FEMS Immunology & Medical Microbiology*, 38(3), 265-271.
- Yu, J.H., Kim, N.Y., Cho, N.-G., Kim, J. H., Kang, Y. A., & Lee, H.G. (2010). Epidemiology of *Campylobacter jejuni* outbreak in a middle school in Incheon, Korea. *Journal of Korean Medical Science*, 25(11), 1595-600.
- Zanetti, F., Varoli, O., Stampi, S., & De Luca, G. (1996). Prevalence of thermophilic *Campylobacter* and *Arcobacter butzleri* in food of animal origin. *International Journal of Food Microbiology*, 33(2-3), 315-21.
- Zanoni, R. G., Debruyne, L., Rossi, M., Revez, J., & Vandamme, P. (2009). *Campylobacter cuniculorum* sp. nov., from rabbits. *International Journal of Systematic and Evolutionary Microbiology*, 59(7), 1666-1671.
- Zhao, C., Ge, B., De Villena, J., Sudler, R., Yeh, E., Zhao, S., & Meng, J. (2001). Prevalence of *Campylobacter* spp., *Escherichia coli*, and *Salmonella* serovars in retail chicken, turkey, pork, and beef from the Greater Washington, DC, area. *Applied and Environmental Microbiology*, 67(12), 5431-5436.
- Zhao, S., Young, S. R., Tong, E., Abbott, J. W., Womack, N., Friedman, S. L., & McDermott, P. F. (2010). Antimicrobial resistance of *Campylobacter* isolates from retail meat in the United States between 2002 and 2007. *Applied and Environmental Microbiology*, 76(24), 7949-56.
- Zilbauer, M., Dorrell, N., Wren, B. W., & Bajaj-Elliott, M. (2008). *Campylobacter jejuni*-mediated disease pathogenesis: an update. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 102(2), 123-9.
- Zorman, T., Heyndrickx, M., Uzunović-Kamberović, S., & Smole Mozina, S. (2006). Genotyping of *Campylobacter coli* and *C. jejuni* from retail chicken meat and humans with campylobacteriosis in Slovenia and Bosnia and Herzegovina. *International Journal of Food Microbiology*, 110(1), 24-33.