

**MONITORING SPATIAL DISTRIBUTION OF NONPATHOGENIC  
*FUSARIUM OXYSPORUM* IN THE RHIZOSPHERE AND ROOTS OF  
BANANA USING  $\beta$ -D-GLUCURONIDASE AND GREEN FLUORESCENT  
PROTEIN**

**By**

**WONG PUI SEE**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfilment of the Requirements for the Degree of Master of Science**

**March 2005**

## **DEDICATION**

I dedicate this study to my parents, sisters and brother for their love and  
affection.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Master of Science

**MONITORING SPATIAL DISTRIBUTION OF NONPATHOGENIC  
*FUSARIUM OXYSPORUM* IN THE RHIZOSPHERE AND ROOTS OF  
BANANA USING  $\beta$ -D-GLUCURONIDASE AND GREEN FLUORESCENT  
PROTEIN**

By

**WONG PUI SEE**

**March 2005**

**Chairman: Professor Sariah Meon, PhD**

**Faculty: Agriculture**

This study was conducted to monitor the spatial distribution of nonpathogenic isolate of *Fusarium oxysporum* (Fo4), using a detectable marker. Fo4 was transformed by GUS gene fusion system (*Escherichia coli*  $\beta$ -D-glucuronidase gene) and GFP (Green Fluorescent Proteins). The GUS was detected by 5-bromo-4-chloro-3-indolyl- $\beta$ -D-glucuronide (X-Gluc). The GFP activity was detected under a fluorescence microscope. There was no different in the cultural and morphological characteristics between the transformed and non-transformed Fo4. The antagonistic activity against *Fusarium oxysporum* f. sp. *cubense* Race 4 (FocR4) was maintained at values within the range of 56.98–41.56% based on the Percentage Inhibition of Radial Growth (PIRG) *in vitro*. DNA polymorphism (RAPD) screening by the primers OPC-15 and OPC-11, confirmed that no mutation had occurred after the transformation and, band GUS-F vs. GUS-R and 5-GFP vs. 3-GFP showed that transformed Fo4 still carried the GUS and GFP gene after five consecutive conidiation/generation cycles in culture.

The frequency of re-isolation on PCNB medium amended with 40 µg/ml X-Gluc and 200 µg/ml hygromycin B was highest ( $45.48 \times 10^3$  cfu/g roots) in the rhizosphere of banana plantlet at day 24, and remained to stabilize at between  $37.24 \times 10^3$  cfu/g roots to  $34.52 \times 10^3$  cfu/g roots until the end of sampling at day 36. The transformed Fo4 can be detected inside the root tissues even 4 days after inoculation although the colony forming units (cfu) (1 cfu/g roots) was substantially lower than that detected on the root surface ( $11.76 \times 10^3$  cfu/g roots), suggesting that they were colonizing the root and living on the root exudates. The population of transformed Fo4 in the non-rhizosphere soil initially fell sharply with the time of sampling, but thereafter remained stable ( $4.38-18.00 \times 10^4$  cfu/g soil) until 36 days of sampling.

Effect of transformed and non-transformed Fo4 on plant growth and development of *Fusarium* wilt was conducted using 10 week-old tissue cultured nursery banana seedlings cv. *Berangan*. The failure to produce *Fusarium* wilt symptom confirmed this strain was nonpathogenic to banana seedlings. The seedlings inoculated with FocR4 were pathogenic to cv. *Berangan* with 100% disease severity. However, the seedlings which had been inoculated with either transformed or non-transformed Fo4 before challenged with FocR4 did not show significant differences ( $P>0.05$ ) in the disease severity, confirmed the stability of the transformed GUS activity and GFP. This study suggests the possibility of using the GUS and GFP genes as visible detectable reporter genes for direct monitoring of the spatial distribution of a potential antagonist in the rhizosphere and roots of banana.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PEMANTAUAN TABURAN SPATIAL *FUSARIUM OXYSPORUM* BUKAN PATOGEN DALAM RIZOSFERA DAN AKAR PISANG MENGGUNAKAN  $\beta$ -D-GLUKURONIDAS DAN PROTEIN FLUORESCENS HIJAU**

Oleh

**WONG PUI SEE**

**Mac 2005**

**Pengerusi: Profesor Sariah Meon, PhD**

**Fakulti: Pertanian**

Kajian ini dijalankan untuk memantau taburan *Fusarium oxysporum* (Fo4) bukan patogen menggunakan gen GUS (*Escherichia coli*  $\beta$ -D-glukuronidas gen) dan GFP (protein fluorescens hijau). GUS dikesan menggunakan 5-bromo-4-kloro-3-indolyl- $\beta$ -D-glukuronida (X-Gluc). Aktiviti GFP dikesan di bawah mikroskop fluorescens. Tiada perbezaan dari segi ciri-ciri kultur dan morfologi di antara Fo4 transforman dan bukan transforman. Aktiviti antagonistic terhadap *Fusarium oxysporum* f. sp. *cubense* Race 4 (FocR4) dikekalkan pada nilai antara 56.98-41.56% berdasarkan Peratusan Perencatan Pertumbuhan Radial (PIRG) secara *in vitro*. Analisis polimorfisme DNA (RAPD) menggunakan primer OPC-15 dan OPC-11 membuktikan tiada mutasi berlaku selepas transformasi, dan jalur GUS-F lawan GUS-R and 5-GFP lawan 3-GFP menunjukkan Fo4 transforman masih membawa gen GUS and GFP sepanjang lima kitaran generasi dalam kultur.

Kekerapan pemencilan semula pada medium PCNB yang mengandungi 40  $\mu\text{g}/\text{ml}$  X-Gluc dan 200  $\mu\text{g}/\text{ml}$  hygromycin B adalah tertinggi ( $45.48 \times 10^3$  cfu/g

akar) di dalam rizosfera pada hari ke-24, dan seterusnya stabil antara  $37.24 \times 10^3$  cfu/g akar ke  $34.52 \times 10^3$  cfu/g akar sehingga akhir tempoh penyampelan pada hari ke-36. Fo4 transforman dikesan dalam tisu akar pada hari ke-4 (1 cfu/g akar) selepas penginokulatan sungguhpun unit pembentukan koloni (cfu) adalah rendah daripada yang dikesan pada rizosfera ( $11.76 \times 10^3$  cfu/g akar), menunjukkan Fo4 transforman boleh hidup pada eksudat akar. Populasi Fo4 transforman di dalam tanah pada mulanya menurun dengan mendadak mengikut tempoh penyampelan, tetapi kekal stabil selepas itu ( $4.38-18.00 \times 10^4$  cfu/g tanah) sehingga hari ke-36 penyampelan.

Kesan Fo4 transforman dan bukan transforman ke atas pertumbuhan dan pembentukan layu *Fusarium* diuji pada anak benih pisang cv. *Berangan* berumur 10 minggu. Tiada pembentukan gejala penyakit layu dicatatkan, membuktikan bahawa ia adalah tidak patogenik. Anak benih pisang cv. *Berangan* yang diinokulat dengan FocR4 mencatatkan 100% pembentukan keterukan penyakit layu *Fusarium*. Tetapi apabila anak pokok diinokulat dengan Fo4 transforman atau bukan transforman terlebih dahulu sebelum dilawan dengan FocR4, tiada perbezaan bererti ( $P>0.05$ ) dicatatkan dalam penilaian keterukan penyakit. Ini membuktikan kestabilan transforman Fo4 dan aktiviti GUS dan GFP. Kajian ini mencadangkan kemungkinan menggunakan gen GUS dan GFP sebagai gen pengesan yang nyata untuk pemantauan taburan spatial antagonis yang berpotensi dalam rizosfera dan akar pisang.

## **ACKNOWLEDGEMENTS**

I am thankful to my parents for giving me the strength and ability to complete this study. I also wish to express my sincerest gratitude and appreciation to Prof. Dr. Sariah Meon, Chairperson of my Supervisory Committee, and Associate Professor Dr. Siti Khalijah Daud and Associate Professor Dr. Nor' Aini Mohd Fadzillah, the other members of the Committee, for their guidance, understanding and invaluable advice proffered me throughout my research and preparation of this thesis.

Many thanks too to all the staff in the Pathology and Microbiology Laboratories; Mr. Johari, Mr. Shamsuddin, Mr. Nazri, Mr. Gani and Mr. Khir for their advice and kind assistance in the preparation of materials. Sincere appreciation is also extended to Adeline, Yasmeen, Fitri and Kevin for sharing and making these years memorable. I am indebted to Prof. Manaf for allowing me to use the fluorescence microscope.

I also appreciate very much the love, support and encouragement from my sisters and brother - Ching, Chit, Kwan and Choong. And also deep gratitude to my best friends, Kate, Hoi Ling, Pui Kit, Mark, Hooi Fang, Yee Chern, Tessa, Phaik Yoke and Tse Sun. For my dear Shi Kwai, thank you for always being there and coloring my life.

I certify that an Examination Committee has met on 30<sup>th</sup> March 2005 to conduct the final examination of Wong Pui See, on her Master of Science thesis entitled "Monitoring the Spatial Distribution of Nonpathogenic *Fusarium oxysporum* in the Rhizosphere and Roots of Banana Using β-D-glucuronidase and Green Fluorescent Protein" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

**Inon Sulaiman, PhD**  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Chairman)

**Zainal Abidin Mior Ahmad, PhD**  
Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Internal Examiner)

**Maheran Abd Aziz, PhD**  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Internal Examiner)

**Ibrahim Omar, PhD**  
Stesen MARDI Bukit Tangga  
(External Examiner)

---

**GULAM RUSUL RAHMAT ALI, PhD**  
Professor/Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirements for the degree of Master of Science. The members of the Supervisory Committee are as follows:

**Sariah Meon, PhD**  
Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Chairman)

**Siti Khalijah Daud, PhD**  
Associate Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Member)

**Nor' Aini Mohd Fadzillah, PhD**  
Associate Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Member)

---

**AINI IDERIS, PhD**  
Professor/Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## **DECLARATION**

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

---

**WONG PUI SEE**

Date:

## TABLE OF CONTENTS

	<b>Page</b>
<b>DEDICATION</b>	II
<b>ABSTRACT</b>	III
<b>ABSTRAK</b>	V
<b>ACKNOWLEDGEMENTS</b>	VII
<b>APPROVALS</b>	VIII
<b>DECLARATION</b>	X
<b>LIST OF TABLES</b>	XIII
<b>LIST OF FIGURES</b>	XIV
<b>LIST OF ABBREVIATIONS</b>	XVII
 <b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	1
<b>2 LITERATURE REVIEW</b>	6
2.1 Banana Plant	6
2.2 <i>Fusarium</i> spp.	7
2.3 <i>Fusarium oxysporum</i> f. sp. <i>cubense</i>	9
2.4 <i>Fusarium</i> Wilt of Banana	12
2.4.1 Behavior of Pathogen	12
2.4.2 Symptoms of <i>Fusarium</i> Wilt	13
2.5 Control of <i>Fusarium</i> Wilt of Banana	15
2.5.1 Cultural Control	16
2.5.2 Chemical Control	18
2.5.3 Biological Control	20
2.6 Transformation System	23
2.6.1 pCAMBIA Vector	23
2.6.2 <i>Escherichia coli</i>	24
2.6.3 <i>Agrobacterium tumefaciens</i>	25
2.6.4 GUS Gene Fusion System	26
2.6.4.1 GUS Transformation	29
2.6.5 Green Fluorescent Proteins	33
2.6.5.1 GFP Transformation	34
<b>3 MATERIALS AND METHODS</b>	
3.1 Antagonistic Activity of <i>Fusarium oxysporum</i> Isolates against <i>Fusarium oxysporum</i> f. sp. <i>cubense</i> Race 4	36
3.2 Transformation System	38
3.2.1 Mini-Preparation Procedure for <i>Escherichia coli</i> Plasmid	38
3.2.2 <i>Agrobacterium tumefaciens</i> -Mediated Transformation	40
3.2.3 Transformation of the Nonpathogenic Fo4	42
3.3 Assessing the Stability of the Transformed Fo4	43
3.3.1 Colony Assay of β-D-Glucuronidase Activity	43

3.3.2	Mitotic Stability Test	44
3.3.2.1	Cultural and Morphological Characteristics of the Transformed Fo4	44
3.3.2.2	Antagonistic Activity against FocR4	45
3.3.3	DNA Polymorphism of the Transformed and Non-transformed Fo4	46
3.4	Colonization and Establishment of Transformed Fo4 in the Rhizosphere and Roots of Banana Plantlets and Seedlings	49
3.5	Effect of Transformed and Non-Transformed Fo4 on Plant Growth and Development of <i>Fusarium</i> Wilt	52
<b>4</b>	<b>RESULTS</b>	
4.1	Antagonistic Activity of <i>Fusarium oxysporum</i> Isolates against <i>Fusarium oxysporum</i> f. sp. <i>cubense</i> Race 4	55
4.2	Transformation of Nonpathogenic Fo4	61
4.3	Assessing the Stability of the Transformed Fo4	62
4.4	Colonization and Establishment of Transformed Fo4 in the Rhizosphere and Roots of Banana Plantlets and Seedlings	69
4.5	Effect of Transformed and Non-Transformed Fo4 on Plant Growth and Development of <i>Fusarium</i> Wilt	78
<b>5</b>	<b>DISCUSSION</b>	90
<b>6</b>	<b>CONCLUSION</b>	104
<b>REFERENCES</b>		106
<b>APPENDICES</b>		A.1
<b>BIODATA OF THE AUTHOR</b>		B.1